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Direct observations of diel biological CO₂ fixation in the oceans

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Abstract

Much of the variability in the surface ocean's carbon cycle can be attributed to the availability of sunlight, through processes such as heat fluxes and photosynthesis, which regulate over a wide range of time scales. The critical processes occurring on timescales of a day or less, however, have undergone few investigations, and most of these have been limited to a time span of several days to months, or exceptionally, for longer periods. Optical methods have helped to infer short-term biological variability, however corresponding investigations of the oceanic CO₂ system are lacking. We employ high-frequency CO₂ and optical observations covering the full seasonal cycle on the Scotian Shelf, Northwestern Atlantic Ocean, in order to unravel diel periodicity of the surface ocean carbon cycle and its effects on annual budgets. Significant diel periodicity occurs only if the water column is sufficiently stable as observed during seasonal warming. During that time biological CO₂ drawdown, or net community production (NCP), is delayed for several hours relative to the onset of photosynthetically available radiation (PAR), due to diel cycles in chlorophyll-*a* concentration and to grazing, both of which, we suggest, inhibit NCP in the early morning hours. In summer, NCP decreases by more than 90 %, coinciding with the seasonal minimum of the mixed layer depth and resulting in the disappearance of the diel CO₂ periodicity in the surface waters.

1 Introduction

Shelf and marginal seas play a crucial role in the global carbon cycle as they link its different compartments (land, ocean, atmosphere) (Ciais et al., 2008; Thomas et al., 2008; Chen and Borges, 2009). As a consequence of their role as an integral link between the compartments, the spatial and temporal variability of the carbon cycle in marginal seas is generally higher than in open ocean environments (Thomas and Schneider, 1999; Schiettecatte, 2006; Chen and Borges, 2009; Omar et al., 2010). In

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addition to natural drivers, anthropogenic processes perturbing natural cycles in each of the carbon cycle compartments affect variability in shelf and marginal seas. Examples of such perturbations include eutrophication, ocean acidification and atmospheric nitrogen deposition (Doney et al., 2007; Thomas et al., 2009; Borges and Gypens, 2010; Cai et al., 2011).

Physical, biological and chemical processes govern the variability of the carbon cycle and the air-sea exchange of CO₂. Although these processes are evident at many temporal and spatial scales, they interact at high frequency, local scales, eventually yielding diurnal, seasonal and longer-term periodicity. Our understanding of the monthly to seasonal variability in the carbon cycle in several shelf and marginal seas has improved, in particular with respect to attaining full annual observational coverage (Chen and Borges, 2009; Omar et al., 2010; Thomas et al., 2004; Shadwick et al., 2010, 2011). At shorter time scales, optical methods have helped to infer short-term biological variability (Siegel et al., 1989; Stramska and Dickey, 1992; Cullen et al., 1992; Gernez et al., 2011; Dall'Olmo et al., 2011), however corresponding investigations of the oceanic CO₂ system are largely lacking. Only a few recent studies have mainly focused on time scales of several days to months (Hood et al., 2001; Bates et al., 2001; Copin-Montegut et al., 2004; Lefevre et al., 2008; Bozec et al., 2011), and investigations of processes at the rate of their occurrence in shelf and marginal seas are still sparse when it comes to full seasonal coverage (Vandemark et al., 2011).

The Scotian Shelf region is located at the Eastern Canadian continental shelf at the boundary between the subpolar and subtropical gyres. This region is thus influenced by water masses of arctic origin via the Labrador and Newfoundland Shelves, by low-salinity waters emanating from the Gulf of St. Lawrence, and by the Gulf Stream (Urrego-Blanco and Sheng, 2012). One of the dominant characteristics on the shelf is the large seasonal amplitude in sea surface temperature (SST) between subzero temperatures in winter to approximately 20°C during summer (Shadwick and Thomas, 2011) (Fig. 1). Recent studies have identified the region as a strong source for atmospheric CO₂ at the annual scale, with an intense, but brief period of CO₂ uptake during

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the spring bloom, which occurs at the annual SST minimum in late March to early April (Shadwick et al., 2011) (Fig. 1). Controls of the seasonal to interannual variability of the surface CO₂ system in the Scotian Shelf region have been inferred from satellite observations, and include the intensity of autumn and winter storms, winter nutrient levels, and the onset of post-winter water column stratification (Greenan et al., 2004, 2008; Shadwick et al., 2010). The processes controlling the variability of the carbon cycle at time scales shorter than the monthly to seasonal scale remain poorly understood, which is the case for most regions of the open oceans, as well as shelf and marginal seas. The study, presented here, sheds light on the role of high frequency processes in controlling the carbon cycle in the surface waters of the Scotian Shelf region over a complete annual cycle.

2 Material and methods

We employ observations from a CARIOCA buoy recording surface water $p\text{CO}_2$, SST, salinity, and complementary parameters at an hourly rate. The buoy has been deployed since April 2007. In order to avoid data gaps due to maintenance, we reconstruct an annual cycle using different years of observations. Detailed descriptions of the buoy location and the corresponding sampling activities have been given in Shadwick et al. (2011). In the present study, we focus on three key periods of the annual cycle: 1: winter (pre-spring bloom to spring bloom transition); 2: late spring to early summer, i.e., the warming period; and 3: the autumn to winter transition (Fig. 1). Diel cycles in $p\text{CO}_2$ with hourly resolution were established by removing the 48-point moving average from the (hourly) observations. In order to investigate the crucial period between spring and summer, when the waters are warming (Fig. 1), we combined the CARIOCA data with observations from the SeaHorse, an autonomous profiler (Greenan et al., 2004, 2008), which records water column profiles of temperature, salinity, photosynthetically available radiation (PAR), and downwelling irradiance ($E_d(\lambda)$). Profiles were recorded with approximately 0.5 m vertical resolution approximately every 2 h between 4 April and 27

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July 2007. In an attempt to resolve the contribution of phytoplankton to NCP during the warming period, we derived chlorophyll-*a* concentration from profiles of Seahorse $E_d(\lambda)$ using the model of Nahorniak et al. (2001). This required calculation of the attenuation of E_d at three wavelengths over a depth interval (5–6 m) that was similar to the depths at which other CARIOCA parameters were measured. Since the sensor wavelengths did not exactly match the wavelengths required by the model, $E_d(\lambda)$ was interpolated to model wavelengths (412, 443, 555 nm). Modelled chlorophyll-*a* (Chl_{mod} ; mg m^{-3}) was then calculated using Nahorniak et al. (2001) formulations that include spectral coefficients to describe the absorption properties of water, phytoplankton and coloured dissolved organic matter and assume a value of 0.8 for the average cosine of downwelling light, μ_d (dimensionless). Chl_{mod} values were compared with discrete, fluorometrically determined chlorophyll-*a* values obtained at the same time and depth from ship-based water samples; R^2 and root mean square error (RMSE) values of 0.89 and 0.83 mg m^{-3} ($N = 8$), respectively, were obtained. These values were then averaged into time bins over the 40-day period to give climatological values for Chl_{mod} every two hours during the hours of daylight during which reliable $E_d(\lambda)$ measurements could be made, which, at this time of year and latitude, corresponded to times between $\sim 06:00\text{h}–16:00\text{LT}$.

3 Results and discussion

The spectral analysis of the buoy data (Fig. 2a,b) revealed a 24-h periodicity for the parameters $p\text{CO}_2$ and SST that occurs only when the waters are warming, i.e., between April and late August. $p\text{CO}_2$ and SST also show significant coherent patterns during this time of the year (Fig. 2b, bottom panel) and are the focus of this paper. Outside of this period, periodicity and significant coherence were only observed, if detectable, at longer time scales (several days), which mirrors the typical frequency of winter storms in the region (Fig. 2c). Other parameters such as salinity do not show significant 24-h periodicity, which means that tidal, lateral and other effects are either not identifiable, or

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act on timescales longer than 24 h. Such processes would be captured by the 48-point moving average, and do not influence the diel cycles.

Surface heat fluxes cause variability in SST at diel and seasonal time scales (Umoh and Thompson, 1994), which in turn drive some of the observed variability of the $p\text{CO}_2$, primarily because of the temperature dependence of the Henry constant. We corrected the observed $p\text{CO}_2$ data ($p\text{CO}_{2,\text{obs}}$) to a daily mean temperature to give $p\text{CO}_{2,\text{temp}}$. The difference between $p\text{CO}_{2,\text{obs}}$ and $p\text{CO}_{2,\text{temp}}$ yielded $p\text{CO}_2$ data that are governed by processes other than temperature within a 24-h period. Since we did not detect processes other than SST variability acting on the 24 h period, the remaining $p\text{CO}_2$ variability can be ascribed to biological activity ($p\text{CO}_{2,\text{bio}}$). The daily and diel variability of the $p\text{CO}_{2,\text{obs}}$ and $p\text{CO}_{2,\text{bio}}$ for three periods of the year is shown in Fig. 3. In the winter period (days 80–95, Fig. 3a), the amplitude of the diel oscillation was small and the $p\text{CO}_2$ relatively constant. With the onset of the spring bloom, at approximately day 90, the diel amplitude drastically increased (Fig. 3a). Throughout both of these periods, $p\text{CO}_{2,\text{obs}}$ and $p\text{CO}_{2,\text{bio}}$ were in phase and tracked each other closely indicating that temperature was not the main driver for the short-term variability during this time of the year. Similarly, in the autumn to winter (Fig. 3b) transition $p\text{CO}_{2,\text{obs}}$ and $p\text{CO}_{2,\text{bio}}$ again revealed in-phase patterns, though with a higher amplitude in autumn (days 305–345) than in winter (days 80–95), which can be ascribed to deepening of the mixed layer and an intrusion of high $p\text{CO}_2$ subsurface waters into the surface mixed layer (Shadwick et al., 2011). Between the days 160–200 (end of June until end of July), i.e., later in the season of surface water warming, the amplitude of the diel oscillation was reduced compared to that observed during the spring bloom (Fig. 3a–c). More importantly, a phase shift was detectable between $p\text{CO}_{2,\text{obs}}$ and $p\text{CO}_{2,\text{bio}}$, with the latter occurring approximately 3 h earlier than the $p\text{CO}_{2,\text{obs}}$ (Fig. 3d). We postulate that the cause for this phase shift might be diel cycles in biological activity. Such diel cycles were only visible, when the water column was sufficiently stable such as during thermal stratification (Fig. 1b; Shadwick et al. (2011), their Fig. 9).

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We can consider the diel cycle of $p\text{CO}_2$ as a composite of a temperature driven component, which follows the diel cycle of SST (Fig. 3d), and a biologically driven component, controlled by the balance of production and respiration of organic matter. The diel biological cycle obtained here can be considered a net biological signal, which shows an increase of the $p\text{CO}_{2,\text{bio}}$, i.e., net respiration, beginning in the evening (approx. 21:00 h), and ending in the morning hours (10:00h–11:00 h), when net community production (NCP) begins to exceed community respiration. NCP, indicated by a negative gradient in the $p\text{CO}_2$ anomaly (Fig. 3d), dominates the system until dusk. Subtracting the respiration signal, computed from the slope of the $p\text{CO}_{2,\text{bio}}$ during nighttime conditions, allows us to estimate the diurnal cycle of gross primary production (GPP). The corresponding respiration rate, assumed to be constant throughout the day, is estimated to be $0.05 \mu\text{mol C (l h)}^{-1}$; the rates of NCP and GPP are $0.26 \mu\text{mol C (l h)}^{-1}$ and $0.31 \mu\text{mol C (l h)}^{-1}$, respectively, both lasting approximately 10 h per day.

The onset of the net photosynthetic CO_2 drawdown occurs approximately 4–5 h after sunrise, near the period maximum of PAR (Fig. 3d) – a phenomenon that has been documented frequently but surprisingly, has rarely been discussed or addressed in detail. Stramska and Dickey (1992), Siegel et al. (1989) or Gernez et al. (2011) report a phase shift of several hours between the onset of PAR and maximum values of either the beam attenuation coefficient (c_p) or dissolved oxygen (O_2) concentrations, the latter two mirroring the build up of photosynthetic organic matter and revealing phasing throughout the diurnal cycle, which is comparable to the phasing of $p\text{CO}_{2,\text{bio}}$. Bozec et al. (2011, their Fig. 5) also show a similar phase shift between dissolved O_2 and $p\text{CO}_2$, relative to PAR, however, these authors conclude that $p\text{CO}_2$ and PAR are out of phase by 180 degrees and O_2 is in phase with PAR. In summary, it appears that there is sufficient evidence in the literature to suggest that the signal of net biological carbon fixation in the water column, as revealed by $p\text{CO}_2$, O_2 or beam attenuation measurements, is detectable at or around peak PAR, which is the ultimate energy source for the biological carbon fixation.

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In order to determine whether this reported periodicity in metrics of phytoplankton photosynthetic activity occurred at our study site, we used the model of Nahorniak et al. (2001) to derive estimates of chlorophyll-*a* concentration (Chl_{mod} ; mg m^{-3}) every two hours during daylight from SeaHorse profiler measurements of multispectral downwelling irradiance, $E_d(\lambda)$. When averaged over the day 160–200 period, a diel cycle in Chl_{mod} was revealed with a difference of $\sim 30\%$ between minimum and maximum values, and where the lowest values corresponded to low PAR levels during early morning and late afternoon, and maximum values to peak PAR at \sim midday (Fig. 3d). When compared with the $p\text{CO}_2$ diel cycles, it was observed that onset of net CO_2 drawdown, indicated by the change in gradient of $p\text{CO}_{2,\text{bio}}$ from positive to negative, coincided with the time of the maximum Chl_{mod} values. In other words, our data suggest that a threshold Chl_{mod} must first be attained before the system achieves net CO_2 drawdown. Others have also reported diel signals in either *Chl-a* or photosynthetic parameters (Cullen et al., 1992; Bruyant et al., 2005), with peak values around midday. It should also be noted that photoinhibition may depress photosynthetic rate at high irradiances, especially near midday, potentially further modulating the exact occurrence of net CO_2 drawdown with respect to PAR levels.

Both our results and those reported in the literature are consistent with the notion of a phase shift between the onset of PAR at daybreak and of net carbon fixation at approximate solar noon. Meso- and microzooplankton, which are active during nighttime, might still exert grazing pressure on phytoplankton during the early morning hours and thereby keep the abundance of phytoplankton low during the initial hours of daylight (Siegel et al., 1989). In addition to grazing effects, the diurnal variability in phytoplankton photosynthetic activity itself also contributes to delaying net carbon fixation until PAR has reached approximately maximum values as discussed above. Furthermore, in order to detect net carbon fixation as a change in the $p\text{CO}_2$ and O_2 concentrations in the water column, primary producers have to compensate for the respiratory activity of the heterotrophs, which in turn might exhibit relatively constant activity rates throughout the diel cycle (Cullen et al., 1992; Gernez et al., 2011). Based on our analysis,

we suggest that grazing and the dependence of the photosynthetic rate on PAR would cause the ecosystem to become autotrophic just before or at maximum PAR values (Fig. 3d). In our study, we were not able to identify any diurnal variability in the water column structure, i.e., in strength and depth of stratification, which might have provoked the phase shift between PAR and $p\text{CO}_{2,\text{bio}}$.

We have obtained the seasonal dynamics of NCP integrating the hourly $p\text{CO}_{2,\text{bio}}$ values (Fig. 4.). The maximum value of NCP is 3.4 mol C m^{-2} , or $271 \mu\text{mol C l}^{-1}$. Of particular interest is the observation that the carbon fixation rate, i.e., the DIC uptake rate by phytoplankton, is fairly constant from the onset of the spring bloom (approx. day 100) to approximately day 180, with DIC uptake rates of approximately $2.5 \mu\text{mol C (ld)}^{-1}$, corresponding to a NCP rate of $0.26 \mu\text{mol C (lh)}^{-1}$, assuming a 10-h photoperiod per day (Fig. 3d). The production rate decreases to about $0.1 \mu\text{mol C (ld)}^{-1}$ (Fig. 4), when the mixed layer depth reaches its minimum. Thereafter, a combination of dramatically reduced availability of nutrients and light inhibition make NCP almost vanish.

4 Conclusions

In summary, we observed a statistically significant diurnal periodicity of the CO_2 system only during the period, when the water is warming. The corresponding increase in water column stability facilitates the appearance of the diurnal cycle. The unraveled diurnal cycles of the surface $p\text{CO}_2$ reveal that net photosynthetic carbon fixation, i.e., NCP, begins approximately 4–5 h after onset of PAR. Grazing and transitional metabolic rates are likely the responsible processes for this phase shift, eventually allowing the ecosystem to be autotrophic for approximately 10 h out of 16 h of daylight.

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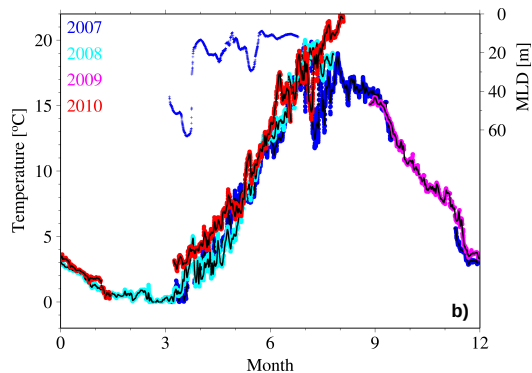
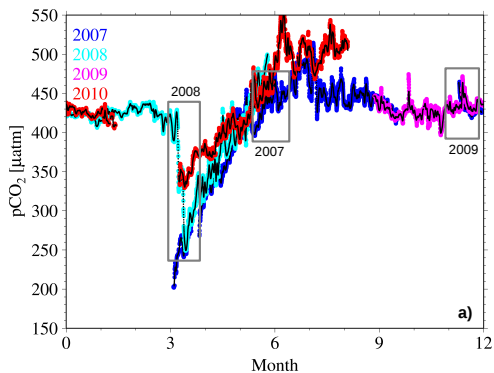


Fig. 1. Composite of available $p\text{CO}_2$ **(a)** and temperature data **(b)** for the years 2007–2010, recorded by the CARIOCA buoy. For the further study we evaluate data from June/July 2007, April 2008 and November/December 2009 as indicated by the gray boxes in a), assuming a climatological annual cycle. The black lines indicate the 48-point moving average. In **(b)**, the depth of the maximum of the Brunt-Väisälä frequency is shown as measure of the mixed layer depth (MLD), computed from the Seahorse data, deployed between April 2007 and July 2007.

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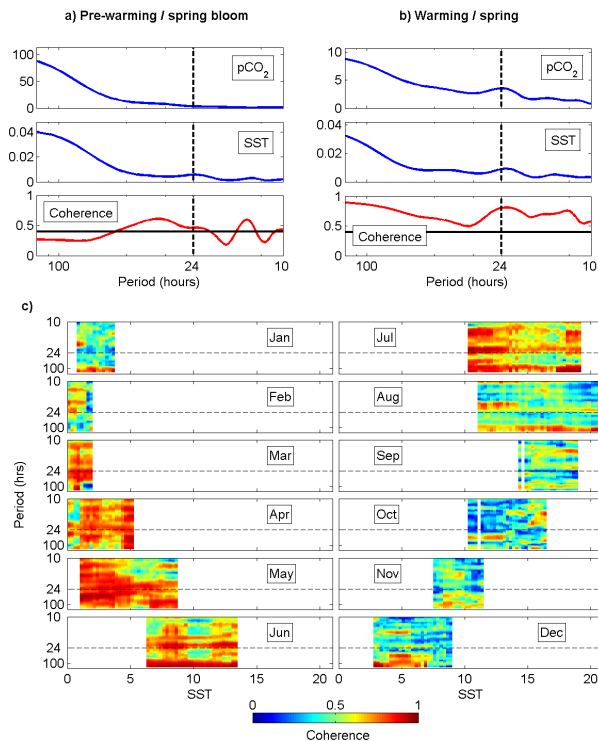


Fig. 2. Spectral analysis of $p\text{CO}_2$ and SST data from the CARIOCA buoy. Power spectra of $p\text{CO}_2$ and SST and their coherence are shown for two periods: **(a)** the pre-warming/spring bloom period and **(b)** the warming period (see Fig. 1a gray boxes). The dashed line on each plot indicates the 24 h period, and the horizontal solid line on the coherence plots shows the 5% significance level based on the degrees of freedom of the cross-spectral estimators. The coherence between $p\text{CO}_2$ and SST is plotted as a function of SST in monthly composites **(c)**, illustrating the temporal evolution of the coherence. The color scale indicates the level of coherence between $p\text{CO}_2$ and SST, and the dashed line shows the 24 h period. High coherence at the 24 h period occurs only during the period when the water is warming.

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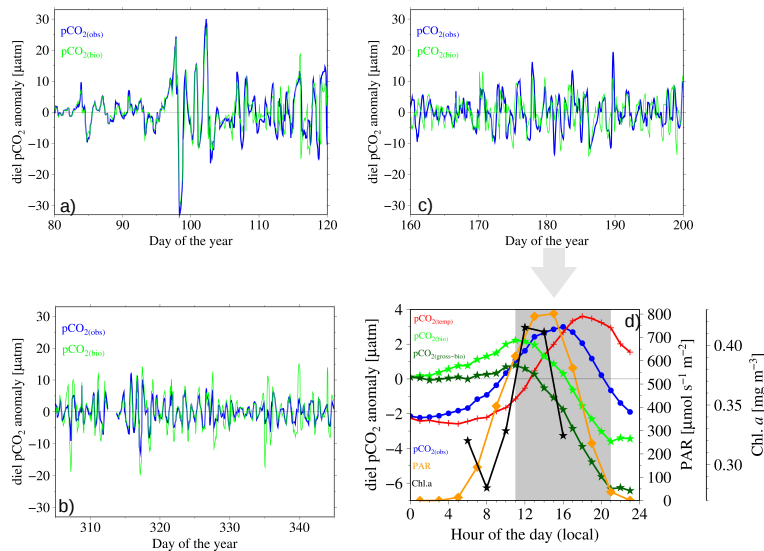


Fig. 3. Diel anomalies and daily cycles of observed $p\text{CO}_2$, biologically controlled $p\text{CO}_2$, PAR, and Chl-*a*. We show diel anomalies of $p\text{CO}_{2,\text{obs}}$ (blue) and $p\text{CO}_{2,\text{bio}}$ (green) for three periods of the year (a–c) as depicted in Fig. 1. Note the phase shift in the warming period (c,d), while outside the warming period (a,b) both parameters show the same timing (i.e., they are independent). Average diel anomalies or daily cycles compiled for the 40-day period from days 160–200 (see Fig. 1) observed $p\text{CO}_2$ ($p\text{CO}_{2,\text{obs}}$, blue), biologically controlled $p\text{CO}_2$ ($p\text{CO}_{2,\text{bio}}$, green), temperature-only controlled $p\text{CO}_2$ ($p\text{CO}_{2,\text{temp}}$, red), $p\text{CO}_{2,\text{bio}}$ corrected for respiratory activity ($p\text{CO}_{2,\text{gross-bio}}$, dark green) and photosynthetic active radiation (PAR, orange). Chlorophyll-*a* (chl-*a*, black) is shown for the hours (6.00 h–16.00 h), when reliable $E_d(\lambda)$ measurements can be made by the Seahorse. The gray-shaded box indicated the period, when net carbon fixation occurs. Note that $p\text{CO}_{2,\text{temp}}$ reflects the daily cycle of temperature. The respiration rate has been computed from the slope of the $p\text{CO}_{2,\text{bio}}$ during night time conditions. The production rates have been computed from the slopes of $p\text{CO}_{2,\text{bio}}$ and $p\text{CO}_{2,\text{gross-bio}}$ during daytime (gray-shaded area), respectively. The time scale reveals local time, i.e., GMT –3 h.

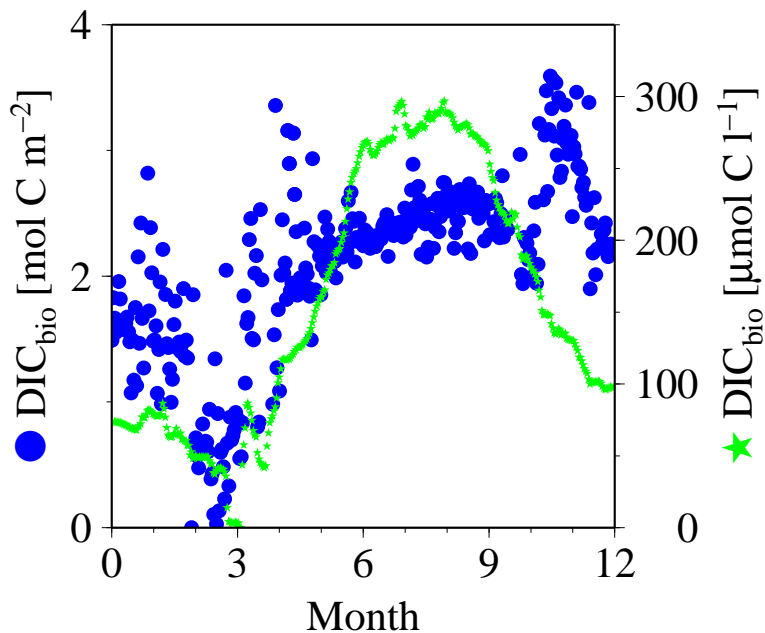


Fig. 4. Annual cycle of biological DIC uptake. We show the annual cycle of biological DIC uptake, integrated from hourly values, shown as mixed layer inventory (left y-axis, blue) and as concentration change (right y-axis, green). The production rate has been computed from the slope of the DIC uptake.

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