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Emissions of BVOC from Lodgepole Pine in response to Mountain Pine Beetle attack in high and low mortality forest stands

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Abstract

In this screening study biogenic volatile organic compound (BVOC) emissions from intact branches of lodgepole pine (*Pinus contorta*) trees were measured from trees at two forested sites that have been impacted differently by the mountain pine beetle (MPB) with one having higher mortality and the other with lower mortality. Differences in the amounts and chemical diversity of BVOC between the two sites and from apparently healthy trees versus trees in different stages of MPB attack are presented, as well as (for one site) observed seasonal variability in emissions. A brief site comparison is made of the hydrological characteristics and prior disturbances (both natural and man-made) at the sites. Trees sampled at the site experiencing high MPB-related tree mortality had lower chemodiversity in terms of monoterpene (MT) emission profiles, while profiles were more diverse at the lower-mortality site. Also at the higher-mortality site, MPB-infested trees in various stages of decline had lower emissions of sesquiterpenes (SQT) compared to healthy trees, while at the site with lower mortality, MPB-survivors had significantly higher SQT emissions during part of the growing season when compared to both uninfested and newly-infested trees. SQT profiles differed between the two sites, and, like monoterpene and oxygenated VOC profiles, varied through the season. For the low-mortality site in which repeated measurements were made over the course of the early summer-late fall, higher chemical diversity was observed in early compared to late-season measurements for all compound classes investigated (MT, oxygenated VOC, and SQT), with the amount of change appearing to correlate to the MPB status of the trees studied. Emissions of methyl-3-buten-2-ol had a distinct seasonal signal but were not much different between healthy or infested trees, except in trees with dead needles, from which emissions of this compound were negligible, and in late-season MPB survivors, in which they were higher than in newly-infested or uninfested trees. Emissions of SQT were significantly higher in the MPB survivors during both mid- and late-season sampling at the low-mortality site. The changes in emissions could have implications for regional air quality and climate through changes in ozone

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and aerosol distributions, although this study was designed as a preliminary screening effort and not enough individuals were sampled for all of the observed differences to be statistically demonstrated. Despite this, the compelling differences in emissions observed between the sites and individual trees with differing MPB-infestation statuses and the potential impacts these have on regional atmospheric chemistry argue for further research in this topic.

1 Introduction

The vast pine forests of Western North America have recently been the stage for an unprecedented epidemic of Mountain Pine Beetle (MPB; *Dendroctonus ponderosae*) infestation, with the latitudinal extent of the attack spanning from Canada to Mexico and extending westward from Nebraska to the Pacific coast. The reasons behind the magnitude and sweeping extent of the current epidemic are not fully known, although several factors may be at work. Years of drought in many parts of the afflicted areas have made trees less fit to ward off MPB attack (Allen et al., 2010; Breshears et al., 2005). Warmer temperatures experienced in recent decades over much of the region have resulted in fewer beetle kill events which usually occur during early and late freezes, as it is during these times when the insects are in their most vulnerable developmental stages (Raffa et al., 2008; Robbins, 2010). Milder temperatures have also created longer growing seasons, which may have allowed more than one generation of MPB to propagate each year (Mitton and Ferrenberg, 2012). Mature forests are also known to be more vulnerable to MPB, while varying forest management strategies (clearcutting, thinning, fire suppression, etc.), may also heighten vulnerabilities (e.g. Pendall et al., 2010; USDA Forest Service, 2011).

There are numerous potential ramifications associated with the large scale die-off of forests in Western North America, including forests becoming a source, rather than a sink for carbon (Kurz et al., 2008), loss of species habitat, changes in local land-atmosphere exchanges (Wiedinmyer et al., 2012), increased fire susceptibility from

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dead and dying timber stands, changes in snowpack and water quality, threats to critical water supplies, etc. (Clow et al., 2011; Pugh et al., 2011). Edburg et al. (2012) have hypothesized a chronological sequence of impacts as a cascade of ecological changes of which biogenic emissions may be an important outcome.

5 Terrestrial vegetation, particularly forests, are a major source of reactive biogenic volatile organic compounds (BVOC) to the atmosphere. These emissions influence atmospheric oxidant chemistry and contribute to secondary aerosol formation, thus playing a role in both climate and air quality. Insect herbivory triggers changes in both the quantities and compositions of BVOC emissions from many types of vegetation.

10 MPB identify and select suitable host trees via olfactory cues from trees, including some particular BVOC emissions (Seybold et al., 2006). Although MPB attacks several pine species, lodgepole pine (*Pinus contorta*) is its main target (Powell and Raffa, 2011). Endophytic pine bark beetles such as MPB are thought to fly mainly within the stem height of potential host trees (Safranyik et al., 2000; Seybold et al., 2006), and thus many ecologically-driven studies have focused on emissions of volatiles (mainly monoterpenes) associated with boles in tree trunks (e.g. Billings et al., 1976; Gara et al., 1993; Pettersson, 2001). A less understood consequence of infestation is its effects on branch-level chemical emissions from afflicted trees, which emit various compound classes in addition to monoterpenes (such as methylbutenol, oxygenated compounds and sesquiterpenes), and which could have substantial implications for regional atmospheric chemistry.

25 The objectives of the current study were two-fold. First, branch-level BVOC emissions were screened in two lodgepole pine forest locations that have been impacted differently by the MPB: one with high tree mortality (up to 70%, which is typical over much of the western Rocky Mountains) and another in which die-off from MPB has been much lower (~5–10%). The second objective was to characterize the seasonal variability among healthy, uninfested trees, and from trees in varying stages of MPB attack at one of the sites. The BVOC emissions measurements described in this paper were performed at two lodgepole pine-dominated forest sites: Chimney Park (CP), Wyoming

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and the University of Colorado Mountain Research Station (MRS). The study was performed as a preliminary screening effort to determine whether site-specific and/or infestation status-related differences in branch-level BVOC emissions were compelling enough to affect fluxes of these compounds to the atmosphere and thus warrant further inquiry and measurements at the sites.

2 Methods

Different classes of trees were sampled depending on the site; descriptions and sampling codes used for each site are presented in Table 1. At CP three classes of trees were sampled: healthy, uninfested trees (referred to hereafter as “Live Green” or “LG” trees), trees infested with the MPB but still containing live green foliage (“Beetle Green” or “BG” trees), and late-stage infested trees whose needles had turned red but had not yet fallen (“Beetle Red” or “BR” trees). At MRS, we sampled BG trees (referred to as “old.beetle” or “OB” trees), apparently healthy uninfested trees before and after being baited with lures and subsequently attacked by MPB (“BB” and “AB”, respectively), and apparently healthy uninfested trees not baited with MPB lures (referred to as “control” or “CT” trees) The sampling design was as follows: each day, three branch enclosures (described later) were sampled, one from each of the sample classes, with three individuals from each group selected for sampling.

2.1 Chimney Park site description and sampling design

Chimney Park (~41°04′ N, 106°07′ W, elev. ~2750 m.a.s.l.) located in Southern Wyoming, USA (Fig. 1) in the Medicine Bow National Forest, is a forested landscape dominated by lodgepole pine and has a long history of experimental research dating back to the 1970’s. At the time of the sampling visit, there were 6 stands within CP actively being used for experimental purposes. Trees growing in or near four of these stands were selected for sampling for this study (Table 2).

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Measurements at CP were made from 15–17 September 2010. Many of the trees growing at CP offered little or no access even to their lowest branches, making the selection of trees for sampling challenging. The enclosures for each day were generally installed on three trees growing as near as possible to each other (Table 2), although it was not always feasible to find a tree from each sampling category growing in the same stand. Due to pump failure on day three, no BR enclosure was installed.

2.2 Mountain Research Station site description and sampling design

The University of Colorado's Mountain Research Station (~40°02' N, 105°32' W, elev. ~2980 m) is located in the Roosevelt National Forest in North-Central Colorado (35 km west of Boulder; Fig. 1) and has been used as an experimental and research forest since at least the 1920's. The MRS site description and extensive climate data is available from the NSF-supported Niwot Ridge Long-Term Ecological Research project and the University of Colorado Mountain Research Station (<http://culter.colorado.edu/NWT/index.html>). The site where BVOC sampling was conducted is dominated by both lodgepole and limber (*P. flexilis*) pine trees. Although MPB infestation has been observed in the MRS site, outbreaks have not been widespread and have mostly focused on the limber pines. Those lodgepole trees that have been hit appear to be surviving the beetle attacks. The reasons for this resilience are not known, but may reflect water status, a resistance to the strains of blue-stain fungus carried by the beetles at the site, or other unknown factors. Some tree mortality from MPB at the MRS has occurred in a smallscale experimental manipulation study that uses commercially available pheromones to attract MPB to specified host trees (Mitton and Ferrenberg, 2012). It is these experimentally-baited trees (along with several unbaited trees) which were sampled at MRS.

The sampling approach taken at MRS was different than that at CP. We had the unique opportunity to sample trees before and after they were infested with MPB, thanks to the tree-baiting experiment fully described by Mitton and Ferrenberg (2012). The objective of the MPB baiting experiment was to ensure MPB attacks for

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documenting MPB flight seasons and generation times between limber and lodgepole pine hosts. Pine trees used in the experiment were selected because they were mature canopy or sub-canopy trees (>20 cm DBH) and had no evidence of prior MPB attacks. The timing of baiting with MPB aggregation pheromone coincided with the 2011 peak flight of MPB at the MRS (Ferrenberg and Mitton, unpublished data).

We planned three 2-day sampling visits to the MRS site from summer to early fall of 2011: 30 June–1 July, 7–8 August and 17–18 September. The first sampling visit was chosen to be at a time before MPB were observed to be flying in the area but after temperatures had warmed up enough to allow the trees to become active in terms of BVOC emissions (generally, this occurs in late May–early June; however, the summer of 2011 was rather cold and rainy, so the pre-baiting measurements were conducted 30 June–1 July). The second visit (7–8 August) was timed to occur just after baiting and close to the week of peak beetle flight in the area, and the third (and final) visit to the site was performed on 17–18 September. This time period was chosen to be close to the same time of year that sampling at CP was performed (for comparison purposes).

2.3 Sampling methods

Branch enclosures at both sites consisted of custom-made Tedlar bags, each of which contained a 1/8" PTFE/stainless steel sampling port located on the edge of the bag furthest from the base of the branch. The bags were placed carefully over the selected branches, ensuring to the best extent possible that needle surfaces were not in contact with the enclosure walls. The enclosures were secured over the branches using ~0.2 cm diameter elastic cords. Enclosure volumes ranged from ~7–10 l, depending on the size of the enclosed branch. Purge air was delivered into the enclosure through a 1/4" PTFE tube (fed into the bag through the bag opening along the branch), connected to a micro-diaphragm pump (KNF, type UNMP830KNDC, Neuberger, Inc., Trenton, NJ, USA) powered by a rechargeable 12 V, 9 Ah sealed battery (WKA12-9F2, Werker, China). The purge air supply pump and battery were placed inside a plastic bin to protect against water intrusion in the case of inclement weather. The pump storage

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bin was located on the ground at the base of the tree being sampled, except in the case of one CP tree, on which the selected sampling branch (the lowest accessible branch on the tree) was high enough above ground (~3.7 m) that the pump enclosure was placed on a ladder. This was done to minimize the length of the enclosure purge air supply line, as it was determined that the length of this line inversely affected the air flow delivery rate of the pump.

The inlet to the pump was fitted with an activated charcoal-filled glass tube (ORBO-32, Supelco, Bellefonte, PA, USA) to scrub the inlet air of ambient VOC and oxidants. One inlet sample was collected at some point in between sample collection at each enclosure, to ensure that ambient VOCs were not influencing the sampled enclosure air. For each quantified and speciated BVOC found in enclosure samples, the reported emission rate of the compound was reduced by the inlet concentration (if detected) from the enclosure inlet sample. For most enclosures and compounds, inlet concentrations were much lower than 1 % of concentrations observed exiting the enclosures. The enclosures were installed on the branches ~16–18 h prior to the start of sampling, in order to allow any installation-induced emissions bursts to subside and to give the enclosure time to equilibrate prior to sampling. As samples were generally collected starting near 12:00 local time, enclosures were installed during the late afternoon/evening of the previous day. Prior to sampling each day, the charcoal inlet scrubber was removed and replaced, and the battery powering the enclosure purge pump was changed.

Temperature inside of the enclosure was measured using either a thermocouple (type K, OMEGA Engineering, Inc., Stamford, CT, USA), or a HOBO temperature sensor (TM6-HE, Onset, Cape Cod, MA, USA). The temperature sensors were fed into the enclosure bag along the base of the branch and shielded from direct solar radiation. Photosynthetically-active radiation (PAR) outside of, (but next to) the branch enclosure was measured using Quantum sensors (using either LI-190, LI-COR Environmental, Lincoln, NE, USA or Apogee SQ-110, Apogee Instruments, Logan, UT, USA). The PAR sensors were mounted onto a leveled tripod at the same height as the branch being measured. Temperature and light data were logged using HOBO dataloggers

(model No. U12-014, H08-004-02, or H08-006-04, Onset, Cape Cod, MA, USA). The HOBO dataloggers were evaluated indoors prior to deployment and were compared with real-time thermocouple temperature readings; temperature responses were within 0.5 °C between each of the sensors/loggers.

Branch enclosure outlet and inlet air samples were collected onto stainless steel cartridges filled with adsorbent. The two-stage cartridges were either custom-filled in-house with a mixture of ~150 mg of Tenax TA (60/80 mesh, Buchem BV, Apeldoorn, The Netherlands) and ~170 mg Carbotrap (20/40 mesh, Sigma-Aldrich Co. LLC, St. Louis, MO, USA), or purchased pre-filled with ~350 mg of Tenax GR (35/60 mesh) and Carbograph 5TD (40/60 mesh; Markes International, Llantrisant, RCT, UK). Samples were collected using custom-made, mass-flow-controlled pumps designed and built in-house. Sampling rates varied from ~150–200 ml min⁻¹ and samples were collected for 30 min.

After sampling was completed, the sampled branches were harvested, dried in a ~57 °C oven for ~48 h, and weighed (both for needle and non-needle biomass). At MRS, repeated measurements were made on dedicated branches from the selected trees during each visit, rather than on different branches. Since emission rates are generally expressed as a function of dry biomass weight, several pictures of each branch were taken at the beginning of sampling in late May to confirm whether any branches had lost needles or twigs over the course of the campaign. None of the branches experienced detectable loss of needles or twigs during the campaign, but by the third sampling period many of the branches had dead or dying needles, and when enclosures were installed during this last visit, care was taken to preserve these needles on the branches (or at least keep them inside of the enclosure bags). During the second and third sampling periods, the number of yellowing and/or completely red needles was recorded for each branch. Final emission rate calculations were made after harvesting and drying the branches (after the final round of sampling) and calculating the expected live biomass dry weight for each of the sample periods.

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It is well-known that pine trees shed needles each year and re-grow new needles; this behavior can affect the calculated emission rates for the trees sampled at MRS during June–July and during August, since the needles sampled from trees during these measurement periods were not harvested and weighed until after the collection of the September samples. Needle longevity among lodgepole pines growing at elevations comparable to that of the MRS site was shown to vary between approximately 10–13 yr (corresponding to an average annual loss rate of $\sim 8\text{--}10\% \text{ yr}^{-1}$), while new foliar growth among lodgepole pine (expressed as annual incremental change in foliated shoot length) was found to be $\sim 6\text{--}7\%$ (Schoettle, 1990). The MRS study took place over a period of ~ 11 weeks, while the estimates of new foliar growth provided by Schoettle (1990) are based on annual changes (although most new foliar growth would be expected to occur at high-altitude sites such as MRS between \sim May–August). Given these observations, we estimate that overall biomass change during the course of the MRS study was likely $< 5\%$. Nonetheless, these potential small changes in biomass between the June–July and August measurement periods should be considered as a caveat when interpreting the emission rates obtained for the June–July and August MRS samples.

2.4 Analytical methods

Air sampling cartridges were thermally desorbed using an Ultra autosampler (Series 2, model ULTRATD, Markes International, Llantrisant, RCT, UK) and analytes were then cryo-focused onto a Unity thermal desorber (Markes International, Llantrisant, RCT, UK) operated in splitless mode. The samples were subsequently injected into a gas chromatograph (GC; model 7890A, Agilent Technologies, Inc., Santa Clara, CA, USA). Flow path temperatures within the Ultra-Unity were maintained at $\sim 175^\circ\text{C}$. The GC was equipped with a flame ionization detector (FID) and a mass-selective detector (MSD; 5975C inert MSD, Agilent Technologies, Inc., Santa Clara, CA, USA), and had an HP-5MS column (30 m \times 0.25 mm, 0.25 μm , Agilent Technologies, Inc., Santa Clara, CA, USA), using nitrogen as a carrier gas.

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The MSD was run simultaneously in both SIM and SCAN modes, allowing detection of both routinely-observed compounds as well as those less frequently found. Compounds detected in samples taken during the 2010 Chimney Park campaign were quantified using the FID along with an external gas-phase standard containing 71 ppbV isoprene and 48 ppbV camphene. In cases where analytes could not be quantified using the FID signal, the relative SIM- or SCAN-mode MSD response of the target ion for the compound was quantified relative to the response for the same (or similar) ion quantified in the external standard. The GC oven heating program used in the analysis of the Chimney Park samples was as follows: start temperature: 35 °C, hold time, 1 min, Ramp 1, 6 °C min⁻¹ → 80 °C (no hold time), Ramp 2, 3 °C min⁻¹ → 155 °C (no hold time), Ramp 3, 10 °C min⁻¹ → 190 °C (no hold time), Ramp 4, 25 °C min⁻¹ → 260 °C, final hold time 5.2 min (total run time, 45 min).

For samples analyzed during the 2011 MRS Ridge campaign, quantifications were performed using the FID and a gas-phase external standard containing 165 ppbV isoprene and 71 ppbV camphene. Additionally, for samples collected in 2011, an internal standard consisting of ~1 ppm trans-decahydronaphthalene was added to each sample analyzed. The GC oven temperature ramping program used for the analysis of the MRS samples was as follows: Start temperature: -30 °C, Hold time, 1 min, Ramp 1, 20 °C min⁻¹ → 0 °C (no hold time), Ramp 2, 6 °C min⁻¹ → 80 °C (no hold time), Ramp 3, 3 °C min⁻¹ → 190 °C (no hold time), Ramp 4, 30 °C min⁻¹ → 260 °C, final hold time 5.9 min (total run time, 60.7 min).

Blank samples were also analyzed for each day of the sampling campaign; both opened and unopened blanks were analyzed to determine potential contributions to detected analytes from background concentrations or sample handling and storage activities. For any compounds detected in the samples that were also found in blanks, the maximum amount of analyte detected in the blank was subtracted from the enclosure samples for that day. For most compounds investigated, an analytical precision of ~3 % was estimated based on repeated analyses of gas-phase standards, while accuracies

of quantitative emission results reported using SIM ion quantitation are estimated at $\pm 15\%$.

2.5 Statistical methods

Two-sample *t*-tests were used to verify whether observed differences in total monoterpene (MT) basal emission rates (BER i.e., emissions normalized to standard conditions of 30 °C) and sesquiterpene (SQT) BERs were significant between individual trees and/or sampling classes (depending on the site and number of samples/individuals screened). Prior to running the *t*-tests, *F*-tests were performed on each variable tested. When the results of the *F*-tests indicated that variance was not significantly different (for $\alpha = 0.01$), variances were pooled for the *t*-test calculations. Statistical tests were not performed for methyl-3-buten-2-ol (MBO) or oxygenated volatile organic compound (OVOC) emissions due to the lack of well-constrained techniques to normalize these emissions to account for light and temperature effects.

K-means cluster analysis of September MT profiles was applied to the live trees at both CP (including LG and BG trees, $n = 6$, but excluding the presumably dead BR trees) and MRS (all trees, $n = 6$, since no dead trees were sampled at MRS) to look for significant differences in MT profiles which would indicate the presence of different chemotypes at the sites. Cluster solutions for each site were tried for the three or five most dominant MT (since these contributed most of the observed variability) and for all MT detected (excluding MT that comprised $<0.1\%$ of average MT composition) after Bäck et al. (2012). The sum of squared error (SSE) for every possible selected cluster solution was tested against the SSE using 250 randomized versions of the original input data. The SSE of strong cluster solutions would be expected to decrease more quickly than the randomized SSE solutions as the number of clusters increases. Principal component analyses (PCA) were also performed for the selected cluster solutions and the clusters plotted based on the PCA results for the first two principal components. Strong clusters would be expected to have good structure and little or no overlap in the PCA plot space.

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3 Results

Emission rates (ERs) and ratio analyses (i.e., the contribution to each compound class by individual compounds) are presented for three or four groups of compounds/compound classes (depending on the site): MBO, MT, SQT, and OVOC (results available for MRS site only). For all compounds, we report ranges of observed ERs and, if appropriate, results from linear regressions. For MT and SQT, BERs and corresponding standard deviations are reported, though it should be noted that there was substantial variability in enclosure temperatures and emissions observed during the study, and the expected exponential relationships between MT and/or SQT emissions and temperature were frequently not observed. BER were calculated using Eq. (5) from Guenther et al. (1993): $BER = ER_T / (\exp[\beta(T - T_s)])$, where ER_T = ER observed at temperature T , $T_s = 30^\circ\text{C}$, and $\beta = 0.1$ for MT (Guenther et al., 1993) and 0.15 for SQT (Duhl et al., 2008). For MBO and OVOC, avg. ERs and associated standard deviations are reported instead of BERs. All ERs and BERs reported in the text are in units of $\mu\text{g C g}_{\text{dw}}^{-1} \text{hr}^{-1}$.

3.1 Chimney Park, WY results

During the course of the three-day sampling campaign, a total of 29 enclosure samples and 7 inlet samples were collected. After analysis of the enclosure samples, it was discovered that 9 of the cartridges used during the CP sampling campaign had damaged adsorbent materials and the emission results obtained from these cartridges were therefore excluded from the final emission rate analysis. Despite this loss of data, several qualitative differences in absolute emission rates emerged between the various studied groups for two of the compound classes observed (MBO and SQT).

The BR trees emitted negligible amounts of MBO (Table 3), which is predictable given the fact that these needles appeared to be dead and MBO is known to be synthesized de novo, largely as a function of photosynthesis (e.g. Gray et al., 2003). There were no obvious class-related differences MT emissions among the BG and LG trees (Table 7)

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or in MBO emissions across the sampling classes. Tree BR1 had the highest avg. MT BER (4.15), followed by BG3 (2.13) and LG3 (1.36), while the remaining five trees had avg. BERs ranging from 0.35–0.54. The high emissions observed in BR1 likely reflect the fact that, as the needles (and possibly also branch and/or twig biomass) on this branch were dead, emissions could have been caused by simple evaporation of stored MT from pools in the tissues of this branch, and/or release of MT caused by rupture of some of the needles and twigs during enclosure installation. The BG and BR trees had lower SQT ERs and BERs than the LG trees, even those LG branches that were subjected to comparable (or even lower) branch enclosure temperatures than their BG or BR counterparts (Fig. 2, Table 3). The LG trees emitted a significantly higher fraction of SQT than the BG trees (Table 7) and had significantly higher SQT BERs than the BG trees, although this relationship was weaker.

Although the total amounts of MT emitted from the various branches studied at CP varied quite a lot from individual to individual, the ratios of these compounds emitted by individual branches were remarkably similar among the LG and BG individuals sampled (Table 4), with β -pinene always comprising the largest component of emissions, followed by α -pinene and/or β -phellandrene (and in one case, 3-carene). The BR trees differed somewhat in the composition of their MTs, with 3-carene being dominant followed by β -pinene and/or β -phellandrene. Within-tree variation in MT composition was very low across the samples, except for the third sample collected from enclosure LG1 (which had experienced substantial heat stress, with enclosure temperatures averaging $\sim 14.6^\circ\text{C}$ above ambient temperatures over the course of the second and third sample collection periods). Only the first two samples from LG1 were included in the average shown for this tree in Table 3, since the MT ratios were very similar between these two samples. The third sample collected from LG1 (after ~ 3.5 h of nearly-continuous heat stress on the branch) exhibited a $\sim 10\%$ reduction in the contribution from 3-carene (compared with the first sample collected), the appearance of $\sim 5\%$ z - β -ocimene (which was not found in any other sample collected during the campaign and may be produced in response to stress), and small changes in the other MTs

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detected. Although the MT ratios for LG1 changed over the course of sampling (presumably in response to heat stress), the SQT ratios remained more or less unchanged despite the heat stress.

K-means cluster analysis applied to the live (i.e., LG and BG) trees for three MT lumping schemes tested (i.e., using either the three or five most dominant MT or all MT detected) and for all cluster solutions evaluated (2–6 clusters) resulted in SSEs that never deviated from the SSEs of the 250 randomized versions of the original input data, indicating the presence of only one MT chemotype at CP (among live trees but irrespective of infestation). The PCA plots obtained for the various cluster solutions and lumping schemes lacked coherent structure and frequently contained overlap between clusters, further evidence that only one chemotype was present in the live trees at CP.

The ratios of SQT observed at CP tell a different story, with SQT compositions being somewhat unique across the three sample classes, and possibly also among individual stands (Table 5) α -farnesene or e - β -farnesene were the dominant compounds among the LG trees, while the BG trees had more heterogeneous SQT profiles, with increased importance of γ - and δ -cadinene, along with aromadendrene and α -cuprenene in BG2. Interestingly, LG3 was similar to BG3; these two trees were growing relatively close to each other (but were still in different stands), while BG2 and BR2 also had similarities (and were growing just a few meters apart). Among the BR trees that were screened, BR1 had SQT ratios comprised almost entirely of γ - and δ -cadinene, while BR2 (which emitted very little SQT) was dominated by γ -cadinene, aromadendrene, and α -cuprenene.

3.2 MRS, CO sampling results

3.2.1 MBO emissions

There was no obvious difference in MBO ERs (Table 3) between any of the classes during either the 30 June–1 July or 7–8 August sampling visits, although on both sampling occasions (including the first, pre-baiting measurement), one of the three samples

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collected from BB2 exhibited an MBO emission burst (~ 2.5 times higher than other samples collected at similar temperatures, with no correlation to PAR). We have excluded these values from the calculation of slopes describing the relationship between MBO emission rate and temperature (below), but the values are reported in the foot-
notes of Table 2. As previously observed in ponderosa pine (Gray et al., 2003) we saw a linear response in MBO emissions to temperature, although the total emission rates and slope of this line differed between these first two visits: during 30 June–1 July sampling, the slope was 0.226 ($R^2 = 0.73$), while for 7–8 August sampling, the slope = 0.046, ($R^2 = 0.73$).

Samples collected on 17–18 September revealed that the two OB trees, CT2, and the secondary branch sampled from AB1 (AB1b) had higher MBO ERs than the other branches screened (Table 3). CT2 and AB2 emitted comparable amounts of MBO to what was measured in AB1b and OB1, although the temperatures these branches experienced were $\sim 5^\circ\text{C}$ warmer than what was observed in enclosures AB1b and OB1. The other trees (CT1, AB1 and AB2) all had lower MBO ERs. One enclosure (OB2) reached $\sim 24^\circ\text{C}$ during the September measurements, with a corresponding MBO ER of ~ 2.25 . Interestingly, the group of branches with negligible MBO emissions in mid-September were also the branches that had experienced heat stress from enclosure overheating in August (although CT2, which had higher MBO emissions, experienced a short duration of heat stress during August sampling; see Heat Stress section below).

3.2.2 MT emissions

During sampling on 30 June–1 July five of the trees screened had relatively low MT BERs (compared to other pine species as reported in Helmig et al., 2007) while CT1 exhibited a higher MT BER (Table 3) and there was significant variability in MT BER among most of the trees (Table 7). Samples collected in early August (Fig. 3) indicated a more or less linear relationship between emissions and temperature and less tree-to-tree variability in BERs (Table 7). The slope obtained when a linear regression was performed on all the data (0.07) had a much lower R^2 value (0.51) than when

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the baited trees were treated with a separate regression analysis. The baited trees exhibited lower MT emissions than both control and old-beetle trees at similar temperatures, and had a lower slope (0.07, $R^2 = 0.80$) than control and old beetle trees (0.11, $R^2 = 0.84$, Table 3). The September samples mostly displayed a linear relationship between temperature and emissions, with a slope of 0.11 ($R^2 = 0.78$), excluding OB1 which had a significantly higher BER than the other branches (Table 7), as well as a secondary enclosure placed on AB1 (AB1b, placed on a previously unsampled branch to evaluate the potential long-term effects of enclosure heat stress that occurred during August measurements, see Heat Stress section below), which had a BER ~2.6 times higher than the other enclosures. The enclosures exhibiting the lowest MT emissions in September were the same enclosures that experienced the most heat stress during the August measurements (see Heat Stress section, below).

3.2.3 MT speciation profiles

MT species (Table 4) identified in each tree during the course of the campaign exhibited significantly more tree-to-tree variability than what was observed at the CP site. During late June/early July, β -pinene was the dominant MT emitted by three of the trees (OB1, OB2 and BB1), β -phellandrene was the dominant MT emitted by both trees selected to be controls (CT1, CT2), and 3-carene was the dominant MT emitted by BB2.

During mid-September sampling, the MT which were dominant in the first sample period returned to being dominant (with 3 of the trees showing higher fractional contributions from these dominant compounds relative to the first sampling visit), except in CT1 (which was also by far the highest MT emitter). The baited trees both exhibited a decrease in the richness of MT chemical species emitted by the end of the campaign, with an average of 10 MT observed on 17–18 September compared to 16 during 30 June–1 July. The control trees both also exhibited a slight decrease (average of 14 MT species observed on 17–18 September compared to 16 during 30 June–1 July), while the old-beetle trees did not show a decrease in MT diversity. A secondary enclosure that was placed on a previously unsampled branch from baited tree #1 (i.e. AB1b)

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during the September measurements had a very similar MT profile as the dedicated branch from this tree.

When k-means clustering was applied to the September MT compositions for all trees, and the resulting SSE for different cluster solutions was tested against the SSE results of 250 randomized versions of the original data, the results indicated strong solutions for 3–5 clusters depending on the lumping scheme used (i.e., using either the three or five most dominant MT or all MT detected), indicating the presence of 3–5 MT chemotypes at MRS in September (irrespective of infestation status). The PCA plots obtained for the various cluster solutions and lumping schemes also indicated strong clusters, further evidence that several chemotypes were present in the live trees at MRS. When three clusters were selected, the resultant chemotypes could be classified according to their dominant MT (β -pinene, β -phellandrene, or 3-carene). When 5 clusters were used, additional “mixed” chemotypes emerged that were formed when the average contribution to total MT from the second-most dominant MT was within a factor of two of the dominant compound.

3.2.4 SQT emissions

During 30 June–1 July sampling, two of the trees (OB2 and CT1) exhibited significantly higher SQT BERs than the other four trees (Tables 3 and 7). During both the early-August and mid-September sampling days, the trees that had survived beetle attacks 1–2 yr prior to sampling (OB1, OB2) had the highest SQT BERs. For August samples, SQT emissions were >2 times higher in the old-beetle trees than in any of the other trees at comparable temperatures (Table 3) and BERs were significantly higher than all other trees, although the relationships were weak (Table 7) SQT emissions measured in September were lower than August emissions for all trees, with the old-beetle branches emitting ~5–40 times more SQT than other trees at similar temperatures (Fig. 4, Table 3); the BERs for the OB trees were also significantly higher than the other trees (Table 7).

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3.2.5 SQT speciation profiles

Table 5 depicts the SQT species identified in each tree during the campaign. Similar to behavior observed in MT profiles at the MRS site, there tended to be a decrease in the fractional contribution of the dominant SQT species emitted in late June/early July as compared with early August samples, except in baited tree #1 and control tree #2 (where the dominant compound from 30 June–1 July samples became more dominant). The second-most dominant compound from 30 June–1 July samples generally became the dominant compound during the early-August measurements. In both the 30 June–1 July and 7–8 August sample periods, *cis*- β -farnesene and α -farnesene were the dominant SQT emitted from all of the trees. Other SQT that tended to be present in quantities $\geq 2\%$ during the first two sampling visits were α -transbergamotene, an unidentified SQT, an in a few enclosures, γ - and δ -cadinene. Similar to observed MT behavior, the baited trees exhibited a decrease in the richness of SQT chemical species emitted by the end of the campaign. Compared to 30 June–1 July sampling, the control trees averaged three fewer SQT species observed in September, while the old-beetle trees averaged one less SQT present during the September visit as compared to the first measurements. Both baited trees had completely different SQT profiles in September (comprised mostly of *e*- β -farnesene), compared with samples from the first two visits and with the other trees. Control tree #1, which had SQT profiles similar to the other trees during the first two measurement periods, also had a radically different SQT profile during the final sampling visit, dominated by δ - and γ -cadinene, a profile that was similar to that of the MPB-infested trees at Chimney Park.

3.2.6 OVOC emissions

Emissions of total OVOC (Table 3) observed during the 30 June–1 July campaign exhibited an exponential temperature response ($R^2 = 0.64$), with an avg. OVOC ER of 0.165 and avg. enclosure temperature of 24.4 °C. 7–8 August OVOC emissions were

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similar to 30 June–1 July values for all trees except AB2. OVOC emissions in mid-September were <0.0004 for all enclosures other than the two OB trees and CT2. Tree OB1 emitted ~ 22 times more OVOC during the 17–18 September measurements than other trees at similar enclosure temperatures while the avg. ER for OB2 and CT2 (which reached the highest temperatures) was 0.003 at an avg. temperature of 18.2°C . There is an uncertainty of $\sim 30\%$ associated with reported OVOC ERs and speciation profiles.

3.2.7 OVOC speciation profiles

OVOC profiles (Table 6) observed during the first visit showed eucalyptol as the dominant compound in BB1 and both of OB trees, while amyl acetate was dominant in CT2. OVOC measured from CT1 was dominated by methyl salicylate and had a significant contribution ($\sim 28\%$) from pinocarvone, which was not found in amounts $>5\%$ in any other tree during the campaign. Linalool was the dominant OVOC observed in the first post-baiting measurements (mid-August) from both baited trees as well as in CT1, while amyl acetate was the major OVOC emitted from OB1 and CT2 (which also emitted $\sim 31\%$ methyl salicylate). OB2 was dominated by eucalyptol and linalool. Camphor dominated September OVOC emissions all of the branches except OB1 and OB2 in which amyl acetate and eucalyptol were the dominant compounds, respectively. As observed for MT and SQT ratios, the old beetle trees also had the highest average OVOC diversity as a group during the final sampling visit.

3.2.8 Effects of heat-stress on emissions

MRS control tree #2 experienced very high temperatures (just over 40°C , the highest observed enclosure temperature; average ambient temperature at this time was $\sim 28^{\circ}\text{C}$) during sampling on 8 August. The enclosure overheating, which lasted for ~ 35 min, occurred between collection of the first and second samples (both of which were within 3°C of ambient values), and was caused by an extended period of direct

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sunlight incident on the enclosure. This offered an opportunity to evaluate the effects of a short-duration of heat stress on both compound ratios and total emissions before and after exposure of the branch to high temperatures, as well as to evaluate longer-term effects on emissions since we returned to the site and sampled all of the branches again in mid-September. Ratios of OVOC did not change substantially after the exposure of the branch to high temperatures, and the branch emitted about the same total amounts of OVOC as other branches (except AB2, which emitted more OVOC than any other branch). In contrast, MT and SQT ratios did change before and after the heat event experienced by the branch. The largest change in MT composition was a ~23% decrease in β -phellandrene and a ~14% increase in α -terpinene, with contributions from other MT also changing following the heat event, but to a lesser extent. The SQT, α -farnesene decreased by ~18% while *cis*- β -farnesene increased by almost 16%.

The total amount of SQT emitted following the heat event was almost the same as the pre-heating emission, despite the fact that the temperature was ~1.3°C warmer in the latter sample. Total MT emitted before and after the heat event exhibited temperature-related differences which were similar to the quantities and temperature-dependencies of MT emitted by OB 1 (which did not experience heat stress), suggesting that short-term heat stress did not substantially affect the quantity of MT emitted.

In AB2 August enclosure temperatures slowly climbed from ~30°C up to ~39°C in the 45 min prior to the collection of the first sample, and thereafter declined back to within ~2–3°C of ambient values for the remainder of the sampling period. MT ratios did not change dramatically, although myrcene became more dominant and carene became less dominant. Limonene also increased slightly. SQT contributions between the first and third samples changed almost identically to what was observed in CT2 following heat stress, while the first two samples collected from AB 2 were quite similar in terms of SQT ratios, so most of the change occurred after enclosure temperatures had cooled down and remained relatively cool for ~1 h. Ratios of OVOC did not change much during the course of sampling.

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5 A second branch from baited tree #1 (“AB1b”) was sampled during the September sampling period to evaluate potential long-term effects on emissions following a period of extreme heat stress, as the primary branch sampled from this tree was exposed to the longest period (~4.5 h) of heat stress during August sampling. MT ratios were similar between this branch and the primary branch sampled from the tree. MT ERs were slightly higher in branch b than in the other branches experiencing similar temperatures (branches AB1 and CT1), but still lower than what was emitted by OB1, which experienced similar enclosure temperatures during September sampling. SQT were not detected from AB1b during September sampling, although they were emitted in tiny amounts from AB1.

4 Discussion

15 This screening study highlights differences in the chemical diversity of emissions from lodgepole pines in two forest sites impacted differently by the MPB, and also presents the seasonal variation observed in BVOC emissions from apparently healthy trees versus trees in different stages of MPB attack. At the Chimney Park, Wyo. site, where MPB-related mortality has been high, we found remarkable homogeneity in MT speciation profiles among the various live lodgepole trees studied, irrespective of whether or not they had been infested by MPB. The ostensibly dead (i.e. red-needled) trees at the Chimney Park site emitted negligible amounts of MBO and had different MT profiles than live trees. This may simply reflect a difference in the MT speciation profiles of woody and foliar tissues. Ratios of SQT detected at that site were more unique to the studied groups (i.e., uninfested trees had a different profile than infested but still living trees etc.), and possibly reflective of the stage of infestation, although within-stand differences may also have been present. The SQT γ - and δ -cadinene seemed to correlate with MPB infestation at CP. Measurements conducted at this site suggest 20 that trees which might be expected to succumb to MPB attack may have lower SQT emission capacities than their non-infested counterparts.

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Lodgepole trees screened at the University of Colorado Mountain Research Station (MRS) site exhibited significantly more diverse MT profiles, even though the trees that were sampled at this site were growing much nearer to each other and were also closer in age than the Chimney Park trees. At the MRS site, lodgepoles appear to be more resilient to the MPB and/or its fungal symbiont, the blue stain fungus (of which there at least two species). Although, to our knowledge, little is known about whether specific chemotypes of *Pinus contorta* are better equipped to survive MPB attack, it has been known for some time that MT emission profiles observed both in tree resin and in emissions are unique between most subspecies/varieties of lodgepole (e.g. Zavarin et al., 1969; Lusebrink et al., 2011). Even within a seemingly homogenous zone, several chemotypes of a given species may be present, and the relative ratios of compounds detected in ambient air have been observed to reflect this diversity (Bäck et al., 2012).

SQT profiles from MRS trees in both late June/early July and early August showed a high degree of homogeneity, although samples collected in mid-September (during the same time of year that the trees at Chimney Park were sampled) suggest that at some point in the ~6 weeks following MPB attack, SQT profiles change in newly-infested trees. Also, the trees that survive attack (from previous years) may eventually return to their “pre-attack” SQT profiles (based on the observation that survivors of MPB attack had profiles similar to uninfested trees). There were no obvious class-related differences in SQT emissions at the MRS in late June/early July sampling, but during both the early-August and mid-September visits to MRS, the trees that had survived beetle attacks 1–2 yr prior to sampling exhibited the highest temperature-adjusted SQT emission capacities, which was opposite the trend observed at CP. The two beetle attack survivors sampled at MRS in early August did not experience enclosure heat stress, while the other four enclosures (placed on two baited and two control trees) did experience varying degrees of enclosure over-heating. However, the mid-September sampling period included an enclosure placed on a previously unsampled branch from one of the baited trees, and results from this enclosure showed no SQT emissions. Further evidence supporting this conclusion is the observation that one of the control

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trees experienced elevated enclosure temperatures for only ~35 min during the August measurements, and still exhibited substantially lower SQT emissions than the trees that had survived MPB attack in prior years. The reasons why elevated SQT emissions were seen in the MPB survivors only during mid- and late-season sampling are unknown, but it could be a response to beetle activity at the site, since MPB were not observed at the site during the early season sampling campaign at MRS.

Emissions of MBO from vegetation are both light- and temperature-driven and decline with needle age (Gray et al., 2003). This was not observed between the late June/early July versus early August campaigns, although MBO emission capacities (at comparable temperatures) were lower in mid-September. The observations that mid-September MBO and MT emissions were lower among branches that had experienced heat stress lead us to conclude that exposure to a relatively short duration (~30 min to several hours) heat event may reduce longer-term MBO and MT (and possibly also SQT) emission capacity, although this effect is not definitive. Both MT and SQT ratios did change in response to heat stress, although this was likely a short-term effect.

There was quite a lot of diversity in OVOC profiles emitted by the MRS trees earlier in the growing season, but this declined towards the end of the season. Other than a tree potentially infested with an unknown pathogen at MRS (see below), there were not significant differences in total OVOC emissions between the trees, except during the last measurement days, when the MPB survivors emitted more OVOC than their counterparts, and had the highest chemical diversity in OVOC (as well as in MT and SQT) during this time. There was a general pattern across all compound classes studied that the number of compounds detected in each class declined during the growing season, with the greatest decline observed in newly-infested trees and the least decline in survivors of MPB-attack from previous years.

We postulate that one of the two trees selected to be a control at the MRS site (CT1) may have been infested with an unknown pathogen. This tree exhibited by far the highest MT emission capacity during the 30 June–1 July sampling visit and among the highest SQT emissions during this visit. By now it has been well-documented that

biotic stressors can enhance emissions of both MT and SQT (Duhl et al., 2008; Huber et al., 2004). Although stress-induced emissions bursts have also been observed following enclosure installation (e.g. Arey et al., 1995), all enclosures were allowed to equilibrate following installation and prior to sampling for a minimum of 16 h to minimize this risk. Enclosure temperatures remained very close to ambient values for all trees sampled during this first visit, so heat stress was ruled out as a cause of the unusual emissions observed from CT1. This tree also emitted methyl salicylate as its dominant OVOC during the first sampling period (unlike any other tree sampled), a compound whose emissions are known to be enhanced in many tree species in response to both biotic and abiotic stressors (e.g. Kännaste et al., 2008; Joó et al., 2011). This tree also emitted ~30–50 times more o-cymene and p-cymenene than any of the other trees sampled (data not shown). Finally, this tree had unusual behavior in both MT and SQT profiles through the course of the growing season. Whereas the dominant MT observed in the early part of the season was also dominant towards the end of the growing season, in this tree alone a different MT was dominant during the final sampling visit. Bäck et al. (2012) found that despite seasonal changes observed in MT profiles among different chemotypes of Scots pine (*Pinus sylvestris* L.), the dominant compound did not cease to be dominant. Also, the SQT profile observed in this tree at the end of the campaign was singular, but was interestingly comprised mostly of the same two compounds found to be associated with MPB infestation at the Chimney Park site (δ - and γ -cadinene).

Some evidence suggests that fire injury in *Pinus contorta* can lessen induced defenses against MPB (Powell and Raffa, 2011). Fires have not been common at MRS in the last few hundred years nor at CP in the last 100 yr (Table 1). None of the trees sampled had obvious fire damage, and so this effect is not suspected to play a role in observed differences between Chimney Park and MRS. It is not known which species of blue stain fungus predominate either site, while it is known that some species are more virulent than others (Lusebrink et al., 2011), and so this factor cannot be excluded when pondering the difference in apparent MPB resilience between the sites.

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Finally, the presence or absence of water stress must be considered when evaluating differences between the sites, as water deficit has been shown to decrease emissions of MT in lodgepole seedlings and may also affect tree resistance to MPB (Lusebrink et al., 2011; Safranyik et al., 2010). We analyzed precipitation data for the 5- and 10-yr period prior to the start of sampling at both the Chimney Park and MRS sites (using PRISM precipitation data, PRISM Climate Group, Oregon State University, <http://prism.oregonstate.edu>, created 4 February 2004), and found that both sites had received at or above 100 % of their 1971–2000 average during the study time periods (Table 1), although it should be noted that average annual precipitation at CP is ~60 % of the average for MRS.

The fact that the sites were relatively moist during the recent period compared to their long-term climatologies is not intended to dispute that severe punctuated drought may have been a possible causative mechanism for the MPB outbreak that caused widespread tree mortality as has been suggested by others (e.g. Bentz et al., 2010; Breshears et al., 2005). Instead, it is only presented to illustrate the recent hydroclimatic context of the summers when BVOC sampling was performed. Lastly, it is noted that the seasonal soil moisture in this region is largely modulated by strong springtime snowmelt followed by a long gradual dry-down throughout the summer growing season with periodic pulses from summer rainfall events. Both sites exhibited this typical behavior (not shown) during the 2010 and 2011 sampling years.

The observations made in this study have unclear consequences on our understanding of feedbacks in plant-insect interactions (since MPB are thought to respond mainly to BVOC emanating from trunks) whereas the potential for MPB-driven effects on regional air quality and SOA formation from both trunk- and canopy-level emission changes is a more obvious use of the data. Since secondary organic aerosol (SOA) yields of different MT and SQT species vary by more than a factor of two (Ng et al., 2006), a change in MT or SQT speciation can result in a substantial change in the amount of SOA produced from these emissions. The change in MT speciation of the newly-infested trees at MRS include one case where the average SOA

yield would increase, due to a higher proportion of 3-carene and δ -limonene, and another case where the average SOA yield would be expected to decrease due to the increase in the contribution of β -pinene. Reactivities of the SQT species (relative to the major atmospheric oxidants) observed in this study are not well constrained, and further inquiry into the aerosol formation potentials of the SQT cadinenes (which may become elevated in MPB-infested trees experiencing MPB-related decline) and farnesenes is warranted. The predicted half-lives of α - and β -farnese with respect to OH and O₃ are ~60–70% of what is predicted for δ - and γ -cadinene (<http://www.epa.gov/opptintr/exposure/pubs/episuite.htm>), therefore the expected SOA yield might be lower if the cadinenes are emitted in increased proportions by declining trees. Also of consequence are the total emissions of the various compound classes, though these impacts need to be more fully qualified before such extrapolations can be made. Amin et al. (2012) observed higher trunk-level concentrations of the MT β -phellandrene and total MT near infested lodgepole trees as compared to healthy trees and suggested a possible increase in SOA from the MPB epidemic due to both increased MT concentrations near infested trees and to the reactivity of β -phellandrene compared with other MT, although it is also necessary to consider that as soon as needles die, there will be significant loss of SOA-formation potential from MBO since this compound is produced de novo. MBO is thought to make substantial contributions to SOA formation in areas where it is emitted in great quantities, such from the pine forests of western North America (Steiner et al., 2007).

A caveat to the results presented in this study is the low number of individuals screened, which limits the validity of making extrapolations based on the data we have presented. However, the stark contrast evident between the two sites speaks for itself, and several statistically significant relationships have been observed. These findings, especially when combined with other relevant studies beg for more research into relationships between lodgepole pine chemodiversity and MBP resilience. Additionally, future measurements should focus on quantifying the potential magnitude and direction

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of change in altered BVOC emissions and thus possible consequences for regional atmospheric chemistry.

5 Conclusions

Whether the relatively high degree of variability in monoterpene profiles observed in lodgepole pine trees growing at the MRS site is related to the MPB-resistance observed there remains uncertain. Owing to the low number of individuals screened, this study also leaves open the question of to what degree emissions and ratios of various BVOC compound classes emitted from the forest are in response to beetle activity versus natural tree-to-tree variation and seasonal changes. We observed fewer compounds in each compound class (monoterpenes, oxygenated VOC, and sesquiterpenes) studied near the end of the growing season as compared to earlier in the season at MRS. Results from our study suggest that in lodgepole pine trees, high proportions of certain SQT (i.e., δ - and γ -cadinene) may be indicative of poor overall fitness, caused by either unknown pathogens (as may have been the case in one tree sampled at MRS), or by the decline following successful MPB infiltration (as seen at Chimney Park). Changes in sesquiterpene composition from newly-infested trees may take weeks to become apparent, and may not persist in trees that survive attack. Our results suggest that lodgepole trees which do survive may have long-term increases in SQT emission capacities, but only during certain times of the growing season, whereas stands of trees that succumb to the MPB/blue stain complex (based on CP results) may have reduced SQT emissions relative to healthy trees (at least towards the end of the growing season). Since SQT have higher SOA yields than most other BVOC emitted by vegetation, this could have significant implications for regional air quality and aerosol formation at certain times of the year. Future research should focus on understanding MPB effects on chemodiversity and conversely the effects of lodgepole pine chemodiversity on MBP resilience, as well as on quantifying the potential magnitude and direction of

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Table 1. Descriptions of trees sampled at CP and MRS, stand characteristics, summary of site disturbance and management histories, and hydrological characteristics of CP and MRS sampling sites.

Site	Tree classes sampled, tree codes for each tree sampled in class	Stand characteristics, management and disturbance histories	Site hydrological characteristics
CP	Un-infested apparently healthy, LG1, LG2, LG3	The sampled stands at CP vary in density and basal area (Table 2), reflecting differences in natural regeneration since the last major disturbances that occurred in the area (early 20th century), including a combination of logging and stand-replacing fire, and more recent management activities (thinning and harvest). Since ~2007, CP has seen rather intense infestation by the MPB that has resulted in as much as 70% tree mortality in some of the stands.	Avg. annual precipitation: 435 mm. In the 5- and 10-yr periods prior to the sampling year (2010), CP received an average of 106% and 110% of normal precipitation, with punctuated deficits in annual precipitation (e.g. 81% of avg. in 2002)
CP	Infested 1–3 yr prior to sampling, green needles, BG1, BG2, BG3		
CP	Infested 1–3 yr prior to sampling and in decline (red needles), BR1, BR2		
MRS	Un-infested trees used as controls, CT1, CT2	The trees growing in and around MRS have not been extensively managed since widespread cutting that continued throughout the 1800s until roughly 1910. Large-scale wildfires are rare in lodgepole forests near the MRS over the past 300–400 yr (Sibold et al., 2006). Infection by dwarf mistletoe (<i>Arceuthobium americanum</i>) is common among lodgepole pines at the MRS, but does not currently appear to significantly affect tree mortality following MPB attack (Ferrenberg, unpublished data). Episodic MPB outbreaks have been recorded for nearby forests in the 1930s and 1970s (Sibold et al., 2007), with the most recent infestations at the MRS beginning in 2007.	Avg. annual precipitation: 730 mm. In the 5- and 10-yr periods prior to the sampling year (2011), MRS received an average of 99% of normal precipitation (during both periods), with punctuated deficits in annual precipitation (e.g. 78% of avg. in 2002)
MRS	Apparently un-infested trees sampled before MPB baiting, BB1, BB2		
MRS	BB trees (from above) sampled after MPB baiting and subsequent infestation, AB1, AB2		
MRS	Trees infested 1–3 yr prior to sampling with no sign of decline, OB1, OB2		



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Table 2. Stand-memberships and characteristics for trees sampled at CP (September 2010) and MRS (June–September 2011).

Stand code (Site code)	tree code(s) of trees sampled within stand and sample dates	Stand characteristics	Avg. distance (m) between trees sampled on same day and other stands
S1 (CP)	LG1 (15 Sep 2010)	Regenerating stand containing almost exclusively LG trees (i.e., little or no MPB infestation). Stand tree age range: 25–30 yr. Avg. stand DBH: 4 cm (DBH of LG1 ~10 cm); avg. stem density: 8670 ha ⁻¹ . Located in stand surrounding S1, >70% of trees infested by MPB (starting in 2008–2009). Age range: 40–80 yr, DBH of BG1: ~15 cm, avg. stem density unknown but likely 1000–2000 ha ⁻¹	located ~160 m W of S3 and ~410 m from S4.
just outside of S1 (CP)	BG1 (15 Sep 2010)		BG1 located ~30 m from LG1 (S1)
S2 (CP)	BR1 (15 Sep 2010)	Age unknown (but probably 40–60 yr), Avg. DBH: 20 cm (DBH of BR1: ~15 cm); avg. stem density: 1160 ha ⁻¹ .	BR1 located ~160 m from BG1 and ~140 m from LG1, ~180 m SW of S3 and ~420 m WSW of S4
S3 (CP)	LG2, BG2, BR2 (16 Sep 2010)	Managed (thinned) stand. Infested by MPB in ~2009., avg. DBH: 14 cm; avg. stem density: 2250 ha ⁻¹	located ~160 m east of S1, avg. distance between S3 trees: 23 m
S4 (CP)	BG3 (17 Sep 2010)	Most heavily-infested stand sampled (site of earliest outbreak at CP starting in ~2007). Mostly contains sparsely-distributed BR trees. Age ≥80 yr, avg. DBH: 25 cm, avg. stem density: 730 ha ⁻¹	S4 located ~240 m ENE of S3,
just outside of S4 (CP)	LG3 (17 Sep 2010)	Age unknown (but probably 50–70 yr), DBH of LG3: ~18 cm; stem density unknown	located 120 m from BG3
(MRS)	(30 Jun–1 Jul 2011, 7–8 Aug 2011, 17–18 Sep 2010)	Avg. DBH = 23 cm, avg. stem density = 1500 ha ⁻¹ . Age distribution is roughly normal with a narrow age span of ~130 yr (Knowles and Grant, 1983)	all MRS trees were growing within 40 m of each other



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Table 3. ER and Temperature (T) ranges observed during the CP and MRS sampling visits; results are grouped according to compound/compound class and (when appropriate) sample class. Avg. Basal ERs (BER) and corresponding standard deviations (σ) are presented for MT and SQT; avg. ER and standard deviations are reported for MBO and OVOC.

Site	Sampling dates	Tree(s) or tree class(es)	Cmpd. or cmpd. class	ER range ($\mu\text{gCg}_{\text{dw}}^{-1}\text{h}^{-1}$)	T range ($^{\circ}\text{C}$)	Avg. BER (σ) if MT or SQT, avg. ER (σ) if MBO or OVOC ($\mu\text{gCg}_{\text{dw}}^{-1}\text{h}^{-1}$)
CP	15–17 Sep 2010	LG, BG	MBO	0–5.38	18.3–35.8	1.16 (1.43)
CP	15–17 Sep 2010	BR	MBO	0–0.001	16.4–20.	0.0007 (0.0005)
CP	15–17 Sep 2010	all	MT	0.08–1.97	16.4–35.8	1.16 (1.47)
CP	15–17 Sep 2010	LG	SQT	0.037–0.235	20.4–35.8	0.165 (0.071)
CP	15–17 Sep 2010	BG	SQT	0.001–0.060	18.3–25.0	0.045 (0.061)
CP	15–17 Sep 2010	BR	SQT	0.001–0.015	16.4–20.0	0.038 (0.038)
MRS	30 Jun–1 Jul 2011	All*	MBO	0.37–13.25	14.2–33.0	4.71 (3.36)
MRS	7–8 Aug 2011	All**	MBO	1.29–19.09	22.9–39.7	8.43 (5.61)
MRS	17–18 Sep 2011	AB1b, OB1	MBO	0.43–0.96	10.5–13.4	0.69 (0.25)
MRS	17–18 Sep 2011	OB2, CT2	MBO	0.37–2.25	15.9–24.0	0.90 (0.69)
MRS	17–18 Sep 2011	AB1, AB2, CT1	MBO	0.03–0.11	12.0–15.7	0.07 (0.03)
MRS	30 Jun–1 Jul 2011	All except CT1	MT	0.05–4.57	14.2–33.0	1.30 (1.26)
MRS	30 Jun–1 Jul 2011	CT1	MT	2.97–11.43	18.9–24.4	15.67 (5.88)
MRS	7–8 Aug 2011	OB, CT	MT	0.17–1.69	22.9–36.6	0.83 (0.32)
MRS	7–8 Aug 2011	AB	MT	0.12–1.42	26.0–39.7	0.40 (0.14)
MRS	17–18 Sep 2011	All, except OB1 & AB1b	MT	0.01–0.67	12.0–24.0	0.36 (0.32)
MRS	17–18 Sep 2011	OB1	MT	0.79–0.92	10.5–12.6	5.41 (0.15)
MRS	17–18 Sep 2011	AB1b	MT	0.10–0.23	11.4–13.4	0.94 (0.40)
MRS	30 Jun–1 Jul 2011	All, except OB2 & CT1	SQT	0.004–0.090	14.2–33.0	0.073 (0.044)
MRS	30 Jun–1 Jul 2011	OB2 & CT1	SQT	0.157–0.340	18.9–26.2	0.707 (0.172)
MRS	7–8 Aug 2011	OB	SQT	0.322–0.992	25.2–33.8	0.932 (0.312)
MRS	7–8 Aug 2011	AB & CT	SQT	0.157–0.760	22.9–39.7	0.260 (0.183)
MRS	17–18 Sep 2011	OB	SQT	0.028–0.151	1.5–24.0	0.434 (0.098)
MRS	17–18 Sep 2011	AB & CT	SQT	0–0.013	11.4–18.0	0.019 (0.023)
MRS	30 Jun–1 Jul 2011	all	OVOC	0.008–0.458	14.2–33.0	0.165 (0.139)
MRS	7–8 Aug 2011	all except AB2	OVOC	0.023–0.439	22.9–39.7	0.161 (0.102)
MRS	7–8 Aug 2011	AB2	OVOC	0.590–3.139	26.0–38.5	1.908 (1.277)
MRS	15–17 Sep 2010	CT1, AB	OVOC	0–0.0004	11.4–15.7	0.0001 (0.0001)
MRS	15–17 Sep 2010	CT2, OB2	OVOC	0.001–0.08	15.9–24.0	0.003 (0.002)
MRS	15–17 Sep 2010	OB1	OVOC	0.007–0.009	10.5–12.6	0.008 (0.001)

* 1 sample from pre-bait.2 was excluded from the reported average ERs (MBO ER = 29.2; T = 28.2).

** 1 sample from pre-bait.2 was excluded from the reported ERs (MBO ER = 27.6; T = 29.0).

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Table 4. Average percent contribution of individual compounds to total observed MT emissions (standard deviations shown in parentheses) at CP and MRS (n.d. = not detected, compounds averaging <0.1% of total MT emissions for all trees sampled at site were excluded), grouped by site and (for CP) sampling class.

Site, trees included, dates, number of samples	tricyclene	α -thujene	α -pinene	camphene	sabinene	β -pinene	β -myrcene	α -phellandrene	3-carene	α -terpinene	δ -limonene	β -phellandrene	γ -terpinene	terpinolene
CP, LG & BG, Sep 2010, 13	0.17 (0.1)	0.16 (0.09)	12.77 (3.36)	2.34 (1.23)	n.d.	50.78 (12.41)	0.93 (0.59)	1.38 (0.62)	10.29 (12.12)	0.85 (0.76)	4.38 (3.11)	14 (5.49)	0.85 (0.9)	1.11 (0.85)
CP, BR, Sep 2010, 4	0.15 (0.12)	0.37 (0.27)	8.29 (3.47)	1.7 (1.38)	n.d.	17.42 (5.72)	2.76 (2.78)	2.56 (1.4)	39.46 (5.69)	1.53 (2.23)	2.76 (0.38)	17.49 (6.93)	0.52 (0.5)	4.99 (5.69)
MRS, BB1, 30 Jun 2011, 3	0.21 (0.06)	0.53 (0.23)	11.61 (2.12)	2.33 (1.19)	1.39 (0.72)	40.53 (2.31)	6.92 (3.37)	0.59 (0.25)	21.07 (6.88)	0.83 (0.01)	2.35 (0.5)	9.62 (12.84)	0.17 (0.15)	1.75 (0.53)
MRS, BB2, 1 Jul 2011, 3	0.21 (0.12)	0.65 (0.13)	8.89 (4.14)	1.03 (0.99)	2.23 (0.45)	17.71 (13.6)	8.41 (1.37)	0.36 (0.09)	39.51 (7.11)	1.59 (0.52)	12.25 (1.4)	n.d.	0.83 (0.33)	6.3 (10.91)
MRS, CT1, 30 Jun 2011, 3	0.48 (0.3)	0.26 (0.07)	4.72 (0.71)	3.2 (1.49)	0.49 (0.02)	21.24 (2.91)	2.95 (0.75)	2.56 (0.57)	8.34 (1.05)	0.67 (0.06)	0.38 (0.67)	53.53 (2.61)	0.18 (0.04)	0.75 (0.08)
MRS, CT2, 1 Jul 2011, 3	0.13 (0.02)	0.4 (0.01)	3.6 (0.82)	0.76 (0.16)	0.71 (0.19)	6.43 (0.23)	3.79 (0.15)	0.88 (0.81)	32.81 (1.82)	1 (0.08)	1.76 (0.06)	45.25 (2.17)	0.49 (0.03)	1.97 (0.25)
MRS, OB1, 30 Jun 2011, 3	0.27 (0.06)	0.55 (0.25)	14.09 (1.33)	2.85 (0.27)	1.58 (0.76)	49.94 (5.18)	6.21 (3.53)	0.36 (0.02)	15.28 (3.23)	1.03 (0.29)	4.34 (0.62)	2.9 (5.03)	0.5 (0.43)	n.d.
MRS, OB2, 1 Jul 2011, 3	0.32 (0.18)	0.41 (0.09)	9.61 (0.99)	2.32 (2.14)	1.54 (1.01)	36.34 (2.18)	4.27 (0.29)	0.85 (0.44)	12.74 (0.68)	0.67 (0.12)	2.48 (0.72)	27.38 (2.27)	0.35 (0.07)	0.6 (0.24)
MRS, AB1, 7 Aug 2011, 3	1.16 (0.22)	0.57 (0.08)	7.33 (0.54)	7.84 (0.67)	0.8 (0.16)	26.59 (0.57)	3.47 (0.33)	0.46 (0.17)	27.28 (0.66)	1.3 (0.2)	4.05 (0.31)	12.13 (0.49)	1.53 (0.39)	4.69 (0.24)
MRS, AB2, 8 Aug 2011, 3	0.7 (0.21)	0.29 (0.02)	4.9 (0.44)	4.85 (0.51)	0.31 (0.13)	12.1 (1.97)	8.68 (4.04)	0.82 (0.14)	29.91 (6.75)	5.94 (1.35)	21.28 (3.37)	1.56 (1.1)	3.86 (3.03)	4.49 (2.24)
MRS, CT1, 7 Aug 2011, 3	1.1 (0.52)	0.31 (0.02)	9.38 (1.07)	6.91 (3.09)	0.14 (0.12)	34.47 (3.41)	1.54 (0.05)	0.34 (0.04)	23.33 (1.26)	0.68 (0.08)	3.65 (0.92)	15.21 (2.17)	0.52 (0.05)	1.79 (0.05)
MRS, CT2, 8 Aug 2011, 2	0.74 (0.78)	0.25 (0.11)	4.6 (1.1)	3.9 (3.96)	0.26 (0.37)	7.49 (5.46)	1.58 (4.95)	5.06 (0.18)	35.6 (9.9)	7.61 (1.61)	2.27 (16.03)	29.14 (0.44)	0.31 (0.4)	0.97 (0.1)
MRS, OB1, 7 Aug 2011, 3	0.36 (0.13)	0.23 (0.08)	9.34 (2.23)	4.04 (1.52)	0.18 (0.32)	35.75 (3.9)	2.34 (1.04)	0.29 (0.03)	19.19 (6.34)	0.49 (0.14)	6 (0.4)	20.09 (2.97)	0.1 (0.18)	1.38 (0.93)
MRS, OB2, 8 Aug 2011, 3	0.43 (0.06)	0.34 (0.03)	8.48 (0.68)	3.33 (0.12)	0.65 (0.14)	29.89 (1.45)	3.95 (0.63)	0.54 (0.04)	25.94 (1.58)	0.83 (0.01)	4.89 (0.09)	18.4 (0.16)	0.48 (0.06)	1.67 (0.24)
MRS, AB1 & AB1b, 17–18 Sep 2011, 5	0.27 (0.11)	0.1 (0.02)	13.07 (2.49)	5.2 (1.45)	n.d.	59.06 (6.82)	0.14 (0.19)	0.33 (0.17)	7.72 (2.56)	0.19 (0.26)	2.22 (0.74)	11.46 (0.81)	n.d.	0.12 (0.08)
MRS, AB2, 18 Sep 2011, 3	n.d. (0.07)	0.08 (0.15)	n.d. (11.84)	n.d. (4.45)	n.d.	n.d. (9.13)	n.d. (42.6)	n.d. (0.88)	n.d. (22.02)	n.d.	n.d. (2.65)	n.d. (1.23)	n.d.	0.69 (0.12)
MRS, CT1, 17 Sep 2011, 3	0.22 (0.07)	0.15 (0.13)	11.84 (2.94)	4.45 (1.85)	n.d.	42.6 (3.17)	0.88 (0.29)	0.46 (0.38)	22.02 (0.5)	n.d.	1.85 (0.29)	14.55 (1.96)	n.d.	0.84 (0.29)
MRS, CT2, 18 Sep 2011, 3	0.18 (0.02)	0.19 (0.03)	5.36 (0.91)	1.8 (0.33)	0.06 (0.05)	9.57 (0.92)	0.8 (0.05)	0.92 (0.1)	22.43 (4.47)	n.d.	3.19 (0.21)	54.85 (2.73)	n.d.	0.64 (0.37)
MRS, OB1, 17 Sep 2011, 3	0.11 (0.03)	0.15 (0.03)	6.61 (0.94)	1.81 (1.18)	0.07 (0.12)	38.01 (5.99)	1.11 (0.16)	0.36 (0.09)	22.99 (3)	0.27 (0.11)	4.65 (1.35)	21.95 (1.02)	0.02 (0.04)	1.84 (0.12)
MRS, OB2, 18 Sep 2011, 3	0.27 (0.15)	0.16 (0.06)	12.29 (5.51)	5.19 (4.12)	0.05 (0.08)	28.33 (24.61)	0.6 (0.23)	0.68 (0.35)	19.47 (3.55)	0.31 (0.29)	4.61 (2.35)	27.42 (10.27)	0.06 (0.11)	0.44 (0.1)

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Table 5. Average percent contribution of individual compounds to total observed SQT emissions (standard deviations shown in parentheses) at CP and MRS (n.d. = not detected). Compounds comprising <0.1 % of total SQT emissions were excluded, asterisk indicates tentative compound identification.

Site, tree, dates, number of samples	α -copaene	β -caryophyllene	α -bergamotene	α -trans-bergamotene	aromadendrene*	c - β -limesene	e - β -limesene	γ -murolene	α -zingiberene*	α -cuprenene	α -limesene	γ -cadinene	δ -cadinene	unidentified _{SQT}
CP, LG1, 15 Sep 2010, 2	n.d.	n.d.	0.07 (0.03)	n.d.	0.36 (0)	n.d.	12.55 (1.47)	n.d.	3.15 (0.13)	n.d.	80.93 (1.96)	0.57 (0.25)	0.65 (0.14)	1.72 (0.19)
CP, LG2, 16 Sep 2010, 2	n.d.	0.08 (0.12)	2.93 (1.97)	n.d.	n.d.	n.d.	53.78 (8.52)	n.d.	0.7 (0.16)	n.d.	37.92 (5.38)	n.d.	n.d.	4.59 (1.12)
CP, LG3, 17 Sep 2010, 2	0.85 (0.13)	n.d.	0.06 (0)	5.12 (0.69)	n.d.	n.d.	2.5 (0.29)	n.d.	n.d.	n.d.	75.67 (3.74)	6.83 (1.29)	3.89 (1.14)	5.07 (0.19)
CP, BG1, 15 Sep 2010, 2	n.d.	n.d.	2.71 (0.1)	n.d.	n.d.	n.d.	40.76 (5.15)	n.d.	4.96 (1.12)	n.d.	2.82 (3.98)	13.9 (6.68)	34.35 (4.07)	n.d. 0.53
CP, BG2, 16 Sep 2010, 3	n.d.	0.89 (1.54)	1.23 (0.31)	24.69 (10.99)	n.d.	n.d.	23.26 (6.48)	n.d.	n.d.	13.32 (13.43)	6.55 (0.11)	15.46 (13.14)	11.99 (10.93)	0.91 (0.31)
CP, BG3, 17 Sep 2010, 2	1.23 (0.87)	n.d.	0.15 (0.01)	2.7 (0.23)	n.d.	n.d.	3.18 (0.04)	n.d.	n.d.	n.d.	64.27 (6.62)	13.14 (2.86)	12.71 (2.96)	2.61 (0.18)
CP, BR1, 15 Sep 2010, 2	n.d.	n.d.	1.4 (0.59)	n.d.	1.77 (0.79)	n.d.	0.95 (1.35)	n.d.	n.d.	n.d.	64.27 (0.7)	43.33 (6.39)	52.34 (1.49)	0.19 (0.26)
CP, BR2, 16 Sep 2010, 2	n.d.	2.39 (0.4)	0.44 (0.62)	n.d.	37.07 (0.15)	n.d.	25.63 (7.21)	n.d.	n.d.	27.35 (0.7)	4.52 (6.39)	2.47 (2.14)	2.38 (1.49)	n.d.
MRS, BB1, 30 Jun 2011, 3	0.92 (0.84)	n.d.	9.16 (1)	1.46 (1.3)	53.09 (0.71)	4.79 (1.68)	0.79 (0.41)	0.8 (0.38)	14.26 (3.11)	14.26 (1.26)	4.38 (1.39)	2.14 (1.39)	8.22 (1.47)	0.92 (0.84)
MRS, BB2, 1 Jul 2011, 3	n.d.	0.47 (0.66)	n.d.	3.98 (2.75)	0.25 (0.26)	34.16 (19.66)	0.88 (1)	0.16 (0.13)	n.d.	57.19 (16.15)	0.31 (0.17)	0.08 (0.05)	2.52 (0.87)	0.47 (0.66)
MRS, CT1, 30 Jun 2011, 3	n.d.	0.02 (0.03)	n.d.	14.27 (0.75)	n.d.	67.01 (1.68)	n.d.	0.82 (0.09)	0.52 (0.08)	9.01 (1.54)	3.5 (0.37)	2.48 (0.24)	2.37 (0.17)	0.02 (0.03)
MRS, CT2, 1 Jul 2011, 3	n.d.	n.d.	n.d.	15.45 (4.06)	2.02 (1.88)	37.74 (1.57)	n.d.	2.3 (1.04)	2.39 (0.99)	24.18 (14.03)	7.66 (3.13)	6.01 (1.78)	2.26 (0.12)	n.d.
MRS, OB1, 30 Jun 2011, 3	n.d.	n.d.	n.d.	7.18 (2.27)	1.3 (1.14)	46.56 (12.78)	0.98 (0.96)	1.01 (0.48)	n.d.	24.3 (22.97)	4.79 (3)	4.09 (1.39)	9.79 (2.76)	n.d.
MRS, OB2, 1 Jul 2011, 3	n.d.	n.d.	n.d.	13.66 (1.45)	0.08 (0.07)	45.6 (4.62)	n.d.	0.26 (0.04)	0.31 (0.06)	30.79 (5.94)	1.17 (0.12)	0.82 (0.08)	7.31 (0.05)	n.d.
MRS, AB1, 7 Aug 2011, 3	n.d.	0.77 (0.08)	n.d.	6.94 (0.34)	0.49 (0.11)	55.51 (2.31)	n.d.	0.34 (0.07)	0.34 (0.04)	24.66 (1.19)	1.02 (0.07)	0.74 (0.09)	9.19 (2.33)	0.77 (0.08)
MRS, AB2, 8 Aug 2011, 3	n.d.	0.17 (0.15)	n.d.	2.77 (0.4)	0.27 (0.35)	46.4 (9.51)	n.d.	0.12 (0.01)	0.13 (0.1)	47.67 (9.2)	0.23 (0.02)	0.34 (0.03)	1.37 (0.33)	0.17 (0.15)
MRS, CT1, 7 Aug 2011, 3	n.d.	0.67 (0.2)	n.d.	6.58 (2.22)	0.17 (0.07)	54.79 (2.75)	n.d.	1.87 (0.46)	1.94 (0.67)	8.66 (0.9)	8.01 (1.87)	6.18 (1.41)	11.14 (2.13)	0.67 (0.2)
MRS, CT2, 8 Aug 2011, 2	n.d.	n.d.	n.d.	1.46 (0.31)	0.17 (0.24)	46.95 (10.43)	n.d.	0.26 (0.22)	0.37 (0.24)	47.22 (13.38)	1.2 (0.98)	1.16 (0.82)	1.22 (0.15)	n.d.
MRS, OB1, 7 Aug 2011, 3	n.d.	n.d.	n.d.	1.88 (0.49)	n.d.	39.28 (8.42)	n.d.	0.13 (0.06)	0.09 (0.08)	51.52 (9.35)	0.5 (0.16)	0.35 (0.11)	6.25 (0.54)	n.d.
MRS, OB2, 8 Aug 2011, 3	n.d.	n.d.	n.d.	6.63 (0.26)	n.d.	39.43 (4.48)	n.d.	0.18 (0.06)	0.21 (0.08)	44.82 (4.45)	0.62 (0.15)	0.45 (0.06)	7.66 (0.61)	n.d.
MRS, AB1 & AB1b, 17–18 Sep 2011, 5	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	80 (54.77)	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
MRS, AB2, 18 Sep 2011, 3	n.d.	n.d.	n.d.	7.39 (1.53)	n.d.	92.61 (1.53)	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
MRS, CT1, 17 Sep 2011, 3	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	3.01 (5.21)	6.12 (10.59)	n.d.	44.56 (8.52)	46.32 (10.22)	n.d.	n.d.
MRS, CT2, 18 Sep 2011, 3	n.d.	n.d.	n.d.	0.65 (0.13)	0.72 (1.24)	74.33 (4.1)	n.d.	0.62 (0.16)	1.48 (0.52)	15.4 (6.03)	2.6 (0.4)	4.19 (1.45)	n.d.	n.d.
MRS, OB1, 17 Sep 2011, 3	n.d.	n.d.	n.d.	4.44 (0.71)	0.99 (0.37)	78 (11.01)	n.d.	n.d.	n.d.	11.51 (11.89)	0.88 (0.16)	1.36 (0.19)	2.82 (0.37)	n.d.
MRS, OB2, 18 Sep 2011, 3	n.d.	n.d.	n.d.	2.89 (0.49)	0.08 (0.13)	65.96 (5.37)	n.d.	0.12 (0.04)	0.16 (0.01)	28.65 (5.25)	0.54 (0.1)	0.67 (0.29)	0.92 (0.06)	n.d.

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Table 6. Average percent contribution of individual compounds to total observed OVOC emissions (standard deviations shown in parentheses) at MRS (n.d. = not detected), asterisk indicates tentative compound identification.

Site, tree, dates, number of samples	amyl acetate*	eucalyptol	linalool	camphor	pinocarvone*	methyl salicylate	cumin aldehyde*	piperitone*
MRS, BB1, 30 Jun 2011, 3	0.2 (0.18)	94.73 (0.87)	1.46 (1.47)	0.28 (0.21)	1.54 (0.21)	1.61 (1.76)	0.1 (0.09)	0.09 (0.09)
MRS, BB2, 1 Jul 2011, 3	0.22 (0.07)	32.73 (7.54)	63.74 (9.9)	0.74 (0.54)	1.62 (0.82)	0.7 (0.71)	0.03 (0.03)	0.23 (0.23)
MRS, CT1, 30 Jun 2011, 3	3.56 (0.63)	n.d.	2.21 (3.83)	6.08 (2.41)	27.63 (8.13)	49.55 (9.05)	7.89 (0.88)	3.08 (2.73)
MRS, CT2, 1 Jul 2011, 3	67.73 (0.93)	17.99 (3.65)	6.54 (3.19)	1.3 (0.31)	0.63 (0.07)	5.19 (0.18)	0.22 (0.05)	0.42 (0.07)
MRS, OB1, 30 Jun 2011, 3	43.98 (9.19)	51.23 (9.52)	n.d.	2.98 (0.5)	0.62 (0.13)	1.19 (0.14)	n.d.	n.d.
MRS, OB2, 1 Jul 2011, 3	0.75 (0.31)	93.15 (0.81)	0.44 (0.18)	0.82 (0.17)	1.85 (0.7)	2.14 (0.35)	0.26 (0.14)	0.59 (0.13)
MRS, AB1, 7 Aug 2011, 3	9.7 (5.9)	6.21 (2.01)	76.24 (8.58)	1.19 (0.51)	4.72 (2.8)	1.16 (0.77)	n.d.	0.78 (0.61)
MRS, AB2, 8 Aug 2011, 3	0.05 (0.05)	0.09 (0)	97.84 (1)	0.38 (0.11)	0.23 (0.4)	1.32 (0.59)	n.d.	0.08 (0.07)
MRS, CT1, 7 Aug 2011, 3	2.44 (0.64)	n.d.	81.2 (8.08)	2.69 (1.08)	6.42 (2.99)	6.85 (2.74)	0.19 (0.34)	0.21 (0.37)
MRS, CT2, 8 Aug 2011, 2	61.44 (5.27)	5.52 (4.26)	n.d.	1.76 (1.1)	n.d.	31.27 (0.09)	n.d.	n.d.
MRS, OB1, 7 Aug 2011, 3	85.8 (1.62)	5.57 (1.3)	0.66 (0.58)	3.05 (0.4)	0.9 (0.19)	4.03 (0.18)	n.d.	n.d.
MRS, OB2, 8 Aug 2011, 3	10.47 (2.41)	46.39 (1.16)	29.54 (3.47)	2.76 (0.43)	3.47 (0.49)	5.17 (2.15)	0.29 (0.25)	1.91 (0.55)
MRS, AB1 & AB1b, 17–18 Sep 2011, 5	n.d.	6.32 (14.14)	n.d.	93.68 (14.14)	n.d.	n.d.	n.d.	n.d.
MRS, AB2, 18 Sep 2011, 3	n.d.	n.d.	n.d.	100.00	n.d.	n.d.	n.d.	n.d.
MRS, CT1, 17 Sep 2011, 3	n.d.	n.d.	n.d.	66.67 (57.74)	n.d.	n.d.	n.d.	n.d.
MRS, CT2, 18 Sep 2011, 3	15.56 (26.96)	9.72 (16.83)	n.d.	74.72 (23.58)	n.d.	n.d.	n.d.	n.d.
MRS, OB1, 17 Sep 2011, 3	49.7 (2.43)	21.48 (3.7)	n.d.	27.11 (1.32)	1.7 (2.95)	n.d.	n.d.	n.d.
MRS, OB2, 18 Sep 2011, 3	n.d.	76.21 (2.53)	n.d.	21.23 (2.41)	2.56 (0.5)	n.d.	n.d.	n.d.

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Table 7. Two-sided *t*-tests results (reported as *p*-values) between individual tree MT and SQT BERs during the first, second, and third sampling visits at MRS, and between sampling classes at CP (bold lettering indicates significance at the 95 % confidence level). The BR sample class at CP as well as August emissions from tree CT-2 at MRS were excluded from statistical analyses due to small sample sizes.

MRS 30 Jun–1 Jul MT						MRS 30 Jun–1 Jul SQT					
tree code	BB2	CT1	CT2	OB1	OB2	tree code	BB2	CT1	CT2	OB1	OB2
BB1	0.7775	0.04641	0.006056	0.2778	0.03485	BB1	0.03432	2.10E-05	0.05103	0.0694	0.04678
BB2		0.04562	0.007741	0.8456	0.03581	BB2		0.00011	0.2163	0.1222	0.02161
CT1			0.01918	0.04581	0.01957	CT1			3.54E-05	2.78E-05	0.3808
CT2				0.004998	0.925	CT2				0.5278	0.01531
OB1					0.02972	OB1					0.01406

MRS 7–8 Aug MT					MRS 7–8 Aug SQT						
tree code	AB2	CT1	CT2	OB1	OB2	tree code	AB2	CT1	CT2	OB1	OB2
AB1	0.8991	0.00986		0.3523	0.003709	AB1	0.132	9.62E-05		0.04251	0.04514
AB2		0.0406		0.3494	0.0293	AB2		0.3302		0.04597	0.08523
CT1				0.6696	0.9846	CT1				0.05665	0.06575
OB1					0.6561	OB1					0.4565

MRS 17–18 Sep 2011 MT						MRS 17–18 Sep SQT					
tree code	AB2	CT1	CT2	OB1	OB2	tree code	AB2	CT1	CT2	OB1	OB2
AB1	0.04071	0.9706	0.0726	4.54E-07	0.1069	AB1	0.09668	0.01253	0.0496	0.01746	0.002582
AB2		0.1572	0.02322	5.56E-07	0.03098	AB2		0.02455	0.0209	0.01776	4.47E-05
CT1			0.1235	9.08E-07	0.05372	CT1			0.04349	0.01721	0.002536
CT2				1.63E-06	0.1342	CT2				0.00273	0.0001341
OB1					4.77E-05	OB1					0.1577

CP 15–17 Sep 2010											
MT BER			MT fraction			SQT BER			SQT fraction		
tree code	BG	BR	tree code	BG	BR	tree code	BG	BR	tree code	BG	BR
LG	0.7853		LG	0.2438		LG	0.09836		LG	0.03284	

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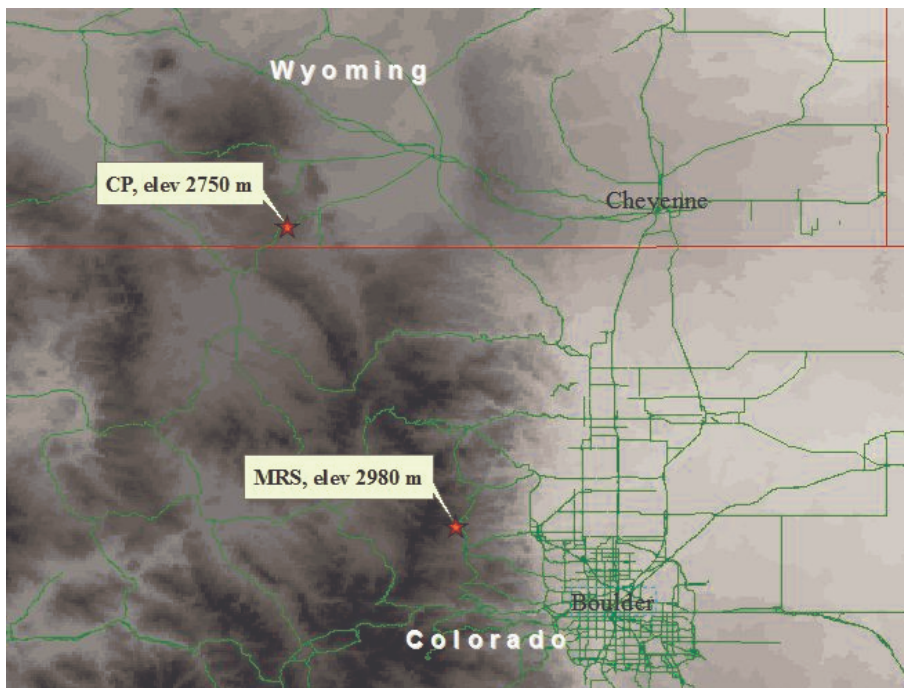


Fig. 1. Location of the Chimney Park and Mountain Research Station sampling sites.

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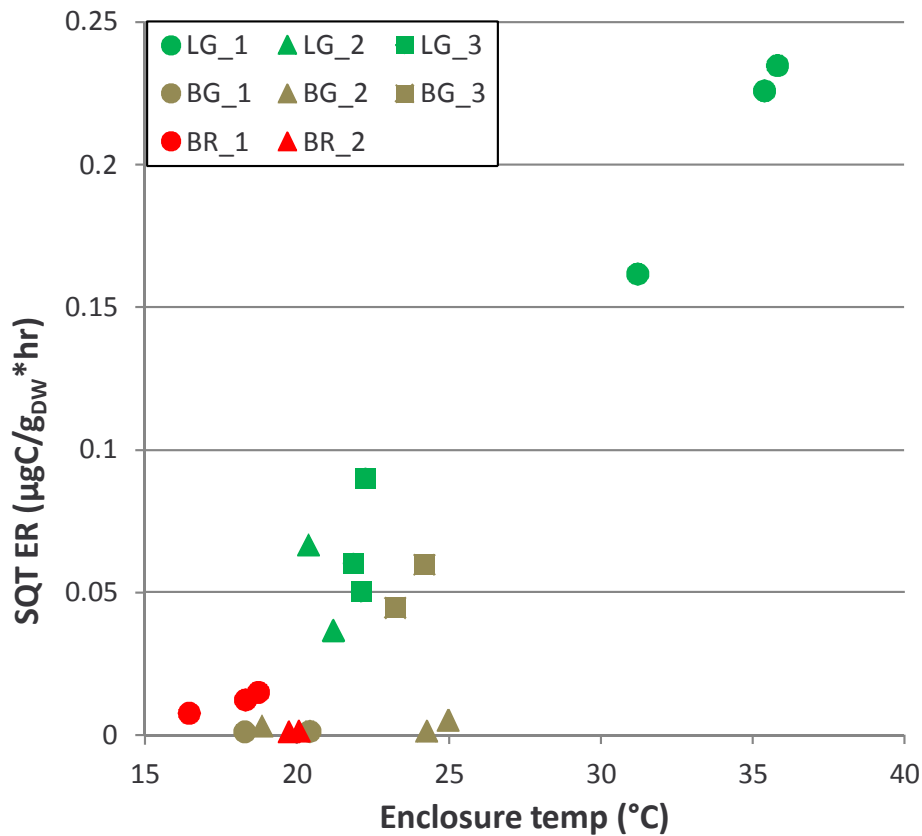


Fig. 2. SQT emissions observed at Chimney Park, WY during mid-September, 2010.

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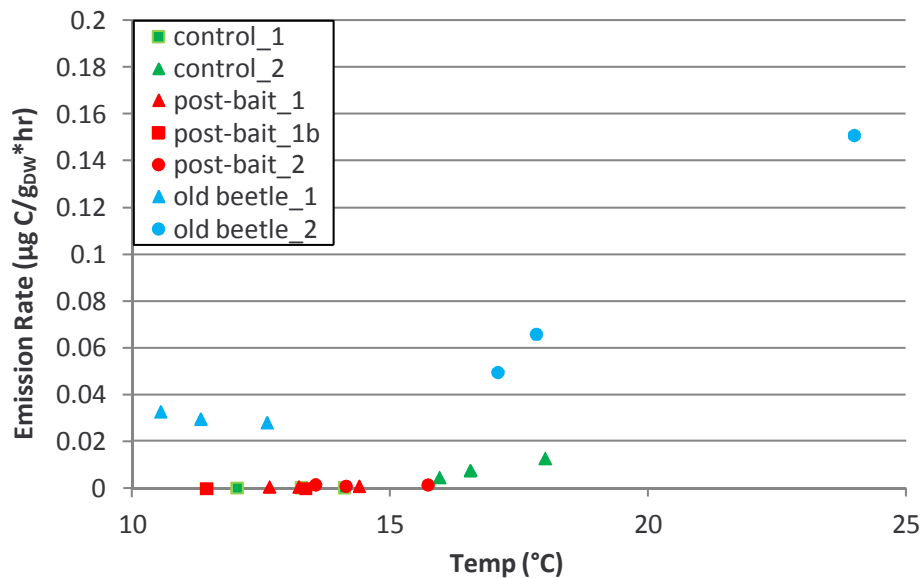


Fig. 4. SQT emissions observed during the last MRS sampling visit (17–18 September), plotted as a function of temperature.

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