#### **Response to Comments**

### Comments are in black font and our responses are in italicized font.

#### **Associate Editor**

Dear Authors.

Even without looking in detail at the main manuscript, more work is needed. I would start with the Table and Figures. Normally these are self-explanatory or mostly, some of yours lack this. These are my suggestions for improvement.

Thank you very much for your helpful suggestions. We have revised the manuscript accordingly.

Table 1 I would not put the 14 sites in alphabetically order, but select a parameter relevant to the explaining the observations in your work, e.g. you could list them in order of low to high precipitation or temperature or the other way.

Based on the correlation in Fig. 3, we have re-ordered them according to latitude. Please see Table 1 for revision.

Fig 1. The placing of the locations and displayed parameters on the USA map is nice, but why do I need to look at them on a map? Is there more C up North more bound Fe-C, maybe indicate what it shows with a view arrows .... is it related to T or precipitation, see comment Table 1 maybe this is the parameter to list Table 1 under.

As similar as for Table 1, there is generally higher TC and TOC in soils with higher latitude. We feel it is good to keep them in the map. We revised the figure caption accordingly to emphasize the trend.

Fig 2 what relationship are shown, if the are there they are not obvious. If there are no relationship, why put in a figure at all.

Fig. 2A shows the contents of reactive iron and OC:Fe molar ratio. Fig. 2B shows the variations in OC:Fe, discussed in the manuscript. Fig. 2B was moved to the supplementary material.

Fig 3 Do we need all the show subfigures? I would prefer a selection of the most pertinent examples which link to the text, with the rest going to supplementary material.

Partial of Fig. 3 was moved to supplementary material.

Fig 4 This is very poor Figure, messy and unclear it gives a bad impression of overall standard of the manuscript, it lets your paper down.

This figure has been improved accordingly. The relationships between labile carbon/uncalibrated Fe-bound organic carbon and texture were kept. Relationship between Fe-bound organic carbon and texture was moved to supplementary material, as there is no significant correlation.

# Fig 5 see comment Fig 3

We kept the spectra for aliphatic carbon in the manuscript, and moved other spectra to supplementary material.

Fig 6. A lot of information but no self-evident message from both Figures, please condense the information more and better.

This figure has been improved by keeping the  $^{13}C$  for Fe-bound OC and non-Fe-bound OC in the manuscript, and moving other data to the supplementary material. Now, it clearly showed the enrichment of  $^{13}C$  in Fe-bound OC.

# 1 Iron-Bound Organic Carbon in Forest Soils: Quantification

# 2 and Characterization

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#### ABSTRACT

Iron oxide minerals play an important role in stabilizing organic carbon (OC) and regulating the 13 biogeochemical cycles of OC on the earth surface. To predict the fate of OC, it is essential to 14 understand the amount, spatial variability, and characteristics of Fe-bound OC in natural soils. In 15 this study, we investigated the concentrations and characteristics of Fe-bound OC in soils 16 collected from 14 forests in the United States, and determined the impact of ecogeographical 17 variables and soil physicochemical properties on the association of OC and Fe minerals. On 18 19 average, Fe-bound OC contributed 37.8% of total OC (TOC) in forest soils. Atomic ratios of OC:Fe ranged from 0.56 to 17.7 with values of 1-10 for most samples, and the ratios indicate the 20 importance of both sorptive and incorporative interactions. The fraction of Fe-bound OC in TOC 21  $(f_{\text{Fe-OC}})$  was not related to the concentration of reactive Fe, which suggests that the importance of 22 23 association with Fe in OC accumulation was not governed by the concentration of reactive Fe. Concentrations of Fe-bound OC and f<sub>Fe-OC</sub> increased with latitude and reached peak values at a 24 site with a mean annual temperature of 6.6 °C. Attenuated total reflectance-Fourier transform 25 infrared spectroscopy (ATR-FTIR) and near-edge X-ray absorption fine structure (NEXAFS) 26 analyses revealed that Fe-bound OC was less aliphatic than non-Fe-bound OC. Fe-bound OC 27

also was more enriched in  $^{13}$ C compared to the non-Fe-bound OC, but C/N ratios did not differ substantially. In summary,  $^{13}$ C-enriched OC with less aliphatic carbon and more carboxylic carbon was associated with Fe minerals in the soils, with values of  $f_{\text{Fe-OC}}$  being controlled by both sorptive and incorporative associations between Fe and OC. Overall, this study demonstrates that Fe oxides play an important role in regulating the biogeochemical cycles of C in forest soils, and uncovers the governing factors for the spatial variability and characteristics of Fe-bound OC.

# 1 Introduction

Soil organic carbon (OC) in forests is a vital component of C biogeochemical cycles (Eswaran et al., 1999). Global warming can potentially accelerate the decomposition of forest soil OC, contributing to greenhouse gas emissions (Steffen et al., 1998). Alternatively, forest soils can act as strong sinks for OC, if appropriate management is implemented, such as forest harvesting and fire treatment (Eswaran et al., 1999; Johnson and Curtis, 2001). Understanding the fate and stability of forest OC is important for evaluating and managing the global C cycle under the framework of climate change.

Currently, there is an information gap concerning the stability and residence time of OC, contributing to the problem that the residence time of OC (ranging from months to hundreds of years) is a major source of uncertainty in modeling and prediction of C cycles (Schmidt et al., 2011; Riley et al., 2014). Many concepts have been proposed to account for OC stabilization and therefore residence times, including molecular recalcitrance, physical occlusion, and chemical protection (Sollins et al., 1996; Krull et al., 2003; Baldock et al., 2004; Mayer et al., 2004; Zimmerman et al., 2004; Schmidt et al., 2011). In general, the stability of OC is regulated by biogeochemical reactions occurring at the interfaces between OC, minerals, and microorganisms, and further knowledge about the mechanism for OC stabilization is critical for building up process-based models to simulate and predict C cycles.

A number of lines of evidence suggest a key importance of iron oxide minerals in the stabilization of OC (Kalbitz et al., 2005; Kaiser and Guggenberger, 2007; Wagai and Mayer, 2007). Iron oxides have a relatively high sorption capacity for OC, with sorption coefficients for OC much higher than that of other metal oxides (Kaiser and Guggenberger, 2007; Chorover and

Amistadi, 2001). Wagai and Mayer (2007) reported Fe-bound OC concentrations in soils up to 22 mg g<sup>-1</sup> soil, contributing up to 40% of total OC (TOC) for most forest soils. Similarly, Lalonde et al. (2012) found that Fe-bound OC contributed 22% of TOC in sediments. Studies have shown that Fe minerals protect OC from degradation and inhibit mineralization of OC (Baldock and Skjemstad, 2000; Kalbitz et al., 2005). There is, however, no systematic study on the occurrence of Fe-bound OC across different forests and its governing factors.

The overall goals of this study were to investigate the spatial variability of Fe-bound OC across forest soils, the factors that control Fe-bound OC concentrations, and the characteristics of Fe-bound OC with respect to the physicochemical properties of soils. In this study, we first quantified the concentration of Fe-bound OC across 14 forest soils in the United States and analyzed the spatial distribution and influences of ecogeographical factors. Second, we investigated the impact of soil physicochemical properties on the Fe-OC associations. Third, we studied molecular characteristics of Fe-bound OC vs. non-Fe-bound OC, including how Fe-OC association influenced the chemical properties of OC and the stable isotope composition. Hence, this study provided a systematic evaluation for the Fe-bound OC in United States forests, the influences of ecological factors on the occurrence of Fe-bound OC, and the effects of association with Fe on the chemical properties of OC.

#### 2. Methods & Materials

### 2.1 Chemicals and materials

Reagents used for Fe reduction experiments include sodium bicarbonate (NaHCO<sub>3</sub>: Sigma-Aldrich, St. Louis, MO, USA), trisodium citrate dihydrate (Na<sub>3</sub>C<sub>6</sub>H<sub>5</sub>O<sub>7</sub>•2H<sub>2</sub>O: Acros Organics, New Jersey, USA), and sodium dithionite (Na<sub>2</sub>S<sub>2</sub>O<sub>4</sub>: Alfa Aesar, Ward Hill, MA, USA). All chemicals used were analytical grade.

#### 2.2 Soil sample collection, primary characterization and pretreatment

Soil samples were collected from 14 forest sites in the United States (Obrist et al., 2011, 2012, 2015). The abbreviations and the basic information for the sites are summarized in Table 1. More detailed information on the sites and sampling protocols can be found in previous publications (Obrist et al., 2011, 2012, 2015). Briefly, two replicate plots at each forest site were

sampled. During 2007-2009, top soils (0-20 cm) from all sites were collected using clean latex gloves and stainless steel sampling equipment. All the samples were immediately transferred to plastic freezer bags and kept on ice before transportation to the laboratory. Soil texture was analyzed by an ASTM 152-type hydrometer at the Soil Forage and Water Analysis Laboratory at Oklahoma State University (Obrist et al., 2011). The soil pH was measured by mixing soil particle with deionized (DI) water in a solid/solution ratio of 1:1 (Kalra, 1995). Soil samples used in the experiments in this study were ground to < 500 µm and freeze-dried after the removal of roots and visible plant material and large particles (>2 mm) by dry sieving.

#### Table 1

# 2.3 Total C (TC), TOC and stable C isotope analyses

TC, TOC and stable C isotopic compositions of soil samples were analyzed using a Eurovector elemental analyzer (Eurovector SPA, Milan, Italy) interfaced to a Micromass IsoPrime stable isotope ratio mass spectrometer (Micromass UK Ltd., Manchester, UK). Acetanilide (71.09 % C by weight) was used as a standard compound to establish a calibration curve between mass of C and the m/z 44 response from the mass spectrometer. In this study, the concentration of TC and TOC were expressed as weight %. Stable C isotope analyses were performed after the method of Werner et al. (1999), with results reported in the usual delta notation in units of ‰ vs. Vienna Pee Dee Belemnite (VPDB). For TOC analysis, soil samples were acidified with 1 M HCl with the solution/solid ratio of 1 mL solution/0.5 g soil and heated at 100°C for 1 hour. The treatment was repeated three times until there was no further effervescence upon acid addition, after which the samples were dried and analyzed. All analyses are based on standard curves with  $R^2$ >0.99. The detection limit for C is 0.2 mg g<sup>-1</sup> soil. The average coefficient of variation for the analysis of C is 20.2%.

### 2.4 Nitrogen (N) analysis

The N concentration of each sample was analyzed using a Eurovector elemental analyzer. Acetanilide (10.36 % N by weight) was used as a standard compound to establish a calibration curve between mass of N and the response of the thermal conductivity detector in the elemental analyzer. Total N and non-Fe-bound N concentrations were measured before and after a Fe

reduction release treatment for each sample. All analyses are based on standard curves with  $R^2>0.99$ . The detection limit for N is 0.2 mg g<sup>-1</sup> soil. The average coefficient of variation for the analysis of N is 20.5%.

# 2.5 Analysis of Fe-bound OC

The concentration of Fe-bound OC was quantified by an established Fe reduction release method, commonly known as DCB extraction involving sodium dithionite, citrate and bicarbonate (Mehra and Jackson, 1960; Wagai and Mayer, 2007; Lalonde et al., 2012). The DCB extraction is assumed to extract most free Fe oxides (i.e. goethite, hematite, ferrihydrite and others) existing in soils, but should not extract structural Fe in clay minerals (Mehra and Jackson, 1960; Wagai and Mayer et al., 2007; Lalonde et al., 2012). In this study, we followed the specific protocol detailed in Lalonde et al. (2012). An aliquot (0.25 g) of soil was mixed with 15 mL of buffer solution at pH 7 (containing 0.11 M bicarbonate and 0.27 M trisodium citrate), and then heated to  $80^{\circ}$ C in a water bath. The reducing agent sodium dithionite was added to the samples with final concentration of 0.1 M, and maintained at  $80^{\circ}$ C for 15 min. The samples were then centrifuged at 10,000 rpm for 10 min, the supernatant was removed, and the residual particles were rinsed using 5 mL of DI water. The rinse/centrifuge process was performed three times. The residual particles were freeze-dried and analyzed for TC and TOC concentrations and  $\delta^{13}$ C composition. The mass of residual particles was used to calculate the OC concentration associated with non-Fe minerals.

The background release of OC during the heating process was measured following the method in Lalonde et al. (2012), where sodium citrate and dithionite were replaced by sodium chloride with the same ionic strength. An aliquot (0.25 g) of dry soil was mixed with 15 mL of 1.6 M NaCl and 0.11 M NaHCO<sub>3</sub>, and heated to 80°C. Then 0.22 g of NaCl was added, and the solution was maintained at 80°C for 15 min. The samples were then centrifuged at 10,000 rpm and rinsed three times, and freeze-dried before analysis. The mass of residual particles was used to calculate the concentration of OC released by heating to 80°C. In preliminary experiments, we found that the solution pH increased rapidly during the heating-extraction process with bicarbonate and sodium chloride only, and the increased pH values facilitated the release of additional OC. Hence, we used a lower initial pH of 6 to compensate for the shift to higher pH during heating. To validate the measurement for the concentration of OC released during heating,

we also tested the release of OC using a phosphate buffer (same ionic strength) in lieu of the bicarbonate buffer, which can maintain a pH of 7 during heating. Our results showed that the concentration of OC released was similar for both the bicarbonate and phosphate buffer extraction reactions (Supplementary Material, Fig. S1).

#### 2.6 Quantification of reactive Fe

The concentration of reactive Fe in soils was determined by analyzing the Fe released during the DCB reduction process. After the reduction treatment, the supernatant of each sample was filtered using a 0.2  $\mu$ m syringe filter (cellulose acetate), and analyzed for Fe concentration by inductively coupled plasma - atomic emission spectroscopy (Varian-Vista AX CCD, Palo Alto, CA, USA) at an optical absorption wavelength of 259.9 nm. All analyses are based on standard curves with  $R^2>0.99$ . The detection limit for Fe is 0.04 mg g<sup>-1</sup> soil. The average coefficient of variation for the analysis of Fe is 25.8%.

# 2.7 Attenuated total reflectance-Fourier transform infrared spectroscopy (ATR-FTIR)

ATR-FTIR analysis to characterize the molecular composition of OC was performed for original soil samples and residual soils after DCB extraction using a Thermo Scientific Nicolet 6700 FTIR (Waltham, MA). Dry soil samples were placed directly on the crystal and forced to contact well with the crystal. Spectra were acquired at the resolution of 4 cm<sup>-1</sup> based on 100 scans. Data collection and baseline correction were accomplished using OMNIC software version 8.3.103.

# 2.8 Near-edge X-ray absorption fine structure (NEXAFS) analysis

For further characterization of chemical structure of OM, carbon (1s) K-edge NEXAFS analyses were performed for select soil samples, i.e. for soils with the highest and lowest values of the fraction of Fe-bound OC to TOC. The soil particles were suspended in DI water and deposited on an Au-coated silicon wafer attached to a Cu sample holder. Before analysis, samples were dried in a vacuum desiccator. The X-ray-based experiments were performed on the Spherical Grating Monochromator (SGM) beamline at the Canadian Light Source (Saskatoon, Canada) (Regier et al., 2007). The energy scale was calibrated using citric acid (absorption at 288.6 eV). Major technical parameters and set-up for the beamline include: X-ray energy ranges

250-2000 eV; 45 mm planer undulator; 1000 μm×100 μm spot size; silicon drift detectors (SDD); a titanium filter before the sample; entrance and exit slit gaps of 249.9 μm and 25 μm (Gillespie et al., 2015). Carbon 1s spectra were acquired by slew scans from 270 to 320 eV at 20 s dwell time and 20 scans per sample on a new spot. For data normalization, I<sub>0</sub> was collected by measuring the scatter of the incident beam from a freshly Au-coated Si wafer using SDD. Before the I<sub>0</sub> normalization, the pre-edge baseline was adjusted to near zero to remove the scatter in the sample data (Gillespie et al., 2015).

#### 3. Results and Discussion

#### 3.1 Concentration of Fe-bound OC

This study covered five major forest types in North America, including Spruce-Fir, Pine, Oak, Chaparral, and Maple-beech-birch forests distributed between 29° and 47° N. For the 14 forest soils, TC concentrations ranged between  $1.5\pm0.1$  and  $8.3\pm2.1\%$  (all percentages given are weight-based), and TOC concentrations ranged between  $1.3\pm0.3$  and  $6.2\pm2.9\%$ , which are comparable to values previously reported for North American forest soils (Wagai and Mayer, 2007; Wilson et al., 2013). Bicarbonate extraction-calibrated Fe-bound OC concentrations ranged from 0.3 to 1.9%, with the fraction of Fe-bound OC to TOC ( $f_{\text{Fe-OC}}$ ) averaging 37.8 $\pm20.0\%$  (Fig. 1, Supplementary Material, Table S1). Forest HL (Maine) had the highest  $f_{\text{Fe-OC}}$  of 57.8%, while forests GS (Florida) and OR (Tennessee) had  $f_{\text{Fe-OC}}$  values below detection limits (i.e., below 0.6%). Based on an estimate that 1502 Pg (Pg=1×10<sup>15</sup> g) of TOC is stored in terrestrial soils (Scharlemann, et al., 2014), scaling up these results to a global estimate would yield 538.5 $\pm271.5$  Pg of Fe-bound OC residing in terrestrial soils.

#### Fig. 1

#### 3.2 Fe-OC association

The values of  $f_{\text{Fe-OC}}$  were influenced not only by the concentration of reactive Fe, but also by the type of association between Fe and OC. In this study, the concentration of reactive Fe in forest soils ranged from 0.1 mg g<sup>-1</sup> to 19.3 mg g<sup>-1</sup>, which is low compared to values of reactive Fe of up to 180 mg g<sup>-1</sup> reported previously (Wagai and Mayer, 2007; Wagai et al., 2013) (Fig. 2). A Mollisol in forest site MS (California) had the highest concentration of reactive Fe, while a

Spodosol in forest site GS (Florida) had the lowest reactive Fe concentration. There was no significant correlation between  $f_{\text{Fe-OC}}$  and the concentration of reactive Fe (Pearson Correlation Coefficient r=-0.418, p=0.137, Supplementary Material, Fig. S2). This suggests that the proportion of Fe-bound OC is not strongly controlled by the reactive Fe concentration.

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#### **Fig. 2.**

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The OC:Fe molar ratio ranged from 0.56 to 17.7 for all 14 soils, with a value between 1 and 10 for 10 soils (Fig. 2). Previous studies have suggested that the OC:Fe molar ratio can be used as an indicator for the type of association between Fe oxides and OC, with lower values indicating sorptive interactions while higher values indicate incorporation of OC within Fe oxides (Wagai et al., 2007; Guggenberger and Kaiser, 2003). The highest sorption capacity measured for OC onto Fe oxide corresponds to an OC:Fe molar ratio = 1.0 (Kaiser and Guggenberger, 2006), but by incorporation and co-precipitation of Fe oxide OC:Fe molar ratio can reach much higher values (Guggenberger and Kaiser, 2003). With OC:Fe molar ratios generally between 1-10 for about two thirds of the forest soils in this study, we propose that incorporation of OC into Fe oxides plays a major role in the accumulation of Fe-bound OC exceeding sorption by at least a factor of 1 to almost 20 (Wagai and Mayer, 2007; Lalonde, 2012). However, for the HT (Michigan), HL (Maine) and TKF (California) forest soils, the OC:Fe molar ratios were even higher than 10 with a maximum value of 17.8 (Fig. 2), implying that incorporation of OC into Fe oxides dominated at these sites. Similar to  $f_{\text{Fe-OC}}$ , OC:Fe ratios were not related to the concentration of reactive Fe and showed large variation for soils with similar concentration of total reactive Fe (Supplementary Material, Fig. S2). This further indicates that the type of interactions between OC and Fe was not governed by the amount of Fe. The OC:Fe ratio is potentially regulated by the mineral phases of Fe, as poorly-crystalline Fe oxides have a higher capacity to bind with OC than crystalline Fe minerals (Eusterhues et al., 2014). When sorption dominates the interactions between OC and Fe, OC:Fe can also be influenced greatly by the particle size and surface area of Fe oxides (Gu et al., 1995). Further investigations are needed to determine the factors that control the OC:Fe ratio, and also f<sub>Fe-OC</sub> values for soils. Nevertheless, the lack of (or poor) relationship shown here between the concentration of Fe-bound OC and Fe

concentrations demonstrates the limitations associated with predicting and modeling the behavior of C in forest soils based on the Fe concentrations in soils alone.

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# 3.3 Spatial variance and ecogeographical factors

We analyzed the influences of ecogeographical factors on the occurrence of Fe-bound OC in forest soils (Fig. 3, Supplementary Material, Fig. S3, Fig. S4). There was a significant correlation between the TOC concentration and latitude (Pearson correlation coefficient r=0.619, p=0.018), a pattern commonly observed due to lower microbial activity and turnover rates of C at higher, colder latitudes (Davidson and Janssens, 2006). The concentration of reactive Fe, if excluding soil MS in California, was also significantly related to latitude (r=0.824, p=0.001). Both concentrations of Fe-bound OC and  $f_{\text{Fe-OC}}$  were also correlated with latitude (r=0.523, p=0.053; r=0.525, p=0.054). Among our samples, the soil in forest HL in Maine, one of the three northern-most site with latitude of 45°, had the highest f<sub>Fe-OC</sub> of 57.8%. In forest GS in Florida with lowest latitude of 29.7°, the  $f_{\text{Fe-OC}}$  was below detection limits, possibly due to the low concentration of reactive Fe (0.08 mg g<sup>-1</sup>). Hence, increase in latitude both increased concentrations of TOC in soil as well concentrations of Fe-bound OC, suggesting increased interactions between Fe oxide and OC at higher latitudes. There were no clear trends in TOC or Fe-OC interactions with longitude. For elevation, we separated two groups of samples, with one group located below 1000 m (asl) and the other group above (mainly around 2000 and 4000 asl). Concentrations of TOC and Fe-bound OC, however, were not significantly different between the two groups. There were no clear trends with precipitation either, although others have reported positive relationships between mean annual precipitation and soil TOC concentration at a global scale (Amundson, 2001). The concentration of Fe-bound OC and f<sub>Fe-OC</sub> reached peak value with mean annual temperatures at 6.6°C, with lower values both at higher and lower temperatures. Temperature dependence of Fe-bound OC can be regulated by effects of temperature on the mineral phase of Fe oxides and OC dynamics. Given that ferrihydrite can incorporate more OC than other crystalline Fe oxides, an increase in temperature favors the transformation of ferrihydrite to other crystalline iron oxides (Gnanaprakash et al., 2007; Zhao et al., 1994). However, an increase in temperature can also accelerate weathering of other minerals, and increased release of silicon can slow the transformation of ferrihydrite (Cornell et al., 1987; White and Blum, 1995). However, there is also evidence that temperature can affect the chemical

composition of soil OC substantially (Conant et al., 2011). For example, increased temperature decreased the content of oxidized functional groups, such as saccharides, which would consequently inhibit the interactions between OC and Fe oxides (Amelung et al., 1997). The overall pattern can result from combined effects of temperature on Fe mineral phase and OC transformation. Further investigations are required to elucidate the mechanism more accurately. Finally, the study covered 7 major soil orders, i.e. Alfisols (sample number n=3), Spodosols (n=4), Mollisols (n=1), Inceptisols (n=2), Entisols (n=2), Gelisols (n=1), and Ultisols (n=1). Although there are limited replications in many of these soil orders, the highest concentration of Fe-bound OC were observed in Spodosols. Regarding  $f_{\text{Fe-OC}}$ , the highest values were also found in Spodosols, possibly indicating a particular importance of Fe-bound OC in this soil type which occupies 3.5% of US land areas and 4% of global ice-free land (Soil Survey Staff, 1999). However, due to the limited number of samples for each soil order, these findings warrant further confirmation.

#### Fig. 3

# 3.4 Impact of soil physicochemical properties on Fe-OC association

Soil texture can potentially influence the accumulation of Fe-bound OC. Figure 4 demonstrates that the fraction of non-calibrated Fe-bound OC showed a significant positive correlation with the fraction of sand (r=0.72, p<0.001), and negative correlations with the fraction of silt (r=-0.697, p<0.001) and clay (r=-0.616, p<0.001). There were similar correlations between labile OC and the fractions of sand (r=0.57, p=0.033), silt (r=-0.51, p=0.062) and clay (r=-0.638, p=0.014). However, the calibrated Fe-bound OC had no significant correlation with any of the texture fractions (Supplementary Material, Fig. S5). These correlations indicate that the labile OC was mainly associated with the sand component of forest soils, but that the soil texture did not affect the Fe-bound OC. There is debate on the relative roles of sand, clay and silt in the stabilization of OC in soil (Percival et al., 2000; Six et al., 2002; Eusterhues et al., 2005; Vogel et al., 2014). Eusterhues et al. (2005) found a relationship between the resistance of organic matter to oxidative degradation and the clay concentration in soils, suggesting the importance of clay minerals in the stabilization and accumulation of soil OC. Reduced chemical potential of soil organic matter in small pores of clay-rich soils also limits

microbial degradation and enhance its stabilization (Riedel and Weber, 2016). In contrast, Percival et al. (2000) found that the clay mineral fraction explained little of the variation in the accumulation of OC across a range of soil types in New Zealand. Vogel et al. (2014) found that less than 20% of clay mineral surfaces were covered by the sorption of OC, indicating that a limited proportion of clay mineral surface contributed towards the stabilization of OC. Our results suggest that the Fe oxide-mediated stabilization of OC was not related to the size/aggregation-based process, although the labile carbon concentrations increased with the fraction of sand in the soils.

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#### Fig. 4

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The Fe-OC association can also be influenced by the soil pH, which affects the mineral phases of Fe oxides and their surface charge, and their interactions with OC. For our soil samples, the soil pH ranged from 4.1 to 6.3, similar to measurements by Wagai and Mayer (2007) for North America soils. There was no significant correlation between the  $f_{\text{Fe-OC}}$  and soil pH, e.g. the HL (Maine) soil with pH of 4.4 had the highest  $f_{\text{Fe-OC}}$  of 57.8%, while the TS(II) (Washington) soil with a similar pH of 4.5 only had a f<sub>Fe-OC</sub> of 7.4%. For soils with pH ranging from 4.9 to 5.8, f<sub>Fe-OC</sub> did not change correspondingly. Contrastingly, values of OC:Fe molar ratios were significantly influenced by the soil pH; except for one outlier sample of TS(II) (Washington) soil, there was a significant negative correlation between the OC:Fe molar ratio and soil pH (r=-0.477, p=0.09) (Supplementary Material, Fig. S6). This may be due to the lower pH values favoring the complexation and precipitation of Fe with OC, while higher pH favors sorptive interactions between Fe minerals and OC (Tipping et al., 2002). If comparing samples with similar pH, the soils with higher TOC had higher OC:Fe molar ratios, e.g. the GS soil (TOC = 1.1%) with pH of 4.7 had an OC:Fe molar ratio = 8.5, while the HT (Michigan) soil (TOC = 3.0%) with similar pH of 4.7 had an OC:Fe molar ratio = 17.1. This was consistent with Schwertmann et al. (1986), who found that the major form of Fe would change from FeO<sub>x</sub> to complexes with OC when there is higher OC supply.

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#### 3.5 Molecular characteristics of Fe-bound OC

The chemical composition of Fe-bound OC can be substantially different from non-Febound OC (Adhikari and Yang, 2015) with broad implications on the C biogeochemical cycles. although such differences so far have received limited attention. We analyzed the difference in chemical composition of Fe-bound OC compared to non-Fe-bound OC using ATR-FTIR analysis (Fig. 5, Supplementary Material, Fig. S7). Overall, there were limited fingerprint peaks for OC, because of the low concentration of TOC and technical challenge for analyzing whole soil particles with FTIR (Calderon et al., 2011; Simonetti et al., 2012). Reeves (2012) demonstrated that FTIR analysis of mineral soils in the ranges of 1600-1750 and 2800-3000 cm<sup>-1</sup> only can be used to study OC. Peaks in the range of 500-1200 cm<sup>-1</sup> indicate the presence of clay or other Fe/Al minerals (Supplementary Material, Fig. S7) (Madejova, 2003; Harsh et al., 2002; Parikh et al., 2014), such as kaolinite or montmorillonite at 850-1200 cm<sup>-1</sup> (Madejova, 2003). Absorption at 850-1200 cm<sup>-1</sup> can also be due to the presence of polysaccharides, but definitive identification of polysaccharides is not possible in the presence of minerals (Senesi et al., 2003; Tandy et al., 2010). The spectra in the range of 1600-1750 cm<sup>-1</sup> normally contain fingerprint peaks for functional groups of amides, carboxylates and aromatics (Parikh et al., 2014), but we did not detect any significant peaks in this range. In the range of 2800-3000 cm<sup>-1</sup>, there were no significant peaks for the original soil samples, but after Fe extraction we detected significant peaks at 2850 and 2930 cm<sup>-1</sup>, which are characteristic for the presence of aliphatic carbon. The substantial differences in spectra before and after Fe extraction indicate that aliphatic OC was enriched in the residual soils after extraction. Other functional groups, such as aromatic carbon and hydrophilic functional groups, were more strongly associated with Fe minerals and removed during the Fe extraction, as hydrophilic functional groups can form inner-sphere coordination complexation with iron oxides, and aromatic carbon has electron donor-acceptor interactions with iron oxides (Gu et al., 1995; Axe and Persson, 2001). This result was consistent with a previous study using ultra-high resolution mass spectrometry, showing the release of more aromatic carbon during the reductive dissolution of Fe oxides (Riedel et al., 2014). Analysis for the chemical nature of Fe-bound OC can be influenced by the potential reaction of natural organic matter with dithionite, which was not noticed in previous studies (Lalonde et al., 2012; Wagai and Mayer, 2007). The most likely reaction between dithionite and organic matter is the reduction of oxidized organic functional groups. Our recent study showed that dithionite could reduce quinone groups in natural organic matter (Adhikari et al., 2016). Most likely, other major

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functional groups, such as carboxylic and carbonyl functional groups, cannot be reduced by dithionite based on their reduction potentials (Bar-Even et al., 2012; Mayhew et al., 1978). Further investigations are needed to elaborate the detailed influences of dithionite reduction on the molecular properties of organic matter.

#### Fig. 5

Furthermore, we analyzed the C 1s NEXAFS spectra of two original, non-extracted soils with the highest and lowest values of  $f_{\text{Fe-OC}}$ , i.e. HL (Maine) ( $f_{\text{Fe-OC}}$ =57.8%) and OR (Tennessee) ( $f_{\text{Fe-OC}}$  non-detectable) (Supplementary Material, Fig. S8). Three major fingerprint peaks were detected for both soils, including peaks at 285.3, 287.0 and 288.7 eV, which are corresponding to aromatic carbon, aliphatic carbon and carboxylic carbon, respectively (Schumacher et al., 2005; Solomon et al., 2005; Lehmann et al., 2008). The OR (Tennessee) soil had a more substantial signal at 287.0 eV than the HL (Maine) soil, indicating a higher aliphatic carbon concentration in the OR (Tennessee) soil compared to the HL (Maine) soil. Ratio of carboxylic carbon to aromatic carbon (peak height) was 3.8 for HL (Maine) and 1.0 for OR (Tennessee), suggesting that the HL (Maine) soil with higher  $f_{\text{Fe-OC}}$  has relatively more carboxylic carbon compared to aromatic carbon. Hence, the C1s NEXAFS spectra suggest that the soil with the higher  $f_{\text{Fe-OC}}$  has higher concentration of carboxylic C, while the soil with the lower  $f_{\text{Fe-OC}}$  value has a higher aliphatic C concentration. This result is consistent with the comparison of ATR-FTIR spectra in soils before and after Fe extraction, providing evidence that Fe oxides are mainly associated with more hydrophilic and carboxylic carbon, while non-Fe-bound OC was more aliphatic.

To further investigate the relationships between soil OC and Fe minerals, we analyzed the stable C isotopic compositions ( $\delta^{13}$ C) of Fe-bound vs. non-Fe-bound OC (i.e., the residual OC after DCB extraction). The  $\delta^{13}$ C for original soil samples ranged from –24.5‰ to –27.5‰, and the values for non-Fe-bound OC were –25.1‰ to –28.0‰. The  $\delta^{13}$ C for Fe-bound OC was calculated by combined isotope-mass balance (equation (1))

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$$\delta^{13}C_{TOC} \times TOC = \delta^{13}C_{labile} \times OC_{labile} + \delta^{13}C'_{Fe-OC} \times OC'_{Fe} + \delta^{13}C_{non-Fe-OC} \times OC_{non-Fe}$$
 (1)

where TOC is the concentration of total organic carbon, OC<sub>labile</sub> is the concentration of labile OC (extractable by bicarbonate buffer), OC<sub>non-Fe</sub> is the concentration of non-Fe-bound OC (residual

OC after Fe extraction), and OC'<sub>Fe</sub> is the concentration of Fe-bound OC (excluded the labile OC); 395

 $\delta^{13}$ C<sub>TOC</sub> is  $\delta^{13}$ C for bulk OC,  $\delta^{13}$ C<sub>labile</sub> is  $\delta^{13}$ C for labile OC,  $\delta^{13}$ C'<sub>Fe-OC</sub> is  $\delta^{13}$ C for Fe-bound OC, 396

 $\delta^{13}$ C<sub>non-Fe-OC</sub> is  $\delta^{13}$ C for non-Fe-bound OC. However, it is difficult to directly resolve the  $\delta^{13}$ C<sub>labile</sub> 397

and  $\delta^{13}$ C'<sub>Fe-OC</sub> using this equation. We simplified it to equation (2): 398

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$$\delta^{^{13}}C_{Fe-OC} = \frac{(\delta^{^{13}}C_{TOC} \times TOC - \delta^{^{13}}C_{non-Fe-OC} \times OC_{non-Fe})}{OC_{Fe}}$$
400 where  $\delta^{^{13}}C_{Fe-OC}$  is  $\delta^{^{13}}C$  for Fe-bound OC (including the labile OC),  $\delta^{^{13}}C_{TOC}$  is  $\delta^{^{13}}C$  for bulk OC,

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 $\delta^{13}$ C<sub>non-Fe-OC</sub> is  $\delta^{13}$ C for non-Fe-bound OC, TOC is the concentration of total organic carbon, 401

OC<sub>non-Fe</sub> is the concentration of non-Fe-bound OC, and OC<sub>Fe</sub> is the concentration of Fe-bound 402

OC. The  $\delta^{13}$ C for Fe-bound OC was heaviest for the TKF (California) soil with a value of – 403

23.0%, and the lightest for the GS (Florida) forest at -27.0%. Across all study sites, Fe-bound 404

OC was relatively enriched in <sup>13</sup>C (1.5±1.2‰ heavier) compared to the non-Fe-bound OC. 405

However, there is also a contribution of labile OC to the Fe-bound OC, where labile OC is the 406

OC extracted during the dithionite-absent extraction described earlier). The  $\delta^{13}$ C value for labile 407

OC can be calculated using equation (3): 408

$$409 \qquad \delta^{13} C_{labile} = \frac{(\delta^{13} C_{TOC} \times TOC - \delta^{13} C_{non-labile} \times OC_{non-labile})}{OC_{labile}}$$
(3)

where  $\delta^{13}$ C<sub>labile</sub> is  $\delta^{13}$ C for labile OC,  $\delta^{13}$ C<sub>TOC</sub> is  $\delta^{13}$ C for bulk OC,  $\delta^{13}$ C<sub>non-labile</sub> is  $\delta^{13}$ C for non-410

labile OC, OC<sub>non-labile</sub> is the concentration of non-labile OC, and OC<sub>labile</sub> is the concentration of

labile OC. Calculated values of  $\delta^{13}$ C<sub>labile</sub> range from -23.4% to -30.3%, and were lighter than the 412

values for  $\delta^{13}$ C<sub>Fe-OC</sub>. Although it is not reliable to quantitatively calculate the  $\delta^{13}$ C for Fe-bound

OC subtracting the influences of labile OC, these results indicate that the true value for  $\delta^{13}C_{Fe-OC}$ 

should be even somewhat heavier than the results presented in Fig. 6 and Supplementary 415

Material, Fig. S9. 416

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Our results demonstrate that Fe-bound OC was enriched in <sup>13</sup>C compared to the non-Febound OC in forest soils, which is consistent with results for sediments, where Fe-bound OC was 1.7±2.8‰ heavier than non-Fe-bound OC (Lalonde et al., 2012) (Fig. 6A). Previous studies showed that <sup>13</sup>C-enriched organic matter in sediments was enriched with O and N (due to the presence of compounds such as proteins and carbohydrate groups), while the microbial biomassderived lipid fraction was relatively <sup>13</sup>C-depleted (Wang et al., 1998; Zelles et al., 1992). Similarly, compound-specific isotopic analyses have shown that oxygen- and nitrogen-rich constituents, such as cellulose, hemi-cellulose and amino acids, are <sup>13</sup>C-enriched compared to

hydrocarbons (Glaser, 2005), and these  $^{13}$ C-enriched oxygen- and nitrogen-rich compounds can associate with Fe oxide extensively through inner-sphere coordination interactions (Parikh et al., 2014). The value of  $\Delta^{13}$ FeoC-nonFeoC = ( $\delta^{13}$ CFe-OC -  $\delta^{13}$ Cnon-Fe-OC) (difference in  $\delta^{13}$ C for Fe-bound OC and non-Fe-bound OC) was inversely correlated with the molar ratio of OC:Fe (r=-0.53, p=0.05, Fig. 6B). These relationships suggest that the enrichment in  $^{13}$ C was to some degree related to the OC:Fe ratio. As discussed previously (section 3.2), lower OC:Fe ratios indicate an increased contribution from sorptive interactions of OC with Fe minerals as compared to incorporation of OC within iron oxides and OC, and these sorptive interactions between oxygen-and nitrogen-rich organic compounds and Fe oxide results in the enrichment of  $^{13}$ C of Fe-bound OC vs. non-Fe-bound OC. Previous studies have attributed the stability of relatively labile and reactive compounds, such as amino acids and sugars, to their interactions with minerals (Schmidt et al., 2011), and our results demonstrated the importance of sorption to Fe minerals in increasing the stability of relatively reactive labile compounds.

# Fig. 6

Nitrogen (N)-containing functional groups are potentially important for the association between OC and Fe oxides, although the concentrations of N are much lower than C (Yang et al., 2012; Barber et al., 2014). The bulk soil contained 0.05-0.45 % N, while the non-Fe-bound component (i.e. the residual solid after DCB extraction) contained 0.06-0.32 % N. Concentrations of Fe-bound N, calculated by difference, ranged up to 0.13 %. However, it is important to note that this number is based without a calibration for labile N that may be removed by the dithionite-free DCB extraction (data not available). There were significant correlations between C and N concentrations for both bulk soils (r=0.847, p<0.001: Supplementary Material, Fig. S10) and the non-Fe-bound residual components (r=0.858, p<0.001: Supplementary Material, Fig. S10), with molar C/N ratios of 14.2±2.6 and 13.7±2.3 for bulk and non-Fe-bound OC, respectively. These C/N values are essentially identical to a previously observed molar C/N ratio = 14.3 for a large set of world-wide soils samples (Cleveland et al., 2007), and a molar C/N ratio = 14.4 for OC-rich samples in China (Tian et al., 2010). This result suggests that C/N ratios for Fe-bound OC did not differ from that of non-Fe-bound OC, assuming that the labile carbon did not have a substantially different C/N ratio.

Therefore, in contrast to the <sup>13</sup>C enrichment observed for Fe-bound OC, the interactions with Fe minerals did not affect the C/N ratio substantially.

#### 4. Conclusion

Overall, this study provided a comprehensive investigation into the amount and characteristics of Fe-bound OC in forest soils as well as the impact of soil physicochemical properties on Fe-bound OC. On average, Fe-bound OC contributed to 37.8% of TOC in forest soils, composing an important component of C cycles in terrestrial ecosystem. The OC:Fe molar ratios in the forest soils studied ranged from 0.56 to 17.7, indicating the importance of both sorptive and incorporative interactions between Fe and OC.  $f_{\text{Fe-OC}}$  increased with latitude, and reached the peak value for soils with an annual mean temperature of 6.6°C, as a result of the temperature dependence of Fe mineral phase and OC transformation. Combined studies of FTIR, NEXAFS, and  $^{13}$ C analysis revealed that Fe-bound OC was less aliphatic, more carboxylic, and more enriched in  $^{13}$ C, compared to non-Fe-bound OC. Assuming Fe-bound OC is relatively stable, Fe oxides serve as a storage reservoir on decadal time scales for hydrophilic and carboxylic OC, which would be otherwise relatively more available for microbial degradation.

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6/1	Figure Captions
672	Figure 1. Concentrations of total carbon (TC), total organic carbon (TOC) and Fe-bound OC in
673	14 forest soils across the United States. TC, TOC, and Fe-bound OC contents are generally
674	higher in soils with higher latitude (also shown in Figure 3). Duplicate measurements were
675	conducted for each of two plots in every forest site. Error bars represent standard deviation of
676	measurements of four replicates for each forest site.
677	Figure 2. Concentration of reactive Fe and OC:Fe molar ratio in the U.S. forest soils.
678	Figure 3. Relationships between TOC, concentration of Fe-bound OC, $f_{\text{Fe-TOC}}$ and
679	ecogeographical parameters including latitude, elevation (asl), and temperature (annual mean).
680	Figure 4. Relationships between the fractions of iron-bound organic carbon (uncalibrated for
681	loss of labile OC) and labile organic carbon and soil texture (i.e., fractions of sand, silt, and clay
682	in forest soils). Values of Pearson correlation coefficients (r) and significance levels were given.
683	Figure 5. Attenuated total reflectance-Fourier transform infrared spectroscopy (ATR-FTIR)
684	analysis for representative forest soils before (black line) and after Fe extraction (red line). Al
685	the spectra are background-calibrated. Among the 14 forest soils sampled in this study, we used
686	five different forest soils, with $f_{\text{Fe-OC}}$ ranging 5.6-57.8%.
687	Figure 6. A. δ <sup>13</sup> C of iron-bound and non-iron bound organic carbon for 14 U.S. forest sites. B
688	Correlation between $\Delta^{13}_{FeOC-nonFeOC}$ and molar ratio of OC:Fe.
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Table 1 Information for the 14 forest sites studied (Obrist et al., 2011, 2012, 2015)												
Forest ID	Abbr.	Location	Soil Order (US)	Soil Class <sup>a</sup> (FAO)	Climate Zone	LAT (°) <sup>b</sup>	LONG (°) <sup>c</sup>	Elevation (m asl)	Precip.d (mm y <sup>-1</sup> )	Temp <sup>e</sup> (°C)		
<b>Gainesville</b>	<b>GS</b>	<mark>Gainesville,</mark>	<b>Spodosols</b>	<b>Podzols</b>	<mark>Humid</mark>	<mark>29.74</mark>	<del>-82.22</del>	<mark>50</mark>	1228	<b>21.7</b>		
		Florida			Subtropical							
Oak Ridge	OR	<mark>Oak Ridge,</mark>	<b>Ultisols</b>	Acrisols	Humid	<mark>35.97</mark>	<del>-84,28</del>		1350	14.5		
A =1,1= 1	AT	Tennessee	A 1.C 1 -	T: - 1- 0-	Subtropical	20.72	02.20	210	1022	12.0		
<b>Ashland</b>	<b>AL</b>	<mark>Ashland,</mark> Missouri	<b>Alfisols</b>	Luvisols & Greyzems	Humid Continental	38.73	<mark>-92.20</mark>	<u> 210</u>	1023	13.9		
Little Valley	<b>LVF</b>	Little	<b>Entisols</b>	Arenosols	Highland	39.12	-119.93	2010	<del>551</del>	<b>5.0</b>		
(post-fire)	LVF	Valley,	LIIIISOIS	Ateliosois	Climate	39.12	-117.73	<u> 2010</u>	<u> </u>	<u>3.0</u>		
(post-inc)		Nevada			Cimate							
Little Valley	LV	Little	<b>Entisols</b>	Arenosols	<b>Highland</b>	39.12	-119.93	2011	<mark>550</mark>	5.0		
Dittie valley	<b></b> •	Valley,	Littisois	7 1101105015	Climate	37.12	117.75	2011	<u> </u>	<b>5.0</b>		
		Nevada										
<b>Marysville</b>	MS	Marysville,	Mollisols	Luvisols	<b>Mediterranean</b>	39.25	-121.28	386	<del>775</del>	<b>16.9</b>		
111W1 J 5 7 111U	1.10	California		24/15015	climate	<u>07.20</u>	121.20	<del>5 5 5</del>	, , , ,	20.5		
<b>Truckee</b>	<b>TKF</b>	Truckee,	<b>Alfisols</b>	Luvisols	Highland	<mark>39.37</mark>	<b>-120.1</b>	1768	<mark>569</mark>	<mark>6.0</mark>		
(post-fire)		California			Climate							
Truckee	<b>TK</b>	Truckee,	<b>Alfisols</b>	<b>Luvisols</b>	<mark>Highland</mark>	<mark>39.37</mark>	<b>-120.1</b>	<b>1767</b>	<mark>568</mark>	<mark>5.9</mark>		
		<b>California</b>			Climate							
Niwot Ridge	<b>NR</b>	Niwot	<b>Alfisols</b>	<b>Cambisols</b>	<mark>Highland</mark>	40.0 <mark>3</mark>	-105.55	<mark>3050</mark>	<mark>800</mark>	1.3		
		Ridge,			Climate							
**		Colorado	a 1 1	n		10.65	0645	0.1.0	0.1.0			
Hart	HT	Hart,	<b>Spodosols</b>	<b>Podzols</b>	Humid	<mark>43.67</mark>	-86.15	<mark>210</mark>	<mark>812</mark>	<mark>7.6</mark>		
D = -41 - 44	DI	Michigan	C 1 1.	D - 1 - 1 - 0	Continental Libraria	44.0	71.20	272	1200	4.5		
Bartlett	<b>BL</b>	<mark>Bartlett,</mark> New	<b>Spodosols</b>	Podzols & Lithosols	<mark>Humid</mark> Continental	<mark>44.0</mark>	<mark>-71.29</mark>	<mark>272</mark>	1300	4.5		
		Hampshire		Littiosois	Continental							
Howland	HL	Howland,	<b>Spodosols</b>	Luvisols	Humid	45.20	<mark>-68.74</mark>	<mark>60</mark>	1040	<mark>6.7</mark>		
110 Widita		Maine Maine	Spoucsois	24110015	Continental	12.20	00.71		10.10	<del>0.7</del>		
Thompson I	<b>TSI</b>	Ravensdale,	<b>Inceptisols</b>	<b>Cambisols</b>	Highland	<mark>47.38</mark>	<del>-121.93</del>	<mark>221</mark>	1141	<mark>9.8</mark>		
		<b>Washington</b>			Climate							
Thompson II	TSII	Ravensdale,	<b>Inceptisols</b>	<b>Cambisols</b>	<mark>Highland</mark>	<mark>47.38</mark>	<del>-121.93</del>	<mark>220</mark>	1140	<mark>9.8</mark>		
		<b>Washington</b>			Climate							

a: Food and Agriculture Organization; b latitude; c: longitude d: annual mean precipitation; e: annual mean temperature.

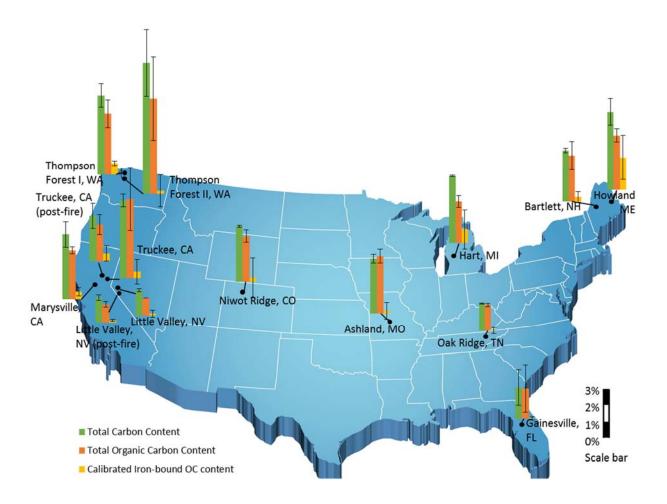
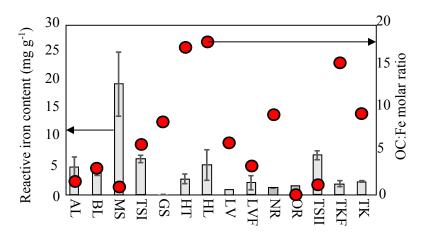
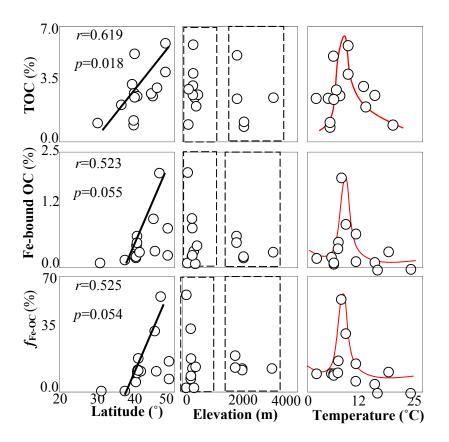


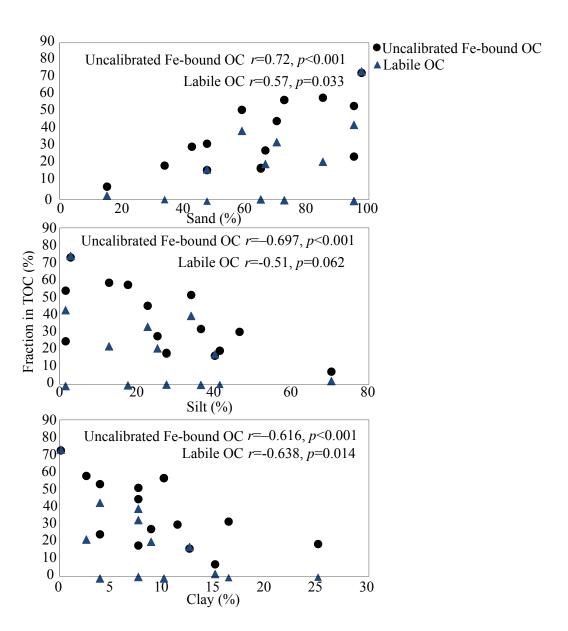
Fig. 1



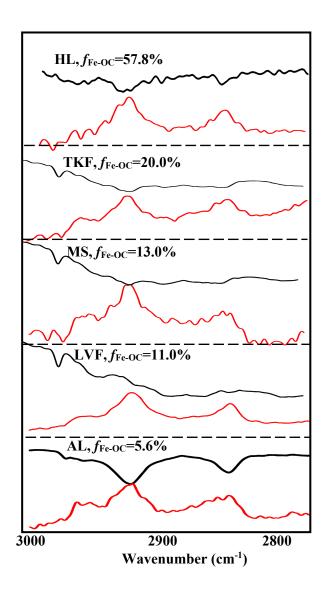
718 Fig. 2



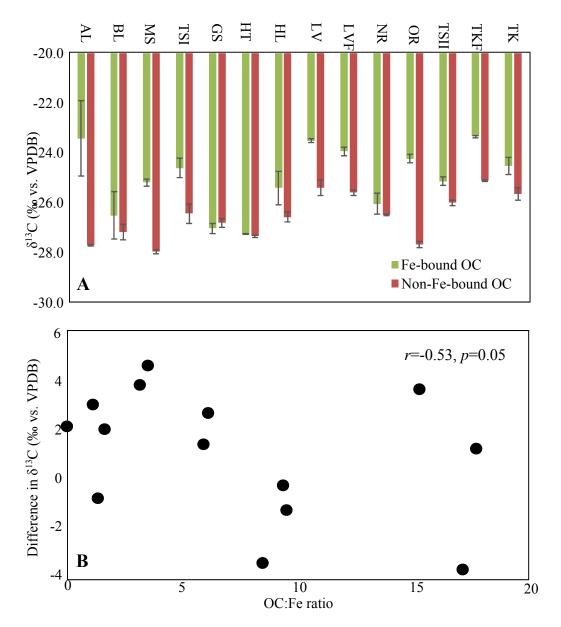
741 Fig. 3



758 Fig. 4759



768 Fig. 5



**Fig. 6**