Iron-Bound Organic Carbon in Forest Soils: Quantification and Characterization

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12 ABSTRACT

Iron oxide minerals play an important role in stabilizing organic carbon (OC) and regulating the 13 biogeochemical cycles of OC on the earth surface. To predict the fate of OC, it is essential to 14 understand the amount, spatial variability, and characteristics of Fe-bound OC in natural soils. In 15 this study, we investigated the concentrations and characteristics of Fe-bound OC in soils 16 collected from 14 forests in the United States, and determined the impact of ecogeographical 17 variables and soil physicochemical properties on the association of OC and Fe minerals. On 18 19 average, Fe-bound OC contributed 37.8% of total OC (TOC) in forest soils. Atomic ratios of OC:Fe ranged from 0.56 to 17.7 with values of 1-10 for most samples, and the ratios indicate the 20 importance of both sorptive and incorporative interactions. The fraction of Fe-bound OC in TOC 21 $(f_{\text{Fe-OC}})$ was not related to the concentration of reactive Fe, which suggests that the importance of 22 23 association with Fe in OC accumulation was not governed by the concentration of reactive Fe. Concentrations of Fe-bound OC and $f_{\text{Fe-OC}}$ increased with latitude and reached peak values at a 24 site with a mean annual temperature of 6.6 °C. Attenuated total reflectance-Fourier transform 25 infrared spectroscopy (ATR-FTIR) and near-edge X-ray absorption fine structure (NEXAFS) 26 analyses revealed that Fe-bound OC was less aliphatic than non-Fe-bound OC. Fe-bound OC 27

also was more enriched in ¹³C compared to the non-Fe-bound OC, but C/N ratios did not differ substantially. In summary, ¹³C-enriched OC with less aliphatic carbon and more carboxylic carbon was associated with Fe minerals in the soils, with values of $f_{\text{Fe-OC}}$ being controlled by both sorptive and incorporative associations between Fe and OC. Overall, this study demonstrates that Fe oxides play an important role in regulating the biogeochemical cycles of C in forest soils, and uncovers the governing factors for the spatial variability and characteristics of Fe-bound OC.

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36 **1 Introduction**

Soil organic carbon (OC) in forests is a vital component of C biogeochemical cycles (Eswaran et al., 1999). Global warming can potentially accelerate the decomposition of forest soil OC, contributing to greenhouse gas emissions (Steffen et al., 1998). Alternatively, forest soils can act as strong sinks for OC, if appropriate management is implemented, such as forest harvesting and fire treatment (Eswaran et al., 1999; Johnson and Curtis, 2001). Understanding the fate and stability of forest OC is important for evaluating and managing the global C cycle under the framework of climate change.

44 Currently, there is an information gap concerning the stability and residence time of OC, contributing to the problem that the residence time of OC (ranging from months to hundreds of 45 years) is a major source of uncertainty in modeling and prediction of C cycles (Schmidt et al., 46 2011; Riley et al., 2014). Many concepts have been proposed to account for OC stabilization and 47 48 therefore residence times, including molecular recalcitrance, physical occlusion, and chemical protection (Sollins et al., 1996; Krull et al., 2003; Baldock et al., 2004; Mayer et al., 2004; 49 50 Zimmerman et al., 2004; Schmidt et al., 2011). In general, the stability of OC is regulated by biogeochemical reactions occurring at the interfaces between OC, minerals, and microorganisms, 51 52 and further knowledge about the mechanism for OC stabilization is critical for building up process-based models to simulate and predict C cycles. 53

A number of lines of evidence suggest a key importance of iron oxide minerals in the stabilization of OC (Kalbitz et al., 2005; Kaiser and Guggenberger, 2007; Wagai and Mayer, 2007). Iron oxides have a relatively high sorption capacity for OC, with sorption coefficients for OC much higher than that of other metal oxides (Kaiser and Guggenberger, 2007; Chorover and Amistadi, 2001). Wagai and Mayer (2007) reported Fe-bound OC concentrations in soils up to 22 mg g⁻¹ soil, contributing up to 40% of total OC (TOC) for most forest soils. Similarly, Lalonde et al. (2012) found that Fe-bound OC contributed 22% of TOC in sediments. Studies have shown that Fe minerals protect OC from degradation and inhibit mineralization of OC (Baldock and Skjemstad, 2000; Kalbitz et al., 2005). There is, however, no systematic study on the occurrence of Fe-bound OC across different forests and its governing factors.

The overall goals of this study were to investigate the spatial variability of Fe-bound OC 64 across forest soils, the factors that control Fe-bound OC concentrations, and the characteristics of 65 Fe-bound OC with respect to the physicochemical properties of soils. In this study, we first 66 quantified the concentration of Fe-bound OC across 14 forest soils in the United States and 67 analyzed the spatial distribution and influences of ecogeographical factors. Second, we 68 investigated the impact of soil physicochemical properties on the Fe-OC associations. Third, we 69 studied molecular characteristics of Fe-bound OC vs. non-Fe-bound OC, including how Fe-OC 70 association influenced the chemical properties of OC and the stable isotope composition. Hence, 71 this study provided a systematic evaluation for the Fe-bound OC in United States forests, the 72 influences of ecological factors on the occurrence of Fe-bound OC, and the effects of association 73 with Fe on the chemical properties of OC. 74

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76 2. Methods & Materials

77 2.1 Chemicals and materials

Reagents used for Fe reduction experiments include sodium bicarbonate (NaHCO₃:
Sigma-Aldrich, St. Louis, MO, USA), trisodium citrate dihydrate (Na₃C₆H₅O₇•2H₂O: Acros
Organics, New Jersey, USA), and sodium dithionite (Na₂S₂O₄: Alfa Aesar, Ward Hill, MA,
USA). All chemicals used were analytical grade.

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83 2.2 Soil sample collection, primary characterization and pretreatment

84 Soil samples were collected from 14 forest sites in the United States (Obrist et al., 2011, 85 2012, 2015). The abbreviations and the basic information for the sites are summarized in Table 1. 86 More detailed information on the sites and sampling protocols can be found in previous 87 publications (Obrist et al., 2011, 2012, 2015). Briefly, two replicate plots at each forest site were

sampled. During 2007-2009, top soils (0-20 cm) from all sites were collected using clean latex 88 gloves and stainless steel sampling equipment. All the samples were immediately transferred to 89 plastic freezer bags and kept on ice before transportation to the laboratory. Soil texture was 90 analyzed by an ASTM 152-type hydrometer at the Soil Forage and Water Analysis Laboratory at 91 Oklahoma State University (Obrist et al., 2011). The soil pH was measured by mixing soil 92 particle with deionized (DI) water in a solid/solution ratio of 1:1 (Kalra, 1995). Soil samples 93 used in the experiments in this study were ground to $< 500 \,\mu\text{m}$ and freeze-dried after the removal 94 of roots and visible plant material and large particles (>2 mm) by dry sieving. 95

96

97 Table 1

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99 2.3 Total C (TC), TOC and stable C isotope analyses

TC, TOC and stable C isotopic compositions of soil samples were analyzed using a 100 Eurovector elemental analyzer (Eurovector SPA, Milan, Italy) interfaced to a Micromass 101 IsoPrime stable isotope ratio mass spectrometer (Micromass UK Ltd., Manchester, UK). 102 Acetanilide (71.09 % C by weight) was used as a standard compound to establish a calibration 103 curve between mass of C and the m/z 44 response from the mass spectrometer. In this study, the 104 105 concentration of TC and TOC were expressed as weight %. Stable C isotope analyses were performed after the method of Werner et al. (1999), with results reported in the usual delta 106 107 notation in units of ‰ vs. Vienna Pee Dee Belemnite (VPDB). For TOC analysis, soil samples were acidified with 1 M HCl with the solution/solid ratio of 1 mL solution/0.5 g soil and heated 108 at 100°C for 1 hour. The treatment was repeated three times until there was no further 109 effervescence upon acid addition, after which the samples were dried and analyzed. All analyses 110 are based on standard curves with $R^2 > 0.99$. The detection limit for C is 0.2 mg g⁻¹ soil. The 111 average coefficient of variation for the analysis of C is 20.2%. 112

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114 2.4 Nitrogen (N) analysis

The N concentration of each sample was analyzed using a Eurovector elemental analyzer. Acetanilide (10.36 % N by weight) was used as a standard compound to establish a calibration curve between mass of N and the response of the thermal conductivity detector in the elemental analyzer. Total N and non-Fe-bound N concentrations were measured before and after a Fe

reduction release treatment for each sample. All analyses are based on standard curves with 119 $R^2 > 0.99$. The detection limit for N is 0.2 mg g⁻¹ soil. The average coefficient of variation for the 120 analysis of N is 20.5%. 121

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2.5 Analysis of Fe-bound OC

The concentration of Fe-bound OC was quantified by an established Fe reduction release 124 method, commonly known as DCB extraction involving sodium dithionite, citrate and 125 bicarbonate (Mehra and Jackson, 1960; Wagai and Mayer, 2007; Lalonde et al., 2012). The DCB 126 extraction is assumed to extract most free Fe oxides (i.e. goethite, hematite, ferrihydrite and 127 others) existing in soils, but should not extract structural Fe in clay minerals (Mehra and Jackson, 128 1960; Wagai and Mayer et al., 2007; Lalonde et al., 2012). In this study, we followed the specific 129 130 protocol detailed in Lalonde et al. (2012). An aliquot (0.25 g) of soil was mixed with 15 mL of buffer solution at pH 7 (containing 0.11 M bicarbonate and 0.27 M trisodium citrate), and then 131 heated to 80°C in a water bath. The reducing agent sodium dithionite was added to the samples 132 with final concentration of 0.1 M, and maintained at 80°C for 15 min. The samples were then 133 134 centrifuged at 10,000 rpm for 10 min, the supernatant was removed, and the residual particles were rinsed using 5 mL of DI water. The rinse/centrifuge process was performed three times. 135 The residual particles were freeze-dried and analyzed for TC and TOC concentrations and δ^{13} C 136 composition. The mass of residual particles was used to calculate the OC concentration 137 associated with non-Fe minerals. 138

The background release of OC during the heating process was measured following the 139 method in Lalonde et al. (2012), where sodium citrate and dithionite were replaced by sodium 140 chloride with the same ionic strength. An aliquot (0.25 g) of dry soil was mixed with 15 mL of 141 1.6 M NaCl and 0.11 M NaHCO₃, and heated to 80°C. Then 0.22 g of NaCl was added, and the 142 solution was maintained at 80°C for 15 min. The samples were then centrifuged at 10,000 rpm 143 and rinsed three times, and freeze-dried before analysis. The mass of residual particles was used 144 145 to calculate the concentration of OC released by heating to 80°C. In preliminary experiments, we found that the solution pH increased rapidly during the heating-extraction process with 146 147 bicarbonate and sodium chloride only, and the increased pH values facilitated the release of 148 additional OC. Hence, we used a lower initial pH of 6 to compensate for the shift to higher pH during heating. To validate the measurement for the concentration of OC released during heating, 149

we also tested the release of OC using a phosphate buffer (same ionic strength) in lieu of the bicarbonate buffer, which can maintain a pH of 7 during heating. Our results showed that the concentration of OC released was similar for both the bicarbonate and phosphate buffer extraction reactions (Supplementary Material, Fig. S1).

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155 **2.6 Quantification of reactive Fe**

The concentration of reactive Fe in soils was determined by analyzing the Fe released during the DCB reduction process. After the reduction treatment, the supernatant of each sample was filtered using a 0.2 μ m syringe filter (cellulose acetate), and analyzed for Fe concentration by inductively coupled plasma - atomic emission spectroscopy (Varian-Vista AX CCD, Palo Alto, CA, USA) at an optical absorption wavelength of 259.9 nm. All analyses are based on standard curves with R^2 >0.99. The detection limit for Fe is 0.04 mg g⁻¹ soil. The average coefficient of variation for the analysis of Fe is 25.8%.

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164 2.7 Attenuated total reflectance-Fourier transform infrared spectroscopy (ATR-FTIR)

ATR-FTIR analysis to characterize the molecular composition of OC was performed for original soil samples and residual soils after DCB extraction using a Thermo Scientific Nicolet 6700 FTIR (Waltham, MA). Dry soil samples were placed directly on the crystal and forced to contact well with the crystal. Spectra were acquired at the resolution of 4 cm⁻¹ based on 100 scans. Data collection and baseline correction were accomplished using OMNIC software version 8.3.103.

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172 **2.8** Near-edge X-ray absorption fine structure (NEXAFS) analysis

173 For further characterization of chemical structure of OM, carbon (1s) K-edge NEXAFS analyses were performed for select soil samples, i.e. for soils with the highest and lowest values 174 of the fraction of Fe-bound OC to TOC. The soil particles were suspended in DI water and 175 deposited on an Au-coated silicon wafer attached to a Cu sample holder. Before analysis, 176 samples were dried in a vacuum desiccator. The X-ray-based experiments were performed on the 177 Spherical Grating Monochromator (SGM) beamline at the Canadian Light Source (Saskatoon, 178 Canada) (Regier et al., 2007). The energy scale was calibrated using citric acid (absorption at 179 288.6 eV). Major technical parameters and set-up for the beamline include: X-ray energy ranges 180

181 250-2000 eV; 45 mm planer undulator; 1000 μ m×100 μ m spot size; silicon drift detectors (SDD); 182 a titanium filter before the sample; entrance and exit slit gaps of 249.9 μ m and 25 μ m (Gillespie 183 et al., 2015). Carbon 1s spectra were acquired by slew scans from 270 to 320 eV at 20 s dwell 184 time and 20 scans per sample on a new spot. For data normalization, I₀ was collected by 185 measuring the scatter of the incident beam from a freshly Au-coated Si wafer using SDD. 186 Before the I₀ normalization, the pre-edge baseline was adjusted to near zero to remove the scatter 187 in the sample data (Gillespie et al., 2015).

188 **3. Results and Discussion**

3.1 Concentration of Fe-bound OC

This study covered five major forest types in North America, including Spruce-Fir, Pine, 190 Oak, Chaparral, and Maple-beech-birch forests distributed between 29° and 47° N. For the 14 191 forest soils, TC concentrations ranged between 1.5±0.1 and 8.3±2.1% (all percentages given are 192 weight-based), and TOC concentrations ranged between 1.3±0.3 and 6.2±2.9%, which are 193 comparable to values previously reported for North American forest soils (Wagai and Mayer, 194 2007; Wilson et al., 2013). Bicarbonate extraction-calibrated Fe-bound OC concentrations 195 196 ranged from 0.3 to 1.9%, with the fraction of Fe-bound OC to TOC (fFe-OC) averaging 37.8±20.0% (Fig. 1, Supplementary Material, Table S1). Forest HL (Maine) had the highest fe-oc of 57.8%, 197 while forests GS (Florida) and OR (Tennessee) had fre-OC values below detection limits (i.e., 198 below 0.6%). Based on an estimate that 1502 Pg (Pg=1×10¹⁵ g) of TOC is stored in terrestrial 199 200 soils (Scharlemann, et al., 2014), scaling up these results to a global estimate would yield 201 538.5±271.5 Pg of Fe-bound OC residing in terrestrial soils.

- 202
- 203 Fig. 1
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205 **3.2 Fe-OC association**

The values of $f_{\text{Fe-OC}}$ were influenced not only by the concentration of reactive Fe, but also by the type of association between Fe and OC. In this study, the concentration of reactive Fe in forest soils ranged from 0.1 mg g⁻¹ to 19.3 mg g⁻¹, which is low compared to values of reactive Fe of up to 180 mg g⁻¹ reported previously (Wagai and Mayer, 2007; Wagai et al., 2013) (Fig. 2). A Mollisol in forest site MS (California) had the highest concentration of reactive Fe, while a Spodosol in forest site GS (Florida) had the lowest reactive Fe concentration. There was no significant correlation between $f_{\text{Fe-OC}}$ and the concentration of reactive Fe (Pearson Correlation Coefficient *r*=-0.418, *p*=0.137, Supplementary Material, Fig. S2). This suggests that the proportion of Fe-bound OC is not strongly controlled by the reactive Fe concentration.

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- 216 Fig. 2.
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The OC:Fe molar ratio ranged from 0.56 to 17.7 for all 14 soils, with a value between 1 218 and 10 for 10 soils (Fig. 2). Previous studies have suggested that the OC:Fe molar ratio can be 219 used as an indicator for the type of association between Fe oxides and OC, with lower values 220 indicating sorptive interactions while higher values indicate incorporation of OC within Fe 221 oxides (Wagai et al., 2007; Guggenberger and Kaiser, 2003). The highest sorption capacity 222 measured for OC onto Fe oxide corresponds to an OC:Fe molar ratio = 1.0 (Kaiser and 223 Guggenberger, 2006), but by incorporation and co-precipitation of Fe oxide OC:Fe molar ratio 224 can reach much higher values (Guggenberger and Kaiser, 2003). With OC:Fe molar ratios 225 generally between 1-10 for about two thirds of the forest soils in this study, we propose that 226 incorporation of OC into Fe oxides plays a major role in the accumulation of Fe-bound OC 227 exceeding sorption by at least a factor of 1 to almost 20 (Wagai and Mayer, 2007; Lalonde, 2012). 228 However, for the HT (Michigan), HL (Maine) and TKF (California) forest soils, the OC:Fe 229 molar ratios were even higher than 10 with a maximum value of 17.8 (Fig. 2), implying that 230 incorporation of OC into Fe oxides dominated at these sites. Similar to $f_{\text{Fe-OC}}$, OC:Fe ratios were 231 not related to the concentration of reactive Fe and showed large variation for soils with similar 232 233 concentration of total reactive Fe (Supplementary Material, Fig. S2). This further indicates that the type of interactions between OC and Fe was not governed by the amount of Fe. The OC:Fe 234 ratio is potentially regulated by the mineral phases of Fe, as poorly-crystalline Fe oxides have a 235 236 higher capacity to bind with OC than crystalline Fe minerals (Eusterhues et al., 2014). When 237 sorption dominates the interactions between OC and Fe, OC:Fe can also be influenced greatly by the particle size and surface area of Fe oxides (Gu et al., 1995). Further investigations are needed 238 239 to determine the factors that control the OC: Fe ratio, and also $f_{\text{Fe-OC}}$ values for soils. Nevertheless, 240 the lack of (or poor) relationship shown here between the concentration of Fe-bound OC and Fe

concentrations demonstrates the limitations associated with predicting and modeling the behaviorof C in forest soils based on the Fe concentrations in soils alone.

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244 3.3 Spatial variance and ecogeographical factors

245 We analyzed the influences of ecogeographical factors on the occurrence of Fe-bound OC in forest soils (Fig. 3, Supplementary Material, Fig. S3, Fig. S4). There was a significant correlation 246 between the TOC concentration and latitude (Pearson correlation coefficient r=0.619, p=0.018), 247 a pattern commonly observed due to lower microbial activity and turnover rates of C at higher. 248 colder latitudes (Davidson and Janssens, 2006). The concentration of reactive Fe, if excluding 249 250 soil MS in California, was also significantly related to latitude (r=0.824, p=0.001). Both concentrations of Fe-bound OC and $f_{\text{Fe-OC}}$ were also correlated with latitude (r=0.523, p=0.053; 251 r=0.525, p=0.054). Among our samples, the soil in forest HL in Maine, one of the three 252 northern-most site with latitude of 45°, had the highest $f_{\text{Fe-OC}}$ of 57.8%. In forest GS in Florida 253 with lowest latitude of 29.7°, the $f_{\text{Fe-OC}}$ was below detection limits, possibly due to the low 254 concentration of reactive Fe (0.08 mg g⁻¹). Hence, increase in latitude both increased 255 concentrations of TOC in soil as well concentrations of Fe-bound OC, suggesting increased 256 interactions between Fe oxide and OC at higher latitudes. There were no clear trends in TOC or 257 258 Fe-OC interactions with longitude. For elevation, we separated two groups of samples, with one group located below 1000 m (asl) and the other group above (mainly around 2000 and 4000 asl). 259 Concentrations of TOC and Fe-bound OC, however, were not significantly different between the 260 two groups. There were no clear trends with precipitation either, although others have reported 261 262 positive relationships between mean annual precipitation and soil TOC concentration at a global scale (Amundson, 2001). The concentration of Fe-bound OC and *f*_{Fe-OC} reached peak value with 263 mean annual temperatures at 6.6°C, with lower values both at higher and lower temperatures. 264 Temperature dependence of Fe-bound OC can be regulated by effects of temperature on the 265 mineral phase of Fe oxides and OC dynamics. Given that ferrihydrite can incorporate more OC 266 than other crystalline Fe oxides, an increase in temperature favors the transformation of 267 ferrihydrite to other crystalline iron oxides (Gnanaprakash et al., 2007; Zhao et al., 1994). 268 However, an increase in temperature can also accelerate weathering of other minerals, and 269 increased release of silicon can slow the transformation of ferrihydrite (Cornell et al., 1987; 270 271 White and Blum, 1995). However, there is also evidence that temperature can affect the chemical

272 composition of soil OC substantially (Conant et al., 2011). For example, increased temperature decreased the content of oxidized functional groups, such as saccharides, which would 273 274 consequently inhibit the interactions between OC and Fe oxides (Amelung et al., 1997). The overall pattern can result from combined effects of temperature on Fe mineral phase and OC 275 transformation. Further investigations are required to elucidate the mechanism more accurately. 276 Finally, the study covered 7 major soil orders, i.e. Alfisols (sample number n=3), Spodosols 277 (n=4), Mollisols (n=1), Inceptisols (n=2), Entisols (n=2), Gelisols (n=1), and Ultisols (n=1). 278 Although there are limited replications in many of these soil orders, the highest concentration of 279 Fe-bound OC were observed in Spodosols. Regarding f_{Fe-OC}, the highest values were also found 280 in Spodosols, possibly indicating a particular importance of Fe-bound OC in this soil type which 281 occupies 3.5% of US land areas and 4% of global ice-free land (Soil Survey Staff, 1999). 282 However, due to the limited number of samples for each soil order, these findings warrant further 283 confirmation. 284

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286 Fig. 3

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288 **3.4 Impact of soil physicochemical properties on Fe-OC association**

Soil texture can potentially influence the accumulation of Fe-bound OC. Figure 4 289 demonstrates that the fraction of non-calibrated Fe-bound OC showed a significant positive 290 291 correlation with the fraction of sand (r=0.72, p<0.001), and negative correlations with the fraction of silt (r=-0.697, p < 0.001) and clay (r=-0.616, p < 0.001). There were similar 292 correlations between labile OC and the fractions of sand (r=0.57, p=0.033), silt (r=-0.51, 293 p=0.062) and clay (r=-0.638, p=0.014). However, the calibrated Fe-bound OC had no significant 294 295 correlation with any of the texture fractions (Supplementary Material, Fig. S5). These correlations indicate that the labile OC was mainly associated with the sand component of forest 296 297 soils, but that the soil texture did not affect the Fe-bound OC. There is debate on the relative roles of sand, clay and silt in the stabilization of OC in soil (Percival et al., 2000; Six et al., 2002; 298 299 Eusterhues et al., 2005; Vogel et al., 2014). Eusterhues et al. (2005) found a relationship between the resistance of organic matter to oxidative degradation and the clay concentration in soils, 300 suggesting the importance of clay minerals in the stabilization and accumulation of soil OC. 301 Reduced chemical potential of soil organic matter in small pores of clay-rich soils also limits 302

303 microbial degradation and enhance its stabilization (Riedel and Weber, 2016). In contrast, Percival et al. (2000) found that the clay mineral fraction explained little of the variation in the 304 305 accumulation of OC across a range of soil types in New Zealand. Vogel et al. (2014) found that less than 20% of clay mineral surfaces were covered by the sorption of OC, indicating that a 306 limited proportion of clay mineral surface contributed towards the stabilization of OC. Our 307 results suggest that the Fe oxide-mediated stabilization of OC was not related to the 308 size/aggregation-based process, although the labile carbon concentrations increased with the 309 fraction of sand in the soils. 310

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- 312 Fig. 4
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314 The Fe-OC association can also be influenced by the soil pH, which affects the mineral phases of Fe oxides and their surface charge, and their interactions with OC. For our soil samples, 315 the soil pH ranged from 4.1 to 6.3, similar to measurements by Wagai and Mayer (2007) for 316 North America soils. There was no significant correlation between the $f_{\text{Fe-OC}}$ and soil pH, e.g. the 317 318 HL (Maine) soil with pH of 4.4 had the highest $f_{\text{Fe-OC}}$ of 57.8%, while the TS(II) (Washington) soil with a similar pH of 4.5 only had a fe-oc of 7.4%. For soils with pH ranging from 4.9 to 5.8, 319 f_{Fe-OC} did not change correspondingly. Contrastingly, values of OC:Fe molar ratios were 320 significantly influenced by the soil pH; except for one outlier sample of TS(II) (Washington) 321 322 soil, there was a significant negative correlation between the OC:Fe molar ratio and soil pH (r=-0.477, p=0.09) (Supplementary Material, Fig. S6). This may be due to the lower pH values 323 324 favoring the complexation and precipitation of Fe with OC, while higher pH favors sorptive interactions between Fe minerals and OC (Tipping et al., 2002). If comparing samples with 325 326 similar pH, the soils with higher TOC had higher OC:Fe molar ratios, e.g. the GS soil (TOC = 1.1%) with pH of 4.7 had an OC:Fe molar ratio = 8.5, while the HT (Michigan) soil (TOC = 1.1%) 327 3.0%) with similar pH of 4.7 had an OC:Fe molar ratio = 17.1. This was consistent with 328 Schwertmann et al. (1986), who found that the major form of Fe would change from FeO_x to 329 330 complexes with OC when there is higher OC supply.

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332 **3.5 Molecular characteristics of Fe-bound OC**

333 The chemical composition of Fe-bound OC can be substantially different from non-Febound OC (Adhikari and Yang, 2015) with broad implications on the C biogeochemical cycles, 334 335 although such differences so far have received limited attention. We analyzed the difference in chemical composition of Fe-bound OC compared to non-Fe-bound OC using ATR-FTIR 336 337 analysis (Fig. 5, Supplementary Material, Fig. S7). Overall, there were limited fingerprint peaks for OC, because of the low concentration of TOC and technical challenge for analyzing whole 338 soil particles with FTIR (Calderon et al., 2011; Simonetti et al., 2012). Reeves (2012) 339 demonstrated that FTIR analysis of mineral soils in the ranges of 1600-1750 and 2800-3000 cm⁻¹ 340 only can be used to study OC. Peaks in the range of 500-1200 cm⁻¹ indicate the presence of clay 341 or other Fe/Al minerals (Supplementary Material, Fig. S7) (Madejova, 2003; Harsh et al., 2002; 342 Parikh et al., 2014), such as kaolinite or montmorillonite at 850-1200 cm⁻¹ (Madejova, 2003). 343 Absorption at 850-1200 cm⁻¹ can also be due to the presence of polysaccharides, but definitive 344 identification of polysaccharides is not possible in the presence of minerals (Senesi et al., 2003; 345 Tandy et al., 2010). The spectra in the range of 1600-1750 cm⁻¹ normally contain fingerprint 346 peaks for functional groups of amides, carboxylates and aromatics (Parikh et al., 2014), but we 347 did not detect any significant peaks in this range. In the range of 2800-3000 cm⁻¹, there were no 348 significant peaks for the original soil samples, but after Fe extraction we detected significant 349 peaks at 2850 and 2930 cm⁻¹, which are characteristic for the presence of aliphatic carbon. The 350 substantial differences in spectra before and after Fe extraction indicate that aliphatic OC was 351 352 enriched in the residual soils after extraction. Other functional groups, such as aromatic carbon and hydrophilic functional groups, were more strongly associated with Fe minerals and removed 353 during the Fe extraction, as hydrophilic functional groups can form inner-sphere coordination 354 complexation with iron oxides, and aromatic carbon has electron donor-acceptor interactions 355 356 with iron oxides (Gu et al., 1995; Axe and Persson, 2001). This result was consistent with a previous study using ultra-high resolution mass spectrometry, showing the release of more 357 aromatic carbon during the reductive dissolution of Fe oxides (Riedel et al., 2014). Analysis for 358 the chemical nature of Fe-bound OC can be influenced by the potential reaction of natural 359 360 organic matter with dithionite, which was not noticed in previous studies (Lalonde et al., 2012; Wagai and Mayer, 2007). The most likely reaction between dithionite and organic matter is the 361 reduction of oxidized organic functional groups. Our recent study showed that dithionite could 362 reduce quinone groups in natural organic matter (Adhikari et al., 2016). Most likely, other major 363

functional groups, such as carboxylic and carbonyl functional groups, cannot be reduced by dithionite based on their reduction potentials (Bar-Even et al., 2012; Mayhew et al., 1978). Further investigations are needed to elaborate the detailed influences of dithionite reduction on the molecular properties of organic matter.

368

369 Fig. 5

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Furthermore, we analyzed the C 1s NEXAFS spectra of two original, non-extracted soils 371 with the highest and lowest values of *f*_{Fe-OC}, i.e. HL (Maine) (*f*_{Fe-OC}=57.8%) and OR (Tennessee) 372 (f_{Fe-OC} non-detectable) (Supplementary Material, Fig. S8). Three major fingerprint peaks were 373 detected for both soils, including peaks at 285.3, 287.0 and 288.7 eV, which are corresponding to 374 aromatic carbon, aliphatic carbon and carboxylic carbon, respectively (Schumacher et al., 2005; 375 Solomon et al., 2005; Lehmann et al., 2008). The OR (Tennessee) soil had a more substantial 376 signal at 287.0 eV than the HL (Maine) soil, indicating a higher aliphatic carbon concentration in 377 the OR (Tennessee) soil compared to the HL (Maine) soil. Ratio of carboxylic carbon to 378 379 aromatic carbon (peak height) was 3.8 for HL (Maine) and 1.0 for OR (Tennessee), suggesting that the HL (Maine) soil with higher f_{Fe-OC} has relatively more carboxylic carbon compared to 380 381 aromatic carbon. Hence, the C1s NEXAFS spectra suggest that the soil with the higher $f_{\text{Fe-OC}}$ has higher concentration of carboxylic C, while the soil with the lower fFe-OC value has a higher 382 383 aliphatic C concentration. This result is consistent with the comparison of ATR-FTIR spectra in soils before and after Fe extraction, providing evidence that Fe oxides are mainly associated with 384 385 more hydrophilic and carboxylic carbon, while non-Fe-bound OC was more aliphatic.

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To further investigate the relationships between soil OC and Fe minerals, we analyzed the stable C isotopic compositions (δ^{13} C) of Fe-bound vs. non-Fe-bound OC (i.e., the residual OC after DCB extraction). The δ^{13} C for original soil samples ranged from –24.5‰ to –27.5‰, and the values for non-Fe-bound OC were –25.1‰ to –28.0‰. The δ^{13} C for Fe-bound OC was calculated by combined isotope-mass balance (equation (1))

$$392 \quad \delta^{13}C_{\text{TOC}} \times \text{TOC} = \delta^{13}C_{\text{labile}} \times \text{OC}_{\text{labile}} + \delta^{13}C'_{\text{Fe-OC}} \times \text{OC'}_{\text{Fe}} + \delta^{13}C_{\text{non-Fe-OC}} \times \text{OC}_{\text{non-Fe}}$$
(1)

where TOC is the concentration of total organic carbon, OC_{labile} is the concentration of labile OC
 (extractable by bicarbonate buffer), OC_{non-Fe} is the concentration of non-Fe-bound OC (residual

395 OC after Fe extraction), and OC'_{Fe} is the concentration of Fe-bound OC (excluded the labile OC); 396 δ^{13} CTOC is δ^{13} C for bulk OC, δ^{13} Clabile is δ^{13} C for labile OC, δ^{13} C'_{Fe-OC} is δ^{13} C for Fe-bound OC, 397 δ^{13} Cnon-Fe-OC is δ^{13} C for non-Fe-bound OC. However, it is difficult to directly resolve the δ^{13} Clabile 398 and δ^{13} C'_{Fe-OC} using this equation. We simplified it to equation (2):

$$399 \quad \delta^{13}C_{\text{Fe-OC}} = \frac{(\delta^{13}C_{\text{TOC}} \times \text{TOC} - \delta^{13}C_{\text{non-Fe-OC}} \times \text{OC}_{\text{non-Fe}})}{OC_{\text{ToC}}}$$
(2)

where $\delta^{13}C_{\text{Fe-OC}}$ is $\delta^{13}C$ for Fe-bound OC (including the labile OC), $\delta^{13}C_{\text{TOC}}$ is $\delta^{13}C$ for bulk OC, 400 $\delta^{13}C_{non-Fe-OC}$ is $\delta^{13}C$ for non-Fe-bound OC, TOC is the concentration of total organic carbon, 401 OC_{non-Fe} is the concentration of non-Fe-bound OC, and OC_{Fe} is the concentration of Fe-bound 402 OC. The δ^{13} C for Fe-bound OC was heaviest for the TKF (California) soil with a value of – 403 23.0‰, and the lightest for the GS (Florida) forest at -27.0‰. Across all study sites, Fe-bound 404 OC was relatively enriched in 13 C (1.5±1.2‰ heavier) compared to the non-Fe-bound OC. 405 However, there is also a contribution of labile OC to the Fe-bound OC, where labile OC is the 406 OC extracted during the dithionite-absent extraction described earlier). The δ^{13} C value for labile 407 OC can be calculated using equation (3): 408

409
$$\delta^{13}C_{\text{labile}} = \frac{\left(\delta^{13}C_{\text{TOC}} \times \text{TOC} - \delta^{13}C_{\text{non-labile}} \times \text{OC}_{\text{non-labile}}\right)}{\text{OC}_{\text{labile}}}$$
(3)

where $\delta^{13}C_{labile}$ is $\delta^{13}C$ for labile OC, $\delta^{13}C_{TOC}$ is $\delta^{13}C$ for bulk OC, $\delta^{13}C_{non-labile}$ is $\delta^{13}C$ for nonlabile OC, OC_{non-labile} is the concentration of non-labile OC, and OC_{labile} is the concentration of labile OC. Calculated values of $\delta^{13}C_{labile}$ range from -23.4% to -30.3%, and were lighter than the values for $\delta^{13}C_{Fe-OC}$. Although it is not reliable to quantitatively calculate the $\delta^{13}C$ for Fe-bound OC subtracting the influences of labile OC, these results indicate that the true value for $\delta^{13}C_{Fe-OC}$ should be even somewhat heavier than the results presented in Fig. 6 and Supplementary Material, Fig. S9.

Our results demonstrate that Fe-bound OC was enriched in ¹³C compared to the non-Fe-417 bound OC in forest soils, which is consistent with results for sediments, where Fe-bound OC was 418 1.7±2.8‰ heavier than non-Fe-bound OC (Lalonde et al., 2012) (Fig. 6A). Previous studies 419 showed that ¹³C-enriched organic matter in sediments was enriched with O and N (due to the 420 421 presence of compounds such as proteins and carbohydrate groups), while the microbial biomassderived lipid fraction was relatively ¹³C-depleted (Wang et al., 1998; Zelles et al., 1992). 422 Similarly, compound-specific isotopic analyses have shown that oxygen- and nitrogen-rich 423 constituents, such as cellulose, hemi-cellulose and amino acids, are ¹³C-enriched compared to 424

hydrocarbons (Glaser, 2005), and these ¹³C-enriched oxygen- and nitrogen-rich compounds can 425 associate with Fe oxide extensively through inner-sphere coordination interactions (Parikh et al., 426 2014). The value of $\Delta^{13}_{\text{FeOC-nonFeOC}} = (\delta^{13}C_{\text{Fe-OC}} - \delta^{13}C_{\text{non-Fe-OC}})$ (difference in $\delta^{13}C$ for Fe-bound 427 OC and non-Fe-bound OC) was inversely correlated with the molar ratio of OC:Fe (r=-0.53, 428 p=0.05, Fig. 6B). These relationships suggest that the enrichment in ¹³C was to some degree 429 related to the OC:Fe ratio. As discussed previously (section 3.2), lower OC:Fe ratios indicate an 430 increased contribution from sorptive interactions of OC with Fe minerals as compared to 431 incorporation of OC within iron oxides and OC, and these sorptive interactions between oxygen-432 and nitrogen-rich organic compounds and Fe oxide results in the enrichment of ¹³C of Fe-bound 433 OC vs. non-Fe-bound OC. Previous studies have attributed the stability of relatively labile and 434 reactive compounds, such as amino acids and sugars, to their interactions with minerals (Schmidt 435 et al., 2011), and our results demonstrated the importance of sorption to Fe minerals in increasing 436 the stability of relatively reactive labile compounds. 437

- 438
- 439 Fig. 6
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Nitrogen (N)-containing functional groups are potentially important for the association 441 between OC and Fe oxides, although the concentrations of N are much lower than C (Yang et al., 442 2012; Barber et al., 2014). The bulk soil contained 0.05-0.45 % N, while the non-Fe-bound 443 444 component (i.e. the residual solid after DCB extraction) contained 0.06-0.32 % N. Concentrations of Fe-bound N, calculated by difference, ranged up to 0.13 %. However, it is 445 important to note that this number is based without a calibration for labile N that may be 446 removed by the dithionite-free DCB extraction (data not available). There were significant 447 448 correlations between C and N concentrations for both bulk soils (r=0.847, p<0.001: Supplementary Material, Fig. S10) and the non-Fe-bound residual components (r=0.858, 449 p < 0.001: Supplementary Material, Fig. S10), with molar C/N ratios of 14.2±2.6 and 13.7±2.3 for 450 bulk and non-Fe-bound OC, respectively. These C/N values are essentially identical to a 451 452 previously observed molar C/N ratio = 14.3 for a large set of world-wide soils samples (Cleveland et al., 2007), and a molar C/N ratio = 14.4 for OC-rich samples in China (Tian et al., 453 2010). This result suggests that C/N ratios for Fe-bound OC did not differ from that of non-Fe-454 bound OC, assuming that the labile carbon did not have a substantially different C/N ratio. 455

456 Therefore, in contrast to the 13 C enrichment observed for Fe-bound OC, the interactions with Fe 457 minerals did not affect the C/N ratio substantially.

458

459 **4. Conclusion**

Overall, this study provided a comprehensive investigation into the amount and characteristics of 460 Fe-bound OC in forest soils as well as the impact of soil physicochemical properties on Fe-461 bound OC. On average, Fe-bound OC contributed to 37.8% of TOC in forest soils, composing an 462 important component of C cycles in terrestrial ecosystem. The OC:Fe molar ratios in the forest 463 soils studied ranged from 0.56 to 17.7, indicating the importance of both sorptive and 464 incorporative interactions between Fe and OC. f_{Fe-OC} increased with latitude, and reached the 465 peak value for soils with an annual mean temperature of 6.6°C, as a result of the temperature 466 dependence of Fe mineral phase and OC transformation. Combined studies of FTIR, NEXAFS, 467 and ¹³C analysis revealed that Fe-bound OC was less aliphatic, more carboxylic, and more 468 enriched in ¹³C, compared to non-Fe-bound OC. Assuming Fe-bound OC is relatively stable, Fe 469 oxides serve as a storage reservoir on decadal time scales for hydrophilic and carboxylic OC, 470 which would be otherwise relatively more available for microbial degradation. 471

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481 **References**

Adhikari, D. and Yang, Y.: Selective stabilization of aliphatic organic carbon by iron oxide, Sci. Rep., 5, 2015.

- 484 Adhikari, D., Poulson, S. R., Sumaila, S., Dynes, J. J., McBeth, J. M., and Yang, Y.: Asynchronous
- reductive release of iron and organic carbon from hematite–humic acid complexes, Chem. Geol., 430, 1320, 2016.
- Amelung, W., Flach, K. W., and Zech, W.: Climatic effects on soil organic matter composition in the
 great plains, Soil Sci. Soc. Am. J., 61, 115-123, 1997.
- Amundson, R.: The carbon budget in soils, Annual Review of Earth and Planetary Sciences, 29, 535-562,
 2001.
- Axe, K. and Persson, P.: Time-dependent surface speciation of oxalate at the water-boehmite (gammaAlOOH) interface: implications for dissolution, Geochim. Cosmochim. Ac., 65, 4481-4492, 2001.
- Baldock, J. A. and Skjemstad, J. O.: Role of the soil matrix and minerals in protecting natural organic
 materials against biological attack, Org. Geochem., 31, 697-710, 2000.
- Barber, A., Lalonde, K., Mucci, A., & Gélinas, Y.: The role of iron in the diagenesis of organic carbon
 and nitrogen in sediments: A long-term incubation experiment. Mar. Chem., 162, 1-9, 2014.
- Bar-Even, A., Flamholz, A., Noor, E., and Milo, R.: Rethinking glycolysis: on the biochemical logic of
 metabolic pathways, Nat. Chem. Biol., 8, 509-517, 2012.
- Batjes, N. H.: Documentation to ISRIC-WISE global data set of derived soil properties on a ¹/₂° by ¹/₂°
 drid (Version 1.0), International Soil Reference and Information Centre, Wageningen, The Netherlands, 1996.
- Calderon, F. J., Reeves, J. B., III, Collins, H. P., and Paul, E. A.: Chemical differences in soil organic
 matter fractions determined by diffuse-reflectance mid-infrared spectroscopy, Soil Sci. Soc. Am. J., 75,
 568-579, 2011.
- 505 Chorover, J. and Amistadi, M. K.: Reaction of forest floor organic matter at goethite, birnessite and 506 smectite surfaces, Geochim. Cosmochim. Ac., 65, 95-109, 2001.
- 507 Cleveland, C. C. and Liptzin, D.: C : N : P stoichiometry in soil: is there a "Redfield ratio" for the 508 microbial biomass?, Biogeochemistry, 85, 235-252, 2007.
- 509 Conant, R. T., Ryan, M. G., Agren, G. I., Birge, H. E., Davidson, E. A., Eliasson, P. E., Evans, S. E., Frey,
- S. D., Giardina, C. P., Hopkins, F. M., Hyvonen, R., Kirschbaum, M. U. F., Lavallee, J. M., Leifeld, J.,
 Parton, W. J., Steinweg, J. M., Wallenstein, M. D., Wetterstedt, J. A. M., and Bradford, M. A.:
 Temperature and soil organic matter decomposition rates synthesis of current knowledge and a way
 forward, Glob. Change Biol., 17, 3392-3404, 2011.
- Cornell, R. M., Giovanoli, R., and Schindler, P. W.: Effect of silicate species on the transformation of
 ferrihydrite into goethite and hematite in alkaline media, Clay. Clay Miner., 35, 21-28, 1987.
- 516 Davidson, E. A. and Janssens, I. A.: Temperature sensitivity of soil carbon decomposition and feedbacks
 517 to climate change, Nature, 440, 165-173, 2006.
- 518 Eswaran, H., Reich, P. F., Kimble, J. M., Beinroth, F. H., Padmanabhan, E., and Moncharoen, P.: Global
- 519 Climate Change and Pedogenic Carbonates. Lal, R. (Ed.), Lewis Publishers, Boca Raton, FL. USA, 1999.

- 520 Eusterhues, K., Hadrich, A., Neidhardt, J., Kusel, K., Keller, T. F., Jandt, K. D., and Totsche, K. U.:
- 521 Reduction of ferrihydrite with adsorbed and coprecipitated organic matter: microbial reduction by
- 522 Geobacter bremensis vs. abiotic reduction by Na-dithionite, Biogeosciences, 11, 4953-4966, 2014.
- Eusterhues, K., Rumpel, C., and Kogel-Knabner, I.: Organo-mineral associations in sandy acid forest
 soils: importance of specific surface area, iron oxides and micropores, Eur. J. Soil Sci., 56, 753-763, 2005.
- 525 Gillespie, A. W., Phillips, C. L., Dynes, J. J., Chevrier, D., Regier, T. Z., and Peak, D.: Advances in using
- 526 soft X-ray spectroscopy for measurement of soil biogeochemical processes, Adv. Agron., 133, 1-32, 2015.
- 527 Glaser, B.: Compound-specific stable-isotope (delta C-13) analysis in soil science, J. Plant Nutr. Soil Sci.,
 528 168, 633-648, 2005.
- 529 Gnanaprakash, G., Mahadevan, S., Jayakumar, T., Kalyanasundaram, P., Philip, J., and Raj, B.: Effect of
- 530 initial pH and temperature of iron salt solutions on formation of magnetite nanoparticles, Mater. Chem.
- 531 Phys., 103, 168-175, 2007.
- Gu, B. H., Schmitt, J., Chen, Z., Liang, L. Y., and McCarthy, J. F.: Adsorption and desorption of different
 organic-matter fractions on iron-oxide, Geochim. Cosmochim. Ac., 59, 219-229, 1995.
- Guggenberger, G. and Kaiser, K.: Dissolved organic matter in soil: challenging the paradigm of sorptive
 preservation, Geoderma, 113, 293-310, 2003.
- Harsh, J. B., Chorover, J., Nizeyimana, E.: Allophane and imogolite. Chap. 9. In: Soil Mineralogy with
 environmental applications, Dixon, J. B., Schulze, D. G.: (Ed.), Book Series SSSA No. 7, Madison, WI.,
 2002.
- Johnson, D. W., and Curtis, P. S.: Effects of forest management on soil C and N storage: meta analysis,
 Forest Ecol. Manag., 140, 227-238, 2001.
- Kaiser, K. and Guggenberger, G.: Sorptive stabilization of organic matter by microporous goethite:
 sorption into small pores vs. surface complexation, Eur. J. Soil Sci., 58, 45-59, 2007.
- Kalbitz, K., Schwesig, D., Rethemeyer, J., and Matzner, E.: Stabilization of dissolved organic matter by
 sorption to the mineral soil, Soil Biol. Biochem., 37, 1319-1331, 2005.
- 545 Kalra, Y. P., Agrawal, H. P., Allen, E., Ashworth, J., Audesse, P., Case, V. W., Collins, D., Combs, S. M.,
- 546 Dawson, C., Denning, J., Donohue, S. J., Douglas, B., Drought, B. G., Flock, M. A., Friedericks, J. B.,
- 547 Gascho, G. J., Gerstl, Z., Hodgins, L., Hopkins, B., Horneck, D., Isaac, R. A., Kelly, P. M., Konwicki, J.,
- Kovar, J., Kowalenko, G., Lutwick, G., Miller, R. O., Munter, R., Murchison, I., Neary, A., Neumann, R.,
 Neville, M., Nolan, C. B., Olive, R., Pask, W., Pastorek, L., Peck, T. R., Peel, T., Ramakers, J., Reid, W.
- 550 S., Rodd, V., Schultz, R., Simard, R., Singh, R. S., Sorrels, J., Sullivan, M., Tran, S., Trenholm, D., Trush,
- 551 J., Tucker, M. R., Turcotte, E., Vanniekerk, A., Vijan, P. N., Villanueva, J., Wang, C., Warncke, D. D.,
- 552 Watson, M. E., Wikoff, L., and Yeung, P.: Determination of pH of soils by different methods -
- 553 collaborative study, J. AOAC Int., 78, 310-324, 1995.
- 554 Krull, E. S., Baldock, J. A., and Skjemstad, J. O.: Importance of mechanisms and processes of the 555 stabilization of soil organic matter for modelling carbon turnover, Funct. Plant Biol., 30, 207-222, 2003.
- Lalonde, K., Mucci, A., Ouellet, A., and Gélinas, Y.: Preservation of organic matter in sediments promoted by iron, Nature, 483, 198-200, 2012.

- Lehmann, J., Solomon, D., Kinyangi, J., Dathe, L., Wirick, S., and Jacobsen, C.: Spatial complexity of soil organic matter forms at nanometer scales, Nature Geosci., 1, 238-242, 2008.
- 560 Madejova, J.: FTIR techniques in clay mineral studies, Vib. Spectrosc., 31, 1-10, 2003.
- Mayer, L. M., Schick, L. L., Hardy, K. R., Wagai, R., and McCarthy, J.: Organic matter in small
 mesopores in sediments and soils, Geochim. Cosmochim. Ac., 68, 3863-3872, 2004.
- Mayhew, S. G.: The redox potential of dithionite and SO⁻² from equilibrium reactions with flavodoxins,
 methyl viologen and hydrogen plus hydrogenase, Eur. J. Biochem., 85, 535-547, 1978.
- Mehra, O. P., and Jackson, M. L.: Iron oxide removal from soils and clays by a dithionite-citrate system
 buffered with sodium bicarbonate, In National Conference on Clay. Clay miner., 7, 317-327, 1960.
- Obrist, D.: Mercury distribution across 14 U.S. forests. part II: patterns of methyl mercury concentrations
 and areal mass of total and methyl mercury, Environ. Sci. Technol., 46, 5921-5930, 2012.
- 569 Obrist, D., Johnson, D. W., Lindberg, S. E., Luo, Y., Hararuk, O., Bracho, R., Battles, J. J., Dail, D. B.,
- 570 Edmonds, R. L., Monson, R. K., Ollinger, S. V., Pallardy, S. G., Pregitzer, K. S., and Todd, D. E.:
- 571 Mercury distribution across 14 US forests. part I: spatial patterns of concentrations in biomass, litter, and
- 572 soils, Environ. Sci. Technol., 45, 3974-3981, 2011.
- Obrist, D., Zielinska, B., and Perlinger, J. A.: Accumulation of polycyclic aromatic hydrocarbons (PAHs)
 and oxygenated PAHs (OPAHs) in organic and mineral soil horizons from four US remote forests,
 Chemosphere, 134, 98-105, 2015.
- Parikh, S. J., Goyne, K. W., Margenot, A. J., Mukome, F. N. D., and Calderon, F. J.: Soil Chemical
 Insights Provided through Vibrational Spectroscopy, Adv. Agron., 126, 1-148, 2014.
- Percival, H. J., Parfitt, R. L., and Scott, N. A.: Factors controlling soil carbon levels in New Zealand
 grasslands: Is clay content important?, Soil Sci. Soc. Am. J., 64, 1623-1630, 2000.
- Reeves, J. B., III: Mid-infrared spectral interpretation of soils: Is it practical or accurate?, Geoderma, 189,
 508-513, 2012.
- Regier, T., Krochak, J., Sham, T. K., Hu, Y. F., Thompson, J., and Blyth, R. I. R.: Performance and
 capabilities of the Canadian Dragon: the SGM beamline at the Canadian Light Source, Nuclear
 Instruments & Methods in Physics Research Section a-Accelerators Spectrometers Detectors and
 Associated Equipment, 582, 93-95, 2007.
- Riedel, T., Iden, S., Geilich, J., Wiedner, K., Durner, W., and Biester, H.: Changes in the molecular
 composition of organic matter leached from an agricultural topsoil following addition of biomass-derived
 black carbon (biochar), Org. Geochem., 69, 52-60, 2014.
- Riedel, T., and Weber, T. K.: The chemical potential of water in soils and sediments, Soil Sci. Soc. Am. J.,
 80, 79-83, 2016.
- Riley, W. J., Maggi, F., Kleber, M., Torn, M. S., Tang, J. Y., Dwivedi, D., and Guerry, N.: Long
 residence times of rapidly decomposable soil organic matter: application of a multi-phase, multicomponent, and vertically resolved model (BAMS1) to soil carbon dynamics, Geoscientific Model
 Development, 7, 1335-1355, 2014.

- Scharlemann, J. P. W., Tanner, E. V. J., Hiederer, R., and Kapos, V.: Global soil carbon: understanding
 and managing the largest terrestrial carbon pool, Carbon Manag., 5, 81-91, 2014.
- 597 Schmidt, M. W. I., Torn, M. S., Abiven, S., Dittmar, T., Guggenberger, G., Janssens, I. A., Kleber, M.,
- 598 Kogel-Knabner, I., Lehmann, J., Manning, D. A. C., Nannipieri, P., Rasse, D. P., Weiner, S., and
- Trumbore, S. E.: Persistence of soil organic matter as an ecosystem property, Nature, 478, 49-56, 2011.
- 600 Schumacher, M., Christl, I., Scheinost, A. C., Jacobsen, C., and Kretzschmar, R.: Chemical heterogeneity
- 601 of organic soil colloids investigated by scanning transmission X-ray microscopy and C-1s NEXAFS
- 602 microspectroscopy, Environ. Sci. Technol., 39, 9094-9100, 2005.
- Schwertmann, U. and Latham, M.: Properties of iron-oxides in some new caledonian oxisols, Geoderma,
 39, 105-123, 1986.
- Senesi, N., D'Orazio, V., and Ricca, G.: Humic acids in the first generation of EUROSOILS, Geoderma,
 116, 325-344, 2003.
- Simonetti, G., Francioso, O., Nardi, S., Berti, A., Brugnoli, E., Lugato, E., and Morari, F.:
 Characterization of humic carbon in soil aggregates in a long-term experiment with manure and mineral
 fertilization, Soil Sci. Soc. Am. J., 76, 880-890, 2012.
- Six, J., Callewaert, P., Lenders, S., De Gryze, S., Morris, S. J., Gregorich, E. G., Paul, E. A., and Paustian,
 K.: Measuring and understanding carbon storage in afforested soils by physical fractionation, Soil Sci.
 Soc. Am. J., 66, 1981-1987, 2002.
- Sollins, P., Homann, P., and Caldwell, B. A.: Stabilization and destabilization of soil organic matter:
 Mechanisms and controls, Geoderma, 74, 65-105, 1996.
- Solomon, D., Lehmann, J., Kinyangi, J., Liang, B. Q., and Schafer, T.: Carbon K-edge NEXAFS and
 FTIR-ATR spectroscopic investigation of organic carbon speciation in soils, Soil Sci. Soc. Am. J., 69,
 107-119, 2005.
- 618 Staff, S. S.: Soil taxonomy: A basic system of soil classification for making and interpreting soil surveys.
 619 2nd edition, Natural Resources Conservation Service. U.S. Department of Agriculture Handbook, 436,
 620 1999.
- Steffen, W., Noble, I., Canadell, J., Apps, M., Schulze, E. D., Jarvis, P. G., Baldocchi, D., Ciais, P.,
 Cramer, W., Ehleringer, J., Farquhar, G., Field, C. B., Ghazi, A., Gifford, R., Heimann, M., Houghton, R.,
- 623 Kabat, P., Korner, C., Lambin, E., Linder, S., Mooney, H. A., Murdiyarso, D., Post, W. M., Prentice, I. C.,
- Raupach, M. R., Schimel, D. S., Shvidenko, A., Valentini, R., and Terrestrial Carbon Working, G.: The terrestrial carbon cycle: implications for the Kyoto Protocol, Science, 280, 1393-1394, 1998.
- Tandy, S., Healey, J. R., Nason, M. A., Williamson, J. C., Jones, D. L., and Thain, S. C.: FT-IR as an
 alternative method for measuring chemical properties during composting, Bioresour. Technol., 101, 54315436, 2010.
- Tian, H., Chen, G., Zhang, C., Melillo, J. M., and Hall, C. A. S.: Pattern and variation of C:N:P ratios in
 China's soils: a synthesis of observational data, Biogeochemistry, 98, 139-151, 2010.
- 631 Tipping, E., Rey-Castro, C., Bryan, S. E., and Hamilton-Taylor, J.: Al(III) and Fe(III) binding by humic
- substances in freshwaters, and implications for trace metal speciation, Geochim. Cosmochim. Ac., 66,
- **633** 3211-3224, 2002.

- 634 Vogel, C., Mueller, C. W., Hoeschen, C., Buegger, F., Heister, K., Schulz, S., Schloter, M., and Koegel-
- Knabner, I.: Submicron structures provide preferential spots for carbon and nitrogen sequestration in soils,
 Nat. Commun., 5, 2014.
- Wagai, R. and Mayer, L. M.: Sorptive stabilization of organic matter in soils by hydrous iron oxides,
 Geochim. Cosmochim. Ac., 71, 25-35, 2007.
- 639 Wagai, R., Mayer, L. M., Kitayama, K., and Shirato, Y.: Association of organic matter with iron and
- aluminum across a range of soils determined via selective dissolution techniques coupled with dissolved
- nitrogen analysis, Biogeochemistry, 112, 95-109, 2013.
- Wang, X. C., Druffel, E. R. M., Griffin, S., Lee, C., and Kashgarian, M.: Radiocarbon studies of organic
 compound classes in plankton and sediment of the northeastern Pacific Ocean, Geochim. Cosmochim.
 Ac., 62, 1365-1378, 1998.
- 645 Werner, R. A., Bruch, B. A., and Brand, W. A.: ConFlo III An interface for high precision delta(13)C
- and delta(15)N analysis with an extended dynamic range, Rapid. Commun. Mass Sp., 13, 1237-1241,1999.
- White, A. F., and Blum, A. E.: Effects of climate on chemical-weathering in watersheds, Geochim.
 Cosmochim. Ac., 59, 1729-1747, 1995.
- Wilson, B. T., Woodall, C.W., and Griffith, D.M.: Imputing forest carbon stock estimates from inventory
 plots to a nationally continuous coverage, Carbon Balance and Manag., 8, 2013.
- Yang, W. H., Weber, K. A., and Silver, W. L.: Nitrogen loss from soil through anaerobic ammonium
 oxidation coupled to iron reduction. Nature Geosci., 5, 538-541, 2012.
- Zelles, L., Bai, Q. Y., Beck, T., and Beese, F.: Signature fatty acids in phospholipids and
 lipopolysaccharides as indicators of microbial biomass and community structure in agricultural soils, Soil
 Biol. and Biochem., 24, 317-323, 1992.
- Zhao, J. M., Huggins, F. E., Feng, Z., and Huffman, G. P.: Ferrihydrite-surface-structure and its effects on
 phase-transformation, Clay. Clay Miner., 42, 737-746, 1994.
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671 Figure Captions

- **Figure 1.** Concentrations of total carbon (TC), total organic carbon (TOC) and Fe-bound OC in 14 forest soils across the United States. TC, TOC, and Fe-bound OC contents are generally higher in soils with higher latitude (also shown in Figure 3). Duplicate measurements were conducted for each of two plots in every forest site. Error bars represent standard deviation of measurements of four replicates for each forest site.
- **Figure 2.** Concentration of reactive Fe and OC:Fe molar ratio in the U.S. forest soils.
- **Figure 3.** Relationships between TOC, concentration of Fe-bound OC, $f_{\text{Fe-TOC}}$ and ecogeographical parameters including latitude, elevation (asl), and temperature (annual mean).
- 680 Figure 4. Relationships between the fractions of iron-bound organic carbon (uncalibrated for
- loss of labile OC) and labile organic carbon and soil texture (i.e., fractions of sand, silt, and clay
- in forest soils). Values of Pearson correlation coefficients (r) and significance levels were given.
- 683 Figure 5. Attenuated total reflectance-Fourier transform infrared spectroscopy (ATR-FTIR)
- analysis for representative forest soils before (black line) and after Fe extraction (red line). All
- the spectra are background-calibrated. Among the 14 forest soils sampled in this study, we used
- 686 five different forest soils, with $f_{\text{Fe-OC}}$ ranging 5.6-57.8%.
- **Figure 6. A**. δ^{13} C of iron-bound and non-iron bound organic carbon for 14 U.S. forest sites. **B**. Correlation between $\Delta^{13}_{FeOC-nonFeOC}$ and molar ratio of OC:Fe.
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Forest ID	Abbr.	Location	Soil Order (US)	Soil Class ^a (FAO)	Climate Zone	LAT (°) ^b	LONG (°) ^c	Elevation (m asl)	Precip. ^d (mm y ⁻¹)	Temp ^e (°C)
Gainesville	GS	Gainesville, Florida	Spodosols	Podzols	Humid Subtropical	29.74	-82.22	50	1228	21.7
Oak Ridge	OR	Oak Ridge, Tennessee	Ultisols	Acrisols	Humid Subtropical	35.97	-84,28		1350	14.5
Ashland	AL	Ashland, Missouri	Alfisols	Luvisols & Greyzems	Humid Continental	38.73	-92.20	210	1023	13.9
Little Valley (post-fire)	LVF	Little Valley, Nevada	Entisols	Arenosols	Highland Climate	39.12	-119.93	2010	551	5.0
Little Valley	LV	Little Valley, Nevada	Entisols	Arenosols	Highland Climate	39.12	-119.93	2011	550	5.0
Marysville	MS	Marysville, California	Mollisols	Luvisols	Mediterranean climate	39.25	-121.28	386	775	16.9
Truckee (post-fire)	TKF	Truckee, California	Alfisols	Luvisols	Highland Climate	39.37	-120.1	1768	569	6.0
Truckee	ТК	Truckee, California	Alfisols	Luvisols	Highland Climate	39.37	-120.1	1767	568	5.9
Niwot Ridge	NR	Niwot Ridge, Colorado	Alfisols	Cambisols	Highland Climate	40.03	-105.55	3050	800	1.3
Hart	HT	Hart, Michigan	Spodosols	Podzols	Humid Continental	43.67	-86.15	210	812	7.6
Bartlett	BL	Bartlett, New Hampshire	Spodosols	Podzols & Lithosols	Humid Continental	44.0	-71.29	272	1300	4.5
Howland	HL	Howland, Maine	Spodosols	Luvisols	Humid Continental	45.20	-68.74	60	1040	6.7
Thompson I	TSI	Ravensdale, Washington	Inceptisols	Cambisols	Highland Climate	47.38	-121.93	221	1141	9.8
Thompson II	TSII	Ravensdale, Washington	Inceptisols	Cambisols	Highland Climate	47.38	-121.93	220	1140	9.8
a: Food and A	gricultu	re Organization	; b latitude; c	: longitude d:	annual mean prec	ipitation	; e: annual	mean tempe	erature.	

Table 1 Information for the 14 forest sites studied (Obrist et al., 2011, 2012, 2015)





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