



1 Iron-Bound Organic Carbon in Forest Soils: Quantification

2 and Characterization

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12 ABSTRACT

Iron oxide minerals play an important role in stabilizing organic carbon (OC) and regulating the 13 biogeochemical cycles of OC on the earth surface. To predict the fate of OC, it is essential to 14 completely understand the amount, spatial variability and characteristics of Fe-bound OC in 15 natural soils. In this study, we investigated the concentrations and characteristics of Fe-bound OC 16 in soils collected from 14 forests in the United States, and determined the impact of 17 ecogeographical variables and soil physicochemical properties on the association of OC and Fe 18 minerals. We found that Fe-bound OC contributed up to 57.8% of total OC (TOC) in forest soils. 19 Atomic ratios of OC:Fe ranged from 0.56 to 17.7 with values of 1-10 for most samples, and these 20 ratios indicate an importance of both sorptive and incorporative interactions. The fraction of Fe-21 bound OC in TOC (f_{Fe-OC}) was not related to the concentration of reactive Fe, which suggests that 22 the importance of association with Fe in OC accumulation was not governed by the concentration 23 of reactive Fe. Concentrations of Fe-bound OC and fre-OC increased with the latitude and reached 24 peak values at a site with a mean annual temperature of 6.6 °C. Attenuated total reflectance-Fourier 25 transform infrared spectroscopy (ATR-FTIR) and near-edge X-ray absorption fine structure 26 27 (NEXAFS) analyses revealed that Fe-bound OC was less aliphatic than non-Fe-bound OC. Fe-





bound OC also was more enriched in ¹³C compared to the non-Fe-bound OC, but C/N ratios did not differ substantially. In summary, ¹³C-enriched OC with less aliphatic carbon and more carboxylic carbon was associated with Fe minerals in the soils, with values of $f_{\text{Fe-OC}}$ being controlled by both sorptive and incorporative associations between Fe and OC. Overall, this study demonstrates that Fe oxides play an important role in regulating the biogeochemical cycles of C in forest soils, and uncovers the governing factors for the spatial variability and characteristics of Fe-bound OC.

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36 **1 Introduction**

Soil organic carbon (OC) in forests contributes 40% of the global carbon (C) mass, and is a vital component of C biogeochemical cycles (Eswaran et. al., 1999). Global warming can potentially accelerate the decomposition of forest soil OC, contributing to greenhouse gas emissions (Steffen et al., 1998). Alternatively, forest soils can act as strong sinks for OC, if appropriate management is implemented, such as forest harvesting and fire treatment (Eswaran et. al., 1999; Johnson and Curtis, 2001). Understanding the fate and stability of forest OC is important for evaluating and managing the global C cycle under the framework of climate change.

44 Currently, there is a large information gap concerning the stability and residence time of OC, contributing to the problem that the residence time of OC (ranging from months to hundreds 45 of years) is a major source of uncertainty in modeling and prediction of C cycles (Schmidt et al., 46 2011; Riley et al., 2014). Many concepts have been proposed to account for OC stabilization and 47 therefore residence times, including molecular recalcitrance, physical occlusion, and chemical 48 protection (Sollins et al., 1996; Krull et al., 2003; Baldock et al., 2004; Mayer et al., 2004; 49 50 Zimmerman et al., 2004; Schmidt et al., 2011). In general, the stability of OC is regulated by biogeochemical reactions occurring at the interfaces between OC, minerals, and microorganisms, 51 and further knowledge about the mechanism for OC stabilization is critical for building up process-52 53 based models to simulate and predict C cycles.

A number of lines of evidence suggest a key importance of iron oxide minerals in the stabilization of OC (Kalbitz et al., 2005; Kaiser and Guggenberger, 2007; Wagai and Mayer, 2007). Iron oxides have a relatively high sorption capacity for OC, with sorption coefficients for OC much higher than that of other metal oxides (Kaiser and Guggenberger, 2007; Chorover and Amistadi,





2001). Wagai and Mayer (2007) reported Fe-bound OC concentrations in soils up to 22 mg g⁻¹ soil,
contributing up to 40% of total OC (TOC) for most forest soils. Similarly, Lalonde et al. (2012)
found that Fe-bound OC contributed 22% of TOC in sediments. Studies have shown that Fe
minerals protect OC from degradation and inhibit mineralization of OC (Baldock and Skjemstad,
2000; Kalbitz et al., 2005). There is, however, no systematic study on the occurrence of Fe-bound
OC across different forests and its governing factors.

The overall goals of this study were to investigate the spatial variability of Fe-bound OC 64 across forest soils and the controlling factors, and to study the characteristics of Fe-bound OC in 65 respect to physicochemical properties of soils. In this study, we first quantified the concentration 66 of Fe-bound OC across 14 forest soils in the United States and analyzed the spatial distribution 67 and influences of ecogeographical factors. Second, we investigated the impact of soil 68 physicochemical properties on the Fe-OC associations. Third, we studied molecular characteristics 69 70 of Fe-bound OC versus non-Fe-bound OC, including how Fe-OC association influenced the chemical properties of OC and their stable isotope composition. Hence, this study provided a 71 systematic evaluation for the Fe-bound OC in United States forests, the influences of ecological 72 factors on the occurrence of Fe-bound OC, and the effects of association with Fe on the chemical 73 74 properties of OC.

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76 2. Methods & Materials

77 2.1 Chemicals and materials

Reagents used for Fe reduction experiments include sodium bicarbonate (NaHCO₃: SigmaAldrich, St. Louis, MO, USA), trisodium citrate dihydrate (Na₃C₆H₅O₇•2H₂O: Acros Organics,
New Jersey, USA), and sodium dithionite (Na₂S₂O₄: Alfa Aesar, Ward Hill, MA, USA). All
chemicals used were analytical grade.

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83 2.2 Soil sample collection, primary characterization and pretreatment

Soil samples were collected from 14 forest sites in the United States (Obrist et al., 2011,
2012, 2015). The abbreviations and the basic information for the sites are summarized in Table 1.
More detailed information on the sites and sampling protocols can be found in previous
publications (Obrist et al., 2011, 2012, 2015). Soil texture was analyzed by an ASTM 152-type





hydrometer at the Soil Forage and Water Analysis Laboratory at Oklahoma State University
(Obrist et al., 2011). The soil pH was measured by mixing soil particle with deionized (DI) water
in a solid/solution ratio of 1:1 (Kalra, 1995). Soil samples used in the experiments in this study
were ground to < 500 µm and freeze-dried.

92

93 Table 1

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95 2.3 Total C (TC), TOC and stable C isotope analyses

96 TC, TOC and stable C isotopic compositions of soil samples were analyzed using a Eurovector elemental analyzer (Eurovector SPA, Milan, Italy) interfaced to a Micromass IsoPrime 97 stable isotope ratio mass spectrometer (Micromass UK Ltd., Manchester, UK). Acetanilide (71.09 % 98 C by weight) was used as a standard compound to establish a calibration curve between mass of C 99 100 and the m/z 44 response from the mass spectrometer. In this study, the concentration of TC and TOC were expressed as weight %. Stable C isotope analyses were performed after the method of 101 102 Werner et al. (1999), with results reported in the usual delta notation in units of % vs. Vienna Pee Dee Belemnite (VPDB). For TOC analysis, soil samples were acidified with 1 M HCl with the 103 104 solution/solid ratio of 1 mL solution/0.5 g soil and heated at 100°C for 1 hour. The treatment was repeated three times until there was no further effervescence upon acid addition, after which the 105 samples were dried and analyzed. 106

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108 2.4 Nitrogen (N) analysis

The N concentration of each sample was analyzed using a Eurovector elemental analyzer. Acetanilide (10.36 % N by weight) was used as a standard compound to establish a calibration curve between mass of N and the response of the thermal conductivity detector in the elemental analyzer. Total N and non-Fe-bound N concentrations were measured before and after a Fe reduction release treatment for each sample.

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115 2.5 Analysis of Fe-bound OC

The concentration of Fe-bound OC was quantified by an established Fe reduction release
method, commonly known as DCB extraction involving sodium dithionite, citrate and bicarbonate
(Mehra and Jackson, 1960; Wagai and Mayer, 2007; Lalonde et al., 2012). The DCB extraction is





assumed to extract most free Fe oxides (i.e. goethite, hematite, ferrihydrite and others) existing in 119 soils, but should not extract structural Fe in clay minerals (Mehra and Jackson, 1960; Wagai and 120 Mayer et al., 2007; Lalonde et al., 2012). In this study, we followed the specific protocol detailed 121 in Lalonde et al. (2012). An aliquot (0.25 g) of soil was mixed with 15 mL of buffer solution at 122 pH 7 (containing 0.11 M bicarbonate and 0.27 M trisodium citrate), and then heated to 80°C in a 123 water bath. The reducing agent sodium dithionite was added to the samples with final 124 concentration of 0.1 M, and maintained at 80°C for 15 min. The samples were then centrifuged at 125 10,000 rpm for 10 min, the supernatant was removed, and the residual particles were rinsed using 126 127 5 mL of DI water. The rinse/centrifuge process was performed three times. The residual particles were freeze-dried and analyzed for TC and TOC concentrations and $\delta^{13}C$ composition. The mass 128 of residual particles were used to calculate the OC concentration associated with non-Fe minerals. 129

The background release of OC during the heating process was measured following the 130 method in Lalonde et al. (2012), where sodium citrate and dithionite were replaced by sodium 131 132 chloride with the same ionic strength. An aliquot (0.25 g) of dry soil was mixed with 15 mL of 1.6 M NaCl and 0.11 M NaHCO₃, and heated to 80°C. Then 0.22 g of NaCl was added, and the solution 133 134 was maintained at 80°C for 15 min. The samples were then centrifuged at 10,000 rpm and rinsed three times, and freeze-dried before analysis. The mass of residual particles was used to calculate 135 136 the concentration of OC released by heating to 80°C. In preliminary experiments, we found that the solution pH increased rapidly during the heating-extraction process with bicarbonate and 137 sodium chloride only, and the increased pH values facilitated the release of additional OC. Hence, 138 we used a lower initial pH of 6 to compensate for the shift to higher pH during heating. To validate 139 the measurement for the concentration of OC released during heating, we also tested the release of 140 OC using a phosphate buffer (same ionic strength) in lieu of the bicarbonate buffer, which can 141 maintain a pH of 7 during heating. Our results showed that the concentration of OC released was 142 similar for both the bicarbonate and phosphate buffer extraction reactions (Supplementary Material, 143 144 Fig. S1).

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146 **2.6 Quantification of reactive Fe**

147 The concentration of reactive Fe in soils was determined by analyzing the Fe released 148 during the DCB reduction process. After the reduction treatment, the supernatant of each sample 149 was filtered using a $0.2 \,\mu m$ syringe filter (cellulose acetate), and analyzed for Fe concentration by





- 150 inductively coupled plasma atomic emission spectroscopy (Varian-Vista AX CCD, Palo Alto,
- 151 CA, USA) at an optical absorption wavelength of 259.9 nm.
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153 **2.7** Attenuated total reflectance-Fourier transform infrared spectroscopy (ATR-FTIR)

ATR-FTIR analysis to characterize the molecular composition of OC was performed for original soil samples and residual soils after DCB extraction using a Thermo Scientific Nicolet 6700 FTIR (Waltham, MA). Dry soil samples were placed directly on the crystal and forced to contact well with the crystal. Spectra were acquired at the resolution of 4 cm⁻¹ based on 100 scans. Data collection and baseline correction were accomplished using OMNIC software version 8.3.103.

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160 2.8 Near-edge X-ray absorption fine structure (NEXAFS) analysis

For further characterization of chemical structure of OM, carbon (1s) K-edge NEXAFS 161 analyses were performed for select soil samples, i.e. for soils with the highest and lowest values 162 of the fraction of Fe-bound OC to TOC. The soil particles were suspended in DI water and 163 deposited on an Au-coated silicon wafer attached to a Cu sample holder. Before analysis, samples 164 were dried in a vacuum desiccator. The X-ray-based experiments were performed on the Spherical 165 Grating Monochromator (SGM) beamline at the Canadian Light Source (Saskatoon, Canada) 166 (Regier et al., 2007). The energy scale was calibrated using citric acid (absorption at 288.6 eV). 167 Major technical parameters and set-up for the beamline include: X-ray energy ranges 250-2000 168 eV; 45 mm planer undulator; 1000 µm×100 µm spot size; silicon drift detectors (SDD); a titanium 169 170 filter before the sample; entrance and exit slit gaps of 249.9 µm and 25 µm (Gillespie et al., 2015). Carbon 1s spectra were acquired by slew scans from 270 to 320 eV at 20 s dwell time and 20 scans 171 per sample on a new spot. For data normalization, I₀ was collected by measuring the scatter of the 172 incident beam from a freshly Au-coated Si wafer using SDD. Before the I_0 normalization, the pre-173 174 edge baseline was adjusted to near zero to remove the scatter in the sample data (Gillespie et al., 2015). 175

176 3. Results and Discussion

3.1 Concentration of Fe-bound OC

This study covered five major forest types in North America, including Spruce-Fir, Pine, Oak, Chaparral, and Maple-beech-birch forests distributed between 29° and 47° N. For the 14





forest soils, TC concentrations ranged between 1.5 ± 0.1 and $8.3\pm2.1\%$ (all percentages given are 180 weight-based), and TOC concentrations ranged between 1.3±0.3 and 6.2±2.9%, which are 181 comparable to values previously reported for North American forests (Wagai and Mayer, 2007; 182 Wilson et al., 2013). Bicarbonate extraction-calibrated Fe-bound OC concentrations ranged from 183 0.3 to 1.9%, with the fraction of Fe-bound OC to TOC ($f_{\text{Fe-OC}}$) averaging 37.8±20.0% (Fig. 1, 184 Supplementary Material, Table S1). Forest HL (Maine) had the highest fre-oc of 57.8%, while 185 forests GS (Florida) and OR (Tennessee) had f_{Fe-OC} values below detection limits (i.e., below 0.6%). 186 Based on an estimate that 1502 Pg (Pg=1×10¹⁵ g) of OC is stored in terrestrial soils (Scharlemann, 187 188 et al., 2014), scaling up these results to a global estimate would yield 538.5±271.5 Pg of Fe-bound OC residing in terrestrial soils. This is a very large pool, e.g. compared to an estimated 19-45 Pg 189 Fe-bound OC that reside in global surface marine sediments (Lalonde et al., 2012). As a major 190 component of the global C pool, Fe-bound OC in terrestrial soils therefore is expected to play a 191 192 key role in global C biogeochemical cycles.

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194 Fig. 1

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196 **3.2 Fe-OC association**

The values of $f_{\text{Fe-OC}}$ were influenced not only by the concentration of reactive Fe, but also 197 by the type of association between Fe and OC. In this study, the concentration of reactive Fe in 198 forest soils ranged from 0.1 mg g⁻¹ to 19.3 mg g⁻¹, which is relatively low compared to values of 199 200 reactive Fe of up to 180 mg g⁻¹ reported previously (Wagai and Mayer, 2007; Wagai et al., 2013) (Fig. 2A). A Mollisol in forest sites MS (California) had the highest concentration of reactive Fe, 201 when a Spodosol in forest site GS (Florida) had the lowest reactive Fe concentration. To our 202 surprise, there was no significant correlation between $f_{\text{Fe-OC}}$ and the concentration of reactive Fe 203 204 (Pearson Correlation Coefficient r=-0.418, p=0.137, Fig. 2B). This suggests that the proportion of Fe-bound OC is not strongly controlled by the reactive Fe concentration. 205

The OC:Fe molar ratio ranged from 0.56 to 17.7 for all 14 soils, with a value between 1 and 10 for 10 soils (Fig. 2A). Previous studies have suggested that the OC:Fe molar ratio can be used as an indicator for the type of association between Fe oxides and OC, with lower values indicating sorptive interactions, while higher values indicate incorporation of OC within Fe oxides (Wagai et al., 2007; Guggenberger and Kaiser, 2003). The highest sorption capacity measured for





OC onto Fe oxide corresponds to an OC:Fe molar ratio = 1.0 (Kaiser and Guggenberger, 2006), 211 but by incorporation and co-precipitation of Fe oxide OC:Fe molar ratio can reach much higher 212 values (Guggenberger and Kaiser, 2003). With OC:Fe molar ratios generally between 1-10 for 213 most of the forest soils in this study, we propose that incorporation of OC into Fe oxides plays a 214 major role in the accumulation of Fe-bound OC exceeding sorption by at least a factor of 1 to 10 215 (Wagai and Mayer, 2007; Lalonde, 2012). However, for the HT (Michigan), HL (Maine) and TKF 216 (California) forest soils, the OC:Fe molar ratios were even higher than 10 with maximum value of 217 17.8 (Fig. 2A), implying that incorporation of OC into Fe oxides dominated at these sites. The 218 219 value of OC:Fe ratio was not related to the concentration of reactive Fe, and varied a lot for soils with similar concentration of total reactive Fe (Fig. 2B). This indicates the type of interactions 220 between OC and Fe was not governed by the amount of Fe. The OC:Fe ratio is potentially regulated 221 by the mineral phases of Fe, as poorly-crystalline Fe oxide has a higher capacity to bind with OC 222 223 than crystalline Fe minerals (Eusterhues et al., 2014). When sorption dominates the interactions between OC and Fe, OC:Fe can also be influenced greatly by the particle size and surface area of 224 Fe oxides (Gu et al., 1995). Further investigations are needed to determine the factors that control 225 the OC:Fe ratio, and also f_{Fe-OC} values for soils. Nevertheless, the lack of (or poor) relationship 226 shown here between the concentration of Fe-bound OC and Fe concentrations demonstrates the 227 limitations associated with predicting and modeling the behavior of C in soils based on the Fe 228 concentrations in soils alone. 229

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233 3.3 Spatial variance and ecogeographical factors

We analyzed the influences of ecogeographical factors on the occurrence of Fe-bound OC in forest soils (Fig. 3). There was a significant correlation between the TOC concentration and latitude (Pearson correlation coefficient p=0.619, r=0.018), a pattern commonly observed due to lower microbial activity and turnover rates of C at higher, colder latitudes (Davidson and Janssens, 2006). Concentration of reactive Fe, if excluding soil MS in California, it was also significantly related to latitude (p=0.824, r=0.001). Both concentrations of Fe-bound OC and f_{Fe-OC} also were correlated with latitude (p=0.523, r=0.053; p=0.525, r=0.054). Among our samples, soil in forest HL in

²³¹ Fig. 2.





Maine, one of the three northern-most site with latitude of 45° , had the highest fre-oc of 57.8%. In 241 forest GS in Florida with lowest latitude of 29.7°, the f_{Fe-OC} were below detection limits, possibly 242 due to the low concentration of reactive Fe (0.08 mg g^{-1}). Hence, increase in latitude both increased 243 concentrations of TOC in soil as well concentrations of Fe-bound OC, suggesting increased 244 interactions between Fe oxide and OC at higher latitudes. There were no clear trends in TOC or 245 Fe-OC interactions with longitude. For elevation, we separated two groups of samples, with one 246 group located below 1000 m (asl) and the other group above (mainly around 2000 and 4000 asl). 247 Concentrations of TOC and Fe-bound OC, however, were not significantly different between the 248 249 two groups. There were no clear trends with precipitation either, although others have reported positive relationships between mean annual precipitation and soil TOC concentration at a global 250 scale (Amundson, 2001). Both concentrations of Fe-bound OC and f_{Fe-OC} reached highest value 251 with mean annual temperatures at 6.6°C. The increased annual mean temperature increased the 252 253 concentration of Fe-bound OC and $f_{\text{Fe-OC}}$ when it was below 6.6°C, then decreased when annual mean temperature was over 6.6°C. Our results therefore suggest a possibly ideal temperature range 254 where Fe-OC interactions may be particularly pronounced. Finally, the study covered 7 major soil 255 orders, i.e. Alfisols (sample number n=3), Spodosols (n=4), Mollisols (n=1), Inceptisols (n=2), 256 Entisols (n=2), Gelisols (n=1), and Ultisols (n=1). Although there are limited replications in many 257 of these soil orders, highest concentration of Fe-bound OC were observed in Spodosols. Regarding 258 $f_{\text{Fe-OC}}$, the ratios also were highest in Spodosols, possibly indicating a particular importance of Fe-259 bound OC in this soil type which occupies 3.5% of US land areas and 4% global ice-free land (Soil 260 261 Survey Staff, 1999). However, due to the limited number of samples for each soil order, these findings warrant further confirmation. 262 263

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264 Fig. 3

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266 **3.4 Impact of soil physicochemical properties on Fe-OC association**

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Soil texture can potentially influence the accumulation of Fe-bound OC. Figure 4 demonstrates that the fraction of non-calibrated Fe-bound OC showed a significant positive correlation vs. fraction of sand (r=0.72, p<0.001), and negative correlations vs. fraction of silt (r=-0.697, p<0.001) and fraction of clay (r=-0.616, p<0.001). There were similar correlations between





labile OC, and the fraction of sand (r=0.72, p<0.001), silt (r=0.72, p<0.001) and clay (r=0.72, 272 p < 0.001). However, the calibrated Fe-bound OC had no significant correlation vs. any of the 273 texture fractions. These correlations indicate that the labile OC was mainly associated with the 274 sand component of forest soils, but that the soil texture did not affect the Fe-bound OC. There has 275 been debate on the relative roles of sand, clay and silt in the stabilization of OC in soil (Percival et 276 al., 2000; Six et al., 2002; Eusterhues et al., 2005; Vogel et al., 2014). Eusterhues et al. (2005) 277 found a relationship between the resistance of organic matter to oxidative degradation and the clay 278 279 concentration in soils, suggesting the importance of clay minerals in the stabilization and 280 accumulation of soil OC. In contrast, Percival et al. (2000) found that the clay mineral fraction explained little of the variation in the accumulation of OC across a range of soil types in New 281 Zealand. Vogel et al. (2014) found that less than 20% of clay mineral surfaces were covered by 282 the sorption of OC, indicating that a limited proportion of clay mineral surface contributed towards 283 284 the stabilization of OC. Our results suggest that the Fe oxide-mediated stabilization of OC was not related to the size/aggregation-based process, although the labile carbon concentrations increased 285 with the fraction of sand in the soils. 286

287

288 Fig. 4

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The Fe-OC association can also be influenced by the soil pH, which affects the mineral 290 phases of Fe oxides and their surface charge, and their interactions with OC. For our soil samples, 291 292 the soil pH ranged from 4.1 to 6.3, similar to measurements by Wagai and Mayer (2007) for North America soils. There was no significant correlation between the $f_{\text{Fe-OC}}$ and soil pH, e.g. the HL 293 (Maine) soil with pH of 4.4 had the highest $f_{\text{Fe-OC}}$ of 57.8%, while the TS(II) (Washington) soil 294 with a similar pH of 4.5 only had a $f_{\text{Fe-OC}}$ of 7.4%. For soils with pH ranging from 4.9 to 5.8, $f_{\text{Fe-OC}}$ 295 296 did not change correspondingly. Contrastingly, values of OC:Fe molar ratios were significantly influenced by the soil pH; expect for one outlier sample of TS(II) (Washington) soil, there was a 297 significant negative correlation between the OC:Fe molar ratio and soil pH (r=-0.477, p=0.09) 298 (Supplementary Material, Fig. S2). This may be due to the lower pH values favoring the 299 300 complexation and precipitation of Fe with OC, while higher pH favors sorptive interactions between Fe minerals and OC (Tipping et al., 2002). If comparing samples with similar pH, the 301 soils with higher TOC had higher OC:Fe molar ratios, e.g. the GS soil (TOC = 1.1%) with pH of 302





4.7 had an OC:Fe molar ratio = 8.5, while the HT (Michigan) soil (TOC = 3.0%) with similar pH
of 4.7 had an OC:Fe molar ratio = 17.1. This was consistent with the concept model that the form
of Fe was dominated by the Fe-OC co-precipitated complex with relatively higher OC supply
(Schwertmann et al., 1986).

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308 **3.5 Molecular characteristics of Fe-bound OC**

The chemical composition of Fe-bound OC can be substantially different from non-Fe-309 bound OC (Adhikari and Yang, 2015) with broad implications on the C biogeochemical cycles, 310 311 although such differences so far have received limited attention. We analyzed the difference in chemical composition of Fe-bound OC compared to non-Fe-bound OC using ATR-FTIR analysis 312 (Fig. 5). Overall, there were limited fingerprint peaks for OC, because of the low concentration of 313 TOC and technical challenge for analyzing whole soil particles with FTIR (Calderon et al., 2011; 314 Simonetti et al., 2012). Reeves (2012) demonstrated that FTIR analysis of mineral soils in the 315 ranges of 1600-1750 and 2800-3000 cm⁻¹ only can be used to study OC. Peaks in the range of 500-316 1200 cm⁻¹ indicate the presence of clav or other Fe/Al minerals (Fig. 5) (Madejova, 2003; Harsh 317 et al., 2002; Parikh et al., 2014), such as kaolinite or montmorillonite at 850-1200 cm⁻¹ (Madejova, 318 2003). Absorption at 850-1200 cm⁻¹ can also be due to the presence of polysaccharides, but 319 definitive identification of polysaccharides is not possible in the presence of minerals (Senesi et 320 al., 2003; Tandy et al., 2010). The spectra in the range of 1600-1750 cm⁻¹ normally contain 321 fingerprint peaks for functional groups of amides, carboxylates and aromatics (Parikh et al., 2014), 322 but we did not detect any significant peaks in this range. In the range of 2800-3000 cm⁻¹, there 323 were no significant peaks for the original soil samples, but after Fe extraction we detected 324 significant peaks at 2850 and 2930 cm⁻¹, which are characteristic for the presence of aliphatic 325 carbon. The substantial differences in spectra before and after Fe extraction indicate that aliphatic 326 327 OC was enriched in the residual soils after extraction. Other functional groups, such as aromatic carbon and hydrophilic functional groups, were more strongly associated with Fe minerals and 328 removed during the Fe extraction, as hydrophilic functional groups can form inner-sphere 329 coordination complexation with iron oxides, and aromatic carbon has electron donor-acceptor 330 331 interactions with iron oxides (Gu et al., 1995; Axe and Persson, 2001).

- 332
- 333 Fig. 5





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Furthermore, we analyzed the C 1s NEXAFS spectra of two original, non-extracted soils 335 with the highest and lowest values of $f_{\text{Fe-OC}}$, i.e. HL (Maine) ($f_{\text{Fe-OC}}=57.8\%$) and OR (Tennessee) 336 (fre-oc non-detectable) (Supplementary Material, Fig. S3). Three major fingerprint peaks were 337 detected for both soils, including peaks at 285.3, 287.0 and 288.7 eV, which are corresponding to 338 aromatic carbon, aliphatic carbon and carboxylic carbon, respectively (Schumacher et al., 2005; 339 Solomon et al., 2005; Lehmann et al., 2008). The OR (Tennessee) soil had a more substantial 340 signal at 287.0 eV than the HL (Maine) soil, indicating a higher aliphatic carbon concentration in 341 342 the OR (Tennessee) soil compared to the HL (Maine) soil. Ratio of carboxylic carbon to aromatic carbon (peak height) was 3.8 for HL (Maine) and 1.0 for OR (Tennessee), suggesting that the HL 343 (Maine) soil with higher *f*_{Fe-OC} has relatively more carboxylic carbon compared to aromatic carbon. 344 Hence, the C1s NEXAFS spectra suggest that the soil with the higher fre-oc has higher 345 concentration of carboxylic C, while the soil with the lower fre-OC value has a higher aliphatic C 346 concentration. This result is consistent with the comparison of ATR-FTIR spectra in soils before 347 and after Fe extraction, providing evidence that Fe oxides are mainly associated with more 348 hydrophilic and carboxylic carbon, while non-Fe-bound OC was more aliphatic. 349

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To further investigate the relationships between soil OC and Fe minerals, we analyzed the stable C isotopic compositions (δ^{13} C) of Fe-bound vs. non-Fe-bound OC (i.e., the residual OC after DCB extraction). The δ^{13} C for original soil samples ranged from –24.5‰ to –27.5‰, and the values for non-Fe-bound OC were –25.1‰ to –28.0‰. The δ^{13} C for Fe-bound OC was calculated by combined isotope-mass balance (equation (1))

$$356 \qquad \delta^{13}C_{\text{TOC}} \times \text{TOC} = \delta^{13}C_{\text{labile}} \times \text{OC}_{\text{labile}} + \delta^{13}C'_{\text{Fe-OC}} \times \text{OC'}_{\text{Fe}} + \delta^{13}C_{\text{non-Fe-OC}} \times \text{OC}_{\text{non-Fe}}$$
(1)

where TOC is the concentration of total organic carbon, OC_{labile} is the concentration of labile OC (extractable by bicarbonate buffer), OC_{non-Fe} is the concentration of non-Fe-bound OC (residual OC after Fe extraction), and OC'_{Fe} is the concentration of Fe-bound OC (excluded the labile OC); $\delta^{13}C_{TOC}$ is $\delta^{13}C$ for bulk OC, $\delta^{13}C_{labile}$ is $\delta^{13}C$ for labile OC, $\delta^{13}C'_{Fe-OC}$ is $\delta^{13}C$ for Fe-bound OC, $\delta^{13}C_{non-Fe-OC}$ is $\delta^{13}C$ for non-Fe-bound OC. However, it is difficult to directly resolve the $\delta^{13}C_{labile}$ and $\delta^{13}C'_{Fe-OC}$ using this equation. We simplified it to equation (2):

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$$\delta^{13}C_{Fe-OC} = \frac{\left(\delta^{12}C_{TOC} \times TOC \cdot \delta^{12}C_{non-Fe-OC} \times OC_{non-Fe}\right)}{OC_{Fe}}$$
(2)





where $\delta^{13}C_{\text{Fe-OC}}$ is $\delta^{13}C$ for Fe-bound OC (including the labile OC), $\delta^{13}C_{\text{TOC}}$ is $\delta^{13}C$ for bulk OC, 364 δ^{13} Cnon-Fe-OC is δ^{13} C for non-Fe-bound OC, TOC is the concentration of total organic carbon, OCnon-365 Fe is the concentration of non-Fe-bound OC, and OCFe is the concentration of Fe-bound OC. The 366 δ^{13} C for Fe-bound OC was heaviest for the TKF (California) soil with a value of -23.0‰, and the 367 lightest for the GS (Florida) forest at -27.0%. Across all study sites, Fe-bound OC was relatively 368 enriched in ¹³C (1.5±1.2‰ heavier) compared to the non-Fe-bound OC. However, there is also a 369 contribution of labile OC to the Fe-bound OC, where labile OC is the OC extracted during the 370 dithionite-absent extraction described earlier). The δ^{13} C value for labile OC can be calculated using 371 372 equation (3): $\delta^{^{13}}C_{labile} = \frac{(\delta^{^{13}}C_{TOC} \times TOC \cdot \delta^{^{13}}C_{non-labile} \times OC_{non-labile})}{OC_{labile}}$ 373 (3)

where $\delta^{13}C_{labile}$ is $\delta^{13}C$ for labile OC, $\delta^{13}C_{TOC}$ is $\delta^{13}C$ for bulk OC, $\delta^{13}C_{non-labile}$ is $\delta^{13}C$ for non-labile OC, OC_{non-labile} is the concentration of non-labile OC, and OC_{labile} is the concentration of labile OC. Calculated values of $\delta^{13}C_{labile}$ range from -23.4% to -30.3%, and were lighter than the values for $\delta^{13}C_{Fe-OC}$. Although it is not reliable to quantitatively calculate the $\delta^{13}C$ for Fe-bound OC subtracting the influences of labile OC, these results indicate that the true value for $\delta^{13}C_{Fe-OC}$ should be even somewhat heavier than the results presented in Fig. 6.

Our results demonstrate that Fe-bound OC was enriched in ¹³C compared to the non-Fe-380 bound OC in forest soils, which is consistent with results for sediments, where Fe-bound OC was 381 1.7±2.8‰ heavier than non-Fe-bound OC (Lalonde et al., 2012) (Fig. 6A). Wang et al. (1998) 382 have shown that ¹³C-enriched organic matter in sediments was enriched with O and N (due to the 383 presence of compounds such as proteins and carbohydrate groups), while the lipid fraction was 384 relatively ¹³C-depleted. Similarly, compound-specific isotopic analyses have shown that O/N-rich 385 constituents, such as cellulose, hemi-cellulose and amino acids, are ¹³C-enriched compared to 386 hydrocarbons (Glaser, 2005), and these ¹³C-enriched O/N-rich compounds can associate with Fe 387 oxide extensively through inner-sphere coordination interactions (Parikh et al., 2014). The value 388 of $\Delta^{13}_{\text{FeOC-nonFeOC}}$ (= $\delta^{13}_{\text{CFe-OC}}$ - $\delta^{13}_{\text{Cnon-Fe-OC}}$) (difference in δ^{13}_{C} for Fe-bound OC and non-Fe-389 bound OC) was inversely correlated with the molar ratio of OC:Fe (r=-0.53, p=0.05, Fig. 6B). 390 These relationships suggest that the enrichment in ¹³C was to some degree related to the OC:Fe 391 ratio, with lower OC:Fe leading to higher enrichment in ¹³C. As discussed previously, lower OC:Fe 392 393 ratios indicate an increased contribution from sorptive interactions of OC with Fe minerals as compared to incorporation of OC within iron oxides and OC, and these sorptive interactions 394





between O/N-rich organic compounds and Fe oxide results in the enrichment of ¹³C of Fe-bound
OC vs. non-Fe-bound OC. Previous studies have attributed the stability of relatively labile and
reactive compounds, such as amino acids and sugars, to their interactions with minerals (Schmidt
et al., 2011), and our results demonstrated the importance of sorption to Fe minerals in increasing
the stability of relatively reactive labile compounds.

400

401 Fig. 6

402

403 Nitrogen (N)-containing functional groups are potentially important for the association between OC and Fe oxides, although the concentrations of N are much lower than C. The bulk soil 404 contained 0.05-0.45 % N, while the non-Fe-bound component (i.e. the residual solid after DCB 405 extraction) contained 0.06-0.32 % N. Concentrations of Fe-bound N, calculated by difference, 406 407 ranged up to 0.13 %. However, it is important to note that this number is based without a calibration for labile N that may be removed by the dithionite-free DCB extraction (data not available). There 408 were significant correlations between C and N concentrations for both bulk soils (r=0.847. p<0.001: 409 Supplementary Material, Fig. S4) and the non-Fe-bound residual components (r=0.858, p<0.001: 410 411 Supplementary Material, Fig. S4), with molar C/N ratios of 14.2±2.6 and 13.7±2.3 for bulk and non-Fe-bound OC, respectively. These C/N values are essentially identical to a previously 412 observed molar C/N ratio = 14.3 for a large set of world-wide soils samples (Cleveland et al., 413 2007), and a molar C/N ratio = 14.4 for OC-rich samples in China (Tian et al., 2010). This result 414 415 suggests that C/N ratios for Fe-bound OC did not differ from that of non-Fe-bound OC, assuming that the labile carbon did not have a substantially different C/N ratio. Therefore, in contrast to the 416 417 ¹³C enrichment observed for Fe-bound OC, the interactions with Fe minerals did not affect the C/N ratio substantially. 418

419

420 4. Conclusion

Fe oxides are recognized as an important mineral phase regulating the amount and characteristics of OM in forest soils. The spatial variability of Fe-bound OC is governed by the geographical factors, such as latitude and annual mean temperature, and also the soil physicochemical properties.





- 424 Chemical composition of Fe-bound OC is substantially different from the rest compartment of soil
- 425 OC. Overall, this study provided a comprehensive investigation into the spatial variance of Fe-
- 426 bound OC in forest soils, its governing factors, and how the Fe-OC associations affect the chemical
- 427 characteristics of OC. As a substantial amount of OC was bound to Fe oxides, and association with
- 428 Fe affected the quality of OC, Fe oxides can regulate the biogeochemical cycles of carbon and its
- 429 response to climate change.
- 430

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- 438

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605 Figure Captions

- 606 Figure 1. Concentrations of total carbon (TC), total organic carbon (TOC) and Fe-bound OC in
- 607 14 forest soils across the United States. Duplicate measurements were conducted for each of two
- plots in every forest site. Error bars represent standard deviation of measurements of four replicates
- 609 for each forest site.
- 610 Figure 2. A Concentration of reactive Fe and OC:Fe molar ratio in US forest soils. B Relationship
- between the fraction of Fe-bound OC in TOC $(f_{\text{Fe-OC}})$ / OC:Fe molar ratio and reactive Fe
- 612 concentration in US forest soils.
- **Figure 3.** Correlation between the TOC, reactive Fe, concentration of Fe-bound OC, *f*_{Fe-TOC}, OC:Fe
- and ecogeographical parameters including latitude, longitude, elevation (asl), precipitation (mean
- annual) and temperature (annual mean).
- 616 Figure 4. Correlation of the fractions of iron-bound organic carbon (uncalibrated and calibrated
- for loss of labile OC) and labile organic carbon vs. fractions of sand, silt, and clay in forest soils.
- **Figure 5**. Attenuated total reflectance-Fourier transform infrared spectroscopy (ATR-FTIR)
- analysis for representative forest soils before (black line) and after Fe extraction (red line). All the
- spectra are background-calibrated. Among the 14 forest soils sampled in this study, we used five
- 621 different forest soils, with $f_{\text{Fe-OC}}$ ranging 5.6-57.8%.
- **Figure 6.** A. δ^{13} C of total organic carbon and non-iron bound organic carbon for 14 U.S. forest
- 623 sites. **B**. Correlation between $\Delta^{13}_{FeOC-nonFeOC}$ and molar ratio of OC:Fe.
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- 625
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- 627



Forest ID	Abb r.	Location	Soil Order	Climate Zone	Precip. ^a (mm y ⁻¹)	Temp ^b (°C)	LAT(°)c	LONG(°) ^d	Elevation (m asl)
Ashland	AL	Ashland, Missouri	Alfisols	Humid Continental	1023	13.9	38.73	-92.20	210
Bartlett	BL	Bartlett, New Hampshire	Spodosols	Humid Continental	1300	4.5	44.0	-71.29	272
Marysvil le	MS	Marysville, California	Mollisols	Mediterranea n climate	775	16.9	39.25	-121.28	386
Gainesvi lle	GS	Gainesville, Florida	Spodosols	Humid Subtropical	1228	21.7	29.74	-82.22	50
Oak Ridge	OR	Oak Ridge, Tennessee	Ultisols	Humid Subtropical	1350	14.5	35.97	-84,28	
Little Valley (post- fire)	LV F	Little Valley, Nevada	Entisols	Highland Climate	551	5.0	39.12	-119.93	2010
Little Valley	LV	Little Valley, Nevada	Entisols	Highland Climate	550	5.0	39.12	-119.93	2011
Truckee (post- fire)	TK F	Truckee, California	Alfisols	Highland Climate	569	6.0	39.37	-120.1	1768
Truckee	ТК	Truckee, California	Alfisols	Highland Climate	568	5.9	39.37	-120.1	1767
Niwot Ridge	NR	Niwot Ridge, Colorado	Alfisols	Highland Climate	800	1.3	40.03	-105.55	3050
Hart	HT	Hart, Michigan	Spodosols	Humid Continental	812	7.6	43.67	-86.15	210
Howland	HL	Howland, Maine	Spodosols	Humid Continental	1040	6.7	45.20	-68.74	60
Thomps on I	TSI	Ravensdale, Washington	Inceptisols	Highland Climate	1141	9.8	47.38	-121.93	221
Thomps on II	TSI I	Ravensdale, Washington	Inceptisols	Highland Climate	1140	9.8	47.38	-121.93	220

a: annual precipitation; b: annual mean temperature; c latitude; d: longitude.

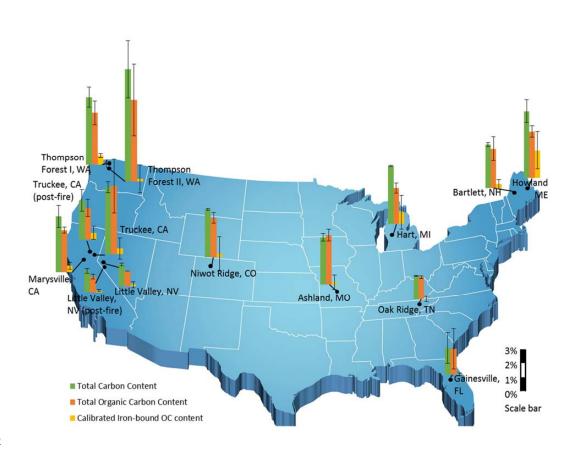
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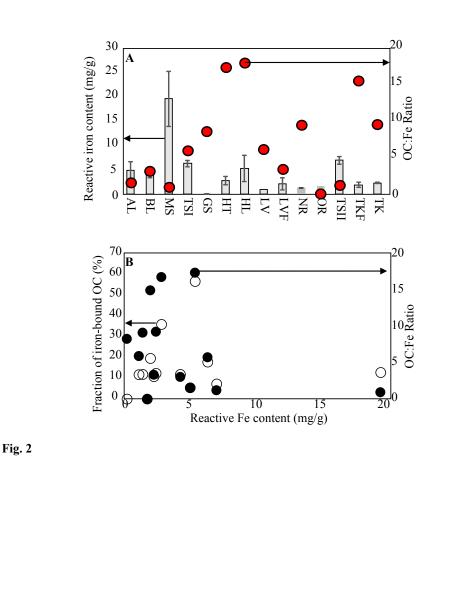




- 633 Fig. 1

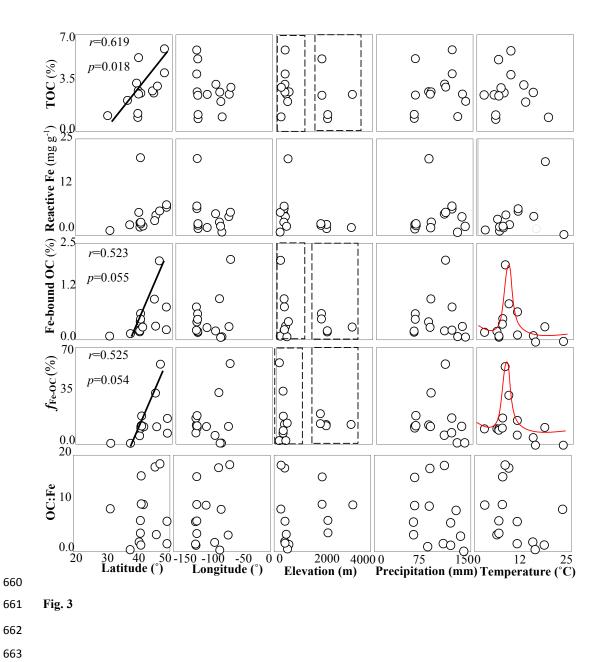








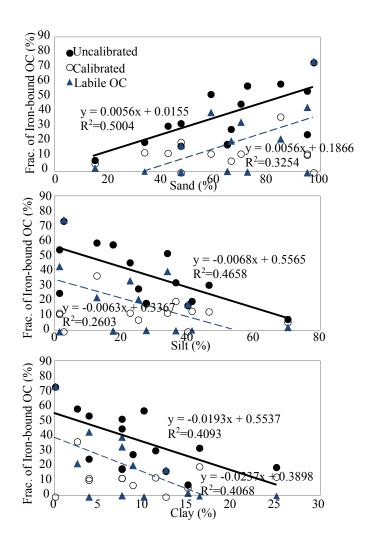




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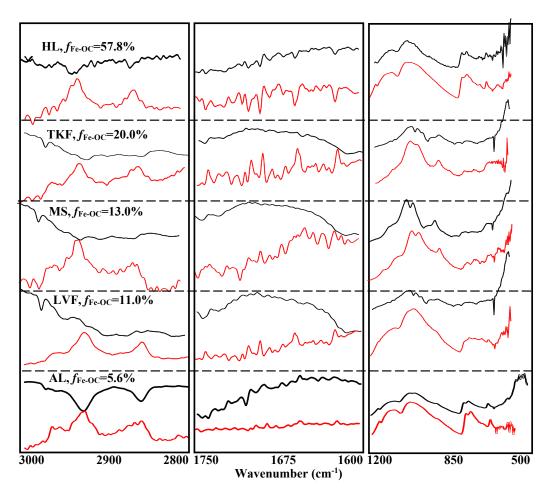




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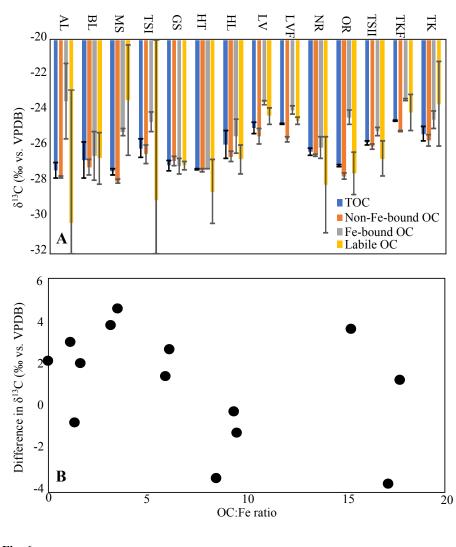




680 Fig. 5







- 692 Fig. 6