

# Underestimation of boreal soil carbon stocks by mathematical soil carbon models linked to soil nutrient status

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**Abstract.** Inaccurate estimate of the largest terrestrial carbon pool, soil organic carbon (SOC) stock, is the major source of uncertainty in simulating feedback of climate warming on ecosystem-atmosphere carbon exchange by process based ecosystem and soil carbon models. Although the models need to simplify complex environmental processes of soil carbon sequestration, in a large  
5 mosaic of environments a missing key driver could lead into a modelling bias in predictions of SOC stock change.

We aimed to evaluate SOC stock estimates of process based models (Yasso07, Q, and CENTURY) against the Swedish forest soil inventory data (3230 samples) organized by recursive partitioning method into distinct soil groups with underlying SOC stock development linked to physicochemical  
10 conditions.

The Yasso07 and Q models that used only climate and litterfall input data and ignored soil properties generally agreed with two third of measurements. However, in fertile sites with high nitrogen deposition, high cation exchange capacity, or moderately increased soil water content, Yasso07 and Q underestimated SOC stocks. Accounting for soil texture (clay, silt, and sand content) and structure  
15 (bulk density) in CENTURY model showed no improvement on carbon stock estimates, as CENTURY deviated in similar manner.

Our analysis suggested that the soils with poorly predicted SOC stocks, as characterized by the high nutrient status and well sorted parent material, indeed have had other predominant drivers of SOC stabilization lacking in the models presumably the mycorrhizal organic uptake and organo-mineral  
20 stabilization processes. Our results imply that the role of soil nutrient status as regulator of organic matter mineralization has to be re-evaluated, since correct steady state SOC stocks are decisive for predicting future SOC change [and soil](#) CO<sub>2</sub> [efflux](#).

## 1 Introduction

In spite of the historical net carbon sink of boreal soils, 500 Pg of carbon since the last ice age  
25 (Rapalee et al., 1998; DeLuca and Boisvenue 2012; Scharlemann et al., 2014), boreal soils could be-  
come a net source of carbon to the atmosphere as a result of long-term climate warming (Kirschbaum  
2000; Amundson 2001). They have the potential to release larger quantities of carbon than all an-  
thropogenic carbon emissions combined (337 Pg) (Boden et al., 2010). In order to preserve the soil  
carbon pool and to utilize the soil carbon sequestration potential to mitigate anthropogenic CO<sub>2</sub>  
30 emissions, mitigation strategies of climate forcing aim to improve soil organic matter management  
(Schlesinger 1999; Smith 2005; Wiesmeier et al., 2014).

Supporting soil management decisions requires an accurate quantification of spatially variable soil  
organic carbon (SOC) stock and SOC stock changes (Scharlemann et al., 2014). The initial level of  
SOC stock is essential in order to estimate SOC stock changes (Palosuo et al., 2012, Todd-Brown  
35 et al., 2014), especially when estimating carbon emissions due to land-use change e.g. afforestation  
of grasslands (Berthrong et al., 2009). Process-oriented soil carbon models like CENTURY, Roth-C,  
Biome-BCG, ORCHIDEE, JSBACH, ROMUL, Yasso07 and Q are important tools for predicting  
SOC stock change, but there are also risks for poor predictions (Todd-Brown et al., 2013, DeLuca  
and Boisvenue 2012). The models need further validation and improvement as they show poor spatial  
40 agreement on fine scale and moderate agreement on regional scale against SOC stock data (Todd-  
Brown et al., 2013; Ortiz et al., 2013). Despite the potentially quantitative importance of CO<sub>2</sub> emis-  
sions the expected change will be small in relation to the SOC stock. Therefore, the uncertainty  
of measurements and/or model estimates could prevent conclusions on SOC stock changes (Palosuo  
et al., 2012; Ortiz et al., 2013; Lethonen et al., 2015a) especially for the soils with largest SOC stocks  
45 which are the most sensitive to carbon loss. Beside large uncertainties, the poor agreement between  
the modelled and measured SOC stocks (Todd-Brown et al., 2013) could also indicate missing biotic  
or abiotic drivers of long-term carbon storage (Schmidt et al., 2011; Averill et al., 2014).

For example ignoring the essential role of soil nutrient availability in ecosystem carbon use ef-  
ficiency (Fernández-Martínez et al., 2014) could lead to missing important controls of plant litter  
50 production and soil organic matter stabilization mechanisms. Soil nutrient status is linked to the  
mobility of nutrients in the water solution (Husson et al., 2013), production, quality and microbial  
decomposition of plant litter (Orwin et al., 2011), and formation of the soil organic matter (SOM).  
The SOM affects soil nutrient status by recycling of macronutrients (Husson et al., 2013), and water  
retention and water availability (Rawls et al., 2003).

55 In spite of state of the art soil carbon modelling based on the amount and quality of plant litter  
“recalcitrance”, affected by climate and/or soil properties as in the Yasso07, Q and CENTURY  
models, these type of process based models do not include mechanisms for SOM stabilization by a)  
the organic nutrient uptake by mycorrhizal fungi; b) humic organic carbon interactions with silt-clay  
minerals; and c) the inaccessibility of deep soil carbon and carbon in soil aggregates to soil biota

60 (Orwin et al., 211; Sollins et al., 1996; Torn et al., 1997; Six et al., 2002; Fan et al., 2008; Dungait  
et al., 2012; Clemente et al., 2011). Although the models do not contain aforementioned mechanisms  
and controls for changes in SOM stabilization processes, they have been parameterized using a wide  
variety of datasets and can treat soil biotic, physicochemical and environmental changes implicitly.  
The Yasso07 model (Tuomi et al., 2009, 2011) is an advanced forest soil carbon model and it is used  
65 for Kyoto protocol reporting of changes in soil carbon amounts for the United Nations Framework  
Convention on Climate Change (UNFCCC) by European countries e.g. Austria, Finland, Norway,  
and Switzerland. The Q model (Ågren et al., 2007) is a mechanistic litter decomposition model  
developed in Sweden and used e.g. to compare results produced with Swedish national inventory data  
(Stendahl et al., 2010, Ortiz et al., 2011) and also with other models at national or global scales (Ortiz  
70 et al., 2013; Yurova et al., 2010). The CENTURY model (Parton et al., 1987, 1994, Adair et al., 2008)  
is one of the most widely applied models and it is used for soil carbon reporting to UNFCCC by  
Canada, Japan, and USA. Although individual parameters and functions vary, mathematical models  
such as Yasso07, Q and CENTURY have similar structures. For example, these models are driven  
by the decomposition rates of litter input and soil organic matter (SOM). Decomposing litter and  
75 SOM is divided into pools based on litter quality, and its transfer from one pool to another is apart  
from model functions and parameters affected by temperature (Q) and/or water (Yasso07), and/or  
soil texture and structure (CENTURY). The Q model does not include explicit moisture function,  
whereas for the Yasso07 and CENTURY models precipitation ~~effects~~ affects decomposition (Tuomi  
et al., 2009; Adair et al., 2008). On the other hand, the models do not explicitly or by default include  
80 mechanisms that reduce decomposition by excessive precipitation/moisture (Falloon et al., 2011).

We hypothesized that (1) soil carbon estimates of the Yasso07, Q, and CENTURY models would  
deviate for soils where SOC stabilization processes not implicitly accounted by the models are pre-  
dominant, (2) the Yasso07 and Q models ignoring soil properties would fail on the nutrient rich sites  
of South-West coast of Sweden and on occasionally paludified clay and silt soils, and (3) the CEN-  
85 TURY model outperforms the Yasso07 and Q models due to fact that it includes soil properties as  
input variables.

We grouped Swedish forest soil inventory data into homogenous groups with specific soil physic-  
ochemical conditions using regression tree and recursive partitioning modelling methods. After that  
we ran the models into a steady state with a litter input which was derived from the Swedish forest  
90 inventory. Thereafter we compared the model estimates against data by groups that were obtained  
from the regression tree model. In discussion we address the reasons why the models deviate and  
indicate directions of further improvements.

## 2 Material and methods

### 2.1 Measurements

95 We analysed data from the Swedish forest soil inventory (SFSI) which is a stratified national grid survey of vegetation and physicochemical properties of soils (SLU, 2011, Olsson et al., 2009). All analysis was done using R software for statistical computing and graphics (R core team 2014). The soil data were identical to dataset used in Stendahl et al. (2010). We restricted our sample plots to minerogenic soils since the Q, Yasso07, and CENTURY models were not developed for use on peat  
100 soils, and only to plots for forest land use with Swedish forest inventory data (SFI). We also excluded samples with total ~~soil organic carbon (SOC)~~ SOC stock below 2.8 and above 470.5 ( $\text{tC ha}^{-1}$ ), i.e. samples with SOC stock below 0.01 and above 99.9 percentile. Measurement data originated from the 1993 to 2002 which constitute a full inventory, and from 2020 sample plots located around Sweden, and in total it including 3230 samples. For each sample plot the weather (years 1961-  
105 2011) and N deposition (years 1999-2001) data was retrieved from the nearest stations of Swedish Meteorological and Hydrological Institute (SMHI) network (Fig. 1). The plots which were linked by the closest distance to the given weather station had the same weather and N deposition data, and the number of soil samples per station ranged between 10 and 70. The mean total SOC stock of samples corresponding to weather stations ranged between 40 and 200 ( $\text{tC ha}^{-1}$ ), and the SOC stock level  
110 increased from the South to North of Sweden (Fig. 1).

Each sample plot contained categorical data from the field survey on the sorting of soil parent material, humus type, soil texture, and soil moisture. In our analysis we reduced categorical classes by basing them on the sorting of soil parent material and humus type (Table 1). We determined numeric values for silt, clay, and sand content from soil texture categories by Albert Atterberg's distribution  
115 of the different grain size fractions in tills and by Lindén's (2002) distributions for sediments (Table 1). We also determined numeric values of volumetric soil water content (SWC) from categorical field data classified according to the depth of the ground water level (WL) and the observations of Tupek et al. (2015) (Table 1).

As typical for soil carbon inventories, the variation of data was large (Table 2). For example, the  
120 mean total SOC stock of all samples was 93 ( $\text{tC ha}^{-1}$ ) while 1st and 99th percentiles were 17 and 309 (Table 2). The mean SOC stock was 33.3 and 66.8 ( $\text{tC ha}^{-1}$ ) for the humus horizon and the mineral soil. The mean values of cation exchange capacity (CEC) 23.9 ( $\text{mmol}_c \text{kg}^{-1}$ ), the base saturation 36.4%, and the C/N ratio 16.5 indicated conditions of medium fertility, although the soils were mostly acidic (mean pH was 5.2). The mean prevailing soil water content (22.3) was typical  
125 for the well-drained forest soils. The mean annual temperatures ranged from below 0 to above 8 °C, and annual precipitation varied between 392 and 1154 mm (Table 2). Total SOC stock for all the samples generally increased for peat and peat like humus forms, for well sorted sediments, for soils with high fraction of silt and clay and with increasing soil moisture (Fig. S1).

### 2.1.1 Biomass and litterfall estimates

130 Forest stand biomass was estimated by allometric biomass functions for stem with bark, branch, foliage, stump, coarse-roots and fine-roots applied to basic tree dimensions (breast height diameter, total height of tree, number of trees) of SFI stands (Marklund 1988; Pettersson and Ståhl 2006; Repola 2008; Lehtonen et al., 2015b). In order to simulate “steady state” soil carbon stock we estimated long term mean forest biomass, referred to as “steady state forest” below.

135 We adopted an actual fraction of photosynthetically active absorbed radiation ( $f_{APAR}$ , Fig. A1) as a relative indicator of a site’s capacity to produce biomass (minimum = 0, maximum = 1). The  $f_{APAR}$  was calculated based on basic tree measurements as in Härkönen et al. (2010) and for the main tree species (pine, spruce, deciduous) it was well correlated with the stand basal area (Appendix A). The steady state forest  $f_{APAR}$  values were assumed to be in a range  
140 between the median and the maximum fraction of the actual state forest  $f_{APAR}$  for a given species, latitudinal degree, and site productivity class (Appendix A).

~~We modelled the steady state biomass by applying the fitted exponential functions between the measured forest biomass components (stem, branch, foliage, stump, coarse-roots, fine-roots) “actual state” and the actual fraction of absorbed radiation ( $f_{APAR}$ ) (Appendix B) to the “steady state” forest  $f_{APAR70}$ . The  $f_{APAR70}$  We selected steady state  $f_{APAR}$  as the 70th percentile ( $f_{APAR70}$ ) out of a range from the 50th to 95th, because the modelled soil carbon distributions with a litter input from the  $f_{APAR70}$  biomass best agreed with the measured soil carbon distributions (Fig. S2). The  $f_{APAR70}$  was the estimated 70th percentile of the actual fraction of absorbed radiation specific for a given species, latitudinal degree, and site productivity class (Fig. B1). We selected the 70th percentile out of a range from the 50th to 95th, because the modelled soil carbon distributions with a litter input from the  $f_{APAR70}$  biomass best agreed with the measured soil carbon distributions (Fig. S2). The ground-~~

We modelled the steady state biomass by applying the fitted exponential functions between the actual state forest biomass components (stem, branch, foliage, stump, coarse-roots, fine-roots, estimated by tree stand measurements and the allometric biomass functions) and the actual fraction of absorbed radiation ( $f_{APAR}$ ) (Appendix B) to the estimated  $f_{APAR70}$  of the steady state forest. The understory vegetation of the steady state forest was estimated by applying our ground vegetation models (Appendix C) to the modelled steady state forest characteristics, and plot specific environmental conditions.

160 In order to derive the litter inputs, annual turnover rate (TR, the fraction of living biomass that is shed onto the ground per year, unitless) of biomass components were applied to the modelled biomass components of the steady state forest. The needle litter TR was a linear function of latitude for pine and spruce and a constant for deciduous species (Ågren et al., 2007). The TR of branches and roots were from Mukkonen and Lehtonen (2004), Lehtonen et al. (2004) and the TR of stump  
165 and stem were from Viro (1955), Mälkönen (1974, 1977) as cited in Liski et al. (2006). For tree fine

170 roots we assumed there was a difference between tree species and between southern and northern Sweden. For pine, spruce, and birch the fine roots TR were 0.811, 0.868, and 1.0 respectively as reported by Maldi (2001) and Kurz et al. (1996), and cited in Liski et al. (2006). Kleja et al. (2008) and Leppälampi-Kujansuu et al. (2014) reported different fine root TR for Southern (1 and 0.83) and Northern Finland (0.5). We interpolated TR according to the mean annual temperature gradient between TR of fine roots in the South and the North. The fine roots TR of 0.811, 0.868, and 1.0 in the warmest southernmost soil plots were thus reduced down to 0.5 in the coldest northernmost soil plots. The understory TR were applied as in Lehtonen et al. (manuscript).

175 The major part of the litter input originated from the tree stand biomass components which were modeled by the non-linear functions with  $R^2$  values close to 0.9 (Fig. B1, Tables A1 and B1). The linear understory vegetation models had low  $R^2$  values (Table C1). However, when the understory models (Appendix C) were applied only to plots close to steady state forest, as in our application, the  $R^2$  values of predicted and observed understory components were larger (Fig. S9). In comparison to major understory litterfall originating from reasonably well predicted dwarf-shrubs and mosses (Fig. S9 and S10), the influence of poorer understory models (for herbs, grass, and lichens) was small on predictions of the understory litter and marginal on predictions of the total forest litterfall (Fig. S10). The main improvement on the accuracy of total litter input was achieved by avoiding the confounding effect of actual forest state by modelling the biomass/litterfall estimates representing the mean long-term conditions (defined by estimated steady state  $f_{APAR70}$ ) for small regions (defined by degree of latitude and productivity class for dominant species, Fig. A1). Thus the estimates accurately reflected the long-term spatial variability in dominant species, nutrient status and climate (Fig. S11) and lacked higher spatial and temporal precision; as attempts for high precision of the estimates applied for the period of the last few thousands of years would be uncertain due to high variation of factors affecting plot history.

### 190 **2.1.2 Correlation analysis**

Overall our data consists of 3230 soil samples and their carbon stocks linked to soil physicochemical variables, stand and ground vegetation biomass and litterfall components, and nearest weather station environmental variables. We performed the Spearman's rank correlation analysis between the total soil carbon stock and the other soil variables, site, climate and vegetation characteristics. As expected the total soil carbon stock most strongly correlated with the measured variables used for its calculation e.g. bulk density, depth of humus and mineral soil, carbon content, and stoniness. These variables were excluded from further regression tree analysis which aimed to group data according to the processes of soil carbon stock development.

### 2.1.3 Regression trees

200 In order to organize SOC data into groups according to the physicochemical soil variables and to better understand the nature of measured data, we generated regression trees of SOC stocks by using recursive partitioning (RPART) (Therneau and Atkinson 1997). RPART is based on developing decision rules for predicting and cross validation of continuous output of soil carbon stocks (regression tree). The classification tree was built by finding a single variable which best splits the data into two  
205 groups. Each sub-group was recursively separated until no improvement could be made to the soil carbon stock estimated by using the split based regression model. The complex resultant regression tree model was cross validated for a nested set of sub trees by computing the estimate of soil carbon stock to trim back the full tree.

When building the regression tree models we excluded variables such as bulk density, carbon  
210 contents of soil layers, soil depth, and stoniness, since these measured variables were used for determining the total soil carbon stock. The selected variables for the RPART data mining were based on the correlations analysis (see 2.1.2.), the processes of soil organic matter formation (e.g. Husson et al., 2013) and decomposition, and represented the soil categorical variables (sorting of parent material, soil texture, long-term soil moisture and humus form), soil physicochemical variables (sand,  
215 clay, and silt content, long-term soil moisture, highly bound water, C/N ratio, pH, CEC of organic, B, BC, and C horizons), climatic variables (annual mean air temperature, annual precipitation sum), and stand and site characteristics (tree species coverage of pine, spruce and deciduous, total foliar litter input, productivity class and N deposition). Alternatively we also ran regression and classification analysis by excluding all measured soil variables because soil variables are often unavailable  
220 for landscape level modelling.

The regression tree model separated the measured total SOC stocks ( $\text{tC ha}^{-1}$ ) into 10 groups. The cation exchange capacity of the BC horizon (CEC,  $\text{mmol}_c \text{kg}^{-1}$ ) divided all the samples into 2/3 of lower SOC stock groups (means between 65 and 130  $\text{tC ha}^{-1}$ ) and 1/3 of larger groups (means between 86 and 269  $\text{tC ha}^{-1}$ ) (Fig. 2a). The group of the smallest SOC stock consisted of 959  
225 samples compared to 8 samples of the group with the largest SOC stocks.

Two-thirds of the smaller SOC stocks were subdivided by CEC and the type sorting of soil parent material (sorted or unsorted). One-third of the larger SOC stocks was subdivided by the C/N ratio, CEC, N deposition among others. Roughly generalized, groups from left to right or from 1 to 10 formed a gradient in levels of SOC stock, moisture, nutrient status, and production (Fig. 2, Table S1).

230 The alternative regression tree model was built with variables other than soil properties. The regression tree with the annual mean air temperature, the annual precipitation sum and the percentage of pine trees in the stand, and the nitrogen deposition separated measured SOC stocks ( $\text{tC ha}^{-1}$ ) into five groups (Fig. S3). Colder groups with smaller SOC stocks (means 67 and 85) also had less litter input (below 3  $\text{tC ha}^{-1}$ ) and low productivity class (height of trees at 100 years of age, H100

235 < 20 m) (Table S2). Nitrogen deposition only slightly impacted the higher productivity class of soils and litter input (Table S2).

## 2.2 Soil carbon stock modelling

The Q model (Rolff and Ågren, 1999) is a continuous mechanistic litter decomposition model describing change of soil organic matter over time. The decomposition rate for the branch, stem, needle, 240 fine root, and woody litter fractions is controlled by the temperature, litter quality, microbial growth and litter invasion rate. The model has been calibrated for seven climatic regions of Sweden in order to account for Swedish temperature and precipitation gradients (Ortiz et al., 2011) (Table 3). The Q model was applied in several studies of SOC stock and change estimation in Sweden (e.g. Stendahl et al., 2010; Ortiz et al., 2013; Ågren et al., 2007). The Q model was run for seven Swedish climatic 245 regions (Ortiz et al., 2011). The mean regional parameterization from the calibration of the ~~2012~~ 2011 Q model was used for the plot simulations. Thus, the simulations in each region represent variations in climate and litter input and not parameter variations. The steady state soil carbon stocks are estimated in the model using the equation for steady state soil carbon stock which is derived from the decomposition functions with constant amounts and quality of litter input.

250 The Yasso07 model (Tuomi et al., 2009; 2011) is ~~an advanced forest soil carbon model~~ one of the most widely applied SOC models. The model was calibrated based on almost 10 000 measurements of litter decomposition from Europe, North and South America (Table 3). The required annual inputs of litterfall, its size and chemical composition, temperature and precipitation determine the decomposition and sequestration rates of soil organic matter. Yasso07 estimates SOC stock to a 255 depth of 1 m (organic and mineral layers), change of SOC stock, and heterotrophic soil respiration. ~~The Yasso07 model, which is used for soil carbon Kyoto protocol reporting by several European countries, i.e. Austria, Finland, Norway, and Switzerland, is one of the most widely applied SOC model.~~ Species specific chemical composition of different litter compartments of Yasso07 were used according to Liski et al. (2009). The initial soil organic matter of Yasso07 was zero. The simulated 260 soil carbon stock corresponding to a steady-state between the litter input and decomposition was achieved by a Yasso07 spin-up run of 10 000 years. Yasso07 runs used litter inputs of the steady state forest biomasses (see 2.1.1.) and climate variables (annual air temperature, monthly temperature amplitude, and annual precipitation). The global parameter values of decomposition rates, flow rates, and other dependencies of Yasso07 soil carbon model were adopted from Tuomi et al. (2011) 265 and the estimates of Yasso07 SOC stocks were used in comparison with measurements and other models. We did not use the SOC stocks simulated with the more recent Yasso07 parameters based on the litter decomposition data from the Nordic countries (Rantakari et al., 2012), because the SOC stocks simulated with the global parameter values produced better fit with SFSI measurements.

The CENTURY mathematical model originally developed for grassland systems (Parton et al., 270 1987) has been since modified for various ecosystems including boreal forests (Nalder and Wein



2006). The CENTURY is also one of the most widely applied models ~~and it is used for soil carbon reporting to UNFCCC by Canada, Japan, and USA~~. The soil organic matter in the model consists of active, slow, and passive pools which have different TR (Table 3). The decomposition rates are modified by temperature and moisture, and in addition the decomposition rates of the slow and passive pools rely on lignin to N and C to N ratios, while the active pool decomposition rate relies on soil texture. The model simulates soil organic matter to a depth of 20 cm. The model simulates plant production and pools of living biomass, while TR for biomass pools determine the litterfall inputs to soil. To compare the performance of the soil sub-model with other soil carbon dynamics models, Q and Yasso07, we only used the CENTURY soil sub-model. We used the same litterfall inputs as used by the Q and Yasso07 simulations, which were estimated by our litterfall modelling (see 2.1.1.). For CENTURY we adopted general parameters from the parameter file “tree.100”, parameters of site “AND H\_J\_ANDREWS” for conifers, and site “CWT Coweeta” for deciduous trees. The nitrogen dynamics in our CENTURY model application were held constant. The CENTURY SOC stocks simulation were run with steady state forest litter inputs, site specific soil parameters (specific bulk density, sand, silt, and clay content) and climate variables (monthly air temperature, monthly precipitation). The simulated steady state SOC stocks were estimated by a spin-up run of 5 000 years. The number of years to reach steady state was sought empirically on 100 random sites, and differs from Yasso07 because running CENTURY was computationally more demanding.

### 3 Results

290 The distributions of Yasso07, Q, and CENTURY model estimates of total SOC stocks ( $\text{tC ha}^{-1}$ ) were in agreement for 2/3 of the measured data with lower SOC stock (Fig. 3, distributions of groups 1, 2, and 4). The remaining 1/3 of data was underestimated by models. This 1/3 of data was separated into 7 physicochemical soil groups (means of groups in range from 104 to exceptionally large 269  $\text{tC ha}^{-1}$ , see Fig. 3, distributions of groups 3, and 5-10). The linear regression of mean levels of all 10 physicochemical soil groups (weighted by the number of samples in each group) between the modelled and measured SOC stocks showed smaller underestimation of Yasso07 compared to the CENTURY and Q models (Fig. 4). The weighted root mean square error (RMSE) was 31.6 ( $\text{tC ha}^{-1}$ ) for Yasso07 and 41.7 and 38.8 for CENTURY and Q respectively. The proportion of explained variance was larger for Q ( $r^2 = 0.58$ ) than for Yasso07 and CENTURY ( $r^2 = 0.42$  and 0.39) (Fig. 4). The deviation of the distributions of CENTURY SOC stocks, simulated using soil bulk density, sand, silt, and clay content, were similar as for Yasso07 and Q estimates for 10 physicochemical soil groups (Fig. 3).

As expected, the models clearly showed less variation than the measurements. The shift of the mean values from the center of distribution, the width of confidence intervals of means, and the width of the tails of distributions were clearly larger for the measurements than for the modelled es-

timates (Fig. 3). The modelled distributions agreed for the poor-medium fertility soils with low and medium measured SOC stocks, low and medium ~~cation-exchange-capacity (CEC)~~CEC, unsorted parent material, low temperatures and low production (groups 1, 2, and 4) (Fig. 2, Table S1, Fig. 3). Disagreement between modelled and measured SOC stock distributions were formed on fertile soils  
310 with sorted parent material (groups 3 and 5), soils with higher water content (groups 3, 5, and 10), where nitrogen deposition was large (groups 7 and 8), and where ~~cation-exchange-capacity (CEC)~~CEC was median or large (Fig. 2, Fig. 3). The largest deviation between the measured and modelled distributions was found for the relatively small physicochemical groups of soils (3%) typical for highly bound water and peat humus types (groups 8 and 10) (Fig. 2, Fig. 3). The distributions  
315 of measured total SOC stocks ( $\text{tC ha}^{-1}$ ) generally increased for the groups with higher nutrient status (Fig. 3, Fig. S4). The distributions of SOC stocks in mineral soil were larger than those in humus horizon, and distributions of mineral SOC stocks increased with fertility slightly more than distributions of SOC stocks in humus horizon (Fig. S4).

After excluding all the soil physicochemical characteristics from the recursive partitioning, the  
320 SOC stock distributions of 5 groups regression tree model (Fig. S3, Table S2) were in agreement between the measurements and model estimates for 3 groups (77% of samples) and deviated for 2 groups (23%) (Fig. S5).

The models underestimated distributions on sites with high ( $> 10 \text{ kgN ha}^{-1} \text{ y}^{-1}$ ) nitrogen deposition (21% of samples) and on sites with warm and dry climate (2% of samples) (Fig. S5). The  
325 modelled SOC stock distributions agreed with measurements for all models on sites with cold annual temperatures  $< 3 \text{ }^\circ\text{C}$  in northern sites (low-C.cold.pine, low-C.cold.other) (Fig. S5). However, for warmer conditions in middle Sweden on sites with low nitrogen deposition SOC stock distributions only Yasso07 predictions agreed with the measurements but were underestimated for CENTURY and Q estimates.

330 The variation of density functions of modeled SOC stocks for 10 physicochemical groups (Fig. 3) was similar to the variation of the total annual plant litter input ( $\text{tC ha}^{-1}$ ) (Fig. S6). The mean levels of annual plant litter input and mean SOC stocks for 10 groups were strongly correlated (the proportion of explained variance of weighted linear regressions ranged between 0.85 for Yasso07 and 0.96 for the CENTURY and Q models). None of the models was able to explain the spatial  
335 variations for any of the physicochemical groups well (Fig. S7). Model estimates were correlated better between Yasso07 and CENTURY with an  $r^2$  range from 45 to 66%, whereas  $r^2$  values with Q estimates and the other two models ranged from 12 to 36% (Fig. S8).

## 4 Discussion

### 4.1 SOC stock distributions linked to mechanisms of SOM stabilization

340 It has been suggested that process based soil carbon models with the current formulation lacking major soil environmental and biological controls of decomposition would fail for conditions where these controls predominate (Schmidt et al., 2011; Averill et al., 2014). Although, the effect of the soil properties on SOC stocks e.g. soil nutrient status in the widely used models such as Yasso07, Q, and CENTURY have not previously been quantitatively evaluated. We found that in comparison  
345 with Swedish forest soil inventory (~~SFSI~~) data, the models based on the amount and quality of inherent structural properties of plant litter (Q, Yasso07, and CENTURY) produced accurate SOC stock estimates for 2/3 of northern boreal forest soils in Sweden. Two-thirds of the distributions of SOC stocks measurements of SFSI agreed with distributions of SOC stock estimates of the Q, Yasso07, and CENTURY soil carbon models (Fig. 3, distributions of groups 1, 2, and 4). However,  
350 the SOC stocks underestimation by these models for one third of the data (Fig. 3, distributions of groups 3, and 5-10) indicated that some drivers other than molecular structure, especially site nutrient status, play an important role in higher SOC stocks sequestration.

Some level of deviation from measurements and poorly explained spatial variation (Fig. S7) was expected from the uncertainties of the SOC measurements, annual plant litter inputs and climate  
355 variability for the model SOC stock change estimates (Ortiz et al., 2013; Lehtonen et al., 2015a). For the long-term SOC stock development the model uncertainties are less known than for the short-term litter decomposition. Previously reported fine scale comparison also showed poor agreement between Earth system models and the Northern Circumpolar Soil Carbon Database (Todd-Brown et al., 2013), although drivers of the deviation still remained open. Our results showed that if mod-  
360 els strongly depend on the litter inputs (Fig. S6) then the spatial differences between measured and modeled SOC stock distributions could be linked to sites with rich nutrient status through cation exchange capacity, C/N ratio, N deposition, drainage (sorting of parent material) among other factors (Fig. 2 and 3). Additionally, when the soil properties were excluded from the regression, the estimates of SOC stocks also deviated for the fertile groups (Fig. S5). However, the rich nutrient  
365 status for these groups was linked to differences in species composition, N deposition, and climate (temperature, precipitation) instead of soil properties (Fig. S3).

Larger net soil carbon accumulation in nutrient rich sites could be attributed to the relative differences in litterfall components (relatively more leaves and branches with higher N content than fine roots) ~~and to the reduced microbial demand for N from fine roots and SOM~~, and to higher N  
370 availability and carbon use efficiency of decomposers, reduction of respiration per unit of C uptake (Ågren et al. 2001, Manzoni et al. 2012, Fernández-Martínez et al., 2014). Largest deviation between measured and modeled data in our study was found for fertile presumably N rich and fresh to fresh-moist sites. The soils with large N deposition were also highly productive and showed high

to exceptionally high SOC stocks (Fig. 2, Fig. 3, soil groups 7 and 8). This was in agreement with fertilization and modelling study of Franklin et al. (2003) showing an increase in soil C accumulation with N addition. Our forest biomass and litterfall estimates were based on forest inventory and modeling, but the site nutrient status was only partially reflected in the amount of biomass/litterfall and its quality. The quality was only reflected through the biochemical differences between species and plant litter components. The relative differences between the biomass/litterfall components or between C/N ratios of litterfall in relation to site fertility are not accounted by the current biomass models, but soil fertility could be considered in an attempt of SOC stock modelling. For example the proportion of acid -, water -, and ethanol-soluble and non-soluble litter inputs for Yasso07 could be re-evaluated by allowing it to vary depending on site fertility, in addition to currently used variation specific for species and the litter components.

The litter decomposition and SOC stabilization rates in Yasso07, Q, and CENTURY based on the litter quality “recalcitrance” originating from the litter bag mass loss measurements have major drawbacks. The mass loss from the litter bags is assumed to be fully mineralized, although the litterbags are subjected to non-negligible leaching (Rantakari et al., 2012; Kammer and Hagedorn, 2011). The SOC stabilization represented in models by the remaining litter mass is thus underestimated due to the fraction of particulate organic matter and dissolved organic carbon that is lost from the litterbags but later immobilized e.g. through organo-mineral stabilization. The use of stable isotopes seems to determine the field carbon mineralization and accumulation rates from the labile (high C quality and N concentration) or recalcitrant (low C quality and N concentration) litter more accurately than litter bags (Kammer and Hagedorn, 2011).

Higher amount of more recalcitrant fine roots compared to more labile leaves (Xia et al., 2015) heavily increased the soil carbon sequestration in CENTURY model simulations which was in line with McCormack et al. (2015). Though, the contribution of fine roots to SOC stabilization is still not settled due to the significant role of mycorrhizal fungi in SOC accumulation (Averill et al., 2014; Orwin et al., 2011). Xia et al. (2015) claimed that more recalcitrant fine roots contribute to stable SOC more than leaf litter, because fine roots degrade slower. This would be supported by the fact if the precursors of fine roots that are degraded by fungi are more stable than the precursors of leaves degraded by microbes. However, more recalcitrant plant litter has been also suggested to stabilize less SOC stocks (Kammer and Hagedorn, 2011). This is a result of recalcitrant litter satisfying less of the microbial N demands promoting respiration and reducing the long-term production of microbial products, precursors for the organo-mineral stabilization (Cotrufo et al., 2013, Castellano et al., 2015). According to the microbial efficiency-matrix (MEM) stabilization mechanism (Cotrufo et al., 2013) fertile sites with relatively more labile plant litter, but with larger absolute production and larger microbial activity than poor sites, would in long-term stabilize more carbon through organo-mineral stabilization. Our results supported MEM stabilization theory by showing larger car-

410 bon stocks in mineral soil than in humus horizon, and by relatively more SOC stocks in mineral soil in fertile groups than in poor conditions (Fig. S4).

Expanding on the CENTURY model structure, the MySCaN model incorporating the organic nutrient uptake by mycorrhizal fungi estimated positive effect on SOC accumulation, relatively larger in poor than in fertile sites (Orwin et al., 2011). ~~Ignoring Therefore, not accounting for~~ the organic  
415 nutrient uptake by mycorrhizal fungi by the Yasso07, Q, and CENTURY models probably led to the underestimation of SOC stocks in ~~medium-highly productive soils~~sites with higher nutrient status. This hypothesis needs to be tested in further studies. We did not have all input data and the source code to include MySCaN into our model intercomparison. The spatial trends of N and P data of litter in Sweden that would be needed to make such study were not available. However, adjusting biomass  
420 turnover rates, used for the litter input estimation, in dependence to site fertility would lead into larger inputs for fertile sites and increased SOC stock accumulation as a result of increasing plant productivity and inputs. It is well established that SOM increases soil fertility by improving the soil water and nutrient holding capacity; recycling of SOM increases CEC, humic substances and nutrient availability for plant resulting in larger biomass/litter production (Zandonadi et al., 2013). As an  
425 alternative to adjusting turnover rates with site fertility, we suggest that a feedback link in models between increasing fertility due to SOC stock accumulation (e.g. due to increased CEC relative to humus, increased nitrogen availability), increasing litter inputs, and reduced rates of SOC decomposition per unit of litter input (e.g. through satisfying more microbial N demand with less respiration, limited oxygen in increased moisture conditions) would also increase SOC stock accumulation.

430 Increased moisture and more frequent water saturation due to SOC accumulation limits soil oxygen availability and slows rates of microbial decomposition which increases the rate of SOC stabilization. The CENTURY model has an optional function that represents the reduction of decomposition caused by anaerobic conditions. The function becomes active when a controlling parameter, “drain”, is changed, and the value of the parameter has to be arbitrarily determined through parameter fitting against SOC data (e.g. Raich et al., 2000). The function is meant for anaerobic conditions  
435 in poorly drained soils, and therefore is not applicable to (most of) our sites. In addition, tuning a specific parameter to reproduce the SOC data was beyond the scope of this study. Our results, which were derived from mostly well drained soils, suggest that high SOC stocks may be partly caused by reduction of decomposition at increased water content. Detailed modelling of soil water conditions  
440 requires specific functions and many parameters, which are not included in simpler SOC models like Q and Yasso07. However, appropriate modelling of soil water conditions and reduction of decomposition in wet conditions (not necessarily at saturation) would potentially improve the performance of SOC models in particular for soils with high SOC stocks.

## 4.2 Intercomparison of models

445 The similarities between the variations of modeled SOC stocks and litterfall inputs for the soil groups  
with different fertilities (Fig. 3, Fig. S6) could be expected for the Yasso07 and Q models which  
ignore the soil properties. These models run organic matter decomposition and humus stabilization  
with litterfall, temperature and/or precipitations input data. Litter quality as input in Yasso07 and Q  
implicitly includes some information on soil properties, but as we saw litter quality hardly mapped  
450 any of soil fertility. Unexpectedly the low impact of soil properties on the estimates was seen also  
in the relatively more complex model CENTURY (accounting for the plot specific bulk density,  
sand, silt, and clay content in addition to litter input, temperature and precipitation data). Contrary  
to our expectation, the CENTURY model still heavily depended on the amount of litter input, and  
its variations of the estimated SOC stocks distributions were similar to those for the Yasso07 and Q  
455 models. In testing multiple soil carbon models with same litter inputs Palosuo et al. (2012) observed  
larger variation in modeled SOC stocks at the early stage of the litter decomposition (10 years)  
but later on at 100 years the variation decreased. Although the variations were similar between the  
models, the estimated CENTURY SOC stocks distributions were slightly lower than the Yasso07  
estimates. CENTURY in its original configuration simulated SOC stock up to 20 cm soil depth  
460 (Metherell et al., 1993) whereas the Yasso07, Q, and measured SOC stocks data represented up to  
100 cm of the soil (Tuomi et al., 2009, Stendahl et al., 2010). In Yasso07 model parameters were  
calibrated based on soil age chronosequence data of SOC stocks for soil depths up to 30 cm, which  
was assumed to represent 60% of the total SOC stocks up to 100 cm soil depth (Liski et al., 1998,  
2005 as cited by Tuomi et al., 2009). Therefore, if 40-50% of the missing deep carbon were added on  
465 top of the original CENTURY estimates as is done for Yasso07, the SOC stock levels for CENTURY  
would be larger than those for the Yasso07 and Q models.

Although estimated SOC stocks of CENTURY were generally lower than those of Yasso07, the  
correlation between CENTURY and Yasso07 estimates was stronger than for Q model compared to  
two other models (Fig. S8). The reason was probably similar global parameterizations of Yasso07  
470 and CENTURY whereas Q was specifically parameterized and applied for the regions in Sweden  
(Ågren and Hyvönen 2003, Ortiz et al., 2013). Furthermore the Q model SOC stock estimates  
were more sensitive to differences in species coverage e.g. to pine and spruce (Ågren and Hyvö-  
nen 2003) and formed two distinct point cloud distributions (one for pine and broadleaves, the other  
for spruce) when compared with the CENTURY or Yasso07 estimates (Fig. S8). In spite of similar-  
475 ities in Yasso07 and CENTURY SOC stocks estimates, Yasso07 through species specific litterfall  
solubility (Liski et al., 2009) was more sensitive to species coverage than CENTURY which treated  
conifers in a single group (Metherell et al., 1993). Pine and other species (spruce) coverage was  
shown to affect measured low and median SOC stocks of colder climate if the soil properties were  
not considered (Fig. S5). Therefore the pattern of increased accumulation of SOC stock on sites  
480 with larger spruce coverage partially observed in distribution of Yass07 estimates, and missing in

the CENTURY estimates, could be related to the slightly lower solubility/decomposability of spruce compared to pine litterfall. However, the CENTURY model SOC stocks were also highly sensitive to accurate estimation of fineroots litterfall (Mc Cormack et al., 2015) typically increasing with colder climate and increasing the C/N ratio of the organic layer (Lehtonen et al., 2015b) which is driven by the dominant tree species (Cools et al., 2014).

Large SOC stocks measurements on sites with high long-term nitrogen deposition over 10 kgN ha<sup>-1</sup> y<sup>-1</sup> (Fig. 3 and Fig. S4) were underestimated by the Q, Yasso07, and CENTURY models. A positive correlation between nitrogen deposition and SOC stocks measurements in Sweden had been previously reported by Olsson et al. (2009), and the modelling study by Svensson et al. (2008) indicated that Swedish soil carbon was decreasing in the North and increasing in the South mainly as a result of different nitrogen inputs. The Q and Yasso07 models do not have nitrogen processes. As for CENTURY, it is reported that large N input could enhance plant productivity and then increase SOC (Raich et al., 2000). The purpose of the study was to evaluate the performance of soil carbon models against the same SOC data using the same litter input, and therefore only the soil carbon submodel was used and the feedback of nitrogen input to plant productivity was not included in this study. However, as in the case of drainage discussed above, the original CENTURY incorporates more detailed processes than the relatively simpler soil carbon models, Q and Yasso07, do, and hence the original CENTURY could potentially reproduce a wider range of SOC if it was parameterized in detail.

## 500 5 Conclusions

~~The~~ In this study we presented the reasons to re-evaluate the connection between the soil nutrient status and performance of widely applied soil carbon models (Yasso07, Q, and CENTURY). As previously described in detail, our simulation was based on the widely used process based SOC models, accurate driving data including litter inputs, and massive SOC data points (Swedish inventory data, N=3230). The models differed in the main controls and functions and their performance was expected to depend on model complexity (CENTURY outperforming Q and Yasso07). The intercomparison of SOC stocks between Yasso07, Q, and CENTURY models and Swedish soil carbon inventory data revealed that these process based mathematical models developed for predicting short-term SOC stock changes ~~such as Yasso07, Q, or CENTURY can all~~ in their current state ~~can~~ predict accurate long-term SOC stocks for most soils. ~~However, for the~~ The estimates of Yasso07 fitted generally better to measurements than those of CENTURY making the use of the Yasso07 model which requires fewer parameters and less input data more preferable over CENTURY. However, because the models with their current formulation lack nutrient status related controls of decomposition and soil carbon accumulation, they underestimated for conditions where the high nutrient status predominate, in our application for medium-highly ~~fertile soils the~~

~~accumulation of stable SOC by models based on extrapolation of initial plant litter decomposition into the long-term leads to underestimation. Therefore productive sites of Southern Sweden.~~

520 Through the intercomparison of three different widely-used SOC models with massive data points, we identified that re-evaluating of the impact of nutrient status would improve the model development towards their accuracy. Particularly, the relationship between the soil nutrient status and the mechanism of soil organo-mineral carbon stabilization needs to be ~~evaluated~~, re-evaluated by soil carbon models considering only the uppermost soil layers (e.g. CENTURY), because larger SOC stocks were found in the mineral than in the humus soil horizon. We suggest evaluating enhanced microbial transformation of soil organic matter and the mycorrhizal organic nutrient uptake ~~and in~~ relation to larger plant biomass/litter production in nutrient rich sites resulting to higher SOC stock accumulation ~~. For in deeper soil layers. In addition for~~ the organo-mineral carbon stabilization, we suggest further model development accounting for the soil nutrient status through evaluating the effect of topography on sorting of the parent material, and its silt and clay complexes. ~~If modelscan be further developed to-~~

530 Our study is very useful for developing accurate soil carbon and Earth system models. Furthermore, developing accurate models that would account for the ~~processes that affect the soil nutrient status as one of the key controls affecting the~~ soil organic matter production and ~~stabilization than the soil carbon stock estimates, needed when GHG inventories are used to estimate emissions and sinks due to land-use change, and soil carbon stock management would be improved.~~

535 ~~The estimates of Yasso07 fitted generally better to measurements than those of CENTURY making the use of the Yasso07 model which requires fewer parameters and less input data more preferable over CENTURY. If CENTURY estimates would be scaled from 20 cm up to 1m the underestimation with data would improve, although the deviation in fertile soils would be similar. Furthermore when running soil carbon models such as those which obtain litter inputs based on~~  
540 ~~current stand measurements, when past forest stand dynamics are unknown, we suggest using litter inputs from the steady state forest estimated as 70th percentile of the maximum current state forest biomass for a given species, latitude and productivity class. As models heavily depend on the litter input and its quality, a more accurate litter input would also improve the soil organic carbon~~ stock estimates  
545 SOC stabilization improves estimation of feedback of global warming on SOC stock temperature sensitivity and soil CO<sub>2</sub> efflux, national reporting of soil carbon stock changes for UNFCCC, and implications of decisions mitigating the climate change effects on soil carbon stocks.

## Appendix A: Models of fraction of absorbed radiation for actual and steady state forest

The fraction of photosynthetically active absorbed radiation ( $f_{APAR}$ ) for actual state forest was calculated based on basic tree measurements of Swedish forest inventory data as in Härkönen et al. (2010). For the main tree species  $f_{APAR}$  was also well correlated with the stand basal area



( $r^2$  was 0.85, 0.86, and 0.88 for pine, spruce, and deciduous stands respectively, coefficients of regressions in Table A1). The actual state forest  $f_{APAR}$  varied between 0 and maximum close to 1 (Fig. A1).

The steady state forest  $f_{APAR}$  values were assumed to be in range between the median and the maximum fraction of actual state forest  $f_{APAR}$  for given species, latitudinal degree, and site productivity class (indicated by the height of largest tress at 100 years of stands age). The steady state forest  $f_{APAR}$  values were set to 70th percentile of maximum  $f_{APAR}$  ( $f_{APAR70}$ ) for given species, latitudinal degree, and site productivity class. We selected 70th percentile out of range from 50th to 95th, because the modelled soil carbon distributions with the litter input from biomass of  $f_{APAR70}$  best agreed with measured soil carbon distributions (Fig. S2). The  $f_{APAR70}$  values specific for pine, spruce, and deciduous stands were first modelled by regression models with latitude ( $f_{APAR70LAT}$ ) (Table A2) and then reduced by the difference between the modelled  $f_{APAR70}$  by regression models with productivity class (H100) ( $f_{APAR70H100}$ ) (Table A1) and maximum  $f_{APAR70H100}$  ( $f_{APAR70} = f_{APAR70LAT} + f_{APAR70H100} - \text{maximum } f_{APAR70H100}$ ). The  $f_{APAR70}$  values equaled the  $f_{APAR70LAT}$  values only for the maximum productivity class, otherwise it was reduced.

### Appendix B: Models of forest dry weight biomass ( $\text{kg ha}^{-1}$ ) with $f_{APAR}$ .

We fitted species specific exponential regression models between the biomass components (stem, branch, foliage, stump, coarse-roots, fine-roots) of actual state forest and the actual fraction of absorbed radiation ( $f_{APAR}$ ) (statistics of the regression models in Table B1). The biomass components derived with allometric models (measured) and those derived with  $f_{APAR}$  models (modeled) showed strong correlations (Fig. B1). In order to model the longterm mean forest biomass “steady state forest biomass” we applied the  $f_{APAR}$  biomass models to the modeled  $f_{APAR70}$  values.

### Appendix C: Models of understory vegetation.

We used Swedish forest inventory –ground vegetation coverage (%) data visually monitored between 1993 and 2002 on 2440 plots around Sweden with altogether 4472 observations separately for species of –forest floor vegetation /or their classes (Table S3). –In order to derive the ground vegetation biomass and to apply the coverage/biomass conversion functions (Aleksi-Lehtonen, unpublished results Lehtonen et al., manuscript), we grouped the species coverage observations into five functional types (dwarf-shrubs, herbs, grasses, moss, and lichen) (Table S3). The applied coverage/biomass conversion functions estimated separately the above- and below-ground biomass components for dwarf-shrubs, herbs, and grasses, and total biomass for moss, and lichen.

Except the understory coverage, the forest inventory data also contained basic tree dimensions (diameter and height of trees) and stand variables (species dominance, age, basal area, site productivity class indicated by the height of largest tress at 100 years of stands age), and also we linked

585 the plots by their closest proximity to ~~Swedish Meteorological and Hydrological Institute (SMHI)~~  
SMHI weather stations with weather data (air temperature, precipitation) and location attributes of  
the weather stations (latitude, longitude, altitude).

We built linear ~~ground-vegetation-models for~~ dry weight biomass of understory vegetation  
(kg ha<sup>-1</sup>) ~~models~~ in a two level selection of the predictors from stand, weather and location vari-  
590 ables. First, we selected the predictors into linear models by using R package “Mass” and its stepwise  
model selection by exact AIC (Venables and Ripley, 2002). Second, we refined the model by using  
“relaimpo” R package estimating usefulness (Grömping, 2006), or relative importance for each of  
the predictors in the model, and by selecting only predictors with relative importance  $\geq 0.1$ . The  
general form of the models was:

$$595 \quad y_i = a + b_1x_1 + \dots + b_nx_n + \varepsilon, \quad (C1)$$

Where  $y_i$  is the understory dry weight biomass (kg ha<sup>-1</sup>),  $x_1 \dots x_n$  are the predictors,  $a, b_1 \dots b_n$   
are parameters of the  $i^{th}$  understory functional type (Table C1), and  $\varepsilon$  is the residual error. Statistics  
of the models are shown in Table C1. Scatter plots between the measured coverage derived biomass  
and modelled dry weight biomass (kg ha<sup>-1</sup>) of the functional types of ground vegetation for the  
600 forests in their actual state close to the estimated steady state are shown on Fig. S9. ~~Statistics of the~~  
~~models are shown in Table C1.~~

### Code and data availability

The source codes of the Yasso07, Q and CENTURY models used in this paper are available  
through the supplementary material. Data used in this study can be available directly by contacting  
605 the authors.

*Acknowledgements.* This work was part of the ~~project~~ Metla project 7509 ‘Improving soil carbon estimation  
of GHG inventory’ funded by the Finnish Ministry of the Environment and the Ministry of Agriculture and  
Forestry, ~~and Academy of Finland mobility projects 276300 and 276602.~~

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**Table 1.** Description of the Swedish Forest Soil Inventory (SFSI) data reduction of soil sorting of parent material and humus types; SFSI conversion estimate of soil classes of soil moisture to numerical representation of soil water content according to observations from Tupek et al. (2015); and SFSI conversion estimate of classes to numerical representation of soil texture (sand, silt, and clay content for sediments by Lindén (2002) and for tills by Albert Atterberg’s distribution of the different grain size fractions).

<b>SORTING PARENT MATERIAL</b>		<b>HUMUS TYPE</b>		<b>MOISTURE</b>		
SFSI	REDUCED	SFSI	REDUCED	SFSI	SFSI	NUMERIC
Bedrock	Bedrock	Moder	No-peat		Water	Long-term
Poorly sorted sediments	Unsorted	Mor 1	No-peat		level (m)	moisture %
Tills	Unsorted	Mor 2	No-peat	Dry	<2	10
Well sorted sediments	Sorted	Mull	No-peat	Fresh	1-2	20
		Mull-Moder	Peat	Fresh-moist	<1	30
		Peat	Peat	Moist	<0.5	50
		Peat-Mor	Peat			

<b>TEXTURE</b>						
SFSI	NUMERIC SEDIMENTS			TILLS		
	Sand %	Silt %	Clay %	Sand %	Silt %	Clay %
Bedrock	0	0	0	0	0	0
Boulder	0	0	0	0	0	0
Gravel	10	0	0	10	0	0
Coarse-sand	40	5	0	40	5	0
Sand	80	10	0	45	10	0
Fine-sand	70	25	5	55	15	0
Coarse-silt	50	40	10	65	20	5
Fine-silt	10	75	15	55	35	10
Clay	0	65	35	0	85	15
Peat	0	0	0	0	0	0

**Table 2.** Descriptive characteristics (mean, confidence interval, 1<sup>st</sup>, 50<sup>th</sup>, and 99<sup>th</sup> percentile) of selected variables (n = 3230 samples). The values of the bulk density, cation exchange capacity, base saturation, C/N ratio, and pH are shown only for BC soil horizon (fixed 45–50 cm depth from the ground surface) due to the strong correlation to the total soil carbon stock. The productivity class (H100, m) is an approximation of the site fertility expressed as the height of trees at 100 years of age. Stand and understory biomass, and litter input are modelled values for approximated steady state conditions based on actual state measurements.

	Mean	CI	1 <sup>st</sup> percentile	50 <sup>th</sup> percentile	99 <sup>th</sup> percentile
Total soil carbon stock (tC ha <sup>-1</sup> )	93.24	1.95	17.02	79.68	308.68
Humus carbon stock (tC ha <sup>-1</sup> )	33.29	1.17	3.89	22.82	176.66
Mineral soil carbon stock (tC ha <sup>-1</sup> )	66.82	1.7	6.92	54.81	273.91
Depth of humus (cm)	10.52	0.27	1	8	36
Depth of soil (cm)	93.37	0.6	18	99	99
Stoniness (%)	39.91	0.54	3.96	42.37	65.05
Bulk density of BC (g dm <sup>-3</sup> )	1267.1	5.5	790.55	1294.9	1522.13
Cation exchange capacity of BC (mmol <sub>c</sub> kg <sup>-1</sup> )	23.94	1.28	1.53	12.33	203.25
Base saturation of BC (%)	36.44	1.02	4.33	25.73	100
C/N ratio of BC	16.5	0.35	3.33	14.98	62.45
pH of BC	5.17	0.02	4.36	5.08	7.26
Silt content (%)	19.98	0.57	0	15	85
Clay content (%)	3.16	0.25	0	0	35
Sand content (%)	51.25	0.63	0	55	80
Long-term soil moisture (%)	22.36	0.2	10	20	30
Mean air temperature (°C)	4.63	0.09	-0.44	5.34	8.47
Total precipitation (mm)	697.87	7.13	392.54	637.11	1154.55
Nitrogen deposition (kgN ha <sup>-1</sup> y <sup>-1</sup> )	7.17	0.14	2.35	6.56	17.67
Productivity class (H100, m)	23.61	0.21	12	23	36
Total stand biomass (tC ha <sup>-1</sup> )	56.02	1.39	1.34	51.14	156.52
Total understory biomass (tC ha <sup>-1</sup> )	2.69	0.05	0.96	2.37	6.02
Total litterfall input (tC ha <sup>-1</sup> )	3.17	0.03	1.65	3.07	5.28



**Table 3.** Description of models and data inputs relevant for this study.

Model	Yasso07	Q	CENTURY v. 4.0 soil submodel
Time step	Year	Year	Month
Parameters	General (world wide litter bags)	Seven Swedish regions	Two forest sites (evergreen and deciduous)
Carbon pools	Labile (acid -, water -, and ethanol-soluble and non-soluble), recalcitrant (humus)	Cohorts (foliage, stems, branches, coarse roots, fine roots, "grass"), soil organic	Litter (surface structural and metabolic, belowground str. and met.), surface microbial, soil organic matter (active, slow and passive)
Biomass	Biomass components estimated by allometric biomass functions and provided stand data for litter input estimation		
Litter amount	Annual or monthly fractions of biomass components (species specific, same total litter inputs for all models)		
Litter quality	Litterature based solubilities	Estimated cohorts qualities	C/N ratios and lignin/N ratios
Temperature air	Annual mean, monthly amplitude	Annual mean	Max and min monthly mean
Precipitation	<del>Monthly</del> - <del>Annual</del> total	-	Monthly total
Soil properties	-	-	Bulk density, sand, silt, and clay content
Soil depth (m)	1	-	0.2

**Table A1.** Parameter estimates and their standard errors of the  $f_{APAR}$  regressions with the stand basal area (BA,  $\text{m}^2 \text{ha}^{-1}$ ), and the  $f_{APAR70LAT}$  and  $f_{APAR70H100}$  regressions with the latitude (LAT,  $^\circ$ ) and with the productivity class (H100, m) for Scots pine, Norway spruce, and deciduous stands.

$f_{APAR} = a * BA / (b + BA)$	a $\pm$ SE	b $\pm$ SE	c $\pm$ SE	adj. $R^2$
pine	0.996 $\pm$ 0.029	11.754 $\pm$ 0.811		0.85
spruce	1.167 $\pm$ 0.034	10.668 $\pm$ 0.870		0.86
deciduous	1.129 $\pm$ 0.064	7.407 $\pm$ 1.149		0.88
$f_{APAR70LAT} = LAT / (a + b * LAT) + c$				
pine	-9.976e+03 $\pm$ 3.691e+03 <sup>a</sup>	1.430e+02 $\pm$ 5.416e+01 <sup>b</sup>	7.220e-01 $\pm$ 1.819e-02	0.92
spruce	-2.689e+03 $\pm$ 3.507e+03 <sup>c</sup>	3.533e+01 $\pm$ 5.025e+01 <sup>d</sup>	9.654e-01 $\pm$ 9.221e-02	0.74
$f_{APAR70LAT} = a + b * LAT$				
deciduous	1.363 $\pm$ 0.282	-0.009 $\pm$ 0.005 <sup>e</sup>		0.26
$f_{APAR70H100} = a * e^{(b/H100)}$				
pine	0.85565 $\pm$ 0.01917	-5.22016 $\pm$ 0.40807		0.89
spruce	0.96726 $\pm$ 0.01009	-2.85354 $\pm$ 0.21634		0.86
deciduous	0.93991 $\pm$ 0.02331	-2.63462 $\pm$ 0.50325		0.51

$p < 0.001$  for all parameters except for <sup>a</sup> 0.023, <sup>b</sup> 0.024, <sup>c</sup> 0.461, <sup>d</sup> 0.498, and <sup>e</sup> 0.076.

**Table B1.** Parameter estimates and their standard errors for the coefficients of the dry weight biomass ( $\text{kg ha}^{-1}$ ) models with the fraction of absorbed radiation ( $y = ab^{f_{APAR}}$ ) for Scots pine, Norway spruce, and deciduous stands.

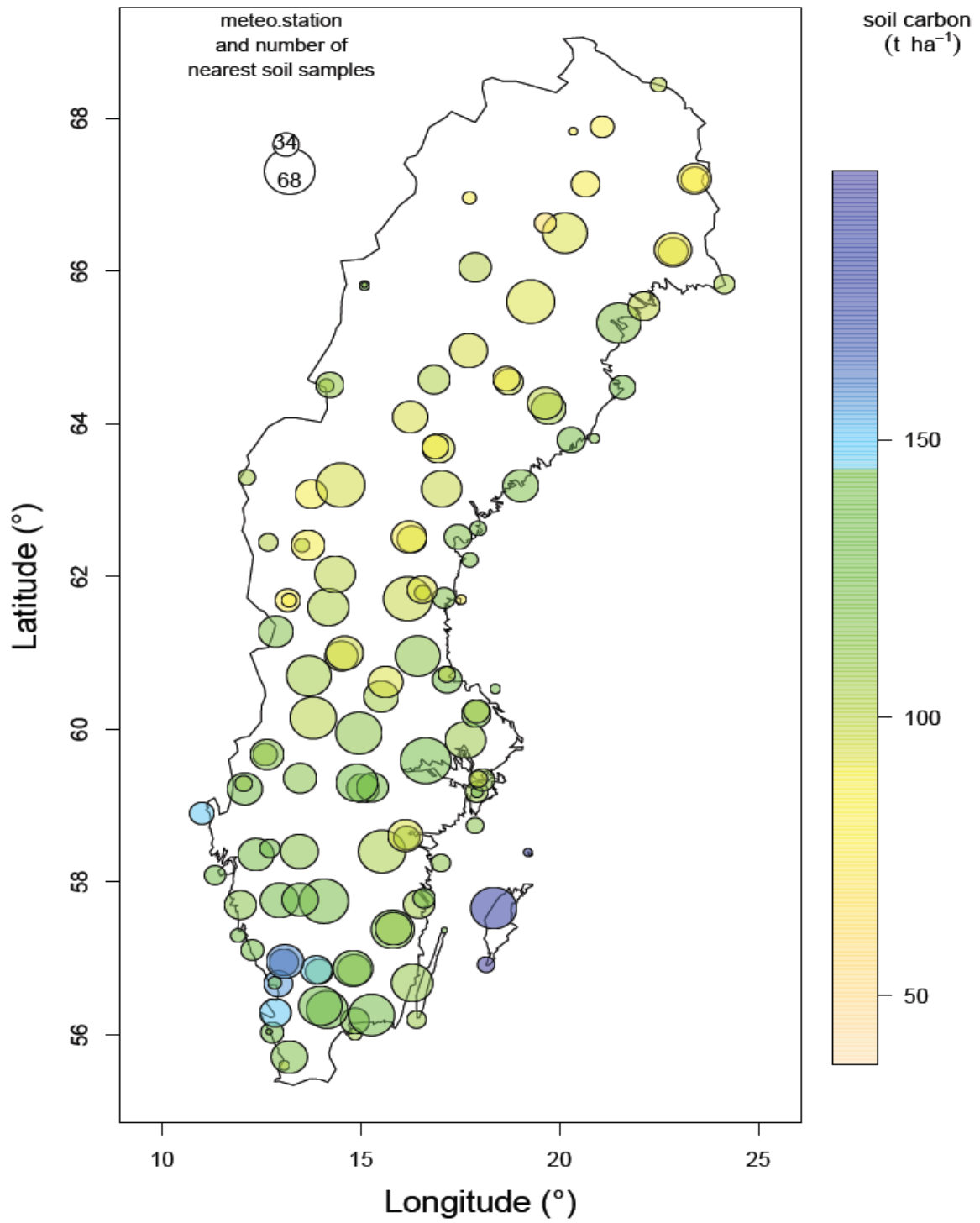
$y = ab^{f_{APAR}}$	species	$a \pm \text{SE}$	$b \pm \text{SE}$	$adj. R^2$
branch	pine	610.23±21.043	121.592±5.967	0.917
	spruce	877.265±34.535	54.157±2.457	0.918
	deciduous	289.719±26.464	155.506±15.838	0.892
fineroot	pine	422.031±12.675	20.51±0.914	0.836
	spruce	316.675±13.816	15.186±0.78	0.799
	deciduous	452.632±27.715	14.499±1.032	0.823
foliage	pine	361.428±24.095	86.091±8.223	0.714
	spruce	766.324±40.277	33.323±2.033	0.827
	deciduous	141.11±28.347	70.629±15.992	0.56
root	pine	703.163±26.166	183±9.62	0.918
	spruce	628.686±32.37	113.435±6.665	0.903
	deciduous	358.635±33.267	149.85±15.506	0.888
stem and bark	pine	1793.215±83.818	253.676±16.658	0.889
	spruce	974.029±72.348	229.024±19.259	0.856
	deciduous	971.587±97.632	160.858±18.015	0.876
stump	pine	231.701±10.273	214.429±13.394	0.893
	spruce	170.77±10.331	129.219±8.907	0.877
	deciduous	79.779±8.388	215.511±25.165	0.874

$p < 0.001$  for all parameters.

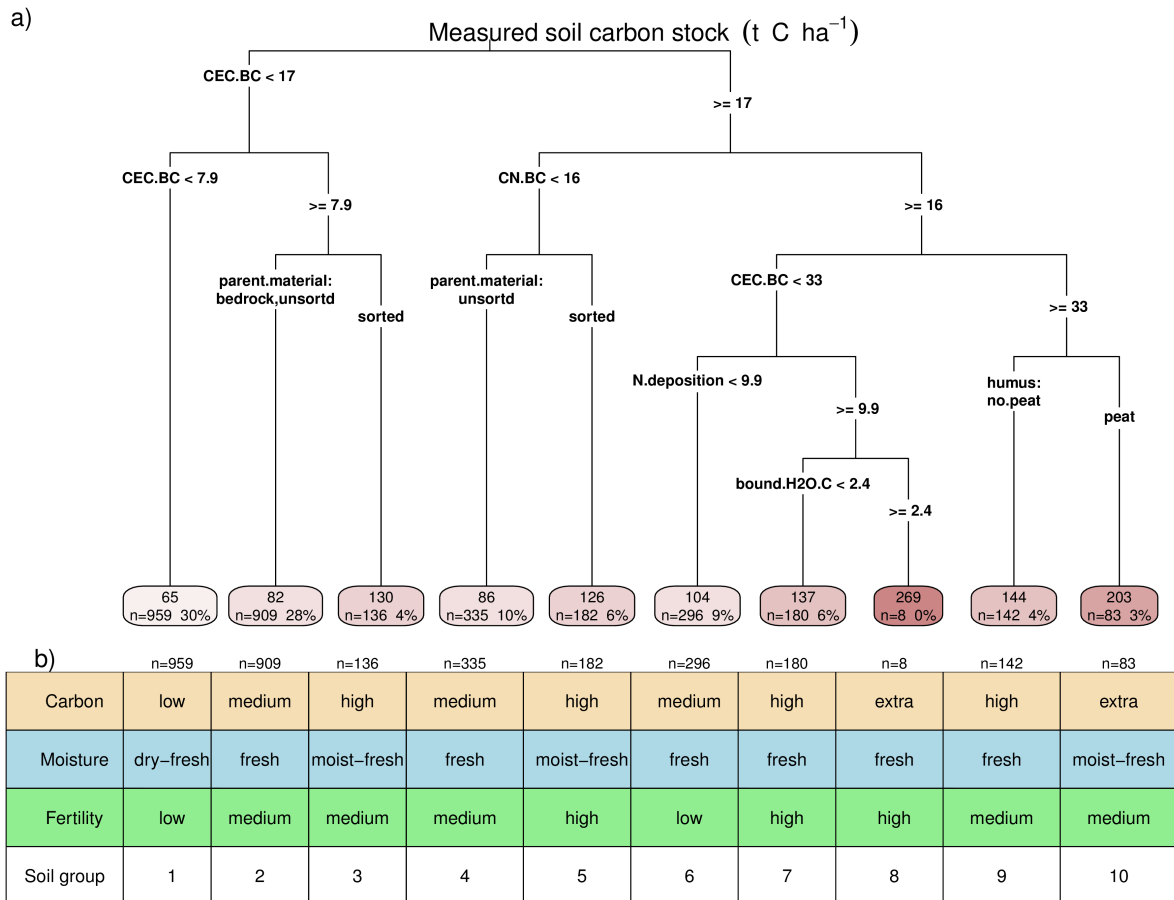
**Table C1.** Parameter estimates and their standard errors for the coefficients of the forest ground-understory vegetation dry weight biomass ( $W$ , kg ha<sup>-1</sup>) models (Eq. C1) for functional types (1-dwarfshrubs, 2-herbs, 3-grasses, 4-mosses and 5-lichens) with intercept (a) and n number of predictors (b1- age (years), b2 – basal area (m<sup>2</sup> ha<sup>-1</sup>), b3 – annual air temperature (°C), b4 - latitude (°), b5 – H100 (height of trees at 100 years of age, m), b6 – H100 of spruce trees (m), b7 – H100 of pine trees (m), b8- pine dominance (0/1), b9-spruce dominance (0/1)). For the latin names of species included into understory functional types see Table S3.

W		a±SE	b1±SE	b2±SE	b3±SE	b4±SE	b5±SE	b6±SE	b7±SE	b8±SE	b9±SE	adj. R <sup>2</sup>
Above-ground	1	24.28±0.32	0.13±0.01	-0.43±0.02						7.13±0.33		0.29
	2	-82.13±6.8			-0.1±0.1 <sup>a</sup>	1.23±0.1		0.77±0.03				0.12
	3	4.07±0.30		-0.16±0.01				0.27±0.01		-1.36±0.15		0.21
	4	32.9±0.62					-0.78±0.04		0.48±0.06	3.66±0.3	5.76±0.29	0.22
	5	19.91±0.57		-0.13±0.01					-0.45±0.02		6.31±0.29	0.25
	total	43.68±0.29	0.12±0.01	-0.41±0.01						6.34±0.3		0.30
Below-ground	1	-256.3±3.5	0.1±0.01	-0.35±0.02		5.05±0.06				8.56±0.35		0.75
	2	-89.34±7.85			-0.03±0.1 <sup>b</sup>	1.4±0.12		0.78±0.04		-4.97±0.27		0.19
	3	5.97±0.37		-0.19±0.01				0.32±0.01		-1.78±0.19		0.21
	total	-251.9±3.3		-0.2±0.01		5.15±0.05						0.7
Total		-222.7±4.0	0.12±0.01	-0.44±0.02		4.9±0.07						0.67

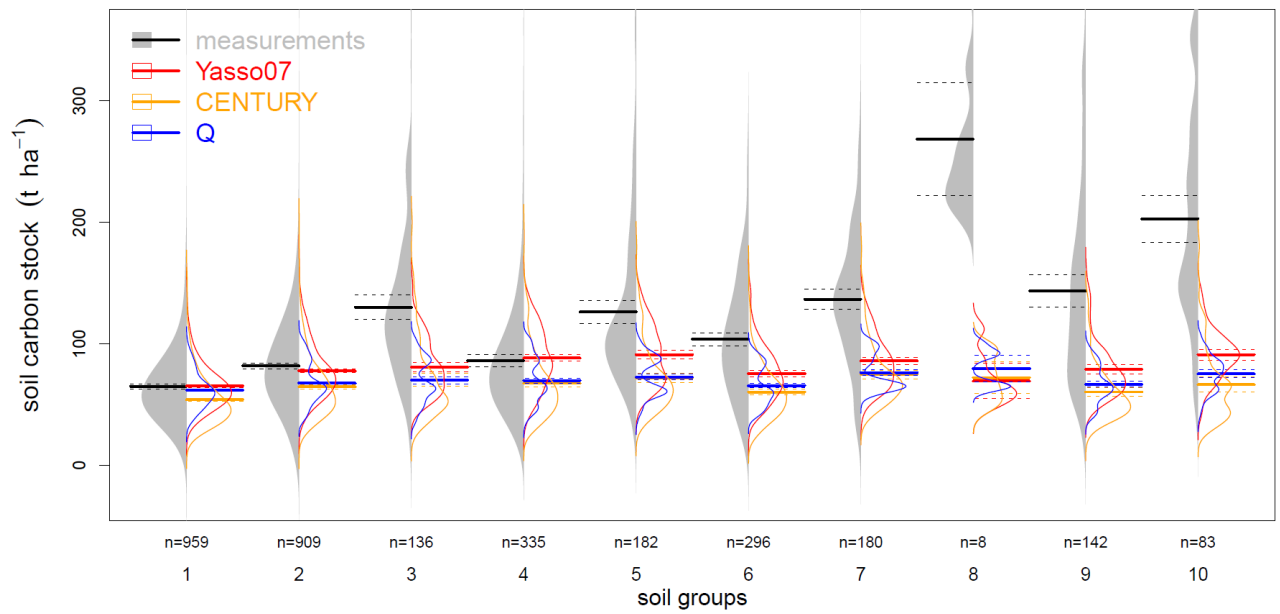
$p < 0.001$  for all parameters except for <sup>a</sup> $p = 0.44$ , and <sup>b</sup> $p = 0.84$ .



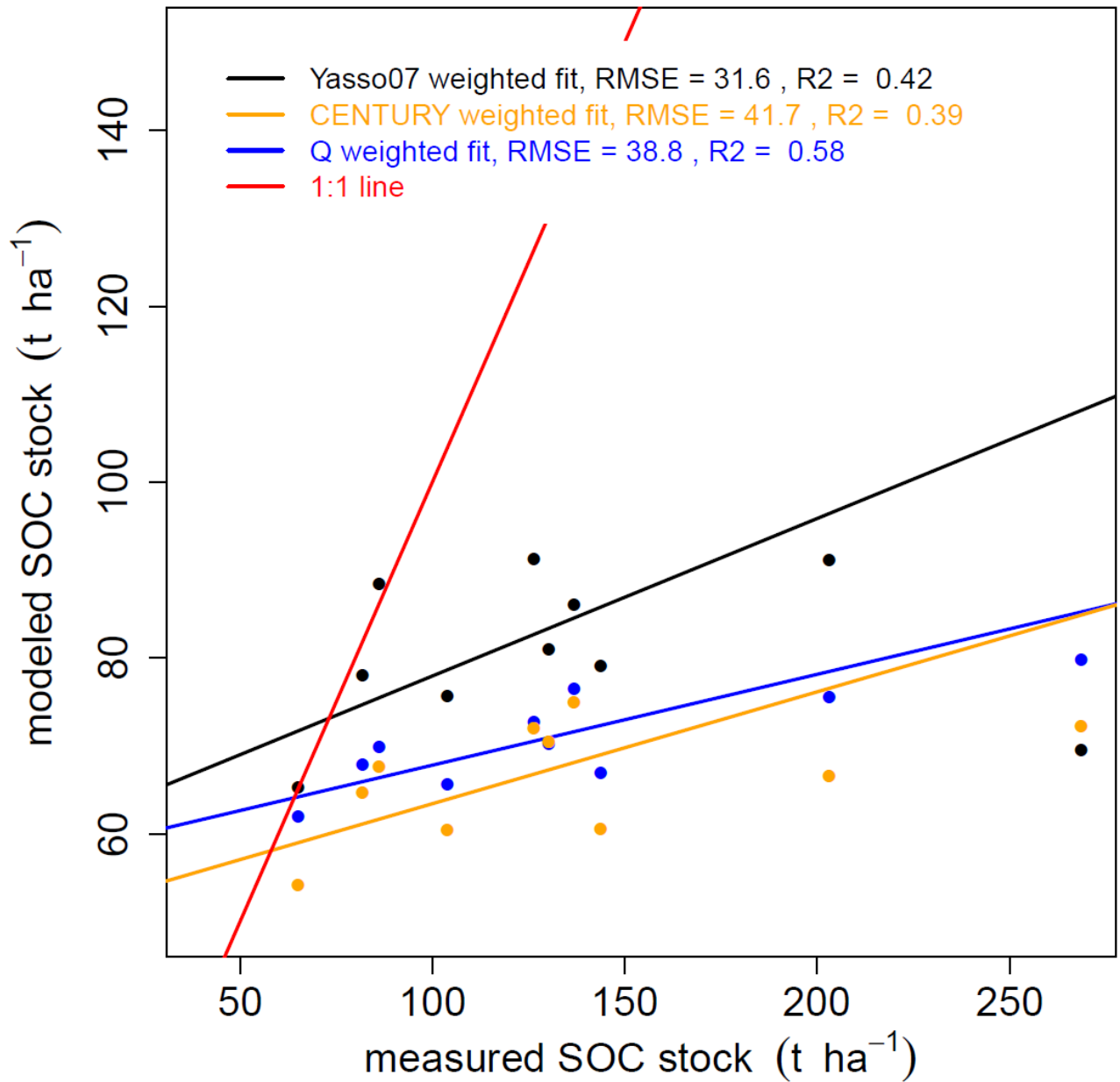
**Figure 1.** Geographical locations of meteorological stations with corresponding number of nearest soil samples (n, size of the circle) and their mean measured soil organic carbon stock (tC ha<sup>-1</sup>, color of the circle) across Sweden.



**Figure 2.** a) Classification/regression tree for the measured soil carbon stock (tC ha<sup>-1</sup>), soil physicochemical properties and site environmental characteristics; the cation exchange capacity of BC horizon (CEC.BC, (mmol<sub>c</sub> kg<sup>-1</sup>)), the C/N ratio (CN.BC), the nitrogen deposition (N.deposition kgN ha<sup>-1</sup> y<sup>-1</sup>), the highly bound soil water of C horizon (bound.H2O.C, %), and soil class variables as type of sorted or unsorted soil parent material and humus type. Note that variables used to calculate the soil carbon stock (bulk density, carbon content, depth, and stoniness) were excluded from the regression tree analysis. The values in the leaves of the tree show for the distinct environmental conditions mean soil carbon stock (tC ha<sup>-1</sup>), number and percentage of samples. b) The interpretation of 10 physicochemical soil groups of the regression tree model into the levels of carbon, soil moisture, and fertility roughly increasing from left to right.

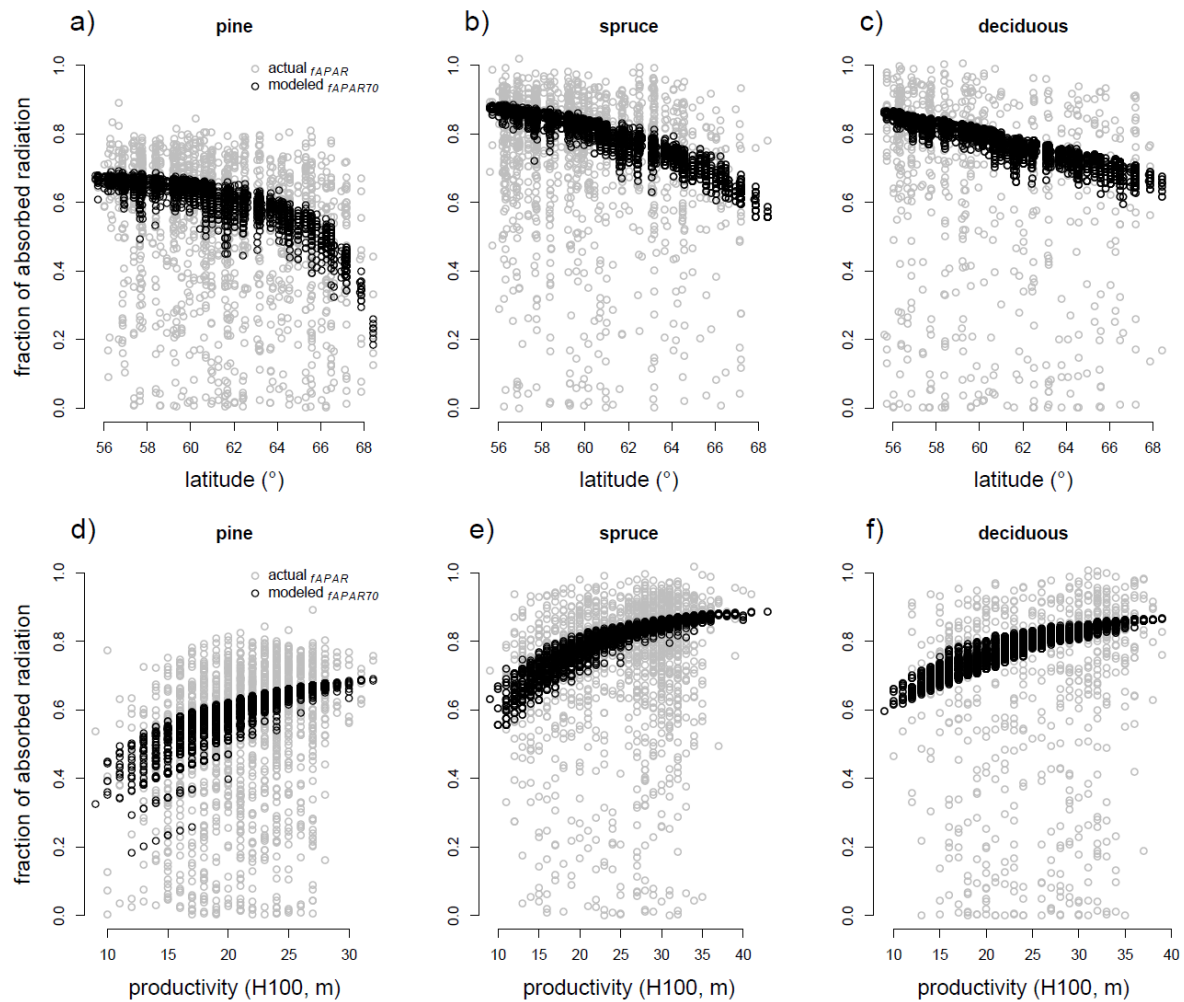


**Figure 3.** Bean plot of density functions for 10 physicochemical groups of the soil carbon (tC ha<sup>-1</sup>) measurements (grey fill) and estimates simulated by the soil carbon models Yasso07, CENTURY, and Q with the litter input derived from the steady state forest. The thin lines are the density distributions. The thick lines are the group means and dashed lines are their confidence intervals. The n is number of samples. For description of group levels of SOC stocks, moisture, and fertility see Fig.2 and Table S1.

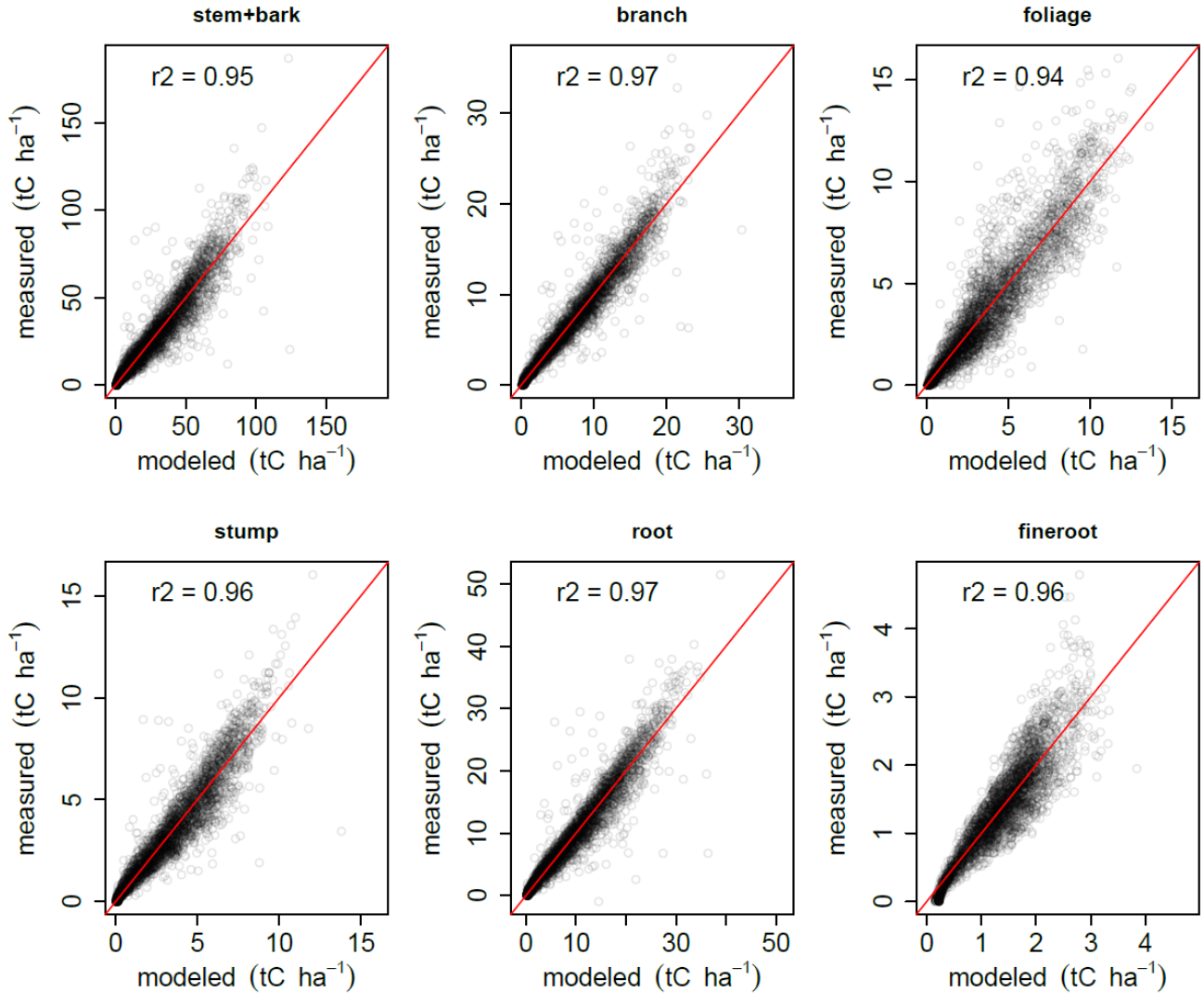


**Figure 4.** Scatter plot between mean measured and mean modeled soil organic carbon stocks (tC ha<sup>-1</sup>) for 10 physicochemical groups for Yasso07, CENTURY and Q models. Data were fitted with weighted linear regression (lines). The number of samples in each group was used as weights for fitting and also as weights for the weighted mean of squared differences between the modeled and measured values (MSE, tC ha<sup>-1</sup>). The RMSE is the square root of MSE. The  $r^2$  is the proportion of explained variance.





**Figure A1.** Actual state fraction of absorbed radiation ( $f_{APAR}$ , estimated as in Härkönen et al., 2010) (measured actual  $f_{APAR}$ ) and steady state  $f_{APAR}$  (modeled max  $f_{APAR70}$ ) which was set to 70th percentile of maximum  $f_{APAR}$  for given species, latitudinal degree, and site productivity class. Panels a), b), and c) show relation between  $f_{APAR}$  and latitude ( $^{\circ}$ ) for forest stands dominant by Scots pine, Norway spruce and deciduous species, whereas panels d), e), and f) show relation between  $f_{APAR}$  and site productivity class (H100, height of dominant trees at 100 years in meters).



**Figure B1.** Scatter plots for the dry weight tree biomass components ( $W_i$ , tC ha<sup>-1</sup>) between "modelled" (estimated based on fraction of absorbed radiation,  $f_{APAR}$ , and our  $f_{APAR}$  models) and "measured" (estimated based on basic tree stand dimensions and allometric biomass models) and "modelled" (estimated based on fraction of absorbed radiation). The  $r^2$  values represent the coefficient of determination indicating how close the modeled values fit the measured values.

# **SUPPLEMENTARY MATERIAL: Underestimation of boreal soil carbon stocks by mathematical soil carbon models linked to soil nutrient status**

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## **1 Source codes of the Yasso07, Q, and CENTURY model**

### **1.1 Yasso07 model**

[https://code.google.com/p/yasso07ui/source/browse/trunk/y07\\_subroutine.f90](https://code.google.com/p/yasso07ui/source/browse/trunk/y07_subroutine.f90)

MODULE yasso

10 IMPLICIT NONE

CONTAINS

    SUBROUTINE mod5c(a,t,cl,init,inf,s,leac,z)

        !components separately

    IMPLICIT NONE

15     !\*\*\*\*\* &

        !GENERAL DESCRIPTION FOR ALL THE MEASUREMENTS

   !\*\*\*\*\* &

   !returns the model prediction for given parameters

   ! 1-16 matrix A entries: 4\*k, 12\*p

20     !17-19 Climate-dependence parameters: b1, b2, g1

   !20-22 Leaching parameters: f1, f2, f3 IGNORED IN THE Y07-UI

```

!23-25 Woody parameters
!26-27 Humus parametens: kH, pH
REAL,DIMENSION(27),INTENT(IN) :: a !parameters
25 REAL,INTENT(IN) :: t,s,leac !time,size,leaching
REAL,DIMENSION(3),INTENT(IN) :: cl !climatic conditions
REAL,DIMENSION(5),INTENT(IN) :: init
REAL,DIMENSION(5),INTENT(IN) :: inf !infall
REAL,DIMENSION(5),INTENT(OUT) :: z
30 REAL,DIMENSION(5,5) :: m,mt,m2,mi
INTEGER :: i
REAL,PARAMETER :: pi=3.1415926535
REAL :: tem
REAL,DIMENSION(5) :: te
35 REAL,DIMENSION(5) :: z1,z2
!temperature annual cycle approximation
te(1)=cl(1)+4*cl(3)*(1/SQRT(2.0)-1)/pi
te(2)=cl(1)-4*cl(3)/SQRT(2.0)/pi
te(3)=cl(1)+4*cl(3)*(1-1/SQRT(2.0))/pi
40 te(4)=cl(1)+4*cl(3)/SQRT(2.0)/pi
tem=0.0
DO i=1,4 !Annual cycle, different models
    tem=tem+EXP(a(17)*te(i)+a(18)*te(i)**2.0)/4.0 !Gaussian
END DO
45 !Precipitation dependence
tem=tem*(1.0-EXP(a(19)*cl(2)/1000))
!Size class dependence - - no effect if sc = 0.0
m(1,1)=a(1)*tem*MIN(1.0,(1.0+a(23)*s+a(24)*s**2.0)**a(25))
m(2,2)=a(2)*tem*MIN(1.0,(1.0+a(23)*s+a(24)*s**2.0)**a(25))
50 m(3,3)=a(3)*tem*MIN(1.0,(1.0+a(23)*s+a(24)*s**2.0)**a(25))
m(4,4)=a(4)*tem*MIN(1.0,(1.0+a(23)*s+a(24)*s**2.0)**a(25))
!Calculating matrix M, normal
m(2,1)=a(5)*ABS(m(2,2))
m(3,1)=a(6)*ABS(m(3,3))
55 m(4,1)=a(7)*ABS(m(4,4))
m(5,1)=0.0
m(1,2)=a(8)*ABS(m(1,1))
m(3,2)=a(9)*ABS(m(3,3))

```

```

m(4,2)=a(10)*ABS(m(4,4))
60 m(5,2)=0.0
m(1,3)=a(11)*ABS(m(1,1))
m(2,3)=a(12)*ABS(m(2,2))
m(4,3)=a(13)*ABS(m(4,4))
m(5,3)=0.0
65 m(1,4)=a(14)*ABS(m(1,1))
m(2,4)=a(15)*ABS(m(2,2))
m(3,4)=a(16)*ABS(m(3,3))
m(5,4)=0.0
m(5,5)=a(26)*tem !no size effect in humus
70 DO i=1,4
    m(i,5)=a(27)*ABS(m(i,i)) !mass flows EWAN -> H
END DO
!Leaching
m(1,1)=m(1,1)+leac*c1(2)/1000
75 m(2,2)=m(2,2)+leac*c1(2)/1000
m(3,3)=m(3,3)+leac*c1(2)/1000
m(4,4)=m(4,4)+leac*c1(2)/1000
!DY solution starts here...
DO i=1,5
80 z1(i)=DOT_PRODUCT(m(:,i),init)+inf(i)
END DO
mt=m*t
CALL matrixexp(mt,m2)
DO i=1,5
85 z2(i)=DOT_PRODUCT(m2(:,i),z1)-inf(i)
END DO
CALL inverse(m,mi)
DO i=1,5
z1(i)=DOT_PRODUCT(mi(:,i),z2)
90 END DO
z=z1
CONTAINS
SUBROUTINE matrixexp(a,b)
IMPLICIT NONE
95 !returns approximated matrix exponential

```

```

!Taylor approximation.. another algorithm perhaps?
REAL,DIMENSION(5,5),INTENT(IN) :: a
REAL,DIMENSION(5,5),INTENT(OUT) :: b
REAL,DIMENSION(5,5) :: c,d
100 REAL :: p,normiter
INTEGER :: i,q,j
q=10
b=0.0
DO i=1,5
105   b(i,i)=1.0
END DO
normiter=2.0
j=1
CALL matrix2norm(a, p)
110 DO
    IF(p< normiter)THEN
        EXIT
    END IF
    normiter=normiter*2.0
115   j=j+1
END DO
c=a/normiter
b=b+c
d=c
120 DO i=2,q
    d=MATMUL(c,d)/REAL(i)
    b=b+d
END DO
DO i=1,j
125   b=MATMUL(b,b)
END DO
END SUBROUTINE matrixexp
SUBROUTINE matrix2norm(a,b)
IMPLICIT NONE
130   !returns matrix 2-norm with cartesian vector x,
      !| | x| | = 1
      !square matrix input (generalize if necessary)

```

```

REAL,DIMENSION(5,5),INTENT(IN) :: a
REAL,INTENT(OUT) :: b
135  INTEGER :: n,i
      n=SIZE(a(1,:))
      b=0.0
      DO i=1,n
          b=b+SUM(a(:,i))*2.0
140  END DO
      b=SQRT(b)
END SUBROUTINE matrix2norm
SUBROUTINE inverse(a,b)
IMPLICIT NONE
145  !returns an inverse of matrix a
      !(column elimination strategy)
      !input has to be a square matrix, otherwise erroneous
REAL,DIMENSION(5,5),INTENT(IN) :: a
REAL,DIMENSION(5,5),INTENT(OUT) :: b
150  REAL,DIMENSION(5,5) :: c
      INTEGER :: n,m,i,j
      n=SIZE(a(1,:))
      m=SIZE(a(:,1))
      IF (m/=n) THEN
155  WRITE(*,*) " Does not compute."
          WRITE(*,*) " No square matrix input."
          WRITE(*,*) " Error in function: inverse"
      ELSE
!       ALLOCATE(b(n,n),c(n,n))
160  c=a
          b=0.0
          DO i=1,n !setting b a unit matrix
              b(i,i)=1.0
          END DO
165  DO i=1,n
          !what if diagonal values are zeros?
          IF(c(i,i)==0.0)THEN!case of singlar matrix, is it?
              b(i,:)=0.0
              c(i,:)=0.0
          END IF
      END DO

```

```

170         b(:,i)=0.0
           c(:,i)=0.0
           !           b(i,i)=1.0
           !           c(i,i)=1.0
           ELSE
175         b(i,:)=b(i,+)/c(i,i)
           c(i,:)=c(i,+)/c(i,i)
           END IF
           DO j=1,i-1
           b(j,:)=b(j,)-b(i,)*c(j,i)
180         c(j,:)=c(j,)-c(i,)*c(j,i)
           END DO
           DO j=i+1,n
           b(j,:)=b(j,)-b(i,)*c(j,i)
           c(j,:)=c(j,)-c(i,)*c(j,i)
185         END DO
           END DO
           IF (c(n,n)==0.0) THEN
           b(n,:)=0.0
           b(:,n)=0.0
190         !           b(n,n)=1.0
           ELSE
           b(n,:)=b(n,+)/c(n,n)
           END IF
           !now, b is supposed to be the requested inverse
195         END IF
           END SUBROUTINE inverse
           END SUBROUTINE mod5c
           END MODULE yasso

```

## 200 1.2 Q model

```

!***** &
!   Main program to calculate carbon store in forest soils.
!   SPRUCE
!***** &
205 !   Edited by Carina Ortiz, Version 2015-11-16

```



```

! Understory vegetation, variable temp, variable litter
  real, dimension(1) :: yr(0:0),carb(0:0),
    + totc(0:0),u0y(0:0)
  real, dimension(1,5) :: lit(0:0,5),cssn(0:0,1),
210   + vtemp(0:0,1:244),cssnund(0:0,1),ssi0b(0:0,1),
    + ssi0scr(0:0,1),ssi0s(0:0,1),alfanss(0:0,1),
  z(0:0,1),u0(0:0,1)
  * ,ssi0b(0:0,5),ssi0scr(0:0,5),ssi0s(0:0,5)
  * + ssi0scr(0:0,5),cssnund(0:0,5)
215   real, dimension (244,0:0) :: nefr !(jj,ii)
  real, dimension (244,0:0) :: br !(jj,ii)
  real, dimension (244,0:0) :: st !(jj,ii)
  real, dimension (244,0:0) :: stpcr !(jj,ii)
  real, dimension (244,0:0) :: und !(jj,ii)
220  real, dimension (244,0:0) :: vtemp !(jj,ii)
! needlesfineroots jj columns(MCSIMS), rows(yr) ii
  real i0, u0sum, ts
  character (len=10) :: dumtext
  integer :: r
225 ! Monte Carlo Simulations
! Read file with parameter setups
! Open parameter file
OPEN (unit=111, file='lparvaraccregsprucemean.dat',status='old')
  read(111,*) dumtext
230 ! Open output file
  open(unit=11, file='q_soil.dat')
! Read matrix file with litter production in N simulations
! (Monte Carlo), each fraction one separate file
  open(unit=1111,file='biom70reg1SpruceNeedlesandfineroots.dat',
235   + status='old')
  read(1111,*) dumtext
  read(1111,*) nefr
  open(unit=11111,file='biom70reg1SpruceBranches.dat',
240   + status='old')
  read(11111,*) dumtext
  read(11111,*) br

```

```

open(unit=1111111,file='biom70reg1SpruceStumpCoarseroots.dat',
+   status='old')
245   read(1111111,*) dumtext
      read(1111111,*) stpcr
      open(unit=1111111,file='biom70reg1SpruceStem.dat',
+   status='old')
      read(1111111,*) dumtext
250   read(1111111,*) st
open(unit=11111111,file='biom70reg1SpruceUnderveg.dat ',
+   status='old')
      read(11111111,*) dumtext
      read(11111111,*) und
255 ! Read variable temperature file yearly means
open(unit=1, file='reftempreg1spruce.dat',status='old')
      read(1,*) dumtext
      read(1,*) vtemp
! Estimates the mean temperature for steady state
260 *   ts=ts+vtemp
*   ii=ii+1
*   temp=ts/ii !mean temp
! Main loop Number of Monte Carlo simulations
DO ii=1,244
265 read(111,*) q0n,q0w,e0,etal1,beta,maxb,maxs,u00,u01
! Temperature for steady state.
! First row temperature, only for one step
do jj=0,0!first row is dumtext !
      vtempsite(jj,ii) = vtemp(ii,jj)!
270   u0(jj,1)=u00+u01*vtempsite(jj,ii)
      do i=0,0
          u0y(i)=u00+u01*vtempsite(jj,ii) ! each year new temp.
      enddo
      enddo
275 ! Set some parameters for decomposition functions
      ssi0n=0
      fC=0.5
      do jj=0,0
          alfanss(jj,1)=fC*beta*etal1*u0(jj,1)*q0n**beta

```

```

280   z(jj,1)=(1.-e0)/(beta*etall*e0)
      end do
! Create litter matrix
      do jj=0,0!first row is dumtext
          lit(jj,1) = nefr(ii,jj)
285     !litter fraction 1 needles & fine roots
          lit(jj,2) = br(ii,jj)
          !litter fraction 2 branches & roots
          lit(jj,3) = st(ii,jj)
          !litter fraction 3 stems
290     lit(jj,4) = stpcr(ii,jj)
          !litter fraction 4 stump & coarse roots
          lit(jj,5) = und(ii,jj)
          !litter fraction 5 understorey
      enddo
295 !Calculates steady state litter fraction for needles
      do jj=0,0
          cssn(jj,1)=(lit(jj,1))*1/(alfanss(jj,1)*(z(jj,1)-1))
      enddo
!Steady state understorey vegetation
300   do jj=0,0
          cssnund(jj,1)=(lit(jj,5))*1/(alfanss(jj,1)*(z(jj,1)-1))
      enddo
!Steady state input branches
      do jj=0,0
305     ssi0b(jj,1)=lit(jj,2)
      enddo
!Steady state input stump and coarse roots
      do jj=0,0
          ssi0scr(jj,1)=lit(jj,4)
310     enddo
!Steady state input stem
      do jj=0,0
          ssi0s(jj,1)=lit(jj,3)
      enddo
315 ! Fixed parameters
      fC=0.5

```

```

!      Call for soil decomposition model
      call soildecomp(fC,u0,u0y,etall,e0,beta,ssi0b,ssi0s,ssi0scr,
+
+          q0n,cssn,q0w,maxb,maxs,itend,lit,carb,nitr,
320 +          cssnund)
! WRITE TO OUTPUTFILE
* do i=0,0
* write(11,*) carb(i)
* enddo
325 ! End main loop
* enddo !input litter loop
ENDDO !parameter loop
*! Close Temperature file
close(unit=1)
330 ! Close litterfiles
close(unit=1111)
      close(unit=11111)
close(unit=111111)
close(unit=1111111)
335 close(unit=11111111)
! Close OUTPUT document
close(unit=11)
! Close parameterrange file
close(unit=111)
340 !***** &
!      Subroutine to calculate carbon stores in forest soils
!      Version 2015-11-16
!      Undervegetation, variable temp, variable litter
      subroutine soildecomp (fC,u0,u0y,etall,e0,beta,ssi0b,ssi0s,
345 + ssi0scr,q0n,cssn,q0w,maxb,maxs,itend,lit,carb,nitr,cssnund)
      real, dimension(1) :: carb(0:0), totc(0:0),
+          u0y(0:0)
      real, dimension(1,5) :: lit(0:0,5), vtemp(0:0,1:225)
      real, dimension (225,0:0) :: nefr !(jj,ii)
350 real, dimension (225,0:0) :: br !(jj,ii)
      real, dimension (225,0:0) :: st !(jj,ii)
      real, dimension (225,0:0) :: stpcr !(jj,ii)
      real, dimension (225,0:0) :: und !(jj,ii)

```

```

    real, dimension (225,0:0) :: vtemp !(jj,ii)
355 * needlesfineroots jj columns(MCSIMS), rows(yr) ii
        real i0, u0sum, t, gnv, gnold, gbv, gbold, gsv, gsold
        dimension gn(0:0), gb(0:0), gs(0:0)
        dimension hn(0:0), hb(0:0), hs(0:0), qn(0:0)
*       real, dimension (101,100):: a(0:100,1:100)
360     integer :: r
!       Litter and OM decomposition parameters
z=(1.-e0)/(beta*etall*e0)
zn=1./(beta*etall*e0)
alfa =fC*beta*etall*u0*q0w**beta
365 !Alfa with constant temp!
alfav=fC*beta*etall*q0w**beta
!Alfa with variable temperature vtemp!
        alfan=fC*beta*etall*u0*q0n**beta
        !Old alfa with constant temp!
370 alfanv=fC*beta*etall*q0n**beta
!Alfa with variable temperature vtemp!
        alfa0n=fN/fC-(beta*etall*e0+e0-1)*(fn/fc-r0)/(beta*etall*e0-1)
!       Old SOM from steady state org.mat. is calculated and summed
        do i=0,0
375     u0sum=0
        do k=1,i !No decomposition the same year as we assess
            u0sum=u0sum+u0y(k)
            !Integral of u0 when using variable temperature
        enddo
380     gnold= (1+alfanv*u0sum)**(1-(1-e0)/
!Decomposition of old needles
        + (e0*etall*beta))
t=(i)
!       Call for decomposition fuctions
385 !       that calculates the remaining mass of the o.m.
        gsold=gold(t,maxs,alfa,alfav,u0sum,z)
        gbold=gold(t,maxb,alfa,alfav,u0sum,z)
!       Old SOM is summed
        carb(i)=
390     + gnold*cssn !part of o.m left and the steady state input

```

```

in fraction
    + +gnold*cssnund
    + +gbold*ssi0b
    + +gsold*ssi0s
395    + +gsold*ssi0scr
write(11,*) carb (i)
enddo
! New SOM from is calculated
! and summed with the olds steady state carbon
400 do i=0,0
    do mt=0,i
        u0sum=0
        if (i.ne.mt) then
            !No decomposition the same year as we assess
405    do k=(mt+1),i
                u0sum=u0sum+u0y(k)
                !Integral of u0 when using variable temperature
            enddo
        endif
410    gnv=1./(1.+alfanv*u0sum)**z
        !Decomposition of new needles!
        t=(i-mt)
        gbv=gbran(t,maxb,alfa,alfav,u0sum,z)
        !Decomposition of new branches!
415    if (gbv.gt.1) then
            gbv=1
        endif
        gsv=gbran(t,maxs,alfa,alfav,u0sum,z)
        !Decomposition of new stems!
420    if (gbs.gt.1) then
            gbs=1
        endif
! SOM from old steady state and new litter is summed
carb(i)=carb(i)+
425    + lit(mt,1)*gnv+
    + lit(mt,2)*gbv+
    + (lit(mt,3)+lit(mt,4))*gsv+

```

```

+    lit(mt,5)*gmv
enddo
430    enddo
!    Litter fractions are 1=needles 2=branches
!    3=stem 4=stump & course roots 5=under vegetation
9991  continue
      return
435 999  end
!    Function for decomposition of old steady state carbon
!    for branches (2) and stems(3+4)
      function gold(t,itmax,alfa,alfav,u0sum,z)
!Decomposition of old branches & stem!
440  tmax=real(itmax)
      if (t.le.tmax) then
gold=-2.*(1+alfav*u0sum)**(2.-z)/(tmax*alfa**2*(1.-z)*(2.-z))
+2.*(1+alfav*u0sum)**(3.-z)-1.)
+ / (tmax**2*alfa**3*(1.-z)*(2.-z)*(3.-z))
445  + +(1.-t/tmax)**3*tmax/3.- (1.-t/tmax)**2/(alfa*(1.-z))
+ +2.*(tmax-t)/(tmax**2*alfa**2*(1.-z)*(2.-z))
      return
      else
gold=-2.*(1+alfav*u0sum)**(2.-z)/(tmax*alfa**2*(1.-z)*(2.-z))
450  + 2.*(1+(alfav*u0sum-alfa*(tmax)))**(3.-z)-
+ (1+alfav*u0sum)**(3.-z))
+ / (tmax**2*alfa**3*(1.-z)*(2.-z)*(3.-z))
      return
      endif
455  end
      function gbran(t,itmax,alfa,alfav,u0sum,z)
!Decomposition of new branches!
      tmax=real(itmax)
      if (t.le.tmax) then
460  gbran=2.*((1+alfav*u0sum)**(1.-z)-(1.-t/tmax))/(tmax*alfa*(1.-z))
+ +2.*(1-(1+alfav*u0sum)**(2.-z))/((tmax**2)*(alfa**2)*
+ (1.-z)*(2.-z))+(1.-t/tmax)**2
      return
      else

```

```

465     gbran=2.*(1.+alfav*u0sum)**(1.-z)/(tmax*alfa*(1.-z))
+ +2.*((1.+(alfav*u0sum-alfa*tmax))**(2.-z)-
+ (1.+alfav*u0sum)**(2.-z))/
+ ((tmax**2)*(alfa**2)*(1.-z)*(2.-z))
     return
470   endif
     end

```

### 1.3 CENTURY model

```

#####
475 ##
##   SOC sub-model of the CENTURY version 4.0
##
#####
#
480 # Coded in R by Shoji Hashimoto (shojih@ffpri.affrc.go.jp)
# edited by Boris Tupek (boris.tupek@luke.fi)
# original model available at
# https://www.nrel.colostate.edu/projects/century/obtain2.htm
#####
485 # Related source files in the original CENTURY model
# Please see /original/source/*.f
#
# adjlig.f, anerob.f, csa_detiv.f, csa_main.f, cycle.f, declig.f
# decomp.f, eachyr.f, h2olos.f, litdec.f, partit.f
490 # pevap.f, prelim.f, simsom.f, somdec.f, tcalc.f
# wdeath.f, woodec.f, and so on.
#
#####
# Simplification:
495 #
# only for forest ecosystem (not grass, savanna etc)
# no irrigation
# not floating C/N ratio for plant organs.
# cnr_max=cnr_min=cnr_initial in tree.100
500 # no mineral N cycling: constant N at surface soil

```



```

# (xNmineral in f_site.100)
# drain=1, anerb=1
# ideo=2 in fix.100 (water function for calculating defac)
# no CO2 effect
505 #
#####
# A bug in the original CENTURY
#
# a bug (please see calfc_wtpt function below)
510 # The difference in results was small,
# but it depends on the climate and soil.
#
# BFix<-0: with bug as the original CENTURY
# BFix<-1: the bug was fixed
515 rm(list=ls())
options(digits=12)
BFix<-1
#DEFINE number of years for spinup simulations!
TSTART=1
520 TEND=500 # 5000 for steady state
#####
# Read data
#####
# parameters from fix.100 in the original CENTURY
525 # environments (site specific temperature,
# precipitation from SMHI), site.100 in the original CENTURY
# parameters describing site conditions(site specific sand,
# silt,clay,bulk density from SFSI data)
# see file site.100 in the original CENTURY
530 # parameters describing tree,
# see tree.100 in the original CENTURY
# "AND H_J ANDREWS" for conifers
# "Coweeta" for deciduous
# initial conditions from site.100
535 ## READ SITE SPECIFIC data #####
#general parameters (fix.100)
parameters.names<-c("adepl","adep2","adep3","adep4","adep5",

```

```

"adep6", "adep7", "adep8", "adep9", "adep10",
"awt11", "awt12", "awt13", "awt14", "awt15",
540 "awt16", "awt17", "awt18", "awt19", "awt110",
"damr11", "damr21", "damrmn", "dec11,
Asrfstr_0", "dec21, Asrfmet_0", "dec12,
Abelstr_0", "dec22, Abelmet_0", "dec31,
Asrfmic_0", "dec32, kactv_0", "dec5, kslow_0",
545 "dec4, kpass_0", "Edepth", "Elitst",
"Fwloss1", "Fwloss2", "Fwloss3", "Fwloss4",
"ntspm, CYCL", "OMLECH(1)", "OMLECH(2)",
"OMLECH(3)", "P1CO2A1", "P1CO2A2", "P1CO2B1",
"P1CO2B2", "P2CO2", "P3CO2", "pabres",
550 "Peftxa", "Peftxb", "pligst1", "pligst2",
"PMCO21", "PMCO22", "PmnTmp", "PmxBio",
"PmxTmp", "PS1CO21", "PS1CO22", "PS1S31",
"PS1S32", "PS2S31", "PS2S32", "Rsplig",
"spl1", "spl2", "strmax1", "strmax2",
555 "teff1", "teff2", "teff3", "Tmelt1", "Tmelt2")
parameters.values <-c(15,15,15,15,30,30,30,30,0,0,0.8,
0.6,0.4,0.3,0.2,0.2,0.2,0.2,0,0,0,
0.02,15,3.9,14.8,4.9,18.5,6,7.3,
0.2,0.0045,0.2,0.4,0.8,0.8,0.65,
560 0.9,4,0.03,0.12,60,0.6,0.17,0,
0.68,0.55,0.55,100,0.25,0.75,3,
3,0.55,0.55,0.004,600,-0.0035,
0.45,0.55,0.003,0.032,0.003,
0.009,0.3,0.85,0.013,5000,
565 5000,0,0.125,0.07,-8,4)
parameters <- data.frame(V1=parameters.values,
V2=parameters.names)
#initial parameters (site.100)
init.names<-c("xsrfstr", "xsrfmet", "xsrfmic", "xbelstr",
570 "xbelmet", "xactv", "xslow", "xpass",
"xwood1", "xwood2", "xwood3",
"rwcf_initial1", "rwcf_initial2",
"rwcf_initial3", "rwcf_initial4",
"rwcf_initial5", "rwcf_initial6",

```

```

575         "rwcf_initial7", "rwcf_initial8",
           "rwcf_initial9", "rwcf_initial10",
           "asmos1", "asmos2", "asmos3", "asmos4",
           "asmos5", "asmos6", "asmos7", "asmos8",
           "asmos9", "asmos10", "asmos11", "snql",
580         "snow", "srfstrlig", "belstrlig")
init.values <-c(240, 60, 60, 186.5, 113.4, 130, 2570,
               1596, 500, 500, 500, 0.5, 0.5, 0.5, 0.5,
               0.5, 0.5, 0.5, 0.5, 0.5, 0.5, 0.2,
               0.2, 0.2, 0.2, 0.2, 0.2, 0.2, 0.2,
585               0.2, 0.2, 0.2, 0, 0, 0.275, 0.354)
init <- data.frame(V1=init.values, V2=init.names)
#site (site.100)
site.parameters.names <-c("sitlat", "sitlog",
                          "sand", "silt", "clay", "bd",
590                          "nlayer", "nlaypg", "drain",
                          "basef", "stormf",
                          "SWFLAGflag_fc_wtpt (0useactual, 1.0Guputa)",
                          "AWILT1", "AWILT2", "AWILT3",
                          "AWILT4", "AWILT5", "AWILT6",
595                          "AWILT7", "AWILT8", "AWILT9",
                          "AWILT10",
                          "AFIEL1", "AFIEL2", "AFIEL3",
                          "AFIEL4", "AFIEL5", "AFIEL6",
                          "AFIEL7", "AFIEL8", "AFIEL9",
600                          "AFIEL10", "elev", "xNmineral")
#site sand, silt, clay, bulk density
site.parameters.values <-c(59.36, 13.47, 0.55, 0.15, 0, 1.226,
                          8, 5, 1, 0.5, 0.9, 1, 0.2, 0.2, 0.2, 0.2,
                          0.2, 0.2, 0.2, 0.2, 0.2, 0.2, 0.3, 0.3,
605                          0.3, 0.3, 0.3, 0.3, 0.3, 0.3, 0.3, 0.3,
                          50, 1.65)

site.parameters <- data.frame(V1=site.parameters.values,
                             V2=site.parameters.names)
#climate environment (site.100)
610 envi.parameters.names <-c("Prec (1) cm", "Prec (2) cm",
                             "Prec (3) cm", "Prec (4) cm", "Prec (5) cm",

```

```

        "Prec (6) cm", "Prec (7) cm",
        "Prec (8) cm", "Prec (9) cm", "Prec (10) cm",
        "Prec (11) cm", "Prec (12) cm",
615    "Tmin (1) degree", "Tmin (2) degree",
        "Tmin (3) degree", "Tmin (4) degree",
        "Tmin (5) degree", "Tmin (6) degree",
        "Tmin (7) degree", "Tmin (8) degree",
        "Tmin (9) degree", "Tmin (10) degree",
620    "Tmin (11) degree", "Tmin (12) degree",
        "Tmax (1) degree", "Tmax (2) degree",
        "Tmax (3) degree", "Tmax (4) degree",
        "Tmax (5) degree", "Tmax (6) degree",
        "Tmax (7) degree", "Tmax (8) degree",
625    "Tmax (9) degree", "Tmax (10) degree",
        "Tmax (11) degree", "Tmax (12) degree")
envi.parameters.values <-c(3.395,2.695,2.884,3.051,3.306,
        4.471,4.623,6.016,5.494,5.221,
        5.659,3.858,-6.647,-7.235,-4.201,
630    -0.121,4.97,9.538,11.74,11.038,7.266,
        3.379,-1.027,-5.32,-0.782,-0.375,3.613,
        9.369,15.549,19.758,21.351,20.219,
        15.489,9.915,4.286,0.584)
envi.parameters <- data.frame(V1=envi.parameters.values,
635    V2=envi.parameters.names)
#tree
tree.parameters.names <-c("cerfor(1:2:3,1,1),cnr_fol",
        "cerfor(1:2:3,3,1),cnr_bra",
        "cerfor(1:2:3,4,1),cnr_ste",
640    "cerfor(1:2:3,2,1),cnr_fir",
        "cerfor(1:2:3,5,1),cnr_cor",
        "DECW1,kwood1_0,bra", "DECW2,
        kwood2_0,ste", "DECW3,kwood3_0,cor",
        "forrtf", "leafdr1", "leafdr2",
645    "leafdr3", "leafdr4", "leafdr5",
        "leafdr6", "leafdr7", "leafdr8",
        "leafdr9", "leafdr10", "leafdr11",
        "leafdr12",

```

```

        "wdlig1,cfol_lig","wdlig3,cbra_lig",
650     "wdlig4,cste_lig","wdlig2,cfir_lig",
        "wdlig5,ccor_lig",
        "wooddr1fol","wooddr3bra",
        "wooddr4ste","wooddr2fir", "wooddr5cor")
tree.parameters.values <-c(20,99,140,40,83,1.5,0.5,0.6,
655     0.5,0,0,0,0.002,0.006,0.012,
        0.013,0.039,0.175,0.664,0.066,
        0.023,0.223,0.25,0.25,0.25,0.25,
        1,0.01,0.002,0.04,0.004)
tree.parameters <- data.frame(V1=tree.parameters.values,
660     V2=tree.parameters.names)
# biomass components gC.m-2
biomass.in <- data.frame(id=1,
        foliage.tot70=795.954,
        branch.tot70=1241.235,
665     wood.tot70=5110.385,
        fineroot.tot70=251.318,
        root.tot70=1652.101)
# litter components gC.m-2
litter.in <- data.frame(id=1,
670     foliage.lit.tot70=116.804,
        branch.lit.tot70=15.515,
        wood.lit.tot70=12.447,
        fineroot.lit.tot70=131.778,
        root.lit.tot70=20.651)
675 # Define objects from SITE SPECIFIC PARAMETERS: #####
# environment(meteo), site, and tree parameters #####
#site specific parameters
envi <- envi.parameters
tree <- tree.parameters
680 site <- site.parameters
## define environment #####
# prec: monthly precipitation, cm
# atempmin: monthly minimum air temperature
# atempmax: monthly maximum air temperature
685 prec<-matrix(0,nrow=12,ncol=1)

```

```

atempmin<-matrix(0,nrow=12,ncol=1)
atempmax<-matrix(0,nrow=12,ncol=1)
for(m in 1:12)
{
690   prec[m]<-envi[m,1]
      atempmin[m]<-envi[m+12,1]
      atempmax[m]<-envi[m+24,1]
}
## define site parameters #####
695 # awilt: wilting point
# afield: field capacity
sitlat<-site[1,1]
sitlog<-site[2,1]
sand<-site[3,1]
700 silt<-site[4,1]
clay<-site[5,1]
bd<-site[6,1]
#use mean soil parameters for swedish soils
#(if soil data is not available)
705 if (is.na(bd)){
      bd<-1.2
}
if(sum(sand,silt,clay)==0){
710   clay<-0.179
      bd<-0.029
}
nlayer<-as.integer(site[7,1])
nlaypg<-as.integer(site[8,1])
715 drain<-site[9,1]
basef<-site[10,1]
stormf<-site[11,1]
flag_fc_wtpt<-as.integer(site[12,1])
awilt<-matrix(0,nrow=10,ncol=1)
720 afield<-matrix(0,nrow=10,ncol=1)
for(i in 1:10)
{

```

```

    awilt[i]<-site[12+i,1]
    afiel[i]<-site[22+i,1]
725 }
    elev<-site[33,1]
    xNmineral<-site[34,1]
    ## define init parameters #####
    # xsrfstr: surface structural
730 # xsrfmet: surface metabolic
    # xsrfmic: surface microbe
    # xbelstr: belowground structural
    # xbelmet: belowground metabolic
    # xactv: actic pool
735 # xslow: slow pool
    # xpass: passive pool
    # xwood1: branch litter
    # xwood2: stem litter
    # xwood3: coarse root litter
740 # rwcfc: volumetric soil water content
    # asmos: soil water content of the ith soil layer cmh2o
    xsrfstr<-init[1,1]
    xsrfmet<-init[2,1]
    xsrfmic<-init[3,1]
745 xbelstr<-init[4,1]
    xbelmet<-init[5,1]
    xactv<-init[6,1]
    xslow<-init[7,1]
    xpass<-init[8,1]
750 xwood1<-init[9,1]
    xwood2<-init[10,1]
    xwood3<-init[11,1]
    tawood <- xwood1 + xwood2
    tbwood <- xwood3
755 talit <- xsrfstr + xsrfmet + xsrfmic
    tblit <- xbelstr + xbelmet
    somsc <- xactv + xslow + xpass
    somtc <- xactv + xslow + xpass + xbelstr + xbelmet
    rwcfc<-matrix(0.1,nrow=10,ncol=1)

```

```

760 for(j in 1:nlayer)
  {
    rwcfc[j]<-init[11+j,1]
  }
  asmos<-matrix(0.1,nrow=11,ncol=1)
765 for(j in 1:(nlayer+1))
  {
    asmos[j]<-init[21+j,1]
  }
  snlq<-init[33,1]
770 snow<-init[34,1]
  srfstrlig<-init[35,1]
  belstrlig<-init[36,1]
  ##
  ## define tree parameters #####
775 # CN ratio of foliage, branch stem, fine roots, coarse roots
  # Decomposition constant
  # Translocation of N
  # Lignin ratios
  # Death rate
780 cnr_fol<-tree[1,1]
  cnr_bra<-tree[2,1]
  cnr_ste<-tree[3,1]
  cnr_fir<-tree[4,1]
  cnr_cor<-tree[5,1]
785 kwood1<-tree[6,1]
  kwood2<-tree[7,1]
  kwood3<-tree[8,1]
  forrtf<-tree[9,1]
  leafdr<-matrix(0,nrow=12,ncol=1)
790 for(j in 1:12)
  {
    leafdr[j]<-tree[j+9,1]
  }
  cfol_lig<-tree[22,1]
795 cbra_lig<-tree[23,1]
  cste_lig<-tree[24,1]

```



```

cfir_lig<-tree[25,1]
ccor_lig<-tree[26,1]
wooddr<-matrix(0,nrow=5,ncol=1)
800 for(j in 1:5)
    {
        wooddr[j]<-tree[j+26,1]
    }
    ## define main (FIX) parameters #####
805 # A: decomposition constant
    # k: decomposition constant
    adep<-matrix(0.1,nrow=10,ncol=1)
    for(j in 1:10)
    {
810     adep[j]<-parameters[j,1]
    }
    #
    #
    awt1<-matrix(0,nrow=10,ncol=1)
815 for(j in 1:10)
    {
        awt1[j]<-parameters[10+j,1]
    }
    damr11<-parameters[21,1]
820 damr21<-parameters[22,1]
    damrmn<-parameters[23,1]
    Asrfstr<-parameters[24,1]
    Asrfmet<-parameters[25,1]
    Abelstr<-parameters[26,1]
825 Abelmet<-parameters[27,1]
    Asrfmic<-parameters[28,1]
    kactv<-parameters[29,1]
    kslow<-parameters[30,1]
    kpass<-parameters[31,1]
830 Edepth<-parameters[32,1]
    elitst<-parameters[33,1]
    fwloss1<-parameters[34,1]
    fwloss2<-parameters[35,1]

```

```

fwloss3<-parameters[36,1]
835 fwloss4<-parameters[37,1]
CYCL<-as.integer(parameters[38,1])
omlech<-matrix(0,nrow=3,ncol=1)
omlech[1]<-parameters[39,1]
omlech[2]<-parameters[40,1]
840 omlech[3]<-parameters[41,1]
P1CO2A1<-parameters[42,1]
P1CO2A2<-parameters[43,1]
P1CO2B1<-parameters[44,1]
P1CO2B2<-parameters[45,1]
845 Psrfmic<-P1CO2A1
Pactv<-P1CO2A2+P1CO2B2*sand
Pslow<-parameters[46,1]
Ppass<-parameters[47,1]
pabres<-parameters[48,1]
850 Peftxa<-parameters[49,1]
Peftxb<-parameters[50,1]
pligst1<-parameters[51,1]
pligst2<-parameters[52,1]
Psrfstr<-parameters[53,1]
855 Psrfmet<-parameters[54,1]
Pbelstr<-Psrfstr
Pbelmet<-Psrfmet
PmnTmp<-parameters[55,1]
PmxBio<-parameters[56,1]
860 PmxTmp<-parameters[57,1]
PS1CO21<-parameters[58,1]
PS1CO22<-parameters[59,1]
ps1s31<-parameters[60,1]
ps1s32<-parameters[61,1]
865 ps2s31<-parameters[62,1]
ps2s32<-parameters[63,1]
RSPLIG<-parameters[64,1]
spl1<-parameters[65,1]
spl2<-parameters[66,1]
870 strmax1<-parameters[67,1]

```

```

strmax2<-parameters[68,1]
teff1<-parameters[69,1]
teff2<-parameters[70,1]
teff3<-parameters[71,1]
875 Tmelt1<-parameters[72,1]
Tmelt2<-parameters[73,1]
#Biomass data from Swe Forest and Soil Inventory #####
#biomass components gC.m-2
#biomass.in
880 pools.bfol<- biomass.in[1,2]
pools.bbba<-biomass.in[1,3]
pools.bste<-biomass.in[1,4]
pools.bfir<-biomass.in[1,5]
pools.bcor<-biomass.in[1,6]
885 #Litterfall SITE SPECIFIC data
litter.in <- litter.in
# Initialization #####
stempave<-0.0
defac<-0.0
890 pet<-0.0
anerb<-0.0
CO2out<-0.0
leaching<-0.0
pet<-matrix(0,nrow=12,ncol=1)
895 avh2o<-matrix(0.0,nrow=3,ncol=1)
amov<-matrix(0.0,nrow=11,ncol=1)
tran<-0.0
evap<-0.0
streaml<-0.0
900 cleach<-0.0
tcleach<-0.0
#####
#
## Functions of the CENTURY #####
905 #
#####
## function (calpet ) #####

```

```

## potential evapotranspiration
calpet<-function()
910 {
    # Linacre(1977) from CENTURY source
    #As in the CENTURY
    elev<-0.0
    ave<-matrix(0,nrow=12,ncol=1)
915  ave[1]<-(atempmax[1]+atempmin[1])/2.0
    highest<-ave[1]
    lowest<-ave[1]
    for(k in 2:12)
    {
920  ave[k]<-(atempmax[k]+atempmin[k])/2.0
        if(ave[k]>highest)
        {
            highest<-ave[k]
        }
925  if(ave[k]<lowest)
        {
            lowest<-ave[k]
        }
    }
930  if(lowest< (-10.0))
    {
        lowest<- (-10.0)
    }
    ra<-abs(highest-lowest)
935  for(k in 1:12)
    {
        if(atempmin[k]<(-10.0))
        {
            tr<-atempmax[k]-(-10.0)
940  }
        else
        {
            tr<-atempmax[k]-atempmin[k]
        }
    }
}

```

```

945     t<-tr/2.0+atempmin[k]
        tm<-t+0.006*elev
        td<-0.0023*elev+0.37*t+0.53*tr+0.35*ra-10.9
        e<-((700.0*tm/(100.0-abs(sitlat)))+15.0*td)/(80.0-t)
        monpet<-(e*30.0)/10.0
950     if(monpet < 0.5)
        {
            pet[k]<<-0.5*fwloss4
        }
        else
955     {
            pet[k]<<-monpet*fwloss4
        }
    }
}
960 ## function (calstemp) #####
## soil temperature
calstemp<-function(month)
{
    #For Forest only (e.g. no savana)
965     stdead<-0.0
        bio<-(pools.bfol)*2.5+stdead+(xsrfstr+xsrfmet)*2.0*elitst
        if(bio>PmxBio)
        {
            bio<-PmxBio
970     }
    else {
        bio<-bio
    }
    stempmax <<-atempmax[month]+
975         (25.4/(1+18.0*exp(-0.20*atempmax[month]))) *
            (exp(PmxTmp*bio)-0.13)
    stempmin <<-atempmin[month]+PmnTmp*(bio)-1.78
    stempave <<-(stempmax+stempmin)/2.0
}
980 ## function (calfc_wtpt) #####
## field capacity and wilting point

```

```

calfc_wtpt<-function()
{
  #From CENTURY source
985  #swflag lets the model user choose between using actual data
  #for awilt and afiel or equations from Gupta and Larson (1979)
  #or Rawls et al (1982).
  #swflag=0
  #Use actual data
990  #swflag=1
  #Use G&L for both awilt (-15 bar) and afiel (-0.33 bar)
  #swflag=2
  #Use G&L for both awilt (-15 bar) and afiel (-0.10 bar)
  #swflag=3
995  #Use Rawls for both awilt (-15 bar) and afiel (-0.33 bar)
  #swflag=4
  #Use Rawls for both awilt (-15 bar) and afiel (-0.10 bar)
  #swflag=5
  #Use Rawls for afiel (-0.33 bar) and actual data for awilt
1000 #swflag=6
  #Use Rawls for afiel (-0.10 bar) and actual data for awilt
  fcsa<-c( 0.3075,    0.5018,    -0.20,    -0.30,    -0.19,    0.31)
  fcsi<-c( 0.5886,    0.8548,     0.0,     0.0,     0.0,     0.0)
  fccl<-c( 0.8039,    0.8833,     0.36,     0.23,     0.0,     0.0)
1005 fcom<-c( 2.208E-03, 4.966E-03, 0.0299,  0.0317, 0.0210, 0.0260)
  fcbd<-c(-0.1434,   -0.2423,    0.0,     0.0,     0.0,     0.0)
  fcwp<-c( 0.0,      0.0,      0.0,     0.0,     0.72,    0.41)
  fcin<-c( 0.0,      0.0,      0.2576,  0.4118, 0.2391, 0.4103)
  wpsa<-c(-0.0059,   -0.0059,    0.0,     0.0,     0.0,     0.0)
1010 wpsi<-c( 0.1142,    0.1142,    0.0,     0.0,     0.0,     0.0)
  wpcl<-c( 0.5766,    0.5766,    0.50,    0.50,    0.0,     0.0)
  wpom<-c( 2.228E-03, 2.228E-03, 0.0158,  0.0158, 0.0,     0.0)
  wpbd<-c( 0.02671,   0.02671,   0.0,     0.0,     0.0,     0.0)
  wpwp<-c( 0.0,      0.0,      0.0,     0.0,     1.0,    1.0)
1015 wpin<-c( 0.0,      0.0,      0.0260,  0.0260, 0.0,     0.0)
  #print(somsc)
  ompc <- somsc*1.724/(10000*bd*Edepth)
  swflag<-flag_fc_wtpt
}

```

```

for(lyr in 1:nlayer)
1020 {
    #Please note:
    #In the original CENTURY model,
    #somsc was not calculated before the call of the prelim.f,
    #so afiel is calculated using somsc=ompc=0.
1025 #This is a bug of the original CENTURY model
    if(BFix==0)
    {
        ompc<-0.0
    }
1030 afiel[lyr] <<- fcsa[swflag]*sand + fcsi[swflag]*silt +
                fccl[swflag]*clay + fcom[swflag]*ompc +
                fcdb[swflag]*bd + fcwp[swflag]*awilt[lyr] +
                fcin[swflag]
    awilt[lyr] <<- wpsa[swflag]*sand + wpsi[swflag]*silt+
1035 wpcl[swflag]*clay + wpom[swflag]*ompc +
                wpbd[swflag]*bd + wpwp[swflag]*awilt[lyr] +
                wpin[swflag]
    ompc<-ompc*0.85
}
1040 }
## function (calwater) #####
## soil water content
calwater<-function(month)
{
1045 #Initialize
    add<-0.0
    amelt<-0.0
    asimx<-0.0
    avh2o[1]<<-0.0
1050 avh2o[2]<<-0.0
    avh2o[3]<<-0.0
    avap<-0.0
    evl<-0
    pevp<-0.0
1055 pttr<-0.0

```

```

    rwc1<-0.0
    tran<<-0.0
    trap<-0.01
    aabs<-0.0
1060  evsnow<-0.0
    evap<<-0.0
    petrem<-pet[month]
    awwt<-matrix(0.0,nrow=11,ncol=1)
    #CO2 effect was not included here
1065  co2val<-1.0
    irract<-0.0
    inputs<-prec[month]+irract
    winputs<-inputs
    atempave<<-(atempmax[month]+atempmin[month])/2.0
1070  aliv<-pools.bfol*2.5
    alit<-(xsrfstr+xsrfmet)*2.0
    adead<-0.0
    #*****
    #Snow
1075  #Snowfall
    if(atempave <= 0.0)
    {
        #snow <- snow + prec[month]
        snow <<- snow + inputs
1080  winputs<-0.0
    }
    # melt
    if(atempave >= Tmelt1)
    {
1085  melt <- Tmelt2 *(atempave -Tmelt1)
        if(melt>snow)
        {
            melt<-snow
        }
1090  snow <<-snow-melt
    ##.....
    if((atempave > 0.0) && (snow > 0.0))

```



```

    {
      snlq<<-snlq+inputs
1095    }
    snlq<<-snlq+melt
    if(snlq >= (0.05*snow))
    {
      add<-snlq -0.05*snow
1100    snlq<<-snlq-add
    }
  }
  if(snow > (0.0))
  {
1105    evsnow<-petrem*0.87
    snow1<-snow+snlq
    if(evsnow > snow1)
    {
      evsnow<-snow1
1110    }
    snow<<-snow-evsnow*(snow/snow1)
    snlq<<-snlq-evsnow*(snlq/snow1)
    evap<<-evap+evsnow
    petrem<-petrem-evsnow/0.87
1115    if(petrem < 0.0)
    {
      petrem<-0.0
    }
  }
1120  if(snow <= 0.0)
  {
    sd<-aliv+adead
    if(sd > 800.0)
    {
1125    sd<-800.0
    }
    if(alit > 400.0)
    {
      alit<-400.0

```

```

1130     }
        aint<-(0.0003 * alit +0.0006 *sd) *fwloss1
        aabs<-0.5*exp((-0.002*alit)-(0.004*sd))*fwloss2
        if((aabs+aint)*inputs<0.4*petrem)
        {
1135         evl<-(aabs+aint)*winputs
        }
        else
        {
            evl<-0.4*petrem
1140     }
        evap<<-evap+evl
        add<-add+winputs -evl
        trap<-petrem-evl
    }
1145 if(atempave < 2.0)
    {
        pttr<-0.0
    }
    else
1150 {
        pttr<-petrem *0.65 *(1.0 -exp(-0.020 *aliv)) *co2val
    }
    if(pttr <= trap){trap<-pttr}
    if(trap <= 0.0){trap<-0.01}
1155 ##.....
    #hpttr is not included
    pevp<-petrem -trap -evl
    if(evap<0.0){pevp<-0.0}
    if((trap-0.01) < add)
1160 {
        #print(add)
        tran<<- trap-0.01
    }
    else
1165 {
        tran<<- add
    }

```

```

    }
    trap<-trap-tran
    add<-add-tran
1170   strm<-0.0
        base<-0.0
        stream1<<-0.0
        for(j in 1:nlayer)
        {
1175     asmos[j]<<-asmos[j]+add
            afl<-adep[j]*afiel[j]
            if(asmos[j]>afl)
            {
1180     amov[j]<<-asmos[j]-afl
            asmos[j]<<-afl
            if(j == nlayer)
            {
                strm<-amov[j]*stormf
            }
1185     }
            else
            {
                amov[j]<<-0.0
            }
1190     add<-amov[j]
        }
        asmos[nlayer+1]<<-asmos[nlayer+1]+add-strm
        base<-asmos[nlayer+1]*basef
        asmos[nlayer+1]<<-asmos[nlayer+1]-base
1195     stream1<<-strm+base
        asimx<-asmos[1]
        rwcl<-0.0
        tot<-0.0
        tot2<-0.0
1200     for(j in 1:nlayer)
        {
            avw<-asmos[j]-awilt[j]*adep[j]
            if(avw < 0.0)

```

```

    {
1205     avw<-0.0
    }
    awwt[j]<-avw*awt1[j]
    tot<-tot+avw
    tot2<-tot2+awwt[j]
1210 }
    if(tot<trap)
    {
        trap<-tot
    }
1215 else
    {
        trap<-trap
    }
    if(tot2 > 0.0)
1220 {
        for(j in 1:nlayer)
        {
            avinj<-asmos[j]-awilt[j]*adep[j]
            if(avinj < 0.0)
1225 {
                avinj<-0.0
            }
            trl<-(trap*awwt[j])/tot2
            if(trl > avinj)
1230 {
                trl<-avinj
            }
            asmos[j]<<-asmos[j]-trl
            #if(year==5 && month==1){cat(asmos[j], trl,"\n")}
1235 avinj<-avinj-trl
            tran<<-tran+trl
            rwcf[j]<-(asmos[j]/adep[j]-awilt[j])/(afiel[j]-awilt[j])
            if(j<=nlaypg)
            {
1240     avh2o[1]<<-avh2o[1]+avinj
            }
        }
    }

```

```

    }
    avh2o[2]<<-avh2o[2]+avinj
    if(j <= (2))
    {
1245     avh2o[3]<<-avh2o[3]+avinj
    }
  }
}
fwlos<-0.25
1250 evmt<-(rwc[1]-fwlos)/(1.0-fwlos)
if(evmt <= (0.01))
{
  evmt<-0.01
}
1255 evlos<-evmt*pevp*aabs*0.10
avinj<-asmos[1]-awilt[1]*adep[1]
if(avinj < 0.0)
{
  avinj<-0.0
1260 }
if(evlos > avinj)
{
  evlos<-avinj
}
1265 asmos[1]<<-asmos[1]-evlos
evap<<-evap+evlos
avhsm<-(asmos[1]+rwc1*asimx)/(1.0+rwc1)
rwc[1]<<-(avhsm/adep[1]-awilt[1])/(afiel[1]-awilt[1])
avh2o[1]<<-avh2o[1]-evlos
1270 avh2o[2]<<-avh2o[2]-evlos
avh2o[3]<<-avh2o[3]-evlos
}
## function (caldefac) #####
## decomposition factor
1275 caldefac<-function()
{
  if(snow > 0.0)

```

```

    {
      stempave<-0.0
1280   }
      # Cal defac
      tfunc<-teff1+teff2*exp(teff3 * stempave)
      rprpet <<- (avh2o[3] + prec[month] ) / pet[month]
      /* ideo in fix.100 in Century control linear 1 or ratio 2 option
1285   /* this is ideo==2
      if(rprpet > 9.0 )
      {
        wfunc<<-1.0
      }
1290   else
      {
        wfunc<<-1.0/(1.0+30.0*exp(-8.5*rprpet))
      }
      #if(wfunc>1.0)
1295   #{
      #  wfunc<<-1.0
      #}
      defac<<-tfunc*wfunc
      ##
1300   # If you want to use the defac from the original CENTURY, then.
      # defac<<-centdefac[1+(year-1)*12+month,2]
      #*** Cal anerb
      anerb <<- 1.0
    }
1305 ## functions to Divide litter inputs#####
      ##
      ## function (calcenturyinput) #####
      ## litter inputs into each soil carbon pool:1
      calcenturyinput<-function()
1310 {
      if(flows.lfinfol>0.0)
      {
        #centurypartit(1, cnr_srflit)
        centurypartit(1, cnr_fol)
      }
    }

```

```

1315 }
      else
      {
        usrfstr <<-0.0
        usrfmet <<-0.0
1320    usrfstr_lig <<- 0.0
      }
      if(flows.lfinfir>0.0)
      {
        #centurypartit(2, cnr_bellit)
1325    centurypartit(2, cnr_fir)
      }
      else
      {
        ubelstr <<- 0.0
1330    ubelmet <<- 0.0
        ubelstr_lig <<- 0.0
      }
      uwood1 <<- flows.lfinbra
      uwood2 <<- flows.lfinste
1335    uwood3 <<- flows.lfincor
      uwood1_lig <<- flows.lfinbra * cbra_lig
      uwood2_lig <<- flows.lfinste * cste_lig
      uwood3_lig <<- flows.lfincor * ccor_lig
    }
1340 ## function (centurypartit) #####
      ## litter inputs into each soil carbon pool:2
      centurypartit<-function(p, cnr)
      {
        #translocation
1345    #forrtf
        if(p==1)
        {
          cpart<- flows.lfinfol
          epart<- flows.lfinfol*(1.0/cnr)*(1.0-forrtf)
1350    #amax1
          if(cpart/pabres > 1.0)

```

```

    {
      s<-cpart/pabres
    }
1355 else
    {
      s<-1.0
    }
    #damr11<-0.0
1360 dirabs<- damr11 * xNmineral * s
    if((epart+dirabs) <= 0.0)
    {
      rcetot<-0.0
    }
1365 else
    {
      rcetot<-cpart/(epart+dirabs)
    }
    if(rcetot < damrmn)
1370 {
      dirabs<-cpart/damrmn-epart
    }
    if(dirabs<0.0)
    {
1375     dirabs<-0.0
    }
    frlign<- cfol_lig
  }
  else if (p==2)
1380 {
    cpart<- flows.lfinfir
    #epart<- flows.lfinfir*(1.0/cnr)*(1.0-forrtf)
    epart<- flows.lfinfir*(1.0/cnr)
    #amax1
1385 if(cpart/pabres > 1.0)
    {
      s<-cpart/pabres
    }
  }

```



```

else
1390 {
    s<-1.0
}
dirabs<- damr21 * xNmineral * s
if((epart+dirabs) <= 0.0)
1395 {
    rcetot<-0.0
}
else
{
1400    rcetot<-cpart/(epart+dirabs)
}
if(rcetot < damrmn)
{
    dirabs<-cpart/damrmn-epart
1405 }
if(dirabs<0.0)
{
    dirabs<-0.0
}
1410 frlign<- cfir_lig
}
else
{
    printf("error")
1415 }
###
frn<- (epart + dirabs)/(cpart*2.5)
rlnres<-frlign/frn
frmet<- spl1 -spl2 *rlnres
1420 if(frlign > (1.0-frmet))
{
    frmet<- 1.0-frlign
}
if(frmet<0.20)
1425 {

```

```

    frmet<-0.20
  }
  caddm <- cpart*frmet
  if(caddm < 0.0)
1430 {
    caddm<-0.0
  }
  cadds <- cpart-caddm
  fligst <- frlign/(cadds/cpart)
1435 if(fligst > 1.0)
  {
    fligst <- 1.0
  }
  if(p==1)
1440 {
    usrfstr <<- flows.lfinfol *(1.0-frmet)
    usrfmet <<- flows.lfinfol *frmet
    usrfstr_lig <<- fligst
  }
1445 else if (p==2)
  {
    ubelstr <<- flows.lfinfir *(1.0-frmet)
    ubelmet <<- flows.lfinfir *frmet
    ubelstr_lig <<- fligst
1450 }
  }
#####
## functions (calcentury)
## to calculate soil carbon dynamics
1455 ## *****
calcentury<-function()
{
  uwood1<-flows.lfinbra
  uwood2<-flows.lfinste
1460 uwood3<-flows.lfincor
  #*****
  # ** Dead branch = Wood 1

```

```

#strlig=(xwood1*wood1strlig+uwood1_lig)/(xwood1+uwood1)
#wood1strlig= strlig
1465 strlig <-cbra_lig
if(xwood1>0.000001)
{
  tcflow <- xwood1*defac*kwood1*exp(-pligst1*strlig)*DT
  if(tcflow>xwood1)
1470 {
    tcflow<-xwood1
  }
}
else
1475 {
  tcflow<-0.0
}
tsom2_fwood1 <- tcflow * strlig
#*Respiration associated with decomposition to som2
1480 co2los <- tsom2_fwood1 * RSPLIG
CO2out <<- CO2out+co2los
#*Net C flow to SOM2
tsom2_fwood1 <- tsom2_fwood1 - co2los
tsom1_fwood1 <- tcflow - tsom2_fwood1 - co2los
1485 #*Respiration associated with decomposition to som1
co2los <- tsom1_fwood1 * PS1CO21
tsom1_fwood1 <- tsom1_fwood1 -co2los
CO2out<<-CO2out+co2los
#*****
1490 xwood1_new <- xwood1 + uwood1 - tcflow
#*****
# ** Dead Stem = Wood 2
#strlig=(xwood2*wood2strlig+uwood2_lig)/(xwood2+uwood2)
#wood2strlig= strlig
1495 strlig<-cste_lig
if(xwood2>0.000001)
{
  tcflow<- xwood2*defac*kwood2*exp(-pligst1*strlig)*DT
  if(tcflow>xwood2)

```

```

1500     {
           tcflow<-xwood2
         }
       }
     else
1505     {
           tcflow<-0.0
         }
       tsom2_fwood2 <- tcflow * strlig
       /*Respiration associated with decomposition to som2
1510     co2los <- tsom2_fwood2 * RSPLIG
       CO2out<<-CO2out+co2los
       /*Net C flow to SOM2
       tsom2_fwood2 <- tsom2_fwood2 - co2los
       tsom1_fwood2 <- tcflow - tsom2_fwood2 - co2los
1515     /*Respiration associated with decomposition to som1
       co2los <- tsom1_fwood2 * PS1CO21
       tsom1_fwood2 <- tsom1_fwood2 -co2los
       CO2out<<-CO2out+co2los
       #*****
1520     xwood2_new <- xwood2 + uwood2 - tcflow
       #*****
       # ** Dead Coarse root = Wood 3
       #strlig=(xwood3*wood3strlig+uwood3_lig)/(xwood3+uwood3)
       #wood3strlig= strlig
1525     strlig<-ccor_lig
       if(xwood3>0.000001)
         {
           tcflow<- xwood3*defac*kwood3*exp(-pligst2*strlig)*anerb*DT
           if(tcflow>xwood3)
1530           {
               tcflow<-xwood3
             }
         }
       }
     else
1535     {
           tcflow<-0.0
         }
       }

```

```

}
tsom2_fwood3 <- tcflow * strlig
#*Respiration associated with decomposition to som2
1540 co2los <- tsom2_fwood3 * RSPLIG
CO2out<<-CO2out+co2los
#*Net C flow to SOM2
tsom2_fwood3 <- tsom2_fwood3 - co2los
tsom1_fwood3 <- tcflow - tsom2_fwood3 - co2los
1545 #*Respiration associated with decomposition to som1
co2los <- tsom1_fwood3 * PS1CO21
tsom1_fwood3 <- tsom1_fwood3 -co2los
CO2out<<-CO2out+co2los
#*****
1550 xwood3_new <- xwood3 + uwood3 - tcflow
#*****
# ** surface structural
#strlig=(pools.xlig1_fol + pools.xlig1_bra +
#      pools.xlig1_ste)/(pools.talit)
1555 #srfstrlig = xsrfstr*srfstrlig/xsrfstr
#strlig=(xsrfstr*srfstrlig + usrfstr_lig)/(xsrfstr+usrfstr)
strlig<-(xsrfstr*srfstrlig + usrfstr_lig*usrfstr)/
      (xsrfstr + usrfstr)
srfstrlig <<- strlig
1560 if(xsrfstr>0.000001)
{
  if(xsrfstr>strmax1)
  {
    mass<-strmax1
1565   }
  else
  {
    mass<-xsrfstr
  }
1570 tcflow <-mass*defac*Asrfstr*exp(-pligst1*strlig)*DT
if(tcflow>xsrfstr)
{
  tcflow<-xsrfstr

```

```

    }
1575 }
    else
    {
        tcflow<-0.0
    }
1580 tsom2_fsrfstr <- tcflow * strlig
    #*Respiration associated with decomposition to som2
    co2los <- tsom2_fsrfstr * RSPLIG
    CO2out <<- CO2out+co2los
    #*Net C flow to SOM2
1585 tsom2_fsrfstr <- tsom2_fsrfstr - co2los
    tsom1_fsrfstr <- tcflow - tsom2_fsrfstr - co2los
    #*Respiration associated with decomposition to som1
    co2los <- tsom1_fsrfstr * PS1CO21
    tsom1_fsrfstr <- tsom1_fsrfstr -co2los
1590 CO2out <<- CO2out+co2los
    #*****
    xsrfstr_new <- xsrfstr + usrfstr - tcflow
    #*****
    # ** soil structural
1595 #strlig=(pools.xlig1_fir+pools.xlig1_cor)/(pools.tblit)
    #belstrlig = xbelstr*belstrlig/xbelstr
    strlig<-(xbelstr*belstrlig + ubelstr_lig*ubelstr)/
        (xbelstr + ubelstr)
    belstrlig<<-strlig
1600 if(xbelstr>0.000001)
    {
        if(xbelstr>strmax2)
        {
            mass<-strmax2
1605 }
        else
        {
            mass<-xbelstr
        }
1610 tcflow<-mass*defac*Abelstr*exp(-pligst2*strlig)*anerb*DT

```

```

        if(tcflow>xbelstr)
        {
            tcflow<-xbelstr
        }
1615 }
else
{
    tcflow<-0.0
}
1620 tsom2_fbelstr <- tcflow * strlig
    #*Respiration associated with decomposition to som2
    co2los <- tsom2_fbelstr * RSPLIG
    CO2out <<- CO2out+co2los
    #*Net C flow to SOM2
1625 tsom2_fbelstr <- tsom2_fbelstr - co2los
    tsom1_fbelstr <- tcflow - tsom2_fbelstr - co2los
    #*Respiration associated with decomposition to som1
    co2los <- tsom1_fbelstr * PS1CO22
    tsom1_fbelstr <- tsom1_fbelstr -co2los
1630 CO2out<<-CO2out+co2los
    #*****
    xbelstr_new <- xbelstr + ubelstr - tcflow
    #*****
    # ** surface metab
1635 if(xsrmet>0.000001)
    {
        tcflow<-xsrmet * defac * Asrmet * DT
        if(tcflow>xsrmet)
        {
1640         tcflow<-xsrmet
        }
    }
else
{
1645     tcflow<-0.0
}
co2los<-tcflow*Psrmet

```

```

tsom1_fsrfmet <- tcflow-co2los
CO2out <<- CO2out+co2los
1650 xsrfmet_new <- xsrfmet +usrfmet -tcflow
#*****
# ** belowground metab
if(xbelmet>0.000001)
{
1655   tcflow<-xbelmet * defac * Abelmet* anerb * DT
   if(tcflow>xbelmet)
   {
     tcflow<-xbelmet
   }
1660 }
else
{
   tcflow<-0.0
}
1665 co2los<-tcflow*Pbelmet
tsom1_fbelmet<-tcflow-co2los
CO2out<<-CO2out+co2los
xbelmet_new<-xbelmet + ubelmet -tcflow
#*****
1670 #**** surface microbe
if(xsrfmic>0.000001)
{
   tcflow <- xsrfmic * defac * Asrfmic *DT
   if(tcflow>xsrfmic)
1675   {
     tcflow <- xsrfmic
   }
}
else
1680 {
   tcflow<-0.0
}
co2los<-tcflow*Psrfmic
tsom2_fsrfmic<-tcflow-co2los

```



```

1685 CO2out<<-CO2out+co2los
      xsrfmic_new <- xsrfmic + tsom1_fsrfstr + tsom1_fsrfmet +
                    tsom1_fwood1 + tsom1_fwood2 -tcflow
      #xsrfmic_new= -tcflow + xsrfmic + tsom1_fsrfstr+tsom1_fsrfmet
      #xsrfmic_new= -tcflow + xsrfmic + tsom1_fsrfmet
1690 #*****
      #**** active
      eftext <- Peftxa + Peftxb * sand
      if(xactv>0.000001)
      {
1695   tcflow<-xactv* defac * eftext * kactv * anerb *DT
       if(tcflow>xactv)
       {
         tcflow<-xactv
       }
1700 }
      else
      {
        tcflow<-0.0
      }
1705 co2los<-tcflow*Pactv
      #*cfsfs2=tcflow-co2los
      #*tcflow=tcflow-co2los
      CO2out<<-CO2out+co2los
      fpsls3 <- psls31 + psls32 * clay
1710 tsom3_factv<-tcflow * fpsls3
      #leaching
      if(amov[2]>0.0)
      {
        orglch<-omlech[1]+omlech[2]*sand
1715 t<-1.0-(omlech[3]-amov[2])/omlech[3]
        if(t>1.0)
        {
          linten<-1.0
        }
1720 else
      {

```

```

        linten<-t
    }
    cleach<<-tcflow * orglch * linten
1725 }
else
{
    cleach<<-0.0
}
1730 tcleach<<-tcleach+cleach
tsom2_factv<-tcflow -co2los -tsom3_factv -cleach
#* Updated at the end.
#xactv_new = xactv + tsom1_fbelstr +tsom1_fbelmet +
#          tsom1_fwood3 +tsom1_fslow +tsom1_fpass -tcflow
1735 xactv_new <- xactv + tsom1_fbelstr +tsom1_fbelmet +
          tsom1_fwood3 -tcflow
#*****
#**** Slow
if(xslow>0.000001)
1740 {
    tcflow<-xslow *defac * kslow * anerb *DT
    if(tcflow>xslow)
    {
        tcflow<-xslow
1745 }
    }
else
{
    tcflow<-0.0
1750 }
co2los<-tcflow*Pslow
#*cfsfs2=tcflow-co2los
#*tsom3_fslow=tcflow-co2los
CO2out<<-CO2out+co2los
1755 xslow_new <- xslow + tsom2_fsrfstr +tsom2_fsrfmic +
          tsom2_fbelstr +tsom2_factv + tsom2_fwood1 +
          tsom2_fwood2 + tsom2_fwood3 -tcflow
fps2s3 <- ps2s31 + ps2s32 * clay

```

```

tsom3_fslow<-tcflow * fps2s3
1760 tsom1_fslow<-tcflow -co2los -tsom3_fslow
#*****
#**** Passive
if(xpass>0.000001)
{
1765   tcflow<-xpass *defac  * kpass* anerb *DT
   if(tcflow>xpass)
   {
     tcflow<-xpass
   }
1770 }
else
{
  tcflow<-0.0
}
1775 co2los<-tcflow*Ppass
#*cfsfs2=tcflow-co2los
tsom1_fpass<-tcflow-co2los
CO2out<<-CO2out+co2los
xpass_new <- xpass + tsom3_factv +tsom3_fslow -tcflow
1780 #*****
#***** Active new
#xactv_new = xactv + tsom1_fpass
#xactv_new = xactv + tsom1_fslow
xactv_new <- xactv_new + tsom1_fslow + tsom1_fpass
1785 #*****
#***** UPDATE
xsrfstr <<- xsrfstr_new
xsrfmet <<- xsrfmet_new
xsrfmic <<- xsrfmic_new
1790 xbelstr <<- xbelstr_new
xbelmet <<- xbelmet_new
xactv <<- xactv_new
xslow <<- xslow_new
xpass <<- xpass_new
1795 xwood1 <<- xwood1_new

```

```

xwood2 <<- xwood2_new
xwood3 <<- xwood3_new
#*****
somsc <<- xactv + xslow + xpass
1800 talit <<- xsrfstr + xsrfmet + xsrfmic
tblit <<- xbelstr + xbelmet
somtC <<-xactv + xslow + xpass + xbelstr + xbelmet
tawood <<- xwood1 + xwood2
tbwood <<- xwood3
1805 }
#end of functions
## calculate field capacity, wilting point #####
awilt
afiel
1810 if(flag_fc_wtpt>0.0)
{
  somsc <- xactv + xslow + xpass
  calfc_wtpt()
}
1815 awilt
afiel
## Initialize soil water condition #####
# essential to calculate deep asmos correctly
pet
1820 calpet()
pet
for(month in 1:12)
{
  calwater(month)
1825 }
obj.s <- ls()
#obj.s
#####
## MAIN CENTURY SIMULATION #####
1830 carbon.out <- NULL
for(s in 1:1){
  id<-s

```

```

soil.carbon.year.out <-NULL
for(year in TSTART:TEND){
1835   CO2out<-0.0
       calpet()
       #month loop
       for(month in 1:12){
           #month=1
1840   tcleach<-0.0
       DT<-1.0/(12.0*CYCL)
       ##.....
       #Litterfall SITE SPECIFIC data
       flows.lfinfol<-litter.in [1,2]*leafdr[month]*(1.0/CYCL)
1845   flows.lfinbra<-litter.in [1,3]*(1.0/(12*CYCL))
       flows.lfinste<-litter.in [1,4]*(1.0/(12*CYCL))
       flows.lfinfir<-litter.in [1,5]*(1.0/(12*CYCL))
       flows.lfincor<-litter.in [1,6]*(1.0/(12*CYCL))
       talit <-xsrfstr + xsrfmet +xsrfmic
1850   tblit <- xbelstr + xbelmet
       tawood <- xwood1 + xwood2
       tbwood <- xwood3
       somsc <- xactv + xslow + xpass
       somtc <- xactv + xslow + xpass + xbelstr + xbelmet
1855   ##.....
       calstemp(month)
       calwater(month)
       if(snow>0.0)
       {
1860   stempmax <-0.0
       stempmin <-0.0
       stempave <-0.0
       }
       ##.....
1865   caldefac()
       calcenturyinput()
       # CENTURY CARBON FUNCTION SIMULATIONS
       # updated 4 times per month (CYCL=4)
       for(i in 1:CYCL)

```

```

1870      {
          calcentury()
        }
#end of centurycal CYCL loop
      }
1875 #end of month loop ()
      ##.....
      ## site specific output of CENTURY carbon
      if(year==year) #TEND)
      {
1880      soil.carbon0 <- data.frame(id,year, month,
                                   xsrfstr, xsrfmet,
                                   xsrfmic, xbelstr, xbelmet,
                                   xactv, xslow, xpass, somsc,
                                   xwood1, xwood2, xwood3,
1885      CO2out, somtc)
      }

      soil.carbon.year.out <- rbind(soil.carbon.year.out,
                                   soil.carbon0)
1890  }
#end of year loop
      carbon.out <-rbind(carbon.out,soil.carbon.year.out )
      }
#end of site.group for loop
1895 options(digit=12)
century.out <-carbon.out[,c("id","year","month",
                           "CO2out","somsc",
                           "xbelstr","xbelmet",
                           "xactv","xslow","xpass",
1900      "somtc")]
#convert gC.m-2 to tC.ha-1 by 1/1e6*1e4
century.out[,4:11]<-century.out[,4:11]/100
##.....
#plot components of soil carbon stock
1905 #somtc <- xactv + xslow + xpass + xbelstr + xbelmet
#figure

```

```

par(mfrow=c(1,1), mar=c(5,5,1,1))
plot(century.out$year,century.out$somtc,
      log="y", ylim=c(0.3,round(max(century.out$somtc),1)+50),
1910   ylab="CENTURY soil carbon pools (tC/ha)",
      xlab="year")
lines(century.out$year,century.out$somtc,
      lwd=2)
lines(century.out$year,century.out$xactv,
1915   col="blue", lwd=2)
lines(century.out$year,century.out$xslow,
      col="red", lwd=2)
lines(century.out$year,century.out$ypass,
      col="orange", lwd=2)
1920 lines(century.out$year,century.out$xbelstr,
      col="grey", lwd=2)
lines(century.out$year,century.out$xbelmet,
      col="magenta", lwd=2)
legend("bottomright",
1925   c("total","active","slow","passive",
        "bg.structural","bg.metabolic"),
      col=c("black","blue","red","orange","grey","magenta"),
      pch=c(1,NA,NA,NA,NA,NA),
      lwd=2, lty=1, border="white",bg="white")
1930

```

**Table S1.** Statistical characteristics (mean, standard error) of basic variables for groups of soils derived by recursive partitioning including soil variables (see Fig. 2a), compared with interpretation of carbon, moisture, and fertility of groups.

N of soil samples in groups		959	909	136	335	182	296	180	8	142	83
Total SOC (tC ha <sup>-1</sup> )	Mean	65.1	81.8	130.2	86.2	126.4	103.9	136.8	268.6	143.7	203.1
	SE	1	1.3	5.1	2.6	4.8	2.8	4.2	23.7	6.7	9.8
SOC mineral (tC ha <sup>-1</sup> )	Mean	45.4	56.4	86.9	68.5	98.4	73.2	92.4	230.6	108.8	153.3
	SE	1.6	2.1	8.2	4.9	9.1	4.5	7	33	12.8	21.6
C/N	Mean	13.4	18.6	15.4	10.7	8.1	23.8	21.7	23.1	23.3	32.6
	SE	0.3	0.4	0.8	0.2	0.2	0.5	0.5	1.9	0.6	2
H100 (m)	Mean	20.8	24.3	24.7	25.4	26.6	22.2	29.9	31	23.6	24
	SE	0.2	0.2	0.5	0.4	0.4	0.3	0.3	0.5	0.5	0.6
Total Litter (tC ha <sup>-1</sup> )	Mean	2.7	3.3	3.4	3.5	3.6	3.1	4	3.8	3.2	3.5
	SE	0	0	0.1	0.1	0.1	0	0.1	0.2	0.1	0.1
Temperature air (C)	Mean	3.3	4.8	5.1	5.1	5.8	4.5	7.3	7.3	5.3	6.4
	SE	0.1	0.1	0.2	0.1	0.1	0.1	0.1	0.2	0.2	0.2
Long-term moisture (%)	Mean	20.2	22.4	26	23.6	26.2	22.9	22.9	21.3	21.8	25.7
	SE	0.2	0.2	0.6	0.3	0.5	0.4	0.4	1.3	0.5	0.7
Precipitation (mm y-1)	Mean	698.8	712.9	697.1	644.1	630.3	693	817.2	1173.2	687.9	619.4
	SE	5.7	7	18.8	10.4	12.5	10.8	21.2	162	18.1	27
CEC (mmol <sub>c</sub> kg <sup>-1</sup> )	Mean	4.7	12.1	11.6	49.2	91.5	24	23.5	24.5	59.7	98.7
	SE	0.1	0.1	0.2	2.6	5.1	0.3	0.3	1.7	3.2	7.6
pH	Mean	5.2	5.1	5.1	5.5	5.6	5	4.8	4.6	4.9	6
	SE	0	0	0	0	0.1	0	0	0.1	0.1	0.2
Clay content (%)	Mean	0.8	1.1	4.2	5.9	21.5	1.7	2.4	2.5	2.4	8
	SE	0.1	0.1	0.5	0.5	1	0.3	0.3	1.3	0.4	1.2
Silt content (%)	Mean	15.1	14.5	29	27.2	57.8	16.4	17.9	18.8	17.1	32.5
	SE	0.3	0.3	1.5	1.2	1.4	0.7	0.7	3.2	1	3
Carbon		low	medium	high	medium	high	medium	high	extra	high	extra
Moisture		dry-fresh	fresh	moist-fresh	fresh	moist-fresh	fresh	fresh	fresh	fresh	moist-fresh
Fertility		low	medium	medium	medium	high	low	high	high	medium	medium
Soil group		1	2	3	4	5	6	7	8	9	10

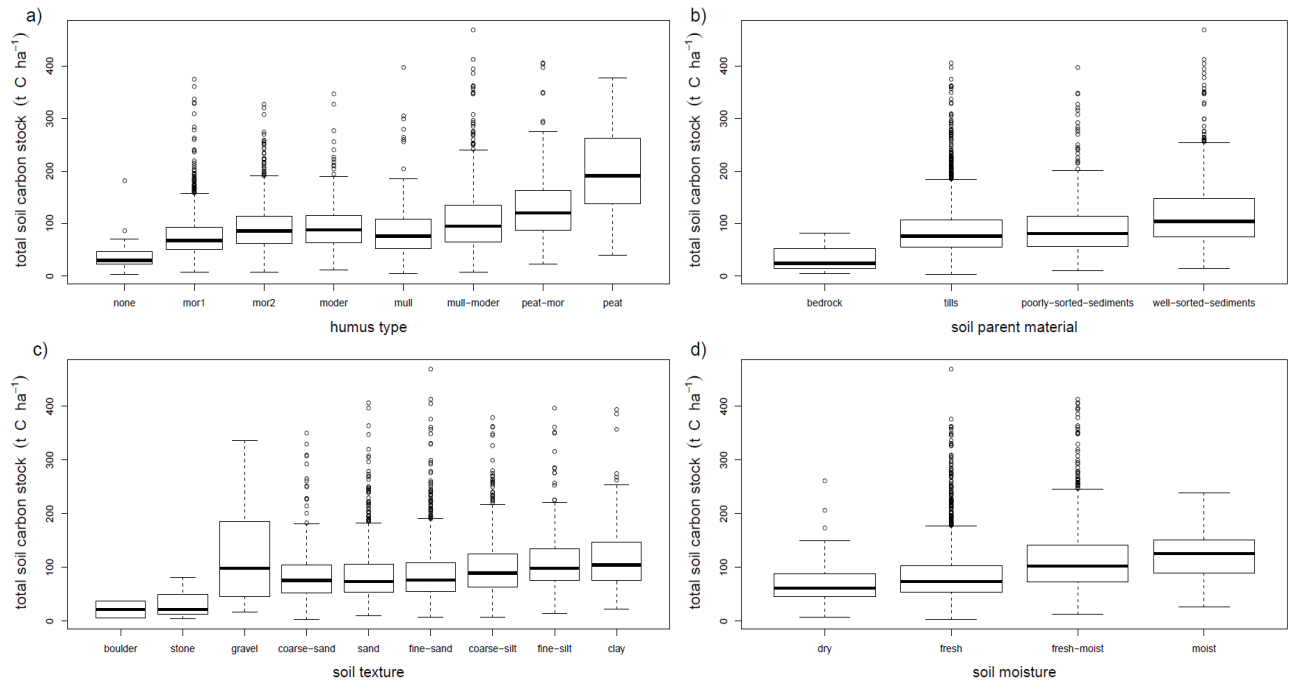


**Table S2.** Statistical characteristics (mean, standard error) of basic environmental and soil variables for regression tree of data groups classified by recursive partitioning with data excluding soil variables (see Fig. 3).

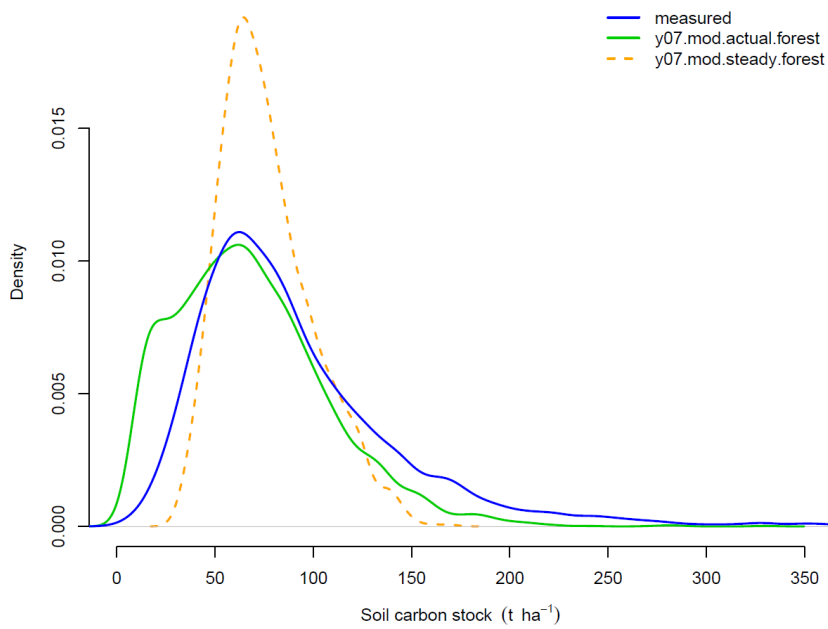
Number of samples		735.0	932.0	796.0	711.0	56.0
Total soil carbon (tC ha <sup>-1</sup> )	Mean	67.1	85.4	96.5	120.1	179.0
	SE	1.4	1.5	2.0	2.4	12.5
C/N	Mean	15.0	15.8	16.8	18.1	35.2
	SE	0.3	0.3	0.4	0.4	3.2
H100 (m)	Mean	19.0	19.7	27.0	30.1	18.7
	SE	0.1	0.1	0.1	0.2	0.3
Total Litter (tC ha <sup>-1</sup> )	Mean	2.3	2.8	3.6	4.0	3.0
	SE	0.0	0.0	0.0	0.0	0.1
Soil water content (%)	Mean	20.2	24.2	22.0	22.6	22.3
	SE	0.2	0.2	0.2	0.2	1.0
Temperature air (C)	Mean	2.4	2.7	6.4	7.2	7.2
	SE	0.1	0.1	0.0	0.0	0.0
pH	Mean	5.2	5.2	5.3	4.9	7.2
	SE	0.0	0.0	0.0	0.0	0.1
Sand content (%)	Mean	52.7	51.5	47.2	55.1	36.2
	SE	0.5	0.5	0.8	0.6	4.0
Clay content (%)	Mean	0.8	2.6	6.1	3.0	5.8
	SE	0.1	0.2	0.4	0.2	1.1
Silt content (%)	Mean	14.6	19.2	25.3	20.0	29.5
	SE	0.3	0.5	0.8	0.5	4.1
Group acronym		low-C cold.pine	median-C cold.other	median-C warm.rainy low-N	hig-C warm.rainy high-N	extra-C warm.dry

**Table S3.** Species and classes of ground vegetation grouped into functional types (1-dwarfshrubs, 2-herbs, 3-grasses, 4-mosses and 5-lichens).

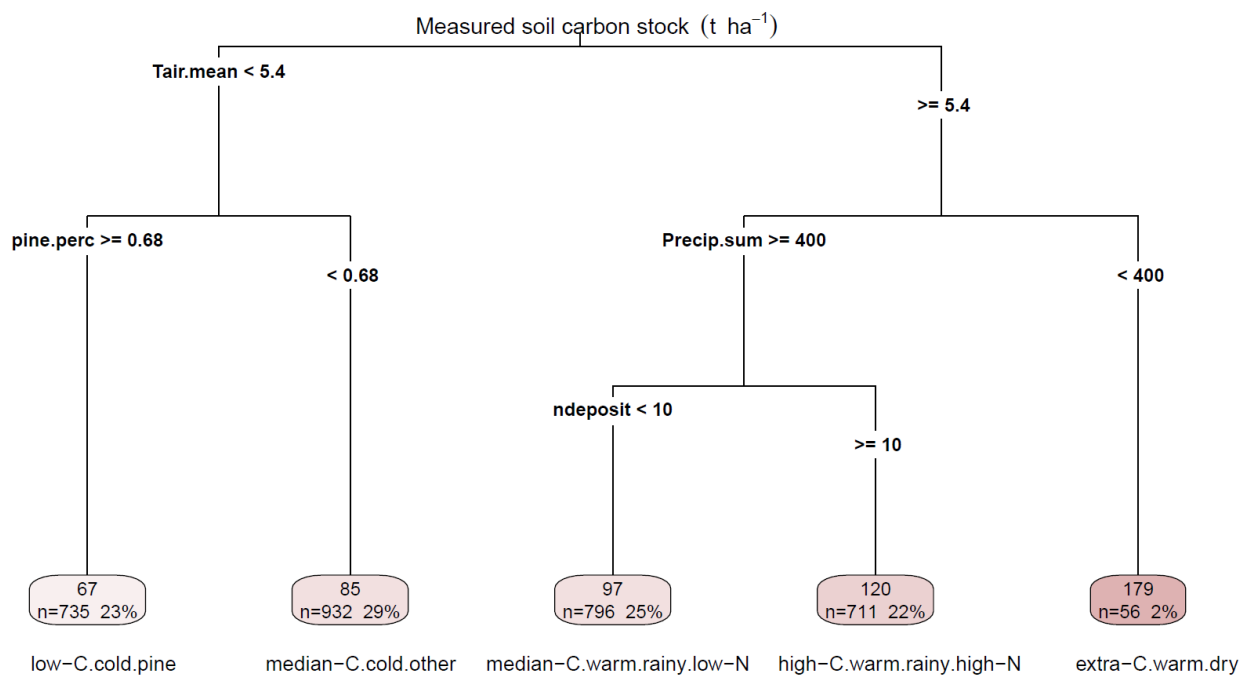
Functional type	Ground vegetation
1	<i>Vaccinium myrtillus</i> , <i>Vaccinium vitis-idaea</i> , <i>Arctostaphylos uva-ursi</i> , <i>Empetrum nigrum</i> ssp., <i>Calluna vulgaris</i> , <i>Erica tetralix</i> , <i>Vaccinium uliginosum</i> , <i>Rhododendron tomentosum</i> , <i>Andromeda polifolia</i> , <i>Vaccinium oxycoccus/microcarpum</i> , Other field layer plants
2	<i>Gymnocarpium dryopteris</i> , <i>Oxalis acetosella</i> , <i>Anemone nemorosa</i> , <i>Maianthemum bifolium</i> , <i>Chamaenerion angustifolium</i> , <i>Anthriscus sylvestris</i> , <i>Melampyrum pratense/sylvaticum</i> , <i>Equisetum sylvaticum</i> , <i>Menyanthes trifoliata</i> , <i>Rubus chamaemorus</i> , <i>Phegopteris connectilis</i> , <i>Hepatica nobilis</i> , <i>Geum rivale</i> , <i>Urtica dioica</i> , <i>Rumex acetosa</i> , <i>Stellaria nemorum</i> , <i>Stellaria holostea</i> , <i>Silene dioica</i> , <i>Aconitum lycoctonum</i> subsp. <i>septentrionale</i> , <i>Actaea erythrocarpa</i> , <i>Trollius europaeus</i> , <i>Cardamine bulbifera</i> , <i>Filipendula ulmaria</i> , <i>Mercurialis perennis</i> , <i>Sanicula europaea</i> , <i>Aegopodium podagraria</i> , <i>Librar</i> , <i>Galium odoratum</i> , <i>Lamium galeobdolon</i> , <i>Stachys sylvatica</i> , <i>Cirsium palustre</i> , <i>Cirsium heterophyllum</i> , <i>Lactuca alpina</i> , <i>Lactuca muralis</i> , <i>Crepis paludosa</i> , <i>Paris quadrifolia</i> , <i>Neottia ovata</i> , <i>Geranium sylvaticum</i> , <i>Rubus idaeus</i> , Other large grown ferns
3	Broad-leaved grass, Narrow-leaved grass, <i>Carex globularis</i> , Other sedges
4	<i>Pteridium aquilinum</i> , <i>Lycopodiaceae</i> , <i>Spagnum</i> spp, <i>Polytrichum commune</i> , <i>Pleurozium schreberi</i> , <i>Hylocomium splendens</i> , Other bryophytes
5	<i>Cladonia</i> , <i>Stereocaulon</i> spp, <i>Cladina</i> spp, <i>Cladonia</i> and <i>Cladina</i> , Other lichens



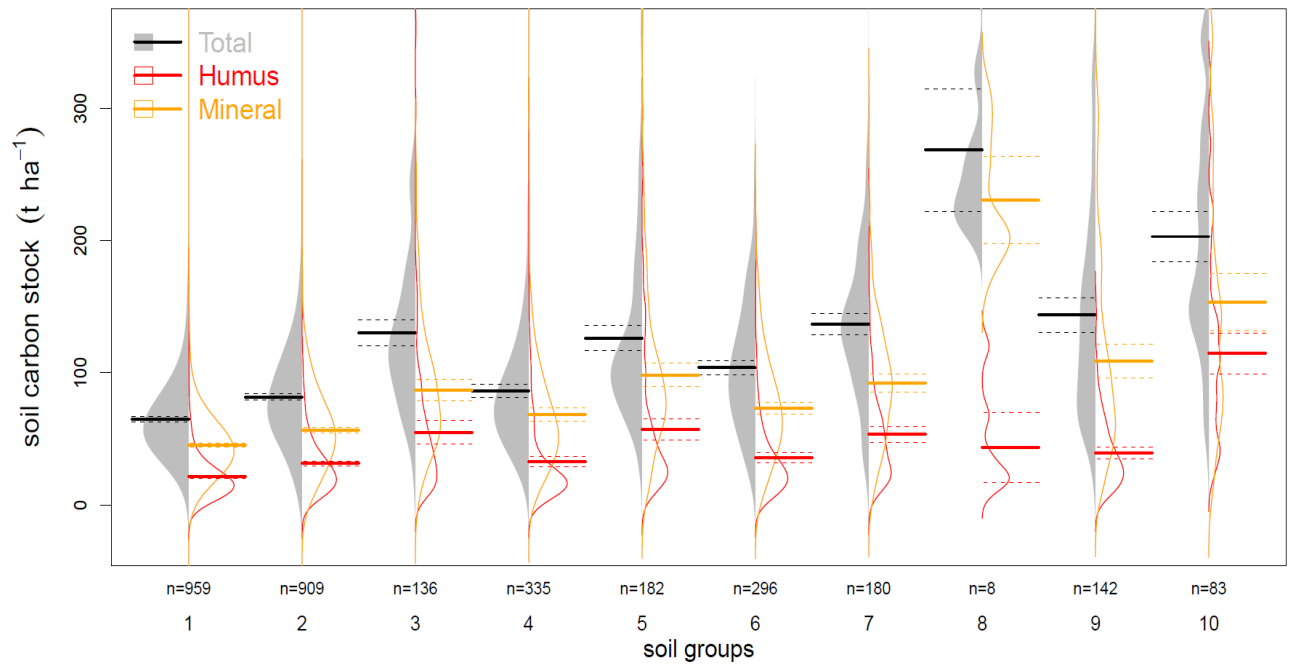
**Figure S1.** Boxplot main levels (minimum, 1st quartile, median, 3rd quartile, maximum, and dots for outliers) of the total soil carbon stock (tC ha<sup>-1</sup>) for SFSI categorical data on a) humus type, b) soil parent material, c) soil texture, and d) soil moisture.



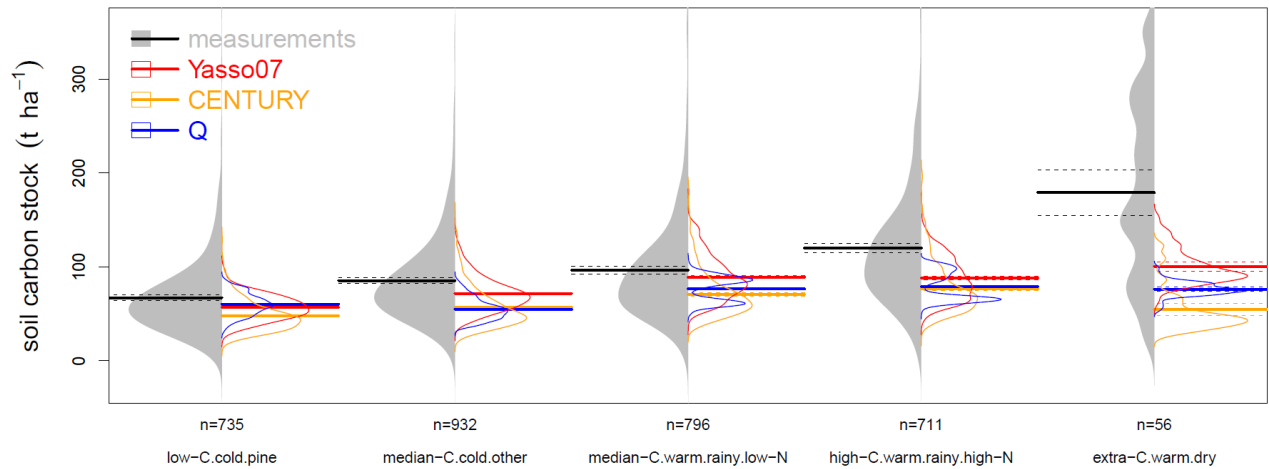
**Figure S2.** Density function of soil carbon stock measurements (measured) and the simulated soil carbon by the soil carbon model Yasso07 run with actual state forest and litter inputs (y07.mod.actual.forest) and with the steady state forest and litter inputs (y07.mod.steady.forest).



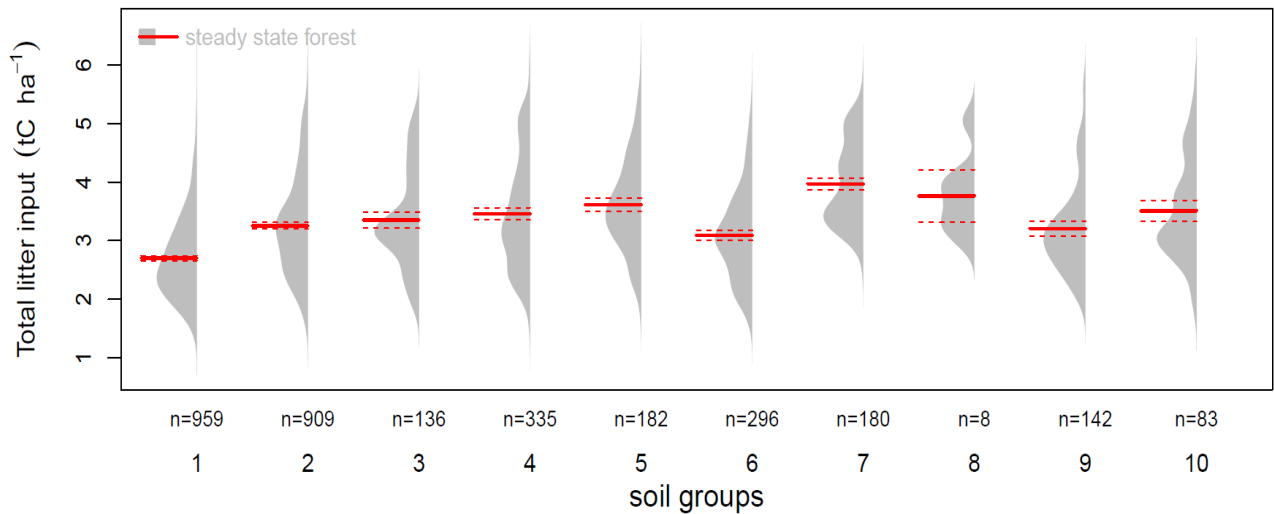
**Figure S3.** Classification/regression tree for the measured soil carbon stock ( $\text{tC ha}^{-1}$ ) and site environmental characteristics excluding soil physicochemical properties; the annual air temperature ( $T_{\text{air.mean}}$ ,  $^{\circ}\text{C}$ ), the fraction of pine trees of the total canopy (pine.perc), the annual precipitation sum (Precip.sum, mm), and the nitrogen deposition (ndeposit,  $\text{kgN ha}^{-1} \text{y}^{-1}$ ). The values in the leaves of the tree show for the distinct environmental conditions mean soil carbon stock ( $\text{tC ha}^{-1}$ ), number and percentage of samples. The group acronyms are shown below the leaves of the regression tree.



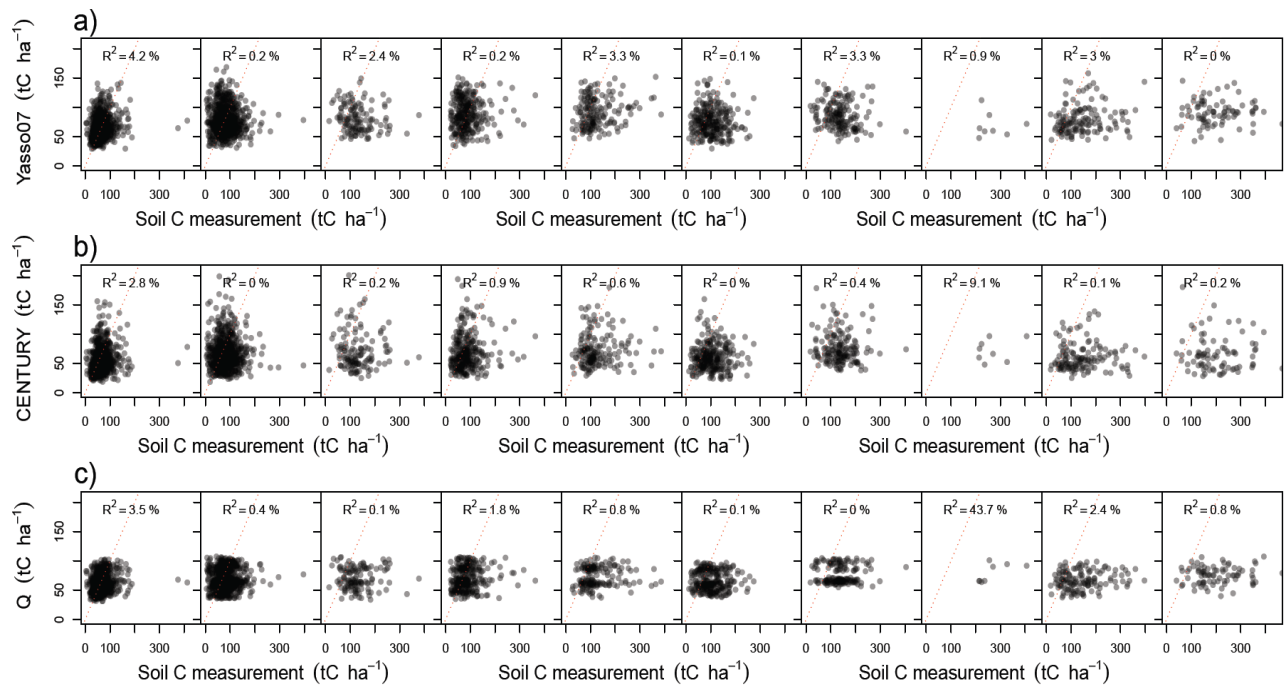
**Figure S4.** Density functions for 10 physicochemical groups of the soil carbon (SOC) stock (tC ha<sup>-1</sup>) Swedish forest soil inventory measurements for soil depth up to 1 m (total, grey fill) and for the soil humus horizon and mineral soil horizon. The thin lines are the density distributions. The thick lines are the group means and dashed lines are their confidence intervals. The n is number of samples. For description of group levels of SOC stocks, moisture, and fertility see Fig.2 and Table S1.



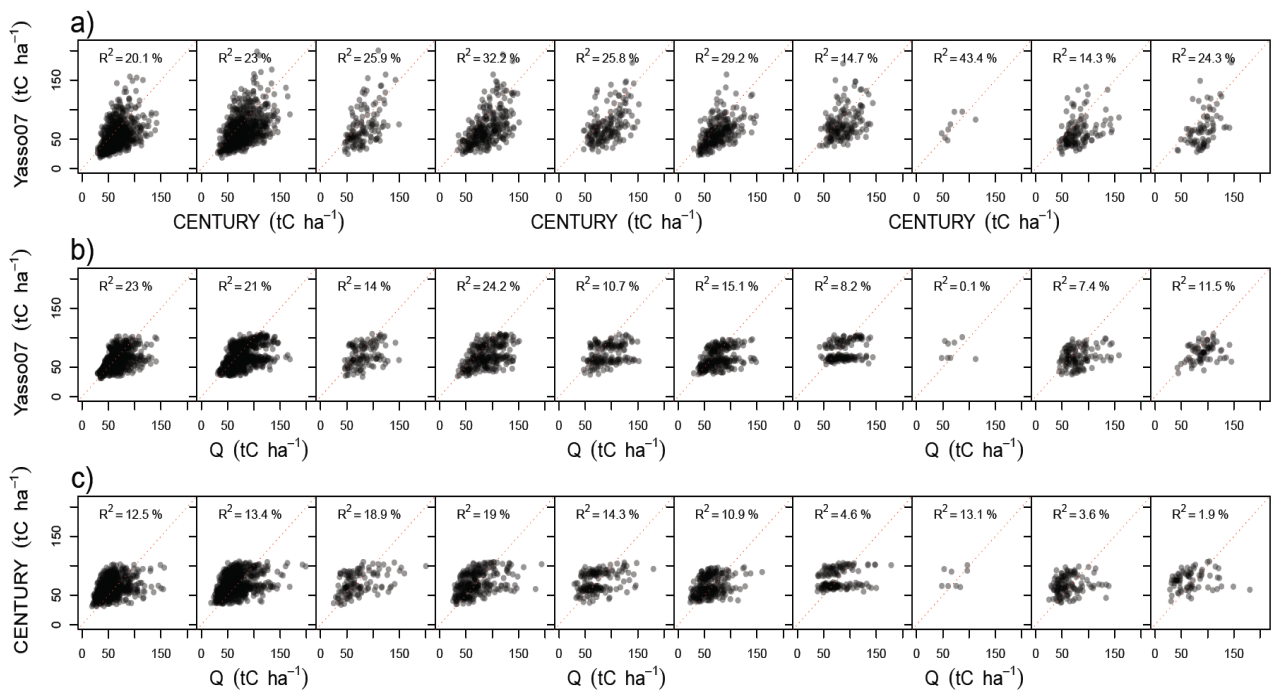
**Figure S5.** Bean plot of density functions for 5 groups of the soil carbon ( $tC\ ha^{-1}$ ) measurements (C.measured, grey fill) and soil carbon estimates simulated by the soil carbon models Yasso07, CENTURY, and Q with the litter input derived from the steady state forest. The thin lines are the density distributions. The thick lines are the group means and dashed lines are their confidence intervals. The n is number of samples. For description of group acronyms based on levels of SOC stocks, temperature, percentage of pine in canopy, precipitation, and nitrogen deposition see Fig.3 and Table S2.



**Figure S6.** Density functions for 10 physicochemical groups of the total annual plant litter input ( $tC\ ha^{-1}$ ) of steady state forest. The thick lines are the group means and dashed lines are their confidence intervals. The n is number of samples. For description of group levels of SOC stocks, moisture, and fertility see Fig.2 and Table S1.

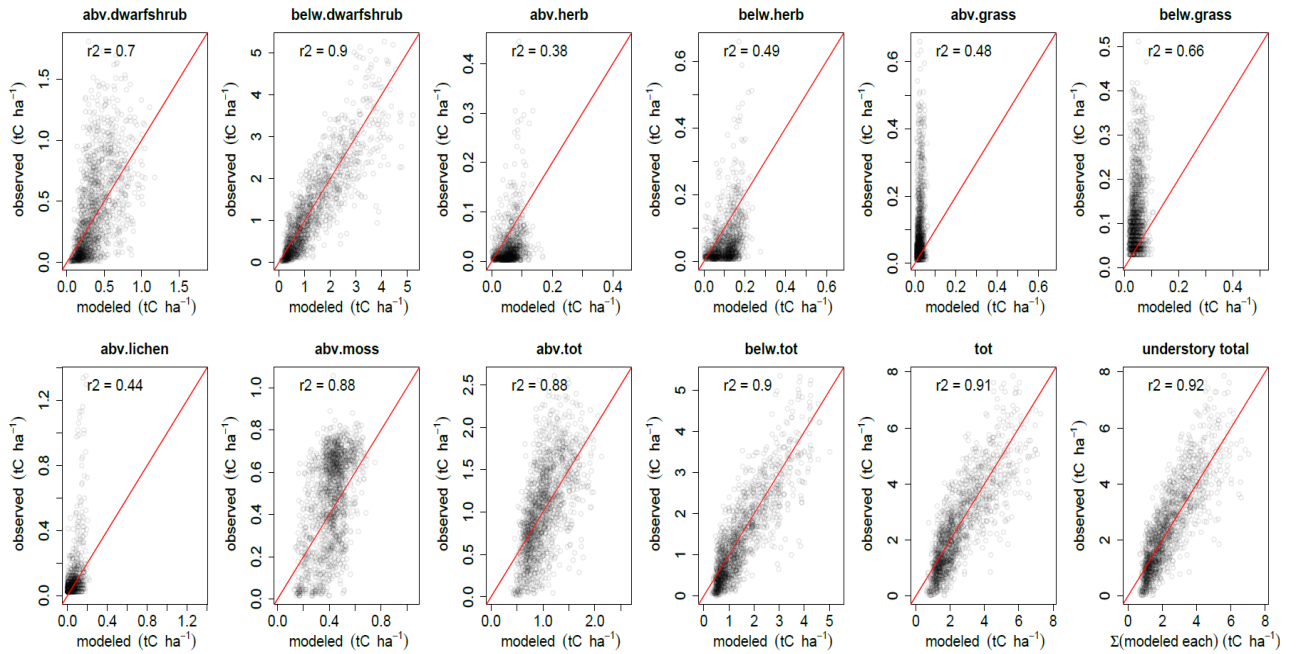


**Figure S7.** Scatter plots between model soil organic carbon stock ( $\text{tC ha}^{-1}$ ) measurements and a) Yasso07 and CENTURY, b) Yasso07 and Q, and c) CENTURY and Q for 10 physicochemical groups of Fig.4.

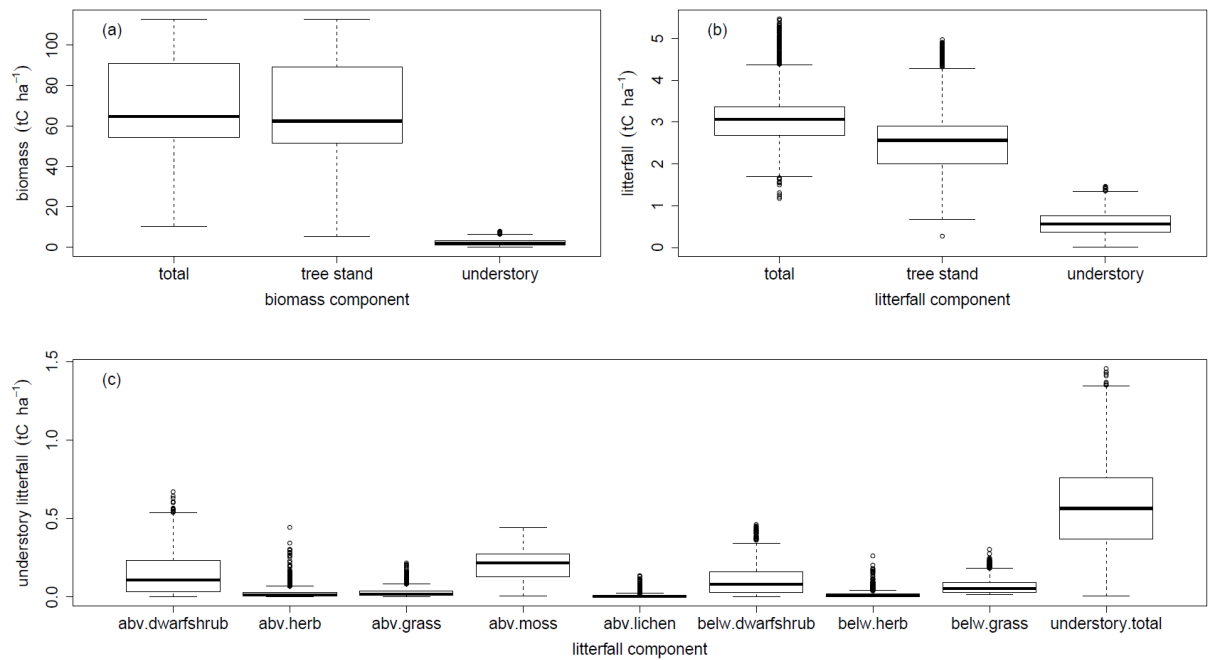


**Figure S8.** Scatter plots between model soil organic carbon estimates ( $\text{tC ha}^{-1}$ ) of a) Yasso07 and CENTURY, b) Yasso07 and Q, and c) CENTURY and Q for 10 physicochemical groups of Fig.4.

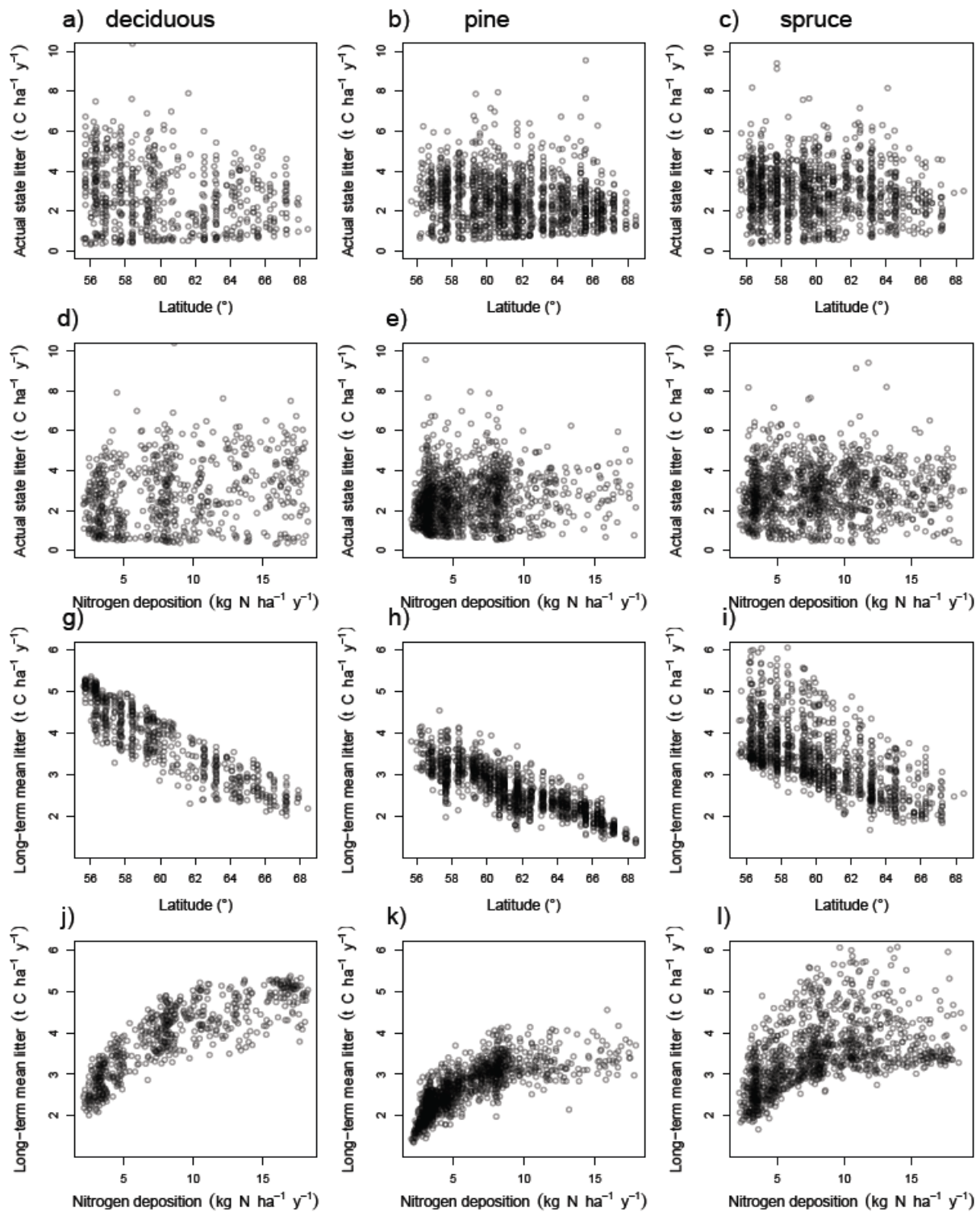




**Figure S9.** Scatter plots for the dry weight biomass ( $\text{tC ha}^{-1}$ ) of the functional types of understory vegetation for Swedish Forest Inventory plots in actual state being close to the estimated long-term mean conditions “steady state”. On the x-axis is the biomass modelled by the understory vegetation dry weight biomass ( $\text{tC ha}^{-1}$ ) models and on the y-axis is the observed coverage multiplied by the coverage/biomass conversion functions. The abbreviations “abv”, “belw”, and “tot” mean aboveground, belowground and total. The last panel for “understory total” shows high agreement between the sums of each modeled functional types and the sums of all functional types. The  $r^2$  values represent the coefficient of determination indicating how close the modeled values fit the observed values.



**Figure S10.** The tree stand and understory forest (a) biomass, (b) litterfall, and (c) understory litterfall (all in  $tC\ ha^{-1}$ ) for Swedish Forest Inventory plots with available understory coverage observations and in their actual state close to the estimated long-term mean conditions “steady state”.



**Figure S11.** Scatterplots between Latitude (°) and the actual state forest litterfall (tC ha<sup>-1</sup> y<sup>-1</sup>) a), b), c) or long-term mean “steady state” forest litterfall (tC ha<sup>-1</sup> y<sup>-1</sup>), g) h), i); and scatterplots between Nitrogen deposition (kgN ha<sup>-1</sup> y<sup>-1</sup>) and the actual state forest litterfall (tC ha<sup>-1</sup> y<sup>-1</sup>) d), e), f) or long-term mean “steady state” forest litterfall (tC ha<sup>-1</sup> y<sup>-1</sup>) j), k), l) for deciduous species, Scots pine, and Norway spruce dominated stands.