- Sediment characteristics as an important factor for revealing carbon 1
- storage in Zostera marina meadows: a comparison of four European 2 areas
- 4

5	Martin Dahl ^{1*} , Diana Deyanova ¹ , Silvia Gütschow ¹ , Maria E. Asplund ² , Liberatus D. Lyimo ^{1,3} ,
6	Ventzislav Karamfilov ⁴ , Rui Santos ⁵ , Mats Björk ¹ , Martin Gullström ¹

- 7
- 8 ¹ Seagrass Ecology & Physiology Research Group, Department of Ecology, Environment and Plant Sciences, Stockholm
- 9 University, SE-106 91 Stockholm, Sweden
- 10 ² The Sven Lovén Center for Marine Sciences, University of Gothenburg, Kristineberg 566, SE-451 78 Fiskebäckskil,
- 11 Sweden
- 12 ³ School of Biological Science, University of Dodoma, Box 338 Dodoma, Tanzania
- 13 ⁴ Institute for Biodiversity and Ecosystem Research at the Bulgarian Academy of Sciences, 2, Gagarin Street, 1113
- 14 Sofia, Bulgaria
- ⁵ ALGAE -Marine Ecology Research Group, CCMar Center of Marine Sciences, Faro, Portugal 15
- 16
- 17 *Correspondence author. E-mail: martin.dahl@su.se
- 18
- 19

20 Abstract. The seagrass ecosystem is an important natural carbon sink but the efficiency varies greatly depending on 21 species composition and environmental conditions. What causes this variation is not fully known and could have 22 important implications for management and protection of the seagrass habitat to continue to act as a natural carbon 23 sink. Here, we assessed sedimentary organic carbon in Zostera marina meadows (and adjacent unvegetated sediment) 24 in four distinct areas of Europe (Gullmar Fjord on the Swedish west coast, Askö in the Baltic Sea, Sozopol in Black Sea 25 and Ria Formosa in southern Portugal) down to \sim 35 cm depth. We also tested how sedimentary organic carbon in Z. 26 marina meadows relates to different sediment characteristics, a range of seagrass-associated variables and water 27 depth. The carbon storage varied among areas, where the Gullmar Fiord had a 15 times higher carbon storage 28 compared to Askö and Sozopol. We found that a high proportion of fine grain size, high porosity and low density of the 29 sediment is strongly related to high carbon content in Z. marina sediment. We suggest that sediment characteristics 30 should be highlighted as an important factor when evaluating high priority areas in management of Z. marina generated 31 carbon sinks.

- 32
- 33 Keywords: Carbon storage variability, Zostera marina, grain size, sediment characteristics, natural carbon sinks.
- 34

35 1. Introduction

36

37 Seagrass ecosystems are considered highly efficient natural carbon sinks (Mcleod et al., 2011) but there is a large 38 variation in the capacity to store carbon, depending on species composition and habitat characteristics (Lavery et al., 39 2013; Rozaimi et al., 2013). While the carbon sequestration efficiency is quite well documented for many seagrass 40 species (e.g. Kennedy et al., 2010; Fourgurean et al., 2012) the effects of different factors influencing intraspecific 41 variation has only recently been investigated. To get a more accurate estimate of the global seagrass carbon sink 42 capacity cause-effect relationships need to be better understood, and as seagrass loss is accelerating (Waycott *et al.*, 43 2009) information on habitat characteristics affecting carbon storage are of importance for an efficient protection and 44 management strategy to increase carbon storage capacity (Duarte et al., 2011).

45

46 There are several environmental factors (e.g. water depth and hydrodynamic processes) and seagrass habitat variables 47 (e.g. canopy height and shoot density) that influence the carbon storage in seagrass sediments (Samper-Villarreal et al., 48 2016). For example, seagrass meadows at shallower depths are known to have a high accumulation of sedimentary 49 carbon (Serrano et al., 2015), which could be associated with higher primary production and larger standing biomass 50 stock (Serrano et al., 2014). Dense meadows have the ability to stabilize the sediment (and thereby preventing it from 51 eroding) (Suvkerbuvk et al., 2015) and seagrass habitats with a high canopy can trap a high amount of suspended 52 particles and thus potentially increase the sedimentation of organic matter (Fonseca and Cahalan, 1992; Hendriks et 53 al., 2008). Further, as the belowground biomass largely contributes to the carbon storage due to its high production, 54 fast turnover and higher decay-resistant lignin content compared to the leaves (Duarte et al., 1998; Klap et al., 2000) a 55 large root-rhizome system could render a higher carbon storage (Kenworthy and Thaver, 1984). In the coastal 56 environment, sediment grain size is known to influence the aggregation of organic particles with finer grain sizes 57 increasing the organic matter content of the sediment (Mayer, 1994b). By reducing water velocity and facilitating 58 sedimentation processes a seagrass meadow could increase the amount of fine particles, which thus promote a high 59 carbon storage. Grain size has recently been shown to influence carbon storage in some seagrass areas (Röhr et al., 2016, Serrano et al., 2016), especially in meadows with a low contribution of autochthonous derived carbon, although
the influence of grain size on carbon storage is not universal for all seagrass species and habitats (Serrano et al., 2016).
Grain size is also strongly related to sediment porosity and density, which influence the oxygen conditions in the
sediment. Oxygen levels together with the microbial community composition, biomass carbon and nutrient content are
important factors for the degradation rate of organic matter in the sediment (Benner et al., 1984; Deming and Harass,
1993; Enriquez et al., 1993) and therefore influencing the carbon sequestration process.

66

67 Zostera maring L is the most widely spread seagrass species in the northern hemisphere, with a distribution in Europe 68 stretching from the southern Black Sea and the gulf of Cádiz (southern Portugal) up to Iceland and the northern parts of Norwav (Green and Short, 2003). The plant biomass is generally larger at higher latitudes (Short et al., 2007) because 69 70 of more optimal growth temperatures (Moore and Short, 2003). Large seagrass populations can be found along the 71 Swedish west coast and at the east coast of Denmark (Baden and Boström, 2001: Olesen and Sand-Jensen, 1994), where 72 they form extensive meadows with shoots over 1 m in length. Due to its wide distribution Z. marina populations have 73 adapted to a large range of environmental conditions, with potential differences in carbon storage capacity. The species 74 can tolerate salinity ranging from 5 to 35 (Boström et al., 2003) and a depth distribution from the intertidal down to 75 30 m depending on water clarity (Phillips and Meñez, 1988). Zostera marina also grows in various substrates, from 76 coarser stone-sand bottoms to finer silt and clay sediment. In this study, we aim to assess and compare carbon storage 77 in Z. marina meadows at four different areas in Europe as well as to examine relationships between sediment organic 78 carbon content and several explanatory predictors including seagrass structural complexity, carbon and nitrogen 79 content of the seagrass biomass and sediment characteristics (i.e. sediment porosity and density, and grain size) in 80 order to determine factors influencing the storage capacity of *Z. marina* meadows in these areas.

81

- 82 2. Methods
- 83

84 2.1 Study sites

85

86 This study was conducted in four different areas in Europe (the Swedish Skagerrak and Baltic coasts, Black Sea in 87 Bulgaria and the southern coast of Portugal; Table 1 and Fig. 1) from June to October 2013 with one complimentary 88 field sampling performed in October 2014. The different study areas cover a range of environmental and physical 89 conditions (e.g. salinity and temperature) for Z. maring in Europe. In each area, two meadows and one unvegetated 90 area (reference site) were sampled, except for Portugal with one additional unvegetated area and the Baltic Sea where 91 one meadow and one unvegetated area were added (Table 1). The sampling on the Swedish west coast were carried 92 out off the Sven Lovén Centre for Marine Sciences – Kristineberg in the Gullmar Fjord (58°20'N, 11°33'E; Table 1). The 93 area is comprised of small islands and shallow bays making it highly productive and a suitable environment for seagrass 94 growth with many sheltered soft bottoms covered by extended Z. marina beds. In the Baltic Sea, samples were collected 95 in the area around the Askö Laboratory in Stockholm Archipelago (58°49'N, 17°39'E). The Baltic Sea is a brackish water 96 system and the salinity is about 5-6 outside Askö, which is on the distribution limit for *Z. marina* (Boström et al., 2003). 97 Low salinity is known to negatively affect production and growth of the plant (Salo et al., 2014). In the Baltic Sea, Z. 98 marina grows at approximately 2-5 m depths (sometimes together with Ruppia maritima) and on more coarse 99 sediment compared to the Skagerrak area (Baden and Boström, 2001). In the Black Sea, sampling was carried out in 100 two sites around the Laboratory of Marine Ecology in Sozopol, Bulgaria (42°25'N, 27°41E). The salinity is around 17 101 and commonly Z. marina grows in mixed stands with Z. noltii. The Ropotamo (Rt) site is situated in the vicinity of the 102 Ropotamo river mouth. Ria Formosa (Algarve Marine Sciences Centre – Faro) is located in southern Portugal (36°59'N, 103 $7^{\circ}52'W$) and is a coastal lagoon with large intertidal areas and a tidal fluctuation of 2-3 m. This is the only area in the 104 present study with pronounced tidal variation, and the water depth for the Portugal sites was standardized to mean 105 low water (MLW) by calculating the difference between the measured water depth and the tide at the time of 106 measurement. The tide values were obtained from Ria Formosa tidal station (Faro-Olhão) with the mean water level

- as reference depth. Ria Formosa is a lagoon with scarce *Z. marina* distribution (which at times grows together with *Cymodocea nodosa*) and apart from one other area in Portugal (Óbidos Lagoon) the only one that still harbor *Z. marina*,
 which has decreased drastically during the past 20 years (Cunha et al., 2013).
- 110

111 2.2 Sediment sampling and biometrical measurements

112

113 At each site, six sediment cores were taken with a push corer (h=50 cm, $\phi=8$ cm) at a distance of 10-30 m apart from 114 each other. The edge of the corer was sharpened to easier press down the core into the sediment and to reduce the 115 shortening (compression) of the sediment collected (Serrano *et al.*, 2012). However, due to the difference in sediment 116 compactness between sites the length of the sediment core varied (because of difficulties in pressing down the core in 117 coarser sediment). Each core was sliced into a maximum of six segments (0-2.5 cm, 2.5-5 cm, 5-12.5 cm, 12.5-25 cm, 118 25-37.5 cm. 37.5-45 cm) with the majority of samples lacking the deepest segment. The corers were stored vertical 119 prior to slicing the sediment into depth segments. We examined the influence of core shortening in the Skagerrak area. 120 where the compression is expected to be the highest in our study due to the soft sediment and high porosity (Glew et 121 al. 2001), by measuring the length of the outer and inner edge of the corer from the edge of the core to the sediment 122 surface when pressed down into the sediment (n=6). The effect of core shortening was derived from the difference 123 between the inner and outer length of the corer and compression was calculated to be 8 %. This has not been corrected 124 for in the data and is further addressed in the discussion as a source of error. Within a few meters from each core at 125 the seagrass sites, shoot height (cm, n=20) was measured, percentage seagrass coverage (n=10) were estimated (in 0.5 126 x 0.5 m squares) and biomass samples (n=3) were collected (0.25 x 0.25 m). The biomass samples were used for 127 estimating above- and belowground seagrass biomass (as dry weight) and for counting number of shoots. Before 128 weighing the seagrass was cleaned and epiphytes removed, and the dry weight was measured after 24-48 h in 60°C 129 until constant weight. One out of the three biomass samples collected around each core were analyzed for carbon and 130 nitrogen content (n = 6 for each meadow). The sediment samples were cleaned from roots and rhizomes, larger shells 131 and benthic organisms, and homogenized prior of drying. The sediment was dried in 60°C for approximately 48 h until

132 the weight was constant. Before drying a sediment sample it was divided into two subsamples, one for analysis of 133 carbon and nitrogen content, and the other for grain size analysis. A mixing mill (Retch 400 mm) was used to grind the 134 sediment into a fine powder to further homogenize the subsample used for analysis of carbon and nitrogen content. 135 The carbon and nitrogen contents in biomass and sediment were analyzed using an organic elemental analyzer (Flash 136 2000, Thermo Fischer scientific). Prior to analysis for C_{org} content the sediment samples were pre-treated with 1 M HCl 137 (direct addition until the reaction of carbonate was complete) to remove inorganic carbon and dried at 60°C for 24 h. 138 Total nitrogen (N_T) was derived from untreated sediment samples was used to estimate the nitrogen content due to 139 possible alteration of the nitrogen values when treated with HCl (Harris et al., 2001). Sediment porosity was given as 140 percentage (%) by calculating sediment wet weight minus dry weight divided by the sample volume, whereas sediment 141 density (g DW mL⁻¹) was derived from dividing the dry weight of the sediment by the volume of the sample. A literature 142 survey for measurements of sediment carbon content in Z. marina meadows in Europe and other temperate regions 143 was conducted using Web of Science and Google Scholar with the search words "Zostera maring, sediment, organic". 144 Additionally, grev literature including thesis work was also used as well as unpublished data from colleagues.

145

146 2.3 Grain size analysis

147

148 Three sediment cores in each habitat were used for particle size analysis and each depth section was separately 149 analyzed. Prior to analysis the total dry weight of sediment for each section was determined and 100 ml of 0.05 M 150 Na₄P₂O₇ was added to break down aggregates of clay particles. All of the sediment samples were dry-sieved for 10 min 151 using a sieving tower (CISA electromagnetic sieve shaker, Spain) (including sieves of 0.074 mm, 0.125 mm, 0.25 mm, 152 0.5 mm, 1 mm and 2 mm) and the sediment of each sieve was weighed to determine the weight of the separate fractions 153 (the average weight of the samples was 97 g). In depth sections with high organic carbon content (>0.5%), the organic 154 matter was removed prior to dry sieving, through oxidation with 35% H₂O₂, as the organic matter content leads to 155 aggregation of particles (Gee and Bauder, 1986). When the reaction with H_2O_2 had ceased the samples were centrifuged 156 for a minimum of 20 min at 4500 RPM, in which the supernatant was carefully removed using a pipette, and

subsequently the samples were washed in distilled water and centrifuged again to remove H₂O₂ residues. After dry seiving, some of the samples from the Skagerrak and Ria Formosa areas had to be analysed with hydrometer for an accurate estimate of total grain size due to a high proportion of finer fractions (>15% was assessed as %<0.074 mm) in those sediments. The samples were once more treated with 0.05 M Na₄P₂O₇ and placed in a 1L cylinder containing distilled water and kept in suspension. At fixed time intervals (1, 2, 4, 10, 20, 50, 100, 200, 400 and 1000 min) the hydrometer was inserted and the concentration of sediment (g L⁻¹) was noted. The mean grain size was presented in phi (ϕ) units.

164

165 2.4 Statistical analysis

166

167 To test for differences in sedimentary carbon storage ($%C_{org}$ and g C_{org} cm⁻²) and grain size particles >0.074 mm among 168 areas, between Z. maring and unvegetated areas (habitat) and among sediment depths, nested general linear mixed 169 model ANOVAs were performed using site as random factor and with habitat nested in area and sediment depth nested 170 in core. In those cases where the ANOVA models were significant, Tukey's HSD post hoc test was used to determine 171 significant differences between specific areas and between habitats (Z. maring meadows vs. unvegetated areas). Prior 172 to analysis all data were checked for normal distribution using the Shapiro-Wilk normality test and homogeneity of 173 variances using Levene's test. When assumptions were not met the data was $\log_{10}(x+1)$ transformed. Partial Least 174 Square (PLS) regression technique (by modeling of projections of latent structures; Wold et al., 2001) and Principal 175 Component Analysis (PCA) were conducted in SIMCA 13.0.3 (UMETRICS) to test the influence of sediment 176 characteristics, water depth and seagrass-related variables on sediment carbon content (mean % C for the top 25 cm 177 of sediment). The advantage of using PLS modeling is that it can handle collinear explanatory data as well as a large 178 number of predictors. All cores were standardized to a depth of 25 cm for the sediment characteristics (porosity, 179 density, grain size and organic carbon content) prior to the PLS- and PCA analyses. Some of the cores at Askö (both 180 seagrass- and unvegetated sites) lacked the 12.5-25 cm depth segment and in these cases logarithmic regressions were used to extrapolate the data down to 25 cm depth (Torö [T] %Corg; y=-0.87ln[x]+0.3845, g Corg cm⁻²; y=-181

- 182 0.001ln[x]+0.0052, Torö [Tr] [r], %C_{org}; y=-0.032ln[x]+0.2225, g C_{org} cm⁻²; y=-0.0002ln[x]+0.0053). The carbon content
- 183 in seagrass meadows decreases logarithmically with sediment depth in general (Fourqurean et al., 2012) due to
- degradation and remineralization of organic material with time (Burdige, 2007; Henrichs, 1992),

- 186 3. Results
- 187

188 3.1 Variation in sedimentary carbon storage

189

190 The Z. marina meadows had significantly higher sedimentary carbon content (both in % Corg and g Corg cm⁻²) compared 191 to the unvegetated areas (P < 0.001; Table 2). Within the different areas only Gullmar Fiord and Ria Formosa showed 192 significantly different values compared to their respective unvegetated areas (P < 0.001) while Askö and Sozopol did 193 not show any between-habitat differences (Fig. 2). In terms of % Corg and g Corg cm⁻². Gullmar Fiord was significantly 194 different from all other areas (P < 0.05) whereas Ria Formosa were significantly different to Sozopol (P < 0.05) but not 195 to Askö, and no difference was seen between Sozopol and Askö (Table 2; Fig. 2). The highest amount of sedimentary 196 carbon was seen in the Gullmar Fjord, followed by Ria Formosa, Askö and Sozopol (Table 4). There were no significant 197 differences in either % C_{org} or g C_{org} cm⁻² among different sediment depths (Table 2; Fig. 3).

198

3.2 Influence of sediment characteristics and seagrass-associated variables on carbon storage

200

201 When the relationship between % Corg and explanatory variables (Tables 2, 3 and 4) was examined in a PLS (Partial 202 least square) regression model the sediment characteristics explained most of the model (with a variance of importance 203 value >1) where the proportion of sediment particles < 0.074 mm (%) was the most important, followed by sediment 204 porosity (%), sediment density (g DW mL⁻¹) and mean grain size (ϕ) (Fig. 4). These variables characterizing the 205 sediment were all positively correlated to % C_{org} except sediment density that showed a negative relationship with % 206 C_{org} . The cumulative fraction explaining the % C_{org} variation (R_v^2 cum) of the predictor variables combined was 0.81 207 and the models cross-validated variance (O^2 statistics) showed high predictability with O^2 -value of 0.79, thus larger 208 than the significant level of 0.05. The results of the model with g C_{org} cm⁻² (not shown here) as response variable were 209 highly similar to the results of % C_{org} (Q² = 0.77, R_y^2 cum = 0.78) with the same predictor variables (i.e. sediment 210 characteristics) explaining most of the % Corg variation and correlated in the same way. All seagrass-associated

variables showed a positive relationship with % C_{org} except for belowground (Bg) biomass N (%), which was the least influential variable in the model. In general, the seagrass-associated variables showed a lower contribution to the overall model compared to the sediment characteristics. Water depth (m) was also negatively correlated to % C_{org} but was, as with the seagrass-associated variables, of minor importance (Fig. 4).

215

216 The amount of sediment particles <0.0074 mm was significantly higher in Z. marina meadows compared to unvegetated 217 areas (P < 0.001; Table 2). This was true for all of four areas when comparing to respective unvegetated areas (P < 0.001; Table 2). 218 0.05). Sediment grain size particles < 0.074 mm were significantly different between areas (P < 0.001: Table 2), where 219 Gullmar Fiord and Ria Formosa showed significantly higher values compared to the other areas (P < 0.001), while there 220 were no significant differences between Askö and Sozopol (Table 4). There was no difference in grain size particles 221 <0.074 in terms of sediment depth (Table 2). Mean grain size (ϕ) and sediment particles < 0.074 mm (%) both showed 222 strong positive linear relationship with % C_{org} in Z. maring meadows (mean phi (ϕ), $R^2 = 0.74$, P < 0.001; sediment 223 particles < 0.074 mm (%), $R^2 = 0.91$, P < 0.001; Fig. 5a and b). For unvegetated areas, mean grain size (ϕ) did not show 224 any relationship with % C_{org} (linear regression, $R^2 = 0.009$, P < 0.40; Fig 5c) but was positively related to sediment 225 particles <0.074 mm (%) (linear regression, $R^2 = 0.42$, P < 0.001; Fig. 5d). The sediment density (g DW mL⁻¹) had a 226 negative effect on % C_{org} in the seagrass sites (linear regression, $R^2 = 0.84$, P < 0.001) and sediment porosity (%) was 227 positively related to % C_{org} (linear regression, R² = 0.80, P < 0.001; Fig. 6a and b). There was no significant relationship 228 between % C_{org} and sediment density (g DW mL⁻¹) in unvegetated areas while sediment porosity (%) was significantly 229 influencing % C_{org} but showed a low R₂-value (linear regression, R² = 0.08, P < 0.001; Fig. S2).

230

- for the carbon content while the sedimentary carbon in Ria Formosa was more related to seagrass cover (%) and dry
- 237 weight belowground biomass (m²). The sedimentary organic carbon content in seagrass sites in Baltic- and Black Seas
- were also more related to the seagrass-associated variables, such as dry weight aboveground biomass (m²) and shoot
- density (m²), but also water depth (m) for one of the sites (Storsand, S).

242 4. Discussion

243

244 In this assessment of four Z. marina areas in Europe, we found a large variation in organic carbon storage where the 245 carbon-rich sediment of the Gullmar Fjord on the Swedish west coast was 15 times higher compared to levels in the 246 Baltic- and Black Seas. Along with recent studies (Lavery et al., 2013; Samper-Villarreal et al., 2016), this study shows 247 that the environmental conditions play an essential role in determining the carbon sink capacity. Here we demonstrate 248 that sediment characteristics influence carbon storage in Z. marina meadows, where high sedimentary organic carbon 249 corresponds with high content of fine grain size, high sediment porosity and low sediment density. Seagrass meadows 250 situated in areas characterized by these sediment properties are therefore suggested to have a high potential as natural 251 carbon sinks.

252

253 Overall Z. marina meadows showed higher carbon content than nearby unvegetated areas, with the exception of the 254 seagrass meadows with the lowest carbon storage, which illustrates just as previous studies have shown (e.g. Kennedy 255 et al. 2010; Mcleod et al. 2011), that the seagrass ecosystem is a significant carbon sink. The mean carbon content of 256 the Gullmar Fiord was higher than estimated global averages (Fourgurean et al., 2012; Kennedy et al., 2010). 257 demonstrating the high carbon capacity of the area. The comparison with other Z. marina meadows in Europe and USA 258 also showed that the Swedish Skagerrak coast (e.g. the Gullmar Fjord) has an overall high carbon storage capacity 259 (Table 5). The lowest carbon content were found in the Baltic- and Black Sea (no previous studies on sedimentary 260 carbon content could be found for the Black Sea; Table 5). This could be related to less suitable physical conditions of 261 the Brackish environment with lower salinity, which may negatively affect plant growth and meadow productivity (Salo 262 et al., 2014), in combination with growing in more exposed areas with coarser (sandy) sediment, as seen in the Z. 263 *marina* meadows at Askö, where the most sheltered bays with finer grain sizes are dominated by brackish water plants. 264 such as Potamogeton pectinatus and Zannichellia palustris (Idestam-Almquist, 2000). Meadows situated in more 265 exposed areas could result in a high export of the produced organic matter, as suggested by Röhr et al., (2016) instead 266 of the carbon being accumulated in the sediment, leading to a low carbon storage potential of the area. This could also

267 be true for the meadows in Ria Formosa, the only area in this study with a pronounced tide, where the higher 268 hydrodynamic forces could also lead to increased sediment erosion. Although the meadows at Sozopol and Askö were 269 dominated by Z. marina also smaller seagrass species (i.e. Zostera noltii and Ruppia maritima) were found in the 270 meadows; smaller species with lower canopy and belowground biomass could also be part of the explanation to lower 271 sedimentary carbon concentrations as trapping of suspended particles (Fonseca and Cahalan, 1992) and the 272 belowground biomass production contribute to the accumulation of carbon (Duarte et al., 1998). The trapping of fine-273 grained particles and prevention of sediment particle resuspention (by reducing the water velocity) in the canopy are 274 also likely the reason why the Z. maring meadows had substantially higher amount of smaller grain size particles 275 compared to the unvegetated areas. Due to the fact that core shortening was not corrected for in our sediment samples 276 there might be a margin of error up to 8% in our data. The influence of compression is most likely highest in the 277 Skagerrak area, where the sediment is soft and has a high porosity (Glew et al., 2001), but given the large variation in 278 carbon storage a reduction of 8% in sedimentary carbon content will not undermine our general conclusion.

279

280 A high carbon content in Zostera marina sediment seems to be related to the sediment characteristics of the area. A 281 high proportion of finer grain size particles leads to preservation and accumulation of organic matter (Keil et al., 1994; 282 Mayer, 1994a, 1994b) due to a higher surface area on fine-grained particles, leading to an aggregation of organic matter 283 (Bergamaschi et al., 1997). Finer grain sizes in combination with high organic matter and nutrient content, as seen in 284 the Gullmar Fiord sites, could cause a depletion of oxygen in the sediment because of increased oxygen consumption 285 by detritivore organisms (detritivores) and decreased permeability (Pollard and Moriarty, 1991; Wilson et al., 2008). 286 which slows down the degradation process of organic matter (Hedges and Keil, 1995). Sediment grain size has recently 287 been described as a strong predictor for carbon storage in another blue carbon habitat, i.e. saltmarshes (Kelleway et 288 al., 2016), and for seagrass meadows, the finer grain sized particles has shown to influence sedimentary carbon content 289 in some seagrass habitats (Röhr et al., 2016), while in others it seems less important (Samper-Villarreal *et al.*, 2016). 290 The relations between carbon storage and various sediment characteristics are more pronounced in meadows with 291 low seagrass biomass and high proportion of finer particle sizes, while in meadows with larger seagrass species, i.e.

292 *Posidonia* spp. and *Amphibolis* spp., having high amount of autochthonously derived sedimentary carbon, the mud and 293 silt content was shown to have little influence (Serrano et al., 2016). Compared to *Posidonia* spp. and *Amphibolis* spp. 294 the smaller sized Z. maring plants will potentially contribute less to the sediment organic matter pool, which might be 295 the reason to why the proportion of fine sediment particles was strongly coupled to a high carbon content in the present 296 study. Other factors have previously shown to be of importance, such as water depth, meadow productivity, 297 sedimentation rate, trapping of fine-grained sediment and organic matter (Serrano et al., 2015), and while these factors 298 were not seen or accounted for in this study they may also be relevant when determining areas of high carbon storage 299 potential. The grain size is directly linked to the sediment porosity and density where the organic carbon has a negative 300 effect on sediment density (Avnimelech et al., 2001: Gullström et al., submitted). This was also seen in our study as 301 higher sedimentary carbon values were found in areas with lower sediment density (and hence higher porosity). For 302 these reasons, we suggest that sediment characteristics of the area where Z. marina meadows are situated is relevant 303 for revealing the carbon storage potential.

304

305 A high organic content in the sediment could, however, cause a depletion of oxygen (Holmer, 1999) and at too low 306 oxygen levels seagrass can no longer maintain the aerobic conditions of the rhizosphere, eventually leading to seagrass 307 mortality (Terrados et al., 1999) with consequences for the carbon storage capacity. The seagrasses could adapt to 308 lower oxygen concentrations by reducing the shoot density (Folmer et al., 2012) and thereby lower the oxygen demand 309 of the root-rhizome system, which may explain why the areas with high proportion of fine grain size particles in this 310 study had the lowest shoot density. High canopy height, high shoot density and shallow depths are generally considered 311 to increase sedimentation rates and thus promote accumulation of finer grain size particles (Bos et al., 2007; Fonseca 312 and Cahalan, 1992; Peralta et al., 2008). This implies that aboveground seagrass structure and water depth should 313 influence the sediment carbon storage, however, in our study these variables were of minor influence. The influence of 314 seagrass meadow structure on sediment composition is complex and hard to predict, and may be highly influenced by 315 environmental conditions (van Katwijk et al., 2010). The carbon storage in Z. marina meadows in our study was clearly 316 related to sediments with high proportion of fine grain size particles, high porosity and low density. In areas with less

317 fine-sized sediment particles other variables, such as above- and below-ground seagrass biomass, seagrass cover and 318 shoot density, have a more pronounced influence on carbon storage levels. For example, the influence of belowground 319 biomass and seagrass cover on sedimentary carbon content in Ria Formosa could be due to the stabilizing properties 320 of dense meadows (Suykerbuyk et al., 2015), the binding of sediment by the root-rhizome system (Christianen et al., 321 2013) and the high lignin content of the belowground biomass (Klap et al., 2000), which results in more decay-resistant 322 carbon and a slower decomposition (Cowie and Hedges, 1984; Ertel and Hedges, 1985). Seagrass biomass and cover 323 are generally highly dynamic and act on a shorter time-scale than the sedimentary carbon storage processes, therefore 324 estimates of present seagrass meadow properties may not be fully representative over decades or centuries, which is 325 the likely time-scale for carbon storage in the sediment. The age of the sediment and the rate of accumulation of organic 326 matter are factors that vary between sites where a higher sedimentation rate increases the amount of organic carbon 327 and could be a potential explanation to variation in carbon storage among seagrass meadows (Serrano et al, 2015).

328

329 The continuous loss of seagrass areas (Waycott et al., 2009) leads to a decline in natural carbon sinks (Dahl et al., 2016; 330 Marbà et al., 2015), and to ensure efficient management, factors for high carbon storage capacity should be evaluated. 331 Several environmental and seagrass-related factors have shown to be of importance, i.e. water depth (Serrano et al., 332 2014), meadow size (Ricart et al., 2015), hydrodynamics and seagrass canopy complexity (Samper-Villarreal et al., 333 2016). In our study, the main factors related to high carbon storage were the sediment density and porosity, and 334 amount of fine grain size particles in the sediment, whereas the seagrass-associated variables had a minor influence. 335 Therefore, we highlight that the sediment characteristics is an important factor for a high carbon storage potential in 336 these types of Z. marina meadows, and should be taking into consideration (together with other relevant factors) when 337 evaluating high priority areas for protection of efficient carbon storage Z. marina areas.

338

339 Data availability

- All data is presented in the manuscript and supplementary figures. Figure S1, shows the variance of importance (VIP)
 for the response variables in the PLS model (see Fig. 4). Figure S2, semi-log plots showing the relationship between
 sediment porosity and density, and sedimentary organic carbon for unvegetated areas.
- 344

345 Author contribution

346

The design of this study was carried out by Martin Dahl, Martin Gullström, Mats Björk and Diana Deyanova. The
collection of data was done by Martin Dahl, Diana Deyanova, Liberatus D. Lyimo, Martin Gullström, Maria E. Asplund
and Ventzislav Karamfilov, and was analyzed by Martin Dahl, Martin Gullström, Mats Björk, Silvia Gütschow and Maria
E. Asplund. Martin Dahl prepared the manuscript with contribution from Martin Gullström, Mats Björk, Maria E.
Asplund, Diana Deyanova, Ventzislav Karamfilov, Silvia Gütschow and Rui Santos.

352

353 Acknowledgement

354

We thank Bruno Dias Duarte Fragoso, Dimitar Berov, Atanas Tanev, Diana Perry, Tom Staveley and Kuzman Dimov for their assistance in the field, and Stefania Klayn and Yoana Georgieva for helping out with preparing the sediment prior to analysis. We are also grateful to Alan Koliji for his contribution on the grain size analysis. This study was partly funded by the Swedish International Development Cooperation Agency (Sida) through the Bilateral Marine Science Program between Sweden and Tanzania.

- 360
- 361

362 References

Avnimelech, Y., Ritvo, G., Meijer, L. E. and Kochba, M.: Water content, organic carbon and dry bulk density in flooded
sediments, Aquac. Eng., 25(1), 25–33, doi:10.1016/S0144-8609(01)00068-1, 2001.

Baden, S. P. and Boström, C.: The leaf canopy of seagrass beds: faunal community structure and function in a salinity
gradient along the Swedish coast, in Ecological Studies, vol. 151, edited by K. Reise, pp. 213–236, Springer Berlin /
Heidelberg., 2001.

Benner, R., Maccubbin, A. E. and Hodson, R. E.: Anaerobic biodegradation of the lignin and polysaccharide components
of lignocellulose and synthetic lignin by sediment microflora, Appl. Environ. Microbiol., 47(5), 998–1004, 1984.

Bergamaschi, B. a., Tsamakis, E., Keil, R. G., Eglinton, T. I., Montluçon, D. B. and Hedges, J. I.: The effect of grain size and
surface area on organic matter, lignin and carbohydrate concentration, and molecular compositions in Peru Margin
sediments, Geochim. Cosmochim. Acta, 61(6), 1247–1260, doi:10.1016/S0016-7037(96)00394-8, 1997.

Bos, A. R., Bouma, T. J., de Kort, G. L. J. and van Katwijk, M. M.: Ecosystem engineering by annual intertidal seagrass beds:
sediment accretion and modification, Estuar. Coast. Shelf Sci., 74(1-2), 344–348, doi:10.1016/j.ecss.2007.04.006, 2007.

Boschker, H., Wielemaker, A., Schaub, B. and Holmer, M.: Limited coupling of macrophyte production and bacterial
carbon cycling in the sediments of *Zostera* spp. meadows, Mar. Ecol. Prog. Ser., 203, 181–189,
doi:10.3354/meps203181, 2000.

Boström, C., Baden, S. and Krause-Jensen, D.: The seagrasses of Scandinavia and the Baltic sea, in World atlas of
seagrasses: present status and future conservation, edited by E. . Green and F. T. Short, pp. 27–37, California Press,
California, 2003.

Boström, C., O'Brien, K., Roos, C. and Ekebom, J.: Environmental variables explaining structural and functional diversity
of seagrass macrofauna in an archipelago landscape, J. Exp. Mar. Bio. Ecol., 335, 52–73,
doi:10.1016/j.jembe.2006.02.015, 2006.

Burdige, D. J.: Preservation of organic matter in marine sediments: Controls, mechanisms, and an imbalance in sediment
 organic carbon budgets?, Chem. Rev., 107(2), 467–485, doi:10.1021/cr050347q, 2007.

- Christianen, M. J. A., van Belzen, J., Herman, P. M. J., van Katwijk, M. M., Lamers, L. P. M., van Leent, P. J. M. and Bouma, T.
 J.: Low-canopy seagrass beds still provide important coastal protection services, PLoS One, 8(5), doi:10.1371/journal.pone.0062413, 2013.
- Cowie, G. L. and Hedges, J. I.: Carbohydrates sources in a coastal marine environment, Geochim. Cosmochim. Acta, 48, 2075–2087, 1984.

Cunha, A. H., Assis, J. F. and Serrão, E. A.: Seagrasses in Portugal: a most endangered marine habitat, Aquat. Bot., 104, 193–203, doi:10.1016/j.aquabot.2011.08.007, 2013.

Dahl, M., Deyanova, D., Lyimo, L. D., Näslund, J., Samuelsson, G. S., Mtolera, M. S. P., Björk, M. and Gullström, M.: Effects
of shading and simulated grazing on carbon sequestration in a tropical seagrass meadow, J. Ecol., n/a-n/a,
doi:10.1111/1365-2745.12564, 2016.

- Dale, N. G.: Bacteria in intertidal sediment: Factors related to their distribution, Limnol. Oceanogr., 19(3), 509–519,
 1974.
- Deming, J. W. and Harass, J. A.: The early diagenesis of organic matter: bacterial activity, in Organic Geochemstry:
 principles and application, edited by M. H. Engel and S. A. Macko., 1993.
- Duarte, C. M., Merino, M., Agawin, N. S. R., Uri, J., Fortes, M. D., Gallegos, M. E., Marbá, N. and Hemminga, M. A.: Root
 production and belowground seagrass biomass, Mar. Ecol. Prog. Ser., 171, 97–108, 1998.
- 402 Duarte, C. M., Kennedy, H., Marbà, N. and Hendriks, I.: Assessing the capacity of seagrass meadows for carbon burial:
 403 current limitations and future strategies, Ocean Coast. Manag., 83, 32–38, doi:10.1016/j.ocecoaman.2011.09.001, 2011.
- Enriquez, S., Duarte, C. M. and Sand-Jensen, K.: Patterns in decomposition rates among photosynthetic organisms: the
 importance of detritus C :N :P content, Oecologia, 94, 457–471, 1993.
- Ertel, J. R. and Hedges, J. I.: Sources of sedimentary humic substances: vascular plant debris, Geochim. Cosmochim. Acta,
 49(10), 2097–2107, doi:10.1016/0016-7037(85)90067-5, 1985.
- Folmer, E. O., Geest, M., Jansen, E., Olff, H., Michael Anderson, T., Piersma, T. and Gils, J. a.: Seagrass-sediment feedback:
 an exploration using a non-recursive structural equation model, Ecosystems, 15(8), 1380–1393, doi:10.1007/s10021012-9591-6, 2012.
- Fonseca, M. S., Thayer, G. W., Chester, A. J. and Foltz, C.: Impact of Scallop Harvesting on Eelgrass (*Zostera marina*)
 Meadows : Implications for Management, North Am. J. Fish., 4, 37–41, doi:10.1577/1548-8659(1984)4, 1984.
- Fonseca, M. S. and Cahalan, J. A.: A preliminary evaluation of wave attenuation by four species of seagrass, Estuar. Coast.
 Shelf Sci., 35, 565–576, 1992.
- Fourqurean, J. W., Duarte, C. M., Kennedy, H., Marbà, N., Holmer, M., Mateo, M. A., Apostolaki, E. T., Kendrick, G. a., KrauseJensen, D., McGlathery, K. J. and Serrano, O.: Seagrass ecosystems as a globally significant carbon stock, Nat. Geosci.,
 5(7), 505–509, doi:10.1038/ngeo1477, 2012.
- Fredriksen, S., De Backer, A., Boström, C. and Christie, H.: Infauna from *Zostera marina* L. meadows in Norway.
 Differences in vegetated and unvegetated areas, Mar. Biol. Res., 6(February 2015), 189–200, doi:10.1080/17451000903042461, 2010.
- Gee, G. W. and Bauder, J. W.: Particle size analysis, in Methods of Soil Analysis: Part 1—Physical and Mineralogical
 Methods, edited by A. Klute, pp. 383–409, American Society of Agronomy, Madison., 1986.
- Green, E. P. and Short, F. T.: World atlas of seagrasses: present status and future conservation, University of California
 Press, Berkley USA., 2003.

- Gullström, M., Baden, S. and Lindegarth, M.: Spatial patterns and environmental correlates in leaf-associated epifaunal
 assemblages of temperate seagrass (*Zostera marina*) meadows, Mar. Biol., 159, 413–425, doi:10.1007/s00227-0111819-z. 2012.
- Gullström, M., Lyimo, L. D., Dahl, M., Samuelsson, G. S., Eggertsen, M., Anderberg, E., Rasmusson, L. M., Linderholm, L.
 W., Knudby, A., Bandeira, S., Mtwana Nordlund, L., Björk, M.:Blue carbon storage in tropical seagrass meadows
 influenced by plant structure, sediment composition and landscape configuration. Submitted manuscript.
- Harris, D., Horwáth, W. R. and van Kessel, C.: Acid fumigation of soils to remove carbonates prior to total organic carbon
 or CARBON-13 isotopic analysis, Soil Sci. Soc. Am. J., 65(July 2000), 1853–1856, doi:10.2136/sssaj2001.1853, 2001.
- Hedges, J. I. and Keil, R. G.: Sedimentary organic matter preservation: an assessment and speculative synthesis, Mar.
 Chem., 49(2-3), 81–115, doi:10.1016/0304-4203(95)00008-F, 1995.
- Hendriks, I. E., Sintes, T., Bouma, T. J. and Duarte, C. M.: Experimental assessment and modeling evaluation of the effects
 of the seagrass *Posidonia oceanica* on flow and particle trapping, Mar. Ecol. Prog. Ser., 356, 163–173,
 doi:10.3354/meps07316, 2008.
- Henrichs, S. M.: Early diagenesis of organic matter in marine sediments: progress and perplexity, in Marine Chemistry,
 vol. 39, pp. 119–149., 1992.
- Holmer, M.: The effect of oxygen depletion on anaerobic organic matter degradation in marine sediments, Estuar. Coast.
 Shelf Sci., 48(3), 383–390, doi:10.1006/ecss.1998.0424, 1999.
- Holmer, M. and Laursen, L.: Effect of shading of *Zostera marina* (eelgrass) on sulfur cycling in sediments with
 contrasting organic matter and sulfide pools, J. Exp. Mar. Bio. Ecol., 270, 25–37, doi:10.1016/S0022-0981(02)00015-1,
 2002.
- Holmer, M., Carta, C. and Andersen, F.: Biogeochemical implications for phosphorus cycling in sandy and muddy
 rhizosphere sediments of *Zostera marina* meadows (Denmark), Mar. Ecol. Prog. Ser., 320, 141–151,
 doi:10.3354/meps320141, 2006.
- Idestam-Almquist, J.: Dynamics of submersed aquatic vegetation on shallow soft bottoms in the Baltic Sea, J. Veg. Sci.,
 11(3), 425-432, doi:10.2307/3236635, 2000.
- Jephson, T., Nyström, P., Moksnes, P. O. and Baden, S. P.: Trophic interactions in *Zostera marina* beds along the Swedish
 coast, Mar. Ecol. Prog. Ser., 369, 63–76, doi:10.3354/meps07646, 2008.
- Jerling, L., Lindhe., A.; Mjukbottenvegetation vid Trosa skärgård, Three-year project (MSc thesis), Dept of. Botany,
 Stockholm University, 1976.
- van Katwijk, M. M., Bos, A. R., Hermus, D. C. R. and Suykerbuyk, W.: Sediment modification by seagrass beds:
 muddification and sandification induced by plant cover and environmental conditions, Estuar. Coast. Shelf Sci., 89, 175–
 181, doi:10.1016/j.ecss.2010.06.008, 2010.

- Keil, R., Montluçon, D., Prahl, F. and Hedges, J.: Sorptive preservation of labile organic matter in marine sediments,
 Nature, 370(JANUARY 1994), 549–552, doi:10.1038/370549a0, 1994.
- Kelleway, J. J., Saintilan, N., Macreadie, P. I. and Ralph, P. J.: Sedimentary factors are key predictors of carbon storage in
 SE Australian saltmarshes, Ecosystems, (C), doi:10.1007/s10021-016-9972-3, 2016.

Kennedy, H., Beggins, J., Duarte, C. M., Fourqurean, J. W., Holmer, M., Marbà, N. and Middelburg, J. J.: Seagrass sediments
as a global carbon sink: isotopic constraints, Global Biogeochem. Cycles, 24(4), GB4026, doi:10.1029/2010GB003848,
2010.

- Kenworthy, W. J. and Thayer, G. W.: Production and decomposition of the roots and rhizomes of seagrasses, *Zostera marina* and *Thalassia testudinum*, in temperate and subtropical marine ecosystems, Bull. Mar. Sci., 35(3), 364–379, 1984.
- Kenworthy, W. J., Gallegos, C. L., Costello, C., Field, D. and di Carlo, G.: Dependence of eelgrass (*Zostera marina*) light
 requirements on sediment organic matter in Massachusetts coastal bays: Implications for remediation and restoration,
 Mar. Pollut. Bull., 83(2), 446–457, doi:10.1016/j.marpolbul.2013.11.006, 2014.
- Klap, V. A., Hemminga, M. A. and Boon, J. J.: Retention of lignin in seagrasses: angiosperms that returned to the sea, Mar.
 Ecol. Prog. Ser., 194, 1–11, 2000.
- Lavery, P. S., Mateo, M.-Á., Serrano, O. and Rozaimi, M.: Variability in the carbon storage of seagrass habitats and its
 implications for global estimates of blue carbon ecosystem service., PLoS One, 8(9), e73748,
 doi:10.1371/journal.pone.0073748, 2013.
- Marbà, N., Arias-Ortiz, A., Masqué, P., Kendrick, G. A., Mazarrasa, I., Bastyan, G. R., Garcia-Orellana, J. and Duarte, C. M.:
 Impact of seagrass loss and subsequent revegetation on carbon sequestration and stocks, J. Ecol., 103, 296–302,
 doi:10.1111/1365-2745.12370, 2015.
- 478 Mayer, L. M.: Relationships between mineral surfaces and organic carbon concentrations in soils and sediments, Chem.
 479 Geol., 114, 347–363, doi:10.1016/0009-2541(94)90063-9, 1994a.
- 480 Mayer, L. M.: Surface area control of organic carbon accumulation in continental shelf sediments, Geochim. Cosmochim.
 481 Acta, 58(4), 1271–1284, doi:10.1016/0016-7037(94)90381-6, 1994b.
- Mcleod, E., Chmura, G. L., Bouillon, S., Salm, R., Björk, M., Duarte, C. M., Lovelock, C. E., Schlesinger, W. H. and Silliman, B.
 R.: A blueprint for blue carbon: toward an improved understanding of the role of vegetated coastal habitats in sequestering CO₂, Front. Ecol. Environ., 9(10), 552–560, doi:10.1890/110004, 2011.
- Moore, K. A. and Short, F. T.: *Zostera*: biology, ecology, and management, in Seagrasses: Biology, Ecology and
 Conservation, edited by A. W. D. Larkum, R. J. W. Orth, and C. M. Duarte, pp. 361–386, Springer, Dordrecht., 2003.
- 487 Olesen, B. and Sand-Jensen, K.: Biomass-density patterns in the temperate seagrass *Zostera marina*, Mar. Ecol. Prog.
 488 Ser., 109, 283–292, doi:10.3354/meps111283, 1994.

- Peralta, G., Van Duren, L. A., Morris, E. P. and Bouma, T. J.: Consequences of shoot density and stiffness for ecosystem
 engineering by benthic macrophytes in flow dominated areas: a hydrodynamic flume study, Mar. Ecol. Prog. Ser., 368,
 103–115, doi:10.3354/meps07574, 2008.
- 492 Phillips, R. C. and Meñez, E. G.: Seagrasses, Smithsonian Contributions To the Marina Sciences, nr 34, Washington DC,
 493 USA., 1988.
- Pollard, P. C. and Moriarty, D. J. W.: Organic carbon decomposition, primary and bacterial productivity, and sulphate
 reduction, in tropical seagrass beds of the Gulf of Carpentaria, Australia, Mar. Ecol. Prog. Ser., 69(1-2), 149–159,
 doi:10.3354/meps069149, 1991.
- 497 Ricart, A. M., York, P. H., Rasheed, M. A., Pérez, M., Romero, J., Bryant, C. V. and Macreadie, P. I.: Variability of sedimentary 498 Pollut. organic carbon in patchy seagrass landscapes. Mar. Bull. 100(1).476-482. 499 doi:10.1016/j.marpolbul.2015.09.032, 2015.
- Rozaimi, M., Serrano, O. and Lavery, P. S.: Comparison of carbon stores by two morphologically different seagrasses, J.
 R. Soc. West. Aust., 96(October), 81–83, 2013.
- Rueda, J. L. and Salas, C.: Molluscs associated with a subtidal *Zostera marina* L. bed in southern Spain: Linking seasonal
 changes of fauna and environmental variables, Estuar. Coast. Shelf Sci., 79(1), 157–167,
 doi:10.1016/j.ecss.2008.03.018, 2008.
- Salo, T., Pedersen, M. F. and Boström, C.: Population specific salinity tolerance in eelgrass (*Zostera marina*), J. Exp. Mar.
 Bio. Ecol., 461, 425–429, doi:10.1016/j.jembe.2014.09.010, 2014.
- Samper-Villarreal, J., Lovelock, C. E., Saunders, M. I., Roelfsema, C. and Mumby, P. J.: Organic carbon in seagrass
 sediments is influenced by seagrass canopy complexity, turbidity, wave height, and water depth, Limnol. Oceanogr.,
 n/a-n/a, doi:10.1002/lno.10262, 2016.
- Serrano, O., Mateo, M. A., Renom, P. and Julià, R.: Characterization of soils beneath a *Posidonia oceanica* meadow,
 Geoderma, 185-186, 26-36.
- 512 Serrano, O., Lavery, P. S., Rozaimi, M. and Mateo, M. Á.: Influence of water depth on the carbon sequestration capacity
 513 of seagrasses, Global Biogeochem. Cycles, 28, 950–961, doi:10.1002/2014GB004872.Received, 2014.
- Serrano, O., Ricart, A. M., Lavery, P. S., Mateo, M. A., Arias-Ortiz, A., Masque, P., Steven, A., and Duarte, C. M.; Key
 biogeochemical factors affecting soil carbon storage in *Posidonia* meadows. Biogeosciences Discuss., 12, 18913-18944,
 2015.
- Serrano, O., Lavery, P. S., Duarte, C. M., Kendrick, G. A., Calafat, A., York, P., Steven, A. and Macreadie, P.: Can mud (silt and clay) concentration be used to predict soil organic carbon content within seagrass ecosystems?, Biogeosciences Discuss., (January), 1–24, doi:10.5194/bg-2015-598, 2016.
- Short, F. T., Carruthers, T., Dennison, W. and Waycott, M.: Global seagrass distribution and diversity: A bioregional
 model, J. Exp. Mar. Bio. Ecol., 350(1-2), 3–20, doi:10.1016/j.jembe.2007.06.012, 2007.

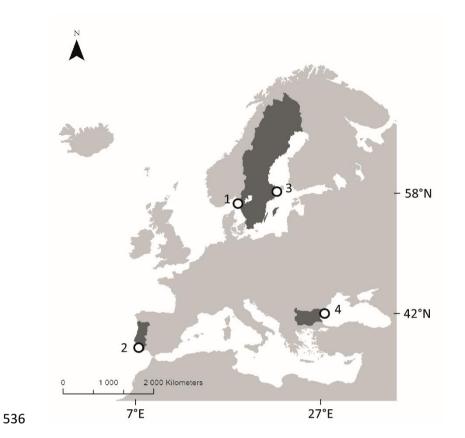
Suykerbuyk, W., Bouma, T. J., Govers, L. L., Giesen, K., de Jong, D. J., Herman, P., Hendriks, J. and van Katwijk, M. M.:
Surviving in changing seascapes: sediment dynamics as bottleneck for long-term seagrass presence, Ecosystems, doi:10.1007/s10021-015-9932-3, 2015.

Terrados, J., Duarte, C. M., Kamp-Nielsen, L., Agawin, N. S. R., Gacia, E., Lacap, D., Fortes, M. D., Borum, J., Lubanski, M.
and Greve, T.: Are seagrass growth and survival constrained by the reducing conditions of the sediment?, Aquat. Bot.,
65(1-4), 175–197, doi:10.1016/S0304-3770(99)00039-X, 1999.

Waycott, M., Duarte, C. M., Carruthers, T. J. B., Orth, R. J., Dennison, W. C., Olyarnik, S., Calladine, A., Fourqurean, J. W.,
Heck, K. L., Hughes, A. R., Kendrick, G. A., Kenworthy, W. J., Short, F. T. and Williams, S. L.: Accelerating loss of seagrasses
across the globe threatens coastal ecosystems., Proc. Natl. Acad. Sci. U. S. A., 106(30), 12377–12381,
doi:10.1073/pnas.0905620106, 2009.

Wilson, A. M., Huettel, M. and Klein, S.: Grain size and depositional environment as predictors of permeability in coastal
marine sands, Estuar. Coast. Shelf Sci., 80(1), 193–199, doi:10.1016/j.ecss.2008.06.011, 2008.

Wold, S., Sjöström, M. and Eriksson, L.: PLS-regression: A basic tool of chemometrics, Chemom. Intell. Lab. Syst., 58(2),
109–130, doi:10.1016/S0169-7439(01)00155-1, 2001.



537 Figure 1. The four study regions, Gullmar Fjord (Skagerrak, Sweden) (1), Ria Formosa (gulf of Cádiz, Portugal) (2), Askö

538 (Baltic Sea, Sweden) (3) and Sozopol (Black Sea, Bulgaria) (4).

Area	Site	Vegetation	Coordinates	Mean depth (m)
Gullmar Fj	ord (Skagerrak, Sweden)			
	Finnsbo (F)	Z marina	58°17'55N, 11°29'34E	2.8
	Kristineberg (K)	Z. marina	58°14'53N, 11°26'51E	3.0
	Rödberget (Rö) (r)	Unvegetated	58°15'06N, 11°27'54E	2.5
Ria Formos	sa (gulf of Cádiz, Portugal) ¹			
	Culatra channel (C)	Z. marina/ C. nodosa	37°00'14N 7°49'36W	1.9
	Ilha da Culatra (I)	Z. marina	36°59'50N, 7°49'41W	1.0
	Culatra channel (Cr) (r)	Unvegetated	37°00'15N, 7°49'33W	2.6
	Ilha da Culatra (Ir) (r)	Unvegetated	36°59'51N, 7°49'40W	1.8
Askö (Balt	ic Sea, Sweden)			
	Torö (T)	Z. marina/R. maitima	58°48'14N, 17°47'32E	3.2
	Långskär (L)	Z. marina/R. maitima	58°48'00N, 17°40'48E	2.2
	Storsand (S)	Z. marina	58°48'26N, 17°41'40E	3.8
	Torö (Tr) (r)	Unvegetated	58°48'21N, 17°47'31E	6
	Godahoppsudden (Gh) (r)	Unvegetated	58°48'09N, 17°42'24E	2.9
Sozopol (B	Black Sea, Bulgaria)			
1	Ropotamo (Rt)	Z. marina/ Z. noltii	42°19'49N, 27°45'20E	2.7
	Gradina (G)	Z. marina/ Z. noltii	42°25'39N, 27°39'05E	4.2
	Bay of Sozopol (r)	Unvegetated	42°24'42N, 27°39'48E	5.7

r = reference site (unvegetated area) ¹Depth values standardized to mean low water (MLW).

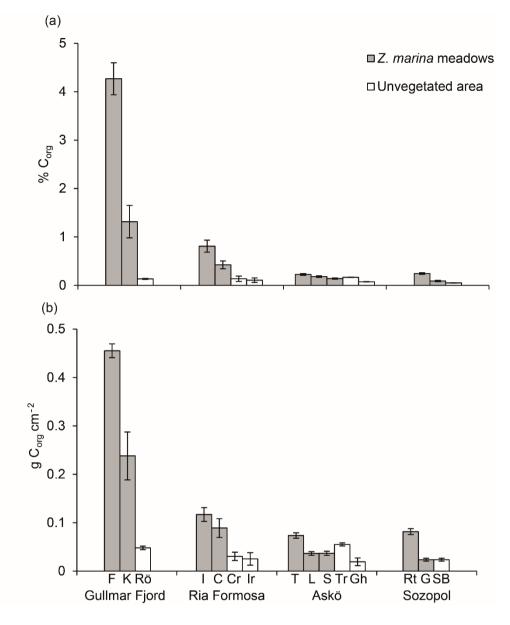


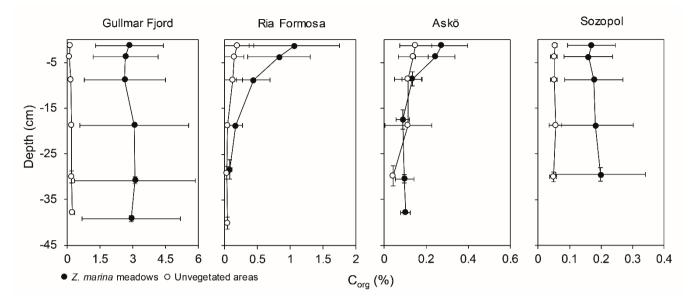


Figure 2. Mean (±SE) % C_{org} (a) and g C_{org} cm⁻² (b) in sediment (for 0-25 cm sediment depth). The percent organic carbon
(% C_{org}) is presented as a mean of the content of the top 25 cm sediment, while carbon per unit area (g C_{org} cm⁻²) is the
total (accumulated) amount of carbon in the top 25 cm of sediment. For full names of the sites see Table 1.

- 550 Table 2. Summary of nested general linear mixed model ANOVAs for sediment carbon content and sediment grain size
- 551 (% C_{org}, g C_{org} cm⁻² and sediment grain size particles <0.074 mm). The factor Habitat is comparing Z. marina meadows and
- 552 unvegetated areas. Bold values indicates significant values (P<0.05).

Source of variation			% C _{org}		g	C _{org} cm	-2	Grain Siz	ze (<0.0	74 mm)
	df	MS	F	р	MS	F	р	MS	F	р
-										
Area	3	1.0734	81.00	<0.001	0.000	98.36	< 0.001	4.6387	78.32	<0.001
Habitat (Area)	4	1.1603	87.55	<0.001	0.000	112.23	< 0.001	4.7868	80.82	<0.001
Core	5	0.0188	1.42	0.218	0.000	2.30	0.045	0.0415	0.70	0.498
Sediment depth (Core)	24	0.0107	0.81	0.724	0.000	1.51	0.059	0.0677	1.14	0.328
Residual	378	0.0133			0.000			0.0592		

554



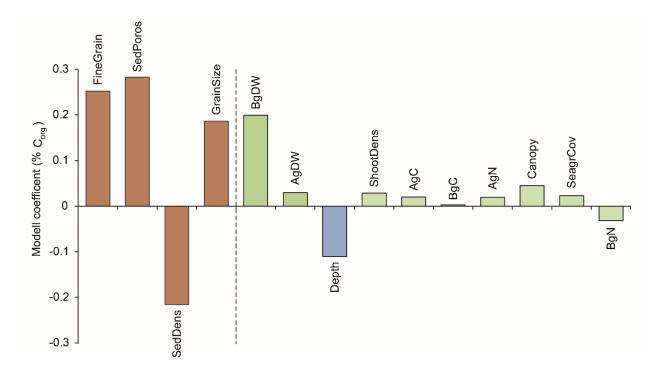
557 Figure 3. Mean sedimentary carbon (% Corg ± SD) depth profiles grouped for the different regions showed as mean slice depth. Note that the scale on the x-axes differs among the different depth profiles due to large variation in carbon content among areas.

	Aboveground bioma	2		Aboveground biomass	d biomass			Belowground biomass	d biomass		
Areas	(m ⁻²) height (cm	0	cover (%)	% C	% N	C:N	g DW m ⁻²	% C	% N	C:N	g DW m ⁻²
Gullmar Fjord	Gullmar Fjord 157.9 ± 43.8 81.4 ± 18.2 36.9 ± 14.0 38.8 ± 0.7 2.1 ± 0.3 18.4 ± 2.0	81.4 ± 18.2	36.9 ± 14.0	38.8±0.7 2.	1±0.3 1		39.4 ± 31.1	34.2 ± 1.4	1.1 ± 0.1	30.2 ± 2.8	$\pm 1.4 1.1 \pm 0.1 30.2 \pm 2.8 253.0 \pm 86.0$
Ria Formosa	$264.9 \pm 107.8 32.5 \pm 4.4 79.1 \pm 10.6 34.4 \pm 1.2 1.4 \pm 0.1 25.2 \pm 1.2$	32.5 ± 4.4	79.1 ± 10.6	34.4 ± 1.2 1.	4±0.1 2		108.3 ± 58.5	30.8 ± 3.2	1.1 ± 0.1	1.1±0.1 37.8±4.5	494.2 ± 230.2
Askö	338.1 ± 160.3	51.7 ± 12.4	47.6 ± 18.1	37.2 ± 2.0 1.	8±0.3 2	20.9 ± 2.9	$338.1 \pm 160.3 \hspace{.1in} 51.7 \pm 12.4 \hspace{.1in} 47.6 \pm 18.1 \hspace{.1in} 37.2 \pm 2.0 \hspace{.1in} 1.8 \pm 0.3 \hspace{.1in} 20.9 \pm 2.9 \hspace{.1in} 255.7 \pm 193.4$	32.8 ± 2.9	1.1 ± 0.1	1.1 ± 0.1 31.5 ± 3.2	205.9 ± 88.7
Sozopol	$419.6 \pm 315.3 \ \ 63.5 \pm 11.2 \ \ 63.5 \pm 11.1 \ \ 36.0 \pm 1.3 \ \ 1.9 \pm 0.3$	63.5 ± 11.2	63.5 ± 11.1	36.0±1.3 1.	9±0.3 1	19.6 ± 3.2	122.0 ± 110.8	30.3 ± 3.0	$\pm 3.0 0.8 \pm 0.2$	28.6 ± 5.7	86.4±80.6
8											

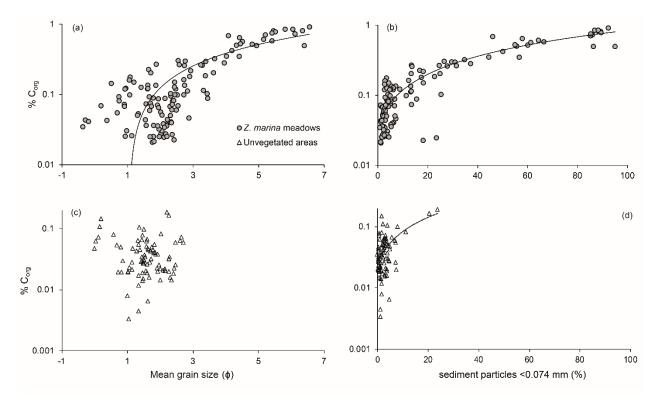
Table 3. Seagrass meadow variables (mean ± SD) for the different areas.

Areas	% Corg	g C _{org} cm ⁻²	Sediment porosity (%)	Sediment density (g DW mL ⁻¹)	%N	C:N	Mean grain size (ф)	Sediment particles <0.074 mm (%)
Gullmar Fjord	2.79 ± 0.5	0.35 ± 0.041	67.0 ± 14.1	0.71 ± 0.33	0.28 ± 0.16	9.39 ± 1.26	4.89 ± 0.93	62.8 ± 25.6
Ria Formosa	0.61 ± 0.09	0.10 ± 0.012	43.0 ± 5.4	1.13 ± 0.14	0.08 ± 0.01	7.15 ± 0.83	2.34 ± 0.56	17.9 ± 5.8
Askö	0.18 ± 0.01	0.05 ± 0.005	31.9 ± 2.4	1.4 ± 0.15	0.03 ± 0.01	5.60 ± 1.27	1.19 ± 0.79	3.7 ± 0.6
Sozopol	0.17 ± 0.02	0.05 ± 0.009	41.8 ± 5.2	1.25 ± 0.04	0.05 ± 0.04	3.22 ± 1.25	2.08 ± 0.27	2.6 ± 1.9

in the different areas. Mean grain size is presented with phi (ϕ) units. Table 4. Seagrass sediment data as mean (± SD for all variables except carbon content, which is presented with ± SE) for the depth profile (0-25 cm)



564 Figure 4. PLS (Partial Least Square) regression model coefficient plot for % Corg in sediment (using a mean of the carbon 565 content for the top 25 cm sediment). The predictor variables are ranked in level of importance (left to right) where the four 566 variables left of the striped bar having a VIP-value >1 (i.e. FineGrain, SedPoros, SedDens and GrainSize) and hence 567 significantly influencing % Corg. Brown = sediment characteristics, green = seagrass-associated variables and blue = water 568 depth; Variables included in the model were FineGrain (sediment particles <0.074 mm, %), SedPoros (sediment porosity, 569 %), SedDens (sediment density, g DW mL⁻¹), GrainSize (mean grain size, ϕ), Bg and Ag DW (belowground [roots and 570 rhizomes] and aboveground [shoots] biomass dry weight, m²), Depth (water depth, m), ShootDens (shoot density, m²), Ag 571 and Bg biomass C and N (biomass carbon and nitrogen content, %), Canopy (shoot height, cm) and SeagrCov (seagrass 572 cover, %).



574

Figure 5. Semi-log plots (log₁₀[x+1]) showing the relationship between % C_{org} and grain size. The % C_{org} is presented with a log scale as it gave the best fit of the models. Grain size is shown as mean grain size (ϕ) and sediment particles <0.074 mm (%) for *Z. marina* meadows (a and b) and unvegetated areas (c and d). The % C_{org} was positively linked to both sediment particles <0.074 mm (%) (R² = 0.91, *P* < 0.001) and mean grain size (ϕ) (R² = 0.74, *P* < 0.001) for *Z. marina* meadows but for unvegetated area only sediment particles < 0.074 mm (%) showed this relationship with % C_{org} (R² = 0.42, *P* < 0.001).

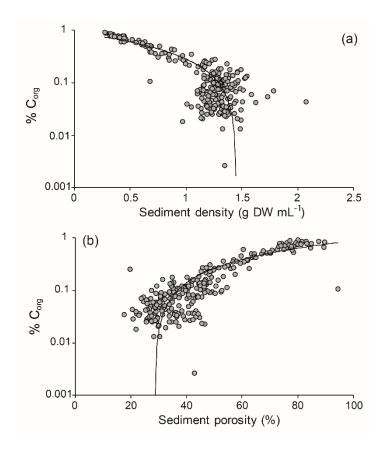
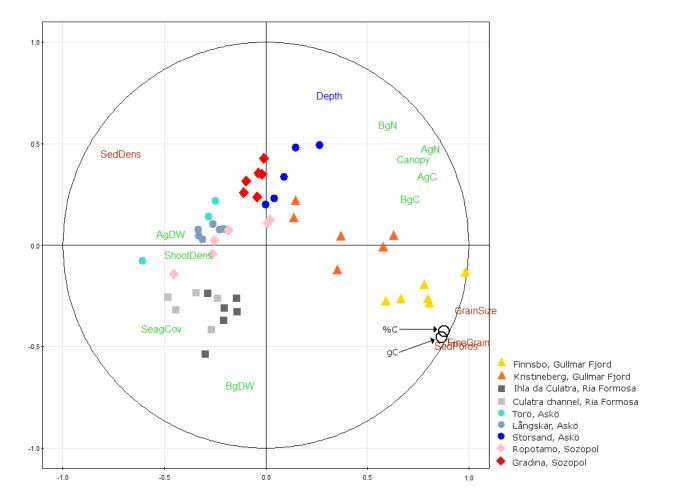




Figure 6. Semi-log plots (log₁₀[x+1]) for sediment density (a) and sediment porosity (b) in relation to % C_{org} for the *Z. marina* sites. The sediment density (g DW mL⁻¹) was negatively influencing the amount of organic carbon ($R^2 = 0.84$, *P* < 0.001), while there was a positive relation between sediment porosity (%) and % C_{org} ($R^2 = 0.80$, *P* < 0.001).



586

Figure 7. PCA (Principal Component Analysis) showing the nine seagrass sites, the two response variables (sedimentary % C_{org} and g C_{org} cm⁻²) and predictor variables (14 in total). The percent organic carbon (% C_{org}) is presented as a mean of the content of the top 25 cm sediment, while carbon per unit area (g C_{org} cm⁻²) is the total (accumulated) amount of carbon in the top 25 cm of sediment. The colors of the letters represent different groups of predictor variables; brown = sediment characteristics, green = seagrass-associated variables, blue = water depth. Black circles are the response variables, i.e. organic carbon (%C = % C_{org} and gC = g C_{org} cm⁻²). For explanations to the abbreviations of predictor variables see Fig. 4.

^aDichromate titration method (Gaudette et al. 1974) ¹Number of meadows for each area and sediment core depth. *Studies presenting % OM if both % C_{org} and % OM are included on the same row.

 $^+$ Converted values (partly or all) from % OM to % C_{org} (conversion factor: 0.43)

		Longitude	Sediment core	C _{orz} (%) ±	OM (%) ±	Water depth	2	1
Area	Countries	(N)	depth (cm)	SD	SD	(m) ± SD	Sites ¹	References
Norwegian Sea	Norway	63°21'	0-10	0.49ª		8.0	1	Fredriksen et al., 2010
Skagerrak	Sweden	57°48'- 59°00'	0-5	$5.67 \pm 3.92^{+}$	25.2*	2.0 ± 0.4	50	Jephsen et al., 2009*, Gullström <i>e</i> t al. 2012
	Sweden	58°14'- 58°17'	0-5	2.74 ± 1.49		2.9 ± 0.9	2	This study
	Norway		0-10	0.68±0.38ª		4.0	З	Fredriksen et al., 2010
Kattegat	Denmark	54°58'- 56°49'	0-5	$1.68\pm2.05^{+}$	$4.13\pm5.14^*$	2.7 ± 0.10	11	Boschker et al., 2000, Röhr et al., 2016*
			11-15	$0.54 \pm 0.60^{+}$	1.26 ± 1.41	2.3 ± 2.0	6	Fredriksen et al.,2006, Holmer & Laursen 2002, Holmer et al. 2006
Baltic Sea	Sweden, Finland	55°23'- 60°21'	0-5	$0.46\pm0.30^{+}$	1.07 ± 0.69	2.3 ± 0.7	13	Jerling and Lindhe, 1976, Jephsen et al., 2009, Röhr et al., 2016
	Sweden	58°49'	0-5	0.26 ± 0.11		3.8 ± 1.1	3	This study
	Sweden, Finland	55°23'- 60°21'	8-10	$0.36\pm0.30^{+}$	0.86 ± 0.70	3.3 ± 0.9	14	Boström et al., 2006
North Sea	Netherlands	51°34'- 53°25'	0-5	0.73		Intertidal	1	Boschker et al., 2006
			0-20	$0.90\pm0.60^{\ast}$	2.10 ± 1.40		ы	van Katwijk et al., 2010
Black sea	Bulgaria	42°19'- 42°22'	0-5	$\textbf{0.16} \pm \textbf{0.07}$		4.2 ± 1.6	2	This study
North Atlantic Ocean	France, USA	34°43'- 44°42'	0 -5	$1.92 \pm 2.15^{+}$	4.59±5.60*	2.1 ± 0.9	9	Foneseca et al., 1984*, Boschker et al., 2006, Kenworthy et al., 2014*, Dale 1974
	USA		11-21	2.08		Intertidal	1	Dale 1974
	Portugal	36°59'- 37°00'	0-5	0.94 ± 0.59		1.3 ± 1.0	2	This study
Mediterranean	Spain	36°44'	0-5	0.90+	2.30	13	1	Rueda and Salas 2008

dichromate titration was used (Gaudette et al. 1974).

sediment with <0.2 %0M. All studies have determined % 0M to % Corg by LOI (Loss on ignition) or using a CN elemental analyzer except a where

were only % OM was presented a conversion factor of 0.43 was used to convert % OM to % Corg as calculated by Fourqurean et al. (2012) for seagrass

Table 5. Summary of literature data on organic carbon (% Corg) and % OM (organic matter) content in seagrass sediment in Europe and USA. In studies

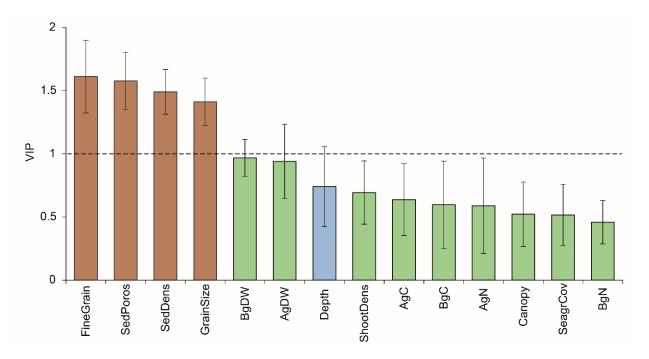


Figure S1. VIP-values (variance of importance) for independent variables used in the PLS model testing relationships to
carbon content (mean % C for the top 25 cm sediment). The variables are listed in the level of importance and those with
VIP-values >1 (dashed line) has a significant influence on the model. Brown = sediment characterisitcs, green = seagrassassociated variables and blue = water depth. FineGrain (sediment particles <0.074 mm, %), SedPoros (sediment porosity,
%), SedDens (sediment density, g DW mL⁻¹), GrainSize (mean grain size, \$\phi\$), Bg and Ag DW (belowground biomass dry
weight, m²), Depth (water depth, m), ShootDens (shoot density, m²), Ag and Bg biomass C and N (biomass carbon and
nitrogen content, %), Canopy (shoot height, cm), SeagrCov (seagrass cover, %) were used as predictor variables.

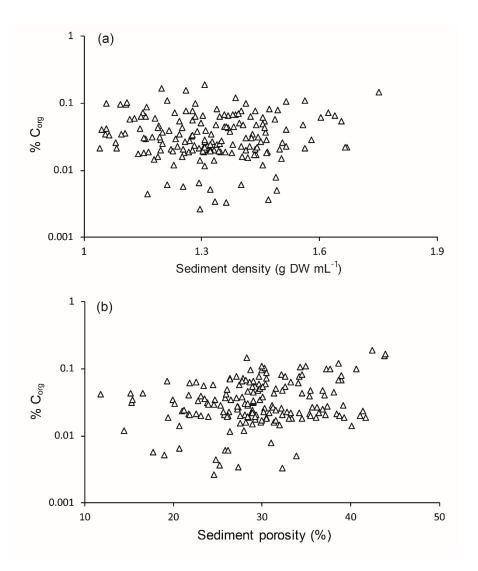


Figure S2. Semi-log plots ($log_{10}[x+1]$) for sediment density (g DW mL⁻¹) (a), and sediment porosity (%) (b) in relation to organic carbon content (% C_{org}) for unvegetated areas. There was no significant relationship between sediment density and organic carbon. The sediment porosity was, however, positively linked to sedimentary organic carbon but had a low R²-value (linear regression, R² = 0.08, *P* < 0.001).