



1 **Provenance of branched GDGTs in the Tagus River drainage basin and its**
2 **outflow in the Atlantic Ocean over the Holocene**

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4 *Lisa Warden^a, Jung-Hyun Kim^{a,b}, Claudia Zell^{a,c}, Geert-Jan Vis^d, Henko de Stigter^e, Jérôme Bonnin^f, and Jaap S.*
5 *Sinninghe Damsté^{a,g}*

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7 ^a*NIOZ Netherlands Institute for Sea Research, Department of Marine Microbiology and*
8 *Biogeochemistry, and Utrecht University, P.O. Box 59, 1790 AB Den Burg, the Netherlan*

9 ^b*Department of Marine Science and Convergence Technology, Hanyang University ERICA campus,*

10 *55 Hanyangdaehak-ro, Sangnok-gu, Ansan-si, Gyeonggi-do 426-791, South Korea*

11 ^c*Pierre and Marie Curie University - Paris 6 · METIS - UMR 7619 France*

12 ^d*TNO, Geological Survey of the Netherlands, P.O. Box 80015, 3508 TA Utrecht, the Netherlands*

13 ^e*NIOZ Netherlands Institute for Sea Research, Department of Ocean Systems Sciences, and Utrecht*

14 *University, PO Box 59, 1790 AB Den Burg, the Netherlands*

15 ^f*UMR-EPOC 5805 CNRS, Université de Bordeaux, Allée Geoffroy St Hilaire, 33615 Pessac, France*

16 ^g*Utrecht University, Faculty of Geosciences, PO Box 80021, 3508 TA Utrecht, the Netherlands*

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18 *Correspondence to: Lisa Warden (lisa.warden@nioz.nl)*

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23 **Keywords:** Tagus River basin, Holocene, temperature reconstructions, branched glycerol

24 dialkyl glycerol tetraethers, paleoclimate



25 **Abstract**

26 The distributions of branched glycerol dialkyl glycerol tetraethers (brGDGTs), which
27 are transported from the soils where they are predominantly produced to marine sediments via
28 rivers, have been applied in reconstructing mean annual air temperatures (MAT) and pH of
29 soils. However, paleoclimate reconstructions using sedimentary brGDGTs have proven
30 difficult in arid regions including the Iberian Peninsula. Recently, six novel 6-methyl
31 brGDGTs have been described using new analytical methods (in addition to the nine 5-methyl
32 brGDGTs previously used for climate reconstructions), and so new pH and MAT calibrations
33 have been developed that in a set of global soil samples were shown to improve the accuracy
34 of reconstructions, especially in arid regions. Because of this we decided to apply the new
35 method to separate the 5- and 6-methyl isomers along with the novel calibrations to a sample
36 set that runs in a transect from source to sink along the Tagus River and out to the deep ocean
37 off the Portuguese margin and spans the last 6,000 years in order to determine if it improves
38 paleoclimate reconstructions in this area. We found that although pH reconstructions in the
39 soils were improved using the new calibration, MAT reconstructions were not much better
40 even with the separation of the 5- and 6-methyl brGDGTs. This confirmed the conclusion of
41 previous studies that the amount of aquatically produced brGDGTs is overwhelming the soil
42 derived ones in marine sediments and complicating MAT reconstructions in the region.
43 Additionally, the new separation revealed a strong relationship between the new degree of
44 cyclization (DC') of the brGDGTs and MAT not seen before that could be making
45 temperature reconstructions in this and other arid regions difficult.

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50 **1. Introduction**

51 Understanding past climate variability is important for predicting future climate change
52 as well as how ecosystems, organisms and human society could be affected. The validation of
53 climate proxies is imperative for the correct interpretation of climate archives and therefore
54 also for the climate models building on these past climate data. Terrestrial environments play
55 an important role in global climate, however, continental climate reconstructions are hindered
56 by the lack of continental temperature proxies. In the future, changes in terrestrial climate are
57 likely to have a large impact on human society just as they had in the past (e.g. Haug et al.,
58 2003). Availability of trustworthy temperature data from the terrestrial environment will be
59 essential for the development of reliable climate models.

60 The distribution of branched glycerol dialkyl glycerol tetraethers (brGDGTs, Fig. S1), a
61 group of membrane-spanning lipids that occur in heterotrophic bacteria (Pancost and
62 Sinninghe Damsté, 2003; Weijers et al., 2010) pervasive in peat (Weijers et al., 2006) and
63 worldwide in soils has proven useful as a tool to obtain high resolution, continental
64 temperature reconstructions (Weijers et al., 2007a; Schouten et al., 2008; Bendle et al., 2010).
65 BrGDGTs are biosynthesized by bacteria (Sinninghe Damsté et al., 2011; 2014) living in soils
66 and the distribution of brGDGTs in soils is affected by growth temperature and pH (Weijers et
67 al., 2006). More specifically, the degree of methylation of the brGDGTs (expressed as
68 methylation of branched tetraethers index, MBT) relates to mean annual air temperature
69 (MAT), and to a lesser extent soil pH, whereas the degree of cyclization (DC) of the
70 brGDGTs (also expressed as the cyclization of branched tetraethers index, CBT) correlates
71 solely with soil pH (Weijers et al., 2007a). MBT has recently been amended to become MBT'
72 by eliminating the brGDGTs that rarely occur in soils (Peterse et al., 2012). These
73 observations led to the development of a continental paleoclimate proxy based on the
74 distribution of brGDGTs that has been applied in paleosoils (Peterse et al., 2009; Weijers et



75 al., 2007a; 2011). Branched GDGTs that are produced in soils are washed by runoff into
76 streams and rivers where they are transported to and deposited in river sediment and in coastal
77 marine sediment that are under the influence of major river systems. In this way, brGDGTs
78 have been used as recorders of the continental paleoclimate (Weijers et al., 2007b; Bendle et
79 al., 2010; Hren et al., 2010; Keating-Bitonti et al., 2011).

80 Complications using brGDGTs as a proxy for MAT have arisen in some settings. In
81 marine sediments receiving a low input of soil organic matter (OM), it was found that the
82 distribution of brGDGTs and the reconstructed temperatures were quite different from that
83 observed in regional soils (Peterse et al., 2009). Peterse et al. (2012) found in arid regions
84 temperature is no longer an important control on the distribution of brGDGTs and therefore
85 MAT reconstructions in these areas should be interpreted with care. In the Iberian Peninsula,
86 Menges et al. (2013) found that MBT' was not correlated to MAT but instead correlated with
87 the aridity index (AI), a parameter for water availability in soils, and mean annual
88 precipitation (MAP). In drainage basins with varying soil sources that had different MATs
89 (i.e. mountainous vs. lowland), it was found that the provenance of the soil matter must be
90 considered when interpreting MAT reconstructions (Bendle et al., 2010). In-situ production of
91 brGDGTs can occur within the river systems (Yang et al., 2012; Zell et al., 2013, De Jonge et
92 al., 2014b) and cause brGDGT distributions and MAT reconstructions that differ from those
93 in the soils of the source area. These complications make it vital to investigate how varying
94 environmental conditions, the transport of these terrestrially derived fossilized lipids, and in-
95 situ production affect the implementation of brGDGTs for paleoclimate reconstructions.

96 Recently a set of six new brGDGT isomers that differ in the position of the methyl
97 groups were identified and described (De Jonge et al., 2013). The relative abundance of these
98 novel, 6-methyl brGDGTs are strongly dependent on pH and so by excluding them from the
99 MBT' index (newly defined as MBT'_{5ME}) the correlation with MAT is improved (De Jonge et



100 al., 2014a). The CBT index was also redefined in this study, as CBT', to include all of the pH
101 dependent 6-methyl brGDGTs and consequently yielded a higher correlation with soil pH as a
102 result (De Jonge et al., 2014a). De Jonge et al. (2014a) also developed, based on a dataset of
103 globally distributed soils, a new pH calibration taking into account the new CBT' as well as
104 new MAT calibrations, defined as MAT_{mr} and MAT_{mrs}. In a global soil set they were shown
105 to improve the accuracy of reconstructions, especially in arid regions. These indices and
106 calibrations were applied in a coastal sediment core in the Northern Kara Sea off Siberia in a
107 study emphasizing the importance of examining the provenance of brGDGTs when using
108 these lipids for paleoclimate reconstructions (De Jonge et al., 2015).

109 A comprehensive study has been previously performed on the present day transport of
110 brGDGTs in the Tagus River basin from source to sink (Zell et al., 2014). The results from
111 this study demonstrated that the distribution of brGDGTs in the riverine suspended particulate
112 matter (SPM) did not reflect that of the soils, implying that due to the aquatic production in
113 river and marine environments the use of brGDGTs for paleoclimate reconstructions in the
114 region would be complicated (Zell et al., 2014; 2015). Here we examine if the assessment of
115 the provenance of brGDGTs in the Tagus River basin can be improved by the application of
116 the analytical methods allowing the separation of the 5- and 6-methyl brGDGTs (De Jonge et
117 al., 2013). In addition, we examine if the provenance of brGDGTs changed over the Holocene
118 and if the distribution of brGDGTs in the past reflected continental sources and thus past
119 temperature and pH of the soils in the drainage basin of the river. To this end we compare the
120 down core brGDGT distributions in Holocene sediments retrieved from four locations along a
121 transect in the Tagus River basin, which includes the river floodplain (Tagus River Floodplain
122 core), the offshore mudbelt (Mudbelt core), and marine sediments from the canyons (Lisbon
123 Canyon Head core and Lower-Sétubal canyon core) (Fig. 1), and compare them to brGDGT
124 distributions of soil and river SPM from the Tagus River watershed. This allows insight into



125 the potential and limitations of using the novel MAT_{mrs}/CBT' proxies for climate
126 reconstruction in this region and in river systems in general.

127

128 1. Study Area

129 The Tagus River drains the central part of the Spanish Plateau with an E-W orientation
130 (Benito et al., 2003). The waters originate at an elevation of about 1600 m altitude in eastern
131 Spain at the Iberian Range and the mouth of the river feeds into the Atlantic Ocean near
132 Lisbon (Vis and Kasse, 2009). At 1,200 km long the Tagus River is the longest river of the
133 Iberian Peninsula and it occupies 82×10^3 km² making it the third largest in catchment area
134 (Benito et al., 2003). The Tagus Basin is surrounded by mountains on three sides with the
135 Iberian Range to the east, the Central Range to the north, and the Toledo Mountains to the
136 south. Present-day mean discharge at the Tagus River mouth is $400 \text{ m}^3 \text{ s}^{-1}$ (Vale and Catarino,
137 1996; Vaz et al., 2011) and the largest contribution of draining tributaries comes from the
138 Central Range in the North (Benito et al., 2003). The Tagus River is characterized by extreme
139 seasonal and annual variability, including periods of flooding with 30 times the mean
140 discharge and an annual discharge cycle characterized by two peaks in the winter (December
141 and then again February to March) and a discharge minimum in the summer (August) (Benito
142 et al., 2003). Since the 1940s dams have been built along the expanse of the Tagus River for
143 water supply, hydropower, and flood prevention (Dias et al., 2002), which have likely
144 impacted the transport of brGDGTs in the Tagus River system since their construction.

145 Where the Tagus River debouches into the Atlantic Ocean, the narrow continental shelf
146 and steep continental slope are deeply incised by the Lisbon-Setúbal canyon system. The head
147 of the Lisbon branch of that canyon system is located 13 km offshore from the Tagus River
148 mouth at 120 m water depth. From that point, the canyon descends over a length of 165 km
149 until it opens out onto the Tagus Abyssal Plain at 4860 m (Lastras et al., 2009). Even though



150 the shelf is very narrow, sparse amounts of continental organic matter and clastic sediment
151 reach the deep ocean in this region (Jouanneau et al., 1998; de Stigter et al., 2011; Vis et al.,
152 in press). This is because the Lisbon-Setúbal canyon is not a very dynamic system and has a
153 weak down-canyon transport of sediments (Jouanneau et al., 1998; Jesus et al., 2010; de
154 Stigter et al., 2011). A part of the continental shelf in this region is covered by mud deposits,
155 which originate predominantly from the Tagus estuary (Jouanneau et al., 1998). According to
156 this same study, the mouth of the much smaller Sado River is located further to the southeast
157 and contributes only a relatively minor sediments volume to the shelf mud deposits.

158 Generally, the climate of the Tagus River Basin is characterized by seasonal variability
159 and is considered continental Mediterranean (Le Pera and Arribas, 2004). Summers in the
160 Tagus region are hot and dry and the winters are relatively mild and wet (Benito et al., 2003).
161 During the summers, the climate regime in the Tagus Basin is controlled by the Azores high
162 and in the winter by the westerlies (Benito et al., 2003). The MAT in the interior regions of
163 the Tagus River basin varies from the highlands to the lowlands of the inner basin from 7.5 to
164 12.5°C, respectively and can increase up to 16°C along the Atlantic Coast (Le Pera and
165 Arribas, 2004). The mean annual precipitation in the lowlands of the inner basin is mostly
166 below 500 mm making it an arid region, however, some of the highest altitudes of the
167 mountainous areas have a larger mean annual precipitation ranging from 750-1200 mm (Le
168 Pera and Arribas, 2004).

169 The Iberian Peninsula is located between two major pressure systems, the Azores High
170 and the Iceland Low, which make up the North Atlantic Oscillation (NAO). This climate
171 phenomenon is caused by the varying pressure gradient in the North Atlantic and greatly
172 influences climate conditions all over Europe (Hurrell, 1995; Hurrell and VanLoon, 1997).
173 Because of the Iberian Peninsula's advantageous position for studying the shifting NAO, the
174 climate in this region has been intensively investigated (Zorita et al., 1992; Rodó et al., 1997;



175 Trigo et al., 2004). Many of these studies are from an oceanic perspective, obtaining sea
176 surface temperatures from marine sediments using the alkenone unsaturation indices
177 (Abrantes et al., 2005, 2009; Rodrigues et al., 2009), coccolithophore assemblages (Cachao
178 and Moita, 2000; Palumbo et al., 2013), and stable isotopic oxygen composition of
179 foraminifera (Lebreiro et al., 2006 ; Bartels-Jónsdóttir et al., 2006; 2009). The terrestrial
180 climate has been examined using continental paleoarchives such as speleothems (Munoz-
181 Garcia et al., 2007; Martin-Chivelet et al., 2011; Stoll et al., 2013), tree rings (Andreu et al.,
182 2007; Linan et al., 2012), and pollen (Huntley and Prentice, 1988; Lebreiro et al., 2006; Davis
183 et al., 2003; Fletcher et al., 2007; Corella et al., 2013). The integrated continental and marine
184 approach can give complimentary information to past climate in a region and by using the
185 same proxy on the continent, in the ocean, and at the ocean-continent interface we would
186 perhaps obtain a clearer picture of continental climate processes in an area rather than using
187 separate studies or a multi-proxy approach.

188

189 2. Material and Methods

190 **Sample collection.** Soil samples, riverbank sediment samples, and river SPM from the Tagus
191 River basin (Fig. 1b) were collected previously (Zell et al., 2014). These samples were
192 complemented with four long sediment cores collected along a transect running from the
193 Tagus River to the lower continental slope (Fig. 1). The Tagus River Floodplain core
194 (0501.029) was collected in a low-energy backswamp of the present-day floodplain of the river
195 at ~4 km west of the Tagus channel (Table 1). The sediment was collected using an Edelman
196 hand auger for sediment above the groundwater table and a gauge for sediment below the
197 groundwater table (Vis et al., 2008). The sediments were wrapped in the field for laboratory
198 analyses. The other three cores were collected using a piston corer, during campaigns in May
199 2007 and March 2011 with RV *Pelagia* conducted by the NIOZ - Royal Netherlands Institute



200 for Sea Research. The coring site for the Mudbelt core (64PE332-30-2) was to the west of the
201 Tagus Estuary mouth, for the Lisbon Canyon Head core (64PE332-44-2) it was to the east of
202 the Tagus Estuary mouth, and for the Lower Setúbal Canyon core (64PE269-39) it was on the
203 crest of the northern levee of the lower Setúbal Canyon (Table 1).

204

205 **Age models.** The accelerated mass spectrometry (AMS) ^{14}C measurements of the three
206 marine sediment cores were carried out at the BETA analytic laboratory (USA) on benthic or
207 planktonic forams, gastropods or shells fragments (Table 2). As for the Tagus River
208 Floodplain core, the radiocarbon dating material was performed for a previous study and
209 consisted of mostly terrestrial botanical macrofossils, but other bulk material was used as well
210 (Vis et al., 2008). In order to establish consistent chronologies for the four sediment cores, all
211 the AMS dates were calibrated into calendar ages using the CALIB 7.0, available at
212 <http://radiocarbon.pa.qub.ac.uk/calib> (Stuiver et al., 1998). The calibration data and curve
213 selection used for the three marine sediment cores was Marine13 and for the Tagus River
214 Floodplain core IntCal13 was used (Reimer et al., 2013). All radiocarbon dates mentioned
215 have age spans at the 2σ range and are expressed as calibrated ages (cal. BP) (Table 2, Fig.
216 S2).

217

218 **Bulk isotope data** Prior to bulk carbon isotope analysis, sediment was decalcified using a 2 N
219 HCL solution for approximately 18 h. The sediment was rinsed three times using double-
220 distilled water and then freeze dried again. Total organic carbon (TOC) and $\delta^{13}\text{C}_{\text{TOC}}$ (Table 3)
221 were measured in duplicate using the Flash 2000 series Organic Elemental Analyzer (Thermo
222 Scientific) equipped with a TCD detector. The $\delta^{13}\text{C}_{\text{TOC}}$ is expressed in relation to the Vienna
223 PeeDee Belemnite (VPDB) standard and the isotope analysis precision was 0.1‰.



224

225 **Lipid extraction and GDGT analysis.** Between 1-3 g of freeze dried sediment was extracted
226 using the DionexTM accelerated solvent extraction (ASE) with dichloromethane
227 (DCM):methanol (9:1, v/v) as the solvent at a temperature of 100°C and a pressure of 1500
228 psi for 5 min with 60 % flush and purge 60 seconds. The extract was then collected and dried
229 using Caliper Turbovap®LV. Next, using DCM, the lipid extract was dried over a column of
230 anhydrous Na₂SO₄ and then blown down under a gentle stream of N₂. In order to quantify
231 GDGTs, 1 µg of an internal standard (C₄₆ GDGT; Huguet et al., 2006) was added to the total
232 lipid extract before it was separated over a column of Al₂O₃ (activated for 2 h at 150°C) into
233 three fractions using hexane:DCM (9:1, v:v) for the apolar fraction, hexane:DCM (1:1, v:v)
234 for the ketone fraction and DCM:MeOH (1:1, v:v) for the polar fraction. The polar fraction,
235 which contained the GDGTs, was dried under a N₂ stream and then re-dissolved in
236 hexane:isopropanol (99:1, v:v) at a concentration 10 mg ml⁻¹. Finally it was passed through a
237 0.45 µm PTFE filter and analysed with high performance liquid chromatography-atmospheric
238 pressure positive ion chemical ionization–mass spectrometry (HPLC-APCI-MS) with a
239 separation method that allows the separation of 5- and 6-methyl brGDGTs (Hopmans et al.,
240 2015). For the study of Zell et al., (2014) the samples were split into two different fractions
241 before the analysis, the intact polar lipid (IPL) fraction and core lipid (CL) fractions. For the
242 purposes of this study the IPL and CL fractions of the river SPM were analyzed separately on
243 the HPLC-APCI-MS for GDGTs (Hopmans et al., 2015) and then the amount of GDGTs
244 found in the CL and IPL fractions were combined. After analysis some of the GDGT based
245 indices were recalculated for the entire sample set.

246



247 **Calculation of GDGT-based proxies.** The Roman numerals refer to the GDGTs indicated in
 248 Fig. S1. The 5-methyl brGDGTs and 6-methyl brGDGTs are distinguished by an accent on
 249 the 6-methyl brGDGTs. The GDGT indicated by IV is crenarchaeol, the isoprenoid GDGT
 250 specific to Thaumarchaeota (Sinninghe Damsté et al., 2002).

251 The BIT Index (Hopmans et al., 2003) was calculated using the following formulae that
 252 specifically includes the novel 6-methyl brGDGTs according to De Jonge et al. (2015):

$$253 \text{ BIT index} = (Ia+IIa+IIIa+IIa'+IIIa')/(Ia+IIa+IIIa+IIa'+IIIa'+IV) \quad (1)$$

254 The isomer ratio (IR) signifies the quantity of the penta- and hexamethylated 6-Me brGDGTs
 255 compared to the total brGDGTs and was calculated according to De Jonge et al., (2015):

$$256 \text{ IR} = (IIa'+IIb'+IIc'+IIIa'+IIIb'+IIIc')/(IIa+IIb+IIc+IIIa+IIIb+IIIc+IIa'+IIb'+IIc'+IIIa'+IIIb'+IIIc') \quad (2)$$

258 The relative abundance of the penta- and hexamethylated 6-methyl brGDGTs are calculated
 259 according to (De Jonge et al., 2014b):

$$260 \text{ IR}_{II} = IIa'/(IIa+IIa') \quad (3)$$

$$261 \text{ IR}_{III} = IIIa'/(IIIa+IIIa') \quad (4)$$

262 The $\text{MBT}'_{5\text{Me}}$ (which excludes the 6-methyl brGDGTs) was used to calculate MAT according
 263 to De Jonge et al., (2014a):

$$264 \text{ MBT}'_{5\text{Me}} = (Ia+Ib+Ic)/(Ia+Ib+Ic+IIa+IIb+IIc+IIIa) \quad (5)$$

$$265 \text{ MAT} = -8.57 + 31.45 * \text{MBT}'_{5\text{Me}} \quad (6)$$

266 The equation to determine DC (Sinninghe Damsté et al., 2009) was reformulated to
 267 specifically include the pentamethylated 6-methyl brGDGTs:



$$268 \quad DC' = (Ib+IIb+IIb')/(Ia+Ib+IIa+IIb+IIa'+IIb') \quad (7)$$

269 To calculate pH and MAT the novel MAT_{mr} /CBT' calibration was used (De Jonge et al.,
 270 2014a):

$$271 \quad CBT' = {}^{10}\log[(Ic+IIa'+IIb'+IIc'+IIIa'+IIIb'+IIIc')/(Ia+IIa+IIIa)] \quad (8)$$

$$272 \quad pH = 7.15 + 1.59 * CBT' \quad (9)$$

$$273 \quad MAT_{mr} = 7.17 + 17.1 * [Ia] + 25.9 * [Ib] + 34.4 * [Ic] - 28.6 * [IIa] \quad (10)$$

$$274 \quad MAT_{mrs} = 5.58 + 17.91 * [Ia] - 18.77 * [IIa] \quad (11)$$

275

276 **Statistical analysis.** Using R software package for statistical analysis we performed principal
 277 component analysis (PCA) based on the correlation matrix. We performed the PCA on the
 278 fractional abundances of all 15 of the 5- and 6-methyl brGDGTs for the entire sample set
 279 along the transect from the land to the ocean.

280

281 3. Results

282 We report bulk and brGDGT data for four cores covering Holocene sedimentation in the
 283 Tagus River Basin and its outflow into the Atlantic. We compare these data with new results
 284 acquired through an improved LC method able to distinguish between the 5- and 6-methyl
 285 brGDGTs (De Jonge et al., 2013) on the soils, riverbank sediments, and SPM samples
 286 previously obtained by Zell et al., (2014).

287

288 4.1 Bulk parameters of the sediments



289 The age-depth models for the marine sediment cores (Fig. S2, Table 1) are based on
290 radiocarbon dating of picked foraminifera, gastropods, and shell fragments. The data show
291 that of the four sediment cores from the transect the Tagus River Floodplain sediments date to
292 6.7 cal. kyrs. BP, the Mudbelt sediments date to 5.8 cal. kyrs. BP, the Lisbon Canyon Head
293 sediments date to 8.7 cal. kyrs. BP, and the Lower Setúbal Canyon penetrated the oldest strata
294 (13.0 cal. kyrs. BP). Reported values for sediments from each location were averaged over the
295 interval 0-6.0 cal. kyrs. BP, so as to avoid a bias in the data since not all of the sediment cores
296 covered more than 6.0 kyrs.

297 The bulk carbon isotope data for the Tagus River SPM, riverbank sediments, and soils
298 has been previously discussed in Zell et al., (2014). The TOC values for the Tagus River
299 Floodplain sediments are relatively high and also highly variable with a range of 1.5-16 wt. %
300 and a mean of 6.5 ± 4.3 wt. % (average \pm standard deviation) and the mean $\delta^{13}\text{C}_{\text{TOC}}$ was -
301 27.0 ± 1.0 ‰ (Fig. 2; Table 3). In the Mudbelt sediments the TOC is less variable than in the
302 Tagus River Floodplain sediments, ranging from 0.6-1.2 wt. % and with an average of
303 0.9 ± 0.2 wt. % (Fig. 2; Table 3). The average $\delta^{13}\text{C}_{\text{TOC}}$ in the Mudbelt sediments, -24.3 ± 0.2 ‰,
304 is higher than in the Tagus River Floodplain sediments. The average $\delta^{13}\text{C}_{\text{TOC}}$ of the Lisbon
305 Canyon Head sediments, -23.0 ± 0.6 ‰, is higher than the Mudbelt sediments and the TOC
306 content is similar to that of the Mudbelt sediments, ranging from 0.25-1.5 wt. % with the
307 mean of 0.9 ± 0.3 wt. % (Fig. 2; Table 3). The average $\delta^{13}\text{C}_{\text{TOC}}$ values in the Lower Setúbal
308 Canyon sediments (-23.4 ± 1.5 ‰) are similar to those of the Lisbon Canyon Head sediments
309 with a TOC content ranging from 0.51-0.85 wt. % with a mean value of 0.65 ± 0.14 wt. % (Fig.
310 2; Table 3).

311

312 **4.2 Concentrations and distribution of GDGTs**



313 *Tagus Soils and Riverbank Sediments.* The average concentration of crenarchaeol is
314 higher in the riverbank sediments ($\sim 8.7 \pm 7.8 \mu\text{g gOC}^{-1}$) than in the soils ($\sim 1.4 \pm 1.1 \mu\text{g gOC}^{-1}$)
315 (Fig. 3a-b; Table 3). The same trend is true for the brGDGTs with the average concentration
316 being higher in the riverbank sediments ($\sim 33.9 \pm 24.5 \mu\text{g gOC}^{-1}$) than the soils ($\sim 6.8 \pm 6.5 \mu\text{g}$
317 gOC^{-1}) (Fig. 3a-b; Table 3). The values of the BIT index were similar to those previously
318 reported (Zell et al., 2014) for both the soils and riverbank sediments and ranged from 0.3 to
319 1.0 with an average of 0.7 ± 0.2 (Fig. 3c; Table 3). The re-analysis of the brGDGTs in the soils
320 reveals that the relative abundance of the novel 6-methyl brGDGTs is highly variable (ranging
321 from 0.13-0.92) and can be quite high; the average values for the IR are 0.6 ± 0.3 (Fig. 3e;
322 Table 3). IR is even higher but less variable for the riverbank sediments with an average of
323 0.7 ± 0.1 (Fig. 3e; Table 3). In general the penta- and hexamethylated brGDGTs show the same
324 ratio of 5- and 6-methyl isomers (Fig. 4), however, in soils from an altitude of >350 m the 6-
325 methyl brGDGTs are especially dominant (Fig. S4). Values for the new MBT'_{5me} index,
326 which excludes the 6-methyl brGDGTs (cf. De Jonge et al., 2014a), of the soils and riverbank
327 sediments are quite similar with an average of 0.5 ± 0.1 in both cases (Fig. 3f; Table 3). The
328 DC' ratio deviates between the soils and the riverbank sediments (Fig. 3d; Table 3). The DC'
329 for the soils is highly variable but on average low (0.2 ± 0.1); for the riverbank sediments it is
330 higher with an average of 0.4 ± 0.1 .

331 *Tagus River SPM.* The SPM was obtained from the Tagus Estuary near the mouth of
332 the Tagus River once a month over the course of a year (excluding the month of August).
333 Data from the Tagus River SPM showed that the summed brGDGT and crenarchaeol
334 concentrations in the river SPM varied throughout the year and were on average $45 \pm 23 \mu\text{g}$
335 gOC^{-1} , and $9.8 \pm 6.8 \mu\text{g OC}^{-1}$, (Figs. 3a-b; Table 3), respectively, resulting in only small
336 variations in the BIT index (i.e. 0.8 ± 0.1 ; Fig. 3c; Table 3). The distribution of brGDGTs (Fig.



337 5c) was relatively constant throughout the year as is evident from the values for MBT'_{5me}

338 (0.5 ± 0.0), DC' (0.3 ± 0.0), and IR (0.6 ± 0.0) for the river SPM (Figs. 3d-f; Table 3).

339 *Tagus River Floodplain sediments.* The average crenarchaeol concentration is fairly
340 low in the Tagus River Floodplain sediments, $2.8\pm 1.7 \mu\text{g gOC}^{-1}$, conversely, the average sum
341 of the brGDGTs in the sediments, $70\pm 26 \mu\text{g gOC}^{-1}$, is the largest out of the entire transect
342 (Figs. 3a-b; Table 3). The BIT index is fairly high and constant throughout the sediment core
343 with an average value of 0.9 ± 0.0 (Fig. 3c; Table 3). The distribution of brGDGTs (Fig. 5d) is
344 somewhat similar to that of the riverine SPM (Fig. 4c) and shows no major changes over the
345 Holocene. The Tagus River Floodplain sediments has the lowest average values for MBT'_{5me} ,
346 0.4 ± 0.1 , and IR, 0.4 ± 0.0 , of all the sediment records in the transect (Figs. 3e-f). The mean
347 DC' throughout the sediments in this sample set is 0.4 ± 0.1 (Fig. 3d; Table 3).

348 *Mudbelt sediments.* The average concentration of the brGDGTs, $25\pm 14 \mu\text{g gOC}^{-1}$, is
349 lower than in the Tagus River Floodplain sediments, however, the concentration of
350 crenarchaeol, $170\pm 50 \mu\text{g gOC}^{-1}$, is higher in the Mudbelt sediments (Figs. 3a-b; Table 3). This
351 results in a lower mean value of the BIT index (i.e. 0.09 ± 0.03 ; Fig. 3c; Table 3). The brGDGT
352 distribution is relatively constant over the Holocene and is fairly similar to that of the Tagus
353 River floodplain sediments with slightly higher fractional abundances of Ia and IIIa' (cf. Figs.
354 5d-e; Table 3). The average value of the MBT'_{5me} (0.5 ± 0.0) is similar to the Tagus River SPM
355 value (Fig. 3f). The average value of the DC' is 0.3 ± 0.1 and the mean value of the IR is
356 0.5 ± 0.0 (Figs. 3d-e; Table 3).

357 *Lisbon Canyon Head sediments.* The average sum of the brGDGTs, $31\pm 9.3 \mu\text{g gOC}^{-1}$,
358 is about the same in the Lisbon Canyon Head sediments as in the Mudbelt sediments but the
359 amount of crenarchaeol, $390\pm 130 \mu\text{g gOC}^{-1}$, is larger in the Lisbon Canyon Head sediments
360 (Figs. 3a-b; Table 3). This results in lower BIT values (0.1 ± 0.0) than in the Mudbelt



361 sediments (Fig. 3c; Table 3). The average brGDGT distribution (Fig. 5f) is fairly similar to
362 that of the Tagus Floodplain and Mudbelt sediments and is relatively constant over the
363 Holocene. The average of the MBT'_{5me} (0.5 ± 0.0) is statistically identical to that in the
364 Mudbelt sediments (Fig. 3f; Table 3). However, the average IR, 0.6 ± 0.0 , and DC' , 0.4 ± 0.0 ,
365 are both a bit higher (Figs. 3d-e; Table 3).

366 *Lower Setúbal Canyon sediments.* The concentrations of the brGDGTs in these most
367 distal sediments are quite low, on average $16 \pm 5.5 \mu\text{g OC}^{-1}$ (Fig. 3a; Table 3), while the
368 amount of crenarchaeol in this sediment core is the highest out of the entire transect at
369 $470 \pm 200 \mu\text{g gOC}^{-1}$ (Fig. 3b; Table 3). This results in a low average BIT index value of
370 0.02 ± 0.01 (Fig. 3c; Table 3). The average distribution of brGDGTs in these sediments (Fig.
371 5g) is different from the marine sediments from the other two sites, with a higher fractional
372 abundance of IIIa'. However, another component with the same molecular ion eluted at
373 around the same time as IIIa' in the Lower Setúbal Canyon sediments (which we determined
374 was not the “mixed 5,6-dimethyl isomer”; cf. Weber et al., 2015), complicating integration
375 and quantification. This indicates that the brGDGT results from these sediments must be
376 interpreted with some caution. The average MBT'_{5me} (0.6 ± 0.1) and DC' (0.4 ± 0.1) are fairly
377 similar to the Lisbon Canyon Head sediments averages but the average IR (0.7 ± 0.0) is the
378 highest of all sediments (Figs. 3d-f; Table 3).

379

380 4.3 PCA

381 In order to determine the variation in the distribution of brGDGTs, we performed
382 principal component analysis (PCA) on the distributions of brGDGTs of all the samples
383 examined. Most variation is explained by principal component 1 (PC1; 29.8 %) and is clearly
384 related to the fractional abundance of the 5-methyl versus 6-methyl brGDGTs (Fig. 6a). With



385 the exception of IIIc (which is typically a minor brGDGT with a fractional abundance of <1
386 %; Fig. 5), all of the 5-methyl brGDGTs score positively on PC1 and the 6-methyl brGDGTs
387 score negatively. For the overall data set, PC1 is highly negatively correlated with the IR ratio
388 (Fig. 7a, $R^2=0.78$). PC2 explains 25.6 % of the variance of the PCA. Branched GDGTs that
389 score positively on PC2 are generally comprised of cyclized and more methylated brGDGTs
390 (Fig. 6a). With the exception of IIIc (which is typically a minor brGDGT with a fractional
391 abundance of <1 %; Fig. 4), all of the tetra- and penta-methylated brGDGTs containing no
392 cyclopentane moiety (i.e. Ia, IIa, and IIa') score negatively on PC2. Consequently, PC2 is
393 highly positively correlated with DC' for the whole data set (Fig. 7b, $R^2=0.84$).

394

395 4. Discussion

396 5.1 Environmental parameters affecting brGDGT distribution in Tagus soils

397 Evident from the earlier study by Zell et al. (2014) was that the distribution of the
398 brGDGTs in Tagus soils varies widely. The primary environmental parameters influencing
399 brGDGT distributions in soil (Weijers et al., 2006), i.e. MAT and pH, did differ substantially
400 in the Tagus River basin. MAT varies from 10-17°C and pH from 5.5-8.6 (Zell et al., 2014)
401 and both parameters show a distinct correlation with altitude ($R^2=0.93$ and 0.73, respectively).
402 Applying the brGDGT global soil calibration of Peterse et al. (2012), Zell et al. (2014) arrived
403 at unrealistically low (0-10°C) estimated MATs using the brGDGT distributions. This was
404 attributed to the arid conditions in the region ($MAP < 800 \text{ mm yr}^{-1}$), which has in other studies,
405 including one that analyzed soils from the Iberian peninsula, been indicated as a likely cause
406 for the discrepancy between actual and reconstructed MAT using brGDGT distributions
407 (Peterse et al., 2012; Dirghangi et al., 2013; Menges et al., 2013). Our re-analysis of the soils
408 taking into account the novel 6-methyl brGDGTs now provides the possibility to re-evaluate



409 these data. It is clear that the fractional abundances of the novel 6-methyl brGDGTs vary to a
410 large extent. The IR_{II} and IR_{III} vary from 0.1 to 0.9 (Fig. 4) and some of the soils score very
411 negatively on PC1 (Fig. 6b), which is predominantly determined by the fractional abundance
412 of the 6-methyl brGDGTs. From the global soil brGDGT dataset (De Jonge et al., 2014a) it
413 was evident that the main factor influencing the fractional abundance of the 6-methyl
414 brGDGTs is soil pH with an increased abundance in high pH soils. In the Tagus River basin
415 soil pH indeed shows a large variation, i.e. from 5.5 to 8.6, and this likely explains the large
416 variation in IR. When we calculate the pH from the brGDGT distribution using the new
417 equation (9) of De Jonge et al. (2014a), which is based predominantly on the fractional
418 abundances of 6-methyl brGDGTs, we find a highly significant correlation between measured
419 and reconstructed pH ($R^2=0.89$) following the 1:1 line (Fig. 9a). Differences in soil pH also
420 affect the degree of cyclization of brGDGTs (Weijers et al., 2007a; De Jonge et al., 2014b)
421 and indeed we find a significant positive correlation between DC' and soil pH ($R^2=0.74$). The
422 effect of MAT is not clearly revealed in the dataset. For the global soil brGDGT dataset a
423 strong relationship exists between MAT and MBT'_{5Me} (De Jonge et al., 2014a). Although we
424 observe substantial variation for MBT'_{5Me} in soils (i.e. 0.3-0.7; Fig. 3f) for this dataset, we do
425 not observe a statistically significant relationship of MAT with MBT'_{5Me}. Also, reconstructed
426 MATs are far too low, i.e. 0.5–13°C using equation (6) and 2.6–11°C using equation (10).
427 Evidently, the “cold bias” of the brGDGT distributions in the soils of the Tagus river basin
428 (Zell et al., 2014) is not solved when 5- and 6-methyl brGDGTs are individually quantified.

429 Previously it was postulated that in this region aquatic in-situ production and arid
430 conditions are complicating the use of brGDGTs for climate reconstructions (Menges et al.,
431 2013; Zell et al., 2014). Within the soil sample set a strong negative relationship exists
432 between the DC' and the measured MAT in the Tagus basin ($R^2=0.79$), whereas the degree of
433 cyclization up until this point has only been reported to be related to pH and not to MAT



434 (Weijers et al., 2007a). Conversely, though, the MAT_{mrs} reconstructed values for the soils
435 have a positive correlation with DC' ($R^2=0.51$) and it is lower than with the measured MAT.
436 Although at this point we are unsure if this association occurs in other arid areas as well, we
437 do believe this strong relationship between the DC' and the MAT could be affecting the
438 applicability of brGDGTs for temperature reconstructions in this region.

439

440 **5.2 Provenance of brGDGTs in the Tagus River and its outflow**

441 The application of brGDGTs in marine sediments influenced by river outflows for
442 reconstruction of the continental paleoclimate (e.g. Weijers et al., 2006) rests on the premise
443 that the distribution of the brGDGTs produced in the soils must be conserved throughout
444 riverine transport to the sediments where they are archived. Therefore, we compare brGDGT
445 distributions and concentrations from the rest of the sample set in the source-to-sink transect
446 to determine if the soil signal is conserved during transport in the Tagus River basin. The
447 PCA results (Fig. 6b) indicate that for the most part the distribution of brGDGTs from the
448 river SPM and sediments along the transect is not similar to those from the soils or the Tagus
449 Watershed. Sediments from three of the sample sets in the transect, the Tagus River
450 Floodplain sediments, the Mudbelt sediments and the Lisbon Canyon Head sediments, all plot
451 differently from the soils, and although the distributions of the Lower Setúbal Canyon
452 sediments and the Tagus River SPM plot closer, there is still an offset from the soils. The
453 Tagus Riverbank sediments plot the most closely to that of the soils in the Tagus River basin,
454 however, again a slight offset still exists. So, even without considering the effects of
455 environmental parameters on brGDGT distributions, we can already conclude that the
456 brGDGTs in the sediments and river SPM only reflect the distribution of brGDGTs in the



457 Tagus soils by a minor extent and, thus, it is unlikely that Tagus soils are a major source for
458 brGDGTs in the marine sediments.

459 Using PCA (Fig. 6) we tried to determine what factors are causing the variation in the
460 distribution of brGDGTs in the Tagus River basin. PC1 is primarily related to the
461 predominance of 5-methyl versus 6-methyl brGDGTs (Fig. 6a) and thus pH (cf. De Jonge et
462 al., 2014a). This was confirmed for the soil data set where the calculated pH based on the
463 fractional abundance of predominantly 6-methyl brGDGTs shows a good correspondence
464 with measured pH (see Sect. 5.1). De Jonge et al. (2014b) showed that in the SPM of the
465 alkaline waters of the river Yenisei 6-methyl brGDGTs also predominate, indicating that pH
466 in all kinds of environmental settings determines the ratio between 5 and 6-methyl brGDGTs.
467 The Tagus riverbank sediments, river SPM and the Lower Setúbal Canyon sediments score
468 mostly negatively on PC1 as do soils from higher altitudes (>350 m) (Fig. 6b). The Mudbelt
469 sediments, Lisbon Canyon Head sediments, the Tagus River Floodplain sediments, and the
470 lower altitude soils (<350 m) have similar abundances of the 5- and 6-methyl brGDGTs or
471 higher abundances of the 5-methyl brGDGTs and plot mostly positively on PC1. Since the
472 Tagus River Floodplain sediments, the Mudbelt sediments, and the Lisbon Canyon Head
473 sediments do not have a predominance of 6-methyl brGDGTs, this indicates that either they
474 received an equal contribution of soil derived organic matter from the lower altitude soils in
475 the region (<350 m) as from the higher altitude region (>350m) or, more likely, that in-situ
476 production of brGDGTs is a large source of brGDGTs in these sample sets.

477 PC2 also explains a substantial part of the variance in the dataset (25.6 %, Fig. 6b) and
478 is correlated with DC' ($R^2=0.84$, Fig. 7b). Since pH is also the main driver of DC' (Weijers et
479 al., 2007a), it suggests that differences in pH are also responsible for the variance seen in
480 PC2. The samples that standout are the sediments from the Lower Setúbal Canyon core,
481 which are the most marine sediments in the sample set, and plot most positively, and the



482 lowest altitude soils (28-344 m), which plot the most negatively. These latter soils are
483 characterized by a low measured pH. The oldest (11.6-13.0 kys BP) sediments of the Lower
484 Setúbal Canyon score most positively on PC2. A high degree of cyclization of brGDGTs has
485 been observed previously in marine sediments from a Svalbard fjord and attributed to marine
486 in-situ production in the alkaline pore waters of marine sediments (Peterse et al., 2009;
487 Weijers et al, 2014). Re-analysis of the Svalbard sediments for brGDGTs actually showed that
488 this cyclization affects the tetra- and pentamethylated brGDGTs to a much larger extent than
489 that of the hexamethylated brGDGTs (Sinninghe Damsté et al., unpublished results) and the
490 same observation can be made for the sediments of the Lower Setúbal Canyon (Fig. 5g).
491 Evidently, the high degree of cyclization of brGDGTs as a response to pH is not as clearly
492 seen in the soils since the high altitude, high pH soils from the Tagus watershed (Fig. S3c) do
493 not exhibit the pattern (i.e. fractional abundance of I Ib' larger than that of IIa') observed in
494 the Lower Setúbal Canyon sediments (Fig. 5g). This pattern is, to a lesser degree, also seen in
495 the sediments of the Lisbon Canyon Head core (Fig. 5f). As mentioned earlier, the Lower
496 Setúbal Canyon sediments also display a predominance of 6-methyl brGDGTs over the 5-
497 methyl counterparts, especially with regards to the hexamethylated brGDGTs. In the Lower
498 Setúbal Canyon sediments IIIa' is by far the most abundant brGDGT, consisting of 29 % of
499 the entire brGDGT pool (Fig. 5g). This is comparable to Svalbard sediments (Sinninghe
500 Damsté et al., unpublished results) where IIIa' is also the most abundant brGDGT. Taken
501 together this clearly indicates the influence of in-situ production in the Lower Setúbal Canyon
502 sediments. However, the degree of cyclization for Ia-c and IIa-c is not as high as observed for
503 the Svalbard sediments, which still suggests some allochthonous input of brGDGTs even in
504 these remote marine sediments.

505 Another way to determine if in-situ production is a factor affecting the brGDGT
506 distribution in aquatic environments is by the calculation of reconstructed pH values. If in-situ



507 production is heavily contributing to the brGDGT pool, then the reconstructed pH values
508 should reflect that of the aquatic environment in which they were produced. The average
509 reconstructed pH of the sample sets in the transect are relatively high with a clear trend to
510 higher values with increasing distance from the river mouth (Fig. 8a), which would be in line
511 with increased in situ production of brGDGTs in the alkaline pore waters of marine
512 sediments. However, these values are still within the range of the measured (5.5-8.5) and
513 reconstructed (Fig. 8a) pH of the soils and so this does not prove in-situ production as a major
514 contributor of brGDGTs in these sample sets. Conversely, the newly calculated DC', also a
515 reflection of pH, is quite variable throughout the sample sets in the transect except for in the
516 river SPM where it is fairly constant (Fig. 3d; Table 3). Since the DC' is lowest in the soils
517 (0.2 ± 0.1) and then higher in the rest of the samples in the transect (0.3-0.4), this suggests in-
518 situ production is an issue (cf. Zell et al., 2014) in all of the sample sets (Fig. 3d; Table 3).

519

520 **5.3 brGDGTs as indicators of terrestrial OM transport by the Tagus River**

521 Classically, the assessment of the contribution of terrestrial OM to marine sediments is
522 performed by measuring $\delta^{13}\text{C}_{\text{TOC}}$ (Hedges and Oades, 1997 and references cited therein). In
523 the earlier study of the Tagus River system, Zell et al. (2014) determined that the average
524 $\delta^{13}\text{C}_{\text{TOC}}$ of the riverine SPM ($\sim -29 \pm 0.8\text{‰}$), like the Tagus soils, are consistent with a
525 predominant C_3 higher plants origin (Fry and Sherr, 1984). Additionally, this study found the
526 $\delta^{13}\text{C}_{\text{TOC}}$ in marine surface sediments off the Portuguese coast in front of the Tagus River
527 increase with increasing distance offshore by an increased contribution of ^{13}C -enriched
528 marine OM. This trend is also evident for the Holocene sediments studied here. The most
529 terrestrial sediments of the transect, i.e. from the Tagus River Floodplain, also have a $\delta^{13}\text{C}_{\text{TOC}}$
530 value ($\sim -27 \pm 1.0\text{‰}$; Fig. 2a; Table 3) consistent with a predominant C_{origin} of higher plants.



531 Moving offshore, the less negative $\delta^{13}\text{C}_{\text{TOC}}$ values of the Mudbelt sediments ($-24\pm 0.2\text{‰}$), the
532 Lisbon Canyon Head sediments ($-23\pm 0.6\text{‰}$) and the Lower Setúbal Canyon sediments ($-$
533 $23\pm 1.5\text{‰}$) all indicate that the majority of the TOC off the Portuguese shelf is of marine
534 origin (Fig. 2a; Table 3). So, as Zell et al. (2014) found with marine surface sediments off the
535 Portuguese coast, the $\delta^{13}\text{C}_{\text{TOC}}$ (‰) averages from the sediments in our transect also increase
536 with increasing distance offshore, demonstrating that the present trend in the $\delta^{13}\text{C}_{\text{TOC}}$ signal
537 remained the same over the Holocene.

538 Zell et al. (2014) previously showed that in the present day Tagus River system the
539 amount of brGDGTs ($\mu\text{g gOC}^{-1}$) increases from the soils to the riverbank sediment to the river
540 SPM and explained this increase as proof of riverine in-situ production of brGDGTs.
541 Concentrations of summed brGDGTs in surface sediments in transects from the Portuguese
542 coast rapidly declined with increasing distance from the coast, suggesting that brGDGTs
543 could still be used as tracer for terrestrial organic matter (Zell et al., 2015). The trends
544 observed in these earlier studies are confirmed here for the Holocene. The Tagus River
545 Floodplain sediments have the highest concentration of brGDGTs ($67\pm 26\ \mu\text{g gOC}^{-1}$) in the
546 entire transect, much higher than in the soils (Fig. 3b; Table 3). However, the sediments in
547 this core are somewhat atypical for the Tagus Floodplain as some layers consist of peat as a
548 result of the low-energy backswamp conditions in the vicinity, which could explain the
549 difference in brGDGT concentrations from the surrounding soils. This could also be due to
550 the addition of aquatically produced brGDGTs from the river during times of flooding
551 although it should be noted that the concentration of brGDGTs is even higher than in riverine
552 SPM (Fig. 3b). The summed brGDGT concentration decreases and is fairly similar among the
553 Mudbelt sediments ($25\pm 14\ \mu\text{g gOC}^{-1}$) and the Lisbon Canyon Head sediments ($31\pm 9.3\ \mu\text{g}$
554 gOC^{-1}), and then decreases further moving away from the coastline to the Lower Setúbal
555 Canyon sediments ($16\pm 5.5\ \mu\text{g gOC}^{-1}$) demonstrating the decrease in input of riverine



556 brGDGTs moving away from the shoreline (Fig. 3b). However, even though the sum of the
557 brGDGTs are lower in the marine sediment than in the Tagus River Floodplain sediments, the
558 amount of brGDGTs in all four sediment cores are higher than in the Tagus soils ($\sim 6.8 \pm 6.5 \mu\text{g}$
559 gOC^{-1}) indicating the origin of the brGDGTs in the sediment cores are not all soil derived and
560 pointing instead to riverine in-situ production as well as possibly in aquatic sediments (Fig.
561 3b).

562 A previous study by Zell et al., (2015) determined that in the surface sediments off the
563 coast of Portugal the BIT index is influenced by both declining brGDGT concentrations and
564 increased crenarchaeol production with increasing distance from the coast. For the Holocene
565 sediments studied here, the average concentration of crenarchaeol in the Tagus River
566 Floodplain sediments is low ($2.8 \pm 1.7 \mu\text{g gOC}^{-1}$) and similar to that of the Tagus soils (1.4 ± 1.1
567 $\mu\text{g gOC}^{-1}$; Fig. 3a; Table 3). The crenarchaeol concentration increases in the sediments with
568 increasing distance from the shoreline, signifying the increase in marine production with
569 water depth and distance from the coast (Fig. 3a). Consequently, the BIT index is the highest
570 in the Tagus River Floodplain sediments (0.94 ± 0.03) out of the entire transect (Fig. 3c) and
571 then the BIT index decreases within the sediments along the transect with increasing distance
572 from the Portuguese coast potentially signifying a decrease in terrestrial input moving away
573 from the shoreline. A moderate negative correlation exists between the BIT index and $\delta^{13}\text{C}_{\text{TOC}}$
574 values for the entire sample set ($R^2=0.55$, Fig. 10), demonstrating that as $\delta^{13}\text{C}_{\text{TOC}}$ values
575 become less terrestrial, the BIT index indicates less terrestrial input, so both parameters
576 corroborate a decrease in terrestrial influence with distance from the coast confirming the
577 results from previous studies (Zell et al., 2014; 2015).

578



579 **5.4 Factors affecting the application of brGDGTs for paleoclimate reconstructions off**
580 **the Iberian Peninsula**

581 Despite the caveats with respect to in-situ production of brGDGTs in aquatic
582 environments as described in the previous section, we tested how the new soil calibration
583 based on individually quantified 5-methyl and 6-methyl brGDGTs (De Jonge et al., 2014a)
584 performed to reconstruct continental MAT in this region. For this comparison we will
585 consider the present day MAT of the entire Tagus River basin, $14.6\pm 2.2^\circ\text{C}$ (Zell et al., 2014),
586 assuming that soil derived brGDGTs from along the whole river basin are contributing to the
587 marine sediments. The assumption that the brGDGTs from the entire Tagus River basin are
588 being contributed to oceanic sediments is probably invalid for modern times as the
589 construction of dams along the Tagus River, which began in the 1940s, most likely prevents
590 part of the terrestrial material from upstream making it downstream and out off the coast of
591 Portugal. However, since we are not looking at marine surface sediments in this study but
592 instead sediments deposited during the Holocene, the placement of dams in the river should
593 not affect our results except for with the riverine SPM. Despite the separation of the 5- and 6-
594 methyl brGDGT isomers and the application of the new proxy, the reconstructed MATs using
595 both riverine SPM and Holocene sediments is still substantially lower than 14.6°C (Figs. 8b
596 and 8c), as noted for the soils (see Sect. 5.1). Using the MAT_{mr} calibration the Lisbon Canyon
597 Head sediments the average temperature ($12.4\pm 0.5^\circ\text{C}$) comes closest to the modern day MAT
598 in the region and using the MAT_{mrs} calibration the Lower Setúbal Canyon sediments
599 ($11.2\pm 0.7^\circ\text{C}$) has the most similar average temperature (Figs. 8b and 8c).

600 Even though we used the new calibration to reconstruct MAT, it should be noted that
601 the low BIT values (< 0.15 ; Fig. 3c) of the Holocene sediments deposited at the three marine
602 sites indicates that there were probably not enough soil-derived brGDGTs making it out to
603 ocean and being deposited in the sediments over the Holocene for reliable climate



604 reconstructions (cf. Weijers et al., 2014). By looking at the summed concentration of
605 brGDGTs along the entire transect, it is apparent that since it is lowest in the soils that
606 although the BIT index seems high enough for MAT reconstructions in the riverbank
607 sediments and river SPM, the origin of the brGDGTs may not be solely soil derived and this
608 could be complicating reconstructions throughout the transect as was discussed previously
609 (see Sect. 5.2). This further supports earlier conclusions from preceding studies (Yang et al.,
610 2012; Zell et al., 2013) stating that the amount and origin of brGDGTs in a system needs to be
611 examined along with the BIT index when determining if brGDGTs can be applied for MAT
612 reconstruction.

613

614 **6. Conclusions**

615 We have established that the distribution of brGDGTs varies greatly within the Tagus
616 River basin (Fig. 5) and although this may be partly explained by the varying contributions of
617 higher altitude, which contain a greater proportion of 6-methyl isomers, versus lower altitude
618 soils in the sample sets it is more likely due to the contribution of aquatically produced
619 brGDGTs in some of the sample sets. In order to use sedimentary brGDGTs for paleoclimate
620 reconstructions, the distribution of brGDGTs in the soils must be related to the MAT and
621 conserved throughout riverine transport to the sediments where they are deposited, however,
622 our results corroborate previous studies stating that most of the terrestrial matter is not making
623 it out to the ocean and being deposited in sediments close to shore. The lack of soil derived
624 OM in offshore sediments along with the substantial input of aquatically produced brGDGTs
625 is complicating MAT reconstructions from sedimentary, marine brGDGTs in this region.

626 Additionally, we confirm the findings of Zell et al., (2014; 2015) that in-situ production
627 of brGDGTs is occurring in the river and marine systems of the Tagus River basin and go on



628 to show that there are indications that it occurred in the past as well. Although in-situ
629 production is complicating environmental reconstructions using marine sediments, another
630 issue is that accurate MAT reconstructions using brGDGTs cannot currently be performed on
631 the soils, even with the separation of the 6-methyl brGDGTs from the 5-methyl isomers using
632 the new method and calibrations. Previous studies have concluded that paleoclimate
633 reconstructions in arid regions using brGDGTs are complicated due to a breakdown in the
634 relationship with MBT' and MAT (Peterse et al., 2012; Menges et al., 2013). In this study we
635 confirm that there is not a strong relationship between the MBT'_{5me} and measured MAT in
636 this arid region. However, we also do not observe the same relationship with MAP and
637 MBT'_{5me} that has been previously reported between MAP and MBT' in arid regions and has
638 been implicated in making reconstructions difficult. Instead, we see a strong relationship with
639 the DC' and measured MAT in the area not observed before. We also see a predominance of
640 6-methyl isomers, previously only reported in river SPM, in the Tagus soils from greater than
641 350 m altitude. Although this might be a characteristic of arid soils and related to MAP since
642 it is below 550 mm yr⁻¹ in most of the soil samples above 350 m, the two highest elevation
643 soil samples, which both have a MAP above 550 mm yr⁻¹, also demonstrate this trend. Future
644 studies need to be performed in arid environments to determine if a strong relationship
645 between MAT and DC' as well as a predominance of 6-methyl isomers are characteristics of
646 arid regions and contributing to the complications found using brGDGTs for paleoclimate
647 reconstructions. Also, higher elevation environments should be further studied to determine if
648 a predominance of 6-methyl brGDGTs is a feature of higher altitudes and complicating
649 climate reconstructions.

650 Because of these unique features in this region, perhaps the development of a local
651 calibration could assuage difficulties in using brGDGTs as a paleoclimate proxy for soils in
652 the Tagus River basin. This would not, however, solve the issue of in-situ produced brGDGTs



653 overwhelming the amount of soil derived brGDGTs in aquatic sediments. We did find that the
654 new CBT' and pH calibration do an excellent job reconstructing pH in the soils of the Tagus
655 basin and since pH is related to other environmental factors such as MAP this will be useful
656 for paleoclimate reconstructions in terrestrial sites over the Iberian peninsula where in-situ
657 production is not a complicating factor.

658

659 **Author contribution**

660 J.H. Kim and J.S.S. Damsté designed the study which was carried out by L. Warden who
661 completed bulk carbon isotope and brGDGT analysis on samples along with C. Zell. C. Zell,
662 H. Stigter, G.J. Vis and J.H. Kim collected samples for this study. J. Bonnin picked forams for
663 dating. L. Warden and J.S.S. Damsté prepared the manuscript with contributions from all co-
664 authors.

665

666 **Acknowledgements**

667 The research leading to these results has received funding from the European Research
668 Council under the European Union's Seventh Framework Programme (FP7/2007-2013) / ERC
669 grant agreement n° [226600]. JSSD is supported by funding from the Netherlands Earth
670 System Science Center (NESSC) though a gravitation grant from the Dutch ministry for
671 Education, Culture and Science. We thank the captain and crew of the R/V Pelagia for their
672 support with sampling. We thank J. Ossebaar and E.C. Hopmans for their help with the
673 HPLC-APCI-MS analysis, J. Ossebaar for his help with the element analyzer, and L.A.C.
674 Rosales and A. Mets for help with sample preparation.

675



676 **Data Availability**

677 The data presented in this paper can be found in Table 3.

678 **Figure captions**

679 **Figure 1** The location of the study area on the Iberian Peninsula with the stations where the
680 four sediment cores were sampled (indicated by black squares) along a transect from the
681 Tagus River to off the Portuguese continental margin as well as the river SPM sampling site
682 (indicated by a white diamond), riverbank sediment sampling sites (indicated by red circles),
683 and soil sampling sites (indicated by black circles). The River SPM, riverbank sediments and
684 soil samples were all collected for a previous study. Digital elevation data from Jarvis et al.
685 (2006) and bathymetry from IOC-IHO-BODC (2003).

686 **Figure 2** Boxplots of (a) $\delta^{13}\text{C}_{\text{TOC}}$ (‰) of the organic carbon and (b) TOC (wt. %) for each
687 sample set along the Tagus River source to sink transect. The increasing $\delta^{13}\text{C}_{\text{TOC}}$ values in the
688 sediment core locations with increasing distance from the coast indicates that more of the
689 organic carbon in these sediments is marine derived.

690 **Figure 3** Boxplots of (a) crenarchaeol concentrations ($\mu\text{g gOC}^{-1}$), (b) sum of brGDGTs (μg
691 gOC^{-1}), (c) BIT index, (d) DC', (e) IR, (f) $\text{MBT}'_{5\text{me}}$ for each sample set in the transect from
692 the land to the ocean off the Portuguese coast.

693 **Figure 4** Isomer ratio for the non-cyclized pentamethylated brGDGT (IR_{II}) plotted against
694 that of the non-cyclized hexamethylated brGDGT (IR_{III}).

695 **Figure 5** Average distribution of brGDGTs for each sample set along the transect of samples
696 that runs from the land to the ocean off the coast of Lisbon. Evident from this figure is that the
697 distribution of brGDGTs within this sample set varies greatly. Distributions of brGDGTs in
698 marine sediments only reflects the distribution of the brGDGTs from the Tagus soils to a



699 minor extent. The color of the bars reflects the brGDGT structure as labeled in the legend and
700 the range indicated with the error bars equals 2xs the standard deviation.

701 **Figure 6** Principal component analysis based on the fractional abundances of the 15
702 brGDGTs of samples in the transect that runs from inland to off the coast of Portugal plotting
703 a) the scores of the brGDGT compounds on the first two principal components (PC) and b)
704 the scores of the samples from each sample set used in this study.

705 **Figure 7** Scatter plots of (a) PC1 against the IR ($R^2=0.78$) and (b) PC2 against DC' ($R^2=0.84$)
706 for the entire set of samples used in this study.

707 **Figure 8** Boxplots of all the sample sets within the transect from the land to the deep ocean
708 off the Portuguese coast for (a) reconstructed pH, (b) MAT_{mrs} ($^{\circ}\text{C}$) and (c) MAT_{mr} ($^{\circ}\text{C}$). Red
709 dotted line indicates estimated present day MAT for the Tagus River basin (14.6°C).

710 **Figure 9** Panels a-c show scatterplots of the Tagus soil samples for a) reconstructed and
711 measure pH ($R^2=0.89$), b) reconstructed MAT_{mr} ($^{\circ}\text{C}$) and measured MAT ($^{\circ}\text{C}$) ($R^2=0.27$), c)
712 reconstructed MAT_{mrs} ($^{\circ}\text{C}$) and measured MAT ($^{\circ}\text{C}$) ($R^2=0.38$). For panels b-c the soil
713 samples from an altitude greater than 350m are indicated in black and those from an altitude
714 below 350m are indicated in green. Panels d-f show scatter plots of the Tagus riverbank
715 sediments for d) reconstructed and measured pH ($R^2=0.14$), e) reconstructed MAT_{mr} ($^{\circ}\text{C}$) and
716 measured MAT ($^{\circ}\text{C}$) ($R^2=0.31$), f) reconstructed MAT_{mrs} ($^{\circ}\text{C}$) and measured MAT ($^{\circ}\text{C}$)
717 ($R^2=0.23$). Panel g is a scatter plot showing the reconstructed and measured pH for the Tagus
718 River SPM samples ($R^2=0.09$).

719 **Figure 10** Scatterplot of the BIT index and $\delta^{13}\text{C}_{\text{TOC}}$ (‰ VPDB) for the entire Portuguese
720 sample set ($R^2=0.55$). In general, higher BIT values correlate with more depleted $\delta^{13}\text{C}_{\text{TOC}}$
721 indicating a more terrestrial signal and lower BIT values correlate with less depleted $\delta^{13}\text{C}_{\text{TOC}}$
722 indicating a more marine signal.



723

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945 **Table 1** provides the stations, sediment core names, locations of sampling and water depth for
946 each sediment core used in this study.

Station	Core name	Latitude [N]	Longitude [W]	Water depth [m]
0501.029	Tagus River Floodplain	39° 23' 07.80"	08° 31' 55.56"	0
64PE332-30-2	Tagus Mudbelt	38° 39' 02.20"	09° 28' 07.68"	82
64PE332-44-2	Lisbon Canyon Head	38° 30' 20.19"	09° 15' 04.87"	259
64PE269-39-4	Lower Setúbal Canyon	38° 13' 12.00"	10° 10' 00.00"	4217

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961 **Table 2** summarizes the data used to determine an age depth model for the sediment samples
 962 in this study. ^aData is from Vis et al., (2008).

Sediment core	Lab code	Depth in core [cm]	Mean depth in core [cm]	Uncorrected AMS ¹⁴ C ages [yr BP]	Analytical error ($\pm 1\sigma$) [yrs]	Ages ($\Delta R = 0$ yr) ($\pm 2\sigma$) [cal yr BP]	Ages [cal yr BP]	Analyzed material
0501.029 ^a		0-2	1				0	
0501.029 ^a		331-334	332.5	1136	38	964-1150	1057	Roots of fraction > 125 μ m
0501.029 ^a		331-334	332.5	1022	37	901-1001	951	Total organic fraction > 125 μ m
0501.029 ^a		604-607	605.5	3089	38	3209-3383	3296	Terrestrial botanical macrofossils
0501.029 ^a		711-712	711.5	4129	42	4530-4821	4676	Terrestrial botanical macrofossils
0501.029 ^a		1024-1029	1026.5	5790	40	6485-6676	6581	Terrestrial botanical macrofossils
0501.029 ^a		1046-1050	1048	5900	45	6633-6805	6719	Terrestrial botanical macrofossils
64PE332-30-2		0-2	1				0	
64PE332-30-2	BETA 348791	20-22	21	500	30	40-236	138	Gastropod fragments
64PE332-30-2	BETA 348792	428-430	429	1730	30	1221-1349	1285	<i>Ammonia beccarii</i> (benthic forams)
64PE332-30-2	BETA 348793	678-680	679	2320	30	1848-2032	1940	Gastropod
64PE332-30-2	BETA 317911	976-978	977	5370	30	5643-5849	5746	Bivalve shell fragments
64PE332-44-2		0-2	1				0	
64PE332-44-2	BETA 317906	521-523	521.5	2330	30	1858-2044	1951	Mixed planktonic forams
64PE332-44-2	BETA 317907	770-772	771.5	5390	30	5664-5865	5765	Gastropod
64PE332-44-2	BETA 317908	924.5-926.5	925.5	8160	40	8515-8798	8657	Mixed planktonic forams
64PE269-39-4		0	0				0	
64PE269-39-4	BETA 330562	5	5	930	30	486-608	547	<i>G. bulloides</i> (planktonic forams)
64PE269-39-4	BETA 330563	100	100	4980	50	5205-5466	5336	<i>G. bulloides</i> (planktonic forams)
64PE269-39-4	BETA 330564	200	200	10190	40	11092-11271	11182	<i>G. bulloides</i> (planktonic forams)
64PE269-39-4	BETA 348794	280	280	11540	40	12865-13150	13008	<i>G. bulloides</i> (planktonic forams)



963 **Table 3.** Concentrations of GDGTs and brGDGT based indices for each sample set along the
 964 transect. Values in red were previously published in Zell et al., (2014).

Sample name	Age (cal. kyrs. BP)	TOC (wt. %)	$\delta^{13}C_{TOC}$ (‰ VPDB)	Crenarc haeol ($\mu\text{g gOC}^{-1}$)	Sum brGDGTs ($\mu\text{g gOC}^{-1}$)	BIT index	MBT _{5me} '	DC'	IR	IR IIa	IR IIIa
Tagus soils											
TRS-8b	n/a	3.0	-27.8	2.2	19.2	0.88	0.51	0.13	0.39	0.36	0.43
TRS-7	n/a	5.0	-27.5	0.0	7.5	1.00	0.56	0.01	0.13	0.12	0.17
TRS-9	n/a	0.5	-27.2	0.1	7.8	0.99	0.68	0.01	0.14	0.14	0.15
TRS-3	n/a	0.7	-29.0	0.6	4.1	0.87	0.40	0.05	0.41	0.43	0.39
TRS-4	n/a	0.7	-28.7	1.2	5.3	0.81	0.42	0.04	0.39	0.41	0.30
TRS-5	n/a	2.2	-28.4	2.4	15.3	0.85	0.29	0.10	0.37	0.37	0.35
TRS-10	n/a	1.5	-28.5	1.4	1.6	0.49	0.60	0.19	0.84	0.88	0.83
TRS-12	n/a	0.2	-25.1	4.0	2.4	0.34	0.62	0.22	0.87	0.87	0.91
TRS-14b	n/a	0.8	-25.3	1.7	2.1	0.48	0.57	0.32	0.85	0.89	0.86
TRS-13	n/a	6.9	-27.0	0.9	1.2	0.54	0.33	0.13	0.74	0.80	0.71
TRS-15	n/a	0.9	-25.7	1.9	2.0	0.48	0.46	0.21	0.86	0.90	0.86
TRS-16	n/a	0.1	-24.8	1.6	3.0	0.62	0.56	0.19	0.92	0.93	0.93
TRS-20	n/a	0.1	-26.1	0.6	3.8	0.84	0.52	0.38	0.83	0.87	0.83
TRS-19	n/a	0.1	-25.7	0.7	19.7	0.95	0.56	0.44	0.84	0.85	0.89
Tagus Riverbank sediments											
TRS-6	n/a	1.7	-26.3	16.7	76.0	0.77	0.53	0.30	0.54	0.46	0.59
TRS-8a	n/a	0.2	-26.5	10.8	68.2	0.83	0.55	0.26	0.77	0.71	0.83
TRS 2a	n/a	3.3	-23.6	3.4	12.8	0.70	0.63	0.42	0.79	0.81	0.77
TRS 2b	n/a	0.5	-25.1	8.1	55.1	0.80	0.57	0.45	0.67	0.72	0.72
TRS 1a	n/a	0.9	-26.9	9.7	18.8	0.62	0.54	0.21	0.68	0.68	0.66
TRS1b	n/a	1.5	-27.6	26.1	19.7	0.28	0.64	0.53	0.72	0.82	0.80
TRS-11	n/a	1.3	-27.0	6.3	3.9	0.33	0.48	0.24	0.75	0.79	0.75
TRS-14a	n/a	3.0	-27.1	1.2	21.7	0.92	0.38	0.44	0.55	0.56	0.68
TRS-17	n/a	3.7	-29.6	1.6	37.8	0.93	0.41	0.51	0.70	0.65	0.66
TRS-22	n/a	0.6	-29.8	3.1	24.8	0.86	0.41	0.40	0.80	0.75	0.82
Tagus River SPM											
TR 2 Sup July	n/a	2.6	-29.2	6.2	33.1	0.80	0.51	0.28	0.60	0.53	0.64
TR 3#1 –Sup Sept.	n/a	1.8	-28.4	5.6	20.7	0.73	0.52	0.30	0.59	0.51	0.61
TR4 #1 Oct.	n/a	2.5	-30.9	8.6	38.1	0.77	0.52	0.30	0.61	0.54	0.62
TR5 #1 Sup Nov.	n/a	1.3	-28.9	5.8	54.8	0.85	0.49	0.25	0.62	0.55	0.68
TR 6 #1 Sup Dec.	n/a	2.4	-29.4	11.5	86.8	0.85	0.47	0.26	0.63	0.56	0.67
TR7 #1 Sup Jan.	n/a	2.4	-29.8	9.8	53.6	0.77	0.48	0.27	0.63	0.56	0.67
TR8 #1 Sup Feb.	n/a	1.0	-29.4	16.8	46.9	0.69	0.50	0.27	0.58	0.51	0.62
TR9 #1 Sup Mar.	n/a	2.2	-29.0	6.3	21.9	0.72	0.50	0.29	0.58	0.51	0.61
TR10 #1 Sup Apr.	n/a	1.9	-28.5	0.8	36.5	0.96	0.52	0.29	0.58	0.50	0.61
TR11 #1 Sup May	n/a	1.7	-28.5	26.3	80.8	0.69	0.52	0.29	0.56	0.48	0.59
TR12 #1 Sup June	n/a	1.3	-27.8	9.5	22.8	0.65	0.55	0.23	0.51	0.48	0.58
Tagus River Floodplain											



sediments (0501.029) depth (cm)											
10.0	0.0	1.7	-26.3	2.1	29.7	0.92	0.34	0.31	0.39	0.39	0.44
95.0	0.3	2.6	-26.5	6.4	66.8	0.89	0.36	0.24	0.43	0.43	0.45
195.0	0.6	1.5	-27.4	2.5	111.8	0.97	0.34	0.37	0.38	0.40	0.43
241.0	0.8	1.5	-26.0	5.2	62.0	0.90	0.40	0.24	0.35	0.36	0.38
341.5	1.0	11.2	-27.5	0.9	47.6	0.97	0.45	0.39	0.36	0.34	0.37
401.0	1.5	12.1	-27.7	1.2	90.6	0.98	0.38	0.38	0.40	0.36	0.46
453.0	2.0	8.0	-26.7	3.0	120.9	0.97	0.34	0.35	0.43	0.43	0.50
542.0	2.8	7.7	-27.3	3.2	91.9	0.95	0.37	0.39	0.40	0.41	0.42
577.0	3.1	4.6	-27.2	3.3	84.2	0.95	0.43	0.35	0.38	0.37	0.41
641.0	3.8	5.3	-28.1	1.0	59.6	0.97	0.52	0.39	0.34	0.35	0.39
681.0	4.3	6.6	-28.6	1.0	52.3	0.97	0.46	0.45	0.37	0.37	0.42
741.0	4.9	4.2	-24.4	4.2	74.3	0.92	0.43	0.45	0.38	0.38	0.36
862.0	5.6	16.2	-26.8	2.2	58.4	0.94	0.45	0.45	0.46	0.43	0.52
982.0	6.3	8.7	-27.7	1.5	46.5	0.95	0.46	0.49	0.37	0.36	0.37
1041.0	6.7	5.2	-27.2	4.4	48.2	0.87	0.48	0.45	0.40	0.39	0.41
Mudbelt sediments (64PE332-30-2) depth (cm)											
1.0	0.0	1.2	-23.7	200.9	28.5	0.09	0.58	0.35	0.51	0.45	0.54
25.0	0.2	1.0	-24.2	194.7	40.7	0.13	0.54	0.28	0.49	0.43	0.52
53.0	0.2	0.9	-24.5	189.3	36.4	0.13	0.52	0.26	0.47	0.44	0.50
75.0	0.3	1.0	-24.4	206.5	43.8	0.14	0.52	0.26	0.48	0.43	0.51
101.0	0.4	0.9	-24.3	228.9	44.4	0.13	0.53	0.27	0.48	0.43	0.51
151.0	0.5	0.9	-24.3	194.8	40.2	0.13	0.53	0.27	0.49	0.44	0.53
201.0	0.6	1.0	-24.5	187.9	31.9	0.11	0.51	0.27	0.47	0.43	0.50
248.0	0.8	0.6	-24.5	224.0	58.0	0.14	0.53	0.26	0.46	0.42	0.49
297.0	0.9	1.1	-24.5	168.3	30.6	0.12	0.50	0.26	0.47	0.42	0.51
347.0	1.1	1.0	-24.1	202.5	24.3	0.08	0.52	0.30	0.50	0.45	0.53
397.0	1.2	1.1	-24.1	180.4	27.3	0.10	0.49	0.30	0.48	0.43	0.52
429.0	1.3	1.2	-24.0	77.3	8.7	0.08	0.50	0.34	0.53	0.45	0.57
496.0	1.5	1.0	-24.3	143.8	15.1	0.07	0.52	0.32	0.53	0.46	0.57
546.0	1.6	0.9	-24.1	11.2	1.4	0.08	0.52	0.33	0.51	0.45	0.55
596.0	1.7	1.1	-27.3	141.3	14.8	0.07	0.51	0.34	0.52	0.45	0.56
645.0	1.9	1.1	-29.9	108.7	12.8	0.08	0.52	0.34	0.52	0.45	0.55
680.0	2.0	1.1	-27.4	142.1	17.4	0.08	0.54	0.31	0.50	0.44	0.54
741.0	2.7	0.8	-27.3	176.1	16.0	0.06	0.52	0.35	0.53	0.46	0.55
791.0	3.4	0.7	-24.5	240.8	19.8	0.05	0.50	0.37	0.55	0.48	0.58
840.0	4.0	0.9	-24.4	139.3	11.0	0.05	0.48	0.37	0.54	0.48	0.55
890.0	4.6	0.6	-24.6	180.2	14.2	0.05	0.48	0.39	0.55	0.47	0.57
977.0	5.7	0.7	-27.5	125.8	11.8	0.06	0.46	0.40	0.53	0.46	0.52
Lisbon Canyon Head sediments (64PE332-44-2) depth (cm)											
1.0	0.0	1.5	22.9	420.5	27.1	0.04	0.60	0.41	0.55	0.48	0.56
45.0	0.2	1.2	-22.8	409.9	47.3	0.07	0.58	0.40	0.55	0.48	0.57
85.0	0.3	1.1	-23.4	440.4	48.1	0.07	0.54	0.36	0.53	0.46	0.55



130.5	0.5	1.2	-23.4	429.6	41.2	0.06	0.55	0.36	0.53	0.46	0.57
187.5	0.7	1.2	-23.7	308.4	27.6	0.06	0.56	0.36	0.53	0.45	0.56
221.5	0.8	1.1	-24.4	157.9	35.1	0.12	0.54	0.28	0.47	0.42	0.51
278.5	1.0	1.2	-23.2	283.6	22.7	0.05	0.54	0.36	0.51	0.45	0.54
326.5	1.2	1.0	-22.6	361.7	31.1	0.05	0.53	0.42	0.56	0.47	0.60
371.5	1.4	1.1	-22.6	374.1	26.5	0.05	0.53	0.41	0.55	0.47	0.58
429.0	1.6	1.0	-22.6	399.6	29.2	0.05	0.55	0.42	0.56	0.49	0.59
480.0	1.8	1.1	-23.4	242.3	18.0	0.05	0.54	0.41	0.55	0.48	0.57
502.0	1.9	1.1	-23.9	365.1	25.4	0.04	0.52	0.42	0.56	0.49	0.58
522.0	2.0	1.0	-23.4	376.8	30.9	0.05	0.55	0.39	0.55	0.48	0.57
550.0	2.4	1.0	-23.0	363.1	26.7	0.05	0.54	0.43	0.57	0.50	0.59
570.0	2.7	0.7	-22.6	493.6	34.3	0.04	0.53	0.43	0.57	0.50	0.59
630.0	3.6	0.7	-22.4	389.6	25.7	0.04	0.53	0.44	0.58	0.51	0.60
686.0	4.5	0.5	-22.4	640.2	40.7	0.04	0.54	0.45	0.59	0.51	0.60
728.0	5.1	0.5	-22.1	100.1	6.7	0.04	0.50	0.45	0.57	0.51	0.58
771.0	5.8	0.7	-23.3	620.7	37.3	0.04	0.53	0.44	0.58	0.50	0.59
805.0	6.4	0.3	-22.1	414.0	25.9	0.04	0.52	0.45	0.58	0.51	0.58
869.5	7.6	0.3	-22.3	538.9	35.7	0.04	0.54	0.41	0.57	0.50	0.56
925.5	8.7	0.3	-23.1	409.4	30.1	0.04	0.52	0.46	0.59	0.51	0.58
Lower Setúbal Canyon sediments (64PE269-39-4) depth (cm)											
1.0	0.0	0.8	-23.5	265.3	20.3	0.05	0.68	0.38	0.73	0.60	0.77
20.0	1.3	0.7	-22.3	506.5	15.6	0.02	0.60	0.39	0.77	0.60	0.80
40.0	2.3	0.6	-22.0	751.5	18.1	0.02	0.58	0.39	0.73	0.61	0.75
60.0	3.3	0.8	-22.2	540.7	16.2	0.02	0.56	0.46	0.75	0.61	0.74
80.0	4.3	0.8	-22.3	460.5	13.5	0.02	0.56	0.41	0.75	0.63	0.77
100.0	5.3	0.8	-22.8	204.7	7.3	0.03	0.57	0.37	0.74	0.66	0.75
120.0	6.5	0.7	-22.8	199.2	8.0	0.03	0.57	0.38	0.77	0.66	0.78
140.0	7.7	0.5	-26.1	433.3	13.6	0.02	0.51	0.41	0.73	0.67	0.74
160.0	8.8	0.5	-22.8	395.6	16.9	0.03	0.45	0.43	0.69	0.64	0.64
180.0	10.0	0.4	-24.9	709.2	25.6	0.02	0.48	0.43	0.69	0.58	0.65
200.0	11.2	0.5	-25.7	703.9	22.1	0.02	0.54	0.41	0.66	0.52	0.64
220.0	11.6	n/a	n/a	n/a	n/a	0.02	0.48	0.47	0.66	0.52	0.59
240.0	12.1	n/a	n/a	n/a	n/a	0.01	0.56	0.57	0.70	0.56	0.63
260.0	12.6	n/a	n/a	n/a	n/a	0.03	0.58	0.49	0.71	0.58	0.64
280.0	13.0	n/a	n/a	n/a	n/a	0.01	0.52	0.48	0.69	0.57	0.65

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Figure 1

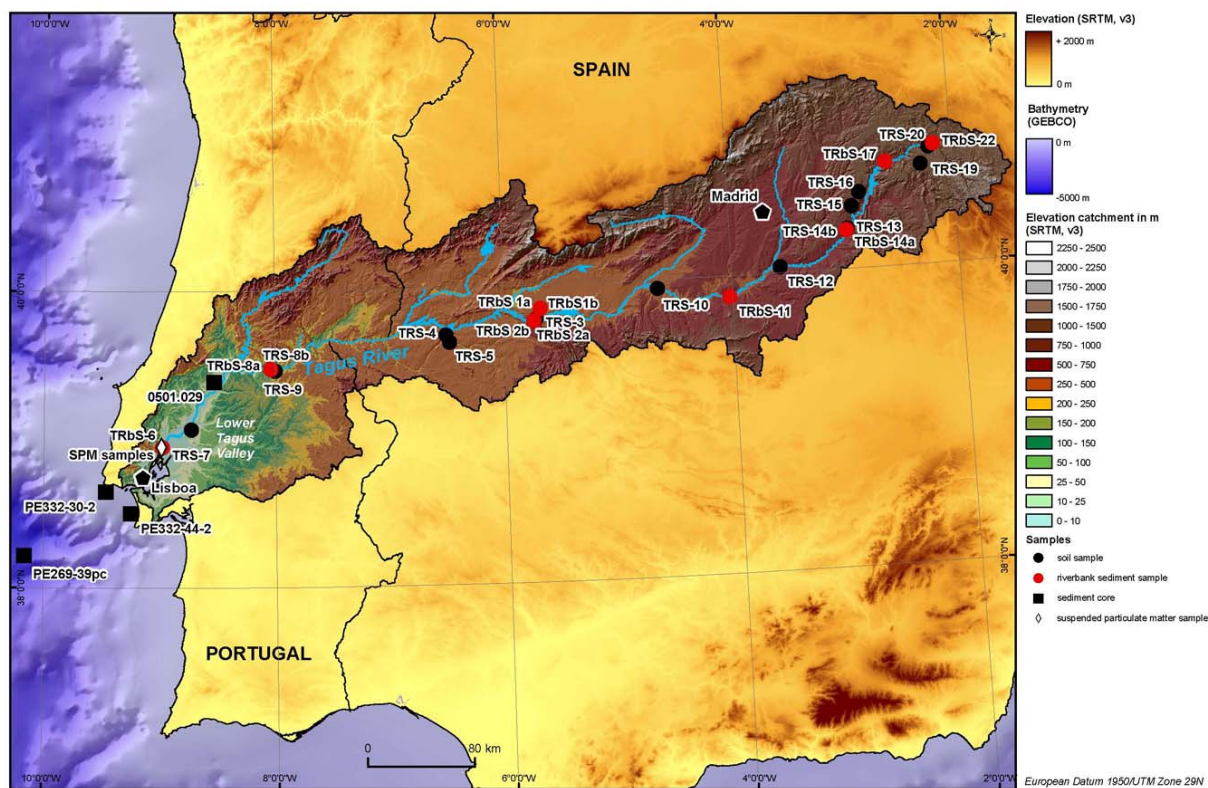




Figure 2

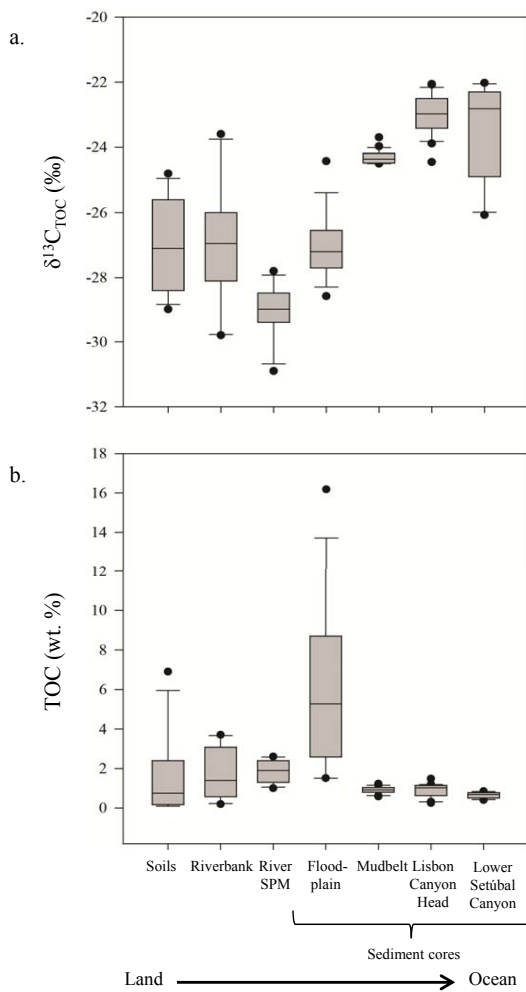




Figure 3

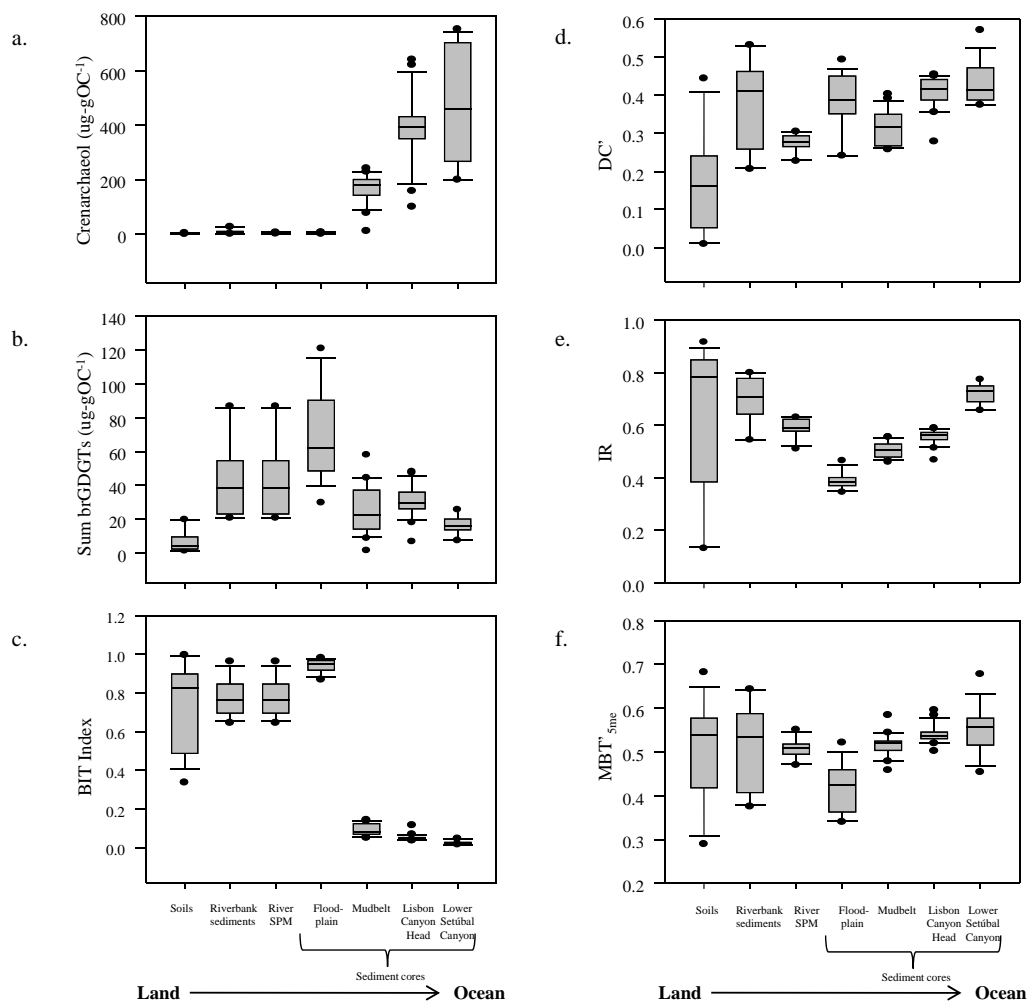




Figure 4

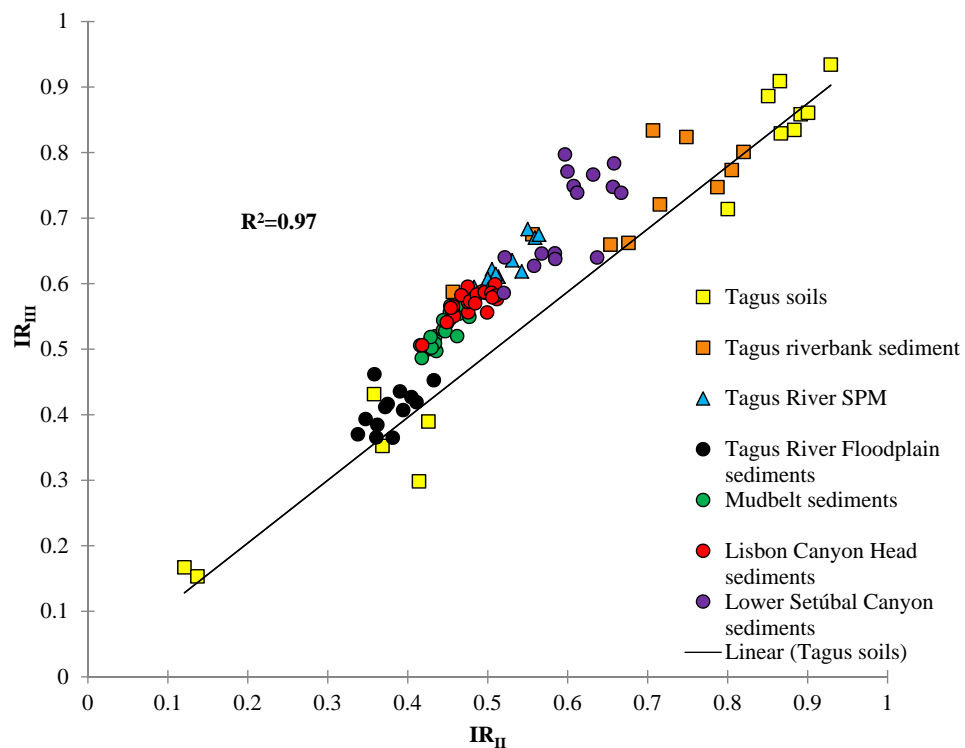




Figure 5

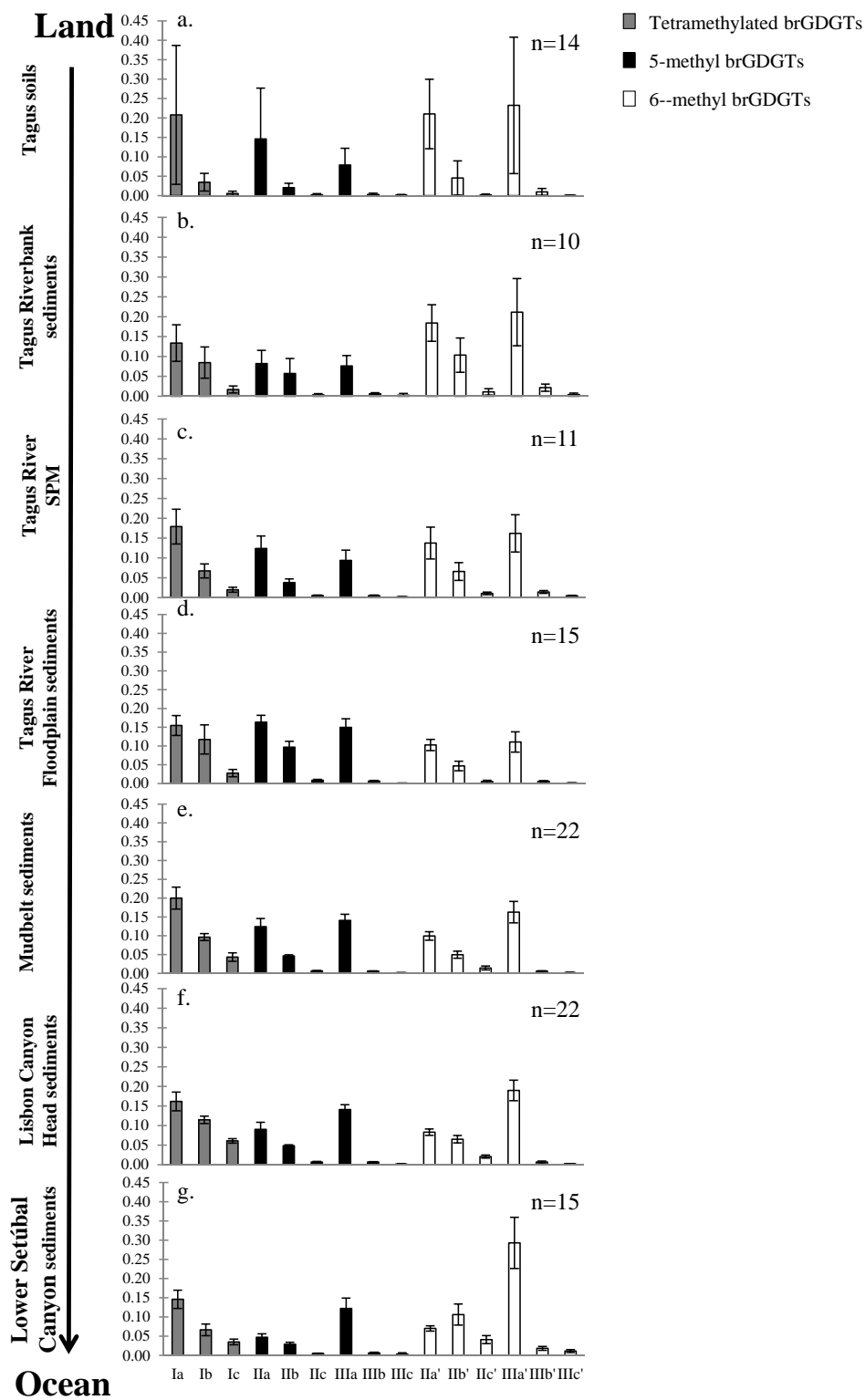




Figure 6

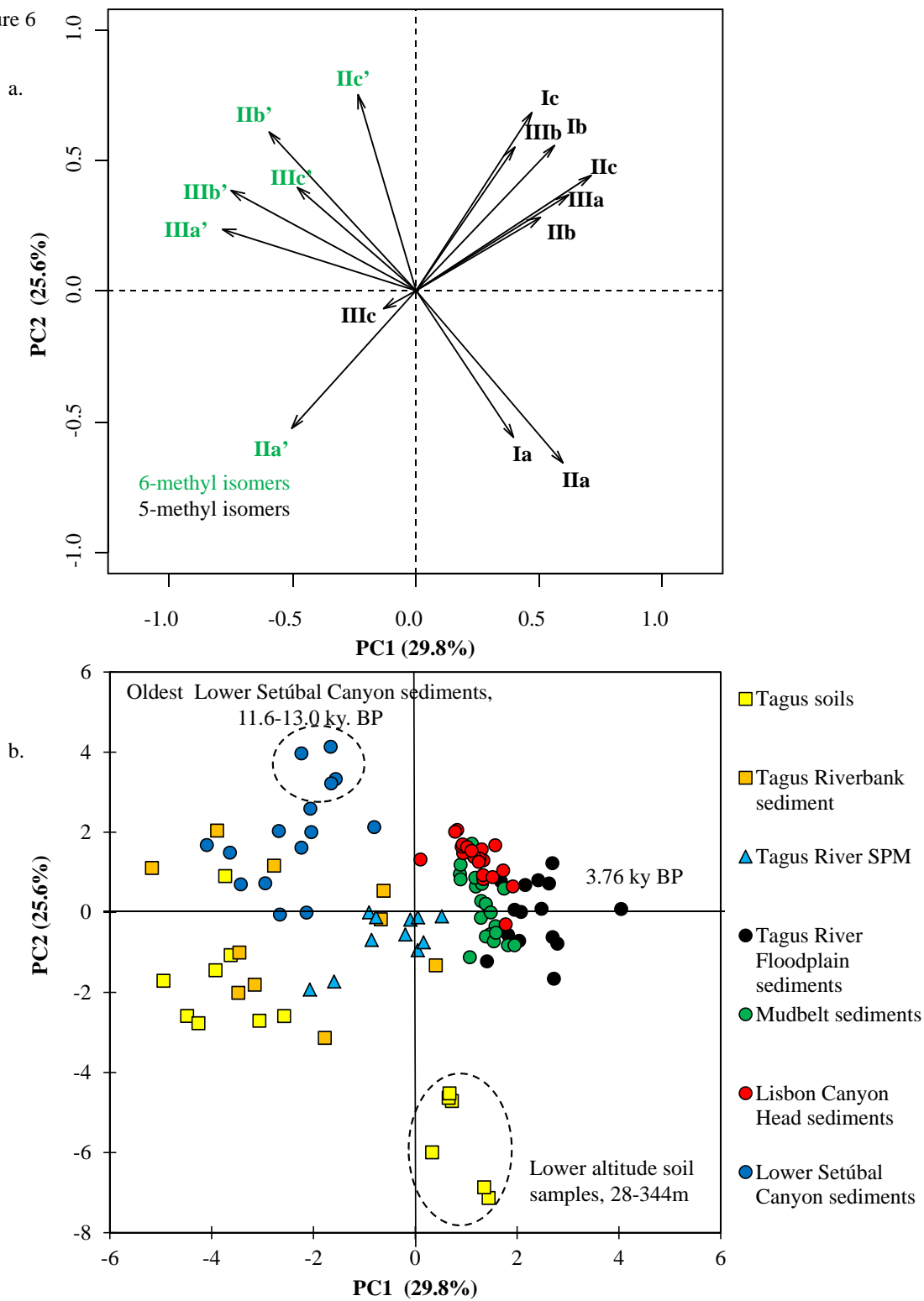




Fig. 7

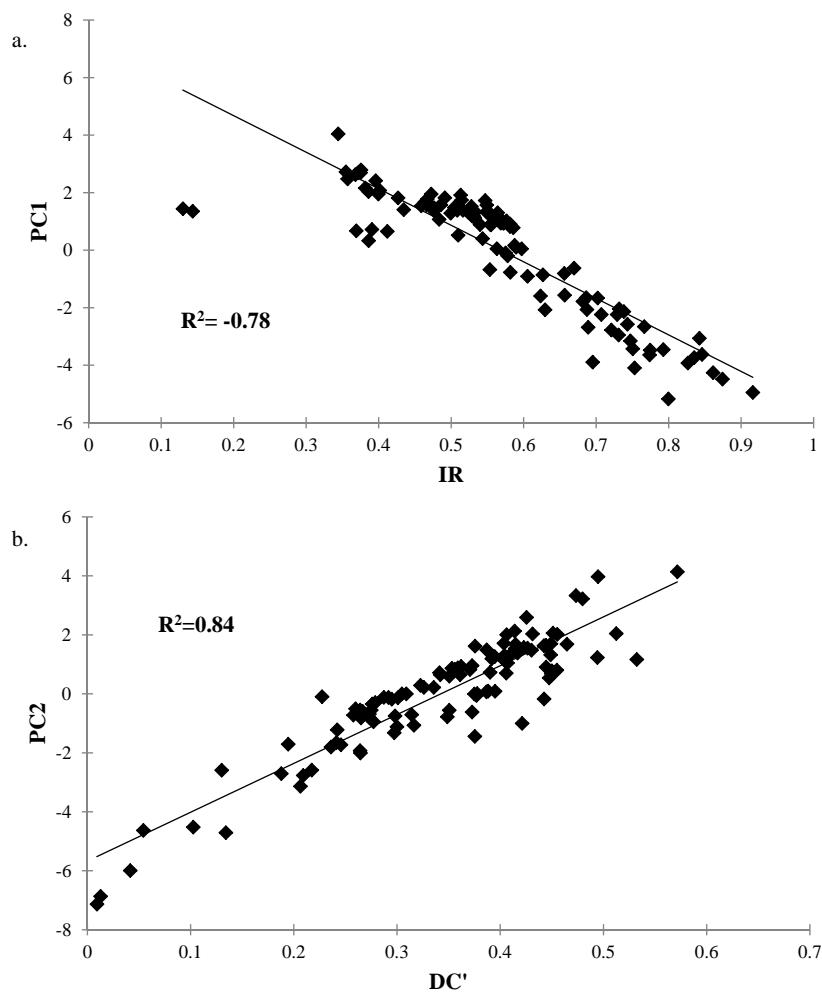




Figure 8

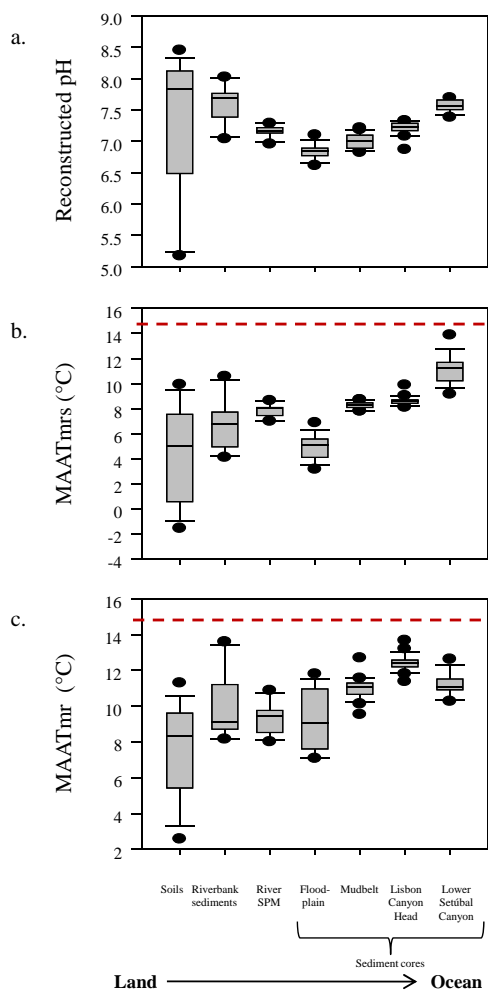




Figure 9

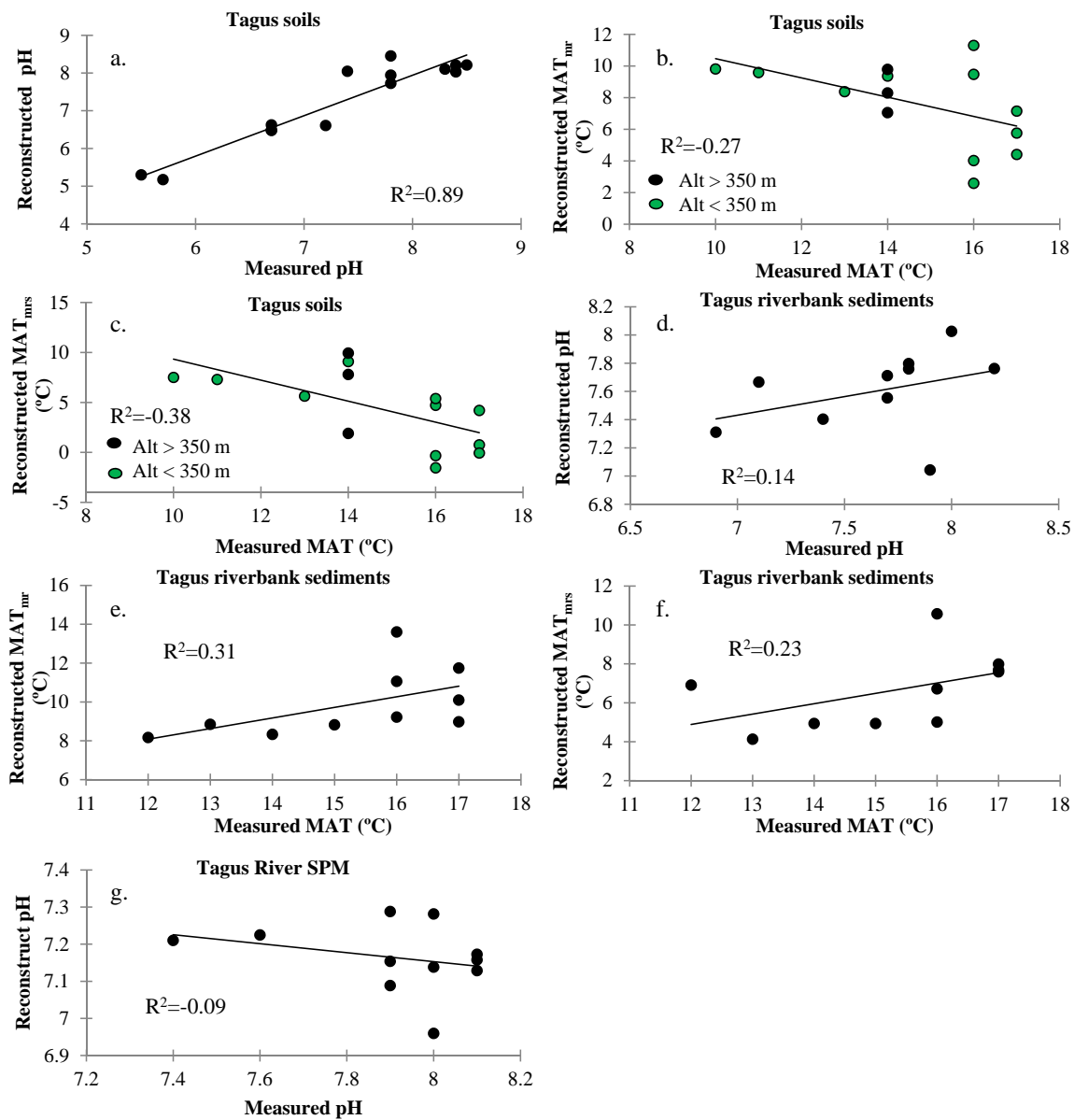




Figure 10

