

1 **Effect of the silica content of diatom prey on the production, decomposition and**
2 **sinking of fecal pellets of the copepod *Calanus sinicus***

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11

12 **Abstract**

13 The effects of changing the amount of silica in the cell wall of diatom prey, on
14 the production, decomposition rate and sinking velocity of fecal pellets of the
15 calanoid copepod, *Calanus sinicus*, were examined. Using different light intensities to
16 control the growth of the diatom *Thalassiosira weissflogii* also led to the
17 accumulation of different amounts of biogenic silica. Copepods were then fed with
18 either low (~1600 cells L⁻¹) or high (~8000 cells L⁻¹) concentrations of this diatom.
19 Copepods fed on a high concentration of diatoms with high-silica content, exhibited a
20 lower grazing rate and lower fecal pellet production rate than those fed on a high
21 concentration of diatoms with low-silica content. However, there was no difference in
22 either the grazing or fecal pellet production rates at low prey concentrations with
23 high- or low-silica content. The size of the fecal pellets produced was only affected
24 by the prey concentration, and not by the silica content of prey. In addition, the
25 degradation rate of the fecal pellets was much higher for copepods fed a low-silica
26 diet than for those fed on a high-silica diet. Significantly lower densities and sinking
27 rates only occurred in the fecal pellets of copepods fed a low-silica diet and a low
28 prey concentration. Calculating the L-ratio (the ratio of degradation rate:sinking rate)
29 for each group indicated that the fecal pellets produced by copepods fed on highly
30 silicified diatoms are likely to transport both biogenic silica and organic carbon to the
31 deep layer; whereas those produced following the consumption of low-silica diatoms
32 are likely to decompose in the mixing layer.

33

34

35 **Introduction**

36 In the marine environment, zooplankton fecal pellets constitute ~~athe~~ the main vehicle
37 for transporting biogenic elements to the sediments, although a substantial proportion
38 of this flux is recycled or repackaged in the water column by microbial decomposition
39 and zooplankton coprophagy (Turner, 2002; 2015). Diatoms are ~~among~~ the most
40 abundant phytoplankton, and they represent ~~athe~~ the main component in the diet of
41 zooplankton in marine environments. Studies show that zooplankton with a diatom
42 diet usually produce fecal pellets that sink faster than those on other diets (Feinberg
43 and Dam, 1998). Dagg et al. (2003) reported that the contribution of fecal pellets to
44 the flux of particulate organic carbon (POC) and biogenic silica (bSi) is higher during
45 the spring diatom bloom than during the summer within the Antarctic Polar Front
46 region. Similarly, Goldthwait and Steinberg (2008) reported an increase in
47 mesozooplankton biomass and fecal production and flux inside cyclonic and
48 mode-water eddies. However, González et al. (2007) reported a negative correlation
49 between the vertical carbon flux of diatoms and the production of fecal material in a
50 time-series study in the upwelling waters off Chile.

51 The quantity and characteristics of the fecal pellets produced by zooplankton
52 depend on several factors. The pellet production rate is reported to be affected by the
53 rate of ingestion and assimilation efficiency (Butler and Dam, 1994; Besiktepe and
54 Dam, 2002). It has also been demonstrated that the type of diet can affect the
55 characteristics of the fecal pellets produced; including size, density and sinking rates
56 (e.g., Feinberg and Dam, 1998 and ref. therein). In addition, the decomposition rate of
57 pellets varies with water temperature, as well as with both microbial and metazoan
58 activity (Poulsen and Iversen, 2008; Svensen et al., 2012). Factors that contribute to
59 the sinking velocity of the pellets include size, density and shape, all of which can
60 vary dramatically both among different zooplankton species and within the same

61 zooplankton species feeding on different types of prey (Fowler and Small, 1972;
62 Turner, 1977; Feinberg and Dam, 1998). Turbulence in the water column, the
63 presence or absence of a peritrophic membrane, and the production of microbial gas
64 within a peritrophic membrane might also affect the sinking rate of pellets (Honjo and
65 Roman, 1978; Bathmann et al., 1987). Indeed, the sinking rate and decomposition rate
66 are the two most important parameters used, to determine whether a pellet will or will
67 not be successfully transported into deeper water before its contents are degraded. For
68 example, a slowly-sinking pellet is more likely to decompose and become part of the
69 recycled materials before it exits the euphotic zone (Dagg and Walser, 1986).

70 The cell wall (frustrule) of diatoms is composed of two silicate shells, which are
71 believed to act as a defense mechanism to prevent ingestion by grazers (Pondaven et
72 al., 2007); thus different levels of silicification of the frustrule might affect the grazing
73 rate of copepods (Friedrichs et al., 2013, Liu et al., in revision). The silica content of
74 the cell wall of diatoms is not only species-specific, but it is also affected by
75 environmental parameters such as light, temperature, salinity, pH, nutrients and trace
76 metals (Martin-Jézéquel et al., 2000 and ref. therein; Claquin et al., 2002; Vrieling et
77 al., 2007; Herve et al., 2012; Liu et al., in revision). Although the frustule has no
78 nutritional value for zooplankton, it is thought to provide ballast, which is especially
79 advantageous when the fecal pellets are sinking. Hence, pellets with a high diatom
80 biomass generally exhibit higher levels of export of POC (Armstrong et al., 2002;
81 François et al., 2002; Klaas and Archer, 2002). Thus, the content of the zooplankton
82 diet (and therefore the type and concentration of ballast minerals ingested) might
83 strongly affect the sinking velocity of the fecal pellets produced, and hence the
84 vertical flux of biogenic silica and carbon.

85 Most of the studies describing the production rates and characteristics of
86 copepod fecal pellets have focused on aspects such as food types (Feinberg and Dam,

87 1998), or the different periods of phytoplankton blooms (Butler and Dam, 1994).
88 There are currently no reports that describe the effect of the silica content of diatoms
89 on the production, degradation and sinking of fecal pellets. Liu et al. (~~under-~~
90 ~~review~~2016) recently demonstrated that the diatom *Thalassiosira weissflogii*, when
91 grown at different light levels, contains varying amounts of silica, and that the small
92 calanoid copepod *Parvocalanus crassirostris*, when fed on diatoms containing high
93 levels of silica exhibited a reduced feeding rate, and stagnant growth as well as low
94 egg production and hatching success. In this study we used the same diatom species
95 with different silica content as prey to study the characteristics of the fecal pellets
96 produced by the herbivorous copepod, *Calanus sinicus*.

97

98 **Materials and Methods**

99 **Copepod and prey culture conditions.** The herbivorous copepod *Calanus*
100 *sinicus* was collected from the coastal waters around Hong Kong in February 2013.
101 ~~Copepods~~They were maintained on a 14 h light:10 h dark cycle at 23.5°C in 2 L glass
102 containers with 0.2 µm-filtered seawater. The copepods were fed a mixed algal diet
103 consisting of *Rhodomonas* sp. and *Thalassiosira weissflogii* at a concentration of
104 ~5000 cells L⁻¹; this food suspension was supplied to the cultures twice a week and
105 the whole culture seawater was replaced every week. The copepods were maintained
106 for more than one month prior to the start of the experiment to ensure that all the
107 adults were grown in approximately the same conditions and were of approximately
108 the same age.

109 The diatom, *T. weissflogii*, was maintained in exponential growth in *f/2* medium
110 (Guillard, 1975), under light intensities of either 15 µmol photons s⁻¹ m⁻² or 200 µmol
111 photons s⁻¹ m⁻² to generate cells with different cellular silica contents (Liu et al., ~~under-~~
112 ~~review~~2016). The diatom cultures were transferred every 4 or 8 days for the high- and

113 low-light batches, respectively. After two transfers the amount of biogenic silica in
114 the diatom cells was measured using a modified version of the method described by
115 Paasche (1980), following the procedures described more recently by Grasshoff et al.
116 (1999). Cells were collected on a 1 μm polycarbonate filter (47mm diameter) and
117 washed with 10 ml autoclaved seawater and 0.01M HCl during filtration to remove
118 the intercellular silicate pools. The folded filter was immediately placed into a 15 ml
119 polypropylene tube and stored at -80°C . Hydrolysis was carried out using 4ml of 5%
120 NaOH, digested at 85°C for 2 hours. After cooling, 0.72 ml of 1.0 M HCl was added
121 to each tube, lowering the pH to 3-4. Silicic acid concentration of samples was
122 determined colourmetrically, through the formation of blue coloured silicocomplexes.

123

124 **Experimental design.** Active adult female *Calanus sinicus* with intact
125 appendages were selected and starved for 24 hours before an experiment. A total of
126 seven experiments was conducted to determine fecal pellet production, degradation
127 and sinking, and in each experiment these parameters were measured both at low and
128 high food concentrations, and at high and low levels of silica contained in the diatom
129 prey (Table 1). In each experiment, the copepods were fed with the same species of
130 diatom (i.e., *T. weissflogii*), at either ca. 1600 cells L^{-1} (low concentration), or ca.
131 8000 cell L^{-1} (high concentration), the latter being above the food saturation level
132 according to Frost (1972). The abundance and volume of diatoms were measured
133 (triplicate subsamples) using a Beckman Coulter Z2 Particle Counter and Size
134 Analyzer.

135 In the fecal pellet production experiments, five replicate bottles containing one
136 copepod per bottle, and two control bottles without a grazer, were used. All the bottles
137 were filled with 100 ml freshly-prepared media consisting of 0.2 μm prefiltered
138 seawater and suspensions of the respective prey for each treatment. All incubations

139 were conducted at 23.5°C and in the dark for 24 hours. At the end of the incubation
140 period, a 2 ml sample was collected from each bottle and fixed with acid Lugol's at a
141 final concentration of 2%, for subsequent diatom quantification. The remaining water
142 was collected in a 50 ml polypropylene tube and fixed with glutaraldehyde at a final
143 concentration of 1%, for further quantification of the fecal pellets.

144 In order to obtain fresh pellets for the degradation experiments, two plastic
145 beakers were prepared for the high- and low-silica content prey. Each beaker
146 contained 7-8 copepods and 700 ml culture medium, prepared as described for the
147 production experiments. After 12 hours of incubation (except for experiment # 3,
148 which was incubated for 18 hours), the medium was sieved through a 40 µm mesh to
149 collect the fecal pellets and then rinsed with autoclaved 0.22 µm filtered seawater. At
150 least 20 intact fecal pellets were selected using a glass Pasteur pipette under a
151 stereomicroscope and poured into a 250 ml polycarbonate bottle containing 200 ml of
152 2 µm pre-filtered sea water taken from the field. The number of replicate bottles and
153 the incubation period of each experiment are show in Table 2. All the bottles were put
154 on a roller at 0.4 rpm in the dark at 23.5°C and then at the end of the respective
155 incubation times, the whole water of each bottle was collected in a plastic bottle and
156 fixed with glutaraldehyde at a final concentration of 1% for further fecal pellet
157 analysis.

158 Experiments to estimate the fecal pellet sinking rate were conducted by obtaining
159 fecal pellets using the degradation experiment procedure (described above) but with
160 an incubation time of 24 hours. After collecting all the fecal pellets from the beakers,
161 50 intact pellets were selected and suspended in 260 ml 0.2 µm prefiltered autoclaved
162 seawater. The fecal pellet sinking rate was measured using a SETCOL chamber (49
163 cm height, 2.6 cm inner diameter) made by 4 mm Plexiglas (Bienfang, 1981), filled
164 with well-mixed pellet-containing seawater. The chamber was allowed to settle for 6

165 min, and then the whole column of water was collected from outflow tubes in a
166 top-to-bottom order. The water was collected in a plastic bottle and fixed with
167 glutaraldehyde as described above, for subsequent fecal pellet analysis.

168

169 **Determining the number and size of fecal pellets.** The water samples
170 containing the fecal pellets in the 50-ml polypropylene tubes were allowed to settle
171 for 24 hours. The upper water was then removed smoothly and the remainder was
172 poured into the well of a 6-well plate and the number of pellets was counted using an
173 inverted microscope (Olympus IX51) at 100× magnification. Only intact fecal pellets
174 and fragments with end points were counted. The total number of fecal pellets was
175 then calculated to include all of the intact fecal pellets plus half of the pellet fragments.

176 Images of at least 30 intact fecal pellets were acquired with a CCD camera ([Model 4.2,](#)
177 [Diagnostic Instrument Inc., USA](#)), after which the length and width of each fecal
178 pellet was measured and the volume was calculated making the assumption that they
179 are cylindrical in shape.

180

181 **Calculating the fecal pellet degradation rate.** The rate of degradation of the
182 fecal pellets was calculated from the loss of fecal pellet equation, described by:

$$183 \quad N_t = N_0 e^{-rt}$$

184 where N is the total number of fecal pellets in the incubation bottle at the
185 beginning (N_0) and end of the experiment (N_t); t is the incubation time (in days); and r
186 is the degradation rate (d^{-1}). The degradation rate estimated in this study only
187 considered the effect of microbial organisms and assumed that the loss rate was
188 exponential.

189

190 **Calculating the fecal pellet sinking velocity.** The rate that fecal pellets sank
191 was calculated from the formula reported by Bienfang et al. (1982), which was
192 originally used to measure the average sinking rate of phytoplankton. Thus:

$$193 \quad S = \frac{N_s}{N_T} \times \frac{L}{t}$$

194 where S is the average sinking velocity; L is the height of the sinking column; t is the
195 duration of the trial; N_T is the total number of fecal pellets within the settling water
196 volume; and N_s is the total number of fecal pellets that settled during the trial time.

197 In addition, the density of the fecal pellets was calculated using the
198 semi-empirical equation deduced by Komar (1980), as follows:

$$199 \quad w_s = 0.079 \frac{1}{\mu} (\rho_s - \rho) g L^2 \left(\frac{L}{D} \right)^{-1.664}$$

200 where w_s is the sinking velocity of the fecal pellets; μ and ρ are the fluid viscosity and
201 density, respectively; L and D are the length and diameter of the fecal pellets,
202 respectively, assuming they are in the cylindrical shape; g is the acceleration of
203 gravity; and ρ_s is the density of fecal pellet.

204

205 **Results**

206 **Grazing response**

207 The cellular silica content of first and second generation *T. weissflogii* when
208 cultured at high and low light intensities is shown in Fig. 1. After two transfers the
209 cellular biogenic silica content was significantly different (t-test, $p < 0.05$; Fig. 1) when
210 comparing the high-light and low-light culture conditions. The silica content of high-
211 and low-silica diatoms used in all the experiments was consistent and the differences
212 between the two treatments were all statistically significant (Table 1). Other cellular
213 parameters, such as cell size and carbon and nitrogen contents, were also measured
214 for selected samples (data not shown), and the results were consistent with those
215 reported in a previous study (Liu et al., 2016), which showed no significant difference
216 between the two types of prey.

217 The grazing response of *C. sinicus* to diatoms with different silica contents
218 showed similar patterns between high (ca. 8000 cells ml^{-1}) and low (ca. 1600 cells
219 ml^{-1}) prey concentration (Fig. 2). At high concentrations of prey, *C. sinicus* grazed the
220 diatoms with low cellular silica content two times faster than when they had a high
221 silica content (t-test, $p < 0.05$). The same trend was also observed at low concentrations
222 of the prey, although in this case the difference was not statistically significant. In
223 addition, the rate of clearance was significantly higher for the low-silica prey than for
224 the high-silica prey at both low and high prey concentrations (t-test, $p < 0.05$). These
225 results indicate that the silica content of diatoms can affect the grazing activity of
226 copepods.

227

228 **Fecal pellet production**

229 The rate of fecal pellet production varied both with the silica content and the
230 concentration of the prey (Fig. 3A). At a high prey concentration, *C. sinicus* that were

231 fed on low-silica prey produced significantly higher amounts of fecal pellets (192 ± 32
232 FP ind⁻¹ d⁻¹) than those fed on high-silica prey (113 ± 47 FP ind⁻¹ d⁻¹, $p < 0.05$); which
233 corresponds well with the rate of ingestion (Fig. 2A and 3A). At a low prey
234 concentration, however, the production of fecal pellets by *C. sinicus* fed with the low
235 and high-silica prey was not significantly different (Fig. 3A). In addition, the size of
236 the fecal pellets was only affected by the concentration of the prey, and not by the
237 silica content of the prey (Fig. 3B). Thus, the fecal pellets produced in the high
238 concentration of prey groups had a mean length and width of 582.4 ± 98.7 μm and
239 72.5 ± 4.5 μm , respectively, which are significantly larger than the size of those
240 produced in the low concentration of prey groups, which had an average length and
241 width of 352.4 ± 54.7 μm and 59.6 ± 6.8 μm , respectively (~~t-test~~ANOVA, $p < 0.05$).

242

243 **Fecal pellet degradation rate and sinking rate**

244 The degradation rate of fecal pellets was significantly different when the
245 copepods fed on diatoms with different silica content (Table 2). The degradation rate
246 of the fecal pellets produced from the low-silica prey was approximately 4-5-fold
247 higher than that of the pellets generated from the high-silica prey, irrespective of the
248 prey concentration or the period of degradation incubation. In addition, the
249 degradation rate of the fecal pellets from low prey concentration was significantly
250 higher than ones from high prey concentration after an incubation period of 24 hr (p
251 < 0.05 , ~~t-test~~ANOVA). Furthermore, the degradation rate obtained following 48 h
252 incubation was significantly higher than that following just 24 h incubation (only high
253 prey concentration experiments) for both the high ($p < 0.05$, t-test) and low ($p < 0.01$,
254 t-test) silica prey (Table 2), indicating an acceleration of degradation in the second
255 day of incubation.

256 The sinking rate of fecal pellets was also different for the high and low prey

257 concentrations (Fig. 4). At a high concentration of prey, the sinking rates of the pellets
258 produced by the high- and low-silica prey (i.e., 3.05 and 3.13 cm min⁻¹, respectively),
259 were not significantly different. However, at a low prey concentration, the sinking
260 rate of pellets from the high-silica-content prey (i.e., 2.59 cm min⁻¹) was
261 significantly greater (t-test, p<0.01) than that of pellets from the low-silica-content
262 prey (i.e., 0.53 cm min⁻¹). The average density of the fecal pellets was calculated as
263 being 1.093-1.095 g cm⁻³ at the high prey concentration, and 1.035-1.097 g cm⁻³ at the
264 low prey concentration. The variation in the calculated density of fecal pellets is
265 consistent with the pattern of sinking rate, with the lowest density occurred in fecal
266 pellets from low-silica prey at the low prey concentration (Fig. 4).

267

268 Discussion

269 The grazing activity of copepods varies not only with the concentration of the
270 prey but also with the nutritional quality of the prey. In our study, the grazing and
271 clearance rates determined with the varying food concentrations, followed a similar
272 trend to that described in the literature (e.g., Frost, 1972). In addition, the grazing
273 activity was affected by the cellular silica content of the prey, as has been observed
274 with other copepod species (Liu et al., [under review 2016](#)). Silicification ~~is~~ has been
275 suggested to be one of the strategies that is used by diatoms to protect them from
276 ingestion by grazers (Pondaven et al., 2007). Friedrichs et al. (2013) examined the
277 mechanical strength of the frustules of three diatom species and measured the feeding
278 efficiency of copepods on these diatoms. Their results showed that the diatom species
279 with the more weakly-silicified frustules and the highest growth rate was the least
280 stable and was fed upon the most, whereas the species with the most complex frustule
281 exhibited the greatest stability and was fed upon the least. Within the same species of
282 diatom, different growth rates have resulted in different amounts of silica in the

283 frustule (Claquin et al., 2002). This results in higher copepod ingestion and clearance
284 rates for diatoms with a low silica content when compared with those for diatoms with
285 a higher silica content (Liu et al., ~~2016~~under review). The results obtained in ~~this new-~~
286 ~~the current~~ study are consistent with those reported by Friedrichs et al. (2013) and Liu
287 et al. (~~2016~~under review).

288 Previous studies indicate that while there is a linear relationship between the
289 ~~ingestion rate and the~~ total number of fecal pellets produced ~~in per~~ unit time ~~and the-~~
290 ~~ingestion rate~~ (Ayukai and Nishizawa, 1986; Ayukai, 1990), there is a high level of
291 variation among different diets (Båamstedt et al., 1999 and ~~the references~~ therein;
292 Besiktepe and Dam, 2002). In addition, the size of ~~the~~ fecal pellets increases as the
293 concentration of the food increases, such that they reach a maximum size when the
294 concentration of food is above the saturation level (Dagg and Walser, 1986; Butler
295 and Dam, 1994). Our results confirmed these previous findings and demonstrated that
296 the size of ~~the fecal~~ pellets produced was only affected by the concentration of prey,
297 and ~~fecal pellets~~~~they~~ did not show any significant size differences when comparing
298 prey of high and low cellular silica content. Butler and Dam (1994) reported that
299 when sufficient food was available, the size of the fecal pellets varied with the
300 nutritional quality (e.g., the C:N ratio) of the prey. Since diatoms with different silica
301 content (generated by varying the light intensity) do not differ in their cellular C:N
302 ratio (Claquin et al., 2002; Liu et al., ~~under review~~2016), ~~they these ratios diddo~~ not
303 affect the size of the pellets produced.

304 The degradation rate and sinking velocity of the fecal pellets are highly
305 dependent on the characteristics of the pellets, which are in turn affected by the
306 quality and quantity of the food ingested (Feinberg and Dam, 1998; Turner, 2002;
307 2015 and references therein). For example, it is known that the decomposition rate of
308 the fecal pellets is affected by diet, pellet size and the producer of the pellets (e.g.,

309 Shek and Liu, 2010), but no research ~~mentions have addressed~~ the degradation rates
310 of fecal pellets produced by prey under different stoichiometric conditions. Hansen et
311 al. (1996) estimated the degradation rate of fecal pellets produced from diets of
312 *Thalassiosira weissflogii*, a diatom; *Rhodomonas baltica*, a nanoflagellate; or
313 *Heterocapsa triquetra*, a dinoflagellate. ~~F~~~~They showed that the~~ fecal pellets produced
314 from a diet of the diatom species presented the slowest rate of degradation when
315 compared with those produced from diets of the nanoflagellate or dinoflagellate
316 species. Similarly, Olesen et al. (2005) compared the degradation rate of fecal pellets
317 produced on a diet of the diatom, *Skeletonema costatum*, or the nanoflagellate,
318 *Rhodomonas salina*, and reported a similar trend but higher degradation rates than
319 Hansen et al. (1996). The relationship between the surface:volume ratio and the
320 degradation rate of fecal pellets was used to explain the variation in the degradation
321 rate of pellets produced with different diets. Our results (Table 2) were higher than
322 those reported by Hansen et al. (1996), which were 0.024 d⁻¹ for *T. weissflogii*, but ~~our~~
323 ~~results~~~~they~~ showed a similar trend to those summarized by Olesen et al. (2005)
324 (dashed line in Fig. 5), in that there was an increase in the degradation rate with the
325 increase in fecal pellet surface:volume ratio, although the degradation rates that we
326 measured, exceeded the predicted rates in most cases, particularly ~~those for fecal~~
327 ~~pellets~~ produced with low--Si diatom prey (Fig. 5). The generally higher rates in our
328 study might be caused by the higher temperature that we used when compared with
329 the previous studies (i.e., 23.5°C in our study *versus* 17°C and 18°C in Olesen et al.,
330 2005 and Hansen et al., 1996, respectively), but the ~~differences in predator and prey~~
331 ~~quality, particular the role of~~ cellular Si content ~~in this study~~, cannot be ignored.

332 The sinking rate of fecal pellets is usually considered to be related to their size
333 and density, which is in turn dependent on the concentration and composition of the
334 prey (Bienfang, 1980; Urban et al., 1993; Feinberg and Dam, 1998). We also

335 demonstrated that fecal pellet size, sinking rate and density were correlated with the
336 concentration of prey (Fig. 3B, 4), especially in the low-silica diatom prey treatment.
337 Using the ratio of ingestion rate : fecal pellet production rate ratio as an index to
338 compare the diatom content per fecal pellet, no differences were found in pellets
339 produced from diets of the same silica content (Fig. 6), indicating that prey
340 concentration does not affect the package content of the fecal pellets. On the other
341 hand, copepods were shown to pack fewer hard-shelled (i.e., high-Si) diatoms into
342 each fecal pellet in comparison to the soft-shelled (i.e., low-Si) diatoms, although
343 these data were not significantly different statistically (Fig. 6).

344 The fecal pellets of copepods are formed in the midgut surrounded by a
345 peritrophic membrane, which is believed to protect the gut wall from the sharp edges
346 of the prey's cell wall. Moreover, the different sizes of fecal pellets with similar prey
347 content per fecal pellet are thought to result from the decreasing gut passage time
348 with the increasing of food concentration. A high prey concentration results in the
349 food passing through the gut more quickly and results in incomplete digestion,
350 whereas a low prey concentration allows the food to be kept in the intestinal tract for a
351 longer time and therefore digestion is relatively more complete. We showed that the
352 silica content of the diatom cell wall determines the density and sinking rate of the
353 fecal pellets when the prey concentration was low due to complete digestion. In
354 addition, we showed that only the low concentration of low-Si prey group, resulted in
355 a significantly lower fecal pellet density and sinking rate. In previous studies, the
356 sinking rate and density of the fecal pellets of *Calanus* were shown to be 70-171 m
357 day⁻¹ and 1.07-1.17 g cm⁻³, respectively (Bienfang, 1980; Urban et al., 1993), which
358 are considerably higher than our results (Fig. 4). We suggest that these differences
359 might be caused by the differences in methodology used (Griffin, 2000).

360 To compare the combined effects of sinking and degradation rates for each

361 treatment, the reciprocal length scale, or L-ratio, which is the fraction of pellet
362 degradation per unit length traveled, was calculated (Feinberg and Dam, 1998). The
363 product of the L-ratio multiplied by the depth of the mixed layer can then be used to
364 provide the degree of degradation of a pellet within this layer. The results from such
365 calculations suggest that some diets might result in pellets that are substantially
366 recycled within the epipelagic layer whereas others result in pellets that are exported
367 out of the mixed layer in a relatively non-degraded manner. It should be pointed out,
368 however, ~~that~~ the degradation rates we calculated are likely to be highly
369 underestimated due to the absence of zooplankton activities. For example, it has been
370 reported that copepod ingestion of entire fecal pellets (i.e., coprophagy~~ia~~) or ~~the~~ only
371 partial break down of fecal pellets might dramatically reduce the overall downward
372 transport of fecal material and thus increase its retention in the epipelagic layer
373 (Lampitt et al., 1990; Gonzalez and Smetacek, 1994; Svensen et al., 2012). For the
374 same reason, plus the absence of turbulence in our experimental set-up, our sinking
375 rate measurements are likely to be overestimated. Nevertheless, the L-ratio provides a
376 relative indicator of the export efficiency of the fecal pellets produced on diatom diets
377 of different silica content and can be used for a comparison with copepod fecal pellets
378 produced with other diets. Our results also show that pellets produced from high silica
379 content diatoms are more likely to sink out of the mixed layer before being degraded,
380 when compared with pellets from low silica content diatoms. On the other hand, fecal
381 pellets produced from a low concentration of prey with low ~~-~~Si content are the most
382 likely to be degraded in the ~~mixed euphotic~~ layer (Table 3). Our results suggest that
383 the grazing activity of copepods might result in organic matter being mostly recycled
384 in the ~~mixed~~ layer during the fast ~~-~~growth period of diatoms (e.g., at the beginning
385 of the bloom), whereas it could accelerate the export of POC to the deep ocean by
386 producing fast ~~-~~sinking fecal pellets during the slow ~~-~~growth period of diatoms (e.g.,

387 during the senescent stage of the diatom bloom).

388 In conclusion, the silica content of the cell wall of diatoms can affect the grazing
389 activity of copepods and influence the rates of production-~~rate~~, decomposition-~~rate~~
390 and sinking-~~rate~~ of their fecal pellets. Our findings suggest that it is not only the
391 nutritional quality, but also the digestion process of copepods that can result in the
392 different characteristics of the pellets produced. In addition, it is a combination of
393 both degradation and sinking rates, (which are affected by the abundance and cellular
394 silica content of the diatom prey among other physicochemical factors), that
395 determines the efficiency of the downward export of biogenic silica and organic
396 carbon by fecal pellets.

397

398

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402

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534 Table 1. Summary of the concentration and cellular silica content of the diatom prey
 535 in each experiment.

536

Expt.	Measurements	[Prey]	Silica level	Initial prey density (cells mL ⁻¹)	Cellular silica (pg SiO ₂ cell ⁻¹)
1	Fecal pellet production	High	High	8194 ± 166.9	55.7 ± 1.7
		High	Low	7976 ± 8.5	38.2 ± 1.4
2		Low	High	1640 ± 28.3	51.7 ± 1.9
		Low	Low	1490 ± 84.9	31.4 ± 6.6
3		High	High	8194 ± 166.9	55.7 ± 1.7
		High	Low	7976 ± 8.5	38.2 ± 1.4
4	Fecal pellet degradation*	High	High	7499 ± 63.6	58.9 ± 2.4
		High	Low	7344 ± 169.7	33.4 ± 4.3
5		Low	High	1640 ± 28.3	51.7 ± 1.9
		Low	Low	1490 ± 84.9	31.4 ± 6.6
6	Fecal pellet	High	High	8114 ± 138.0	56.5 ± 5.9
		High	Low	7904 ± 124.7	27.0 ± 0.6
7	sinking	Low	High	1790 ± 48.1	52.1 ± 1.3
		Low	Low	1545 ± 75.0	30.3 ± 3.1

537 The incubation time of the 3 fecal pellet degradation experiments can be found in
 538 Table 3.

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541 Table 2. Degradation rate of the fecal pellets produced by *C. sinicus* after they were
 542 fed on diatoms with different silica content.

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Prey concentration	Incubation period	Silicon status of prey	n	Degradation rate (day ⁻¹)
High	48 hr	HSi	3	0.21±0.15
		LSi	3	0.91±0.17
High	24 hr	HSi	4	0.03±0.04
		LSi	4	0.15±0.02
Low	24 hr	HSi	3	0.08±0.04
		LSi	2	0.38±0.03

544 HSi: high silica content, LSi: low silica content.

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549 Table 3. The L-ratio (m⁻¹), determined as the mean degradation rate constant (t⁻¹),
 550 divided by the mean sinking rate (m d⁻¹), for each treatment.

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Prey silica content	High food concentration	Low food concentration
High Si	3.91×10 ⁻⁴	7.56×10 ⁻⁴
Low Si	1.09×10 ⁻³	1.65×10 ⁻²

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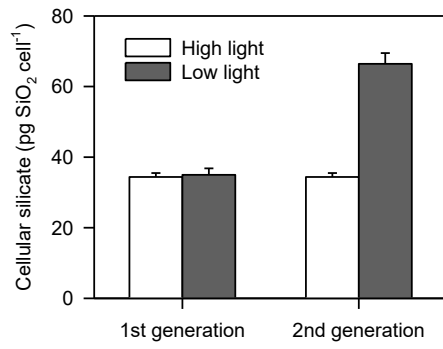
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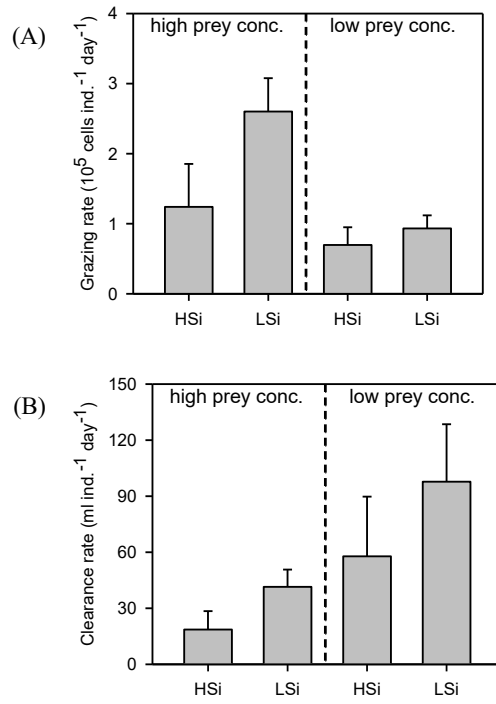
559 Fig. 1. The cellular silica content of *T. weissflogii* grown under different light
560 intensities. The error bars show one standard deviation (n=3).



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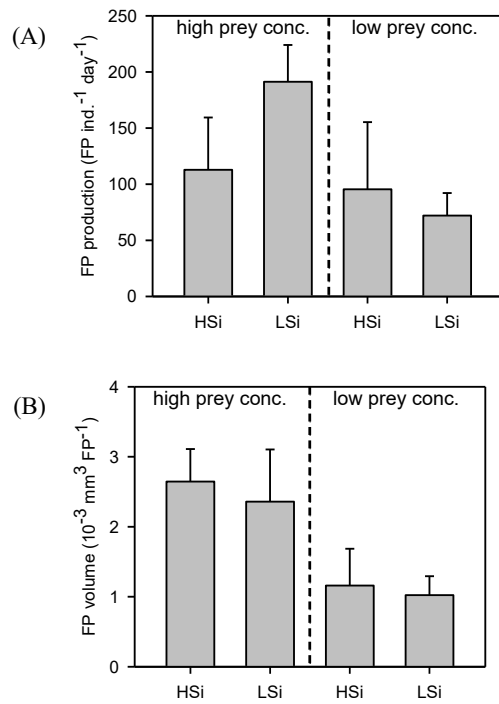
563 Fig. 2. Grazing rate (A) and clearance rate (B) of *C. sinicus* fed on diatoms with
564 different silica content. HSi and LSi are high and low silica diatom prey, respectively.
565 The error bars show one standard deviation (n=5).



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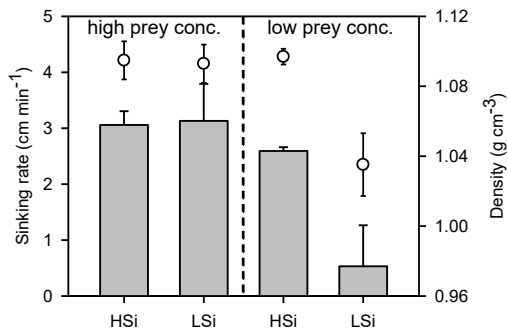
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568 Fig. 3. The rate of fecal pellet production (A), and the average volume of each fecal
569 pellet (B), produced by *C. sinicus*. HSi and LSi indicate high and low silica diatom
570 prey, respectively. The error bars show one standard deviation (n = 5).



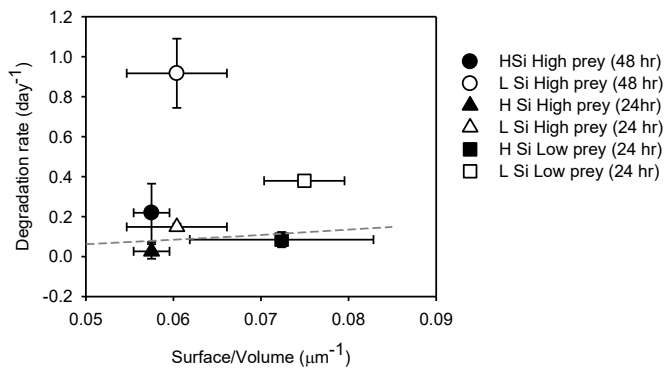
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574 Fig. 4. The sinking rate (bars) and calculated density (open dots) of the fecal pellets
575 generated by *C. sinicus* produced following each treatment. HSi and LSi are high and
576 silica diatom prey, respectively. The errors bar show one standard deviation (n=3).



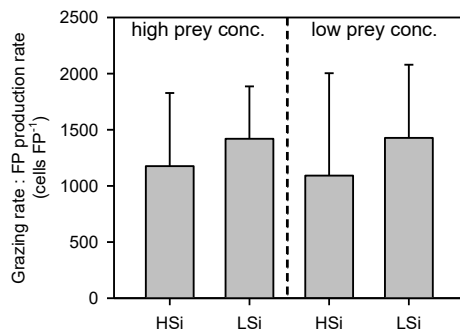
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583 Fig. 5. The relationship between degradation rates and surface:volume ratio of fecal
584 pellets from different experimental treatments. HSi and LSi are high and low silica
585 content diatoms, respectively; high and low prey are high and low prey concentrations,
586 respectively; 48 hr and 24 hr are the incubation periods used for the degradation
587 experiments. The error bars show ± 1 standard deviation and the dashed line shows the
588 relationship curve generalized by Olesen et al. (2005).
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593 Fig. 6. The grazing rate: fecal pellet production rate ratio of each treatment. HSi and
594 LSi are the high and low silica diatom prey, respectively. The error bars show one
595 standard deviation.



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