

**Fate of terrigenous organic matter across the Laptev Sea
from the mouth of the Lena River to the deep sea of the
Arctic interior**

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Abstract

Ongoing global warming in high latitudes may cause an increasing supply of permafrost-derived organic carbon through both river discharge and coastal erosion to the Arctic shelves. Mobilized permafrost carbon can be either buried in sediments, transported to the deep sea or degraded to CO₂ and outgassed, potentially constituting a positive feedback to climate change.

This study aims to assess the fate of terrestrial organic carbon (TerrOC) in the Arctic marine environment by exploring how it changes in concentration, composition and degradation status across the wide Laptev Sea shelf. We analyzed a suite of terrestrial biomarkers as well as source-diagnostic bulk carbon isotopes ($\delta^{13}\text{C}$, $\Delta^{14}\text{C}$) in surface sediments from a Laptev Sea transect spanning more than 800 km from the Lena River mouth (< 10 m water depth) across the shelf to the slope and rise (2000-3000 m water depth). These data provide a broad view on different TerrOC pools and their behavior during cross-shelf transport. The concentrations of lignin phenols, cutin acids and high-molecular weight (HMW) wax lipids (tracers of vascular plants) decrease by 89-99 % along the transect. Molecular-based degradation proxies for TerrOC (e.g., the carbon preference index of HMW lipids, the HMW acids/alkanes ratio and the acid/aldehyde ratio of lignin phenols) display a trend to more degraded TerrOC with increasing distance from the coast. We infer that the degree of degradation of permafrost-derived TerrOC is a function of the time spent under oxic conditions during protracted cross-shelf transport. Future work should therefore seek to constrain cross-shelf transport times in order to compute a TerrOC degradation rate and thereby help to quantify potential carbon-climate feedbacks.

1 Introduction

Amplified global warming in high latitudes has raised growing concern about potential positive carbon-climate feedbacks. Arctic soils store about half of the global soil organic carbon (Tarnocai et al., 2009), with 60 % of this in perennally frozen grounds (Hugelius et al., 2014). With ongoing climate change these vast carbon reservoirs become increasingly vulnerable. Mobilization and transport of old terrigenous organic carbon (TerrOC) into the Arctic Ocean is expected to intensify (Gustafsson et al., 2011) through enhancing river discharge (McClelland et al., 2008) with augmenting sediment loads (Gordeev, 2006; Syvitski, 2002) and accelerating coastal erosion (Günther et al., 2013). This material can be buried in the sediments of the Arctic shelves, transported across the margin towards deeper basins or degraded and re-introduced into the modern carbon cycle as CO₂, thereby not only providing a potential positive feedback to global warming, but also causing severe ocean acidification (Semiletov et al., 2016). The fate of permafrost-released TerrOC in the marine environment is thus crucial for future climate projections, yet insufficiently understood (Vonk and Gustafsson, 2013).

The East Siberian Arctic Shelf (ESAS) is with a width of > 800 km the world's largest continental shelf. It receives TerrOC both from the erosion of the East Siberian shoreline, largely consisting of organic-rich, late-Pleistocene ice-complex deposits (Yedoma), and via the Great Russian Arctic Rivers, which drain extensive areas of continuous and discontinuous permafrost. The Laptev Sea is a representative for the TerrOC dominated Siberian shelf seas, since its main organic carbon input originates from substantial coastal erosion (as observed in the Buor-Khaya Bay, Sánchez-García et al., 2011; Semiletov et al., 2011; Vonk et al., 2012) and the Lena River, the main fluvial sediment source for the entire ESAS (Holmes et al., 2002).

Previous studies have focused on near-shore areas and the inner shelf (e.g. Bröder et al., 2016; Charkin et al., 2011; Feng et al., 2015; Karlsson et al., 2011; Salvadó et al., 2015; Sánchez-García et al., 2011; Semiletov et al., 2005, 2012, 2013; Tesi et al., 2014; Vonk et al., 2010, 2012, 2014; Winterfeld et al., 2015b, 2015a). They reported large fractions of old

64 TerrOC in particulate organic carbon (POC) and surface sediments close to the coast, using
65 different approaches such as applying carbon-isotope-based source apportionment (e.g.
66 Gustafsson et al., 2011; Semiletov et al., 2005; Vonk et al., 2010, 2012, 2014; Salvadó et al.,
67 2015, for the iron-associated OC fraction in the sediment) and by analyzing terrigenous
68 biomarkers in both surface sediments (e.g. Feng et al., 2013; Stein and Macdonald, 2004;
69 Tesi et al., 2014, 2016) and POC in the water column (e.g. Charkin et al., 2011; Karlsson et
70 al., 2011; Winterfeld et al., 2015a). This is the first study that encompasses sampling stations
71 along the entire transect from the Lena River mouth, across the wide Laptev Sea shelf, to the
72 continental slope and rise. Our major objective was to gain new insights regarding the
73 behavior of different TerrOC pools, in particular investigating potential degradation of
74 permafrost-released material along the land-shelf-basin continuum. The Laptev Sea and
75 adjacent East Siberian Sea are among the widest continental margins on Earth (Jakobsson
76 et al., 2004). The resulting long cross-shelf transport and thereby time spent in oxic
77 sediments might exert first order control on TerrOC degradation (e.g. Keil et al., 2004). Our
78 study area is thus well suited to test hypotheses on the fate of permafrost carbon in terms of
79 carbon-climate feedback. We have therefore characterized TerrOC in surface sediments
80 along the Laptev Sea transect on both bulk and molecular level, exploiting source-diagnostic
81 bulk carbon isotopes ($\delta^{13}\text{C}$, $\Delta^{14}\text{C}$) as well as an extensive biomarker suite (lignin phenols and
82 cutin acids obtained by alkaline CuO oxidation and high-molecular-weight solvent-extractable
83 lipids, such as *n*-alkanes and *n*-alkanoic acids).

2 Material and Methods

2.1 Study area

The Laptev Sea is the shallowest of the Arctic shelf seas with an average depth of 48 m (Jakobsson et al., 2004). It spans over 498,000 km² with a volume of 24,000 km³ and is located between the Kara Sea and Severnaya Zemlya in the West and the East Siberian Sea and the New Siberian Islands in the East. The main sources of particulate organic carbon (POC) for the Laptev Sea are terrigenous, both from coastal erosion and river runoff (Sánchez-García et al., 2011; Stein and Macdonald, 2004). Marine primary production is limited to on average two ice-free months per year and therefore generally low. Nutrient-poor waters on the Siberian shelves resulting from a strong stratification further impede phytoplankton growth (Sakshaug, 2004).

The destabilization of Pleistocene Ice-Complex Deposits along the coastline is a main sediment source for the Laptev Sea (Rachold et al., 2000). The total POC input from coastal erosion to Laptev and East Siberian Sea is estimated to be between 4.0 Tg yr⁻¹ (Semiletov, 1999; Stein and Fahl, 2000) and 22 ± 8 Tg yr⁻¹ (including net subsea permafrost-carbon erosion, Vonk et al., 2012).

The Lena River is estimated to provide 20.7 Tg of sediment per year (Holmes et al., 2002), i.e. > 70 % of the total riverine input to the Laptev Sea (Gordeev, 2006) with an average water discharge of 588 km³ yr⁻¹ (Holmes et al., 2012). It drains a watershed of ~2.46 x 10⁶ km² (Holmes et al., 2012), of which 77 % is underlain by continuous permafrost (Amon et al., 2012). Water discharge peaks in June, during the spring flood, when about 75 % of total organic carbon is delivered (Rachold et al., 2004). Total POC discharge by the Lena River can be up to 0.38 Tg yr⁻¹ (Semiletov et al., 2011).

Sediment transport pathways are largely influenced by the prevailing atmospheric conditions: During cyclonic summers (i.e. positive phase of the Arctic Oscillation), northerly winds predominate, strengthening the Siberian Coastal Current, which transports Lena River water masses along the coast towards the East Siberian Sea; whereas during anticyclonic summers (i.e. negative phase of the Arctic Oscillation and mainly southerly winds) the Lena

River plume is exported onto the mid-shelf and towards the deep part of the Arctic Ocean (Charkin et al., 2011; Dmitrenko et al., 2008; Guay et al., 2001; Wegner et al., 2013; Weingartner et al., 1999). Sediment transport in the Laptev Sea is strongly seasonal. The main transport of Lena River water with high concentrations of suspended particulate matter (SPM) towards the mid-shelf takes place shortly after river-ice breakup (Wegner et al., 2005). During the ice-free summer, SPM circulates between inner and mid-shelf with very little material escaping over the shelf break to the deeper parts of the Arctic Ocean. Significant sediment export is suggested to happen during freeze-up through SPM that is incorporated in sea ice and then transported across the continental margin (Dethleff, 2005; Eicken et al., 1997) or through the formation of dense bottom water resulting from brine ejection (Dethleff, 2010; Ivanov and Golovin, 2007). Hardly any sediment transport occurs beneath the ice cover.

Holocene-scale linear sedimentation rates for the Laptev Sea vary between 0.12 and 0.59 mm yr⁻¹ according to ¹⁴C dating of marine bivalves (Stein and Fahl, 2004, and citations therein), whereas centennial-scale ²¹⁰Pb-derived rates for the more recent Laptev Sea can be up to 1.3 mm yr⁻¹ (Vonk et al., 2012). These rates do not seem to be correlated with water depth on the shelf, but values for the continental slope and rise tend to be on the lower end (0.12-0.38 mm yr⁻¹) (Stein and Fahl, 2004, and citations therein).

2.2 Sampling

Sediment sampling locations span from close to the Lena River mouth and in the Buor-Khaya Bay, across the shelf, to the continental slope and rise, covering a transect of > 800 km with water depths increasing by more than two orders of magnitude. Samples SW-1, SW-2, SW-3, SW-4, SW-6, SW-14, SW-23 and SW-24 were collected during the SWERUS-C3 expedition on IB *ODEN* during summer 2014 using an Oktopus multicorer (8 Plexiglas tubes, 10 cm diameter). All other samples were collected during the International Siberian Shelf Study (ISSS-08) expedition onboard the RV *Yacob Smirnitskyi* during summer 2008. The YS-4, YS-6, YS-13 and YS-14 samples were taken with a GEMAX gravity corer (2 Plexiglas

tubes, 9 cm diameter); YS-9 and TB-46 were collected with a Van Veen grab sampler. For the grab samples only surface sediments (uppermost cm) were subsampled and used in this study. Sediment cores were cut into 1 cm slices within 24 hours after sampling. To account for lower sediment accumulation rates on the rise, for SW-1, SW-2, SW-3 and SW-4 a higher resolution of 0.5 cm for the top 10 cm was chosen. The depositional age for all samples is thus between ~8 and ~70 years (depending on which sedimentation rates are employed). All samples were kept frozen throughout the expedition and freeze-dried upon arrival to Stockholm University laboratories. See Semiletov and Gustafsson (2009) for more information on the ISSS-08 expedition. For exact sampling locations see Table 1.

2.3 Surface area

All surface area analyses have been performed on a Micromeritics Gemini VII Surface Area and Porosity analyzer. Freeze-dried subsamples of ~0.7 g were furnace-dried at 400 °C for 12 h and gently cooled down to room temperature to remove all organic material. Keil and Cowie (1999) have shown that this method yields statistically similar results to the method using removal with sodium pyrophosphate/ hydrogen peroxide (Mayer, 1994). The samples were then desalted by repeated mixing with 50 ml of MilliQ water and centrifugation (20 min at 8000 rpm), followed by further freeze-drying. Directly prior to analysis they were degassed in a Micromeritics FlowPrep 060 Sample Degas System for 2 h at 200 °C under a constant nitrogen flow. Each analysis was initiated by measuring the free space in the vial. The specific surface areas were derived from 6 pressure-point measurements (relative pressure $p/p_0 = 0.05-0.3$, equilibration time 5 s) with nitrogen as adsorbing gas (Brunauer et al., 1938). The instrumental precision was $0.1-0.3 \text{ m}^2 \text{ g}^{-1}$, which corresponds to a relative uncertainty of about 1 %. The performance of the instrument was monitored with the surface area reference material Carbon Black ($21.0 \pm 0.75 \text{ m}^2 \text{ g}^{-1}$) provided by Micromeritics.

2.4 X-Ray Fluorescence

The mineral composition of ~1 g freeze-dried, homogenized subsamples was also characterized with a wavelength dispersive sequential Philips PW2400 X-ray Fluorescence (XRF) spectrometer. Prior to the analysis, sediment samples were combusted for 12h at 450 °C to remove the organic fraction. The XRF was operated under vacuum conditions on samples prepared as glass beads using lithium tetraborate and melted with a fluxer Claisse Fluxy (~1150°C) (Mercone et al., 2001). The relative error was less than 0.6 % for major elements and less than 3 % for trace elements. In this study only SiO₂, Al₂O₃ and CaO are reported.

2.5 Bulk elemental and carbon isotope analysis

Concentration and $\delta^{13}\text{C}$ isotopic composition of total organic carbon (TOC) were determined at the Stable Isotope Laboratory, Department of Geological Sciences, Stockholm University. Homogenized subsamples of ~10 mg were repeatedly acidified (HCl, 1.5 M, Ag capsules) to remove carbonates (Nieuwenhuize et al., 1994). TOC concentrations and $\delta^{13}\text{C}$ isotopic composition were measured simultaneously with a Carlo Erba NC2500 elemental analyzer connected via a split interface to a Finnigan MAT Delta V mass spectrometer. TOC concentrations were blank corrected and the relative error was < 1 %. Stable isotope data are reported relative to VPDB using the $\delta^{13}\text{C}$ notation.

Radiocarbon analyses of acidified samples were conducted at the US National Ocean Sciences Accelerator Mass Spectrometry (NOSAMS) Facility of the Woods Hole Oceanographic Institution, USA, according to their standard routines (Pearson et al., 1998). The relative error of the measurements was < 0.5 %. Radiocarbon data are reported using the $\Delta^{14}\text{C}$ notation following Stuiver and Polach (1977).

2.6 Biomarkers

2.6.1 CuO-oxidation products

Microwave-assisted alkaline CuO oxidation was performed according to the method established by Goñi and Montgomery (2000). Homogenized subsamples of 100-400 mg of

sediment (corresponding to 2-5 mg OC) were combined with 300 mg of copper(II) oxide and 50 mg of ferrous ammonium sulfate and oxidized under oxygen-free conditions (degassed NaOH, 8 wt %) at 150 °C for 90 min using an UltraWAVE Milestone 215 Microwave oven. After oxidation, known amounts of trans-cinnamic acid and ethyl vanillin were added as recovery standards. Samples were acidified to pH 1 with HCl (12 M) and repeatedly extracted with ethyl acetate. Anhydrous Na₂SO₄ was added to remove remaining water. The solvent was evaporated and extracts re-dissolved in pyridine. For quantification, subsamples were derivatized with bis-trimethylsilyl-trifluoroacetamide (BSTFA) + 1 % trimethylchlorosilane (TMCS) and analyzed on a gas-chromatograph mass spectrometer in full scan mode (GC-MS, Agilent) using a DB5-MS capillary column (60 m x 250 µm, 0.25 µm stationary phase thickness, Agilent J&W) with a temperature profile of initially 60 °C followed by a ramp of 5 °C min⁻¹ until reaching and holding 300 °C for 5 min. The quantification of lignin phenols, benzoic acids, and p-hydroxybenzenes was achieved by comparison to the response factors (key ions) of commercially available standards. For cutin-derived products, fatty acids and dicarboxylic acids the response factor of trans-cinnamic acid was used as in Goñi et al. (1998).

2.6.2 Solvent-extractable lipids

Wax lipids were extracted by means of accelerated solvent extraction (Dionex ASE 300) using dichloromethane:methanol (9:1) according to the method described by Wiesenberger et al. (2004). Pre-rinsed stainless-steel vessels were loaded with ~3 g of freeze-dried sediment, filled up with pre-combusted glass beads and pre-combusted glass fiber filters at both ends. Two extraction cycles were performed per sample applying a static pressure of 1500 psi and a temperature of 80 °C for 5 min after a heating phase of 5 min. The flush volume was 50 % of the 34 ml cell size with a purging time of 100 s. Extracts were further purified (addition of activated Cu for sulfur and anhydrous Na₂SO₄ for water removal) and then separated into a neutral and an acid fraction using BondElut cartridges (bonded phase NH₂, Varian), eluting with dichloromethane:isopropanol (2:1) for

223 the neutral and methyl *tert*-butyl ether with 4 % acetic acid for the acid fraction according to
224 the method described by van Dongen et al. (2008a). The neutral fraction was further
225 separated into a polar and a non-polar fraction with an Al₂O₃ column. For each of the three
226 compound classes *n*-alkanes (neutral non-polar fraction), *n*-alkanols (neutral polar fraction)
227 and *n*-alkanoic acids (acid fraction) ~10 mg of one internal standard, d₅₀-tetracosane, 2-
228 hexadecanol and d₃₉-eicosanoic acid respectively, were added to the sediment samples prior
229 to extraction. All fractions were then analyzed on a GC–MS (Agilent) using the same column
230 and temperature program as for the CuO products. The polar and acid fractions were
231 derivatized with BSTFA + 1 % TMCS prior to analysis. Quantification was performed using a
232 5-point calibration curve with commercially available standards. Here, we only report data for
233 high-molecular weight (HMW) *n*-alkanes and *n*-alkanoic acids, where HMW refers to carbon
234 chain-lengths of ≥ 23 for *n*-alkanes and ≥ 24 for *n*-alkanoic acids.

3 Results and Discussion

The fate of permafrost-released terrigenous organic carbon (TerrOC) across the Laptev Sea shelf is controlled by competing processes. Degradation and sorting, as well as replacement of TerrOC by autochthonous marine organic matter all co-occur to varying degrees during cross-shelf transport. To disentangle their effects on the fate of permafrost-released TerrOC we first report changes in bulk sediment and OC properties and then focus on differences on the molecular level.

3.1 Characterization of the transect on a bulk level

Bulk total organic carbon (TOC) concentrations decreased across the shelf with highest values (~2 %) at shallow water depths and lowest values on the shelf edge (~0.8 %); at high water depths (> 2000 m) concentrations were slightly higher (~1 %). TOC values and the general pattern were in accordance with previous data from the Laptev Sea (Semiletov et al., 2005; Shakhova et al., 2015; Stein and Fahl, 2004; Vonk et al., 2012) and within the same range of those measured for the North American Arctic margin (Goni et al., 2013).

Normalizing TOC concentrations to the mineral-specific surface area (SA) helps to understand the influence of physical sorting and preferential deposition on the observed TOC trends since SA is correlated to the sediment grain size to a first order approximation. To test if the mineral surface area is altered by the input of autochthonous organisms with siliceous or carbonaceous skeleton (e.g. silicoflagellates/diatoms or foraminifera/shells respectively), the mineral composition of the sediments was examined by X-ray fluorescence analysis.

There were no apparent trends with water depth for either $\text{SiO}_2/\text{Al}_2\text{O}_3$ or $\text{CaO}/\text{Al}_2\text{O}_3$; therefore, marine production is not expected to have a measureable effect and SA can thus be regarded as a conservative parameter. This was also confirmed by low biogenic silica concentrations for the Laptev Sea reported earlier (< 1.4 %, Mammone, 1998).

The relationship between TOC and SA has been widely studied on continental margins (e.g. Blair and Aller, 2012; Keil et al., 1994; Mayer, 1994). The TOC/SA ratios of typical river suspended sediments range between 0.4 and 1 mg m^{-2} (Mayer, 1994). TOC/SA ratios > 1

263 mg m⁻² have been found in areas with high TOC supply (e.g. river outlets) and where the
264 deposited organic matter had spent little time under oxic conditions (short oxygen exposure
265 time, OET) (Mayer et al., 2002). Ratios < 0.4 mg m⁻² generally correspond to sediments from
266 deeper parts of the ocean and long OETs (e.g. Aller and Blair, 2006). Accordingly, the
267 TOC/SA values along the Laptev Sea transect displayed a strong decrease from 2.2 and 1.7
268 mg m⁻² close to the Lena River delta (water depths of 11 and 7 m, respectively) to about 0.3
269 mg m⁻² at water depths greater than 2000 m (Fig. 2A), proposing extensive TOC loss during
270 cross-shelf transport.

271 Bulk TOC isotopes have been broadly used to distinguish between organic matter sources.
272 Radiocarbon isotopes (¹⁴C) convey information about the age of organic material, with
273 younger OC having higher $\Delta^{14}\text{C}$ values. Marine organic matter produced primarily from CO₂
274 is expected to have modern ¹⁴C signatures, whereas permafrost-derived TerrOC has aged
275 both on land and during transport and has thus more depleted ¹⁴C values. The $\Delta^{14}\text{C}$ values
276 for our Laptev Sea transect were generally low (< -280 ‰, Fig. 2B), suggesting a significant
277 input of pre-aged TerrOC (as in Vonk et al., 2012). Bulk TOC showed less depleted $\Delta^{14}\text{C}$
278 signatures with increasing distance from land on the shelf (from about -500 ‰ to about -340
279 ‰ on the outer shelf, Fig. 2B), reflecting a dilution of older TerrOC with younger marine
280 material. On the slope and rise, however, $\Delta^{14}\text{C}$ values decreased again to about -410 ‰.
281 This difference may be a result of ageing during lateral transport and/or after deposition due
282 to lower accumulation rates on slope and rise. The range between -340 ‰ and -410 ‰
283 corresponds to a $\Delta^{14}\text{C}$ age difference of about 900 years; however, the depositional age
284 differences between shelf and slope samples were estimated to be less than 80 years (see
285 Section 2.2). Ageing after burial alone does therefore not explain the difference in $\Delta^{14}\text{C}$. Keil
286 et al. (2004) estimated a lateral transport time of 1800 years across the Washington margin
287 (158 km) from $\Delta^{14}\text{C}$ data of bulk OC in surface sediments. For the > 200 km distance
288 between mid-shelf and rise a bulk ageing of 900 years does therefore not seem
289 unreasonable. It has to be taken into account, however, that mainly the TerrOC fraction of
290 the bulk OC is subject to such protracted lateral transport. Transport times would thus have

to be significantly higher in order to explain this age difference for the entire bulk OC. One indication supporting this hypothesis of protracted lateral transport of TerrOC is the degradation status of TerrOC at the deep stations. All molecular degradation proxies point towards highly reworked material (see Section 3.3), suggesting that only the most refractory TerrOC fraction is found at great water depths off the continental margin. Alternatively, the lower $\Delta^{14}\text{C}$ values at high water depths may be the consequence of more effective degradation of marine organic matter throughout the water column, resulting in a comparatively lower input of young autochthonous material. However, this latter scenario is not supported by the stable carbon isotopic signature.

For stable carbon isotopes (^{13}C), terrigenous sources are generally more depleted than marine organic matter (Fry and Sherr, 1984). In this study, values for $\delta^{13}\text{C}$ of TOC ranged between -26.5 ‰ and -22.3 ‰. The trend towards more enriched TOC with increasing distance from the coast (Fig. 2B) can be explained by a growing proportion of marine organic matter. However, the $\delta^{13}\text{C}$ signature of the marine source appeared to be heavier than typical marine planktonic material in that region (-26.7 ± 1.2 ‰, Panova et al., 2015; -24 ± 3 ‰, Vonk et al., 2012, and citations therein). One possible explanation for this discrepancy is an underestimated influence of ice algae that were reported to have highly enriched $\delta^{13}\text{C}$ values between -15 to -18 ‰ (Schubert and Calvert, 2001). Significant seafloor deposition of ice algal biomass has been observed previously for the Arctic basins (Boetius et al., 2013).

Another option would be a more refractory, isotopically-enriched marine endmember (-21.2 ‰) as suggested by Magen et al. (2010). They argue that lighter isotopes are preferentially consumed by bacteria, which in turn enriches the remaining marine organic matter. Following their reasoning, the more enriched values observed for this transect may be interpreted as an increasing proportion of refractory marine organic matter.

Winterfeld et al. (2015b) analyzed surface water particulate organic carbon (POC) in the Lena River delta and found a mean $\delta^{13}\text{C}$ of -29.6 ± 1.5 ‰. Karlsson et al. (2011) reported similarly depleted $\delta^{13}\text{C}$ values for POC from the Buor-Khaya Bay (-29.0 ± 2.0 ‰), while their mean value for sedimentary OC for the same stations was significantly more enriched (-25.9

± 0.4 ‰) and agreed well with our data for the shallow stations (-26.2 ± 0.3 ‰, stations YS-13, YS-14 and TB-46). Lena River POC $\delta^{13}\text{C}$ values from high-discharge periods agree well with the more enriched values we found for the shallow stations (Rachold and Hubberten, 1998). Stein and Fahl (2004), Semiletov et al. (2011, 2012) and Vonk et al. (2012) presented similar $\delta^{13}\text{C}$ ranges and trends for sediments from parts of the Laptev Sea as is reported in the current study for the entire width of the Laptev Sea shelf. For the Arctic Amerasian Continental shelf, Naidu et al. (2000) reported contrasts in absolute $\delta^{13}\text{C}$ values comparing surface sediment samples from different regions, but all commonly displayed an increasing trend for $\delta^{13}\text{C}$ values across the shelf, suggesting a growing fraction of marine organic matter with increasing distance from the coast.

Combining TOC/SA ratios with stable isotope signatures ($\delta^{13}\text{C}$) may serve to disentangle two different processes, which occur synchronously during cross-shelf transport (as in Keil et al. 1997a): 1.) The net loss of TerrOC and 2.) the replacement of TerrOC with autochthonous marine OC. Net loss of TerrOC, caused by either degradation or hydrodynamic sorting during transport, has been quantified previously using TOC/SA ratios (e.g. Aller and Blair, 2006; Keil et al., 1997a). The carrying-capacity of inorganic particles for OC is assumed to be a function of the SA (Mayer, 1994); a decrease in TOC/SA values can therefore be regarded as TOC net loss.

Replacement of TerrOC with autochthonous marine OC does not change this ratio. But since marine OC is known to be isotopically enriched in $\delta^{13}\text{C}$ over TerrOC, this process is recorded by an increasing isotopic signature. Along the Laptev Sea transect, both processes seemed to play an important role (Fig. 2C). High TOC/SA values close to the Lena River decreased sharply outbound in the nearshore regime, pointing to extensive net loss, while the increase in $\delta^{13}\text{C}$ values was minor in this area. Once TOC/SA ratios were $< 0.8 \text{ mg m}^{-2}$ (water depths $> 20 \text{ m}$), the isotopic changes and thus the replacement of TerrOC with marine OC became increasingly important. Similar trends were observed for the Amazon River delta (Keil et al., 1997b).

However, the TOC/SA trend in the shallower sediments is likely driven by both degradation of OC bound to the mineral matrix during cross-shelf transport and sorting of vascular plant fragments that are retained in the inner shelf. A recent study (Tesi et al., 2016) has shown that ~50 % of the total OC pool in the inner Laptev shelf surface sediments exists in the form of large vascular plant fragments. They are trapped close to the coast due to their size and resulting settling (Stoke's law), while the OC bound to the fine mineral matrix is more buoyant and transported offshore towards deeper waters.

3.2 Molecular indicators of organic matter sources

3.2.1 Biomarker distributions

The abundances of different source-diagnostic molecular proxies have been extensively investigated to elucidate complex carbon-cycling mechanisms. In this study, a biomarker suite of CuO oxidation products and solvent-extractable lipids was analyzed in order to gain more insights on TerrOC sources and degradation status along the Laptev Sea transect. All biomarker concentrations were normalized to the sediment-specific surface area (SA) instead of OC content to avoid the signals being overshadowed by other carbon pools. As shown by the lack of water-depth-related changes in the mineral composition (Section 3.1), mineral-matrix dilution by biogenic material is negligible.

Lignin-derived phenols have been widely used to trace TerrOC in the marine environment (e.g. Ertel and Hedges, 1984; Goñi and Hedges, 1995; Hedges and Mann, 1979). The lignin macro-molecule is only synthesized in vascular plants (and certain seaweed species that are not existing in the study area) to render stability to the cell walls. Lignin-derived phenols are typically grouped by phenol type (V: vanillyl phenols, i.e. vanillin, acetovanillone, and vanillic acid; S: syringyl phenols, i.e. syringaldehyde, aceto syringone, and syringic acid; C: cinnamyl phenols, i.e. p-coumaric and ferulic acids). Total lignin refers to the sum of the three groups. Across the shelf, lignin loadings decreased substantially with increasing distance from the coast/water depth ($45 \mu\text{g m}^{-2}$ close to the coast, $0.43 \pm 0.09 \mu\text{g m}^{-2}$ for the deep stations; loss of $99.1 \pm 0.2 \%$, Fig. 3A).

Cutin-derived hydroxy fatty acids are another compound class obtained from CuO oxidation, which have been used in parallel with lignin phenols (e.g. Goñi et al., 2000; Prahl et al., 1994). They are mainly associated with the soft tissues of vascular plants such as leaves and needles. Cutin acid loadings displayed a similar trend as lignin phenols ($11 \mu\text{g m}^{-2}$ close to the coast, $0.061 \pm 0.010 \mu\text{g m}^{-2}$ for the deep stations; loss of $99.4 \pm 0.1 \%$, Fig. 3A). Similar values and sharp declines with increasing distance from the coast for lignin and cutin have been observed for the whole East Siberian Arctic Shelf (ESAS) (Tesi et al., 2014) (Fig. 4 for comparison of lignin phenol concentrations with literature values for different Arctic margins). A recent study (Winterfeld et al., 2015a) for the Buor-Khaya Bay (5.8-17 m water depth) reported lignin phenol concentrations on the same order of magnitude, up to 40 % higher for the shallowest samples, and decreasing with increasing depth. For the Beaufort Sea shelf, Goñi et al. (2000) found a less drastic decline in lignin phenols and cutin acids going from 5 m water depth to 210 m, which likely reflected both lower concentrations in the shallow waters (factor of ~ 2), and a narrower and steeper shelf. Lignin phenols were also higher at greater water depths than on the ESAS. This may reflect the differences in bathymetry: since the Beaufort Sea shelf is not as wide as, but steeper than, the ESAS, lateral transport is possibly faster, leaving less time for organic matter to be degraded along the way. A comparison between different shelf-slope systems across the North American Arctic margin (Goni et al., 2013) revealed very low lignin and cutin concentrations for the Canadian Archipelago, Lancaster Sound and Davis Strait, whereas both concentrations and trends with water depth for the Beaufort Sea, Chuckchi Sea and Bering Sea were similar to the results from this study. An exception to these patterns was Barrow Canyon, where at water depths of > 2000 m lignin and cutin concentrations were as high as the ones observed close to the Lena River delta, pointing to efficient rapid TerrOC transfer with comparably short oxygen exposure times through this active canyon (Goni et al., 2013) (Fig. 4 and Fig. S1). Solvent extractable high-molecular weight (HMW) *n*-alkanes and *n*-alkanoic acids make up the major part of epicuticular leaf waxes (Eglinton and Hamilton, 1967) and have been

broadly employed as TerrOC biomarkers (for the Arctic Ocean e.g. van Dongen et al., 2008; Yunker et al., 1995, 2005). HMW wax lipids in this study also presented a decreasing trend with increasing water depth/distance from the coast, but to a lesser extent than lignin phenols or cutin acids (HMW *n*-alkanes: $1.1 \mu\text{g m}^{-2}$ close to the coast, $0.12 \pm 0.02 \mu\text{g m}^{-2}$ for the deep stations; HMW *n*-alkanoic acids: $12 \mu\text{g m}^{-2}$ close to the coast, $0.42 \pm 0.29 \mu\text{g m}^{-2}$ for the deep stations; loss of $89 \pm 2 \%$ and $96 \pm 3 \%$, respectively, Fig. 3B).

Previous studies in the same area reported similar lipid biomarkers concentrations, which confirm the magnitude of the decreasing trends with increasing water depth (Karlsson et al., 2011; Vonk et al., 2010) (Fig. S1). HMW *n*-alkane concentrations in the Beaufort and the Chuckchi Sea (Belicka et al., 2004; Yunker et al., 1993) are in accordance with the ones measured on the ESAS, but the shallowest sample on the Beaufort Shelf is ~2 times lower than the shallow ESAS samples (Fig. S1). This might imply that sediments transported by the Mackenzie River to the Beaufort Shelf have lower TerrOC concentrations than Lena River transported sediments. For the Mackenzie Shelf, Goñi et al. (2000) used lignin phenols and cutin acids to estimate a terrigenous $\delta^{13}\text{C}$ endmember and therewith derived a terrigenous contribution of almost 80 % for the shallowest sediments, while rough estimates from C/N and $\delta^{13}\text{C}$ data suggested that TerrOC made up only 30-50 % of the organic carbon (Macdonald et al., 2004). For the Lena Delta, source apportionment calculations using $\delta^{13}\text{C}$ and $\Delta^{14}\text{C}$ data attributed up to 83 % of the organic carbon to terrigenous sources (Vonk et al., 2012).

All TerrOC biomarker loadings displayed a strong decrease across the shelf, but their relative losses differ substantially between compound classes (Fig. 3C). These findings agree with previous results for the ESAS (Tesi et al., 2014), where similar differences between biomarkers were reported. A somewhat larger decrease was observed for lignin than for cutin, in contrast to this study. The different extents of biomarker losses for the different compound classes may either be attributed to preferential degradation of lignin phenols and cutin acids, implying that they are more labile than HMW *n*-alkanes and *n*-alkanoic acids, or sorting during transport, suggesting that they are associated with a sediment fraction that is

430 hydraulically more retained and carried less efficiently to the outer shelf/slope. A recent study
 431 (Tesi et al., 2016) aimed to disentangle these two processes by analyzing different fractions
 432 of bulk surface sediments from three transects (yet with only three stations each) across the
 433 ESAS. The fractions were separated according to density (1.8 g cm^{-3} cutoff), size ($>63 \text{ }\mu\text{m}$,
 434 $38\text{-}63 \text{ }\mu\text{m}$, $< 38 \text{ }\mu\text{m}$) and settling velocity (1 m d^{-1} cutoff). The highest lignin phenol
 435 abundance was found in low-density plant fragments ($26\text{-}55 \text{ mg g}^{-1} \text{ OC}$). These large
 436 particles have a higher settling velocity (Stokes' law) and are therefore hydraulically retained
 437 close to the coast. Cutin acids and plant wax lipids were mainly associated with the high-
 438 density fine ($< 38 \text{ }\mu\text{m}$, $> 1 \text{ m d}^{-1}$) and ultrafine ($< 38 \text{ }\mu\text{m}$, $< 1 \text{ m d}^{-1}$) mineral fractions. Within
 439 the fine and ultrafine fractions, which made up about 95 % of the organic carbon on the outer
 440 shelf, they found drastic losses of all biomarkers with increasing distance from the coast,
 441 which they attributed to degradation during the protracted cross-shelf transport. Relative
 442 decreases appeared to depend on the number of functional groups of the compound class:
 443 $98 \pm 1 \text{ \%}$ for lignin phenols, $97 \pm 1 \text{ \%}$ for cutin acids, $96 \pm 1 \text{ \%}$ for HMW *n*-alkanoic acids and
 444 $89 \pm 4 \text{ \%}$ for HMW *n*-alkanes. According to that study, the steep cross-shelf gradients
 445 observed here for lignin phenols can be attributed to both hydrodynamic sorting close to the
 446 coast and degradation during transport. From the data in the current study alone, the two
 447 processes occurring in parallel - degradation and sorting during cross-shelf transport - cannot
 448 be disentangled. However, using the data from (Tesi et al., 2016), we can make a rough
 449 correction for the sorting part to derive an estimate of the net extent of degradation. For the
 450 shallowest station in their study (same as here, TB-46), about 75 % of the lignin phenols
 451 were associated with the low density fraction that was retained close to the coast. If we thus
 452 assume only 25 %, i.e. 11 of the $45 \text{ }\mu\text{g m}^{-2}$ to be associated with the fine fraction that is
 453 actually transported across the shelf, we obtain a reduction by $96 \pm 1 \text{ \%}$ that can be
 454 attributed to degradation (compared to 99.1 % net loss). These results agree with the values
 455 presented in (Tesi et al., 2016). For the other compounds analyzed 55-74 % are associated
 456 with the fine fraction even for the shallowest station and they therefore experience sorting to
 457 a lesser extent.

Degradation after burial is assumed to play only a minor role. Differences in sedimentation ages are expected to be small (Section 2.1) and a study on centennial-scale sediment cores from the East Siberian Sea (Bröder et al., 2016) detected no significant TerrOC degradation (as recorded by biomarker loss) with increasing sediment depth. Also in that study, lignin phenol and cutin acid loadings were on average 20 times higher on the inner than on the outer shelf, whereas for HMW *n*-alkanoic acids and *n*-alkanes the difference between inner and outer shelf was only a factor of ~3-5. Contrasts between the stations were found to be larger than down-core changes. This may be due to the fact that the cores in that study only encompassed about one century of sedimentation ages, while the protracted cross-shelf transport likely requires much longer timescales.

3.2.2 Lignin Phenol sources

Relative distributions of different lignin phenol classes reveal more information on TerrOC sources since they are specific to different plant types. Syringyl phenols are not produced by gymnosperm (non-flowering) plants; elevated syringyl to vanillyl ratios (i.e. S/V > 1, Hedges and Parker, 1976) are therefore attributed to more lignin phenols from angiosperm (flowering) plants. These ratios have to be handled with care, though, because the preferential degradation of syringyl phenols by white- and brown-rot fungi on land can also alter S/V ratios (Hedges et al., 1988). S/V values for the Laptev Sea transect increased with increasing water depth from ~0.65 for the inner shelf to ~1.0 for the slope/rise sediments (Fig. 5A). This trend can either be explained by preferential degradation of gymnosperm material or sorting during transport. Tesi et al. (2014) measured generally lower values for S/V (ESAS average: 0.47, for only Lena watershed dominated locations: 0.42) recording no trend with water depth (Fig. S1 for comparisons with other studies). Their deepest station was located at only 69 m water depth, though, whereas in this study sediments from down to 3146 m water depth were analyzed. S/V ratios in Buor-Khaya Bay surface sediments (Winterfeld et al., 2015a) were also lower (0.43 ± 0.02 on average) and displayed no trend with water depth. Within the water depth interval they studied (5.8-17 m), however, the

486 samples analyzed in this study had also quite homogeneous S/V ratios (0.64 ± 0.02). Two
 487 sediment cores from the East Siberian Sea (Bröder et al., 2016) showed also lower S/V
 488 values (inner shelf surface sediment: 0.62, outer shelf surface sediment: 0.50) displaying no
 489 clear trends over time/down-core. For the Beaufort Sea shelf Goñi et al. (2000) detected
 490 rather high values (0.54-1.71), which (besides the very high value at 61 m water depth)
 491 agree with the data from this study. Other transects across the North American Arctic margin
 492 (Goni et al., 2013) had slightly lower S/V ratios with no observed trends with water depth.
 493 The ratio of cinnamyl to vanillyl phenols (C/V) is associated with the relative contributions of
 494 woody versus soft material, because only non-woody vascular plants synthesize cinnamyl
 495 phenols (Hedges and Mann, 1979a). This ratio admittedly decreases with ongoing
 496 degradation (Opsahl and Benner, 1995) and may therefore not be used as an unambiguous
 497 source indicator. We observed that C/V values strongly decreased across the Laptev Sea
 498 Shelf from ~0.5 (close to the Lena River outlet) to ~0.1 (on the slope/rise, Fig. 5B), which
 499 may reflect the preferential degradation of soft tissues. This trend is not likely caused by
 500 hydrodynamic sorting, since typically the larger, low-density, woody plant fragments are
 501 retained in shallower water, whereas finer material is transported further across the shelf
 502 (e.g. Keil et al., 1994; Tesi et al., 2016).
 503 C/V ratios in Buor-Khaya Bay sediments (Winterfeld et al., 2015a) in shallow waters were on
 504 average lower and more homogeneous (0.17 ± 0.03) than those measured in this study (0.41
 505 ± 0.12 for the corresponding depth interval) (Fig. S1 for comparisons with other studies). C/V
 506 values for the entire ESAS were on average 0.15 (0.14 ± 0.07 for only Lena dominated
 507 waters) with no water depth trend (Tesi et al., 2014). In shallow sediment cores from the East
 508 Siberian Sea, Bröder et al. (2016) measured C/V ratios of 0.20 (inner shelf) and 0.13 (outer
 509 shelf) for the surface sediments with no significant trend over sediment depth. For the
 510 Mackenzie Shelf C/V values ranged between 0.16 and 0.32 and slightly increased with
 511 increasing water depth (Goñi et al., 2000). In contrast, in the Bering Sea, Chuckchi Sea,
 512 Barrow Canyon, Canadian Archipelago, Lancaster sound and Davis Strait there were no C/V
 513 trends observed (Goni et al., 2013), with lower values in the Canadian part (0.10 ± 0.12) and

highest values on the Beaufort Sea slope, where values slightly decreased with increasing depth (0.39 ± 0.07).

A comparison to the S/V-C/V signatures of potential Arctic plant end-members (compiled by Amon et al., 2012, and citations therein, Tesi et al., 2014, and Winterfeld et al., 2015a) showed that lignin phenols likely derive from both angio- and gymnosperm soft tissues in the shallower samples, closely matching with willow (*Salix*) tissues measured by Winterfeld et al. (2015a). With increasing water depths, angiosperm wood became the most important source material, while gymnosperm wood, grasses and mosses did not appear to contribute significantly to the overall lignin phenol fingerprint (Fig. 5C). As discussed earlier, this trend may well be a result of preferential degradation and sorting during cross-shelf transport and not derive from actual changes in source material.

3.3 Degradation status of organic matter

During degradation, syringyl and vanillyl phenol aldehydes are oxidized to carboxylic acids of the same phenol group. Increasing Sd/SI and Vd/VI ratios can therefore qualitatively indicate ongoing degradation of lignin phenols (Ertel and Hedges, 1984; Hedges et al., 1988). For fresh plant material typical acid-to-aldehyde ratios are around 0.1-0.2 (Hedges et al., 1988). Winterfeld et al. (2015a), however, found values as high as Sd/SI = 0.80 and Vd/VI = 0.67 for a moss species (*Aulacomnium turgidum*), Sd/SI = 0.87 for larch (*Larix*) needles and Sd/SI = 0.49 Vd/VI = 0.41 for wild rosemary (*Ledum palustre*). Sedges (*Carex spp.*), dwarf birch (*Betula nana*) and willow (*Salix*) range between Sd/SI = 0.13-0.24 and Vd/VI = 0.18-0.23. The ratio of CuO oxidation-derived 3,5-dihydroxybenzoic acid to vanillyl phenols (3,5-Bd/V) also serves as a proxy for degradation as 3,5-Bd is formed during humification likely occurring in soils (Gordon and Goñi, 2004; Hedges et al., 1988; Prahl et al., 1994; Tesi et al., 2014). For this reason, this proxy can trace mineral rich soil organic matter in contrast to vascular plant debris (e.g. Dickens et al., 2007; Prahl et al., 1994) as well as degradation during cross shelf transport (Tesi et al., 2016).

541 Sd/SI, Vd/VI and 3,5-Bd/V all increased along the transect, implying more degraded material
 542 with increasing residence time in the shelf system (Fig. 6A). There appeared to be no
 543 differences between outer shelf/slope and rise, which may indicate that TerrOC on the slope
 544 is already highly reworked. In contrast, Tesi et al. (2014) found no correlation between Sd/SI
 545 or Vd/VI and distance from the coast, while 3,5-Bd/V significantly increased with increasing
 546 distance from the coast (Fig. S2 for comparisons with other studies). Sd/SI values for the
 547 Buor-Khaya Bay from Winterfeld et al. (2015a) were slightly higher (1.04 ± 0.24) than
 548 samples from the corresponding water depths in this study (0.66 ± 0.15), whereas Vd/VI
 549 values were significantly higher (1.28 ± 0.30 compared to 0.59 ± 0.14). Measurements for the
 550 Mackenzie Shelf agreed with the ones in this study (Sd/SI = 0.81 ± 0.25 compared to $1.01 \pm$
 551 0.33 for the corresponding water depths; Vd/VI = 0.69 ± 0.14 to 0.86 ± 0.26 ; 3,5-Bd/V = 0.19
 552 ± 0.04 to 0.31 ± 0.15), but did not show a trend with water depth (Goñi et al., 2000).
 553 Tesi et al. (2016) observed lower acid/aldehyde ratios for the lignin-rich low-density fraction
 554 compared to the other fractions (high-density with different grain sizes and settling velocities)
 555 in coastal surface sediments from the ESAS. With increasing distance from the coast, these
 556 values increased, whereas for the other fractions there were no apparent trends. These
 557 findings were interpreted as relatively fresh lignin in the low-density fraction (rich in large
 558 plant fragments) compared to the relatively degraded lignin that had likely experienced
 559 leaching and adsorbed to the fine mineral fractions (i.e. mineral bound OC). Bulk 3,5-Bd/V
 560 values are potentially affected by both sorting and degradation, as they increased with
 561 decreasing particle size (fine and ultrafine fractions had the most degraded signal and are
 562 preferentially transported to the outer shelf) and across the shelf in each of the fractions.
 563 The carbon preference indices for HMW *n*-alkanes and HMW *n*-alkanoic acids have also
 564 been widely applied as degradation proxies for plant waxes in marine sediments (for the
 565 ESAS, e.g. van Dongen et al., 2008; Fahl and Stein, 1997; Fernandes and Sicre, 2000; Vonk
 566 et al., 2010). It measures the ratio of odd-to-even numbers of carbon chain-lengths of HMW
 567 lipids and is based on the preference of odd carbon chain-lengths for HMW *n*-alkanes in
 568 fresh plant material (even carbon chain-lengths for HMW *n*-alkanoic acids; Eglinton and

Hamilton, 1967). With ongoing degradation this preference is lost and the CPI approaches 1 (Bray and Evans, 1961).

We observed that the HMW *n*-alkane CPI presented a similar pattern as the lignin phenol based degradation indices. However, the HMW *n*-alkanoic acid CPI did not show as much of a degradation trend (HMW *n*-alkane CPI: ~5.7 close to the coast, ~2.2 for the deep stations; HMW *n*-alkanoic acids: ~5.4 close to the coast, ~4.1 for the deep stations; Fig. 6B). Karlsson et al. (2011) measured lipid CPIs in the Buor-Khaya Bay with 10-80 km distance to the coast and obtained similar results to this ~800 km cross-shelf study, with higher values closer to the river delta (Fig. S2 for comparisons with other studies). Their data appears to have a wider spread, though, which might be due to either the narrower dynamic range. Fahl and Stein (1997) also reported a large range of *n*-alkane CPI values (< 0.2- > 5) for Laptev Sea sediments. Fernandes and Sicre (2000) analyzed sediments from the Kara Sea and from the major rivers discharging into this sea, Ob and Yenisey rivers. In the marine environment and the Ob River, they observed HMW *n*-alkane CPI values between 4.8 and 5.3, similar to those found at shallow water depths in this study. For the Yenisey River and mixing zone, they found higher CPI values, pointing to fresher material being transported there. Vonk et al. (2010) recorded HMW *n*-alkane CPI values for sediments along the East Siberian Sea Kolyma paleoriver transect (across the East Siberian Sea) shelf that decreased from > 7.5 to < 4.0 with increasing distance from the river mouth, overall higher than in this study but confirming the general trend to more degraded material on the outer shelf. Tesi et al. (2016) found HMW *n*-alkanoic acid CPI values to decrease with decreasing particle size with no significant trends across the shelf in all but the low-density fraction, which is largely retained close to the shore. The HMW *n*-alkane CPI values in that study, however, showed no systematical differences between different fractions, but an overall decreasing trend with increasing distance from the coast.

When undergoing degradation, HMW *n*-alkanoic acids may also lose their functional groups, turning them into HMW *n*-alkanes (Meyers and Ishiwatari, 1993). The slightly decreasing ratio of HMW *n*-alkanoic acids to *n*-alkanes also hints at more degraded material with

597 increasing water depth, although, due to a rather large variability, this trend is not significant.
598 For the Buor-Khaya Bay surface sediments Karlsson et al. (2011) obtained similar results
599 (0.48-10.7, here 1.1-10.9) with higher values closer to the river delta (Fig. S2 for
600 comparisons with other studies). Along the Kolyma paleoriver transect, Vonk et al. (2010)
601 measured HMW *n*-alkanoic acid to *n*-alkane ratios between 1 and 6 with no clear trend with
602 increasing distance from the river mouth. Tesi et al. (2016) found decreasing values with
603 increasing distance from the coast with no differences between the fractions. Two sediment
604 cores from inner and outer East Siberian Sea recording about one century of sedimentation
605 showed no clear trend in CPI or HMW *n*-alkanoic acid/*n*-alkane towards more degraded
606 TerrOC with increasing sediment depth (Bröder et al., 2016), but displayed a similar
607 difference between inner and outer shelf as seen in this study. This contrasting behavior for
608 cross-shelf and down-core trends may be caused by significantly different timescales for the
609 two processes: about one century in situ/after burial compared to potentially several millennia
610 long lateral transport. Furthermore, the degradation efficiency is likely higher under the oxic
611 conditions prevailing during cross-shelf lateral transport (Keil et al., 2004), than in the anoxic
612 conditions that predominate below a few millimeters of sediments on the ESAS (e.g. Boetius
613 and Damm, 1998). Comparing in situ to transport-related oxygen exposure times on the wide
614 Arctic shelves could potentially resolve the observed discrepancies.

4 Concluding remarks and future research directions

Across the Laptev Sea from the Lena River mouth to the deep sea of the Arctic interior a considerable loss of terrigenous organic matter has been observed on both bulk and molecular level. All terrigenous biomarkers display a massive decline with increasing water depth along this high-resolution transect due to hydrodynamic sorting and degradation during transport. Terrigenous organic matter (TerrOC) seems to be also qualitatively more degraded on the outer shelf, slope and rise compared to inner shelf and coastal areas. These results corroborate and expand previous findings for the East Siberian Arctic Shelf, showing that the shelf seas in this region function as an active reactor for TerrOC. Since the East Siberian Arctic Shelf belongs to the widest and shallowest continental margins on Earth, cross-shelf transport times and thus the time spent in oxic sediments are expected to be comparatively long. This stands in contrast to e.g. the Mackenzie basin, which is thought to act as a geological sink for organic carbon due to its TerrOC burial (Hilton et al., 2015). For narrower Arctic shelves in general, where transport times can be expected to be much shorter, organic matter transfer towards the deeper basins appears to be much more efficient, with high TerrOC concentrations in surface sediments even at greater water depths (e.g. Barrow Canyon, Goni et al., 2013). It can therefore be assumed that the cross-shelf transport time exerts first-order control over the extent of TerrOC degradation. With ongoing global warming, rising permafrost-derived organic carbon input from river-sediment discharge and coastal erosion is expected to reach the marine environment. It is therefore crucial to better constrain cross-shelf transport times in order to determine a TerrOC degradation rate and thereby contribute to quantifying potential carbon-climate feedbacks.

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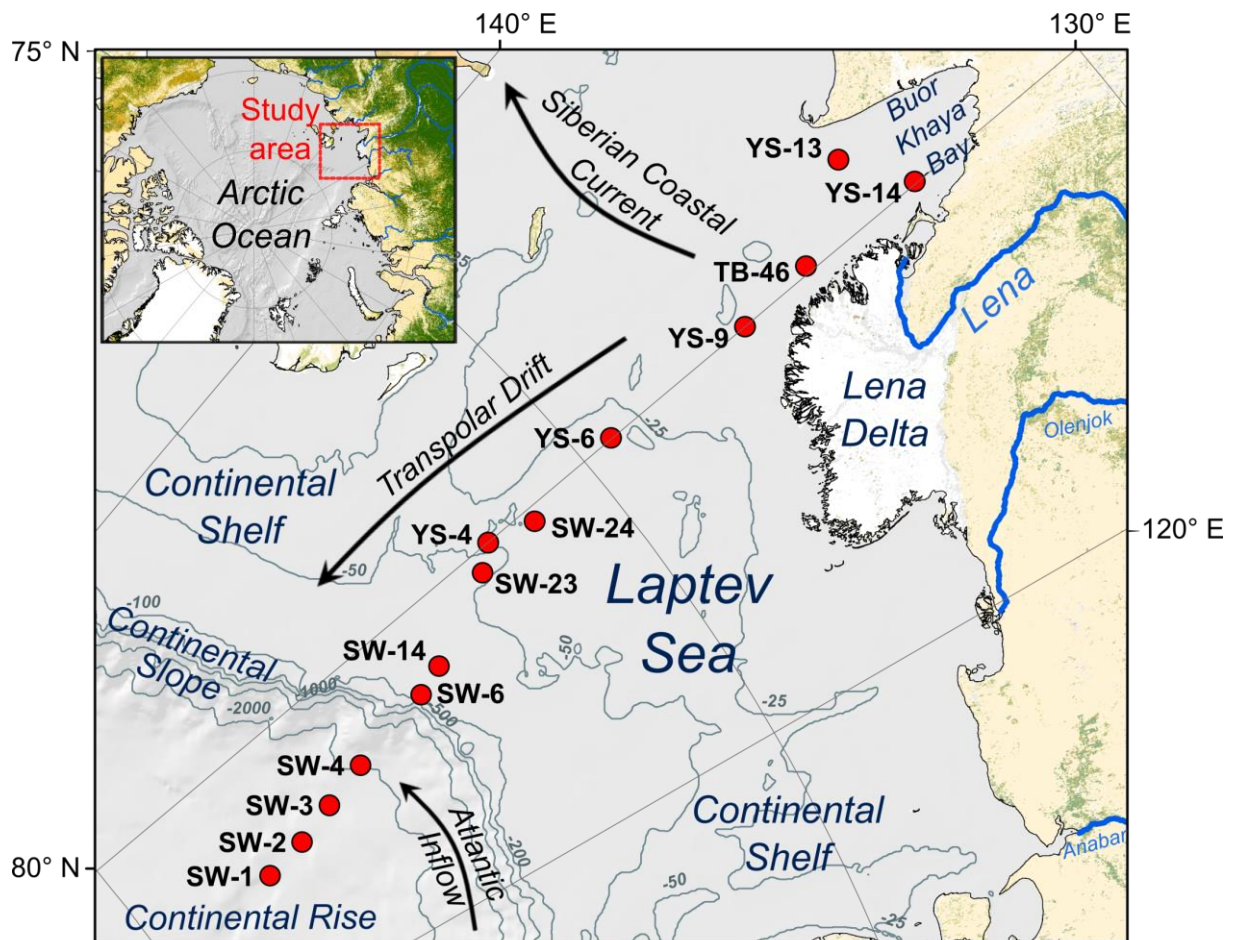


Figure 1: Map of the study area in the Laptev Sea. Red filled circles mark the sediment sampling sites. The transect reaches from the Lena River mouth and the Buor-Khaya Bay (water depths ~10 m) across the Laptev Sea Shelf (mean depth ~50 m) to the slope/shelf break and rise (water depths ~3000 m). Arrows show the directions of the prevailing ocean currents.

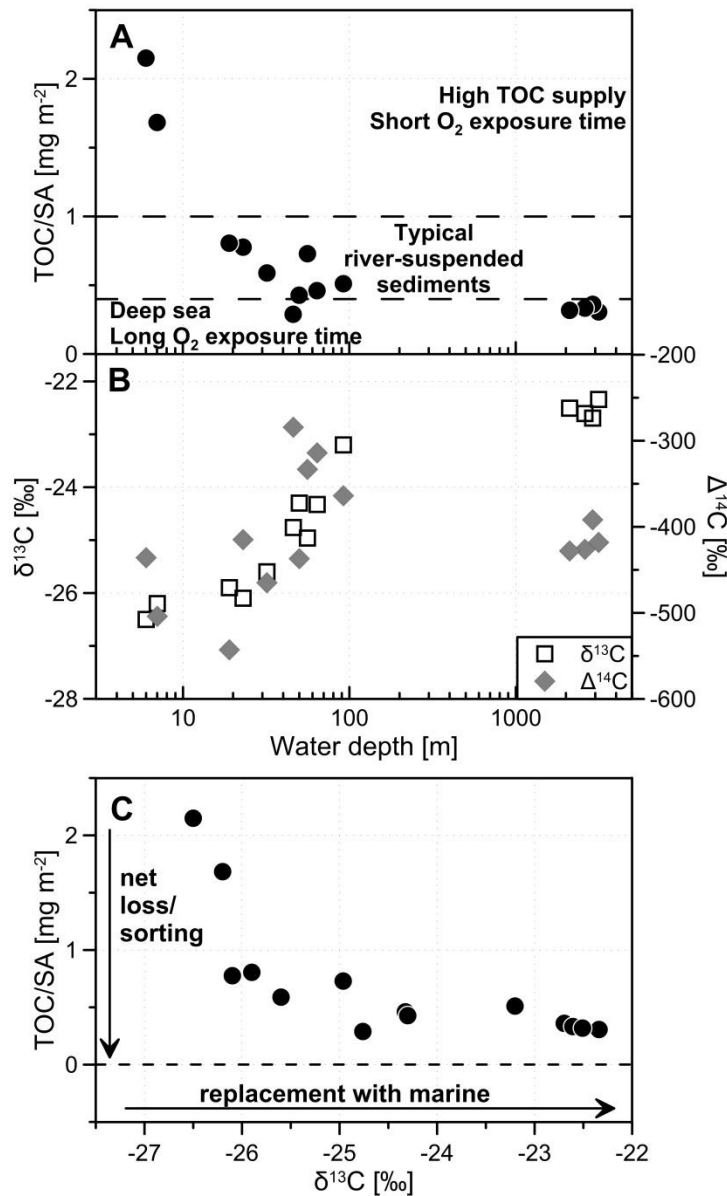


Figure 2: (A) The ratio of total organic carbon (TOC) to mineral surface area (SA). Typical values for deep sea, river-suspended sediments and high TOC supply are taken from Blair and Aller (2012). (B) The stable carbon isotopic signal ($\delta^{13}\text{C}$, open boxes) and the radiocarbon isotopic signal ($\Delta^{14}\text{C}$, filled diamonds). (C) The relationship between TOC/SA and $\delta^{13}\text{C}$ can help to disentangle two processes occurring simultaneously during cross-shelf transport: The net loss (i.e. degradation) or sorting (i.e. hydraulically retaining) of TerrOC leads to a shift towards lower TOC/SA ratios, whereas the replacement/dilution with marine OC shifts the isotopic signature towards higher values.

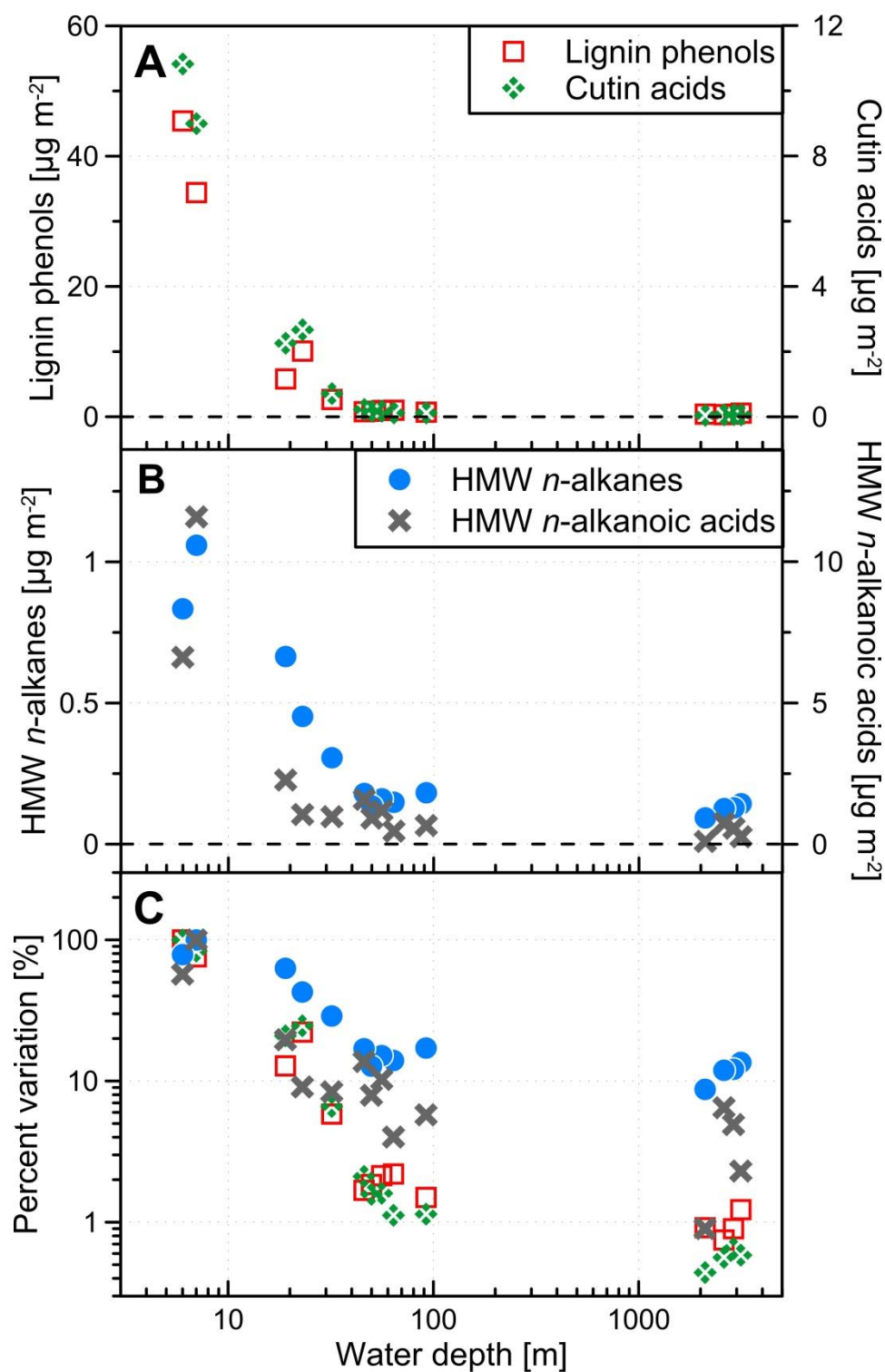


Figure 3: Terrigenous biomarker loadings across the shelf: (A) lignin phenols and cutin acids, (B) HMW *n*-alkanes and HMW *n*-alkanoic acids. (C) Comparison between the different biomarkers along the transect: lignin phenols, cutin acids, HMW *n*-alkanoic acids and *n*-alkanes where each is normalized to respective highest value (corresponding to 100 %).

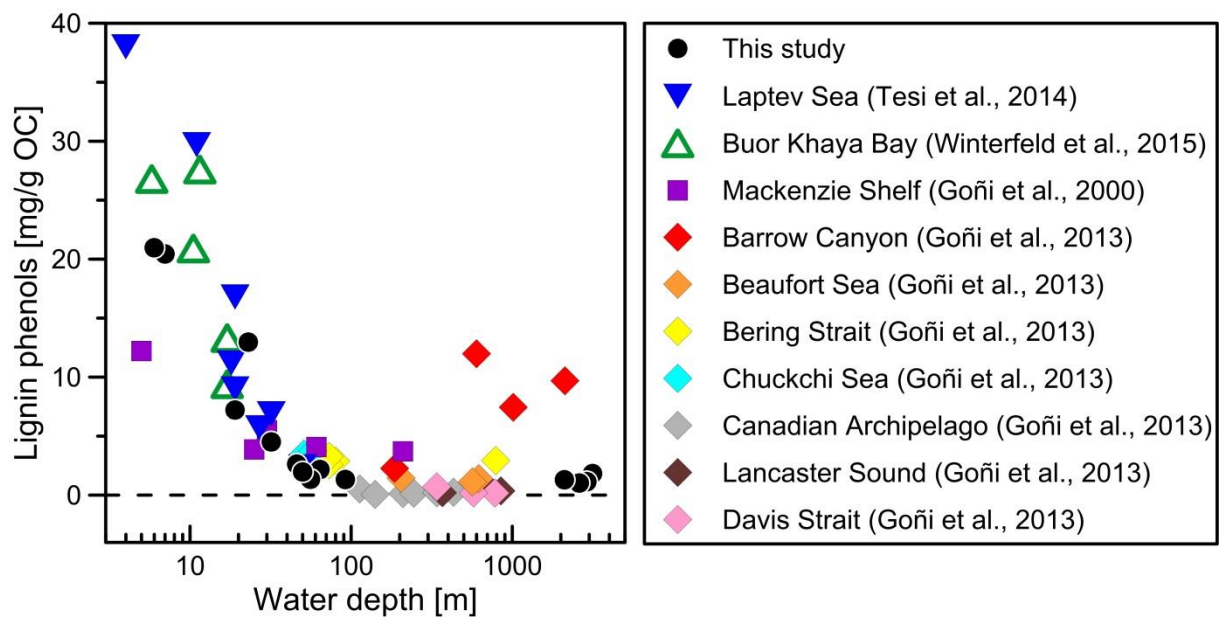


Figure 4: A comparison of lignin phenol data from this project to values from published studies around the Arctic Ocean. Similar decreasing trends with increasing water depth are observed for all systems but Barrow Canyon, where elevated lignin phenols concentrations are found even at depth of > 1000 m.

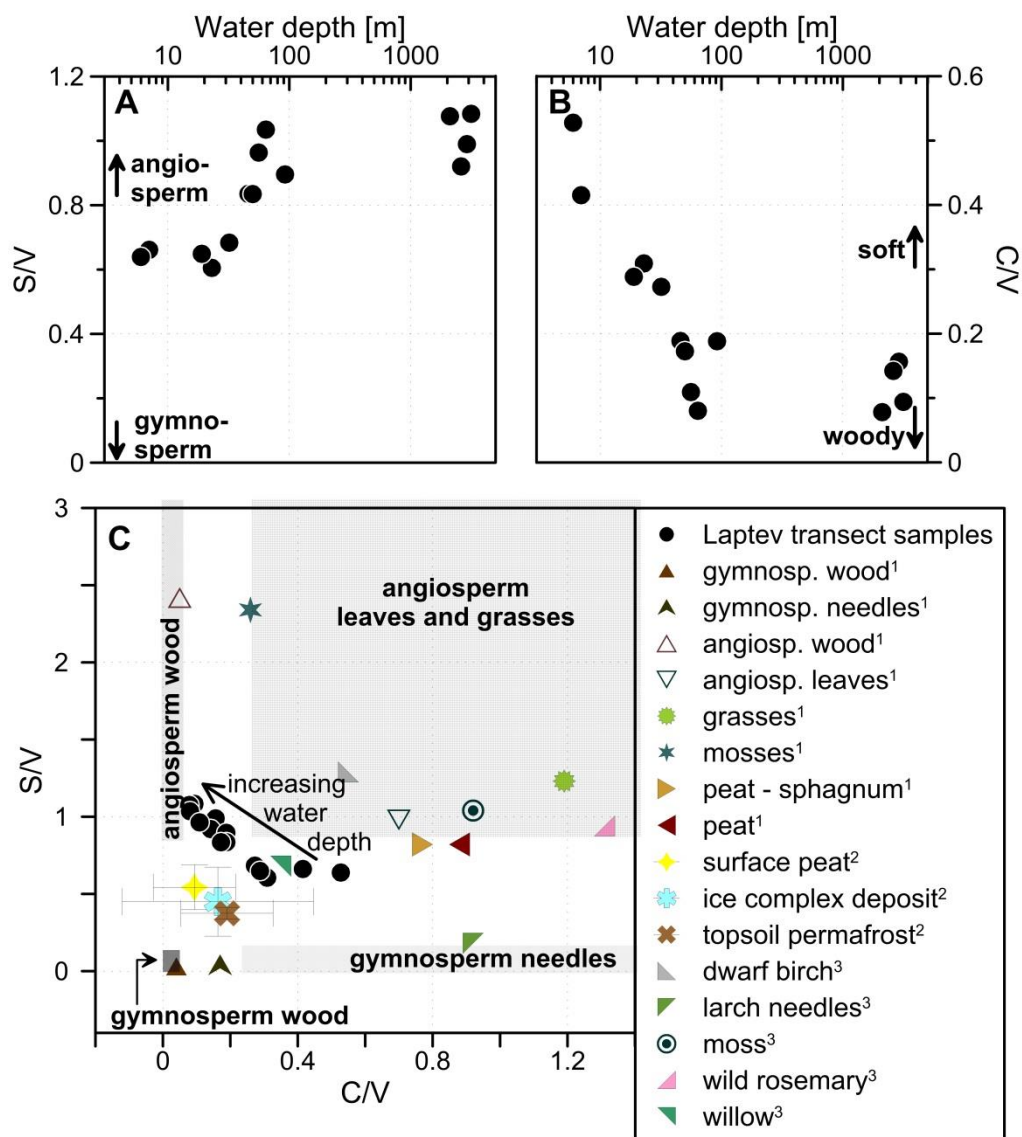


Figure 5: The lignin phenol composition carries source information: (A) an increasing ratio of syringyl to vanillyl phenols (S/V) suggests relatively more angiosperm material. (B) A decreasing ratio of cinnamyl to vanillyl phenols (C/V) implies an increasing relative contribution of woody material compared to soft tissues. (C) Comparison of S/V and C/V with the end-members for different Arctic plants as compiled from different studies by Amon et al. (2012, and citations therein, here marked with ¹); ice-complex deposit and topsoil permafrost as determined by Tesi et al. (2014, here marked with ²) and more plant species measured by Winterfeld et al. (2015a, here marked with ³). The boxes indicate typical ranges of S/V and C/V for different vascular plant tissues in different locations (e.g. Goñi et al., 2000).

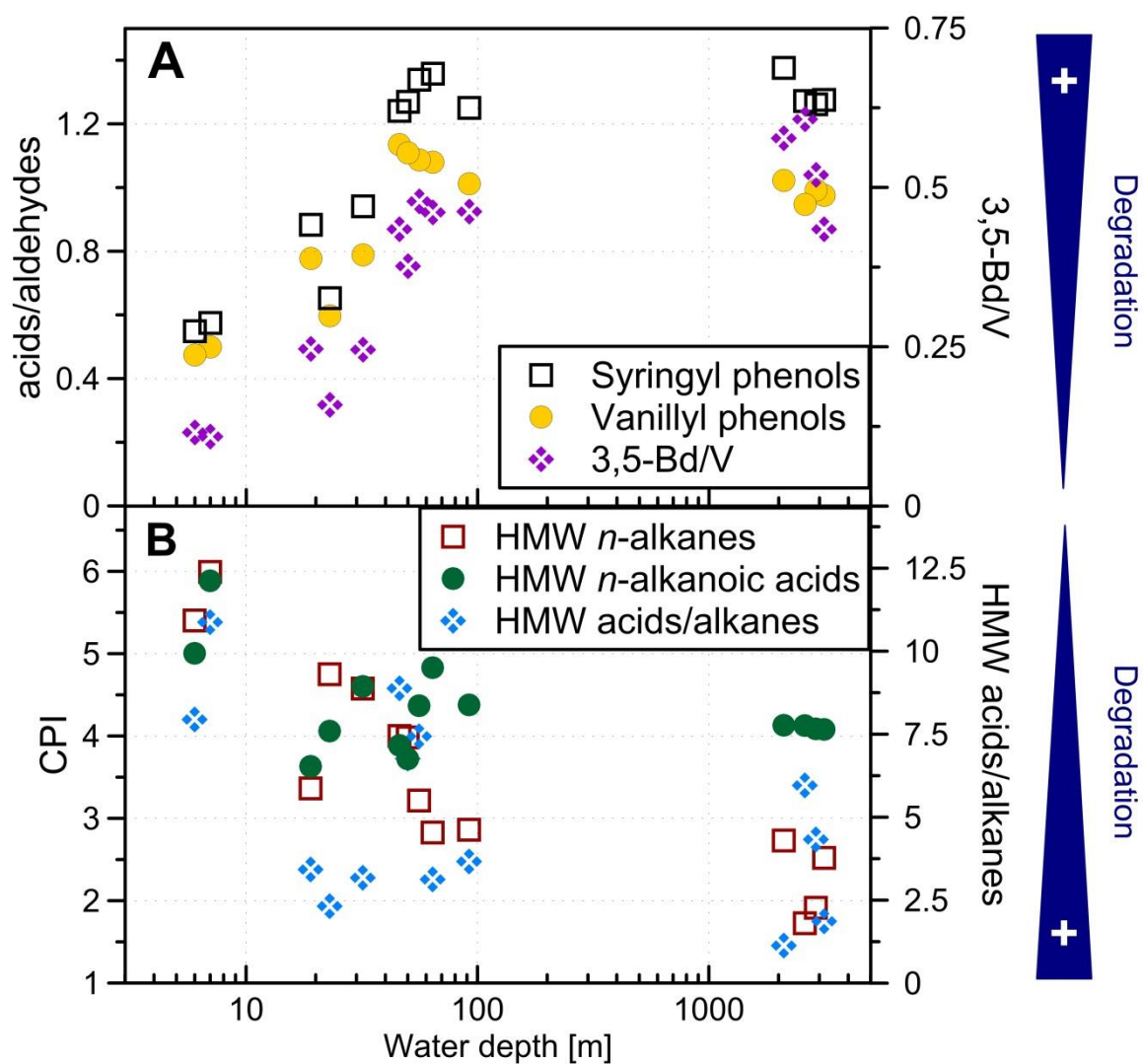


Figure 6: Degradation proxies for TerrOC, blue triangles point toward lower extent of degradation: (A) CuO-oxidation derived ratios Sd/SI, Vd/VI and 3,5-Bd/V. (B) Carbon preference indices (CPI) of HMW *n*-alkanes and *n*-alkanoic acids and the ratio of HMW *n*-alkanoic acids to HMW *n*-alkanes.

984 **Table 1: List of surface sediment samples from the Laptev Sea transect**

ID	Sample type	Lat ° N	Long ° E	Water depth m	OC mg g ⁻¹	SA m ² g ⁻¹	δ ¹³ C ‰	Δ ¹⁴ C ‰	SiO ₂ wt %	Al ₂ O ₃ wt %	CaO wt %
SW-1	0-0.5cm	78.942	125.243	3146	10.4	34.0	-22.34	-418	60.3	16.5	2.4
SW-2	0-0.5cm	78.581	125.607	2900	13.8	38.3	-22.70	-392	57.8	17.2	2.1
SW-3	0-0.5cm	78.238	126.150	2601	10.6	31.8	-22.61	-426	62.1	16.0	1.6
SW-4	0-0.5cm	77.855	126.664	2106	13.2	41.5	-22.51	-428	56.6	17.5	1.3
SW-6	0-1cm	77.142	127.378	92	7.6	14.9	-23.20	-364	72.0	12.6	1.7
SW-14	0-1cm	76.894	127.798	64	8.9	19.4	-24.33	-314	71.3	12.5	1.5
SW-23	0-1cm	76.171	129.333	56	15.8	21.7	-24.96	-333	68.9	13.6	1.4
YS-4	0-1cm	75.987	129.984	50	13.4 ^a	31.4	-24.76 ^a	-284 ^a	63.8	15.1	1.3
SW-24	0-1cm	75.599	129.558	46	10.7	37.0	-24.30	-437	62.5	15.4	1.2
YS-6	0-1cm	74.724	130.016	32	18.6 ^a	31.6	-25.60 ^a	-465 ^a	62.1	16.1	1.3
YS-9	Grab	73.366	129.997	23	13.1 ^b	16.9	-26.10 ^b	-415 ^b	70.8	14.0	1.3
YS-13	0-1cm	71.968	131.701	19	18.9 ^a	23.5	-25.90 ^a	-543 ^a	61.6	17.4	0.8
YS-14	0-1cm	71.630	130.050	7	19.1 ^a	11.4	-26.20 ^a	-504 ^a	69.6	15.0	1.6
TB-46	Grab	72.700	130.180	6	25.8 ^a	12.0 ^c	-26.50 ^a	-436 ^a	67.9	15.2	1.8

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986 ^a Data from Vonk et al. (2012); ^b data from Tesi et al. (2016); ^c data from Karlsson et al.
987 (2014).

988 **Table 2: Biomarker results for surface sediment samples from the Laptev Sea transect**

ID	Lignin $\mu\text{g m}^{-2}$	Cutin $\mu\text{g m}^{-2}$	HMW* alkanes $\mu\text{g m}^{-2}$	HMW** acids $\mu\text{g m}^{-2}$	S/V	C/V	Sd/SI	Vd/VI	3,5Bd/V	CPI alk	CPI acids alk	acids/ alk
SW-1	0.56	0.063	0.14	0.27	1.1	0.09	1.3	0.98	0.43	2.5	4.1	1.9
SW-2	0.41	0.070	0.13	0.57	0.99	0.16	1.3	0.99	0.52	1.9	4.1	4.3
SW-3	0.34	0.061	0.13	0.75	0.92	0.14	1.3	0.95	0.61	1.7	4.1	6.0
SW-4	0.42	0.048	0.093	0.10	1.1	0.08	1.4	1.0	0.58	2.7	4.1	1.1
SW-6	0.68	0.12	0.18	0.67	0.90	0.19	1.2	1.0	0.46	2.9	4.4	3.7
SW-14	1.0	0.12	0.15	0.46	1.0	0.08	1.4	1.1	0.46	2.8	4.8	3.1
SW-23	0.97	0.17	0.16	1.2	0.96	0.11	1.3	1.1	0.48	3.2	4.4	7.4
YS-4	0.84	0.17	0.13	0.92	0.83	0.17	1.3	1.1	0.38	4.0	3.7	6.8
SW-24	0.76	0.23	0.18	1.6	0.84	0.19	1.2	1.1	0.43	4.0	3.9	8.9
YS-6	2.7	0.71	0.31	0.97	0.68	0.27	0.94	0.79	0.25	4.6	4.6	3.2
YS-9	10	2.7	0.45	1.1	0.60	0.31	0.65	0.60	0.16	4.7	4.1	2.3
YS-13	5.8	2.3	0.64	2.3	0.65	0.29	0.88	0.78	0.25	3.4	3.6	3.4
YS-14	34	9.0	1.1	12	0.66	0.42	0.57	0.50	0.11	6.0	5.9	11
TB-46	45	11	0.83 ^d	6.6 ^d	0.64	0.53	0.55	0.47	0.12	5.4 ^d	5.0 ^d	7.9 ^d

989

990 * carbon chain-lengths 23-34; ** carbon chain-lengths 24-30.

991 ^d recalculated data from Karlsson et al. (2011).