



1 **Fate of terrigenous organic matter across the Laptev Sea**
2 **from the mouth of the Lena River to the deep sea of the**
3 **Arctic interior**

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14 **Abstract**

15 Ongoing global warming in high latitudes may cause an increasing supply of permafrost-
16 derived organic carbon through both river discharge and coastal erosion to the Arctic
17 shelves. Here it can be either buried in sediments, transported to the deep sea or degraded
18 to CO₂ and outgassed, potentially constituting a positive feedback to climate change.
19 This study aims to assess the fate of terrestrial organic carbon (TerrOC) in the Arctic marine
20 environment by exploring how it changes in concentration, composition and degradation
21 status across the wide Laptev Sea shelf. We analyzed a suite of terrestrial biomarkers as
22 well as source-diagnostic bulk carbon isotopes ($\delta^{13}\text{C}$, $\Delta^{14}\text{C}$) in surface sediments from a
23 Laptev Sea transect spanning more than 800 km from the Lena River mouth (~10 m water
24 depth) across the shelf to the slope and rise (2000-3000 m water depth). These data provide
25 a broad view on different TerrOC pools and their behavior during cross-shelf transport. The
26 concentrations of lignin phenols, cutin acids and high-molecular weight (HMW) wax lipids
27 (tracers of vascular plants) decrease by 89-99 % along the transect. Molecular-based
28 degradation proxies for TerrOC (e.g., the carbon preference index of HMW lipids, the HMW
29 acids/alkanes ratio and the acid/aldehyde ratio of lignin phenols) display a trend to more
30 degraded TerrOC with increasing distance from the coast. We infer that the degree of
31 degradation of permafrost-derived TerrOC is a function of the time spent under oxic
32 conditions during protracted cross-shelf transport. Future work should therefore seek to
33 constrain cross-shelf transport times in order to compute a TerrOC degradation rate and
34 thereby help to quantify potential carbon-climate feedbacks.



35 1 Introduction

36 Amplified global warming in high latitudes has raised growing concern about potential
37 positive carbon-climate feedbacks. Arctic soils store about half of the global soil organic
38 carbon (Tarnocai et al., 2009), with 60 % of this in perennially frozen grounds (Hugelius et
39 al., 2014). With ongoing climate change these vast carbon reservoirs become increasingly
40 vulnerable. Mobilization and transport of old terrigenous organic carbon (TerrOC) into the
41 Arctic Ocean is expected to intensify (Gustafsson et al., 2011) through enhancing river
42 discharge (McClelland et al., 2008) with augmenting sediment loads (Gordeev, 2006;
43 Syvitski, 2002) and accelerating coastal erosion (Günther et al., 2013). This material can be
44 buried in the sediments of the Arctic shelves, transported across the margin towards deeper
45 basins or degraded and re-introduced into the modern carbon cycle as CO₂, thereby not only
46 providing a potential positive feedback to global warming, but also causing severe ocean
47 acidification (Semiletov et al., 2016). The fate of permafrost-released TerrOC in the marine
48 environment is thus crucial for future climate projections, yet insufficiently understood (Vonk
49 and Gustafsson, 2013).

50 The East Siberian Arctic Shelf (ESAS) is with a width of > 800 km the world's largest
51 continental shelf. It receives TerrOC both from the erosion of the East Siberian shoreline,
52 largely consisting of organic-rich, late-Pleistocene ice-complex deposits (Yedoma), and via
53 the Great Russian Arctic Rivers, which drain extensive areas of continuous and
54 discontinuous permafrost. The Laptev Sea is a representative for the TerrOC dominated
55 Siberian shelf seas, since its main organic carbon input originates from substantial coastal
56 erosion (as observed in the Buor-Khaya Bay, Sánchez-García et al., 2011; Semiletov et al.,
57 2011; Vonk et al., 2012) and the Lena River, the main fluvial sediment source for the entire
58 ESAS (Holmes et al., 2002).

59 Previous studies have focused on near-shore areas and the inner shelf (e.g. Bröder et al.,
60 2016; Charkin et al., 2011; Feng et al., 2015; Karlsson et al., 2011; Salvadó et al., 2015;
61 Sánchez-García et al., 2011; Semiletov et al., 2005, 2012; Tesi et al., 2014; Winterfeld et al.,
62 2015b, 2015a; Vonk et al., 2010, 2012, 2014). They reported large fractions of old TerrOC in



63 particulate organic carbon (POC) and surface sediments close to the coast, using different
64 approaches such as applying carbon-isotope-based source apportionment (e.g. (Gustafsson
65 et al., 2011; Semiletov et al., 2005; Vonk et al., 2010, 2012, 2014) and by analyzing
66 terrigenous biomarkers in both surface sediments (e.g. Feng et al., 2013; Stein and
67 Macdonald, 2004; Tesi et al., 2014, 2016) and POC in the water column (e.g. Charkin et al.,
68 2011; Karlsson et al., 2011; Winterfeld et al., 2015a). This is the first study that encompasses
69 sampling stations along the entire transect from the Lena River mouth, across the wide
70 Laptev Sea shelf, to the continental slope and rise. Our major objective was to gain new
71 insights in the behavior of different TerrOC pools, in particular investigating potential
72 degradation of permafrost-released material along the land-shelf-basin continuum. We have
73 therefore characterized TerrOC in surface sediments along the Laptev Sea transect on both
74 bulk and molecular level, exploiting source-diagnostic bulk carbon isotopes ($\delta^{13}\text{C}$, $\Delta^{14}\text{C}$) as
75 well as an extensive biomarker suite (lignin phenols and cutin acids obtained by alkaline CuO
76 oxidation and high-molecular-weight solvent-extractable lipids, such as *n*-alkanes and *n*-
77 alkanolic acids).



78 2 Material and Methods

79 2.1 Study area

80 The Laptev Sea is the shallowest of the Arctic shelf seas with an average depth of 48 m
81 (Jakobsson et al., 2004). It spans over 498,000 km² with a volume of 24,000 km³ and is
82 located between the Kara Sea and Severnaya Zemlya in the West and the East Siberian Sea
83 and the New Siberian Islands in the East. The main sources of particulate organic carbon
84 (POC) for the Laptev Sea are terrigenous, both from coastal erosion and river runoff
85 (Sánchez-García et al., 2011; Stein and Macdonald, 2004). Marine primary production is
86 limited to on average two ice-free months per year and therefore generally low. Nutrient-poor
87 waters on the Siberian shelves resulting from a strong stratification further impede
88 phytoplankton growth (Sakshaug, 2004).

89 The destabilization of Pleistocene Ice-Complex Deposits along the coastline is a main
90 sediment source for the Laptev Sea (Rachold et al., 2000). The total POC input from coastal
91 erosion to Laptev and East Siberian Sea is estimated to be between 4.0 Tg yr⁻¹ (Semiletov,
92 1999; Stein and Fahl, 2000) and 22 ± 8 Tg yr⁻¹ (including net subsea permafrost-carbon
93 erosion, Vonk et al., 2012).

94 The Lena River is estimated to provide 20.7 Tg of sediment per year to the Laptev Sea
95 (Holmes et al., 2002) with an average water discharge of 588 km³ yr⁻¹ (Holmes et al., 2012).
96 It drains a watershed of ~2.46 x 10⁶ km² (Holmes et al., 2012), of which 77 % is underlain by
97 continuous permafrost (Amon et al., 2012). Water discharge peaks in June, during the spring
98 flood, when about 75 % of total organic carbon is delivered (Rachold et al., 2004). Total POC
99 discharge by the Lena River can be up to 0.38 Tg yr⁻¹ (Semiletov et al., 2011).

100 Sediment transport pathways are largely influenced by the prevailing atmospheric conditions:
101 During cyclonic summers (i.e. positive phase of the Arctic Oscillation), northerly winds
102 predominate, strengthening the Siberian Coastal Current, which transports Lena River water
103 masses along the coast towards the East Siberian Sea; whereas during anticyclonic
104 summers (i.e. negative phase of the Arctic Oscillation and mainly southerly winds) the Lena
105 River plume is exported onto the mid-shelf and towards the deep part of the Arctic Ocean



(Charkin et al., 2011; Dmitrenko et al., 2008; Guay et al., 2001; Wegner et al., 2013; Weingartner et al., 1999). Sediment transport in the Laptev Sea is strongly seasonal. The main transport of Lena River water with high concentrations of suspended particulate matter (SPM) towards the mid-shelf takes place shortly after river-ice breakup (Wegner et al., 2005). During the ice-free summer, SPM circulates between inner and mid-shelf with very little material escaping over the shelf break to the deeper parts of the Arctic Ocean. Significant sediment export is suggested to happen during freeze-up through SPM that is incorporated in sea ice and then transported across the continental margin (Dethleff, 2005; Eicken et al., 1997) or through the formation of dense bottom water resulting from brine ejection (Dethleff, 2010; Ivanov and Golovin, 2007). Hardly any sediment transport occurs beneath the ice cover.

Holocene-scale linear sedimentation rates for the Laptev Sea vary between 0.12 and 0.59 mm yr⁻¹ according to ¹⁴C dating of marine bivalves (Stein and Fahl, 2004, and citations therein), whereas centennial-scale ²¹⁰Pb-derived rates for the more recent Laptev Sea can be up to 1.3 mm yr⁻¹ (Vonk et al., 2012). These rates do not seem to be correlated with water depth on the shelf, but values for the continental slope and rise tend to be on the lower end (0.12-0.38 mm yr⁻¹) (Stein and Fahl, 2004, and citations therein).

123

124 2.2 Sampling

Sediment sampling locations span from close to the Lena River mouth and in the Buor-Khaya Bay, across the shelf, to the continental slope and rise, covering a transect of > 800 km with water depths increasing by more than two orders of magnitude. Samples SW-1, SW-2, SW-3, SW-4, SW-6, SW-14, SW-23 and SW-24 were collected during the SWERUS-C3 expedition on IB *ODEN* during summer 2014 using an Oktopus multicorer (8 Plexiglas tubes, 10 cm diameter). All other samples were collected during the International Siberian Shelf Study (ISSS-08) expedition onboard the RV *Yacob Smirnitskyi* during summer 2008. The YS-4, YS-6, YS-13 and YS-14 samples were taken with a GEMAX gravity corer (2 Plexiglas tubes, 9 cm diameter); YS-9 and TB-46 were collected with a Van Veen grab sampler. For



the grab samples only surface sediments (uppermost cm) were subsampled and used in this study. Sediment cores were cut into 1 cm slices within 24 hours after sampling. To account for lower sediment accumulation rates on the rise, for SW-1, SW-2, SW-3 and SW-4 a higher resolution of 0.5 cm for the top 10 cm was chosen. The depositional age for all samples is thus between ~8 and ~70 years (depending on which sedimentation rates are employed). All samples were kept frozen throughout the expedition and freeze-dried upon arrival. See Semiletov and Gustafsson (2009) for more information on the ISSS-08 expedition. For exact sampling locations see Table 1.

2.3 Surface area

All surface area analyses have been performed on a micromeritics Gemini VII Surface Area and Porosity analyzer. Freeze-dried subsamples of ~0.7 g were furnace-dried at 400 °C for 12 h and gently cooled down to room temperature. They were then desalted by repeated mixing with 50 ml of MilliQ water and centrifugation (20 min at 8000 rpm), followed by further freeze-drying. Directly prior to analysis they were degassed in a Micromeritics FlowPrep 060 Sample Degas System for 2 h at 200 °C under a constant nitrogen flow. Each analysis was initiated by measuring the free space in the vial. The specific surface areas were derived from 6 pressure-point measurements (relative pressure $p/p_0 = 0.05$ -0.3, equilibration time 5 s) with nitrogen as adsorbing gas (Brunauer et al., 1938). The instrumental error was 0.1-0.3 m² g⁻¹, which corresponds to a relative error of about 1 %. The performance of the instrument was monitored with the surface area reference material Carbon Black (21.0 ± 0.75 m² g⁻¹) provided by Micromeritics.

2.4 X-Ray Fluorescence

The mineral composition of ~1 g freeze-dried, homogenized subsamples was characterized with a wavelength dispersive sequential Philips PW2400 X-ray Fluorescence (XRF) spectrometer. The XRF was operated under vacuum conditions on samples prepared as glass beads using lithium tetraborate and melted with a fluxer Claisse Fluxy (~1150°C)



162 (Mercone et al., 2001). The relative error was less than 0.6 % for major elements and less
163 than 3 % for trace elements. In this study only SiO₂, Al₂O₃ and CaO are reported.

164

165 2.5 Bulk elemental and carbon isotope analysis

166 Total organic carbon (TOC) concentrations and $\delta^{13}\text{C}$ isotopic composition were determined
167 at the Stable Isotope Laboratory, Department of Geological Sciences, Stockholm University.
168 Homogenized subsamples of ~10 mg were repeatedly acidified (HCl, 1.5 M, Ag capsules) to
169 remove carbonates (Nieuwenhuize et al., 1994). TOC concentrations and $\delta^{13}\text{C}$ isotopic
170 composition were measured simultaneously with a Carlo Erba NC2500 elemental analyzer
171 connected via a split interface to a Finnigan MAT Delta V mass spectrometer. TOC
172 concentrations were blank corrected and the relative error was < 1 %. Stable isotope data
173 are reported relative to VPDB using the $\delta^{13}\text{C}$ notation.

174 Radiocarbon analyses of acidified samples were conducted at the US National Ocean
175 Sciences Accelerator Mass Spectrometry (NOSAMS) Facility of the Woods Hole
176 Oceanographic Institution, USA, according to their standard routines (Pearson et al., 1998).
177 The relative error of the measurements was < 0.5 %. Radiocarbon data are reported using
178 the $\Delta^{14}\text{C}$ notation following Stuvier and Polach (1977).

179

180 2.6 Biomarkers

181 2.6.1 CuO-oxidation products

182 Microwave-assisted alkaline CuO oxidation was performed according to the method
183 established by Goñi and Montgomery (2000). Homogenized subsamples of 100-400 mg of
184 sediment (corresponding to 2-5 mg OC) were combined with 300 mg of copper(II) oxide and
185 50 mg of ferrous ammonium sulfate and oxidized under oxygen-free conditions (degassed
186 NaOH, 8 wt %) at 150 °C for 90 min using an UltraWAVE Milestone 215 Microwave. After
187 oxidation, known amounts of trans-cinnamic acid and ethyl vanillin were added as recovery
188 standards. Samples were acidified to pH 1 with HCl (12 M) and repeatedly extracted with
189 ethyl acetate. Anhydrous Na₂SO₄ was added to remove remaining water. The solvent was



190 evaporated and extracts re-dissolved in pyridine. For quantification, subsamples were
191 derivatized with bis-trimethylsilyl-trifluoroacetamide (BSTFA) + 1 % trimethylchlorosilane
192 (TMCS) and analyzed on a gas-chromatograph mass spectrometer in full scan mode (GC-
193 MS, Agilent) using a DB5-MS capillary column (60 m x 250 μ m, 0.25 μ m stationary phase
194 thickness, Agilent J&W) with a temperature profile of initially 60 °C followed by a ramp of 5
195 °C min⁻¹ until reaching and holding 300 °C for 5 min. The quantification of lignin phenols,
196 benzoic acids, and p-hydroxybenzenes was achieved by comparison to the response factors
197 (key ions) of commercially available standards. For cutin-derived products, fatty acids and
198 dicarboxylic acids the response factor of trans-cinnamic acid was used as in Goñi et al.
199 (1998).

200

201 2.6.2 Solvent-extractable Lipids

202 Wax lipids were extracted by means of accelerated solvent extraction (Dionex ASE 300)
203 using dichloromethane:methanol (9:1) according to the method described by Wiesenberget
204 al. (2004). Pre-rinsed stainless-steel vessels were loaded with ~3 g of freeze-dried sediment,
205 filled up with pre-combusted glass beads and pre-combusted glass fiber filters at both ends.
206 Two extraction cycles were performed per sample applying a static pressure of 1500 psi and
207 a temperature of 80 °C for 5 min after a heating phase of 5 min. The flush volume was 50 %
208 of the 34 ml cell size with a purging time of 100 s.

209 Extracts were further cleaned (addition of activated Cu for sulfur and anhydrous Na₂SO₄ for
210 water removal) and then separated into a neutral and an acid fraction using BondElut
211 cartridges (bonded phase NH₂, Varian), eluting with dichloromethane:isopropanol (2:1) for
212 the neutral and methyl *tert*-butyl ether with 4 % acetic acid for the acid fraction according to
213 the method described by van Dongen et al. (2008a). The neutral fraction was further
214 separated into a polar and a non-polar fraction with an Al₂O₃ column. For each of the three
215 compound classes *n*-alkanes (neutral non-polar fraction), *n*-alkanols (neutral polar fraction)
216 and *n*-alkanoic acids (acid fraction) ~10 mg of one internal standard, d₅₀-tetracosane, 2-
217 hexadecanol and d₃₉-eicosanoic acid respectively, were added to the sediment samples prior



218 to extraction. All fractions were then analyzed on a GC–MS (Agilent) using the same column
219 and temperature program as for the CuO products. The polar and acid fractions were
220 derivatized with BSTFA + 1 % TMCS prior to analysis. Quantification was performed using a
221 5-point calibration curve with commercially available standards.



3 Results and Discussion

The fate of permafrost-released terrigenous organic carbon (TerrOC) across the Laptev Sea shelf is controlled by competing processes. Degradation and sorting, as well as replacement of TerrOC by autochthonous marine organic matter all co-occur to varying degrees during cross-shelf transport. To disentangle their effects on the fate of permafrost-released TerrOC we first report changes in bulk sediment and OC properties and then focus on differences on the molecular level.

3.1 Characterization of the transect on a bulk level

Bulk total organic carbon (TOC) concentrations decreased across the shelf with highest values (~2 %) at shallow water depths and lowest values on the shelf edge (~0.8 %); at high water depths (> 2000 m) concentrations were slightly higher (~1 %). TOC values and the general pattern were in accordance with previous data from the Laptev Sea (Semiletov et al., 2005; Shakhova et al., 2015; Stein and Fahl, 2004; Vonk et al., 2012) and within the same range of those measured for the North American Arctic margin (Goni et al., 2013).

Normalizing TOC concentrations to the mineral-specific surface area (SA) helps to understand the influence of physical sorting and preferential deposition on the observed TOC trends since SA is correlated to the sediment grain size to a first order approximation. To test if the mineral surface area is altered by the input of autochthonous organisms with siliceous or carbonaceous skeleton (e.g. silicoflagellates/diatoms or foraminifera/shells respectively), the mineral composition of the sediments was examined by X-ray fluorescence analysis.

There were no apparent trends with water depth for either $\text{SiO}_2/\text{Al}_2\text{O}_3$ or $\text{CaO}/\text{Al}_2\text{O}_3$; therefore, marine production is not expected to have a measureable effect and SA can thus be regarded as a conservative parameter. This was also confirmed by low biogenic silica concentrations for the Laptev Sea reported earlier (< 1.4 %, Mammone, 1998).

The relationship between TOC and SA has been widely studied on continental margins (e.g. Blair and Aller, 2012; Keil et al., 1994; Mayer, 1994). The TOC/SA ratios of typical river suspended sediments range between 0.4 and 1 mg m^{-2} . TOC/SA ratios > 1 mg m^{-2} have



250 been found in areas with high TOC supply (e.g. river outlets) and where the deposited
251 organic matter had spent little time under oxic conditions (short oxygen exposure time, OET).
252 Ratios $< 0.4 \text{ mg m}^{-2}$ generally correspond to sediments from deeper parts of the ocean and
253 long OETs. Accordingly, the TOC/SA values along the Laptev Sea transect displayed a
254 strong decrease from 2.2 and 1.7 mg m^{-2} close to the Lena River delta (water depths of 11
255 and 7 m, respectively) to about 0.3 mg m^{-2} at water depths greater than 2000 m (Fig. 2A),
256 proposing extensive TOC loss during cross-shelf transport.

257 Bulk TOC isotopes have been broadly used to distinguish between organic matter sources.
258 Radiocarbon isotopes (^{14}C) convey information about the age of organic material, with
259 younger OC having higher $\Delta^{14}\text{C}$ values. Marine organic matter produced primarily from CO_2
260 is expected to have modern ^{14}C signatures, whereas permafrost-derived TerrOC has aged
261 both on land and during transport and has thus more depleted ^{14}C values. The $\Delta^{14}\text{C}$ values
262 for our Laptev Sea transect were generally low ($< -280 \text{ ‰}$, Fig. 2B), suggesting a significant
263 input of pre-aged TerrOC (as in Vonk et al., 2012). Bulk TOC showed less depleted $\Delta^{14}\text{C}$
264 signatures with increasing distance from land on the shelf (from about -500 ‰ to about -340
265 ‰ on the outer shelf, Fig. 2B), reflecting a dilution of older TerrOC with younger marine
266 material. On the slope and rise, however, $\Delta^{14}\text{C}$ values decreased again to about -410 ‰ .
267 This difference may be a result of ageing during lateral transport and/or after deposition due
268 to lower accumulation rates on slope and rise. The range between -340 ‰ and -410 ‰
269 corresponds to a $\Delta^{14}\text{C}$ age difference of about 900 years; however, the depositional age
270 differences between shelf and slope samples were estimated to be less than 80 years (see
271 Section 2.2). Ageing after burial alone does therefore not explain the difference in $\Delta^{14}\text{C}$. Keil
272 et al. (2004) estimated a lateral transport time of 1800 years across the Washington margin
273 (158 km) from $\Delta^{14}\text{C}$ data of bulk OC in surface sediments. For the $> 200 \text{ km}$ distance
274 between mid-shelf and rise a bulk ageing of 900 years does therefore not seem
275 unreasonable. It has to be taken into account, however, that mainly the TerrOC fraction of
276 the bulk OC is subject to such protracted lateral transport. Transport times would thus have
277 to be significantly higher in order to explain this age difference for the entire bulk OC.



278 For stable carbon isotopes (^{13}C), terrigenous sources are generally more depleted than
279 marine organic matter (Fry and Sherr, 1984). In this study, values for $\delta^{13}\text{C}$ of TOC ranged
280 between -26.5‰ and -22.3‰ . The trend towards more enriched TOC with increasing
281 distance from the coast (Fig. 2B) can be explained by a growing proportion of marine organic
282 matter. However, the $\delta^{13}\text{C}$ signature of the marine source appeared to be heavier than typical
283 marine planktonic material in that region ($-26.7 \pm 1.2\text{‰}$, Panova et al., 2015; $-24 \pm 3\text{‰}$,
284 Vonk et al., 2012, and citations therein). One possible explanation for this discrepancy is an
285 underestimated influence of ice algae that were reported to have highly enriched $\delta^{13}\text{C}$ values
286 between -15 to -18‰ (Schubert and Calvert, 2001). Significant seafloor deposition of ice
287 algal biomass has been observed previously for the Arctic basins (Boetius et al., 2013).
288 Another option would be a more refractory, isotopically-enriched marine endmember (-21.2
289 ‰) as suggested by Magen et al. (2010). They argue that lighter isotopes are preferentially
290 consumed by bacteria, which in turn enriches the remaining marine organic matter. Following
291 their reasoning, the more enriched values observed for this transect may be interpreted as a
292 increasing proportion of refractory marine organic matter.
293 Winterfeld et al. (2015b) analyzed surface water particulate organic carbon (POC) in the
294 Lena River delta and found a mean $\delta^{13}\text{C}$ of $-29.6 \pm 1.5\text{‰}$. Karlsson et al. (2011) reported
295 similarly depleted $\delta^{13}\text{C}$ values for POC from the Buor-Khaya Bay ($-29.0 \pm 2.0\text{‰}$), while their
296 mean value for sedimentary OC for the same stations was significantly more enriched (-25.9
297 $\pm 0.4\text{‰}$) and agreed well with our data for the shallow stations ($-26.2 \pm 0.3\text{‰}$, stations YS-
298 13, YS-14 and TB-46). Lena River POC $\delta^{13}\text{C}$ values from high-discharge periods agree well
299 with the more enriched values we found for the shallow stations (Rachold and Hubberten,
300 1998). Stein and Fahl (2004), Semiletov et al. (2011, 2012) and Vonk et al. (2012) presented
301 similar $\delta^{13}\text{C}$ ranges and trends for sediments from parts of the Laptev Sea as is reported in
302 the current study for the entire width of the Laptev Sea shelf. For the Arctic Amerasian
303 Continental shelf, Naidu et al. (2000) reported contrasts in absolute $\delta^{13}\text{C}$ values comparing
304 surface sediment samples from different regions, but all commonly displayed an increasing



305 trend for $\delta^{13}\text{C}$ values across the shelf, suggesting a growing fraction of marine organic matter
306 with increasing distance from the coast.
307 Combining TOC/SA ratios with stable isotope signatures ($\delta^{13}\text{C}$) may serve to disentangle two
308 different processes, which occur synchronously during cross-shelf transport (as in Keil et al.
309 1997a): 1.) The net loss of TerrOC and 2.) the replacement of TerrOC with autochthonous
310 marine OC.
311 Net loss of TerrOC, caused by either degradation or hydrodynamic sorting during transport,
312 has been quantified previously using TOC/SA ratios (Aller and Blair, 2006; Keil et al., 1997a).
313 The carrying-capacity of inorganic particles for OC is assumed to be a function of the SA
314 (Mayer, 1994); a decrease in TOC/SA values can therefore be regarded as TOC net loss.
315 Replacement of TerrOC with autochthonous marine OC does not change this ratio. But since
316 marine OC is known to be isotopically enriched in $\delta^{13}\text{C}$ over TerrOC, this process is recorded
317 by an increasing isotopic signature. Along the Laptev Sea transect, both processes seemed
318 to play an important role (Fig. 2C). High TOC/SA values close to the Lena River decreased
319 sharply, pointing to extensive net loss, while the increase in $\delta^{13}\text{C}$ values was minor in this
320 area. Once TOC/SA ratios were $< 0.8 \text{ mg m}^{-2}$ (water depths $> 20 \text{ m}$), the isotopic changes
321 and thus the replacement of TerrOC with marine OC became increasingly important. Similar
322 trends were observed for the Amazon River delta (Keil et al., 1997b).
323 However, the TOC/SA trend in the shallower sediments is likely driven by both degradation
324 of OC bound to the mineral matrix during cross-shelf transport and sorting of vascular plant
325 fragments that are retained in the inner-shelf. A recent study (Tesi et al., 2016) has shown
326 that ~50 % of the total OC pool in the inner Laptev shelf surface sediments exists in the form
327 of large vascular plant fragments. They are trapped close to the coast due to their size
328 (Stoke's law), while the OC bound to the fine mineral matrix is more buoyant and transported
329 offshore towards deeper waters.

330

331 3.2 Molecular indicators of organic matter sources

332 3.2.1 Biomarker distributions



333 The abundances of different source-diagnostic molecular proxies have been extensively
334 investigated to elucidate complex carbon-cycling mechanisms. In this study, a biomarker
335 suite of CuO oxidation products and solvent-extractable lipids was analyzed in order to gain
336 more insights on TerrOC sources and degradation status along the Laptev Sea transect. All
337 biomarker concentrations were normalized to the sediment-specific surface area (SA)
338 instead of OC content to avoid the signals being overshadowed by other carbon pools. As
339 shown by the lack of water depth related changes in the mineral composition (Section 3.1),
340 mineral-matrix dilution by biogenic material is negligible.

341 Lignin-derived phenols have been widely used to trace TerrOC in the marine environment
342 (e.g. Ertel and Hedges, 1984; Goñi and Hedges, 1995; Hedges and Mann, 1979). The lignin
343 macro-molecule is only synthesized in vascular plants (and certain seaweed species that are
344 not existent in the study area) to render stability to the cell walls. Across the shelf, lignin
345 loadings decreased substantially with increasing distance from the coast/water depth ($45 \mu\text{g}$
346 m^{-2} close to the coast, $0.43 \pm 0.09 \mu\text{g m}^{-2}$ for the deep stations; loss of $99.1 \pm 0.2 \%$, Fig. 3A).

347 Cutin-derived hydroxy fatty acids are another compound class obtained from CuO oxidation,
348 which have been used in parallel with lignin phenols (e.g. Goñi et al., 2000; Prahl et al.,
349 1994). They are mainly associated with the soft tissues of vascular plants such as leaves and
350 needles. Cutin acid loadings displayed a similar trend as lignin phenols ($11 \mu\text{g m}^{-2}$ close to
351 the coast, $0.061 \pm 0.010 \mu\text{g m}^{-2}$ for the deep stations; loss of $99.4 \pm 0.1 \%$, Fig 3A).

352 Similar values and sharp declines with increasing distance from the coast for lignin and cutin
353 have been observed for the whole East Siberian Arctic Shelf (ESAS) (Tesi et al., 2014). A
354 recent study (Winterfeld et al., 2015a) for the Buor-Khaya Bay (5.8-17 m water depth)
355 reported lignin phenol concentrations on the same order of magnitude, up to 40 % higher for
356 the shallowest samples, and decreasing with increasing depth. For the Beaufort Sea shelf,
357 Goñi et al. (2000) found a less drastic decline in lignin phenols and cutin acids going from 5
358 m water depth to 210 m, which likely reflected both lower concentrations in the shallow
359 waters (factor of ~2), and a narrower and steeper shelf. Lignin phenols were also higher at
360 greater water depths than on the ESAS. This may reflect the differences in bathymetry: since



the Beaufort Sea shelf is not as wide as, but steeper than the ESAS, lateral transport is possibly faster, leaving less time for organic matter to be degraded along the way. A comparison between different shelf-slope systems across the North American Arctic margin (Goni et al., 2013) revealed very low lignin and cutin concentrations for the Canadian Archipelago, Lancaster Sound and Davis Strait, whereas both concentrations and trends with water depth for the Beaufort Sea, Chuckchi Sea and Bering Sea were similar to the results from this study. An exception to these patterns was Barrow Canyon, where at water depths of > 2000 m lignin and cutin concentrations were as high as the ones observed close to the Lena River delta, pointing to efficient rapid TerrOC transfer with comparably short oxygen exposure times through this active canyon (Goni et al., 2013).

Solvent extractable high-molecular weight (HMW) *n*-alkanes and *n*-alkanoic acids make up the major part of epicuticular leaf waxes (Eglinton and Hamilton, 1967) and have been broadly employed as TerrOC biomarkers (for the Arctic Ocean e.g. van Dongen et al., 2008; Yunker et al., 1995, 2005). HMW wax lipids in this study also presented a decreasing trend with increasing water depth/distance from the coast, but to a lesser extent than lignin phenols or cutin acids (HMW *n*-alkanes, carbon chain-lengths of 23-34: $1.1 \mu\text{g m}^{-2}$ close to the coast, $0.12 \pm 0.02 \mu\text{g m}^{-2}$ for the deep stations; HMW *n*-alkanoic acids, chain-lengths 24-30: $12 \mu\text{g m}^{-2}$ close to the coast, $0.42 \pm 0.29 \mu\text{g m}^{-2}$ for the deep stations; loss of $89 \pm 2 \%$ and $96 \pm 3 \%$, respectively).

Previous studies in the same area reported similar lipid biomarkers concentrations, which confirm the magnitude of the decreasing trends with increasing water depth (Karlsson et al., 2011; Vonk et al., 2010). HMW *n*-alkane concentrations in the Beaufort and the Chuckchi Sea (Belicka et al., 2004; Yunker et al., 1993) are in accordance with the ones measured on the ESAS, but the shallowest sample on the Beaufort Shelf is ~2 times lower than the shallow ESAS samples. This might imply that sediments transported by the Mackenzie River to the Beaufort Shelf have lower TerrOC concentrations than Lena River transported sediments. For the Mackenzie Shelf Goñi et al. (2000) used lignin phenols and cutin acids to estimate a marine $\delta^{13}\text{C}$ endmember and therewith derived a terrigenous contribution of



almost 80 % for the shallowest sediments, while rough estimates from C/N and $\delta^{13}\text{C}$ data suggested that TerrOC made up only 30-50 % of the organic carbon (Macdonald et al., 2004). For the Lena Delta, source apportionment calculations using $\delta^{13}\text{C}$ and $\Delta^{14}\text{C}$ data attributed up to 83 % of the organic carbon to terrigenous sources (Vonk et al., 2012). All TerrOC biomarker loadings displayed a strong decrease across the shelf, but their relative losses differ substantially between compound classes (Fig. 3C). These findings agree with previous results for the ESAS (Tesi et al., 2014), where similar differences between biomarkers were reported. A larger decrease was observed for lignin than for cutin, in contrast to this study. The different extents of biomarker losses for the different compound classes may either be attributed to preferential degradation of lignin phenols and cutin acids, implying that they are more labile than HMW *n*-alkanes and *n*-alkanoic acids, or sorting during transport, suggesting that they are associated with a sediment fraction that is hydraulically retained and carried less efficiently to the outer shelf/slope. A recent study (Tesi et al., 2016) aimed to disentangle these two processes by analyzing different fractions of bulk surface sediments from three transects (with three stations each) on the ESAS. The fractions were separated according to density (1.8 g cm^{-3} cutoff), size ($>63 \mu\text{m}$, $38\text{--}63 \mu\text{m}$, $< 38 \mu\text{m}$) and settling velocity (1 m d^{-1} cutoff). The highest lignin phenol abundance was found in low-density plant fragments ($26\text{--}55 \text{ mg g}^{-1} \text{ OC}$). These large particles have a higher settling velocity (Stokes' law) and are therefore hydraulically retained close to the coast. Cutin acids and plant wax lipids were mainly associated with the mineral high-density fine ($< 38 \mu\text{m}$, $> 1 \text{ m d}^{-1}$) and ultrafine ($< 38 \mu\text{m}$, $< 1 \text{ m d}^{-1}$) fractions. Within the fine and ultrafine fractions, which made up about 95 % of the organic carbon on the outer shelf, they found drastic losses of all biomarkers with increasing distance from the coast, which they attributed to degradation during the protracted cross-shelf transport. Relative decreases appeared to depend on the number of functional groups of the compound class: $98 \pm 1 \%$ for lignin phenols, $97 \pm 1 \%$ for cutin acids, $96 \pm 1 \%$ for HMW *n*-alkanoic acids and $89 \pm 4 \%$ for HMW *n*-alkanes. According to that study, the steep cross-shelf gradients observed here for lignin phenols can be attributed to both hydrodynamic sorting close to the coast and degradation



417 during transport. The other compounds analyzed are mainly affected by degradation and
418 experience sorting to a lesser extent.
419 Degradation after burial is assumed to play only a minor role in this study. Differences in
420 sedimentation ages are expected to be small (Section 2.1) and a study on centennial-scale
421 sediment cores from the East Siberian Sea (Bröder et al., 2016) detected no significant
422 TerrOC degradation (as recorded by biomarker loss) with increasing sediment depth. Also in
423 that study, lignin phenol and cutin acid loadings were on average 20 times higher on the
424 inner than on the outer shelf, whereas for HMW *n*-alkanoic acids and *n*-alkanes the
425 difference between inner and outer shelf was only a factor of ~3-5. Contrasts between the
426 stations were found to be larger than down-core changes. This may be due to the fact that
427 the cores in that study only encompassed about one century of sedimentation ages, while
428 the protracted cross-shelf transport possibly requires much longer timescales.

429

430 3.2.2 Lignin Phenol sources

431 Relative distributions of different lignin phenol classes reveal more TerrOC source
432 information since they are specific to different plant types. Syringyl phenols are not produced
433 by gymnosperm (non-flowering) plants; elevated syringyl to vanillyl ratios (i.e. S/V > 1,
434 Hedges and Parker, 1976) are therefore attributed to more lignin phenols from angiosperm
435 (flowering) plants. These ratios have to be handled with care, though, because the
436 preferential degradation of syringyl phenols by white- and brown-rot fungi on land can also
437 alter S/V ratios (Hedges et al., 1988). S/V values for the Laptev Sea transect increased with
438 increasing water depth from ~0.65 for the inner shelf to ~1.0 for the slope/rise sediments
439 (Fig. 5A). This trend can either be explained by preferential degradation of gymnosperm
440 material or sorting during transport. Tesi et al. (2014) measured generally lower values for
441 S/V (ESAS average: 0.47, for only Lena watershed dominated locations: 0.42) recording no
442 trend with water depth. Their deepest station was located at only 69 m water depth, though,
443 whereas in this study sediments from down to 3146 m water depth were analyzed. S/V ratios
444 in Buor-Khaya Bay surface sediments (Winterfeld et al., 2015a) were also lower (0.43 ± 0.02)



on average) and displayed no trend with water depth. Within the water depth interval they studied (5.8-17 m), however, the samples analyzed in this study had also quite homogeneous S/V ratios (0.64 ± 0.02). Two sediment cores from the East Siberian Sea (Bröder et al., 2016) showed also lower S/V values (inner shelf surface sediment: 0.62, outer shelf surface sediment: 0.50) displaying no clear trends over time/down-core. For the Beaufort Sea shelf Goñi et al. (2000) detected rather high values (0.54-1.71), which (besides the very high value at 61 m water depth) agree with the data from this study. Other transects across the North American Arctic margin (Goni et al., 2013) had slightly lower S/V ratios with no observed trends with water depth.

The ratio of cinnamyl to vanillyl phenols (C/V) is associated with the relative contributions of woody versus soft material, because only non-woody vascular plants synthesize cinnamyl phenols (Hedges and Mann, 1979a). This ratio admittedly decreases with ongoing degradation (Opsahl and Benner, 1995) and may therefore not be used as an unambiguous source indicator. We observed that C/V values strongly decreased across the Laptev Sea Shelf from ~0.5 (close to the Lena River outlet) to ~0.1 (on the slope/rise, Fig. 5B), which may reflect the preferential degradation of soft tissues. This trend is not likely caused by hydrodynamic sorting, since typically the larger, low-density, woody plant fragments are retained in shallower water, whereas finer material is transported further across the shelf (e.g. Keil et al., 1994; Tesi et al., 2016).

C/V ratios in Buor-Khaya Bay sediments (Winterfeld et al., 2015a) in shallow waters were on average lower and more homogeneous (0.17 ± 0.03) than those measured in this study (0.41 ± 0.12 for the corresponding depth interval). C/V values for the entire ESAS were on average 0.15 (0.14 ± 0.07 for only Lena dominated waters) with no water depth trend (Tesi et al., 2014). In shallow sediment cores from the East Siberian Sea, Bröder et al. (2016) measured C/V ratios of 0.20 (inner shelf) and 0.13 (outer shelf) for the surface sediments with no significant trend over sediment depth. For the Mackenzie Shelf C/V values ranged between 0.16 and 0.32 and slightly increased with increasing water depth (Goñi et al., 2000). In contrast, in the Bering Sea, Chuckchi Sea, Barrow Canyon, Canadian Archipelago,



473 Lancaster sound and Davis Strait there were no C/V trends observed (Goni et al., 2013), with
474 lower values in the Canadian part (0.10 ± 0.12) and highest values on the Beaufort Sea
475 slope, where values slightly decreased with increasing depth (0.39 ± 0.07).

476 A comparison to the S/V-C/V signatures of potential Arctic plant end-members (compiled by
477 Amon et al., 2012, and citations therein, Tesi et al., 2014, and Winterfeld et al., 2015a)
478 showed that lignin phenols likely derive from both angio- and gymnosperm soft tissues in the
479 shallower samples, closely matching with willow (*Salix*) tissues measured by Winterfeld et al.
480 (2015a). With increasing water depths, angiosperm wood became the most important source
481 material, while gymnosperm wood, grasses and mosses did not appear to contribute
482 significantly to the overall lignin phenol fingerprint (Fig. 5C). As discussed earlier, this trend
483 may well be a result of preferential degradation and sorting during cross-shelf transport and
484 not derive from actual changes in source material.

485

486 3.3 Degradation status of organic matter

487 During degradation, syringyl and vanillyl phenol aldehydes are oxidized to carboxylic acids of
488 the same phenol group. Increasing Sd/SI and Vd/VI ratios can therefore qualitatively indicate
489 ongoing degradation of lignin phenols (Ertel and Hedges, 1984; Hedges et al., 1988). For
490 fresh plant material typical acid-to-aldehyde ratios are around 0.1-0.2 (Hedges et al., 1988).
491 Winterfeld et al. (2015a), however, found values as high as Sd/SI = 0.80 and Vd/VI = 0.67 for
492 a moss species (*Aulacomnium turgidum*), Sd/SI = 0.87 for larch (*Larix*) needles and Sd/SI =
493 0.49 Vd/VI = 0.41 for wild rosemary (*Ledum palustre*). Sedges (*Carex spp.*), dwarf birch
494 (*Betula nana*) and willow (*Salix*) range between Sd/SI = 0.13-0.24 and Vd/VI = 0.18-0.23.
495 The ratio of CuO oxidation-derived 3,5-dihydroxybenzoic acid to vanillyl phenols (3,5-Bd/V)
496 also serves as a proxy for degradation as 3,5-Bd is formed during humification likely
497 occurring in soils (Gordon and Goñi, 2004; Hedges et al., 1988; Prah et al., 1994; Tesi et al.,
498 2014). For this reason, this proxy can trace mineral rich soil organic matter in contrast to
499 vascular plant debris (e.g. Dickens et al., 2007; Prah et al., 1994) as well as degradation
500 during cross shelf transport (Tesi et al., 2016).



Sd/SI, Vd/VI and 3,5-Bd/V all increased along the transect, implying more degraded material with increasing residence time in the shelf system. There appeared to be no differences between outer shelf/slope and rise, which may indicate that TerrOC on the slope is already highly reworked. In contrast, Tesi et al. (2014) found no correlation between Sd/SI or Vd/VI and distance from the coast, while 3,5-Bd/V significantly increased with increasing distance from the coast. Sd/SI values for the Buor-Khaya Bay from Winterfeld et al. (2015a) were slightly higher (1.04 ± 0.24) than samples from the corresponding water depths in this study (0.66 ± 0.15), whereas Vd/VI values were significantly higher (1.28 ± 0.30 compared to 0.59 ± 0.14). Measurements for the Mackenzie Shelf agreed with the ones in this study (Sd/SI = 0.81 ± 0.25 compared to 1.01 ± 0.33 for the corresponding water depths; Vd/VI = 0.69 ± 0.14 to 0.86 ± 0.26 ; 3,5-Bd/V = 0.19 ± 0.04 to 0.31 ± 0.15), but did not show a trend with water depth (Goñi et al., 2000).

Tesi et al. (2016) found no difference in acid/aldehyde ratios between different density, grain size or settling velocity fractions of surface sediments from three (shorter) transects on the ESAS; it can thus be assumed that these proxies are not affected by hydrodynamic sorting during transport. Degradation-caused changes were limited to the lignin-rich low-density fraction, where Vd/VI and Sd/SI increased with increasing distance from the coast. Bulk 3,5-Bd/V values are potentially affected by both sorting and degradation, as they increased with decreasing particle size (fine and ultrafine fractions had the most degraded signal and are preferentially transported to the outer shelf) and across the shelf in each of the fractions.

The carbon preference indices for HMW *n*-alkanes and HMW *n*-alkanoic acids have also been widely applied as degradation proxies for plant waxes in marine sediments (for the ESAS, e.g. van Dongen et al., 2008; Fahl and Stein, 1997; Fernandes and Sicre, 2000; Vonk et al., 2010). It measures the ratio of odd-to-even numbers of carbon chain-lengths of HMW lipids and is based on the preference of odd carbon chain-lengths for HMW *n*-alkanes in fresh plant material (even carbon chain-lengths for HMW *n*-alkanoic acids; Eglinton and Hamilton, 1967). With ongoing degradation this preference is lost and the CPI approaches 1 (Bray and Evans, 1961).



We observed that the HMW *n*-alkane CPI presented the same pattern as the lignin phenol based degradation indices, however, the HMW *n*-alkanoic acid CPI did not show as much of a degradation trend (HMW *n*-alkane CPI: ~5.7 close to the coast, ~2.2 for the deep stations; HMW *n*-alkanoic acids: ~5.4 close to the coast, ~4.1 for the deep stations). Karlsson et al. (2011) measured lipid CPIs in the Buor-Khaya Bay with 10-80 km distance to the coast and obtained similar results to this ~800 km cross-shelf study, with higher values closer to the river delta. Their data appears to have a wider spread, though, which might be due to either the narrower dynamics range, or a different definition of high- molecular weight: in this study, carbon chain lengths of ≥ 23 for *n*-alkanes and ≥ 24 for *n*-alkanoic acids were defined as HMW, whereas Karlsson et al. (2011) used ≥ 21 for both compound classes. Fahl and Stein (1997) also reported a large range of *n*-alkane CPI values (< 0.2 - > 5) for Laptev Sea sediments. Fernandes and Sicre (2000) analyzed sediments from the Kara Sea and from the major rivers discharging into this sea, Ob and Yenisey rivers. In the marine environment and the Ob River, they observed HMW *n*-alkane CPI values between 4.8 and 5.3, similar to those found at shallow water depths in this study. For the Yenisey River and mixing zone, they found higher CPI values, pointing to fresher material being transported there. Vonk et al. (2010) recorded HMW *n*-alkane CPI values for sediments along the East Siberian Sea Kolyma paleoriver transect (across the East Siberian Sea) shelf that decreased from > 7.5 to < 4.0 with increasing distance from the river mouth, overall higher than in this study but confirming the general trend to more degraded material on the outer shelf. Tesi et al. (2016) found HMW *n*-alkanoic acid CPI values to decrease with decreasing particle size with no significant trends across the shelf in all but the low-density fraction, which is largely retained close to the shore. The HMW *n*-alkane CPI values in that study, however, showed no systematical differences between different fractions, but an overall decreasing trend with increasing distance from the coast.

When undergoing degradation, HMW *n*-alkanoic acids may also lose their functional groups, turning them into HMW *n*-alkanes (Meyers and Ishiwatari, 1993). The slightly decreasing ratio of HMW *n*-alkanoic acids to *n*-alkanes also hints at more degraded material with



557 increasing water depth, although, due to a rather large variability, this trend is not significant.
558 For the Buor-Khaya Bay surface sediments Karlsson et al. (2011) obtained similar results
559 (0.48-10.7, here 1.1-10.9) with higher values closer to the river delta. Along the Kolyma
560 paleoriver transect, Vonk et al. (2010) measured HMW *n*-alkanoic acid to *n*-alkane ratios
561 between 1 and 6 with no clear trend with increasing distance from the river mouth. Tesi et al.
562 (2016) found decreasing values with increasing distance from the coast with no differences
563 between the fractions. Two sediment cores from inner and outer East Siberian Sea recording
564 about one century of sedimentation showed no clear trend in CPI or HMW *n*-alkanoic acid/*n*-
565 alkane towards more degraded TerrOC with increasing sediment depth (Bröder et al., 2016),
566 but displayed a similar difference between inner and outer shelf as seen in this study. This
567 contrasting behavior for cross-shelf and down-core trends may be caused by significantly
568 different timescales for the two processes: about one century in situ/after burial compared to
569 potentially several millennia long lateral transport. Furthermore, the degradation efficiency is
570 possibly higher under the oxic conditions prevailing during cross-shelf lateral transport (Keil
571 et al., 2004), than in the anoxic conditions that predominate below a few millimeters of
572 sediments on the ESAS (e.g. Boetius and Damm, 1998). Comparing in situ to transport-
573 related oxygen exposure times on the wide Arctic shelves could potentially resolve the
574 observed discrepancies.



4 Concluding remarks and future research directions

Across the Laptev Sea from the Lena River mouth to the deep sea of the Arctic interior a considerable loss of terrigenous organic matter has been observed on both bulk and molecular level. All terrigenous biomarkers display a massive decline with increasing water depth along this high-resolution transect due to hydrodynamic sorting and degradation during transport. Terrigenous organic matter (TerrOC) seems to be also qualitatively more degraded on the outer shelf, slope and rise compared to inner shelf and coastal areas. These results corroborate and expand previous findings for the East Siberian Arctic Shelf, showing that the shelf seas in this region function as an active reactor for TerrOC. This stands in sharp contrast to e.g. the Mackenzie basin, which is thought to act as a geological sink for organic carbon due to its efficient TerrOC burial (Hilton et al., 2015). For narrower Arctic shelves in general, where transport times can be expected to be much shorter, organic matter transfer towards the deeper basins appears to be much more efficient, with high TerrOC concentrations in surface sediments even at greater water depths (e.g. Barrow Canyon, Goni et al., 2013). It can therefore be assumed that the cross-shelf transport time exerts first-order control over the extent of TerrOC degradation. With ongoing global warming, rising permafrost-derived organic carbon input from river-sediment discharge and coastal erosion is expected to reach the marine environment. It is therefore crucial to better constrain cross-shelf transport times in order to determine a TerrOC degradation rate and thereby contribute to quantifying potential carbon-climate feedbacks.

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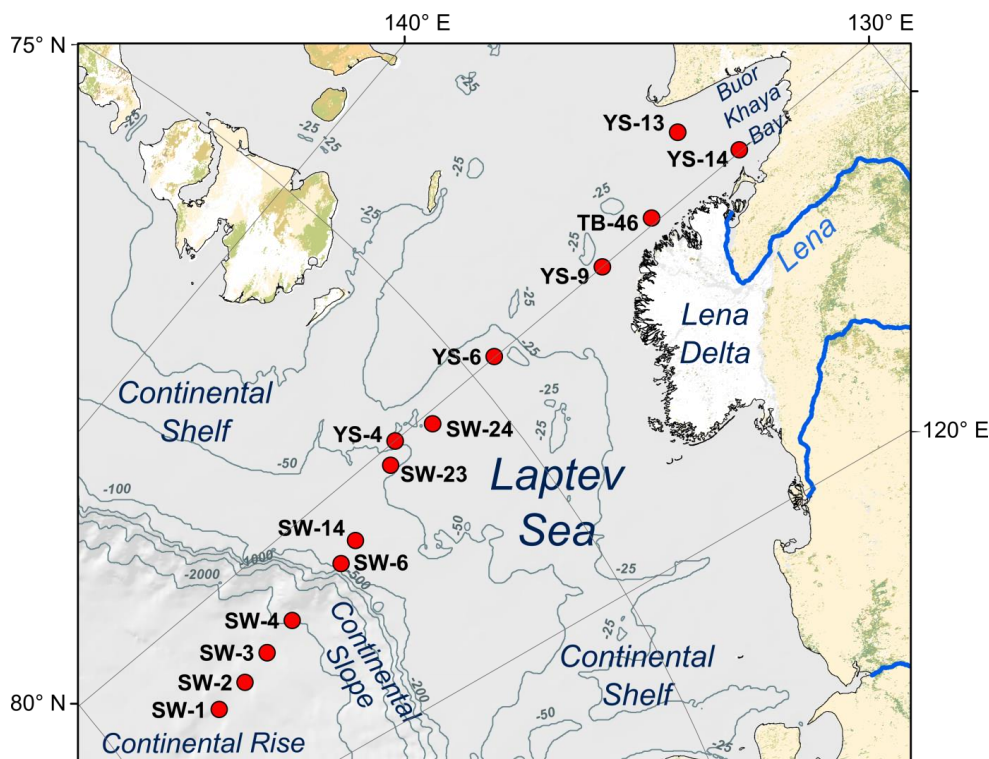
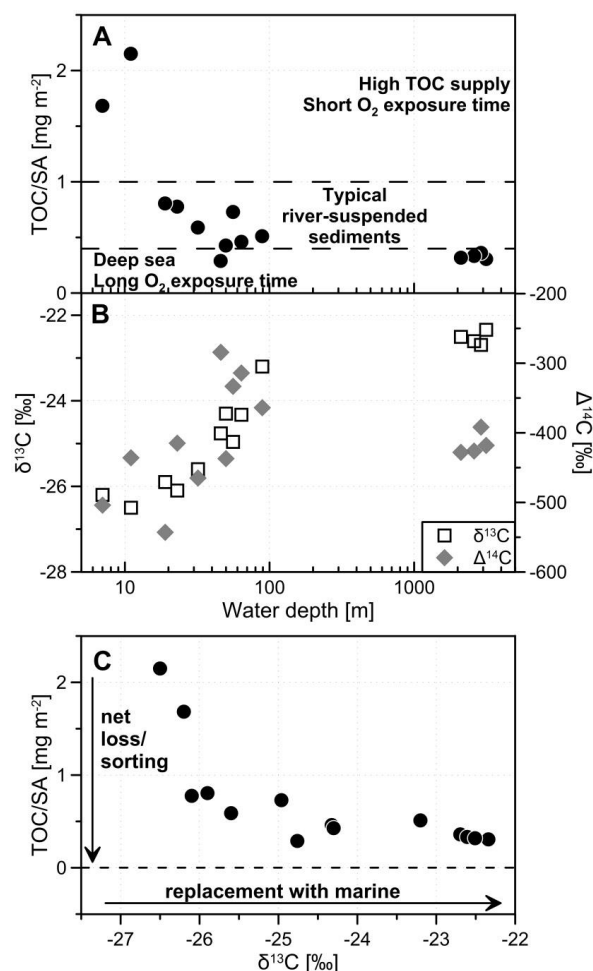
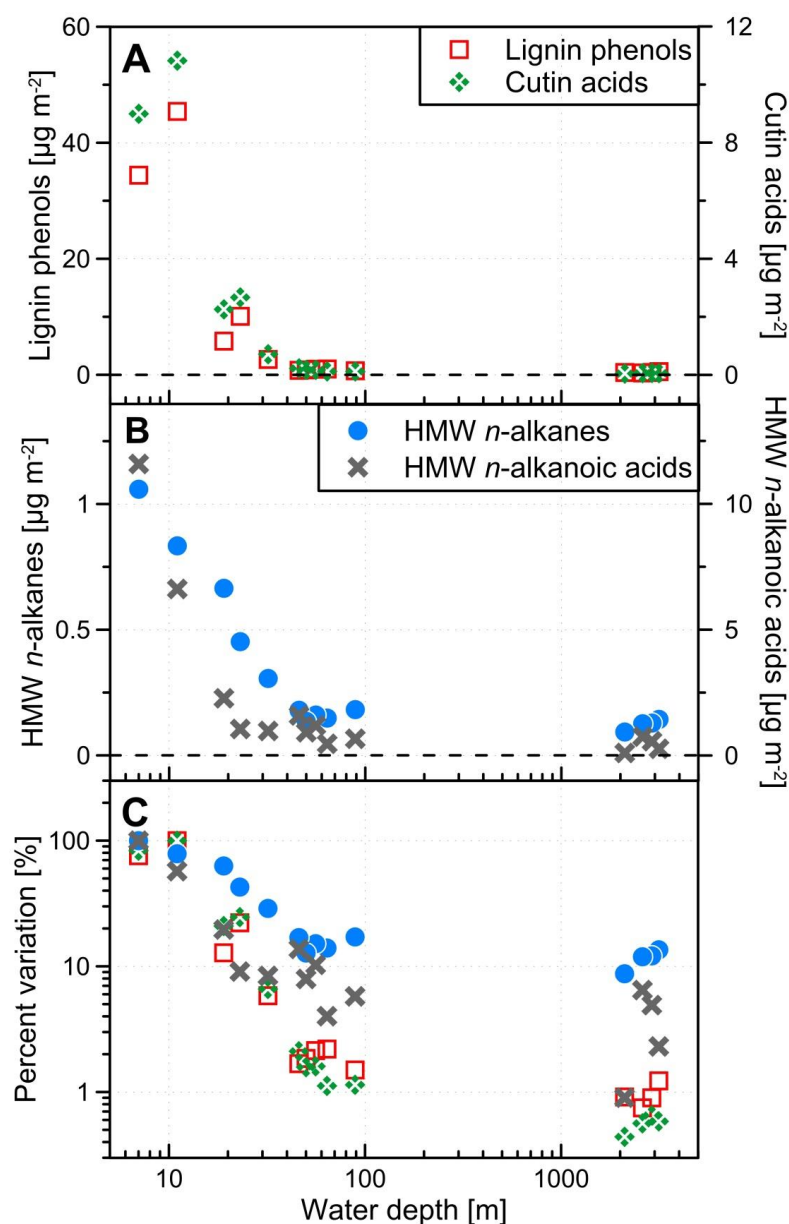


Figure 1: Map of the study area in the Laptev Sea. Red filled circles mark the sediment sampling sites. The transect reaches from the Lena River mouth and the Buor-Khaya Bay (water depths ~10 m) across the Laptev Sea Shelf (mean depth ~50 m) to the slope/shelf break and rise (water depths ~3000 m).

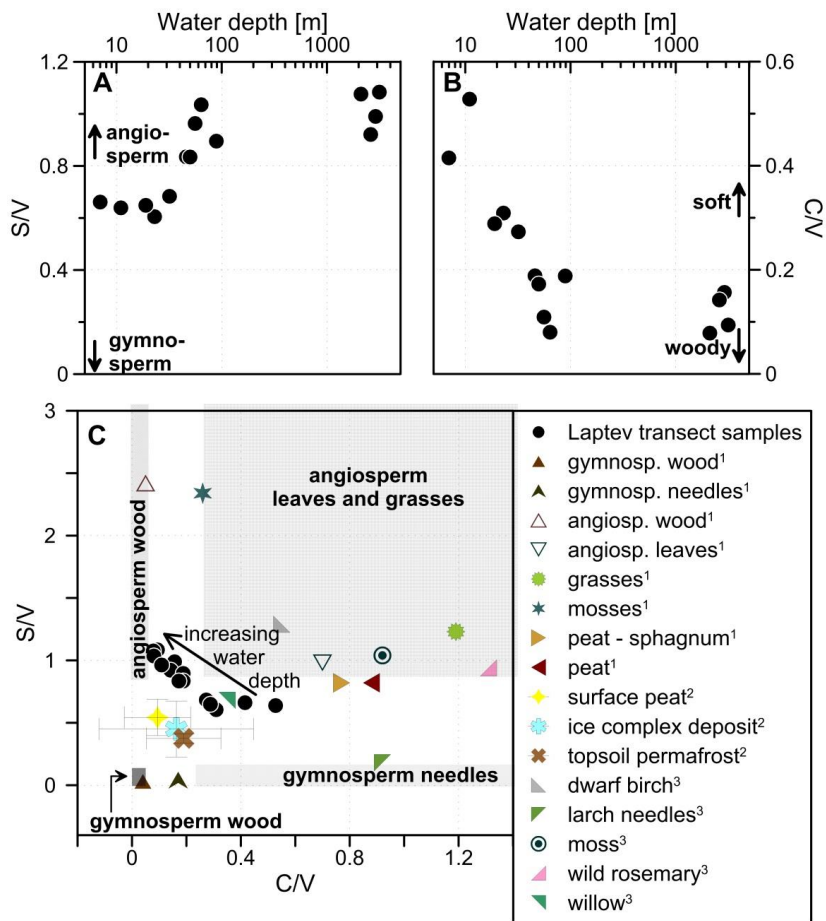


898
 899 Figure 2: (A) The ratio of total organic carbon (TOC) to surface area (SA) decreases with
 900 increasing water depth by a factor of 7 or 86%. (B) The stable carbon isotopic signal (δ¹³C)
 901 increases with increasing water depth from -26 ‰ to -22 ‰, suggesting a change in source
 902 material from terrigenous to marine dominated. The radiocarbon isotopic signal (Δ¹⁴C)
 903 increases toward the outer shelf, supporting an increase in fresh marine organic carbon. The
 904 slope sediments show an older (more depleted) Δ¹⁴C signal, possibly due to ageing during
 905 transport and in situ. (C) The relationship between TOC/SA and δ¹³C can help to disentangle
 906 two processes occurring simultaneously during cross-shelf transport: The net loss (i.e.
 907 degradation) or sorting (i.e. hydraulically retaining) of TerrOC leads to a shift towards lower
 908 TOC/SA ratios, whereas the replacement/dilution with marine OC shifts the isotopic signature
 909 towards higher values.



910

911 Figure 3: Terrigenous biomarker loadings display a strong decrease with increasing water
 912 depth: (A) for lignin phenols and cutin acids by a factor of 130 and 230 respectively, (B) for
 913 HMW *n*-alkanes and HMW *n*-alkanoic acids by a factor of 12 and 44, respectively. (C)
 914 Comparison between the different biomarkers along the transect: lignin phenols, cutin acids,
 915 HMW *n*-alkanoic acids and *n*-alkanes where each is normalized to respective highest value
 916 (corresponding to 100 %).



917

918 Figure 4: The lignin phenol composition carries source information: (A) an increasing ratio of
 919 syringyl to vanillyl phenols (S/V) suggests relatively more angiosperm material on the outer
 920 shelf/slope possibly due to selective degradation or sorting during transport. (B) The ratio of
 921 cinnamyl to vanillyl phenols (C/V) decreases with increasing water depths, which implies an
 922 increasing relative contribution of woody material compared to soft tissues. (C) Comparison
 923 of S/V and C/V with the end-members for different Arctic plants as compiled from different
 924 studies by Amon et al. (2012, and citations therein, here marked with ¹); ice-complex deposit
 925 and topsoil permafrost as determined by Tesi et al. (2014, here marked with ²) and more
 926 plant species measured by Winterfeld et al. (2015a, here marked with ³). The boxes indicate
 927 typical ranges of S/V and C/V for different vascular plant tissues in different locations (e.g.
 928 Goñi et al., 2000). The surface sediment lignin phenol compositions along the Laptev
 929 transect appear to be a mix of angio- and gymnosperm soft tissues most similar to willow and
 930 shift towards angiosperm wood with increasing water depth.

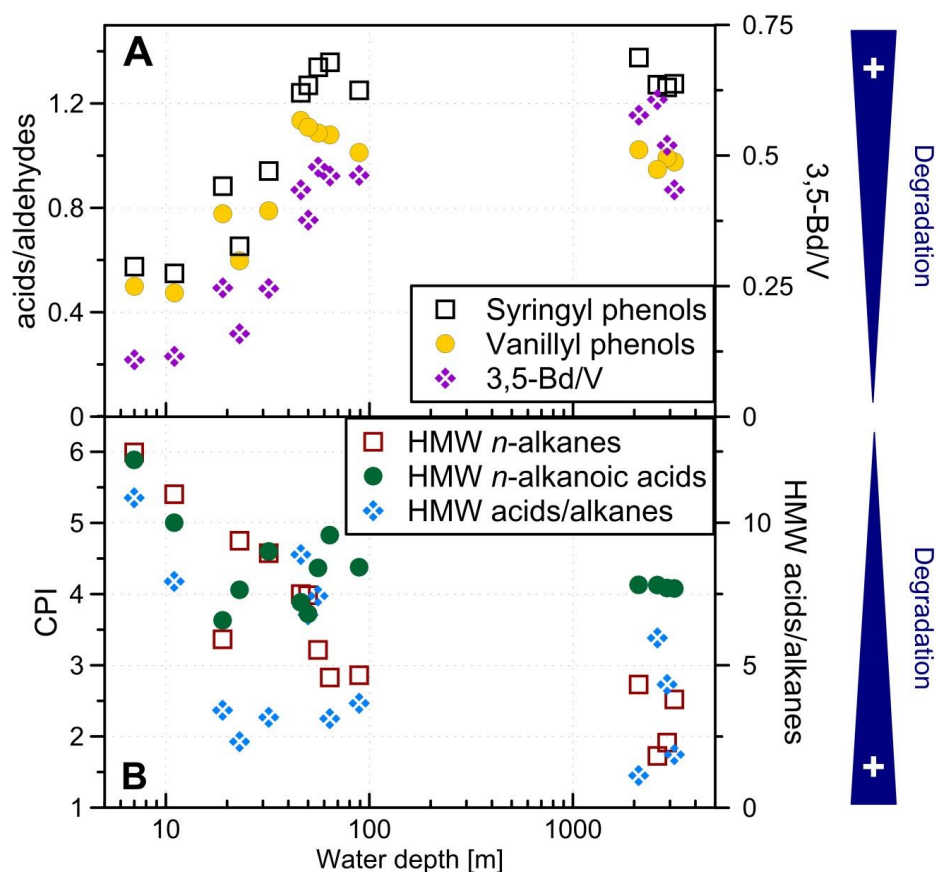


Figure 5: Degradation proxies for TerrOC, blue triangles point toward lower extent of degradation: (A) CuO-oxidation derived ratios Sd/SI, Vd/VI and 3,5-Bd/V all display a trend toward more degraded material with increasing distance from the shore with no difference between outer shelf and slope/rise sediments. (B) The carbon preference indices (CPI) of HMW *n*-alkanes and *n*-alkanoic acids show the same trend, yet not as pronounced for the *n*-alkanoic acids. The ratio of HMW *n*-alkanoic acids to HMW *n*-alkanes has a wider scatter, but also hints at more degraded material with increasing water depth.



940 **Table 1: List of surface sediment samples from the Laptev Sea transect.**

ID	Sample type	Lat ° N	Long ° E	Water depth m	OC mg·g ⁻¹	SA m ² ·g ⁻¹	δ ¹³ C ‰	Δ ¹⁴ C ‰	SiO ₂ wt %	Al ₂ O ₃ wt %	CaO wt %
SW-1	0-0.5cm	78.942	125.243	3146	10.4	34.0	-22.34	-418	60.3	16.5	2.4
SW-2	0-0.5cm	78.581	125.607	2900	13.8	38.3	-22.70	-392	57.8	17.2	2.1
SW-3	0-0.5cm	78.238	126.150	2601	10.6	31.8	-22.61	-426	62.1	16.0	1.6
SW-4	0-0.5cm	77.855	126.664	2106	13.2	41.5	-22.51	-428	56.6	17.5	1.3
SW-6	0-1cm	77.142	127.378	89	7.6	14.9	-23.20	-364	72.0	12.6	1.7
SW-14	0-1cm	76.894	127.798	64	8.9	19.4	-24.33	-314	71.3	12.5	1.5
SW-23	0-1cm	76.171	129.333	56	15.8	21.7	-24.96	-333	68.9	13.6	1.4
YS-4	0-1cm	75.987	129.984	50	13.4 ^a	31.4	-24.76 ^a	-284 ^a	63.8	15.1	1.3
SW-24	0-1cm	75.599	129.558	46	10.7	37.0	-24.30	-437	62.5	15.4	1.2
YS-6	0-1cm	74.724	130.016	32	18.6 ^a	31.6	-25.60 ^a	-465 ^a	62.1	16.1	1.3
YS-9	Grab	73.366	129.997	23	13.1 ^b	16.9	-26.10 ^b	-415 ^b	70.8	14.0	1.3
YS-13	0-1cm	71.968	131.701	19	18.9 ^a	23.5	-25.90 ^a	-543 ^a	61.6	17.4	0.8
YS-14	0-1cm	71.630	130.050	7	19.1 ^a	11.4	-26.20 ^a	-504 ^a	69.6	15.0	1.6
TB-46	Grab	72.700	130.180	11	25.8 ^a	12.0 ^c	-26.50 ^a	-436 ^a	67.9	15.2	1.8

941

942 ^a Data from Vonk et al. (2012); ^b data from Tesi et al. (2016); ^c data from Karlsson et al.
 943 (2014).



944 **Table 2: Biomarker results for surface sediment samples from the Laptev Sea**
 945 **transect.**

ID	Lignin $\mu\text{g m}^{-2}$	Cutin $\mu\text{g m}^{-2}$	HMW* alkanes $\mu\text{g m}^{-2}$	HMW** acids $\mu\text{g m}^{-2}$	S/V	C/V	Sd/SI	Vd/VI	3,5Bd/V	CPI alk	CPI acids/ alk	acids/ alk
SW-1	0.56	0.063	0.14	0.27	1.1	0.09	1.3	0.98	0.43	2.5	4.1	1.9
SW-2	0.41	0.070	0.13	0.57	0.99	0.16	1.3	0.99	0.52	1.9	4.1	4.3
SW-3	0.34	0.061	0.13	0.75	0.92	0.14	1.3	0.95	0.61	1.7	4.1	6.0
SW-4	0.42	0.048	0.093	0.10	1.1	0.08	1.4	1.0	0.58	2.7	4.1	1.1
SW-6	0.68	0.12	0.18	0.67	0.90	0.19	1.2	1.0	0.46	2.9	4.4	3.7
SW-14	1.0	0.12	0.15	0.46	1.0	0.08	1.4	1.1	0.46	2.8	4.8	3.1
SW-23	0.97	0.17	0.16	1.2	0.96	0.11	1.3	1.1	0.48	3.2	4.4	7.4
YS-4	0.84	0.17	0.13	0.92	0.83	0.17	1.3	1.1	0.38	4.0	3.7	6.8
SW-24	0.76	0.23	0.18	1.6	0.84	0.19	1.2	1.1	0.43	4.0	3.9	8.9
YS-6	2.7	0.71	0.31	0.97	0.68	0.27	0.94	0.79	0.25	4.6	4.6	3.2
YS-9	10	2.7	0.45	1.1	0.60	0.31	0.65	0.60	0.16	4.7	4.1	2.3
YS-13	5.8	2.3	0.64	2.3	0.65	0.29	0.88	0.78	0.25	3.4	3.6	3.4
YS-14	34	9.0	1.1	12	0.66	0.42	0.57	0.50	0.11	6.0	5.9	11
TB-46	45	11	0.83 ^d	6.6 ^d	0.64	0.53	0.55	0.47	0.12	5.4 ^d	5.0 ^d	7.9 ^d

946

947 * carbon chain-lengths 23-34; ** carbon chain-lengths 24-30.

948 ^d recalculated data from Karlsson et al. (2011).