

**bg-2016-230**

**The metabolic response of thecosome pteropods from the North Atlantic and North Pacific Oceans to high CO<sub>2</sub> and low O<sub>2</sub>**

A. E. Maas, G. L. Lawson, and Z. A. Wang

**We would like to thank the referee for their appreciation of our approach and their helpful comments for improving the clarity of the text. We have responded to each suggestion on a topically grouped point by point basis with referee comments in plain text and author response in bold.**

- Section 2.4: How many organisms were incubated per respiration chamber

**Each chamber contained one individual pteropod. The text has been modified to read:**

**“Post-gut-clearance, healthy organisms were put into separate glass syringe respiration chambers, one individual per chamber, with a known volume of 0.2 µm filtered seawater and 25 mg L<sup>-1</sup> each of streptomycin and ampicillin.”**

how many replicates were possible to set up? Is N in table 4 the number of replicates? Please clarify.

- Is N consistent with number of replicates?

**The number of replicates was different for each species, treatment and region. The N in table 4 (previously labeled as the number of individuals) is indeed the number of replicates. In the case of this experimental design individuals and replicates are synonymous. The word ‘individuals’ has been replaced with ‘replicates’ for clarity.**

The discussion is in general ok, but I miss some consideration relating to the specific life stage(s) these experiments were done on. Was it all the same life stages? If not, could that affect results and conclusions?

Of what stage were the pteropods (all adults?)? If different stages, how could that affect results and conclusions drawn?

Were all individuals of the same developmental stage?

**To address this point, text is now included in the methods, results, discussion and conclusion specifically stating that the work was done only with adult life stages.**

**“Species were targeted specifically for their abundance and the likelihood of their presence in both ocean basins and only adult individuals were used.”**

**“Only relatively large adult specimens were used in respiration trials, in part to avoid any confounding effects of ontogeny and in part to ensure a measurable change in oxygen levels.”**

**We agree with the reviewer that juveniles of all the species we studied could potentially be more sensitive to the tested conditions. There is experimental evidence from our group that the veliger stage of *L. retroversa* is more susceptible to CO<sub>2</sub> (Thabet et al. 2015) and clearly as projections are made about population-level responses to changing environmental conditions the sensitivities of all life stages must be considered. The vertical distribution of pteropod juveniles has not been well documented (in part because good morphological keys to that stage do not exist), but it would be interesting to determine whether they are absent from depths where high CO<sub>2</sub> / low O<sub>2</sub> is present naturally. Interestingly many diel vertically migratory species exhibit different vertical distribution throughout ontogeny in relation to midwater regions of high CO<sub>2</sub> / low O<sub>2</sub> (i.e. Wishner et al. 2013; Maas et al. 2014). It seems logical that juvenile pteropods would avoid regions of high CO<sub>2</sub> during initial shell formation as this is a period of sensitivity as has been shown in some pteropods (Thabet et al. 2015) and in other species (Kurihara 2008; Kroeker et al. 2013; Waldbusser et al. 2015).**

Since the text only focuses on adults we have chosen not to discuss this thoroughly; we did, however, reiterate our focus on the adult life stage at the start of the discussion:

**“This study reveals that short term exposure to low O<sub>2</sub> and high CO<sub>2</sub>, similar to what would be experienced by individuals in the Pacific during diel vertical migration, does not influence the oxygen consumption of adult individuals of most of the thecosome pteropod species examined from either the Atlantic or Pacific.”**

And made mention the importance of the consideration of the various sensitivities of different life stages as:

**“Furthermore, although adult individuals may show no change in metabolic rate, there is evidence that juvenile stages of many calcifying species are typically more sensitive to CO<sub>2</sub> exposure (i.e. Connell et al. 2013; Waldbusser et al. 2015) and emerging evidence supports the idea that eggs, veligers and juveniles of *L. retroversa* and *L. helicina* are more vulnerable to acidification than adults (Lischka et al. 2011; Thabet et al. 2015; Manno et al. 2016). Thus, although adults may be capable of surviving short term exposure, as acidity in surface waters increase there may be population level stress due to ontogenetic sensitivity.”**

Please give also size ranges of different species incubated.

- L319: Size ranges of all species would be really helpful to see!

- Section 3.1: Please include information on the size ranges of the different pteropod species.

Table 4: - Please include a size range for all different species

**The size range (average mass and standard error) for the pteropods is already reported in Table 4, and it seems unnecessary to report the range (smallest to largest). If the reviewer instead means size in length, the information is not available for some of the samples which have been used for other purposes. Furthermore, the species documented here vary substantially in shape and have different growth patterns, making it impossible to report a standard size (length/diameter, width). It seems that if length is what the reviewer refers to, this is tied into the previous concern about life stage and we hope that the changes made to the text are sufficient to allay the reviewers' concerns.**

- Section 2.6: Assumptions proved in case of significant results found for *L. retroversa*?

**We assume that the reviewer refers to the assumptions of normality and heterogeneity of variance – please correct if otherwise. These tests were performed and *L. retroversa* met all assumptions. Text has been added on line 370 as follows:**

**“The datasets were tested for normality and homoscedasticity.” Information about the statistical assumptions has been added to Table 5.**

- Section 3.2: This section needs some clarification with respects mentioning of geographical positions, temperatures. . . in the text and the respective table.

- L409/410: Can not find geographical position in table 2?

- L415: geographical position not found in Table 2?

- L432/433: 400 or 385 m? geographical position can not be found in Table 1.

**Text has been added to clarify the geographical positions of each station in the text (latitude, longitude and station number are now specifically described). The range reported in the text describes collectively all of the stations found in the portion of the study region, and hence is a summary of numbers found in Table 2 rather than re-stating exact values from that Table. In addition to the above changes, text has been added to the legend in table 2 to help point out the geographical locations of the hydrographic regimes. Line 1024:**

**“Each basin was characterized by multiple hydrographic regimes (see text and Fig 2); transitions between regimes are denoted by dashed horizontal lines.”**

- L411: 250 or 209 m?

**The sentence was intended to indicate that at all stations in this hydrographic regime, the oxygen fell below 130 within the range of the organisms. To better clarify this point the text has been changed to (Line 420-421):**

**“At these stations O<sub>2</sub> fell below 10% (~130 μmol kg<sup>-1</sup>) at depths less than ~250 m”**

- L412: 110 or 130 m?

**It was between 108.3-131 m. To clarify the sentence was changed to (Line 421-422):**

**“At these stations in the northern part of the transect, pH fell below 7.7 at depths less than 130 m,...”**

- L417: 10–17\_C?

**The change has been made.**

- L421–425: How can I see that *Clio pyramidata* experienced these conditions (in Table1)?

**Table 1 reports that *Clio pyramidata* is a known vertical migrator with a typical range of 0-500 m, and can be found as deep as 1500 m. Based on this information, we would expect that its range similarly extends 500 m in the Pacific, which would put it into conditions of 10% O<sub>2</sub>, 800 μatm pCO<sub>2</sub> and aragonite undersaturation in the Pacific as per the values documented in Table 2. To clarify we have referenced Table 2 in the sentence (which as a note has been moved to the discussion as per the request of reviewer 2).**

- L427: 200 μmol kg<sup>-1</sup> corresponds to what % air saturation O<sub>2</sub>?

The information has been added to Line 436-438 as:

**“In contrast to the Pacific, along the entire Atlantic transect O<sub>2</sub> concentration was above ~200 μmol kg<sup>-1</sup> (~15%) in the top 500 m, while pCO<sub>2</sub> never reached 800 μatm and aragonite undersaturation never occurred throughout the top 1000 m.”**

- L428/429: Would it be possible to indicate the dominant hydrographic regimes in

Fig. 1, would be helpful in connection with the sampling location of different pteropod species.

**An attempt was made to modify the figure, but there was no way to show the regimes clearly on what is already a busy figure. We hope that reviewer agrees that with the new additions to table 2 and the text there is now enough information to make this clear.**

- L437: below 5\_C? I calculated 5.2\_C?

**The sentence was intended to indicate that at all stations in this hydrographic regime, temperatures within the 25-100 m all reached below 5 C. The sentence has been modified as:**

**“Stations conducted in this water were typified by a temperature and salinity anomaly with temperatures falling below 5°C from 25-100 m and a salinity signature < 33, contrasting significantly with the surface salinities of the northern portion (~34) and southern portion (~36) of the Atlantic transect.**

- Could salinity be included in Table 2?

**Salinity has been added to Table 2 and the table caption has been modified to reflect the change.**

- L478/480: This sentence is unclear. According to Table 3, ar was never below 1? And 1.2 is not under-saturated? Please clarify!

The point of the sentence was that omega reached a minimum of 1.2 and hence *approached*, but didn't reach, under-saturation. To clarify the sentence has been changed as follows:

**"The experimental conditions of the high CO<sub>2</sub> treatments reached their lowest value in the middle part of the transect ( $\Omega_{Ar} = 1.2$  at mid-latitudes; Table 3), where cold northern waters of low salinity were encountered. Experimental  $\Omega_{Ar}$  had a range of 1.5-2.0 for the rest of the transect in the Atlantic."**

- L482: In situ values are meant here, right? Maybe indicate in the text, easier for the reader.

**We meant the experimental and have now indicated this in line 492-494 as:**

**"The values of experimental  $\Omega_{Ar}$  were lower overall in the Pacific, although the high CO<sub>2</sub> treatments also never reached under-saturation ( $\Omega_{Ar}$  1.3-1.8)."**

- Section 3.4: As indicated earlier, please clarify how many replicates were measured and how many individuals were incubated per chamber and experiment and species.

**The word 'individuals' has been replaced with 'replicates' in table 4 for clarity. We do not think it is appropriate to re-state that the method was to place single individuals in a chamber in the results, and hope that the reviewer agrees that the clarification in the methods and table are sufficient to clarify the point.**

- L526: Fig 4 not 4A

**The correction has been made**

- L552/553: What stage of *L. helicina* was it? Could the high mortality also be associated with life cycle issues and less so with temperature, i.e. die off after reproduction?

**It is possible, but based on our previous experiences with *Limacina spp.* these organisms are capable of laying eggs throughout their later adult life and typically don't die due to spawning (Maas pers. obs.; Thabet et al. 2015). It is true that they do eventually reach their largest size class and then die. An analysis of the available size class data of all the individuals does not support the idea that the older or younger (larger or smaller respectively – pteropods grow continuously throughout their lives) were differentially susceptible. Live individuals ranged in mass from 0.5-10.4 mg, while dead individuals were 0.9-5 mg. We did not measure individuals from the high mortality events, however, so size may have played a role? In any case, a sentence about alternative hypotheses has been added (Line 569-571):**

**"Alternative hypotheses are that these were population reaching senescence, or that they were collected in a hydrographic regime with low food availability."**

- L617: How likely is it that O<sub>2</sub> saturation below 10% resulted in a substantial difference compared to the results obtained at 10% O<sub>2</sub> saturation? In other words, any idea where a critical threshold level could lie?

**The lower than 10% O<sub>2</sub> saturation that was documented in the wild was never explicitly tested in our experiments. Our lowest experiment only ever reached ~8% oxygen over the course of the respiration. Since these were end point measurements there is no way to determine the P<sub>crit</sub>. Based on previous observations we would hypothesize that the Pacific populations have a lower P<sub>crit</sub> than the Atlantic. Importantly some of these species in the Eastern Tropical Pacific have been shown to survive and respire at 1% O<sub>2</sub> (Maas et al. 2012). An explicit study of differences in P<sub>crit</sub> between the ocean basins using the more modern optical spot sensors (such as in (Kiko et al. 2016; Maas et al. 2016), would be informative and productive.**

In general, please next time indicate numbers of figures and tables directly on the page where figures and tables are shown. The way they are presented here led to a continuous turning around of printed pages and searching for a particular one.

**We apologize that the reviewer found the lack of labels on the figures and tables confusing and will be sure to take this into consideration during our next submission.**

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A. E. Maas, G. L. Lawson, and Z. A. Wang

**We would like to thank the referee for their helpful feedback on this, and a previous version, of this manuscript. We have responded to each suggestion on a point by point basis with referee comments in plain text and author response in bold.**

Comments:

Abstract: The first sentence is a bit confusing. It reads like the burning of fossil fuels directly cause a decrease in O<sub>2</sub>.

**The sentence has been modified to:**

**“As anthropogenic activities directly and indirectly increase carbon dioxide (CO<sub>2</sub>) and decrease oxygen (O<sub>2</sub>) concentrations in the ocean system, it becomes important to understand how different populations of marine animals will respond.”**

L72: This sentence is misleading and tends to indicate that with OA all oceans will be undersaturated.

**The sentence has been modified to:**

**“In some regions, as ocean acidification continues, the water becomes undersaturated and corrosive, meaning that, in the absence of compensating biological action, conditions will favor the dissolution of the CaCO<sub>3</sub> found in the shells and skeletons of ...”**

L79: Probably the change in saturation state is not the only driver (DIC/proton ratios, pH itself, ..). I would replace this by “modifications of the carbonate chemistry”.

**Good point – the sentence has been modified to:**

**“Perturbations of seawater carbonate chemistry can also affect the ability of some calcifying animals...”**

L101: It would be good to compare this value with what is found in the other oceans.

**The sentence has been modified to provide context for the value as:**

**“On top of this natural process, ocean acidification also plays a role: the pH of the upper water column in the North Pacific is decreasing by ~0.002 pH units per year (Byrne et al. 2010; Chu et al. 2016), similar to the global average of 0.0022 pH units per year (Williams et al. 2015). Such a change corresponds to a total CO<sub>2</sub>, or dissolved inorganic carbon (DIC), increase of 1–2 μmol kg<sup>-1</sup> yr<sup>-1</sup> (Peng et al. 2003; Sabine et al. 2008; Sabine and Tanhua 2010; Chu et al. 2016).”**

L146-148: You mention it in the discussion, but it could be interesting to indicate here that they are potentially different species.

**Text has been added as:**

**“The taxonomy of thecosomes has recently begun to be revisited using molecular and paleontological tools (i.e. Hunt et al. 2010; Jennings et al. 2010; Janssen 2012; Maas et al. 2013) and there is growing evidence of cryptic speciation for some pteropod groups (Gasca and Janssen 2014; Burrige et al. 2015). It thus should be noted that the inter-basin comparisons performed here may be of cryptic congeners rather than conspecific populations. Using these organisms, which are presumably adapted to their local conditions, we can test whether species or congeners exhibit a population-specific physiological response to these environmental conditions indicative of different sensitivities.”**

L-226: Add "Surface" before "carbonate chemistry".

**The change has been made.**

L340: The effect is probably minor, but pteropod calcification and excretion can change the TA.

**Very true. The sentence has been modified to reflect the uncertainty that is contributed by these processes, but with the assumption that on the timescale of the experiments the influence would be minor:**

**"TA of experimental water was assumed to have been constant over the course of each experiment as water was filtered (0.2  $\mu\text{m}$ ) and antibiotic treated (thus microbial activities were kept at minimum). Although pteropod aerobic respiration, excretion, and calcification within a respiration chamber could influence TA, these are presumed to have not had a significant influence over the time scales in question."**

L350: Could the difference in TA be due to the bubbling that caused evaporation?

**The bubbling was the same among the different batches of water, and thus it seems likely that the error in TA due to evaporation would have been consistent throughout.**

L456: Increased not decreased?

**Increased. Thanks for the catch!**

The results section contains a large part of methods and discussion. It reads well but I wonder if for clarity the methods and discussion statements should be moved to the corresponding sections.

**In previous versions of the manuscript reviewers found presentation of some of the results confusing. The methods were thus partially repeated in the results section to make sure that the reader understands how the data were collected as it is presented. This was in particular with regards to the carbonate chemistry uncertainty and error. Based on earlier drafts we feel that it is best to retain this material in the same place (results).**

**We have gone through the rest of the results, however, and removed other text that is more discussion based as per the reviewer's recommendations. Specifically, the text that mentions cryptic species has been moved to the introduction and some comments about the distribution of the species to the discussion.**

Burridge AK, Goetze E, Raes N, Huisman J, Peijnenburg KT (2015) Global biogeography and evolution of *Cuvierina* pteropods. BMC evolutionary biology 15 doi 10.1186/s12862-015-0310-8

Byrne RH, Mecking S, Feely RA, Liu X (2010) Direct observations of basin-wide acidification of the North Pacific Ocean. Geophys Res Lett 37: L02601

Chu SN, Wang ZA, Doney SC, Lawson GL, Hoering KA (2016) Changes in anthropogenic carbon storage in the Northeast Pacific in the last decade. Journal of Geophysical Research: Oceans 121 doi 10.1002/2016JC011775

Gasca R, Janssen AW (2014) Taxonomic review, molecular data and key to the species of Creseidae from the Atlantic Ocean. Journal of Molluscan Studies 80: 35-42

Hunt B, Strugnell J, Bednarsek N, Linse K, Nelson RJ, Pakhomov E, Seibel B, Steinke D, Würzberg L (2010) Poles Apart: The "Bipolar" Pteropod Species *Limacina helicina* Is Genetically Distinct Between the Arctic and Antarctic Oceans. PLoS ONE 5: e9835

Janssen AW (2012) Late Quaternary to Recent holoplanktonic Mollusca (Gastropoda) from bottom samples of the eastern Mediterranean Sea: systematics, morphology. Bollettino Malacologico 48: 1-105

- Jennings RM, Bucklin A, Ossenbrügger H, Hopcroft RR (2010) Species diversity of planktonic gastropods (Pteropoda and Heteropoda) from six ocean regions based on DNA barcode analysis. *Deep Sea Research Part II: Topical Studies in Oceanography* 57: 2199-2210
- Maas AE, Blanco-Bercial L, Lawson GL (2013) Reexamination of the species assignment of Diacavolinia pteropods using DNA barcoding. *PLoS ONE* 8: e53889 doi:10.1371/journal.pone.0053889
- Peng T-H, Wanninkhof R, Feely RA (2003) Increase of anthropogenic CO<sub>2</sub> in the Pacific Ocean over the last two decades. *Deep Sea Research Part II: Topical Studies in Oceanography* 50: 3065-3082
- Sabine CL, Feely RA, Millero FJ, Dickson AG, Langdon C, Mecking S, Greeley D (2008) Decadal changes in Pacific carbon. *J Geophys Res Oceans* 113: -
- Sabine CL, Tanhua T (2010) Estimation of anthropogenic CO<sub>2</sub> inventories in the ocean. *Annu Rev Mar Sci* 2: 175-198
- Williams NL, Feely RA, Sabine CL, Dickson AG, Swift JH, Talley LD, Russell JL (2015) Quantifying anthropogenic carbon inventory changes in the Pacific sector of the Southern Ocean. *Marine Chemistry* 174: 147-160



1 **The metabolic response of thecosome pteropods from the North**  
2 **Atlantic and North Pacific Oceans to high CO<sub>2</sub> and low O<sub>2</sub>**

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32 **Abstract.** As anthropogenic activities, ~~notably the burning of fossil fuels, directly and indirectly~~  
33 increase carbon dioxide (CO<sub>2</sub>) and ~~result in a decrease in~~ oxygen (O<sub>2</sub>) concentrations in the  
34 ocean system, it becomes important to understand how different populations of marine animals  
35 will respond. Water that is naturally ~~lower~~low in pH, with a high concentration of carbon dioxide  
36 (hypercapnia) and a low concentration of oxygen, occurs at shallow depths (200-500 m) in the  
37 North Pacific Ocean, whereas similar conditions are absent throughout the upper water column  
38 in the North Atlantic. This contrasting hydrography provides a natural experiment to explore  
39 whether differences in environment cause populations of cosmopolitan pelagic calcifiers,  
40 specifically the aragonitic-shelled pteropods, to have a different physiological response when  
41 exposed to hypercapnia and low O<sub>2</sub>. Using closed-chamber end-point respiration experiments,  
42 eight species of pteropods from the two ocean basins were exposed to high CO<sub>2</sub> (~800 μatm)  
43 while six species were also exposed to moderately low O<sub>2</sub> (10%, or ~130 μmol kg<sup>-1</sup>) and a  
44 combined treatment of low O<sub>2</sub>/high CO<sub>2</sub>. None of the species tested showed a change in  
45 metabolic rate in response to high CO<sub>2</sub> alone. Of those species tested for an effect of O<sub>2</sub>, only  
46 *Limacina retroversa* from the Atlantic showed a response to the combined treatment, resulting in  
47 a reduction in metabolic rate. Our results suggest that pteropods have mechanisms for coping  
48 with short-term CO<sub>2</sub> exposure and ~~suggest~~ that there can be interactive effects between stressors  
49 on the physiology of these open ocean organisms that correlate with natural exposure to low O<sub>2</sub>  
50 and high CO<sub>2</sub>; these are considerations that should be taken into account in projections of  
51 organismal sensitivity to future ocean conditions.

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60 **Key Words: ocean acidification, zooplankton, respiration**  
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## 62 1. Introduction

63 Ocean acidification, a result of the dissolution of anthropogenically-produced carbon dioxide  
64 (CO<sub>2</sub>) into sea water, is increasingly considered to be one of the most pervasive human changes  
65 to the marine system (Doney et al., 2009; Gruber, 2011; Halpern et al., 2008). The pH of the  
66 ocean surface has already dropped by ~0.1 units relative to preindustrial levels and is predicted  
67 to drop another 0.3-0.4 pH units in the next one hundred years (Bopp et al., 2013; Haugan and  
68 Drange, 1996; IPCC, 2013). As CO<sub>2</sub> dissolves in the ocean, it causes changes in seawater  
69 carbonate chemistry, notably increasing hydrogen ion concentration and decreasing the  
70 concentration of carbonate ions. As a consequence of the changing equilibria, there is a reduction  
71 in pH and in the saturation state of calcium carbonate (CaCO<sub>3</sub>), including the biogenic forms of  
72 calcite and aragonite. ~~As~~In some regions, as ocean acidification continues, ~~eventually~~ the water  
73 becomes undersaturated and corrosive, meaning that, in the absence of compensating biological  
74 action, conditions will favor the dissolution of the CaCO<sub>3</sub> found in the shells and skeletons of  
75 calcifying organisms, with aragonite being more sensitive than calcite (Millero, 2007).

76 Ocean acidification, therefore, impacts calcifying species on multiple fronts. Changes in  
77 environmental pH can modify the acid-base balance of intra- and extracellular fluids of marine  
78 organisms, which may result in reduced fitness or outright mortality (Seibel and Fabry, 2003;  
79 Seibel and Walsh, 2001; Widdicombe and Spicer, 2008). ~~Changes in CaCO<sub>3</sub> saturation~~  
80 state~~Perturbations of seawater carbonate chemistry~~ can also affect the ability of some calcifying  
81 animals to create and maintain calcium carbonate structures with implications for energetics,  
82 survival, competition and biogeochemical export (Fabry et al., 2008; Riebesell et al., 2000; Ries  
83 et al., 2009). Understanding the long-term effects of this increase in ocean acidity on both  
84 organisms and ecosystems has, therefore, become of great concern. Important and outstanding  
85 research goals are to understand how changing CO<sub>2</sub> impacts current populations and to predict  
86 whether these populations will be able to adapt to the rate and severity of the rising  
87 anthropogenic CO<sub>2</sub> inputs (e.g. Dam, 2013; Kelly and Hofmann, 2013; Sunday et al., 2011).

88 One approach to understanding the response of marine animals to ~~increasing~~ acidification  
89 is to examine places where animals already experience conditions of elevated CO<sub>2</sub>  
90 (hypercapnia). By comparing individuals that inhabit regions of high CO<sub>2</sub> with those that never  
91 experience high levels naturally, insight can be gained into the potential for adaptation of species  
92 to high CO<sub>2</sub> over evolutionary timescales. The ocean chemistry of the northwest Atlantic and the

93 northeast Pacific Oceans provides such a natural experiment. High CO<sub>2</sub> concentrations are  
94 generally absent from the upper water column in the Atlantic (Wanninkhof et al., 2010). In  
95 contrast there currently are hypercapnic conditions, where the water is undersaturated with  
96 respect to aragonite, in the upper water column in parts of the Pacific.

97         The source of hypercapnia in the Pacific Ocean is a combined result of ocean circulation  
98 coupled with the biological processes, leading the old deep waters of the Pacific to be some of  
99 the most CO<sub>2</sub> rich in the ocean (Broecker et al., 1982). On top of this natural process, ocean  
100 acidification also plays a role: the pH of the upper water column in the North Pacific is  
101 decreasing by ~~on the order of 0.001–0.002~~ pH units per year (~~Byrne et al., 2010~~);  
102 corresponding (Byrne et al., 2010; Chu et al., 2016), similar to the global average of 0.0022 pH  
103 units per year (Williams et al., 2015). Such a change corresponds to a total CO<sub>2</sub>, or dissolved  
104 inorganic carbon (DIC), increase of 1–2 μmol kg<sup>-1</sup> yr<sup>-1</sup> (Chu et al., 2016; Peng et al., 2003;  
105 Sabine et al., 2008; Sabine and Tanhua, 2010). Although the surface waters in these regions are  
106 typically well oxygenated and with a pH > 8, animals that live at or migrate to depth experience  
107 increasingly low oxygen (O<sub>2</sub>), pH, ~~undersaturation~~under-saturation with respect to calcium  
108 carbonate, and elevated CO<sub>2</sub> (Seibel, 2011). Historically these regions, which occur in many  
109 ocean basins, were in fact known more for their low O<sub>2</sub> than for their high CO<sub>2</sub> and were termed  
110 oxygen minimum zones (OMZs). These carbon maximum/oxygen minimum zones are extensive  
111 in the North Pacific Ocean, whereas similar conditions are rare in much of the Atlantic (Paulmier  
112 et al., 2011). Closely related taxa and cosmopolitan species in these two regions therefore  
113 experience very different pH levels, as well as CO<sub>2</sub> and O<sub>2</sub> concentrations in their normal  
114 distribution. Independent from high CO<sub>2</sub>, the reduced O<sub>2</sub> at depth in these OMZs has a profound  
115 impact on zooplankton distribution (i.e.: Escribano et al., 2009; Maas et al., 2014; Wishner et al.,  
116 2008) and can have important implications for the physiology of zooplankton (Childress and  
117 Seibel, 1998; Rosa and Seibel, 2008; Seibel, 2011).

118         Thecosome pteropods are an interesting group for investigating planktonic exposure and  
119 response to hypercapnia and low O<sub>2</sub>. Broadly distributed throughout the open ocean, species of  
120 thecosomes found in shallow waters of temperate and polar seas can become a numerically  
121 dominant member of the zooplankton community (Bednaršek et al., 2012a; Hunt et al., 2008; van  
122 der Spoel, 1967). As such, they can be an important part of the food chain (Armstrong et al.,  
123 2005; Hunt et al., 2008; Karnovsky et al., 2008), and contribute substantially to carbon flux

124 (Bauerfeind et al., 2009; Fabry and Deuser, 1991; Manno et al., 2010; Noji et al., 1997). Bearing  
125 thin shells of aragonite, one of the less stable forms of biogenic calcium carbonate, the  
126 calcification of thecosomes has been shown to be impacted by exposure to conditions replicating  
127 the projected changes in surface water pH and saturation state of the future ocean in the next 100  
128 years (Comeau et al., 2009; Lischka et al., 2011; Manno et al., 2012). Furthermore, recent  
129 assessments have shown that their shells already bear the mark of acidification are degraded in  
130 upwelling and polar regions- characterized by under-saturated conditions with respect to  
131 aragonite (Bednaršek et al., 2014a, b; Bednarsek and Ohman, 2015; Bednaršek et al., 2012b).  
132 Studies of metabolism and behavior, however, reveal a complex sensitivity to pH, dependent  
133 upon natural pre-exposure and the presence of interactive stressors (Comeau et al., 2010a; Maas  
134 et al., 2012a; Manno et al., 2012; Seibel et al., 2012).

135 Previous work has shown that some tropical and sub-tropical thecosome species undergo  
136 diel vertical migrations into persistent and pronounced regions of low O<sub>2</sub> and hypercapnia in the  
137 Eastern tropical North Pacific. These species showed no change in metabolic rate (O<sub>2</sub>  
138 consumption) when exposed to high CO<sub>2</sub> (1000 µatm), revealing the ability of some groups of  
139 thecosome to maintain aerobic metabolism in acidified waters for short periods of time. The one  
140 species in ~~this~~ the region that does not migrate, however, responded with a suppression of  
141 metabolism when exposed to high CO<sub>2</sub> (Maas et al., 2012a). This work in the Eastern tropical  
142 North Pacific provides evidence that there may be the potential for environmental adaptation of  
143 thecosomes to high CO<sub>2</sub>, but provides no insight into the combined effects of CO<sub>2</sub> with low O<sub>2</sub>.  
144 Although research into this topic is underway for other calcifying organisms in coastal habitats  
145 (Gobler et al., 2014; Melzner et al., 2013), in the open ocean our understanding remains limited.

146 The objective of this study, therefore, was to compare the effect of high CO<sub>2</sub> and low O<sub>2</sub>  
147 on thecosome pteropods from the northwest Atlantic and the northeast Pacific Oceans. One of  
148 the benefits of this comparison is that there are a number of species of thecosomes that have  
149 cosmopolitan distributions occupying both basins and that are known to be diel vertical  
150 migrators (Table 1; Bé and Gilmer, 1977; van der Spoel, 1967). Thus populations in the Pacific  
151 would naturally experience hypercapnia and low O<sub>2</sub> in their daytime deep-~~water~~ habitat in the  
152 Pacific, while in contrast, those from the Atlantic would rarely experience the same  
153 environmental stressors. The taxonomy of thecosomes has recently begun to be revisited using  
154 molecular and paleontological tools (i.e. Hunt et al., 2010; Janssen, 2012; Jennings et al., 2010;

155 [Maas et al., 2013](#)) and there is growing evidence of cryptic speciation for some pteropod groups  
156 ([Burrige et al., 2015](#); [Gasca and Janssen, 2014](#)). ~~deep water environmental stressors. Using~~  
157 ~~these organisms, which are presumably adapted to their local conditions, we can test whether~~  
158 ~~species~~It thus should be noted that the inter-basin comparisons performed here may be of cryptic  
159 ~~congeners rather than conspecific populations. Using these organisms, which are presumably~~  
160 ~~adapted to their local conditions, we can test whether species or congeners~~ exhibit a population-  
161 specific physiological response to these environmental conditions indicative of different  
162 sensitivities.

163

## 164 2. Methods

165 Thecosome pteropods caught during cruises to the northwest Atlantic and northeast Pacific were  
166 exposed aboard ship to manipulated conditions of moderately high CO<sub>2</sub> and/or low O<sub>2</sub> for short  
167 durations (< 18 h). After this exposure their metabolic rates were measured and then compared to  
168 determine whether there were species- or region-specific responses to the treatments.

### 169 2.1 Sampling

170 Animals were collected on two cruises, the first on August 7<sup>th</sup> – September 1<sup>st</sup> 2011 in the  
171 northwest Atlantic aboard the R/V *Oceanus*, and the second in the northeast Pacific from August  
172 9<sup>th</sup> – September 18<sup>th</sup> 2012 aboard the R/V *New Horizon*. The majority of the sampling in the  
173 Atlantic took place along a three-part ‘z’-shaped transect running between 35°N 52°W and 50°N  
174 42°W, as well as at sites during transit to and from port (Fig. 1). The first portion of this cruise  
175 track corresponded to a segment of the World Ocean Circulation Experiment / Climate and  
176 Ocean: Variability, Predictability and Change project (WOCE/CLIVAR) line A20. In the North  
177 Pacific the main sampling took place along a two-part transect running between 50°N 150°W  
178 and 33.5°N 135°W, corresponding to a portion of WOCE/CLIVAR line P17N, as well as at sites  
179 during transit to and from port (Fig. 1).

180 Sampling was part of a larger interdisciplinary project employing a suite of tools to  
181 explore the natural distribution and hydrographic environment of the thecosomes. The sampling  
182 design included underway measurements of hydrography, carbonate chemistry and multi-  
183 frequency acoustic ~~backscatter~~[backscattering](#). Comprehensive sampling of the water column  
184 was conducted at pre-determined stations using a depth-stratified 1-m<sup>2</sup> Multiple Opening/Closing  
185 Net and Environmental Sensing System with 150 µm mesh nets (MOCNESS; Wiebe et al.,

186 1985), a towed broadband echosounder, video plankton recorder casts, and profiles with a 24-  
187 [place](#) 10-L Niskin bottle rosette and associated conductivity, temperature and depth (CTD)  
188 package.

189 ~~Hydrographic profiles associated with this study were collected of temperature, O<sub>2</sub> and~~  
190 ~~salinity using the CTD-Rosette-Niskin bottle package at stations along the main survey transects~~  
191 ~~(Fig. 1).~~ This CTD was equipped with dual temperature and conductivity sensors, a Digiquartz  
192 pressure sensor, a SBE43 dissolved oxygen sensor, a biospherical underwater photosynthetically  
193 active radiation (PAR) sensor with surface reference, a Wet Labs C-Star transmissometer (660  
194 nm wavelength), and a Wet Labs ECO-AFL fluorometer.

195 Hydrographic profiles associated with this study were collected of temperature, O<sub>2</sub> and  
196 salinity using the CTD-Rosette-Niskin bottle package at stations along the main survey transects  
197 (Fig. 1). Where CTD casts were unavailable, at stations conducted during the transits to and  
198 from port, an expendable bathythermograph (XBT) was deployed to determine the temperature  
199 of the water column. Bottle samples of carbonate parameters, nutrients, and other parameters  
200 were collected at selected water depths using the CTD-Rosette package.

## 201 **2.2 Environmental Carbonate Chemistry**

202 Discrete pH samples were directly collected from the 10-L Niskin ~~bottle~~[bottles](#) into 10 cm  
203 cylindrical optical cells and measured within 4 h of collection (Clayton and Byrne, 1993;  
204 Dickson et al., 2007). These pH samples were analyzed spectrophotometrically on an Agilent  
205 8453 spectrophotometer at a control temperature ( $25.0 \pm 0.1^\circ\text{C}$ ) following the method detailed in  
206 Dickson (2007) and ~~in~~ Clayton and Byrne (1993) using m-cresol purple as the indicator. The pH  
207 results in total scale have been corrected for indicator impurity (Liu et al., 2011) and indicator  
208 perturbation to seawater samples. The pH measurements have a precision better than 0.001 and  
209 an accuracy of  $\sim 0.002$ .

210 Nutrient samples (nitrate/nitrite, phosphate, silicate, and ammonia) were collected in 20  
211 mL plastic bottles after filtration through a 0.22 $\mu\text{m}$  Pall capsule filter and kept frozen until  
212 analysis. Nutrient samples were analyzed either at the WHOI Nutrient Analytical Facility or the  
213 University of California, Santa Barbara, using a Lachat Instruments QuickChem 8000 four-  
214 channel continuous flow injection system, following standard colorimetric methods approved by  
215 U.S. Environmental Protection Agency.

216 Discrete samples were also taken for dissolved inorganic carbon (DIC) and total  
217 alkalinity (TA). These were collected in 250mL-250 mL Pyrex borosilicate glass bottles after  
218 being filtered with a 0.45  $\mu\text{m}$  in-line capsule filter and poisoned with saturated mercuric chloride  
219 (Dickson et al., 2007). DIC samples were analyzed on a DIC auto-analyzer (AS-C3, Apollo  
220 SciTech, Bogart, USA) via sample acidification, followed by non-dispersive infrared  $\text{CO}_2$   
221 detection (LiCOR 7000: Wang et al., 2013; Wang and Cai, 2004). The instrument was calibrated  
222 with certified reference material (CRM) from Dr. A.G. Dickson at the Scripps Institution of  
223 Oceanography. The DIC measurements have a precision and accuracy of  $\pm 2.0 \mu\text{mol kg}^{-1}$ . TA  
224 measurements were made with an Apollo SciTech alkalinity auto-titrator, a Ross combination  
225 pH electrode, and a pH meter (ORION 3 Star) based on a modified Gran titration method with a  
226 precision and accuracy of  $\pm 2.0 \mu\text{mol kg}^{-1}$  (Wang and Cai, 2004).

227 The remaining water column carbonate system parameters, including aragonite saturation  
228 state and  $\text{pCO}_2$  were calculated from DIC-pH pairs at in situ nutrient, temperature, salinity and  
229 pressure using the software CO2Sys (Pierrot et al., 2006) and the dissociation constants of  
230 Mehrbach et al. (1973), refitted by Dickson and Millero (1987), and the  $\text{KHSO}_4$  dissociation  
231 constant from Dickson (1990). Depths for  $\text{pH}=7.7$ ,  $\text{pCO}_2=800 \mu\text{atm}$  and aragonite saturation  
232 state of 1 were then linearly interpolated using the closest available measurements.

233 Surface water  $\text{pCO}_2$  was continuously measured throughout both cruises using an  
234 automated underway system (Model 8050, General Oceanics Inc., USA) based on headspace air-  
235 seawater equilibration followed by infrared detection (LiCOR 7000). This system was calibrated  
236 every 1-2 hours with three  $\text{CO}_2$  gas standards traceable to World Meteorological Organization  
237  $\text{CO}_2$  Mole Fraction Scale. These underway  $\text{pCO}_2$  measurements have a precision and accuracy of  
238  $\sim \pm 1 \mu\text{atm}$ . Measurements made by the underway system provide insight into the surface  
239 carbonate chemistry parameters at stations made in transit where bottle samples were not  
240 collected.

### 241 **2.3 Specimen Capture**

242 Thecosome species were chosen for physiological study opportunistically as they appeared in net  
243 samples at successive stations. Species were targeted specifically for their abundance and the  
244 likelihood of their presence in both ocean basins: and only adult individuals were used. Most  
245 individuals were collected with a 1-m diameter, 150- $\mu\text{m}$  mesh Reeve net with a  $\sim 25$  L cod-end in  
246 the Atlantic and a similar 1-m diameter, Reeve net equipped with 330- $\mu\text{m}$  mesh in the Pacific.



247 Use of the Reeve net with its large and heavy cod-end in combination with slow haul rates  
248 (typically 5-10 m min<sup>-1</sup>) allowed for a gentle collection of the delicate thecosomes, consistently  
249 supplying animals in good condition with undamaged shells and external mantle appendages.  
250 Net tows were made at night when animals were expected to congregate at shallow depths, were  
251 ~1 h in duration in an effort to minimize the handling time of the organisms, and reached a  
252 maximal depth between 100–150 m. Depths were targeted that had a high chlorophyll *a* peak  
253 during CTD casts, high acoustic backscattering on the echosounder, and/or where thecosomes  
254 had been abundantly sampled at the same station using the MOCNESS. Occasionally,  
255 individuals of less abundant species were collected from the nets of the MOCNESS for  
256 physiological study, but only if their shells were undamaged and they were swimming normally.

257 Post-capture, individuals were transferred to filtered water in densities of < 15 ind. L<sup>-1</sup>  
258 and kept for at least 8 h in temperature controlled waterbaths to allow for gut clearance.  
259 Temperatures for experimentation (20, 15 or 10°C) were chosen to be generally representative of  
260 the waters from which the animals were sampled, based on the vertical distributions and  
261 hydrographic conditions documented with the stratified MOCNESS sampling. Chosen  
262 temperatures were typically the average of the water temperature between 25-100 m, although in  
263 the middle section of the Atlantic cruise experimental temperatures were reflective of the 25–50  
264 m average due to the particularly shallow vertical distribution of the dominant species (*Limacina*  
265 *retroversa*) sampled in this region. This was to ensure that experiments were occurring at  
266 physiologically relevant and, presumably, natural temperatures for each species. After gut  
267 clearance, individuals that were in good condition (i.e., swimming and with shell intact) were  
268 used for oxygen consumption experiments.

#### 269 **2.4 Experimental Exposures and Oxygen Consumption Rate**

270 Post-gut-clearance, healthy individuals/animals were put into separate glass syringe respiration  
271 chambers, one individual per chamber, with a known volume of 0.2 µm filtered seawater and 25  
272 mg L<sup>-1</sup> each of streptomycin and ampicillin. This minimized the microbial respiration effects on  
273 the measurements of carbonate chemistry and O<sub>2</sub> consumption rates by pteropods during the  
274 experiments. The inclusion of antibiotics, a method which has previously been used with  
275 thecosomes to prevent bacterial growth in respiration experiments (Maas et al., 2012b), was  
276 shown during the Pacific cruise to have no effect on the O<sub>2</sub> consumption of at least *Limacina*  
277 *helicina*, for the exposure durations associated with these experiments (Howes et al., 2014). The

278 volume of water in the treatments was chosen to complement the size of the organism and  
279 temperature of the experiment and ranged between 15-50 mL in 2011 and 8-20 mL in 2012. For  
280 every 3-5 treatment chambers, a “control” respiration chamber (experimental seawater with  
281 antibiotics and without pteropods) was set up to monitor microbial activity and to provide water  
282 for characterization of the starting conditions.

283 Filtered seawater for experimental exposures was collected during both cruises in batches  
284 at approximately weekly intervals from the surface; experimental water thus began with  
285 chemical properties (notably including TA, DIC, pH, as well as salinity) reflective of the local  
286 environment and was then manipulated to modify CO<sub>2</sub> and/or O<sub>2</sub> concentrations. Manipulations  
287 were achieved by bubbling 1 L batches of collected seawater with gas mixes (certified accurate  
288 to ± 2%) for 45–60 min with one of two oxygen (21 and 10% O<sub>2</sub>) levels crossed with two CO<sub>2</sub>  
289 (nominally 380 ~~ppm~~µatm and 800 ~~ppm~~µatm) levels. At the time of the experiment, surface air  
290 pCO<sub>2</sub> conditions were on average ca. 380 ppm, dictating our ambient (*i.e.*, low carbon, LC)  
291 conditions. In 2011 the ambient condition (~21% O<sub>2</sub> and 380 µatm CO<sub>2</sub>) was achieved by  
292 bubbling with an ambient clean air line, while in 2012 it was achieved by a certified 380 ppm gas  
293 mix.

294 The experimentally modified concentrations mimic the CO<sub>2</sub> and O<sub>2</sub> levels that would be  
295 experienced by thecosomes within the top 400 m of the Pacific Ocean, and reflect the average  
296 projected atmospheric CO<sub>2</sub> level for the open ocean in the year 2100 (A2 emissions scenario,  
297 IPCC, 2007). This resulted in four total treatments: low (*i.e.*, ambient) CO<sub>2</sub>, high oxygen  
298 (LC/HO) representative of current ambient surface ocean conditions; high carbon, high oxygen  
299 (HC/HO), replicating what we expect the average future surface ~~oceans~~ocean to resemble; low  
300 CO<sub>2</sub>, low oxygen (LC/LO); and high carbon, low oxygen (HC/LO), which is similar to what  
301 organisms in the Pacific would experience during a diel vertical migration into the local oxygen  
302 minimum zone. The goal of this design was to allow us to compare directly the Atlantic and  
303 Pacific thecosomes to see whether exposure to 800 µatm pCO<sub>2</sub> and/or 10% O<sub>2</sub> resulted in  
304 different outcomes. The level of low O<sub>2</sub> chosen for this study was well above the threshold that  
305 has been designated as stressful for non-specialized metazoan life (< 2 mg O<sub>2</sub> L<sup>-1</sup> or 60 µmol O<sub>2</sub>  
306 kg<sup>-1</sup>; Vaquer-Sunyer and Duarte, 2008), in order to test the non-lethal effect of moderately low  
307 O<sub>2</sub> on individuals from the two ocean basins. Calculations based on the salinity and temperature  
308 of the water indicated that bubbling with 10% O<sub>2</sub> achieved conditions of 10–13% O<sub>2</sub> saturation

309 atby the start of experiments. Subsequent analyses (see below) also confirmed that intended CO<sub>2</sub>  
310 concentrations were achieved for all treatments within reasonable ranges, with the exception of  
311 the LC/LO Atlantic treatment. In this case, the gas cylinder was evidently improperly mixed by  
312 the manufacturer and analyses suggested a ca. 100 ppm CO<sub>2</sub> concentration. The results for this  
313 treatment are still presented but should be interpreted as a distinct treatment.

314 Oxygen consumption was measured following similar techniques as described in Marsh  
315 and Manahan (1999). Briefly, at the conclusion of the experiment water was withdrawn from  
316 treatment or control chambers using an airtight 500 µL Hamilton syringe and injected past a  
317 Clarke-type microcathode (part #1302, Strathkelvin Instruments, North Lanarkshire, United  
318 Kingdom) attached to an O<sub>2</sub> meter (part #782) in a water-jacketed injection port (part #MC100).  
319 This was done three times, allowing the reading to stabilize for at least 30 seconds before a  
320 measurement was taken. Generally, the change in oxygen consumption was between 3–25% of  
321 the control value. In high oxygen experiments, if the oxygen level fell below 70% of air  
322 saturation they were excluded from the analysis. Animals

323 Following exposure, animals were removed from the chamber, blotted dry and frozen in  
324 liquid nitrogen. These individuals were later weighed using a microbalance ( $\pm 0.0001$  g) and the  
325 resulting mass specific O<sub>2</sub> consumption rates are reported in  $\mu\text{moles (g wet weight)}^{-1} \text{ h}^{-1}$ . Wet  
326 weights are here used as they are more relevant for physiological understanding of animal  
327 function (Childress et al., 2008) but dry weights can be estimated from these using the wet  
328 weight to dry weight relationships developed previously for pteropods (Ikeda, 2014). To  
329 replicate the duration of exposure that would be experienced by most thecosomes in the Pacific  
330 undergoing a daily migration to depth, the experiments were targeted to last 6–12 h. In practice,  
331 experiments ranged from 6–18 h for normoxic and 3–10 h for low O<sub>2</sub> trials. This variation in  
332 duration resulted from balancing the need to elicit a measureable change in O<sub>2</sub> concentration with  
333 preventing extreme O<sub>2</sub> depletion of the chambers (< 6% oxygen saturation) and accounting for  
334 multiple species of variable size and metabolic rate.

## 335 **2.5 Experimental Carbonate Chemistry**

336 Carbonate chemistry of the treatments was characterized in most cases via measurements of DIC  
337 and TA of experimental seawater, unless indicated otherwise. The process of measuring the O<sub>2</sub> in  
338 the treatments used up a large portion of the water and then the chamber was unsealed and  
339 disturbed to remove the animal, rendering it impractical to measure the carbonate chemistry

340 directly from the respiration chambers. DIC measurements were thus taken from the control  
341 syringes within 18 h of the end of each experiment and used to represent the starting point of the  
342 carbonate chemistry conditions the animals experienced. Water samples were allowed to come to  
343 room temperature (> 6 h) before analysis. DIC was measured using the same system as that used  
344 for the hydrographic characterization (see above). Estimates of the effect of CO<sub>2</sub> production via  
345 respiration in treatment chambers on DIC were made using a respiratory quotient of 0.8 M of  
346 CO<sub>2</sub> per 1 M of O<sub>2</sub> consumed -(Mayzaud, 1976) to characterize the ending conditions of the  
347 experiments.

348 Due to the small volumes of water in the experimental chambers, it was not possible to  
349 measure both DIC and TA from the control syringes. Instead, TA samples intended to be  
350 representative of the starting experimental conditions were collected via siphoning from each  
351 batch of ~~filtered and antibiotic treated~~ collected surface water. These samples were subsequently  
352 measured based on the analytical method described above (Wang and Cai 2004). TA of  
353 experimental water was assumed to have been constant over the course of each experiment as  
354 water was filtered (0.2 μm) and antibiotic treated (thus microbial activities were kept at  
355 minimum), ~~and~~). Although pteropod aerobic respiration does, excretion, and calcification within  
356 a respiration chamber could influence TA, these are presumed to have not change TA in had a  
357 significant way influence over the time scales in question.

358 In some instances, however, measured TA from the batches of experimental water was  
359 substantially dissimilar to that of the surface measurements made from nearby in-situ surface  
360 bottle samples collected with the CTD (> 20 μmol kg<sup>-1</sup>; see section 3.3). Calculated pCO<sub>2</sub> values  
361 in these cases were also significantly different from batches of experimental water collected from  
362 other locations, but bubbled with the same CO<sub>2</sub> gas tank. These differences are more than 10  
363 times the measurement precision/accuracy and 5 times the uncertainty of duplicate sampling and  
364 measurements during the cruises. They are also beyond the likely level of TA variation due to  
365 differences in sampling location (geographic and in depth) between the in situ bottle samples and  
366 experimental water batches and rather are likely a consequence of the difficulties associated with  
367 cleanly siphoning the experimental water batches (i.e.g., contamination during sampling). For  
368 completeness, the carbonate chemistry system parameters for the experimental water, including  
369 aragonite saturation state and pCO<sub>2</sub>, are reported based on calculations using DIC-TA pairs using  
370 both the ~~in situ and~~ experimental TA and the in situ measurements from the CTD bottle samples;

371 in those cases where the TA measurements diverged substantially ( $> 20 \mu\text{mol kg}^{-1}$ ), however, we  
372 base our interpretations on the in-situ measured TA at nearby CTD stations instead of the values  
373 of experimental water. In those circumstances where batch water was taken from test stations  
374 during transit to/from the main study regions and CTD bottle data were unavailable, the  
375 experimental TA was checked using calculated TA values using DIC from the LC/HO treatments  
376 and  $\text{pCO}_2$  from the underway measurements.

## 377 **2.6 Statistics**

378 Oxygen consumption rates were tested for significant differences between groups ~~with~~  
379 ~~Bonferroni pairwise post-hoc comparisons~~ using SPSS. Univariate General Linear Models  
380 (GLM) were conducted to determine the effect of  $\text{CO}_2$  level,  $\text{O}_2$  level, and their interactive effect  
381 using the log transformed oxygen consumption with log transformed wet mass as a covariate  
382 separately for each species (2 factor design; " $\text{CO}_2 \times \text{O}_2$ "). In the Atlantic this full factorial design  
383 was confounded by the incorrect gas mixture so each treatment was tested independently (1  
384 factor design; "treatment"). Species that were collected during both years/basins, and  
385 experiments conducted on species at multiple temperatures, were analyzed separately so that the  
386 effect of variations in mass between seasons and the changes in metabolic rate at different  
387 temperatures would not confound the analysis. The datasets were tested for normality and  
388 homoscedasticity and, in cases where significance was found in the GLM they were explored  
389 with Bonferroni pairwise post-hoc comparisons.

390 For some species the temperature of experimentation was different among stations within  
391 a basin. For analyses with these species when comparing species between ocean basins, we  
392 applied a standard temperature coefficient ( $Q_{10}$ ) to compare across temperatures. The adjusted  
393 rates ( $R_f$ ) were calculated at  $15^\circ\text{C}$  using a  $Q_{10}$  of 2 according to the equation:

$$394 R_f = R_i * \left( Q_{10}^{\left( \frac{15 - T_i}{10} \right)} \right)$$

395 where  $R_i$  is the original metabolic rate measured at the original temperature ( $T_i$ ). Although  
396 previous work with thecosomes has shown that  $Q_{10}$  is species-specific (Maas et al., 2011; Maas  
397 et al., 2012b; Seibel et al., 2007), for many of the species used in this study there are no  
398 published estimates of  $Q_{10}$ . Thus, this coefficient value was chosen as it is mid-range for the  
399 published  $Q_{10}$  of non-polar thecosome species as recently compiled by Ikeda (2014; 1.3-2.7) and

400 is consistent with estimates of average  $Q_{10}$  for marine ectotherms, which typically fall between  
401 2-3 (Hochachka and Somero, 2002; Seibel and Drazen, 2007).

### 403 3. Results

#### 404 3.1 Specimen Capture

405 ~~Following currently accepted taxonomy, individuals from a total of eight species were collected~~  
406 ~~over the course of the two cruises for physiological studies. The taxonomy of thecosomes has~~  
407 ~~recently begun to be revisited using molecular and paleontological tools (i.e. Hunt et al., 2010;~~  
408 ~~Janssen, 2012; Jennings et al., 2010; Maas et al., 2013), however, Following currently accepted~~  
409 ~~morphology-based taxonomy, adult individuals from a total of eight species of pteropods were~~  
410 ~~collected over the course of the two cruises for physiological studies. Only relatively large adult~~  
411 ~~specimens were used in respiration trials, in part to avoid any confounding effects of ontogeny~~  
412 ~~and in part to ensure a measurable change in oxygen levels. ~~and there is growing evidence of~~~~  
413 ~~cryptic speciation for some pteropod groups (BurrIDGE et al., 2015; Gasca and Janssen, 2014). It~~  
414 ~~thus should be noted that these inter-basin comparisons may be of cryptic congeners rather than~~  
415 ~~conspecific populations.~~

416 We collected two species of thecosome pteropods exclusively from the Atlantic,  
417 *Limacina retroversa* (Fleming, 1823), a subpolar species, which is absent from the North Pacific,  
418 and *Diacria trispinosa* (Blainville, 1821), which can be found in temperate and tropical regions  
419 of the Atlantic, Pacific and Indian Oceans. Although present in both the North Atlantic and  
420 Pacific, the polar to sub-polar species *Limacina helicina* (Phipps, 1774), was only sampled in the  
421 Pacific transect. Collections of this species consisted of intermixed formae, the high spiraled  
422 *Limacina helicina helicina acuta* (van der Spoel, 1967), the lower spiraled *Limacina helicina*  
423 *helicina pacifica* (van der Spoel, 1967), and a forma that bore resemblance to both in a mixed  
424 morphology. Since both the assemblage and morphology of these formae were mixed they were  
425 tested as one population/species. In both ocean basins we collected *Styliola subula* (Quoy and  
426 Gaimard, 1827), *Cavolinia inflexa* (Lesueur, 1813) and *Clio pyramidata* (Linnaeus, 1767).  
427 There is some morphological and molecular evidence that *Cuvierina columnella* (Rang, 1827) is  
428 actually multiple distinct species, now including *Cuvierina atlantica* and *Cuvierina pacifica*  
429 (BurrIDGE et al., 2015; Janssen, 2005), and we tested individuals of these species from their  
430 respective ocean basins.

### 3.2 Hydrography

Two hydrographic regimes were evident along the North Pacific study transect (Table 2; Fig. 2). The northern-most stations, ~~including portions of the transit from port and stations from~~ (50°N 150°W to 47 °N 144.6°W; ~~stations T2-T7, 3-7; Fig. 1~~) were coldest, with ~~the~~ temperatures between 25-100 m ranging from 5-10°C. ~~In this area~~ At these stations O<sub>2</sub> fell below 10% (~130 μmol kg<sup>-1</sup>) ~~by at depths less than ~250 m. In this northern part of the transect,~~ pH fell below 7.7 ~~by at depths less than~~ 130 m, and pCO<sub>2</sub> had already reached 800 μatm by ~200 m. Individuals in this area experienced an Ω<sub>Ar</sub> = 1 between 160-185 m, well within the typical diel vertically migratory range of both of the species found in the region (*C. pyramidata* and *L. helicina*). At stations from more southern latitudes, ~~from~~ (47 °N 144.6°W to 33.5°N 135°W; ~~stations 15-34, T9-T10; Fig. 1~~), temperatures at depths between 25-100 m were higher, ranging between 10-~~15~~17°C, representative of the transition zone into the North Pacific Gyre. Along this portion of the transect O<sub>2</sub> concentration consistently fell below 10% by depths ~~between~~of 340 and 400 m. The depth at which pH fell below 7.7 increased gradually from ~150 to 230 m as latitude decreased. ~~Similarly~~Correspondingly, the depth at which pCO<sub>2</sub> in this area reached 800 μatm ~~deepened from~~was 330 to 440 m, and the aragonite saturation horizon ~~transitioned from~~ 330 m to 430 m depth. The depth at which species would experience a pH below 7.7 was within the inhabited depth range known from the literature for all of the species tested in this portion of the study region, but only the species *Clio pyramidata*, with a typical vertical range of 0-500 m (Table 2), would be likely experienced to experience 10% O<sub>2</sub>, 800 μatm pCO<sub>2</sub> and aragonite ~~undersaturation~~under-saturation in its typical distribution ~~in this portion of the Pacific transect~~ (Table 1).

In contrast to the Pacific, along the entire Atlantic transect O<sub>2</sub> concentration was above ~200 μmol kg<sup>-1</sup> (~15%) in the top 500 m, while pCO<sub>2</sub> never reached 800 μatm and aragonite ~~undersaturation~~under-saturation never occurred throughout the top 1000 m. There were three dominant hydrographic regimes in the Atlantic (Table 2; Fig. 2). In the northeastern part of the sampling region (50°N 42°W to 44.9 °N 42°W; ~~stations 21-31; Fig. 1~~), where the Gulf Stream meets the Labrador Current, average temperatures at 25-100 m were near 15°C and pH only fell below 7.7 at depths exceeding 400 m. Similarly, in the southwest part of the sampling region (from 42°N 52°W to 36°N 52°W; ~~stations 3-13; Fig. 1~~), corresponding to the Sargasso Sea and through the Gulf Stream, pH only fell below 7.7 at depths exceeding 450 m, although the upper

462 water column was warmer, with average temperatures ~~being of~~ 20°C. There was a third water  
463 mass type, typical of colder fresher shelf waters, at station 32 and in an intrusion off the Grand  
464 Banks at stations 17 and 19. ~~This Stations conducted in this~~ water ~~was were~~ typified by a  
465 temperature and salinity anomaly with ~~average~~ temperatures ~~falling~~ below 5°C from 25-100 m  
466 and a salinity signature < 33, contrasting significantly with the surface salinities of the northern  
467 portion (~34) and southern portion (~36) of the Atlantic transect. As a consequence, these  
468 stations contained water of the lowest pH, with surface waters reaching 7.7 at depths shallower  
469 than 200 m. ~~Based on previous knowledge of the vertical distributions of the thecosomes used in~~  
470 ~~this study, only the species *Clio pyramidata* would ever experience a pH below 7.7 in this overall~~  
471 ~~Atlantic study region and none of the thecosomes studied would experience 800 µatm pCO<sub>2</sub> or~~  
472 ~~under saturation within their vertical range (Table 1).~~

### 473 3.3 Carbonate Chemistry of Experiments

474 Bubbling with CO<sub>2</sub> levels of ~380 and ~800 ppm resulted in a distinct separation of carbonate  
475 chemistry between treatments during the experiments in both oceans (Table 3). Due to pre-  
476 existing differences in the carbonate chemistry of the seawater collected in each ocean, TA  
477 ~~concentrations were different~~differed between the two basin treatments. In the Atlantic the DIC  
478 of the ambient CO<sub>2</sub> treatments ranged from 2030-2090 µmol kg<sup>-1</sup> and the high CO<sub>2</sub> treatments  
479 from 2140-2220 µmol kg<sup>-1</sup>, with an average difference between treatments of similar temperature  
480 and salinity of 132 µmol kg<sup>-1</sup>. Surface TA in the region decreased from ~2370 µmol kg<sup>-1</sup> in the  
481 southern part of the transect to 2300 µmol kg<sup>-1</sup> in the northern latitudes. In the Pacific the DIC of  
482 the ambient CO<sub>2</sub> treatment ranged from 1930-2020 µmol kg<sup>-1</sup> and the high CO<sub>2</sub> treatment from  
483 2030-2110 µmol kg<sup>-1</sup>, with an average difference of 90.7 µmol kg<sup>-1</sup> between the treatments.  
484 Surface TA in this basin was 2150 µmol kg<sup>-1</sup> in the most northern collection and had  
485 ~~decreased~~increased to 2200 µmol kg<sup>-1</sup> by the transect mid-point.

486 Calculations of pCO<sub>2</sub> based on these measurements of DIC and TA suggested that target  
487 pCO<sub>2</sub> levels were generally attained and were consistent between the two cruises, with the  
488 exception of the LC/LO treatment in the Atlantic. In this case, there was a substantial deviation  
489 from the intended pCO<sub>2</sub>, suggesting values ranging from 99-139 µatm in contrast to a range of  
490 311-391 µatm for the LC/HO in the Atlantic and 283-409 µatm for LC/HO and 295-397 µatm in  
491 the LC/LO in the Pacific. Evidently, this indicates improper mixing of the gas concentration in  
492 the Atlantic LC/LO gas cylinder by the manufacturer. The calculations for the high CO<sub>2</sub>



493 treatments were more consistent between cruises, with pCO<sub>2</sub> for the HC/HO being 585-868  $\mu\text{atm}$   
494 for pCO<sub>2</sub> and the HC/LO being 755-783 in the Atlantic, while in the Pacific the HC/HO  
495 treatment was between 520-740  $\mu\text{atm}$  and the HC/LO 546-766  $\mu\text{atm}$ . The variability in  
496 calculated pCO<sub>2</sub> values likely represents variations in bubbling time, temperature, and the degree  
497 to which the water reached saturation relative to the gas mixtures. ~~The variability within each~~  
498 ~~distinct treatment may also reflect, to some degree, what pteropods may experience under that~~  
499 ~~particular mean condition, i.e. low vs. high CO<sub>2</sub>.~~

500 As a consequence of the natural differences in seawater carbonate chemistry, in particular  
501 the TA differences between two ocean basins, there were inherent differences in the aragonite  
502 saturation state between the Pacific and Atlantic treatments (Table 3). In the Atlantic,  $\Omega_{\text{Ar}}$  of the  
503 ambient CO<sub>2</sub> treatment ranged from 2.4-3.5, except for the LC/LO treatment ( $\Omega_{\text{Ar}}$  4.0-5.5), which  
504 was bubbled with an incorrect gas ~~mixture~~mixture as discussed above. Comparatively In  
505 comparison, in the Pacific the ambient CO<sub>2</sub> condition had a lower range of  $\Omega_{\text{Ar}}$  (2.2-2.4) for both  
506 the LC/HO and the LC/LO treatments. The experimental conditions of the high CO<sub>2</sub> treatments  
507 ~~in the Atlantic only approached under saturation~~reached their lowest value in the middle part of  
508 the transect ( $\Omega_{\text{Ar}} = 1.2$  at mid-latitudes; Table 3), where cold northern waters of low salinity were  
509 encountered ~~and~~. Experimental  $\Omega_{\text{Ar}}$  had a range of 1.5-2.0 for the rest of the transect in the  
510 Atlantic. The values of experimental  $\Omega_{\text{Ar}}$  were lower overall in the Pacific, although the high  
511 CO<sub>2</sub> treatments also never reached under-saturation ( $\Omega_{\text{Ar}}$  1.3-1.8). ~~The~~In general, the  
512 manipulation of carbonate chemistry in ~~general~~this study successfully created two distinct ranges  
513 for both pCO<sub>2</sub> and aragonite saturation state ( $\Omega_{\text{Ar}}$ ) ~~in this study.~~

514 It is important to acknowledge that the production of CO<sub>2</sub> via respiration of the organisms  
515 within the chambers would modify the carbonate chemistry of the treatments over the duration of  
516 the experiments. Based on the average respiration rate, ~~and using a respiratory quotient of 0.8~~  
517 ~~(Mayzaud, 1976)~~, we estimate an average DIC production of  $\sim 18.0 \mu\text{mol kg}^{-1}$  by the end of an  
518 experiment. Applying such a change to the experimental conditions in the northeast Pacific,  
519 where seawater is more sensitive to changes in DIC due to a lower buffering capacity compared  
520 to the Atlantic (i.e., a worst case scenario),  $\Omega_{\text{Ar}}$  would only change by  $<0.1$  in both the LC and  
521 HC experimental chambers over the course of the respiration experiments. Although this is an  
522 appreciable effect, we nonetheless retain a wide separation between the ambient and high CO<sub>2</sub>  
523 treatments and in no cases would the treatments reach under-saturation as a consequence of this

524 biological activity. As such, for simplicity the results reported in Table 3 do not include this  
525 [variability correction for respiration](#).

### 526 **3.4 Oxygen Consumption Rate**

#### 527 **3.4.1 Effect of CO<sub>2</sub>**

528 Varying availability and abundances of the different thecosome pteropod species in the net  
529 samples precluded all species being exposed to the full factorial design but individuals of all  
530 species were tested under the low CO<sub>2</sub>, high oxygen (LC/HO) and high carbon, high oxygen  
531 (HC/HO) treatments (Fig. 3, Table 4). To explore differences in metabolism attributable to a  
532 response to CO<sub>2</sub>, the log transformed wet mass was used in a GLM as a covariate comparing the  
533 log transformed oxygen consumption (response variable) under low and high CO<sub>2</sub> conditions;  
534 each population within a species that was sampled in both basins or run at multiple experimental  
535 temperatures, was examined separately. There was no significant effect of CO<sub>2</sub> for any species in  
536 either basin.

#### 537 **3.4.2 Effect of basin**

538 Following this assessment, we were interested in determining whether there were  
539 between basin differences in metabolic rate. As such we ran a GLM using log transformed  
540 metabolic rates for the three species that were found in both basins, normalized to 15 °C to  
541 account for differences in experimental temperature by applying a standard temperature  
542 coefficient. With the log-transformed wet mass as a covariate, we tested for an effect of basin,  
543 CO<sub>2</sub> and an interactive term. *Clio pyramidata* had a similar metabolic rate between basins. In  
544 contrast, *Cavolinia inflexa* ( $F_{1,20}=10.358$ ,  $p=0.004$ ) and *Styliola subula* ( $F_{1,23}=11.817$ ,  $p=0.002$ )  
545 both had a significantly lower metabolic rate in the Pacific, although no interactive effect of CO<sub>2</sub>.

#### 546 **3.4.2 Effect of O<sub>2</sub>**

547 For the species where enough individuals were collected to provide experimental  
548 replicates to explore the interactive effects of CO<sub>2</sub> and O<sub>2</sub> we also ran a species and basin  
549 specific GLM exploring the effect of treatment (Fig. 3, Table 5). *Clio pyramidata*, the only  
550 species we were able to test in both basins showed no significant effect of high CO<sub>2</sub>, low O<sub>2</sub> or  
551 the interactive treatment in either basin. In the Pacific, *L. helicina* and *C. inflexa* similarly  
552 showed no significant change in metabolic rate as a consequence of any of the treatments. In  
553 contrast, in the Atlantic, there was a significant effect of treatment for *L. retroversa* and a  
554 Bonferroni post-hoc analysis comparing the treatments found that the high CO<sub>2</sub>, low O<sub>2</sub> (HC/LO)

555 treatment was significantly lower than all other treatments (Fig. 4A4;  $F_{3,38}=17.836$ ,  $p<0.001$ ; a  
556 ~60% reduction in the average mass specific metabolic rate in comparison with the LC/HO  
557 treatment; Table 4). *Cuvierina atlantica* was tested at both 15 and 20 °C in the Atlantic, so to  
558 make comparisons among these experiments a temperature coefficient was applied and rates  
559 were normalized to 15 °C, after which no significant effect of any treatment was found for this  
560 species.

561

## 562 4. Discussion

563 This study reveals that short term exposure to low O<sub>2</sub> and high CO<sub>2</sub>, similar to what would be  
564 experienced by individuals in the Pacific during diel vertical migration, does not influence the  
565 oxygen consumption of adult individuals of most of the thecosome pteropod species examined  
566 from either the Atlantic or Pacific. The only species ~~which~~that had a significant change in  
567 respiration in response to any of the treatments was *Limacina retroversa* from the Atlantic, which  
568 responded to the combined effect of low O<sub>2</sub> and high CO<sub>2</sub> with a reduction in oxygen  
569 consumption rate.

### 570 4.1 Experimental Design

571 A factor that should be considered when interpreting our results is the dynamic hydrographic  
572 conditions that the animals experience naturally between and within the ocean basins.  
573 Thecosomes of multiple species were found at a range of temperatures, salinities and carbonate  
574 chemistries, meaning that they experienced a range of pH and aragonite saturation states in their  
575 natural habitat. When comparing animals from multiple locations, we chose to use local water in  
576 order to replicate these natural conditions and to manipulate exclusively the CO<sub>2</sub> concentration,  
577 as this is the factor that is changing due to anthropogenic activity. This approach, however, does  
578 not control for the other parameters of the carbonate chemistry system, which will vary between  
579 regions. Despite this fact, there was a clean distinction between treatments, notably in terms of  
580 aragonite saturation state as well as CO<sub>2</sub> concentration, ~~that~~which provides insight into the effect  
581 of moderate short duration exposure to CO<sub>2</sub>.

582 It is also important to note that the individuals of *L. helicina* from the Pacific experiments  
583 did occasionally have very high mortality during the period prior to experimentation (>80% at  
584 transit station T2 and T5, decreasing substantially to the northwest and along the main Pacific  
585 transect). These individuals, which are polar/sub-polar organisms and are typically found

586 between -2 to 10 °C (Lalli and Gilmer, 1989), were collected from water that was likely near the  
587 upper limit of their optimal temperatures; although alternate possibilities are that these were a  
588 population reaching senescence, or that they were collected in a hydrographic regime with low  
589 food availability. Animals collected from these sites that were used in subsequent respiration  
590 experiments may therefore have been taken from an already stressed population ~~of individuals~~  
591 and should be recognized as such.

## 592 **4.2 Carbon Dioxide Effect**

593 Hydrographic profiles collected in the Pacific coincident to sampling of thecosomes, indicate  
594 that organisms in the northern portion of the study region would experience conditions of high  
595 CO<sub>2</sub> and low O<sub>2</sub> in the upper ~450 m of their distribution (~~Chu et al., in review~~), ~~unlike~~ (Chu et  
596 al., 2016). Based on previous knowledge of the vertical distributions of the thecosomes used in  
597 this study, only the species *Clio pyramidata* would ever experience a pH below 7.7 and none of  
598 the thecosomes studied would experience 800 µatm pCO<sub>2</sub> or under-saturation within their  
599 vertical range in the Atlantic study region and (Table 1). Despite these environmental  
600 differences, we found no significant effect of increasing CO<sub>2</sub> alone on the respiration rates of any  
601 of the species from either ocean basin. These results increase the published evidence that short  
602 term (6-18 h) exposure to enhanced CO<sub>2</sub> without synergistic stressors has no significant effect on  
603 the metabolic rate of many species of thecosome pteropods. Thus far, there are only two species  
604 that have been documented to show a change in metabolism based on exposure to manipulated  
605 CO<sub>2</sub> alone: *Limacina antarctica* (789-1000 µatm, 24 h: Seibel et al., 2012) and *Diacria*  
606 *quadridentata* (1000 µatm, 6-18 h: Maas et al., 2012a). The metabolic rates of all other species  
607 yet studied, including *Hyalocylis striata*, *Clio pyramidata*, *Diacavolinia longirostris*, *Creseis*  
608 *virgula* (6-18 h: Maas et al., 2012a), and *Limacina helicina* (24 h: Comeau et al., 2010a), were  
609 not significantly affected by short term exposure to high CO<sub>2</sub>, although the latter species showed  
610 an increase in metabolic rate when high CO<sub>2</sub> was combined with high temperatures. Our results,  
611 which increase the geographic coverage for *L. helicina* and *C. pyramidata* and provide the first  
612 data about the species *C. pacifica*, *C. atlantica*, *L. retroversa*, *D. trispinosa*, *C. inflexa* and *S.*  
613 *subula*, corroborate these earlier findings.

614 One interpretation of these results is that physiological responses may have occurred, but  
615 involved the reallocation of resources to different tissues or metabolic pathways; this  
616 redistribution could serve to maintain the thecosome total energy budget, and subsequently

617 would not significantly change the metabolic rate of the individuals. A transcriptomic study done  
618 with individuals of *Clio pyramidata* as a companion project to the present work in fact suggested  
619 that expression of some genes was influenced by CO<sub>2</sub> exposure even though metabolic rate ~~is~~was  
620 not (Maas et al., 2015), perhaps suggesting some re-allocation among energetic demands. If this  
621 is the case it indicates that, to some degree, the short-term exposure to high CO<sub>2</sub> concentration is  
622 within the physiological tolerance of the tested species. Alternative hypotheses are that the  
623 duration of exposure was too short or the severity of the CO<sub>2</sub> treatment too minimal to elicit a  
624 measurable response. It is possible, for example, that some processes, like biomineralization,  
625 may be influenced by high CO<sub>2</sub>, but only after a longer exposure duration. Finally, it may be that  
626 changes in respiration rate were subtle, requiring a much greater sample size to identify in light  
627 of biological variability, but exploration of this hypothesis would require a dedicated experiment  
628 to collect more individuals and likely a smaller number of species.

629         This possible tolerance to short term CO<sub>2</sub> exposure may be due to the fact that within  
630 their distribution or diel migrational range there are conditions, or perhaps seasons, where the  
631 natural hydrography causes many species of thecosome to experience conditions of high  
632 CO<sub>2</sub>/low pH, and the species are therefore adapted to this range of exposure. The Arctic species  
633 *L. helicina* and subarctic species *L. retroversa*, for instance, are thought to inhabit waters which  
634 have been shown to reach a concentration of > 950 µatm CO<sub>2</sub> and to be undersaturated with  
635 respect to aragonite during the winter season in Kongsfjord, Svalbard (Lischka and Riebesell,  
636 2012). These conditions are pervasive throughout the upper water column, meaning that *L.*  
637 *helicina* and *L. retroversa*, which are not strong diel migrators, would experience seasonal under-  
638 saturation in these polar regions. The more temperate and tropical open ocean thecosomes,  
639 including *C. pyramidata*, *C. inflexa* and *S. subula* are all currently believed to be circumglobal  
640 and most, to varying degrees, diel migratory (Table 1; Bé and Gilmer, 1977; van der Spoel,  
641 1967). Populations are therefore likely to encounter high CO<sub>2</sub> in sub-surface waters in regions  
642 associated with OMZs, including much of the North Pacific and off the coast of Northern Africa.  
643 The ability to cope with high CO<sub>2</sub> for short durations may have been selected for over time as a  
644 natural consequence of the types of unavoidable environmental variability experienced by these  
645 planktonic populations.

#### 646         **4.3 Low O<sub>2</sub> and Combined Effects**

647 In the Pacific Ocean, none of the species for which we had enough individuals to perform the  
648 low O<sub>2</sub> study (*L. helicina*, *C. pyramidata*, and *C. inflexa*) had a significant change in metabolic  
649 rate under low (10%) O<sub>2</sub>, even when combined with enhanced CO<sub>2</sub>. These results indicate that  
650 the O<sub>2</sub> levels were above the concentration below which these species can no longer sustain their  
651 routine metabolic activity (Pcrit; Hochachka and Somero, 2002) and that any changes in  
652 physiology associated with the treatments required no increased energetic expenditure or  
653 metabolic reduction. As subsurface waters throughout the cruise were frequently below 10% O<sub>2</sub>  
654 (< ~130 μmol kg<sup>-1</sup>), this indicates that these species may be naturally adapted to coping with low  
655 O<sub>2</sub> conditions.

656 In the Atlantic, examination of the effects of low O<sub>2</sub> is ~~confound~~confounded by an  
657 unfortunate and accidentally low level of CO<sub>2</sub> (~130 μatm) in the LC/HO treatment (Table 3).  
658 Tests of the effect of high CO<sub>2</sub> (HC/HO) and the interactive (HC/LO) treatments nonetheless  
659 remain valid, and for *L. retroversa*, exposure to HC/LO caused a large and significant reduction  
660 in metabolic rate. Suppression in metabolic rate is a common tactic for surviving unfavorable  
661 conditions (Guppy and Withers, 1999; Seibel, 2011). Although metabolic depression is generally  
662 survivable in the short term, over longer time scales there are often implications for growth,  
663 reproduction and survival (reviewed in: Pörtner, 2010; Seibel, 2011). In the Atlantic, our  
664 measured in situ O<sub>2</sub> levels were never below 15% (~200 μmol kg<sup>-1</sup>). In contrast with the other  
665 species studied, which in at least some portions of their geographic range are occasionally found  
666 in association with subsurface low O<sub>2</sub> combined with hypercapnia, *L. retroversa* lives  
667 exclusively in the sub-polar North Atlantic Ocean and the Southern Circumpolar Current. As  
668 such this is the only species in this study in which no population is likely to experience  
669 conditions of low O<sub>2</sub> and high CO<sub>2</sub> together naturally anywhere in its distribution. Its inability to  
670 maintain metabolic rate during this interactive exposure may be a short-term metabolic response  
671 to environmental conditions that are unsustainable over longer time periods. As a consequence of  
672 the very low CO<sub>2</sub> in the LC/LO treatment, it is impossible to determine whether the metabolic  
673 suppression for *L. retroversa* in the HC/LO was in response to reduced O<sub>2</sub> availability alone or to  
674 the interactive effect of low O<sub>2</sub> with high CO<sub>2</sub>. In the LC/LO treatment any change in respiration  
675 due to low O<sub>2</sub> could have been masked by a change in the energy budget as a response to the low  
676 (equivalent to pre-industrial atmospheric conditions) levels of CO<sub>2</sub>. The results suggest that  
677 further work in the Atlantic is warranted to disentangle these stressors and to determine whether

678 the observed change in metabolic rate was solely a consequence of O<sub>2</sub> availability or truly a  
679 synergistic effect.

680 Interestingly, although the temperature coefficients were not species-specific and may  
681 not, therefore, perfectly normalize the dataset, one trend revealed by their use was a significant  
682 difference in the normalized metabolic rates between individuals of the species ~~such as~~ *S. subula*  
683 and *C. inflexa* from the Atlantic and Pacific Oceans. The comparatively lower metabolic rates  
684 from the Pacific may be a real response to the lower availability of O<sub>2</sub> for aerobic metabolism.  
685 Having a slower routine rate of O<sub>2</sub> consumption may be the result of a more efficient respiratory  
686 mechanism or an adaptation for coping with occasional exposures to the relatively high CO<sub>2</sub> and  
687 low O<sub>2</sub> conditions found in the northeast Pacific Ocean.

688

## 689 5. Conclusions

690 Thecosomes pteropods are thought to be some of the most sensitive of the oceanic zooplankton  
691 species to acidification. The responses we documented in the face of short-term CO<sub>2</sub> exposure  
692 and low O<sub>2</sub> reveal interesting patterns about basin scale differences in sensitivity, possibly  
693 associated with adaptation to local environmental conditions. Importantly, our results indicate  
694 that short-term exposure to high CO<sub>2</sub> does not have an effect on the respiration rate of multiple  
695 species of temperate and sub-polar thecosome species from both the North Atlantic and Pacific  
696 Oceans, irrespective of recent likely environmental exposure. The lack of effect of CO<sub>2</sub> as a  
697 single-stressor on metabolic rate in adult organisms of various species has been seen in a number  
698 of studies (reviewed in: Dupont et al., 2010; Kroeker et al., 2013), although there are many other  
699 metrics that have been shown to be more consistently affected. As such, thecosomes may have  
700 physiological coping mechanisms that allow them to maintain their energy budget for short  
701 periods of time in the face of high CO<sub>2</sub> via the re-allocation of their energetic resources. Over  
702 longer time periods, however, this could reduce their scope for growth and reproduction,  
703 negatively impacting the fitness of the population as has been demonstrated with other marine  
704 calcifiers (i.e.: Dupont et al., 2013; Melzner et al., 2013; Stumpp et al., 2011). Testing these  
705 hypotheses remains difficult as thecosomes are hard to maintain in captivity and there are no  
706 published studies of individuals kept fed and exposed to CO<sub>2</sub> in laboratory conditions for long  
707 durations (reviewed in: Howes et al., 2014; Thabet et al., 2015). Keeping individuals well fed is  
708 a critical factor since high food availability has been suggested to modulate the effect of high

709 CO<sub>2</sub> exposure in both thecosomes (Seibel et al., 2012) and ~~in~~ other calcifying species (Thomsen  
710 et al., 2013). Comparative short-term studies of wild caught animals such as the present  
711 experiments, therefore, currently give us the best insight into the sensitivity of these open-ocean  
712 populations, and the ability to predict how they will respond to the expected changes in the ocean  
713 environment.

714 Furthermore, although adult individuals may show no change in metabolic rate, there is  
715 evidence that juvenile stages of many calcifying species are typically more sensitive to CO<sub>2</sub>  
716 exposure (i.e. Connell et al., 2013; Waldbusser et al., 2015) and emerging evidence supports the  
717 idea that eggs, veligers and juveniles of *L. retroversa* and *L. helicina* are more vulnerable to  
718 acidification than adults (Lischka et al., 2011; Manno et al., 2016; Thabet et al., 2015). Thus,  
719 although adults may be capable of surviving short-term exposure, as acidity in surface waters  
720 increases there may be population level stress due to ontogenetic sensitivity.

721 These findings also draw attention to the consequences of the high degree of vertical  
722 variability in the open ocean environment, with animals in the Pacific found migrating between  
723 deep waters, undersaturated with respect to aragonite, and the surface (~~Lawson, unpublished~~  
724 ~~data; Chu et al., in review; Maas et al., 2012a~~)(~~Lawson, unpublished data; Chu et al., 2016;~~  
725 ~~Maas et al., 2012a~~). Recent studies in the California Current system indicate that thecosome  
726 shells show signs of in situ dissolution when associated with ~~water masses~~waters that are  
727 undersaturated with respect to aragonite (~~Bednaršek et al., 2014b; Bednarsek and Ohman,~~  
728 ~~2015~~)(~~Bednaršek et al., 2014b; Bednarsek and Ohman, 2015~~). Although our short duration  
729 experiments do not directly address the effect of longer-term exposure to high CO<sub>2</sub>, it does  
730 remind us that as open ocean environments respond to anthropogenic change there may be  
731 vertical refugia from ~~OA~~ocean acidification stress as well as regions where animals may already  
732 experience high CO<sub>2</sub>. As surface waters acidify, the ability to endure short-duration exposure and  
733 to migrate in both the Atlantic and Pacific populations may provide mechanisms for mitigating  
734 detrimental effects of acidification exposure. The potential compression of vertical habitat  
735 associated with the shoaling of the aragonite compensation depth, however, may have  
736 implications for predator/prey interactions, carbon pumping and other ecosystem functions  
737 (Bednarsek and Ohman, 2015; Seibel, 2011). Furthermore, it is clear that thecosome shells are  
738 highly sensitive to dissolution (Comeau et al., 2012; Lischka and Riebesell, 2012; Manno et al.,



739 2012) and there could be fitness and ecological consequences of dissolution in regions with  
740 vertical variation in carbonate chemistry.

741 Finally, as concerns about increasing CO<sub>2</sub> drive further explorations of comparative  
742 organismal physiology in the marine system, it is important to recognize that often the exposure  
743 of animals to increased CO<sub>2</sub> will occur in concert with expanding regions of low O<sub>2</sub>. This has  
744 been explored in the coastal environment where the interaction of acidification with  
745 eutrophication and associated low O<sub>2</sub> is comparatively well studied (Cai et al., 2011; Melzner et  
746 al., 2013); and in theoretical frameworks (Gruber, 2011; Pörtner, 2010; Sokolova, 2013).  
747 Experiments in the open ocean environment, however, are only beginning to be conducted and  
748 their implications explored. This study suggests that to make accurate predictions about how  
749 populations will respond to climate change and adequately understand the factors affecting  
750 organismal response, further investigations of the interactive effects of low O<sub>2</sub> and hypercapnia  
751 should consider natural environmental variability, population biogeography and phylogenetic  
752 sensitivity.

753 **Data availability**

754 Cruise data for the project is available via ~~BCO-DMO~~[the National Science Foundation's](#)  
755 [Biological and Chemical Oceanography Data Management Office \(BCO-DMO\)](#) under the  
756 project “Horizontal and Vertical Distribution of Thecosome Pteropods in Relation to Carbonate  
757 Chemistry in the Northwest Atlantic and Northeast Pacific” ([http://www.bco-](http://www.bco-dmo.org/project/2154)  
758 [dmo.org/project/2154](http://www.bco-dmo.org/project/2154)). The raw data for the respiration experiments are included in this  
759 deposition (DOI: 10.1575/1912/6421).

760

761 **Author contributions**

762 A. Maas and G. Lawson designed the experiments. All co-authors participated in oceanographic  
763 cruises and collection of samples. A. Maas conducted all of the experiments and statistical  
764 analyses. Z.A. Wang advised on the ~~design~~[manipulation](#) of ~~the~~ carbonate chemistry ~~analysis~~ and  
765 provided the measurements of both the hydrographic and experimental conditions. A. Maas  
766 prepared the manuscript with contributions from ~~all~~[both](#) co-authors.

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778 **References**

- 779 Armstrong, J. L., Boldt, J. L., Cross, A. D., Moss, J. H., Davis, N. D., Myers, K. W., Walker, R.  
 780 V., Beauchamp, D. A., and Haldorson, L. J.: Distribution, size, and interannual, seasonal and  
 781 diel food habits of northern Gulf of Alaska juvenile pink salmon, *Oncorhynchus gorbuscha*,  
 782 Deep Sea Research Part II: Topical Studies in Oceanography, 52, 247-265, 2005.
- 783 Bauerfeind, E., Nöthig, E. M., Beszczynska, A., Fahl, K., Kaleschke, L., Kreker, K., Klages, M.,  
 784 Soltwedel, T., Lorenzen, C., and Wegner, J.: Particle sedimentation patterns in the eastern  
 785 Fram Strait during 2000-2005: Results from the Arctic long-term observatory  
 786 HAUSGARTEN, Deep Sea Research (Part I, Oceanographic Research Papers), 56, 1471-  
 787 1487, 2009.
- 788 Bé, A. W. H. and Gilmer, R. W.: A zoogeographic and taxonomic review of Euthecosomatous  
 789 Pteropoda. In: Oceanic Micropalaeontology, Ramsay, A. (Ed.), Academic Press, London,  
 790 1977.
- 791 Bednaršek, N., Feely, R., Reum, J., Peterson, B., Menkel, J., Alin, S., and Hales, B.: *Limacina*  
 792 *helicina* shell dissolution as an indicator of declining habitat suitability owing to ocean  
 793 acidification in the California Current Ecosystem, Proceedings of the Royal Society of  
 794 London B: Biological Sciences, 281, 20140123, 2014a.
- 795 Bednaršek, N., Feely, R., Reum, J., Peterson, B., Menkel, J., Alin, S., and Hales, B.: *Limacina*  
 796 *helicina* shell dissolution as an indicator of declining habitat suitability owing to ocean  
 797 acidification in the California Current Ecosystem, Proceedings of the Royal Society B:  
 798 Biological Sciences, 281, 20140123, 2014b.
- 799 Bednaršek, N., Možina, J., Vogt, M., O'Brien, C., and Tarling, G.: The global distribution of  
 800 pteropods and their contribution to carbonate and carbon biomass in the modern ocean, Earth  
 801 System Science Data, 4, 167-186, 2012a.
- 802 Bednaršek, N. and Ohman, M.: Changes in pteropod distributions and shell dissolution across a  
 803 frontal system in the California Current System, Marine Ecology Progress Series, 523, 93-  
 804 103, 2015.
- 805 Bednaršek, N., Tarling, G., Bakker, D., Fielding, S., Jones, E., Venables, H., Ward, P., Kuzirian,  
 806 A., Lézé, B., and Feely, R.: Extensive dissolution of live pteropods in the Southern Ocean,  
 807 Nature Geoscience, 5, 881-885, 2012b.
- 808 Bigelow, H. B.: Plankton of the offshore waters of the Gulf of Maine, Govt. print. off., 1924.
- 809 Bopp, L., Resplandy, L., Orr, J. C., Doney, S. C., Dunne, J. P., Gehlen, M., Halloran, P., Heinze,  
 810 C., Ilyina, T., and Séférian, R.: Multiple stressors of ocean ecosystems in the 21st century:  
 811 projections with CMIP5 models, Biogeosciences Discussions, 10, 3627-3676, 2013.
- 812 Broecker, W. S., Peng, T.-H., and Beng, Z.: Tracers in the Sea, Lamont-Doherty Geological  
 813 Observatory, Columbia University, Palisades, NY, 1982.
- 814 Burrige, A. K., Goetze, E., Raes, N., Huisman, J., and Peijnenburg, K. T.: Global biogeography  
 815 and evolution of *Cuvierina* pteropods, BMC evolutionary biology, 15, 2015.
- 816 Byrne, R. H., Mecking, S., Feely, R. A., and Liu, X.: Direct observations of basin-wide  
 817 acidification of the North Pacific Ocean, Geophys Res Lett, 37, L02601, 2010.
- 818 Cai, W.-J., Hu, X., Huang, W.-J., Murrell, M. C., Lehrter, J. C., Lohrenz, S. E., Chou, W.-C.,  
 819 Zhai, W., Hollibaugh, J. T., and Wang, Y.: Acidification of subsurface coastal waters  
 820 enhanced by eutrophication, Nature Geoscience, 4, 766-770, 2011.
- 821 Childress, J. J. and Seibel, B. A.: Life at stable low oxygen levels: adaptations of animals to  
 822 oceanic oxygen minimum layers, Journal of Experimental Biology, 201, 1223-1232, 1998.

823 Childress, J. J., Seibel, B. A., and Thuesen, E. V.: N-specific metabolic data are not relevant to  
824 the ‘visual interactions’ hypothesis concerning the depth-related declines in metabolic rates:  
825 Comment on Ikeda et al.(2006), *Mar Ecol Prog Ser*, 373, 187-191, 2008.

826 Chu, S. N., Wang, Z. A., Doney, S. C., Lawson, G. L., and Hoering, K. A.: Changes in  
827 anthropogenic carbon storage in the Northeast Pacific in the last decade, *Journal of*  
828 ~~Geophysical Research—Ocean~~, *in review*, *in review*: *Oceans*, 121, 2016.

829 Clayton, T. D. and Byrne, R. H.: Spectrophotometric seawater pH measurements - Total  
830 hydrogen ion concentration scale calibration of *m*-cresol purple and at-sea results, *Deep-Sea*  
831 *Res. (I)*, 40, 2115-2129, 1993.

832 Comeau, S., Alliouane, S., and Gattuso, J.-P.: Effects of ocean acidification on overwintering  
833 juvenile Arctic pteropods *Limacina helicina*, *Marine Ecology Progress Series*, 456, 279-284,  
834 2012.

835 Comeau, S., Gorsky, G., Jeffree, R., Teysse, J., and Gattuso, J. P.: Impact of ocean acidification  
836 on a key Arctic pelagic mollusc (*Limacina helicina*), *Biogeosciences*, 6, 1877-1882, 2009.

837 Comeau, S., Jeffree, R., Teysse, J. L., and Gattuso, J. P.: Response of the Arctic pteropod  
838 *Limacina helicina* to projected future environmental conditions, *PLoS One*, 5, e11362,  
839 2010a.

840 Connell, S. D., Kroeker, K. J., Fabricius, K. E., Kline, D. I., and Russell, B. D.: The other ocean  
841 acidification problem: CO<sub>2</sub> as a resource among competitors for ecosystem dominance,  
842 *Philosophical Transactions of the Royal Society B: Biological Sciences*, 368, 20120442,  
843 2013.

844 Dam, H. G.: Evolutionary Adaptation of Marine Zooplankton to Global Change, *Annual Review*  
845 *of Marine Science*, 5, 349-370, 2013.

846 Dickson, A. G.: Thermodynamics of the dissociation of boric acid in synthetic seawater from  
847 273.15 to 318.15 K, *Deep Sea Research Part A. Oceanographic Research Papers*, 37, 755-  
848 766, 1990.

849 Dickson, A. G. and Millero, F. J.: A comparison of the equilibrium constants for the dissociation  
850 of carbonic acid in seawater media, *Deep Sea Research Part A. Oceanographic Research*  
851 *Papers*, 34, 1733-1743, 1987.

852 Dickson, A. G., Sabine, C. L., and Christian, J. R.: Guide to best practices for ocean CO<sub>2</sub>  
853 measurements, *PICES special publication*, 3, 2007.

854 Doney, S. C., Fabry, V. J., Feely, R. A., and Kleypas, J. A.: Ocean acidification: the other CO<sub>2</sub>  
855 problem, *Marine Science*, 1, 169-192, 2009.

856 Dupont, S., Dorey, N., Stumpp, M., Melzner, F., and Thorndyke, M.: Long-term and trans-life-  
857 cycle effects of exposure to ocean acidification in the green sea urchin *Strongylocentrotus*  
858 *droebachiensis*, *Marine biology*, 160, 1835-1843, 2013.

859 Dupont, S., Dorey, N., and Thorndyke, M.: What meta-analysis can tell us about vulnerability of  
860 marine biodiversity to ocean acidification?, *Estuarine, Coastal and Shelf Science*, 89, 182-  
861 185, 2010.

862 Escribano, R., Hidalgo, P., and Krautz, C.: Zooplankton associated with the oxygen minimum  
863 zone system in the northern upwelling region of Chile during March 2000, *Deep Sea*  
864 *Research Part II: Topical Studies in Oceanography*, 56, 1083-1094, 2009.

865 Fabry, V. J. and Deuser, W. G.: Aragonite and magnesian calcite fluxes to the deep Sargasso  
866 Sea, *Deep Sea Research Part A. Oceanographic Research Papers*, 38, 713-728, 1991.

867 Fabry, V. J., Seibel, B. A., Feely, R. A., and Orr, J. C.: Impacts of ocean acidification on marine  
868 fauna and ecosystem processes, *ICES Journal of Marine Science*, 65, 414-432, 2008.

869 Gasca, R. and Janssen, A. W.: Taxonomic review, molecular data and key to the species of  
870 Creseidae from the Atlantic Ocean, *Journal of Molluscan Studies*, 80, 35-42, 2014.

871 Gobler, C. J., DePasquale, E. L., Griffith, A. W., and Baumann, H.: Hypoxia and acidification  
872 have additive and synergistic negative effects on the growth, survival, and metamorphosis of  
873 early life stage bivalves, *PLoS ONE*, 9, e83648, 2014.

874 Gruber, N.: Warming up, turning sour, losing breath: ocean biogeochemistry under global  
875 change, *Philosophical Transactions of the Royal Society A*, 369, 1980-1996, 2011.

876 Guppy, M. and Withers, P.: Metabolic depression in animals: physiological perspectives and  
877 biochemical generalizations, *Biological Reviews*, 74, 1-40, 1999.

878 Halpern, B. S., Walbridge, S., Selkoe, K. A., Kappel, C. V., Micheli, F., D'Agrosa, C., Bruno, J.  
879 F., Casey, K. S., Ebert, C., Fox, H. E., Fujita, R., Heinemann, D., Lenihan, H. S., Madin, E.  
880 M. P., Perry, M. T., Selig, E. R., Spalding, M., Steneck, R., and Watson, R.: A global map of  
881 human impact on marine ecosystems, *Science*, 319, 948-952, 2008.

882 Haugan, P. M. and Drange, H.: Effects of CO<sub>2</sub> on the ocean environment, *Energy Conversion  
883 and Management*, 37, 1019-1022, 1996.

884 Hochachka, P. W. and Somero, G. N.: Biochemical adaptation: mechanism and process in  
885 physiological evolution, Oxford University Press, New York, 2002.

886 Howes, E. L., Bednaršek, N., Büdenbender, J., Comeau, S., Doubleday, A., Gallager, S. M.,  
887 Hopcroft, R. R., Lischka, S., Maas, A. E., and Bijma, J.: Sink and swim: a status review of  
888 thecosome pteropod culture techniques, *Journal of Plankton Research*, 36, 299-315, 2014.

889 Hunt, B., Strugnell, J., Bednarsek, N., Linse, K., Nelson, R. J., Pakhomov, E., Seibel, B.,  
890 Steinke, D., and Würzberg, L.: Poles Apart: The “Bipolar” Pteropod Species *Limacina  
891 helicina* Is Genetically Distinct Between the Arctic and Antarctic Oceans, *PLoS ONE*, 5,  
892 e9835, 2010.

893 Hunt, B. P. V., Pakhomov, E. A., Hosie, G. W., Siegel, V., Ward, P., and Bernard, K.: Pteropods  
894 in Southern Ocean ecosystems, *Progress in Oceanography*, 78, 193-221, 2008.

895 Ikeda, T.: Metabolism and chemical composition of marine pelagic gastropod molluscs: a  
896 synthesis, *Journal of Oceanography*, 70, 289-305, 2014.

897 IPCC: Climate Change 2013. The Physical Science Basis. Working Group I Contribution to the  
898 Fifth Assessment Report of the Intergovernmental Panel on Climate Change, Cambridge  
899 University Press, Cambridge, UK, 2013.

900 IPCC: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment  
901 Report of the Intergovernmental Panel on Climate Change, Cambridge University Press,  
902 Cambridge, United Kingdom and New York, NY, USA, 996, 2007, 2007.

903 Janssen, A. W.: Development of Cuvierinidae (Mollusca, Euthecosomata, Cavolinoidea) during  
904 the Cainozoic: a non-cladistic approach with a re-interpretation of Recent taxa, *BASTERIA-  
905 LISSE-*, 69, 25, 2005.

906 Janssen, A. W.: Late Quaternary to Recent holoplanktonic Mollusca (Gastropoda) from bottom  
907 samples of the eastern Mediterranean Sea: systematics, morphology, *Bollettino  
908 Malacologico*, 48, 1-105, 2012.

909 Jennings, R. M., Bucklin, A., Ossenbrügger, H., and Hopcroft, R. R.: Species diversity of  
910 planktonic gastropods (Pteropoda and Heteropoda) from six ocean regions based on DNA  
911 barcode analysis, *Deep Sea Research Part II: Topical Studies in Oceanography*, 57, 2199-  
912 2210, 2010.

- 913 Karnovsky, N. J., Hobson, K. A., Iverson, S., and Hunt, G. L.: Seasonal changes in diets of  
914 seabirds in the North Water Polynya: a multiple-indicator approach, *Marine Ecology*  
915 *Progress Series*, 357, 99, 2008.
- 916 Kelly, M. W. and Hofmann, G. E.: Adaptation and the physiology of ocean acidification,  
917 *Functional Ecology*, 27, 980–990, 2013.
- 918 Kroeker, K. J., Kordas, R. L., Crim, R., Hendriks, I. E., Ramajo, L., Singh, G. S., Duarte, C. M.,  
919 and Gattuso, J. P.: Impacts of ocean acidification on marine organisms: quantifying  
920 sensitivities and interaction with warming, *Global change biology*, 19, 1884–1896, 2013.
- 921 Lalli, C. M. and Gilmer, R. W.: *Pelagic Snails: The Biology of Holoplanktonic Gastropod*  
922 *Mollusks*, Stanford University Press, Stanford, CA, 1989.
- 923 Lischka, S., Büdenbender, J., Boxhammer, T., and Riebesell, U.: Impact of ocean acidification  
924 and elevated temperatures on early juveniles of the polar shelled pteropod *Limacina helicina*:  
925 mortality, shell degradation, and shell growth, *Biogeosciences*, 13, 919–932, 2011.
- 926 Lischka, S. and Riebesell, U.: Synergistic effects of ocean acidification and warming on  
927 overwintering pteropods in the Arctic, *Global Change Biology*, 18, 3517–3528, 2012.
- 928 Liu, X., Patsavas, M. C., and Byrne, R. H.: Purification and characterization of meta-cresol  
929 purple for spectrophotometric seawater pH measurements, *Environmental Science &*  
930 *Technology*, 45, 4862–4868, 2011.
- 931 Maas, A. E., Blanco-Bercial, L., and Lawson, G. L.: Reexamination of the species assignment of  
932 *Diacavolinia* pteropods using DNA barcoding, *PLoS ONE*, 8, e53889, 2013.
- 933 Maas, A. E., Elder, L. E., Dierssen, H. M., and Seibel, B. A.: Metabolic response of Antarctic  
934 pteropods (Mollusca: Gastropoda) to food deprivation and regional productivity, *MEPS*, 441,  
935 129–139, 2011.
- 936 Maas, A. E., Frazar, S. L., Outram, D. M., Seibel, B. A., and Wishner, K. F.: Fine-scale vertical  
937 distribution of macroplankton and micronekton in the Eastern Tropical North Pacific in  
938 association with an oxygen minimum zone, *Journal of Plankton Research*, 36, 1557–1575,  
939 2014.
- 940 Maas, A. E., Lawson, G. L., and Tarrant, A. M.: Transcriptome-wide analysis of the response of  
941 the thecosome pteropod *Clio pyramidata* to short-term CO<sub>2</sub> exposure, *Comparative*  
942 *Biochemistry and Physiology Part D: Genomics and Proteomics*, 2015. 1–9, 2015.
- 943 Maas, A. E., Wishner, K. F., and Seibel, B. A.: The metabolic response of pteropods to  
944 acidification reflects natural CO<sub>2</sub>-exposure in oxygen minimum zones, *Biogeosciences*, 9,  
945 747–757, 2012a.
- 946 Maas, A. E., Wishner, K. F., and Seibel, B. A.: Metabolic suppression in thecosomatous  
947 pteropods as an effect of low temperature and hypoxia in the Eastern Tropical North, *Marine*  
948 *Biology*, 159, 1955–1967, 2012b.
- 949 Manno, C., Morata, N., and Primicerio, R.: *Limacina retroversa*'s response to combined effects  
950 of ocean acidification and sea water freshening, *Estuarine, Coastal and Shelf Science*, 113,  
951 163–171, 2012.
- 952 Manno, C., [Peck, V. L., and Tarling, G. A.: Pteropod eggs released at high pCO<sub>2</sub> lack resilience](#)  
953 [to ocean acidification, \*Scientific reports\*, 6, 2016.](#)
- 954 [Manno, C., Tirelli, V., Accornero, A., and Fonda Umani, S.: Importance of the contribution of](#)  
955 [Limacina helicina faecal pellets to the carbon pump in Terra Nova Bay \(Antarctica\), \*Journal\*](#)  
956 [of Plankton Research](#), 32, 145–152, 2010.
- 957 Marsh, A. G. and Manahan, D. T.: A method for accurate measurements of the respiration rates  
958 of marine invertebrate embryos and larvae, *Marine Ecology Progress Series*, 184, 1–10, 1999.

959 Mayzaud, P.: Respiration and nitrogen excretion of zooplankton. IV. The influence of starvation  
960 on the metabolism and the biochemical composition of some species, *Marine Biology*, 37,  
961 47-58, 1976.

962 Mehrbach, C., Culberson, C., Hawley, J., and Pytkowicz, R.: Measurement of the apparent  
963 dissociation constants of carbonic acid in seawater at atmospheric pressure, *Limnology and*  
964 *Oceanography*, 18, 897-907, 1973.

965 Melzner, F., Thomsen, J., Koeve, W., Oschlies, A., Gutowska, M. A., Bange, H. W., Hansen, H.  
966 P., and Körtzinger, A.: Future ocean acidification will be amplified by hypoxia in coastal  
967 habitats, *Marine Biology*, 160, 1875-1888, 2013.

968 Millero, F. J.: The marine inorganic carbon cycle, *Chemical Reviews*, 107, 308-341, 2007.

969 Noji, T. T., Bathmann, U. V., Bodungen, B., Voss, M., Antia, A., Krumbholz, M., Klein, B.,  
970 Peeken, I., Noji, C. I. M., and Rey, F.: Clearance of picoplankton-sized particles and  
971 formation of rapidly sinking aggregates by the pteropod, *Limacina retroversa*, *Journal of*  
972 *Plankton Research*, 19, 863-875, 1997.

973 Paulmier, A., Ruiz-Pino, D., and Garçon, V.: CO<sub>2</sub> maximum in the oxygen minimum zone  
974 (OMZ), *Biogeosciences*, 8, 239-252, 2011.

975 Peng, T.-H., Wanninkhof, R., and Feely, R. A.: Increase of anthropogenic CO<sub>2</sub> in the Pacific  
976 Ocean over the last two decades, *Deep Sea Research Part II: Topical Studies in*  
977 *Oceanography*, 50, 3065-3082, 2003.

978 Pierrot, D., Lewis, E., and Wallace, D.: Co2sys DOS Program developed for CO<sub>2</sub> system  
979 calculations, Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory,  
980 US Department of Energy. ORNL/CDIAC-105. , 2006. 2006.

981 Pörtner, H. O.: Oxygen-and capacity-limitation of thermal tolerance: a matrix for integrating  
982 climate-related stressor effects in marine ecosystems, *Journal of Experimental Biology*, 213,  
983 881-893, 2010.

984 Riebesell, U., Zondervan, I., Rost, B., Tortell, P. D., Zeebe, R. E., and Morel, F. M. M.: Reduced  
985 calcification of marine plankton in response to increased atmospheric CO<sub>2</sub>, *Nature*, 407, 364-  
986 367, 2000.

987 Ries, J. B., Cohen, A. L., and McCorkle, D. C.: Marine calcifiers exhibit mixed responses to  
988 CO<sub>2</sub>-induced ocean acidification, *Geology*, 37, 1131-1134, 2009.

989 Rosa, R. and Seibel, B. A.: Synergistic effects of climate-related variables suggest future  
990 physiological impairment in a top oceanic predator, *Proceedings of the National Academy of*  
991 *Sciences*, 105, 20776-20780, 2008.

992 Sabine, C. L., Feely, R. A., Millero, F. J., Dickson, A. G., Langdon, C., Mecking, S., and  
993 Greeley, D.: Decadal changes in Pacific carbon, *J. Geophys. Res. Oceans*, 113, -, 2008.

994 Sabine, C. L. and Tanhua, T.: Estimation of anthropogenic CO<sub>2</sub> inventories in the ocean, *Annu.*  
995 *Rev. Mar. Sci.*, 2, 175-198, 2010.

996 Seibel, B. A.: Critical oxygen levels and metabolic suppression in oceanic oxygen minimum  
997 zones, *Journal of Experimental Biology*, 214, 326-336, 2011.

998 Seibel, B. A. and Drazen, J. C.: The rate of metabolism in marine animals: environmental  
999 constraints, ecological demands and energetic opportunities, *Philos Trans R Soc Lond B Biol*  
1000 *Sci*, 362, 2061-2078, 2007.

1001 Seibel, B. A., Dymowska, A., and Rosenthal, J.: Metabolic temperature compensation and co-  
1002 evolution of locomotory performance in pteropod molluscs, *Integrative and Comparative*  
1003 *Biology*, 47, 880-891, 2007.

1004 Seibel, B. A. and Fabry, V. J.: Marine biotic response to elevated carbon dioxide, *Advances in*  
1005 *Applied Biodiversity Science*, 4, 59-67, 2003.

1006 Seibel, B. A., Maas, A. E., and Dierssen, H. M.: Energetic plasticity underlies a variable  
1007 response to ocean acidification in the pteropod, *Limacina helicina antarctica*, *PLoS ONE*,  
1008 7, e30464, 2012.

1009 Seibel, B. A. and Walsh, P. J.: Potential impacts of CO<sub>2</sub> injection on deep-sea biota, *Science*,  
1010 294, 319-320, 2001.

1011 Sokolova, I. M.: Energy-limited tolerance to stress as a conceptual framework to integrate the  
1012 effects of multiple stressors, *Integrative and Comparative Biology*, 53, 597-608, 2013.

1013 Stumpff, M., Wren, J., Melzner, F., Thorndyke, M., and Dupont, S.: CO<sub>2</sub> induced seawater  
1014 acidification impacts sea urchin larval development I: Elevated metabolic rates decrease  
1015 scope for growth and induce developmental delay, *Comparative Biochemistry and*  
1016 *Physiology-Part A: Molecular & Integrative Physiology*, 160, 331-340, 2011.

1017 Sunday, J. M., Crim, R. N., Harley, C. D. G., and Hart, M. W.: Quantifying rates of evolutionary  
1018 adaptation in response to ocean acidification, *PloS one*, 6, e22881, 2011.

1019 Thabet, A. A., Maas, A. E., Lawson, G. L., and Tarrant, A. M.: Life cycle and early development  
1020 of the thecosomatous pteropod *Limacina retroversa* in the Gulf of Maine, including the effect  
1021 of elevated CO<sub>2</sub> levels, *Marine Biology*, 162, 2235-2249, 2015.

1022 Thomsen, J., Casties, I., Pansch, C., Körtzinger, A., and Melzner, F.: Food availability outweighs  
1023 ocean acidification effects in juvenile *Mytilus edulis*: laboratory and field experiments,  
1024 *Global Change Biology*, 19, 1017-1027, 2013.

1025 van der Spoel, S.: Euthecosomata: A group with remarkable developmental stages (Gastropoda,  
1026 Pteropoda), *Noorduijn en Zoon*, Gorinchem, 1967.

1027 Vaquer-Sunyer, R. and Duarte, C. M.: Thresholds of hypoxia for marine biodiversity,  
1028 *Proceedings of the National Academy of Sciences*, 105, 15452-15457, 2008.

1029 [Waldbusser, G. G., Hales, B., Langdon, C. J., Haley, B. A., Schrader, P., Brunner, E. L., Gray,](#)  
1030 [M. W., Miller, C. A., Gimenez, I., and Hutchinson, G.: Ocean acidification has multiple](#)  
1031 [modes of action on bivalve larvae, \*PloS one\*, 10, e0128376, 2015.](#)

1032 Wang, Z. A., Bienvenu, D. J., Mann, P. J., Hoering, K. A., Poulsen, J. R., Spencer, R. G., and  
1033 Holmes, R. M.: Inorganic carbon speciation and fluxes in the Congo River, *Geophys Res*  
1034 *Lett*, 40, 511-516, 2013.

1035 Wang, Z. A. and Cai, W.-J.: Carbon dioxide degassing and inorganic carbon export from a  
1036 marsh-dominated estuary (the Duplin River): A marsh CO<sub>2</sub> pump, *Limnology and*  
1037 *Oceanography*, 49, 341-354, 2004.

1038 Wanninkhof, R., Doney, S. C., Bullister, J. L., Levine, N. M., Warner, M., and Gruber, N.:  
1039 Detecting anthropogenic CO<sub>2</sub> changes in the interior Atlantic Ocean between 1989 and 2005,  
1040 *Journal of Geophysical Research: Oceans*, 115, 2010.

1041 Widdicombe, S. and Spicer, J. I.: Predicting the impact of ocean acidification on benthic  
1042 biodiversity: What can animal physiology tell us?, *Journal of Experimental Marine Biology*  
1043 *and Ecology*, 366, 187-197, 2008.

1044 Wiebe, P., Morton, A., Bradley, A., Backus, R., Craddock, J., Barber, V., Cowles, T., and Flierl,  
1045 G.: New development in the MOCNESS, an apparatus for sampling zooplankton and  
1046 micronekton, *Marine Biology*, 87, 313-323, 1985.

1047 [Williams, N. L., Feely, R. A., Sabine, C. L., Dickson, A. G., Swift, J. H., Talley, L. D., and](#)  
1048 [Russell, J. L.: Quantifying anthropogenic carbon inventory changes in the Pacific sector of](#)  
1049 [the Southern Ocean, \*Marine Chemistry\*, 174, 147-160, 2015.](#)



1050 Wishner, K. F., Gelfman, C., Gowing, M. M., Outram, D. M., Rapien, M., and Williams, R. L.:  
1051 Vertical zonation and distributions of calanoid copepods through the lower oxycline of the  
1052 Arabian Sea oxygen minimum zone, *Progress in Oceanography*, 78, 163-191, 2008.  
1053

1054

1055 Table 1: Environmental preferences and diel vertical migratory patterns for the species used in  
1056 this study based on previously published data (Bé and Gilmer, 1977; Lalli and Gilmer, 1989).

1057 Data includes published full ranges at which organisms have been found, as well as previous

1058 authors' estimates of the preferred (optimal) ranges of each species, when available. Note that

1059 these are based on relatively sparse observations of broadly distributed species, many of which

1060 may be cryptic congeners, and thus should be treated as estimates.

Species	(optimal) temp (°C)	(optimal), depth (m)	migrator?
<i>Cuvierina atlantica</i>	18 to 26	100-250	possible
<i>Cuvierina pacifica</i>	Only recently established as a separate species, the habits are assumed to be similar to the Atlantic congener.		
<i>Cavolinia inflexa</i>	16 to 28	0-250	no
<i>Clio pyramidata</i>	7 to 27	(0-500), <1500	yes
<i>Limacina helicina</i>	(-2 to 10)	(50-100), <300	possible
<i>Limacina retroversa</i>	(7 to 12)	(20-30), < 150	possible
<i>Styliola subula</i>	(18 to 22)	50-300	yes
<i>Diacria trispinosa</i>	9 to 28	30-200	no

1061

1062 Table 2: The hydrography and location for each station where animals for experiments were  
1063 collected. Each basin was characterized by multiple hydrographic regimes (see text and Fig 2);  
1064 transitions between regimes are denoted by dashed horizontal lines. At stations along the main  
1065 transect the depth (m) at which O<sub>2</sub> decreased below 130 μmol O<sub>2</sub> kg<sup>-1</sup> (~10%) ~~and %~~, the average  
1066 temperature from 25-100 m (°C) and the average salinity from 25-100 m were derived from CTD  
1067 casts. At a few stations (denoted via <sup>a</sup>) in the Atlantic there was warm water at the surface and  
1068 cold fresher water below. The only species in this region, *Limacina retroversa*, has an optimum  
1069 temperature between 7-12 °C (Bigelow, 1924) and was generally found above 50 m (Lawson,  
1070 unpublished data). At these sites the average temperature and salinity is reported first for  
1071 between 25-100 m and then also for 25-50 m to reflect the conditions likely experienced by the  
1072 pteropods. pCO<sub>2</sub> and Ω<sub>Ar</sub> were calculated from measured pH and DIC bottle samples. We  
1073 interpolated linearly the depths (m) at which the pH decreased below 7.7, pCO<sub>2</sub> reached 800  
1074 μatm, and aragonite saturation (Ω<sub>Ar</sub>) reached 1 from the discrete measurements at adjacent  
1075 depths. At stations conducted while in transit to the main study transects (denoted by prefix T)  
1076 the average temperature from 25-100 m (°C) was documented from XBT casts. At these transit  
1077 stations no O<sub>2</sub> or carbonate chemistry data were available (noted with a dash). The species  
1078 caught at each station and used in this study are demarcated with a star (\*).  
1079

Year	Station	Latitude (°N)	Longitude (°W)	average temp 25-100 m	average salinity 25-100 m	depth of 130 $\mu\text{mol O}_2 \text{ kg}^{-1}$	depth of pH 7.7	depth of 800 $\mu\text{atm}$	depth of $\Omega_{\text{Ar}} = 1$	<i>C. atlantica</i>	<i>C. pacifica</i>	<i>C. inflexa</i>	<i>C. pyramidata</i>	<i>L. helicina</i>	<i>L. retroversa</i>	<i>S. subula</i>	<i>D. trispinosa</i>
2011 Atlantic	32	49.1	-44.3	5.3, 9.0	<u>34.4</u>	NA	74.1	NA	NA						*		
	31	50.0	-42.0	14	<u>35.8</u>	NA	385.4	NA	NA								*
	30	49.6	-41.9	14.1	<u>35.8</u>	NA	452.8	NA	NA	*							*
	26	47.5	-42.0	13.3	<u>35.2</u>	NA	644.9	NA	NA	*			*				
	24	46.5	-42.0	14.5	<u>35.5</u>	NA	453.9	NA	NA	*			*				
	21	44.9	-42.0	16.5	<u>36.2</u>	NA	501.1	NA	NA				*				*
	19	44.0	-44.9	4.9, 11.2	<u>33.4</u>	NA	181.0	NA	NA						*		
	17	43.0	-47.8	1.8, 8.1	<u>33.2</u>	NA	143.1	NA	NA						*		
	13	40.9	-52.0	20.7	<u>36.5</u>	NA	756.7	NA	NA	*		*					*
	10	47.5	-52.0	19.4	<u>35.9</u>	NA	466.9	NA	NA	*		*					*
	8	38.5	-52.0	22.8	<u>36.5</u>	NA	805.7	NA	NA	*		*					*
	3	36.0	-52.0	21.4	<u>36.6</u>	NA	937.7	NA	NA	*							
	2012 Pacific	T2	45.6	-128.5	-	::	-	-	-	-				*	*		
T3		46.6	-133.5	-	::	-	-	-	-					*			
T4		47.7	-138.5	6.4	::	-	-	-	-				*				
T5		45.7	-129.8	10.0	::	-	-	-	-					*			
T6		46.6	-134.9	9.5	::	-	-	-	-					*			
T7		47.6	-140.2	8.6	::	-	-	-	-				*				
3		49.0	-148.2	6.2	<u>32.7</u>	209	128.9	193.7	168.5								
6		47.5	-145.6	7.1	<u>32.7</u>	235	108.3	199.2	159.1				*	*			
7		47.0	-144.6	7.8	<u>32.7</u>	256	131.0	214.0	185.1				*				
15		43.1	-138.1	10.9	<u>32.9</u>	363	199.5	368.2	334.8				*				
18		41.5	-135.8	13.7	<u>33.0</u>	340	147.3	331.7	380.6				*				
21		39.9	-135.0	12.7	<u>33.1</u>	348	162.0	332.2	302.8		*						
24		38.6	-135.0	14.7	<u>33.3</u>	402	222.8	411.8	372.7		*		*				
30		35.6	-135.0	16.2	<u>33.3</u>	349	200.7	437.8	425.1		*	*	*				
32		34.4	-135.1	16.5	<u>33.3</u>	348	202.9	439.2	432.0		*	*	*				
34	33.6	-135.0	17.4	<u>34.0</u>	368	233.3	370.1	352.4			*	*			*		
T9	33.7	-133.6	17.0	::	-	-	-	-		*	*	*					
T10	33.8	-133.2	15.9	::	-	-	-	-		*	*	*					

Table 3: Carbonate chemistry during manipulation experiments. The manipulation experiments were conducted at multiple temperatures (T.) and salinities (S.) based on the conditions the organisms were caught in. As described in more detail in the text, DIC measurements were made of water drawn from the control chambers while TA was measured for batches of experimental water (denoted as xpt. TA). In situ TA (i.s. TA), based on nearby CTD bottle sampling at the surface, is also shown. At test stations conducted while in transit to/from the main study regions, where bottle samples of in situ TA were unavailable, underway pCO<sub>2</sub> values and the LC/HO DIC were used to calculate in situ TA (denoted with \*). In some instances, measurements of experimental TA differed by >20 μmol kg<sup>-1</sup> from nearby in situ measurements of surface TA. This difference greatly exceeds expected variability based on measurement uncertainty and spatial (geographic and vertical) offsets in the locations of experimental water collection relative to the nearest CTD cast; in these circumstances, the experimental TA was likely erroneous due to sampling errors/issues (e.g., contamination). For completeness, and to aid in identification of erroneous experimental TA values, calculations of carbonate chemistry parameters, including aragonite saturation state ( $\Omega_{Ar}$ ) and pCO<sub>2</sub> were made based on DIC and both experimental TA and in situ TA. In further data analysis and interpretation, calculations based on experimental TA are given preference except those few instances where experimental TA differed from in situ by >20 μmol kg<sup>-1</sup> (bold denotes preferred calculations). Calculated saturation state and pCO<sub>2</sub> are reported as the average and standard deviation per batch of water. Note that the LC/LO gas tank in 2011 (in italics) appears to have been improperly mixed by the manufacturer as calculations suggested it contained a much lower CO<sub>2</sub> level than the intended 380 μatm; it should consequently be considered an entirely separate treatment from the 2011 LC/HO (were/where CO<sub>2</sub> levels were based on bubbling with an ambient air line).

	Treatment	T. °C	S.	i.s. TA ( $\mu\text{mol kg}^{-1}$ )	xpt. TA ( $\mu\text{mol kg}^{-1}$ )	DIC ( $\mu\text{mol kg}^{-1}$ )	i.s. $\Omega\text{Ar}$	i.s. pCO <sub>2</sub> ( $\mu\text{atm}$ )	xpt. $\Omega\text{Ar}$	xpt. pCO <sub>2</sub> ( $\mu\text{atm}$ )	
2011	380 $\mu\text{atm CO}_2$ /	10	33	2300.3	<b>2307.3</b>	2094.4	2.3 ± 0.2	336.2 ± 37.7	<b>2.4 ± 0.2</b>	<b>324.8 ± 35.8</b>	
Atlantic	21% O <sub>2</sub>	15	33	2300.3	<b>2307.3</b>	2066.5	2.6 ± 0.7	404.5 ± 172.7	<b>2.7 ± 0.7</b>	<b>390.8 ± 164.5</b>	
		15	35	<b>2296.4</b>	2354.5	2066.4	<b>2.5 ± 0.1</b>	<b>382.3 ± 20.4</b>	3.1 ± 0.1	297.7 ± 14.3	
		20	34	2353.4*	<b>2345.8</b>	2028.6	3.6 ± 0.2	302.8 ± 31.6	<b>3.5 ± 0.2</b>	<b>311.6 ± 32.9</b>	
		20	34	2366.0	<b>2367.2</b>	2077.5	3.3 ± 0.1	363.1 ± 23.2	<b>3.3 ± 0.1</b>	<b>361.4 ± 23.1</b>	
		10	33	2300.3	<b>2307.3</b>	1919.7	4.0	139.0	<b>4.1</b>	<b>135.5</b>	
	380 $\mu\text{atm CO}_2$ / 10% O <sub>2</sub>	15	33	2300.3	<b>2307.3</b>	1774.8	5.5 ± 0.6	101.2 ± 23.9	<b>5.6 ± 0.6</b>	<b>99.0 ± 23.3</b>	
		15	35	<b>2296.4</b>	2354.5	1852.7	<b>4.6</b>	<b>139.2</b>	5.3	116.1	
		800 $\mu\text{atm CO}_2$ / 21% O <sub>2</sub>	10	33	2300.3	<b>2307.3</b>	2219.7	1.2 ± 0.2	779.9 ± 114.0	<b>1.2 ± 0.2</b>	<b>742.4 ± 106.8</b>
		15	33	2300.3	<b>2307.3</b>	2208.0	1.3	908.7	<b>1.4</b>	<b>867.8</b>	
		15	35	<b>2296.4</b>	2354.5	2139.5	<b>1.9</b>	<b>585.2</b>	2.4	434.4	
800 $\mu\text{atm CO}_2$ / 10% O <sub>2</sub>	20	34	2353.4*	<b>2345.8</b>	2176.9	2.1 ± 0.1	651.8 ± 23.4	<b>2.1 ± 0.1</b>	<b>678.2 ± 24.8</b>		
	20	34	2366.0	<b>2367.2</b>	2212.7	1.9 ± 0.4	786.0 ± 196.0	<b>1.9 ± 0.4</b>	<b>780.9 ± 194.2</b>		
	15	33	2300.3	<b>2307.3</b>	2186.2	1.5 ± 0.2	788.7 ± 157.6	<b>1.5 ± 0.2</b>	<b>754.9 ± 148.3</b>		
	15	35	<b>2296.4</b>	2354.5	2179.6	<b>1.5 ± 0.3</b>	<b>782.9 ± 164.6</b>	2.0 ± 0.3	558.2 ± 103.9		
	2012	380 $\mu\text{atm CO}_2$ /	10	32.1	2151.9*	<b>2142.8</b>	1934.8	2.2 ± 0.1	285.2 ± 21.4	<b>2.3 ± 0.1</b>	<b>283.0 ± 21.2</b>
	Pacific	21% O <sub>2</sub>	10	33.5	2208.0	<b>2222.7</b>	2001.9	2.4 ± 0.6	302.2 ± 100.9	<b>2.4 ± 0.6</b>	<b>303.3 ± 101.4</b>
			15	32.5	<b>2182.6*</b>	2095.7	1983.4	<b>2.2 ± 0.0</b>	<b>388.1 ± 5.5</b>	1.4 ± 0.0	646.7 ± 11.5
15			33.5	2208.0	<b>2222.7</b>	2020.8	2.3 ± 0.2	407.7 ± 52.1	<b>2.3 ± 0.2</b>	<b>409.1 ± 52.4</b>	
380 $\mu\text{atm CO}_2$ / 10% O <sub>2</sub>		10	32.5	<b>2182.6*</b>	2095.7	1973.9	<b>2.3 ± 0.1</b>	<b>295.5 ± 20.0</b>	1.4 ± 0.1	489.2 ± 41.2	
		15	33.5	2208.0	<b>2222.7</b>	2017.5	2.3	3956.0	<b>2.3</b>	<b>397.4</b>	
800 $\mu\text{atm CO}_2$ / 21% O <sub>2</sub>		10	32.1	2151.9*	<b>2142.8</b>	2026.3	1.4 ± 0.1	525.0 ± 35.0	<b>1.4 ± 0.1</b>	<b>519.7 ± 34.5</b>	
		10	33.5	2208.0	<b>2222.7</b>	2120.6	1.3	628.2	<b>1.3</b>	<b>631.2</b>	
		15	32.5	<b>2182.6*</b>	2095.7	2031.7	<b>1.8 ± 0.1</b>	<b>527.6 ± 50.9</b>	1.0 ± 0.1	952.4 ± 115.1	
		15	33.5	2208.0	<b>2222.7</b>	2112.2	1.4 ± 0.2	736.0 ± 96.0	<b>1.4 ± 0.2</b>	<b>739.4 ± 96.6</b>	
800 $\mu\text{atm CO}_2$ / 10% O <sub>2</sub>		10	32.5	<b>2182.6*</b>	2095.7	2066.5	<b>1.4 ± 0.1</b>	<b>545.5 ± 65.1</b>	0.8 ± 0.1	1056.0 ± 151.6	
	15	33.5	2208.0	<b>2222.7</b>	2118.3	1.4	762.4	<b>1.4</b>	<b>766.0</b>		

Table 4: The average wet mass (mass; g) and mass-specific oxygen consumption rate (MO<sub>2</sub>; μmol O<sub>2</sub> g<sup>-1</sup> h<sup>-1</sup>) ± the standard error (SE) for each treatment (Treat.) and species. The ~~number~~ numbers of individuals/replicates (N) per treatment are reported and the species are arranged by temperature (Temp; °C) as well as the year and basin of collection.

Year	Temp.	Species	Treat.	N	mass	±SE	MO <sub>2</sub>	±SE	
2011 Atlantic	10	<i>Limacina retroversa</i>	LC/HO	12	.00281	0.00037	10.33	1.17	
			HC/HO	13	.00284	0.00031	10.10	0.56	
			LC/LO	9	.00274	0.00026	8.12	0.66	
			HC/LO	9	.00377	0.00053	4.21	0.55	
	15	<i>Clio pyramidata</i>	LC/HO	10	.01944	0.00408	7.81	0.71	
			HC/HO	8	.01410	0.00435	8.55	1.48	
			LC/LO	9	.02363	0.00867	6.63	1.21	
			HC/LO	8	.03945	0.00467	6.99	0.45	
		<i>Cuvierina atlantica</i>	LC/HO	8	.04493	0.00264	5.05	0.63	
			LC/LO	10	.04636	0.00252	3.25	0.28	
			HC/LO	10	.05040	0.00219	4.29	0.37	
			<i>Diacria trispinosa</i>	LC/HO	8	.03718	0.00316	4.44	0.56
	20	<i>Cuvierina atlantica</i>	LC/HO	9	.01876	0.00396	4.31	0.85	
			HC/HO	9	.01683	0.00284	4.53	1.13	
		<i>Cavolinia inflexa</i>	LC/HO	8	.00626	0.00104	14.30	1.48	
			HC/HO	4	.00508	0.00049	13.81	1.39	
<i>Styliola subula</i>		LC/HO	10	.00400	0.00038	13.96	1.80		
		HC/HO	8	.00289	0.00035	15.95	0.87		
2012 Pacific		10	<i>Limacina helicina</i>	LC/HO	7	.00140	0.00026	5.26	1.17
				HC/HO	8	.00149	0.00021	5.51	0.69
	LC/LO			6	.00300	0.00058	4.91	0.69	
	HC/LO			10	.00296	0.00038	7.18	1.45	
	<i>Clio pyramidata</i>		LC/HO	9	.02646	0.00258	5.43	0.45	
			HC/HO	8	.02355	0.00369	4.39	0.60	
			LC/LO	14	.01459	0.00185	5.58	0.81	
			HC/LO	12	.01250	0.00245	5.72	1.14	
	15	<i>Cuvierina pacifica</i>	LC/HO	4	.01829	0.00563	3.41	0.56	
			HC/HO	7	.02130	0.00636	3.53	0.57	
		<i>Cavolinia inflexa</i>	LC/HO	5	.01330	0.00062	3.53	0.44	
			HC/HO	8	.01556	0.00149	3.34	0.41	
			LC/LO	4	.01405	0.00185	2.41	0.33	
			HC/LO	2	.01855		3.98		
		<i>Styliola subula</i>	LC/HO	6	.00360	0.00044	5.30	1.20	
			HC/HO	4	.00220	0.00029	7.73	2.14	
<i>Clio pyramidata</i>	LC/HO	4	.03020	0.0037	3.82	0.66			
	HC/HO	5	.02904	0.00329	3.21	0.27			

Table 5: Statistical results of the univariate general linear models (GLM) for each species were analyzed separately by year and are listed by relative to the temperature of the experiment (Temp.; °C). For species studied at multiple temperatures (denoted by \*), the metabolic rates were adjusted to 15°C using a  $Q_{10} = 2$  to allow for direct comparison. The effect of the independent factors of CO<sub>2</sub> level (CO<sub>2</sub>), O<sub>2</sub> level (O<sub>2</sub>), their interactive effect (Int.) and the covariate of mass were analyzed in regards to the metabolic rate and reported as *p*-values for the Pacific (mean mass specific metabolic rate values found in Table 4). For the Atlantic, each treatment was tested as independent (Treat.) due to the accidentally low CO<sub>2</sub> condition in the LC/LO gas mixture. We report whether the data met the assumption of normality of the residuals with Shapiro-Wilk (norm.; for p under 0.05 the assumption is not met) and heterogeneity of variance (var.; for p under 0.05 the assumption is not met) and denote in bold where the dataset did not fully meet these statistical assumptions. Note that for the sole case where the treatment or CO<sub>2</sub> effect was significant (*L. retroversa*) all assumptions were met.

Year	Temp.	Species	Effect on metabolic rate						
			CO <sub>2</sub>	O <sub>2</sub>	Int.	Treat.	Mass	norm.	var.
2011 Atlantic	10	<i>Limacina retroversa</i>				<0.001	<0.001	0.542	0.522
	15	<i>Clio pyramidata</i>				0.295	<0.001	0.079	0.263
		<i>Cuvierina atlantica</i> *				0.174	<0.001	0.972	< <b>0.001</b>
		<i>Diacria trispinosa</i>	.731				<0.001	0.802	0.885
		<i>Cavolinia inflexa</i>	.677				.008	0.498	0.876
		<i>Styliola subula</i>	.791				.040	.922	<b>0.014</b>
2012 Pacific	10	<i>Limacina helicina</i>	.464	.323	.914		.007	<b>0.045</b>	<b>0.026</b>
	15	<i>Clio pyramidata</i> *	.255	.156	.726		.018	< <b>0.001</b>	0.068
		<i>Cuvierina pacifica</i>	.709				<0.001	0.639	0.357
		<i>Cavolinia inflexa</i>	.309	.717	.219		.113	0.581	0.28
		<i>Styliola subula</i>	.763				.668	0.353	0.325



## Figure legends

**Figure 1: Cruise tracks and animal sampling.** Thecosomes were collected during the night at stations along the main survey transect (solid line) and at stations during transit (dashed line) during cruises to the northwest Atlantic in 2011 and northeast Pacific in 2012. The shapes correspond to the species caught at each station and used in this study. Blue (10 °C), grey (15 °C) and red (20 °C) boxes around the station numbers (#) correspond to the temperature that was representative of 25-100 m at each station (Table 2) and used in the experiments with animals from that station.

**Figure 2: Hydrography of sampling regions.** Hydrographic profiles of stations representative of the specific water mass types from the northern (P-T5, P-6, A-26), middle (P-18, A-19) and southern (P-32, A-8) portions of the Pacific (P) and Atlantic (A) study transects (station locations: Fig. 1). At station P-T5, the temperature profile (grey) was from an XBT cast because no CTDs were conducted during transits. For all stations along the main transects, left-hand plots show temperature (grey), salinity (black) and oxygen (black dotted) measured via sensors on the CTD and binned to 1 m depth intervals. Middle plots show TA (black) and DIC (grey) ~~from~~ from discrete bottle samples (dots show depths of bottle samples). Right-hand plots show pCO<sub>2</sub> (black) and aragonite saturation state ( $\Omega_{Ar}$ ; grey) calculated based on TA and DIC measurements.

**Figure 3: Thecosome respirometry.** Mean metabolic rate and standard error ( $\mu\text{mol O}_2 \text{ g}^{-1} \text{ h}^{-1}$ ) of thecosomes exposed to low (i.e., ambient) CO<sub>2</sub> and normal levels of O<sub>2</sub> (light blue; LC/HO), high CO<sub>2</sub> and normal O<sub>2</sub> levels (dark blue; HC/HO), low CO<sub>2</sub> and low O<sub>2</sub> (light red; LC/LO), or high CO<sub>2</sub> and low O<sub>2</sub> (dark red; HC/LO). The species and temperature of the experiment are reported below the x-axis. Significance is reported based on a basin, species, and temperature specific GLM which tested for the effect of treatment on O<sub>2</sub> consumption with a Bonferroni post-hoc analysis: (Table 5). In the Atlantic analysis each treatment was tested independently, while in the Pacific CO<sub>2</sub> and O<sub>2</sub> were treated as factors. For each species and temperature, treatments are reported as non-significant (N.S.) or, in the case of significance, by letters that indicate which treatments are statistically similar (same letter) or different (different letter) at a p-value < 0.05.

Note that for *C. atlantica* the metabolic rates of individuals respired at 20° C were converted to 15°C using a temperature coefficient of 2 (see methods) for this GLM analysis.

**Figure 4:** Log transformed metabolic rates ( $\mu\text{mol O}_2 \text{ h}^{-1}$ ) for *L. retroversa* at 10 °C, not normalized to mass, plotted against the log transformed wet mass (mg) of individuals exposed to low CO<sub>2</sub> and normal levels of O<sub>2</sub> (black circles; LC/HO), high CO<sub>2</sub> and normal O<sub>2</sub> levels (dark grey diamonds; HC/HO), low CO<sub>2</sub> and low O<sub>2</sub> (white circles; LC/LO), or high CO<sub>2</sub> and low O<sub>2</sub> (light grey diamonds; HC/LO).