

## Response Letter

Michael Bahn  
Co-Editor-in-Chief  
Biogeosciences

Dear Dr. Bahn,

Thank you very much for offering us the chance to revise our manuscript “Initial shifts in nitrogen impact on ecosystem carbon fluxes in an alpine meadow: patterns and causes” (bg-2016-436). We have carefully considered the valuable comments and suggestions from you and the reviewer. The manuscript has been revised accordingly. Here are our detailed responses to the reviews. Please note that the comments from the reviewers are in *italics* followed by our responses in **bold** text.

Sincerely,

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## Reply to Report #1

*I reviewed this paper previously, but am not satisfied with the responses of the authors. The following points that I raised previously were not incorporated in an appropriate way.*

**Response: We feel very sorry that the reviewer was not satisfied with our revision. In this version, we have tried our best to carefully study the reviewer's suggestions and revised the manuscript accordingly. The specific comments are addressed below. Hope the reviewer is satisfied with our revision this time.**

*1) The partitioning issue is mentioned (last lines of p14), but in a way that doesn't clearly indicate how it may have influenced the results. I have the feeling that the authors did not really understand my suggestion that the response of  $R_{mic}$  from these collars that exclude roots indicates the N response of  $R_{mic}$  in the absence of plant C inputs. This could be used as an advantage (mechanistic insight into microbial responses in the absence of plant effects) but instead authors only defend their approach by saying that it has been used in many other studies.*

**Response: Thank the reviewer for the thoughtful comments. We have added some discussions about the limitations of the approach we used in the Discussion (Page 14, Line 20- Page 15, Line 4).**

**"This approach cuts roots and excludes effects of changes in plant C allocation on microbial respiration. Soil moisture content may also change in the deep collars, which likely affects microbial respiration. However, this method is at an advantage to explore mechanisms of microbial responses in the absence of plant effects, which is a common and useful technique to partition the components of ER and widely used in previous studies (Wan et al., 2005; Zhou et al., 2007)"**

*On the other hand, potential artifacts should also be acknowledged. I wonder if the negative relationship between  $R_{mic}$  and N addition in year 2 may be related to the increase of  $R_{mic}$  with N addition in year 1. Is it possible that microbes changed from being N limited in year 1 to being C limited in year 2 - something that may not be the case in the presence of plant C inputs.*

**Response: The reviewer is right. The results may be influenced by the approach we used. However, we do not think  $R_{mic}$  is C limited in the second year. First, both the control treatment and the N addition treatment excluded plant C inputs, the negative relationship between  $R_{mic}$  and N addition could be driven by other factors besides C inputs. Second, plant biomass increased under N addition treatments in year 2, so the supply of C was not decreasing.**

*2) More importantly: the authors' statement that pH is driving  $R_{mic}$  responses is not well supported. Previously, authors had not tested any other variable, and just used the relationship between  $R_{mic}$  and pH to argue that pH was driving the  $R_{mic}$  response. In the revised version, they made 2 extra correlations:  $R_{mic}$  vs  $NH_4$  and  $R_{mic}$  vs  $NO_3$ .*

*Correlation coefficients were 0.77 for pH effect, 0.68 for NH<sub>4</sub> effect and 0.76 for NO<sub>3</sub> effect. This minor difference in correlation coefficients is then used to support that pH was most important, but in my view, it is not obviously better than NO<sub>3</sub> (and I think NO<sub>3</sub> + NH<sub>4</sub> should have been tested too – but in a better approach). More decent statistics (e.g. stepwise regression analysis) would be needed to separate the pH effect from the NO<sub>3</sub> effect, and if these cannot be distinguished, it should at least be shown and discussed*

**Response: We greatly appreciate the reviewer’s thoughtful comments. We used stepwise regression analysis to re-analyze the data and found the same result. Only soil pH was left in the model. Thus, the result of stepwise regression analysis confirmed that soil pH was the most important factor driving changes in R<sub>mic</sub>.**

*3) I asked to use AIC instead of R<sup>2</sup> to compare goodness of fit for quadratic vs linear functions. While authors indicate the AIC results in the response letter, they do not use it in the manuscript but use again R<sup>2</sup>. Even if the results are the same, this needs to be corrected.*

**Response: Thanks for the valuable suggestions. We have added the AIC results as supplementary material in the revised paper.**

*4) I asked to improve the comparison with other experiments by considering the fact that initial nutrient availability may have been very different, and responses likely differ in nutrient-poor vs nutrient-rich systems. They did not do this, and in the response letter only show that they measured NH<sub>4</sub> and NO<sub>3</sub> and that it increased with N addition. This is not what I asked for. When comparing the N response of different ecosystems, the original N status should be considered. An infertile system likely responds very differently than a fertile system. If the fertility of other systems (to which the current study is compared) is unknown, authors should at least mention the relevance of it.*

**Response: Thank the reviewer very much for the thoughtful suggestions. We have added some discussions about the initial nutrient availability in the Discussion (Page 13, Line 9-10).**

**“The differences could also relate to the initial nutrient availability in different sites. Ecosystems with high N availability may reach N saturation at low rate of N addition, if there are no other limited factors.”**

*In addition, also language editing is still needed. Even though authors say the manuscript has been checked by a native English speaker, I found several grammatical errors (e.g. missing articles in several places, just like in the previous version )*

**Response: Thanks for the suggestion. We have carefully checked the language throughout the manuscript.**

# Initial shifts in nitrogen impact on ecosystem carbon fluxes in an alpine meadow: patterns and causes

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## Abstract

Increases in nitrogen (N) deposition can greatly stimulate ecosystem net carbon (C) sequestration through positive N-induced effects on plant productivity. However, how net ecosystem CO<sub>2</sub> exchange (NEE) and its components respond to different N addition rates remains unclear. Using a N addition gradient experiment (six levels: 0, 2, 4, 8, 16, 32 gN m<sup>-2</sup> year<sup>-1</sup>) in an alpine meadow on the Qinghai-Tibetan Plateau, we explored the responses of different ecosystem C fluxes to a N addition gradient and revealed mechanisms underlying the dynamic responses. Results showed that NEE, ecosystem respiration (ER), and gross ecosystem production (GEP) all increased linearly with N addition rates in the first year of treatment, but shifted to N saturation responses in the second year with the highest NEE ( $-7.77 \pm 0.48$   $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) occurring under N addition rate of 8 gN m<sup>-2</sup> year<sup>-1</sup>. The saturation responses of NEE and GEP were caused by N-induced accumulation of standing litter, which limited light availability for plant growth, under high N addition. The saturation response of ER was mainly due to the N-induced saturation response of aboveground plant respiration and decreasing soil microbial respiration along the N addition gradient, while decreases in soil microbial respiration under high N addition were caused by N-induced

reductions in soil pH. We also found that various components of ER, including aboveground plant respiration, soil respiration, root respiration, and microbial respiration, responded differentially to the N addition gradient. These results reveal temporal dynamics of N impacts and the rapid shift of ecosystem C fluxes from N limitation to N saturation. Our findings bring evidence of short-term initial shifts of responses of ecosystem C fluxes to increases in N deposition, which should be considered when predicting long-term changes in ecosystem net C sequestration.

## 1 Introduction

Anthropogenic reactive nitrogen (N) inputs to the terrestrial biosphere has increased more than threefold over the past century and is predicted to increase further (Lamarque et al., 2005; Galloway et al., 2008). Because of the strong coupling of ecosystem carbon (C) and N cycles, excess N deposition could have significant impacts on ecosystem C cycle (LeBauer and Treseder, 2008; Liu and Greaver, 2010; Lu et al., 2011). Ecosystem net C sequestration is predicted to increase or have no significant change under rising N deposition (Nadelhoffer et al., 1999; Magnani et al., 2007; Reay et al., 2008; Niu et al. 2010; Lu et al., 2011; Fernandez-Martinez et al., 2014). However, we have limited understanding on the dynamic N responses of C sequestration in terrestrial ecosystems, which is crucial for model projection of the future terrestrial C cycle under rising N deposition (Reay et al., 2008).

Although N addition generally enhances plant growth and ecosystem net primary productivity (NPP) based on global syntheses of N addition experiments (LeBauer and Treseder, 2008; Xia and Wan, 2008; Lu et al., 2011), the responses of ecosystem C fluxes vary with N loading rates (Liu and Greaver, 2010; Lu et al., 2011). According to the N saturation hypothesis, NPP is assumed to slowly increase with N

addition rates first, then get at the maximum value at N saturation point and finally decline with further increase of N input (Aber et al., 1989; Lovett and Goodale, 2011). During this process, NPP shifts from a N limited, a N intermediate, to a N saturation stage as N deposition increases. Similarly, ~~the response of~~ net ecosystem CO<sub>2</sub> exchange (NEE) and its components of gross ecosystem production (GEP) and ecosystem respiration (ER) may also respond nonlinearly to increasing N loading rates (Fleischer et al., 2013; Gomez-Casanovas et al., 2016; Tian et al., 2016). In the N limited stage, low rates of N addition could stimulate ecosystem productivity (Aber et al., 1989), GEP (Fleischer et al., 2013; Gomez-Casanovas et al., 2016), and ER (Hasselquist et al., 2012; Zhu et al., 2016), while in the N saturation stage, high doses of N addition could have negative effects on GEP and ER (Treseder, 2008; Janssens et al., 2010; Maaroufi et al., 2015). The unbalanced responses of GEP and ER may lead to changes in NEE.

Moreover, ER can be divided into aboveground plant respiration, belowground plant respiration (root respiration), and soil microbial respiration. These components of ER could be affected by plant aboveground biomass, root biomass, soil organic matter and microbial biomass C, which may respond variously to N addition (Phillips and Fahey, 2007; Hasselquist et al., 2012). For example, root respiration would be enhanced or not significantly changed under N addition, while soil microbial respiration may be suppressed by N addition (Zhou et al., 2014). The different responses of various components of ER to N addition will also consequently change the response of NEE. Nevertheless, there is limited knowledge on how various components of NEE respond differentially to N addition gradient. In addition, the N responses of ecosystem C fluxes may shift ~~with-over~~ time because of changes in plant community structure and other limiting factors (Niu et al., 2010). It is not clear when and how ecosystem C fluxes get N saturated under increasing N input. The mechanisms underlying the saturation response of C fluxes

are even far from clear, which hinders us from accurately predicting the C cycle in response to rising N deposition.

In this study, we explored the responses of various ecosystem C cycle processes to a N addition gradient in an alpine meadow on the Qinghai-Tibetan Plateau. The Qinghai-Tibetan Plateau has an area of 2.5 million km<sup>2</sup> with alpine meadow covering 35 % of this area, and it is sensitive to environmental change and human activities (Chen et al., 2013). The objectives of this study were to explore how different components of ecosystem C fluxes respond to increasing N loading gradient. Specifically, we addressed the following questions: (i) how do NEE and its components respond to the N addition gradient in the alpine meadow? (ii) whether various C cycle processes can get N saturated? If so, at which N addition level they are saturated and how do the responses shift with time? and (iii) what are the mechanisms underlying N saturation responses of different C cycle processes?

## 2 Materials and methods

### 2.1 Study site

The study site is located in an alpine meadow in Hongyuan County, Sichuan Province, China, which is on the eastern Qinghai-Tibetan Plateau (32°48' N, 102°33' E). The altitude is ~3500 m. Long-term (1961-2013) mean annual precipitation is 747 mm with approximately 80 % occurring in May to September. Long-term mean annual temperature is 1.5 °C with monthly mean temperature ranging from -9.7 °C in January to 11.1 °C in July. The dominant species in this alpine meadow are *Deschampsia caespitosa* (Linn.) Beauv., *Kobresia setchwanensis* Hand. -Mazz., *Carex schneideri* Nelmes, and *Anemone rivularis* Buch.-Ham.. The vegetation cover of this grassland is over 90 %. The soil in the study site is classified

as Mat Cry-gelic Cambisol according to the Chinese classification, with surface soil bulk density being  $0.89 \text{ g cm}^{-3}$ . The soil organic C content and total N content are  $37 \text{ gC kg}^{-1}$  and  $3.5 \text{ gN kg}^{-1}$ , respectively. The background N deposition is ranging from  $0.87$  to  $1.38 \text{ gN m}^{-2} \text{ year}^{-1}$  on the eastern Qinghai-Tibetan Plateau, and the natural N deposition rate in China is ranging from  $0.11$  to  $6.35 \text{ gN m}^{-2} \text{ year}^{-1}$  (Lü and Tian, 2007).

## 2.2 Experimental design

We conducted a N addition experiment with six levels of N addition rate ( $0, 2, 4, 8, 16, 32 \text{ gN m}^{-2} \text{ year}^{-1}$ ) in early 2014. The six N treatments were represented by N0 (control), N2, N4, N8, N16, and N32, respectively. The treatments were randomly assigned with five replications, so there were totally 30 plots. Each plot was  $8 \times 8 \text{ m}$ , and the distance between any two adjacent plots was  $3 \text{ m}$ . The N addition treatments started from May, 2014. In 2014 and 2015, N was applied by hand as  $\text{NH}_4\text{NO}_3$  ( $>99\%$ ) every month from May to September (i.e. during the growing season) before rainfall. The N amount was same in each month. In order to distribute dry  $\text{NH}_4\text{NO}_3$  evenly in the plots, we mixed dry  $\text{NH}_4\text{NO}_3$  with enough amounts of soil to apply.

## 2.3 Ecosystem C cycle properties and soil pH measurement

Ecosystem C fluxes were measured using a transparent static chamber ( $0.5 \times 0.5 \times 0.5 \text{ m}$ ) attached to an infrared gas analyzer (LI-6400XT; LI-COR Environmental, Lincoln, Nebraska, USA) in the field. During each measurement, the chamber was positioned over a square steel frame, which was permanently inserted into soil and offered a flat base for the chamber. Inside the chamber, two electric fans were



mounted in order to mix the chamber atmosphere. The measurements were conducted twice per month on clear, sunny days from May to September in 2014 and 2015. Nine consecutive recordings of CO<sub>2</sub> concentration were taken on each base at 10-second intervals. CO<sub>2</sub> flux rates were determined from the time-courses of the concentrations to calculate net ecosystem CO<sub>2</sub> exchange (NEE). After the measurement of NEE, the chamber was covered by an opaque cloth and the CO<sub>2</sub> measurement was repeated. As the second measurement eliminated light, the CO<sub>2</sub> flux value obtained represented ecosystem respiration (ER). Gross ecosystem production (GEP) was calculated as the difference between NEE and ER. Negative or positive NEE and GEP values represent net C uptake or release, respectively. The detailed methods have also been described in Niu et al. (2008) and Niu et al. (2013).

Soil respiration (SR) was assessed following the measurement of NEE and ER. It was also measured with LI-6400XT attaching a soil CO<sub>2</sub> flux chamber (991 cm<sup>3</sup> in total volume; LI-6400-09; LI-COR Environmental, Lincoln, Nebraska, USA). A PVC collar (10.5 cm in diameter and 5 cm in height) was permanently installed 2-3 cm into the soil. The soil respiration chamber attached to LI-6400XT was placed on each PVC collar for 1-2 minutes to measure SR. Living plants inside the collars were removed regularly by hand to eliminate aboveground plant respiration. Soil heterotrophic respiration (i.e. soil microbial respiration, R<sub>mic</sub>) was measured using the same method as soil respiration. Differently, the PVC collar was 40 cm in height and installed 36-38 cm into the soil. As >90 % of plant roots were distributing in the topsoil (0-20 cm), 40-cm-long PVC collars could cut off old plant roots and prevented new roots from growing inside the collars. Plants in the collars were completely removed by hand to exclude C supply. The experiment was conducted in early 2014 and the measurements of CO<sub>2</sub> fluxes above these 40-cm-long PVC collars began in late July in 2014, leaving enough time for the remaining plant roots

inside the collars to die. Thus CO<sub>2</sub> fluxes in those deep collars represented R<sub>mic</sub>. The method was same to Wan et al. (2005) and Zhou et al. (2007). Root respiration (R<sub>root</sub>) was calculated by value of SR minus R<sub>mic</sub>. Aboveground plant respiration (R<sub>above</sub>) was calculated by ER minus SR, and ecosystem plant respiration (R<sub>plant</sub>) was calculated as the difference between ER and R<sub>mic</sub>. All the measurements of ecosystem C fluxes were simultaneous.

Soil samples were collected from the topsoil (0-10 cm) of the 30 plots on August 15, 2014 and August 14, 2015. Two soil cores (8 cm in diameter and 10 cm in depth) were taken at least 1 m from the edge in each plot, and then completely mixed to get a composite sample. The soil samples were sieved by a 2 mm mesh and then were air-dried for chemical analysis. Soil pH was determined with a glass electrode in a 1:2.5 soil:water solution (w/v).

## 2.4 Statistical analysis

Repeated-measures ANOVA (RMANOVA) was used to examine N addition effects on each ecosystem C flux over the growing season in 2014 and 2015. When we evaluate N addition effects on the different components of ER and their proportions, we averaged their values across the year and then used one-way ANOVA to test the differences among treatments. To test the response pattern of ecosystem C cycle properties to the N addition gradient, we fitted the response parameter to linear or quadratic functions which had higher R<sup>2</sup>. [We also compared AIC between the functions \(Table S1\)](#). Simple linear regression analyses were used to evaluate relationships of ER with its components and NEE across the two years.  $\Delta R_{mic}$  and  $\Delta pH$  were calculated by data in different N addition treatments minus data in the control treatment. All data were tested for normal distribution before statistical analysis. The a posteriori

comparisons were performed by DUNCAN test, and the effects were considered to be significantly different if  $P < 0.05$ . All statistical analyses were conducted with SAS V.8.1 software (SAS Institute Inc., Cary, North Carolina, USA).

## 5 3 Results

### 3.1 NEE and its components in response to the N addition gradient

Net ecosystem CO<sub>2</sub> exchange (NEE) varied throughout the growing seasons in both 2014 and 2015. The maximum rates of net CO<sub>2</sub> uptake (indicated by large negative values of NEE) occurred in July in both years (Fig. 1a,d). N addition had a significant impact on NEE in 2014 ( $P=0.020$ ) and a marginally significant effect in 2015 ( $P=0.059$ ) (Table 1). Mean NEE across months had different responses to the N addition gradient between the two years (Fig. 1a,d). It increased linearly with N addition rates in 2014 (Fig. 2a), but shifted to a saturating response with N addition rates in 2015 (Fig. 2d). The largest NEE was  $-7.77 \pm 0.48 \mu\text{mol m}^{-2} \text{s}^{-1}$  under  $8 \text{ gN m}^{-2} \text{ year}^{-1}$  addition rate (N8) in 2015.

The N addition gradient had significant effects on ER ( $P=0.033$  and  $0.006$ , respectively) and GEP ( $P=0.002$  and  $0.038$ , respectively) in both 2014 and 2015 (Table 1). Similar to NEE, both ER and GEP showed linear responses to N addition rates in 2014 but shifted to saturation responses in 2015 (Fig. 2). On average, ER was enhanced by 0.9-16.1 % in 2014 and 7.9-23.7 % in 2015 under different N addition treatments. GEP was increased by 2.4-19.2 % in 2014 and 6.7-20.5 % in 2015 under different N addition levels, with maximal values being  $-24.40 \pm 0.48 \mu\text{mol m}^{-2} \text{s}^{-1}$  under  $32 \text{ gN m}^{-2} \text{ year}^{-1}$  in 2014 and  $-15.38 \pm 0.72 \mu\text{mol m}^{-2} \text{s}^{-1}$  under  $16 \text{ gN m}^{-2} \text{ year}^{-1}$  in 2015 (Fig. 2).

### 3.2 Components of ecosystem respiration in response to the N addition gradient

We divided ER into aboveground plant respiration ( $R_{\text{above}}$ ), soil respiration (SR), root respiration ( $R_{\text{root}}$ ), and microbial respiration ( $R_{\text{mic}}$ ), and found that different ER components showed diverse responses to the N addition gradient. Mean SR across months was not significantly changed by the N addition gradient in 2014 (Table 1; Fig. 3). However, in 2015, it ranged from  $4.98 \pm 0.33 \mu\text{mol m}^{-2} \text{s}^{-1}$  to  $6.23 \pm 0.23 \mu\text{mol m}^{-2} \text{s}^{-1}$  under different N addition levels, with significant reduction under high N addition levels of 16 and 32  $\text{gN m}^{-2} \text{year}^{-1}$  ( $P=0.010$ ; Fig. 3). Additionally, the relationship between SR and N addition rates was not significant in 2014 (Fig. 3a), while SR leveled off under high N addition rates in 2015 (Fig. 3c). Interestingly,  $R_{\text{mic}}$  increased linearly with N addition rates in 2014 (Fig. 3b), while it decreased with N addition rates in 2015 (Fig. 3d). Comparing among various components of ER, only  $R_{\text{mic}}$  showed distinctively inverse responses to N addition rates between years.  $R_{\text{above}}$  increased with increasing N addition rates in 2014 (Fig. 4b) but got the maximum value at N16 in 2015 (Fig. 4e). By contrast,  $R_{\text{root}}$  decreased with increasing N addition rates in 2014 (Fig. 4c), while it had no statistically significant response to the N addition gradient in 2015 (Fig. 4f).

In addition, the proportions of different efflux components to ER differed in response to the N addition gradient between years (Fig. 5). The proportions of  $R_{\text{above}}$  to ER kept increasing with N addition rates in 2014 but got saturated at N16 in 2015 (Fig. 5a,d). The proportions of  $R_{\text{root}}$  to ER ranged from  $31.90 \pm 6.69 \%$  in N0 plots to  $11.18 \pm 1.28 \%$  in N32 plots in 2014 (Fig. 5b), but was not significantly different among N addition levels in 2015 (Table 1; Fig. 5e). In 2014, the contributions of  $R_{\text{mic}}$  to ER did not significantly change under N addition treatments (Table 1; Fig. 5c), whereas they declined along the N addition gradient in 2015 (Fig. 5f).

### 3.3 Causes for the N saturation responses of ecosystem C fluxes

In order to examine the causes for the N saturation responses of NEE and ER in 2015, we examined the relationship between ER and its various components and also NEE. The results showed that ER had significantly positive correlation with  $R_{\text{above}}$  (Fig. 6a) but not with  $R_{\text{root}}$  (Fig. 6b), and had significantly negative correlation with  $R_{\text{mic}}$  (Fig. 6c). Moreover, NEE closely correlated with ER (Fig. 6d). We further explored the causes for decreasing  $R_{\text{mic}}$  with N addition in 2015 and found that N addition significantly reduced soil pH in 2015 (Fig. 7a). N-induced reduction in soil microbial respiration ( $\Delta R_{\text{mic}}$ ) was positively dependent on N-induced reduction in soil pH ( $\Delta \text{pH}$ ) in 2015 (Fig. 7b), but not in 2014.

10

## 4 Discussion

### 4.1 Nitrogen saturation responses of ecosystem C fluxes and the causes

Our results showed that initial ecosystem C fluxes (NEE, ER, and GEP) in 2014 suggested ecosystem N limitation, whereas in 2015 these C fluxes clearly suggested N saturation under high N addition rates.

15 These findings not only extend the N saturation hypothesis for the response of NPP to N addition (Aber et al., 1998; Aber et al., 1989; Lovett and Goodale, 2011), but also provide comprehensive evidence of potential relationships between various ecosystem C fluxes and ecosystem N dynamics. Previous N addition studies used only one level of N addition and found that NEE showed a positive (Niu et al., 2010; Huff et al., 2015) or no significant response (Harpole et al., 2007; Bubier et al., 2007) to N addition. Using  
20 one level of N addition only might not be enough to capture or quantify complex ecosystem responses to N addition. By using a N addition gradient experiment, this study comprehensively showed the saturation

responses of NEE and its components to different N loading rates.

The N saturation response of NEE in 2015 was mainly attributed to the saturation responses of ER and GEP (Fig. 2), while the N saturation response of ER was likely caused by the saturation response of aboveground plant respiration and decreasing soil microbial respiration along the N addition gradient.

5 The decrease of aboveground plant respiration under N32 treatment was primarily due to that N addition stimulated plant growth and thus standing litter accumulation after plant senescence (Fig. S1-S2). In 2014, plant aboveground biomass (AGB) was stimulated under high N addition treatment, especially AGB of grasses (Fig. S2). In this grassland, grasses usually have higher height than other plants. The accumulation of grasses standing litter under N32 treatment limited light condition for other plants and negatively  
10 influenced plant growth in the early growing season in 2015. Therefore, GEP and NEE did not keep increasing at the highest N addition rate, leading to N saturation response. The N-induced light limitation for plant growth was also observed in other ecosystems, like temperate grassland (Niu et al., 2010; Kim and Henry, 2013). Moreover, our results showed that most components of ER had similar response patterns between the two years except soil microbial respiration that increased in 2014 but decreased in  
15 2015 along with N addition rates. Thus, we propose that soil microbial respiration might play a key role in mediating the N saturation effects for ER and thus NEE, which is not reported in previous studies. The decline of microbial respiration under high N addition conditions was primarily due to the N-induced reduction in soil pH (Fig. 7). Although many factors can influence soil microbial respiration, such as soil N availability and microbial community structure (Janssens et al., 2010), previous studies with similar N  
20 addition gradient suggested that soil pH was the most important driver for responses of microbes under high N addition rates (Liu et al., 2014; Song et al., 2014; Chen et al., 2016). N addition can lead to soil

acidification and bring negative impacts on soil microbial growth and activities (Liu et al., 2014; Tian et al., 2016). In this study, the decreased soil pH may cause toxicity effects on microbial activity (Treseder, 2008; Zhou et al., 2012) and thus reduces microbial respiration after two years of N addition.

#### 5 **4.2 The time and N threshold for the saturation responses**

Our findings demonstrate that N responses of ecosystem C fluxes shifted from linear response to saturation response over the two years of treatments (Fig. 2). A recent study revealed that ecosystem C fluxes exhibited saturating responses to N addition during two consecutive measurement years in a temperate grassland (Tian et al., 2016). However, their measurement was conducted after ten years of N addition treatments (similar N addition rates with our study), so it did not capture the early response signals of ecosystem C exchange. Results of another N addition gradient experiment carried out in three marsh ecosystems showed that aboveground plant biomass increased linearly with N addition rates after seven months of treatment, but showed saturating responses after 14 months of N addition (Vivanco et al., 2015). Taken together with our results, it suggests that N saturation of ecosystem C fluxes might happen within couple years of N input. The different responses between years in this study are not likely due to climate differences, because temperature and precipitation were not significantly different between 2014 and 2015. We acknowledge that our findings are just based on the short-term study, while long-time experiment may capture more robust patterns on N saturation and the underlying mechanisms, but the findings of the initial shift of N responses are helpful to better understand the dynamics of ecosystem in response to external N input.

The N saturation threshold for ecosystem C fluxes of this alpine meadow is approximately  $8 \text{ gN m}^{-2}$

<sup>2</sup> year<sup>-1</sup>. This level is much higher than that in an alpine steppe on the Qinghai-Tibetan Plateau (Liu et al., 2013). In Liu et al.'s study, biomass N concentration, soil N<sub>2</sub>O flux, N-uptake efficiency and N-use efficiency showed saturating responses at N addition rate of 4 gN m<sup>-2</sup> year<sup>-1</sup>. The discrepancy is probably caused by different precipitation at the two sites. The precipitation is 747 mm in our study site and is 415 mm in their study site. The lower precipitation may constrain ecosystem's response to N addition in Liu et al. (2013). Likewise, the N saturation load in our alpine meadow is higher than that in an alpine dry meadow in Colorado (Bowman et al., 2006) and is comparable with that in a temperate steppe of Eurasian grasslands which found a saturation N addition rate of approximately 10.5 gN m<sup>-2</sup> year<sup>-1</sup> (Bai et al., 2010). The differences could also relate to the initial nutrient availability in different sites. Ecosystems with high N availability may reach N saturation at low rate of N addition, if there are no other limited factors. The higher saturation levels indicate that this alpine meadow is more limited by N comparing with other resources. Furthermore, the N critical load for causing changes in ecosystem C cycle processes is around 2 gN m<sup>-2</sup> year<sup>-1</sup> in this alpine meadow. In the first year, ecosystem C exchanges were not significantly different between N0 and N2 treatments, but C fluxes were greater in N2 plots than that in N0 plots in the second year (Fig. 1). This threshold for triggering changes in ecosystem C fluxes is comparable to that in another alpine meadow on the mid-south of the Tibetan Plateau (Zong et al., 2016). Considering that atmospheric wet N deposition is ranging from 0.87 to 1.38 gN m<sup>-2</sup> year<sup>-1</sup> on the eastern Qinghai-Tibetan Plateau (Lü and Tian, 2007), our estimate on N critical load suggests that ecosystem C cycle may be largely affected under future N deposition in the alpine meadow of Qinghai-Tibetan Plateau.

#### **4.3 Diverse responses of C flux components to the N addition gradient**



The components of ER showed diverse responses to the N addition gradient (Fig. 4,5). For example, in 2014, aboveground plant respiration and its proportion to ER increased, but belowground plant respiration and its proportion to ER decreased with N addition amounts (Fig. 4b,c, Fig.5a,b). To our knowledge, no previous study examined different components of ER in response to N addition gradient. Some studies  
5 conducted in alpine grassland demonstrated that N addition had no significant effects on ER (Jiang et al., 2013; Gong et al., 2014), since aboveground biomass did not respond to N addition in their studies. In this study, comparing to the control treatment (without N addition), greater plant growth and aboveground biomass under N addition enhanced aboveground plant respiration and thus stimulated ER. The lack of N effect on soil respiration (SR) in 2014 may be attributed to the counteractive responses of soil microbial  
10 respiration and root respiration to N addition. In the first year, N addition ameliorated the nutrient limitation for microbes, thus soil microbial activity and biomass increased in short term (Treseder, 2008) and subsequently stimulated microbial respiration (Peng et al., 2011). On the other hand, N addition could reduce belowground biomass allocation (Haynes and Gower, 1995), leading to decrease in root respiration. The increase of soil microbial respiration partly offsets the decrease of root respiration. As a result, SR  
15 had no significant difference among N treatments in the first year. However, in the second year, soil microbial respiration declined under high N addition levels, in combination with the low root respiration, resulting in decreases of SR under N16 and N32 treatments. This decrease in SR was also observed in other ecosystems under long-term or high levels of N addition (Yan et al., 2010; Zhou and Zhang, 2014; Maaroufi et al., 2015). We are fully aware that there are some limitations for the partitioning technique,  
20 by which we used deep versus shallow collars to partition root from microbial respiration. This approach cuts roots and excludes effects of changes in plant C allocation on microbial respiration. Soil moisture

content may also change in the deep collars, which likely affects microbial respiration. However, microbial respiration responses are not solely plant-mediated (Janssens et al., 2010). This method is at an advantage to explore mechanisms of microbial responses in the absence of plant effects, which we used is ~~still~~ a common and useful technique to partition the components of ER, ~~which is~~ and widely used in previous studies (Wan et al., 2005; Zhou et al., 2007).

## 5 Conclusions

Based on a field N addition gradient experiment, this study tested N saturation theory against multiple C cycle processes. We found that ecosystem C fluxes of NEE, GEP, and ER shifted from linear responses to saturation responses over two years of N addition. The saturation responses of NEE and ER were mainly caused by the N-induced saturation response of aboveground plant respiration and decreasing soil microbial respiration along the N addition gradient. N-induced reduction in soil pH was the main mechanism underlying declines in microbial respiration under high N addition. We also revealed that various components of ER, including aboveground plant respiration, soil respiration, root respiration, and microbial respiration, responded differentially to the N addition gradient. The findings suggest that the C cycle processes have differential responses to N addition between aboveground and belowground plant parts, and between plants and microbes. Our findings provide experimental evidences for the dynamic N responses of ecosystem C cycle, which is helpful for parameterizing biogeochemical models and guiding ecosystem management in light of future increasing N deposition.

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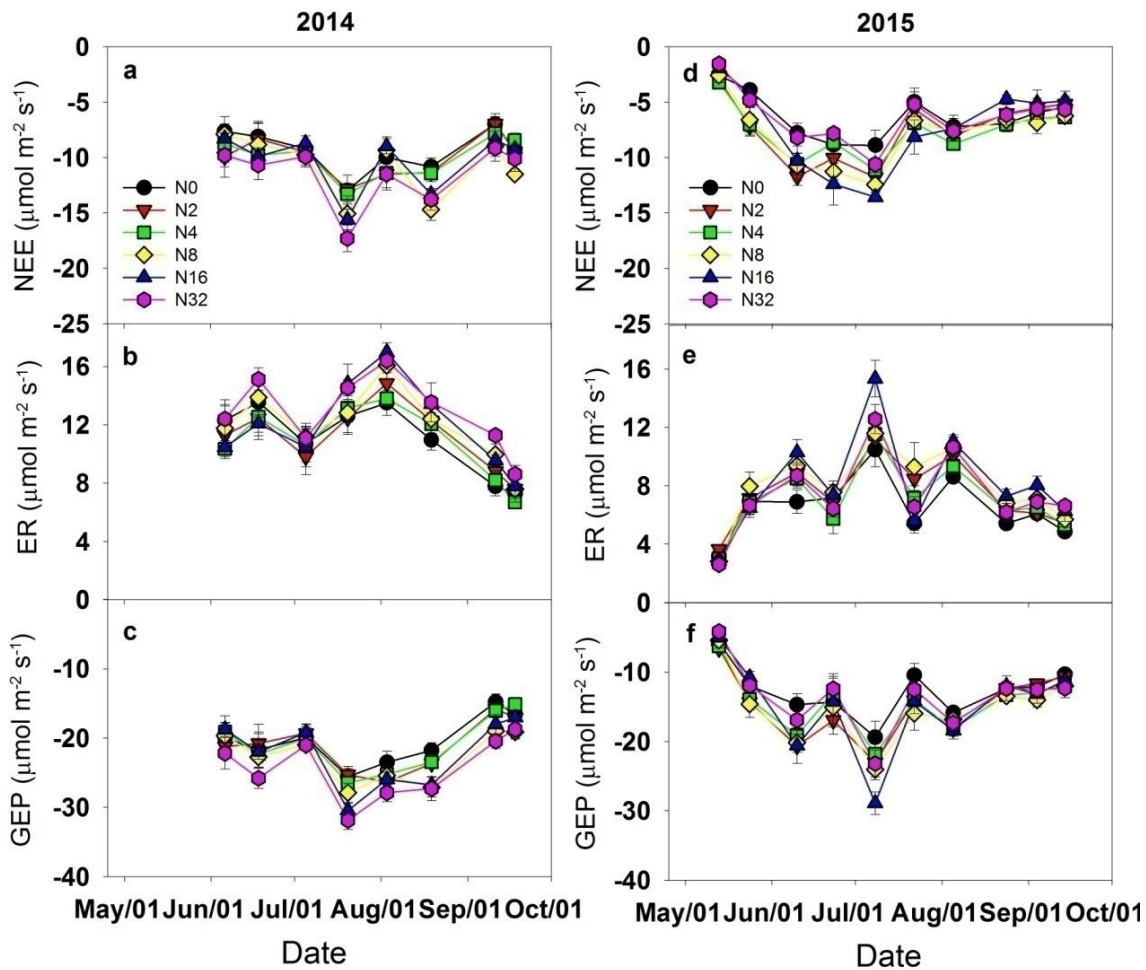
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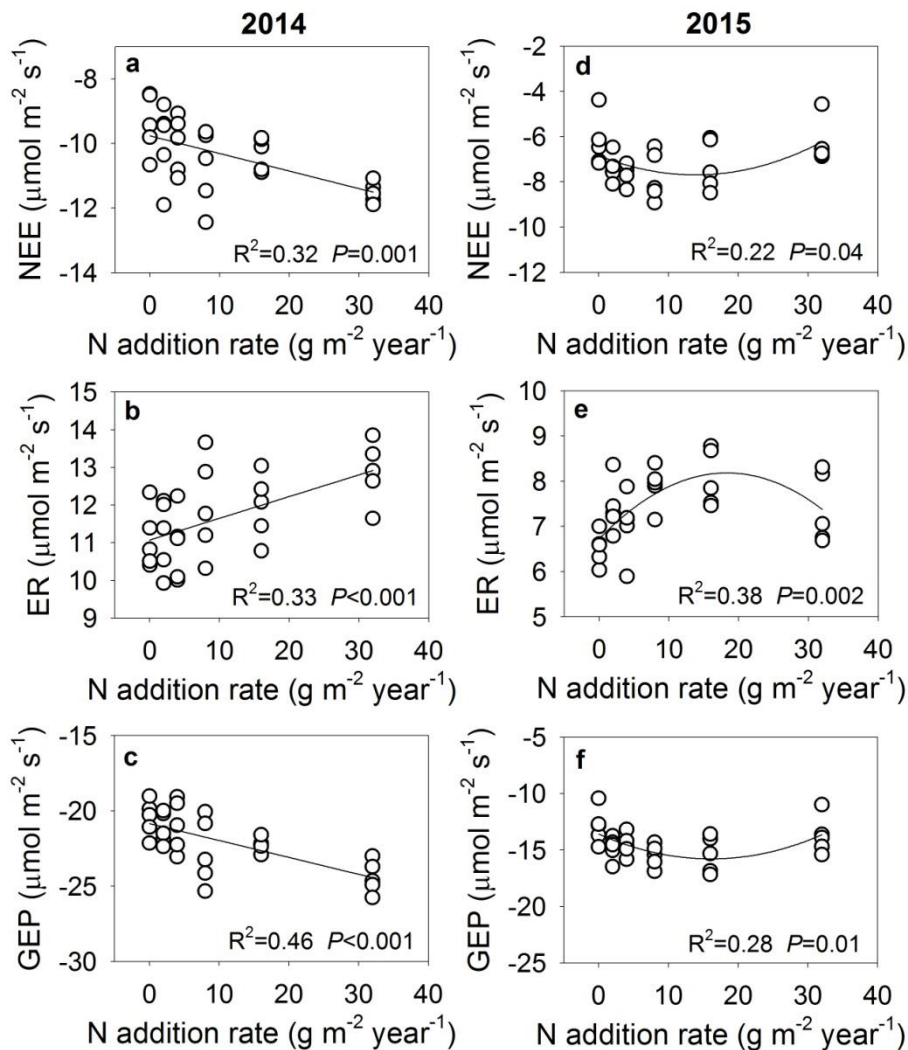
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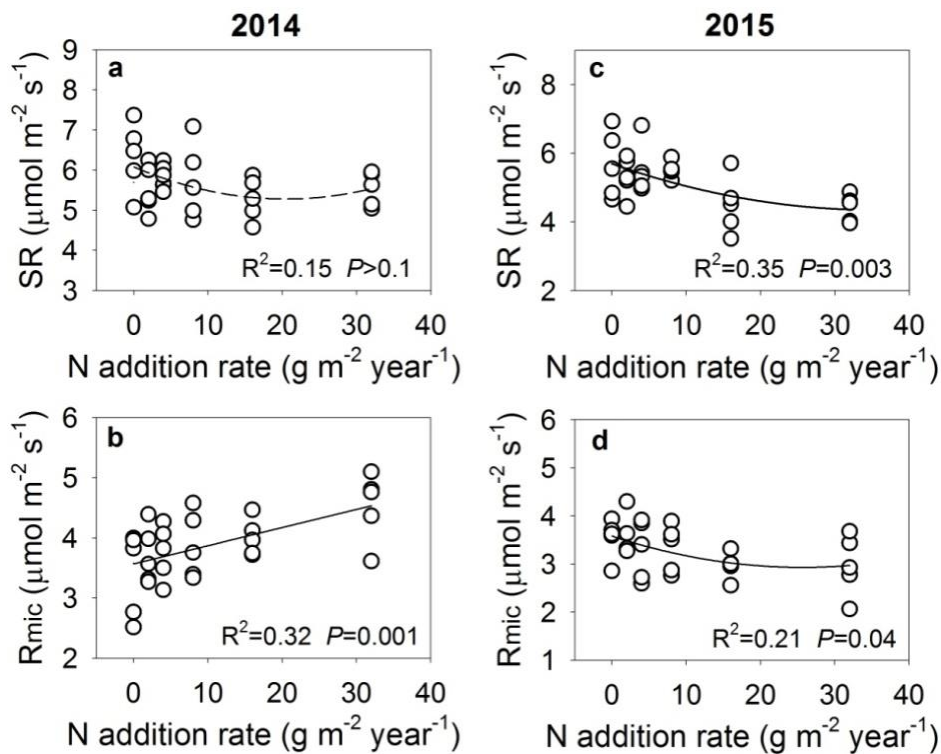
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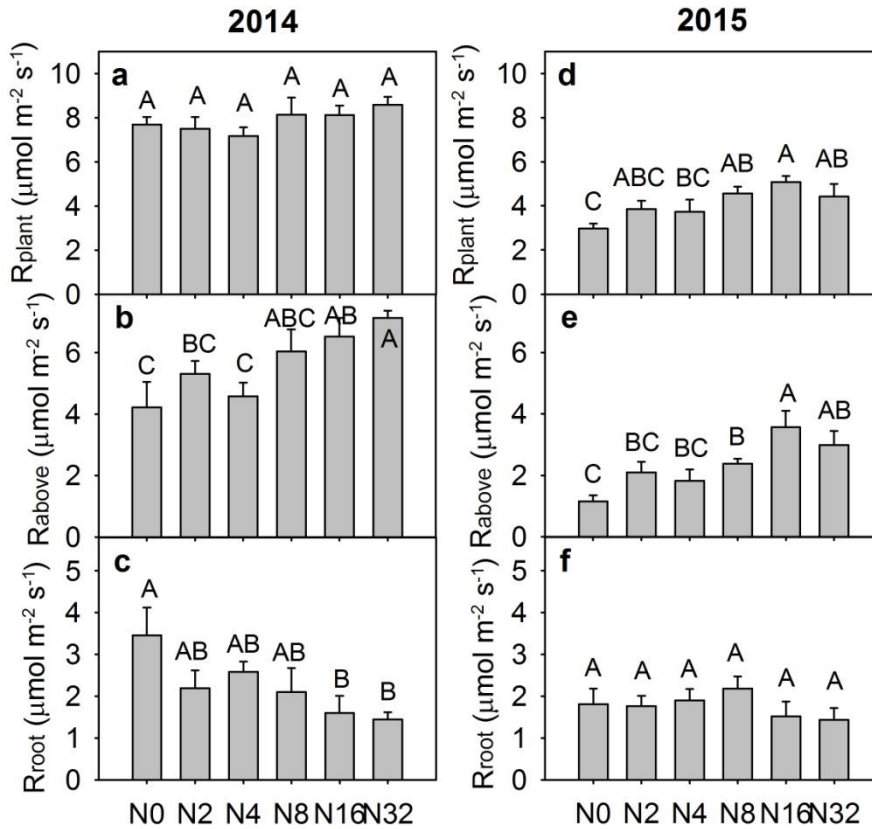
**Figure 1. Seasonal dynamics of net ecosystem CO<sub>2</sub> exchange (NEE) (a, d), ecosystem respiration (ER) (b, e), and gross ecosystem production (GEP) (c, f) in 2014 and 2015. N0, N2, N4, N8, N16, N32 represent N addition rate of 0, 2, 4, 8, 16, 32 gN m<sup>-2</sup> year<sup>-1</sup>, respectively.**



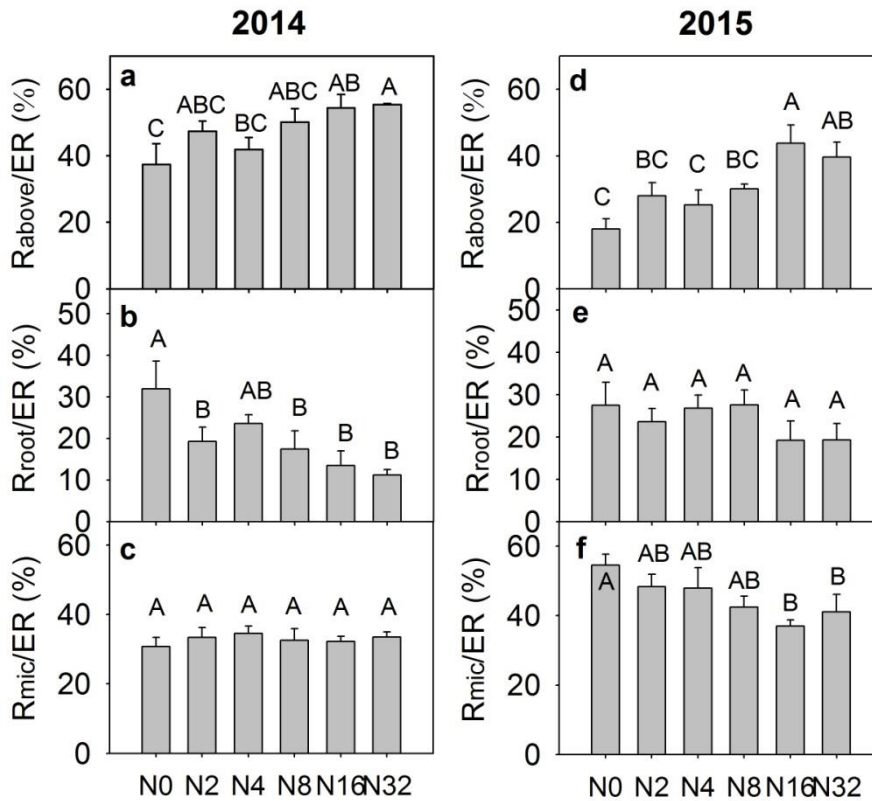
**Figure 2. Relationships between N addition rate and net ecosystem CO<sub>2</sub> exchange (NEE) (a, d), ecosystem respiration (ER) (b, e), and gross ecosystem production (GEP) (c, f) in 2014 and 2015.**



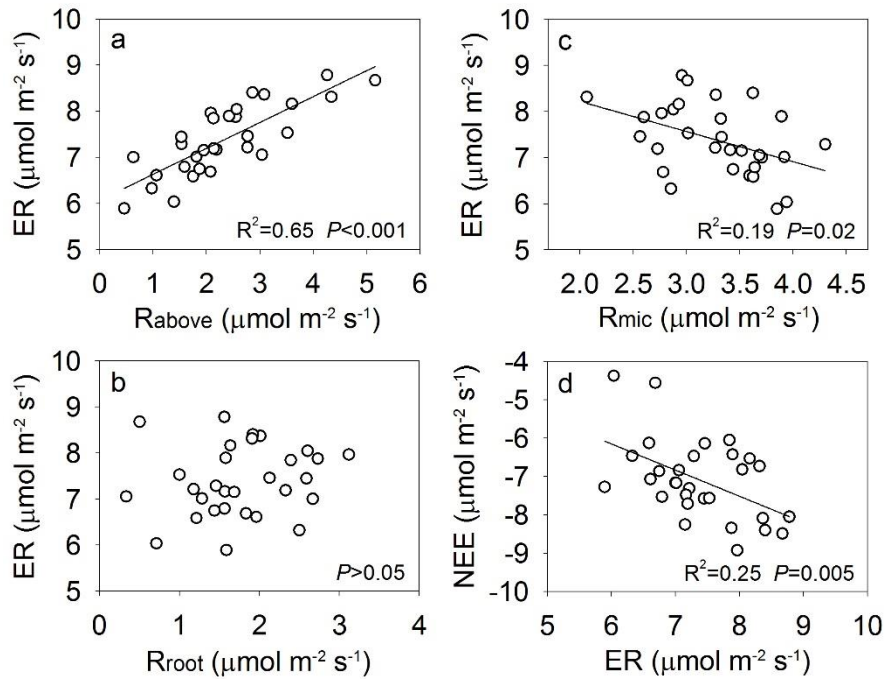
**Figure 3. Relationships between N addition rate and soil respiration (SR) (a, c), and soil microbial respiration ( $R_{\text{mic}}$ ) (b, d) in 2014 and 2015.**



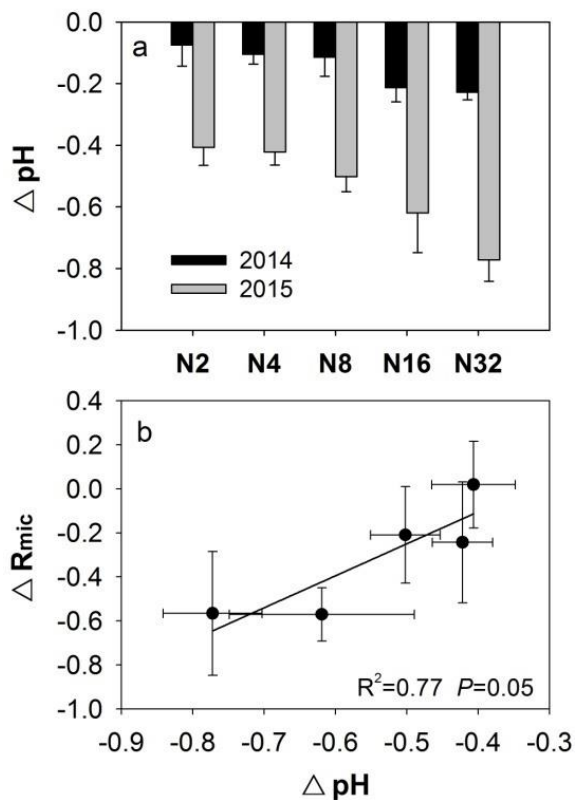
**Figure 4. Plant respiration and its components in response to the N addition gradient in 2014 and 2015 (mean  $\pm$ SE, n = 5).  $R_{\text{plant}}$ : plant respiration (a, d),  $R_{\text{above}}$ : aboveground plant respiration (b, e),  $R_{\text{root}}$ : plant root respiration (c, f). N0, N2, N4, N8, N16, N32 represent N addition rate of 0, 2, 4, 8, 16, 32 gN m<sup>-2</sup> year<sup>-1</sup>, respectively.**



**Figure 5.** The contributions of different source components to ecosystem respiration (ER) in response to the N addition gradient in 2014 and 2015 (mean  $\pm$  SE, n = 5). **R<sub>above</sub>**: aboveground plant respiration, **R<sub>root</sub>**: plant root respiration, **R<sub>mic</sub>**: soil microbial respiration. N0, N2, N4, N8, N16, N32 represent N addition rate of 0, 2, 4, 8, 16, 32 gN m<sup>-2</sup> year<sup>-1</sup>, respectively.



**Figure 6. Relationships between aboveground plant respiration ( $R_{\text{above}}$ ), root respiration ( $R_{\text{root}}$ ), soil microbial respiration ( $R_{\text{mic}}$ ) and ecosystem respiration (ER) (a,b,c), ER and net ecosystem  $\text{CO}_2$  exchange (NEE) (d) across all plots in 2015.**



**Figure 7. N-induced changes in soil pH ( $\Delta$ pH) (a) (mean  $\pm$  SE, n = 5) and the dependence of N-induced changes in soil microbial respiration ( $\Delta$ R<sub>mic</sub>) on N-induced changes in soil pH ( $\Delta$ pH) in 2015 (b). N2, N4, N8, N16, N32 represent N addition rate of 2, 4, 8, 16, 32 gN m<sup>-2</sup> year<sup>-1</sup>, respectively.**



**Table 1. Results (F and P values) of one-way ANOVA on the effects of nitrogen addition on ecosystem C fluxes in 2014 and 2015. NEE: net ecosystem CO<sub>2</sub> exchange, ER: ecosystem respiration, GEP: gross ecosystem production, SR: soil respiration, R<sub>mic</sub>: soil microbial respiration, R<sub>plant</sub>: plant respiration, R<sub>above</sub>: aboveground plant respiration, R<sub>root</sub>: plant root respiration.**

df	NEE		ER		GEP		SR		R <sub>mic</sub>		
	F	P	F	P	F	P	F	P	F	P	
2014	5	3.35	0.020	2.95	0.033	5.37	0.002	1.56	0.209	1.49	0.246
2015	5	2.50	0.059	4.35	0.006	2.83	0.038	3.94	0.010	1.40	0.259

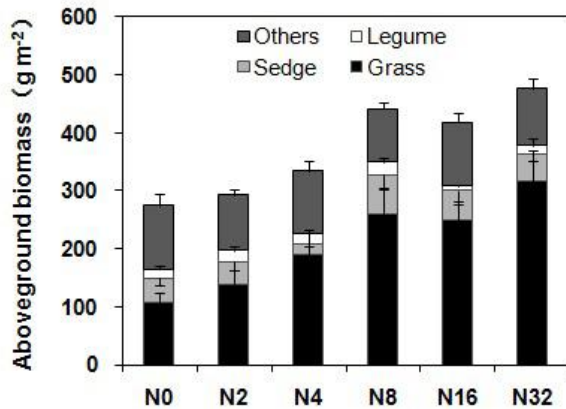
  

df	R <sub>plant</sub>		R <sub>above</sub>		R <sub>root</sub>		R <sub>above</sub> /ER		R <sub>root</sub> /ER		R <sub>mic</sub> /ER		
	F	P	F	P	F	P	F	F	F	P	F	P	
2014	5	1.06	0.409	3.84	0.011	2.64	0.049	3.08	0.027	3.56	0.015	0.28	0.919
2015	5	3.25	0.022	5.38	0.002	0.78	0.573	5.54	0.002	0.97	0.456	2.46	0.062

# Initial shifts in nitrogen impact on ecosystem carbon fluxes in an alpine meadow: patterns and causes

Bing Song, Jian Sun, Qingping Zhou, Ning Zong, Linghao Li, Shuli Niu

## Supplementary Material



**Figure S1.** Plant aboveground biomass in response to the N addition gradient in 2014. N0, N2, N4, N8, N16, N32 represent N addition rate is 0, 2, 4, 8, 16, 32 gN m<sup>-2</sup> year<sup>-1</sup>, respectively.



**Figure S2.** N-induced accumulation of standing litter in high N addition plots (photos were taken on June 13).

**Table S1** Comparisons of Akaike information criterion (AIC) among functions describing the relationships between NEE, ER, GEP, SR and  $R_{mic}$  (Y) and N addition rate (X). Quadratic functions work better than linear ones for ecosystem C fluxes in 2015. NEE: net ecosystem CO<sub>2</sub> exchange, ER: ecosystem respiration, GEP: gross ecosystem production, SR: soil respiration,  $R_{mic}$ : soil microbial respiration.

	<u>Functions in 2014</u>		<u>Functions in 2015</u>	
	<u>Linear<sup>1</sup></u>	<u>Quadratic<sup>2</sup></u>	<u>Linear<sup>1</sup></u>	<u>Quadratic<sup>2</sup></u>
<u>NEE</u>	<u>78.39</u>	<u>80.26</u>	<u>88.69</u>	<u>84.82</u>
<u>ER</u>	<u>71.68</u>	<u>73.48</u>	<u>90.12</u>	<u>82.30</u>
<u>GEP</u>	<u>77.96</u>	<u>79.86</u>	<u>87.69</u>	<u>77.68</u>
<u>SR</u>	<u>87.88</u>	<u>87.34</u>	<u>79.43</u>	<u>78.18</u>
<u>R<sub>mic</sub></u>	<u>78.33</u>	<u>80.27</u>	<u>85.15</u>	<u>84.48</u>

1) linear model:  $Y = b_1 + b_2 \times X$       2) quadratic model:  $Y = b_1 + b_2 \times X + b_3 \times X^2$