1 **Small phytoplankton contribution to the total primary production in**

2 **the Amundsen Sea**

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Abstract

Small-sized phytoplankton is anticipated to be more important for phytoplankton community in a recent changing ocean condition. However, little information on the contribution of small-sized phytoplankton to overall phytoplankton production is currently available in the Amundsen Sea. To determine the contributions of small-sized phytoplankton to total biomass and primary production, carbon and nitrogen uptake rates of total and small-sized phytoplankton were obtained from 12 productivity stations in the Amundsen Sea. The daily carbon uptake rates of total phytoplankton averaged in this study were 0.42 g C m^{-2} d^{-1} (S.D. = ± 0.30 g C m^{-2} d^{-1}) and 0.84 g C m^{-2} d^{-1} (S.D. = ± 0.18 g C m^{-2} d^{-1}) whereas the daily total nitrogen (nitrate and ammonium) uptake rates were 0.12 g N m^{-2} d^{-1} (S.D. = ± 0.09 g N m^{-2} d^{-1}) and 0.21 g N m^{-2} d^{-1} (S.D. = ± 0.11 g N m^{-2} d^{-1}), respectively for non-polynya and polynya regions, which were within the ranges reported previously. Small phytoplankton contributed 26.9 % and 27.7 % to the total carbon and nitrogen uptake rates of phytoplankton in this study, respectively, which were relatively higher than the chlorophyll-a contribution (19.4 %) of small phytoplankton. For a comparison of different regions, the contributions for chlorophyll-a concentration and primary production of small phytoplankton averaged from all the non-polynya stations were 42.4 % and 50.8 %, which were significantly higher than those (7.9 % and 14.9 %, respectively) in polynya region. A strong negative correlation ($r^2 = 0.790$, $p < 0.05$) was found between the contributions of small phytoplankton and the total daily primary production of phytoplankton in this study. This finding implies that daily primary production decreases as small phytoplankton contribution increases, which is mainly due to the lower carbon uptake rate of small phytoplankton than large phytoplankton. Under ongoing environmental changes caused by global warming, a potential decrease of total primary production would be led by increasing contribution of small phytoplankton in the Amundsen Sea.

Keywords: Phytoplankton, Primary production, Polynya, Amundsen Sea

1. Introduction

The Amundsen Sea is located in the West Antarctica between the Ross Sea and Bellingshausen Sea, which is one of the least-biologically studied regions in the Southern Ocean. Recently several international research programs (KOPRI Amundsen project, iSTAR, ASPIRE, and DynaLiFe) were launched to understand this remote area. Field-measurement data revealed that annual primary production of phytoplankton reaching to 220 g C m^{-2} y^{-1} in the Amundsen Sea polynya is as high as that of Ross Sea polynya (200 g C m^{-2} y^{-1}) which was previously known for the highest productivity region in the Southern Ocean (Lee et al., 2012). Given the fact that the chl-*a* concentration averaged from all the chl-a measured stations was twice higher than that of the only productivity-measured stations, Lee et al., (2012) argued that the annual production in the Amundsen Sea polynya could be even two-fold higher than that of Ross Sea polynya.

Over the past several decades a rapid climate change has been detected and subsequently physical changes have occurred in the marine ecosystem in the western Antarctic Peninsula (WAP) (Rückamp et al. 2011; Ducklow et al. 2007). Recent studies revealed that the Thwaites Glacier in Pine Island Bay is retreating fast and the ice volume loss in the nearby Getz Ice shelf is accelerating (Joughin et al., 2014; Paolo et al., 2015). Shoaling warm Circumpolar Deep Water is believed to be a main reason for the ice sheet mass loss largely caused by ice shelf basal melt underside of the ice shelves (Yager et al. 2012; Schmidtko et al. 2014). The climate change from a cold-dry polar-type to a warm-humid sub-Antarctic-type drives subsequent changes in ocean biological productivity along the WAP shelf over the recent three decades (Montes-Hugo et al. 2009).

Phytoplankton as the base of oceanic food webs can be an indicator for changes in marine ecosystems responding to current climate changes (Moline et al., 2004; Wassman et al., 2011; Arrigo et al., 2015). In an expecting warmer ocean condition, small-sized phytoplankton is anticipated to contribute more to total phytoplankton community and thus marine ecosystems (Morán et al., 2010; Li et al., 2009; Lee et al., 2013). In consistent, Li et al. (2009) found increasing small-sized phytoplankton in the Canada Basin in the Arctic Ocean under freshening surface waters which results in a stronger stratification and

69 lower nutrient supply in the upper water column. Moreover, in the Antarctic Ocean, Moline et al. (2004)

70 found a consistent transition from large diatoms to small cryptophytes associated with glacial melt water

71 in the coastal waters along the Antarctic Peninsula in response to a regional warming trend. This change

72 in dominant phytoplankton community from large to small cells will likely cause further alteration of

73 higher trophic levels and subsequent food web (Moline et al., 2004). However, little information on the

74 contribution of small-sized phytoplankton to primary production is available in the Antarctic Ocean

75 (Saggiomo et al. 1998), especially in the Amundsen Sea with a rapid melting of ice shelf (Yager et al.

76 2012; Schmidtko et al. 2014). Thus, the main objective in this study is to determine what extend small-

77 sized phytoplankton contributes to overall total biomass and primary production in the Amundsen Sea to

78 lay the groundwork for the future monitoring of marine ecosystem change responding to ongoing changes

79 in environmental conditions.

80

81 **2. Materials and methods**

82 *2.1. Samplings*

83   Water samples were collected for carbon and nitrogen uptake measurements of phytoplankton in

84 the Amundsen Sea during the KOPRI Amundsen cruise from 1 to15 January, 2014 onboard the Korean

85 Research Icebreaker ship *Araon*. Using a dual stable isotope technique (Lee et al., 2012; Kim et al., 2015),

86 the experiments of carbon and nitrogen uptake rates of phytoplankton were conducted at 12 selected

87 productivity stations including 2 revisited-stations (St. 3-1 and St. 19-1) when on-deck incubations were

88 available during daytime at oceanographic survey stations. Based on sea ice concentration data from

89 National Snow & Ice Data Center during the cruise period (Fig. 1), our study region was further separated

90 into polynya and non-polynya areas for comparison based on sea ice distribution and concentration during

91 the cruise period. Four stations (St. 1, St. 2, St. 3, and St. 3-1) among the 12 stations were belong to non-

92 polynya region and the rest of the stations were belong to polynya region.

93   After 6 light depths (100, 50, 30, 12, 5, and 1% penetration of the surface irradiance, PAR) were

94 determined with an LI-COR underwater 4π light sensor, water samples for the uptake experiments as

95   well as biological and chemical property analysis were obtained from a CTD-rosette sampler system

96   equipped with 24 10-L Niskin bottles.

97   *2.2. Total and size-fractionated chlorophyll-a concentration*

98   Water samples for total and size-fractionated chlorophyll-a concentrations of phytoplankton were

99   obtained at the 12 productivity stations. Total chlorophyll-a concentrations were measured at six different

100   light depths (100, 50, 30, 12, 5 and 1% of PAR). For size-fractionated chlorophyll-a concentrations, water

101   samples were collected at three light depths (100, 30, and 1 %). Water samples (0.3–0.5 L) for total

102   chlorophyll-a concentrations were filtered using Whatman glass fiber filters (GF/F; 25 mm). For different

103   size-fractionated chlorophyll-a concentrations water samples (0.7–1 L) were passed sequentially through

104   20 and 5 µm Nucleopore filters (47 mm) and 0.7 µm GF/F filters (47 mm). After the filters were extracted

105   using the method described by Kim et al. (2015), all chlorophyll-a concentrations were subsequently

106   determined onboard using a Trilogy fluorometer (Turner Designs, USA). The methods and calculations

107   for chlorophyll-a were based on Parsons et al. (1984).

108   *2.3. Carbon and nitrogen uptake experiments*

109   Carbon and nitrogen uptake experiments of phytoplankton were executed by a ^{13}C-^{15}N dual

110   isotope tracer technique previously applied for the Amundsen Sea (Lee et al. 2012; Kim et al. 2015). In

111   this study, basically we followed same procedure of Lee et al. (2012). In brief, six light depths (100, 50,

112   30, 12, 5, and 1%) were determined with an LI-COR underwater 4π light sensor (LI-COR Inc., Lincoln,

113   Nebraska, USA) lowered with CTD/rosette sampler. Water sample from each light depth was transferred

114   into different screened polycarbonate incubation bottle (1 L) which matches with each light depth. The

115   productivity bottles were incubated in large polycarbonate material incubators cooled with running

116   surface seawater on deck under natural light conditions, after the water samples were inoculated with

117   labeled carbon (NaH^{13}CO$_3$) and nitrate (K^{15}NO$_3$) or ammonium (^{15}NH$_4$Cl) substrates. After 4–5 h

118   incubations, the incubated waters were well mixed and distributed into two filtration sets for the carbon

119   and nitrogen uptake rates of total (> 0.7 µm) and small-sized cells (< 5 µm). The incubated waters (0.3 L)

120   for total uptake rates were filtered through pre-combusted GF/F filters (24 mm diameter), whereas waters

121   samples (0.5 L) for the uptake rates of small-sized cells were passed through 5 μm Nuclepore filters (47

122   mm) to remove large-sized cells (> 5 μm) and then the filtrate was passed through GF/F (24 mm) for the

123   small-sized cells (Lee et al., 2013). The values for large phytoplankton in this study were obtained from

124   the difference between small and total fractions (Lee et al., 2013). The filters were immediately preserved

125   at -80℃ until mass spectrometric analysis. After acid fuming overnight to remove carbonate, the

126   concentrations of particulate organic carbon (POC) and nitrogen (PON) and the abundance of ^{13}C and

127   ^{15}N were determined by a Finnigan Delta+XL mass spectrometer at the Alaska Stable Isotope Facility,

128   USA.

129   All contribution results of small phytoplankton in this study were estimated from comparison of

130   small phytoplankton to total phytoplankton integral values from 100 to 1 % light depth at each station

131   based on the trapezoidal rule. Daily carbon and nitrogen uptake rates of phytoplankton were based on our

132   hourly uptake rates measured in this study and a 24-h photoperiod per day during the summer period in

133   the Amundsen Sea (Lee et al., 2012).

134

135   **3. Results**

136   3.1. Chlorophyll-a, POC, and PON contributions of small phytoplankton

137   The total (large + small phytoplankton) chlorophyll-a concentration integrated from six different

138   light depths ranged from 11.1 to 80.3 mg chl-a m^{-2} (mean ± S.D. = 57.4 ± 25.2 mg chl-a m^{-2}), whereas

139   small (< 5 µm) chlorophyll-a concentration ranged from 3.9 to 9.4 mg chl-a m^{-2} (mean ± S.D. = 5.7 ± 1.7

140   mg chl-a m^{-2}) in this study (Fig. 2). The average contribution of small phytoplankton to the total

141   chlorophyll-a concentration was 19.4 % (S.D. = ± 26.0 %) ranging from 4.9 to 76.5 %. In the Amundsen

142   Sea, large phytoplankton (> 5 μm) were generally predominant (approximately 80 %) during the cruise

143   period in 2014 based on different-sized chlorophyll-a concentrations. For a regional comparison, the

144   average contributions of small phytoplankton to the total chlorophyll-a concentration were 42.4 % (S.D. =

145 ± 37.2 %) and 7.9 % (S.D. = ± 3.5 %) for non-polynya and polynya regions, respectively. The

146 chlorophyll-a contribution of small phytoplankton was larger in the non-polynya stations than the polynya

147 stations although they were not significantly different (t-test, p = 0.16).

148   The total integral POC concentration of phytoplankton displayed no large spatial variation

149 ranging from 4.72 to 9.22 mg C m^{-2} (Fig. 3). In comparison, the total integral PON concentration of

150 phytoplankton ranged from 0.76 to 1.74 mg C m^{-2}. The POC contribution of small phytoplankton ranged

151 from 30.7 to 65.5 % (mean ± S.D. = 41.1 ± 10.6 %), whereas the PON contribution ranged similarly from

152 30.8 to 67.2 % (mean ± S.D. = 41.3 ± 11.5 %) in the Amundsen Sea (Fig. 3). Specifically, the POC and

153 PON contributions of small phytoplankton averaged from all the polynya stations were 36.9 % (S.D. = ±

154 4.6 %) and 37.0 % (S.D. = ± 6.9 %), respectively. In comparison, the POC and PON contributions of

155 small phytoplankton averaged from the non-polynya stations were 49.5 % (S.D. = ± 14.4 %) and 50.0 %

156 (S.D. = ± 15.1 %), respectively.

157 3.2. Carbon uptake rate contributions of small phytoplankton

158   The total daily carbon uptake rates of phytoplankton (large + small phytoplankton) integrated

159 from six different light depths ranged from 150.4 to 1213.4 mg C m^{-2} d^{-1} with an average of 696.5 mg C

160 m^{-2} d^{-1} (S.D. = ± 298.4 mg C m^{-2} d^{-1}) in this study (Fig. 4). In contrast, the rates of small phytoplankton

161 ranged between 58.6 and 266.4 mg C m^{-2} d^{-1} with an average of 124.9 mg C m^{-2} d^{-1} (S.D. = ± 62.4 mg C

162 m^{-2} d^{-1}). Small phytoplankton contributed 26.9 % (S.D. = ± 29.3%) to total daily carbon uptake rate of

163 total phytoplankton.

164   Specifically, the total daily carbon uptake rates of phytoplankton ranged from 150.4 to 796.4 mg

165 C m^{-2} d^{-1} with an average of 415.0 mg C m^{-2} d^{-1} (S.D. = ± 298.2 mg C m^{-2} d^{-1}) in non-polynya region,

166 whereas they ranged from 654.8 to 1213.4 mg C m^{-2} d^{-1} with an average of 837.3 mg C m^{-2} d^{-1} (S.D. = ±

167 184.1 mg C m^{-2} d^{-1}) in polynya region. The total daily carbon uptake rates of phytoplankton were

168 significantly higher (t-test, p < 0.05) in polynya stations than non-polynya stations. The rates of small

169 phytoplankton ranged between 58.6 and 193.6 mg C m^{-2} d^{-1} with an average of 126.5 mg C m^{-2} d^{-1} (S.D. =

170 ± 55.2 mg C m^{-2} d^{-1}) in non-polynya region, whereas they ranged from 62.2 to 266.4 mg C m^{-2} d^{-1} with an

171 average of 124.1 mg C m^{-2} d^{-1} (S.D. = ± 69.3 mg C m^{-2} d^{-1}) in polynya region. The daily carbon uptake

172 rates of small phytoplankton were not significantly different (t-test, p > 0.05) between polynya and non-

173 polynya stations. The average contributions of small phytoplankton to total daily carbon uptake rates were

174 50.8 % (S.D. = ± 42.8 %) and 14.9 % (S.D. = ± 8.4 %), respectively for non-polynya and polynya regions.

175 The average contributions were largely different between polynya and non-polynya regions but they were

176 not statistically significant (t-test, p > 0.05).

177 3.3. Nitrogen uptake rate contributions of small phytoplankton

178 The total daily nitrate uptake rates of phytoplankton (large + small phytoplankton) ranged from

179 34.0 to 174.2 mg N m^{-2} d^{-1} with an average of 93.7 mg N m^{-2} d^{-1} (S.D. = ± 43.2 mg N m^{-2} d^{-1}), whereas the

180 rates of small phytoplankton ranged from 6.1 to 40.9 mg N m^{-2} d^{-1} with an average of 19.0 mg N m^{-2} d^{-1}

181 (S.D. = ± 11.3 mg N m^{-2} d^{-1}) in this study (Fig. 5). Small phytoplankton contributed 21.5 % (S.D. = ±

182 11.1 %) to total daily nitrate uptake rates. In comparison, the total daily ammonium uptake rates of

183 phytoplankton ranged from 12.4 to 173.8 mg N m^{-2} d^{-1} with an average of 86.7 mg N m^{-2} d^{-1} (S.D. = ±

184 75.9 mg N m^{-2} d^{-1}), whereas the rates of small phytoplankton ranged from 9.1 to 81.1 mg N m^{-2} d^{-1} with

185 an average of 25.7 mg N m^{-2} d^{-1} (S.D. = ± 21.1 mg N m^{-2} d^{-1}) in this study (Fig. 6). Small phytoplankton

186 contributed 38.7 % (S.D. = ± 24.9 %) to total daily ammonium uptake rates. The contributions of small

187 phytoplankton were significantly higher in ammonium uptake rate than nitrate uptake rate (t-test, p <

188 0.05).

189 Specifically for different regions, the total daily nitrate uptake rates of phytoplankton ranged from

190 34.0 to 142.1 mg N m^{-2} d^{-1} with an average of 71.9 mg N m^{-2} d^{-1} (S.D. = ± 48.4 mg N m^{-2} d^{-1}) in non-

191 polynya region whereas they ranged from 44.2 to 174.2 mg N m^{-2} d^{-1} with an average of 104.6 mg N m^{-2}

192 d^{-1} (S.D. = ± 39.0 mg N m^{-2} d^{-1}) in polynya region, respectively. In comparison, the daily nitrate uptake

193 rates of small phytoplankton ranged from 7.5 to 26.6 mg N m^{-2} d^{-1} with an average of 16.7 mg N m^{-2} d^{-1}

194 (S.D. = ± 7.8 mg N m^{-2} d^{-1}) and from 6.1 to 40.9 mg N m^{-2} d^{-1} with an average of 20.1 mg N m^{-2} d^{-1} (S.D.

195 = ± 13.1 mg N m^{-2} d^{-1}), respectively for non-polynya and polynya regions. The contributions of small

196 phytoplankton to the total daily nitrate uptake rates were 28.2 % (S.D. = ± 15.9 %) in non-polynya region

197 and 18.1 % (S.D. = ± 6.8 %) in polynya region, respectively. The total daily ammonium uptake rates of

198 total phytoplankton ranged between 12.3 and 106.1 mg N m^{-2} d^{-1} (mean ± S.D. = 49.7 ± 41.2 mg N m^{-2} d^{-1})

199 in non-polynya region and between 18.1 and 269.3 mg N m^{-2} d^{-1} (mean ± S.D. = 105.2 ± 84.6 mg N m^{-2} d^{-}

200 ^{1}) in polynya region. In comparison, the rates of small phytoplankton ranged between 9.1 and 22.4 mg N

201 m^{-2} d^{-1} (mean ± S.D. = 15.8 ± 6.4 mg N m^{-2} d^{-1}) in non-polynya region and between 9.9 and 81.1 mg N m^{-}

202 ^{2} d^{-1} (mean ± S.D. = 30.7 ± 24.5 mg N m^{-2} d^{-1}) in polynya region. Small phytoplankton contributed 52.8 %

203 (S.D. = ± 40.5 %) and 31.6 % (S.D. = ± 10.1 %) to the total daily ammonium uptake rates in non-polynya

204 and polynya regions, respectively which were not significantly different (t-test, p = 0.37).

205 The total integral daily nitrogen uptake rate (nitrate + ammonium uptake rates) of phytoplankton

206 ranged from 46.4 to 443.5 mg N m^{-2} d^{-1} (mean ± S.D. = 180.4 ± 106.7 mg N m^{-2} d^{-1}) in this study. For

207 non-polynya and polynya regions, they ranged from 46.4 to 248.1 mg N m^{-2} d^{-1} (mean ± S.D. = 121.6 ±

208 89.3 mg N m^{-2} d^{-1}) and from 91.7 to 443.5 mg N m^{-2} d^{-1} (mean ± S.D. = 209.8 ± 107.3 mg N m^{-2} d^{-1}),

209 respectively. In comparison, the total integral daily nitrogen uptake rate of small phytoplankton ranged

210 from 16.6 to 46.6 mg N m^{-2} d^{-1} (mean ± S.D. = 32.5 ± 13.2 mg N m^{-2} d^{-1}) and from 17.6 to 122.0 mg N m^{-}

211 ^{2} d^{-1} (mean ± S.D. = 50.8 ± 32.4 mg N m^{-2} d^{-1}) for non-polynya and polynya regions, respectively. Small

212 phytoplankton contributed 36.2 % (S.D. = ± 23.0 %) to the total integral daily nitrogen uptake rates in

213 non-polynya region, whereas they contributed 23.5 % (S.D. = ± 6.0 %) for polynya region. The integral

214 daily nitrogen uptake rates and contributions of small phytoplankton were not statistically different

215 between non-polynya and polynya regions.

216

4. Discussion and conclusion

218 The total daily carbon uptake rates of phytoplankton averaged for non-polynya and polynya

219 regions were 0.42 g C m^{-2} d^{-1} (S.D. = ± 0.30 g C m^{-2} d^{-1}) and 0.84 g C m^{-2} d^{-1} (S.D. = ± 0.18 g C m^{-2} d^{-1}),

220 respectively in this study. According to the previous reports in the Amundsen Sea (Lee et al., 2012; Kim

221 et al., 2015), the total daily carbon uptake rates ranged from 0.2 to 0.12 g C m^{-2} d^{-1} in non-polynya region.

222 Our rate (0.42 g C m^{-2} d^{-1}) in non-polynya region is somewhat higher than those reported previously but

223 they are not significantly different (t-test, p = 0.77). In comparison, our total daily carbon uptake rate in

224 polynya region (0.84 g C m^{-2} d^{-1}) is within the range between Lee et al. (2012; 2.2 g C m^{-2} d^{-1}) and Kim et

225 al. (2015; 0.2 g C m^{-2} d^{-1}). The carbon uptake rates of phytoplankton in Lee et al. (2012) and Kim et al.

226 (2015) were measured during December 21, 2010-January 23, 2011 and February 11 to March 14, 2012,

227 respectively. Our measurements in this study were executed mainly during January 1-15, 2014.

228 For the Amundsen polynya region, a large seasonal variation in the total daily carbon uptake rate of

229 phytoplankton was already reported by Kim et al. (2015) and Arrigo et al. (2012) based on filed-measured

230 data and satellite-derived approach, respectively. It is appeared that this seasonal variation largely

231 depends on the bloom stage of phytoplankton which peaks during the late December-January and

232 terminates at late February (Arrigo and van Dijken 2003; Arrigo et al., 2012; Kim et al., 2015). The

233 carbon uptake rates of phytoplankton in Lee et al. (2012) and Kim et al. (2015) were measured during

234 December 21 January 23, 2010 and February 11 to March 14, 2012, respectively. Our measurements in

235 this study were executed mainly during January 1-15, 2014.

236 The total daily nitrogen uptake rates of phytoplankton were 0.12 g N m^{-2} d^{-1} (S.D. = ± 0.09 g N m^{-}

237 ^{2} d^{-1}) and 0.21 g N m^{-2} d^{-1} (S.D. = ± 0.11 g N m^{-2} d^{-1}) for non-polynya and polynya regions, respectively

238 in this study. Previous studies reported that the total daily nitrogen uptake rates in non-polynya region

239 were 0.24 g N m^{-2} d^{-1} during Dec. 21, 2010-Jan. 23, 2011 in 2010/2011 and 0.04 g N m^{-2} d^{-1} during Feb. 11

240 to Mar. 14, 2012 in 2012 whereas the uptake rates in polynya region were 0.93 g N m^{-2} d^{-1} in 2010/2011

241 and 0.06 g N m^{-2} d^{-1} in 2012 in the Amundsen Sea (Lee et al., 2012; Kim et al., 2015). Our total daily

242 nitrogen uptake rates of phytoplankton in non-polynya and polynya regions were between the two

243 previous studies (Lee et al., 2012; Kim et al., 2015). Based on the nitrate and ammonium uptake rates in

244 this study, *f*-ratios (nitrate uptake rate/nitrate+ammonium uptake rates) averaged for non-polynya and

245 polynya regions were 0.62 (S.D. = ± 0.08) and 0.54 (S.D. = ± 0.20), respectively. These ratios also were

246 between the two previous studies. Although they were not significant different because of a large spatial

247 variation, larger *f*-ratios in non-polynya than in polynya region are consistent with the results of the

248 previous studies (Lee et al., 2012; Kim et al., 2015). At this point, we do not have a solid explanation for

249 that but a further future study is needed for the higher *f*-ratio mechanism in non-polynya region.

250   The percent contributions of small phytoplankton in terms of chlorophyll-a, POC/PON, daily

251 carbon and nitrogen uptake rates are shown in Table 1. The overall contribution of small phytoplankton to

252 the total chlorophyll-a concentration for all the productivity stations was 19.4 % (S.D. = ± 26.0 %) which

253 is significantly (*t*-test, $p < 0.05$) lower than the POC contribution (41.1 ± 10.6 %). This is consistent with

254 the result in the Chukchi Sea, Arctic Ocean reported by Lee et al. (2013). They explained that higher POC

255 content per chlorophyll-a unit of small phytoplankton could cause their higher POC contribution (Lee et

256 al., 2013). Given C/N ratio (mean ± S.D. = 6.6 ± 0.6) and $\delta^{13}C$ (mean ± S.D. = -25.9 ± 1.0 ‰) of sample

257 filters attained for POC and PON in this study, our filtered samples are believed to be mainly

258 phytoplankton-originated POC and PON (Kim et al., 2016). Thus, a significant potential overestimated

259 POC contribution of non-phytoplankton materials could be excluded for the higher POC contribution than

260 chlorophyll-a contribution of small phytoplankton. Therefore, small phytoplankton contributions based on

261 conventional assessments of chlorophyll-a concentration might lead an underestimated contribution of

262 small phytoplankton (Lee et al., 2013). In fact, several authors argued that chlorophyll-a concentration

263 might be not a good index for phytoplankton biomass since it largely depends on environmental factors

264 such as nutrient and light conditions as well as dominant groups and physiological status of

265 phytoplankton (Desortová 1981; Behrenfeld et al., 2005; Kruskopf and Flynn, 2006; Behrenfeld and Boss

266 2006). However, the effects of non-phytoplankton carbon materials such as extracellular carbon mucilage

267 can not be completely excluded for the POC contribution as discussed below.

268   The overall contributions of carbon and nitrogen (nitrate and ammonium) uptake rates of small

269 phytoplankton at all the productivity stations in this study are similar with 26.9 % and 27.7 %,

270 respectively. These contributions are relatively higher than the chlorophyll-a contribution of small

271 phytoplankton but they are not statistically different (t-test, $p > 0.05$). In general, the contribution of daily

272 ammonium uptake rate of small phytoplankton are significantly (t-test, $p < 0.05$) higher than the

273 contribution of daily nitrate uptake rate of small phytoplankton at all the stations in this study. It is well-

274 known for the ammonium preference of small phytoplankton in various regions (Koike et al., 1986;

275 Tremblay et al., 2000, Lee et al., 2008; Lee et al., 2013).

276   In terms of the contributions in different regions, all the contributions (Chlorophyll-a, POC/PON,

277 carbon and nitrogen uptake rates) of small phytoplankton were higher in non-polynya region than in

278 polynya region (Table 1). In addition, the chlorophyll-a contribution of small phytoplankton (mean ± S.D.

279 = 7.9 ± 3.5 %) was significantly (t-test, $p < 0.01$) lower than the POC contribution (mean ± S.D. = 36.9 ±

280 4.6 %) in polynya region, whereas they were not statistically different in non-polynya region (Table 1).

281 This indicates that small phytoplankton contributed more to the total POC than the chlorophyll-a

282 concentration in the polynya region. We do not have species compositions of phytoplankton in this study,

283 but previous results reported that *Phaeocystis* sp. are dominant in the Amundsen Sea polynya region

284 ~~whereas diatoms are relatively dominant in the non-polynya regions~~ (Lee et al., 2012). Generally,

285 *Phaeocystis* spp. release a large portion (up to 46 %) of extracellular carbon mucilage which makes their

286 colonial form (Matrai et al., 1995). This non-phytoplankton carbon material without chlorophyll-a might

287 cause a higher POC contribution of small phytoplankton in the polynya region during this study. In fact,

288 the contribution of the daily carbon uptake rates of small phytoplankton (14.9 ± 8.4 %) was not as high as

289 the POC contribution (36.9 ± 4.6 %) in the polynya region. The chlorophyll-a contributions of small

290 phytoplankton were lower than that of the daily carbon uptake rate in this study, which is consistent with

291 the results from polynya and marginal ice zone stations in the Ross Sea, Antarctica during austral spring

292 and summer (Saggiomo et al., 1998). They reported that the chlorophyll-a and primary production

293 contributions of pico-phytoplankton (< 2 μm) were 29 % and 40 % at polynya stations whereas the

294 contributions were 17 % and 32 % at marginal ice zone stations, respectively. In polynya region, they

295 found much higher contributions in chlorophyll-a and primary production of small phytoplankton than

296 those in this study although their size of small phytoplankton is somewhat smaller than our size (< 5 μm).

297   We found a strong negative correlation ($r^2 = 0.790$, $p < 0.05$) between the productivity

298 contributions of small phytoplankton and total daily carbon uptake rates of phytoplankton in the

299 Amundsen Sea (Fig. 7), which implies that daily primary production decreases as small phytoplankton

300 contribution increases. This is mainly because of the relatively lower carbon uptake rate of small

301 phytoplankton than large phytoplankton in the Chukchi Sea, Arctic Ocean reported by Lee et al. (2013).

302 Moline et al. (2004) suggested that further warming air temperatures will increase inputs of glacial

303 melting water and subsequently increase the contributions of small phytoplankton over large

304 phytoplankton community (Moline et al. 2004). If these small phytoplankton were dominant under

305 ongoing more melting conditions of glaciers, a potential increasing contribution of small phytoplankton

306 might cause a subsequent decrease in the total primary production of phytoplankton in the Amundsen Sea

307 based on this study in Figure 7.

308 In respect to food quality of small phytoplankton as a basic food source to herbivores,

309 macromolecular compositions such as proteins, lipids, and carbohydrates as photosynthetic-end products

310 will be needed for better understanding alterations of marine ecosystem in response to ongoing

311 environmental changes (Lee et al., 2013). According to Kang et al. (accepted), small phytoplankton

312 assimilate more food materials and calorific contents per unit of chlorophyll-a concentration and thus

313 provide more contributions in respect to energy aspect than other phytoplankton community in the

314 East/Japan Sea. In conclusion, monitoring the contributions of small-sized phytoplankton to total biomass

315 and primary production of total phytoplankton community could be important as a valuable indicator to

316 sense future changes in marine ecosystem under ongoing various climate-associated environmental

317 changes. Moreover, further detailed studies for macromolecular compositions of small phytoplankton will

318 be necessary for the anticipating small-dominant ecosystem under warming oceans.

319

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324

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Table caption

Table 1. Contributions (%) ~~of chlorophyll-a, POC, PON, and carbon and nitrogen uptake rates)~~ of small phytoplankton in the Amundsen Sea. Contributions of chlorophyll-a, POC, PON, and carbon and nitrogen uptake rates were derived from water euphotic column-integrated values averaged from stations.

403 **Figure captions**

404 Fig. 1. Sampling locations in the Amundsen Sea. Red closed circles represent productivity stations. Sea

405   ice concentration data during the cruise period from Nimbus-7 SMMR and DMSP SSM/I-

406   SSMIS Passive Microwave data provided by National Snow & Ice Data Center.

407 Fig. 2. Water column-integrated chlorophyll-a concentration at the productivity stations in the Amundsen

408   Sea.

409 Fig. 3. Water column-integrated concentrations of POC and PON of small and large phytoplankton.

410 Fig. 4. Water column-integrated daily carbon uptake rates of small and large phytoplankton.

411 Fig. 5. Water column-integrated daily nitrate uptake rates of small and large phytoplankton.

412 Fig. 6. Water column-integrated daily ammonium uptake rates of small and large phytoplankton.

413 Fig. 7. Relationship between productivity contributions of small phytoplankton and the total daily carbon

414   uptake rates of phytoplankton (large + small). The total daily carbon uptake rates were

415   transformed into natural logs for a linear regression.

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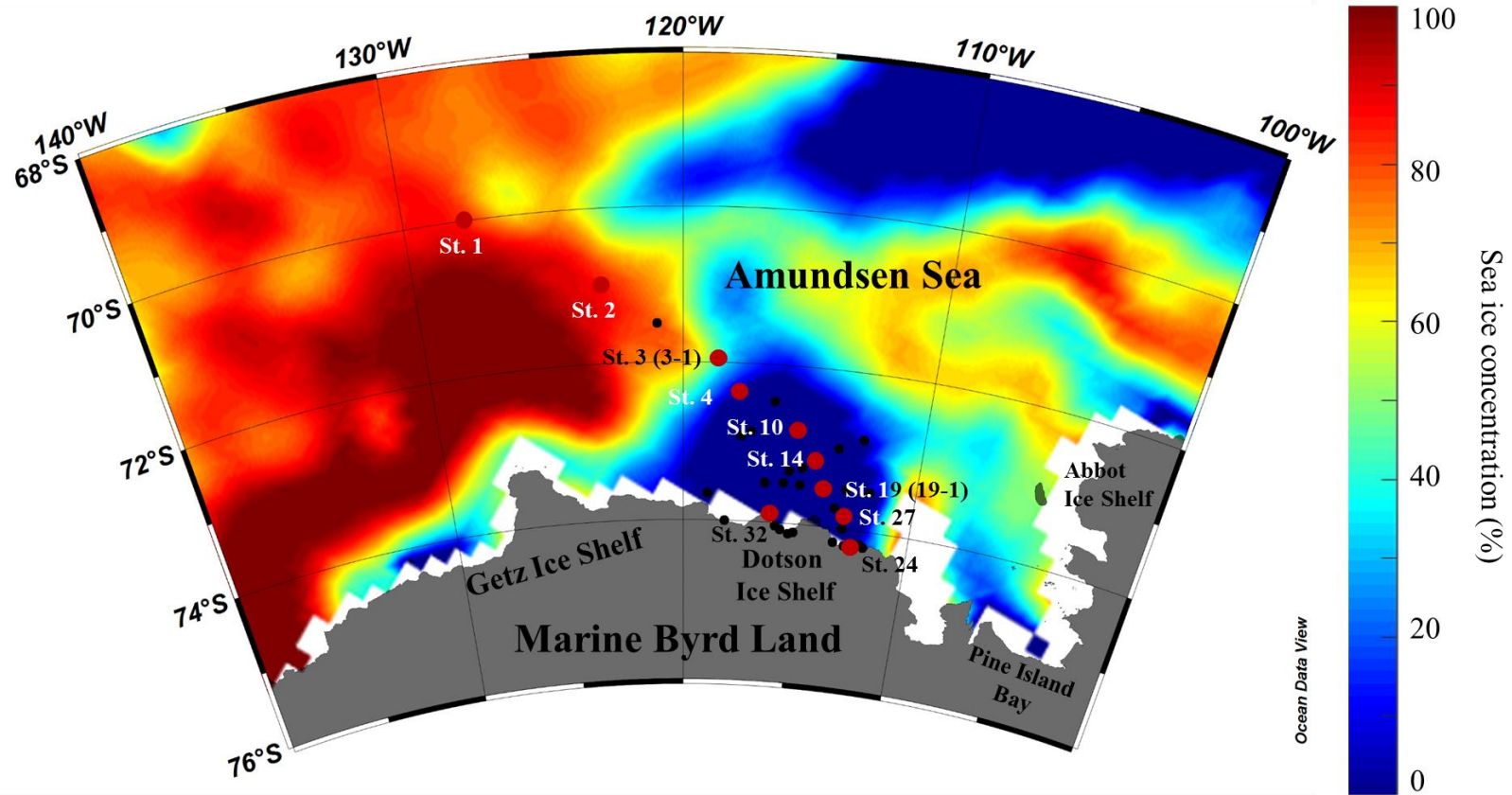
417

418

Table 1. Contributions (%) of small phytoplankton in the Amundsen Sea. Contributions of chlorophyll-a, POC, PON, and carbon and nitrogen uptake rates were derived from water euphotic column-integrated values averaged from stations.

	Chlorophyll-a	POC	PON	Daily carbon uptake rate	Daily nitrate uptake rate	Daily ammonium uptake rate	Total nitrogen uptake rate
All stations	19.4 ± 26.0	41.1 ± 10.6	41.3 ± 11.5	26.9 ± 29.3	21.5 ± 11.1	38.7 ± 24.9	27.7 ± 14.4
Non-polynya	42.4 ± 37.2	49.5 ± 14.4	50.0 ± 15.1	50.8 ± 42.8	28.2 ± 15.9	52.8 ± 40.5	36.2 ± 23.0
Polynya	7.9 ± 3.5	36.9 ± 4.6	37.0 ± 6.9	14.9 ± 8.4	18.1 ± 6.8	31.6 ± 10.1	23.5 ± 6.0

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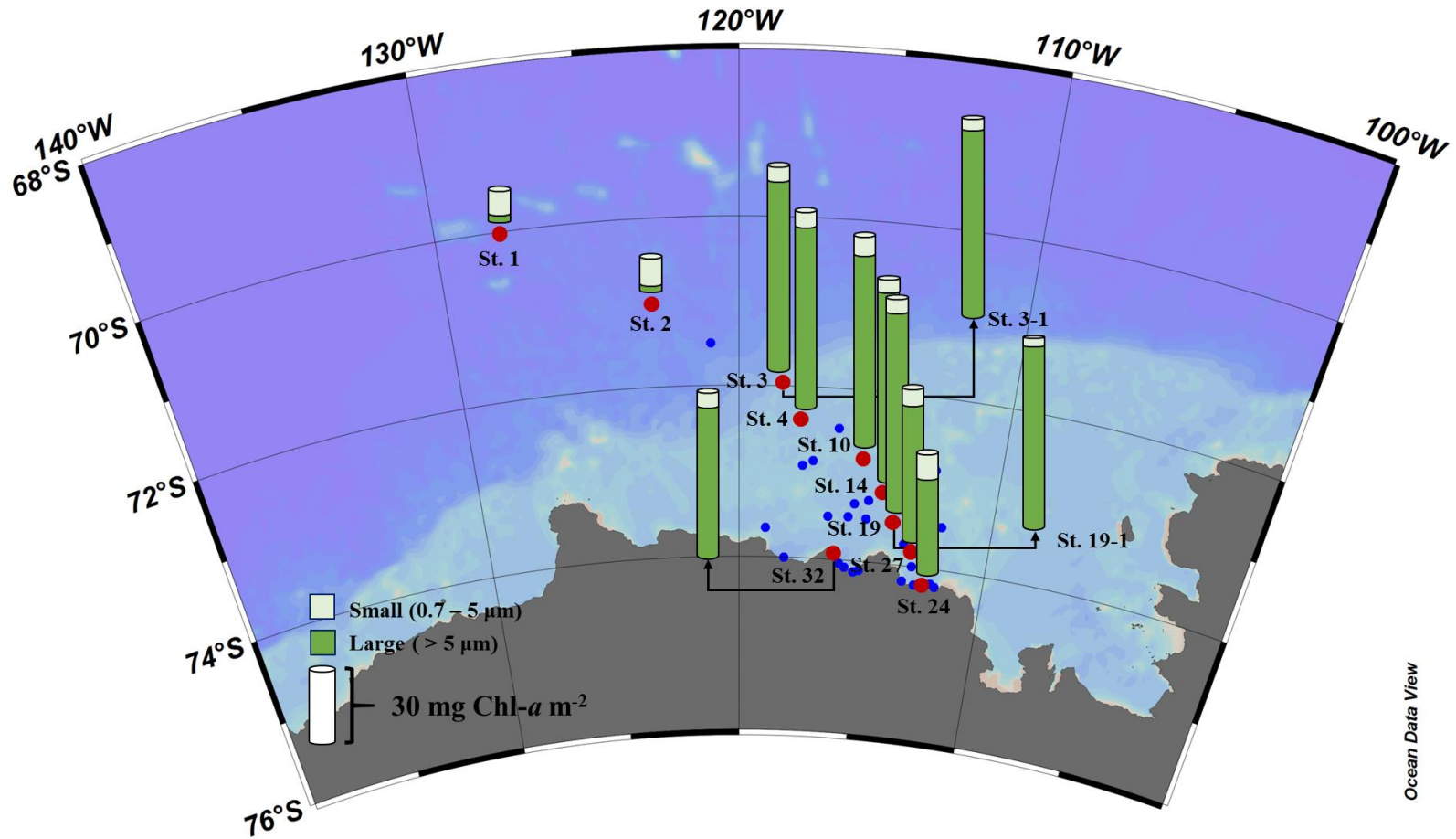


Fig. 2. Water column-integrated chlorophyll-a concentration at the productivity stations in the Amundsen Sea.

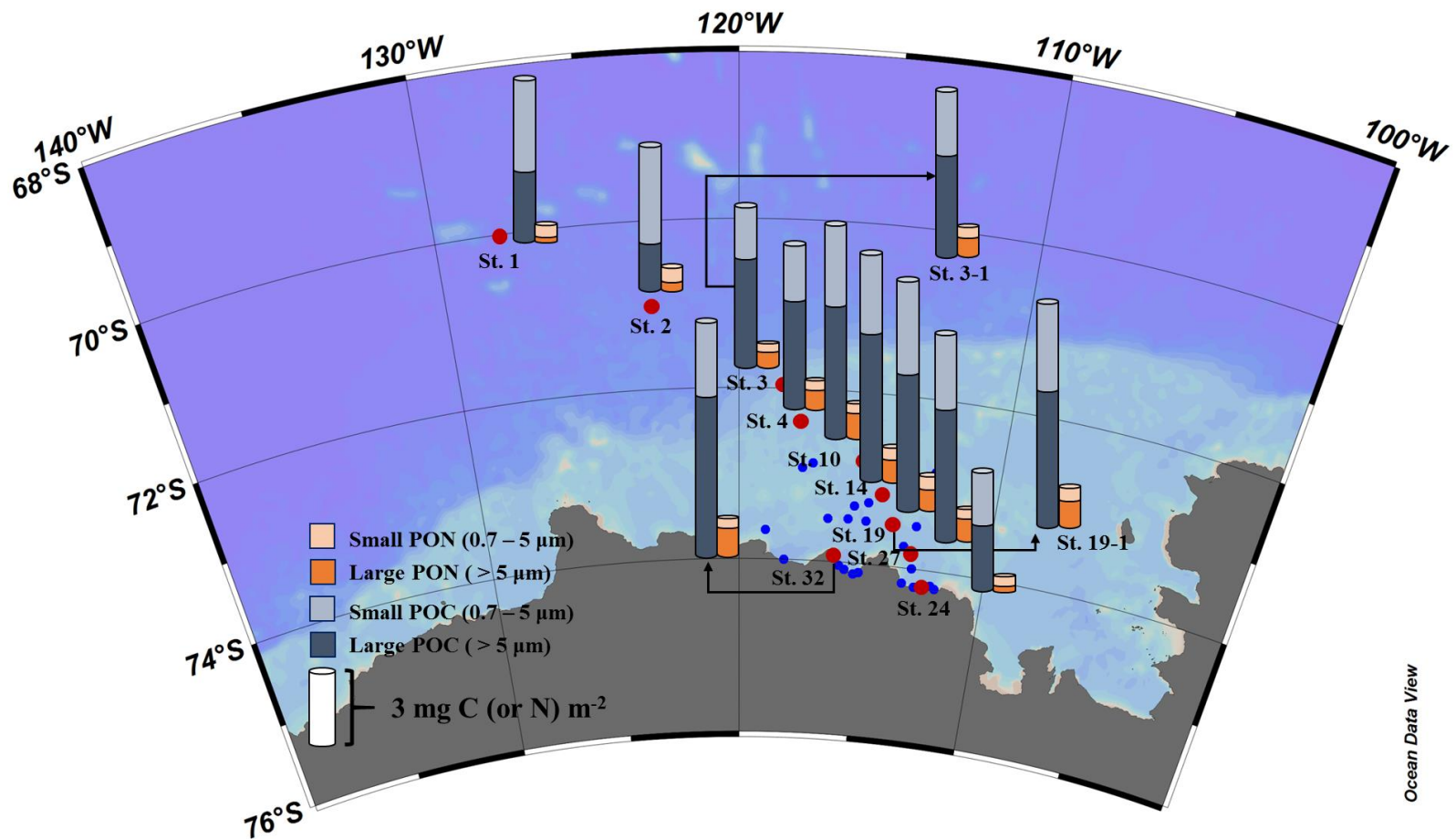



Fig. 3. Water column-integrated concentrations of POC and PON of small and large phytoplankton.

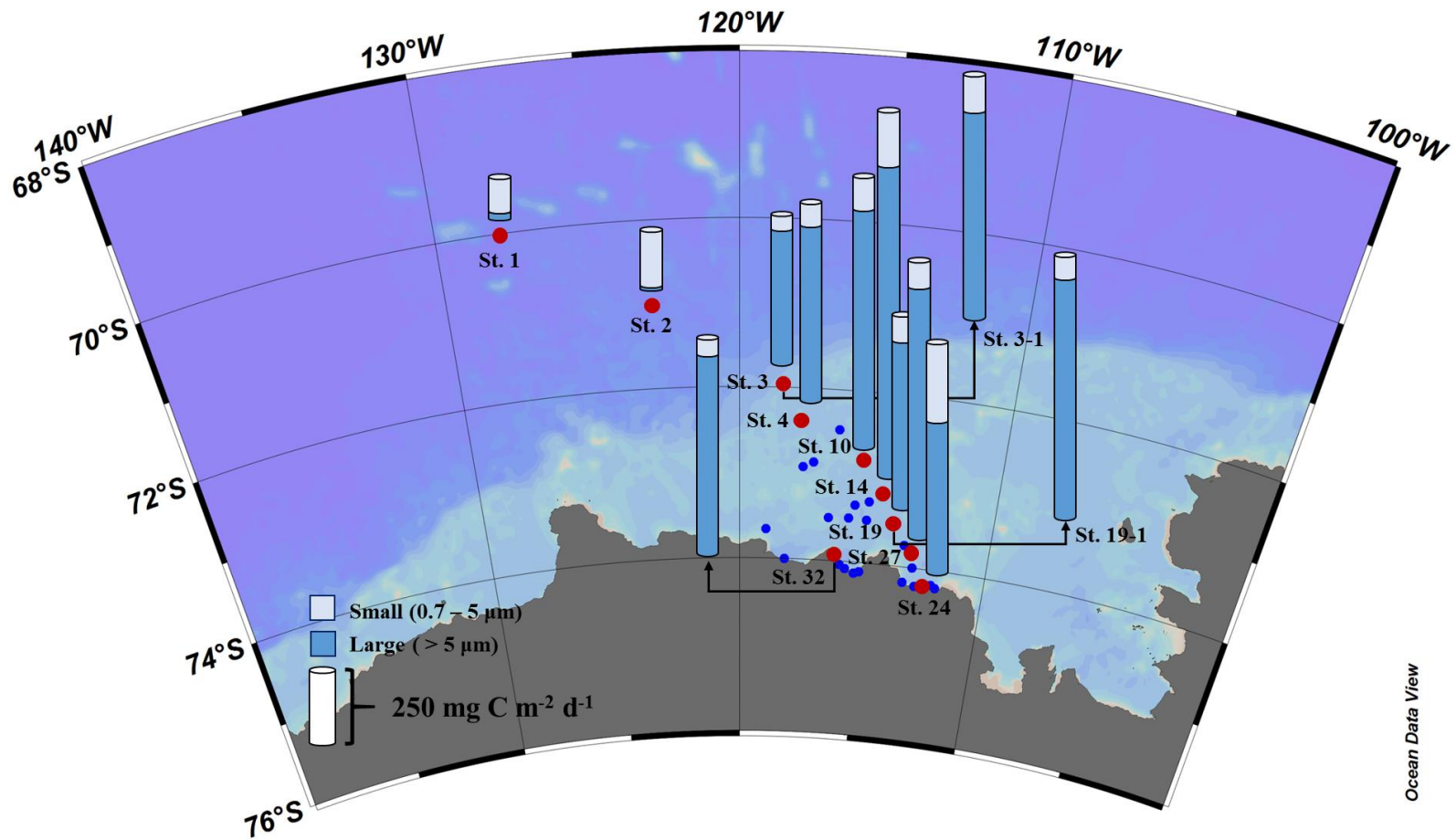



Fig. 4. Water column-integrated daily carbon uptake rates of small and large phytoplankton.

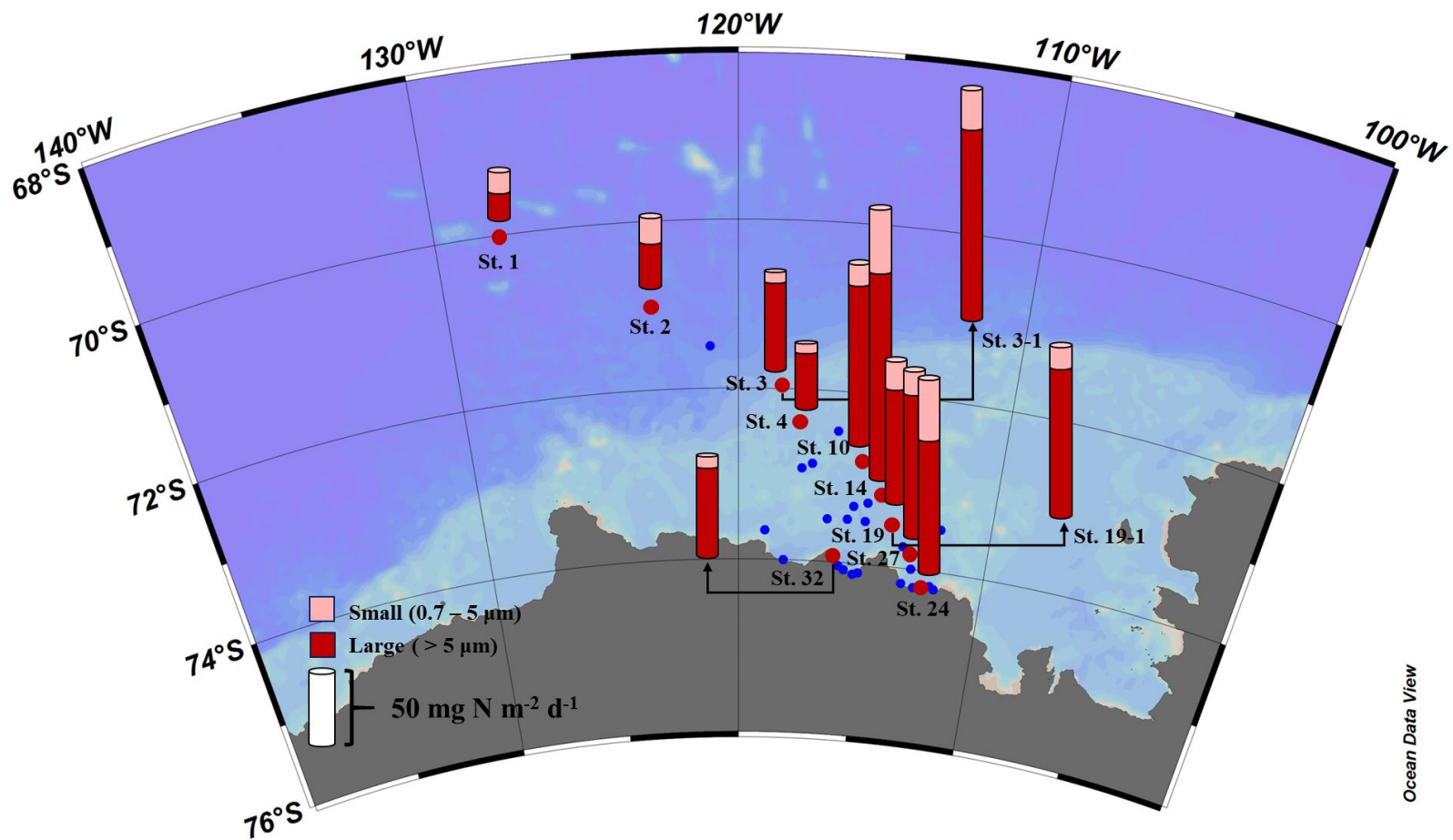



Fig. 5. Water column-integrated daily nitrate uptake rates of small and large phytoplankton.

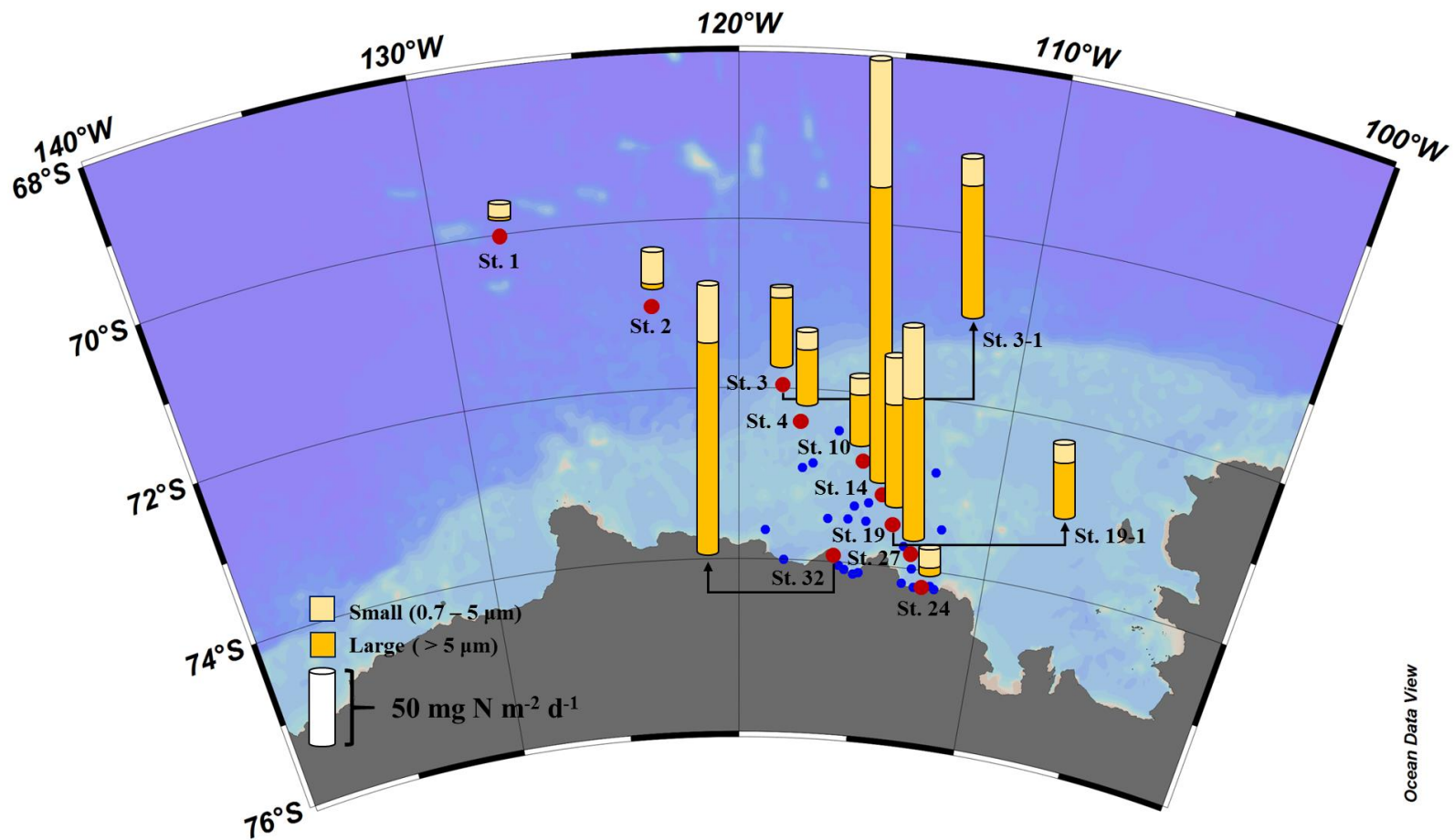
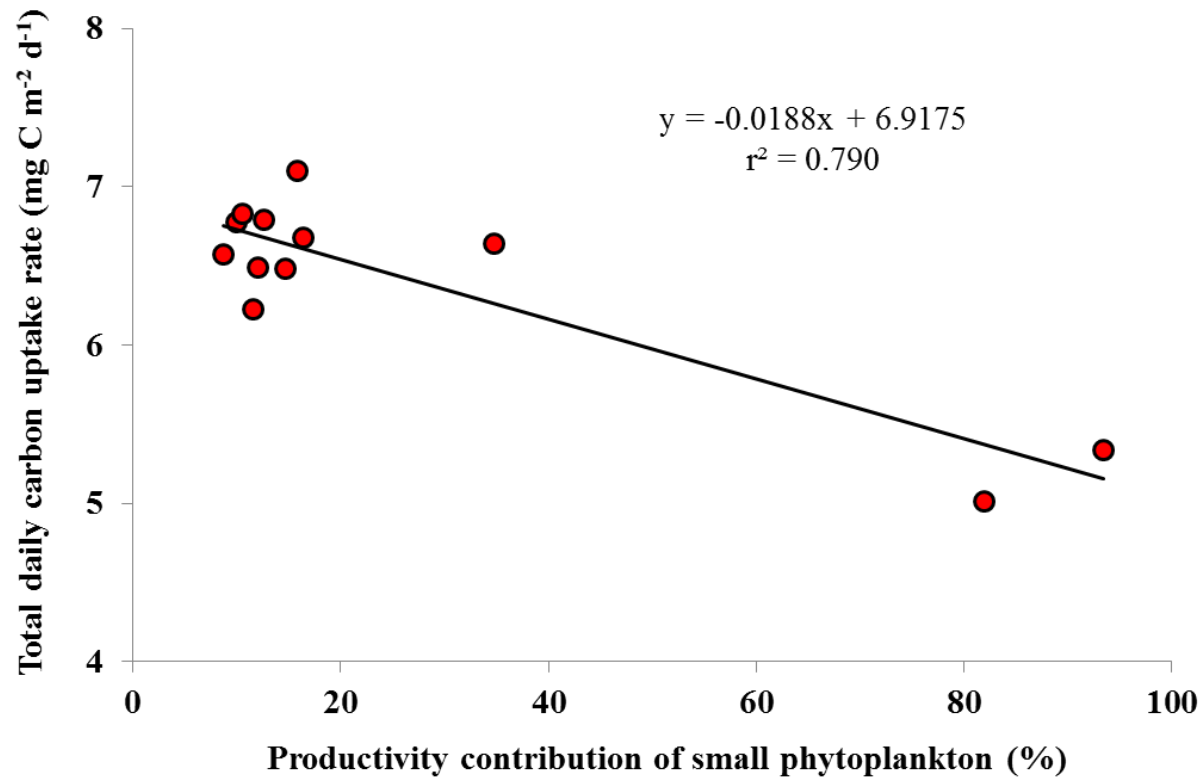



Fig. 6. Water column-integrated daily ammonium uptake rates of small and large phytoplankton.

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