

1 **Interactive comment on “Small phytoplankton contribution to the total**
2 **primary production in the Amundsen Sea” by Sang H. Lee et al.**

3 **Anonymous Referee #1**

4

5 Received and published: 17 November 2016

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7 The manuscripts “Small phytoplankton contribution to the total primary production in the Amundsen
8 Sea” by Lee et al. presents observational data in Amundson during 1-14 January 2014 cruise and
9 discussed an important issue on the small phytoplankton contribution to the total primary production. I
10 found the data and discussion deserved for publication in BG with minor revision. I suggest the authors to
11 improve description of the differences between non-polynya and polynya regions, maybe a regroup those
12 stations in order to make the conclusions stronger.

13 → Since our study region was separated into polynya and non-polynya areas based on sea ice
14 concentration data from National Snow & Ice Data Center during the cruise period (Fig. 1) as we
15 mentioned that in line 88-92, page 4, regrouping those stations based on the result outcome is rather
16 arbitrary. Therefore, we would like to stick with the previous two groups based on sea ice concentration.

17

18 It is also important to include time period of measurements when discuss comparison with other studies in
19 many places in the manuscript. Here are some details: L223-225: “our total 223 daily carbon uptake rate
20 in 224 polynya region (0.84 g C m⁻² d⁻¹) is within the range between Lee et al. (2012; 2.2 g C m⁻² d⁻¹)
21 and Kim et 225 al. (2015; 0.2 g C m⁻² d⁻¹).” The wide range of carbon uptake rates are mainly due to the
22 different measurement timing (or location). This is an example where it is necessary to add which month
23 (not just year) the data were measured when comparing those numbers.

24 → Yes, the different carbon uptake rates among different studies are mainly due to the different
25 measurement timing. We indicated the time period of the measurement for each study for the comparison
26 of the rates in line 225-227 and line 239-240, page 10.

27

28 L274 states “small phytoplankton were higher in non-polynya region (Table 1)”. L281 states ‘diatoms are
29 relatively dominant in the non-polynya regions (Lee et al., 2012)’. Please explain why they are different
30 as we normally think diatom is large phytoplankton.

31 → We are not saying diatom is small phytoplankton in this paragraph. As we mentioned in line 143-145,
32 the average contributions of small phytoplankton to the total chlorophyll-a concentration were 42.4 %
33 (S.D. = ± 37.2 %) for non-polynya based on different sizes of chlorophyll-a concentration which

34 indicating there were still some large amount of small phytoplankton ($< 5\mu\text{m}$) although they were not
35 dominant group. Since it is rather confused, we rephrased it in line 284, page 12.

36
37 In Fig 2-4, small phytoplankton were lower in non-polynya stations 3 and 3-1, higher in 1 and 2. Stations
38 1 and 2 had very low production and its ratio may not represent the ratio when bloom occurs in those
39 locations. It is necessary to note whether the ratios in Table 1 is the average of ratios in each station or
40 calculated from the average of chl-a, PP.

41 → The ratios in Table 1 are the euphotic water column values averaged from all stations, non-polynya
42 station, and polynya stations. We clarified that in the caption of Table 1.

43
44 L315: 'anticipating small-dominant ecosystem under warming oceans'. We have found increasing small
45 phytoplankton due to warming Arctic, but in Amundson, small phytoplankton contribution was found to
46 be higher under ice (cold) rather than in polynya (warm) in this study. It looks like we are heading to
47 large-dominant ecosystem under warming ocean in Amundson.

48 → Polynya and non-polynya regions are different systems with different environmental conditions so that
49 we can not simply say that. That is a main reason for why we separated them in this study. Actually, the
50 data in Figure 7 included all stations from polynya and non-polynya regions.

51

52 **Interactive comment on “Small phytoplankton contribution to the total**
53 **primary production in the Amundsen Sea” by Sang H. Lee et al.**

54 **Anonymous Referee #2**

55 Received and published: 12 December 2016

56

57 General Comments:

58 The manuscript “Small phytoplankton contribution to the total primary production in the Amundsen Sea”
59 by Lee et al. presents size-fractionated chlorophyll, particulate organic carbon/nitrogen, and
60 carbon/nitrogen uptake rates in the Amundsen Sea to characterize the contribution of small phytoplankton.

61 As the authors state, this type of data is lacking in the Amundsen Sea, yet is invaluable for understanding
62 how the region might be altered by climate change. I commend the authors on the collection of a unique
63 dataset, and given the importance of the data, would be excited to see this manuscript published in
64 Biogeosciences. However, it is my opinion that it should be reconsidered after major revisions for the
65 following reasons:

66 - There are strong statements re. the future importance and driving mechanism of small phytoplankton in
67 the Amundsen Sea based on limited evidence from that region, and rather extrapolated from other regions
68 (more northern Western Antarctic Peninsula and Arctic Ocean). Ultimately I feel that the focus should
69 primarily be on establishing a baseline dataset for the region on small phytoplankton, rather than
70 predictions that cannot be supported by the data presented (i.e. data from one year) and instead are based
71 on data from other regions.

72 → We agree with the reviewer’s opinion. So, we modified our manuscript to delete the prediction parts in
73 line 36-38, page 2 and line 309-314, page 13.

74 - There is seemingly an inconsistency (or at best, a lack of explanation) between the demonstrated
75 importance of small phytoplankton outside the polynya region and the claim that small phytoplankton
76 will grow in importance with climate change (won’t the non-polynya region decrease in size with
77 increased warming?).

78 → Polynya and non-polynya regions are different systems with different environmental conditions so that
79 we can not simply say that. That is a main reason for why we separated them in this study. Actually, the
80 data in Figure 7 included all stations from polynya and non-polynya regions. Anyway, we modified our
81 manuscript to delete the prediction parts.

82 - There is a large focus on the comparison of data inside and outside of the polynya region, but with
83 limited justification for this comparison, or discussion of how the polynya may be altered by climate

84 change. Ultimately I agree that this comparison is valuable, but primarily in the context of establishing a
85 baseline dataset for the region.

86 → We agree with the reviewer's opinion. So, we modified our manuscript to delete the prediction parts in
87 line 36-38, page 2 and line 309-314, page 13.

88

89 - The Results section needs to be reorganized (see suggestions below).

90 → We reorganized as reviewer suggested throughout the result section.

91 - There are numerous grammatical errors, some of which I have identified in the "Technical
92 Corrections" section.

93 → We checked and revised the grammatical errors throughout the text.

94

95 Specific Comments:

96 - Lines 54-62: I think it is important to indicate that Ducklow et al. (2007) and Montes-Hugo et al. (2009)
97 detail the western Antarctic Peninsula (WAP) that is a focus of the LTER (north of $\sim 68^{\circ}\text{S}$), and do not
98 include the Amundsen Sea region.

99 → We indicated that in line 56-57, page 3.

100 - Line 71: "in response to a regional warming trend" - I think this wording is too strong. Moline et al.
101 (2004) note the association between cryptophytes and low salinity water (likely glacial meltwater), and
102 hypothesize that cryptophytes will increase in importance given the predicted regional warming trend.
103 Regarding the association between cryptophytes and glacial meltwater, Moline et al. (2004) suggest that
104 this is salinity driven (they cite studies demonstrating cryptophytes tolerate/prefer lower salinity water), a
105 point that Moline made nearly a decade earlier (Moline and Prezelin 1996, MEPS).

106 → We deleted the sentence.

107 - Lines 71-73: re. an example of food web alteration due to a shift in phytoplankton community
108 composition to smaller cells at least provide the example that krill do not feed efficiently on cryptophytes
109 (see Moline et al. 2004 for references).

110 → We further discussed on that in line 321-325, page 13.

111 - Line 79: "environmental conditions" - could this not simply be referred to as climate change?

112 → Yes, it could. We revised it in line 81-82, page 4.

113 - Line 82: Consider renaming, e.g. "Water samples".

114 → We renamed it in line 85, page 4.

115 - Results section: this section is very tedious to read. Perhaps that is unavoidable given the results
116 presented (essentially a long list of averages and standard deviations). However, I think it would benefit
117 tremendously from some reorganization. All statistics should be reported in a consistent manner, e.g.

118 range followed by mean +/- SD in parentheses. Additionally, each topic has the same info presented, e.g.
 119 total/small cells, % contribution, inside/outside polynya. I think it would help guide the reader if this info
 120 was presented in a consistent order for each topic.

121 → We revised the result section as suggested.

122 - Lines 273-275: The authors present strong evidence that small phytoplankton contribute more in the
 123 non-polynya region than the polynya region. How might we expect the polynya to be altered with climate
 124 change? It seems reasonable to expect that the non-polynya region will decrease in size, and thus reduce
 125 the contribution of small phytoplankton. This is inconsistent with the stated motivation and implications
 126 of the paper (i.e. an increase in the contribution of small phytoplankton, and resulting decrease in primary
 127 production), and needs to be addressed.

128 → Actually, polynya and non-polynya regions are different systems with different environmental
 129 conditions so that we can not simply expect that. Actually, our non-polynya stations were not an ice free
 130 open ocean in this study (see Figure 1). Increasing polynya region altered with climate change could
 131 cause different conditions from previous original conditions. That is a main reason for why we separated
 132 them in this study. The data in Figure 7 included all stations from polynya and non-polynya regions.

133 - Lines 299-304: the prediction of Moline et al. (2004) for an increase in the contribution of smaller
 134 phytoplankton with expanding meltwater is for the portion of the WAP that is a focus of the LTER (north
 135 of 68S), and did not explicitly include the Amundsen Sea region. Do the authors have any evidence
 136 specific to their region of interest for a potential shift to smaller phytoplankton, as well as a driving
 137 mechanism? If not, I do not think they can make strong statements re. the future of Amundsen Sea
 138 phytoplankton community composition, as well as its impact on primary production (using the
 139 relationship in Fig. 7).

140 → We deleted the sentence.

141 - Lines 305-315: this discussion should include the fact that krill do not efficiently feed on small
 142 phytoplankton (see Moline et al. 2004 for references).

143 → We further discussed on that in line 323-325, page 13.

144

145 Technical Corrections:

146 - Lines 18-19: "Small-sized phytoplankton : : : ocean condition." - rephrase.

147 → We rephrased that in line 18-19, page 2.

148 - Line 45: refer to Fig. 1.

149 → We referred to Fig. 1 in line 45, page 3.

150 - Line 65: "In an expecting : : :" - rephrase.

151 → We rephrased that in line 67, page 3.

- 152 - Line 67: “In consistent : : :” - rephrase.
- 153 → We rephrased that in line 69-71, page 3.
- 154 - Line 73: “higher trophic levels” and “subsequent food web” are redundant.
- 155 → We deleted subsequent food web in line 322, page 13.
- 156 - Line 76: “what extend” - rephrase.
- 157 → We rephrased that in line 79, page 4.
- 158 - Line 78: “marine ecosystem : : : ongoing changes” - rephrase.
- 159 → We rephrased that in line 81-82, page 4.
- 160 - Lines 83-88: refer to Fig. 1 in here somewhere.
- 161 → We referred to Fig. 1 in line 87, page 4.
- 162 - Lines 91-22: “were belong” - rephrase.
- 163 → We rephrased that in line 95, page 5.
- 164 - Line 95: “biological and chemical property” - please be specific.
- 165 → Actually I tried to mention that other researchers collected water samples for their own biological and
- 166 chemical research. We deleted that since it might be confused.
- 167 - Lines 109-113: the information re. the isotope tracer technique, light depths, and light
- 168 sensor was already provided.
- 169 → We deleted the same information in line 113-117, page 5.
- 170 - Lines 137-138: “integrated from six different light depths” - change to “depth integrated”?
- 171 → We changed it to depth integrated in line 141, page 6, line 163, page 7, and line 183, page 8.
- 172 - Lines 141-143: “In the Amundsen Sea : : : 2014 : : :” – unnecessary info (the cruise location and date
- 173 has already been specified).
- 174 → We deleted unnecessary info in line 146-147, page 7.
- 175
- 176

177 **Small phytoplankton contribution to the total primary production in**
178 **the Amundsen Sea**

179
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192

193 **Abstract**

194 Small-sized phytoplankton ~~is~~are anticipated to be more important ~~for phytoplankton community~~
195 in a recent~~ly~~ changing ocean condition. However, little information on the contribution of small-sized
196 phytoplankton to overall phytoplankton production is currently available in the Amundsen Sea. To
197 determine the contributions of small-sized phytoplankton to total biomass and primary production, carbon
198 and nitrogen uptake rates of total and small-sized phytoplankton were obtained from 12 productivity
199 stations in the Amundsen Sea. The daily carbon uptake rates of total phytoplankton averaged in this study
200 were $0.42 \text{ g C m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.30 \text{ g C m}^{-2} \text{ d}^{-1}$) and $0.84 \text{ g C m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.18 \text{ g C m}^{-2} \text{ d}^{-1}$) whereas
201 the daily total nitrogen (nitrate and ammonium) uptake rates were $0.12 \text{ g N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.09 \text{ g N m}^{-2}$
202 d^{-1}) and $0.21 \text{ g N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.11 \text{ g N m}^{-2} \text{ d}^{-1}$), respectively for non-polynya and polynya regions,
203 which were within the ranges reported previously. Small phytoplankton contributed 26.9 % and 27.7 % to
204 the total carbon and nitrogen uptake rates of phytoplankton in this study, respectively, which were
205 relatively higher than the chlorophyll-a contribution (19.4 %) of small phytoplankton. For a comparison
206 of different regions, the contributions for chlorophyll-a concentration and primary production of small
207 phytoplankton averaged from all the non-polynya stations were 42.4 % and 50.8 %, which were
208 significantly higher than those (7.9 % and 14.9 %, respectively) in polynya region. A strong negative
209 correlation ($r^2 = 0.790$, $p < 0.05$) was found between the contributions of small phytoplankton and the
210 total daily primary production of phytoplankton in this study. This finding implies that daily primary
211 production decreases as small phytoplankton contribution increases, which is mainly due to the lower
212 carbon uptake rate of small phytoplankton than large phytoplankton. ~~Under ongoing environmental~~
213 ~~changes caused by global warming, a potential decrease of total primary production would be led by~~
214 ~~increasing contribution of small phytoplankton in the Amundsen Sea.~~

215

216

217 Keywords: Phytoplankton, Primary production, Polynya, Amundsen Sea

218

219 1. Introduction

220 The Amundsen Sea is located in the West Antarctica between the Ross Sea and Bellingshausen
221 Sea (Fig. 1), which is one of the least-biologically studied regions in the Southern Ocean. Recently
222 several international research programs (KOPRI Amundsen project, iSTAR, ASPIRE, and DynaLiFe)
223 were launched to understand this remote area. Field-measurement data revealed that annual primary
224 production of phytoplankton reaching to $220 \text{ g C m}^{-2} \text{ y}^{-1}$ in the Amundsen Sea polynya is as high as that
225 of Ross Sea polynya ($200 \text{ g C m}^{-2} \text{ y}^{-1}$) which was previously known for the highest productivity region in
226 the Southern Ocean (Lee et al., 2012). Given the fact that the ~~chl-a~~chlorophyll-a concentration averaged
227 from all the chlorophyll-a-measured stations was twice higher than that of the only productivity-
228 measured stations, Lee et al., (2012) argued that the annual production in the Amundsen Sea polynya
229 could be even two-fold higher than that of Ross Sea polynya.

230 Over the past several decades a rapid climate change has been detected and subsequently
231 physical changes have occurred in the marine ecosystem in the western Antarctic Peninsula (WAP)
232 mainly based on the results from Palmer Antarctic Long-Term Ecological Research project which focused
233 on the north of $\sim 69^\circ \text{S}$ (Rückamp et al. 2011; Ducklow et al. 2007; Montes-Hugo et al. 2009). Recent
234 studies revealed that the Thwaites Glacier in Pine Island Bay is retreating fast and the ice volume loss in
235 the nearby Getz Ice shelf is accelerating (Joughin et al., 2014; Paolo et al., 2015). Shoaling warm
236 Circumpolar Deep Water is believed to be a main reason for the ice sheet mass loss largely caused by ice
237 shelf basal melt underside of the ice shelves (Yager et al. 2012; Schmidtko et al. 2014). The climate
238 change from a cold-dry polar-type to a warm-humid sub-Antarctic-type drives subsequent changes in
239 ocean biological productivity along the WAP shelf over the recent three decades (Montes-Hugo et al.
240 2009).

241 Phytoplankton as the base of oceanic food webs can be an indicator for changes in marine
242 ecosystems responding to ~~current climate~~environmental changes (Moline et al., 2004; Wassman et al.,
243 2011; Arrigo et al., 2015). For example, a shift in phytoplankton community structure from large diatoms
244 to relatively small cryptophytes could be tightly associated with changes in glacial melt-water runoff and

245 ~~reduced surface water salinity (Moline et al., 2004). In an expecting warmer ocean condition, small sized~~
246 ~~phytoplankton is anticipated to contribute more to total phytoplankton community and thus marine~~
247 ~~ecosystems (Morán et al., 2010; Li et al., 2009; Lee et al., 2013). In consistent, Li et al. (2009) found~~
248 ~~increasing small sized phytoplankton in the Canada Basin in the Arctic Ocean under freshening surface~~
249 ~~waters which results in a stronger stratification and lower nutrient supply in the upper water column.~~
250 ~~Moreover, in the Antarctic Ocean, Moline et al. (2004) found a consistent transition from large diatoms to~~
251 ~~small cryptophytes associated with glacial melt water in the coastal waters along the Antarctic Peninsula~~
252 ~~in response to a regional warming trend. This change in dominant phytoplankton community from large~~
253 ~~to small cells will likely cause further alteration of higher trophic levels and subsequent food web (Moline~~
254 ~~et al., 2004).~~ ~~However~~ To date, little information on the contribution of small-sized phytoplankton to
255 primary production is available in the Antarctic Ocean (Saggiomo et al. 1998), especially in the
256 Amundsen Sea with a rapid melting of ice shelf (Yager et al. 2012; Schmidtko et al. 2014). Thus, the
257 main objective in this study is to determine that to what extent ~~dt~~ small-sized phytoplankton contributes to
258 overall total biomass and primary production in the Amundsen Sea to lay the groundwork for ~~the future~~
259 monitoring of marine ecosystem ~~change~~ responding to ~~ongoing~~ changes in environmental conditions.

260

261 **2. Materials and methods**

262 *2.1. Water ~~S~~sampleings*

263 Water samples were collected for carbon and nitrogen uptake measurements of phytoplankton in
264 the Amundsen Sea (Fig. 1) during the KOPRI Amundsen cruise from 1 to 15 January, 2014 onboard the
265 Korean Research Icebreaker ship *Araon*. Using a dual stable isotope technique (Lee et al., 2012; Kim et
266 al., 2015), the experiments of carbon and nitrogen uptake rates of phytoplankton were conducted at 12
267 selected productivity stations including 2 revisited-stations (St. 3-1 and St. 19-1) when on-deck
268 incubations were available during daytime at oceanographic survey stations. Based on sea ice
269 concentration data from National Snow & Ice Data Center during the cruise period (Fig. 1), our study
270 region was further separated into polynya and non-polynya areas for comparison based on sea ice

271 distribution and concentration during the cruise period. Four stations (St. 1, St. 2, St. 3, and St. 3-1)
272 among the 12 stations ~~were~~ belong to non-polynya region and the rest of the stations ~~were~~ belong to
273 polynya region.

274 After 6 light depths (100, 50, 30, 12, 5, and 1% penetration of the surface irradiance, PAR) were
275 determined with an LI-COR underwater 4π light sensor, water samples for the uptake experiments ~~as~~
276 ~~well as biological and chemical property analysis~~ were obtained from a CTD-rosette sampler system
277 equipped with 24 10-L Niskin bottles.

278 2.2. Total and size-fractionated chlorophyll-a concentration

279 Water samples for total and size-fractionated chlorophyll-a concentrations of phytoplankton were
280 obtained at the 12 productivity stations. Total chlorophyll-a concentrations were measured at six different
281 light depths (100, 50, 30, 12, 5 and 1% of PAR). For size-fractionated chlorophyll-a concentrations, water
282 samples were collected at three light depths (100, 30, and 1 %). Water samples (0.3–0.5 L) for total
283 chlorophyll-a concentrations were filtered using Whatman glass fiber filters (GF/F; 25 mm). For different
284 size-fractionated chlorophyll-a concentrations water samples (0.7–1 L) were passed sequentially through
285 20 and 5 μm Nucleopore filters (47 mm) and 0.7 μm GF/F filters (47 mm). After the filters were extracted
286 using the method described by Kim et al. (2015), all chlorophyll-a concentrations were subsequently
287 determined onboard using a Trilogy fluorometer (Turner Designs, USA). The methods and calculations
288 for chlorophyll-a were based on Parsons et al. (1984).

289 2.3. Carbon and nitrogen uptake experiments

290 ~~Carbon and nitrogen uptake experiments of phytoplankton were executed by a ^{13}C - ^{15}N dual~~
291 ~~isotope tracer technique previously applied for the Amundsen Sea (Lee et al. 2012; Kim et al. 2015). In~~
292 ~~this study, basically we followed same procedure of Lee et al. (2012). In brief, six light depths (100, 50,~~
293 ~~30, 12, 5, and 1%) were determined with an LI-COR underwater 4π light sensor (LI-COR Inc., Lincoln,~~
294 ~~Nebraska, USA) lowered with CTD/rosette sampler. Water sample from each light depth was transferred~~
295 into different screened polycarbonate incubation bottle (1 L) which matches with each light depth. The

296 productivity bottles were incubated in large polycarbonate material incubators cooled with running
297 surface seawater on deck under natural light conditions, after the water samples were inoculated with
298 labeled carbon ($\text{NaH}^{13}\text{CO}_3$) and nitrate (K^{15}NO_3) or ammonium ($^{15}\text{NH}_4\text{Cl}$) substrates. After 4–5 h
299 incubations, the incubated waters were well mixed and distributed into two filtration sets for the carbon
300 and nitrogen uptake rates of total ($> 0.7 \mu\text{m}$) and small-sized cells ($< 5 \mu\text{m}$). The incubated waters (0.3 L)
301 for total uptake rates were filtered through pre-combusted GF/F filters (24 mm diameter), whereas waters
302 samples (0.5 L) for the uptake rates of small-sized cells were passed through 5 μm Nuclepore filters (47
303 mm) to remove large-sized cells ($> 5 \mu\text{m}$) and then the filtrate was passed through GF/F (24 mm) for the
304 small-sized cells (Lee et al., 2013). The values for large phytoplankton in this study were obtained from
305 the difference between small and total fractions (Lee et al., 2013). The filters were immediately preserved
306 at -80°C until mass spectrometric analysis. After acid fuming overnight to remove carbonate, the
307 concentrations of particulate organic carbon (POC) and nitrogen (PON) and the abundance of ^{13}C and
308 ^{15}N were determined by a Finnigan Delta+XL mass spectrometer at the Alaska Stable Isotope Facility,
309 USA.

310 All contribution results of small phytoplankton in this study were estimated from comparison of
311 small phytoplankton to total phytoplankton integral values from 100 to 1 % light depth at each station
312 based on the trapezoidal rule. Daily carbon and nitrogen uptake rates of phytoplankton were based on our
313 hourly uptake rates measured in this study and a 24-h photoperiod per day during the summer period in
314 the Amundsen Sea (Lee et al., 2012).

315

316 3. Results

317 3.1. Chlorophyll-a, POC, and PON contributions of small phytoplankton

318 The depth-integrated total (large + small phytoplankton) chlorophyll-a concentration integrated
319 from six different light depths ranged from 11.1 to 80.3 mg chl-a m^{-2} (mean \pm S.D. = 57.4 ± 25.2 mg chl-a
320 m^{-2}), whereas small ($< 5 \mu\text{m}$) chlorophyll-a concentration ranged from 3.9 to 9.4 mg chl-a m^{-2} (~~mean \pm~~

321 ~~S.D. = 5.7 ± 1.7 mg chl-a m⁻²~~ in this study (Fig. 2). The ~~average~~ contribution of small phytoplankton to
 322 the total chlorophyll-a concentration was ~~4.9-76.5 %~~ ~~19.4 %~~ (~~S.D. = 19.4 ± 26.0 %~~) ~~ranging from 4.9 to~~
 323 ~~76.5 %~~. ~~In the Amundsen Sea,~~ Large phytoplankton (> 5 µm) were generally predominant
 324 (approximately 80 %) ~~during the cruise period in 2014~~ based on different-sized chlorophyll-a
 325 concentrations. For a regional comparison, the average contributions of small phytoplankton to the total
 326 chlorophyll-a concentration were 42.4 % (~~S.D. = ± 37.2 %~~) and 7.9 % (~~S.D. = ± 3.5 %~~) for non-polynya
 327 and polynya regions, respectively (Table 1). The chlorophyll-a contribution of small phytoplankton was
 328 larger in the non-polynya regionstations than the polynya regionstations although they were not
 329 significantly different (t-test, p = 0.16).

330 The total integral POC concentration of phytoplankton displayed no large spatial variation
 331 ranging from 4.72 to 9.22 mg C m⁻² (Fig. 3). In comparison, the total integral PON concentration of
 332 phytoplankton ~~ranged from was~~ 0.76 ~~to~~ 1.74 mg C m⁻². The POC contribution of small phytoplankton
 333 ~~ranged from was~~ 30.7 ~~to~~ 65.5 % (~~mean ± S.D. = 41.1 ± 10.6 %~~), whereas the PON contribution ~~ranged~~
 334 ~~similarly from~~ ~~was~~ 30.8 ~~to~~ 67.2 % (~~mean ± S.D. = 41.3 ± 11.5 %~~) in the Amundsen Sea (Fig. 3).
 335 Specifically, the POC and PON contributions of small phytoplankton averaged from all the productivity
 336 stations in the polynya regionstations were 36.9 % (~~S.D. = ± 4.6 %~~) and 37.0 % (~~S.D. = ± 6.9 %~~),
 337 respectively. ~~In comparison, the POC and PON contributions of small phytoplankton averaged from the~~
 338 ~~non-polynya stations, whereas they~~ were 49.5 % (~~S.D. = ± 14.4 %~~) and 50.0 % (~~S.D. = ± 15.1 %~~),
 339 respectively in the non-polynya region (Table 1).

340 3.2. Carbon uptake rate contributions of small phytoplankton

341 The depth-integrated total daily carbon uptake rates of phytoplankton (large + small
 342 phytoplankton) ~~integrated from six different light depths ranged from was~~ 150.4 ~~to~~ 1213.4 mg C m⁻² d⁻¹
 343 ~~with an average of~~ (696.5 mg C m⁻² d⁻¹ (~~S.D. = ± 298.4 mg C m⁻² d⁻¹~~) in this study (Fig. 4). In contrast,
 344 the rates of small phytoplankton ranged between 58.6 and 266.4 mg C m⁻² d⁻¹ ~~with an average of~~ (124.9
 345 ~~mg C m⁻² d⁻¹~~ (~~S.D. = ± 62.4 mg C m⁻² d⁻¹~~). Small phytoplankton contributed 26.9 % (~~S.D. = ± 29.3%~~) to
 346 total daily carbon uptake rate of total phytoplankton.

347 Specifically, the total daily carbon uptake rates of phytoplankton ~~ranged from was~~ 150.4 to 796.4
 348 mg C m⁻² d⁻¹ ~~with an average of~~ (415.0 mg C m⁻² d⁻¹ (S.D. = ± 298.2 mg C m⁻² d⁻¹) in the non-polynya
 349 region, whereas ~~they ranged from it was~~ 654.8 to 1213.4 mg C m⁻² d⁻¹ ~~with an average of~~ (837.3 mg C m⁻²
 350 d⁻¹ (S.D. = ± 184.1 mg C m⁻² d⁻¹) in the polynya region. The total daily carbon uptake rates of
 351 phytoplankton were significantly higher (t-test, p < 0.05) in the polynya regionstations than the non-
 352 polynya regionstations. The rates of small phytoplankton ~~ranged between was~~ 58.6 and 193.6 mg C m⁻²
 353 d⁻¹ ~~with an average of~~ (126.5 mg C m⁻² d⁻¹ (S.D. = ± 55.2 mg C m⁻² d⁻¹) in the non-polynya region,
 354 whereas ~~they ranged from it was~~ 62.2 to 266.4 mg C m⁻² d⁻¹ ~~with an average of~~ (124.1 mg C m⁻² d⁻¹ (S.D.
 355 = ± 69.3 mg C m⁻² d⁻¹) in the polynya region. The daily carbon uptake rates of small phytoplankton were
 356 not significantly different (t-test, p > 0.05) between the polynya and non-polynya stations. The average
 357 contributions of small phytoplankton to total daily carbon uptake rates were 50.8 % (S.D. = ± 42.8 %) and
 358 14.9 % (S.D. = ± 8.4 %), respectively for the non-polynya and polynya regions (Table 1). The average
 359 contributions were largely different between the polynya and non-polynya regions but they were not
 360 statistically significant (t-test, p > 0.05).

361 3.3. Nitrogen uptake rate contributions of small phytoplankton

362 The depth-integrated total daily nitrate uptake rates of phytoplankton (large + small
 363 phytoplankton) ~~ranged from was~~ 34.0 to 174.2 mg N m⁻² d⁻¹ ~~with an average of~~ (93.7 mg N m⁻² d⁻¹ (S.D. =
 364 ± 43.2 mg N m⁻² d⁻¹), whereas the rates of small phytoplankton ~~ranged from was~~ 6.1 to 40.9 mg N m⁻² d⁻¹
 365 ~~with an average of~~ (19.0 mg N m⁻² d⁻¹ (S.D. = ± 11.3 mg N m⁻² d⁻¹) in this study (Fig. 5). Small
 366 phytoplankton contributed 21.5 % (S.D. = ± 11.1 %) to total daily nitrate uptake rates. In comparison, the
 367 total daily ammonium uptake rates of phytoplankton ranged from 12.4 to 173.8 mg N m⁻² d⁻¹ ~~with an~~
 368 ~~average of~~ (86.7 mg N m⁻² d⁻¹ (S.D. = ± 75.9 mg N m⁻² d⁻¹), whereas the rates of small phytoplankton
 369 ranged from 9.1 to 81.1 mg N m⁻² d⁻¹ ~~with an average of~~ (25.7 mg N m⁻² d⁻¹ (S.D. = ± 21.1 mg N m⁻² d⁻¹)
 370 in this study (Fig. 6). Small phytoplankton contributed 38.7 % (S.D. = ± 24.9 %) to total daily ammonium
 371 uptake rates. The contributions of small phytoplankton were significantly higher in ammonium uptake
 372 rate than nitrate uptake rate (t-test, p < 0.05).

Specifically for different regions, the total daily nitrate uptake rates of phytoplankton ranged from 34.0 to 142.1 mg N m⁻² d⁻¹ with an average of 71.9 mg N m⁻² d⁻¹ (S.D. = ± 48.4 mg N m⁻² d⁻¹) in the non-polynya region whereas they ranged from 44.2 to 174.2 mg N m⁻² d⁻¹ with an average of 104.6 mg N m⁻² d⁻¹ (S.D. = ± 39.0 mg N m⁻² d⁻¹) in the polynya region, respectively. In comparison, the daily nitrate uptake rates of small phytoplankton ranged from 7.5 to 26.6 mg N m⁻² d⁻¹ with an average of 16.7 mg N m⁻² d⁻¹ (S.D. = ± 7.8 mg N m⁻² d⁻¹) and from 6.1 to 40.9 mg N m⁻² d⁻¹ with an average of 20.1 mg N m⁻² d⁻¹ (S.D. = ± 13.1 mg N m⁻² d⁻¹), respectively for the non-polynya and polynya regions. The contributions of small phytoplankton to the total daily nitrate uptake rates were 28.2 % (S.D. = ± 15.9 %) in the non-polynya region and 18.1 % (S.D. = ± 6.8 %) in the polynya region, respectively (Table 1). The total daily ammonium uptake rates of total phytoplankton ranged between 12.3 and 106.1 mg N m⁻² d⁻¹ (mean ± S.D. = 49.7 ± 41.2 mg N m⁻² d⁻¹) in the non-polynya region and between 18.1 and 269.3 mg N m⁻² d⁻¹ (mean ± S.D. = 105.2 ± 84.6 mg N m⁻² d⁻¹) in the polynya region. In comparison, the rates of small phytoplankton ranged between 9.1 and 22.4 mg N m⁻² d⁻¹ (mean ± S.D. = 15.8 ± 6.4 mg N m⁻² d⁻¹) in the non-polynya region and between 9.9 and 81.1 mg N m⁻² d⁻¹ (mean ± S.D. = 30.7 ± 24.5 mg N m⁻² d⁻¹) in the polynya region. Small phytoplankton contributed 52.8 % (S.D. = ± 40.5 %) and 31.6 % (S.D. = ± 10.1 %) to the total daily ammonium uptake rates in the non-polynya and polynya regions, respectively which were not significantly different (t-test, p = 0.37).

The total integral daily nitrogen uptake rate (nitrate + ammonium uptake rates) of phytoplankton ranged from 46.4 to 443.5 mg N m⁻² d⁻¹ (mean ± S.D. = 180.4 ± 106.7 mg N m⁻² d⁻¹) in this study. For the non-polynya and polynya regions, they ranged from 46.4 to 248.1 mg N m⁻² d⁻¹ (mean ± S.D. = 121.6 ± 89.3 mg N m⁻² d⁻¹) and from 91.7 to 443.5 mg N m⁻² d⁻¹ (mean ± S.D. = 209.8 ± 107.3 mg N m⁻² d⁻¹), respectively. In comparison, the total integral daily nitrogen uptake rates of small phytoplankton ranged from 16.6 to 46.6 mg N m⁻² d⁻¹ (mean ± S.D. = 32.5 ± 13.2 mg N m⁻² d⁻¹) and from 17.6 to 122.0 mg N m⁻² d⁻¹ (mean ± S.D. = 50.8 ± 32.4 mg N m⁻² d⁻¹) for the non-polynya and polynya regions, respectively. Small phytoplankton contributed 36.2 % (S.D. = ± 23.0 %) to the total integral daily nitrogen uptake rates in the non-polynya region, whereas they contributed 23.5 % (S.D. = ±

399 6.0 %) for the polynya region (Table 1). The integral daily nitrogen uptake rates and contributions of
 400 small phytoplankton were not statistically different between the non-polynya and polynya regions.

402 4. Discussion and conclusion

403 The total daily carbon uptake rates of phytoplankton averaged for the non-polynya and polynya
 404 regions were $0.42 \text{ g C m}^{-2} \text{ d}^{-1}$ (~~S.D. = ± 0.30 g C m⁻² d⁻¹~~) and $0.84 \text{ g C m}^{-2} \text{ d}^{-1}$ (~~S.D. = ± 0.18 g C m⁻² d⁻¹~~),
 405 respectively in this study. According to the previous reports in the Amundsen Sea (Lee et al., 2012; Kim
 406 et al., 2015), the total daily carbon uptake rates ranged from 0.2 to 0.12 g C m⁻² d⁻¹ in the non-polynya
 407 region. Our rate ($0.42 \text{ g C m}^{-2} \text{ d}^{-1}$) in the non-polynya region is somewhat higher than those reported
 408 previously but they are not significantly different (t-test, $p = 0.77$). In comparison, our total daily carbon
 409 uptake rate in the polynya region ($0.84 \text{ g C m}^{-2} \text{ d}^{-1}$) is within the range between Lee et al. (2012; 2.2 g C
 410 $\text{m}^{-2} \text{ d}^{-1}$) and Kim et al. (2015; $0.2 \text{ g C m}^{-2} \text{ d}^{-1}$). The carbon uptake rates of phytoplankton in Lee et al.
 411 (2012) and Kim et al. (2015) were measured during December 21, 2010-January 23, 2011 and February
 412 11 to March 14, 2012, respectively. Our measurements in this study were executed mainly during January
 413 1-15, 2014. For the Amundsen polynya region, a large seasonal variation in the total daily carbon uptake
 414 rate of phytoplankton was already reported by Kim et al. (2015) and Arrigo et al. (2012) based on filed-
 415 measured data and satellite-derived approach, respectively. It is appeared that this seasonal variation
 416 largely depends on the bloom stage of phytoplankton which peaks during the late December-January and
 417 terminates at late February (Arrigo and van Dijken 2003; Arrigo et al., 2012; Kim et al., 2015). ~~The~~
 418 ~~carbon uptake rates of phytoplankton in Lee et al. (2012) and Kim et al. (2015) were measured during~~
 419 ~~December 21-January 23, 2010 and February 11 to March 14, 2012, respectively. Our measurements in~~
 420 ~~this study were executed mainly during January 1-15, 2014.~~

421 The total daily nitrogen uptake rates of phytoplankton were $0.12 \text{ g N m}^{-2} \text{ d}^{-1}$ (~~S.D. = ± 0.09 g N m⁻²~~
 422 d^{-1}) and $0.21 \text{ g N m}^{-2} \text{ d}^{-1}$ (~~S.D. = ± 0.11 g N m⁻² d⁻¹~~) for non-polynya and polynya regions, respectively
 423 in this study. Previous studies reported that the total daily nitrogen uptake rates in non-polynya region
 424 were $0.24 \text{ g N m}^{-2} \text{ d}^{-1}$ during Dec. 21, 2010-Jan. 23, 2011~~in 2010/2011~~ and $0.04 \text{ g N m}^{-2} \text{ d}^{-1}$ during Feb. 11

425 | ~~to Mar. 14, 2012~~~~in 2012~~ whereas the uptake rates in polynya region were $0.93 \text{ g N m}^{-2} \text{ d}^{-1}$ in 2010/2011
426 | and $0.06 \text{ g N m}^{-2} \text{ d}^{-1}$ in 2012 in the Amundsen Sea (Lee et al., 2012; Kim et al., 2015). Our total daily
427 | nitrogen uptake rates of phytoplankton in non-polynya and polynya regions were between the two
428 | previous studies (Lee et al., 2012; Kim et al., 2015). Based on the nitrate and ammonium uptake rates in
429 | this study, f -ratios (nitrate uptake rate/nitrate+ammonium uptake rates) averaged for non-polynya and
430 | polynya regions were 0.62 (~~S.D. = ± 0.08~~) and 0.54 (~~S.D. = ± 0.20~~), respectively. These ratios also were
431 | between the two previous studies. Although they were not significant different because of a large spatial
432 | variation, larger f -ratios in non-polynya than in polynya region are consistent with the results of the
433 | previous studies (Lee et al., 2012; Kim et al., 2015). At this point, we do not have a solid explanation for
434 | that but a further future study is needed for the higher f -ratio mechanism in non-polynya region.

435 | The percent contributions of small phytoplankton in terms of chlorophyll-a, POC/PON, daily
436 | carbon and nitrogen uptake rates are shown in Table 1. The overall contribution of small phytoplankton to
437 | the total chlorophyll-a concentration for all the productivity stations was 19.4% (~~S.D. = ± 26.0 %~~) which
438 | is significantly (t-test, $p < 0.05$) lower than the POC contribution ($41.1 \pm 10.6 \%$). This is consistent with
439 | the result in the Chukchi Sea, Arctic Ocean reported by Lee et al. (2013). They explained that higher POC
440 | content per chlorophyll-a unit of small phytoplankton could cause their higher POC contribution (Lee et
441 | al., 2013). Given C/N ratio (~~mean ± S.D. =~~ 6.6 ± 0.6) and $\delta^{13}\text{C}$ (~~mean ± S.D. =~~ $-25.9 \pm 1.0 \text{ ‰}$) of sample
442 | filters attained for POC and PON in this study, our filtered samples are believed to be mainly
443 | phytoplankton-originated POC and PON (Kim et al., 2016). Thus, a significant potential overestimated
444 | POC contribution of non-phytoplankton materials could be excluded for the higher POC contribution than
445 | chlorophyll-a contribution of small phytoplankton. Therefore, small phytoplankton contributions based on
446 | conventional assessments of chlorophyll-a concentration might lead an underestimated contribution of
447 | small phytoplankton (Lee et al., 2013). In fact, several authors argued that chlorophyll-a concentration
448 | might be not a good index for phytoplankton biomass since it largely depends on environmental factors
449 | such as nutrient and light conditions as well as dominant groups and physiological status of
450 | phytoplankton (Desortová 1981; Behrenfeld et al., 2005; Kruskopf and Flynn, 2006; Behrenfeld and Boss

451 2006). However, the effects of non-phytoplankton carbon materials such as extracellular carbon mucilage
452 can not be completely excluded for the POC contribution as discussed below.

453 The overall contributions of carbon and nitrogen (nitrate and ammonium) uptake rates of small
454 phytoplankton at all the productivity stations in this study are similar with 26.9 % and 27.7 %,
455 respectively. These contributions are relatively higher than the chlorophyll-a contribution of small
456 phytoplankton but they are not statistically different (t-test, $p > 0.05$). In general, the contribution of daily
457 ammonium uptake rate of small phytoplankton ~~are~~ is significantly (t-test, $p < 0.05$) higher than the
458 contribution of daily nitrate uptake rate of small phytoplankton at all the stations in this study. It is well-
459 known for the ammonium preference of small phytoplankton in various regions (Koike et al., 1986;
460 Tremblay et al., 2000, Lee et al., 2008; Lee et al., 2013).

461 In terms of the contributions in different regions, all the contributions (~~C~~chlorophyll-a, POC/PON,
462 carbon and nitrogen uptake rates) of small phytoplankton were higher in the non-polynya region than in
463 the polynya region (Table 1). In addition, the chlorophyll-a contribution of small phytoplankton (~~mean \pm~~
464 ~~S.D. =~~ 7.9 ± 3.5 %) was significantly (t-test, $p < 0.01$) lower than the POC contribution (~~mean \pm~~
465 ~~S.D. =~~ 36.9 ± 4.6 %) in the polynya region, whereas they were not statistically different in the non-polynya
466 region (Table 1). This indicates that small phytoplankton contributed more to the total POC than the
467 chlorophyll-a concentration in the polynya region. We do not have species compositions of phytoplankton
468 in this study, but previous results reported that *Phaeocystis* sp. are dominant in the Amundsen Sea
469 polynya region ~~whereas diatoms are relatively dominant in the non-polynya regions~~ (Lee et al., 2012).
470 Generally, *Phaeocystis* spp. release a large portion (up to 46 %) of extracellular carbon mucilage which
471 makes their colonial form (Matrai et al., 1995). This non-phytoplankton carbon material without
472 chlorophyll-a might cause a higher POC contribution of small phytoplankton in the polynya region during
473 this study. In fact, the contribution of the daily carbon uptake rates of small phytoplankton (14.9 ± 8.4 %)
474 was not as high as the POC contribution (36.9 ± 4.6 %) in the polynya region. The chlorophyll-a
475 contributions of small phytoplankton were lower than ~~that~~ those of the daily carbon uptake rate in this
476 study, which is consistent with the results from polynya and marginal ice zone stations in the Ross Sea,

477 Antarctica during austral spring and summer (Saggiomo et al., 1998). They reported that the chlorophyll-a
478 and primary production contributions of pico-phytoplankton ($< 2 \mu\text{m}$) were 29 % and 40 % at polynya
479 stations whereas the contributions were 17 % and 32 % at marginal ice zone stations, respectively. In the
480 polynya region, they found much higher contributions in chlorophyll-a and primary production of small
481 phytoplankton than those in this study although their size of small phytoplankton is somewhat smaller
482 than our size ($< 5 \mu\text{m}$).

483 We found a strong negative correlation ($r^2 = 0.790$, $p < 0.05$) between the productivity
484 contributions of small phytoplankton and total daily carbon uptake rates of phytoplankton in the
485 Amundsen Sea (Fig. 7), which implies that daily primary production decreases as small phytoplankton
486 contribution increases. This is mainly because of the relatively lower carbon uptake rate of small
487 phytoplankton than large phytoplankton in the Chukchi Sea, Arctic Ocean reported by Lee et al. (2013).
488 ~~Moline et al. (2004) suggested that further warming air temperatures will increase inputs of glacial~~
489 ~~melting water and subsequently increase the contributions of small phytoplankton over large~~
490 ~~phytoplankton community (Moline et al. 2004). If these small phytoplankton were dominant under~~
491 ~~ongoing more melting conditions of glaciers, a potential increasing contribution of small phytoplankton~~
492 ~~might cause a subsequent decrease in the total primary production of phytoplankton in the Amundsen Sea~~
493 ~~based on this study in Figure 7.~~

494 In respect to food quality of small phytoplankton as a basic food source to herbivores,
495 macromolecular compositions such as proteins, lipids, and carbohydrates as photosynthetic-end products
496 will be needed for better understanding alterations of small cells-dominant marine ecosystem in response
497 to ~~ongoing~~ environmental changes (Lee et al., 2013). According to Kang et al. (accepted), small
498 phytoplankton— assimilate more food materials and calorific contents per unit of chlorophyll-a
499 concentration and thus provide more contributions in respect to energy aspect than other phytoplankton
500 community in the East/Japan Sea. However, ~~This change in dominant phytoplankton community from~~
501 ~~large to small cells will likely cause further alteration of higher trophic levels because of prey size itself~~
502 ~~available to higher trophic grazers and subsequent food web (Moline et al., 2004).~~ In conclusion,

503 monitoring the contributions of small-sized phytoplankton to total biomass and primary production of
504 total phytoplankton community could be important as a valuable indicator to sense future environmental
505 changes in marine ecosystem ~~under ongoing various climate associated environmental changes. Moreover,~~
506 ~~further detailed studies for macromolecular compositions of small phytoplankton will be necessary for the~~
507 ~~anticipating small dominant ecosystem under warming oceans.~~

508

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513

514 **5. References**

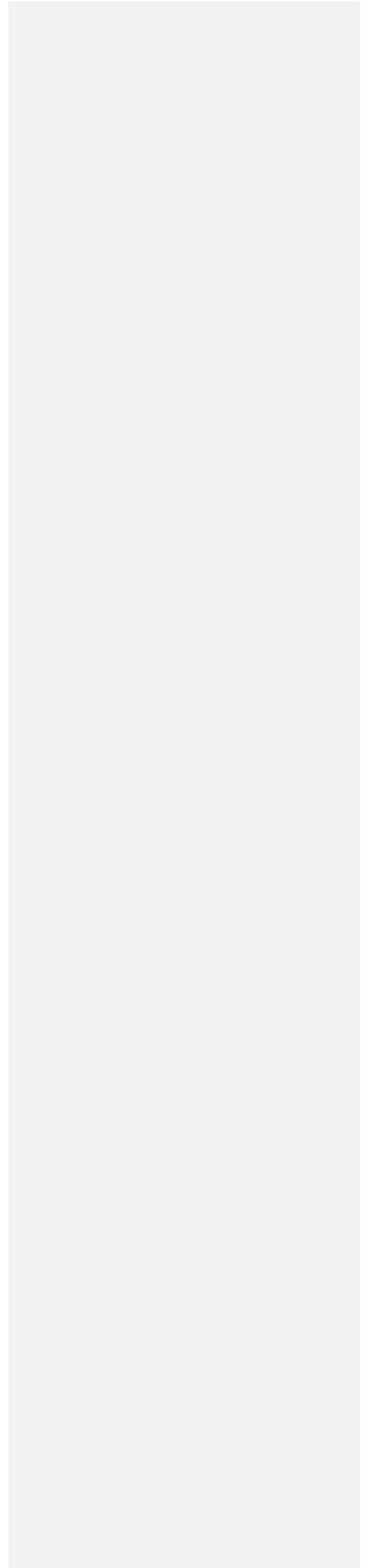
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587

588



590 **Table caption**

591 Table 1. Contributions (%) ~~of chlorophyll-a, POC, PON, and carbon and nitrogen uptake rates~~ of small
592 phytoplankton in the Amundsen Sea. Contributions of chlorophyll-a, POC, PON, and carbon and nitrogen
593 uptake rates were derived from water euphotic column-integrated values averaged from stations.

594

595 **Figure captions**

596 Fig. 1. Sampling locations in the Amundsen Sea. Red closed circles represent productivity stations. Sea
597 ice concentration data during the cruise period from Nimbus-7 SMMR and DMSP SSM/I-
598 SSMIS Passive Microwave data provided by National Snow & Ice Data Center.

599 Fig. 2. Water column-integrated chlorophyll-a concentration at the productivity stations in the Amundsen
600 Sea.

601 Fig. 3. Water column-integrated concentrations of POC and PON of small and large phytoplankton.

602 Fig. 4. Water column-integrated daily carbon uptake rates of small and large phytoplankton.

603 Fig. 5. Water column-integrated daily nitrate uptake rates of small and large phytoplankton.

604 Fig. 6. Water column-integrated daily ammonium uptake rates of small and large phytoplankton.

605 Fig. 7. Relationship between productivity contributions of small phytoplankton and the total daily carbon
606 uptake rates of phytoplankton (large + small). The total daily carbon uptake rates were
607 transformed into natural logs for a linear regression.

608

609

610

Table 1. Contributions (%) of small phytoplankton in the Amundsen Sea. Contributions of chlorophyll-a, POC, PON, and carbon and nitrogen uptake rates were derived from water euphotic column-integrated values averaged from stations.

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	Chlorophyll-a	POC	PON	Daily carbon uptake rate	Daily nitrate uptake rate	Daily ammonium uptake rate	Total nitrogen uptake rate
All stations	19.4 ± 26.0	41.1 ± 10.6	41.3 ± 11.5	26.9 ± 29.3	21.5 ± 11.1	38.7 ± 24.9	27.7 ± 14.4
Non-polynya	42.4 ± 37.2	49.5 ± 14.4	50.0 ± 15.1	50.8 ± 42.8	28.2 ± 15.9	52.8 ± 40.5	36.2 ± 23.0
Polynya	7.9 ± 3.5	36.9 ± 4.6	37.0 ± 6.9	14.9 ± 8.4	18.1 ± 6.8	31.6 ± 10.1	23.5 ± 6.0

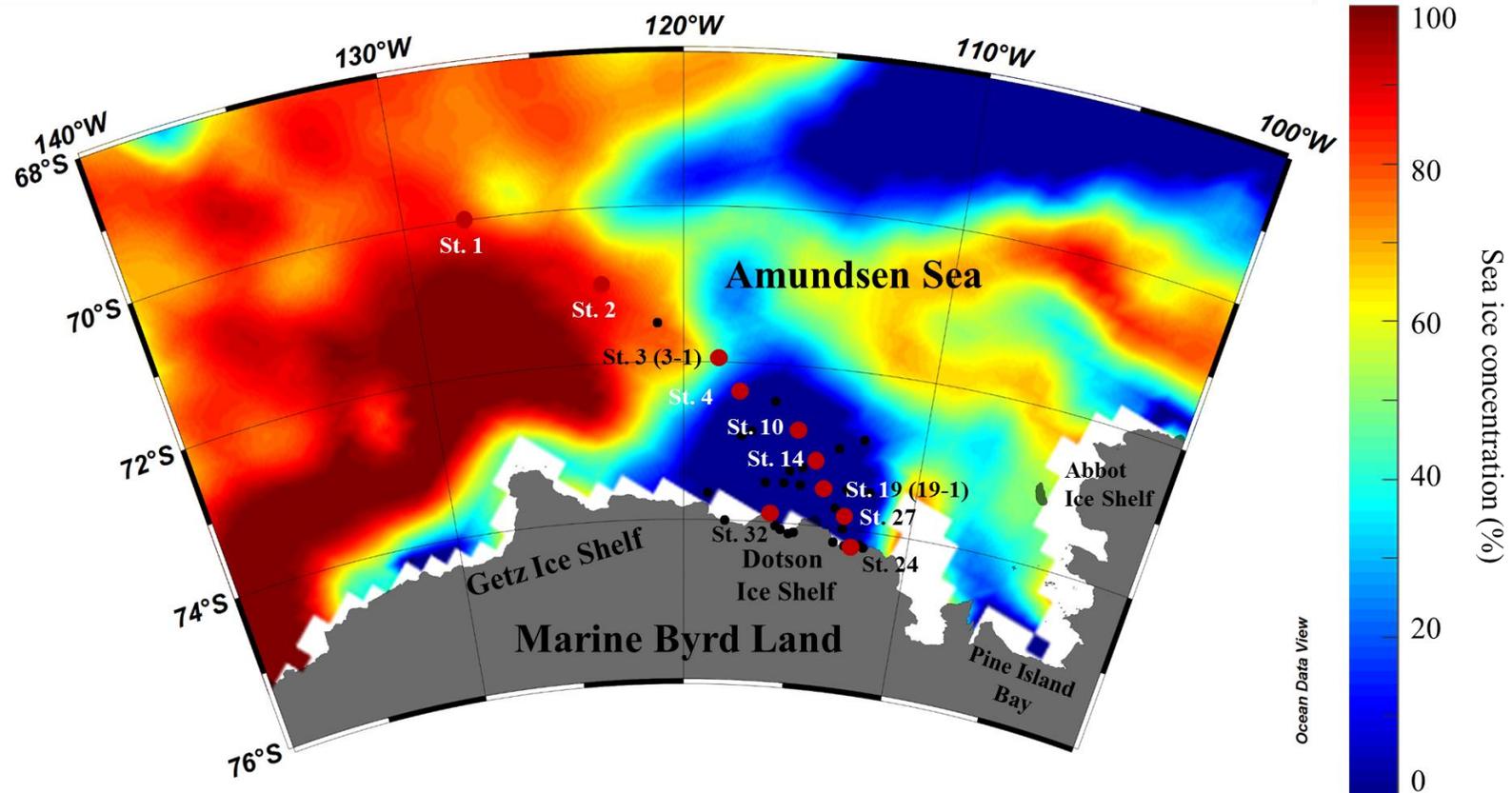


Fig. 1. Sampling locations in the Amundsen Sea. Red closed circles represent productivity stations. Sea ice concentration data during the cruise period from Nimbus-7 SMMR and DMSP SSM/I-SSMIS Passive Microwave data provided by National Snow & Ice Data Center.

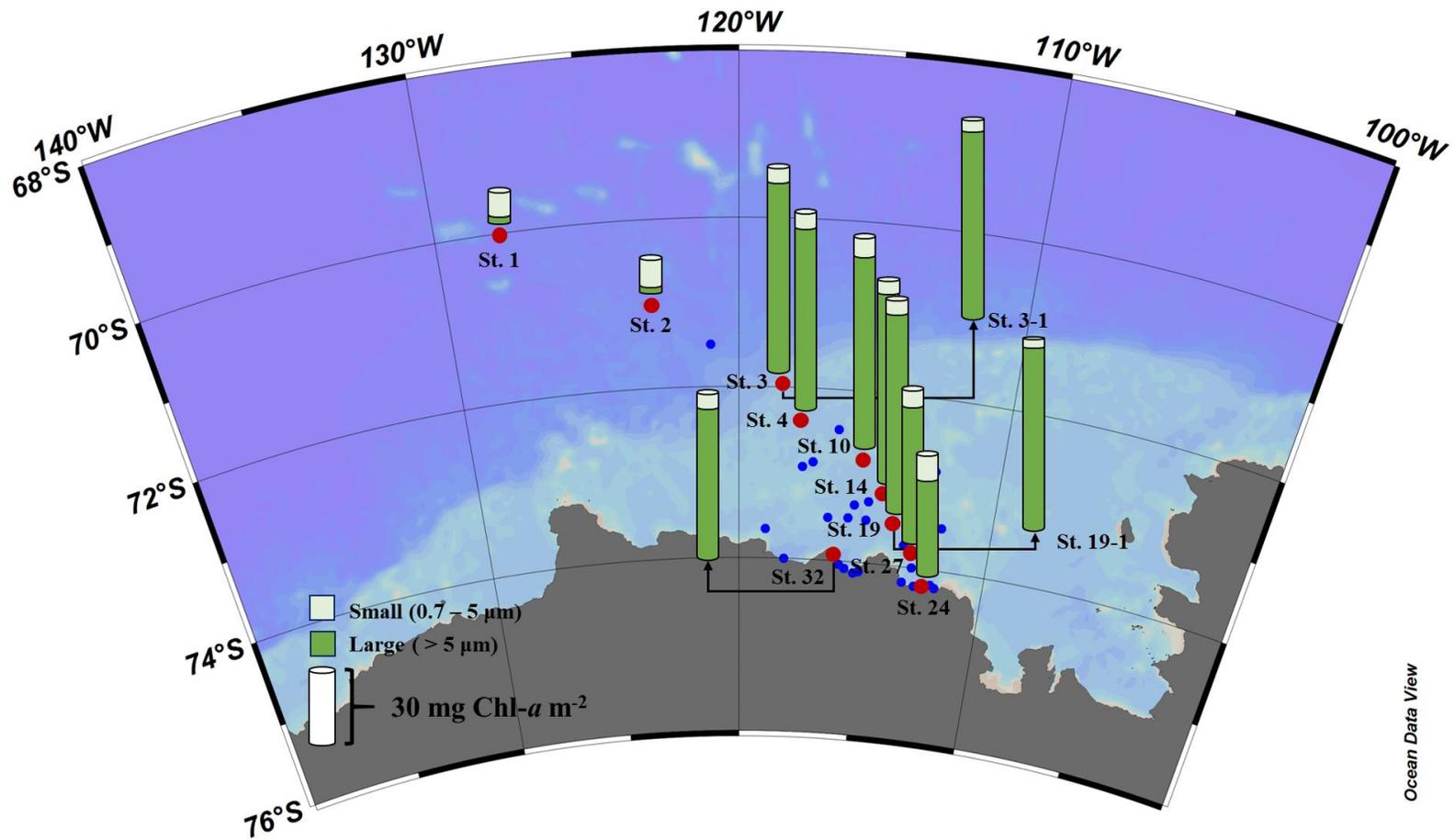


Fig. 2. Water column-integrated chlorophyll-a concentration at the productivity stations in the Amundsen Sea.

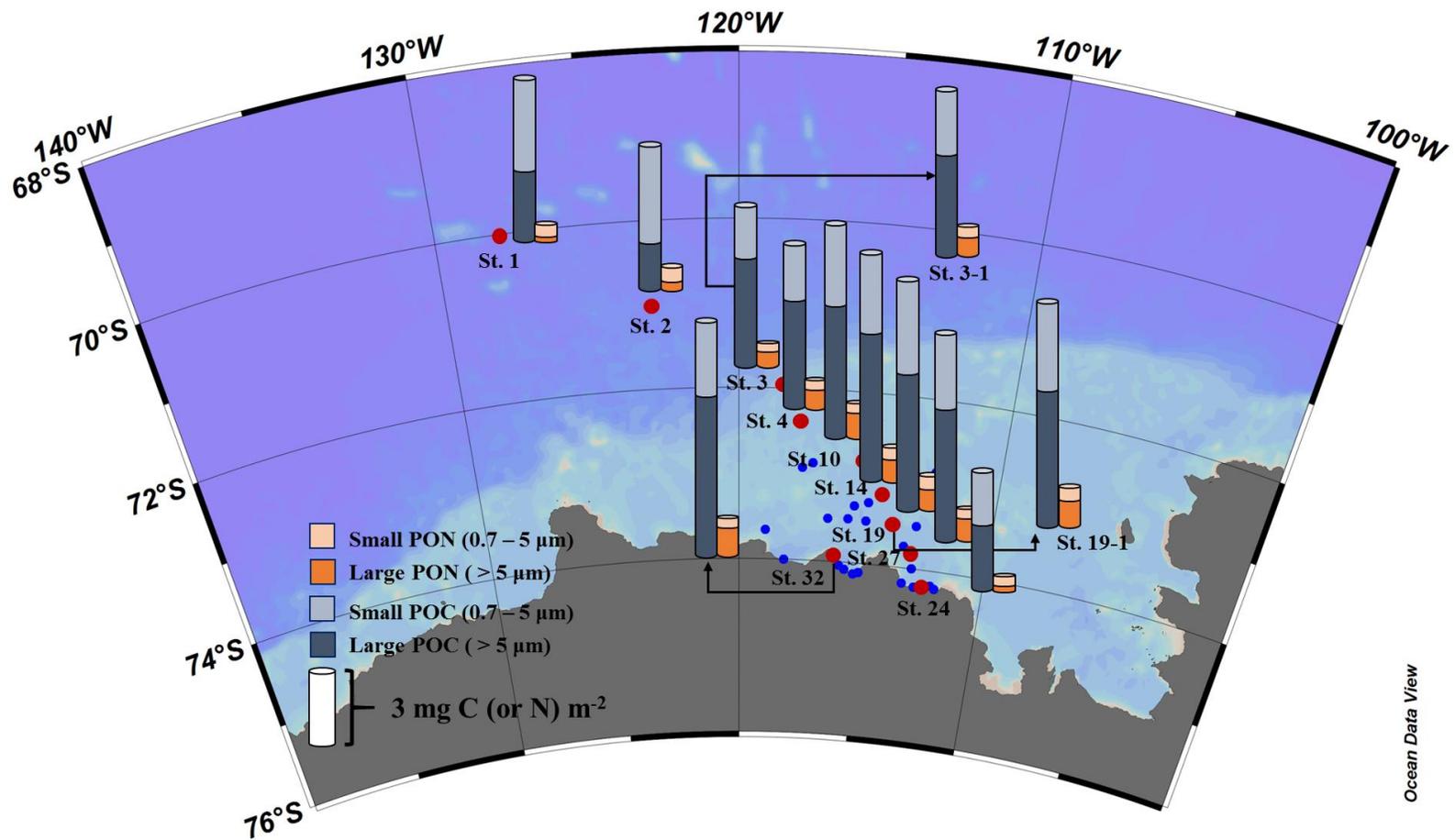


Fig. 3. Water column-integrated concentrations of POC and PON of small and large phytoplankton.

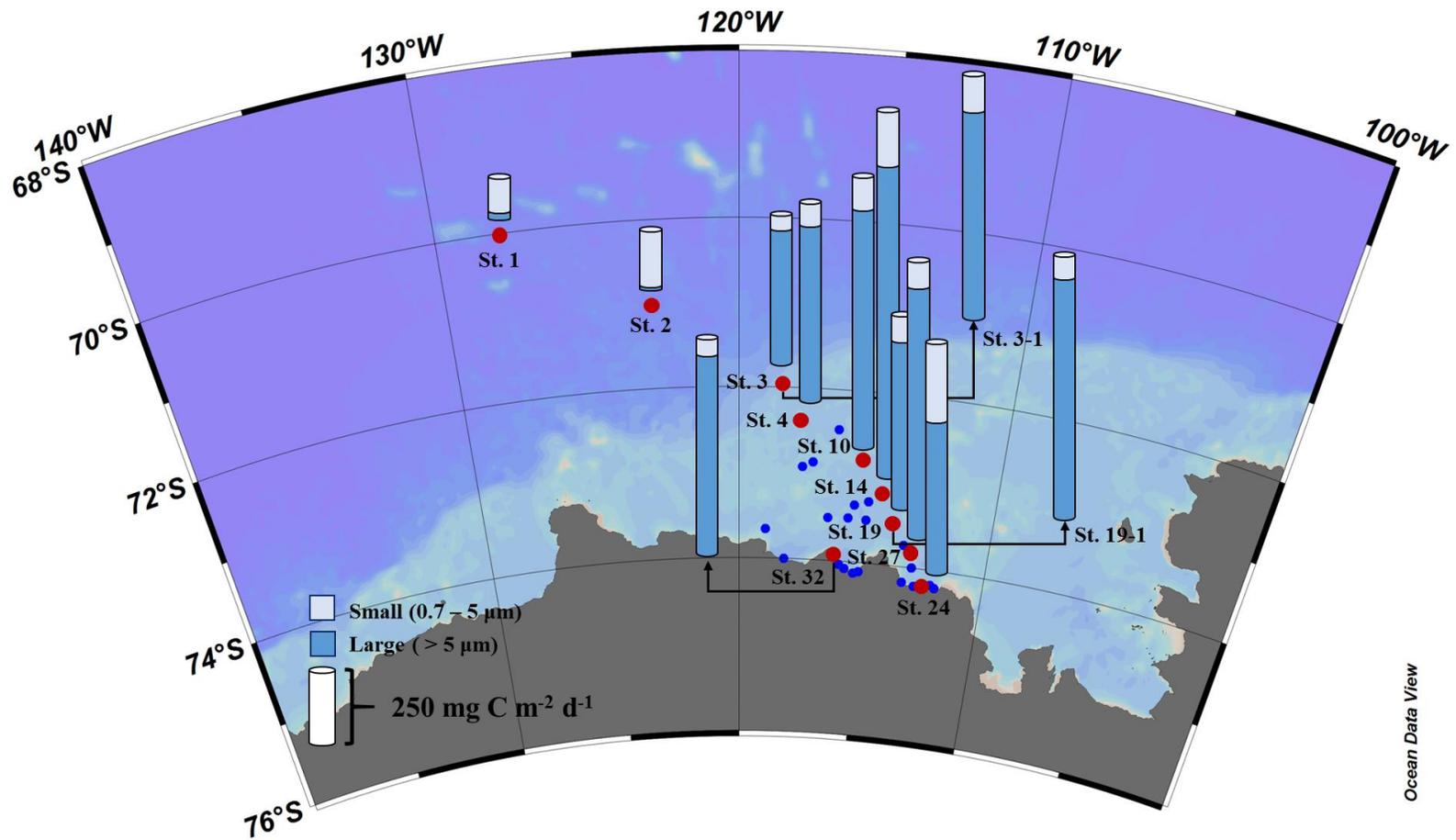


Fig. 4. Water column-integrated daily carbon uptake rates of small and large phytoplankton.

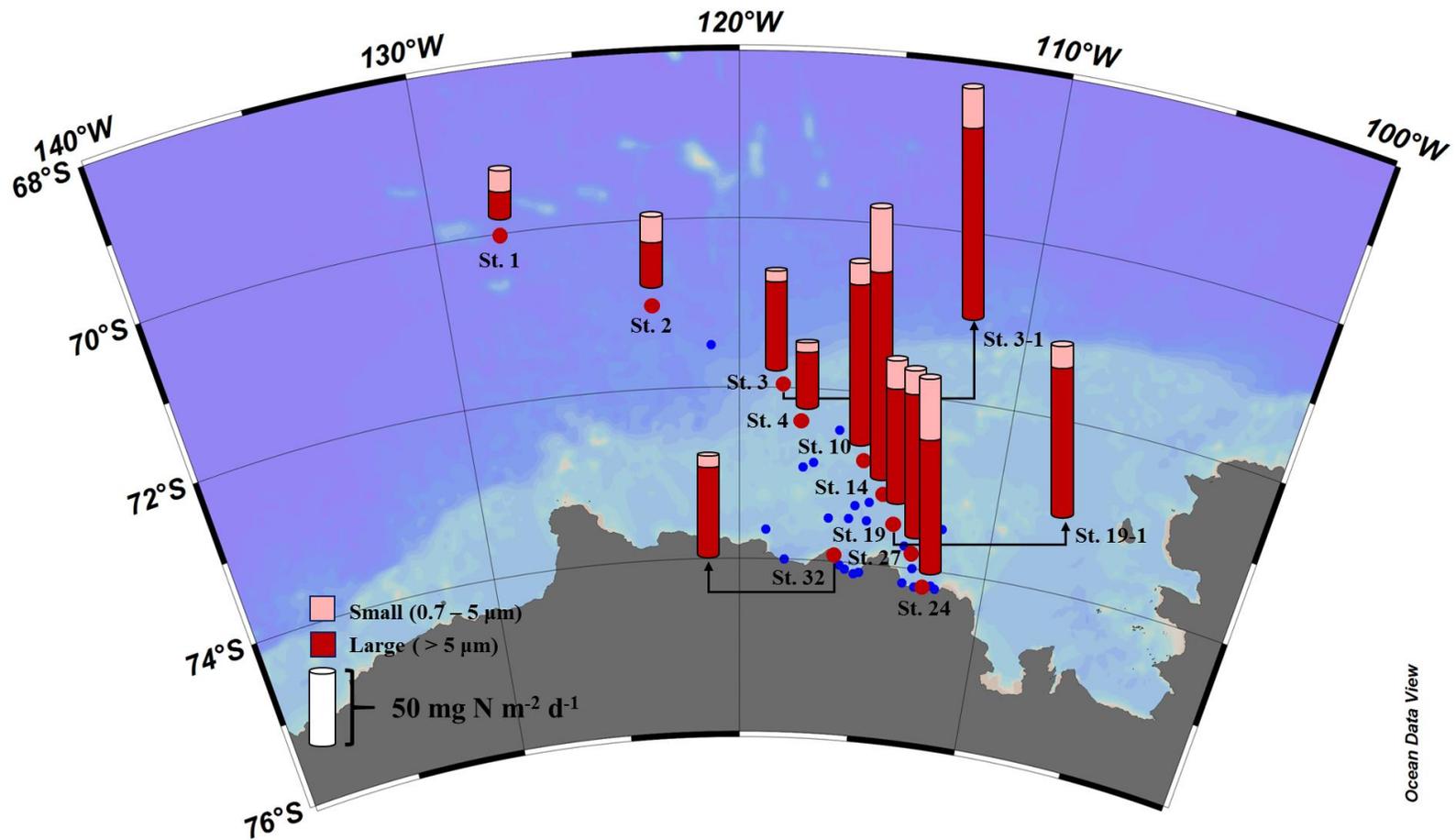


Fig. 5. Water column-integrated daily nitrate uptake rates of small and large phytoplankton.

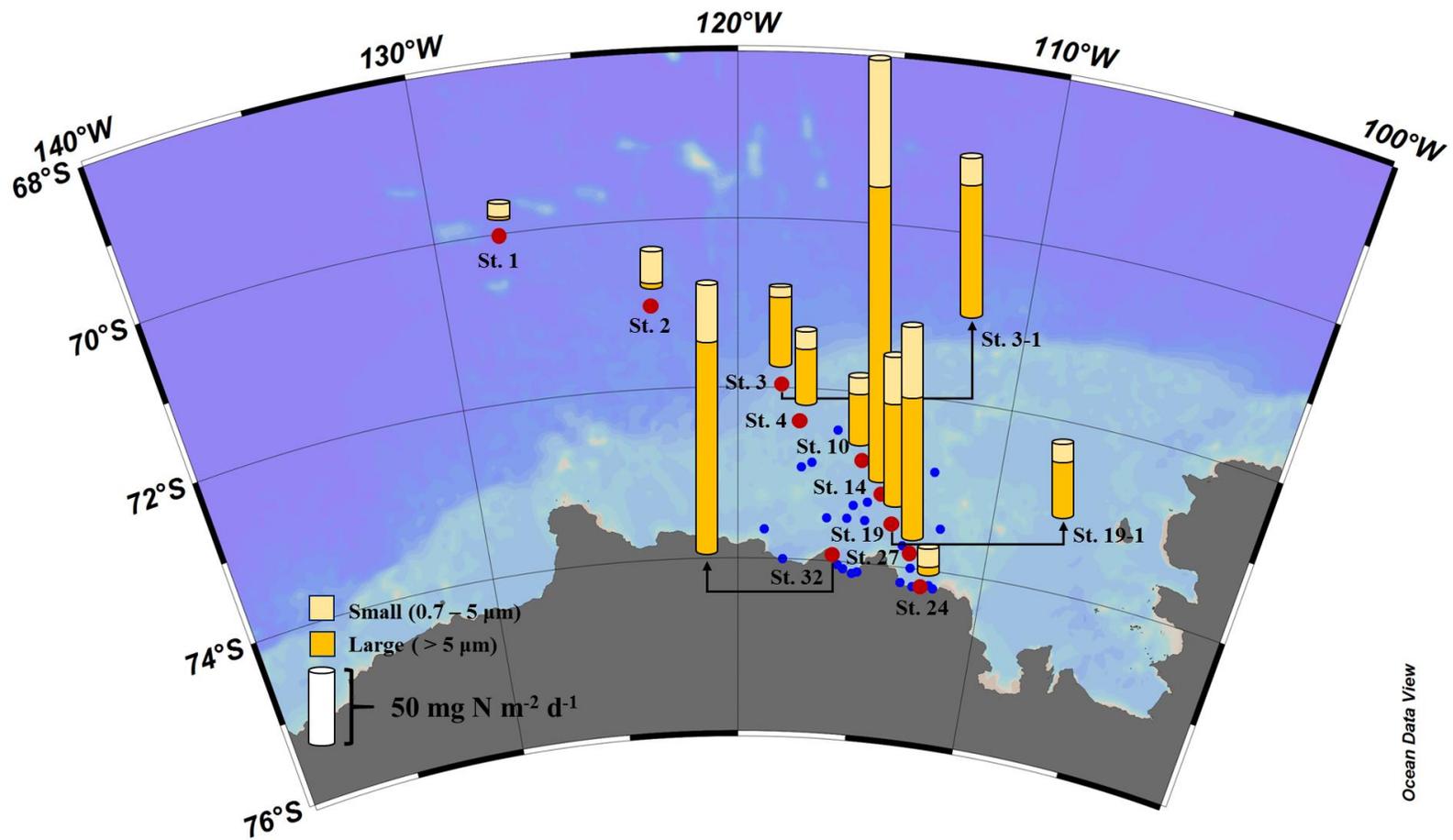


Fig. 6. Water column-integrated daily ammonium uptake rates of small and large phytoplankton.

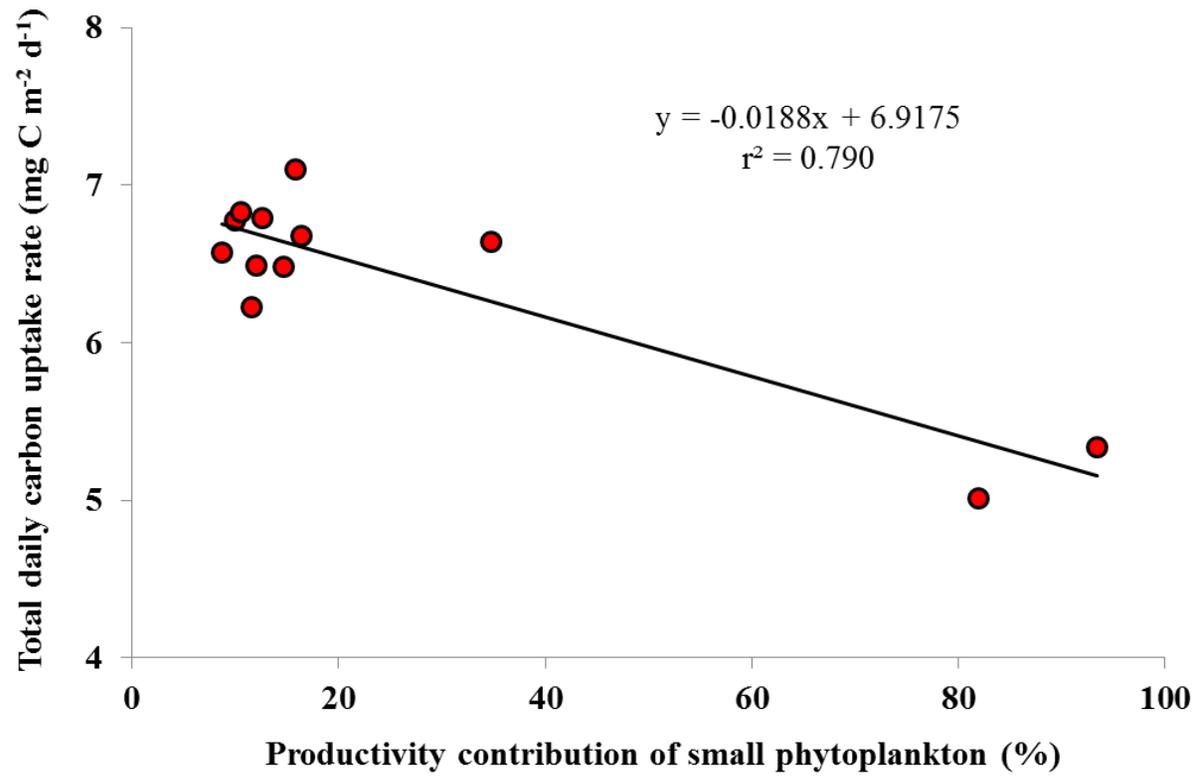


Fig. 7. Relationship between productivity contributions of small phytoplankton and the total daily carbon uptake rates of phytoplankton (large + small). The total daily carbon uptake rates were transformed into natural logs for a linear regression.