

1 **Small phytoplankton contribution to the total primary production in**
2 **the Amundsen Sea**

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17 **Abstract**

18 Small-sized phytoplankton are anticipated to be more important in a recently changing ocean
19 condition. However, little information on the contribution of small-sized phytoplankton to overall
20 phytoplankton production is currently available in the Amundsen Sea. To determine the contributions of
21 small-sized phytoplankton to total biomass and primary production, carbon and nitrogen uptake rates of
22 total and small-sized phytoplankton were obtained from 12 productivity stations in the Amundsen Sea.
23 The daily carbon uptake rates of total phytoplankton averaged in this study were $0.42 \text{ g C m}^{-2} \text{ d}^{-1}$ (S.D. =
24 $\pm 0.30 \text{ g C m}^{-2} \text{ d}^{-1}$) and $0.84 \text{ g C m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.18 \text{ g C m}^{-2} \text{ d}^{-1}$) whereas the daily total nitrogen
25 (nitrate and ammonium) uptake rates were $0.12 \text{ g N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.09 \text{ g N m}^{-2} \text{ d}^{-1}$) and $0.21 \text{ g N m}^{-2} \text{ d}^{-1}$
26 1 (S.D. = $\pm 0.11 \text{ g N m}^{-2} \text{ d}^{-1}$), respectively for non-polynya and polynya regions, which were within the
27 ranges reported previously. Small phytoplankton contributed 26.9 % and 27.7 % to the total carbon and
28 nitrogen uptake rates of phytoplankton in this study, respectively, which were relatively higher than the
29 chlorophyll-a contribution (19.4 %) of small phytoplankton. For a comparison of different regions, the
30 contributions for chlorophyll-a concentration and primary production of small phytoplankton averaged
31 from all the non-polynya stations were 42.4 % and 50.8 %, which were significantly higher than those
32 (7.9 % and 14.9 %, respectively) in polynya region. A strong negative correlation ($r^2 = 0.790$, $p < 0.05$)
33 was found between the contributions of small phytoplankton and the total daily primary production of
34 phytoplankton in this study. This finding implies that daily primary production decreases as small
35 phytoplankton contribution increases, which is mainly due to the lower carbon uptake rate of small
36 phytoplankton than large phytoplankton.

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39 **Keywords:** Phytoplankton, Primary production, Polynya, Amundsen Sea

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41 **1. Introduction**

42 The Amundsen Sea is located in the West Antarctica between the Ross Sea and Bellingshausen
43 Sea (Fig. 1), which is one of the least-biologically studied regions in the Southern Ocean. Recently
44 several international research programs (KOPRI Amundsen project, iSTAR, ASPIRE, and DynaLiFe)
45 were launched to understand this remote area. Field-measurement data revealed that annual primary
46 production of phytoplankton reaching to $220 \text{ g C m}^{-2} \text{ y}^{-1}$ in the Amundsen Sea polynya is as high as that
47 of Ross Sea polynya ($200 \text{ g C m}^{-2} \text{ y}^{-1}$) which was previously known for the highest productivity region in
48 the Southern Ocean (Lee et al., 2012). Given the fact that the chlorophyll-a concentration averaged from
49 all the chlorophyll-measured stations was twice higher than that of the only productivity-measured
50 stations, Lee et al., (2012) argued that the annual production in the Amundsen Sea polynya could be even
51 two-fold higher than that of Ross Sea polynya.

52 Over the past several decades a rapid climate change has been detected and subsequently
53 physical changes have occurred in the marine ecosystem in the western Antarctic Peninsula (WAP)
54 mainly based on the results from Palmer Antarctic Long-Term Ecological Research project which focused
55 on the north of $\sim 69^\circ \text{S}$ (Ducklow et al. 2007; Montes-Hugo et al. 2009). Recent studies revealed that the
56 Thwaites Glacier in Pine Island Bay is retreating fast and the ice volume loss in the nearby Getz Ice shelf
57 is accelerating (Joughin et al., 2014; Paolo et al., 2015). Shoaling warm Circumpolar Deep Water is
58 believed to be a main reason for the ice sheet mass loss largely caused by ice shelf basal melt underside of
59 the ice shelves (Yager et al. 2012; Schmidtke et al. 2014). The climate change from a cold-dry polar-type
60 to a warm-humid sub-Antarctic-type drives subsequent changes in ocean biological productivity along the
61 WAP shelf over the recent three decades (Montes-Hugo et al. 2009).

62 Phytoplankton as the base of oceanic food webs can be an indicator for changes in marine
63 ecosystems responding to environmental changes (Moline et al., 2004; Wassman et al., 2011; Arrigo et al.,
64 2015). For example, a shift in phytoplankton community structure from large diatoms to relatively small
65 cryptophytes could be tightly associated with changes in glacial melt-water runoff and reduced surface
66 water salinity (Moline et al., 2004). To date, little information on the contribution of small-sized

67 phytoplankton to primary production is available in the Antarctic Ocean (Saggiomo et al. 1998),
68 especially in the Amundsen Sea with a rapid melting of ice shelf (Yager et al. 2012; Schmidtko et al.
69 2014). Thus, the main objective in this study is to determine that to what extent small-sized
70 phytoplankton contributes to overall total biomass and primary production in the Amundsen Sea to lay the
71 groundwork for monitoring of marine ecosystem responding to changes in environmental conditions.

72

73 **2. Materials and methods**

74 *2.1. Water samples*

75 Water samples were collected for carbon and nitrogen uptake measurements of phytoplankton in
76 the Amundsen Sea (Fig. 1) during the KOPRI Amundsen cruise from 1 to 15 January, 2014 onboard the
77 Korean Research Icebreaker ship *Araon*. Using a dual stable isotope technique (Lee et al., 2012; Kim et
78 al., 2015), the experiments of carbon and nitrogen uptake rates of phytoplankton were conducted at 12
79 selected productivity stations including 2 revisited-stations (St. 3-1 and St. 19-1) when on-deck
80 incubations were available during daytime at oceanographic survey stations. Based on sea ice
81 concentration data from National Snow & Ice Data Center during the cruise period (Fig. 1), our study
82 region was further separated into polynya and non-polynya areas for comparison based on sea ice
83 distribution and concentration during the cruise period. Four stations (St. 1, St. 2, St. 3, and St. 3-1)
84 among the 12 stations belong to non-polynya region and the rest of the stations belong to polynya region.

85 After 6 light depths (100, 50, 30, 12, 5, and 1% penetration of the surface irradiance, PAR) were
86 determined with an LI-COR underwater 4π light sensor, water samples for the uptake experiments were
87 obtained from a CTD-rosette sampler system equipped with 24 10-L Niskin bottles.

88 *2.2. Total and size-fractionated chlorophyll-a concentration*

89 Water samples for total and size-fractionated chlorophyll-a concentrations of phytoplankton were
90 obtained at the 12 productivity stations. Total chlorophyll-a concentrations were measured at six different
91 light depths (100, 50, 30, 12, 5 and 1% of PAR). For size-fractionated chlorophyll-a concentrations, water
92 samples were collected at three light depths (100, 30, and 1 %). Water samples (0.3–0.5 L) for total

93 chlorophyll-a concentrations were filtered using Whatman glass fiber filters (GF/F; 25 mm). For different
94 size-fractionated chlorophyll-a concentrations water samples (0.7–1 L) were passed sequentially through
95 20 and 5 μm Nucleopore filters (47 mm) and 0.7 μm GF/F filters (47 mm). After the filters were extracted
96 using the method described by Kim et al. (2015), all chlorophyll-a concentrations were subsequently
97 determined onboard using a Trilogy fluorometer (Turner Designs, USA). The methods and calculations
98 for chlorophyll-a were based on Parsons et al. (1984).

99 *2.3. Carbon and nitrogen uptake experiments*

100 Water sample from each light depth was transferred into different screened polycarbonate
101 incubation bottle (1 L) which matches with each light depth. The productivity bottles were incubated in
102 large polycarbonate material incubators cooled with running surface seawater on deck under natural light
103 conditions, after the water samples were inoculated with labeled carbon ($\text{NaH}^{13}\text{CO}_3$) and nitrate (K^{15}NO_3)
104 or ammonium ($^{15}\text{NH}_4\text{Cl}$) substrates. After 4–5 h incubations, the incubated waters were well mixed and
105 distributed into two filtration sets for the carbon and nitrogen uptake rates of total ($> 0.7 \mu\text{m}$) and small-
106 sized cells ($< 5 \mu\text{m}$). The incubated waters (0.3 L) for total uptake rates were filtered through pre-
107 combusted GF/F filters (24 mm diameter), whereas waters samples (0.5 L) for the uptake rates of small-
108 sized cells were passed through 5 μm Nucleopore filters (47 mm) to remove large-sized cells ($> 5 \mu\text{m}$) and
109 then the filtrate was passed through GF/F (24 mm) for the small-sized cells (Lee et al., 2013). The values
110 for large phytoplankton in this study were obtained from the difference between small and total fractions
111 (Lee et al., 2013). The filters were immediately preserved at -80°C until mass spectrometric analysis.
112 After acid fuming overnight to remove carbonate, the concentrations of particulate organic carbon
113 (POC) and nitrogen (PON) and the abundance of ^{13}C and ^{15}N were determined by a Finnigan Delta+XL
114 mass spectrometer at the Alaska Stable Isotope Facility, USA.

115 All contribution results of small phytoplankton in this study were estimated from comparison of
116 small phytoplankton to total phytoplankton integral values from 100 to 1 % light depth at each station
117 based on the trapezoidal rule. Daily carbon and nitrogen uptake rates of phytoplankton were based on our

118 hourly uptake rates measured in this study and a 24-h photoperiod per day during the summer period in
119 the Amundsen Sea (Lee et al., 2012).

120

121 **3. Results**

122 3.1. Chlorophyll-a, POC, and PON contributions of small phytoplankton

123 The depth-integrated total (large + small phytoplankton) chlorophyll-a concentration ranged from
124 11.1 to 80.3 mg chl-a m⁻² (mean ± S.D. = 57.4 ± 25.2 mg chl-a m⁻²), whereas small (< 5 µm) chlorophyll-
125 a concentration ranged from 3.9 to 9.4 mg chl-a m⁻² (5.7 ± 1.7 mg chl-a m⁻²) in this study (Fig. 2). The
126 contribution of small phytoplankton to the total chlorophyll-a concentration was 4.9-76.5 % (19.4 ±
127 26.0 %). Large phytoplankton (> 5 µm) were generally predominant (approximately 80 %) based on
128 different-sized chlorophyll-a concentrations. For a regional comparison, the average contributions of
129 small phytoplankton to the total chlorophyll-a concentration were 42.4 % (± 37.2 %) and 7.9 % (± 3.5 %)
130 for non-polynya and polynya regions, respectively (Table 1). The chlorophyll-a contribution of small
131 phytoplankton was larger in the non-polynya region than the polynya region although they were not
132 significantly different (t-test, p = 0.16).

133 The total integral POC concentration of phytoplankton displayed no large spatial variation
134 ranging from 4.72 to 9.22 mg C m⁻² (Fig. 3). In comparison, the total integral PON concentration of
135 phytoplankton was 0.76-1.74 mg C m⁻². The POC contribution of small phytoplankton was 30.7-65.5 %
136 (41.1 ± 10.6 %), whereas the PON contribution was 30.8-67.2 % (41.3 ± 11.5 %) in the Amundsen Sea
137 (Fig. 3). Specifically, the POC and PON contributions of small phytoplankton averaged from all the
138 productivity stations in the polynya region were 36.9 % (± 4.6 %) and 37.0 % (± 6.9 %), respectively,
139 whereas they were 49.5 % (± 14.4 %) and 50.0 % (± 15.1 %), respectively in the non-polynya region
140 (Table 1).

141 3.2. Carbon uptake rate contributions of small phytoplankton

142 The depth-integrated total daily carbon uptake rate of phytoplankton (large + small phytoplankton)
143 was 150.4-1213.4 mg C m⁻² d⁻¹ (696.5 ± 298.4 mg C m⁻² d⁻¹) in this study (Fig. 4). In contrast, the rates of

144 small phytoplankton ranged between 58.6 and 266.4 mg C m⁻² d⁻¹ (124.9 ± 62.4 mg C m⁻² d⁻¹). Small
145 phytoplankton contributed 26.9 % (± 29.3%) to total daily carbon uptake rate of total phytoplankton.

146 Specifically, the total daily carbon uptake rate of phytoplankton was 150.4-796.4 mg C m⁻² d⁻¹
147 (415.0 ± 298.2 mg C m⁻² d⁻¹) in the non-polynya region, whereas it was 654.8-1213.4 mg C m⁻² d⁻¹ (837.3
148 ± 184.1 mg C m⁻² d⁻¹) in the polynya region. The total daily carbon uptake rates of phytoplankton were
149 significantly higher (t-test, p < 0.05) in the polynya region than the non-polynya region. The rate of small
150 phytoplankton was 58.6-193.6 mg C m⁻² d⁻¹ (126.5 ± 55.2 mg C m⁻² d⁻¹) in the non-polynya region,
151 whereas it was 62.2-266.4 mg C m⁻² d⁻¹ (124.1 ± 69.3 mg C m⁻² d⁻¹) in the polynya region. The daily
152 carbon uptake rates of small phytoplankton were not significantly different (t-test, p > 0.05) between the
153 polynya and non-polynya stations. The average contributions of small phytoplankton to total daily carbon
154 uptake rates were 50.8 % (± 42.8 %) and 14.9 % (± 8.4 %), respectively for the non-polynya and polynya
155 regions (Table 1). The average contributions were largely different between the polynya and non-polynya
156 regions but they were not statistically significant (t-test, p > 0.05).

157 3.3. Nitrogen uptake rate contributions of small phytoplankton

158 The depth-integrated total daily nitrate uptake rate of phytoplankton (large + small phytoplankton)
159 was 34.0-174.2 mg N m⁻² d⁻¹ (93.7 ± 43.2 mg N m⁻² d⁻¹), whereas the rate of small phytoplankton was 6.1-
160 40.9 mg N m⁻² d⁻¹ (19.0 ± 11.3 mg N m⁻² d⁻¹) in this study (Fig. 5). Small phytoplankton contributed 21.5 %
161 (± 11.1 %) to total daily nitrate uptake rates. In comparison, the total daily ammonium uptake rates of
162 phytoplankton ranged from 12.4 to 173.8 mg N m⁻² d⁻¹ (86.7 ± 75.9 mg N m⁻² d⁻¹), whereas the rates of
163 small phytoplankton ranged from 9.1 to 81.1 mg N m⁻² d⁻¹ (25.7 ± 21.1 mg N m⁻² d⁻¹) in this study (Fig. 6).
164 Small phytoplankton contributed 38.7 % (± 24.9 %) to total daily ammonium uptake rates. The
165 contributions of small phytoplankton were significantly higher in ammonium uptake rate than nitrate
166 uptake rate (t-test, p < 0.05).

167 For different regions, the total daily nitrate uptake rates of phytoplankton were 34.0-142.1 mg N
168 m⁻² d⁻¹ (71.9 ± 48.4 mg N m⁻² d⁻¹) in the non-polynya region and 44.2-174.2 mg N m⁻² d⁻¹ (104.6 ± 39.0
169 mg N m⁻² d⁻¹) in the polynya region, respectively. In comparison, the daily nitrate uptake rates of small

170 phytoplankton were 7.5-26.6 mg N m⁻² d⁻¹ (16.7 ± 7.8 mg N m⁻² d⁻¹) and 6.1-40.9 mg N m⁻² d⁻¹ (20.1 ±
171 13.1 mg N m⁻² d⁻¹), respectively for the non-polynya and polynya regions. The contributions of small
172 phytoplankton to the total daily nitrate uptake rates were 28.2 % (± 15.9 %) in the non-polynya region and
173 18.1 % (± 6.8 %) in the polynya region, respectively (Table 1). The total daily ammonium uptake rates of
174 total phytoplankton were 12.3 and 106.1 mg N m⁻² d⁻¹ (49.7 ± 41.2 mg N m⁻² d⁻¹) in the non-polynya
175 region and 18.1-269.3 mg N m⁻² d⁻¹ (105.2 ± 84.6 mg N m⁻² d⁻¹) in the polynya region. In comparison, the
176 rates of small phytoplankton ranged between 9.1 and 22.4 mg N m⁻² d⁻¹ (15.8 ± 6.4 mg N m⁻² d⁻¹) in the
177 non-polynya region and between 9.9 and 81.1 mg N m⁻² d⁻¹ (30.7 ± 24.5 mg N m⁻² d⁻¹) in the polynya
178 region. Small phytoplankton contributed 52.8 % (± 40.5 %) and 31.6 % (± 10.1 %) to the total daily
179 ammonium uptake rates in the non-polynya and polynya regions, respectively which were not
180 significantly different (t-test, p = 0.37).

181 The total integral daily nitrogen uptake rate (nitrate + ammonium uptake rates) of phytoplankton
182 was 46.4-443.5 mg N m⁻² d⁻¹ (180.4 ± 106.7 mg N m⁻² d⁻¹) in this study. For the non-polynya and polynya
183 regions, they were 46.4-248.1 mg N m⁻² d⁻¹ (121.6 ± 89.3 mg N m⁻² d⁻¹) and 91.7-443.5 mg N m⁻² d⁻¹
184 (209.8 ± 107.3 mg N m⁻² d⁻¹), respectively. In comparison, the total integral daily nitrogen uptake rates of
185 small phytoplankton were 16.6 to 46.6 mg N m⁻² d⁻¹ (32.5 ± 13.2 mg N m⁻² d⁻¹) and 17.6-122.0 mg N m⁻²
186 d⁻¹ (50.8 ± 32.4 mg N m⁻² d⁻¹) for the non-polynya and polynya regions, respectively. Small
187 phytoplankton contributed 36.2 % (± 23.0 %) to the total integral daily nitrogen uptake rates in the non-
188 polynya region, whereas they contributed 23.5 % (± 6.0 %) for the polynya region (Table 1). The integral
189 daily nitrogen uptake rates and contributions of small phytoplankton were not statistically different
190 between the non-polynya and polynya regions.

191

192 **4. Discussion and conclusion**

193 The total daily carbon uptake rates of phytoplankton averaged for the non-polynya and polynya
194 regions were 0.42 g C m⁻² d⁻¹ (± 0.30 g C m⁻² d⁻¹) and 0.84 g C m⁻² d⁻¹ (± 0.18 g C m⁻² d⁻¹), respectively in
195 this study. According to the previous reports in the Amundsen Sea (Lee et al., 2012; Kim et al., 2015), the

196 total daily carbon uptake rates ranged from 0.2 to 0.12 g C m⁻² d⁻¹ in the non-polynya region. Our rate
197 (0.42 g C m⁻² d⁻¹) in the non-polynya region is somewhat higher than those reported previously but they
198 are not significantly different (t-test, p = 0.77). In comparison, our total daily carbon uptake rate in the
199 polynya region (0.84 g C m⁻² d⁻¹) is within the range between Lee et al. (2012; 2.2 g C m⁻² d⁻¹) and Kim et
200 al. (2015; 0.2 g C m⁻² d⁻¹). The carbon uptake rates of phytoplankton in Lee et al. (2012) and Kim et al.
201 (2015) were measured during December 21, 2010-January 23, 2011 and February 11 to March 14, 2012,
202 respectively. Our measurements in this study were executed mainly during January 1-15, 2014. For the
203 Amundsen polynya region, a large seasonal variation in the total daily carbon uptake rate of
204 phytoplankton was already reported by Kim et al. (2015) and Arrigo et al. (2012) based on field-measured
205 data and satellite-derived approach, respectively. It is appeared that this seasonal variation largely
206 depends on the bloom stage of phytoplankton which peaks during the late December-January and
207 terminates at late February (Arrigo and van Dijken 2003; Arrigo et al., 2012; Kim et al., 2015). The total
208 daily nitrogen uptake rates of phytoplankton were 0.12 g N m⁻² d⁻¹ (\pm 0.09 g N m⁻² d⁻¹) and 0.21 g N m⁻² d⁻¹
209 (\pm 0.11 g N m⁻² d⁻¹) for non-polynya and polynya regions, respectively in this study. Previous studies
210 reported that the total daily nitrogen uptake rates in non-polynya region were 0.24 g N m⁻² d⁻¹ during Dec.
211 21, 2010-Jan. 23, 2011 and 0.04 g N m⁻² d⁻¹ during Feb. 11 to Mar. 14, 2012 whereas the uptake rates in
212 polynya region were 0.93 g N m⁻² d⁻¹ in 2010/2011 and 0.06 g N m⁻² d⁻¹ in 2012 in the Amundsen Sea (Lee
213 et al., 2012; Kim et al., 2015). Our total daily nitrogen uptake rates of phytoplankton in non-polynya and
214 polynya regions were between the two previous studies (Lee et al., 2012; Kim et al., 2015). Based on the
215 nitrate and ammonium uptake rates in this study, *f*-ratios (nitrate uptake rate/nitrate+ammonium uptake
216 rates) averaged for non-polynya and polynya regions were 0.62 (\pm 0.08) and 0.54 (\pm 0.20), respectively.
217 These ratios also were between the two previous studies. Although they were not significant different
218 because of a large spatial variation, larger *f*-ratios in non-polynya than in polynya region are consistent
219 with the results of the previous studies (Lee et al., 2012; Kim et al., 2015). At this point, we do not have a
220 solid explanation for that but a further future study is needed for the higher *f*-ratio mechanism in non-
221 polynya region.

222 The percent contributions of small phytoplankton in terms of chlorophyll-a, POC/PON, daily
223 carbon and nitrogen uptake rates are shown in Table 1. The overall contribution of small phytoplankton to
224 the total chlorophyll-a concentration for all the productivity stations was 19.4 % (\pm 26.0 %) which is
225 significantly (t-test, $p < 0.05$) lower than the POC contribution (41.1 ± 10.6 %). This is consistent with the
226 result in the Chukchi Sea, Arctic Ocean reported by Lee et al. (2013). They explained that higher POC
227 content per chlorophyll-a unit of small phytoplankton could cause their higher POC contribution (Lee et
228 al., 2013). Given C/N ratio (6.6 ± 0.6) and $\delta^{13}\text{C}$ (-25.9 ± 1.0 ‰) of sample filters attained for POC and
229 PON in this study, our filtered samples are believed to be mainly phytoplankton-originated POC and PON
230 (Kim et al., 2016). Thus, a significant potential overestimated POC contribution of non-phytoplankton
231 materials could be excluded for the higher POC contribution than chlorophyll-a contribution of small
232 phytoplankton. Therefore, small phytoplankton contributions based on conventional assessments of
233 chlorophyll-a concentration might lead an underestimated contribution of small phytoplankton (Lee et al.,
234 2013). In fact, several authors argued that chlorophyll-a concentration might be not a good index for
235 phytoplankton biomass since it largely depends on environmental factors such as nutrient and light
236 conditions as well as dominant groups and physiological status of phytoplankton (Desortová 1981;
237 Behrenfeld et al., 2005; Kruskopf and Flynn, 2006; Behrenfeld and Boss 2006). However, the effects of
238 non-phytoplankton carbon materials such as extracellular carbon mucilage can not be completely
239 excluded for the POC contribution as discussed below.

240 The overall contributions of carbon and nitrogen (nitrate and ammonium) uptake rates of small
241 phytoplankton at all the productivity stations in this study are similar with 26.9 % and 27.7 %,
242 respectively. These contributions are relatively higher than the chlorophyll-a contribution of small
243 phytoplankton but they are not statistically different (t-test, $p > 0.05$). In general, the contribution of daily
244 ammonium uptake rate of small phytoplankton is significantly (t-test, $p < 0.05$) higher than the
245 contribution of daily nitrate uptake rate of small phytoplankton at all the stations in this study. It is well-
246 known for the ammonium preference of small phytoplankton in various regions (Koike et al., 1986;
247 Tremblay et al., 2000, Lee et al., 2008; Lee et al., 2013).

248 In terms of the contributions in different regions, all the contributions (chlorophyll-a, POC/PON,
249 carbon and nitrogen uptake rates) of small phytoplankton were higher in the non-polynya region than in
250 the polynya region (Table 1). In addition, the chlorophyll-a contribution of small phytoplankton ($7.9 \pm$
251 3.5%) was significantly (t-test, $p < 0.01$) lower than the POC contribution ($36.9 \pm 4.6 \%$) in the polynya
252 region, whereas they were not statistically different in the non-polynya region (Table 1). This indicates
253 that small phytoplankton contributed more to the total POC than the chlorophyll-a concentration in the
254 polynya region. We do not have species compositions of phytoplankton in this study, but previous results
255 reported that *Phaeocystis* sp. are dominant in the Amundsen Sea polynya region (Lee et al., 2012).
256 Generally, *Phaeocystis* spp. release a large portion (up to 46 %) of extracellular carbon mucilage which
257 makes their colonial form (Matrai et al., 1995). This non-phytoplankton carbon material without
258 chlorophyll-a might cause a higher POC contribution of small phytoplankton in the polynya region during
259 this study. In fact, the contribution of the daily carbon uptake rates of small phytoplankton ($14.9 \pm 8.4 \%$)
260 was not as high as the POC contribution ($36.9 \pm 4.6 \%$) in the polynya region. The chlorophyll-a
261 contributions of small phytoplankton were lower than those of the daily carbon uptake rate in this study,
262 which is consistent with the results from polynya and marginal ice zone stations in the Ross Sea,
263 Antarctica during austral spring and summer (Saggiomo et al., 1998). They reported that the chlorophyll-a
264 and primary production contributions of pico-phytoplankton ($< 2 \mu\text{m}$) were 29 % and 40 % at polynya
265 stations whereas the contributions were 17 % and 32 % at marginal ice zone stations, respectively. In the
266 polynya region, they found much higher contributions in chlorophyll-a and primary production of small
267 phytoplankton than those in this study although their size of small phytoplankton is somewhat smaller
268 than our size ($< 5 \mu\text{m}$).

269 We found a strong negative correlation ($r^2 = 0.790$, $p < 0.05$) between the productivity
270 contributions of small phytoplankton and total daily carbon uptake rates of phytoplankton in the
271 Amundsen Sea (Fig. 7), which implies that daily primary production decreases as small phytoplankton
272 contribution increases. This is mainly because of the relatively lower carbon uptake rate of small
273 phytoplankton than large phytoplankton in the Chukchi Sea, Arctic Ocean reported by Lee et al. (2013).

274 In respect to food quality of small phytoplankton as a basic food source to herbivores, macromolecular
275 compositions such as proteins, lipids, and carbohydrates as photosynthetic-end products will be needed
276 for better understanding small cells-dominant marine ecosystem in response to environmental changes
277 (Lee et al., 2013). According to Kang et al. (accepted), small phytoplankton assimilate more food
278 materials and calorific contents per unit of chlorophyll-a concentration and thus provide more
279 contributions in respect to energy aspect than other phytoplankton community in the East/Japan Sea.
280 However, this change in dominant phytoplankton community from large to small cells will likely cause
281 further alteration of higher trophic levels because of prey size itself available to higher trophic grazers
282 (Moline et al., 2004). In conclusion, monitoring the contributions of small-sized phytoplankton to total
283 biomass and primary production of total phytoplankton community could be important as a valuable
284 indicator to sense environmental changes in marine ecosystem

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286 **Acknowledgments**

287 We thank the captain and crew members of the Korean Research Icebreaker, *Araon*, for their
288 outstanding assistance during the cruise. This research was supported by the Korea Polar Research
289 Institute (KOPRI; PP15020).

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365 **Table caption**

366 Table 1. Contributions (%) of small phytoplankton in the Amundsen Sea. Contributions of chlorophyll-a,
367 POC, PON, and carbon and nitrogen uptake rates were derived from water euphotic column-integrated
368 values averaged from stations.

369

370 **Figure captions**

371 Fig. 1. Sampling locations in the Amundsen Sea. Red closed circles represent productivity stations. Sea

372 ice concentration data during the cruise period from Nimbus-7 SMMR and DMSP SSM/I-

373 SSMIS Passive Microwave data provided by National Snow & Ice Data Center.

374 Fig. 2. Water column-integrated chlorophyll-a concentration at the productivity stations in the Amundsen

375 Sea.

376 Fig. 3. Water column-integrated concentrations of POC and PON of small and large phytoplankton.

377 Fig. 4. Water column-integrated daily carbon uptake rates of small and large phytoplankton.

378 Fig. 5. Water column-integrated daily nitrate uptake rates of small and large phytoplankton.

379 Fig. 6. Water column-integrated daily ammonium uptake rates of small and large phytoplankton.

380 Fig. 7. Relationship between productivity contributions of small phytoplankton and the total daily carbon

381 uptake rates of phytoplankton (large + small). The total daily carbon uptake rates were

382 transformed into natural logs for a linear regression.

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Table 1. Contributions (%) of small phytoplankton in the Amundsen Sea. Contributions of chlorophyll-a, POC, PON, and carbon and nitrogen uptake rates were derived from water euphotic column-integrated values averaged from stations.

	Chlorophyll-a	POC	PON	Daily carbon uptake rate	Daily nitrate uptake rate	Daily ammonium uptake rate	Total nitrogen uptake rate
All stations	19.4 ± 26.0	41.1 ± 10.6	41.3 ± 11.5	26.9 ± 29.3	21.5 ± 11.1	38.7 ± 24.9	27.7 ± 14.4
Non-polynya	42.4 ± 37.2	49.5 ± 14.4	50.0 ± 15.1	50.8 ± 42.8	28.2 ± 15.9	52.8 ± 40.5	36.2 ± 23.0
Polynya	7.9 ± 3.5	36.9 ± 4.6	37.0 ± 6.9	14.9 ± 8.4	18.1 ± 6.8	31.6 ± 10.1	23.5 ± 6.0

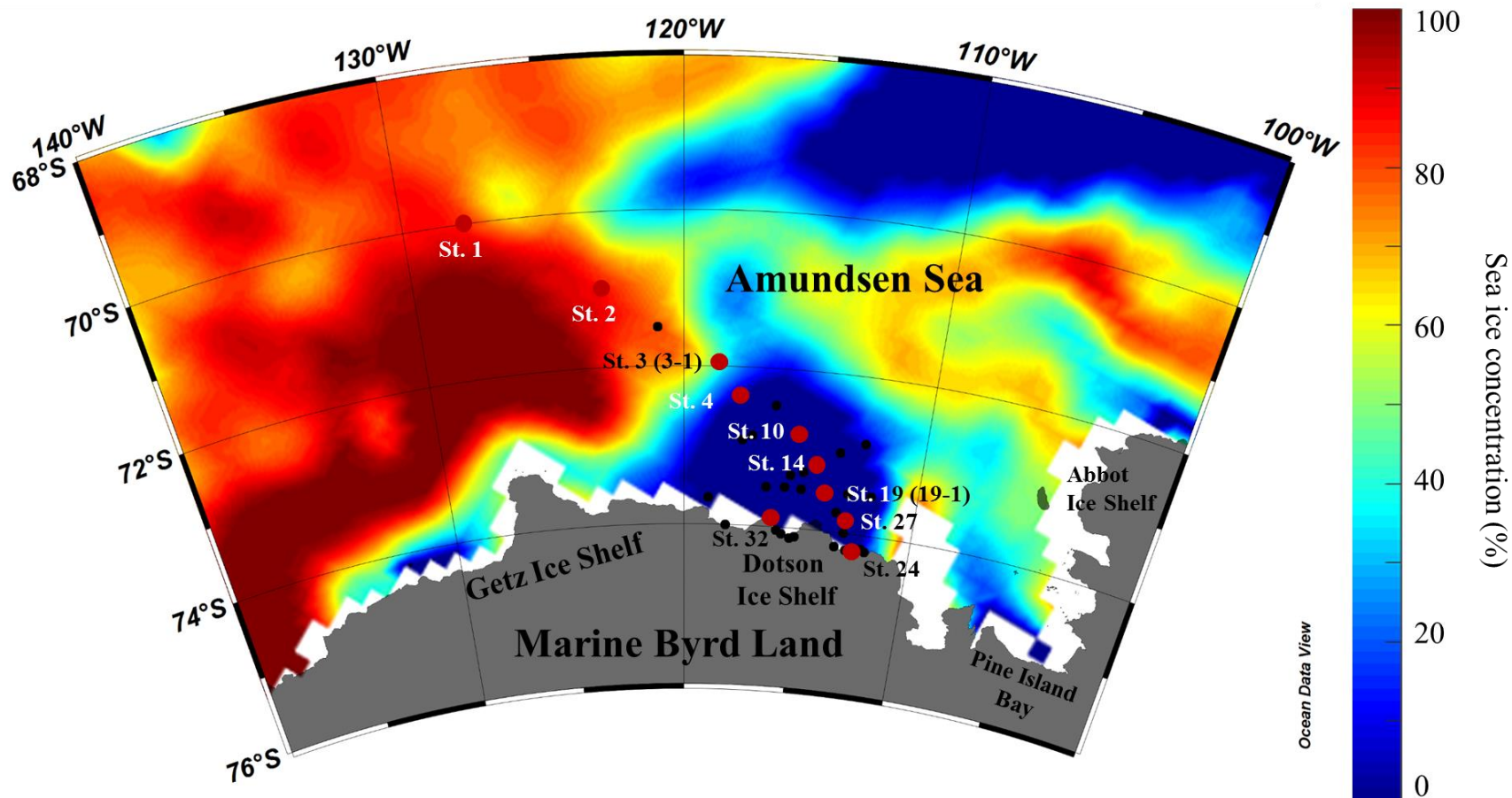


Fig. 1. Sampling locations in the Amundsen Sea. Red closed circles represent productivity stations. Sea ice concentration data during the cruise period from Nimbus-7 SMMR and DMSP SSM/I-SSMIS Passive Microwave data provided by National Snow & Ice Data Center.

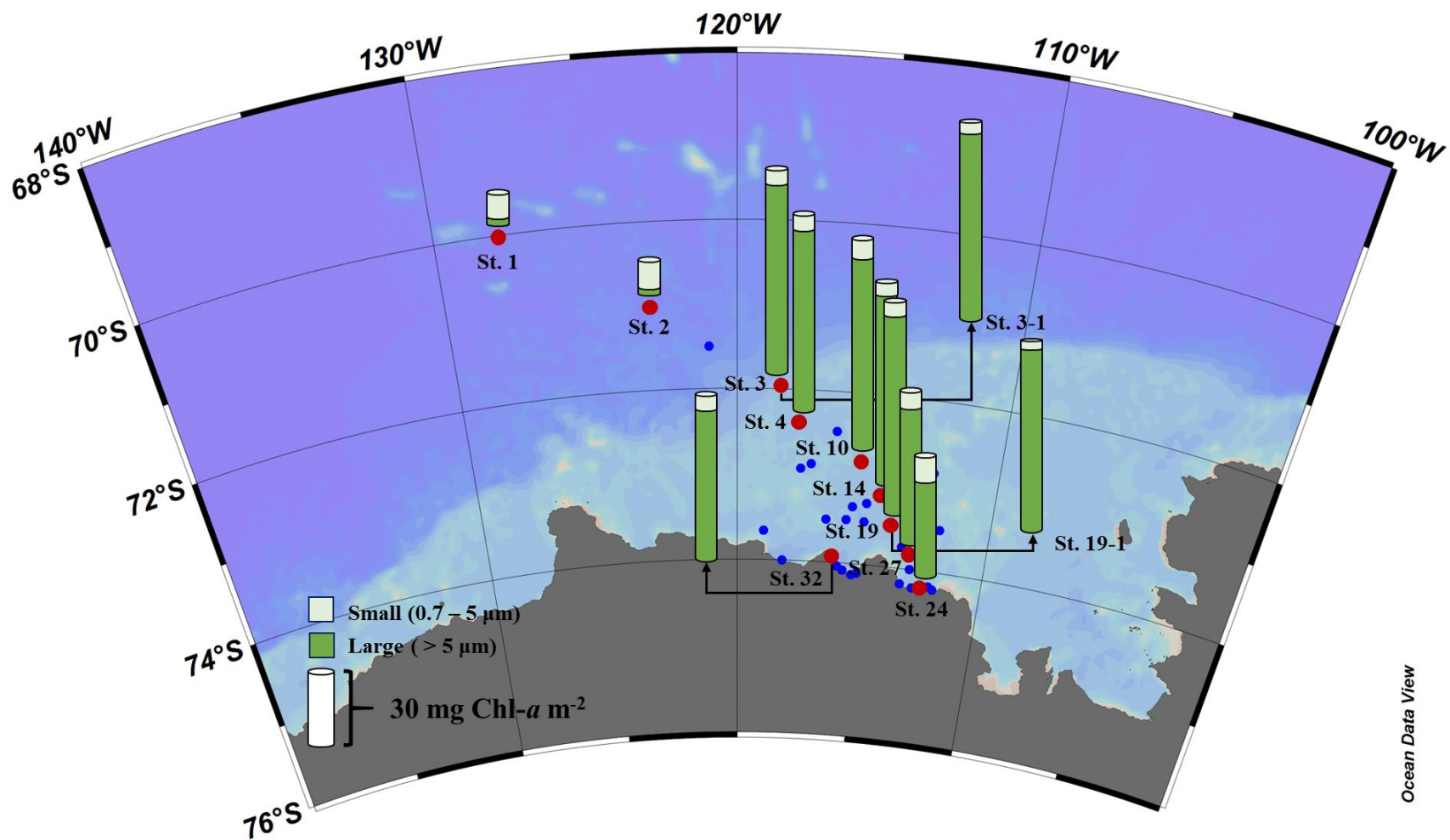


Fig. 2. Water column-integrated chlorophyll-a concentration at the productivity stations in the Amundsen Sea.

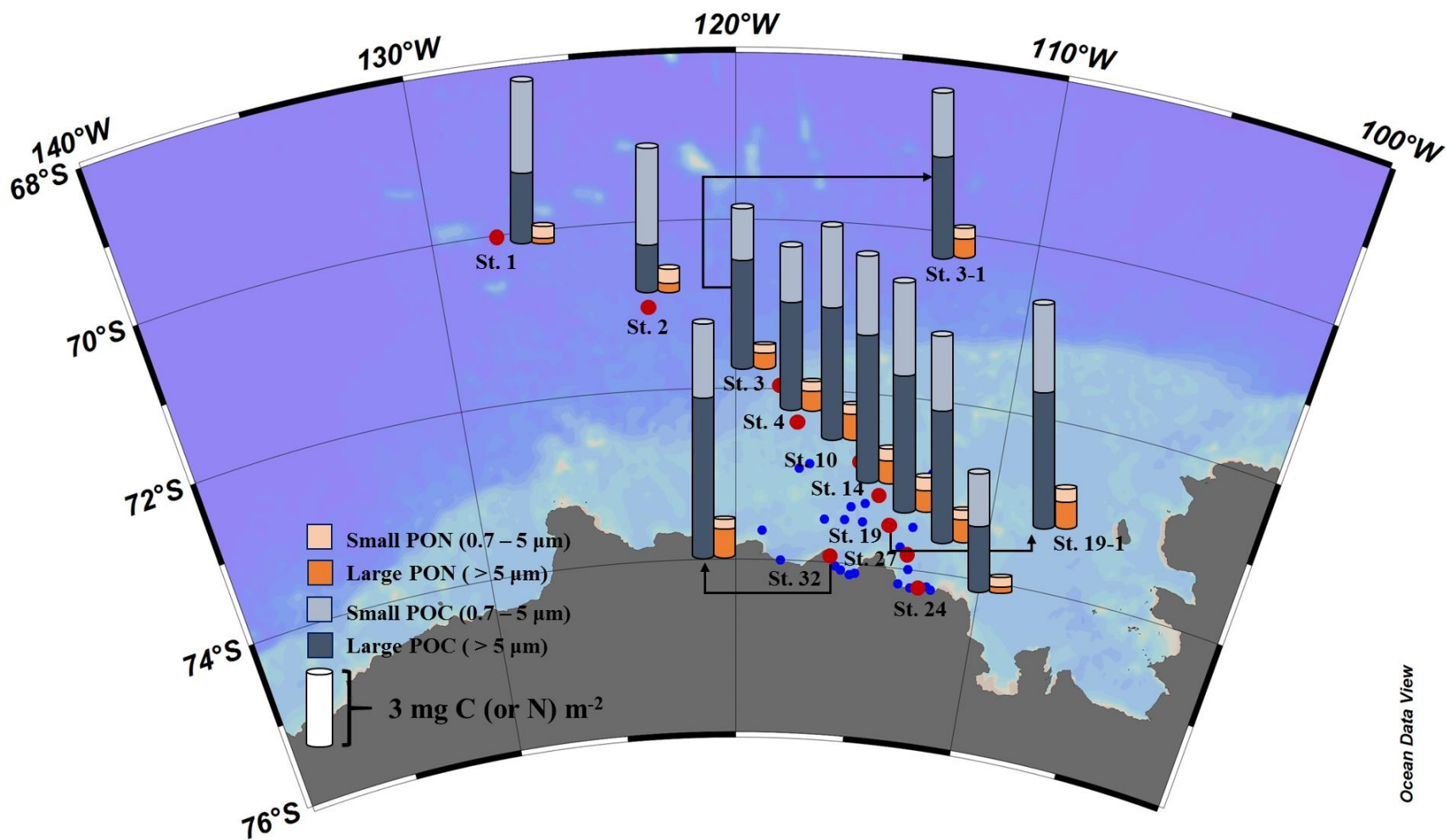


Fig. 3. Water column-integrated concentrations of POC and PON of small and large phytoplankton.

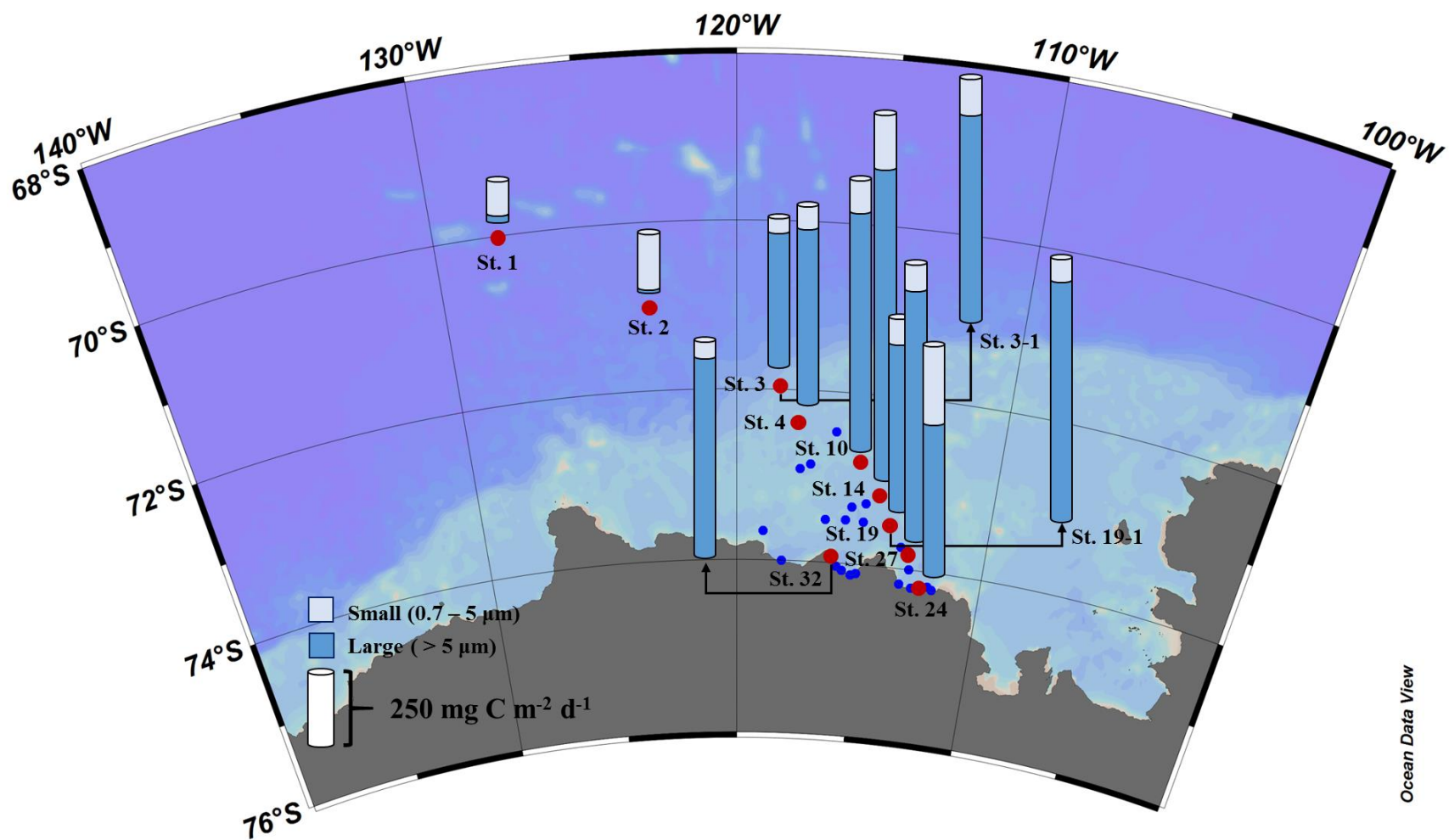


Fig. 4. Water column-integrated daily carbon uptake rates of small and large phytoplankton.

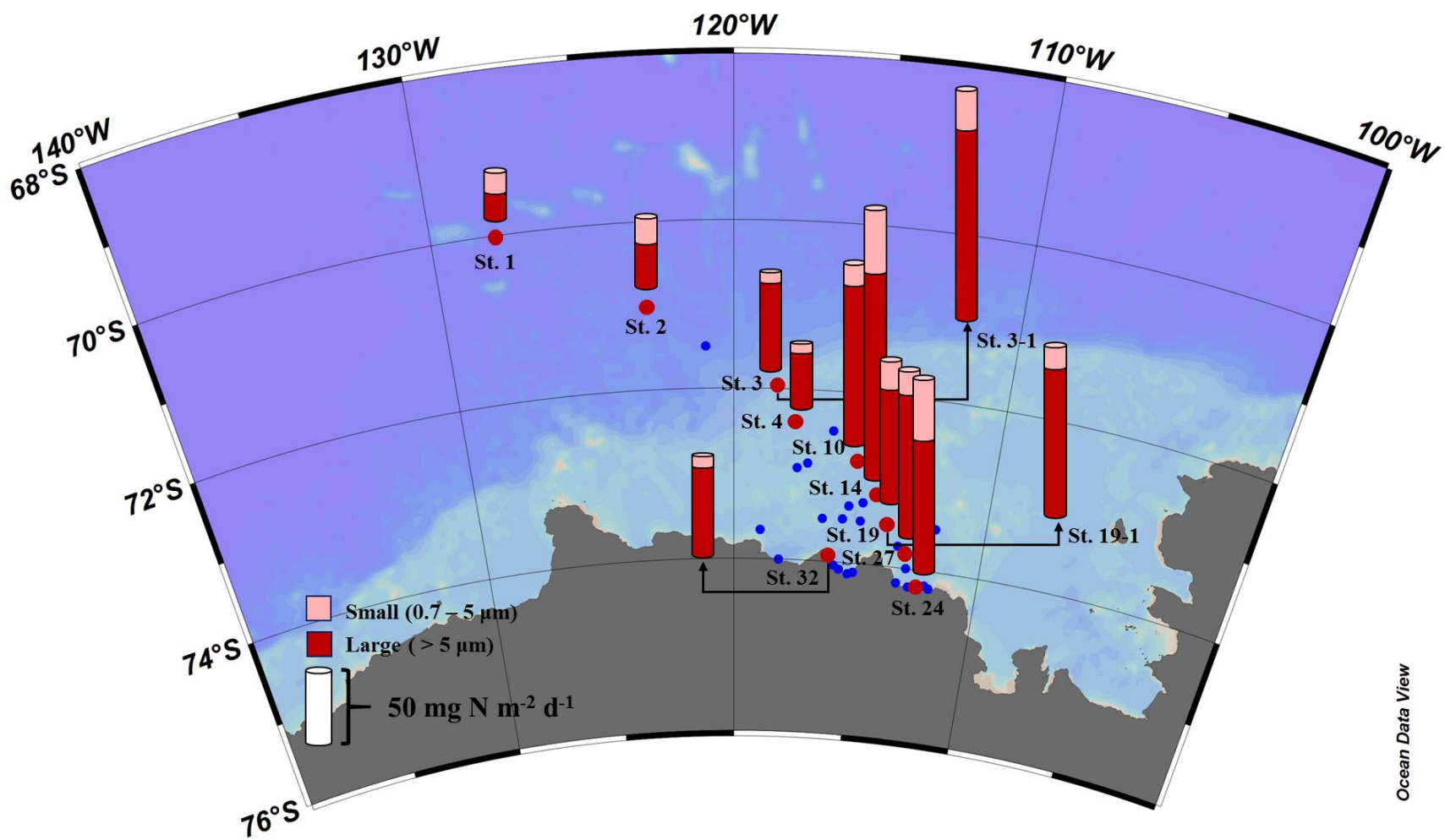


Fig. 5. Water column-integrated daily nitrate uptake rates of small and large phytoplankton.

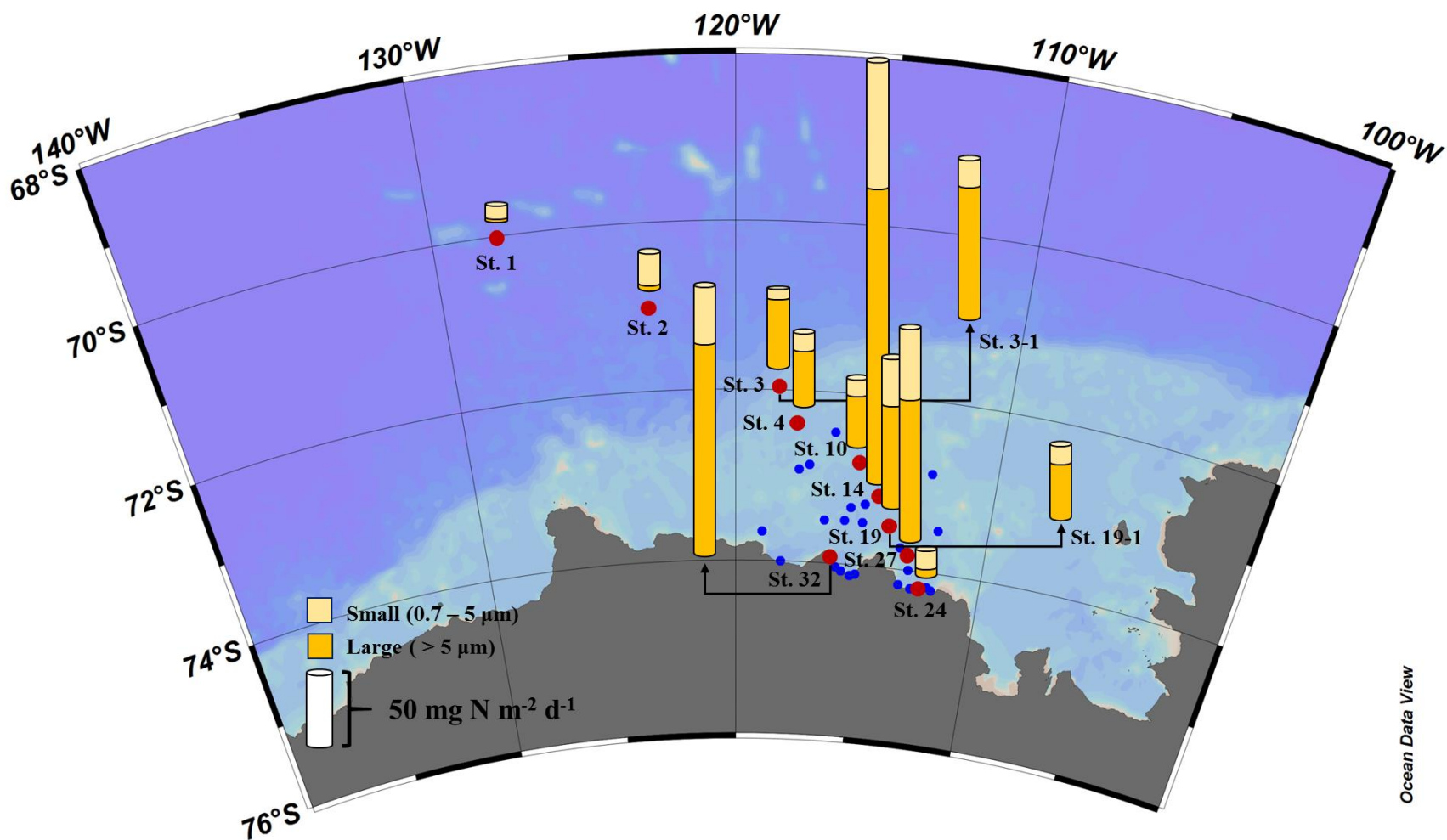


Fig. 6. Water column-integrated daily ammonium uptake rates of small and large phytoplankton.

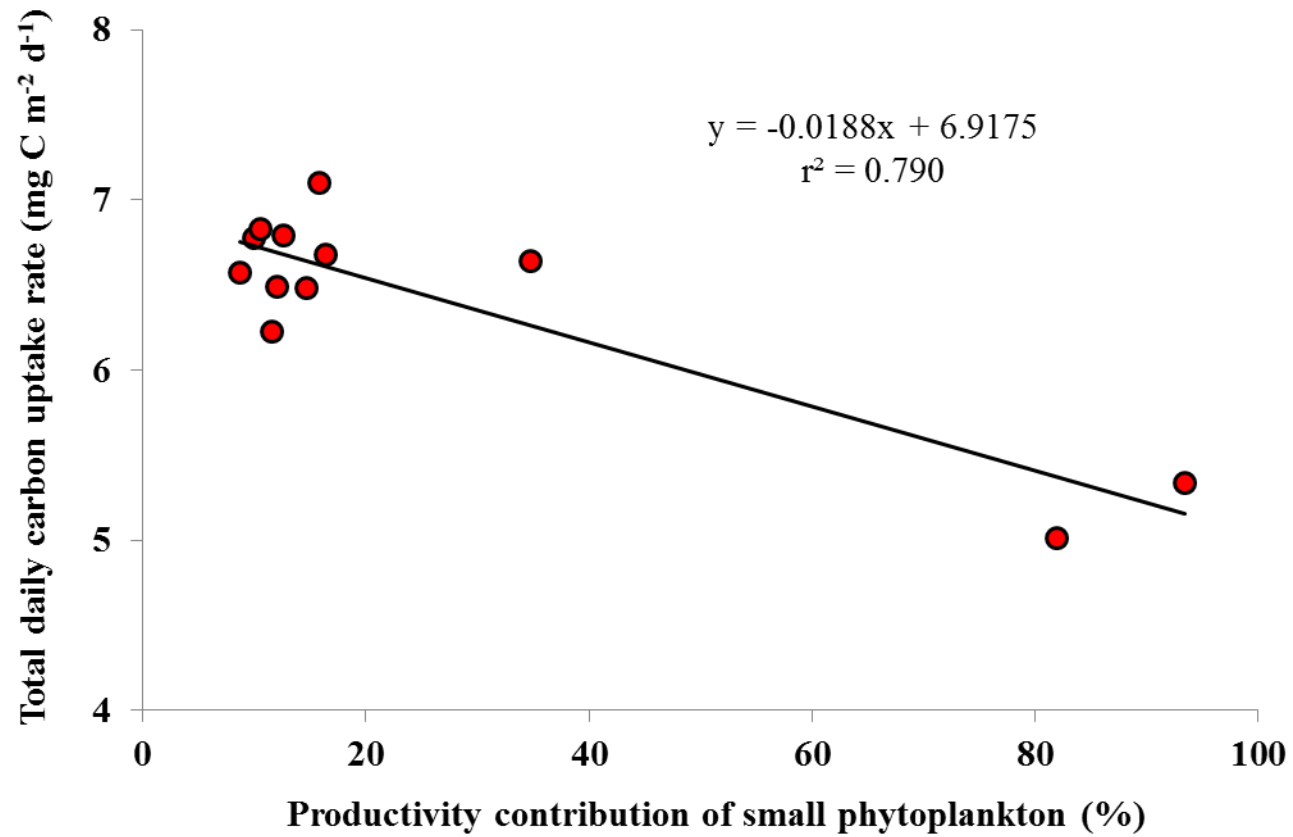


Fig. 7. Relationship between productivity contributions of small phytoplankton and the total daily carbon uptake rates of phytoplankton (large + small). The total daily carbon uptake rates were transformed into natural logs for a linear regression.