

1 **Small phytoplankton contributions to the standing stocks and the total**
2 **primary production in the Amundsen Sea**

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17 Abstract

18 Small phytoplankton are anticipated to be more important in a recently warming and freshening
19 ocean condition. However, little information on the contribution of small phytoplankton to overall
20 phytoplankton production is currently available in the Amundsen Sea. To determine the contributions of
21 small phytoplankton to total biomass and primary production, carbon and nitrogen uptake rates of total
22 and small phytoplankton were obtained from 12 productivity stations in the Amundsen Sea. The daily
23 carbon uptake rates of total phytoplankton averaged in this study were $0.42 \text{ g C m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.30 \text{ g C}$
24 $\text{m}^{-2} \text{ d}^{-1}$) and $0.84 \text{ g C m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.18 \text{ g C m}^{-2} \text{ d}^{-1}$) for non-polynya and polynya regions,
25 respectively, whereas the daily total nitrogen (nitrate and ammonium) uptake rates were $0.12 \text{ g N m}^{-2} \text{ d}^{-1}$
26 (S.D. = $\pm 0.09 \text{ g N m}^{-2} \text{ d}^{-1}$) and $0.21 \text{ g N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.11 \text{ g N m}^{-2} \text{ d}^{-1}$), respectively for non-polynya
27 and polynya regions, all of which were within the ranges reported previously. Small phytoplankton
28 contributed 26.9 % and 27.7 % to the total carbon and nitrogen uptake rates of phytoplankton in this study,
29 respectively, which were relatively higher than the chlorophyll-a contribution (19.4 %) of small
30 phytoplankton. For a comparison of different regions, the contributions for chlorophyll-a concentration
31 and primary production of small phytoplankton averaged from all the non-polynya stations were 42.4 %
32 and 50.8 %, which were significantly higher than those (7.9 % and 14.9 %, respectively) in polynya
33 region. A strong negative correlation ($r^2 = 0.790$, $p < 0.05$) was found between the contributions of small
34 phytoplankton and the total daily primary production of phytoplankton in this study. This finding implies
35 that daily primary production decreases as small phytoplankton contribution increases, which is mainly
36 due to the lower carbon uptake rate of small phytoplankton than large phytoplankton.

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39 **Keywords:** Phytoplankton, Primary production, Polynya, Amundsen Sea

40

41 **1. Introduction**

42 The Amundsen Sea is located in the West Antarctica between the Ross Sea and Bellingshausen
43 Sea (Fig. 1), which is one of the least-biologically studied regions in the Southern Ocean. Recently,
44 several international research programs (KOPRI Amundsen project, iSTAR, ASPIRE, and DynaLiFe)
45 were launched to improve the understanding of this remote area. Field-measurement data revealed that
46 annual primary production of phytoplankton reaching to $220 \text{ g C m}^{-2} \text{ y}^{-1}$ in the Amundsen Sea polynya is
47 as high as that of Ross Sea polynya ($200 \text{ g C m}^{-2} \text{ y}^{-1}$) which was previously known for the highest
48 productivity region in the Southern Ocean (Lee et al., 2012). Given the fact that the chlorophyll-a
49 concentration averaged from all the chlorophyll-a measured stations was twice as high as that of the only
50 productivity-measured stations, Lee et al. (2012) argued that the annual production in the Amundsen Sea
51 polynya could be even two times higher than that of Ross Sea polynya.

52 Over the past several decades, a rapid climate change has been detected and subsequently
53 physical changes have occurred in the marine ecosystem in the western Antarctic Peninsula (WAP),
54 which was mainly based on the results from Palmer Antarctic Long-Term Ecological Research project
55 focusing on the north of $\sim 69^\circ \text{S}$ (Ducklow et al. 2007; Montes-Hugo et al. 2009). Recent studies revealed
56 that the Thwaites Glacier in Pine Island Bay is retreating fast and the ice volume loss in the nearby Getz
57 Ice shelf is accelerating (Joughin et al., 2014; Paolo et al., 2015). Shoaling warm Circumpolar Deep
58 Water is believed to be a main cause of the ice sheet mass loss through the ice shelf basal melt underside
59 of the ice shelves (Yager et al. 2012; Schmidtko et al. 2014). Climate change from a cold-dry polar-type
60 to a warm-humid sub-Antarctic-type has driven subsequent changes in ocean biological productivity
61 along the WAP shelf over the recent three decades (Montes-Hugo et al. 2009).

62 Phytoplankton, as the base of oceanic food webs, can be an indicator for changes in marine
63 ecosystems responding to environmental changes (Moline et al., 2004; Wassman et al., 2011; Arrigo et al.,
64 2015). For example, a recurrent shift in phytoplankton community structure from large diatoms to
65 relatively small cryptophytes could be tightly associated with changes in glacial melt-water runoff
66 (Moline et al., 2004). To date, little information on the contribution of small phytoplankton to primary

67 production is available in the Antarctic Ocean (Saggiomo et al. 1998), especially in the Amundsen Sea
68 with a rapid melting of ice shelf (Yager et al. 2012; Schmidtko et al. 2014). Thus, our main objective in
69 this study is to determine contributions of small phytoplankton to the overall total biomass and primary
70 production of phytoplankton in the Amundsen Sea for monitoring marine ecosystem responding to
71 environmental condition change.

72

73 **2. Materials and methods**

74 *2.1. Total and size-fractionated chlorophyll-a concentration*

75 Water samples for total and size-fractionated chlorophyll-a concentrations of phytoplankton were
76 obtained at the 12 productivity stations in the Amundsen Sea (Fig. 1) during the KOPRI Amundsen cruise
77 from 1 to 15 January, 2014 onboard the Korean Research Icebreaker ship *Araon*. Based on the sea ice
78 concentration data from National Snow & Ice Data Center during the cruise period in 2013 (Fig. 1), our
79 study region was further separated into polynya and non-polynya areas for comparison. Four stations (St.
80 1, St. 2, St. 3, and St. 3-1) among the 12 stations belong to the non-polynya region and the rest of the
81 stations belong to the polynya region. St. 3 and St. 3-1 were on the fringe of the polynya area,
82 experiencing approximately 30% sea ice cover. Following the definition of polynya as an area of open
83 water within sea ice zone, we grouped them into non-polynya regions. Six different light depths (100, 50,
84 30, 12, 5, and 1% penetration of the surface irradiance, PAR) were determined with an LI-COR
85 underwater 4π light sensor. Total chlorophyll-a concentrations were measured at the six different light
86 depths (100, 50, 30, 12, 5 and 1% of PAR). For size-fractionated chlorophyll-a concentrations, water
87 samples were collected at three light depths (100, 30, and 1 %). Water samples (0.3–0.5 L) for total
88 chlorophyll-a concentrations were filtered using Whatman glass fiber filters (GF/F; 25 mm). For different
89 size-fractionated chlorophyll-a concentrations water samples (0.7–1 L) were passed sequentially through
90 20 and 5 μm Nucleopore filters (47 mm) and 0.7 μm GF/F filters (47 mm). After the filters were extracted
91 using the method described by Kim et al. (2015), all chlorophyll-a concentrations were subsequently

92 determined onboard using a Trilogy fluorometer (Turner Designs, USA). The methods and calculations
93 for chlorophyll-a were based on Parsons et al. (1984).

94 *2.2. Carbon and nitrogen uptake experiments*

95 Water samples were collected for and carbon and nitrogen uptake measurements of
96 phytoplankton. Using a dual stable isotope technique (Lee et al., 2012; Kim et al., 2015), the experiments
97 of carbon and nitrogen uptake rates of phytoplankton were conducted at 12 selected productivity stations
98 including 2 revisited-stations (St. 3-1 and St. 19-1) when on-deck incubations were available during
99 daytime at oceanographic survey stations. Water samples from the 6 light depths for the uptake
100 experiments were obtained from a CTD-rosette sampler system equipped with 24 10-L Niskin bottles.
101 Water sample from each light depth was transferred into different screened polycarbonate incubation
102 bottles (1 L) which matched each light depth. The bottles were placed in large polycarbonate incubators
103 cooled with running surface seawater on deck under natural light conditions for 4-5 hours, after the water
104 samples in the incubation bottles were inoculated with labeled carbon ($\text{NaH}^{13}\text{CO}_3$) and nitrate (K^{15}NO_3)
105 or ammonium ($^{15}\text{NH}_4\text{Cl}$) substrates. After 4–5 h incubations, the incubated waters were well mixed and
106 distributed into two filtration sets for the carbon and nitrogen uptake rates of total ($> 0.7 \mu\text{m}$) and small-
107 sized cells ($< 5 \mu\text{m}$). Small-sized cells $< 5 \mu\text{m}$ are generally defined as small phytoplankton in
108 comparison to large diatoms ($> 5 \mu\text{m}$) (Robineau et al., 1994; reference therein). The incubated waters
109 (0.3 L) for total uptake rates were filtered through pre-combusted GF/F filters (25 mm diameter), whereas
110 waters samples (0.5 L) for the uptake rates of small phytoplankton were passed through $5 \mu\text{m}$ Nuclepore
111 filters (47 mm) to remove large phytoplankton cells ($> 5 \mu\text{m}$) and then the filtrate was passed through pre-
112 combusted GF/F (25 mm) for the small phytoplankton (Lee et al., 2013). The values for large
113 phytoplankton in this study were obtained from the difference between small and total fractions (Lee et al.,
114 2013). The filters were immediately preserved at -80°C until mass spectrometric analysis. After acid
115 fuming overnight to remove carbonate, the concentrations of particulate organic carbon (POC) and nitrogen
116 (PON) and the abundance of ^{13}C and ^{15}N were determined by a Finnigan Delta+XL mass spectrometer at

117 the Alaska Stable Isotope Facility, USA. All contribution results of small phytoplankton in this study
118 were estimated from comparison of small phytoplankton to total phytoplankton integral values from 100
119 to 1 % light depth at each station based on the trapezoidal rule. Daily carbon and nitrogen uptake rates of
120 phytoplankton were based on our hourly uptake rates measured in this study and a 24-h photoperiod per
121 day during the summer period in the Amundsen Sea (Lee et al., 2012).

122

123 **3. Results**

124 3.1. Chlorophyll-a, POC, and PON contributions of small phytoplankton

125 The depth-integrated total (large + small phytoplankton) chlorophyll-a concentration was 11.1-
126 80.3 mg chl-a m⁻² (mean ± S.D. = 57.4 ± 25.2 mg chl-a m⁻²) in this study (Fig. 2). The contribution of
127 small phytoplankton to the total chlorophyll-a concentration was 4.9-76.5 % (19.4 ± 26.0 %). Large
128 phytoplankton (> 5 µm) were generally predominant (approximately 80 %) based on different-sized
129 chlorophyll-a concentrations. For a regional comparison, the average contributions of small
130 phytoplankton to the total chlorophyll-a concentration were 42.4 % (± 37.2 %) and 7.9 % (± 3.5 %) for
131 non-polynya and polynya regions, respectively (Table 1). The chlorophyll-a contribution of small
132 phytoplankton was larger in the non-polynya region than in the polynya region although they were not
133 significantly different (t-test, p > 0.05).

134 The depth-integrated total POC concentration of phytoplankton showed no large spatial variation
135 ranging from 4.72 to 9.22 mg C m⁻² (7.40 ± 1.55 mg C m⁻²) (Fig. 3). In comparison, the depth-integrated
136 total PON concentration of phytoplankton was 0.76-1.74 mg N m⁻² (1.33 ± 0.32 mg N m⁻²). The POC
137 contribution of small phytoplankton was 30.7-65.5 % (41.1 ± 10.6 %), whereas the PON contribution was
138 30.8-67.2 % (41.3 ± 11.5 %) in the Amundsen Sea (Fig. 3). Specifically, the POC and PON contributions
139 of small phytoplankton averaged from all the productivity stations in the polynya region were 36.9 % (±
140 4.6 %) and 37.0 % (± 6.9 %), respectively, whereas they were 49.5 % (± 14.4 %) and 50.0 % (± 15.1 %),
141 respectively in the non-polynya region (Table 1). The POC and PON contributions of small

142 phytoplankton were not statistically different between the polynya and non-polynya regions (t-test, $p >$
143 0.05).

144 3.2. Carbon uptake rate contributions of small phytoplankton

145 The depth-integrated total daily carbon uptake rate of phytoplankton (large + small phytoplankton)
146 was 150.4-1213.4 mg C m⁻² d⁻¹ (696.5 ± 298.4 mg C m⁻² d⁻¹) in this study (Fig. 4). In contrast, the rate of
147 small phytoplankton was 58.6-266.4 mg C m⁻² d⁻¹ (124.9 ± 62.4 mg C m⁻² d⁻¹). Small phytoplankton
148 contributed 26.9 % ($\pm 29.3\%$) to total daily carbon uptake rate of total phytoplankton.

149 Specifically, the total daily carbon uptake rate of phytoplankton was 150.4-796.4 mg C m⁻² d⁻¹
150 (415.0 ± 298.2 mg C m⁻² d⁻¹) in the non-polynya region, whereas it was 654.8-1213.4 mg C m⁻² d⁻¹ (837.3
151 ± 184.1 mg C m⁻² d⁻¹) in the polynya region. The total daily carbon uptake rates of phytoplankton were
152 significantly higher (t-test, $p < 0.05$) in the polynya region than the non-polynya region. The rate of small
153 phytoplankton was 58.6-193.6 mg C m⁻² d⁻¹ (126.5 ± 55.2 mg C m⁻² d⁻¹) in the non-polynya region,
154 whereas it was 62.2-266.4 mg C m⁻² d⁻¹ (124.1 ± 69.3 mg C m⁻² d⁻¹) in the polynya region. The daily
155 carbon uptake rates of small phytoplankton were not significantly different (t-test, $p > 0.05$) between the
156 polynya and non-polynya stations. The average contributions of small phytoplankton to total daily carbon
157 uptake rates were 50.8 % (± 42.8 %) and 14.9 % (± 8.4 %), respectively for the non-polynya and polynya
158 regions (Table 1). The average contributions were largely different between the polynya and non-polynya
159 regions but they were not statistically significant (t-test, $p > 0.05$).

160 3.3. Nitrogen uptake rate contributions of small phytoplankton

161 The depth-integrated total daily nitrate uptake rate of phytoplankton (large + small phytoplankton)
162 was 34.0-174.2 mg N m⁻² d⁻¹ (93.7 ± 43.2 mg N m⁻² d⁻¹), whereas the rate of small phytoplankton was 6.1-
163 40.9 mg N m⁻² d⁻¹ (19.0 ± 11.3 mg N m⁻² d⁻¹) in this study (Fig. 5). Small phytoplankton contributed 21.5 %
164 (± 11.1 %) to total daily nitrate uptake rates. In comparison, the total daily ammonium uptake rate of
165 phytoplankton was 12.4-173.8 mg N m⁻² d⁻¹ (86.7 ± 75.9 mg N m⁻² d⁻¹), whereas the rate of small
166 phytoplankton was 9.1-81.1 mg N m⁻² d⁻¹ (25.7 ± 21.1 mg N m⁻² d⁻¹) in this study (Fig. 6). Small
167 phytoplankton contributed 38.7 % (± 24.9 %) to total daily ammonium uptake rates. The contributions of

168 small phytoplankton were significantly higher in ammonium uptake rate than nitrate uptake rate (t-test, p
169 < 0.05).

170 For different regions, the total daily nitrate uptake rates of phytoplankton were 34.0-142.1 mg N
171 $\text{m}^{-2} \text{d}^{-1}$ ($71.9 \pm 48.4 \text{ mg N m}^{-2} \text{d}^{-1}$) in the non-polynya region and 44.2-174.2 mg N $\text{m}^{-2} \text{d}^{-1}$ (104.6 ± 39.0
172 $\text{mg N m}^{-2} \text{d}^{-1}$) in the polynya region, respectively. In comparison, the daily nitrate uptake rates of small
173 phytoplankton were 7.5-26.6 mg N $\text{m}^{-2} \text{d}^{-1}$ ($16.7 \pm 7.8 \text{ mg N m}^{-2} \text{d}^{-1}$) and 6.1-40.9 mg N $\text{m}^{-2} \text{d}^{-1}$ ($20.1 \pm$
174 $13.1 \text{ mg N m}^{-2} \text{d}^{-1}$), respectively for the non-polynya and polynya regions. The contributions of small
175 phytoplankton to the total daily nitrate uptake rates were 28.2 % (± 15.9 %) in the non-polynya region and
176 18.1 % (± 6.8 %) in the polynya region, respectively (Table 1). The total daily ammonium uptake rates of
177 total phytoplankton were 12.3 and 106.1 mg N $\text{m}^{-2} \text{d}^{-1}$ ($49.7 \pm 41.2 \text{ mg N m}^{-2} \text{d}^{-1}$) in the non-polynya
178 region and 18.1-269.3 mg N $\text{m}^{-2} \text{d}^{-1}$ ($105.2 \pm 84.6 \text{ mg N m}^{-2} \text{d}^{-1}$) in the polynya region. In comparison, the
179 rates of small phytoplankton were 9.1-22.4 mg N $\text{m}^{-2} \text{d}^{-1}$ ($15.8 \pm 6.4 \text{ mg N m}^{-2} \text{d}^{-1}$) in the non-polynya
180 region and 9.9-81.1 mg N $\text{m}^{-2} \text{d}^{-1}$ ($30.7 \pm 24.5 \text{ mg N m}^{-2} \text{d}^{-1}$) in the polynya region. Small phytoplankton
181 contributed 52.8 % (± 40.5 %) and 31.6 % (± 10.1 %) to the total daily ammonium uptake rates in the
182 non-polynya and polynya regions, respectively which were not significantly different (t-test, $p > 0.05$).

183 The total integral daily nitrogen uptake rate (nitrate + ammonium uptake rates) of phytoplankton
184 was 46.4-443.5 mg N $\text{m}^{-2} \text{d}^{-1}$ ($180.4 \pm 106.7 \text{ mg N m}^{-2} \text{d}^{-1}$) in this study. For the non-polynya and polynya
185 regions, they were 46.4-248.1 mg N $\text{m}^{-2} \text{d}^{-1}$ ($121.6 \pm 89.3 \text{ mg N m}^{-2} \text{d}^{-1}$) and 91.7-443.5 mg N $\text{m}^{-2} \text{d}^{-1}$
186 ($209.8 \pm 107.3 \text{ mg N m}^{-2} \text{d}^{-1}$), respectively. In comparison, the total integral daily nitrogen uptake rates of
187 small phytoplankton were 16.6-46.6 mg N $\text{m}^{-2} \text{d}^{-1}$ ($32.5 \pm 13.2 \text{ mg N m}^{-2} \text{d}^{-1}$) and 17.6-122.0 mg N $\text{m}^{-2} \text{d}^{-1}$
188 ($50.8 \pm 32.4 \text{ mg N m}^{-2} \text{d}^{-1}$) for the non-polynya and polynya regions, respectively. Small phytoplankton
189 contributed 36.2 % (± 23.0 %) to the total integral daily nitrogen uptake rates in the non-polynya region,
190 whereas they contributed 23.5 % (± 6.0 %) for the polynya region (Table 1). The integral daily nitrogen
191 uptake rates and contributions of small phytoplankton were not statistically different between the non-
192 polynya and polynya regions.

193

194 4. Discussion and conclusion

195 The total daily carbon uptake rates of phytoplankton averaged for the non-polynya and polynya
196 regions were $0.42 \text{ g C m}^{-2} \text{ d}^{-1}$ ($\pm 0.30 \text{ g C m}^{-2} \text{ d}^{-1}$) and $0.84 \text{ g C m}^{-2} \text{ d}^{-1}$ ($\pm 0.18 \text{ g C m}^{-2} \text{ d}^{-1}$), respectively in
197 this study. According to the previous reports in the Amundsen Sea (Lee et al., 2012; Kim et al., 2015), the
198 total daily carbon uptake rates ranged from 0.2 to $0.12 \text{ g C m}^{-2} \text{ d}^{-1}$ in the non-polynya region. Our rate
199 ($0.42 \text{ g C m}^{-2} \text{ d}^{-1}$) in the non-polynya region is somewhat higher than those reported previously but they
200 are not significantly different (t-test, $p > 0.05$). In comparison, our total daily carbon uptake rate in the
201 polynya region ($0.84 \text{ g C m}^{-2} \text{ d}^{-1}$) is lower than that ($2.2 \text{ g C m}^{-2} \text{ d}^{-1}$) of Lee et al. (2012) and higher than
202 that ($0.2 \text{ g C m}^{-2} \text{ d}^{-1}$) of Kim et al. (2015). The carbon uptake rates of phytoplankton in Lee et al. (2012)
203 and Kim et al. (2015) were measured during December 21, 2010 to January 23, 2011 and February 11 to
204 March 14, 2012, respectively. Our measurements in this study were executed mainly during January 1-15,
205 2014. For the Amundsen polynya region, a large seasonal variation in the total daily carbon uptake rate of
206 phytoplankton was already reported by Kim et al. (2015) and Arrigo et al. (2012) based on field-measured
207 data and satellite-derived approach, respectively. Generally, late December is the time of peak uptake rate
208 in this region (Arrigo et al., 2012). Previous studies reported that the total daily nitrogen uptake rates in
209 non-polynya region were $0.24 \text{ g N m}^{-2} \text{ d}^{-1}$ during Dec. 21, 2010 to Jan. 23, 2011 and $0.04 \text{ g N m}^{-2} \text{ d}^{-1}$
210 during Feb. 11 to Mar. 14, 2012, whereas the uptake rates in polynya region were $0.93 \text{ g N m}^{-2} \text{ d}^{-1}$ in
211 2010/2011 and $0.06 \text{ g N m}^{-2} \text{ d}^{-1}$ in 2012 in the Amundsen Sea (Lee et al., 2012; Kim et al., 2015). Our
212 total daily nitrogen uptake rates of phytoplankton in non-polynya ($0.12 \pm 0.09 \text{ g N m}^{-2} \text{ d}^{-1}$) and polynya
213 regions ($0.21 \pm 0.11 \text{ g N m}^{-2} \text{ d}^{-1}$) were between the ranges of two previous studies (Lee et al., 2012; Kim
214 et al., 2015). Based on the nitrate and ammonium uptake rates in this study, *f*-ratios (nitrate uptake
215 rate/nitrate+ammonium uptake rates) averaged for non-polynya and polynya regions were 0.62 (± 0.08)
216 and 0.54 (± 0.20), respectively. These ratios were also between the ranges of two previous studies.
217 Although they were not significantly different because of a large spatial variation, larger *f*-ratios in non-
218 polynya than in polynya region are consistent with the results of the previous studies (Lee et al., 2012;

219 Kim et al., 2015). At this point, we do not have a solid explanation for that but a further future study is
220 needed for the higher *f*-ratio mechanism in non-polynya region.

221 The percent contributions of small phytoplankton to chlorophyll-a, POC/PON, daily carbon and
222 nitrogen uptake rates are shown in Table 1. The result of significantly higher chlorophyll-a contribution
223 than the POC contribution of small phytoplankton is consistent with the result in the Chukchi Sea, Arctic
224 Ocean reported by Lee et al. (2013). They explain that higher POC content per chlorophyll-a unit of small
225 phytoplankton could have caused the higher POC contribution in their study (Lee et al., 2013). Given C/N
226 ratio (6.6 ± 0.6) and $\delta^{13}\text{C}$ (-25.9 ± 1.0 ‰) of sample filters attained for POC and PON in this study, our
227 filtered samples are believed to be mainly phytoplankton-originated POC and PON (Kim et al., 2016).
228 Thus, a significant potential overestimated contribution of POC caused by non-phytoplankton materials
229 could be excluded for the higher POC contribution than chlorophyll-a contribution of small
230 phytoplankton. Therefore, small phytoplankton contributions based on conventional assessments of
231 chlorophyll-a concentration might lead to an underestimated contribution of small phytoplankton (Lee et
232 al., 2013). In fact, several authors argue that chlorophyll-a concentration might not be a good index for
233 phytoplankton biomass since it depends largely on environmental factors such as nutrient and light
234 conditions, as well as dominant groups and physiological status of phytoplankton (Desortová 1981;
235 Behrenfeld et al., 2005; Kruskopf and Flynn, 2006; Behrenfeld and Boss 2006). However, the effects of
236 non-phytoplankton carbon materials such as extracellular carbon mucilage can not be completely
237 excluded for the POC contribution as discussed below.

238 The overall contributions of carbon (26.9 %) and nitrogen (27.7 %) uptake rates of small
239 phytoplankton at all the productivity stations in this study are relatively higher than the chlorophyll-a
240 contribution of small phytoplankton but they are not statistically different (t-test, $p > 0.05$). In general, the
241 contribution of daily ammonium uptake rate of small phytoplankton is significantly (t-test, $p < 0.05$)
242 higher than the contribution of daily nitrate uptake rate of small phytoplankton at all the stations in this
243 study. This is well-known for the ammonium preference of small phytoplankton in various regions (Koike
244 et al., 1986; Tremblay et al., 2000, Lee et al., 2008; Lee et al., 2013).

245 In terms of the contributions in different regions, all the contributions (chlorophyll-a, POC/PON,
246 carbon and nitrogen uptake rates) of small phytoplankton were higher in the non-polynya region than in
247 the polynya region (Table 1). In addition, the chlorophyll-a contribution of small phytoplankton ($7.9 \pm$
248 3.5%) was significantly (t-test, $p < 0.05$) lower than the POC contribution ($36.9 \pm 4.6 \%$) in the polynya
249 region, whereas they were not statistically different in the non-polynya region (Table 1). This indicates
250 that small phytoplankton contributed more to the total POC than to the chlorophyll-a concentration in the
251 polynya region. We do not have species compositions of phytoplankton in this study, but previous results
252 reported that *Phaeocystis* sp. are dominant in the Amundsen Sea polynya region (Lee et al., 2012).
253 Generally, *Phaeocystis* spp. release a large amount (up to 46 %) of extracellular carbon mucilage which
254 constitutes their colonial form (Matrai et al., 1995). This non-phytoplankton carbon material without
255 chlorophyll-a might have caused a high POC contribution of small phytoplankton in the polynya region in
256 this study. In fact, the contribution of the daily carbon uptake rates of small phytoplankton ($14.9 \pm 8.4 \%$)
257 was not as high as the POC contribution ($36.9 \pm 4.6 \%$) in the polynya region. The chlorophyll-a
258 contributions of small phytoplankton were lower than those of the daily carbon uptake rate in this study,
259 which is consistent with the results from polynya and marginal ice zone stations in the Ross Sea,
260 Antarctica during austral spring and summer (Saggiomo et al., 1998). They reported that the chlorophyll-a
261 and primary production contributions of pico-phytoplankton ($< 2 \mu\text{m}$) were 29 % and 40 % at polynya
262 stations, whereas the contributions were 17 % and 32 % at marginal ice zone stations, respectively. In the
263 polynya region, they found much higher contributions in chlorophyll-a and primary production of small
264 phytoplankton than those in this study, although their size of the small phytoplankton is somewhat
265 smaller than our size ($< 5 \mu\text{m}$).

266 In conclusion, we found a strong negative correlation ($r^2 = 0.502$, $p < 0.05$) between the
267 productivity contributions of small phytoplankton and total daily carbon uptake rates of total
268 phytoplankton in the Amundsen Sea (Fig. 7), which indicates that daily primary production decreases as
269 small phytoplankton contribution increases. With respect to food quality of small phytoplankton as a
270 basic food source to herbivores, macromolecular compositions such as proteins, lipids, and carbohydrates

271 as photosynthetic-end products will be needed for a better understanding of a small cells-dominant marine
272 ecosystem in response to environmental changes (Lee et al., 2013). According to Kang et al. (accepted),
273 small phytoplankton assimilate more food materials and calorific contents per unit of chlorophyll-a
274 concentration and thus provide more contributions in respect to energy aspect than do other
275 phytoplankton communities in the East/Japan Sea. However, this change in dominant phytoplankton
276 community from large to small cells will likely cause further alteration in the higher trophic levels
277 because of the prey size available to higher trophic grazers (Moline et al., 2004). Monitoring the
278 contributions of small phytoplankton to total biomass and primary production of total phytoplankton
279 community is important, as it provides a valuable indicator to sense environmental changes and
280 consequently their potential influence on higher trophic animals in marine ecosystem.

281

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286

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364 **Table caption**

365 Table 1. Percent contributions (%) of small phytoplankton to depth-integrated total concentrations of
366 chlorophyll-a, POC, PON, and carbon and nitrogen uptake rates in the Amundsen Sea.

367 **Figure captions**

368 Fig. 1. Sampling locations in the Amundsen Sea. Red closed circles represent productivity stations. Sea
369 ice concentration data during the cruise period in 2013 from Nimbus-7 SMMR and DMSP
370 SSM/I-SSMIS Passive Microwave data provided by National Snow & Ice Data Center.

371 Fig. 2. Water column-integrated chlorophyll-a concentrations (mg chl-a m^{-2}) of small ($0.7\text{-}5\ \mu\text{m}$) and
372 large ($> 5\ \mu\text{m}$) phytoplankton at the productivity stations in the Amundsen Sea.

373 Fig. 3. Water column-integrated concentrations of POC (mg C m^{-2}) and PON (mg N m^{-2}) of small ($0.7\text{-}5$
374 μm) and large ($> 5\ \mu\text{m}$) phytoplankton.

375 Fig. 4. Water column-integrated daily carbon uptake rates ($\text{mg C m}^{-2}\ \text{d}^{-1}$) of small ($0.7\text{-}5\ \mu\text{m}$) and large ($>$
376 $5\ \mu\text{m}$) phytoplankton.

377 Fig. 5. Water column-integrated daily nitrate uptake rates ($\text{mg N m}^{-2}\ \text{d}^{-1}$) of small ($0.7\text{-}5\ \mu\text{m}$) and large ($>$
378 $5\ \mu\text{m}$) phytoplankton.

379 Fig. 6. Water column-integrated daily ammonium uptake rates ($\text{mg N m}^{-2}\ \text{d}^{-1}$) of small ($0.7\text{-}5\ \mu\text{m}$) and
380 large ($> 5\ \mu\text{m}$) phytoplankton.

381 Fig. 7. Relationship between productivity contributions (%) of small phytoplankton and the total daily
382 carbon uptake rates ($\text{mg C m}^{-2}\ \text{d}^{-1}$) of phytoplankton (large + small). The total daily carbon
383 uptake rates were transformed into natural logs for a linear regression. Red circles represent for
384 the data obtained in 2013 (this study). Yellow circles representing for the 2012 data
385 (unpublished) were included for a better regression.

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Table 1. Percent contributions (%) of small phytoplankton to depth-integrated total concentrations of chlorophyll-a, POC, PON, and carbon and nitrogen uptake rates in the Amundsen Sea.

	Chlorophyll-a	POC	PON	Daily carbon uptake rate	Daily nitrate uptake rate	Daily ammonium uptake rate	Total nitrogen uptake rate
All stations	19.4 ± 26.0	41.1 ± 10.6	41.3 ± 11.5	26.9 ± 29.3	21.5 ± 11.1	38.7 ± 24.9	27.7 ± 14.4
Non-polynya	42.4 ± 37.2	49.5 ± 14.4	50.0 ± 15.1	50.8 ± 42.8	28.2 ± 15.9	52.8 ± 40.5	36.2 ± 23.0
Polynya	7.9 ± 3.5	36.9 ± 4.6	37.0 ± 6.9	14.9 ± 8.4	18.1 ± 6.8	31.6 ± 10.1	23.5 ± 6.0

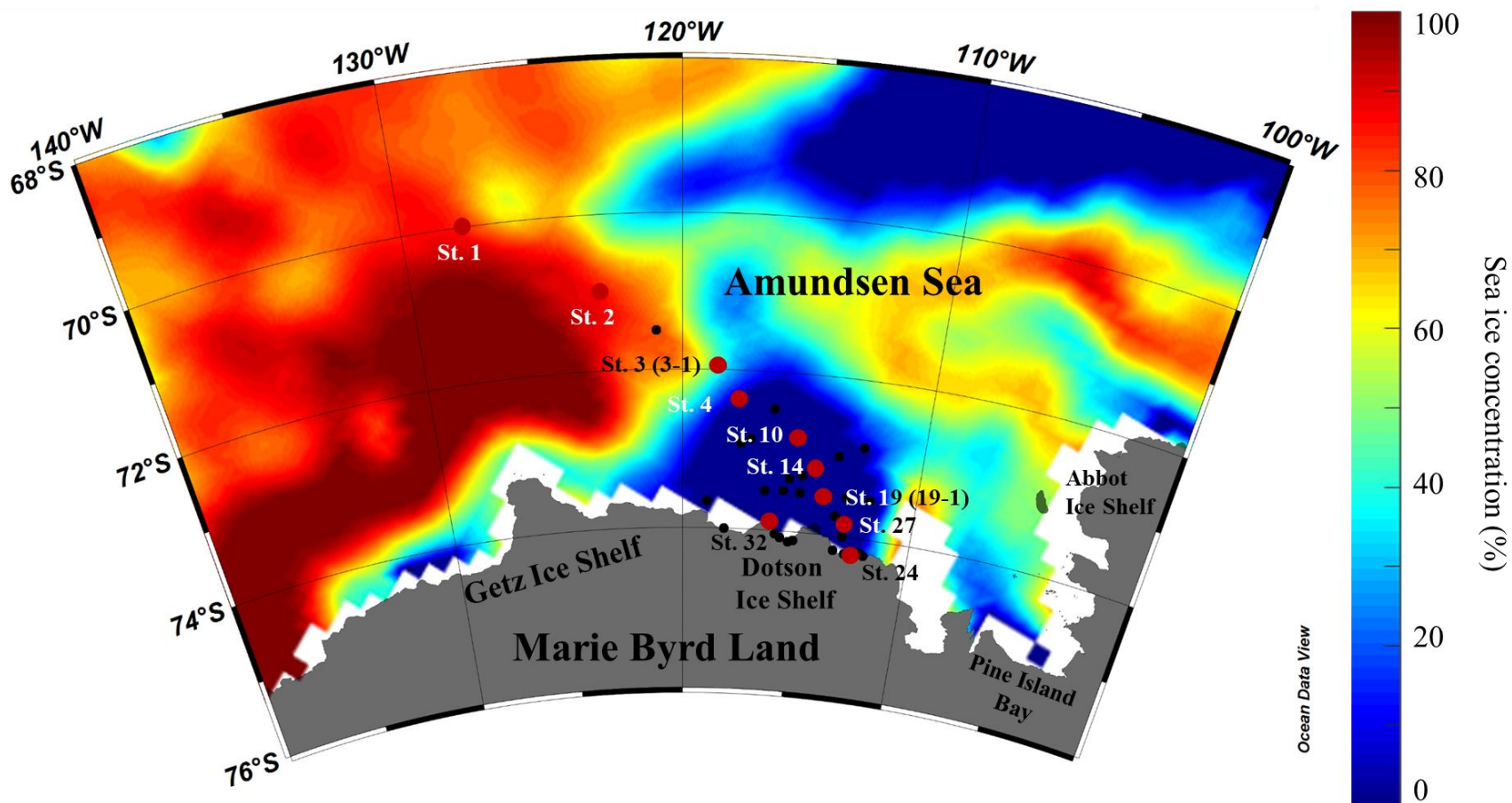


Fig. 1. Sampling locations in the Amundsen Sea. Red closed circles represent productivity stations. Sea ice concentration data during the cruise period in 2013 from Nimbus-7 SMMR and DMSP SSM/I-SSMIS Passive Microwave data provided by National Snow & Ice Data Center.

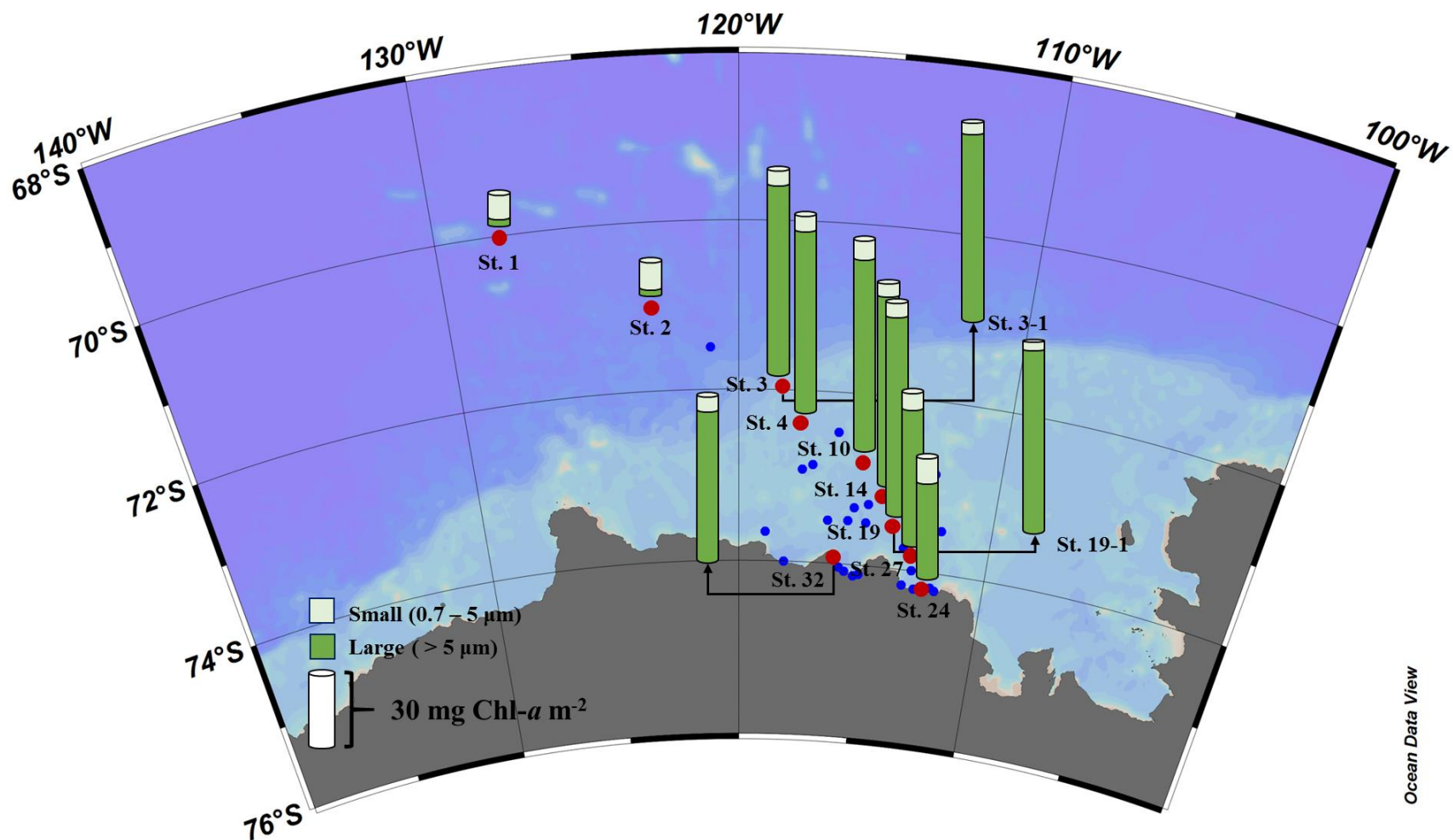


Fig. 2. Water column-integrated chlorophyll-a concentrations (mg chl-a m^{-2}) of small ($0.7\text{--}5\ \mu\text{m}$) and large ($> 5\ \mu\text{m}$) phytoplankton at the productivity stations in the Amundsen Sea.

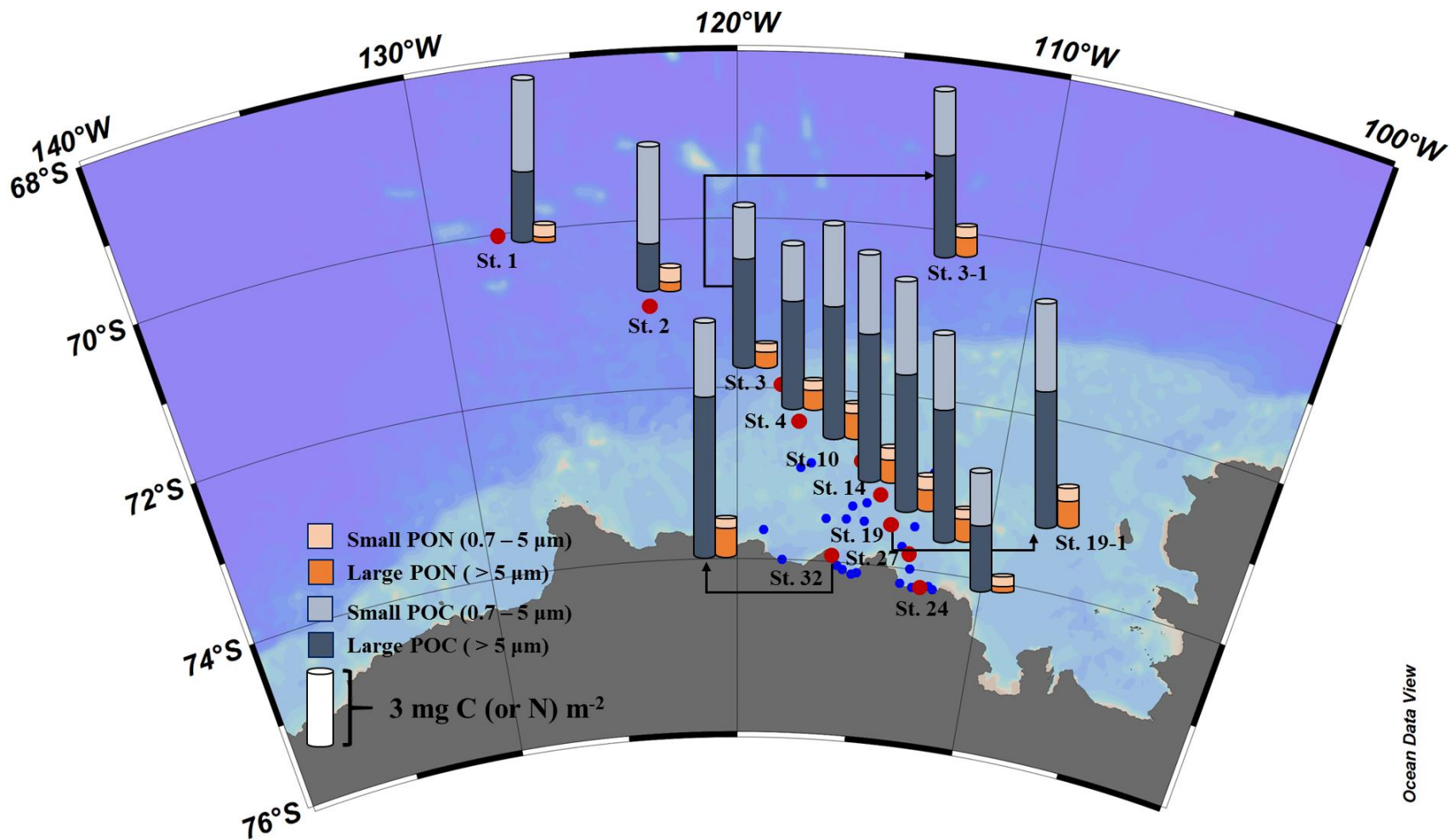


Fig. 3. Water column-integrated concentrations of POC (mg C m^{-2}) and PON (mg N m^{-2}) of small ($0.7-5 \mu\text{m}$) and large ($> 5 \mu\text{m}$) phytoplankton.

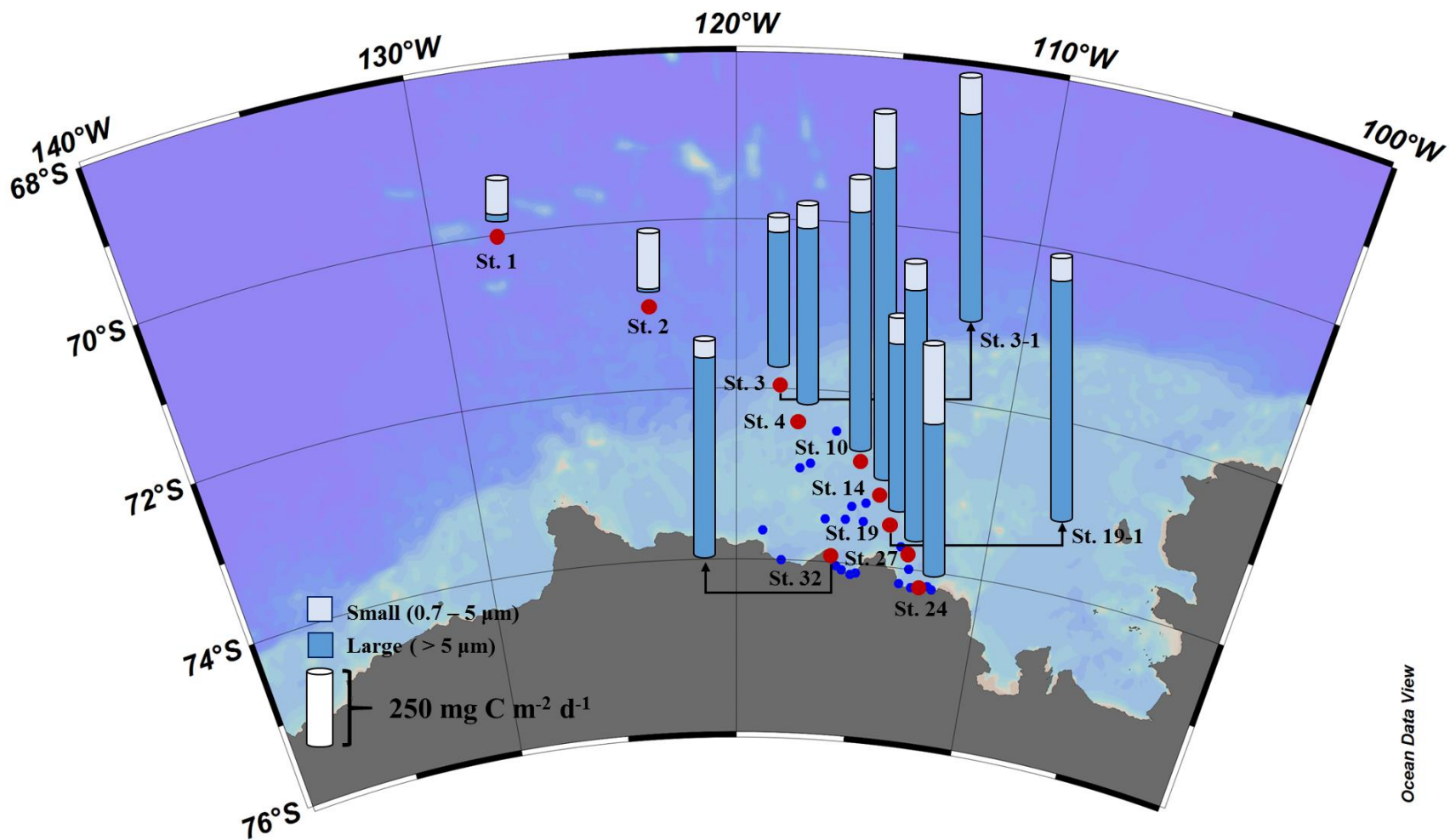


Fig. 4. Water column-integrated daily carbon uptake rates ($\text{mg C m}^{-2} \text{d}^{-1}$) of small ($0.7\text{-}5 \mu\text{m}$) and large ($> 5 \mu\text{m}$) phytoplankton.

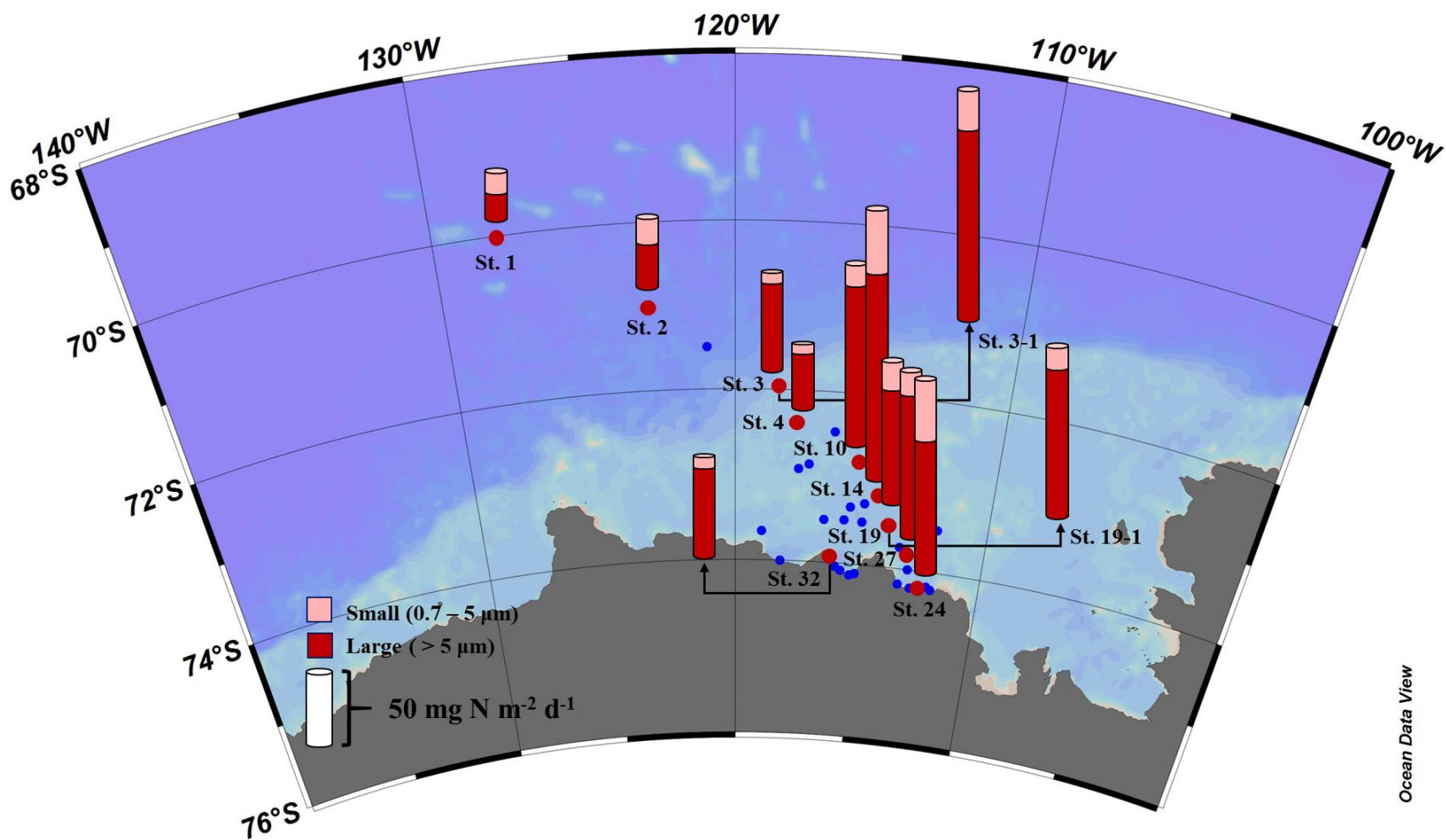
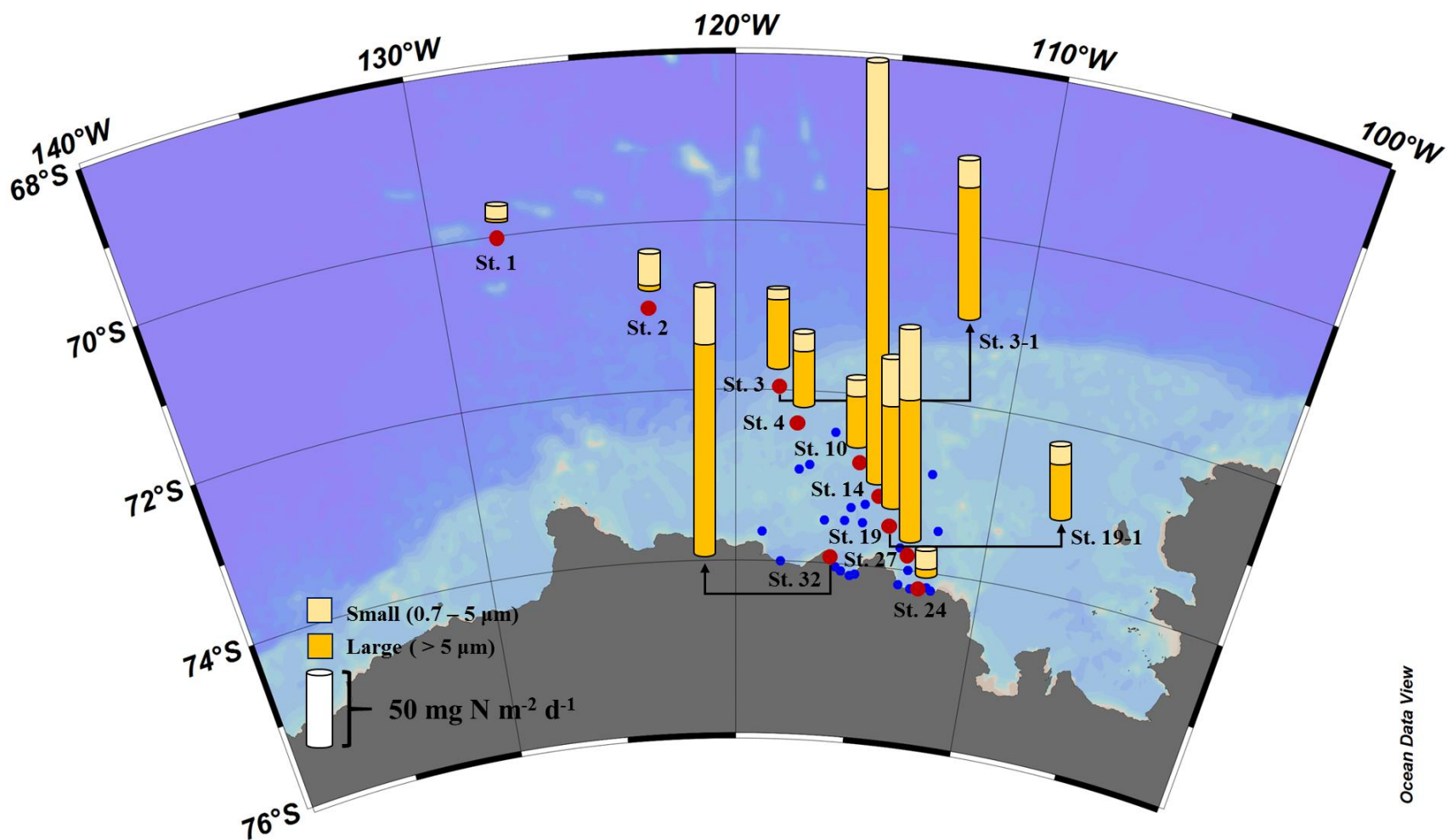


Fig. 5. Water column-integrated daily nitrate uptake rates ($\text{mg N m}^{-2} \text{d}^{-1}$) of small ($0.7\text{--}5 \mu\text{m}$) and large ($> 5 \mu\text{m}$) phytoplankton.



Ocean Data View

Fig. 6. Water column-integrated daily ammonium uptake rates ($\text{mg N m}^{-2} \text{d}^{-1}$) of small ($0.7\text{--}5 \mu\text{m}$) and large ($> 5 \mu\text{m}$) phytoplankton.

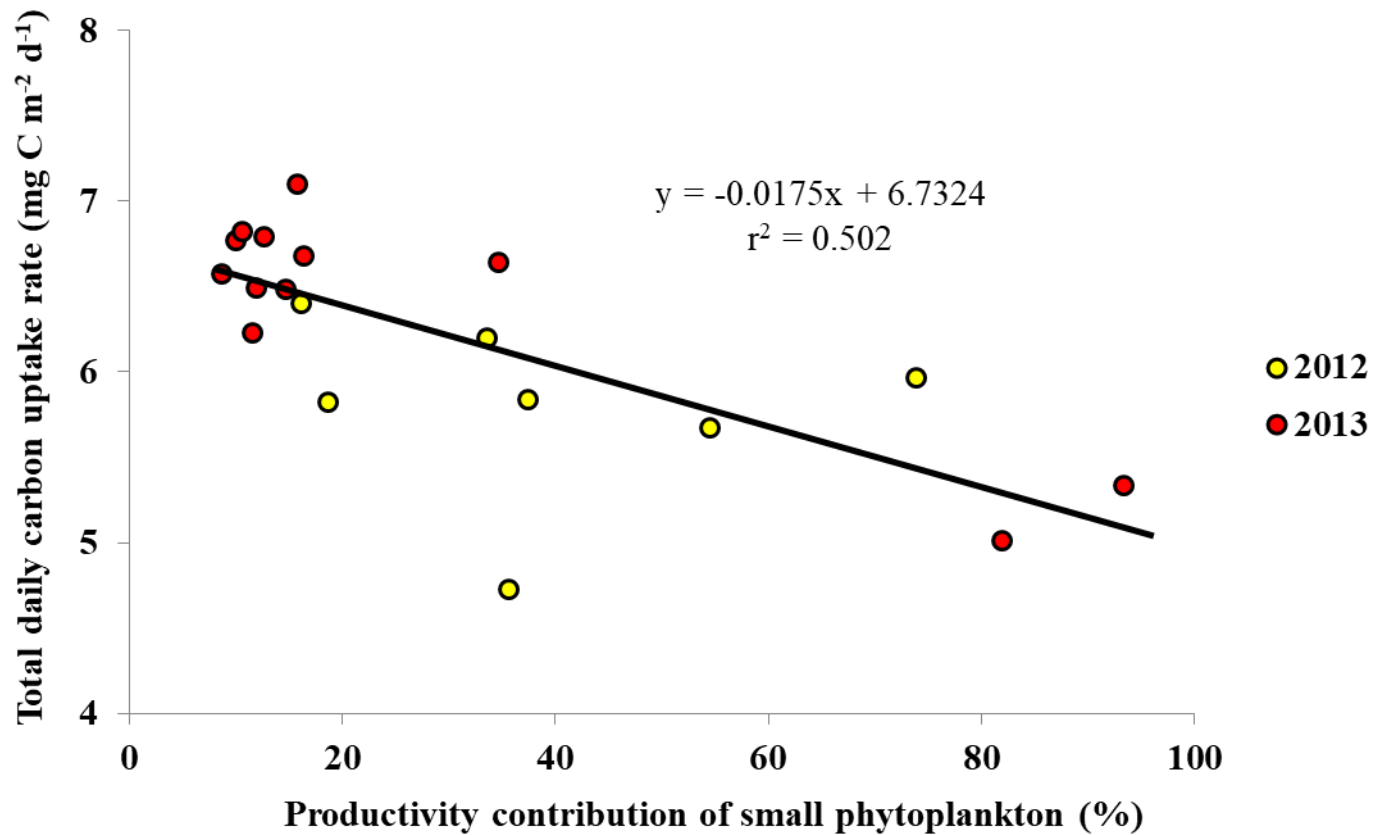


Fig. 7. Relationship between productivity contributions (%) of small phytoplankton and the total daily carbon uptake rates (mg C m⁻² d⁻¹) of phytoplankton (large + small). The total daily carbon uptake rates were transformed into natural logs for a linear regression. Red circles represent for the data obtained in 2013 (this study). Yellow circles representing for the 2012 data (unpublished) were included for a better regression.