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1 **Small phytoplankton contribution to the total primary production in**
2 **the Amundsen Sea**

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17 **Abstract**

18 Small-sized phytoplankton is anticipated to be more important for phytoplankton community in
19 a recent changing ocean condition. However, little information on the contribution of small-sized
20 phytoplankton to overall phytoplankton production is currently available in the Amundsen Sea. To
21 determine the contributions of small-sized phytoplankton to total biomass and primary production, carbon
22 and nitrogen uptake rates of total and small-sized phytoplankton were obtained from 12 productivity
23 stations in the Amundsen Sea. The daily carbon uptake rates of total phytoplankton averaged in this study
24 were $0.42 \text{ g C m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.30 \text{ g C m}^{-2} \text{ d}^{-1}$) and $0.84 \text{ g C m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.18 \text{ g C m}^{-2} \text{ d}^{-1}$) whereas
25 the daily total nitrogen (nitrate and ammonium) uptake rates were $0.12 \text{ g N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.09 \text{ g N m}^{-2}$
26 d^{-1}) and $0.21 \text{ g N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.11 \text{ g N m}^{-2} \text{ d}^{-1}$), respectively for non-polynya and polynya regions,
27 which were within the ranges reported previously. Small phytoplankton contributed 26.9 % and 27.7 % to
28 the total carbon and nitrogen uptake rates of phytoplankton in this study, respectively, which were
29 relatively higher than the chlorophyll-a contribution (19.4 %) of small phytoplankton. For a comparison
30 of different regions, the contributions for chlorophyll-a concentration and primary production of small
31 phytoplankton averaged from all the non-polynya stations were 42.4 % and 50.8 %, which were
32 significantly higher than those (7.9 % and 14.9 %, respectively) in polynya region. A strong negative
33 correlation ($r^2 = 0.790$, $p < 0.05$) was found between the contributions of small phytoplankton and the
34 total daily primary production of phytoplankton in this study. This finding implies that daily primary
35 production decreases as small phytoplankton contribution increases, which is mainly due to the lower
36 carbon uptake rate of small phytoplankton than large phytoplankton. Under ongoing environmental
37 changes caused by global warming, a potential decrease of total primary production would be led by
38 increasing contribution of small phytoplankton in the Amundsen Sea.

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41 **Keywords:** Phytoplankton, Primary production, Polynya, Amundsen Sea

42



43 1. Introduction

44 The Amundsen Sea is located in the West Antarctica between the Ross Sea and Bellingshausen
45 Sea, which is one of the least-biologically studied regions in the Southern Ocean. Recently several
46 international research programs (KOPRI Amundsen project, iSTAR, ASPIRE, and DynaLiFe) were
47 launched to understand this remote area. Field-measurement data revealed that annual primary production
48 of phytoplankton reaching to $220 \text{ g C m}^{-2} \text{ y}^{-1}$ in the Amundsen Sea polynya is as high as that of Ross Sea
49 polynya ($200 \text{ g C m}^{-2} \text{ y}^{-1}$) which was previously known for the highest productivity region in the Southern
50 Ocean (Lee et al., 2012). Given the fact that the chl-*a* concentration averaged from all the chl-*a* measured
51 stations was twice higher than that of the only productivity-measured stations, Lee et al., (2012) argued
52 that the annual production in the Amundsen Sea polynya could be even two-fold higher than that of Ross
53 Sea polynya.

54 Over the past several decades a rapid climate change has been detected and subsequently
55 physical changes have occurred in the marine ecosystem in the western Antarctic Peninsula (WAP)
56 (Rückamp et al. 2011; Ducklow et al. 2007). Recent studies revealed that the Thwaites Glacier in Pine
57 Island Bay is retreating fast and the ice volume loss in the nearby Getz Ice shelf is accelerating (Joughin
58 et al., 2014; Paolo et al., 2015). Shoaling warm Circumpolar Deep Water is believed to be a main reason
59 for the ice sheet mass loss largely caused by ice shelf basal melt underside of the ice shelves (Yager et al.
60 2012; Schmidtko et al. 2014). The climate change from a cold-dry polar-type to a warm-humid sub-
61 Antarctic-type drives subsequent changes in ocean biological productivity along the WAP shelf over the
62 recent three decades (Montes-Hugo et al. 2009).

63 Phytoplankton as the base of oceanic food webs can be an indicator for changes in marine
64 ecosystems responding to current climate changes (Moline et al., 2004; Wassman et al., 2011; Arrigo et
65 al., 2015). In an expecting warmer ocean condition, small-sized phytoplankton is anticipated to contribute
66 more to total phytoplankton community and thus marine ecosystems (Morán et al., 2010; Li et al., 2009;
67 Lee et al., 2013). In consistent, Li et al. (2009) found increasing small-sized phytoplankton in the Canada
68 Basin in the Arctic Ocean under freshening surface waters which results in a stronger stratification and



69 lower nutrient supply in the upper water column. Moreover, in the Antarctic Ocean, Moline et al. (2004)
70 found a consistent transition from large diatoms to small cryptophytes associated with glacial melt water
71 in the coastal waters along the Antarctic Peninsula in response to a regional warming trend. This change
72 in dominant phytoplankton community from large to small cells will likely cause further alteration of
73 higher trophic levels and subsequent food web (Moline et al., 2004). However, little information on the
74 contribution of small-sized phytoplankton to primary production is available in the Antarctic Ocean
75 (Saggiomo et al. 1998), especially in the Amundsen Sea with a rapid melting of ice shelf (Yager et al.
76 2012; Schmidtko et al. 2014). Thus, the main objective in this study is to determine what extend small-
77 sized phytoplankton contributes to overall total biomass and primary production in the Amundsen Sea to
78 lay the groundwork for the future monitoring of marine ecosystem change responding to ongoing changes
79 in environmental conditions.

80

81 **2. Materials and methods**

82 *2.1. Samplings*

83 Water samples were collected for carbon and nitrogen uptake measurements of phytoplankton in
84 the Amundsen Sea during the KOPRI Amundsen cruise from 1 to 15 January, 2014 onboard the Korean
85 Research Icebreaker ship *Araon*. Using a dual stable isotope technique (Lee et al., 2012; Kim et al., 2015),
86 the experiments of carbon and nitrogen uptake rates of phytoplankton were conducted at 12 selected
87 productivity stations including 2 revisited-stations (St. 3-1 and St. 19-1) when on-deck incubations were
88 available during daytime at oceanographic survey stations. Based on sea ice concentration data from
89 National Snow & Ice Data Center during the cruise period (Fig. 1), our study region was further separated
90 into polynya and non-polynya areas for comparison based on sea ice distribution and concentration during
91 the cruise period. Four stations (St. 1, St. 2, St. 3, and St. 3-1) among the 12 stations were belong to non-
92 polynya region and the rest of the stations were belong to polynya region.

93 After 6 light depths (100, 50, 30, 12, 5, and 1% penetration of the surface irradiance, PAR) were
94 determined with an LI-COR underwater 4π light sensor, water samples for the uptake experiments as



95 well as biological and chemical property analysis were obtained from a CTD-rosette sampler system
96 equipped with 24 10-L Niskin bottles.

97 *2.2. Total and size-fractionated chlorophyll-a concentration*

98 Water samples for total and size-fractionated chlorophyll-a concentrations of phytoplankton were
99 obtained at the 12 productivity stations. Total chlorophyll-a concentrations were measured at six different
100 light depths (100, 50, 30, 12, 5 and 1% of PAR). For size-fractionated chlorophyll-a concentrations, water
101 samples were collected at three light depths (100, 30, and 1 %). Water samples (0.3–0.5 L) for total
102 chlorophyll-a concentrations were filtered using Whatman glass fiber filters (GF/F; 25 mm). For different
103 size-fractionated chlorophyll-a concentrations water samples (0.7–1 L) were passed sequentially through
104 20 and 5 μm Nucleopore filters (47 mm) and 0.7 μm GF/F filters (47 mm). After the filters were extracted
105 using the method described by Kim et al. (2015), all chlorophyll-a concentrations were subsequently
106 determined onboard using a Trilogy fluorometer (Turner Designs, USA). The methods and calculations
107 for chlorophyll-a were based on Parsons et al. (1984).

108 *2.3. Carbon and nitrogen uptake experiments*

109 Carbon and nitrogen uptake experiments of phytoplankton were executed by a ^{13}C - ^{15}N dual
110 isotope tracer technique previously applied for the Amundsen Sea (Lee et al. 2012; Kim et al. 2015). In
111 this study, basically we followed same procedure of Lee et al. (2012). In brief, six light depths (100, 50,
112 30, 12, 5, and 1%) were determined with an LI-COR underwater 4π light sensor (LI-COR Inc., Lincoln,
113 Nebraska, USA) lowered with CTD/rosette sampler. Water sample from each light depth was transferred
114 into different screened polycarbonate incubation bottle (1 L) which matches with each light depth. The
115 productivity bottles were incubated in large polycarbonate material incubators cooled with running
116 surface seawater on deck under natural light conditions, after the water samples were inoculated with
117 labeled carbon ($\text{NaH}^{13}\text{CO}_3$) and nitrate (K^{15}NO_3) or ammonium ($^{15}\text{NH}_4\text{Cl}$) substrates. After 4–5 h
118 incubations, the incubated waters were well mixed and distributed into two filtration sets for the carbon
119 and nitrogen uptake rates of total ($> 0.7 \mu\text{m}$) and small-sized cells ($< 5 \mu\text{m}$). The incubated waters (0.3 L)



120 for total uptake rates were filtered through pre-combusted GF/F filters (24 mm diameter), whereas waters
121 samples (0.5 L) for the uptake rates of small-sized cells were passed through 5 μm Nuclepore filters (47
122 mm) to remove large-sized cells ($> 5 \mu\text{m}$) and then the filtrate was passed through GF/F (24 mm) for the
123 small-sized cells (Lee et al., 2013). The values for large phytoplankton in this study were obtained from
124 the difference between small and total fractions (Lee et al., 2013). The filters were immediately preserved
125 at -80°C until mass spectrometric analysis. After acid fuming overnight to remove carbonate, the
126 concentrations of particulate organic carbon (POC) and nitrogen (PON) and the abundance of ^{13}C and
127 ^{15}N were determined by a Finnigan Delta+XL mass spectrometer at the Alaska Stable Isotope Facility,
128 USA.

129 All contribution results of small phytoplankton in this study were estimated from comparison of
130 small phytoplankton to total phytoplankton integral values from 100 to 1 % light depth at each station
131 based on the trapezoidal rule. Daily carbon and nitrogen uptake rates of phytoplankton were based on our
132 hourly uptake rates measured in this study and a 24-h photoperiod per day during the summer period in
133 the Amundsen Sea (Lee et al., 2012).

134

135 3. Results

136 3.1. Chlorophyll-a, POC, and PON contributions of small phytoplankton

137 The total (large + small phytoplankton) chlorophyll-a concentration integrated from six different
138 light depths ranged from 11.1 to 80.3 mg chl-a m^{-2} (mean \pm S.D. = $57.4 \pm 25.2 \text{ mg chl-a m}^{-2}$), whereas
139 small ($< 5 \mu\text{m}$) chlorophyll-a concentration ranged from 3.9 to 9.4 mg chl-a m^{-2} (mean \pm S.D. = 5.7 ± 1.7
140 mg chl-a m^{-2}) in this study (Fig. 2). The average contribution of small phytoplankton to the total
141 chlorophyll-a concentration was 19.4 % (S.D. = ± 26.0 %) ranging from 4.9 to 76.5 %. In the Amundsen
142 Sea, large phytoplankton ($> 5 \mu\text{m}$) were generally predominant (approximately 80 %) during the cruise
143 period in 2014 based on different-sized chlorophyll-a concentrations. For a regional comparison, the
144 average contributions of small phytoplankton to the total chlorophyll-a concentration were 42.4 % (S.D. =



145 ± 37.2 %) and 7.9 % (S.D. = ± 3.5 %) for non-polynya and polynya regions, respectively. The
146 chlorophyll-a contribution of small phytoplankton was larger in the non-polynya stations than the polynya
147 stations although they were not significantly different (t-test, $p = 0.16$).

148 The total integral POC concentration of phytoplankton displayed no large spatial variation
149 ranging from 4.72 to 9.22 mg C m⁻² (Fig. 3). In comparison, the total integral PON concentration of
150 phytoplankton ranged from 0.76 to 1.74 mg C m⁻². The POC contribution of small phytoplankton ranged
151 from 30.7 to 65.5 % (mean \pm S.D. = 41.1 ± 10.6 %), whereas the PON contribution ranged similarly from
152 30.8 to 67.2 % (mean \pm S.D. = 41.3 ± 11.5 %) in the Amundsen Sea (Fig. 3). Specifically, the POC and
153 PON contributions of small phytoplankton averaged from all the polynya stations were 36.9 % (S.D. = \pm
154 4.6 %) and 37.0 % (S.D. = ± 6.9 %), respectively. In comparison, the POC and PON contributions of
155 small phytoplankton averaged from the non-polynya stations were 49.5 % (S.D. = ± 14.4 %) and 50.0 %
156 (S.D. = ± 15.1 %), respectively.

157 3.2. Carbon uptake rate contributions of small phytoplankton

158 The total daily carbon uptake rates of phytoplankton (large + small phytoplankton) integrated
159 from six different light depths ranged from 150.4 to 1213.4 mg C m⁻² d⁻¹ with an average of 696.5 mg C
160 m⁻² d⁻¹ (S.D. = ± 298.4 mg C m⁻² d⁻¹) in this study (Fig. 4). In contrast, the rates of small phytoplankton
161 ranged between 58.6 and 266.4 mg C m⁻² d⁻¹ with an average of 124.9 mg C m⁻² d⁻¹ (S.D. = ± 62.4 mg C
162 m⁻² d⁻¹). Small phytoplankton contributed 26.9 % (S.D. = ± 29.3 %) to total daily carbon uptake rate of
163 total phytoplankton.

164 Specifically, the total daily carbon uptake rates of phytoplankton ranged from 150.4 to 796.4 mg
165 C m⁻² d⁻¹ with an average of 415.0 mg C m⁻² d⁻¹ (S.D. = ± 298.2 mg C m⁻² d⁻¹) in non-polynya region,
166 whereas they ranged from 654.8 to 1213.4 mg C m⁻² d⁻¹ with an average of 837.3 mg C m⁻² d⁻¹ (S.D. = \pm
167 184.1 mg C m⁻² d⁻¹) in polynya region. The total daily carbon uptake rates of phytoplankton were
168 significantly higher (t-test, $p < 0.05$) in polynya stations than non-polynya stations. The rates of small
169 phytoplankton ranged between 58.6 and 193.6 mg C m⁻² d⁻¹ with an average of 126.5 mg C m⁻² d⁻¹ (S.D. =
170 ± 55.2 mg C m⁻² d⁻¹) in non-polynya region, whereas they ranged from 62.2 to 266.4 mg C m⁻² d⁻¹ with an



171 average of $124.1 \text{ mg C m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 69.3 \text{ mg C m}^{-2} \text{ d}^{-1}$) in polynya region. The daily carbon uptake
172 rates of small phytoplankton were not significantly different (t-test, $p > 0.05$) between polynya and non-
173 polynya stations. The average contributions of small phytoplankton to total daily carbon uptake rates were
174 50.8 % (S.D. = $\pm 42.8 \%$) and 14.9 % (S.D. = $\pm 8.4 \%$), respectively for non-polynya and polynya regions.
175 The average contributions were largely different between polynya and non-polynya regions but they were
176 not statistically significant (t-test, $p > 0.05$).

177 3.3. Nitrogen uptake rate contributions of small phytoplankton

178 The total daily nitrate uptake rates of phytoplankton (large + small phytoplankton) ranged from
179 34.0 to $174.2 \text{ mg N m}^{-2} \text{ d}^{-1}$ with an average of $93.7 \text{ mg N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 43.2 \text{ mg N m}^{-2} \text{ d}^{-1}$), whereas the
180 rates of small phytoplankton ranged from 6.1 to $40.9 \text{ mg N m}^{-2} \text{ d}^{-1}$ with an average of $19.0 \text{ mg N m}^{-2} \text{ d}^{-1}$
181 (S.D. = $\pm 11.3 \text{ mg N m}^{-2} \text{ d}^{-1}$) in this study (Fig. 5). Small phytoplankton contributed 21.5 % (S.D. = \pm
182 11.1 %) to total daily nitrate uptake rates. In comparison, the total daily ammonium uptake rates of
183 phytoplankton ranged from 12.4 to $173.8 \text{ mg N m}^{-2} \text{ d}^{-1}$ with an average of $86.7 \text{ mg N m}^{-2} \text{ d}^{-1}$ (S.D. = \pm
184 $75.9 \text{ mg N m}^{-2} \text{ d}^{-1}$), whereas the rates of small phytoplankton ranged from 9.1 to $81.1 \text{ mg N m}^{-2} \text{ d}^{-1}$ with
185 an average of $25.7 \text{ mg N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 21.1 \text{ mg N m}^{-2} \text{ d}^{-1}$) in this study (Fig. 6). Small phytoplankton
186 contributed 38.7 % (S.D. = $\pm 24.9 \%$) to total daily ammonium uptake rates. The contributions of small
187 phytoplankton were significantly higher in ammonium uptake rate than nitrate uptake rate (t-test, $p <$
188 0.05).

189 Specifically for different regions, the total daily nitrate uptake rates of phytoplankton ranged from
190 34.0 to $142.1 \text{ mg N m}^{-2} \text{ d}^{-1}$ with an average of $71.9 \text{ mg N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 48.4 \text{ mg N m}^{-2} \text{ d}^{-1}$) in non-
191 polynya region whereas they ranged from 44.2 to $174.2 \text{ mg N m}^{-2} \text{ d}^{-1}$ with an average of $104.6 \text{ mg N m}^{-2}$
192 d^{-1} (S.D. = $\pm 39.0 \text{ mg N m}^{-2} \text{ d}^{-1}$) in polynya region, respectively. In comparison, the daily nitrate uptake
193 rates of small phytoplankton ranged from 7.5 to $26.6 \text{ mg N m}^{-2} \text{ d}^{-1}$ with an average of $16.7 \text{ mg N m}^{-2} \text{ d}^{-1}$
194 (S.D. = $\pm 7.8 \text{ mg N m}^{-2} \text{ d}^{-1}$) and from 6.1 to $40.9 \text{ mg N m}^{-2} \text{ d}^{-1}$ with an average of $20.1 \text{ mg N m}^{-2} \text{ d}^{-1}$ (S.D.
195 = $\pm 13.1 \text{ mg N m}^{-2} \text{ d}^{-1}$), respectively for non-polynya and polynya regions. The contributions of small
196 phytoplankton to the total daily nitrate uptake rates were 28.2 % (S.D. = $\pm 15.9 \%$) in non-polynya region



197 and 18.1 % (S.D. = ± 6.8 %) in polynya region, respectively. The total daily ammonium uptake rates of
198 total phytoplankton ranged between 12.3 and 106.1 mg N m⁻² d⁻¹ (mean \pm S.D. = 49.7 ± 41.2 mg N m⁻² d⁻¹)
199 in non-polynya region and between 18.1 and 269.3 mg N m⁻² d⁻¹ (mean \pm S.D. = 105.2 ± 84.6 mg N m⁻² d⁻¹)
200 ¹) in polynya region. In comparison, the rates of small phytoplankton ranged between 9.1 and 22.4 mg N
201 m⁻² d⁻¹ (mean \pm S.D. = 15.8 ± 6.4 mg N m⁻² d⁻¹) in non-polynya region and between 9.9 and 81.1 mg N m⁻²
202 ² d⁻¹ (mean \pm S.D. = 30.7 ± 24.5 mg N m⁻² d⁻¹) in polynya region. Small phytoplankton contributed 52.8 %
203 (S.D. = ± 40.5 %) and 31.6 % (S.D. = ± 10.1 %) to the total daily ammonium uptake rates in non-polynya
204 and polynya regions, respectively which were not significantly different (t-test, p = 0.37).

205 The total integral daily nitrogen uptake rate (nitrate + ammonium uptake rates) of phytoplankton
206 ranged from 46.4 to 443.5 mg N m⁻² d⁻¹ (mean \pm S.D. = 180.4 ± 106.7 mg N m⁻² d⁻¹) in this study. For
207 non-polynya and polynya regions, they ranged from 46.4 to 248.1 mg N m⁻² d⁻¹ (mean \pm S.D. = $121.6 \pm$
208 89.3 mg N m⁻² d⁻¹) and from 91.7 to 443.5 mg N m⁻² d⁻¹ (mean \pm S.D. = 209.8 ± 107.3 mg N m⁻² d⁻¹),
209 respectively. In comparison, the total integral daily nitrogen uptake rate of small phytoplankton ranged
210 from 16.6 to 46.6 mg N m⁻² d⁻¹ (mean \pm S.D. = 32.5 ± 13.2 mg N m⁻² d⁻¹) and from 17.6 to 122.0 mg N m⁻²
211 ² d⁻¹ (mean \pm S.D. = 50.8 ± 32.4 mg N m⁻² d⁻¹) for non-polynya and polynya regions, respectively. Small
212 phytoplankton contributed 36.2 % (S.D. = ± 23.0 %) to the total integral daily nitrogen uptake rates in
213 non-polynya region, whereas they contributed 23.5 % (S.D. = ± 6.0 %) for polynya region. The integral
214 daily nitrogen uptake rates and contributions of small phytoplankton were not statistically different
215 between non-polynya and polynya regions.

216

217 4. Discussion and conclusion

218 The total daily carbon uptake rates of phytoplankton averaged for non-polynya and polynya
219 regions were 0.42 g C m⁻² d⁻¹ (S.D. = ± 0.30 g C m⁻² d⁻¹) and 0.84 g C m⁻² d⁻¹ (S.D. = ± 0.18 g C m⁻² d⁻¹),
220 respectively in this study. According to the previous reports in the Amundsen Sea (Lee et al., 2012; Kim
221 et al., 2015), the total daily carbon uptake rates ranged from 0.2 to 0.12 g C m⁻² d⁻¹ in non-polynya region.
222 Our rate (0.42 g C m⁻² d⁻¹) in non-polynya region is somewhat higher than those reported previously but



223 they are not significantly different (t -test, $p = 0.77$). In comparison, our total daily carbon uptake rate in
224 polynya region ($0.84 \text{ g C m}^{-2} \text{ d}^{-1}$) is within the range between Lee et al. (2012; $2.2 \text{ g C m}^{-2} \text{ d}^{-1}$) and Kim et
225 al. (2015; $0.2 \text{ g C m}^{-2} \text{ d}^{-1}$). For the Amundsen polynya region, a large seasonal variation in the total daily
226 carbon uptake rate of phytoplankton was already reported by Kim et al. (2015) and Arrigo et al. (2012)
227 based on field-measured data and satellite-derived approach, respectively. It is appeared that this seasonal
228 variation largely depends on the bloom stage of phytoplankton which peaks during the late December-
229 January and terminates at late February (Arrigo and van Dijken 2003; Arrigo et al., 2012; Kim et al.,
230 2015). The carbon uptake rates of phytoplankton in Lee et al. (2012) and Kim et al. (2015) were measured
231 during December 21-January 23, 2010 and February 11 to March 14, 2012, respectively. Our
232 measurements in this study were executed mainly during January 1-15, 2014.

233 The total daily nitrogen uptake rates of phytoplankton were $0.12 \text{ g N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.09 \text{ g N m}^{-2}$
234 d^{-1}) and $0.21 \text{ g N m}^{-2} \text{ d}^{-1}$ (S.D. = $\pm 0.11 \text{ g N m}^{-2} \text{ d}^{-1}$) for non-polynya and polynya regions, respectively
235 in this study. Previous studies reported that the total daily nitrogen uptake rates in non-polynya region
236 were $0.24 \text{ g N m}^{-2} \text{ d}^{-1}$ in 2010/2011 and $0.04 \text{ g N m}^{-2} \text{ d}^{-1}$ in 2012 whereas the uptake rates in polynya
237 region were $0.93 \text{ g N m}^{-2} \text{ d}^{-1}$ in 2010/2011 and $0.06 \text{ g N m}^{-2} \text{ d}^{-1}$ in 2012 in the Amundsen Sea (Lee et al.,
238 2012; Kim et al., 2015). Our total daily nitrogen uptake rates of phytoplankton in non-polynya and
239 polynya regions were between the two previous studies (Lee et al., 2012; Kim et al., 2015). Based on the
240 nitrate and ammonium uptake rates in this study, f -ratios (nitrate uptake rate/nitrate+ammonium uptake
241 rates) averaged for non-polynya and polynya regions were 0.62 (S.D. = ± 0.08) and 0.54 (S.D. = ± 0.20),
242 respectively. These ratios also were between the two previous studies. Although they were not significant
243 different because of a large spatial variation, larger f -ratios in non-polynya than in polynya region are
244 consistent with the results of the previous studies (Lee et al., 2012; Kim et al., 2015). At this point, we do
245 not have a solid explanation for that but a further future study is needed for the higher f -ratio mechanism
246 in non-polynya region.

247 The percent contributions of small phytoplankton in terms of chlorophyll-*a*, POC/PON, daily
248 carbon and nitrogen uptake rates are shown in Table 1. The overall contribution of small phytoplankton to



249 the total chlorophyll-a concentration for all the productivity stations was 19.4 % (S.D. = ± 26.0 %) which
250 is significantly (*t*-test, $p < 0.05$) lower than the POC contribution (41.1 ± 10.6 %). This is consistent with
251 the result in the Chukchi Sea, Arctic Ocean reported by Lee et al. (2013). They explained that higher POC
252 content per chlorophyll-a unit of small phytoplankton could cause their higher POC contribution (Lee et
253 al., 2013). Given C/N ratio (mean \pm S.D. = 6.6 ± 0.6) and $\delta^{13}\text{C}$ (mean \pm S.D. = -25.9 ± 1.0 ‰) of sample
254 filters attained for POC and PON in this study, our filtered samples are believed to be mainly
255 phytoplankton-originated POC and PON (Kim et al., 2016). Thus, a significant potential overestimated
256 POC contribution of non-phytoplankton materials could be excluded for the higher POC contribution than
257 chlorophyll-a contribution of small phytoplankton. Therefore, small phytoplankton contributions based on
258 conventional assessments of chlorophyll-a concentration might lead an underestimated contribution of
259 small phytoplankton (Lee et al., 2013). In fact, several authors argued that chlorophyll-a concentration
260 might be not a good index for phytoplankton biomass since it largely depends on environmental factors
261 such as nutrient and light conditions as well as dominant groups and physiological status of
262 phytoplankton (Desortová 1981; Behrenfeld et al., 2005; Kruskopf and Flynn, 2006; Behrenfeld and Boss
263 2006). However, the effects of non-phytoplankton carbon materials such as extracellular carbon mucilage
264 can not be completely excluded for the POC contribution as discussed below.

265 The overall contributions of carbon and nitrogen (nitrate and ammonium) uptake rates of small
266 phytoplankton at all the productivity stations in this study are similar with 26.9 % and 27.7 %,
267 respectively. These contributions are relatively higher than the chlorophyll-a contribution of small
268 phytoplankton but they are not statistically different (*t*-test, $p > 0.05$). In general, the contribution of daily
269 ammonium uptake rate of small phytoplankton are significantly (*t*-test, $p < 0.05$) higher than the
270 contribution of daily nitrate uptake rate of small phytoplankton at all the stations in this study. It is well-
271 known for the ammonium preference of small phytoplankton in various regions (Koike et al., 1986;
272 Tremblay et al., 2000, Lee et al., 2008; Lee et al., 2013).

273 In terms of the contributions in different regions, all the contributions (Chlorophyll-a, POC/PON,
274 carbon and nitrogen uptake rates) of small phytoplankton were higher in non-polynya region than in



275 polynya region (Table 1). In addition, the chlorophyll-a contribution of small phytoplankton (mean \pm S.D.
276 = 7.9 ± 3.5 %) was significantly (t-test, $p < 0.01$) lower than the POC contribution (mean \pm S.D. = $36.9 \pm$
277 4.6 %) in polynya region, whereas they were not statistically different in non-polynya region (Table 1).
278 This indicates that small phytoplankton contributed more to the total POC than the chlorophyll-a
279 concentration in the polynya region. We do not have species compositions of phytoplankton in this study,
280 but previous results reported that *Phaeocystis* sp. are dominant in the Amundsen Sea polynya region
281 whereas diatoms are relatively dominant in the non-polynya regions (Lee et al., 2012). Generally,
282 *Phaeocystis* spp. release a large portion (up to 46 %) of extracellular carbon mucilage which makes their
283 colonial form (Matrai et al., 1995). This non-phytoplankton carbon material without chlorophyll-a might
284 cause a higher POC contribution of small phytoplankton in the polynya region during this study. In fact,
285 the contribution of the daily carbon uptake rates of small phytoplankton (14.9 ± 8.4 %) was not as high as
286 the POC contribution (36.9 ± 4.6 %) in the polynya region. The chlorophyll-a contributions of small
287 phytoplankton were lower than that of the daily carbon uptake rate in this study, which is consistent with
288 the results from polynya and marginal ice zone stations in the Ross Sea, Antarctica during austral spring
289 and summer (Saggiomo et al., 1998). They reported that the chlorophyll-a and primary production
290 contributions of pico-phytoplankton ($< 2 \mu\text{m}$) were 29 % and 40 % at polynya stations whereas the
291 contributions were 17 % and 32 % at marginal ice zone stations, respectively. In polynya region, they
292 found much higher contributions in chlorophyll-a and primary production of small phytoplankton than
293 those in this study although their size of small phytoplankton is somewhat smaller than our size ($< 5 \mu\text{m}$).

294 We found a strong negative correlation ($r^2 = 0.790$, $p < 0.05$) between the productivity
295 contributions of small phytoplankton and total daily carbon uptake rates of phytoplankton in the
296 Amundsen Sea (Fig. 7), which implies that daily primary production decreases as small phytoplankton
297 contribution increases. This is mainly because of the relatively lower carbon uptake rate of small
298 phytoplankton than large phytoplankton in the Chukchi Sea, Arctic Ocean reported by Lee et al. (2013).
299 Moline et al. (2004) suggested that further warming air temperatures will increase inputs of glacial
300 melting water and subsequently increase the contributions of small phytoplankton over large



301 phytoplankton community (Moline et al. 2004). If these small phytoplankton were dominant under
302 ongoing more melting conditions of glaciers, a potential increasing contribution of small phytoplankton
303 might cause a subsequent decrease in the total primary production of phytoplankton in the Amundsen Sea
304 based on this study in Figure 7.

305 In respect to food quality of small phytoplankton as a basic food source to herbivores,
306 macromolecular compositions such as proteins, lipids, and carbohydrates as photosynthetic-end products
307 will be needed for better understanding alterations of marine ecosystem in response to ongoing
308 environmental changes (Lee et al., 2013). According to Kang et al. (accepted), small phytoplankton
309 assimilate more food materials and calorific contents per unit of chlorophyll-a concentration and thus
310 provide more contributions in respect to energy aspect than other phytoplankton community in the
311 East/Japan Sea. In conclusion, monitoring the contributions of small-sized phytoplankton to total biomass
312 and primary production of total phytoplankton community could be important as a valuable indicator to
313 sense future changes in marine ecosystem under ongoing various climate-associated environmental
314 changes. Moreover, further detailed studies for macromolecular compositions of small phytoplankton will
315 be necessary for the anticipating small-dominant ecosystem under warming oceans.

316

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395 **Table caption**

396 Table 1. Contributions (%) of chlorophyll-a, POC, PON, and carbon and nitrogen uptake rates) of small

397 phytoplankton in the Amundsen Sea.

398



399 **Figure captions**

400 Fig. 1. Sampling locations in the Amundsen Sea. Red closed circles represent productivity stations. Sea
401 ice concentration data during the cruise period from Nimbus-7 SMMR and DMSP SSM/I-
402 SSMIS Passive Microwave data provided by National Snow & Ice Data Center.

403 Fig. 2. Water column-integrated chlorophyll-a concentration at the productivity stations in the Amundsen
404 Sea.

405 Fig. 3. Water column-integrated concentrations of POC and PON of small and large phytoplankton.

406 Fig. 4. Water column-integrated daily carbon uptake rates of small and large phytoplankton.

407 Fig. 5. Water column-integrated daily nitrate uptake rates of small and large phytoplankton.

408 Fig. 6. Water column-integrated daily ammonium uptake rates of small and large phytoplankton.

409 Fig. 7. Relationship between productivity contributions of small phytoplankton and the total daily carbon
410 uptake rates of phytoplankton (large + small). The total daily carbon uptake rates were
411 transformed into natural logs for a linear regression.

412

413

414



Table 1. Contributions (%) of small phytoplankton in the Amundsen Sea.

	Chlorophyll-a	POC	PON	Daily carbon uptake rate	Daily nitrate uptake rate	Daily ammonium uptake rate	Total nitrogen uptake rate
All stations	19.4 ± 26.0	41.1 ± 10.6	41.3 ± 11.5	26.9 ± 29.3	21.5 ± 11.1	38.7 ± 24.9	27.7 ± 14.4
Non-polynea	42.4 ± 37.2	49.5 ± 14.4	50.0 ± 15.1	50.8 ± 42.8	28.2 ± 15.9	52.8 ± 40.5	36.2 ± 23.0
Polynya	7.9 ± 3.5	36.9 ± 4.6	37.0 ± 6.9	14.9 ± 8.4	18.1 ± 6.8	31.6 ± 10.1	23.5 ± 6.0

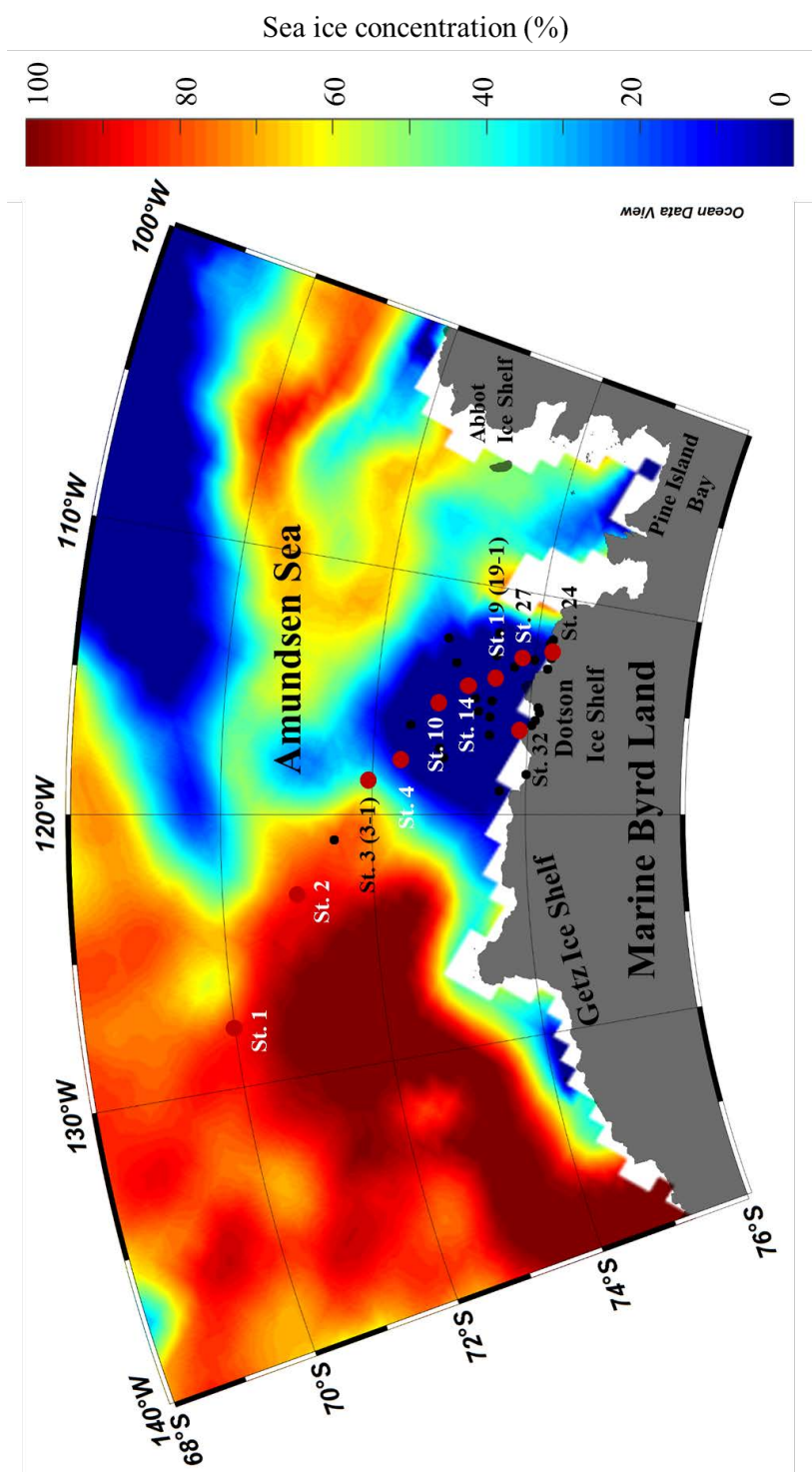


Fig. 1. Sampling locations in the Amundsen Sea. Red closed circles represent productivity stations. Sea ice concentration data during the cruise period from Nimbus-7 SMMR and DMSP SSM/I-SSMIS Passive Microwave data provided by National Snow & Ice Data Center.

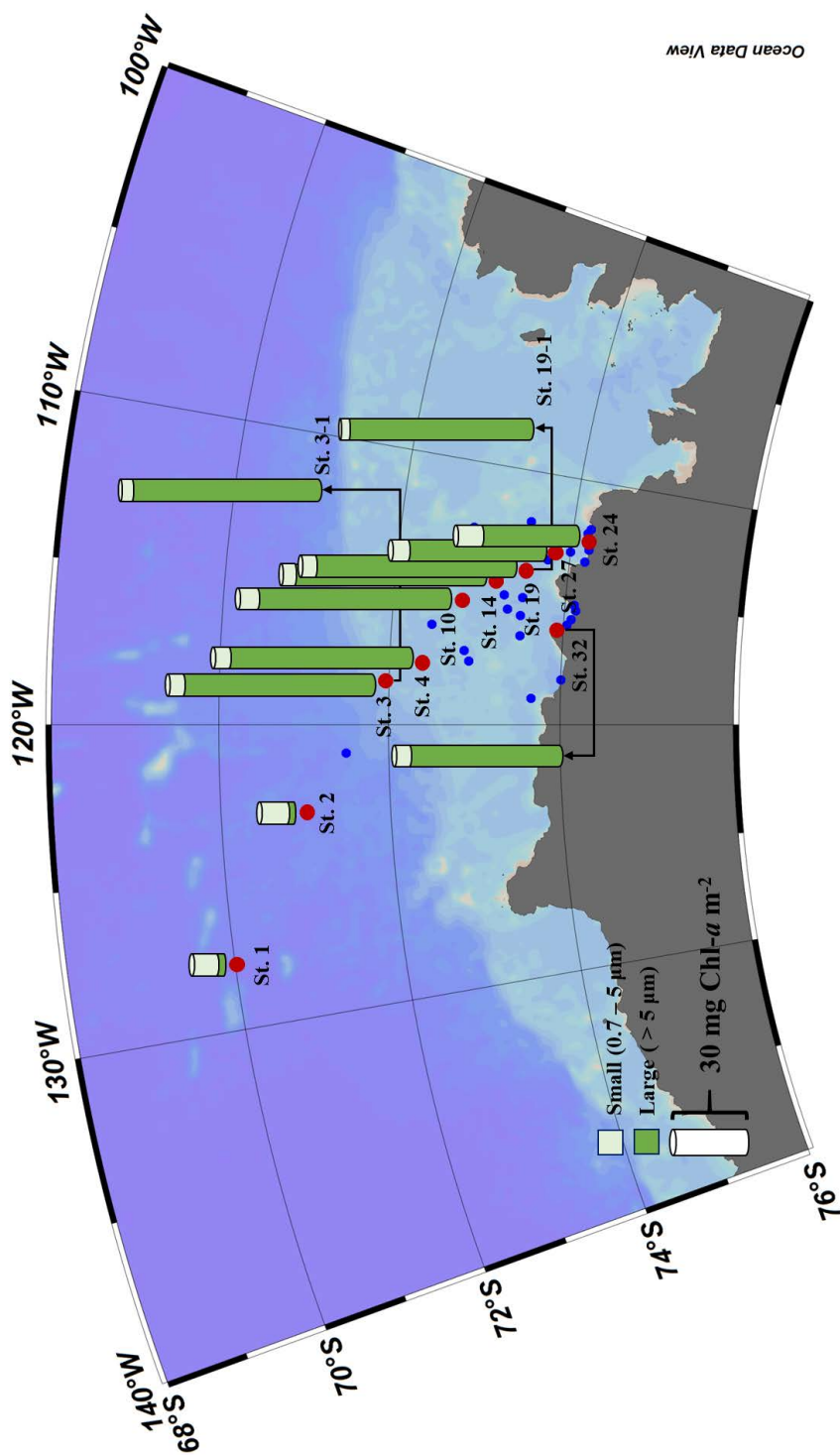


Fig. 2. Water column-integrated chlorophyll-a concentration at the productivity stations in the Amundsen Sea.

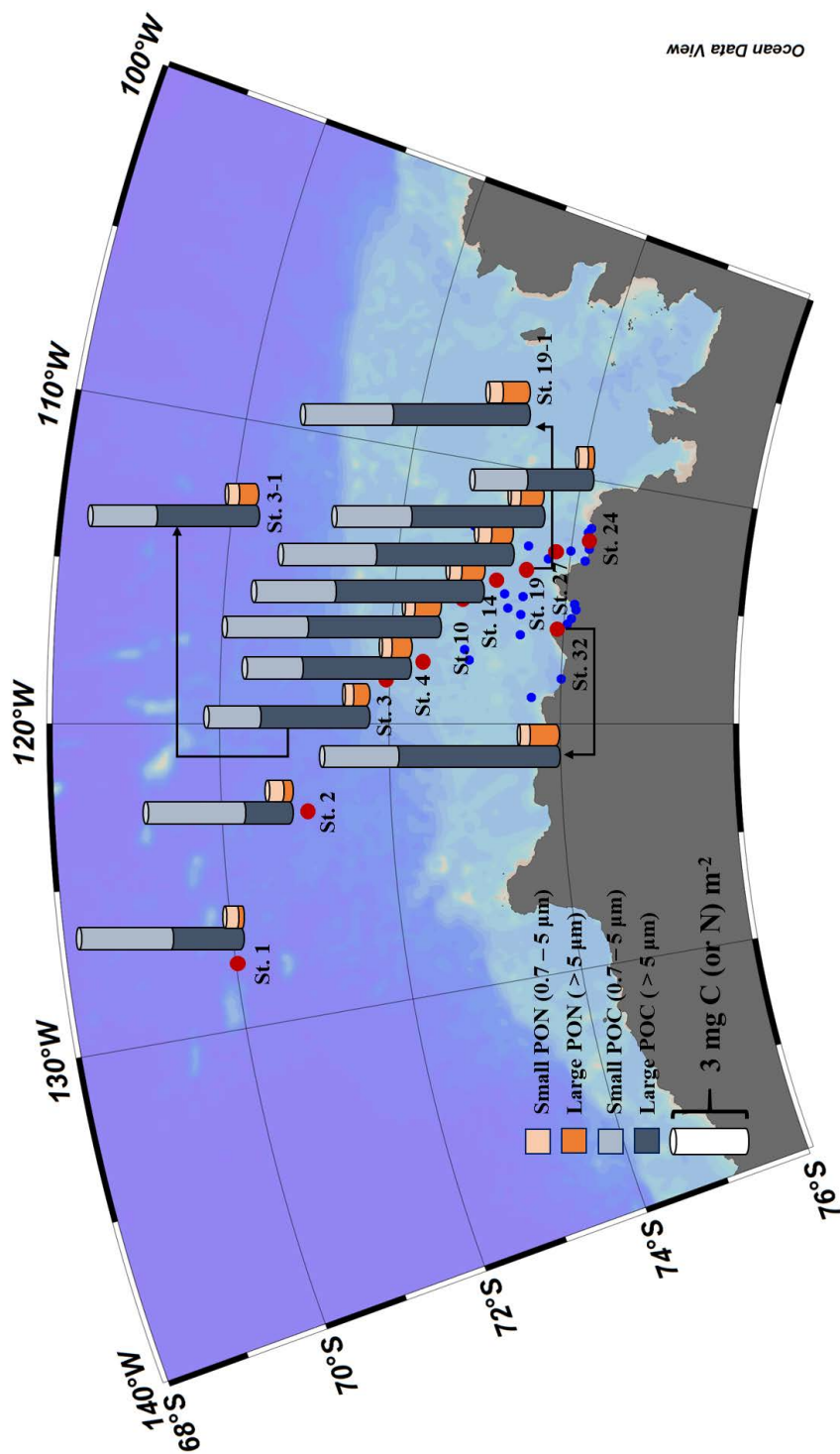


Fig. 3. Water column-integrated concentrations of POC and PON of small and large phytoplankton.

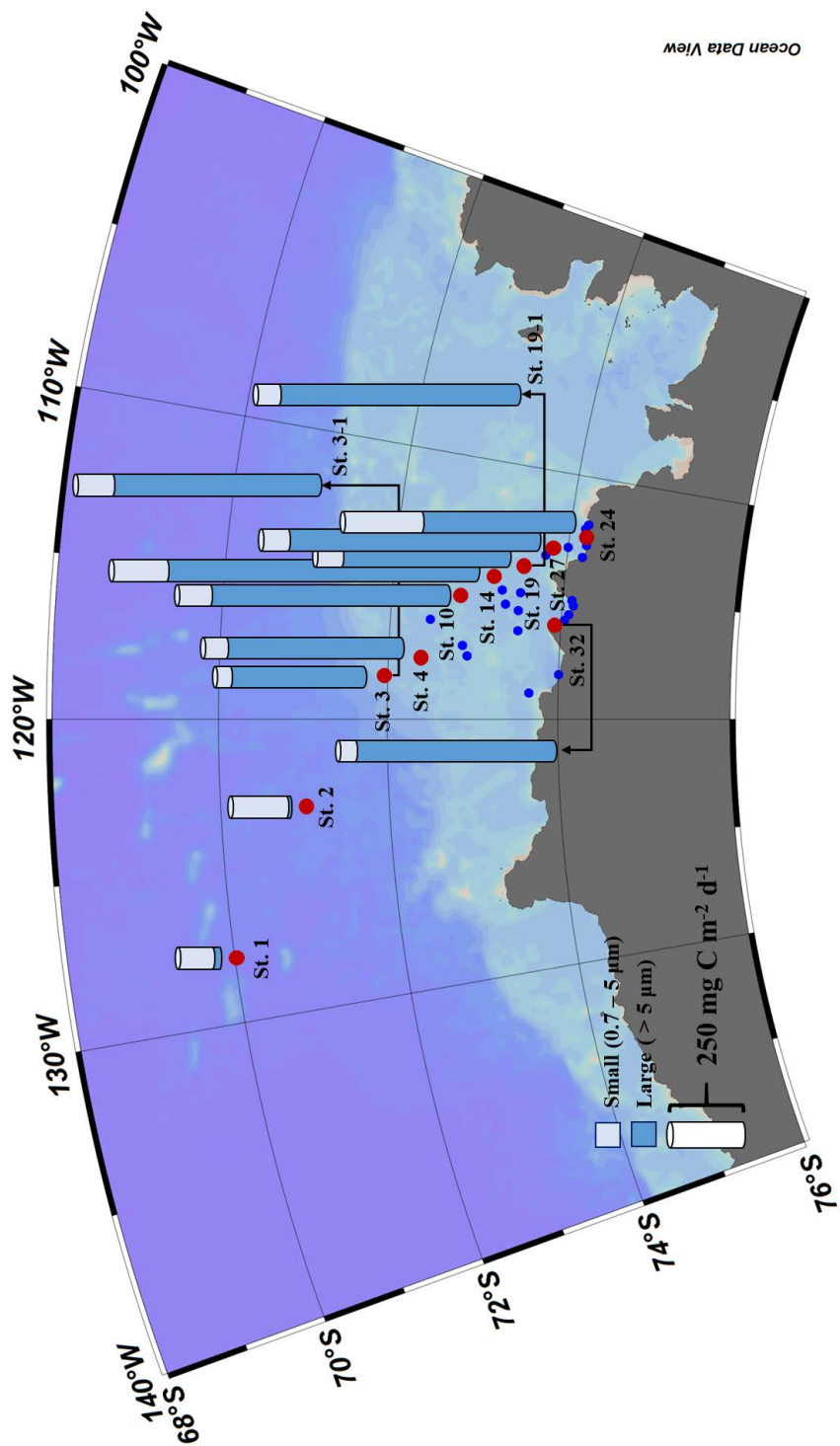


Fig. 4. Water column-integrated daily carbon uptake rates of small and large phytoplankton.

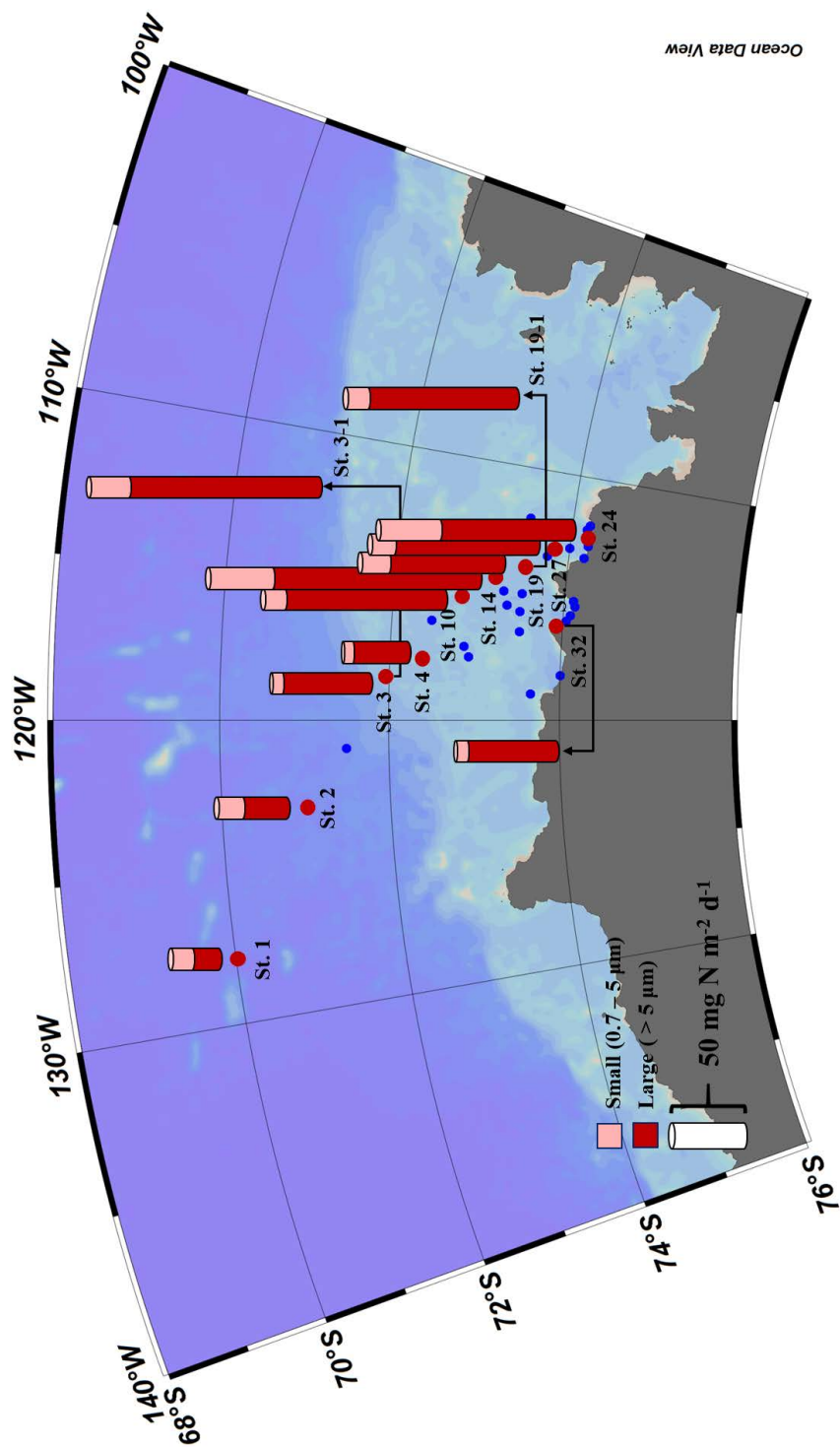


Fig. 5. Water column-integrated daily nitrate uptake rates of small and large phytoplankton.

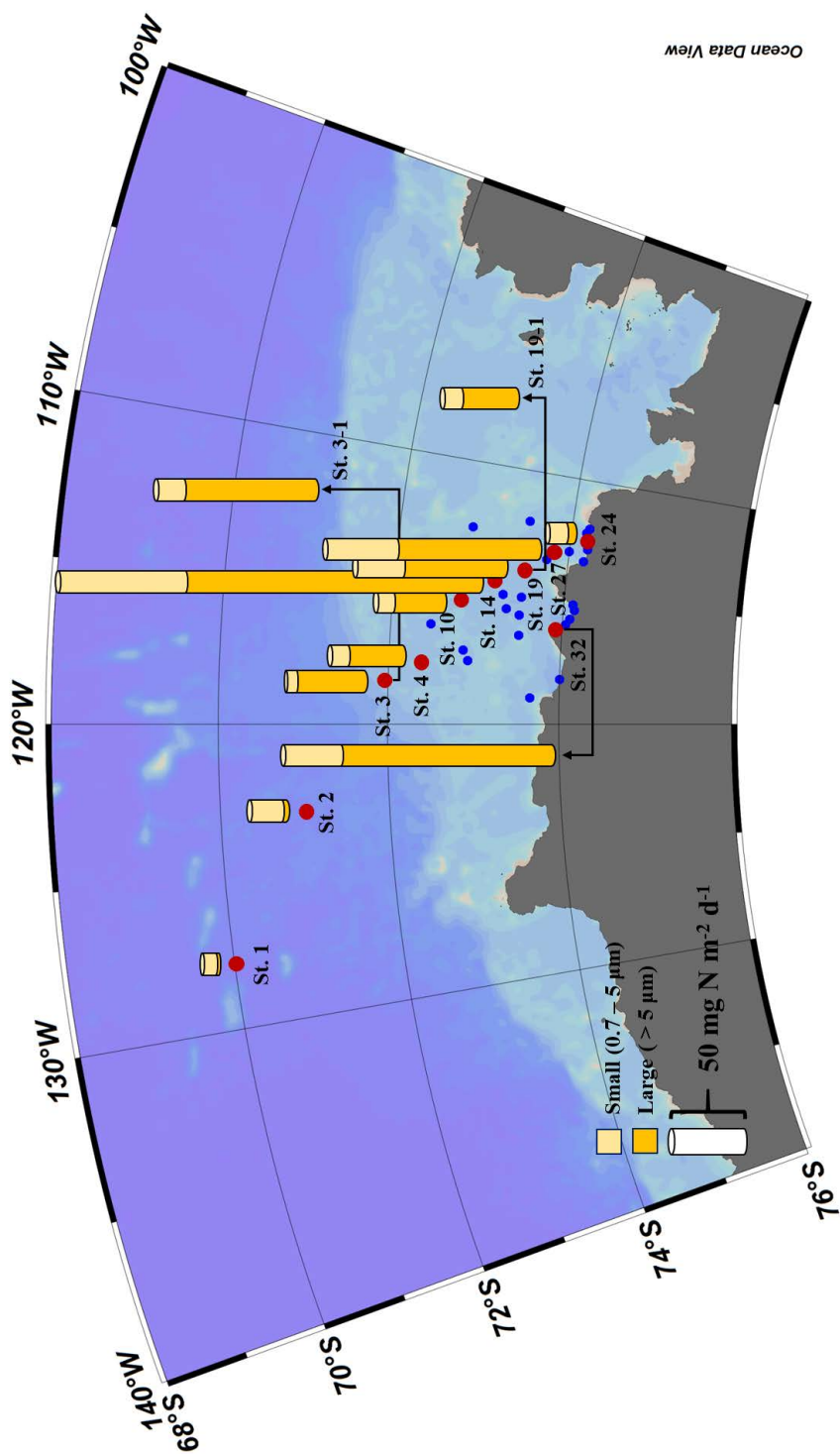


Fig. 6. Water column-integrated daily ammonium uptake rates of small and large phytoplankton.

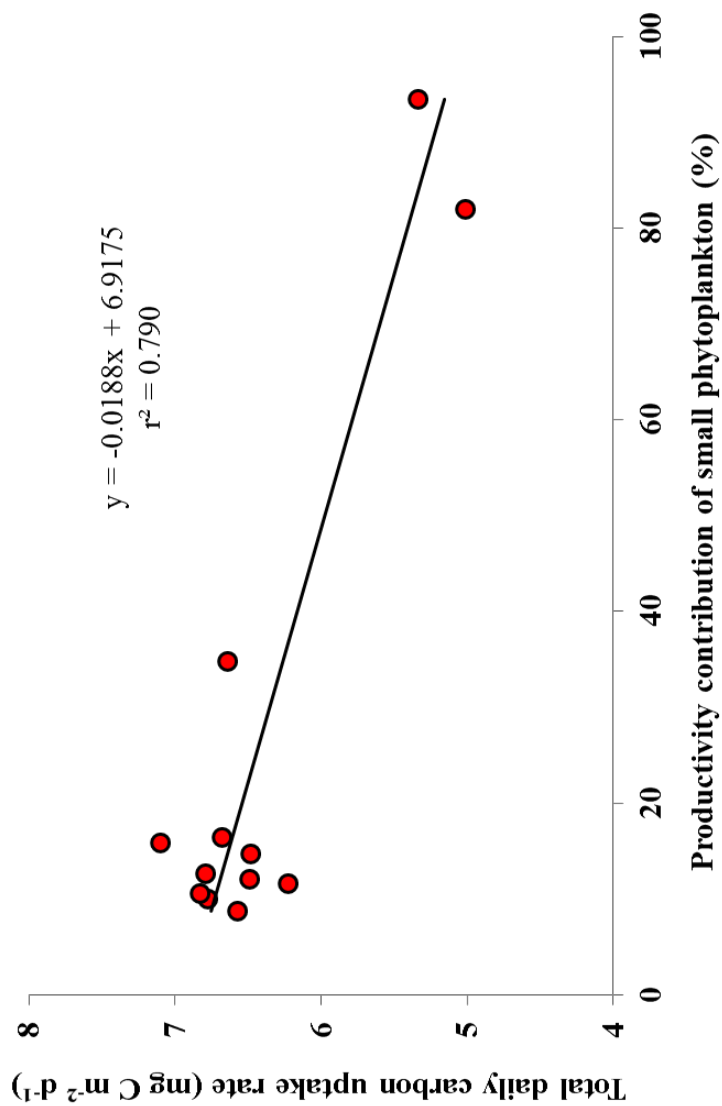


Fig. 7. Relationship between productivity contributions of small phytoplankton and the total daily carbon uptake rates of phytoplankton (large + small). The total daily carbon uptake rates were transformed into natural logs for a linear regression.