Dear Editor,

Thank you for considering the second revision of our manuscript "Phosphorus addition mitigates N_2O and CH_4 emissions in N-saturated subtropical forest, SW China" (bg-2016-470) by Yu et al., for publication in Biogeosciences.

We have revised our manuscript (R2), based on the reviewers' comments. In the point-to-point response, the changes refer to the line numbers in the manuscript version with marked-up changes (attached following the response).

We thank you again for handling our manuscript.

Best regards, Longfei Yu on behalf of all coauthors longfei.yu@nmbu.no

Comments from Reviewer #2

1) Scientific Significance:

The primary objective of this study is to determine how phosphorus availability influences the efflux of N2O and CH4 in a nitrogen-saturated pine-dominated subtropical forest. While we have a fair understanding how the availability of nitrogen influences these processes, we have a very poor understanding how phosphorus can directly, or indirectly influence greenhouse gas production. Prior research on this subject has been inconclusive and this study will help add to the body of literature to advance our understanding of soil nutrient availability and climate forcing.

2) Scientific Quality:

Much improve from the early versions – mixed model is the way to go. My primary concern is the low replication (just three 20 x 20 m plots for each treatment) and perhaps overall discussion needs to be tempered.

R: We thank the reviewer for the critical but constructive comment. Our study explores the effect of phosphorus (P) on N cycling in the N-saturated Tieshanping (TSP) forest (Huang et al., 2015). Indeed, our field observations indicate that P application significantly reduces N_2O and CH_4 emissions from upland soils at TSP. We agree that the current field set-up has its limitation and that our findings cannot be readily scaled up. For this, further research is needed, including work at other sites. The experiment was done in triplicate, as more replicates were not feasible given the available man-power and budget. In terms of replication, our experiment, investigating GHG emissions, does not differ much from others, which also used 20 x 20 m² plots in triplicates (Muller et al., 2015; Martinson et al., 2013) or even 10 x 10 m² plots in triplicates (Zheng et al., 2016; Wang et al., 2014) for each treatment.

3) Presentation Quality:

The discussion needs minor revisions. A fair amount of the discussion is just a summary of the results with modest interpretation or explanation. While it is fine to report how these results are similar/dissimilar with other work, it needs to go beyond that and provide new insights and tieback with the present study. What do the result consistent with this study have in common, but uncommon with work that is inconsistent? More effort is needed in the discussion to explain the 'why' of the results. Certain interesting passages need to be developed (e.g. lines 313-315)

R: We thank the reviewer for the general suggestion. In the revised version, we now expand our explanation of the results. Major changes are made in three sections of the discussion:

1) P effect on soil NO₃⁻ concentration (line 301-310): we attribute the attenuation of soil NO₃⁻ concentration to P stimulation of N uptake by plants and soil microorganisms. Based on the reported increase of N mineralization and nitrification rates in a similar site from South China, we suggest that microbial uptake of N is less likely than the plant uptake. Although this is not supported by our measurement of tree biomass and foliar N during two years, we provide an alternative explanation, which is N uptake by understory vegetation.

2) Comparison of P effect on N_2O emission with other studies (line 324-335): In comparison with TSP, the slow response of N_2O emission to P addition at the Ecuadorian site is suggested to be a result of both low ambient N deposition and low P dose. The Chinese site (Dinghushan) receives similar inorganic N input via throughfall as TSP. But we suggest the Dinghushan site to be less N-rich than TSP, based on the lower KCl-extractable inorganic N content in soil and lower flux of N leaching at Dinghushan. This point is now clearly made in the discussion.

3) We improve our explanation of N inhibition effects on CH_4 uptake by elaborating on known CH_4/NH_4^+ substrate competition for methane monooxygenase (line 372-373) and NO_3^-/NO_2^- toxicity for methanotrophs (line 377-382). In the discussion of P effect on CH_4 uptake, we focus mainly on the direct

P effects on methantrophs and indirect P effects by reducing inorganic N levels in soils (line 386-398). The indirect mechanism seems more likely, as support by our observation.

Line 363: I question that the study actual demonstrates that high deposition has shifted these soils to regional hotspots for N2O and CH4 production – this was not tested. Also, this is difficult to say without historical data or somehow experimental removing/reducing N deposition. Yes, adding P has reduced NO3 concentration, but this result is fairly modest, especially at 20 cm.

R: We agree with the referee that historic data on the relationship between atmogenic N deposition and N concentrations in soil water as well as emission rates of N_2O and CH_4 would be needed, to ultimately prove the connection between elevated N deposition and N_2O and CH_4 emissions in the sub-tropical forest region. As such data do not exist for TSP, our conclusions are solely based on comparison with similar research sites and on manipulation experiments at TSP. For example, Huang et al (2015) showed that a doubling of N deposition at TSP caused a doubling of N leaching (as nitrate), simultaneously in increasing N_2O emission (Liu WJ, personal communication). Our manuscript adds further evidence to this, as we find that P addition causes a significant decline in NO_3^- concentrations in soil water in the surface horizons (O/A and AB; Fig. 1), while emission rates of N_2O and CH_4 decrease (Figs. 4 and 6). Also other studies have reported strong correlation between NO_3^- concentration in soil water and N_2O emission rates (Gundersen et al., 2012).

Earlier studies at the TSP site indicate chronically elevated N deposition levels, high inorganic N concentration in soil water and strong soil acidification (Zhu et al., 2013; Larssen et al., 2011; Huang et al., 2015). These factors contribute to enhanced N_2O emission and reduced CH_4 uptake in soils (Liu et al., 2010; Le Mer and Roger, 2010). Among other studies in Southern Chinese forests (Tang et al., 2006, Fang et al., 2009; Zhang et al., 2008), our study reports the highest N_2O fluxes and lowest CH_4 uptake rates (or even net emission).

In addition, our recent ¹⁵N tracer study at TSP (Yu et al., 2017), has shown that the enriched ¹⁵N signals in emitted N_2O is identical to those in soil NO_3^- extracted from O/A horizon rather than AB horizon. This denotes the importance of NO_3^- availability in surface soils to N_2O emission. Also for the CH₄ oxidation, the N inhibition effect should be most important in surface soil (Bodelier and Laanbroek, 2004), where methanotrophic activity is more active with more aerobic condition. Therefore, our observation of significant decreases in NO_3^- concentration from O/A and AB horizons do support that P addition migrates the N_2O and CH₄ emissions from soil.

Comments from Reviewer #3

The manuscript by Longfei Yu and others presents a replicated forest fertilization experiment in an acidified and N-saturated Masson pine-dominated forest at TieShanPing, SW China. The experiment tests the role of mineral P fertilization in regulating nitrous oxide and methane emissions (uptake). Researchers measured soil water NO3- concentrations, N2O emissions, CH4 emissions (uptake), forest productivity, litter fall, litter chemistry, and soil characteristics prior to and after a one-time fertilization with 79 kg P ha-1 (NaH2PO4).

The Author's found that P fertilization results in declines in soil NO3- and suppressed emissions of N2O, and CH4 (not immediately, but over the long term). With P addition TSP soils switched from a CH4 source to a sink. Elevated biomass production was not observed over the 18-month experimental period. However, understory biomass was not assessed. Based on these results, Authors hypothesized that P additions resulted in increase NO3- uptake by plants and microbes leaving less for denitrification. Also, P addition was thought to lessen the NH4+ inhibition of methane oxidation.

Overall comments:

This manuscript is well written and presents a topic that is of interest generally to the readers of Biogeosciences. I have a couple of concerns that should be addressed prior to publication. First, I feel that these results and their interpretation would be easier to follow if there were a set of explicitly stated hypotheses. There is one hypothesis stated in the Abstract (that concerns the results), but not in the main body of the manuscript.

R: Following the reviewer's recommendation, we have changed our objectives to more explicit hypotheses. We now present three key hypotheses: 1) P addition stimulates tree growth; 2) P addition decreases soil inorganic N availability and 3) P addition reduces N₂O and CH₄ emissions (line 105-109).

Regarding the description of the experimental design and sampling, the description in the methods doesn't seem to reflect the data that is presented for N2O and CH4 (L167-179). From the methods, I gather that N2O and CH4 were measured a total of four times, but clearly more data points are presented. Please clarify in the text how frequently measurements were made over the entire experiment. Other clarifying points are made below in the line-by-line comments.

R: Thanks for the suggestion. To avoid confusion, we now describe the frequency of gas emission sampling together with the soil pore water sampling. In line 178-180, we add "The measurements were conducted bi-monthly in the dry and dormant season and monthly during the growing season, simultaneously with the sampling of soil pore water". In line 181, we put "also" before the description of "short-term" intensive sampling.

With the Results, at times the text is confusing because the treatment effects of P additions and the seasonal/temporal patterns are explained simultaneously. I would recommend some minor reorganizing of this information. Perhaps start with the overall seasonal patterns and then state the treatment effects or the opposite.

R: We appreciate reviewer #3's advice. We have now reorganized the results of N_2O and CH_4 fluxes, by starting with a paragraph for seasonal/temporal patterns of fluxes from Reference plots (line 243-255). The P effect on N_2O and CH_4 fluxes are then presented in another two paragraphs.

Line-by-line comments: L16: Change GHG to green house gas R: OK.

L18: If this is a single fertilizer event is it necessary to have the unit yr-1? R: We have revised it as "Here, we report N_2O and CH_4 emissions together with soil N and P data for a period of 18 months following a single P addition (79 kg P ha⁻¹, as NaH_2PO_4 powder)" (line 17-19).

L20: Rephrase this sentence to read "We observed a significant decline in soil water NO3- concentrations (5 and 20 cm depths) and in soil N2O emissions following P addition."

L21: It is unclear if this number is the amount of reduction or if it represents the total emission. Please clarify

R: Agree. This number for emission reduction is now deleted. See line 20-22.

L23-24: The "As for N2O" is a confusing way to begin this sentence. Can you revise to something like "P addition significantly decreased CH4 emissions, turning TSP soils from a net source to a net sink." I'm sure the Authors will have a more eloquent way of conveying that message.

R: It is now changed to "P addition significantly decreased CH_4 emissions and turned the soil from a net source into a net sink" (line 24-26)

L26-27: It's my preference to put this caveat in the discussion or that it's rephrased. The current wording suggests that you measured understory and that there was an increase in understory biomass. R: This sentence is now rephrased, as "Within the 1.5 years after P addition, no significant increase of forest growth was observed and P stimulation of forest N uptake by understory vegetation remains to be confirmed" (line 27-30)

L48-49: 'frequently shifting aerobic conditions' is awkward please revise. Perhaps this is better put in terms of aerobic and anaerobic? R: OK (line 50-53).

P4 L56: Consider changing 'mineral' to 'inorganic' R: Done.

L100: The hypothesis is stated in the abstract but not the main text. Please include in text prior to the objectives.

R: Please refer to our earlier reply to the overall comments. See also line 105-109.

L105: It's unclear why the study site name is in quotations. L116: TSP hilltop is not intuitive. Please explain in text

R: The quotation is removed now. "TSP hilltop" refers to upland soils from the hillslopes at TSP, which has been previously documented by Zhu et al., 2013b. Here, we rephrase "at TSP hilltop" as "on the hillslopes" (line 123-124).

L121-122: Can you state over what time period the decline in growth has occurred here?

R: Wang et al. (2007) and Li et al. (2014) reported the decline in forest growth at TSP during 2001-2004 and 2004-2012, respectively. So we rephrase the sentence as "Strong soil acidification has been reported to cause severe decline in forest growth at TSP since 2001 (Li et al., 2014; Wang et al., 2007)" (line 129-131).

L128: Rather than an *, please use \times R: OK. Change is made in line 136.

L141: Can you report the Na+ concentrations of the Reference plots? L157: Change (2 mm) to (2mm \times 2mm)

R: We add " $(0.52-1.31 \text{ mg L}^{-1})$ " for the reference plots (line 149). More specific values for different soil depths could be found in Table S2 of the support information.

L165: I think part of the instrument name is missing. Should this be 'inductively coupled plasma atomic emission spectroscopy'? For all makes/models of equipment here and throughout, please add the location information.

R: OK. See line 174-175.

L171: Change to "...into 12 mL pre-evacuated glass vials... (Chromacol, UK)." L172: I would recommend splitting this into another sentence: "Vials were over pressurized to avoid contamination during sample transport."

R: It is modified as "20 ml gas samples were injected into pre-evacuated glass vials (12 ml) crimp-sealed with butyl septa (Chromacol, UK), maintaining overpressure to avoid contamination during shipment", as shown in line 182-184.

L173: Is 'Mixing ratio' what you mean or should this be 'Fluxes of ...' or 'Concentrations of...' R: Yes. Mixing ratio is the direct result that we obtain from gas analysis on gas chromatograph coupled with ECD. Mixing ratios of N_2O and CH_4 are commonly expressed as ppb or ppm, similar to "concentration". For fluxes, we need further calculation based on the change of gas mixing ratio with time, as described later in the same paragraph.

L190: Please specify if the same trees were measured at each time point, this is critical to the interpretation of these data.

R: Yes, they had been marked since the first measurement. This is now explained (line 204).

L194: Rather than 'sum of precipitation' can this be termed 'daily total precipitation'? Please provide the time period over which precipitation and temperature were measured. R: OK (line 207-208).

L198: I gather from the methods that gas samples were collected from August 2013 forward, but only during the month of May (2, 7, 10, and 12). This doesn't reflect all of the data points that are shown in Figures 2 and 5. I would insist that the Authors add clarity to the methods or only show data that were collected in this study.

R: Please refer to the earlier reply to the overall comments of the reviewer. Changes are made accordingly in line 178-180.

L206: It is unclear if fluxes of 'litterfall' nutrients were scaled to the biomass production. Or was litter biomass a component of the overall biomass calculation? L210-213: For tree growth, how were the 3 different time points treated? Please be specific.

R: The fluxes of litterfall were not used for tree biomass evaluation. Instead, we used allometric models (specifically for masson pine; Li et al. 2011 and Zeng et al. 2008) to estimate the tree biomass, based on measured diameters at breast height (DBH). We specify now that DBH is used for tree biomass estimation (line 205).

The data for tree growth refer to Table S3, which includes tree biomass, 500-needle weight and needle nutrient contents. The three samplings were tested for treatment effect separately with one-way ANOVA (line 223-226).

L226: The phrase 'sum of charge of dissolved base cations is unclear', at any rate, it would be more appropriate to say that charge was significantly different between fertilized and unfertilized. I am curious if the 'charge' decreases in the P treatment because of the increase of Na+. Can you please address? R: We thank the reviewer for the suggestion. This phrase is changed to "the overall cationic charge" (line 240-241). The increase in total cationic charge due to Na⁺ addition should be modest, while the charge of dissolved of Ca^{2+} is far more important. P addition stimulated the uptake of N, thus resulting in a decline in soil NO₃⁻ concentration. As NO₃⁻ is the major anion in soil water, the decrease of NO₃⁻ concentration leads to a decline in cation concentration (Figure S3). Thus, the observed decrease in cationic charge is a direct effect of the decline in mobile anions (line 409-411).

L232-234: Please report the block effect here. L240: Was there a significant block effect that could be reported here?

R: The block effects were clearly described in the sentences that follow, in line 247-249 for N_2O and in line 250-254 for CH_4 .

L236-238: Rephrase to read: The P addition resulted in a 50% (average 3 kg N ha-1 yr-1) reduction of cumulative N2O emissions (Fig. 3). Please add +/- Stderror if it is available R: We have revised it to "The P addition resulted in a 50% (3 kg N ha⁻¹ yr⁻¹ on average) reduction of cumulative N₂O emission (Fig. 3)" (line 257-259).

L238: Change was to were. R: "effects" is changed to "effect" (line 259).

L245: Should this unit be CH4-C here and throughout? Also can you add +/- Stderror here? R: The units for N₂O and CH₄ fluxes have been checked throughout the manuscript and corrected as g N₂O-N m⁻² hr⁻¹ and g CH₄-C m⁻² hr⁻¹, respectively. For annual emission, they are described as kg N₂O-N ha⁻¹ yr⁻¹ and kg CH₄-C ha⁻¹ yr⁻¹. Here, the CH₄ flux data show a skewed distribution, thus having large standard deviations. Since the treatment effect on CH₄ fluxes had been tested for significance with mixed-effect models, it is not necessary to include standard error here. L250: What does 138 t ha-1 represent? Is it an average across both years and both treatments? I'm not sure how informative that is. Based on your supplemental data, it looks as if biomass was actually lower in the P addition treatment compared to the Reference treatment.

R: Agree. This value denotes the mean for both years and in both treatments. Based on statistical results, P treatment exhibited no difference from the reference. In the text, we have now described the change as "insignificant" and deleted the value for tree biomass (line 271-272).

L252: The 500g needle weight does not need to be reported here. R: Agree.

L253: This sentence needs to be clarified to indicate the mechanism responsible for differences in needle chemical composition. "Linked" is vague.

R: It is rephrased as "Between the two samplings in 2013 and 2014, we found differences in chemical composition of the pine needles, but the difference between the Reference and P treatment was not significant" (line 273-275).

L253: 'hardly' is a vague word, please replace. L273: Change mineral to inorganic R: OK. "hardly" is changed to "not" now (line 276).

L275-279: This sentence is complex and confusing. Please revise, as it seems to contradict your former statement.

R: OK. We have simplified our discussion, with additional details for support. "These findings are consistent with a number of previous studies (Baral et al., 2014; Hall and Matson, 1999; Mori et al., 2014), which attributed the reduction of N_2O emissions in P-treated soils to decreased NO_3^- availability and thus less denitrification. The attenuation of soil NO_3^- by P addition at TSP may reflect stimulated N uptake by plants and/or soil microorganisms." (299-303).

L273: This paragraph is long and difficult to follow. I believe that the Authors could find a way to make it more streamlined and easier to follow. R: See the next comment.

L283-292: I think it would be better to put your study into context of others that used similar additions. Perhaps the reference to moderate P additions is a bit of a distraction. I would recommend revising to focus on more similar studies.

R: This paragraph has now been divided into two paragraphs. The first one has been rephrased and focuses only on the key points of our experiment (296-310). The second one compares the fertilization effects of N, P and N+P on N₂O emissions among forests with different N status, involving a space-for-time comparison. While our main discussion still addresses the comparison to similar studies in Southern China, we have modified the comparison with the Ecuadorian study, and focus on different responses of N₂O emissions to P addition in forests with different N status (315-345). See more details in our response to comments from Reviewer #2.

L306: Likewise, the point of this paragraph is not entirely clear. As well, it is unclear if the referenced studies are also covering the short-term (~10day) span of time that is referenced in this manuscript.

R: To make it more concise, we have rewritten this paragraph, starting with studies that found that P addition increases N_2O emission. Then we discuss the possible mechanisms for P stimulation of N_2O emissions, and emphasize that the P effect on denitrifier activity could be rather fast (Mori et al., 2013c). However, we did not see any P effect on N_2O emission from the intensive observation shortly after P application at TSP. This may be attributed to large denitrification potentials in TSP soils, supported by a lab-incubation study (Zhu et al., 2013c). See the changes in line 346-359.

L324: Change production to 'CH4 production' just to be clear this isn't primary production R: OK. See line 370.

L353: Change apparently to 'may have' R: OK. See line 407.

L355: I liked the nice flow and organization of this paragraph! R: Much appreciated.

L375-376: Is there a citation from your previous work that you can add here? As is, these data don't provide obvious evidence for this.

R: We have added Zhu et al. (2013b) as a reference (line 429). For more details, see our reply to reviewer#2's comments.

L377: Can this statement be qualified by stating 'to overall reduce' R: Yes, it can. See the changes made in line 430.

L379: GHG is used here and in the abstract, but is not explicitly defined. Please do so. L388: References are not alphabetized consistently.

R: For GHG, please refer to our previous response to the comment on line 16. References have been checked and corrected as requested.

Tables and Figures: L603: Please change 'Background' to 'Ambient' R: OK. See line 675.

Table 1. Was 5.0 mg kg-1 the detection limit of the instrument for PH2O? If so, please just indicate this in the footer of the table rather than dedicating an entire column to the < 5.0 information. R: OK. This is now changed (line 675-681).

Table 2. Here and throughout, please be consistent that the P treatment is '+P'. Also it is unclear what the letters indicate in terms of significance. Should they not indicate significant differences among the Ref and +P treatment? Or is this across all time points? If so, the analysis should more appropriately be a repeated measures analysis.

R: The different letters indicate significant difference between the Reference and P treatment (line 682-685). This is tested by one-way ANOVA for each sampling, which has been specified in statistics (line 223-226).

Figure 4 and Figure 6: The letters indicating significance are somewhat unnecessary here. The point could be made in either the figure legend or with an asterisk centered above the two boxes. In both figures, I would recommend adding the statistical test that you used.

R: We thank the reviewer's suggestion. We would prefer keeping both the letters indicating significance and adding statistical methods to the figure captions. See line 699 and line 707.

Figure S6: Litter is spelled incorrectly in the axis title R: OK. Revised.

Phosphorus addition mitigates N₂O and CH₄ emissions in N saturated subtropical forest, SW China

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11 Abstract

Chronically elevated nitrogen (N) deposition has led to severe nutrient imbalance in forest soils. 12 Particularly in tropical and subtropical forest ecosystems, increasing N loading has aggravated 13 phosphorus (P) limitation of biomass production, and has resulted in elevated emissions of 14 nitrous oxide (N₂O) and reduced uptake of methane (CH₄), both of which are important 15 16 greenhouse gases. Yet, the interactions of N and P and their effects on GHG-greenhouse gas emissions remain understudiedelusive. Here, we report N₂O and CH₄ emissions together with 17 soil N and P data for a period of 18 months following a single P addition (79 kg P ha⁻¹- yr^{-1} , 18 applied as NaH₂PO₄ powder) to an N-saturated, Masson pine-dominated forest soil at 19 20 TieShanPing (TSP), Chongqing, SW China. We observed a significant decline in both in NO₃⁻ concentrations in soil water (at 5- and 20-cm depths) and in soil N_2O emissions, following P 21 <u>application</u>, the latter by 3 kg N ha⁻¹ yr⁻¹. We hypothesize that enhanced N uptake by plants and 22 23 soil microbes in response to P addition, results-resulted in less available NO₃⁻ for denitrification. By contrast to most other forest ecosystems, TSP is a net source of CH₄. As for N₂O, P addition 24 significantly decreased CH₄ emissions, <u>and turning turned</u> the soil from a net source into a net 25 sink- Based on our observation and previous studies in South America and China, we believe 26 that P addition relieves N-inhibition of CH₄ oxidation. Within the 1.5 years after P addition, no 27 28 significant increase of forest growth was observed at TSP, but and P stimulation of forest N uptake by understory vegetation remains to be confirmedwe cannot exclude that understory 29 vegetation increased. Our study indicates that P fertilization of N-saturated, subtropical forest 30 soils may mitigates N₂O and CH₄ emissions, in addition to alleviating nutrient imbalances and 31 32 reducing losses of N through NO_3^- leaching.

Key Word: N₂O and CH₄ emission, N saturation, Phosphate fertilization, soil CH₄ uptake, acid
forest soil.

35 **1 Introduction**

Anthropogenic activities have transformed the terrestrial biosphere into a net source of CH₄, N₂O 36 and CO₂, leading to increased radiative forcing (Montzka et al., 2011; Tian et al., 2016). During 37 the last decade, atmospheric concentrations of CO₂, CH₄, N₂O have increased at rates of 1.9 ppm 38 yr⁻¹, 4.8 and 0.8 ppb yr⁻¹, respectively (Hartmann et al., 2013). In China, the exponential increase 39 of reactive nitrogen (N) input into the biosphere since the 1970s has likely led to more carbon (C) 40 being sequestered in the biosphere (Cui et al., 2013; Shi et al., 2015). However, enhanced 41 emissions of N₂O and CH₄ due to chronic N pollution potentially offset the cooling effect by C 42 sequestration (Liu and Greaver, 2009; Tian et al., 2011). 43

Microbial nitrification and denitrification in soils account for about 60% of N₂O emissions 44 45 globally (Ciais et al., 2013; Hu et al., 2015). Although, microbial activity is often restricted in low pH soils of unproductive forests, surprisingly large N₂O emissions have been reported from 46 acid, upland forest soils in South China (Zhu et al., 2013b). Reported average N₂O fluxes in 47 humid, subtropical forests range from 2.0 to 5.4 kg N₂O-N ha⁻¹ yr⁻¹ (Fang et al., 2009; Tang et al., 48 2006; Zhu et al., 2013b), which by far exceeds global averages for temperate or tropical forest 49 ecosystems (Werner et al., 2007; Zhuang et al., 2012). This has been attributed to frequently 50 51 shifting shifts between aerobic and anaerobic conditions in soils aeration conditions during monsoonal summers, promoting both-alternating nitrification and denitrification (Zhu et al., 52 2013b) and to-causing large soil NO₃⁻ concentrations due to efficient cycling of deposited N in 53 acid subtropical soils (Yu et al., 2016). 54

55 Chronically elevated rates of N deposition (30-65 kg<u>N</u> ha⁻¹ yr⁻¹; Xu et al., 2015) have resulted in 56 strong nutrient imbalances in southern Chinese forests, aggravating phosphorus (P) limitation

(Du et al., 2016). Phosphorous deficiency in N-saturated forests restricts forest growth and thus 57 constrains-limits its capability to retain N (Huang et al., 2015; Li et al., 2016), resulting in ample 58 amounts of mineral-inorganic N (NH₄⁺ and NO₃⁻) being present in the soil solution. Accordingly, 59 Hall & Matson (1999) observed larger N₂O emission in P-limited than in N-limited tropical 60 forests after one year of after repeated N-addition of N. Likewise, previous N manipulation 61 studies in forests of South China reported pronounced stimulation of N₂O emissions by N 62 addition (Chen et al., 2016; Wang et al., 2014; Zheng et al., 2016), supporting the idea that P 63 limitation causes forests to be more susceptible to N saturation and N2O-N loss. In an N-limited 64 tropical montane forest in southern Ecuador, P addition alone (10 kg P ha⁻¹ yr⁻¹) had no effect on 65 N₂O emissions during the first two years. However, N₂O emission was smaller when P was 66 added together with N (50 kg N ha⁻¹ yr⁻¹) than in treatments with N addition alone (Martinson et 67 al., 2013). After continued fertilization for three years, also P addition alone reduced N₂O 68 emissions at these sites (Müller et al., 2015). In tropical China, with high N deposition (~ 36 kg 69 <u>N</u> ha⁻¹ yr⁻¹; Mo et al., 2008), P addition (150 kg P ha⁻¹ yr⁻¹) to an old-growth forest revealed a 70 similar pattern, with no initial effect on N₂O emissions (0-2 years) but a significant longer term 71 effect (3 to 5 years) on N₂O emissions (Chen et al., 2016; Zheng et al., 2016). In a secondary 72 tropical forests in South China, Wang et al. (2014) found no effect on N₂O emissions of P alone 73 (100 kg P ha⁻¹ yr⁻¹), and in treatments combining P with N (100 kg N ha⁻¹ yr⁻¹), N₂O emissions 74 increased during the wet season. Meanwhile, they observed a significant increase in soil 75 microbial biomass after P addition, which is in line with previous findings in tropical forest soils 76 of South China (Liu et al., 2012). Thus, they attributed the stimulating effect of P addition on 77 N₂O emissions to the larger nitrification and denitrification potential of the increased soil 78

microbial biomass. This was also proposed by Mori et al. (2014), based on results from a shortterm incubation study with P addition, excluding plant roots.

As the sole biogenic sink for CH₄, upland soils play an important role in balancing terrestrial 81 CH₄ emissions (Ciais et al., 2013; Dutaur and Verchot, 2007). Atmospheric CH₄ uptake in soil is 82 mediated by the activity of methanotrophic bacteria, which oxidize CH₄ to CO₂ to gain energy 83 for growth. Well-drained forest and grassland soils are dominated by yet uncultured, high-84 affinity methanotrophs residing in the upper soil layers (Le Mer and Roger, 2010). In addition to 85 edaphic factors (pH and nutrients), parameters affecting the diffusion of CH₄ into the soil (soil 86 structure, moisture, temperature) are believed to be the major controllers for CH₄ uptake (Smith 87 et al., 2003). A number of studies have shown that excess N affects CH₄ fluxes in forest soils 88 (Liu and Greaver, 2009; Veldkamp et al., 2013; Zhang et al., 2008b). In general, N addition 89 90 promotes CH₄ uptake in N-limited soils by enhancing growth and activity of methanotrophs, 91 whereas excessive N input and N saturation inhibit CH₄ oxidation on an enzymatic level by substrate competition between CH4 and NH4⁺ (Aronson and Helliker, 2010; Bodelier and 92 Laanbroek, 2004). P addition experiments in N-enriched soils have shown positive effects on 93 CH₄ uptake (Mori et al., 2013a; Zhang et al., 2011), but the underlying mechanisms, i.e. whether 94 P addition affects the methanotrophic community in soils directly or alleviates the N-inhibition 95 96 effect on CH₄ oxidation through enhanced N uptake (Mori et al., 2013b; Veraart et al., 2015), 97 remain unresolved.

Subtropical forests in South China show strong signs of N saturation, with exceedingly high
NO₃⁻ concentrations in soil water (Larssen et al., 2011; Zhu et al., 2013b). Little is known about
how P addition affects N cycling and N₂O emission in these acidic, nutrient-poor soils. Likewise,
the importance of increased mineral-inorganic N concentrations for soil-atmosphere exchange of

102	CH ₄ , and how this is affected by P fertilization remain to be elucidated for soils of the subtropics.
103	Here, we assessed N ₂ O and CH ₄ fluxes in an N-saturated subtropical forest in SW China under
104	ambient N deposition and studied the effects of P addition on emission rates, nutrient availability
105	and tree growth. The objectives were i)We hypothesized that i) P addition stimulates forest
106	growth; ii) stimulated forest growth results in increased N uptake by trees and understory
107	vegetation, and thus decreases the soil inorganic N concentration; iii) P addition reducesto
108	quantify soil N ₂ O and CH ₄ -emission and promotes CH ₄ uptakes, ii) to investigate the effect of P
109	addition on N ₂ O and CH ₄ emission.

110 **2 Materials and Methods**

111 **2.1 Site description**

The study site "TieShanPing" (TSP) is a 16.2 ha subtropical forest (29° 380 N, 106° 410 E; 450 112 m a.s.l.), about 25 km northeast of Chongqing, SW China. TSP is a naturally regenerated, 113 114 secondary mixed coniferous-broadleaf forest, which developed after clear cutting in 1962 (Larssen et al., 2011). The forest stand is dominated by Masson pine (Pinus massoniana) and has 115 a density of about 800 stems ha⁻¹ (Huang et al., 2015). TSP has a monsoonal climate, with mean 116 annual precipitation of 1028 mm, and a mean annual temperature of 18.2 °C (Chen and Mulder, 117 2007a). Most of the precipitation (> 70%) occurs during summer periods (April to September). 118 Soils are predominantly well-drained, loamy yellow mountain soil, classified as Haplic Acrisol 119 120 (WRB 2014), with a thin O horizon (< 2 cm). In the O/A horizon, soil pH is around 3.7, and the mean C/N and N/P ratios are 17 and 16, respectively. In the AB horizon, which has a slightly 121 higher pH, mean C/N is well above 20. The soil bulk density of the O/A horizon (~ 5 cm) is 122 about 0.75 g cm⁻³. Generally, Ssoil water-filled pore space (10 cm) at TSP hilltopon the 123 hillslopes -generally ranges from 50 to 70% (mean ~ 60%; Zhu et al., 2013b). More details on 124 soil properties are given in Table 1. 125

Annual <u>inorganic</u> N deposition at TSP measured in throughfall varies between 40 and 65 kg ha⁻¹
(dominated by NH₄⁺; Yu et al., 2016), while the annual bulk N deposition is from 20 to 30 kg ha⁻¹
¹ (Chen and Mulder, 2007b). According to regional data, annual P deposition via throughfall is <
0.40 kg ha⁻¹ (Du et al., 2016). Strong soil acidification at TSP-has been reported to resulted incause severe decline in forest growth at TSP since 2001 (Li et al., 2014; Wang et al., 2007), and in-a_decreased in abundance and diversity of ground vegetation (Huang et al., 2015).

Pronounced N saturation with strong NO₃⁻ leaching from the top soil has aggravated P deficiency at TSP (Huang et al., 2015). The total P content in the O/A horizon is ~ 300 mg kg⁻¹, while ammonium lactate-extractable P is smaller than 5 mg kg⁻¹ (Table 1).

135 **2.2 Experimental Design**

Three blocks, each having two 20 m ×* 20 m plots, were established on well drained soils of a 136 gently sloping hillside. Adjacent plots were separated by at least 10-m buffer zone. In each block, 137 plots were randomly assigned to a Reference and a P treatment. On 4 May 2014, a single dose of 138 P fertilizer was applied as solid NaH₂PO₄²H₂O, at a rate of 79.5 kg P ha⁻¹. The amount of P 139 added was estimated from P adsorption isotherms (Supplementary Materials, Table S1 and 140 Figure S1), to ensure significantly increase in soil available P. To apply P fertilizer evenly, we 141 divided each plot into a 5 m * 5 m grid and broadcasted the powdered fertilizer by hand in each 142 grid cell. The P dose applied at TSP was intermediate as compared to the 10 kg P ha⁻¹ yr⁻¹ 143 applied by Müller et al. (2015) to a mountain forest in Ecuador and the 150 kg P ha⁻¹ yr⁻¹ applied 144 by Zheng et al. (2016) to a subtropical forest in South China. 145

The addition of NaH₂PO₄·2H₂O at the P-treated plots also resulted in an input of 59.0 kg ha⁻¹ of sodium (Na). One month after the fertilizer application, Na⁺ concentrations in soil water of the P treatments were about 5 mg L⁻¹ at 5-cm depth and 3 mg L⁻¹ at 20-cm depth (Table S2). Although somewhat larger than in the reference Reference plots (0.52-1.31 mg L⁻¹), the Na⁺ concentration in soil water of the P treatments are unlikely to have exerted a strong significant negative impact on plant and microbial activities.

152 **2.3 Sample collection and analyses**

Within each plot, three ceramic lysimeters (P80; Staatliche Porzellanmanufaktur, Berlin) were 153 installed at 5- and 20-cm soils near the plot centre in August 2013. To obtain water samples, 154 350-ml glass bottles with rubber stoppers were pre-evacuated, using a paddle pump, and 155 connected to the lysimeters for overnight sampling. Between November 2013 and October 2015, 156 we sampled soil pore water bi-monthly in the dry and dormant season and monthly during the 157 growing season. All water samples were kept frozen during storage and transport. Concentrations 158 of NH_4^+ , NO_3^- , potassium (K⁺), calcium (Ca²⁺), and magnesium (Mg²⁺) in soil water were 159 160 measured at the Research Center for Eco-Environmental Sciences (RCEES), Chinese Academy of Sciences, Beijing, using ion chromatography (DX-120 for cations and DX-500 for anions). 161

In August 2013, soils from the O/A (0-3 cm), AB (3-8 cm) and B (8-20 cm) horizons were 162 sampled near the lysimeters for soil analysis. Total P and plant-available P contents were 163 monitored in samples collected from the O/A horizons every six months, starting two days 164 before P addition. Soil samples were kept cold (< 4 °C) during transport and storage. Before 165 analysis, soil samples were air dried and sieved (2 mm). Soil pH was measured in soil 166 suspensions (10 g dry soil and 50 ml deionized water) using a pH meter (PHB-4, Leici, China). 167 Total soil C and N contents were determined on dried and milled samples, using a LECO 168 elemental analyzer (TruSpec[@]CHN, USA). To measure total P, 1 g dry soil was digested with 5 169 ml of 6 M H₂SO₄ (Singh et al., 2005) and measured as ortho-phosphate by the molybdenum blue 170 method (Murphy and Riley, 1962). Ammonium lactate (0.01 M)-extractable P and H₂O-171 extractable P (P_{Al} and P_{H2O}, respectively) were measured as ortho-phosphate after extraction (1.5 172 g dry soil in 50 ml solution) (Singh et al., 2005). Ammonium oxalate (0.2 M)-extractable Fe, Al 173 and P were measured by inductively coupled plasma optical emission spectroscopyinductive 174

175 coupled plasma (<u>ICP-OES7500;</u> Agilent, <u>USA</u>) after extraction (1.5 g dry soil in 50 ml
176 solution).

From August 2013 onwards, we measured N₂O and CH₄ emissions in triplicate close to the 177 lysimeters, using static chambers (Zhu et al., 2013b). The measurements were conducted bi-178 monthly in the dry and dormant season and monthly during the growing season, simultaneously 179 with the sampling of soil pore water. To investigate the immediate effect of P addition on N₂O 180 emissions, we also sampled the gas emissions once before (2 May) and three times (7, 10 and 12 181 May) after the P application. Gas samples (20 ml) were taken 1, 5, 15 and 30 minutes after 182 chamber deployment. and 20 ml gas samples were injected into pre-evacuated glass vials (12 ml) 183 crimp-sealed with butyl septa (Chromacol, UK), maintaining overpressure to avoid 184 contamination during sample transportshipment. Mixing ratios of N₂O and CH₄ were analyzed 185 using a gas chromatograph (Model 7890A, Agilent, USA) at RCEES, equipped with an ECD for 186 detection of N₂O (at 375 °C with 25 ml min⁻¹ Ar/CH₄ as make up gas), a FID for CH₄ (250 °C; 187 20 ml min⁻¹ N₂ as make-up gas) and a TCD for CO₂. Exchange rates between soil and 188 atmosphere (emission/uptake) were calculated from measured concentration change in the 189 chambers over time, applying linear or polynomial fits to the concentration data. Cumulative 190 N₂O emissions over time were estimated by linear interpolation between measurement dates 191 192 (Zhu et al., 2013b).

From October 2013 onwards, litterfall was collected during the first week of every month in five replicates per plot. Litterfall collectors were made of 1 m² nylon nets (1 mm mesh size), held in place by four wooden poles 0.8 m above the ground. Fresh litter was dried at 65°C. In early November 2013 and 2014 (at the end of the growing season), we collected current-year pine needles from several branches of three trees in each plot. The collected needles were dried at

65 °C and the dry weight of 500 needles was determined. A subsample was dried at 80 °C and 198 finely milled prior to chemical analysis at the Chinese Academy of Forestry. Total C and N were 199 measured using an elemental analyzer (FLASH 2000; Thermo Scientific; USA). The contents of 200 K, Ca, Mg and P in the needles were determined by ICP-AES (IRIS Intrepid II; Thermo 201 Scientific; USA) after digesting 0.25 g dry weight samples with 5 ml of ultra-pure nitric acid. In 202 November 2013, and 2014, and in February of 2015, we measured the height and the diameter at 203 breast height (DBH) of 6 to 10 Masson pines (only those marked in November 2013; with DBH > 204 205 5 cm) at each plot. These data were DBH was then used to estimate the standing biomass of Masson pines based on standard allometric equations (Li et al., 2011; Zeng et al., 2008). 206

Daily average air temperature and sum ofdaily total precipitation were monitored from July 2013
to November 2015 by a weather station (WeatherHawk 232, USA) placed on the roof at the local
forest bureau, in about 1 km distance from the sampling site (Yu et al., 2016).

210 **2.4 Statistical analyses**

211 Statistical analyses were performed using R version 3.3.1 (R Core Team, 2016). All data were tested for normality (Kolmogorov-Smirnov's test) and homoscedasticity (Levene's test) before 212 further analysis. If not normally distributed, the data were normalized by logarithmic 213 transformation. Considering heterogeneity among blocks, temporal variabilities of NO₃⁻ 214 concentrations, N₂O and CH₄ fluxes were presented separately for each block. For time series 215 data, we used linear mixed-effect (LME) models, to account for both repeated measurements and 216 within-group variance of a stratification variable (block design). LME models were applied to 217 test the effects of P addition on soil N₂O and CH₄ fluxes, NH₄⁺, NO₃⁻, K⁺, Ca²⁺ and Mg²⁺ 218 concentrations in soil water, as well as litterfall weight (Koehler et al., 2009; Müller et al., 2015). 219

220 The analysis was based on data for plot means (the average of 3 subplot replicates) from three blocks. In LME models, treatments (Reference or P addition) were considered fixed effects, 221 while sampling time and plots were treated as random effects. We then assessed the significance 222 of fixed effects through analysis of variance for LME models. One-way analysis of variance 223 (ANOVA, Turkey post-hoc test) was conducted to examine the treatment effects on soil pH, 224 nutrient contents in organic matter, tree biomass, 500-needle weight and needle nutrient content 225 for each samplingand data of tree growth. Significance levels were set to p < 0.05, if not 226 227 specified otherwise.

228 **3 Results**

229 3.1 Nutrient concentrations in soil and soil water

Addition of P resulted in a significant increase in soil P content in the O/A horizon, both as P_{Al} 230 and total P (Table 2). However, after 15 months, only PAI indicated an enhanced P status, while 231 total soil P did not differ significantly from background values at the reference Reference sites. P 232 addition had no significant effect on soil pH, or soil C and N content. The NO₃⁻ concentration in 233 soil water collected at 5 cm depth varied seasonally, with significantly greater values (30-40 mg 234 N L⁻¹) towards the start of the growing season in 2015 (April, Fig. S2), but not in 2014, likely 235 due to dilution by abundant precipitation in February to March 2014. Addition of P resulted in 236 significantly smaller NO_3^- concentrations in soil water at both 5- and 20-cm depths (Fig. 1b). In 237 general, the concentration of NH_4^+ in soil water was small (< 0.5 mg L⁻¹) and not affected by P 238 addition (Fig. 1a). At both depths, mean soil water concentrations of Mg^{2+} and Ca^{2+} were 239 significantly smaller in the P-treated than the reference Reference plots, and the sum of charge of 240 dissolved base overall cationic charges declined significantly in response to P addition (Fig. S3). 241

242 3.2 N₂O and CH₄ fluxes: effects of P addition

In the Reference plotsDuring the experimental period, N₂O fluxes varied seasonally (Fig. 2), showing a significant relationship with daily precipitation (Fig. S4a), but not with daily mean temperature (Fig. S4b). In the Reference plots, mMean N₂O fluxes were generally below 50 μ g N₂O-N m⁻² hr⁻¹ in the dry, cool season, but reached values of up to 600 μ g N₂O-N m⁻² hr⁻¹ in the growing season (Fig. 2). Cumulative N₂O emissions were estimated with seasonally averaged fluxes, and they differed greatly among the three blocks (Fig. 3)-, of which -block 2 had Tthe greatest annual N₂O emission was observed in the Reference plot (7.9 kg N ha⁻¹) of block 2. <u>CH₄</u> 250fluxes in the Reference plots also varied greatly among blocks (Fig. 5). Net-emission of CH_4 was251observed in summer 2013 (~ 80 µg CH_4 -C m⁻² hr⁻¹) in blocks 1 and 2, whereas block 3 showed252 CH_4 uptake. From spring 2014 until October 2015, CH_4 fluxes were less variable in all blocks,253with values fluctuating around zero. A longer period of net-emission was observed in block 3254during the dry season 2014. The fluxes did not correlate with precipitation or air temperature255(Figs. S5c&d).

Mean N₂O fluxes during the 1.5 years after P addition were significantly smaller in the P treatment than in the Reference (Fig. 4). The P addition resulted in a 50% (3 kg N₂O-N ha⁻¹ yr⁻¹ on average) decrease reduction in of cumulative N₂O emission by about 3 kg N ha⁻¹ - yr⁻¹ on average, which is a 50% reduction (Fig. 3). No immediate effects (within days) of P addition on N₂O emission was observed (Fig. S5).

261 CH4 fluxes varied greatly between blocks (Fig. 5). Net-emission of CH4 was observed in summer 2013 (~ 80 µg C m⁻² hr⁻¹) in blocks 1 and 2, whereas block 3 showed CH₄ uptake. From spring 262 2014 until October 2015, CH4 fluxes were less variable in all blocks, with values fluctuating 263 around zero. A longer period of net-emission was observed in block 3 during the dry season 264 2014. The fluxes did not correlate with either precipitation or air temperature (Fig. S5c&d). In 265 the 1.5 years following P addition, mean CH₄ fluxes indicated net CH₄ emission (~ +3.8 µg CH₄-266 <u>C</u> m⁻² hr⁻¹) in the Reference, whereas net CH₄ uptake (~ -6.5 μ g C<u>H₄-C</u> m⁻² hr⁻¹) was observed in 267 268 the P treatment (Fig. 6). The suppressing effect of P addition on CH₄ emission was significant, in accordance with what was found for NO_3^- concentration and N_2O emission. 269

270 **3.3 The effect of P addition on tree growth**

271	Throughout the 2-year experimental period, we observed no significant change in tree biomass
272	(138 tha^{-1}) in response to P addition (Table S3). Likewise, there was no effect of P treatment on
273	the 500-needle weight (13 g on average). Between the two samplings in 2013 and 2014, we
274	found differences in chemical composition of the pine needles, but this effect was not linked to P
275	additionthe difference between the Reference and P treatment was not significant. Also, the C/N
276	and N/P ratios of the needles (40 and 16, respectively) were hardly-not affected by P addition.
277	Monthly litterfall varied seasonally in both Reference and P treatment (Fig. S6), but no
278	significant difference was found between the two treatments.

279 **4 Discussion**

Background-N₂O emission rates in the Reference plots were relatively large (Fig. 2), with mean 280 values close to 100 μ g N₂O-N m⁻² hr⁻¹ (Fig. 4). This is within the range of N₂O emission rates 281 previously reported for well-drained hillslope soils at TSP (Zhu et al., 2013b), but greater than 282 the rates reported for other forests in South China. For instance, N₂O emission rates averaged to 283 37 µg N₂O-N m⁻² hr⁻¹ in unmanaged sites at Dinghushan (Fang et al., 2009; Tang et al., 2006) 284 and 50 µg N₂O-N m⁻² hr⁻¹ in N-fertilized sites (Zhang et al., 2008a). TSP Reference plots emitted 285 on average 5.3 kg N_2 O-N ha⁻¹ yr⁻¹ (Fig. 3), which is about 10% of the annual N deposition (50 kg 286 \underline{N} ha⁻¹ yr⁻¹) (Huang et al., 2015). These fluxes are well above average fluxes reported for tropical 287 rainforests (Werner et al., 2007). Large N₂O emissions at TSP are likely due to the large N 288 deposition rates (Huang et al., 2015), as suggested by the A similar trends of increasing N₂O 289 290 emissions with increasing N deposition rates has been reported for-indicated by data from a wide range of ecosystems (Liu et al., 2009). Also, warm-humid conditions during monsoonal summers 291 may stimulate N₂O emissions (Ju et al., 2011), as monsoonal rainstorms triggered peak fluxes 292 (Pan et al., 2003). The positive correlation between precipitation and N₂O emission peaks (Fig. 293 S4a) may indicate the importance of denitrification as the dominant N₂O source. This is 294 supported by recent ¹⁵N tracing experiments at TSP (Yu et al., 2017; Zhu et al., 2013a). 295

Addition of P caused a significant decline in soil mineral-inorganic N in soil water (predominantly NO_3^- ; Fig. 2), particularly during summers, when NO_3^- concentrations were relatively high-large (Fig. S2). At the same time, annual N₂O emissions decreased by more than 50% (Figs. 3 and 4). These findings are consistent with a number of previous studies (Baral et al., 2014; Hall and Matson, 1999; Mori et al., 2014), which attributed the reduction of N₂O

301	emissions in P-treated soils to decreased NO3 ⁻ availability and thus less denitrification. decreased
302	mineral N content, most likely as a consequence of The attenuation of soil NO ₃ by P addition at
303	<u>TSP may reflect</u> -stimulated <u>plant N</u> uptake by plants and/or soil microbial assimilation organisms.
304	In a similarly N-rich, tropical forest in South China, Chen et al. (2016) reported a stimulation of
305	net N mineralization and nitrification after six years of bi-monthly P addition, despite reduced
306	soil NO_3^- concentration. Therefore, it is likely that plant uptake plays a more important role in P-
307	induced N retention than immobilization by soil microbes. However, during our study period of
308	two years, we did not find significant increase of N uptake based on tree biomass and foliar N
309	content measurements (Table S3). An alternative explanation could be that P addition stimulated
310	of N uptake by ground vegetation, which remains to be confirmed. It is noteworthy that there was
311	no significant correlation between N ₂ O emission rates and soil water NO ₃ ⁻ concentration in our
312	study (Figs. 2 and S2), suggesting that the suppressing effect of P on N ₂ O emissions was indirect,
313	probably by affecting the competition for mineral N between plant roots and microbes (Zhu et al.,
314	2016).

In contrast to our 1.5-year-study, P-addition experiments in South Ecuador (Martinson et al., 315 2013) and South China (at Dinghushan Biosphere Reserve (Zheng et al., 2016) found no effect 316 of a single P addition on N₂O emission during the first two years after application. However, 317 significant reduction in N2O emission was observed after three to five years with of continuous P 318 319 addition, both at the Ecuadorian and the Chinese site (Chen et al., 2016; Müller et al., 2015). For the montane forest site in Ecuador, the observed delay in N2O emission response to P addition 320 may be explained by the moderate amount of P added (10 kg P ha⁻¹ yr⁻¹; Martinson et al., 2013). 321 Moreover, the experiments were conducted in a forest withthe relatively low ambient N 322 deposition (~ 10 kg N ha⁻¹ yr⁻¹) and small N₂O fluxes (~ 0.36 kg N ha⁻¹ yr⁻¹ in the Reference 323

324	plots) (Martinson et al., 2013; Müller et al., 2015). In addition, the moderate amount of P added
325	(10 kg P ha ⁻¹ yr ⁻¹ ; Martinson et al., 2013) could have resulted in an insignificant P effect in the
326	first two years. By contrast, t The Dinghushan site in South China receives 28-36 kg inorganic N
327	ha ⁻¹ yr ⁻¹ through wet inorganic N depositionby throughfall (Zheng et al., 2016Chen et al., 2016;
328	Fang et al., 2008), which is similar to the inorganic N deposition at our site TSP (Chen and
329	Mulder, 2007b; Huang et al., 2015). <u>However, soil KCl-extractable inorganic N (~ 40 mg N kg⁻¹;</u>
330	Zheng et al., 2016) and NO ₃ ⁻ leaching (~ 20 kg N ha ⁻¹ yr ⁻¹ ; Fang et al., 2008) at the Dinghushan
331	site are several-fold smaller than at our site (~ 100 mg N kg ⁻¹ and ~ 50 kg N ha ⁻¹ yr ⁻¹ ,
332	respectively) (Huang et al., 2015; Zhu et al., 2013b). Also, the mean N ₂ O emission rates in the
333	reference plots (10 μ g m ⁻² h ⁻¹) at Dinghushan were smaller than at TSP (> 50 μ g m ⁻² h ⁻¹ ; Fig. 4).
334	These indicate that Dinghushan forest has stronger N assimilation and is thus less N-rich than
335	<u>TSP forest.</u> They also observed larger N_2O emission rates (~ 0.88 kg N ha ⁻¹ yr ⁻¹ in the Reference
336	plots) than in the Ecuadorian site. However, forests do not always display a straightforward
337	relationship between N deposition and N_2O emissions. Manipulation experiments in the
338	European NITREX project, for instance, revealed a much stronger correlation of N ₂ O emissions
339	with soil NO ₃ ⁻ leaching than with N deposition (Gundersen et al., 2012). Indeed, KCl-extractable
340	mineral N at the Dinghushan site (~ 40 mg kg ⁻¹ ; Zheng et al., 2016) is several fold smaller than
341	at our site (> 100 mg kg ⁻¹ ; Zhu et al., 2013b), indicating that Dinghushan is less N-rich than TSP.
342	This-Therefore, we suggests that the response of N ₂ O emission to P addition might-may depend
343	on the N status of the soil. The fact that numerous studies found apparent suppression of N_2O
344	emission in short-term experiments (< 2 years) in N + P treatments, but not in treatments with P
345	alone, supports this idea (Müller et al., 2015; Zhang et al., 2014b; Zheng et al., 2016).

Other studies have Another study in a secondary in South China reported observed increased N2O 346 emissions during two years upon after P addition, in a secondary mixed forest (Mori et al., 2013c; 347 Wang et al., 2014). In an Acacia mangium plantation, fertilized with P, Mori et al. (2013b&c) 348 found that N2O emissions were stimulated in the short-term but reduced in the long-term. While 349 suppression of N₂O emission by P has been attributed to increased plant N uptake (Mori et al., 350 2014), increased N₂O emission is are generally explained by enhanced microbial biomass-growth 351 (Liu et al., 2012) and denitrification activity (Ehlers et al., 2010; He and Dijkstra, 2015). P 352 stimulation of N2O emission by microbial denitrification should be rather fast, as indicated by 353 Mori et al. (2013c) in a short-term (one week) incubation experiment with soils from an Acacia 354 355 mangium plantation. Unlike Mori et al. (2013c), we did not find increased N₂O emissions measured shortly within a week after P addition at our site -in May 2014 were not different from 356 fluxes in untreated reference plots (Fig. S5). This may indicate that suggest that denitrifier 357 358 community at TSP was not responsive to the P applied, probably because TSP hillslope soils 359 have large denitrification potentials (Zhu et al., 2013c). plant uptake at TSP is more important for the effect of P addition on N₂O emissions than changes in microbial activity, which are expected 360 to occur more rapidly. 361

The Reference plots at TSP showed net CH₄ emission for extended periods of the year (Figs. 5 and 6). Also, long-term CH₄ fluxes sampled between 2012 and 2014 on hillslope <u>soilss</u> near-by (Fig. S7; Zhu et al., unpublished data) showed net CH₄ emission. This is in contrast to the generally reported CH₄ sink function of forested upland soils (Ciais et al., 2013; Dutaur and Verchot, 2007). For example, <u>net CH₄ uptake ratefluxe</u>s reported for well-drained, forest soils in South Chinese China forest range from <u>-</u>30 to <u>-</u>60 µg C<u>H₄-C</u> m⁻² hr⁻¹ (Fang et al., 2009; Tang et al., 2006; Zhang et al., 2014a). Since aerated upland soils typically provide favourable conditions

369	for microbial CH ₄ uptake (Le Mer and Roger, 2010), the net emission observed in our sites is
370	unlikely <u>to be</u> due to enhanced <u>CH₄</u> production, but rather <u>by due to supressed CH₄ consumption</u> .
371	One reason-general explanation for the net CH ₄ emission at TSP could be inhibition of CH ₄
372	oxidation by NH_4^+ , as reported previously which competes with CH_4 for the active site at the
373	methane monooxygenase enzyme (Bodelier and Laanbroek, 2004; Zhang et al., 2014a). The
374	concentration of NH_4^+ in the soil water was rather small (< 0.5 g L ^{-1;} Fig. 1), which does not
375	preclude, however, that NH_4^+ availability from the soil exchangeable pool is may have been high.
376	Zhu et al. (2013b) found extraordinarily high KCL-extractable NH_4^+ in TSP surface soils, likely
377	reflecting the large atmogenic NH_4^+ input at the TSP site (Huang et al., 2015). On the other hand,
378	Reay and Nedwell (2004) found that NO ₃ ⁻ inhibits methanotrophic activity in acidic soils, where
379	<u>NH₃ is scarce. Possible mechanisms are the toxicity of denitrification intermediates (e.g. NO₂;</u>
380	Wang and Ineson, 2003) and the osmotic effect of high NO ₃ ⁻ concentration (Hütsch et al., 1996).
381	This deduction can be supported by the high NO_3^- concentration in the acidic soils at TSP (Figs.
382	<u>1 and S2).</u>

P addition had a significant impact on CH₄ fluxes, changing the soil from a net source to a net 383 sink on an annual basis (Fig. 6). However, the uptake rates of CH₄ in the P treatments remained 384 smaller than those reported for forest soils in tropical China (Tang et al., 2006; Zhang et al., 385 386 2008b). The stimulating effect of P addition on CH₄ uptake is consistent with previous studies 387 (Mori et al., 2013a, 2013b; Zhang et al., 2011), and has been attributed to lessening the NH4⁺alleivating N inhibition of methane oxidation. Unfortunately, we did not measure KCl-388 extractable NH_4^+ in our study, but a decline of available NH_4^+ , which is the substrate for 389 nitrification, is likely as NO3⁻ concentrations in soil water were significantly smaller with in the 390 P-treatments (Fig. 1). P addition may also result in a change of the taxonomic composition of the 391

methane oxidizing community (Mori et al., 2013a; Veraart et al., 2015). Alternatively, CH₄ oxidation may be stimulated by increased CH₄ diffusion into the soil, due to enhanced root growth and increased soil water loss due to transpiration in P-amended plots (Zhang et al., 2011). Given the strong N enrichment of TSP forest (Huang et al., 2015), it is likely that the reason for the observed reduction in CH₄ emissions in response to P fertilization is due to alleviating direct NH₄⁺ inhibition of methane monooxygenase (Veldkamp et al., 2013), rather than to Pstimulation of methanotrophic activity (Veraart et al., 2015).

Shortly after fertilizer application, we observed a modest, albeit significant increase of Na⁺ 399 concentration in soil water (Table S2). Other studies have documented the potential toxicity of 400 excess Na⁺ in soil water to plant and microbial activities (Rengasamy et al., 2003; Wong et al., 401 2008). However, the occurrence of Na⁺ toxicity at the treated plots to a degree affecting N 402 turnover processes in our plots, is unlikely, as Na⁺ concentrations in soil water, within one month 403 after application (Table S2), did not exceed 5 mg L^{-1} , which is far smaller than the values 404 commonly assumed to cause indicate toxicity (40 to 100 mg L⁻¹) (Bernstein 1975). Frequent 405 precipitation at TSP (Yu et al., 2016), both prior and following the addition of NaH₂PO₄.2H₂O 406 (Fig. 2), apparently may have diluted and leached Na⁺, thus preventing toxic effects. 407

P application significantly increased plant-available P in the P-limited TSP soil (Table 2). Meanwhile, concentrations of leachable base cations (K^+ , Mg^{2+} , Ca^{2+}) in soil water decreased (Fig. S3), as expected from the reduction of NO₃⁻ concentrations in the P-treatments, which represent a major decline in mobile anions in the P-treated soils (Mochoge and Beese, 1986). We observed no sign of stimulated forest growth or increased N uptake by <u>plants-trees</u> within the relatively short period of our study (Table S3 and Fig. S6), making it difficult to link the observed reduction in <u>mineral-inorganic</u> N in the soil solution (Fig. 1) to plant growth. When 415 interpreting the observed P effect on NO₃⁻ concentrations in soil water, several aspects need to be 416 considered. Firstly, two years of observation may be too short to detect any significant increase 417 in tree growth, due to NO₃⁻ uptake by plants, given the commonly large variabilities in tree biomass estimates (Alvarez-Clare et al., 2013; Huang et al., 2015). Secondly, a significant 418 proportion of the added P, and of excess N, may have been assimilated by the understory 419 biomassygetation, which was not assessed in this study. Previously, understory 420 biomassvegetation has been reported to quickly respond to P addition (Fraterrigo et al., 2011). 421 422 Thirdly, as long-term N saturation and acidification at TSP have reduced forest health (Lu et al., 2010; Wang et al., 2007), we may not expect immediate response of forest growth to P addition. 423 Large needle N/P ratios (17-22, Table S3) indicated that P limitation for tree growth was not 424 relieved 1.5 years after P addition (Li et al., 2016). Therefore, enhanced N uptake by understory 425 growth and/or soil microbial biomass may have been the main mechanisms responsible for 426 observed NO₃⁻ decline in the P-treated soil (Hall & Matson 1999). 427

428 Overall, o<u>O</u>ur study suggests that N-saturated TSP soils act as a regional hotspot for N₂O<u>(Zhu et</u> 429 <u>al., 2013b)</u> and CH₄ emissions. Within the short experimental period of 1.5 years, P fertilization 430 was shown to significantly decrease NO_3^- concentrations in soil water and to <u>overall</u> reduce<u>-both</u> 431 N₂O and CH₄ emissions. These findings provide a promising starting point for improving forest 432 management towards GHG abatement targets, taking into account the P and N status of 433 subtropical soils in the region.

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Table 1 Background Ambient soil properties of the experimental plots at Tieshanping (TSP).676Values are means and standard deviations in parenthesis $(n = 6)^{\oint}$. Soils were sampled in August6772013.

	Soil Layer	pН	Total C	Total N	Total P	C/N	N/P
			g kg ⁻¹	g kg ⁻¹	mg kg ⁻¹		
	O/A (0-3 cm)	3.7 (0.1)	80.7 (32.3)	4.8 (1.7)	308 (57)	17.0 (2.5)	15.5 (5.7)
Block 1	AB (3-8 cm)	3.8 (0.0)	23.9 (9.3)	1.3 (0.6)	*	20.0 (3.0)	-
	B (8-20 cm)	3.9 (0.2)	8.6 (1.2)	< 0.05	-	-	-
Block 2	O/A (0-3 cm)	3.6 (0.1)	77.6 (13.4)	4.7 (0.8)	297 (44)	16.7 (1.3)	15.7 (2.8)
	AB (3-8 cm)	3.7 (0.1)	20.2 (5.3)	1.0 (0.3)	-	21.4 (3.3)	-
	B (8-20 cm)	3.9 (0.1)	7.1 (1.6)	< 0.05	-	-	-
Block 3	O/A (0-3 cm)	3.6 (0.1)	67.0 (15.5)	3.8 (0.8)	223 (45)	17.4 (0.6)	17.2 (3.7)
	AB (3-8 cm)	3.6 (0.1)	21.0 (7.9)	1.1 (0.5)	-	24.5 (4.6)	-
	B (8-20 cm)	3.8 (0.1)	7.2 (1.5)	< 0.05	-	-	-
	Soil Layer	P _{Al}	Al _{ox}	Fe _{ox}	P _{ox}	P _{ox} /	
		mg kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	mg kg ⁻¹	$(Al_{ox} + Fe_{ox})$.)
Block 1	O/A (0-3 cm)	5.8 (1.4)	1700 (513)	1933 (350)	85.8 (22.6)	0.025 (0.008)	
	AB (3-8 cm)	2.1 (0.6)	1217 (243)	1692 (493)	47.1 (22.0)	0.016 (0.007)	
	B (8-20 cm)	< 1.0	1083 (90)	1158 (249)	29.3 (28.6)	0.012 (0.011)	
Block 2	O/A (0-3 cm)	5.9 (1.0)	1500 (238)	1792 (215)	79.2 (21.5)	0.024 (0.007)	
	AB (3-8 cm)	1.6 (0.4)	925 (149)	1517 (320)	37.2 (10.7)	0.016 (0.006)	
	B (8-20 cm)	< 1.0	892 (209)	1033 (413)	16.1 (10.5)	0.009 (0.007)	
Block 3	O/A (0-3 cm)	4.1 (0.9)	1367 (180)	1667 (168)	50.7 (10.9)	0.017 (0.003)	
	AB (3-8 cm)	4.4 (4.0)	1075 (128)	1350 (150)	24.8 (8.3)	0.010 (0.002)	
	B (8-20 cm)	< 1.0	992 (130)	875 (138)	8.0 (2.0)	0.004 (0.001)	

 P_{H2O} - Water-extractable P, P_{A1} = Ammonium lactate-extractable P,

 $Al_{ox} = Oxalate extractable Al, Fe_{ox} = Oxalate extractable Fe, P_{ox} = Oxalate extractable P.$

 $\frac{}{}$ Water-extractable P was below a detection limit of 5 mg kg⁻¹, thus not presented in table,

681 ^{*} Data not available

Table 2 Soil pH, C, N and P contents in the O/A horizon (0-3 cm) in the References (Ref) and P treatments. Values are means and standard deviations in parenthesis (n = 9). P addition was conducted on 14/05/04, after the first two sampling dates.

		pH	Total C	Total N	C/N	P _{Al}	Total P
			g kg ⁻¹	g kg ⁻¹		mg kg ⁻¹	mg kg ⁻¹
13/08/02	Ref	3.7 (0.1) ^{ab†}	8.3 (2.3) ^{ab}	$0.5 (0.1)^{ab}$	16.9 (1.1) ^b	5.4 (1.4) ^a	292 (46) ^{ab}
	Р	$3.6(0.1)^{b}$	6.7 (2.0) ^b	$0.4 (0.1)^{b}$	17.1 (2.1) ^{ab}	5.1 (1.3) ^a	260 (70) ^b
14/05/02	Ref	$3.7(0.1)^{a}$	12.2 (4.2) ^a	$0.9 (0.3)^{a}$	13.7 (1.5) ^b	19.0 (8.0) ^a	336 (65) ^a
	Р	3.8 (0.2) ^a	9.0 (3.5) ^{ab}	0.7 (0.2) ^{ab}	14.2 (2.8) ^{ab}	13.7 (5.2) ^a	270 (72) ^a
14/05/10	Ref	3.8 (0.1) ^{ab}	9.9 (2.1) ^a	$0.7 (0.2)^{ab}$	$14.0 (0.7)^{b}$	15.4 (7.0) ^b	304 (49) ^b
	Р	$3.9(0.3)^{a}$	8.0 (1.9) ^a	$0.6 (0.1)^{b}$	14.3 (1.3) ^{ab}	174 (114) ^a	572 (242) ^a
14/12/02	Ref	$3.8(0.1)^{a}$	10.5 (3.6) ^a	$0.7 (0.3)^{a}$	14.5 (1.3) ^{ab}	14.2 (7.4) ^b	328 (102) ^b
	Р	3.9 (0.2) ^a	9.5 (2.1) ^a	0.7 (0.1) ^{ab}	14.0 (0.8) ^b	66 (24) ^a	442 (106) ^{ab}
15/08/02	Ref	3.9 (0.2) ^{ab}	8.3 (2.2) ^{ab}	$0.4 (0.1)^{ab}$	$20.5(2.5)^{a}$	13.4 (6.2) ^b	291 (61) ^a
	Р	$4.0(0.2)^{a}$	6.5 (1.9) ^b	$0.3 (0.1)^{b}$	19.7 (2.2) ^{ab}	57 (36) ^a	383 (136) ^a

[†] Different letters indicate significant differences <u>between References and P treatments</u> (p < 0.05).

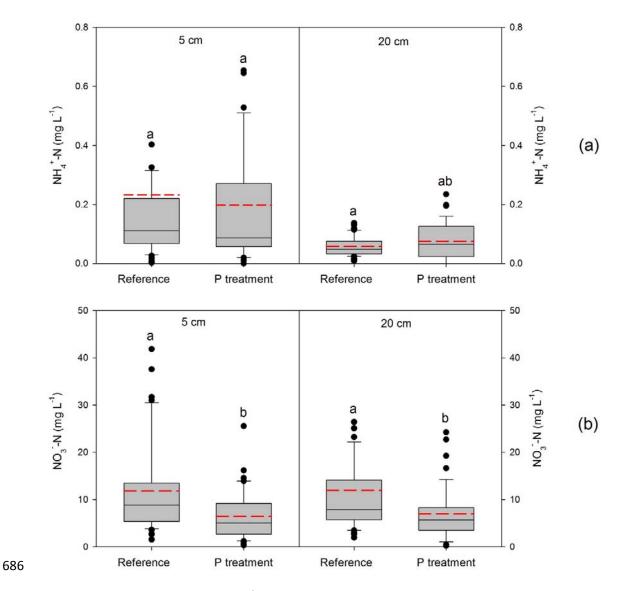


Fig. 1 Box whisker plots of NH_4^+ (a) and NO_3^- (b) concentration in soil water at 5- and 20-cm depths in the References and P treatments, throughout 1.5 years after the P addition; red dashed lines indicate mean values; different letters indicate significant differences (p < 0.05).

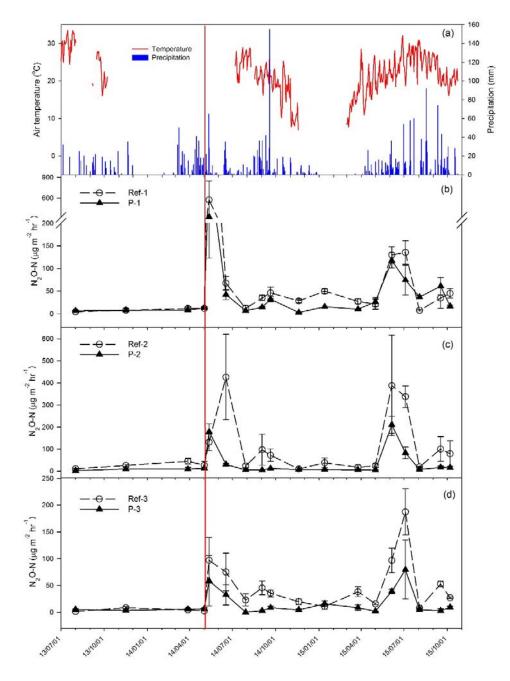
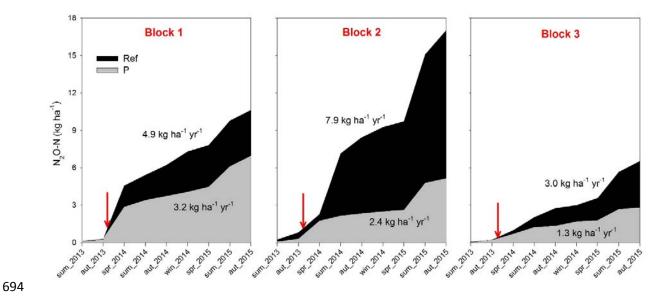
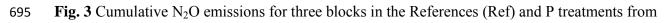
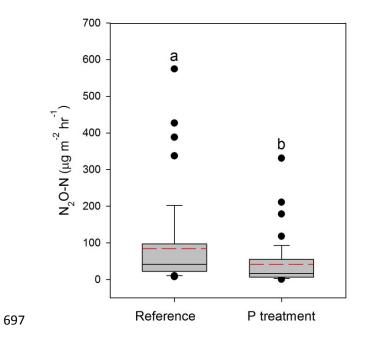


Fig. 2 Daily mean air temperature and precipitation (a), and monthly mean N_2O fluxes (±SE) in the References (Ref) and P treatments in each of the three blocks (b-d); the red vertical line gives the date of P addition (4 May, 2014).





summer 2013 to autumn 2015; the red arrows refer to the date of P addition (4 May, 2014).



698Fig. 4 Box whisker plots for N2O fluxes in the Reference and P treatment throughout 1.5 years699after the P addition; red dashed lines indicate mean values; linear mixed-effect models were used700to test the P treatment effect; different letters indicate significant difference (p < 0.05).

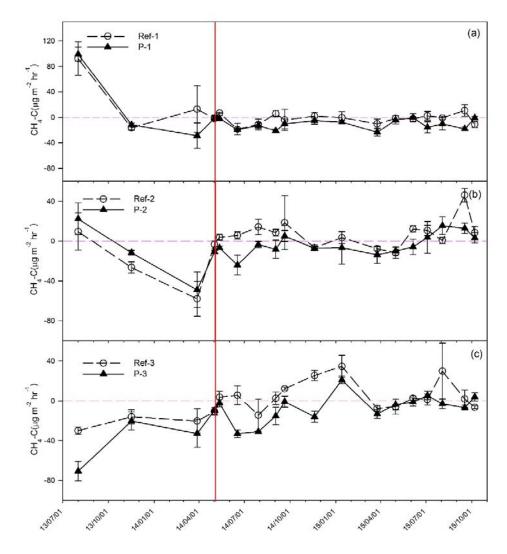
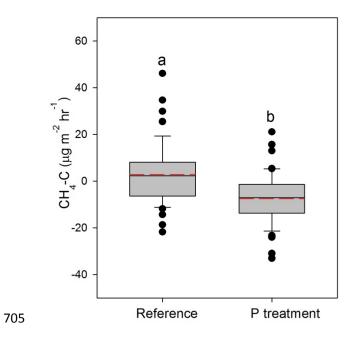


Fig. 5 Monthly mean CH₄ fluxes (±SE) in the References (Ref) and P treatments for three blocks
(a-c); the horizontal broken line indicates zero flux the red vertical line refers to the date of P
addition (4 May, 2014).



706Fig. 6 Box whisker plots of CH_4 fluxes in the Reference and P treatment throughout 1.5 years707after the P addition; red dash lines indicate mean values; linear mixed-effect models were used to708test the P treatment effect; the different letters indicate significant difference (p < 0.05).</td>