

Authors' response to referees

We thank both referees for their supporting comments and good suggestions to improve our manuscript.

During the revision, we recognized a mistake in the conversion of TEP fluxes reported by Martin et al (2011) and Ebersbach et al (2014) into TEP-C fluxes. We corrected this mistake. The change did not affect our interpretations.

Anonymous Referee #1

Received and published: 15 December 2016

Review of bg-2016-508 Particle export fluxes to the oxygen minimum zone of the Eastern

Tropical North Atlantic Anja Engel, Hannes Wagner, Frédéric A. C. Le Moigne, Samuel T. Wilson

The authors present a study of vertical fluxes collected with surface tethered drifting sediment trap from the Eastern Tropical North Atlantic. They collected settling material

from 7 depths; 60 m, 100 m, 150 m, 200 m, 300 m, 400 m, 500 m, and 600 m. Depth between 300 and 500 m were sampling within the oxygen minimum zone. The main findings in the study was that transfer efficiencies in an oxygen minimum zone were higher than expected when only considering temperature dependency for the microbial

degradation of organic matter and that the composition of the organic matter within the settling aggregates had a large impact on the transfer efficiencies. The latter finding

was evident through observations of higher attenuation of amino acids compared to polysaccharide-rich TEP. The manuscript is well written and the data clearly presented.

I only have some minor issues regarding the vertical flux of TEP (see specific comments). I recommend the manuscript for publication in Biogeosciences with minor

revision. Specific comments:

Line 57: Please insert a comma after ".. (Volk and Hoffert, 1985)".

Response: comma was inserted

Line 210: Were the filters for the elemental analyzer wrapped in tin foil or packed in aluminium cups?

Response: The filters were enclosed in tin cups; information was added.

Line 364: The Gum Xanthan flux is per square meter, please correct to $\text{m}^{-2} \text{d}^{-1}$ for both Martin et al. (2011) and Ebersbach et al. (2014).

Response: Units were corrected

Line 388-389: It could also be due to slower sinking velocities. It is not possible to say from b alone which process is driving the values. However, you can say that more

degradation occurred within a depth region, either due to faster degradation or slower settling.

Response: Potential effect of reduced sinking velocity was included

Line 417-419: Looking at figure 3c, I do not see this trend? For deployment #2 there is an increase above the OMZ, then a slight decrease between 200 and 300 m whereafter it is stable and then show a decrease between 500 and 600 m. For deployment #1 it seems like there is no significant changes in TEP flux between 150 and 600 m. So I do not see that there is a clear different between TEP fluxes within the OMZ compared to below.

Response: During both deployments no clear decrease in TEP fluxes was determined for the OMZ. TEP fluxes decreased between 500 and 600m, i.e. below the OMZ; this was more pronounced during the second deployment. We will modify the text to clarify this better.

Line 464-466: This was only observed for deployment #2, not for deployment #1. Deployment#1 showed decreasing ratios already within the OMZ, 400 to 500 m.

Response: Correct, we will modify the text to state that a decrease in TEP-C:POC ratios below the OMZ was only observed during deployment #2.

Anonymous Referee #2

Received and published: 10 February 2017

Particle export fluxes to the oxygen minimum zone of the Eastern Tropical North Atlantic

This paper by Engel et al. describes the flux and composition of particles from surface waters through the oxygen minimum zone of the water column off the coast of Mauritania in the Eastern Tropical North Atlantic. Two deployments of surface-tethered drifting sediment traps collected particles at 7-8 depths including the OMZ from 300-500m. Calculated transfer efficiencies of the total POM and various characterized components of the particles indicate lower attenuation through the OMZ than predicted from seawater temperature. Particle composition data showed highest transfer efficiencies for TEP and lowest for amino acids. The paper is well written and a nice contribution regarding particle flux dynamics in OMZs. I have no major issues with the text and recommend publication after minor revisions detailed below.

Minor comments:

- pg 3, line 57: add comma after citation
- pg 4, line 81: ref LeMoigne et al., 2012 is missing from reference list; LeMoigne et al., 2016 is listed but not cited, perhaps one of these is an error

- 103 - pg 4, line 85: typo, Alldredge is misspelled
104 - pg 4, line 98: typo, change 'be' to 'been'
105 - pg 5, line 130 (and elsewhere throughout): I am not familiar with this spelling of
106 'Mauretania' with an e and could not find other references to Mauritania spelling as
107 such so would recommend changing (certainly if this is an accepted spelling it does
108 not need to be changed)
109 - pg 5, line 131: 'drifting' is written twice
110 - pg 6, line 145: hyphenate 'surface-tethered'
111 - pg 8, line 185: insert 'of' (Aliquots of samples: : :)
112 - pg 9, line 215: remove 'in' (: : :were filtered onto: : :)
113 - pg 9, line 221: hyphenate 'peroxydisulphate-containing'
114 - pg 9, line 231: ref Dittmar et al., 2009 is missing from reference list
115 - pg 11, line 269: close parentheses after 'sample'
116 - pg 17, line 432: opal is incorrectly capitalized
117 - pg 19, line 488: Keil et al., 2016 is missing from reference list (or incorrectly cited
118 here as 2015 is in the list)
119 - pg 20, line 494: insert 'the' (: : :explain the relative: : :)
120 - pg 24, lines 592-600: the two Buesseler et al., 2007 references should be
121 distinguished
122 as a and b here and when cited in the manuscript
123 - pg 26, line 652: Giering reference is formatted incorrectly (begins with 'Sarah')
124 - pg 27, line 675: Kartensen et al., 2008 is not cited in manuscript but is listed here
125 - pg 31, line 760: Ploug and Jorgensen, 1999 is not cited in manuscript but is listed
126 here
127 - pg 34: If possible, it would be nice to include the calculated degradation indices
128 along
129 with the amino acid composition in table 2.

130
131
132 *Response: All suggestions have been considered. Values for the DI's have been*
133 *included in table 2.*
134

135

136

137
138
139 **Particle export fluxes to the oxygen minimum zone of the**
140 **Eastern Tropical North Atlantic**
141
142
143

144 Anja Engel¹, Hannes Wagner¹, Frédéric A. C. Le Moigne¹, Samuel T. Wilson²
145

146 ¹ GEOMAR Helmholtz Centre for Ocean Research Kiel,
147 24105 Kiel, Germany
148

149 ² Daniel K. Inouye Center for Microbial Oceanography: Research and Education, Department
150 of Oceanography, University of Hawaii, Honolulu, HI 96822, USA
151
152
153

154 *Correspondence to:* Anja Engel (aengel@geomar.de)
155
156
157
158
159
160
161

162 **Abstract.** In the ocean, sinking of particulate organic matter (POM) drives carbon export
163 from the euphotic zone and supplies nutrition to mesopelagic communities, the feeding and
164 degradation activities of which in turn lead to export flux attenuation. Oxygen (O₂) minimum
165 zones (OMZs) with suboxic water layers (<5 μmol O₂ kg⁻¹) show a lower carbon flux
166 attenuation compared to well oxygenated waters (>100 μmol O₂ kg⁻¹), supposedly due to
167 reduced heterotrophic activity. This study focuses on sinking particle fluxes through hypoxic
168 mesopelagic waters (<60% μmol O₂ kg⁻¹); these represent ~100 times more ocean volume
169 globally compared to suboxic waters, but have less been studied. Particle export fluxes and
170 attenuation coefficients were determined in the Eastern Tropical North Atlantic (ETNA) using
171 two surface tethered drifting sediment trap arrays with 7 trapping depths located between 100
172 and 600m. Data on particulate matter fluxes were fitted to the normalized power function
173 $F_z = F_{100} (z/100)^{-b}$, with F_{100} being the flux at a depth (z) of 100 m and b being the attenuation
174 coefficient. Higher b -values suggest stronger flux attenuation and are influenced by factors
175 such as faster degradation at higher temperatures. In this study, b -values of organic carbon
176 fluxes varied between 0.74 and 0.80 and were in the intermediate range of previous reports,
177 but lower than expected from seawater temperatures within the upper 500m. During this
178 study, highest b -values were determined for fluxes of particulate hydrolysable amino acids
179 (PHAA), followed by particulate organic phosphorus (POP), nitrogen (PN), carbon (POC),
180 chlorophyll a , and transparent exopolymer particles (TEP), pointing to a sequential
181 degradation of organic matter components during sinking. Our study suggests that in addition
182 to O₂ concentration, organic matter composition co-determines transfer efficiency through the
183 mesopelagial. The magnitude of future carbon export fluxes may therefore also depend on
184 how organic matter quality in the surface ocean changes under influence of warming,
185 acidification, and enhanced stratification.

Formatiert: Tiefgestellt

Gelöscht: -

Gelöscht:

Gelöscht: oxygen

Formatiert: Tiefgestellt

1. Introduction

The biological carbon pump, defined as the export of biologically fixed [carbon dioxide \(CO₂\)](#) from the surface to the deeper ocean mainly in the form of sinking particles (Volk and Hoffert, 1985), influences atmospheric CO₂ concentration and affects ecosystem structure and elemental distributions in the ocean. The total amount of carbon export as well as the efficiency of the biological carbon pump, *i.e.* the ratio between export and primary production, are highly dynamic (Buesseler and Boyd, 2009; Lam et al., 2011). Changes in the efficiency of the biological carbon pump may have been responsible for past atmospheric CO₂ variability between glacial-interglacial transition periods (Kohfeld and Ridgwell, 2009) and play a key role for future climate predictions (Heinze et al., 2015).

Most of the POM being exported below the surface mixed layer (<200m in general) is solubilized and remineralized within the mesopelagic layer, *i.e.* between depths of 200 and 1000 m (Bishop et al., 1978; Suess, 1980). The shallower the carbon remineralization depth, the more likely is CO₂ to exchange with the atmosphere, and hence drive a shorter carbon storage time in the ocean (Volk and Hoffert, 1985; Kwon et al., 2009). Factors driving export flux attenuation in the mesopelagic have therefore a large influence on CO₂ sequestration in the ocean. The vertical profile of sinking particulate organic carbon (POC) flux has often been described by a normalized power function: $F_z = F_{100}(z/100)^{-b}$, where F_z is the particle flux as a function of depth z , F_{100} is the flux at 100 m depth, and b is the flux attenuation coefficient (Martin et al., 1987; hereafter *M87*). The authors of the *M87* study derived an ‘open ocean composite’ for POC export fluxes from North Pacific data with a $F_{100} = 50.3 \text{ mg m}^{-2} \text{ d}^{-1}$ and $b = 0.86$. However strong regional variations of both total export POC fluxes and b values are observed (Martin et al., 1987; Buesseler et al., 2007a; Torres Valdes et al., 2014; Marsay et al., 2015) with several factors proposed to control export flux attenuation. Increased attenuation, *i.e.* higher b -values, have been related to increased temperature (Marsay et al.,

217 2015), zooplankton feeding activity (Lampitt et al., 1990), coprophagy, coprorhexy, and
 218 coprochaly (Belcher et al. 2016), microbial cycling (Giering et al., 2014) and lack of ballast
 219 (LeMoigne et al., 2012). Decreased flux attenuation, *i.e.* lower b -values, and thus higher
 220 transfer efficiencies (T_{eff}) have been associated to high particle sinking velocity depending on
 221 plankton community composition, especially the presence of larger phytoplankton cells
 222 (Buesseler, 1998; Buesseler and Boyd 2009), particle aggregates (Alldredge and Gotschalk,
 223 1989), and fecal pellets (Cavan et al., 2015). Organic polymers, such as transparent
 224 exopolymer particles (TEP) increase the rate of aggregate formation due to their high
 225 stickiness (Alldredge et al., 1993; Engel, 2000; Passow, 2002; Chow et al., 2015) and
 226 supposedly play an important role in particle export fluxes (Passow, 2002; Arrigo, 2007;
 227 Chow et al., 2015). TEP are carbon-rich particles that form from dissolved polysaccharides
 228 (Engel et al., 2004). When included in sinking POM inventories, TEP may increase carbon
 229 relative to nitrogen export fluxes, a mechanism potentially counteracting rising CO_2
 230 concentration in the atmosphere (Schneider et al., 2004; Arrigo, 2007; Engel et al., 2014).
 231 However, TEP themselves are non-sinking due to a high water content and low density
 232 (Azetzu-Scott and Passow, 2004), and little quantitative data are available on TEP export by
 233 sinking particles so far (Passow et al., 2000; Martin et al., 2011; Ebersbach et al., 2014).
 234 Thus, the role of TEP in carbon export is still unresolved.

235 Reduced POC flux attenuation has also been suggested for oxygen minimum zones (OMZs)
 236 (Martin et al., 1987; Haake et al., 1992; Devol and Hartnett, 2001; Van Mooy et al., 2002;
 237 Keil et al., 2015) as a consequence of reduced zooplankton feeding and microbial degradation
 238 activities in suboxic ($<5 \mu\text{mol O}_2 \text{ kg}^{-1}$) waters. So far, the vast majority of mesopelagic
 239 downward POM flux measurements originate from well oxygenated waters ($>100 \mu\text{mol O}_2$
 240 kg^{-1}). In the *M87* study, five sets of drifting sediment traps were deployed in the oxygenated
 241 North Pacific and four sets were deployed in the Eastern Tropical North Pacific (ETNP)
 242 OMZ. The flux attenuation coefficients (b) for the oxygenated North Pacific averaged $0.90 \pm$

Gelöscht: d

244 0.06, while lower b values averaging 0.66 ± 0.24 were measured in the ETNP OMZ. In
245 agreement, Devol and Hartnett (2001) and Van Mooy et al. (2002) observed low particle
246 attenuation in the OMZ of the ETNP off Mexico, yielding b coefficients of 0.36 and 0.40,
247 respectively. Keil et al. (2015) found b values of 0.59-0.63 in the suboxic Arabian Sea. These
248 studies thus indicate that a greater proportion of the sinking POM escapes degradation while
249 sinking through suboxic waters. However, influence of oxygen on organic matter degradation
250 may vary between individual components. For instance, degradation of hydrolysable amino
251 acid under suboxic conditions was found to continue with the same rate as compared to oxic
252 conditions (Van Mooy et al. 2002; Pantoja et al. 2004), suggesting that anaerobic and micro-
253 aerobic bacteria preferentially utilize nitrogen-rich components.

254
255 So far, little is known on sinking POM flux attenuation in hypoxic waters ($<60 \mu\text{mol O}_2 \text{ kg}^{-1}$),
256 which are more widespread ($\sim 4\%$ of ocean volume) compared to suboxic waters ($< 0.05\%$ of
257 ocean volume). Laboratory studies indicated that particle aggregates sinking through hypoxic
258 waters can become suboxic within their interior due to oxygen diffusion limitation and evolve
259 microbial degradation processes typical for suboxic waters (Alldredge and Cohen, 1987;
260 Ploug et al., 1997; Stief et al., 2016). For example, at an ambient O_2 concentration of $60 \mu\text{mol}$
261 kg^{-1} , the O_2 uptake by a 2 mm (diameter) aggregate was diffusion-limited and a 0.5 mm wide
262 anoxic core occurred within its interior (Ploug and Bergkvist, 2015). Since OMZs are
263 expected to expand in the future as a consequence of global warming and altered circulation
264 patterns (Stramma et al., 2008), the role of oxygen in controlling the biological pump
265 efficiency needs to be better constrained for predicting ocean-climate feedbacks. In order to
266 assess what controls carbon flux attenuation and depth-related changes in sinking particle
267 composition in hypoxic waters, we determined downward POM fluxes in the ETNA off the
268 coast of Mauritania, which exhibits an extensive hypoxic OMZ between 300 and 500 m. We
269 used two parallel drifting, surface-tethered sediment trap devices with particle interceptor

Gelöscht: e

Gelöscht: drifting

traps (PITs) at 7-8 different depths between 60-600 m to estimate fluxes to and within the OMZ.

2. Methods

2.1. The Study area

The study was conducted from March 17th to April 16th 2014 during a cruise of the RV METEOR to the ETNA region off the coast of Mauritania (Fig. 1a). The study area included hypoxic waters with minimum values of oxygen concentration of 40 $\mu\text{mol kg}^{-1}$ as determined by CTD (Seabird) casts with two calibrated oxygen sensors at midwater depths of 350-500 m (Fig. 1b) (Visbeck, 2014).

Gelöscht: e

2.2. Sediment trap operation and sample analysis

Free-drifting surface-tethered sediment trap devices were deployed for 196 h during the first deployment and 281h during the second deployment (Fig. 1c). The first trap device was deployed on the 24th of March 2014 (11:00 UTC) at 10.00°N 21.00°W with 12 Particle-Interceptor-Traps (PITs) at each of 8 depths: 60, 100, 150, 200, 300, 400, 500, and 600 m. The device was recovered on the 1st of April 2014 (14:30 UTC) at 10.46°N 21.39°W. The second device was deployed on the 27th of March 2014 (16:00 UTC) at 10.25°N 21°W with 12 PITs at each of 7 depths: 100, 150, 200, 300, 400, 500, and 600 m. The second trap device was recovered on the 8th of April 2014 (09:00 UTC) at 10.63°N 21.50°W. Both devices slowly drifted northwest and were recovered approximately 37 nm away from their deployment location (Fig. 1c). Within the drifting area oxygen concentration in the OMZ resembled the overall pattern of the Mauritanian upwelling with fully hypoxic conditions between 300 and 500 m (Fig. 1d).

Gelöscht:

Gelöscht: e

300 The design of the trap devices and the drifting array basically follows Knauer et al. (1979),
301 with 12 PITs mounted on a polyvinylchloride (PVC) cross frame. The PITs were acrylic tubes
302 with an inside diameter of 7 cm, an outside diameter of 7.6 cm and a height of 53 cm, leading
303 to an aspect ratio of 7.5. The aspect ratio and a baffle system consisting of smaller acrylic
304 tubes attached to the top end of each PIT help to reduce drag-induced movement within the
305 trap (Soutar et al., 1977). PVC crosses with PITs were attached to a free-floating line, which
306 was buoyed at the surface and weighed at the bottom. The surface buoys of the arrays carried
307 GPS/Iridium devices and flashlights.

308 Prior to each deployment, each PIT was filled with 1.5 L filtered surface seawater (0.2 μ m
309 pore size cartridge) collected from the ship's underway seawater system, up to 3/4 of the
310 PITs' height. A brine solution was prepared by dissolving 50 g L⁻¹ sodium chloride with
311 filtered surface seawater and subsequently filtered through a 0.2 μ m cartridge to remove
312 excess particulates. 20 ml of formalin was then added per L of the solution to achieve a brine
313 solution with 2% formalin. The preservative solution was then slowly transferred into each
314 PIT beneath the 1.5 L of filtered seawater using a peristaltic pump. PITs were covered with
315 lids immediately, to minimize contamination before deployment.

316 Sample treatment after trap recovery followed recommendations given by Buesseler et al.
317 (2007**b**). After recovery, all PITs were capped to minimize contamination. The density
318 gradient was visually inspected and found intact at the position of prior to deployment or at a
319 maximum 2 cm above. Then, seawater was pumped out of each PIT using a peristaltic pump
320 down to 2-3 cm above the density gradient. The remaining ~0.6 L were subsequently
321 transferred to canisters, pooled from 11 tubes per depth. 40 ml formalin were added to each
322 canister. Samples from each depth were passed through a 500 μ m nylon mesh. Swimmers
323 were removed from the mesh with forceps under a binocular microscope and the remaining
324 particles, which stuck to the mesh, were transferred back to the sample. Samples were
325 subsequently split into aliquots of the total sample. Therefore, the pooled sample was

transferred into a round 10 L canister and stirred at medium velocity with a magnetic bar. Aliquots were transferred into 0.5 L Nalgene bottles with a flexible tube using a peristaltic pump. Aliquots of samples were filtered under low pressure (<200 mbar) onto different filter types (combusted GF/F 0.7 µm, polycarbonate 0.4 µm, or cellulose acetate 0.8 µm; see below) for different analyses and stored frozen (-20 °C) until analyses.

331

2.2.1. Biogeochemical Analyses

The following parameters were determined: Total particulate mass (TPM), particulate organic carbon (POC), particulate nitrogen (PN), particulate organic phosphorus (POP), biogenic silica (BSi), chlorophyll *a* (Chl *a*), particulate hydrolysable amino acids (PHAA) and transparent exopolymer particles (TEP).

337

TPM was analyzed in triplicate. The following aliquots were filtered in triplicate onto pre-weighed 0.4 µm polycarbonate filters: 800 ml (2 x 400 ml; 8 % of total sample) for the depths of 600 m to 300 m of deployment #1, 400 ml (4 % of total sample) for the depths of 200 m and 150 m of deployment #1 and for all depths of deployment #2, 420 ml (4 % of total sample) for the depth of 100 m and 60 m of deployment #1. Filters were rinsed two times with Milli-Q water, dried at 60 °C for 4 h and stored until weight measurement on a Mettler Toledo XP2U microbalance.

345

POC and PN aliquots were filtered in triplicate onto combusted (8h at 500°C) GF/F filters (Whatmann, 25 mm): 400 ml (4 % of total sample) for the depths of 600 m to 150 m of deployment #1, 420 ml (4 % of total sample) for the depths of 100 m and 60 m of deployment #1, 100 ml (1 % of total sample) for all depths of deployment #2. For the depths of 150 m, 100 m and 60 m of deployment #1, 400 - 420 ml (4 % of total sample) was filtered onto two filters, due to the high particle load at these depths. Filters were exposed to fuming

352 hydrochloric acid in a fuming box over night to remove carbonate and subsequently dried
353 (60°C, 12 h). For analysis, the filters were enclosed in tin cups and analysed using an Euro
354 EA elemental analyzer calibrated with an acetanilide standard. For the depths of 150, 100 and
355 60 m of deployment #1 the sum of both filters was taken.

Gelöscht: wrapped

Gelöscht: foil

356
357 POP was determined in triplicate, except for 60 m depth of deployment #1, which was only
358 determined in duplicate. The following aliquots were filtered onto combusted GF/F filters
359 (Whatmann, 25 mm): 400 ml (4 % of total sample) for the depths of 600 m to 150 m of
360 deployment #1, 420 ml (4 % of total sample) for the depths of 100 m and 60 m of deployment
361 #1, 100 ml (1 % of total sample) for all depths of deployment #2. For the depths of 200 m to
362 60 m of deployment #1, the volume of 400 ml/ 420 ml (4 % of total sample) was filtered onto
363 two filters, due to the high particle load at these shallower depths. Organic phosphorus
364 collected on the filters was digested in the potassium peroxydisulphate-containing substance
365 Oxisolv (Merck) for 30 min in a pressure cooker and measured colorimetrically as ortho-
366 phosphate following the method of Hansen and Koroleff (1999).

Gelöscht: in

Gelöscht:

367
368 PHAA were determined in duplicate. The following aliquots were filtered onto combusted
369 GF/F filters (25 mm): 400 ml (4 % of total sample) for the depths of 600 m to 150 m of
370 deployment #1, 420 ml (4 % of total sample) for the depths of 100 m and 60 m of deployment
371 #1, 100 ml (1 % of total sample) for all depths of deployment #2. For the depths of 150 m,
372 100 m and 60 m of deployment #1, the volume of 400 ml / 420 ml (4 % of total sample) was
373 filtered onto two filters, due to the high particle load at these shallower depths. PHAA
374 analysis was performed according to Lindroth & Mopper (1979) and Dittmar et al. (2009)
375 with some modifications. Duplicate samples were hydrolyzed for 20 h at 100°C with
376 hydrochloric acid (30%, Suprapur, Merck) and neutralized by acid evaporation under vacuum
377 in a microwave at 60°C. Samples were washed with water to remove remaining acid.

382 Analysis was performed on a 1260 HPLC system (Agilent). Thirteen different amino acids
383 were separated with a C18 column (Phenomenex Kinetex, 2.6 μm , 150 x 4.6 mm) after in-line
384 derivatization with o-phthalaldehyde and mercaptoethanol. The following standard amino
385 acids were used: aspartic acid (AsX), glutamic acid (GLX), histidine (His), serine (Ser),
386 arginine (Arg), glycine (Gly), threonine (Thr), alanine (Ala), tyrosine (Tyr), valine (Val),
387 phenylalanine (Phe), isoleucine (Ileu), leucine (Leu), γ - amino butyric acid (GABA). α -
388 amino butyric acid was used as an internal standard to account for losses during handling.
389 Solvent A was 5% acetonitrile (LiChrosolv, Merck, HPLC gradient grade) in
390 sodiumdihydrogenphosphate (Merck, suprapur) buffer (pH 7.0), Solvent B was acetonitrile. A
391 gradient was run from 100% solvent A to 78% solvent A in 50 minutes. The detection limit
392 for individual amino acids was 2 nmol monomer L^{-1} . The precision was <5%, estimated as the
393 standard deviation of replicate measurements divided by the mean. The degradation index
394 (DI) was calculated from the amino acid composition following Dauwe et al. (1999).

395
396 BSi was determined in triplicate. The following aliquots were filtered onto cellulose acetate
397 filters (0.8 μm): 400 ml (4 % of total sample) for the depths of 600 m to 150 m of deployment
398 #1, 420 ml (4 % of total sample) for the depths of 100 m and 60 m of deployment #1, 200 ml
399 (2 x 100 ml; 2 % of total sample) for all depths of deployment #2. Filters were incubated with
400 25 ml NaOH (0.1 M) at 85°C for 2h 15min in a shaking water bath. After cooling of the
401 samples, analysis was conducted according to the method for determination of $\text{Si}(\text{OH})_4$ by
402 Hansen and Koroleff (1999). Fluxes of biogenic opal were calculated assuming a water
403 content of ~10% and therefore the chemical formula $\text{SiO}_2 \times 0.4\text{H}_2\text{O}$ with a density of ~2.1 g
404 cm^{-3} (Mortlock and Fröhlich 1989).

Gelöscht:

405
406 Chl *a* was determined in duplicate. The following aliquots were filtered onto GF/F filters (25
407 mm): 400 ml (4 % of total sample) for the depths of 600 m to 150 m of deployment #1, 420

409 ml (4 % of total sample) for the depths of 100 m and 60 m of deployment #1, 100 ml (1 % of
 410 total sample) for all depths of deployment #2. For the depths of 200 m to 60 m of deployment
 411 #1, the volume of 400 ml / 420 ml (4 % of total sample) was filtered onto two filters, due to
 412 the high particle load at these shallower depths. Samples were analyzed after extraction with
 413 10ml of acetone (90%) on a Turner fluorimeter after Welschmeyer (1994). Calibration of the
 414 instrument was conducted with spinach extract standard (Sigma Aldrich).

415
 416 TEP were determined in quadruplet by microscopy after Engel (2009). Between 3.5 and 10 ml
 417 (0.03-0.1% of total sample) for the depths of deployment #1 and #2 were filtered onto 0.4 μ m
 418 Nuclepore membrane filters (Whatmann) and stained with 1 mL Alcian Blue solution. Filters
 419 were mounted onto Cytoclear© slides and stored at -20 °C until microscopy analysis using a
 420 light microscope (Zeiss Axio Scope A.1) connected to a camera (AxioCAM Mrc). Filters
 421 were screened at 200x magnification. 30 pictures were taken randomly from each filter in two
 422 perpendicular cross sections (15 pictures each; resolution 1040 x 1040 pixel, 8-bit color
 423 depth). Image analysis software WCIF ImageJ (Version 1.44, Public Domain, developed at
 424 the US National Institutes of Health, courtesy of Wayne Rasband, National Institute of Mental
 425 Health, Bethesda, Maryland) was used to semi-automatically analyse particle numbers and
 426 area.

427
 428 The carbon content of TEP (TEP-C) was estimated after Mari (1999) using the size dependent
 429 relationship:

430
 431
$$\text{TEP-C} = a \sum_i (n_i r_i^D), \quad (1)$$

432
 433 with n_i being the number of TEP in the size class i and r_i the mean equivalent spherical radius
 434 of the size class. The constant $a = 0.25 * 10^{-6}$ (μ g C) and the fractal dimension of aggregates

435 D= 2.55 were proposed by Mari (1999). TEP-C was only calculated for the size fraction <5
436 μm including mainly free TEP, because larger TEP included TEP covered aggregates with
437 solid particles. Estimating carbon content of these larger particles would overestimate TEP-C
438 as the volume of the other particles would be included.

439

440 2.3. Calculations and statistics

441 Fluxes of CaCO_3 and lithogenic matter (lith) were calculated as:

442

$$443 [\text{CaCO}_3 + \text{lith}] = [\text{TPM}] - [\text{POM}] - [\text{Opal}], \quad (2)$$

444

445 Total mineral ballast ($\text{ballast}_{\text{total}}$) was calculated as:

446

$$447 [\text{ballast}_{\text{total}}] = [\text{TPM}] - [\text{POM}], \quad (3)$$

448

449 and the percentage of $\text{ballast}_{\text{total}}$ ($\% \text{ballast}_{\text{total}}$) was calculated as:

450

$$451 [\% \text{ballast}_{\text{total}}] = ([\text{TPM}] - [\text{POM}]) / [\text{TPM}] * 100, \quad (4)$$

452

453 The transfer efficiency (T_{eff}) of particulate components was calculated as the ratio of fluxes at
454 600 m to those at 100 m.

455

456 Calculated mean values include replicate measurements of both deployments. Data fits and
457 statistical tests were performed with the software packages Microsoft Office Excel 2010,
458 Sigma Plot 12.0 (Systat) and Ocean Data View (ODV) (Schlitzer, 2013). Weighted-average
459 gridding was used in ODV to display data according to data coverage with automatic scale
460 lengths. The overall significance level was $p < 0.05$.

3. Results and Discussion

3.1. Fluxes of different compounds

Export fluxes of TPM and particulate organic elements determined during both trap deployments showed good overall agreement and a decrease with depth, fitting well to the power law function of *M87* (Fig. 2a-d, Fig.3a-d and Table 1). Averaging fluxes from both deployments yielded a total mass flux of $240 \pm 34 \text{ mg m}^{-2} \text{ d}^{-1}$ at 100 m decreasing to $141 \pm 8.8 \text{ mg m}^{-2} \text{ d}^{-1}$ in the core of the OMZ (400 m) (Fig. 2a). Fluxes of POC, PN and POP at 100 m depth were 73 ± 8.8 , 13 ± 1.4 and $0.67 \pm 0.06 \text{ mg m}^{-2} \text{ d}^{-1}$, respectively, and decreased to 26 ± 4.5 , 3.0 ± 0.41 and $0.19 \pm 0.04 \text{ mg m}^{-2} \text{ d}^{-1}$ at 400 m depth (Fig. 2b-d). The contribution of POC flux to total mass flux (% OC) decreased from about 30% at 60-150 m depth to 17-20% at 400 m depth and showed only a minor decrease below 400 m, to 14-16% at 600 m depth. Similarly, the percentage of PN flux to total mass flux (% N) showed the largest decrease between 60 and 400 m, i.e. from 6.6% to 2.0-2.3%, and less decline below, reaching 1.7-1.8% at 600 m. The percentage of POP flux to total mass flux (% P) decreased from 0.37% at 60 m depth to 0.11-0.16% at 400 m depth, and remained constant below 400 m depth. No previous data are available for POM export fluxes at our study site for direct comparison. However, our trap data compare well to carbon export fluxes estimated from particle size data (i.e. $10-300 \text{ mg C m}^{-2} \text{ d}^{-1}$) reported for 100 m depth in the area off Cape Blanc (Mauritania) by Iversen et al. (2010).

Fluxes of phytoplankton biomass, as indicated from Chl *a*, were similar at 100 m during both deployments, with $104 \pm 1.5 \text{ } \mu\text{g Chl } a \text{ m}^{-2} \text{ d}^{-1}$ during the first and $116 \pm 6.2 \text{ } \mu\text{g m}^{-2} \text{ d}^{-1}$ during the second deployment, but behaved differently below, with a stronger flux attenuation above

the OMZ during the first compared to the second deployment (Fig. 3a). Fluxes within the OMZ core were $35 \pm 0.1 \mu\text{g m}^{-2} \text{d}^{-1}$ (#1) and $53 \pm 0.5 \mu\text{g m}^{-2} \text{d}^{-1}$ (#2) respectively.

Opal fluxes were also similar during both deployments, yielding an average of $47 \pm 3.6 \text{ mg m}^{-2} \text{d}^{-1}$ at 100 m, steadily decreasing to $32 \pm 2.4 \text{ mg m}^{-2} \text{d}^{-1}$ at 400 m depth (Fig. 3b). Similar to Chl *a*, opal fluxes were slightly higher above the OMZ during the second compared to the first deployment, but quite similar or even lower below the OMZ. This may indicate that the second trap device, which drifted more northerly (Fig. 1c), exploited waters of a more recent diatom bloom compared to the first deployment.

Fluxes of $[\text{CaCO}_3 + \text{lith}]$ were similar to opal fluxes during the first deployment ($F_{100}=52 \text{ mg m}^{-2} \text{d}^{-1}$) but considerably lower during the second ($F_{100}=14.8 \text{ mg m}^{-2} \text{d}^{-1}$) (data not shown).

During this study, export fluxes of TEP were estimated from decrease over depth of total particle area and showed the strongest depth attenuation between 60 and 100 m during the first deployment (Fig. 3c). Like Chl *a* fluxes, TEP export fluxes were slightly higher during the second compared to the first deployment. At 100 m depth, average TEP flux was $1860 \pm 46 \text{ cm}^2 \text{ m}^{-2} \text{d}^{-1}$ and decreased to $1190 \pm 52 \text{ cm}^2 \text{ m}^{-2} \text{d}^{-1}$ at 400 m. Using a TEP size to carbon conversion according to Mari (1999) yielded to an average TEP-C ($<5 \mu\text{m}$) flux of $1.73 \pm 0.35 \text{ mg C m}^{-2} \text{d}^{-1}$ at 100m depth, slightly decreasing to $1.64 \pm 0.28 \text{ mg m}^{-2} \text{d}^{-1}$ at 400 m and further to $0.90 \pm 0.32 \text{ mg m}^{-2} \text{d}^{-1}$ at 600 m. Although TEP supposedly play an important role in particle export fluxes (Passow, 2002; Arrigo, 2007; Chow et al., 2015), only a few previous estimates for TEP export fluxes based on sediment traps have been given so far to which we can compare our data. Martin et al. (2011) measured TEP export fluxes during a spring bloom in the Iceland Basin (Northeast Atlantic Ocean) using the PELAGRA neutrally buoyant sediment traps and determined values in the range of 30-120 mg Gum Xanthan Equivalent $\text{m}^{-2} \text{d}^{-1}$. Ebersbach et al. (2014) obtained lower values of 0.03-5.14 mg Gum

515 Xanthan Equivalent $\text{m}^{-2} \text{d}^{-1}$ during the LOHAFEX iron fertilization experiment in the
 516 Southern Ocean. Assuming a conversion factor of $0.63 \mu\text{g C } \mu\text{g}^{-1}$ Gum Xanthan after Engel
 517 (2004), these previous estimates suggest TEP-C export fluxes ranging from 0.02 to 3 mg m^{-2}
 518 d^{-1} for the Southern Ocean and from 19 to $75 \text{ mg m}^{-2} \text{d}^{-1}$ for the North Atlantic spring bloom.
 519 Our data on TEP export fluxes for ETNA region are within the range of these previous
 520 studies, but closer to the lower estimates for the Southern Ocean. It has to be emphasized,
 521 though, that our calculated TEP-C fluxes are likely underestimates, since only suspended, i.e.
 522 ‘free’ TEP $< 5 \mu\text{m}$ were taken into account. TEP-C associated to aggregates cannot be
 523 determined with the applied microscopic technique. Overall, TEP-C export fluxes in the
 524 ETNA were significantly related to Chl *a* fluxes, yielding $[\text{TEP-C, mg m}^{-2} \text{d}^{-1}] = 11.9 [\text{Chl } a;$
 525 $\text{mg m}^{-2} \text{d}^{-1}] + 0.74 (r^2=0.59, n= 15, p<0.01)$.

526
 527 A strong decrease at shallow depth (60–100 m) was also observed for PHAA fluxes during
 528 the first deployment (Fig. 3d). Average PHAA fluxes were $330 \pm 51 \mu\text{mol m}^{-2} \text{d}^{-1}$ at 100 m,
 529 and $90 \pm 20 \mu\text{mol m}^{-2} \text{d}^{-1}$ in the OMZ core at 400 m. These fluxes are equivalent to amino
 530 acid related fluxes of $16.8 \pm 2.6 \text{ mg C m}^{-2} \text{d}^{-1}$ (100 m) and $4.48 \pm 1.0 \text{ mg C m}^{-2} \text{d}^{-1}$ (400 m),
 531 respectively, which are typical values for PHAA-C fluxes in the ocean (Lee and Cronin,
 532 1984). PHAA fluxes decreased slightly within the OMZ, i.e. from 300 to 500 m.

533
 534

535 3.2. Flux attenuation in the ETNA OMZ

536 Fluxes from both deployments were fitted to the exponential decrease model (Martin et al.,
 537 1987) and attenuation coefficients (*b*-values) were estimated for all components (table 1).

538 Higher *b*-values suggest stronger attenuation and may hint to either reduced sinking velocities
 539 of particles or to faster degradation of more labile components. During this study, PHAA
 540 were the most rapidly attenuated components of sinking particles, followed by POP, PN,

Formatiert: Hochgestellt

Gelöscht: ³

Gelöscht: 063

Gelöscht: m

Gelöscht: and Passow

Gelöscht: 1

Gelöscht: of

Gelöscht: 0.3

Gelöscht: a range of

Gelöscht: .

Gelöscht: -

Gelöscht: .

Gelöscht: both

Gelöscht: North Atlantic

Gelöscht: Accordingly

Gelöscht: labile

Gelöscht: during this study

POC, Chl *a*, and TEP (table 1). Attenuation of mineral fluxes was less pronounced than for TPM.

Attenuation coefficient of POC export fluxes was 0.80 during the first and 0.74 during the second deployment. These values are in the intermediate range of previously determined *b*-values for POC attenuation in the mesopelagic, shown to vary between 0.51 as determined in the North Pacific (K2) and 1.59 as determined for the NASG (Buesseler et al., 2007a; Marsay et al., 2015). Based on trap data from fully oxygenated water columns, Marsay et al. (2015) recently suggested a linear relationship between POC flux attenuation and median water temperature within the upper 500m of the water column according to: $b=0.062T+0.303$. Applying this relationship to our study area, with temperature decreasing from 26°C at the surface to 9°C at 500 m and a median temperature value of 12.01°C, would give a *b*-value of 1.05. This estimated *b*-value is higher than the values observed in this study (0.74 - 0.80) and suggests that oxygen deficiency may reduce attenuation of POC fluxes in the ETNA resulting in higher T_{eff} of organic matter though the OMZ's compared to well oxygenated waters.

Differences in flux attenuation coefficients translate into different T_{eff} for individual components, with PHAA being the least and TEP being the most efficiently exported organic component (table 1). In particular, values of T_{eff} for TEP and therewith for TEP-C were about three times higher than for PHAA-C and even clearly higher than for bulk POC, suggesting a preferential export of carbon included in TEP below 100 m. However, a steep decrease of TEP flux was observed between 60 m and 100 m during the first deployment. TEP are produced by a variety of organisms, i.e. different phytoplankton and bacterial species and cannot be considered as of homogenous composition. Several mechanisms may therefore be responsible for a change in TEP transfer efficiency with depth: 1) change of TEP degradability with depth, 2) differences in TEP composition over depth related to association with particles of different settling speed, 3) new production of TEP, abiotically or by bacteria,

583 during solubilization and degradation of sinking particles, 4) capture of suspended TEP by
584 sinking aggregates, or 5) reduced degradation rate of TEP at lower oxygen. In support of the
585 latter hypothesis, an attenuation of TEP fluxes within the OMZ (300-500 m) was not
586 detectable, but rather occurred again below the OMZ.

588 3.3. Changes in POM composition during export

589 POM, assumed to be $2.2 \times [\text{POC}]$ following Klaas and Archer (2002) made the greatest
590 contribution to TPM flux at 60 m, but decreased below. Conversely, [%ballast_{total}] increased
591 with depth, namely from 30% w/w at 60 m to 68% w/w at 600 m.

592 Biogenic opal (density: 2.1 g cm^{-3}) in the ocean is produced mainly by diatoms and
593 radiolarians. During this study, opal made a rather constant contribution to TPM fluxes with
594 20-25% weight below 100 m. Hence, the observed increase in the [%ballast_{total}] with depth
595 was due to an increasing contribution of CaCO_3 and lithogenic material. [CaCO_3 + lith] to
596 TPM increased from 10-15% above 150 m to 45% at 600 m. As a consequence, the ballast
597 ratio, defined as [Opal]:[CaCO_3 +lith] changed from a dominance of opal above the OMZ to a
598 dominance [CaCO_3 +lith] within and below the OMZ (Figure 4). Slight differences were
599 observed between the two deployments. Contribution of opal and of [CaCO_3 +lith] to TPM at
600 100m was almost equal during the first deployment with a share of 18% and 22%,
601 respectively. During the second deployment the contribution of opal to TPM at 100 m was
602 21% but only 6% for [CaCO_3 +lith]. Thus, the higher contribution of opal to TPM fluxes
603 together with higher Chl *a* fluxes indicated that diatomaceous material had a higher share of
604 particles sinking out of the euphotic zone down to the OMZ core during the second compared
605 to the first deployment.

606
607 Molar [POC]:[PN] ratios were close to the Redfield ratio at depths shallower than 100 m,
608 increased to a ratio of 10 at 400 m depth and remained constant between 400 and 600 m

Gelöscht: 0

610 depth (Fig. 5a). [PN]:[POP] ratios were much above Redfield, with values varying between
 611 30 and 45 throughout the water column (Fig. 5b). Also [POC]:[POP] ratios were much higher
 612 than Redfield ratios, and showed an increasing trend down to 300-400 m depth, while
 613 decreasing below (Fig. 5c). These changes in elemental ratios suggested a preferential
 614 remineralization of POP in the upper 300 m, followed by PN and POC deeper down.
 615 The percentage of total organic matter in TPM fluxes decreased from 67% at 100m to 32% at
 616 600m (Fig. 6d). As a consequence of higher T_{eff} of TEP relative to bulk POC, contribution of
 617 TEP-C to POC increased significantly with depth during both deployments ($p < 0.01$; $r^2 = 0.59$,
 618 $n = 15$) and was 2% at 100 m, and 6% within and 5% below the OMZ (Fig. 5e). Because TEP
 619 do not sink by themselves their export to depth depends on their incorporation into settling
 620 aggregates. In a laboratory study, Engel et al. (2009) observed that decomposition of TEP was
 621 faster relative to bulk POC for aggregates formed from calcifying and non-calcifying
 622 *Emiliania huxleyi* cultures. In that experiment, aggregate decomposition was investigated
 623 under oxic conditions. Other studies also showed fast microbial degradation of TEP under
 624 oxic conditions (Bar-Zeev and Rahav, 2015). One possible explanation for increasing [TEP-
 625 C]:[POC] in the hypoxic OMZ of the ETNA region could be that TEP are mostly included in
 626 sinking aggregates, whereas POC could be included in various particle types, such as large
 627 cells, detritus or fecal pellets. Ploug et al. (1997) estimated that carbon turn-over time inside
 628 anoxic aggregates can be strongly reduced. Due to high microbial activity and reduced water
 629 exchange aggregates sinking into hypoxic waters are more likely to experience anoxic
 630 conditions than individual particles (Ploug and Bergkvist, 2015). Thus, TEP settling into
 631 hypoxic waters by aggregates may be exposed to anoxia, and therewith to reduced microbial
 632 degradation, in consequence leading to a preferential TEP transfer through the OMZ. This
 633 may also explain the decrease of [TEP-C]:[POC] ratios below the OMZ at 600 m water depth,
 634 which was, however, only observed during the second deployment. Since PN was more

Gelöscht: observed

rapidly degraded than POC this also implied that the ratio of [PN]:[TEP-C] became lower with depth.

In contrast to [TEP-C]:[POC], values of [PHAA-C]:[POC] in POM fluxes declined during both deployments above the OMZ. However, in the core of the OMZ, at 400 m, [PHAA-C]:[POC] was higher than at 300 and 500 m (Fig. 5f); the same pattern was also observed for [PHAA-N]:[PN] (data not shown). A faster decline in PHAA in sinking particles mainly above but not within the OMZ is different to observations gained for more extensively oxygen-deficient to full anoxic waters of the Eastern Tropical south Pacific (ETSP), which suggested that PHAA are preferentially degraded under low oxygen conditions (Van Mooy et al., 2002). In those studies, total hydrolysable amino acid (THAA) degradation under anoxic conditions was found to continue with the same rate compared to oxic conditions, while degradation of non-amino acid compounds was found to slow down (Pantoja et al., 2004; Van Mooy et al., 2002). A preferential degradation of nitrogen-rich compounds over POC suggests that microbes degrading organic matter under strongly oxygen deficient conditions via denitrification preferentially utilize nitrogen-rich amino acids (Van Mooy et al., 2002). Our data on PHAA do not suggest preferential amino acid loss due to components of sinking POM degradation in the ETNA OMZ. This is in accordance with the absence of microbial N-loss processes/ absence of denitrifying bacteria in ETNA oxygen deficient waters (Löscher et al., 2016). Instead, a slight increase of [PHAA-C]:[POC] in the OMZ may point to higher protein production by bacterial growth as previously observed for mesopelagic waters (Lee and Cronin, 1982, 1984) and may be related to increased growth efficiency of bacteria experiencing low oxygen condition as suggested by Keil et al. (2016).

Among all amino acids determined, Glx, Gly, Gaba and Leu showed the most pronounced variations with depth (Fig. 6a-d, table 2). Whereas Glx and Leu showed a decrease with

depth (Fig. 6a, c), Gly continuously increased. It has been shown that Gly is enriched in the silica-protein complex of diatom frustules (Hecky et al., 1973). Preservation of frustules relative to POM may therefore explain [the](#) relative increase of Gly with depth in sinking particles. Glx has been used as a biomarker (Abramson et al., 2010), since Glx was shown to be enriched in calcareous plankton (Weiner and Erez, 1984). During this study %Mol of Glx was higher during the first deployment, which is in accordance with the observed higher contribution of [CaCO₃+lith] to TPM flux. Gaba has been used as an indicator for bacterial decomposition activity (Lee and Cronin, 1982; Dauwe and Middelburg, 1998; Engel et al., 2009). During this study %Mol Gaba behaved differently during the first compared to the second deployment with similar values within the OMZ, a pattern also observed for opal fluxes (Fig. 3b). Moreover, %Mol of Gaba showed a local peak at 300 m, i.e. within the upper oxycline, and may point to high bacterial activity at this depth. Leu is an essential amino acids and readily taken up by heterotrophic microorganisms. Little change in %Leu in the OMZ core (Fig. 3d) compared to above (<300 m) indicated reduced microbial reworking of organic matter under hypoxic conditions. Another indication of microbial reworking of organic matter can be derived from the Degradation index (DI) (Dauwe et al., 1999). During this study, the DI decreased with increasing depth, but with differences between the deployments (Fig. 7). During #2, DI was slightly higher above the OMZ indicating fresher material. During #1 DI did not decrease within the OMZ, but it continued to decrease from 300 m to 500 m depth during deployment #2. Together with observations on Chl *a* and opal fluxes, as well as changes in ballast ratio, data on DI suggest that the particles of more diatomaceous origin likely continued to decompose under hypoxic conditions.

4. Conclusions

687 Despite an improvement in understanding principle processes and drivers of particle export
688 processes over the past decades, spatial and temporal variability of export fluxes in the ocean
689 are still difficult to predict. This is partly due to the lack of observations in different regions
690 of the mesopelagic realm. Our study is the first to describe fluxes of POM in the hypoxic
691 mesopelagic waters of the ETNA. Our data suggest a higher transfer efficiency than expected
692 from seawater temperature solely, suggesting reduced degradation of organic matter by
693 heterotrophic communities at low oxygen concentration ($<60 \mu\text{mol O}_2 \text{ kg}^{-1}$). The biological
694 carbon pump in high productivity regimes associated to OMZs, i.e. Eastern Boundary
695 Upwelling Systems such as the ETNA region off Mauritania, may therewith be more efficient
696 than in fully oxygenated waters of comparable temperature. In contrast to suboxic systems ($<$
697 $5 \mu\text{mol O}_2 \text{ kg}^{-1}$) a relatively higher loss of amino acids from POM fluxes was not evident for
698 the hypoxic water-column, suggesting microbial N-loss processes were comparatively minor
699 within particles. This, however, requires further investigation since no corresponding rate
700 measurements of denitrification or anammox were conducted during this study. Organic
701 matter composition seems to have a large impact on transfer efficiencies as carbon fluxes
702 associated to amino acids were much more attenuated over depth than carbon fluxes
703 associated to polysaccharide-rich TEP. If these findings are transferable to other oceanic
704 regions, changes in surface ocean organic matter composition in response to climate change
705 may also impact the carbon remineralization depth and therewith may have a feed-back
706 potential to atmospheric CO_2 concentration that yet has to be assessed.

Gelöscht: e

708 5. Competing interest

709 The authors declare that they have no conflict of interest.

711 6. Acknowledgements

713 This study is a contribution to the Collaborative Research Center 754 / SFB
714 Sonderforschungsbereich 754 'Climate-Biogeochemistry Interactions in the Tropical Ocean'.
715 We thank Martin Visbeck, Toste Tanhua, Tobias Hahn, Sunke Schmidtke, and Gerd
716 Krahmann for scientific and technical support as well as for providing oxygen and CTD data.
717 Many thanks go to the shipboard scientific party and crew of Meteor cruise M105. Jon Roa,
718 Ruth Flerus, Scarlett Sett and Tania Klüver are acknowledged for technical assistance. We
719 thank Cindy Lee (Stony Brook University) for helpful advices. FACLM is supported by the
720 DFG Excellence cluster Future Ocean. All data will become available at www.pangea.de
721 upon publication.

722

723

724 **References**

725

726 Abramson, L., Lee, C., Liu, Z.F., Wakeham, S.G., and Szlosek, J.: Exchange between
727 suspended and sinking particles in the northwest Mediterranean as inferred from the organic
728 composition of in situ pump and sediment trap samples. *Limnol. Oceanogr.*, 55, 2, 725-739,
729 doi: 10.4319/lo.2009.55.2.0725, 2010.

730

731 Alldredge, A. L. and Cohen, Y.: Can microscale chemical patches persist in the sea?
732 Microelectrode study of marine snow, fecal pellets. *Science*, 235, 4789, 689-91, 1987.

733

734 Alldredge, A. L., and Gotschalk, C. C.: Direct observation of the mass flocculation of diatom
735 blooms: characteristics, settling velocities and formation of diatom aggregates. *Deep-Sea*
736 *Research*, 36, 159–171, 1989.

737

738 Alldredge, A. L., U. Passow, and Logan, B. E.: The abundance and significance of a class of
739 large, transparent organic particles in the ocean. *Deep-Sea Res.* 40, 1131–1140, 1993.

740

741 Arrigo, K. R.: Carbon cycle - Marine manipulations. *NATURE*, 450, 7169, 491-492, 2007.

742

743 Azetsu-Scott, K., and Passow, U.: Ascending marine particles: Significance of transparent
744 exopolymer particles (TEP) in the upper ocean. *Limnol. Oceanogr.* 49, 3, 741-748, 2004.

745

746 Bar-Zeev, E., and Rahav, E.: Microbial metabolism of transparent exopolymer particles
747 during the summer months along a eutrophic estuary system. *Frontiers in Microbiol.* 6, 403,
748 doi: 10.3389/fmicb.2015.00403, 2015.

749

750

751 Belcher, A., Iversen, M., Manno, C., Henson, S. A., Tarling, G. A., and Sanders, R.: The role
752 of particle associated microbes in remineralization of fecal pellets in the upper mesopelagic of
753 the Scotia Sea, Antarctica. *Limnol. Oceanogr.* 61, 3, 1049-1064, doi: 10.1002/lno.10269 ,
754 2016

755

756 Bishop, J. K. B., Ketten, D. K., and Edmon, J. M.: The chemistry, biology and vertical flux of
757 particulate organic matter from the upper 400 m of the Cape Basin in the southeast Atlantic
758 Ocean. *Deep-Sea Res.* 25, 1121-1161, 1978.

759

760 Buesseler, K. O.: The decoupling of production and particulate export in the surface ocean.
761 *Global Biogeochem. Cycles* 12:297-310, 1998.

762

763 Buesseler, K.O., C.H. Lamborg, P.W. Boyd, P.J. Lam, T.W. Trull, R.R. Bidigare, J.K.B.
764 Bishop, K.L. Casciotti, F. Dehairs, M. Elskens, M. Honda, D.M. Karl, D.A. Siegel, M.W.
765 Silver, D.K. Steinberg, J. Valdes, B. Van Mooy, and Wilson S.: Revisiting carbon flux
766 through the ocean's twilight zone. *Science*, 316, 567–570, 2007[a](#)

767

768 Buesseler, K.O., Antia, A., Chen, M., Fowler, S. W., Gardner, W.D., Gustafsson, O., Harada,
769 K., Michaels, A.F., van der Loeff, M. R., Sarin, M., Steinberg, D. K. and Trull, T.: An
770 assessment of the use of sediment traps for estimating upper ocean particle fluxes. *Journal of*
771 *Marine Research*, 65, 345-416, 2007[b](#).

772

773 Buesseler, K.O., and Boyd P.W.: Shedding light on processes that control particle export and
774 flux attenuation in the twilight zone. *Limnol. Oceanogr.*, 54, 4, 1210–1232, 2009

775

Gelöscht: ,

777 Cavan, E.L., Le Moigne, F. A. C., Poulton, A. J., Tarling, G. A., Ward, P., Daniels, C. J.,
778 Fragoso, G. M., and Sanders, R. J. : Attenuation of particulate organic carbon flux in the
779 Scotia Sea, Southern Ocean, is controlled by zooplankton fecal pellets. Geophys. Res. Lett.,
780 42, 3, 821-830, doi: 10.1002/2014GL062744, 2015

781

782 Chow, J. S., Lee, C. and Engel, A.: The influence of extracellular polysaccharides, growth
783 rate, and free coccoliths on the coagulation efficiency of *Emiliania huxleyi*. Mar. Chem., 175,
784 2015.

785

786 Dauwe, B., and Middelburg, J.J.: Amino acids and hexosamines as indicators of organic
787 matter degradation state in North Sea sediments. Limnol. Oceanogr. 43, 782-798, 1998.

788

789 Dauwe, B., Middelburg, J. J., Herman, P. M. J., and Heip, C. H. R.: Linking diagenetic
790 alteration of amino acids and bulk organic matter reactivity. Limnol. Oceanogr. 44, 1809–
791 1814, 1999.

792

793 Devol, A.H. and Hartnett, H. E.: Role of the oxygen minimum zone in transfer of organic
794 carbon to the deep ocean. Limnol. Oceanogr., 25, 1684-1690, 2001.

795

796 [Dittmar, T., Cherrier, J., and Ludwichowski, K.-U.: The Analysis of Amino Acids in](#)
797 [Seawater. In: Practical Guidelines for the Analysis of Seawater, CRC Press, 2009.](#)

798

799 Ebersbach, F., Assmy, P., Martin, P. et al.: Particle flux characterisation and sedimentation
800 patterns of protistan plankton during the iron fertilisation experiment LOHAFEX in the
801 Southern Ocean. Deep Sea Res. I, 89, 94-103, 2014.

802

Formatiert: Schriftart: Times New Roman

Formatiert: Standard, Block,
Zeilenabstand: Doppelt, Keine
Absatzkontrolle, Tabstopps: 0.99
cm, Links + 1.98 cm, Links + 2.96
cm, Links + 3.95 cm, Links + 4.94
cm, Links + 5.93 cm, Links + 6.91
cm, Links + 7.9 cm, Links + 8.89
cm, Links + 9.88 cm, Links +
10.86 cm, Links + 11.85 cm, Links
+ 12.49 cm, Links + 13.74 cm,
Links + 14.98 cm, Links

Gelöscht: ¶

804 Engel, A.: The role of transparent exopolymer particles (TEP) in the increase in apparent
805 particle stickiness (alpha) during the decline of a diatom bloom. J. Plankton Res., 22, 485-
806 497, 2000.

807
808 Engel, A.: Distribution of transparent exopolymer particles (TEP) in the northeast
809 Atlantic Ocean and their potential significance for aggregation processes Deep Sea Res.
810 I: Oceanographic Research Papers, 51, 1, 83-92, 2004,

811
812 Engel, A., Thoms, S., Riebesell, U., Rochelle-Newall, E. and Zondervan, I.: Polysaccharide
813 aggregation as a potential sink of marine dissolved organic carbon. Nature, 428, 929-932,
814 2004.

815
816 Engel, A., Abramson, L., Szlosek, J., Liu, Z., Stewart, G., Hirschberg, D. and Lee, C.:
817 Investigating the effect of ballasting by CaCO₃ in *Emiliania huxleyi*, II: Decomposition of
818 particulate organic matter. Deep-Sea Res. II, 56, 18, 1408-1419. DOI
819 10.1016/j.dsr2.2008.11.028, 2009.

820
821 Engel, A.: Determination of Marine Gel Particles, in: Practical Guidelines for the Analysis of
822 Seawater, edited, CRC Press, 2009.

823
824 Engel, A., Piontek, J., Grossart, H. P., Riebesell, U., Schulz, K. G. and Sperling, M.: Impact
825 of CO₂ enrichment on organic matter dynamics during nutrient induced coastal phytoplankton
826 blooms. J Plankton Res., 36, 3, 641-657. DOI 10.1093/plankt/fbt125, 2014.

827

Formatiert:
Absatz-Standardschriftart,
Schriftart: Times New Roman

Formatiert: Schriftart: Times New
Roman, Nicht Kursiv

Gelöscht: Engel, A. und Passow, U.:
Carbon and nitrogen content of
transparent exopolymer particles
(TEP) in relation to their Alcian
Blue adsorption. Mar. Ecol. Prog.
Ser., 219, 1-10,
doi:10.3354/meps219001, 2001.¶

Formatiert: Schriftart: Nicht Kursiv

Formatiert: Schriftart: Nicht Kursiv

Formatiert: Schriftart: Nicht Kursiv

836 | [Giering, S. L. C., Sanders, R., Lampitt, R. S., Anderson, T. A., Tamburini, C., Boutrif, M.,](#)
837 | [Zubkov, M. V., Marsay, C. M., Henson, S. A., Saw, K., Cook, K., and Mayor, D. J.](#)
838 | Reconciliation of the carbon budget in the ocean's twilight zone. *Nature*, 507, 480-483, 2014.
839
840 | Haake, B., Ittekkot, V., Ramaswamy, V., Nair, R. R., and Honjo, S. Fluxes of amino acids
841 | and hexosamines to the deep Arabian Sea, *Mar. Chem.*, 40, 291–314, 1992.
842
843 | Hansen, H.P, and Koroleff, F.: Determination of nutrients. In: *Methods of seawater analysis*.
844 | Grasshof, K. (ed), 159–228, 1999.
845
846 | Hecky, R.E., Mopper, K., Kilham, P., and Degens, E. T.: Amino acid and sugar composition
847 | of diatom cell-walls. *Mar. Biol.*, 19, 4, 323-331, doi: 10.1007/BF00348902, 1973.
848
849 | Heinze, C., Meyer, S.,; Goris, N., Anderson, L., Steinfeldt, R., Chang, N., Le Quere, C., and
850 | Bakker, D. C. E. : The ocean carbon sink - impacts, vulnerabilities and challenges. *Earth Syst.*
851 | *Dynamics*, 6, 1, 327-358, doi: 10.5194/esd-6-327-2015, 2015.
852
853 | Iversen, M. H., Nowald, N., Ploug, H., Jackson, G. A., and Fischer, G.: High resolution
854 | profiles of vertical particulate organic matter export off Cape Blanc, Mauritania: Degradation
855 | processes and ballasting effects. *Deep-Sea Res.*, 57, 6, 771-784,
856 | doi:10.1016/j.dsr.2010.03.007, 2010.
857 |
858 | Keil, R. G., Neibauer, J., Biladeau, C., van der Elst, K. and Devol A. H.: A multiproxy
859 | approach to understanding the “enhanced” flux of organic matter through the oxygen deficient
860 | waters of the Arabian Sea. *Biogeosciences*, 13, 2077-2092, 2016,
861 |

Gelöscht: Sarah L. C.

Gelöscht: ,

Gelöscht: ,

Gelöscht: ¶
Karstensen, J., Stramma, L., and
Visbeck, M. (2008) Oxygen minimum
zones in the eastern tropical Atlantic
and Pacific oceans, *Prog. Oceanogr.*,
77, 331–350.¶

Gelöscht: 5

872 Klaas, C., and Archer, D. E.: Association of sinking organic matter with various types of
873 mineral ballast in the deep sea: Implications for the rain ratio. Global Biogeochem. Cycles,
874 16(4), 1116, doi:10.1029/2001GB001765, 2002.

875

876 Knauer, G.A., J.H. Martin, and Bruland, K.W.: Fluxes of particulate carbon, nitrogen, and
877 phosphorus in the upper water column of the northeast Pacific. Deep-Sea Research, 26, 97-
878 108, 1979.

879

880 Kohlfeld, K. E., and Ridgwell, A. Glacial-Interglacial Variability in Atmospheric CO₂. In:
881 Surface Ocean–Lower Atmosphere Processes Geophysical Research Series, 187, American
882 Geophysical Union, doi:10.1029/2008GM000845, 2009.

883

884 Kwon, E. Y., Primeau, F., and Sarmiento J. L.: The impact of remineralization on the air-sea
885 carbon balance, Nat. Geosci., 2, doi:10.1038/ngeo612, 2009.

886

887 Lam, P. J., Doney, S. C. and Bishop J. K. B.: The dynamic ocean biological pump: Insights
888 from a global compilation of particulate organic carbon, CaCO₃ and opal concentrations
889 profiles from the mesopelagic. Global Biogeochem. Cycles., 25, GB3009, doi:
890 10.1029/2010GB003868, 2011.

891

892 Lampitt, R. S., Noji, T. and Von Bodungen, B.: What happens to zooplankton fecal pellets-
893 implications for material flux. Mar. Biol., 104, 1, 15-23, doi: 10.1007/BF01313152, 1990.

894

895 Le Moigne F., Sanders, R., Villa, M., Martin, A., Pabortsava, K., Planquette, H.,
896 Morris, P. J. and Thomalla, S., On the proportion of ballast vs non-ballast associated

Formatiert: Schriftart: Times New Roman, Nicht Kursiv, Englisch (USA)

Formatiert: Schriftart: Times New Roman, Nicht Kursiv, Englisch (USA)

Formatiert: Schriftart: Times New Roman, Nicht Kursiv, Englisch (USA)

Formatiert: Schriftart: Times New Roman, Nicht Kursiv, Englisch (USA)

Formatiert: Block, Zeilenabstand: Doppelt, Tabstopps: 1.25 cm, Links + 2.5 cm, Links + 3.75 cm, Links + 4.99 cm, Links + 6.24 cm, Links + 7.49 cm, Links + 8.74 cm, Links + 9.99 cm, Links + 11.24 cm, Links + 12.49 cm, Links + 13.74 cm, Links + 14.98 cm, Links

Formatiert: Schriftart: Times New Roman, Nicht Kursiv, Englisch (USA)

Formatiert: Schriftart: Times New Roman, Nicht Kursiv, Englisch (USA)

Formatiert: Schriftart: Times New Roman, Nicht Kursiv, Englisch (USA)

Formatiert: Schriftart: Times New Roman, Nicht Kursiv, Englisch (USA)

897 [sinking POC in the surface ocean. Geophys. Res. Letter. doi:10.1029/2012GL052980.](#)

898 [2012,](#)

899 ▼

900 Lee, C., and Cronin, C.: The vertical flux of particulate nitrogen in the sea: Decomposition of
901 amino acids in the Peru upwelling area and the equatorial Atlantic. J. Mar. Res. 40, 227-251,
902 1982.

903

904 Lee, C., and Cronin, C., Particulate amino acids in the sea: Effects of primary productivity
905 and biological decomposition. Journal of Marine Research 42, 1075-1097, 1984.

906

907 Lindroth, P., and Mopper, K.: High performance liquid chromatographic determination of
908 subpicomole amounts of amino acids by precolumn fluorescence derivatization with o-
909 phthaldialdehyde, Analytical Chemistry, 51(11), 1667-1674, 1979.

910

911 Löscher, C., Bange, H. W., Schmitz, R. A., Callbeck, C. M., Engel, A., Hauss, H., Kanzow,
912 T., Kiko, R., Lavik, G., Loginova, A. N., Melzner, F., Meyer, J., Neulinger, S. C., Pahlow,
913 M., Riebesell, U., Schunck, H., Thomsen, S. and Wagner, H.: Water column biogeochemistry
914 of oxygen minimum zones in the eastern tropical North Atlantic and eastern tropical South
915 Pacific Oceans. Biogeosciences (BG), 13 . pp. 3585-3606. DOI 10.5194/bg-13-3585-2016,
916 2016.

917

918 Mari, X.: Carbon content and C:N ratio of transparent exopolymeric particles (TEP) produced
919 by bubbling exudates of diatoms. Mar. Ecol. Progr. Ser., 183, 59-71, 1999.


920

Formatiert: Schriftartfarbe:
Schwarz, Englisch (USA)

Gelöscht: Le Moigne, F. A. C.,
Henson, S. A., Cavan, E., Georges, C.,
Pabortsava, K., Achterberg, E.P.,
Ceballos-Romero, E., Zubkov, M. and
Sanders, R. J.: What causes the inverse
relationship between primary
production and export efficiency in the
Southern Ocean?, Geophys. Res. Lett.,
doi:10.1002/2016GL068480, 2016.¶

930 Marsay, C. M., Sanders, R. J., Henson, S. S., Pabortsava, K., Achterberg, E. P., and Lampitt,
 931 R. S.: Attenuation of sinking particulate organic carbon flux through the mesopelagic ocean.
 932 Proc. Natl. Acad. Sci. U.S.A., 112, 4, 1089-1094, doi: 10.1073/pnas.1415311112, 2015.
 933
 934 Martin, P., Lampitt, R. S., Perry, M. J. Sanders, R. Lee, C., and D'Asaro, E.: Export and
 935 mesopelagic particle flux during a North Atlantic spring diatom bloom. Deep-Sea Research I
 936 58, 338–349, 2011.
 937
 938 Martin, J.H., G.A. Knauer, D.M. Karl and Broenkow W.W.: Vertex – Carbon Cycling in the
 939 Northeast Pacific. Deep-Sea Res. A, 34, 267-285, doi: 10.1016/0198-0149(87)90086-0, 1987.
 940
 941 Mortlock, R.A., and Froelich, P.N.: A simple method for the rapid determina- tion of biogenic
 942 opal in pelagic marine sediments. Deep-Sea Res., Part A, 36, 1415– 1426, 1989.
 943
 944 Pantoja, S., Sepúlveda, J. and González H. E.: Decomposition of sinking proteinaceous
 945 material during fall in the oxygen minimum zone off northern Chile. Deep Sea Research, I,
 946 51, 55-70, 2004.
 947
 948 Passow, U.: Transparent exopolymer particles (TEP) in aquatic environments, Progress in
 949 Oceanography, 55, 287-333, 2002.
 950
 951 Passow, U., Shipe, R. F., Pak, D. K., Brzezinski, M. A., & Alldredge, A. L.: Origin of
 952 transparent exopolymer particles (TEP) and their role in the sedimentation of particulate
 953 matter. Continental Shelf Research, 21, 327–346, 2000.
 954

955 Ploug, H., Kühl M., Buchholz-Cleven, B., and Jørgensen, B.B.: Anoxic aggregates - an
 956 ephemeral phenomenon in the pelagic environment? *Aquat. Microb. Ecol.*, 13, 285–294,
 957 1997.

958 | 

959 Ploug, H., and Bergkvist, J.: Oxygen diffusion limitation and ammonium production within
 960 sinking diatom aggregates under hypoxic and anoxic conditions. *Mar. Chem.* 176, 142-149,
 961 2015.

962

963 Schlitzer, R., Ocean Data View, <http://odv.awi.de>, 2013.

964

965 Schneider, B., Engel, A., and Schlitzer, R.: Effects of depth- and CO₂-dependent C:N ratios
 966 of particulate organic matter (POM) on the marine carbon cycle. *Global Biogeochemical*
 967 *Cycles*, 18, 2, doi:10.1029/2003GB002184, 2004.

968

969 Soutar, A., Kling, S. A., Crill, P. A., Duffrin, E., and Bruland K.W.: Monitoring the marine
 970 environment through sedimentation. *Nature*, 266, 136-139, 1977.

971

972 Stief, P., Kamp, A., Thamdrup, B., and Glud, R.N. : Anaerobic Nitrogen Turnover by Sinking
 973 Diatom Aggregates at Varying Ambient Oxygen Levels. *Frontiers in Microbiol.*, 7, 98 doi:
 974 10.3389/fmicb.2016.00098, 2016.

975

976 Stramma, L., Johnson, G. C., Sprintall, J., and Mohrholz, V.: Expanding Oxygen-Minimum
 977 Zones in the Tropical Oceans, *Science*, 320, 655-658, 2008.

978

979 Suess, E.: Particulate organic carbon flux in the oceans - surface productivity and oxygen
 980 utilization, *Nature* 288, 260–263, 1980.

Gelöscht: ¶
 Ploug, H., and B.B. Jørgensen (1999)
 A net-jet flow system for mass transfer
 and microelectrode studies in sinking
 aggregates. *Mar. Ecol. Prog. Ser.* 176,
 279-290. ¶

987

988 Torres Valdez, S., Painter, . C., Martin, A. P., Sanders, R. and Felden, J.: Data compilation of
 989 fluxes of sedimenting material from sediment traps in the Atlantic Ocean. Earth system
 990 Science Data, 6, 123-145, doi: 10.5194/essd-6-123-2014, 2014.

991

992 Van Mooy, B. A. S., Keil R. G., and Devol, A. H.: Impact of suboxia on sinking particulate
 993 organic carbon: Enhanced carbon flux and preferential degradation of amino acids via
 994 denitrification. Geochim. Cosmochim. Ac., 66, 457-465. doi: 10.1016/s0016-7037(01)00787,
 995 2002.

996

997 Visbeck M.: Oxygen in the Tropical Atlantic OSTRE Second Tracer Survey – Cruise. No.
 998 M105 – March 17 – April 16, 2014 – Mindelo (Cape Verde) – Mindelo (Cape Verde).
 999 METEORBerichte, M105, 49 pp., DFG-Senatskommission für Ozeanographie,
 1000 DOI:10.2312/cr_m105, 2014.

1001

1002 Volk, T., and Hoffert, M. I.: Ocean carbon pumps: Analysis of relative strengths and
 1003 efficiencies in ocean driven atmospheric CO₂ changes. In: Sundquist ET and Broecker WS
 1004 (eds.) The Carbon Cycle and Atmospheric CO₂: Natural Variations Archean to Present,
 1005 Geophysical Monograph Series, vol. 32, pp. 99-110. Washington, DC: American Geophysical
 1006 Union, 1985.

1007

1008 Weiner, S. and Erez, J.: Organic matrix of the shell of the foraminifer *Heterostegina*
 1009 *depressa*. Journal of Foraminifera Res., 14, 3, 206-212, 1984.

1010

1011 Welschmeyer, N. A.: Fluometric analysis of chlorophyll a in the resence of chlorophyll b and
 1012 pheopigments. Limnol. Oceanogr., 39, 1985-1992, 1994.

1013
1014
1015
1016
1017
1018
1019
1020

1021
1022
1023

Tables

Table 1: Fluxes of particulate components at 100m depth (F_{100}) and in the core of the OMZ at 400m (F_{OMZ}), as well as the associated attenuation coefficients (b -values) and transfer efficiencies (T_{eff} , %) over the depth range 100 to 600 m during two traps deployments in the ETNA. All units are in $mg\ m^{-2}\ d^{-1}$ except for TEP fluxes which is reported in total particle area $cm^{-2}\ m^{-2}\ d^{-1}$. Mean values and standard deviations (SD) were calculated from analytical replicates.

Component	F_{100}		F_{OMZ}		b -value		T_{eff} (%)		
	mean	SD	mean	SD	mean	SD	r^2	(600/100 m)	
Mass	I	249	48.9	141	6.8	-0.429	0.090	0.987	41
	II	231	16.3	141	12.1	-0.355	0.033	0.998	52
POC	I	69.4	9.23	23.8	5.4	-0.795	0.031	0.989	23
	II	76.3	8.43	28.1	3.0	-0.741	0.044	0.989	22
PN	I	11.9	1.29	2.76	0.46	-1.013	0.026	0.992	15
	II	13.5	1.12	3.26	0.19	-1.00	0.020	0.990	16
POP	I	0.71	0.07	0.15	0.02	-1.081	0.074	0.992	18
	II	0.64	0.03	0.22	0.02	-0.80	0.034	0.990	23
Opal	I	44.6	1.76	34.0	1.7	-0.0195	0.038	0.987	65
	II	48.6	4.16	30.7	2.0	-0.345	0.052	0.987	44
Chl a	I	0.10	0.00	0.035	0.001	-0.820	0.024	0.990	21
	II	0.12	0.01	0.053	0.005	-0.625	0.082	0.988	24
TEP	I	1650	548	119	36.8	-0.498	0.014	0.548	33
	II	2990	348	1644	95	-0.451	0.069	0.810	37
PHAA-C	I	3.21	-	3.71	0.47	-1.324	0.067	0.994	11
	II	1.28	0.10	5.24	0.79	-0.978	0.096	0.991	14

Gelöscht: —Seitenumbruch—
¶
Formatiert: Links, Tabstopps:
Nicht an 1.25 cm + 2.5 cm + 3.75
cm + 4.99 cm + 6.24 cm + 7.49
cm + 8.74 cm + 9.99 cm + 11.24
cm + 12.49 cm + 13.74 cm +
14.98 cm

1026
1027

Table 2: Composition [\(%Mol\)](#) and degradation index (DI) of PHAA collected at different depths during two trap deployments (#I, #II) in the ETNA region.

Gelöscht: (%Mol)

Depth(m)	AsX	GlX	Ser	Gly	Thr	Arg	Ala	GABA	Tyr	Val	Iso	Phe	Leu	DI
#I														
60	14.15	13.94	8.46	14.29	7.76	5.90	11.94	0.22	0.84	5.69	4.57	4.00	8.26	0.34
100	13.95	13.53	8.29	14.65	7.87	5.77	11.57	0.19	1.64	5.66	4.56	4.07	8.24	0.23
150	14.19	12.73	8.54	15.93	8.10	5.78	11.42	0.31	0.96	5.68	4.44	4.05	7.87	0.29
200	14.17	12.05	9.29	16.02	8.05	5.61	11.69	0.49	1.10	5.65	4.30	4.04	7.54	0.07
300	13.19	11.75	8.58	17.71	7.98	5.31	12.10	0.37	1.82	5.77	4.15	3.83	7.43	0.03
400	14.15	11.77	9.03	18.54	7.94	5.72	10.85	0.46	1.25	5.58	3.93	3.80	6.97	0.02
500	14.06	11.89	9.55	18.70	7.18	6.01	11.02	0.55	1.29	5.19	3.86	3.65	7.05	0.02
600	14.15	13.94	8.46	14.29	7.76	5.90	11.94	0.22	0.84	5.69	4.57	4.00	8.26	0.07
#II														
100	13.89	14.69	8.36	12.94	7.57	5.89	12.26	0.21	0.02	6.13	5.12	4.05	8.86	0.24
150	13.48	14.23	8.46	14.12	7.56	5.68	12.55	0.22	0.00	6.21	5.01	3.85	8.62	0.37
200	13.80	13.90	9.10	14.27	7.20	6.12	11.57	0.27	0.04	6.19	5.07	3.97	8.49	0.13
300	14.58	14.63	8.35	15.16	7.75	5.56	11.75	0.26	0.14	5.62	4.51	3.82	7.88	0.07
400	14.06	13.01	8.72	16.45	7.99	5.55	11.74	0.44	0.79	5.54	4.33	3.77	7.59	0.08
500	14.08	12.90	8.75	16.48	7.59	5.69	11.81	0.37	0.30	5.94	4.62	3.80	7.66	-0.09

600	13.62	12.55	9.16	17.02	7.95	5.75	11.23	0.42	0.38	5.87	4.61	3.88	7.55	-0.04
-----	-------	-------	------	-------	------	------	-------	------	------	------	------	------	------	-------

1029 **Figure captions:**

1030
1031
1032 Figure 1a-d: Map of the study area (A) and depth distribution of oxygen concentration (mol
1033 kg⁻¹) (B) in the Eastern Tropical North Atlantic (ETNA) during the RV Meteor 105 cruise,
1034 when two surface tethered drifting sediment traps (STDT) were deployed (C). Depth
1035 distribution of oxygen concentration (mol kg⁻¹) at stations visited in the deployment area
1036 showed an oxygen minimum zone in the upper mesopelagial (D).

1037
1038 Figure 2a-d: Fluxes of total mass (a) and particulate organic carbon (b; POC), particulate
1039 nitrogen (c; PN), and particulate organic phosphorus (d; POP) during the deployment of two
1040 STDT in the ETNA. Deployments: Solid symbols #I, open symbols #II.

1041
1042 Figure 3a-d: Fluxes of Chlorophyll *a* (a; Chl *a*), opal (b), TEP (c), and PHAA (d) during the
1043 deployment of two STDT in the ETNA. Deployments: Solid symbols #I, open symbols #II.

1044
1045 Figure 4: Changes in mineral ballast ratios of sinking particles with depth during the two
1046 deployments in the ETNA. Deployments: Black bars #I, grey bars #II.

1047
1048 Figure 5a-f: Changes in organic matter composition of particles sinking through the OMZ
1049 during the deployment of two STDT in the ETNA. Deployments: Solid symbols #I, open
1050 symbols #II.

1051
1052 Figure 6a-d: Molar percentages of selected amino acids contained in PHAA during the
1053 deployment of two STDT in the ETNA. Deployments: Solid symbols #I, open symbols #II.

1054

1055 Figure 7: Degradation index (DI) of organic matter in trap collected sinking particles based on
1056 amino acid composition and calculated after Dauwe et al. (1999). Deployments: Black bars
1057 #I, grey bars #II.

1058
1059

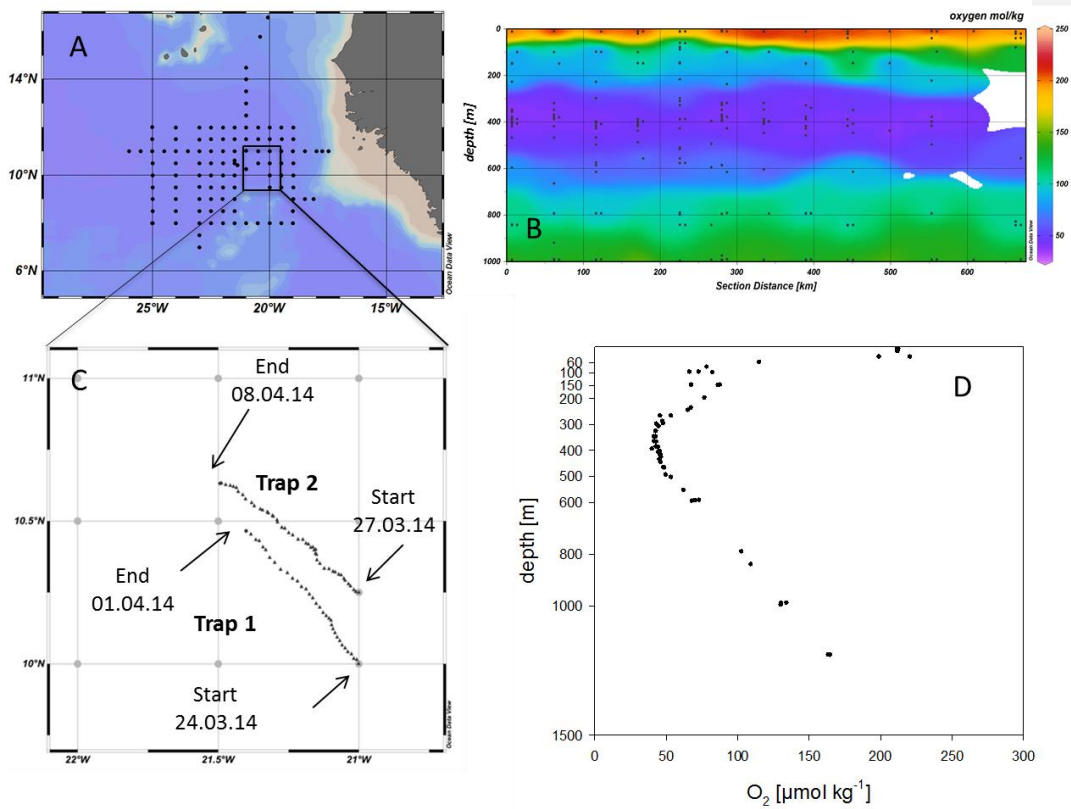
1060

Gelöscht: ¶
¶
¶

1064 **Figures**

1065

Gelöscht: ¶



1066

1067

1068

1069

1070

1071

Figure 1a-d

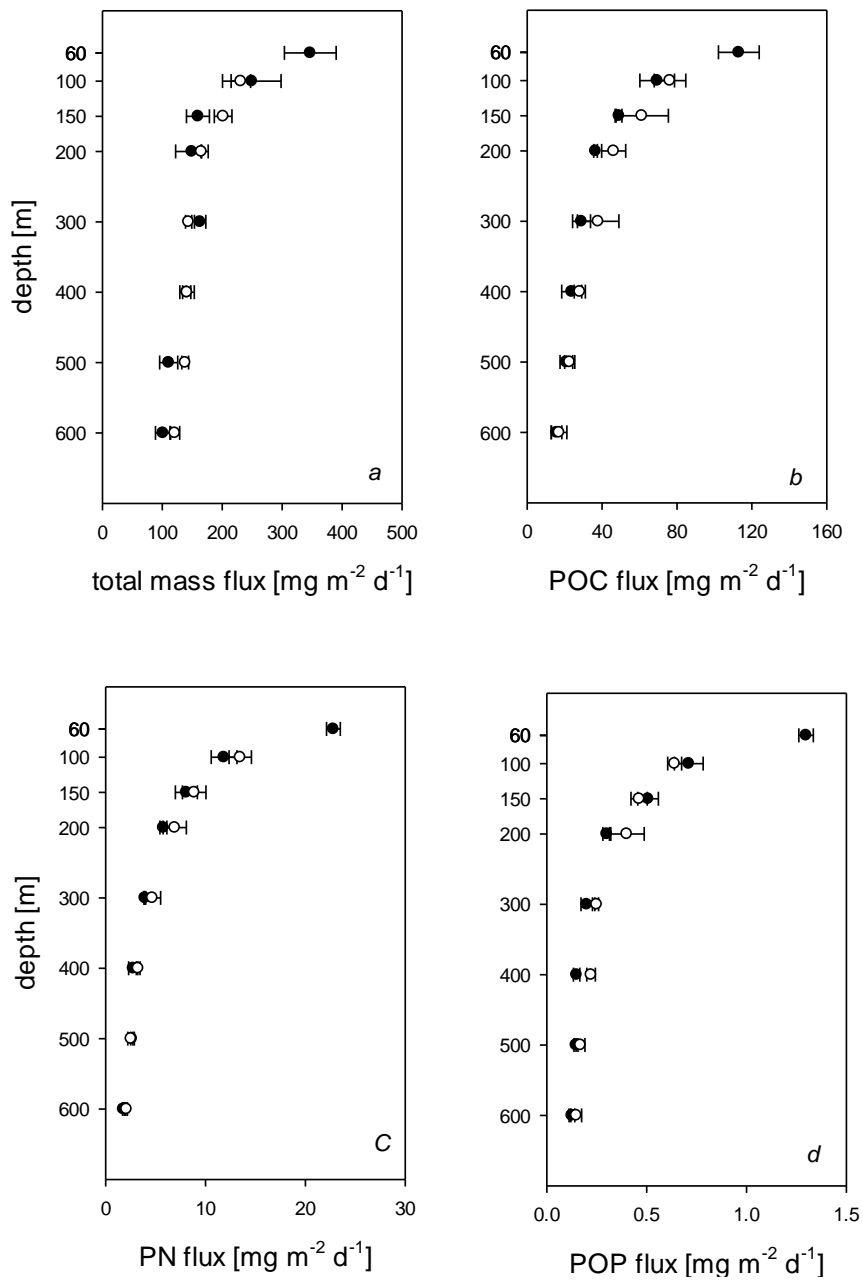


Figure 2a-d

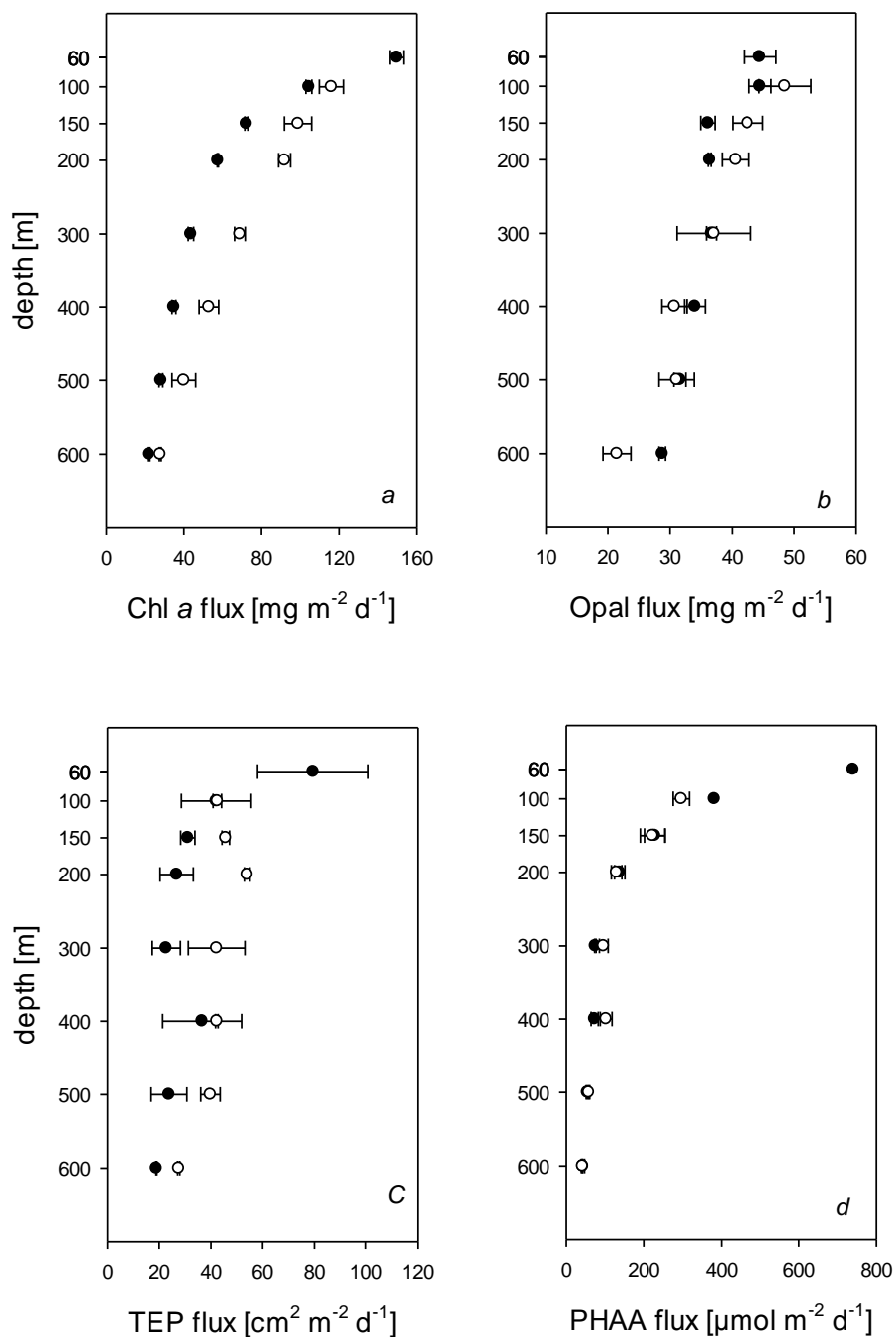


Figure 3a-d

1103
1104
1105
1106
1107
1108
1109
1110
1111
1112
1113
1114
1115
1116
1117
1118
1119
1120
1121
1122

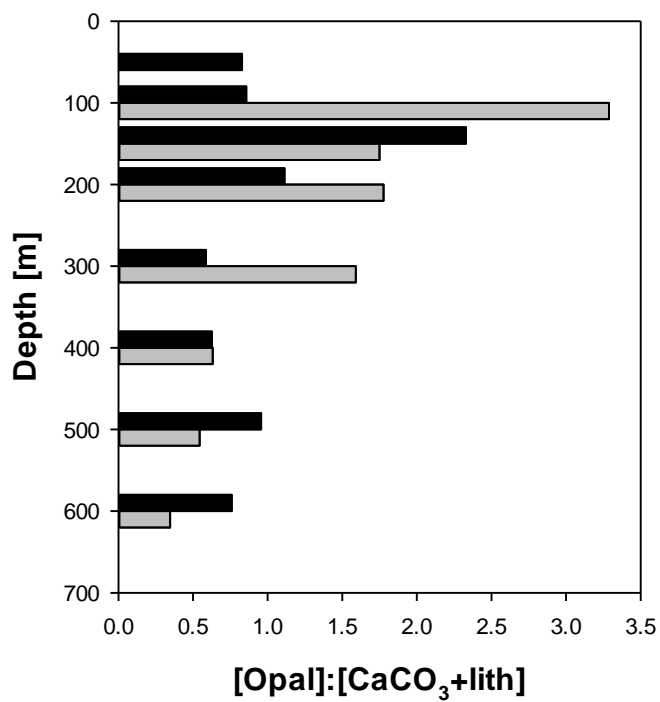


Figure 4

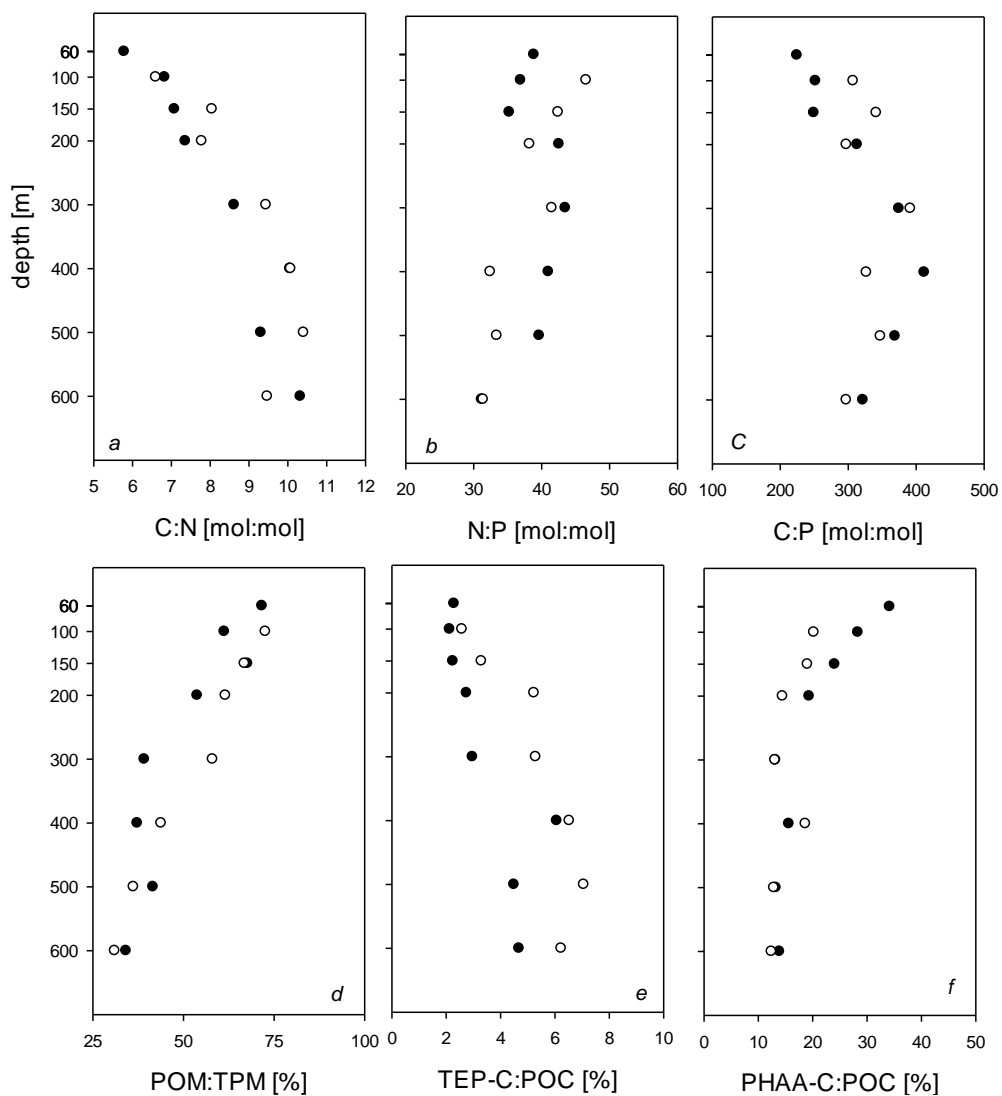


Figure 5a-f

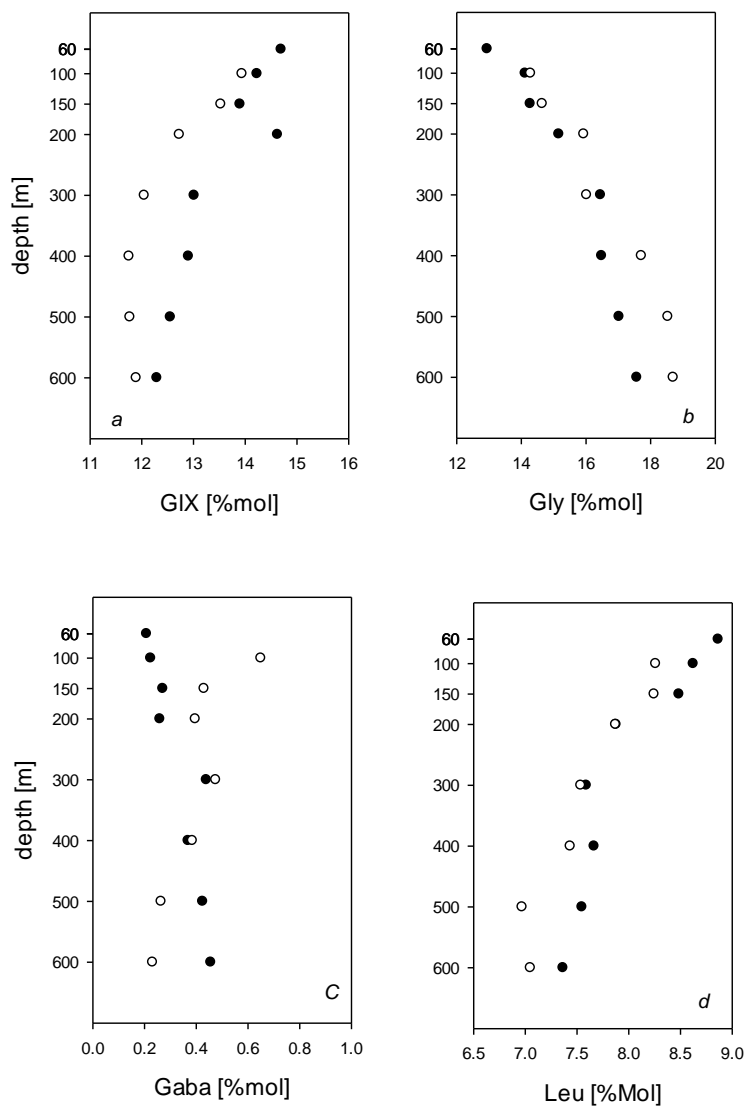
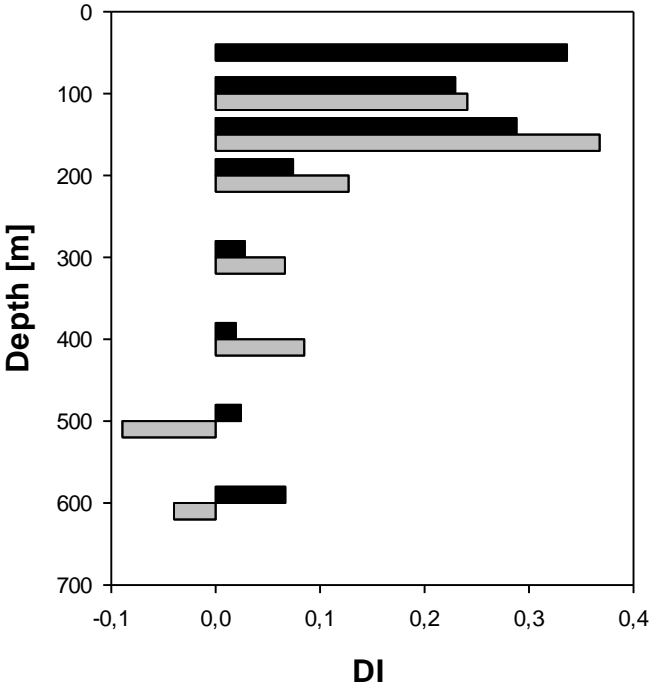


Figure 6a-d

1139

1140



1141

1142

1143

Figure 7