



## Process-based modelling of NH<sub>3</sub> exchange with grazed grasslands

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**Abstract.** In this study the GAG model, a process-based ammonia (NH<sub>3</sub>) emission model for urine patches was extended and applied for the field scale. The new model (GAG\_field) was tested over two modelling periods, for which micrometeorological NH<sub>3</sub> flux data were available. Acknowledging uncertainties in the measurements, the model was able to simulate the main features of the observed fluxes. The temporal evolution of the simulated NH<sub>3</sub> exchange flux was found to be dominated by NH<sub>3</sub> emission from the urine patches, offset by simultaneous NH<sub>3</sub> deposition to areas of the field not affected by urine. The simulations show how NH<sub>3</sub> fluxes over a grazed field in a given day can be affected by urine patches deposited several days earlier, linked to the interaction of volatilization processes with soil pH dynamics. Sensitivity analysis showed that GAG\_field was more sensitive to soil buffering capacity ( $\beta$ ), field capacity ( $\theta_{fc}$ ) and permanent wilting point ( $\theta_{pwp}$ ) than the patch scale model. This can be explained by the different initial soil pH and physical characteristics which determine the maximum volume of urine that can be stored in the NH<sub>3</sub> source layer. It was found that in the case of urine patches with a higher initial soil pH and higher initial soil water content, the sensitivity of NH<sub>3</sub> exchange to  $\beta$  was stronger. Also, in the case of a higher initial soil water content, NH<sub>3</sub> exchange was more sensitive to the changes in  $\theta_{fc}$  and  $\theta_{pwp}$ . The sensitivity analysis showed that the nitrogen content of urine ( $c_N$ ) is associated with high uncertainty in the simulated fluxes. However, model experiments based on  $c_N$  values randomized from an estimated statistical distribution indicated that this uncertainty is considerably smaller in practice. Finally, GAG\_field was tested with a constant soil pH of 7.5. The variation of NH<sub>3</sub> fluxes simulated in this way showed a good agreement with those from the simulations with the original approach, accounting for a dynamically changing soil pH. These results suggest a way for model simplification when GAG\_field is applied later for regional scale.

### List of Symbols

$A_{non}$ (m <sup>2</sup> )	Area of the field unaffected by urine (non-urine area)
$AD$ (ha <sup>-1</sup> )	Animal density
$A_{field}$ (m <sup>2</sup> )	Field area
$A_{patch}$ (m <sup>2</sup> )	Area of a urine patch



$B_{H_2O}$ (dm <sup>3</sup> )	Water budget in the source layer
$B_{H_2O(max)}$ (dm <sup>3</sup> )	Maximal water amount in the source layer
$B_{H_2O}^j$ (dm <sup>3</sup> )	Water budget in the source layer under the urine patches deposited in the $j^{\text{th}}$ time step
$c_N$ (g N dm <sup>-3</sup> )	N content of the urine
$c_N^{Ave}$	Average urinary N concentration in urine patches deposited in the same time step
$c_N^{Dil}$ (g N dm <sup>-3</sup> )	Urine N content after dilution in the soil
$c_N^k$ (g N dm <sup>-3</sup> )	Urinary N concentration in the $k^{\text{th}}$ urine patch
$d$ (m)	Displacement height
$D(c_N)$	Distribution function of urinary nitrogen content
$D_t$	Proportion of the urine-covered area over a $t$ time period on field if there is no overlap between the urine patches
$F_{non}$ (μg N m <sup>-2</sup> s <sup>-1</sup> )	Net NH <sub>3</sub> exchange flux over the non-urine area
$F_g$ (μg N m <sup>-2</sup> s <sup>-1</sup> )	NH <sub>3</sub> exchange flux over the ground
$F_{net}$ (μg N m <sup>-2</sup> s <sup>-1</sup> )	Net NH <sub>3</sub> exchange flux for the whole field
$F_{patch}^j$ (μg N m <sup>-2</sup> s <sup>-1</sup> )	NH <sub>3</sub> emission flux from the urine patches deposited in the $j^{\text{th}}$ time step
$F_t$ (μg N m <sup>-2</sup> s <sup>-1</sup> )	Total NH <sub>3</sub> exchange flux over the canopy
$F_\chi$ (μg N m <sup>-2</sup> s <sup>-1</sup> )	NH <sub>3</sub> exchange flux derived based on measurements with AMANDA
$h$ (m)	Canopy height
$H$ (J m <sup>-2</sup> s <sup>-1</sup> )	Sensible heat flux
$K$	Karman constant
$K$	Parameter representing the uniformity of the excretal distribution on a field
$L$ (m)	Monin-Obukhov length
$LAI$ (m <sup>2</sup> m <sup>-2</sup> )	Leaf area index
$n(t_j)$	Number of urine patches deposited in the $j^{\text{th}}$ time step
$N_t$	Total number of urine patches deposited over a $t$ time period on a field
$p$ (kPa)	Surface atmospheric pressure
$pH(t_0)$	Soil pH before urine patch deposition
$P$ (mm)	Precipitation amount
$PAR$ (μmol m <sup>2</sup> s <sup>-1</sup> )	Photosynthetically active radiation
$P_t$	Proportion of the field covered by urine patches after a $t$ time period
$Q$	Parameter in the calculation of $P_t$
$R_{ac}$ (s m <sup>-1</sup> )	Aerodynamic resistance in the canopy
$R_g$ (s m <sup>-1</sup> )	Resistance on the ground
$REW$ (mm)	Readily evaporable water in the soil
$R_{glob}$ (MJ m <sup>2</sup> h <sup>-1</sup> )	Global radiation / solar radiation
$RH$ (%)	Relative humidity
$Sens_{net}$ (%)	Sensitivity of the total NH <sub>3</sub> exchange over the whole field
$Sens_{patch}$ (%)	Sensitivity of the total NH <sub>3</sub> exchange over the urine patches on the field
$t_i, t_j$	$i^{\text{th}}$ and $j^{\text{th}}$ time steps



$T_{air}$ (°C)	Air temperature at 2 m
$T_{soil}$ (°C)	Soil temperature
$u$ (m s <sup>-1</sup> )	Wind speed
$u_{dir}$ (°)	Angle of the wind direction
$u_*$ (m s <sup>-1</sup> )	Friction velocity
$U_{add}$ (g N)	Urea added to the source layer
$UF$ (animal <sup>-1</sup> day <sup>-1</sup> )	Urination frequency
$W_{rain}$ (dm <sup>3</sup> )	Water input as rain water over the urine patch
$W_{urine}$ (dm <sup>3</sup> )	Volume of urine
$z$ (m)	Height of the NH <sub>3</sub> concentration measurements
$z_w$ (m)	Height of wind measurement
$\beta$ (mol H <sup>+</sup> (pH unit) <sup>-1</sup> dm <sup>-3</sup> )	Soil buffering capacity
$\beta_{patch}$ (mol H <sup>+</sup> (pH unit) <sup>-1</sup> )	Buffering capacity of the source layer
$\Gamma_g$	NH <sub>3</sub> emission potential on the soil surface
$\Gamma_{sto}$	NH <sub>3</sub> emission potential from the stomata
$\Delta z$ (mm)	Thickness of the source layer
$\theta(t_0)$ (m <sup>3</sup> m <sup>-3</sup> )	Soil volumetric water content before urine patch deposition
$\theta_{fc}$ (m <sup>3</sup> m <sup>-3</sup> )	Field capacity
$\theta_{por}$ (m <sup>3</sup> m <sup>-3</sup> )	Porosity
$\theta_{pwp}$ (m <sup>3</sup> m <sup>-3</sup> )	Permanent wilting point
$\theta_{urine}$ (m <sup>3</sup> m <sup>-3</sup> )	Proportion of the source layer that can be filled up by urine
$\sigma, \mu$	Scale parameters of the log-normal distribution
$\Sigma F_{net}$	Total NH <sub>3</sub> exchange over the grazed field
$\Sigma F_{non}$	Total NH <sub>3</sub> exchange over the non-urine area
$\Sigma F_{patch}$	Total NH <sub>3</sub> emission from the urine patches on a grazed field
$X$	Air concentration of NH <sub>3</sub> in the measurement heights of AMANDA
$\chi_a$ (μg N m <sup>-3</sup> )	Air concentration of NH <sub>3</sub> at 1 m height
$\chi_g$ (μg N m <sup>-3</sup> )	Compensation point on the ground
$\chi_p$ (μg N m <sup>-3</sup> )	Compensation point in the soil pores
$\chi_{z0}$ (μg N m <sup>-3</sup> )	Canopy compensation point
$\Psi_H$	Stability function for heat

## 1 Introduction

The global nitrogen (N) cycle has been substantially altered by the emission of reactive nitrogen compounds (N<sub>r</sub>), which is dominated by the emission of ammonia (NH<sub>3</sub>) (Galloway et al., 2008, Fowler et al., 2013). As a result of the strong emission of N<sub>r</sub>, five key environmental threats have been identified: water, air and soil quality, greenhouse balance and ecosystems

5 (Sutton et al., 2011). The main global source of NH<sub>3</sub> emission to the atmosphere is agriculture (EDGAR, 2011), specifically,



the breakdown of animal excreta and fertilizers containing ammonium ( $\text{NH}_4^+$ ). The volatilization of  $\text{NH}_3$  is dependent on meteorology, especially temperature (Flechar et al., 2013, Sutton et al., 2013), which raises the question: how will  $\text{NH}_3$  emission be influenced by climate change? A way to address this question and predict the environmental consequences is to design meteorology-driven  $\text{NH}_3$  emission models for each agricultural source (Sutton et al., 2013). This study represents a step

5 toward this goal by describing an  $\text{NH}_3$  exchange model for grazed fields, accounting for the relevant meteorological drivers. As confirmed by both laboratory and field studies (Farquhar et al., 1980, Sutton et al., 1995), the exchange of  $\text{NH}_3$  between surface and atmosphere is bidirectional. The direction of the net  $\text{NH}_3$  exchange is controlled by the difference in the relative magnitude of atmospheric  $\text{NH}_3$  concentration at two heights above the surface: the so-called ‘compensation point’ (atmospheric  $\text{NH}_3$  concentration right above the surface) and the ambient atmospheric  $\text{NH}_3$  concentration (high above the

10 surface). If the compensation point is the larger of the two  $\text{NH}_3$  is emitted to the atmosphere, whilst if the ambient air concentration is the larger, net deposition takes place, transferring  $\text{NH}_3$  to the surface. The state-of-the-art modelling technique for this bidirectional behaviour is the application of a ‘canopy compensation point’ model (Sutton et al., 1995, Nemitz et al., 2001, Burkhardt et al., 2009, Flechar et al., 2013). These models derive the net  $\text{NH}_3$  emission flux over a canopy by taking into account the  $\text{NH}_3$  exchange with the different sources and sinks within the canopy (e.g. stomata, leaf surface, soil, litter,

15 etc.) as well as the effect of meteorological variables and the canopy on these component  $\text{NH}_3$  fluxes. Over a grazed field the dominant source of  $\text{NH}_3$  is urine rather than dung (Petersen et al., 1998, Laubach et al., 2013). Therefore, the  $\text{NH}_3$  exchange over a grazed field is determined by two main components: the  $\text{NH}_3$  emission from the urine patches and the  $\text{NH}_3$  exchange with the area on the field that is not affected by urine (“non-urine area”). The GAG model (Generation of Ammonia from Grazing, Möring et al., 2016) is a special application of a canopy compensation point model that derives  $\text{NH}_3$

20 volatilization from a unit of  $\text{NH}_3$  source on a grazed field: a single urine patch. GAG calculates  $\text{NH}_3$  emission from a urine patch in a process-based way, simulating the total ammoniacal nitrogen (TAN) and water content under the urine patch as well as the evolution of soil pH. The present paper describes an extension of the GAG model, so that it accounts for the  $\text{NH}_3$  emission from all of the urine patches deposited over a time interval on a grazed field and the  $\text{NH}_3$  exchange with the non-urine area.

25 The primary goal of this model development was to construct a tool that can be used in further studies to gain insights on the effects of meteorological variables on  $\text{NH}_3$  emission from grazing. Furthermore, our aim was to design a model that can be applied to an atmospheric chemistry transport model. Such a model application would serve as a base for future research, investigating how altered climate can affect  $\text{NH}_3$  emission, dispersion and deposition on a larger scale, i.e., regional or global scale. Therefore, simplicity was a key aspect in the model development presented here, while taking into account physical and

30 chemical processes that can be relevant over these larger scales. In the following, firstly, the theoretical background of the field-scale model application is presented (Section 2). Secondly, the equations required for upscaling to a field are provided, as well as the data used in the model evaluation and the methods applied in the sensitivity analysis are introduced (Section 3). This is followed by presentation of the model simulations for two



experimental periods and the outcomes of the sensitivity analysis (Section 4). Finally, we conclude the paper with the discussion of the results and our conclusions (Section 5 and 6).

## 2 Theoretical background

### 2.1 Description of the GAG model

5 The GAG model (Móring et al., 2016) is a process-based  $\text{NH}_3$  emission model for a single urine patch that is capable of simulating the driving soil chemistry by accounting for: the TAN and the water content of the soil under the urine patch (in Fig. 1. TAN budget and water budget, respectively) and the variation of soil pH ( $\text{H}^+$  ion budget in Fig. 1). The TAN and the  $\text{H}^+$  ion budgets are controlled by the hydrolysis of the urea content of urine, as well as the  $\text{NH}_3$  emission from the soil. In the water budget, apart from the liquid content of the incoming urine, precipitation acts as a source term, whilst soil evaporation  
10 is considered as the only sink term.

The GAG\_model is a single layer model, which means that the effective  $\text{NH}_3$  emission occurs only from the urine that a thin top soil layer, the so-called “ammonia source layer” can hold. For further details on the GAG model, see the description in Móring et al. (2016). Hereafter, the original GAG model for patch scale and its extended version for field scale are referred to as GAG\_patch and GAG\_field, respectively.

### 15 2.2 Assumptions for the model application at field scale

Among all the naturally varying factors related to urination events during grazing, the following subsections describe those that are likely to be the most relevant from the point of view of  $\text{NH}_3$  exchange over a grazed field. Firstly, the possible overlap of the patches is examined (Section 2.2.1), then further parameters are discussed that can vary among urination events, such as the area of the patches ( $A_{patch}$ ), the frequency of urination events ( $UF$ ) and the nitrogen content of urine ( $c_N$ ) (Section 2.2.2).  
20 Finally, model assumptions for calculating the total  $\text{NH}_3$  net flux for the field are identified (Section 2.2.3).

#### 2.2.1 Exclusion of the overlap of the urine patches

According to observations (e.g. Betteridge et al., 2010, Moir et al., 2011, Dennis et al., 2013), urine patches over a grazed paddock may overlap. It was found that the overlap can have a large effect on N leaching (Pleasants et al., 2007, Shorten and Pleasants, 2007); however, no studies are available that investigate the effect of overlap in particular on  $\text{NH}_3$  emission from  
25 urine patches.

It is reasonable to assume that the emission flux from the area of the overlap will differ from both the previously and the newly deposited patches due to the differences in the soil chemical properties (Fig. 2). Since urea hydrolysis is in a different stage in the two urine patches, the soil chemistry under them will be different, and their mixture under the overlap is likely to result in a third, different chemical composition. In addition, if patches partly cover each other, the total source area will be smaller  
30 than if they were completely separate, which may influence the total  $\text{NH}_3$  emission from the field. Therefore, it is likely that



the possible overlap of the patches affects  $\text{NH}_3$  emission. However, to predict in every time step of the model which patches will cover each other, and what size the overlap will be, is very difficult. Thus, it would be preferable to neglect the overlap of the patches. To assess the resulting error arising from such a simplification, the difference in the field proportion covered by urine patches was investigated between the two cases: when overlap is assumed and when it is excluded.

- 5 A way to estimate the temporal evolution of the urine-covered proportion of the field is to use a negative binomial distribution function as suggested by Petersen et al. (1956), or the Poisson distribution tested by Romera et al. (2012). Based on the distribution suggested by Petersen et al. (1956), Pakrou and Dillon (2004) determined the proportion of the paddock covered by urine patches ( $P_t$ ) after a  $t$  time period as:

$$P_t = 1 - q^{-K}, \quad (1)$$

- 10 where  $K$  is a parameter that represents the uniformity of the excretal distribution. Following Pakrou and Dillon (2004), a representative value of  $K=7$  was used. The value of  $q$  is calculated as:

$$q = \frac{D_t + K}{K}, \quad (2)$$

in which  $D_t$  is the proportion of the urine-covered area over a  $t$  time period if there is no overlap (Eq. 3), i.e. the total number of the patches ( $N_t$ ) deposited over  $t$  multiplied by  $A_{\text{patch}}$  and divided by the field area ( $A_{\text{field}}$ ).

$$15 \quad D_t = \frac{N_t A_{\text{patch}}}{A_{\text{field}}} \quad (3)$$

Using  $D_t$ , Romera et al. (2012) derived  $P_t$  assuming a Poisson distribution as follows:

$$P_t = 1 - e^{-D_t}, \quad (4)$$

where  $e$  is Euler's constant ( $\sim 2.718$ ).

- 20 To investigate the highest possible difference that the exclusion of overlap can cause, in the following calculation a “worst case scenario” was assumed with the highest possible coverage by urine, i.e. the highest realistic animal density over a field, the largest  $A_{\text{patch}}$  and the highest  $UF$ . The ranges of all these parameters are listed in Table 1 for sheep and cattle, together with their references.

- According to the agricultural statistics of the European Commission for 2010 (EC, 2015), the maximal grazing animal densities on the agricultural holdings Europe-wide were higher than 10 LSU  $\text{ha}^{-1}$  (where LSU stands for livestock unit, which equals to 25 1 dairy cow or 10 sheep). Since no higher values than 10 were identified, 10 LSU  $\text{ha}^{-1}$  was assumed as the maximum. The value of  $N_t$  was calculated as the product of animal density over a hectare ( $A_{\text{field}} = 10\,000 \text{ m}^2$ ) and the maximum daily  $UF$  (urination events per animal per day, Table 1).

- Fig. 3 shows  $P_t$ , using the two different equations, Eq. (1) and (4). These results are very close to each other, with slightly smaller values from Eq. (1). Therefore, for further investigation the  $P_t$  values from Pakrou and Dillon (2004) (Eq. 1) were 30 taken and compared with the no overlap case ( $P_t = D_t$ ). In the case of sheep (Fig. 3a), the difference between  $P_t$  and  $D_t$  became



higher than 5% after the eighth day (and exceeds 10% after the 16<sup>th</sup> day – not shown here), whilst in the case of cattle (Fig. 3b) the same occurred after the 17<sup>th</sup> day.

The great majority of NH<sub>3</sub> is emitted in the first 8 days after the deposition of a urine patch (Sherlock and Goh, 1985). This means that after the eighth day the NH<sub>3</sub> exchange flux over the urine patches will be very close to that of the unaffected area of the field. Presumably, (as suggested by the model results in Móring et al. 2016) at this stage the chemical composition of the soil solution in the source layer under these patches will be also close to that of the initial, unaffected soil. Thus, practically, the patches deposited eight or more days before the given time step can be treated as part of the unaffected area of the field, or in other words, these patches disappear from the field. As a consequence, the total area of the patches grows in the first eight days, then it remains constant while the animals are on the field. Therefore, the probability of overlap after the eighth day will be the same as on the eighth day, since the total area of the patches prone to overlap with the new patches does not change after the eighth day.

Finally, it has to be noted that the results in Fig. 3 illustrate an extreme situation (the “worst case scenario”), and in reality  $P_t$  is much likely to grow rather more slowly. This allows a longer time before the exceedance of the 5% difference in  $P_t$  between the overlap and no-overlap case. Hence, for field-scale application of GAG the effect of overlap between the patches was concluded to be negligible, assuming completely separated urine patches in every time step.

### 2.2.2 Assumptions for $A_{patch}$ , $UF$ and $c_N$

As shown in the previous subsection, the parameters that regulate the extent of the field covered by urine are (i) the number of the animals on the field, (ii)  $A_{patch}$  and (iii)  $UF$ . The first parameter at a field-scale model application is easy to obtain, but the observations of the area of every single urine patch, as well as the number of urinations on an hourly basis, are rather difficult (see the overview of the observation techniques in Dennis et al., 2013).

Therefore, for GAG<sub>field</sub>, a constant  $A_{patch}$  for every individual urination event and a constant  $UF$  were assumed. There are values reported for  $A_{patch}$  in the literature (Table 1), whose average was used in the baseline simulations and with a sensitivity test an estimation was given for the uncertainty resulting from this simplification (Section 4.2.4).

In the literature observational data can be also found for  $UF$  (as shown in Table 1), but the temporal resolution of these data is usually a day. Based on personal communication with farmers, the hourly number of urine patches deposited over a field varies between the grazing and rumination periods and also between day and night. However, for the current modelling study an even distribution of urination events was assumed over the day, dividing the reported average daily  $UF$  by 24 hours. As for  $A_{patch}$ , a sensitivity analysis was carried out for this parameter as well (Section 4.2.4).

Another feature of the individual urination events that strongly influences the subsequent NH<sub>3</sub> volatilization is  $c_N$ . This parameter ranges widely (2 – 20 g N dm<sup>-3</sup>, Whitehead, 1995), not just amongst different animals, but also for different urination events by the same animal (Betteridge et al., 1986, Hoogendoorn et al., 2010). In the baseline simulation a constant average N content was applied. In Section 4.2.4, the response of the model was analysed to this choice of  $c_N$  and also to the uncertainty originating from the temporal variation of this parameter.





### 2.2.3 Assumptions for the calculation of the net NH<sub>3</sub> flux

With all the above assumptions, two types of area can be distinguished over a grazed field: (a) area covered by urine, and (b) area that is not affected by urine, referred to hereafter as “non-urine area” (as shown on Fig. 4). Therefore, it was assumed that the total flux over the field is the sum of the emission from the urine affected area (calculated by GAG) and the exchange with the non-urine area (derived by GAG, assuming constant emission potentials, as explained later, in Section 3.1). Since a grazed field, due to the urine patches, is not a uniform source of NH<sub>3</sub>, an error of the estimation of the total NH<sub>3</sub> flux can originate from the exclusion of the horizontal advection. Although this could be explored by using a dispersion model, since the purpose of this work is to construct a model that can be applied for regional scale, the model was kept at this, lower level of complexity. Finally, the field was assumed to have spatially homogenous physical and soil chemical properties before urine application. This assumption in tandem with the exclusion of the overlap of the urine patches and the horizontal dispersion of NH<sub>3</sub>, leads to the consequence that the total flux over the field is independent of the placement of the patches on the surface.

## 3 Material and methods

### 3.1 Model equations for the field-scale application

Based on the considerations outlined in the previous subsections, for GAG<sub>field</sub> we assumed that physically and chemically identical urine patches are deposited in every time step over the modelling period. To capture the effect of all of the urine patches, in calculating the net NH<sub>3</sub> flux for the whole field ( $F_{net}$ ), an  $n \times n$  matrix can be considered (see Fig. 5, where  $n$  is the number of the time steps in the modelling period). In this matrix  $i$  index denotes the time step for which the given flux is derived and  $j$  shows the time step when the patches were deposited. In this way,  $F_{net}$  in the  $i^{\text{th}}$  time step ( $t_i$ ) can be expressed by Eq. (5).

The first term in the numerator of Eq. (5) represents the NH<sub>3</sub> emitted by the non-urine area: the NH<sub>3</sub> exchange flux over the non-urine area ( $F_{non}$ ) multiplied by the size of this area ( $A_{non}$ ). While the second term in the numerator equals to the total NH<sub>3</sub> emitted from the urine patches, where  $F_{patch}^j$  is the emission flux from the urine patches deposited in the  $j^{\text{th}}$  time step, and  $n(t_j)$  is the number of the patches deposited in the same time step. To calculate  $F_{net}$ , the sum of the two has to be divided by  $A_{field}$  (Eq. 5).

$$F_{net}(t_i) = \frac{F_{non}(t_i)A_{non}(t_i) + \sum_{j=1}^n F_{patch}^j(t_i)n(t_j)A_{patch}}{A_{field}} \quad (5)$$

In the non-urine area, in the absence of any considerable nitrogen input, the soil chemistry is practically undisturbed. Thus, for the non-urine area a modified version of GAG<sub>patch</sub> was applied in which constant soil chemistry was assumed. Based on this,  $F_{non}$  was derived in the same way as the net NH<sub>3</sub> flux ( $F_r$ ) in GAG<sub>patch</sub> described by Eq. (1)-(7) in Möring et al. (2016), together with the following simplifications:





- Since over the non-urine area the dynamic simulation of soil chemistry is not needed, the original version of the two-layer canopy compensation point model by Nemitz et al. (2001) is used. This includes only the original compensation point on the ground ( $\chi_g$ ), instead of the soil resistance and compensation point in the soil assumed for GAG\_patch. As a consequence, for the non-urine area the equation in GAG\_patch for the  $\text{NH}_3$  emission from the soil ( $F_g$ ) changes to:

$$F_g = \frac{\chi_g - \chi_{z_0}}{R_{ac} + R_{bg}}, \quad (6)$$

where  $\chi_{z_0}$  represents the canopy compensation point, and  $R_{ac}$  and  $R_{bg}$  stand for the aerodynamic resistance within the canopy and the quasi-laminar resistance at the ground, respectively. For the parametrization of these variables in GAG\_patch see Möring et al. (2016).

- The value of  $\chi_g$  (Eq. 7) for the non-urine area was calculated similarly to that of the compensation point in the soil pore in GAG\_patch ( $\chi_p$ ), except that the  $\text{NH}_3$  emission potential for the ground ( $\Gamma_g$ ) was handled as a constant (Section 3.2.3) instead of being modelled dynamically as in GAG\_patch in the soil pore. In Eq. (7)  $T_{soil}$  represents the soil temperature.

$$\chi_g = \frac{161500}{T_{soil}} \times \exp\left(\frac{-10380}{T_{soil}}\right) \times \Gamma_g \quad (7)$$

- Since over the non-urine area no N input is assumed, for the emission potential of the stomata ( $\Gamma_{sto}$ ), instead of applying a decay function, like in GAG\_patch, it was treated as constant (Section 3.2.3).

The size of  $A_{non}$  in the given  $t_i$  time step is the area of the field that is not covered by any urine patches:

$$A_{non}(t_i) = A_{field} - \sum_{j=1}^i n(t_j) A_{patch}, \quad (8)$$

where  $n(t_j)$  (Eq. 9) is the number of the urine patches deposited in the  $j^{\text{th}}$  (hourly) time step. This can be expressed as the product of the animal density on the field in  $t_j$  ( $AD(t_j)$ , animals  $\text{ha}^{-1}$ ),  $A_{field}$  (ha) and the daily  $UF$  (urinations  $\text{day}^{-1}$  animal $^{-1}$ ), divided by 24 hours.

$$n(t_j) = \frac{(AD(t_j) \times A_{field} \times UF)}{24} \quad (9)$$

Finally,  $F_{patch}^j(t_i)$  was determined by Eq. (10), which expresses that before the deposition of the urine patch, the area is handled as non-urine area (first condition), and afterwards GAG\_patch calculates the patch emission ( $F_t(t_i)$ , second condition).

$$F_{patch}^j(t_i) = \begin{cases} F_{non}(t_i) & \text{if } i < j \\ F_t(t_i) & \text{otherwise} \end{cases} \quad (10)$$



When calculating  $F_i(t_i)$  a slight modification is also required, regarding the urea added with a single urination ( $U_{add}$ ). At field scale it has to be considered that during the modelling period urine patches may be deposited at the same time as a rain event occurs. A rain event

- i) will dilute the incoming urea solution, and
- 5 ii) may lead to the maximal water content ( $B_{H_2O}(max)$ ) in the  $NH_3$  source layer, which in the model formulation presented by Möring et al. (2016) prevents infiltration, resulting in no N input to the system and consequently no  $NH_3$  emission.

To address the first point, it has to be noted that although over the non-urine area GAG\_field does not simulate the dynamic, temporal evolution of the TAN budget and the soil pH (a constant  $\Gamma_g$  is used as noted above), it does account for the changes in water budget ( $B_{H_2O}$ ) in the source layer. Therefore, the water budget calculated by the GAG\_patch model modified for the non-urine area right before the  $j^{th}$  patch deposition ( $B_{H_2O}^j(t_i = (j - 1))$ ) can be updated by GAG\_patch in the next time step ( $B_{H_2O}^j(t_i = j)$ ). Although the effect of dilution is treated in GAG\_patch, it is defined only for the first time step, when urine is applied to the surface. Therefore, in the field-scale model  $U_{add}$  was calculated for the patch deposited in  $t_j$  as:

$$U_{add}(t_j) = c_N^{Dil}(t_j) \left( B_{H_2O}^j(t_{i=(j)}) - B_{H_2O}^j(t_{i=(j-1)}) \right) \quad (11)$$

where the diluted N concentration in the mixture of rain water and urine ( $c_N^{Dil}$ , Eq. 12) equals to the total amount of N in the urine ( $c_N \times W_{urine}$ ) divided by the sum of the volume of the liquid phase ( $W_{urine} + W_{Rain}(t_i = j)$ ), where  $W_{rain}$  denotes the volume of the infiltrating rain water).

$$c_N^{Dil}(t_j) = \left( \frac{c_N W_{urine}}{W_{urine} + W_{Rain}(t_{i=j})} \right) \quad (12)$$

To avoid the possible error resulting from the second point, it was assumed that the minimum amount of urine that is always allowed to penetrate to the source layer equals to 5% of  $B_{H_2O}(max)$ :

$$20 \left( B_{H_2O}^j(t_{i=(j)}) - B_{H_2O}^j(t_{i=(j-1)}) \right) \geq 0.05 \times B_{H_2O}(max) \quad (13)$$

### 3.2 Dataset used in the baseline simulations and model evaluation

#### 3.2.1 Measurements

GAG\_field was evaluated (Section 4.1) using measurements taken at a grassland site near Easter Bush, UK (see the field specific data in Table 2) by CEH (Centre for Ecology & Hydrology). The field is divided into two halves, the North Field and the South Field, and the instruments were placed on the boundary of the two (Fig. 6). For the site,  $NH_3$  flux measurements are available for a number of years (2001-2007). These fluxes were derived using the aerodynamic gradient method, which calculates the fluxes ( $F_\chi$ ) based on measurements of the vertical gradient of  $NH_3$  air concentration and micrometeorological variables (Eq. 14). In Eq. (14)  $\chi$  denotes the  $NH_3$  air concentration measured at a  $z$  height, whilst  $k$ ,  $u^*$ ,  $d$ ,  $\Psi_H$  and  $L$  stand for the Karman constant, friction velocity, displacement height, the stability function for heat, and the Monin-Obukhov length, respectively.



$$F_{\chi} = -ku_* \frac{\partial \chi}{\partial \left[ \ln(z-d) - \Psi_H \left( \frac{z-d}{L} \right) \right]}, \quad (14)$$

Ammonia concentration measurements were conducted by using a high-resolution NH<sub>3</sub> analyser, AMANDA (Ammonia Measurement by ANnular Denuder sampling with online Analysis, Wyers et al., 1993). During the sampling, gaseous NH<sub>3</sub> is captured in a continuous flow rotating annular wet denuder applying a stripping solution of 3.6 mM sodium hydrogen sulphate (NaHSO<sub>4</sub>). The technique determines the air concentration of NH<sub>3</sub> online by conductivity detection (Milford et al., 2001). The concentration gradients were obtained from concentration measurements at three heights: 0.44, 0.96 and 2.06 m.

The meteorological input variables that are required for a simulation with GAG\_field are the same as for GAG\_patch. From these, air and soil temperature ( $T_{air}$  and  $T_{soil}$ ), relative humidity ( $RH$ ), precipitation ( $P$ ), atmospheric pressure ( $p$ ), global radiation ( $R_{glob}$ ), wind speed ( $u$ ), wind direction ( $u_{dir}$ ) and sensible heat flux ( $H$ ) were observed at Easter Bush. For further details on instrumentation see Milford et al. (2001). Since photosynthetically active radiation ( $PAR$ ,  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) was not measured at the site, it was calculated from  $R_{glob}$  as shown in Eq. (15). According to Emberson et al. (2000),  $PAR$  is 45-50% of  $R_{glob}$  (0.475 in Eq. 15), and it is expressed in  $\mu\text{mol m}^{-2} \text{s}^{-1}$  (to the unit of  $R_{glob}$ ,  $\text{Wm}^{-2}$ , a conversion factor of 4.57 should be applied).

$$PAR = R_{glob} \times 0.475 \times 4.57 \quad (15)$$

15

### 3.2.2 Processing of the measured data for model application

For the baseline simulation and model evaluation (Section 4.1), a subset of the measurement data for 2001-2007 was selected that fulfilled the following criteria:

1. there were animals on the field;
2. grazing started at the beginning of the modelling period;
3. there had been no grazing, fertilizer spreading or grass cutting in the week before the grazing started;
4. there are no significant gaps in the meteorological input data;
5. flux measurements are available for validation.

The second criterion is important because NH<sub>3</sub> fluxes over the field can be affected by emission from urine patches deposited earlier. If the model does not account for these, it may underestimate the fluxes. The management practices listed in the third criterion can also affect the NH<sub>3</sub> exchange in a given time step, as well as fertilization can considerably affect the chemical balance of the soil. The latter would conflict with the model assumption that urine patches are deposited to a non-affected soil. The fourth criterion is necessary, because a continuous input dataset is needed for a simulation, since within GAG\_patch the TAN, the water and the H<sup>+</sup> budgets in a given time step are dependent on the values in the previous time.

As a result of the filtering, two suitable time periods were found: 26/08/2002 00:00 - 03/09/2002 06:00 and 20/06/2003 00:00 - 25/06/2003 05:00. These periods are referred herby to as P2002 and P2003, respectively. In both time intervals cattle were grazing on the South Field. Their number over the two modelling periods is indicated in Table 2.



To prepare the measured datasets for the hourly model application, firstly, the flux measurements were assessed for stability of the AMANDA instrumentation record with periods of obvious instrument malfunction and gaps in data removed (Móring, 2016). All data were then averaged to an hourly time resolution. The time resolution of the ambient air concentration ( $\chi_a$ ),  $u$ ,  $T_{air}$  and  $F_\chi$  (all at 1 m height) as well as  $T_{soil}$  was 15 minutes, whilst it was 30 minutes for  $p$ ,  $R_{glob}$  and  $RH$ . Secondly, in the

5 resulted averaged time series (except in  $F_\chi$ ) gap-filling was carried out. Data were missing from the  $\chi_a$  dataset for the simulation for P2002:

- over 27/08 13:00 – 28/08 13:00,
- on 02/09 at 23:00,
- and over 03/09 13:00 – 17:00.

10 The individual gap was interpolated from the values from the previous and next time step, whilst over the longer periods of missing data in  $\chi_a$  (25 and 5 consecutive hourly time steps), the values were assumed to be zero. In P2003 a single, hourly wind speed was missing at 01:00 on 25/06, which was interpolated based on the data in the neighbouring two time steps.

In the third step of data processing, the measured fluxes were filtered according to the wind direction. As mentioned above, animals were grazing on the South Field and the fluxes were measured at the border line of the two fields (Fig. 6). Therefore,

15 to distinguish the fluxes over the investigated part of the field, only the fluxes were used in the comparison that were associated with wind from the direction of the South Field, between  $135^\circ$  and  $315^\circ$ . The wind blew from this direction in most of the time. In the two modelling periods in P2002 and P2003 the wind direction was the opposite in the 7% and 15% of the hourly time steps, respectively.

In addition, although the  $NH_3$  concentrations measured in the time steps with  $u_{dir}$  from the North Field represents the

20 concentration in the North Field, in order to keep the continuity in the input data, these values were kept in the dataset. If they were substituted with zeros (similarly as it was handled in the gap-filling of  $\chi_a$ ), another type of error would have been added to the input data. Considering the relatively small number of  $u_{dir}$  values from the direction of the North Field, this choice is not anticipated to result in large errors in the  $NH_3$  flux simulations.

### 3.2.3 Model constants

25 The main urine-patch-specific constants defined by Móring et al., (2016) for GAG\_patch, are the soil buffering capacity ( $\beta = 0.021 \text{ mol H}^+(\text{pH unit})^{-1} \text{ dm}^{-3}$ ) and the thickness of the  $NH_3$  source layer ( $\Delta z = 4 \text{ mm}$ ), were not changed in the model experiments with GAG\_field. The other field, urine and site specific constants together with their sources are listed in Table 2.

For the constant  $F_{sto}$  for the non-urine area of the field, the values from the emission potential inventory by Massad et al. (2010)

30 for unfertilized grasslands were averaged. Since in the referenced inventory there were no  $F_g$  estimates for non-fertilized grasslands, it was defined during preliminary simulations with GAG\_field over a time interval when the grassland was not disturbed by any kind of management practice (grazing, fertilizer spreading or grass cutting). The time period of 01/06/2003 00:00 – 09/06/2003 00:00 fulfilled this criteria. These preliminary model experiments indicated a close agreement between the



measured and simulated  $\text{NH}_3$  fluxes with a  $\Gamma_g$  of 3000. Therefore, this value of  $\Gamma_g$  was applied in the baseline simulations with GAG\_field.

### 3.3 Methods used in the sensitivity analysis

#### 3.3.1 Perturbation experiments

5 Similarly to the model perturbation experiments carried out with GAG\_patch (Móring et al., 2016), a sensitivity analysis of GAG\_field to the regulating model parameters (Section 4.2.1-4.2.3) was performed. In addition to the parameters that were investigated for GAG\_patch ( $\Delta z$ ,  $\beta$ ,  $REW$  – readily evaporable water,  $\theta_{fc}$  – field capacity,  $\theta_{pwp}$  – permanent wilting point),  $\Gamma_{sto}$ ,  $\Gamma_g$ , and  $\text{pH}(t_0)$  (soil pH before urine deposition) were also examined. The value of  $\theta_{fc}$  and  $\theta_{pwp}$  express the maximum and the minimum volumetric water content in the soil, including the  $\text{NH}_3$  source layer. For a detailed description of  $REW$ , see Móring et. al (2016).  
10

The perturbation experiments were carried out as follows: the investigated parameter was modified with  $\pm 10\%$  and  $\pm 20\%$ , whilst the other parameters were kept the same. At the end of every simulation, the total  $\text{NH}_3$  exchange was calculated by summing the modelled hourly  $\text{NH}_3$  fluxes in the given modelling period. The difference compared with the baseline simulations was expressed as the percentage of the total  $\text{NH}_3$  exchange in the baseline model integrations (428 g N and 403 g N net emission for the whole field in the baseline simulations for P2002 and P2003, respectively).  
15

When the results from the sensitivity analysis for GAG\_field and GAG\_patch is compared (latter carried out by Móring et al., 2016), two types of lessons can be learned from the differences:

1. how the total  $\text{NH}_3$  exchange responds to the perturbation of the regulating parameters on different scales, and
2. how the sensitivity of total  $\text{NH}_3$  exchange to these parameters differ in the case of a single urine patch (simulated by GAG\_patch) and multiple urine patches (simulated within GAG\_field).  
20

For the first point, answers can be obtained with a simple comparison of the results for the two different scales. In the case of the second point, it has to be considered that the modelling approach for  $\text{NH}_3$  exchange is different in the case of the non-urine area and the urine patches deposited on the field (Section 3.1). As a consequence, the parameters that are used only in the formulation of GAG\_field for the urine patches have an effect on the  $\text{NH}_3$  exchange for the whole field only through the  $\text{NH}_3$  emission from the urine patches. These parameters are  $\Delta z$ ,  $\beta$ ,  $REW$ ,  $\theta_{fc}$ ,  $\theta_{pwp}$ , and  $\text{pH}(t_0)$  (initial soil pH).  
25

The value of  $\Delta z$ ,  $REW$ ,  $\theta_{fc}$ , and  $\theta_{pwp}$  influence the water budget, which is considered in the calculation of the stomatal resistance for both the non-urine area and the patches (Móring et al., 2016). However, preliminary results indicated that without the urine patches (assuming only non-urine area), the change in the total  $\text{NH}_3$  exchange over the field in response to the perturbations applied to these parameters were negligibly small (under 1% in absolute value). Therefore, the effect of  $\Delta z$ ,  $REW$ ,  $\theta_{fc}$ , and  $\theta_{pwp}$  on the total  $\text{NH}_3$  exchange over a grazed field through the non-urine area can be ignored.  
30

In essence, when  $\Delta z$ ,  $\beta$ ,  $REW$ ,  $\theta_{fc}$ ,  $\theta_{pwp}$ , and  $\text{pH}(t_0)$  perturbed, the changes of the total exchange flux are attributed exclusively to the changes in the emission flux over the urine patches. However, the sensitivity of the total net  $\text{NH}_3$  exchange above the



whole field ( $Sens_{net}$ ) and the urine patches ( $Sens_{patch}$ ) are different due to the deposition term to the clean area (as explained below) in the case of the whole field. In the following, it is shown, how  $Sens_{net}$  can be converted to  $Sens_{patch}$ .

Since the net  $NH_3$  exchange over the whole field equals to the sum of the  $NH_3$  emission from the urine patches and the  $NH_3$  exchange over the non-urine area (Fig. 4), the total  $NH_3$  exchange over the whole field ( $\Sigma F_{net}$ , Eq. 16) over a time interval is equal to the sum of the total  $NH_3$  exchange over the non-urine area ( $\Sigma F_{non}$ ) and the total  $NH_3$  emission from the urine patches ( $\Sigma F_{patch}$ ). Therefore, based on Eq. (16), when a urine-patch-related parameter is perturbed, the resulting differences ( $\Delta F$ ) in  $\Sigma F_{patch}$  and  $\Sigma F_{net}$  will be the same.

$$\Sigma F_{net} = \Sigma F_{non} + \Sigma F_{patch} \quad (16)$$

Using  $\Delta F$ ,  $Sens_{patch}$  and  $Sens_{net}$  can be expressed as:

$$Sens_{patch} = \frac{\Delta F}{\Sigma F_{patch}} \quad (17)$$

$$Sens_{net} = \frac{\Delta F}{\Sigma F_{net}} \quad (18)$$

In order to convert the values of  $Sens_{net}$  to  $Sens_{patch}$ , based on Eq. (17) and Eq. (18),  $Sens_{net}$  has to be multiplied by the ratio of  $\Sigma F_{net}$  and  $\Sigma F_{patch}$ . These ratios were approximately 0.5 in the baseline simulations with GAG\_field (0.54 and 0.48 in P2002 and P2003, respectively). Therefore, in order to investigate the difference in the response of  $\Sigma F_{patch}$  in the case of the multiple patches simulated within GAG\_field and the single urine patch simulated by GAG\_patch, this value of 0.5 should be applied to the percentage differences calculated for GAG\_field ( $Sens_{net}$ ) as a multiplying factor.

### 3.3.2 Further methods used in the sensitivity analysis

As explained in Section 2.2.2, for  $c_N$ ,  $A_{patch}$  and  $UF$  constant, average values were applied in the baseline simulations with GAG\_field. However, in reality these parameters can vary amongst different animals, and amongst different urination events as well. To examine the model uncertainty caused by these model assumptions, firstly, a sensitivity analysis was carried out (Section 4.2.4) applying the minimum and the maximum of these parameters as suggested in the literature (Table 1 and 2 – 20 g N dm<sup>-3</sup> for  $c_N$  from Whitehead, 1995).

Since the results indicated that the largest uncertainty is coupled with  $c_N$  (Section 4.2.4), in the case of this parameter further examinations were carried out. In natural conditions, even within an hour, several different urine patches are deposited over the field. For example, calculating with the lowest animal number on the field in the baseline experiment with GAG\_field (17 from Table 2) and the minimal  $UF$  (8 urination day<sup>-1</sup> cattle<sup>-1</sup>, from Table 1), there were at least 5 urine patches deposited in an hour. When the number of urine patches is high enough, it can be assumed that the overall  $c_N$  of all the urine deposited in a given hour is characterized by the average of the  $c_N$  values related to the individual urination events. This can be expressed by Eq. (19), in which  $c_N^{Ave}(t_j)$  represents the average N concentration in the time step  $t_j$ ,  $c_N^k(t_j)$  stands for the N content associated with the k<sup>th</sup> urine patch in  $t_j$ , and  $n(t_j)$  is the number of urine patches deposited in  $t_j$ .



In the baseline simulations with GAG\_field,  $c_N^{Ave}$  was assumed to be  $11 \text{ g N dm}^{-3}$  over the whole modelling period, therefore, it was examined how the model responds to a value of  $c_N^{Ave}$ , which is calculated in every time step according to Eq. (19). To approach this task, firstly  $c_N^k$  values have to be randomized for every urination event from an estimated statistical distribution of  $c_N$ .

$$5 \quad c_N^{Ave}(t_j) = \frac{\sum_{k=1}^{n(t_j)} c_N^k(t_j)}{n(t_j)} \quad (19)$$

Li et al. (2012) fitted a log-normal distribution (Eq. 20) to a  $c_N$  dataset, originating from the observation of two Aberdeen Angus steers over three 24 hour periods (Betteridge et al., 1986). In Eq. (20)  $\sigma$  and  $\mu$  are the scale parameters of the distribution. These, in the fitted distribution by Li et al. (2012), were  $\sigma = 0.786$  and  $\mu = 1.154$ . The mean of  $c_N$  calculated from these values (Eq. 21) was  $4.33 \text{ g N dm}^{-3}$ . In the study of Li et al. (2012), the findings were applied for cows, assuming that the distribution of  $c_N$  is similar with the same  $\sigma$ , but a higher mean  $c_N$ . Based on these, from Eq. (21), Li et al. (2012) derived  $\mu$  of the new distribution for cows and from this they generated a series of samples for  $c_N$ .

$$D(c_N) = \frac{1}{c_N \sigma \sqrt{2\pi}} e^{-\frac{(\ln c_N - \mu)^2}{2\sigma^2}} \quad (20)$$

$$\text{mean}(c_N) = e^{\mu + \frac{\sigma^2}{2}} \quad (21)$$

To test the uncertainty coupled to  $c_N$  in GAG\_field, the following steps were carried out. Firstly, following the method described by Li et al. (2012), based on Eq. (21), a new distribution of  $c_N$  was obtained, assuming a mean  $c_N$  of  $11 \text{ g N dm}^{-3}$ , and  $\sigma = 0.786$ . In this way, the scale parameter  $\mu$  was found to be 2.089. The resulted distribution of  $c_N$  is depicted in Fig. 7. Secondly, in every time step  $c_N^k$  values were randomized from the resulted distribution, and from these,  $c_N^{Ave}$  was derived based on Eq. (19). This resulted in a time series of  $c_N^{Ave}$  values.

In total, 30  $c_N^{Ave}$  time series were generated for both experimental periods (P2002 and P2003) and simulations were performed with GAG\_field, for all of these time series.

Finally, in order to investigate the model response of GAG\_field to a constant value of soil pH, model experiments were performed with different constant values of soil pH (Section 4.2.5).

## 4 Results

### 4.1 Model results derived by GAG\_field

25 The model results for P2002 and P2003 are illustrated in Fig. 4. These model experiments are regarded as the baseline simulations and are discussed in Sections 4.1.1. In addition to the general evaluation of these model results, in Section 4.1.2, the contribution of the  $\text{NH}_3$  emission from the urine patches to the  $\text{NH}_3$  exchange over the whole field is also investigated.





#### 4.1.1 Baseline simulations and model evaluation

In the case of P2002 (Fig. 8a) the model was in a broad accordance with the observations. It captures the characteristic daily variation of  $\text{NH}_3$  exchange detected over 31/08-02/09, with the magnitudes of the modelled and measured generally within  $50 \text{ ng m}^{-2} \text{ s}^{-1}$ . A larger difference occurred on 02/09 when the model clearly underestimated the observations. Discrepancies between the simulated and measured values can be also seen in the first two days of the modelling period and on the fourth day. Nevertheless, on these days the bottom  $\text{NH}_3$  concentration sensor did not work; therefore, the reliability of the flux calculated based only on the concentration measurements at the middle and top level is less certain. In addition, according to the metadata, on 27/08, before the gap in the observed fluxes (Fig. 8a), the stripping solution of the denuder ran out. This could explain the last 2-3 very high measured values beforehand.

In P2003 (Fig. 8b) the simulation generally agreed with the observations within  $50 \text{ ng m}^{-2} \text{ s}^{-1}$ . The match with the observed fluxes was especially close in the second half of 23/06. By contrast, the largest difference was found on 24/06, in the morning, when an emission peak was detected during the measurements at 04:00-08:00. Even though there was a midday peak also in the simulation, it occurred 6 hours later than the maximum in the observation. The increase in measured fluxes on 24/06 was linked to a period of high wind speed (with largest values between 04:00-08:00 AM, not shown here). Although wind speed is included in the model, the larger effect on measured fluxes could imply a proportionately larger effect of turbulence on the fluxes (through atmospheric and within canopy resistances, see the parametrization in Möring et al., 2016) than estimated by the model. In addition, it should be noted that on 20/06 between 11:00 and 15:00 the  $\text{NH}_3$  concentration denuder in the bottom height was not functioning properly, and afterwards it was not operating until 23/06 13:00 PM (in these periods only the remaining two denuders were considered), suggesting uncertainty in the measured dataset.

#### 4.1.2 Contribution of the urine patches to $\text{NH}_3$ exchange over the field

Figure 9 distinguishes the contribution of the urine patches and the non-urine area to the simulated  $\text{NH}_3$  exchange flux for the two modelling periods. It can be seen that the temporal variation of the  $\text{NH}_3$  fluxes over the whole field were dominated by the  $\text{NH}_3$  emission from the urine patches, which was offset by simultaneous  $\text{NH}_3$  deposition to the non-urine area. In the absence of the urine patches in both experiments, deposition would have occurred for most of the time. This illustrates the considerable effect of the presence of grazing animals on  $\text{NH}_3$  exchange over grasslands.

As an example of the differences, it can be seen in Fig. 9a that the  $\text{NH}_3$  emission from the urine patches on 02/09 were almost twice as high as the net exchange over the field. This large difference could explain part of the difference between the simulation and measurements on this day (Fig. 8), if the model overestimated the deposition component of the net flux.

The contribution to the  $\text{NH}_3$  exchange flux was also investigated for the groups of patches deposited in the different time steps (Fig. 10). The ensemble of the fluxes from the different patches show a clear daily variation with  $\text{NH}_3$  emission peaks at midday in both modelling periods. In P2002, these peaks became lower from the fourth day because after the third day instead of the initial 40 animals, only 17 cattle were grazing on the field, depositing fewer urine patches.



In the baseline experiment with GAG\_patch, the first and highest peak in NH<sub>3</sub> emission occurred about 12 hours after the urine application (Móring et al., 2016). By contrast, in the current results using GAG\_field (Fig. 10) it can be observed that in some cases the highest peak over an individually deposited urine patch emerges more slowly, only a day or two days after the urination event. For example, in P2002 (Fig. 10a) from the urine patches deposited on the third day (orange lines) the highest emission occurred on the fourth day, or from the patches deposited on the sixth day (dark green lines) the maximal flux was observed two days later. Further examples from P2003 (Fig. 10.b) are the urination events on the second day (orange lines) from which the highest flux can be observed a day after.

It has to be also noted that NH<sub>3</sub> emission fluxes in a given day can be substantially affected by urine patches deposited several days earlier. For instance, in Fig. 10a, on 02/09 the fluxes originating from the urination events six days before (red lines) are comparable with those from urine patches deposited two days before (dark green lines).

## 4.2 Sensitivity analysis to the regulating model parameters

In the following, first, the results of the perturbation experiments (Table 3) with GAG\_field are discussed (Section 4.2.1-4.2.3). Secondly, in Section 4.2.4 the uncertainty associated with  $c_N$ ,  $A_{patch}$  and  $UF$  is investigated. Finally, model experiments are presented in which GAG\_field was tested with different constant values of soil pH (Section 4.2.5).

### 4.2.1 Sensitivity to $\Delta z$ , $REW$ , $pH(t_0)$ , $\Gamma_{sto}$ and $\Gamma_{soil}$

According to Table 3, compared with the other parameters, for GAG\_field,  $\Sigma F_{net}$  turned out to be the least sensitive to the changes in  $\Delta z$  and  $REW$ . These percentage differences were similar in the case of the perturbation experiments with GAG\_patch, with an overall, slightly weaker sensitivity than was found in the case of GAG\_patch (results are taken from Móring et al., 2016).

In the case of  $pH(t_0)$ ,  $\Sigma F_{net}$  was found to be very sensitive to the  $\pm 10\%$  and  $\pm 20\%$  modifications (Table 3). However, it has to be pointed out that these changes in the value of  $pH(t_0)$  ( $\pm 0.5$  unit for a  $\pm 10\%$  modification and  $\pm 1$  unit for  $\pm 20\%$ ), can be considered as a large increase in the soil pH, taking into account that during intensive urea hydrolysis 2-3 units change can be expected (Fig. 11).

The constant  $\Gamma_{sto}$  and  $\Gamma_{soil}$  affect NH<sub>3</sub> exchange over the whole field exclusively through its effect on the NH<sub>3</sub> exchange over the non-urine area. As the results show (Table 3), the model is only slightly sensitive to  $\Gamma_{sto}$ , whilst  $\Gamma_g$  can have a considerable effect on NH<sub>3</sub> exchange.

### 4.2.2 Sensitivity to $\beta$

In the case of  $\beta$ , strong sensitivity was detected in  $\Sigma F_{net}$  (Table 3). Since  $\beta$  is not used in the parametrization of the NH<sub>3</sub> exchange over the non-urine area, it affects  $\Sigma F_{net}$  exclusively through the urine patches ( $\Sigma F_{patch}$ ). Therefore, the response of the total NH<sub>3</sub> exchange to the perturbation of  $\beta$  over the multiple patches in the two baseline simulations with GAG\_field and the single urine patch in GAG\_patch can be compared. To this, as explained in Section 3.3.1, the multiplying factor of 0.5 has to



be applied to the percentage differences derived for  $\beta$  in the P2002 and P2003 simulations with GAG\_field (Table 3). The values resulting in this way are significantly larger than those reported for GAG\_patch, suggesting a stronger sensitivity of  $\Sigma F_{patch}$  to the variation of  $\beta$  for the multiple patches than for the single patch. The reasons for this large difference between the two cases in the response of  $\Sigma F_{patch}$ , were further investigated in a series of model experiments with GAG\_patch (Table 4).

5 Following Mórning et al. (2016), the effect of buffering on the  $H^+$  ion budget in the  $NH_3$  source layer can be expressed with the term  $(pH(t_i) - pH(t_{i-1})) \times \beta_{patch}$ , where  $\beta_{patch} = \beta \times A_{patch} \times \Delta z$ . Based on these, the main factors that can regulate the governing role of buffering in the evolution of soil pH in the  $NH_3$  source layer and subsequently,  $NH_3$  exchange, are

1)  $pH(t_i) - pH(t_{i-1})$ , and

2)  $\beta_{patch}$ .

10 Considering point 1), if  $pH(t_0)$  is lower, i.e.  $[H^+]$  is higher, during urea hydrolysis more  $H^+$  ion can be consumed. This results in a larger increase in soil pH shortly after the urine patch deposition. In the baseline simulations with GAG\_patch and GAG\_field  $pH(t_0)$  was 6.65 and 4.95, respectively. On Fig. 11 it can be observed that in most of the urine patches deposited in the baseline simulations with GAG\_field, the difference between the initial and maximum soil  
 15 pH was about 3 units, whilst in the case of the baseline experiment with GAG\_patch (with the higher  $pH(t_0)$ ) it was only 2 (Mórning et al., 2016).

These larger changes in soil pH generate a larger buffering effect ( $(pH(t_i) - pH(t_{i-1})) \times \beta_{patch}$ ), i.e. a larger term in the  $H^+$  budget, which makes the system more sensitive to a modification of  $\beta$  through  $\beta_{patch}$ . This was confirmed in the model experiment A (Table 4). In this simulation GAG\_patch was run with the initial pH of 4.95 used in the baseline simulation with GAG\_field. Although the response of  $NH_3$  exchange was relatively weak to the modifications of  $\beta$ , it was stronger than in the original  
 20 perturbation experiment for GAG\_patch (Table 3).

Regarding point 2), the definition of  $\beta_{patch}$  expresses the buffering effect of the solid material of the soil on the liquid content. Since in the model  $\beta_{patch}$  is independent of the liquid content of the soil, within the source layer the same buffering effect takes place even if less urine stored in it. In a smaller amount of urine, the  $H^+$  ion budget (expressed in mol  $H^+$ ) and the variations in it are proportionally smaller too. Therefore, the governing role of the same buffering capacity in the case of a smaller amount  
 25 of urine becomes stronger, resulting in a stronger model sensitivity to  $\beta$ .

The maximum volume of urine that can be stored in the  $NH_3$  source layer ( $\theta_{urine}$ ) can be calculated as the difference of  $\theta_{fc}$  and  $\theta_{pwp}$ . The value of  $\theta_{urine}$  in the baseline experiments with GAG\_field and GAG\_patch were 0.18 and 0.3, respectively. This, based on the above consideration, suggests a stronger response in  $\Sigma F_{patch}$  to the perturbation of  $\beta$  for GAG\_field than GAG\_patch. This effect was explored in the model experiment B (Table 4), in which the

30 baseline simulation with GAG\_patch was performed with  $\theta_{fc}$  and  $\theta_{pwp}$  applied from the baseline experiment with GAG\_field (Table 2). The results show a small difference in  $\Sigma F_{patch}$  in response to the change of  $\beta$ , but it is still larger than in the sensitivity analysis carried out for the baseline simulation with GAG\_patch (Table 3), supporting the effect described above.

When the influence of  $pH(t_0)$  and the soil water content characteristics were examined together (model experiment C, Table 4), their effect added up, reaching a  $\pm 10\%$  difference in  $\Sigma F_{patch}$  when  $\beta$  was modified by  $\pm 20\%$ . The model was tested also with



a higher  $\theta_{pwp}$  (model experiment D, Table 4), assuming that half of the available space for urine in the model soil pore is filled with water, allowing only half of  $\theta_{urine}$  to infiltrate. This can represent a situation on the field when a urine patch is deposited after a rain event, when only half of the soil pore is empty. As expected, due to the smaller amount of urine, with this modification the sensitivity to  $\beta$  became even stronger.

- 5 If the percentage differences for  $\beta$  in GAG\_field reported in Table 3 are multiplied by 0.5, the resulting values (the percentage differences in  $\Sigma F_{patch}$  over the field) are between the values resulted from experiment C and D with GAG\_patch (Table 4). Overall, these findings suggest that over the field scale, the sensitivity of  $\Sigma F_{patch}$  to  $\beta$  over the individual urine patches, deposited in the different time steps, can vary between wide ranges, depending on  $\text{pH}(t_0)$  and the water content of the soil at the time of the given urination event. This varying sensitivity among the urine patches determines the overall sensitivity to  $\beta$
- 10 over the field.

#### 4.2.3 Sensitivity to $\theta_{fc}$ and $\theta_{pwp}$

In the case of  $\theta_{fc}$  and  $\theta_{pwp}$ , the perturbation experiments suggested an extremely strong sensitivity of  $\Sigma F_{net}$  (Table 3). Some of the changes in these parameters resulted in a  $\Sigma F_{net}$  that was double or almost triple (+191% in P2003 when  $\theta_{fc}$  was changed by +20%) of the  $\Sigma F_{net}$  for the baseline simulation. Furthermore, in P2003 when  $\theta_{pwp}$  was modified by +20% the originally positive

15 exchange turned to deposition (-118% less  $\Sigma F_{net}$  than in the baseline experiment).

Neglecting the effect of  $\theta_{fc}$  and  $\theta_{pwp}$  on  $\text{NH}_3$  exchange through the stomatal resistance (Section 3.3.1) in the non-urine area, these parameters influence  $\Sigma F_{net}$  mostly through the urine patches. Therefore, the sensitivity to these parameters over the urine patches in the field-scale experiments is comparable with that over the single urine patch in the baseline simulation with GAG\_patch (Table 3). In order to do the comparison, the multiplying factor of 0.5 has to be applied to the percentage

20 differences in Table 3 for the values derived for P2002 and P2003 with GAG\_field (as explained in Section 3.3.1). Although, in this way, the resulting percentages for P2002 and P2003 become less extreme, they still suggest a substantially stronger sensitivity of  $\Sigma F_{patch}$  to the modifications of  $\theta_{fc}$  and  $\theta_{pwp}$  in GAG\_field than GAG\_patch.

The value of  $\theta_{fc}$  and  $\theta_{pwp}$  influence  $\text{NH}_3$  exchange over a urine patch predominantly through  $\theta_{urine}$ , affecting the amount of urea available for hydrolysis in the  $\text{NH}_3$  source layer. Therefore, the difference in the response of  $\Sigma F_{patch}$  to the changes in  $\theta_{fc}$  and

25  $\theta_{pwp}$  over the two scales, might be caused by the difference in the values of  $\theta_{fc}$  and  $\theta_{pwp}$  used in the baseline simulations with GAG\_field and GAG\_patch. As it was pointed out above, in the baseline simulation with GAG\_patch  $\theta_{urine} = 0.4$ , and over the field scale  $\theta_{urine} = 0.18$ . In the perturbation experiments, when  $\theta_{fc}$  and  $\theta_{pwp}$  are modified this fillable space in the source layer is also affected. As it can be seen in Table 5, the  $\pm 10\%$  and  $\pm 20\%$  modifications of  $\theta_{fc}$  and  $\theta_{pwp}$  resulted in proportionally smaller differences in  $\theta_{urine}$  in the case of GAG\_patch than GAG\_field, suggesting a weaker response in  $\Sigma F_{patch}$  for GAG\_patch.

30 This effect was explored within a series of model experiments with GAG\_patch (Table 6), in which the  $\theta_{fc}$  and  $\theta_{pwp}$  used in the baseline simulation with GAG\_patch (0.4 and 0.1, respectively) were changed to those applied in the baseline simulation with GAG\_field (0.37 and 0.19, respectively). All the other parameters and input variables were kept the same as in the baseline simulation with GAG\_patch. The experiments were carried out in two cases for both  $\theta_{fc}$  and  $\theta_{pwp}$ : 1) when the initial water



content of the soil ( $\theta(t_0)$ ) was assumed to be the  $\theta_{pwp}$  ( $\theta(t_0) = 0.19$ ) and 2) when half of the available space was filled by liquid ( $\theta(t_0) = 0.28$ ), e.g. by rain water from a preceding rainfall.

As it can be seen in Table 6, with the  $\theta(t_0) = \theta_{pwp}$  model setting the sensitivity to both  $\theta_{fc}$  and  $\theta_{pwp}$  became higher than in the case of the original perturbation experiment with GAG\_patch (Table 3). This sensitivity became even stronger when urine was deposited to a half-filled source layer ( $\theta(t_0) = 0.28$ ). If the values from Table 6 are up-scaled to field-scale (dividing them by the factor of 0.5 defined in Section 3.3.1), the resulted percentage differences are similarly high to those observed in the sensitivity test for GAG\_field (Table 3).

These results suggest that depending on the rain events and how they modify the initial water budget in the soil before a urination event, the sensitivity of  $\text{NH}_3$  exchange to the perturbations of  $\theta_{fc}$  and  $\theta_{pwp}$  over the individual urine patches deposited in the modelling period can vary widely. This varying response to  $\theta_{fc}$  and  $\theta_{pwp}$  amongst the urine patches deposited in the field will determine the overall sensitivity to  $\theta_{fc}$  and  $\theta_{pwp}$  over the whole field.

#### 4.2.4 Sensitivity to $c_N$ , $A_{patch}$ and $UF$

As explained in Section 2.2.2, for  $c_N$ ,  $A_{patch}$  and  $UF$  constant, average values were applied in the baseline simulations with GAG\_field. However, in reality these parameters can vary amongst different animals, and amongst different urination events as well. To examine the model uncertainty caused by these model assumptions, firstly, a sensitivity analysis was carried out applying the minimum and the maximum of these parameters as suggested in the literature (Table 1 for  $A_{patch}$  and  $UF$ , and 2 – 20 g N dm<sup>-3</sup> for  $c_N$  from Whitehead, 1995).

According to Table 7, whilst the uncertainty originating from the choice of a constant  $A_{patch}$  and  $UF$  is considerable, the uncertainty coupled with the value of  $c_N$  is extremely large. Although the model shows a large uncertainty associated with  $c_N$ , the close agreement between GAG\_field and the measurements (Fig. 12) suggests that using the same average value in every time step well represents reality. In the following, the reasons of this high uncertainty associated with  $c_N$  is further examined. For this purpose, randomized  $c_N$  time series were generated as described in Section 3.3.2 and using these simulations were performed with GAG\_field.

The ensemble of the simulations derived in this way can be seen in Fig. 12. In both years the largest uncertainty occurred at the peaks of the  $\text{NH}_3$  fluxes. Overall, however, the uncertainties observed in Fig. 12 are much smaller than was suggested by the sensitivity analysis presented above (Table 7). This is because in the sensitivity analysis the two extremes of  $c_N$  were tested, whilst the  $c_N^{Ave}$  values generated from the log-normal distribution of  $c_N$  resulted in a value close to 11 g N dm<sup>-3</sup> applied in the baseline simulation with GAG\_field.

#### 4.2.5 Sensitivity to a constant soil pH

From the point of view of future application of the model for regional scale, computational time could be saved if a constant soil pH over the whole time period could be assumed instead



- of simulating soil pH dynamically for every urine patch deposited in the different time steps. To investigate the effect of such a simplification the baseline simulation with GAG\_field was performed with a constant soil pH of 7.5 (GAGf\_pH7.5). The reason for selecting this value, is that this is the approximate value where the curve of soil pH flattens out in the case of every urine patch deposited in the baseline simulations in GAG\_field (Fig. 11).
- 5 With a fixed value of pH 7.5, the model produced a similar temporal variation in NH<sub>3</sub> flux as with the dynamically changing soil pH in the baseline simulation with GAG\_field (Fig. 13), following relatively closely the fluxes in the baseline simulations. The model was tested with further two constant soil pH values, 7.0 and 8.0 in the experiments GAGf\_pH7.0 and GAGf\_pH8.0, respectively. These simulations resulted in highly different NH<sub>3</sub> exchange fluxes compared to those in the baseline simulations, especially in the case of GAGf\_pH8.0 (Fig. 13).
- 10 Although the results from GAGf\_pH7.5 suggest a possible simplification of the model for larger scale application, GAGf\_8.0 and GAGf\_7.0 implied that the NH<sub>3</sub> exchange fluxes are sensitive to the chosen constant value of soil pH. In GAGf\_pH7.5 that value was applied where the soil pH stabilized under a patch after the intense urea hydrolysis stopped. However, this value might not be the same in every situation. For example, in the case of the baseline experiment with GAG\_patch the curve of soil pH flattened out around pH 7 (Móring et al., 2016). Therefore, further considerations are needed regarding the choice of
- 15 a constant soil pH, which may also be expected to vary with soil type.

## 5 Discussion

### 5.1 Model development and evaluation

- The main source of NH<sub>3</sub> emission from grazed fields - as mentioned above - is the urine patches (Laubach et al., 2013, Petersen et al., 1998). The GAG model (referred to as GAG\_patch in this study), was constructed for a single urine patch by Móring et al. (2016). GAG\_patch is capable of simulating the TAN and the water content of the soil under a urine patch and the variation of soil pH. At a larger scale, over a grazed field, NH<sub>3</sub> exchange is determined by the coupled effect of NH<sub>3</sub> emission from the urine patches and NH<sub>3</sub> exchange with the area of the field that is not affected by urine (non-urine area). Therefore, in this study GAG\_patch was extended and applied at the field scale, by employing it for the urine patches and using a modified version of it for the non-urine area.
- 20 As shown by Móring et al. (2016), the simulations with GAG\_patch with the incorporation of an assumed restart of urea hydrolysis and CO<sub>2</sub> emission resulted in a considerably better representation of the measurements than in the baseline simulation, where these processes were excluded. However, the assumptions for the restart of urea hydrolysis and CO<sub>2</sub> emission were hypothetical or specific for the experimental site. For a general model application these processes would need to be further investigated. Therefore, the possible restart of urea hydrolysis and CO<sub>2</sub> emission were concluded not to be
- 25 implemented to GAG\_field.
- Regarding the model structure and functionality, Móring et al. (2016) provided a comparison for GAG\_patch with the earlier modelling studies for urea affected soils (Sherlock and Goh, 1985, Rachhpal and Nye, 1986) and urine patches (Laubach et
- 30





al., 2012). Its field scale application, GAG\_field is novel among the field scale NH<sub>3</sub> exchange models, considering its dynamic approach for the modelling of soil pH under the urine patches. For the same purpose as GAG\_field, the PaSim ecosystem model by Riedo et al. (2002) and the VOLT' AIR model by Générmont and Cellier (1997) could be used, the latter simulating NH<sub>3</sub> emission related to fertilizer and manure application. Both of these models, however, treat pH as a constant over the whole modelled area and do not account for the characteristics of the temporal development of the NH<sub>3</sub> emission from the individual urine patches. Furthermore, the framework of VOLT' AIR is more complex and requires more input data. Thus, for grazing situations, it is much easier to adapt GAG\_field.

The ultimate goal of the development of GAG\_field was to construct a modelling tool that could be applied to regional (i.e. national or continental) scale. Thus, simplicity was a key aspect of the model development, avoiding extra steps through model simplification during the up-scaling. For this reason, GAG\_field operate with a single soil layer, neglecting the exchange of TAN and the movement of water between the soil layers. Even though the models mentioned above (Générmont and Cellier, 1997, Riedo et al., 2002) apply a more sophisticated, multi-layer approach for the soil, the model code of GAG\_field enables the addition of new modules. For instance, a multi-layer approach for simulating the TAN budget or the water budget in the source layer.

Similarly to GAG\_patch, GAG\_field also accounts for the influence of meteorological variables on NH<sub>3</sub> exchange. This serves as a base of a further study, focusing on the investigation of the meteorological drivers of NH<sub>3</sub> exchange over grazed field. Also, in future work, linking GAG\_field to an atmospheric chemistry transport model, these meteorological effects can be also explored in relation to NH<sub>3</sub> emission, dispersion and deposition on a larger (i.e. regional or global) scale.

Two baseline simulations were performed with GAG\_field over two modelling periods based on data measured at Easter Bush, UK. The modelled and observed NH<sub>3</sub> fluxes were in a reasonably broad agreement. The formulation of GAG\_field allowed us to investigate the NH<sub>3</sub> exchange separately for the urine-affected and unaffected areas, as well as for groups of patches deposited in different time intervals. The results suggested that the temporal evolution of the NH<sub>3</sub> exchange flux over a grazed field is dominated by the NH<sub>3</sub> emission from urine patches and its magnitude is substantially reduced by the simultaneous NH<sub>3</sub> deposition the non-urine area. It was also found that the temporal development of NH<sub>3</sub> emission can be considerably different in urine patches deposited in different time intervals. Moreover, the NH<sub>3</sub> flux over the field in a given day can be largely influenced by urine patches deposited several days earlier.

## 5.2 Sensitivity analysis

It was investigated how the total simulated NH<sub>3</sub> exchange flux responds to an assumed change in the model parameters that regulate NH<sub>3</sub> exchange over the whole field, as well as the TAN content and water content under the urine patches. A series of perturbation experiments was carried out for  $\Delta z$ ,  $REW$ ,  $\Gamma_{sto}$ ,  $\Gamma_g$ ,  $\beta$ ,  $pH(t_0)$ ,  $\theta_{fc}$ ,  $\theta_{pwp}$ . In addition to these analyses, we examined the uncertainty coupled with the selected value for  $A_{patch}$ ,  $UF$  and  $c_N$ , and we also tested GAG\_field with different constant values of soil pH. Although GAG\_field was constructed so that it accounts for the effects of meteorology on the NH<sub>3</sub> exchange over a grazed field, the investigation of the influence of the meteorological variables will be the scope of a future study.





Compared with the other parameters, the total  $\text{NH}_3$  exchange simulated by GAG\_field turned out to be the least sensitive to the changes in  $\Delta z$  and  $REW$ . For  $REW$  GAG\_patch showed a similar, negligibly weak response to the  $\pm 10\%$ ,  $\pm 20\%$  modifications (Móring et al., 2016). In the case of  $\Delta z$ ,  $\text{NH}_3$  exchange was found to be sensitive to the perturbations of this parameter in both the patch-scale and the field-scale experiments, the latter especially in the P2002 simulation.

5 Móring et al. (2016) carried out a model analysis in which the possible extreme values of  $\Delta z$  (calculated as the penetration depth of urine and applied from Laubach et al. 2012), which showed a strong response in the simulated  $\text{NH}_3$  fluxes. However, since the modelled  $\text{NH}_3$  fluxes were in a broad agreement with the measurements in three different model simulations using the same value of  $\Delta z$ , these results suggest that the main governing processes of  $\text{NH}_3$  emission from urine patches might occur in this thin top soil layer ( $\Delta z = 4$  mm) as assumed by Móring et al. (2016). Nevertheless, future work is needed to confirm this  
10 hypothesis, considering how further datasets can help characterize the appropriate thickness of the effective soil emission layer. The constant  $\Gamma_{sto}$  and  $\Gamma_{soil}$  affect  $\text{NH}_3$  exchange over the whole field exclusively through its effect on the  $\text{NH}_3$  exchange over the non-urine area. The results suggested that the model is only slightly sensitive to  $\Gamma_{sto}$ , whilst  $\Gamma_g$  can have a considerable effect on  $\text{NH}_3$  exchange.

Móring et al. (2016) found only a weak sensitivity of the total  $\text{NH}_3$  emission to  $\beta$ . Although the exact same value of  $\beta$  was used  
15 in GAG\_field for both modelling periods as by Móring et al. (2016) in GAG\_patch, at the field scale,  $\text{NH}_3$  exchange was found to be highly sensitive to the same changes in  $\beta$ . It was shown that the dependence of  $\text{NH}_3$  exchange on  $\beta$  is influenced by the soil pH before urine deposition and also by the maximum amount of urine that can be stored in the source layer. According to the results, in the case of the urine patches with higher initial soil pH and higher initial soil water content, the sensitivity of the total net  $\text{NH}_3$  exchange to  $\beta$  is stronger. However, the good agreement found on the field scale between the modelled and the  
20 observed  $\text{NH}_3$  fluxes in both modelling periods, suggests that the natural variability of  $\beta$  might be less than the perturbation applied in the sensitivity analysis. Nevertheless, this requires further experimental investigation.

Móring et al. (2016) showed that the dynamic simulation of soil pH was necessary to represent the first, highest peak in  $\text{NH}_3$  emission after the deposition of a urine patch. This finding can be refined by the current results, suggesting a strong sensitivity in the  $\text{NH}_3$  exchange associated with the value of soil pH before the deposition of the urine patch,  $\text{pH}(t_0)$ . In contrast, the results  
25 for field scale implied that if the value of soil pH after the intensive urea hydrolysis is chosen as a constant (in the presented baseline simulations this was pH 7.5) for the whole modelling period, the  $\text{NH}_3$  fluxes by GAG\_field are similar to those derived with the dynamic chemistry approach.

The apparent contradiction between the results for the two scales can be explained by that in the baseline simulations with GAG\_field (Fig. 11), in most of the urine patches 1.5-2 days after their deposition the soil pH flattened out at 7.5. This means  
30 that in a given hour the total  $\text{NH}_3$  flux over the whole field was mainly affected by urine patches under which the soil pH was about 7.5. These results from the approach with constant soil pH suggests a way for model simplification when it is applied to larger scales. Nevertheless, further considerations are needed to find a generalized approach that determines the applicable value of a constant soil pH.



The sensitivity analysis for both GAG\_patch and GAG\_field showed that the highest uncertainties are associated with the water content of soil at  $\theta_{fc}$  and  $\theta_{pwp}$ . Over the field scale the response of the  $\text{NH}_3$  fluxes was extremely strong to the perturbation of these parameters. This high sensitivity was attributed to the maximum amount of urine that the  $\text{NH}_3$  source layer can hold, which depends on  $\theta_{fc}$  and  $\theta_{pwp}$ , or if the soil volumetric water content is higher than  $\theta_{pwp}$  before a urination event, the initial water content of the soil ( $\theta(t_0)$ ). It was found that in the case of a higher initial soil water content (i.e. less urine in the source layer),  $\text{NH}_3$  exchange was more sensitive to the changes in  $\theta_{fc}$  and  $\theta_{pwp}$ .

The broad agreement between the simulated and measured  $\text{NH}_3$  fluxes suggests that the uncertainty of the measurement of  $\theta_{fc}$  and  $\theta_{pwp}$  might be less than the perturbations applied in the sensitivity analysis ( $\pm 10\%$ ,  $\pm 20\%$ ). However, a regional scale model application would require  $\theta_{fc}$  and  $\theta_{pwp}$  values over a high-resolution grid, which is likely to be coupled with higher uncertainties. Therefore, at regional scale model application, the uncertainty of the input  $\theta_{fc}$  and  $\theta_{pwp}$  datasets has to be assessed when the model results are evaluated.

Finally, according to the results, whilst the uncertainty originating from the choice of a constant  $A_{patch}$  and  $UF$  is considerable, the uncertainty coupled with the value of  $c_N$  is extremely large. Nevertheless, model simulations with randomized N concentrations implied that this uncertainty might be considerably smaller in reality than it was suggested by the sensitivity analysis.

## Conclusions

In this study the GAG model (Móring et al., 2016) for simulating  $\text{NH}_3$  emission from individual urine patches was extended and applied for field scale. The new, field-scale model (GAG\_field) was tested over two modelling periods for a grazed grassland at Easter Bush, UK. Comparison with micrometeorological  $\text{NH}_3$  flux measurements showed that the model reproduced the main features of the observed fluxes.

The simulations indicated that the temporal evolution of the  $\text{NH}_3$  exchange flux over a grazed field is dominated by the  $\text{NH}_3$  emission from the urine patches, which is substantially decreased by the simultaneous  $\text{NH}_3$  deposition the non-urine area. The results presented also showed that the evolution of  $\text{NH}_3$  emission from urine patches deposited in different time steps can be substantially different and that  $\text{NH}_3$  fluxes in a given day can be considerably affected by urine patches deposited several days earlier.

The sensitivity analysis to the regulating model parameters showed that the total  $\text{NH}_3$  flux modelled by GAG\_field is highly sensitive to the buffering capacity ( $\beta$ ), the field capacity ( $\theta_{fc}$ ) and the permanent wilting point ( $\theta_{pwp}$ ). The observed sensitivities turned out to be much higher than was found in the case of GAG\_patch. The different sensitivities over the two scales can be explained by the different initial soil pH and the different soil physical characteristics which determine the maximum volume of urine that can be stored in the  $\text{NH}_3$  source layer. It was found that in the case of urine patches with a higher initial soil pH and higher initial soil water content, the sensitivity of  $\text{NH}_3$  exchange to  $\beta$  was stronger. Also, in the case of a higher initial soil water content,  $\text{NH}_3$  exchange was more sensitive to the changes in  $\theta_{fc}$  and  $\theta_{pwp}$ .



The sensitivity analysis also showed that the nitrogen content of urine ( $c_N$ ) is associated with a high uncertainty. However, model experiments based on  $c_N$  values randomized from an estimated statistical distribution, implied that this uncertainty might considerably smaller in practice.

Finally, GAG\_field was tested with a constant soil pH of 7.5 to see how well a simpler model structure could perform, such for a regional scale application. The variation of  $\text{NH}_3$  fluxes simulated in this way showed a broad agreement with those from the baseline simulations with GAG\_field that accounts for a dynamically changing soil pH. Although there were differences in the detailed time-course of emissions, the overall patterns and magnitude of  $\text{NH}_3$  emissions were similar. These results suggest a way for model simplification when GAG\_field is applied later for regional scale. However, since the  $\text{NH}_3$  exchange fluxes showed a large sensitivity to the value of the applied constant soil pH, further examinations are needed, concerning the choice of this constant value in relation to difference in underlying soil conditions.

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**Table 1. Ranges of the parameters used in the calculation of the urine-covered proportion of a field with an area of 1 ha (= 10 000 m<sup>2</sup>).**

<b>Animal</b>	<b>Sheep</b>	<b>Cattle</b>	<b>Reference</b>
Number of animals on $A_{field}$	1 – 100	0.1 – 10	EC, 2015
Urination frequency ( $UF$ ) (urination animal <sup>-1</sup> day <sup>-1</sup> )	15 – 20	8 – 12	Whitehead, 1995
Patches deposited per day ( $N_i$ )	15 - 2 000	0.8 – 120	-
Patch area ( $A_{patch}$ ) (m <sup>2</sup> )	0.043 - 0.055	0.38 - 0.42	Williams and Haynes, 1994





**Table 2. Urine, soil and site specific constants used in the evaluation of GAG field. The source of the values that were not measured at the site are also indicated. P2002 and P2003 stand for the modelling periods in 2002 and 2003, respectively. Constants used in the model, but not mentioned here were kept the same as defined for the baseline simulation with GAG\_patch (Móring et al., 2016).**

Model constants	Value	Source (if not measured)
<b>Urine specific constants</b>		
$A_{patch}$ (area of a urine patch)	40 dm <sup>2</sup>	Williams and Haynes, 1994 (average value)
$c_N$ (nitrogen content of urine)	11 g N dm <sup>-3</sup>	Whitehead, 1995 (average values)
$W_{urine}$ (volume of urine)	2.5 dm <sup>3</sup>	
<b>Soil specific constants</b>		
$\theta_{fc}$ (field capacity)	0.37	
$\theta_{pwp}$ (permanent wilting point)	0.192	
$\theta_{por}$ (porosity)	0.54	
$pH(t_0)$ (initial soil pH)	4.95	
$\Gamma_g$ (soil emission potential)	3000	Modelled (Section 3.2.3)
$\theta(t_0)$ (initial volumetric water content)	0.356 (P2002) 0.24 (P2003)	
<b>Site specific constants</b>		
Latitude	55.87°	
Longitude	3.03°	
Height above sea level	190 m	
$A_{field}$ (field area)	5.424 ha	
$\Gamma_{sto}$ (stomatal emission potential)	500	Massad et al., 2010 (average value)
$UF$ (urination frequency)	10 animal <sup>-1</sup> day <sup>-1</sup>	Whitehead, 1995 (average values)
$z_w$ (height of wind measurement)	1 m	
$h$ (canopy height)	0.07 m (P2002) 0.08 m (P2003)	
$LAI$ (leaf area index)	0.9 m <sup>2</sup> m <sup>-2</sup> (P2002) <sup>a</sup> 1.1 m <sup>2</sup> m <sup>-2</sup> (P2003) <sup>b</sup>	
Number of cattle on the field	40, 17 (P2002) <sup>c</sup> 50, 52 (P2003) <sup>c</sup>	
$z$ (heights of NH <sub>3</sub> concentration measurements)	0.44 m, 0.96 m, 2.06 m	

<sup>a</sup>There was no measurement in P2002, therefore, the average of the measurements for P2003 was used.

5 <sup>b</sup>The value was measured on 23/06/2003.

<sup>c</sup>The date when the number of animals changed in P2002 and P2003 were 28/08/2002 and 23/06/2003, respectively.



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**Table 3. Results of the perturbation experiments with GAG\_field. The changes in the total NH<sub>3</sub> flux over the field as a response to a change ( $\pm 10\%$  and  $\pm 20\%$ ) in the listed model parameters where expressed as the percentage of the total NH<sub>3</sub> exchange in the baseline simulations with GAG\_field. Results are listed for both modelling periods, P2002 and P2003. As a comparison, the results of the sensitivity analysis carried out by Möring et al. (2016) for GAG\_patch are also indicated. In the column ‘Effect’ the letters denote how the given parameters affect total NH<sub>3</sub> exchange in GAG\_field: through the urine patches (P) or the non-urine area (N).**

Constants	Effect	Model experiment	Change in the total net flux in response to a change of the constants by			
			-20%	-10%	+10%	+20%
$\Delta z$ (thickness of the source layer)	P	P2002	-14%	-6%	+5%	+8%
		P2003	-8	-4%	+2%	-2%
		GAG_patch	-12%	-6%	+5%	+11%
$REW$ (readily evaporable water)	P	P2002	0	0	0	0
		P2003	-3%	-1%	+1%	+2%
		GAG_patch	-3%	-2%	+2%	+4%
$pH(t_0)$ (initial soil pH)	P	P2002	-57%	-30%	+32%	+66%
		P2003	-79%	-42%	+48%	+100%
		GAG_patch	-	-	-	-
$\Gamma_{sto}$ (stomatal emission potential)	N	P2002	-1%	-0.4%	+0.4%	+1%
		P2003	-1%	-0.3%	+0.3%	+1%
		GAG_patch	-	-	-	-
$\Gamma_g$ (soil emission potential)	N	P2002	-17%	-0.8%	+0.8%	+17%
		P2003	-12%	-6%	+6%	+12%
		GAG_patch	-	-	-	-
$\beta$ (soil buffering capacity)	P	P2002	+32%	+15%	-14%	-28%
		P2003	+50%	+24%	-22%	-41%
		GAG_patch	+1%	+1%	-1%	-1%
$\theta_{fc}$ (field capacity)	P	P2002	-119%	-63%	+70%	+148%
		P2003	-153%	-85%	+96%	+191%
		GAG_patch	-18%	-7%	+6%	+9%
$\theta_{pwp}$ (permanent wilting point)	P	P2002	+120%	+57%	-52%	-96%
		P2003	+157%	+76%	-65%	-118%
		GAG_patch	+9%	+5%	-4%	-9%



5 Table 4. Results from simulations with GAG\_patch, testing the effect of  $pH(t_0)$  (initial soil pH),  $\theta_{fc}$  (field capacity) and  $\theta_{pwp}$  (permanent wilting point) on the sensitivity of the total  $NH_3$  emission to  $\beta$ . Input data were applied from the baseline simulation with GAG\_patch (Móring et al., 2016), except for the parameters denoted in the table with a different font style. Bold values are taken from the input data for the baseline simulations with GAG\_field, and italics denote a situation when the water content was assumed to be halfway between the field-scale values of  $\theta_{fc}$  and  $\theta_{pwp}$ . The sensitivity was expressed as the percentage difference in the original  $NH_3$  emission derived with the given model settings with GAG\_patch (listed also in the table for every model experiment).

Model experiment	Model settings			Original emission (g N)	Response of emission to a change in $\beta$ by			
	$pH(t_0)$	$\theta_{fc}$	$\theta_{pwp}$		-20%	-10%	+10%	+20%
A	<b>4.95</b>	0.40	0.10	1.5 g	+5%	+2%	-2%	-5%
B	6.65	<b>0.37</b>	<b>0.19</b>	0.9 g	+3%	+1%	-1%	-2%
C	<b>4.95</b>	<b>0.37</b>	<b>0.19</b>	0.6 g	+11%	+5%	-5%	-10%
D	<b>4.95</b>	<b>0.37</b>	<i>0.28</i>	0.1 g	+42%	+18%	-16%	-30%



**Table 5.** The maximum space in the  $\text{NH}_3$  source layer that can be filled by the incoming liquid ( $\theta_{\text{urine}}$ ) in the baseline experiments with GAG\_patch and GAG\_field, and the percentage it changes when  $\theta_{\text{fc}}$  (field capacity) and  $\theta_{\text{pwp}}$  (permanent wilting point) are modified by  $\pm 10\%$  and  $\pm 20\%$ .

Scale	$\theta_{\text{urine}}$	Percentage difference in $\theta_{\text{urine}}$ as a response to a change in			
		$\theta_{\text{pwp}}$		$\theta_{\text{fc}}$	
		$\pm 10\%$	$\pm 20\%$	$\pm 10\%$	$\pm 20\%$
<b>GAG_patch</b>	0.3	$\pm 3\%$	$\pm 6\%$	$\pm 13\%$	$\pm 26\%$
<b>GAG_field</b>	0.18	$\pm 11\%$	$\pm 22\%$	$\pm 21\%$	$\pm 42\%$



5 **Table 6.** Model results from model experiments with GAG\_patch, testing the effect of the initial water content of the soil ( $\theta(t_0)$ ) on the model sensitivity to  $\theta_{fc}$  (field capacity) and  $\theta_{pwp}$  (permanent wilting point). Input data were applied from the baseline simulation with GAG\_patch, except for  $\theta_{fc}$  and  $\theta_{pwp}$ , which were applied from the baseline simulation with GAG\_field, and  $\theta(t_0)$ , which was modified in the simulations as stated below. The sensitivity was expressed as a percentage difference in the original  $\text{NH}_3$  emission (listed also in the table for every model experiment).

Parameter tested (x)	Model setting $\theta(t_0)$	Original emission (g N)	Response of emission to a change in x by			
			-20%	-10%	+10%	+20%
$\theta_{fc}$	$\theta_{pwp}$	0.9 g	-41%	-20%	+18%	+31%
	0.28	0.4 g	-90%	-47%	+45%	+81%
$\theta_{pwp}$	$\theta_{pwp}$	0.9 g	+33%	+16%	-16%	-31%
	0.28	0.4 g	+67%	+33%	-31%	-58%



**Table 7. Results from the baseline simulations with GAG\_field when the maximum and minimum was applied of the investigated parameters. In every simulation the difference in the total NH<sub>3</sub> exchange was derived, expressed as the percentage of the total exchange in the baseline simulations with GAG\_field.**

Parameters	Min/Max	Change in the total NH <sub>3</sub> exchange	
		P2002	P2003
$A_{patch}$ (dm <sup>2</sup> )	38	-9%	+11%
	40	+9%	-11%
$c_N$ (g N dm <sup>-3</sup> )	2	-187%	-211%
	20	+292%	+403%
$UF$ (urination animal <sup>-1</sup> day <sup>-1</sup> )	8	-38%	-42%
	12	+38%	+42%

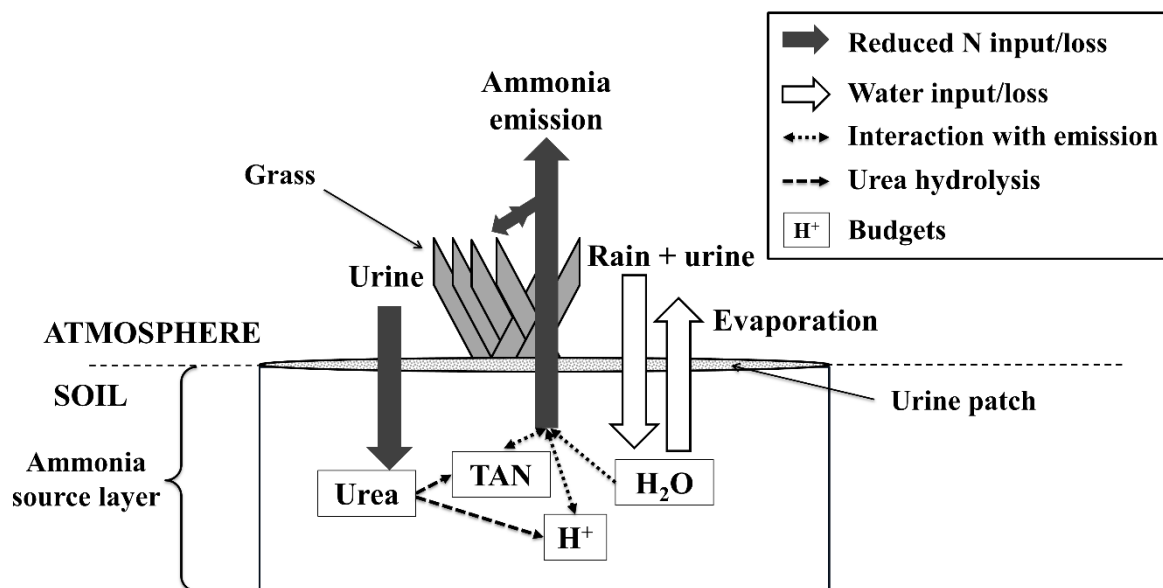
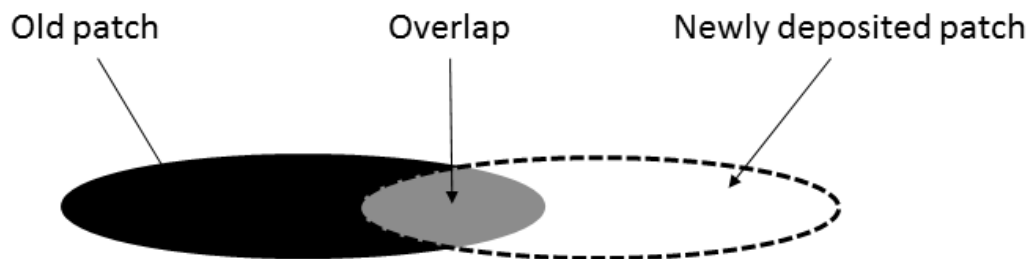
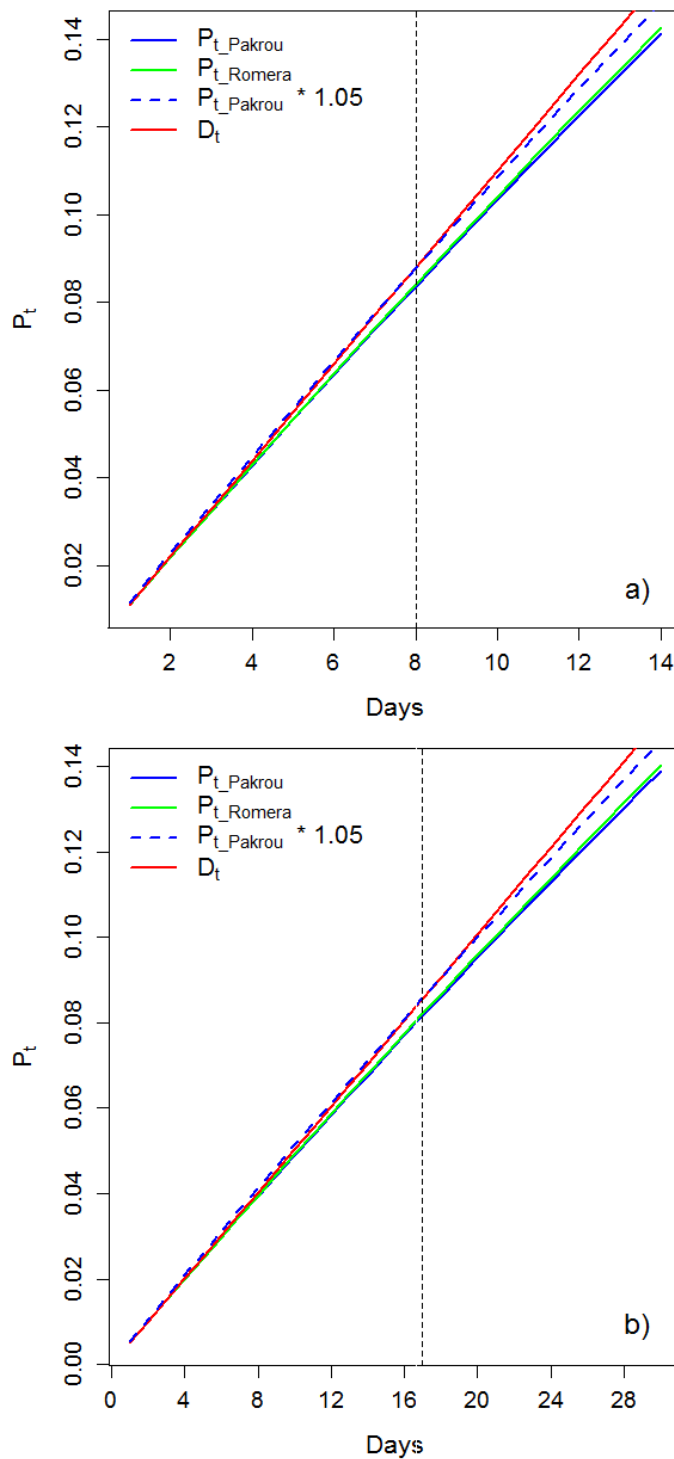


Figure 1. Simplified schematic of the GAG model by Móríng et al. (2016), referred to as GAG\_patch in this study.





**Figure 2.** Difference in the chemical composition of the soil in two urine patches deposited at different times. The different colours of the old and the newly deposited urine patches (black and white, respectively) as well as the overlap between them (grey) show the different soil chemical properties in the different areas.



**Figure 3: Proportion of the field covered by urine patches ( $P_t$ ) calculated for sheep (a) and cattle (b) as suggested by Pakrou and Dillon (2004) ( $P_{t\_Pakrou}$ ), Romera et al. (2012) ( $P_{t\_Romera}$ ) and when there is no overlap between the patches ( $D_t$ ).**

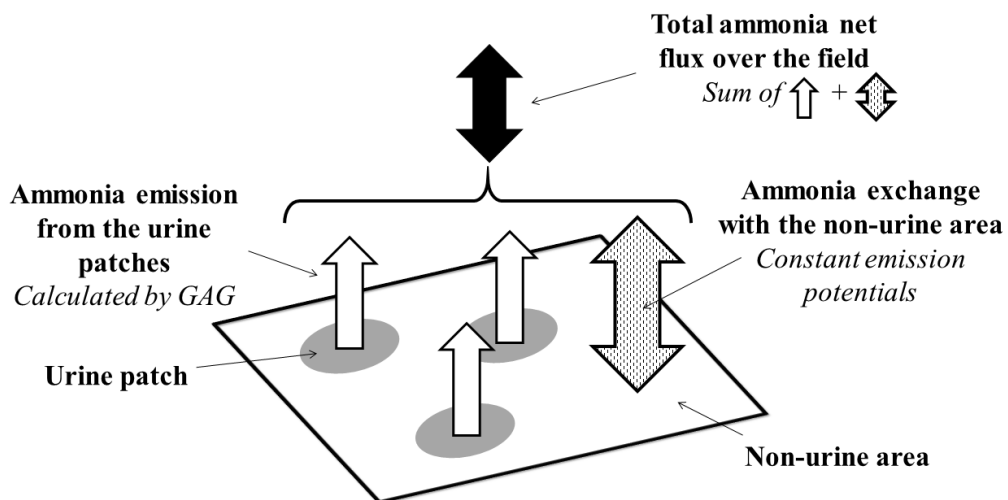


Figure 4. The schematic of GAG<sub>field</sub>. The figure depicts the components of the total net NH<sub>3</sub> flux over the field: NH<sub>3</sub> emission from the urine patches and the NH<sub>3</sub> exchange with the non-urine area.



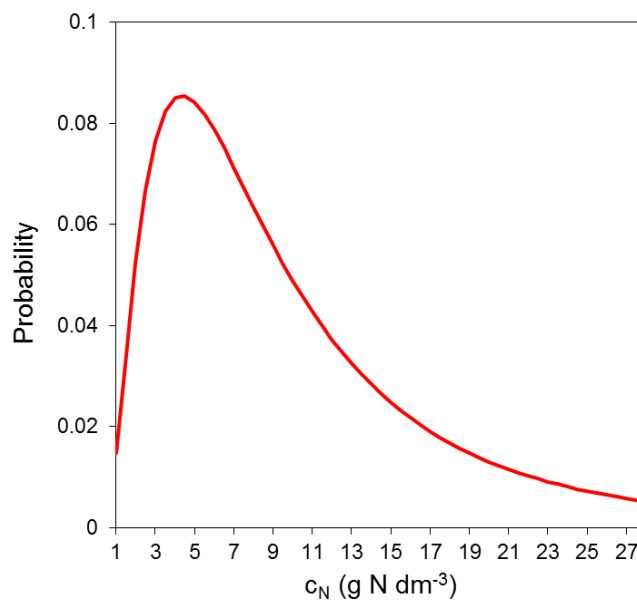
		NH <sub>3</sub> emission from the urine patches					NH <sub>3</sub> exchange with the non-urine area
		Time of the deposition of the urine patches					
		t <sub>j=1</sub>	t <sub>j=2</sub>	t <sub>j=3</sub>	...	t <sub>j=n</sub>	
Time since the beginning of the modelling period	t <sub>i=1</sub>	$F_{patch}^{j=1}(t_{i=1})$	$F_{non}(t_{i=1})$	$F_{non}(t_{i=1})$	...	$F_{non}(t_{i=1})$	$F_{non}(t_{i=1})$
	t <sub>i=2</sub>	$F_{patch}^{j=1}(t_{i=2})$	$F_{patch}^{j=2}(t_{i=2})$	$F_{non}(t_{i=2})$	...	$F_{non}(t_{i=2})$	$F_{non}(t_{i=2})$
	t <sub>i=3</sub>	$F_{patch}^{j=1}(t_{i=3})$	$F_{patch}^{j=2}(t_{i=3})$	$F_{patch}^{j=3}(t_{i=3})$	...	$F_{non}(t_{i=3})$	$F_{non}(t_{i=3})$
	...	...	...	...	...	...	...
	t <sub>i=n</sub>	$F_{patch}^{j=1}(t_{i=n})$	$F_{patch}^{j=2}(t_{i=n})$	$F_{patch}^{j=3}(t_{i=n})$	...	$F_{patch}^{j=n}(t_{i=n})$	$F_{non}(t_{i=n})$
Number of patches deposited in t <sub>j</sub>		n(t <sub>j=1</sub> )	n(t <sub>j=2</sub> )	n(t <sub>j=3</sub> )	...	n(t <sub>j=4</sub> )	0

5 Figure 5. Schematic for the temporal development of NH<sub>3</sub> fluxes (in every i<sup>th</sup> time step, t<sub>i</sub>) as derived by GAG\_field. F<sub>patch</sub><sup>j</sup>(t<sub>i</sub>) stands for the NH<sub>3</sub> flux from the urine patches deposited in the j<sup>th</sup> time step (t<sub>j</sub>), and F<sub>non</sub>(t<sub>i</sub>) stands for the NH<sub>3</sub> flux from the non-urine area. The bottom row shows how many urine patches were deposited in the given j<sup>th</sup> time step (n(t<sub>j</sub>)). Fluxes with striped background are calculated by GAG\_patch, and the fluxes with clear background are calculated by a modified version of GAG\_patch for non-urine area (explained in the text).



**Figure 6.** Satellite photo of the Easter Bush site. The map was generated by Google Maps, indicating the two halves of the field and the place of the instruments on the border of the two denoted by the small yellow rectangle. (The figure is taken from the metadata file by CEH.)

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**Figure 7.** Probability density function of the log-normal distribution generated for the distribution of the nitrogen content of urine ( $c_N$ ). The scale parameters are  $\sigma = 0.786$  and  $\mu = 2.089$ .

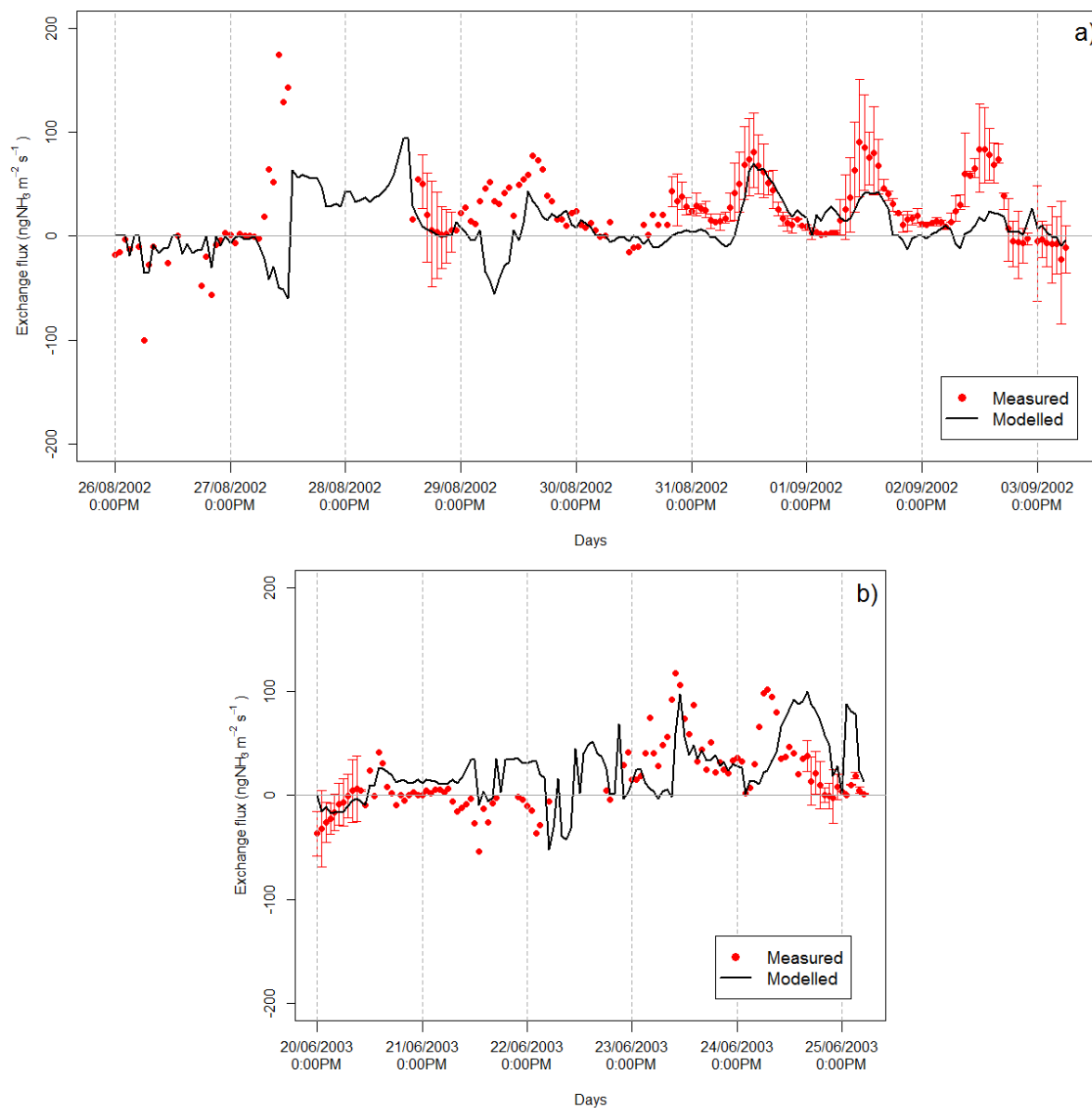
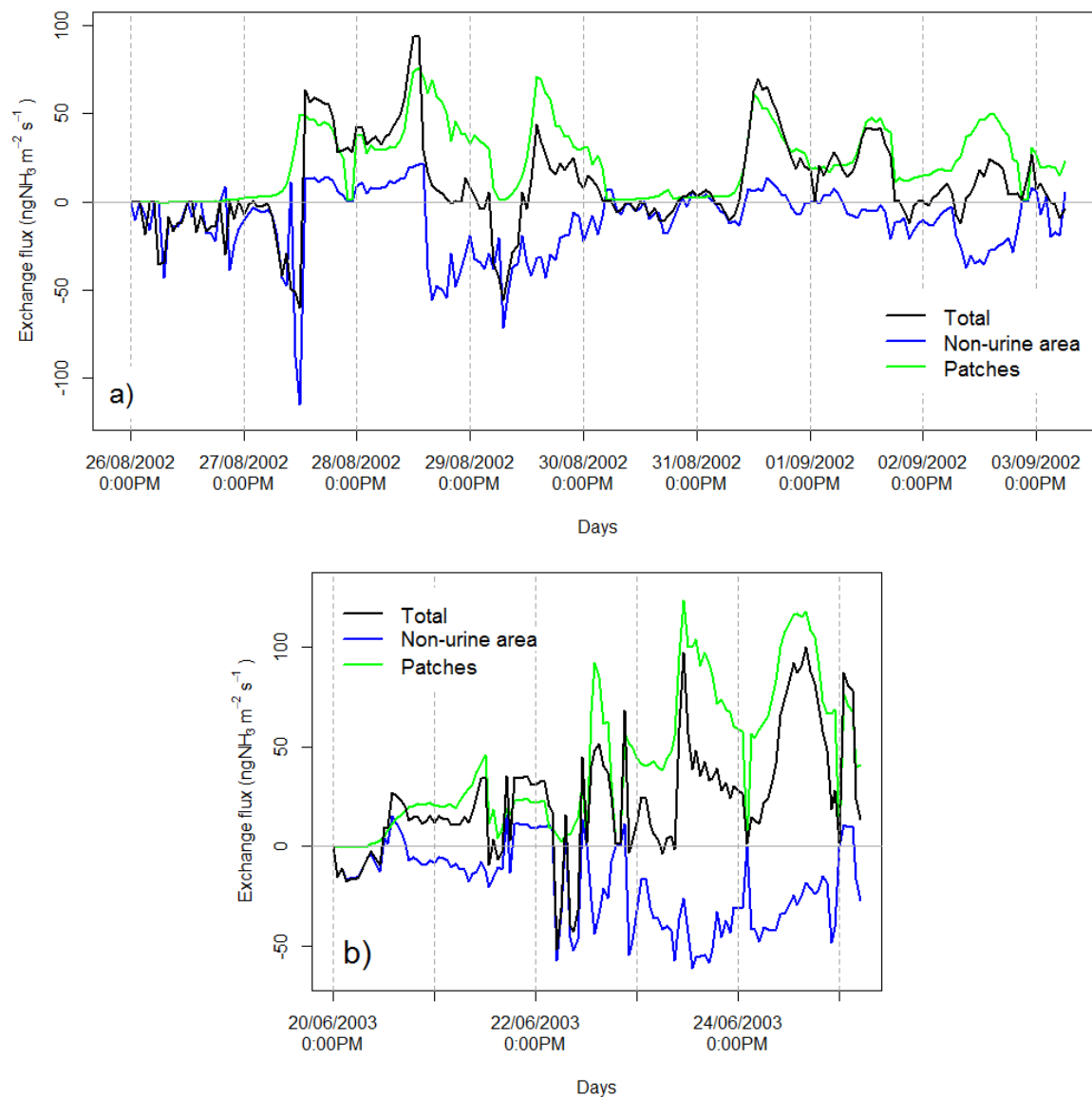


Figure 8. Comparison of the measured and modelled  $\text{NH}_3$  fluxes in the modelling periods P2002 (a) and P2003 (b). The uncertainty of the flux measurements is depicted as error bars. Where the error bars are missing one of the three  $\text{NH}_3$  concentration denuders were malfunctioning or not registering data at all.

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**Figure 9. Simulated NH<sub>3</sub> exchange fluxes over the urine patches, the non-urine area and the whole field in the modelling periods P2002 (a) and P2003 (b).**

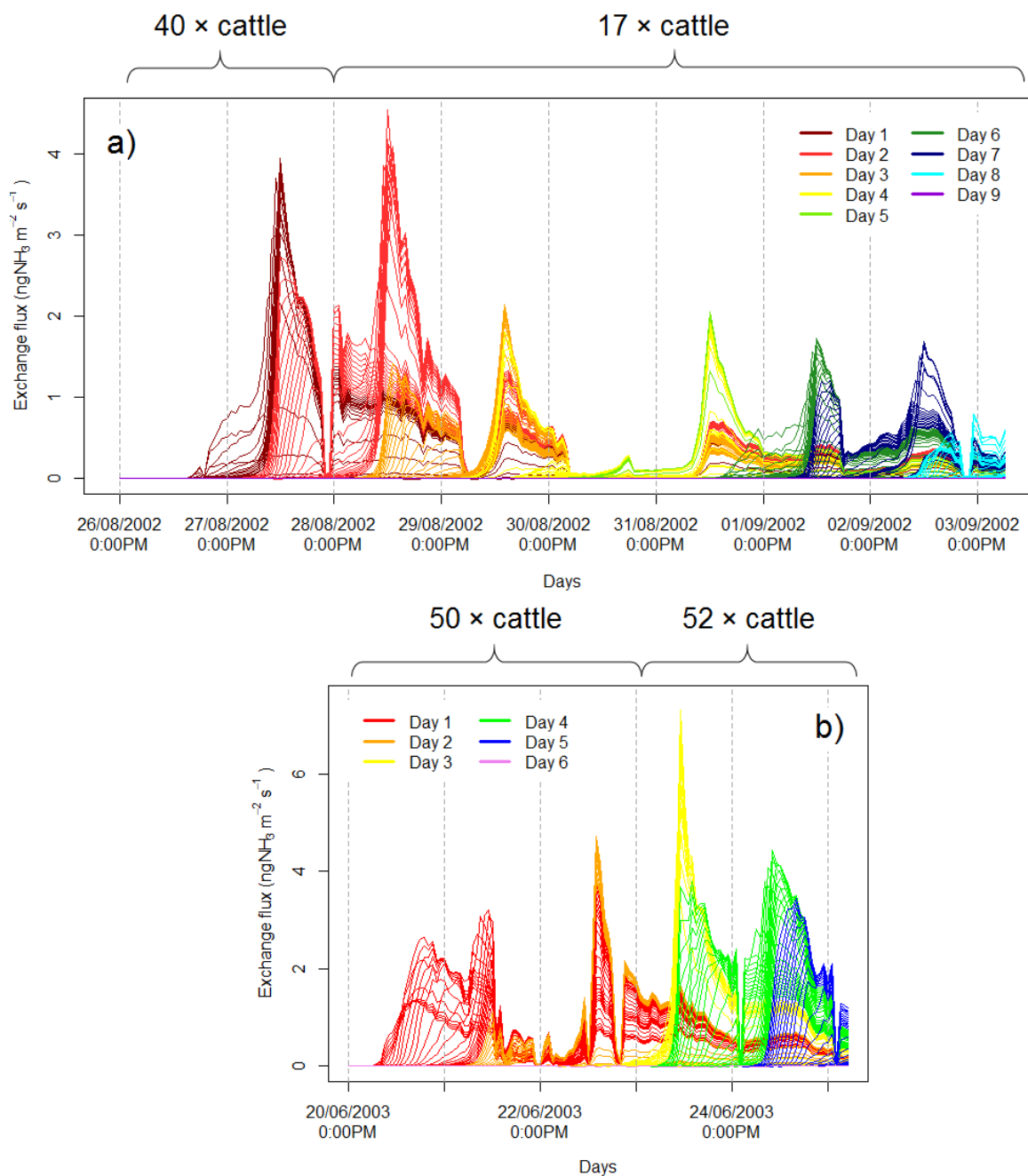
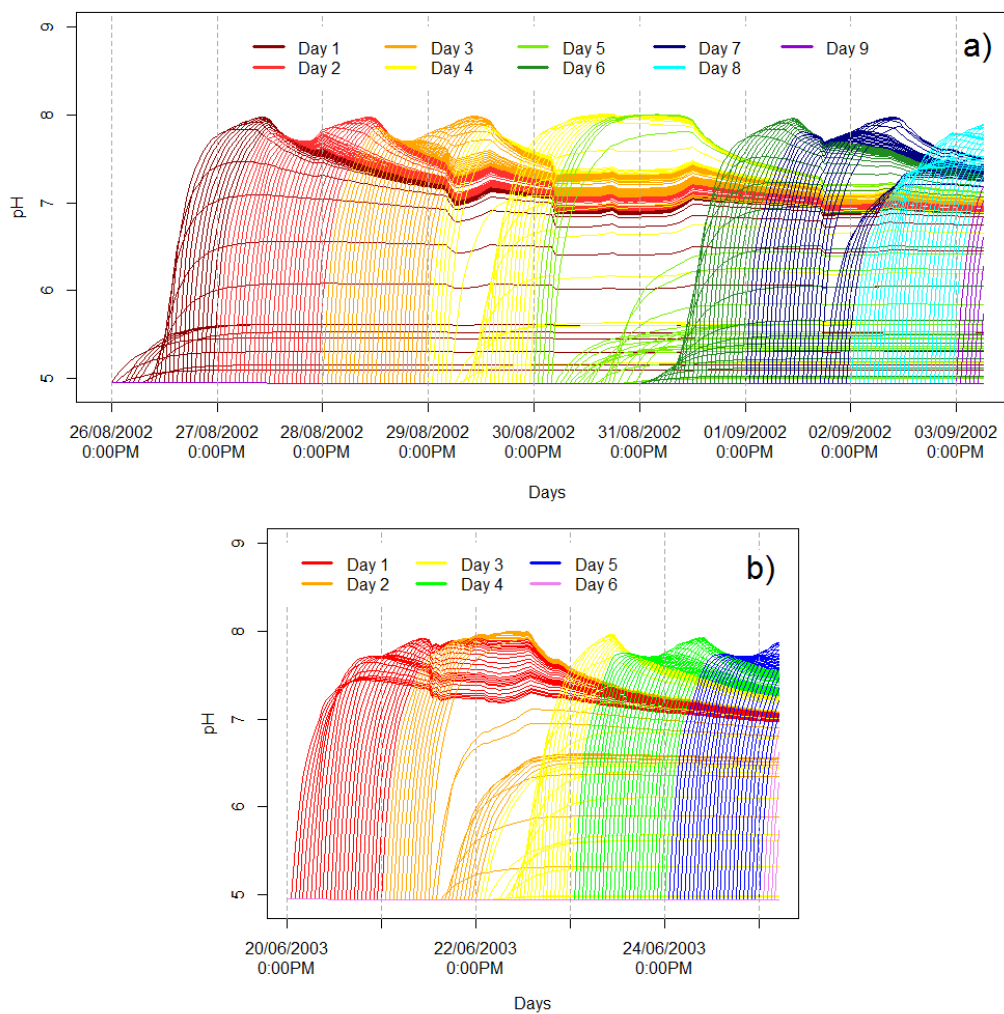


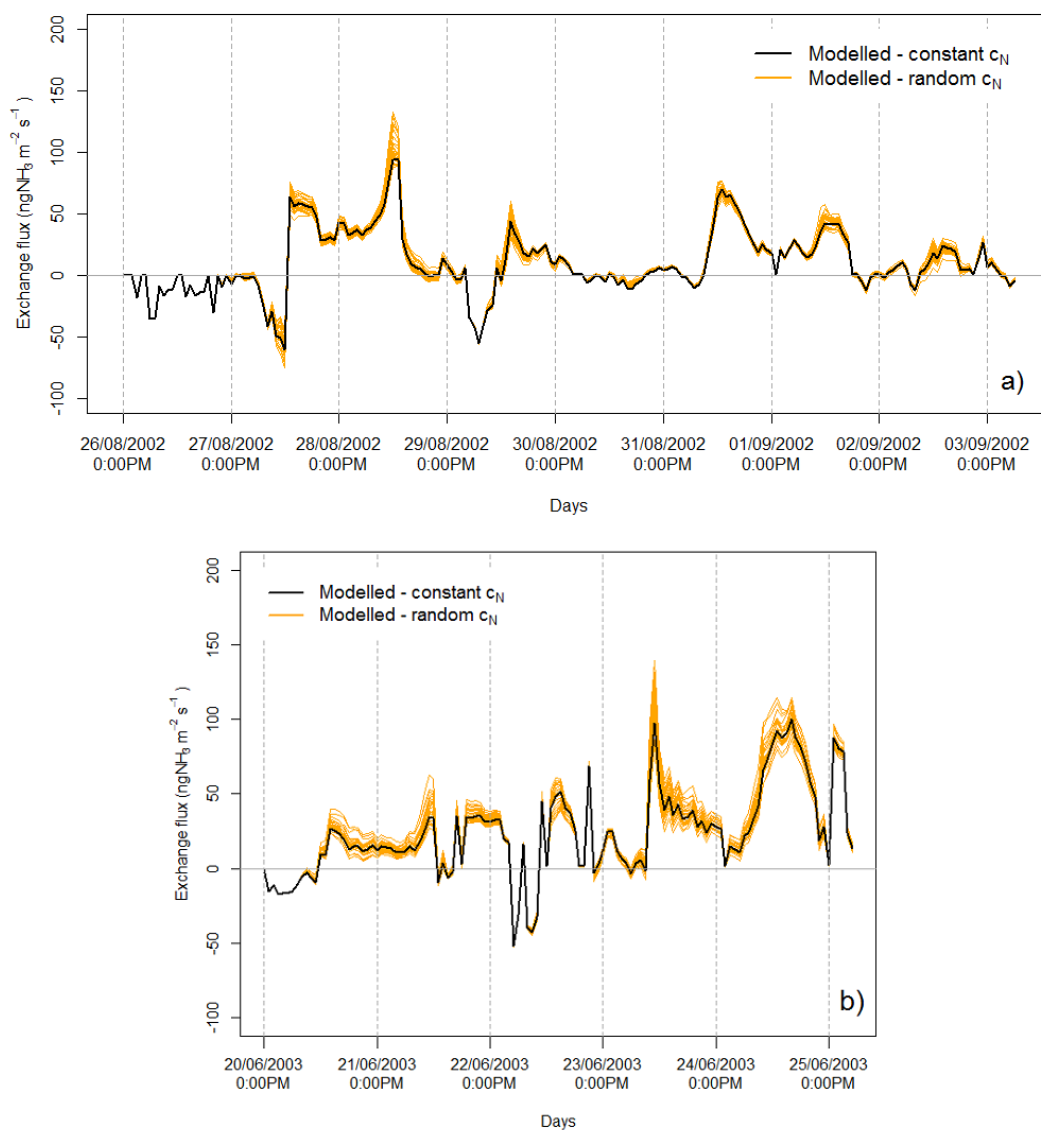
Figure 10. Simulated NH<sub>3</sub> fluxes from urine patches deposited in the same time step in the modelling periods P2002 (a) and P2003 (b). Each line indicates NH<sub>3</sub> fluxes from urine patches deposited in a given time step (expressed for the whole field), while the different colours indicate the days of the urination events. The number above the plots show how many cattle were grazing in the given time intervals.

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**Figure 11. Simulated soil pH in the NH<sub>3</sub> source layer under urine patches deposited in the same time step in the modelling periods, P2002 (a) and P2003 (b) in the baseline experiments with GAG\_field. The different colours indicate the days of the urination events. Each line indicates soil pH under urine patches deposited in a given time step, while the different colours indicate the days of the urination events.**

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**Figure 12.** Simulated  $\text{NH}_3$  exchange fluxes from the baseline simulation with GAG\_field with a constant  $c_N$  (black line), and 30 model experiments in which  $c_N$  was randomized for every time step (orange lines) for the modelling periods P2002 (a) and P2003 (b).

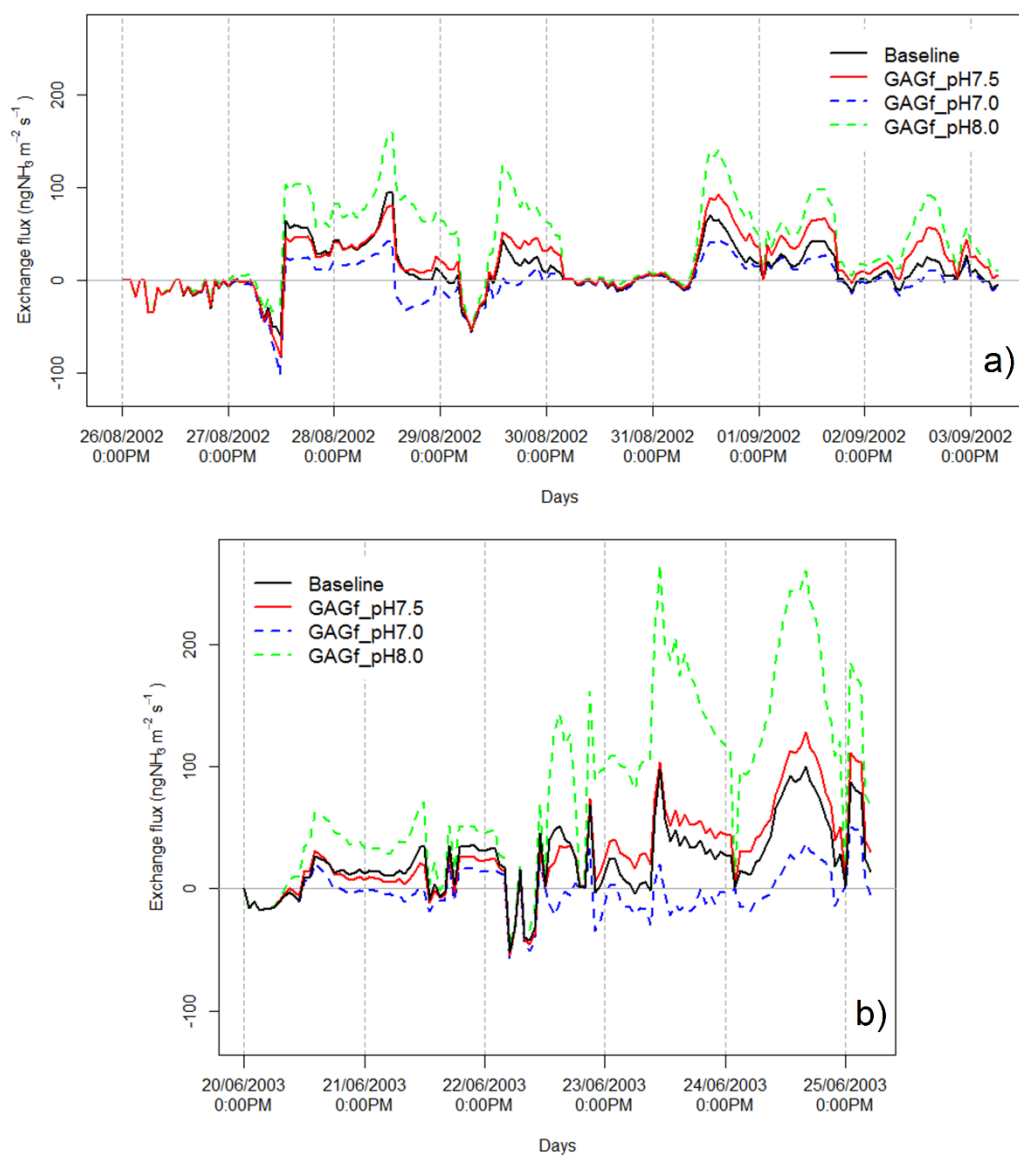


Figure 13.  $\text{NH}_3$  exchange fluxes simulated by GAG\_field with the original dynamic approach for soil pH (Baseline), and when constant values of soil pH were assumed: pH 7.5 (GAGf\_pH7.5), pH 7.0 (GAGf\_pH7.0) and pH 8.0 (GAGf\_pH8.0). Simulations were carried out for both modelling periods, P2002 (a) and P2003 (b).