

1 **Effect of soil saturation on denitrification in a grassland soil**

2 Laura Maritza Cardenas^{1*}, Roland Bol, R.², Dominika Lewicka-Szczebak,³ Andrew Stuart
3 Gregory⁴, Graham Peter Matthews⁵, William Richard Whalley⁴, Thomas Henry Misselbrook¹,
4 David Scholefield¹ and Reinhard Well³

5
6 ¹Rothamsted Research, North Wyke, Okehampton, Devon EX20 2SB, United Kingdom

7 ²Institute of Bio- and Geosciences, IBG-3/Agrosphere, Forschungszentrum Jülich GmbH, 52428
8 Jülich, Germany

9 ³Thünen Institute of Climate-Smart Agriculture, Federal Research Institute for Rural Areas,
10 Forestry and Fisheries, Bundesallee, 50, D-38116 Braunschweig, Germany

11 ⁴Rothamsted Research, Harpenden, Hertfordshire AL5 2JQ, United Kingdom

12 ⁵University of Plymouth, Drake Circus, Plymouth, Devon PL4 8AA, United Kingdom

13 *Correspondence to: Laura M. Cardenas (laura.cardenas@rothamsted.ac.uk)

14

15 **Abstract.** Nitrous oxide (N₂O) is of major importance as a greenhouse gas and precursor of

16 ozone (O₃) destruction in the stratosphere mostly produced in soils. The soil emitted N₂O is

17 generally predominantly derived from denitrification and to a smaller extent, nitrification ~~in soils~~,

18 both processes controlled by environmental factors and their interactions, and are influenced by

19 agricultural management. Soil water content expressed as water filled pore space (WFPS) is a major

20 controlling factor of emissions and its interaction with compaction, has not been studied at the

21 micropore scale. A laboratory incubation was carried out at different saturation levels for a

22 grassland soil and emissions of N₂O and N₂ were measured as well as the isotopocules of N₂O. We

23 found that fluxes variability was larger in the less saturated soils probably due to nutrient

24 distribution heterogeneity created from soil cracks and consequently nutrient hot spots. The results

25 agreed with denitrification as the main source of fluxes at the highest saturations, but nitrification

26 could have occurred at the lower saturation, even though moisture was still high (71% WFSP). The

27 isotopocules data indicated isotopic similarities in the wettest treatments vs the two drier ones. The

28 results agreed with previous findings where it is clear there are 2 N-pools with different dynamics:
29 added N producing intense denitrification, vs soil N resulting in less isotopic fractionation.

30 **Keywords**

31 Grassland, nitrous oxide,- isotopologues, isotopocule, greenhouse gases

33 **1 Introduction**

34 Nitrous oxide (N₂O) is of major importance as a greenhouse gas and precursor of ozone (O₃)
35 destruction in the stratosphere (Crutzen, 1970). Agriculture is a major source of greenhouse gases
36 (GHGs), such as carbon dioxide (CO₂), methane (CH₄) and also N₂O (IPCC, 2006). The application
37 of organic and inorganic fertiliser N to agricultural soils enhances the production of N₂O (Baggs *et*
38 *al.*, 2000). This soil emitted N₂O is predominantly derived from denitrification and to a smaller extent,
39 nitrification in soils (Davidson and Verchot, 2000). Denitrification is a microbial process in which
40 reduction of nitrate (NO₃⁻) occurs to produce N₂O, and N₂ is the final product of this process, benign
41 for the environment, but represents a loss of N in agricultural systems. Nitrification is an oxidative
42 process in which ammonium (NH₄⁺) is converted to NO₃⁻ (Davidson and Verchot, 2000). Both
43 processes are controlled by environmental factors and their interactions, and are influenced by
44 agricultural management (Firestone and Davidson, 1989). It is well recognised that soil water content
45 expressed as water filled pore space (WFPS) is a major controlling factor and as Davidson (1991)
46 illustrated, nitrification is a source of N₂O until WFPS values reach about 70%, after which
47 denitrification dominates. In fact, Firestone and Davidson (1989) gave oxygen supply a ranking of 1
48 in importance as a controlling factor in fertilised soils, above C and N. At WFPS between 45 and
49 75% a mixture of nitrification and denitrification act as N₂O sources. Davidson also suggested that at
50 WFPS values above 90% only N₂ is produced. Several studies have later proposed models to relate
51 WFPS with emissions (Schmidt *et al.*, 2000; Dobbie and Smith, 2001; Parton *et al.*, 2001; del Prado
52 *et al.*, 2006; Castellano *et al.*, 2010) but the “optimum” WFPS for N₂O emissions varies from soil to
53 soil (Davidson, 1991). Soil structure could be influencing this effect and it has been identified to
54 strongly interact with soil moisture (Ball *et al.*, 1999; van Groenigen *et al.*, 2005) through changes in

55 WFPS. Particularly soil compaction due to livestock treading and the use of heavy machinery affect
56 soil structure and emissions as reported by studies relating bulk density to fluxes (Klefoth *et al.*,
57 2014b); and degrees of tillage to emissions (Ludwig *et al.*, 2011).

58 Compaction is known to affect the size of the larger pores (macropores) thereby reducing the
59 soil air volume and therefore increasing the WFPS (for the same moisture content) (van der Weerden
60 *et al.*, 2012). However, little is known about the effect of compaction on the smaller soil pores
61 (micropores) and this could provide valuable information for understanding the simultaneous
62 behaviour of the dynamics of water in the various pore sizes in soil. Such an understanding would
63 lead to the development of better N₂O mitigation strategies via dealing with soil compaction issues.

64 The role of water in soils is closely linked to microbial activity but also relates to the degree
65 of aeration and gas diffusivity in soils (Morley and Baggs, 2010). Water facilitates nutrient supply to
66 microbes and restricts gas diffusion, thereby increasing the residence time of gases in soil, and the
67 chance of further N₂O reduction before it can be released to the atmosphere. This is further aided by
68 the restriction of the diffusion of atmospheric O₂ (Dobbie and Smith, 2001), increasing the potential
69 for denitrification. As a consequence, counteracting effects (high microbial activity vs low diffusion)
70 occur simultaneously making it difficult to predict net processes and corresponding outputs
71 (Davidson, 1991). Detailed understanding of the sources of N₂O and the influence of physical factors,
72 i.e. soil structure and its interaction with moisture, is a powerful ~~tool~~-basis for developing effective
73 mitigation strategies.

74 Isotopocules of N₂O represent the isotopic substitution of the O and/or the two N atoms within
75 the N₂O molecule. The isotopomers of N₂O, are those differing in the peripheral (β) and central N-
76 positions (α) of the linear molecule (Toyoda and Yoshida, 1999) with the intramolecular ¹⁵N site
77 preference (SP; the difference between $\delta^{15}\text{N}^{\alpha}$ - $\delta^{15}\text{N}^{\beta}$) used to identify production processes at the
78 level of microbial species or enzymes involved (Toyoda *et al.*, 2005; Ostrom, 2011). Moreover, $\delta^{18}\text{O}$,
79 $\delta^{15}\text{N}$ and SP of emitted N₂O depend on the denitrification product ratio (N₂O / (N₂+N₂O)), and hence
80 provide insight into the dynamics of N₂O reduction (Well and Flessa, 2009; Lewicka-Szczebak *et al.*,

81 ~~2014; Lewicka-Szczebak *et al.*, 2015; Lewicka-Szczebak *et al.*, 2014).~~ Data reported in the literature
82 ~~provide values for these parameters in relation to the source process for N₂O.~~ Koster *et al.* (2013) for
83 example recently reported $\delta^{15}\text{N}^{\text{bulk}}$ values of N₂O between -36.8‰ and -31.9‰ ~~in~~-under the
84 conditions of their experiment, which are indicative of denitrification according to Perez *et al.* (2006)
85 and Well and Flessa (2009) who proposed the range -54 to -10‰ relative to the substrate. Baggs
86 (2008) summarised that values between -90 to -40‰ are indicative of nitrification. Determination of
87 these values are normally carried out in pure culture studies or in conditions favouring either
88 production or reduction of N₂O (Well and Flessa, 2009). The SP is however considered a better
89 predictor of the N₂O source due to its independence from the substrate signature (Ostrom, 2011).

90 Simultaneous occurrence production and reduction of N₂O as in natural conditions presents
91 a challenge for isotopic factors determination due to uncertainty on N₂ reduction and the co-existence
92 of different microbial communities ~~resulting in other steps of denitrification happening as well~~
93 ~~producing N₂O~~ (Lewicka-Szczebak *et al.*, 2014). Recently, using data from the experiment ~~here~~
94 reported ~~here~~, where soil was incubated under aerobic atmosphere and the complete denitrification
95 process occurs, Lewicka-Szczebak *et al.* (2015) determined fractionation factors associated with N₂O
96 production and reduction using a modelling approach. The analysis comprised measurements of the
97 N₂O and N₂ fluxes combined with isotopocule data. Net isotope effects (η values) are variable to a
98 certain extent as they result from a combination of several processes causing isotopic fractionation
99 (Well *et al.*, 2012). The results generally confirmed the range of values of η (net isotope effects) and
100 $\eta^{18}\text{O}/\eta^{15}\text{N}$ ratios reported by previous studies for N₂O reduction for that part of the soil volume where
101 denitrification was enhanced by the N+C amendment. This did not apply for the other part of the soil
102 volume not reached by the N+C amendment, showing that the validity of published net isotope effects
103 for soil conditions with low denitrification activity still needs to be evaluated.

104 Lewicka-Szczebak *et al.* (2015) observed a clear relationship between ¹⁵N and ¹⁸O isotope
105 effects during N₂O production and denitrification rates. For N₂O reduction, differential isotope effects
106 were observed for two distinct soil pools characterized by different product ratios N₂O / (N₂+N₂O).

107 For moderate product ratios (from 0.1 to 1.0) the range of isotope effects given by previous studies
108 was confirmed and refined, whereas for very low product ratios (below 0.1) the net isotope effects
109 were much smaller. In this paper, we present the results from the gas emissions measurements from
110 soils collected from a long-term permanent grassland soil to assess the impact of different levels of
111 soil saturation on N₂O and N₂ and CO₂ emissions after compaction. CO₂ emissions were measured in
112 addition as an estimate of aerobic respiration and thus of O₂ consumption, which indicates
113 denitrification is promoted. The measurements included the soil isotopomer (¹⁵N_α, ¹⁵N_β and site
114 preference) analysis of emitted N₂O, which in combination with the bulk ¹⁵N and ¹⁸O was used to
115 distinguish between N₂O from bacterial denitrification and other processes (e.g. nitrification and
116 fungal denitrification) (Lewicka-Szczebak, 2016[7a](#)).

117 We conducted measurements at defined saturation of pores size fractions as a prerequisite to
118 model denitrification as a function of water status (Butterbach Bahl *et al.*, 2013 and Müller and
119 Clough, 2014). We have under controlled conditions created a single compaction stress of 200 kPa
120 (typical of soils compacted after grazing) in incremental layers using a uniaxial pneumatic piston to
121 simulate a grazing pressure. We hypothesized that at high water saturation, spatial heterogeneity in
122 of N emissions decreases due to more homogeneous distribution of the soil nutrients and/or anaerobic
123 microsites. We also hypothesized that even at high soil moisture a mixture of nitrification and
124 denitrification can occur. We also aimed to assess how these effects (spatial heterogeneity and source
125 processes) occur in a relatively narrow range of moisture (70-100%). As far as we know there no
126 other studies going to this level of detail. We aimed to understand changes in the ratio N₂O/(N₂O+N₂)
127 at the different moisture levels studied in a controlled manner on soil micro and macropores.
128 Moreover, we used ~~and the behaviour and utility of~~ isotopocule values of N₂O to evaluate if the
129 contribution of bacterial denitrification to the total N₂O flux was affected by moisture status ~~at the
130 different moisture levels studied in a controlled manner on soil micro and macropores.~~

131 **2 Materials and methods**

132 **2.1 Soil used in the study**

133 An agricultural soil, under grassland management since at least 1838 (Barré *et al.*, 2010), was
134 collected from a location adjacent to a long-term ley-arable experiment at Rothamsted Research in
135 Hertfordshire (Highfield, see soil properties in Table 1 and further details in Rothamsted Research,
136 2006; Gregory *et al.*, 2010). The soil had been under permanent cut mixed-species (predominantly
137 *Lolium* and *Trifolium*) vegetation. The soil was sampled as described in Gregory *et al.* (2010). Briefly
138 it was sampled from the upper 150 mm of the profile, air dried in the laboratory, crumbled and sieved
139 (<4 mm), mixed to make a bulk sample and equilibrated at a pre-determined water content (37 g 100
140 g⁻¹; Gregory *et al.*, 2010) in air-tight containers at 4° C for at least 48 hours.

141 **1.2.Preparation of soil blocks**

142 The equilibrated soil was then packed into twelve stainless steel blocks (145 mm diameter; h: 100
143 mm), each of which contained three cylindrical holes (i.d: 50 mm; h: 100 mm each). The cores were
144 packed to a single compaction stress of 200 kPa in incremental layers using a uniaxial pneumatic
145 piston. The three hole- blocks were used to facilitate the compression of the cores. The 200 kPa stress
146 was analogous to a severe compaction event by a tractor (Gregory *et al.*, 2010) or livestock
147 (Scholefield *et al.*, 1985). The total area of the upper surface of soil in each block was therefore 58.9
148 cm² (3 × 19.6 cm²) and the target volume of soil was set to be 544.28 cm³ (3 × 181.43 cm³) with the
149 objective of leaving a headspace of approximately 45 cm³ (3 × 15 cm³) for the subsequent experiment.
150 The precise height of the soil (and hence the volume) was measured using the displacement
151 measurement system of a DN10 Test Frame (Davenport-Nene, Wigston, Leicester, UK) with a
152 precision of 0.001 mm.

153 **2.3 Equilibration of soil cores at different saturations**

154 The soil was equilibrated to four different initial saturation conditions or treatments (t₀) which were
155 based on the likely distribution of water between macropores and micropores. The first treatment was
156 where both the macro- and micropores (and hence the total soil) was fully saturated; the second

157 treatment was where the macropores were half-saturated and the micropores remained fully saturated;
158 the third treatment was where the macropores were fully unsaturated and the micropores again
159 remained fully saturated; and the fourth treatment was where the macropores were fully unsaturated
160 and the micropores were half-saturated. These four treatments are hereafter referred to as SAT/sat;
161 HALFSAT/sat; UNSAT/sat and UNSAT/halfsat, respectively, where upper-case refers to the
162 saturation condition of the macropores and lower-case refers to the saturation condition of the
163 micropores. In order to set these initial saturation conditions, we referred to the gravimetric soil water
164 release characteristic for the soil, as given in Gregory *et al.* (2010) (see supplement 1). To achieve
165 target water contents during the incubation, the amount of liquid added with the C/N amendment (15
166 mL) was taken into account in the total volume of water added. For the SAT/sat and HALFSAT/sat
167 conditions, two sets of three replicate blocks were placed on two fine-grade sand tension tables
168 connected to a water reservoir. For the UNSAT/sat condition a set of three replicate blocks was placed
169 on a tension plate connected to a water reservoir, and the final set of three replicate blocks were placed
170 in pressure plate chambers connected to high-pressure air. All blocks were saturated on their
171 respective apparatus for 24 h, and were then equilibrated for 7 days at the adjusted target matric
172 potentials which were achieved by either lowering the water level in the reservoir (sand tables and
173 tension plate) or by increasing the air pressure (pressure chambers). At the end of equilibration period,
174 the blocks were removed carefully from the apparatus, wrapped in air-tight film, and maintained at 4
175 °C until the subsequent incubation.

176 **2.4 Incubation**

177 The study was carried out under controlled laboratory ~~controlled~~ conditions, using a
178 specialised laboratory denitrification (DENIS) incubation system (Cardenas *et al.*, 2003). Each block
179 containing three cores was placed in an individual incubation vessel of the automated laboratory
180 system in a randomised block design to avoid effect of vessel. The lids for the vessels containing
181 three holes were lined with the cores in the block to ensure that the solution to be applied later would
182 fall on top of each soil core. Stainless steel bulkheads fitted (size for ¼” tubing) on the lids had a

183 three-layered Teflon coated silicone septum (4 mm thick x 7 mm diameter) for supplying the
184 amendment solution by using a gas tight hypodermic syringe. The bulkheads were covered with a
185 stainless steel nut and only open when amendment was applied. The incubation experiment lasted 13
186 days. The incubation vessels with the soils were contained in a temperature controlled cabinet and
187 the temperature set at 20°C. The incubation vessels were flushed from the bottom at a rate of 30 ml
188 min⁻¹ with a He/O₂ mixture (21% O₂, natural atmospheric concentration) for 24 h, or until the system
189 and the soils atmosphere were emitting low background levels of both N₂ and N₂O (N₂ can get down
190 to levels of 280 ppm much smaller than atmospheric values). Subsequently, the He/O₂ supply was
191 reduced to 10 ml min⁻¹ and directed across the soil surface and measurements of N₂O and N₂ carried
192 out at approximately 2 hourly cycles to sample from all the 12 vessels. Emissions of CO₂ were
193 simultaneously measured.

194 **2.5 Application of amendment**

195 An amendment solution equivalent to 75 kg N ha⁻¹ and 400 kg C ha⁻¹ was applied as a 5 ml aliquot a
196 solution containing KNO₃ and glucose to each of the three cores in each vessel on day 0 of the
197 incubation. Glucose is added to optimise conditions for denitrification to occur (Morley and Baggs,
198 2010). The aliquot was placed in a stainless steel container (volume 1.2 l) which had three holes
199 drilled with bulkheads fitted, two to connect stainless steel tubing for flushing the vessel, and the third
200 one to place a septum on a bulkhead to withdraw solution. Flushing was carried out with He for half
201 an hour before the solution was required for application to the soil cores and continued during the
202 application process to avoid atmospheric N₂ contamination (a total of one and a half hours). The
203 amendment solution was manually withdrawn from the container with a glass syringe fitted with a
204 three-way valve onto the soil surface; care was taken to minimise contamination from atmospheric
205 N₂ entering the system. The syringe content was injected to the soil cores via the inlets on the lids
206 consecutively in each lid (three cores) and all vessels, completing a total of 36 applications that lasted
207 about 45 minutes. Incubation continued for twelve days, and the evolution of N₂O, N₂ and CO₂ was
208 measured continuously. At the end of each incubation experiment, the soils were removed from the

209 incubation vessels for further analysis. The three cores in each incubation vessel were ~~pulled~~
210 ~~togetherpooled~~ in one sample and subsamples taken and analysed for mineral N, total N and C and
211 moisture status. ~~The results of the soil analysis for all cores are presented in Table 3..~~

212 **2.6 Gas measurements**

213 Gas samples were directed to the relevant analysers via an automated injection valve fitted with 2
214 loops to direct the sample to two gas chromatographs. Emissions of N₂O and CO₂ were measured by
215 Gas Chromatography (GC), fitted with an Electron Capture Detector (ECD) and separation achieved
216 by a stainless steel packed column (2 m long, 4 mm bore) filled with ‘Porapak Q’ (80–100 mesh) and
217 using N₂ as the carrier gas. The detection limit for N₂O was equivalent to 2.3 g N ha⁻¹ d⁻¹. The N₂ was
218 measured by GC with a He Ionisation Detection (HID) and separation achieved by a PLOT column
219 (30 m long 0.53 mm i.d.), with He as the carrier gas. The detection limit was 9.6 g N ha⁻¹ d⁻¹. The
220 response of the two GCs was assessed by measuring a range of concentrations for N₂O, CO₂ and N₂.
221 Parent standards of the mixtures 10133 ppm N₂O + 1015.8 ppm N₂; 501 ppm N₂O + 253 ppm N₂ and
222 49.5 ppm N₂O + 100.6 ppm N₂ were diluted by means of Mass Flow controllers with He to give a
223 range of concentrations of: for N₂O of up to 750 ppm and for N₂ 1015 ppm. For CO₂ a parent standard
224 of 30,100 ppm was diluted down to 1136 ppm (all standards were in He as the balance gas). Daily
225 calibrations were carried out for N₂O and N₂ by using the low standard and doing repeated
226 measurements. The temperature inside the refrigeration cabinet containing the incubation vessels was
227 logged on an hourly basis and checked at the end of the incubation. The gas outflow rates were also
228 measured and recorded daily, and subsequently used to calculate the flux.

229 **2.7 Measurement of N₂O isotopic signatures**

230 Gas samples for isotopocule analysis were collected in 115 ml serum bottles sealed with grey butyl
231 crimp-cap septa (Part No 611012, Altmann, Holzkirchen, Germany). The bottles were connected by
232 a Teflon tube to the end of the chamber vents and were vented to the atmosphere through a needle, to
233 maintain flow through the experimental system. Dual isotope and isotopocule signatures of N₂O, i.e.
234 δ¹⁸O of N₂O (δ¹⁸O-N₂O), average δ¹⁵N (δ¹⁵N^{bulk}) and δ¹⁵N from the central N-position (δ¹⁵N^α) were

235 analysed after cryo-focussing by isotope ratio mass spectrometry as described previously (Well *et al.*,
236 2008). ^{15}N site preference (SP) was obtained as $\text{SP} = 2 * (\delta^{15}\text{N}^\alpha - \delta^{15}\text{N}^{\text{bulk}})$. Dual isotope and
237 isotopocule ratios of a sample (R_{sample}) were expressed as ‰ deviation from $^{15}\text{N}/^{14}\text{N}$ and $^{18}\text{O}/^{16}\text{O}$
238 ratios of the reference standard materials (R_{std}), atmospheric N_2 and standard mean ocean water
239 (SMOW), respectively:

$$240 \quad \delta X = (R_{\text{sample}}/R_{\text{std}} - 1) \times 1000 \quad [2]$$

241 where $X = ^{15}\text{N}^{\text{bulk}}, ^{15}\text{N}^\alpha, ^{15}\text{N}^\beta$, or ^{18}O

242 **2.8 Data analysis and additional measurements undertaken**

243 The areas under the curves for the N_2O , CO_2 and N_2 data were calculated by using GenStat 11 (VSN
244 International Ltd, Hemel Hempstead, Herts, UK). The resulting areas for the different treatments were
245 analysed by applying analysis of variance (ANOVA). The isotopic ($^{15}\text{N}^{\text{bulk}}$, ^{18}O , and site preference
246 (SP) differences between the four treatment for the different sampling dates were analysed by two-
247 way ANOVA. We also used the Student's t test to check for changes in soil water content over the
248 course of the experiments.

249 Calculation of the relative contribution of the N_2O derived from bacterial denitrification
250 ($\%B_{\text{DEN}}$) was done according to Lewicka-Szczebak *et al.* (2015). The isotopic value of initially
251 produced N_2O , *i.e.* prior to its partial reduction (δ_0) was determined using a Rayleigh model (Mariotti
252 *et al.*, 1982), where δ_0 is calculated using the fractionation factor of N_2O reduction ($\eta_{\text{N}_2\text{O}-\text{N}_2}$) for SP and
253 the fraction of residual N_2O ($r_{\text{N}_2\text{O}}$) which is equal to the $\text{N}_2\text{O}/(\text{N}_2+\text{N}_2\text{O})$ product ratio obtained from
254 direct measurements of N_2 and N_2O flux. An endmember mixing model ~~is~~ was then used to calculate
255 the percentage of bacterial N_2O in the total N_2O flux ($\%B_{\text{DEN}}$) from calculated δ_0 values and the SP
256 and $\delta^{18}\text{O}$ endmember values of bacterial denitrification and fungal denitrification/nitrification. The
257 range in endmember and $\eta_{\text{N}_2\text{O}-\text{N}_2}$ values assumed (adopted from Lewicka-Szczebak, 20167a) to
258 calculated maximum and minimum estimates of $\%B_{\text{DEN}}$ is given in Table 4.

259 Because both, endmember values and $\eta_{\text{N}_2\text{O}-\text{N}_2}$ values are not constant but subject to the given
260 ranges, we calculated here several scenarios using combinations of maximum, minimum and average

261 endmember and $\eta_{\text{N}_2\text{O}-\text{N}_2}$ values (Table 4) to illustrate the possible range of %B_{DEN} for each sample.
262 For occasional cases where %B_{DEN} > 100% the values were set to 100%.

263 At the same time as preparing the main soil blocks, a set of replicate samples was prepared in
264 exactly the same manner, but in smaller cores (i.d: 50 mm; h: 25 mm). On these samples we analysed
265 soil mineral N, total N and C and moisture at the start of the incubation. The same parameters were
266 measured after incubation by doing destructive sampling from the cores. Mineral N (NO₃⁻, NO₂⁻ and
267 NH₄⁺) was analysed after extraction with KCl by means of a segmented flow analyser using a
268 colorimetric technique (Searle, 1984). Total C and N in the air dried soil were ~~analysed~~ determined
269 using a thermal conductivity detector (TCD, Carlo Erba, model NA2000). Soil moisture was
270 determined by gravimetric analysis after drying at 105°C.

271 **3 Results**

272 **3.1 Soil composition**

273 The results after moisture adjustment at the start of the experiment resulted in a range of WFPS of
274 100 to 71% for the 4 treatments (Table 2). The results from the end of the incubation also confirmed
275 that there remained significant differences in soil moisture between the high moisture treatments
276 (SAT/sat and HALFSAT/sat) and the two lower moisture treatments (Table 3; one-way ANOVA,
277 $p < 0.05$). Soil in the two wettest states lost statistically significant amounts of water (10% ($p = 0.006$)
278 and 4.4% ($p < 0.001$) for SAT/sat and HALFSAT/sat, respectively) over the course of the 13-day
279 incubation experiment. This was inevitable as there was no way to hold a high (near-saturation) matric
280 potential once the soil was inside the DENIS assembly, and water would have begun to drain by
281 gravitational forces out of the largest macropores (>30 μm). An additional factor was the continuous
282 He/O₂ delivery over the soil surface which would have caused some drying. We accepted these as
283 unavoidable features of the experimental set-up, but we ~~suggest~~ assume that the main response of the
284 gaseous emissions occurred under the initial conditions, prior to the loss of water over subsequent
285 days. Soil in the two drier conditions had no significant change in their water content over the
286 experimental period ($p = 0.153$ and 0.051 for UNSAT/sat and UNSAT/halfsat, respectively). The

287 results of the initial soil composition were, for mineral N: 85.5 mg NO₃⁻-N kg⁻¹ dry soil, 136.2 mg
288 NH₄⁺-N kg⁻¹ dry soil. The mineral N contents of the soils at the end of the incubation are reported in
289 Table 3 showing that NO₃⁻ was very small in treatments SAT/sat and HALFSAT/sat (~1 mg N kg⁻¹
290 dry soil) compared to UNSAT/sat and UNSAT/halfsat (50-100 mg N kg⁻¹ dry soil) at the end of the
291 incubation. Therefore, there was a significant difference in soil NO₃⁻ between the former, high
292 moisture treatments and the latter drier (UNSAT) treatments which were also significantly different
293 between themselves (p<0.001 for both). The NH₄⁺ content was similar in treatments SAT/sat,
294 HALFSAT/sat and UNSAT/sat (~100 mg N kg⁻¹ dry soil), but slightly lower in treatment
295 UNSAT/halfsat (71.3 mg N kg⁻¹ dry soil), however overall differences were not significant probably
296 due to the large variability on the driest treatment (p>0.05).

297 **3.2 Gaseous emissions of N₂O, CO₂ and N₂**

298 The results for All datasets of N₂O and N₂ emissions showed normal distribution (Fpr.<0.001). The
299 treatments SAT/sat and HALFSAT/sat for all three gases, N₂O, CO₂ and N₂ showed fluxes that were
300 well replicated for all the vessels (see Fig. 1), in contrast for UNSAT/sat and UNSAT/halfsat the
301 emissions between the various replicated vessel in each treatment was not as consistent, leading to a
302 larger within treatment variability in the magnitude and shape of the GHG fluxes measured. The
303 cumulative fluxes also resulted in larger variability for the drier treatments (Table 3).

304 *Nitrous oxide and nitrogen gas.* The general trend was that the N₂O concentrations in the
305 headspace increased shortly after the application of the amendment (Fig. 1). The duration of the N₂O
306 peak for each replicate soil samples was about three days, except for UNSAT/halfsat in which one of
307 the replicate soils exhibit a peak which lasted for about 5 days. The N₂O maximum in the SAT/sat
308 and HALFSAT/sat treatments was of similar magnitude (ea. 5.5 means of 5.5 and 6.5 kg N ha⁻¹ d⁻¹,
309 respectively) ~~and but not~~ those of UNSAT/sat and UNSAT/halfsat ~~also were comparable (at~~
310 ~~around means of 7.1 and 11.9~~ kg N ha⁻¹ d⁻¹, respectively). The N₂ concentrations always increased
311 before the soil emitted N₂O reached the maximum. The lag between both N₂O and N₂ peak for all
312 samples was only few hours. Peaks of N₂ generally lasted just over four days, except in

313 UNSAT/halfsat where one replicate lasted about 6 days (Fig. 1). Unlike in the N₂O data, there was
314 larger within treatment variability in the replicates for all four treatments. The standard deviations of
315 each mean (Table 3) also indicate the large variability in treatments UNSAT/sat and UNSAT/halfsat
316 for both N₂O and N₂.

317 The product ratios, i.e. N₂O/(N₂O+N₂) resulted in a peak just after amendment addition by ca.
318 0.73 (at 0.49 d), 0.65 (at 0.48 d), 0.99 (at 0.35 d) and 0.88 (at 0.42 d) for SAT/sat, HALFSAT/sat,
319 UNSAT/sat and UNSAT/halfsat, respectively, and then decreases gradually until day 3 where it
320 becomes nearly zero for the 2 wettest treatments, and stays stable for the driest treatments between
321 0.1-0.2 (see Table 5 where the daily means of these ratios are presented).

322 The cumulative areas of the N₂O and N₂ peaks analysed by one-way ANOVA resulted in no
323 significant differences between treatments for both N₂O and N₂ (Table 3). Due to the large variation
324 in treatments UNSAT/sat and UNSAT/halfsat we carried out a pair wise analysis by using a weighted
325 t-test (Cochran, 1957). This analysis resulted in treatment differences between SAT/sat and
326 HALFSAT/sat, HALFSAT/sat and UNSAT/sat, SAT/sat and UNSAT/sat, but only at the 10%
327 significance level (P < 0.1 for both N₂O and N₂). ~~It is possible that gases were trapped (particularly in~~
328 ~~the higher saturation treatments) due to low diffusion and thus possibly masked differences in N₂ and~~
329 ~~N₂O production since this fraction of gases was not detected (Harter et al. 2016).~~

330 The results ~~of total N emission (N₂O+N₂) (Table 3) showed that total N emission (N₂O+N₂)~~
331 ~~(Table 3) had a consistent decreasing trend, with decreasing~~ decreased between the highest and lowest
332 soil moisture*s* i.e. from 63.4 for SAT/sat (100% WFPS) to 34.1 kg N ha⁻¹ (71% WFPS) for
333 UNSAT/halfsat. The maximum cumulative N₂O occurred at around 80% WFPS (Fig. 2) whereas the
334 total N₂O+N₂ was largest at about 95% and for N₂ it was our upper treatment at 100% WFPS.

335 *Carbon dioxide.* The background CO₂ ~~values~~ fluxes (before amendment application, i.e. day
336 -1 to day 0) were high at around 30 kg C ha⁻¹ d⁻¹ and variable (not shown). The CO₂ concentrations
337 in the headspace increased within a few hours after amendment application. The maximum CO₂ flux
338 was reached earlier in the drier treatments (about 1-2 days; ~70 kg C ha⁻¹ d⁻¹) compared to the wettest

339 (3 days; $\sim 40 \text{ kg C ha}^{-1} \text{ d}^{-1}$) and former peaks were also sharper (Fig. 1). The cumulative CO_2 fluxes
340 were significantly larger in the two drier unsaturated treatments (ca. $400\text{-}420 \text{ kg C ha}^{-1}$) when
341 compared to the wetter more saturated treatment (ca. $280\text{-}290 \text{ kg C ha}^{-1}$, $P < 0.05$) (Table 3).

342 3.3 Isotopocules of N_2O

343 The $\delta^{15}\text{N}^{\text{bulk}}$ of the soil emitted N_2O in our study differed significantly among the four treatments and
344 between the seven sampling dates ($p < 0.001$ for both); there was also a significant treatment**sampling*
345 date interaction ($p < 0.001$). The maximum $\delta^{15}\text{N}^{\text{bulk}}$ generally occurred on day 3, except for SAT/sat
346 on day 4 (Table 6).

347 The maximum $\delta^{18}\text{O}\text{-N}_2\text{O}$ values were also found on day 3, except for SAT/sat which peaked
348 at day 2 (Table 6). Overall, the $\delta^{18}\text{O}\text{-N}_2\text{O}$ values varied significantly between treatment and sampling
349 dates ($p < 0.001$ for both), but there was no significant treatment**time* interaction ($p > 0.05$).

350 The site preference (SP) for the SAT/sat treatment had an initial maximum value on day 2
351 (6.3‰) which decreased thereafter in the period from day 3 to 5 to a mean SP values of the emitted
352 N_2O of 2.0‰ on day 5, subsequently rising to 8.4‰ on day 12 of the experiment (Table 6). The
353 HALFSAT/sat treatment had the highest initial SP values on day 2 and 3 (both 6.4‰), decreasing
354 again to a value of 2.0‰, but now on day 4 followed by subsequent higher SP values of up to 9.2‰
355 on day 7 (Table 6). The two driest treatments (UNSAT/sat and UNSAT/halfsat) both had an initial
356 maximum on day 3 (11.9‰ and 5.9‰, respectively), and in UNSAT/sat the SP value then decreased
357 to day 7 (3.9‰), but in UNSAT/halfsat treatment after a marginal decrease on day 4 (5.4‰) it then
358 increased throughout the experiment reaching 11.8‰ on day 12 (Table 6). The lowest SP values were
359 generally on day 1 in all treatments. Overall, for all parameters, there was more similarity between
360 the more saturated treatments SAT/sat and HALFSAT/sat, and between the two more dry and aerobic
361 treatments UNSAT/sat and UNSAT/halfsat.

362 The $\text{N}_2\text{O} / (\text{N}_2\text{O} + \text{N}_2)$ ratios vs SP for all treatments in the first two days (when N_2O was
363 increasing and the $\text{N}_2\text{O} / (\text{N}_2\text{O} + \text{N}_2)$ ratio was decreasing) shows a significant negative response of
364 the SP when the ratio increased (Fig. 3). This behaviour suggests that when the emitted gaseous N is

365 dominated by N₂O (ratio close to 1) the SP values will be slightly negative with an intercept of -2‰
366 (Fig. 3), i.e. within the SP range of bacterial denitrification. With decreasing N₂O / (N₂O + N₂) ratio
367 the SP values of soil emitted N₂O were increasing to values up to 8‰. This is in juxtaposition with
368 the situation when the N emissions are dominated by N₂ or N₂O is low, where the SP values of soil
369 emitted N₂O were much higher (Fig. 3), pointing to an overall product ratio related to an ‘isotopic
370 shift’ of 10 to 12.5‰. We fitted 3 functions through this data including a second degree polynomial,
371 a linear and logarithmic function. The fitted logarithmic function in Fig. 3, is in almost perfect
372 agreement with Lewicka-Szczebak *et al.* (2014). Lewicka-Szczebak *et al.* (2014) data fits on the top
373 left of Fig. 3 ~~(their values are for SP and ratio N₂O / (N₂O + N₂): 18.5, 0.18; 10.1, 0.19; 11, 0.28 and~~
374 ~~13.4, 0.24, respectively).~~

375 It has been reported that the combination of the isotopic signatures of N₂O potentially
376 identifies the contribution of processes other than bacterial denitrification (Köster *et al.*, 2015; Wu
377 Di *et al.*, 2016; Deppe *et al.*, 2017). The question arises to which extent the relationships between the
378 δ¹⁸O and δ¹⁵N^{bulk} and between δ¹⁸O and SP within the individual treatments denitrification
379 dynamics. We checked this to evaluate the robustness of isotope effects during N₂O reduction as a
380 prerequisite to calculate the percentage of bacterial denitrification in N₂O production so we have
381 carried out similar analysis with our data. The In our data, maximum δ¹⁸O and SP values, were
382 generally observed at or near the peak of N₂ emissions on days 2-3, independent of the moisture
383 treatment (Table 6 and Fig. 3). δ¹⁵N^{bulk} values of all treatments were mostly negative when N₂O fluxes
384 started to increase (day 1, Fig. 1, Table 6), except for UNSAT/halfsat in which the lowest value was
385 before amendment application, reaching their highest values between days 3 and 4 for when N₂O
386 fluxes were back to the low initial values, and then decreased during the remaining period. δ¹⁸O values
387 increased about 10 - 20‰ after day 1 reaching maximum values on days 2 or 3 in all treatments, while
388 SP increased in parallel, at least by 3‰ (SAT/sat) and up to 12‰ (UNSAT/sat). While δ¹⁸O exhibited
389 a steady decreasing trend after day 3, SP behaved opposite to δ¹⁵N^{bulk} with decreasing values while
390 δ¹⁵N^{bulk} was rising again after days 4 or 5.

391 We further explored the data by looking at the relationships between the $\delta^{18}\text{O}$ and $\delta^{15}\text{N}^{\text{bulk}}$ for
392 all the treatments. The $\delta^{18}\text{O}$ vs $\delta^{15}\text{N}^{\text{bulk}}$ for all treatments is presented separating the data in three
393 periods (Fig. 4): ‘-1’, with $\delta^{18}\text{O}$ vs $\delta^{15}\text{N}^{\text{bulk}}$ values 1 day prior to the moisture adjustment (and N and
394 C application); ‘1-2’, with values in the first 2 days after the addition of water, N and C were added
395 and N_2O emissions were generally increasing in all treatments; and, ‘3-12’, the period in days after
396 moisture adjustment and N and C addition when N_2O emissions generally decreased back to baseline
397 soil emissions. There was a strong and significant relationship ($P < 0.001$ and 0.05 , respectively)
398 between $\delta^{18}\text{O}$ vs $\delta^{15}\text{N}^{\text{bulk}}$ for the high moisture treatments ($R^2 = 0.973$ and 0.923 for SAT/sat and
399 HALFSAT/sat, respectively) at the beginning of the incubation (‘1-2’) when the N_2O emissions are
400 still increasing, in contrast to those of the lower soil moisture treatments that were lower and not
401 significant ($R^2 = 0.294$ and 0.622 , for UNSAT/sat and UNSAT/halfsat, respectively). The
402 relationships between $\delta^{18}\text{O}$ vs $\delta^{15}\text{N}^{\text{bulk}}$ of emitted N_2O for the ‘3-12’ period were significant for
403 SAT/sat and HALFSAT/sat with R^2 values between 0.549 and 0.896 and P values < 0.05 and 0.001 ,
404 respectively (Fig. 4). Regressions were also significant for this period for the driest treatments
405 ($P < 0.001$). Interestingly, with decreasing soil moisture content (Fig. 4a to 4d) the regression lines of
406 ‘1-2’ and ‘3-12’ day period got closer together in the graphs. Overall, the $\delta^{15}\text{N}^{\text{bulk}}$ isotopic distances
407 between the two lines was larger for a given $\delta^{18}\text{O}$ - N_2O value for SAT/sat and HALFSAT/sat (ca.
408 20‰) when compared to the UNSAT/sat and UNSAT/halfsat treatments (ca. 13‰) (Fig. 4). So it
409 seems the $\delta^{15}\text{N}^{\text{bulk}} / \delta^{18}\text{O}$ - N_2O signatures are more similar for the drier soils than the two wettest
410 treatments. In addition, Fig 4 exactly reflects the 2-pool dynamics with increasing $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$
411 while the product ratio goes down (days 2,3), then only $\delta^{15}\text{N}$ continue increasing due to fractionation
412 of the NO_3^- during exhaustion of pool 1 in the wet soil (days 3,4,5), finally as pool 1 is depleted and
413 more and more comes from pool 2, the product ratio increases somewhat, and $\delta^{15}\text{N}$ decreases
414 somewhat since pool 2 is less fractionated and also $\delta^{18}\text{O}$ decreases due to slightly increasing product
415 ratio. Note that the turning points of $\delta^{18}\text{O}$ and product ratio (Table 3 and 4) for the wetter soils almost
416 coincide.

417 Similarly to Fig. 4, $\delta^{18}\text{O}$ vs the SP (Fig. 5) was analysed for the different phases of the
418 experiment. Generally, the slopes (Table 7) for days 1-2 for the three wettest treatments were similar
419 (~ 0.2 - 0.3) following the range of known reduction slopes and also had high and significant ($P < 0.05$)
420 regression coefficients ($R^2 = 0.65, 0.90$ and 0.87 for SAT/sat, HALFSAT/Sat and UNSAT/sat,
421 respectively). The slopes on days 3-5 were variable but slightly similar on days 7-12 (between 0.41 and
422 0.68) for the same three treatments. They were only significant for the 2 driest treatments ($P < 0.05$).
423 On days 7-12 SAT/sat and UNSAT/sat gave significant correlations ($P < 0.001$ and 0.05 , respectively).
424 Figure 5 also shows the “map” for the values of SP and $\delta^{18}\text{O}$ from all treatments. Reduction lines
425 (vectors) represent minimum and maximum routes of isotopocules values with increasing N_2O
426 reduction to N_2 based on the reported range in the ratio between the isotope fractionation factors of
427 N_2O reduction for SP and $\delta^{18}\text{O}$ (Lewicka-Szczebak et al., (20167a)). Most samples are located within
428 the vectors (from Lewicka-Szczebak *et al.* 20167a) area of N_2O production by bacterial denitrification
429 with partial N_2O reduction to N_2 (within uppermost and lowermost N_2O reduction vectors
430 representing the extreme values for the bacterial endmember and reduction slopes). Only a few values
431 of the UNSAT/sat and UNSAT/halfsat treatments are located above that vector area and more close
432 or within the vector area of mixing between bacterial denitrification and fungal
433 denitrification/nitrification.

434 The estimated ranges of the proportion of emitted N_2O resulting from bacterial denitrification
435 ($\%B_{\text{DEN}}$) were on day 1 and 2 after the amendment comparable in all four moisture treatments (Table
436 6). However, during day 3 to 12 the $\%B_{\text{DEN}}$ ranged from 78-100% in SAT/sat and 79-100%
437 HALFSAT/Sat, which was generally higher than that estimated at 54-86% for UNSAT/halfsat
438 treatment. The $\%B_{\text{DEN}}$ of the UNSAT/halfsat in that period was intermediate between SAT/sat and
439 UNSAT/sat with range of range 60-100% (Table 6). The final values were similar to those on day -1
440 except for the UNSAT/sat treatment.

441 **4 Discussion**

442 **4.1 N_2O and N_2 fluxes**

443 The observed decrease in total N emissions with decreasing initial soil moisture reflects the effect of
444 soil moisture as reported in previous studies (Well *et al.*, 2006). The differences when comparing the
445 cumulative fluxes however, were only marginally ($p < 0.1$) significant (Table 3) mostly due to large
446 variability within replicates in the drier treatments (see Fig. 1b). Davidson *et al.* (1991) provided a
447 WFPS threshold for determination of source process, with a value of 60% WFPS as the borderline
448 between nitrification and denitrification as source processes for N₂O production. The WFPS in all
449 treatments in our study was larger than 70%, above this 60% threshold, and referred to as the
450 “optimum water content” for N₂O by Scheer *et al.* (2009), so we can be confident that denitrification
451 was likely to have been the main source process in our experiment. In addition, Bateman *et al.* (2004)
452 observed the largest N₂O fluxes at 70% WFPS on a silty loam soil, lower than the 80% value for the
453 largest fluxes from the clay soil in our study (Fig. 2) suggesting that this optimum value could change
454 with soil type. Further, the maximum total measured N lost (N₂O+N₂) in our study occurred at about
455 95% WFPS (Fig. 2), but not many studies report N₂ fluxes for comparison and we are still missing
456 measurements of nitric oxide (NO) (Davidson *et al.*, 2000) and ammonia (NH₃) to account for the
457 total N losses. It is however possible that the N₂O+N₂ fluxes in the SAT/sat treatment were
458 underestimated due to low diffusivity in the water filled pores (Well *et al.*, 2001). ~~It is possible that~~
459 ~~gases would have been~~ ~~were~~ trapped (particularly in the higher saturation treatments) due to low
460 diffusion and thus possibly masked differences in N₂ and N₂O production since this fraction of gases
461 was not detected (Harter *et al.* 2016). It is worth mentioning that there was some drying during the
462 incubation. The flow of the gas is very slow (10 ml/min) simulating a low wind speed so normally
463 this would dry the soil in field conditions too. It would represent a rainfall event where the initial
464 moisture differs between treatments but some drying occurs due to the wind flow. We believe
465 however, that the effect of drying will be more relevant (and significant relative to the initial moisture)
466 later in the incubation.

467 The smaller standard errors in both N₂O and N₂ data for the larger soil moisture levels (Table
468 3 and Fig. 1) could suggest that at high moisture contents nutrient distribution (N and C) on the top

469 of the core is more homogeneous making replicate cores to behave similarly. At the lower soil
470 moisture for both N₂O and N₂, it is possible that some cracks appear on the soil surface causing
471 downwards nutrient movement, resulting in heterogeneity in nutrient distribution on the surface and
472 increasing variability between replicates, reflected in the larger standard errors of the fluxes. Laudone
473 *et al.* (2011) studied, using a biophysical model, the positioning of the hot-spot zones away from the
474 critical percolation path (described as ‘where air first breaks through the structure as water is removed
475 at increasing tensions’) and found it slowed the increase and decline in emission of CO₂, N₂O and N₂.
476 They found that hot-spot zones further away from the critical percolation path would reach the
477 anaerobic conditions required for denitrification in shorter time, the products of the denitrification
478 reactions take longer to migrate from the hot-spot zones to the critical percolation path and to reach
479 the surface of the system. The model and its parameters can be used for modelling the effect of soil
480 compaction and saturation on the emission of N₂O. They suggest that having determined biophysical
481 parameters influencing N₂O production, it remains to determine whether soil structure, or simply
482 saturation, is the determining factor when the biological parameters are constrained. Furthermore,
483 Clough *et al.* (2013) indicate that microbial scale models need to be included on larger models linking
484 microbial processes and nutrient cycling in order to consider spatial and temporal variation. Kulkarni
485 *et al.* (2008) refers to “hot spots” and “hot moments” of denitrification as scale dependant and
486 highlight the limitations for extrapolating fluxes to larger scales due to these inherent variabilities.
487 Well *et al.* (2003) found that under saturated conditions there was good agreement between laboratory
488 and field measurements of denitrification, and attributed deviations, under unsaturated conditions, to
489 spatial variability of anaerobic microsites and redox potential. Dealing with spatial variability when
490 measuring N₂O fluxes in the field remains a challenge, but the uncertainty could be potentially
491 reduced if water distribution is known. Our laboratory study suggests that soil N₂O and N₂ emission
492 for higher moisture levels would be less variable than for drier soils and suggests that for the former
493 a smaller number of spatially defined samples will be needed to get an accurate field estimate. This
494 applied to a lesser extent to the CO₂ fluxes.

495 Our results, for the two highest water contents (SAT/sat and HALFSAT/sat), indicated that
496 N₂O only contributed 20% of the total N emissions, as compared to 40-50% at the lowest water
497 contents (UNSAT/sat and UNSAT/halfsat, Table 3). This was due to reduction to N₂ at the high
498 moisture level, confirmed by the larger N₂ fluxes, favoured by low gas diffusion which increased the
499 N₂O residence time and the chance of further transformation (Klefoth *et al.*, 2014a). We should also
500 consider the potential underestimation of the fluxes in the highest saturation treatment due to
501 restricted diffusion in the water filled pores (Well *et al.*, 2001). A total of 99% of the soil NO₃⁻ was
502 consumed in the two high water treatments, whereas in the drier UNSAT/sat and UNSAT/halfsat
503 treatments there still was 35% and 70% of the initial amount of NO₃⁻ left in the soil, at the end of the
504 incubation, respectively (Table 3). The total amount of gas lost compared to the NO₃⁻ consumed was
505 almost 3 times for the wetter treatments, and less than twice for the 2 drier ones. This agrees with
506 denitrification as the dominant process source for N₂O with larger consumption of NO₃⁻ at the higher
507 moisture and larger N₂ to N₂O ratios (5.7, 4.7 for SAT/sat and HALFSAT/sat, respectively), whereas
508 at the lower moisture, ratios were lower (1.5 and 1.0 for UNSAT/sat and UNSAT/halfsat,
509 respectively) (Davidson, 1991). This also indicates that with WFPS above the 60% threshold for N₂O
510 production from denitrification, there was an increasing proportion of anaerobic microsites with
511 increase in saturation controlling NO₃⁻ consumption and N₂/N₂O ratios in an almost linear manner.
512 With WFPS values between 71-100 % and N₂/N₂O between 1.0 and 5.7, a regression can be
513 estimated: $Y=0.1723 X - 11.82$ ($R^2=0.8585$), where Y is N₂/N₂O and X is %WFPS. In summary, we
514 propose that heterogeneous distribution of anaerobic microsites could have been the limiting factor
515 for complete depletion of NO₃⁻ and conversion to N₂O in the two drier treatments. In addition, in the
516 UNSAT/halfsat treatment there was a decrease in soil NH₄⁺ at the end of the incubation (almost 50%;
517 Table 3) suggesting nitrification could have been occurring at this water content which also agrees
518 with the increase in NO₃⁻, even though WFPS was relatively high (>71%) (Table 3). It is important
519 to note that as we did not assess gross nitrification, the observed net nitrification based on lowering
520 in NH₄⁺ could underestimate gross nitrification since there might have been substantial N

521 mineralisation during the incubation. However, under conditions favouring denitrification at high soil
522 moisture the typical N₂O produced from nitrification is much lower compared to that from
523 denitrification (Lewicka-Szczebak *et al.*, 20167a) with the maximum reported values for the N₂O
524 yield of nitrification of 1-3 % (e.g. Deppe *et al.*, 2017). If this is the case, nitrification fluxes could
525 not have exceeded 1 kg N with NH₄⁺ loss of < 30 kg * 3% ~1 kg N. This would have represented for
526 the driest treatment, if conditions were suitable only for one day, that nitrification-derived N₂O would
527 have been 6% of the total N₂O produced. Loss of NH₃ was not probable at such low pH (5.6). The
528 corresponding rate of NO₃⁻ production using the initial and final soil contents and assuming other
529 processes were less important in magnitude, would have been < 1 mg NO₃⁻-N kg dry soil⁻¹ d⁻¹ which
530 is a reasonable rate (Hatch *et al.*, 2002). The other three treatments lost similar amounts of soil NH₄⁺
531 during the incubation (23-26%) which could have been due to some degree of nitrification at the start
532 of the incubation before O₂ was depleted in the soil microsites or due to NH₄⁺ immobilisation (Table
533 3) (Geisseler *et al.*, 2010).

534 ~~The CO₂ released in all treatments supports the statement above in relation with the more~~
535 ~~aerobic status of UNSAT/sat and UNSAT/halfsat, because the cumulative CO₂ flux is roughly 1.5~~
536 ~~times higher in the two drier treatments when compared to the wetter ones; but it could have also~~
537 ~~been the result of higher diffusion in the drier treatments.~~

538 A mass N balance, taking into account the initial and final soil NO₃⁻, NH₄⁺, added NO₃⁻ and
539 the emitted N (as N₂O and N₂) results in unaccounted N-loss of 177.2, 177.6, 130.6 and 110.8 mg N
540 kg⁻¹ for SAT/sat, HALFSAT/sat, UNSAT/sat and UNSAT/halfsat, respectively, that could have been
541 emitted as other N gases (such as NO), and some, immobilised in the microbial biomass. NO fluxes
542 reported by Loick *et al.* (2016) for example, result in a ratio N₂O/NO of 0.4. In summary unaccounted-
543 for N loss is two to three times the total measured gas loss (Table 3). In addition, in the SAT/sat
544 treatment there was probably an underestimation of the produced N₂ and N₂O due to restricted
545 diffusion at the high WFPS (e.g. Well *et al.*, 2001).

546 **4.2 Isotopocule trends.**

547 Trends of isotopocule values of emitted N₂O coincided with those of N₂ and N₂O fluxes. The results
548 from the isotopocule data (Table 6 and Fig. 3) also indicated that generally there were more isotopic
549 similarities between the two wettest treatments when compared to the two contrasting drier soil
550 moisture treatments.

551 Isotopocule values of emitted N₂O reflect multiple processes where all signatures are affected
552 by the admixture of several microbial processes, the extent of N₂O reduction to N₂ as well as the
553 variability of the associated isotope effects (Lewicka-Szczebak *et al.*, 2015). Moreover, for δ¹⁸O and
554 δ¹⁵N^{bulk} the precursor signatures are variable (Decock and Six, 2013), for δ¹⁸O the O exchange with
555 water can be also variable (Lewicka-Szczebak *et al.*, 2016**7b**). Since the number of influencing factors
556 clearly exceeds the number of isotopocule values, unequivocal results can only be obtained if certain
557 processes can be excluded or be determined independently, (Lewicka-Szczebak *et al.*, 2015; Lewicka-
558 Szczebak, 2016**7a**). The two latter conditions were fulfilled in this study, i.e. N₂O fluxes were high
559 and several order of magnitude above possible nitrification fluxes, since the N₂O – to- NO₃⁻ ratio
560 yield of nitrification products rarely exceeds 1% (Well *et al.*, 2008; Zhu *et al.*, 2012). Moreover, N₂
561 fluxes and thus N₂O reduction rates were exactly quantified.

562 The estimated values of % B_{DEN} indicate that in the period immediately after amendment
563 application all moisture treatments were similar, reflecting that the microbial response to N and C
564 added was the same and denitrification dominated. This was the same for the rest of the period for
565 the wetter treatments. In the drier treatments, proportions decreased afterwards and were similar to
566 values before amendment application, possibly due to recovery of more aerobic conditions that could
567 have encouraged other processes to contribute. As N₂ was still produced in the driest treatment, (but
568 in smaller amounts), this indicated ongoing denitrifying conditions and thus large contributions to the
569 total N₂O flux from nitrification were not probable~~from nitrification were not probable~~, but some
570 occurred as suggested by NH₄⁺ consumption.

571 The trends observed reflect the dynamics resulting from the simultaneous application of
572 NO₃⁻ and labile C (glucose) on the soil surface as described in previous studies (Meijide *et al.*,

573 2010; Bergstermann *et al.*, 2011) where the same soil was used, resulting in two locally distinct
574 NO_3^- pools with differing denitrification dynamics. In the soil volume reached by the NO_3^- /glucose
575 amendment, denitrification was initially intense with high N_2 and N_2O fluxes and rapid isotopic
576 enrichment of the NO_3^- -N. When the NO_3^- and/or glucose of this first pool were exhausted, N_2 and
577 N_2O fluxes were much lower and dominated by the initial NO_3^- pool that was not reached by the
578 glucose/ NO_3^- amendment and that is less fractionated due to its lower exhaustion by denitrification,
579 causing decreasing trends in $\delta^{15}\text{N}^{\text{bulk}}$ of emitted N_2O .

580 This is also reflected in Fig 4 where N_2O fluxes from both pools exhibited correlations (and
581 mostly significant) between $\delta^{15}\text{N}^{\text{bulk}}$ and $\delta^{18}\text{O}$ due to varying N_2O reduction, but $\delta^{15}\text{N}^{\text{bulk}}$ values in
582 days 1 and 2 - i.e. the phase when Pool 1 dominated - were distinct from the previous and later phase.

583 The fit of $^{15}\text{N}^{\text{bulk}}/^{18}\text{O}$ data to two distinct and distant regression lines can be attributed to
584 two facts: Firstly, in the wet treatment (Fig 4a, b) Pool 1 was probably completely exhausted and
585 there was little NO_3^- formation from nitrification (indicated by final NO_3^- values close to 0, Table 3)
586 whereas the drier treatment exhibited substantial NO_3^- formation and high residual NO_3^- . Hence,
587 there was probably still some N_2O from Pool 1 after day 2 in the dry treatment but not in the wetter
588 ones. Secondly, the product ratios after day 2 of the drier treatments were higher (0.13 to 0.44)
589 compared to the wetter treatments (0.001 to 0.09). Thus the isotope effect of N_2O reduction was
590 smaller in the drier treatments, leading to a smaller upshift of $\delta^{15}\text{N}^{\text{bulk}}$ and thus more negative values
591 after day 2, i.e. with values closer to days 1 +2.

592 This finding further confirms that $\delta^{15}\text{N}/\delta^{18}\text{O}$ patterns are useful to identify the presence of
593 several N pools, e.g. typically occurring after application of liquid organic fertilizers which has
594 been previously demonstrated using isotopocule patterns (Koster *et al.*, 2015).

595 Interestingly, the highest $\delta^{15}\text{N}^{\text{bulk}}$ and $\delta^{18}\text{O}$ values of the emitted N_2O were found in the soils
596 of the HALFSAT/sat treatment, although it may have been expected that the highest isotope values
597 from the N_2O would be found in the wettest soil (SAT/sat) because N_2O reduction to N_2 is favoured
598 under water-saturated conditions due to extended residence time of produced N_2O (Well *et al.*, 2012).

599 However, $N_2O/(N_2+N_2O)$ ratios of the SAT/sat and SAT/halfsat treatments were not different (Table
600 5). Bol *et al.* (2004) also found that some estuarine soils under flooded conditions (akin to our
601 SAT/sat) showed some strong simultaneous depletions (rather than enrichments) of the emitted N_2O
602 $\delta^{15}N^{bulk}$ and $\delta^{18}O$ values. These authors suggested that this observation may have resulted from a flux
603 contribution of an ‘isotopically’ unidentified N_2O production pathway. Another explanation could be
604 complete consumption of some of the produced N_2O in isolated micro-niches in the SAT/sat treatment
605 due to inhibited diffusivity in the fully saturated pores space. N_2 formation in these isolated domains
606 would not affect the isotopocule values of emitted N_2O and this would thus result in lower apparent
607 isotope effects of N_2O reduction in water saturated environments as suggested by Well *et al.* (2012).

608 The SP values obtained were generally below 12‰ in agreement with reported ranges
609 attributed to bacterial denitrification: -2.5 to 1.8‰ (Sutka *et al.*, 2006); 3.1 to 8.9‰ (Well and
610 Flessa, 2009); -12.5 to 17.6‰ (Ostrom, 2011). The SP, believed to be a better predictor of the N_2O
611 source as it is independent of the substrate isotopic signature (Ostrom, 2011), has been suggested as
612 it can be used to estimate N_2O reduction to N_2 in cases when bacterial denitrification can be
613 assumed to dominate N_2O fluxes (Koster *et al.*, 2013; Lewicka-Szczebak *et al.*, 2015). There was a
614 strong correlation between the SP and $N_2O / (N_2O+N_2)$ ratios on the first 2 days of the incubation
615 for all treatments up until the N_2O reached its maximum (Fig. 3) which reflects the accumulation of
616 $\delta^{15}N$ at the alpha position during ongoing N_2O reduction to N_2 . Later on in the experiment beyond
617 day 3, this was not observed probably because in that period the product ratio remained almost
618 unchanged and very low (Table 6). Similar observations have been reported by Meijide *et al.* (2010)
619 and Bergstermann *et al.* (2011), as they also found a decrease in SP during the peak flux period in
620 total N_2+N_2O emissions, but only when the soil had been kept wet prior to the start of the
621 experiment (Bergstermann *et al.*, 2011). These results confirm from 2 independent studies
622 (Lewicka-Szczebak *et al.*, 2014) that there is a relationship between the product ratios and isotopic
623 signatures of the N_2O emitted. The $\delta^{18}O$ vs SP regressions indicate more similarity between the

624 three wettest treatments as well as high regression coefficients, suggesting this SP/ $\delta^{18}\text{O}$ ratio could
625 also be used to help identify patterns for emissions and their sources.

626 **4.3 Link to modelling approaches.**

627 Since isotopocule data could be compared to N_2 and N_2O fluxes, the variability of isotope effects of
628 N_2O production and reduction to N_2 by denitrification could be determined from this data set
629 (Lewicka-Szczebak *et al.*, 2015) and this included modelling the two pool dynamics discussed
630 above. It was demonstrated that net isotope effects of N_2O reduction ($\eta_{\text{N}_2\text{O}-\text{N}_2}$) determined for both
631 NO_3^- pools differed. Pool 1 representing amended soil and resulting in high fluxes but moderate
632 product ratio, exhibited $\eta_{\text{N}_2\text{O}-\text{N}_2}$ values and the characteristic $\eta^{18}\text{O}/\eta^{15}\text{N}$ ratios similar to those
633 previously reported, whereas for Pool 2 characterized by lower fluxes and very low product ratio,
634 the net isotope effects were much smaller and the $\eta^{18}\text{O}/\eta^{15}\text{N}$ ratios, previously accepted as typical
635 for N_2O reduction processes (i.e., higher than 2), were not valid. The question arises, if the poor
636 coincidence of Pool 2 isotopologue fluxes with previous N_2O reduction studies reflects the
637 variability of isotope effects of N_2O reduction or if the contribution of other processes like fungal
638 denitrification could explain this (Lewicka-Szczabak *et al.*, 2017). The latter explanation is
639 evaluated in section 4.3

640 ~~The question arises, if the poor coincidence of Pool 2 isotopocule fluxes with previous N_2O~~
641 ~~reduction studies reflects the variability of isotope effects of N_2O reduction or if the contribution of~~
642 ~~other processes like fungal denitrification could explain this.~~

643 -Liu *et al.* (2016) noted that on the catchment scale potential N_2O emission rates were
644 related to hydroxylamine and NO_3^- , but not NH_4^+ content in soil. Zou *et al.* (2014) found high SP
645 (15.0 to 20.1‰) values at WFPS of 73 to 89% suggesting that fungal denitrification and bacterial
646 nitrification contributed to N_2O production to a degree equivalent to that of bacterial denitrification.

647 To verify the contribution of fungal denitrification and/or hydroxylamine oxidation we can
648 first look at the $\eta_{\text{SP}_{\text{N}_2\text{O}-\text{NO}_3}}$ values calculated in the previous modelling study applied on the same
649 dataset, (Table 1, the final modelling Step, Lewicka-Szczebak *et al.*, 2015). For Pool 1 there are no

650 significant differences between the values of various treatments, SP_0 ranges from (-1.8 ± 4.9) to
651 $(+0.1 \pm 2.5)$. Pool 1 emission was mostly active in days 1-2, hence these values confirm the bacterial
652 dominance in the emission at the beginning of incubation, which originates mainly from the
653 amendment addition and represent similar pathway for all treatments. However, for the Pool 2
654 emission we could observe a significant difference when compared the two wet treatments (SAT/sat
655 and HALFSAT/sat: (-5.6 ± 7.0)) with the UNSAT/sat treatment $(+3.8 \pm 5.8)$. This represents the
656 emission from unamended soil which was dominating after the third day of the incubation and
657 indicates higher nitrification contribution for the drier treatment.

658 **4.4 Contribution of bacterial denitrification.**

659 An endmember mixing approach has been previously used to estimate the fraction of bacterial N_2O
660 ($\%B_{DEN}$), but without independent estimates of N_2O reduction (Zou *et al.*, 2014), but due to the
661 unknown isotopic shift by N_2O reduction, the ranges of minimum and maximum estimates were large,
662 showing that limited information is obtained without N_2 flux measurement.

663 In an incubation study with two arable soils, Koster *et al.* (2013) used $N_2O/(N_2+N_2O)$ ratios
664 and isotopocule values of gaseous fluxes to calculate SP of N_2O production (referred to as SP_0),
665 which is equivalent to SP_0 using the Rayleigh model and published values of $\eta_{N_2O-N_2}$. The
666 endmember mixing approach based on SP_0 was then used to estimate fungal denitrification and/or
667 hydroxylamine oxidation giving indications for a substantial contribution in a clay soil, but not in a
668 loamy soil. Here we presented for the first time an extensive data set with large range in product
669 ratios and moisture to calculate the contribution of bacterial denitrification ($\%B_{DEN}$) of emitted N_2O
670 from SP_0 . The uncertainty of this approach arises from three factors, (i) from the range of SP_0
671 endmember values for bacterial denitrification of -11 to 0 per mil and 30 to 37 for hydroxylamine
672 oxidation/fungal denitrification, (ii) from the range of net isotope effect values of N_2O reduction
673 ($\eta_{N_2O-N_2}$) for SP which vary from -2 to -8 per mil (Lewicka-Szczebak *et al.*, 2015), and iii) system
674 condition (open vs. closed) taken to estimate the net isotope effect (Wu *et al.*, 2016).

675 The observation that %B_{DEN} of emitted N₂O was generally high (63-100%) in the wettest
676 treatment (SAT/sat) was not unexpected. However interestingly %B_{DEN} in the HALFSAT/sat
677 treatment was very similar (71-98%), pointing to the role of the wetter areas of the soil
678 microaggregates contributing to high %B_{DEN} values. The slightly lower values, i.e. down 60% in
679 UNSAT/sat %B_{DEN} range of 60-100%, suggest that the majority of N₂O derived from bacterial
680 denitrification still results from the wetter microaggregates of the soils, despite the fact that the
681 macropores are now more aerobic. Only, when the micropores become partially wet, as in the
682 UNSAT/halfsat treatment, do the more aerobic soil conditions allow a higher contribution of
683 nitrification/fungal denitrification ranging from 0 - 46% (1 - % B_{DEN}, Table 6) on days 3-12 (Zhu *et*
684 *al.*, 2013). Differences in the contribution of nitrification/fungal denitrification between the flux
685 phases when different NO₃⁻ pools were presumably dominating are only indicated in the driest
686 treatment, since 1-%B_{DEN} was higher after day 2 (14 to 46%) compared to days 1+2 (0 to 33 %).
687 This larger share of nitrification/fungal denitrification can be attributed to the increasing
688 contribution from Pool 2 to the total flux as indicated by the modeling of higher SP₀ for Pool 2 (see
689 previous section and Lewicka-Szczebak *et al.* (2015). In addition, indication for elevated
690 contribution of processes other than bacterial denitrification were only evident in the drier
691 treatments during phases before and after N₂, N₂O fluxes were strongly enhanced by glucose
692 amendment. The data supply no clue whether the other processes were suppressed during the anoxia
693 induced by glucose decomposition or just masked by the vast glucose-induced bacterial N₂O fluxes.
694

695 **5 Conclusions**

696 The results from this study demonstrated that at high soil moisture levels, there was less variability
697 in N fluxes between replicates, potentially decreasing the importance of soil hot spots in emissions
698 at these moisture levels. At high moisture there also was complete depletion of nitrate confirming
699 denitrification as the main pathway for N₂O emissions, and due to less diffusion of the produced
700 N₂O, the potential for further reduction to N₂ increased. Under less saturation, but still relatively

701 high soil moisture, nitrification occurred. Isotopic similarities were observed between similar
702 saturation levels and patterns of $\delta^{15}\text{N}/\delta^{18}\text{O}$ and $\text{SP}/\delta^{18}\text{O}$ are suggested as indicators of source
703 processes.

704 **Acknowledgments**

705 The authors would like to thank the technical help from Mark Butler during the laboratory
706 incubation and Andrew Bristow and Patricia Butler for carrying out soil analysis. Also thanks to
707 Dan Dhanoa for advice on statistical analysis, and to Anette Gieseemann and Martina Heuer for help
708 in N_2O isotopic analyses. This study was funded by the UK Biotechnology and Biological Sciences
709 Research Council (BBSRC) with competitive grants BB/E001580/1 and BB/E001793/1.
710 Rothamsted Research is sponsored by the BBSRC.

711

712

713 **Figures**

714 **Figure 1.** Mean of the three replicates for N₂O, N₂ and CO₂ emissions from a. SAT/sat treatment; b.
715 HALFSAT/sat; c. UNSAT/sat; d. UNSAT/halfsat. Grey lines correspond to the standard error of the
716 means.

717 **Figure 2** Total N emissions (N₂O+N₂)-N, N₂O and N₂ vs WFPS. Fitted functions through each
718 dataset are also shown.

719 **Figure 3** Ratio N₂O / (N₂O + N₂) vs. Site Preference (SP) for all for treatments in the first two days.
720 A logarithmic function was fitted through the data, the corresponding equation and correlation
721 coefficient are given.

722 **Figure 4** δ¹⁸O vs δ¹⁵N_{bulk} in all treatments for three periods (day -1 in diamond symbol, days 1-2 in
723 square symbol and days 3-12 in triangle symbol, respectively) in the experiment: a. SAT/sat
724 treatment; b. HALFSAT/sat; c. UNSAT/sat; d. UNSAT/halfsat. Equations of fitted functions and
725 correlation coefficients are shown. Correlations are unadjusted, the P value tests if the slope is
726 different from zero.

727 **Figure 5** Site Preference vs δ¹⁸O in all treatments for three periods (day -1, days 1-2 and days 3-12)
728 in the experiment: a. SAT/sat treatment; b. HALFSAT/sat; c. UNSAT/sat; d. UNSAT/halfsat.
729 Equations of fitted functions and correlation coefficients are in Table 7 for 1-2, 3-5 and 7-12 (5-12
730 for c.). Endmember areas for nitrification, N; bacterial denitrification, D; fungal denitrification, FD
731 and nitrifier denitrification, ND and corresponding vectors or reduction lines (black solid lines) are
732 from Lewicka-Szczebak et al., (20167a), and represent minimum and maximum routes of
733 isotopocule values with increasing N₂O reduction to N₂ based on the reported range in the ratio
734 between the isotope fractionation factors of N₂O reduction for SP and δ¹⁸O (Lewicka-Szczebak et
735 al., 20167a).

736 **Tables**

737 **Table 1** Soil properties of the soil used in the experiment

738 **Table 2** The four saturation conditions used for the soil in the experiment

739 **Table 3** Contents of soil moisture, NO_3^- , NH_4^+ and C:N ratio and cumulative fluxes of N_2O and N_2
740 and CO_2 from all treatments at the end of the incubation.

741 **Table 4** Scenarios with different combinations of $\delta^{18}\text{O}$ and SP endmember values and $\eta\text{N}_2\text{O}-\text{N}_2$
742 values to calculate maximum and minimum estimates of % B_{DEN} (minimum, maximum and average
743 values adopted from Lewicka-Szczebak *et al.*, (2016).

744 **Table 5** Ratios $\text{N}_2\text{O} / (\text{N}_2\text{O} + \text{N}_2)$ for all treatments

745 **Table 6** The temporal trends in $\delta^{15}\text{N}_{\text{bulk}}$, $\delta^{18}\text{O}$, $\delta^{15}\text{N}_\alpha$, SP and % B_{DEN} for all experimental treatments

746 **Table 7** Equations of fitted functions and correlation coefficients corresponding to Figure 5 for Site
747 Preference vs $\delta^{18}\text{O}$ in all treatments for three periods.

748 **References**

- 749 Baggs, E.M., 2008. A review of stable isotope techniques for N₂O source partitioning in soils:
750 recent progress, remaining challenges and future considerations. *Rapid Commun. Mass Sp.*, 22,
751 1664-1672.
- 752 Baggs, E.M., Rees, R.M., Smith, K.A., Vinten, A.J.A., 2000. Nitrous oxide emission from soils
753 after incorporating crop residues. *Soil Use Manage.*, 16, 82-87.
- 754 Ball, B.C., Scott, A., Parker, J.P., 1999. Field N₂O, CO₂ and CH₄ fluxes in relation to tillage,
755 compaction and soil quality in Scotland. *Soil Till. Res.*, 53, 29-39.
- 756 Barré, P., Eglin, T., Christensen, B.T., Ciais, P., Houot, S., Kätterer, T., van Oort, F., Peylin, P.,
757 Poulton, P.R., Romanenkov, V., Chenu, C., 2010. Quantifying and isolating stable soil organic
758 carbon using long-term bare fallow experiments. *Biogeosciences*, 7, 3839-3850.
- 759 Bateman, E., Cadisch, G., Baggs, E., 2004. Soil water content as a factor that controls N₂O
760 production by denitrification and autotrophic and heterotrophic nitrification. *Controlling nitrogen*
761 *flows and losses. 12th Nitrogen Workshop, University of Exeter, UK, 21-24 September 2003*, 290-
762 292.
- 763 Bergstermann, A., Cardenas, L., Bol, R., Gilliam, L., Goulding, K., Meijide, A., Scholefield, D.,
764 Vallejo, A., Well, R., 2011. Effect of antecedent soil moisture conditions on emissions and
765 isotopologue distribution of N₂O during denitrification. *Soil Biol. Biochem.*, 43, 240-250.
- 766 Bol, R., Rockmann, T., Blackwell, M., Yamulki, S., 2004. Influence of flooding on delta N-15,
767 delta O-18, (1)delta N-15 and (2)delta N-15 signatures of N₂O released from estuarine soils - a
768 laboratory experiment using tidal flooding chambers. *Rapid Commun. Mass Sp.*, 18, 1561-1568.
- 769 Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R., Zechmeister-Boltenstern, S. 2013.
770 Nitrous oxide emissions from soils: how well do we understand the processes and their controls?
771 *Phil Trans R Soc B.*, 368: 20130122, <http://dx.doi.org/10.1098/rstb.2013.0122>.
- 772 Cardenas, L.M., Hawkins, J.M.B., Chadwick, D., Scholefield, D., 2003. Biogenic gas emissions
773 from soils measured using a new automated laboratory incubation system. *Soil Biol. Biochem.*, 35,
774 867-870.
- 775 Cardenas, L.M., Thorman, R., Ashlee, N., Butler, M., Chadwick, D., Chambers, B., Cuttle, S.,
776 Donovan, N., Kingston, H., Lane, S., Dhanoa, M.S., Scholefield, D., 2010. Quantifying annual N₂O
777 emission fluxes from grazed grassland under a range of inorganic fertiliser nitrogen inputs. *Agr.*
778 *Ecosyst. Environ.*, 136, 218-226.
- 779 Castellano, M.J., Schmidt, J.P., Kaye, J.P., Walker, C., Graham, C.B., Lin, H., Dell, C.J., 2010.
780 Hydrological and biogeochemical controls on the timing and magnitude of nitrous oxide flux across
781 an agricultural landscape. *Global Change Biol.*, 16, 2711-2720.
- 782 Clough, T.J., Muller, C., Laughlin, R.J., 2013. Using stable isotopes to follow excreta N dynamics
783 and N₂O emissions in animal production systems. *Animal : an international journal of animal*
784 *bioscience*, 7 Suppl 2, 418-426.
- 785 Cochran, W.G. and Cox, G.M., 1957. *Experimental Design*. John Wiley & Sons New York.
- 786 Crutzen, P.J., 1970. Influence of Nitrogen Oxides on Atmospheric Ozone Content. *Quarterly*
787 *Journal of the Royal Meteorological Society*, 96, 320.
- 788 Davidson, E.A., 1991. Fluxes of nitrous oxide and nitric oxide from terrestrial ecosystems. In:
789 *Microbial production and consumption of greenhouse gases: Methane, nitrogen oxides and*
790 *halomethanes*. J.E. Rogers and W.B. Whitman (eds.). American Society for Microbiology,
791 Washington, D.C., pp. 219-235.
- 792 Davidson, E.A., Hart, S.C., Shanks, C.A., Firestone, M.K., 1991. Measuring Gross Nitrogen
793 Mineralization, Immobilization, and Nitrification by N-15 Isotopic Pool Dilution in Intact Soil
794 Cores. *J. Soil Sci.*, 42, 335-349.
- 795 Davidson, E.A., Keller, M., Erickson, H.E., Verchot, L.V., Veldkamp, E., 2000. Testing a
796 conceptual model of soil emissions of nitrous and nitric oxides. *Bioscience*, 50, 667-680.
- 797 Davidson, E.A., Verchot, L.V., 2000. Testing the hole-in-the-pipe model of nitric and nitrous oxide
798 emissions from soils using the TRAGNET database. *Global Biogeochem. Cy.*, 14, 1035-1043.

799 Decock, C., Six, J., 2013. On the potential of delta O-18 and delta N-15 to assess N₂O reduction to
800 N₂ in soil. *Eur. J. Soil Sci.*, 64, 610-620.

801 del Prado, A., Merino, P., Estavillo, J.M., Pinto, M., Gonzalez-Murua, C., 2006. N₂O and NO
802 emissions from different N sources and under a range of soil water contents. *Nutr. Cycl.*
803 *Agroecosys.*, 74, 229-243.

804 Deppe, M., Well, R., Giesemann, A., Spott, O., Flessa, H. 2017. Soil N₂O fluxes and related
805 processes in laboratory incubations simulating ammonium fertilizer depots. *Soil Biol. Biochem.*,
806 104, 68-80.

807 Dobbie, K.E., Smith, K.A., 2001. The effects of temperature, water-filled pore space and land use
808 on N₂O emissions from an imperfectly drained gleysol. *Eur. J. Soil Sci.*, 52, 667-673.

809 Firestone, M.K., Davidson, E.A., 1989. Microbiological basis of NO and N₂O production and
810 consumption in soil. *Exchange of Trace Gases between Terrestrial Ecosystems and the Atmosphere*,
811 47, 7-21.

812 Geisseler, D., Horwath, W.R., Joergensen, R.G., Ludwig, B., 2010. Pathways of nitrogen utilization
813 by soil microorganisms - A review. *Soil Biol. Biochem.*, 42, 2058-2067.

814 Gregory, A.S., Bird, N.R.A., Whalley, W.R., Matthews, G.P., Young, I.M., 2010. Deformation and
815 Shrinkage Effects on the Soil Water Release Characteristic. *Soil Sci. Soc. Am. J.*, 74, 4.

816 Harter, J., Guzman-Bustamente, I., Kuehfuss, S., Ruser, R., Well, R., Spott, O., Kappler, A.,
817 Behrens, S. 2016. Gas entrapment and microbial N₂O reduction reduce N₂O emissions from a
818 biochar-amended sandy clay loam soil. *Scientific Reports*, 6.

819 Hatch, D.J., Sprosen, M.S., Jarvis, S.C., Ledgard, S.F., 2002. Use of labelled nitrogen to measure
820 gross and net rates of mineralization and microbial activity in permanent pastures following
821 fertilizer applications at different time intervals. *Rapid Commun. Mass Sp.*, 16, 2172-2178.

822 IPCC, 2006. 2006 IPCC Guidelines for National Greenhouse Gas Inventories. 2006 IPCC
823 Guidelines for National Greenhouse Gas Inventories, Prepared by the National Greenhouse Gas
824 Inventories Programme, Eggleston H.S., Buendia L., Miwa K., Ngara T. and Tanabe K. (eds).
825 Published: IGES, Japan.

826 Klefoth, R.R., Clough, T.J., Oenema, O., Groenigen, J.W., 2014a. Soil bulk density and moisture
827 content influence relative gas diffusivity and the reduction of nitrogen-15 nitrous oxide. *Vadose*
828 *Zone J.*, 13, 0089-0089.

829 Klefoth, R.R., Clough, T.J., Oenema, O., Van Groenigen, J.-W., 2014b. Soil Bulk Density and
830 Moisture Content Influence Relative Gas Diffusivity and the Reduction of Nitrogen-15 Nitrous
831 Oxide. *Vadose Zone J.*, 13.

832 Koster, J.R., Cardenas, L.M., Bol, R., Lewicka-Szczebak, D., Senbayram, M., Well, R., Giesemann,
833 A., Dittert, K., 2015. Anaerobic digestates lower N₂O emissions compared to cattle slurry by
834 affecting rate and product stoichiometry of denitrification - An N₂O isotopomer case study. *Soil*
835 *Biol. Biochem.*, 84, 65-74.

836 Koster, J.R., Well, R., Dittert, K., Giesemann, A., Lewicka-Szczebak, D., Muhling, K.H.,
837 Herrmann, A., Lammel, J., Senbayram, M., 2013. Soil denitrification potential and its influence on
838 N₂O reduction and N₂O isotopomer ratios. *Rapid Commun. Mass Sp.*, 27, 2363-2373.

839 Kulkarni, M.V., Groffman, P.M., Yavitt, J.B., 2008. Solving the global nitrogen problem: it's a gas!
840 *Frontiers in Ecology and the Environment*, 6, 199-206.

841 Laudone, G.M., Matthews, G.P., Bird, N.R.A., Whalley, W.R., Cardenas, L.M., Gregory, A.S.,
842 2011. A model to predict the effects of soil structure on denitrification and N₂O emission. *J.*
843 *Hydrol.*, 409, 283-290.

844 Lewicka-Szczebak, D., Augustin J., Giesemann A., Well R., 2016a. Quantifying N₂O reduction to
845 N₂ based on N₂O isotopocules - validation with independent methods (Helium incubation and ¹⁵N
846 gas flux method). *Biogeosciences*, 14, 711-732. (accepted manuscript).

847 Lewicka-Szczebak, D., Dyckmans, J., Kaiser, J., Marca, A., Augustin, J., Well, R., 2016b. Oxygen
848 isotope fractionation during N₂O production by soil denitrification. *Biogeosciences*, 13, 1129-1144.

849 Lewicka-Szczebak, D., Well, R., Bol, R., Gregory, A.S., Matthews, G.P., Misselbrook, T., Whalley,
850 W.R., Cardenas, L.M., 2015. Isotope fractionation factors controlling isotopocule signatures of soil-

851 emitted N₂O produced by denitrification processes of various rates. *Rapid Commun. Mass Sp.*, 29,
852 269-282.

853 Lewicka-Szczebak, D., Well, R., Koster, J.R., Fuss, R., Senbayram, M., Dittert, K., Flessa, H.,
854 2014. Experimental determinations of isotopic fractionation factors associated with N₂O production
855 and reduction during denitrification in soils. *Geochim. Cosmochim. Ac.*, 134, 55-73.

856 Liu, S.R., Herbst, M., Bol, R., Gottselig, N., Putz, T., Weymann, D., Wiekenkamp, I., Vereecken,
857 H., Bruggemann, N., 2016. The contribution of hydroxylamine content to spatial variability of N₂O
858 formation in soil of a Norway spruce forest. *Geochim. Cosmochim. Ac.*, 178, 76-86.

859 [Loick, N., Dixon, L., Abalos, D., Vallejo, A., Matthews, G.P., McGeough, K.L., Well, R., Watson,](#)
860 [C.J., Laughlin, R.J., Cardenas, L.M., 2016. Denitrification as a Source of Nitric Oxide Emissions](#)
861 [from a UK Grassland Soil. *Soil Biol. Biochem.*, 95, 1-7.](#)

862 Ludwig, B., Bergstermann, A., Priesack, E., Flessa, H., 2011. Modelling of crop yields and N₂O
863 emissions from silty arable soils with differing tillage in two long-term experiments. *Soil Till. Res.*,
864 112, 114-121.

865 Mariotti, A., Germon, J.C., Leclerc, A., 1982. Nitrogen isotope fractionation associated with the
866 NO₂-N₂O step of denitrification in soils. *Canadian J. Soil Sci.*, 62, 227-241.

867 Meijide, A., Cardenas, L.M., Bol, R., Bergstermann, A., Goulding, K., Well, R., Vallejo, A.,
868 Scholefield, D., 2010. Dual isotope and isotopomer measurements for the understanding of N₂O
869 production and consumption during denitrification in an arable soil. *Eur. J. Soil Sci.*, 61, 364-374.

870 Morley, N., Baggs, E.M., 2010. Carbon and oxygen controls on N₂O and N₂ production during
871 nitrate reduction. *Soil Biol. Biochem.*, 42, 1864-1871.

872 Mualem, Y., 1976. New model for predicting hydraulic conductivity of unsaturated porous-media.
873 *Water Resour. Res.*, 12, 513-522.

874 Muller, C. and Clough, T. J. 2014. Advances in understanding nitrogen flows and transformations:
875 gaps and research pathways. *J. Agric. Sci.*, 152: S34-S44.

876 Ostrom, N., Ostrom, P., 2011. The isotopomers of nitrous oxide: analytical considerations and
877 application to resolution of microbial production pathways. In: Baskaran M (ed). *Handbook*
878 *Environ Isot Geochem*. Springer: Berlin Heidelberg, 453-476.

879 Parton, W.J., Holland, E.A., Del Grosso, S.J., Hartman, M.D., Martin, R.E., Mosier, A.R., Ojima,
880 D.S., Schimel, D.S., 2001. Generalized model for NO_x and N₂O emissions from soils. *J. Geophys.*
881 *Res-Atmos.*, 106, 17403-17419.

882 Perez, T., Garcia-Montiel, D., Trumbore, S., Tyler, S., De Camargo, P., Moreira, M., Piccolo, M.,
883 Cerri, C., 2006. Nitrous oxide nitrification and denitrification N-15 enrichment factors from
884 Amazon forest soils. *Ecol. Appl.*, 16, 2153-2167.

885 Scheer, C., Wassmann, R., Butterbach-Bahl, K., Lamers, J.P.A., Martius, C., 2009. The relationship
886 between N₂O, NO, and N₂ fluxes from fertilized and irrigated dryland soils of the Aral Sea Basin,
887 Uzbekistan. *Plant Soil*, 314, 273-283.

888 Schmidt, U., Thoni, H., Kaupenjohann, M., 2000. Using a boundary line approach to analyze N₂O
889 flux data from agricultural soils. *Nutr. Cycl. Agroecosys.*, 57, 119-129.

890 Scholefield, D., Patto, P.M., Hall, D.M., 1985. Laboratory Research on the Compressibility of 4
891 Topsoils from Grassland. *Soil Till. Res.*, 6, 1-16.

892 Searle, P.L., 1984. The Berthelot or Indophenol Reaction and Its Use in the Analytical-Chemistry of
893 Nitrogen - a Review. *Analyst*, 109, 549-568.

894 Sutka, R.L., Ostrom, N.E., Ostrom, P.H., Breznak, J.A., Gandhi, H., Pitt, A.J., Li, F., 2006.
895 Distinguishing nitrous oxide production from nitrification and denitrification on the basis of
896 isotopomer abundances. *Appl. Environ. Microb.*, 72, 638-644.

897 Toyoda, S., Mutoke, H., Yamagishi, H., Yoshida, N., Tanji, Y., 2005. Fractionation of N₂O
898 isotopomers during production by denitrifier. *Soil Biol. Biochem.*, 37, 1535-1545.

899 Toyoda, S., Yoshida, N., 1999. Determination of nitrogen isotopomers of nitrous oxide on a
900 modified isotope ratio mass spectrometer. *Anal. Chem.*, 71, 4711-4718.

901 van der Weerden, T.J., Kelliher, F.M., de Klein, C.A.M., 2012. Influence of pore size distribution
902 and soil water content on nitrous oxide emissions. *Soil Research*, 50, 125-135.

903 van Genuchten, M.T., 1980. A closed form equation for predicting the hydraulic conductivity of
904 unsaturated soils. *Soil Sci. Soc. Am. J.*, 44, 892-898.

905 van Groenigen, J.W., Kuikman, P.J., de Groot, W.J.M., Velthof, G.L., 2005. Nitrous oxide emission
906 from urine-treated soil as influenced by urine composition and soil physical conditions. *Soil Biol.*
907 *Biochem.*, 37, 463-473.

908 Well, R., Augustin, J., Davis, J., Griffith, S.M., Meyer, K., Myrold, D.D., 2001. Production and
909 transport of denitrification gases in shallow ground water. *Nutr. Cycl. Agroecosys.*, 60, 65-75.

910 Well, R., Augustin, J., Meyer, K., Myrold, D.D., 2003. Comparison of field and laboratory
911 measurement of denitrification and N₂O production in the saturated zone of hydromorphic soils.
912 *Soil Biol. Biochem.*, 35, 783-799.

913 Well, R., Eschenbach, W., Flessa, H., von der Heide, C., Weymann, D., 2012. Are dual isotope and
914 isotopomer ratios of N₂O useful indicators for N₂O turnover during denitrification in nitrate-
915 contaminated aquifers? *Geochim. Cosmochim. Ac.*, 90, 265-282.

916 Well, R., Flessa, H., 2009. Isotopologue signatures of N₂O produced by denitrification in soils.
917 *J. Geophys. Res.-Biogeo.*, 114.

918 Well, R., Flessa, H., Xing, L., Ju, X.T., Romheld, V., 2008. Isotopologue ratios of N₂O emitted
919 from microcosms with NH₄⁺ fertilized arable soils under conditions favoring nitrification. *Soil Biol.*
920 *Biochem.*, 40, 2416-2426.

921 Well, R., Kurganova, I., de Gerenyu, V.L., Flessa, H., 2006. Isotopomer signatures of soil-emitted
922 N₂O under different moisture conditions - A microcosm study with arable loess soil. *Soil Biol.*
923 *Biochem.*, 38, 2923-2933.

924 Wu, D., Koster, J.R., Cardenas, L.M., Bruggemann, N., Lewicka-Szczebak, D., Bol, R., 2016. N₂O
925 source partitioning in soils using N-15 site preference values corrected for the N₂O reduction effect.
926 *Rapid Commun. Mass Sp.*, 30, 620-626.

927 Wu, D., Senbayram, M., Well, R., Bruggemann, N., Pfeiffer, B., Loick, N., Stempfhuber, B.,
928 Dittert, K., Bol, R. (2017) Nitrification inhibitors mitigate N₂O emissions more effectively under
929 straw-induced conditions favoring denitrification. *Soil Biol. Biochem.*, 104, 197-207.

930 Zhu, X., Burger, M., Doane, T.A., Horwath, W.R., 2013. Ammonia oxidation pathways and nitrifier
931 denitrification are significant sources of N₂O and NO under low oxygen availability. *P. Natl. Acad.*
932 *Sci. USA.*, 110, 6328-6333.

933 Zou, Y., Hirono, Y., Yanai, Y., Hattori, S., Toyoda, S., Yoshida, N., 2014. Isotopomer analysis of
934 nitrous oxide accumulated in soil cultivated with tea (*Camellia sinensis*) in Shizuoka, central Japan.
935 *Soil Biol. Biochem.*, 77, 276-291.

936

937

938

939 Table 1. Highfield soil properties

940

Property	Units	Highfield	941 942
Location		Rothamsted Research Herts.	943 944
Grid reference	GB National Grid	TL129130	945
	Longitude	00°21'48"W	946
	Latitude	51°48'18"N	947
Soil type	SSEW ^a group ^c	Paleo-argillic brown earth	948
	SSEW ^a series ^d	Batcombe	949
	FAO ^{bc}	Chromic Luvisol	950
Landuse		Grass; unfertilised; cut	951
pH		5.63	952
Sand (2000-63 µm)	g g ⁻¹ dry soil	0.179	953
Silt (63-2 µm)	g g ⁻¹ dry soil	0.487	954
Clay (<2 µm)	g g ⁻¹ dry soil	0.333	955
Texture	SSEW ^a class ^c	Silty clay loam	956
Particle density	g cm ⁻³	2.436	957
Organic matter	g g ⁻¹ dry soil	0.089	958
Water content for packing	g g ⁻¹ dry soil	0.37	958

959 ^aSoil Survey of England and Wales classification system

960 ^bUnited Nations Food and Agriculture Organisation World Reference Base for Soil Resources classification
961 system (approximation)

962 ^cAvery (1980)

963 ^dClayden & Hollis (1984)

964

965

966
967

Table 2. The four saturation conditions set for the Highfield soil.

Saturation condition	SAT/sat	HALFSAT/sat	UNSAT/sat	UNSAT/ <u>half</u> sat
Macropores	Saturated	Half-saturated	Unsaturated	Unsaturated
Micropores	Saturated	Saturated	Saturated	Half-saturated
<i>As prepared:</i>				
Matric potential, -kPa	4.1	12.3	27.3	136.9
Water content, g 100 g ⁻¹	47.7	42.5	37.2	29.4
Water content, cm ⁻³ 100 cm ⁻³	61.1	54.4	47.7	37.3
Water-filled pore space, %	98	91	82	68
Threshold pore size saturated, μm	73	24	11	2
<i>Final, following amendment:</i>				
Matric potential, -kPa	0	8.6	20.0	78.1
Water content, g 100 g ⁻¹	49.8	44.6	39.3	31.5
Water content, cm ⁻³ 100 cm ⁻³	63.8	57.1	50.4	40.0
Water-filled pore space, %	100	94	85	71
Threshold pore size saturated, μm	all	35	15	4

968
969
970

971
972
973
974

Table 3. Contents of soil moisture, NO₃⁻, NH₄⁺ and C:N ratio and cumulative fluxes of N₂O and N₂ and CO₂ from all treatments at the end of the incubation. Values in brackets are standard deviation of the mean of three values (emissions are expressed per area and soil weight basis).

Treatment	% Mean moisture	NO ₃ ⁻ , mg N kg ⁻¹ dry soil	NH ₄ ⁺ , mg N kg ⁻¹ dry soil	Total C, %	Total N, %	N ₂ O, kg N ha ⁻¹	N ₂ O, mg N kg ⁻¹ dry soil	N ₂ , kg N ha ⁻¹	N ₂ , mg N kg ⁻¹ dry soil	Total emitted N, kg N ha ⁻¹	CO ₂ , kg C ha ⁻¹
SAT/sat	39.8 (1.3)	1.1 (0.4)	104.3 (1.1)	3.61 (0.04)	0.35 (0.004)	9.4 (1.1)	7.8 (0.9)	54.0 (14.0)	44.8 (11.6)	63.4	289.2 (30.4)
HALFSAT/sat	40.2 (0.2)	0.8 (1.0)	104.2 (6.8)	3.64 (0.08)	0.36 (0.004)	10.9 (0.4)	9.0 (0.3)	51.7 (9.0)	42.8 (7.4)	62.6	283.0 (35.5)
UNSAT/sat	36.5 (2.1)	51.2 (37.4)	100.8 (5.7)	3.64 (0.10)	0.36 (0.007)	23.7 (11.0)	20.0 (9.5)	36.0 (28.5)	30.2 (23.7)	59.7	417.6 (57.1)
UNSAT/halfsat	34.3 (1.1)	100.6 (16.1)	71.3 (33.6)	3.53 (0.08)	0.36 (0.01)	16.8 (15.8)	14.0 (13.1)	17.2 (19.4)	14.3 (16.1)	34.1	399.7 (40.6)

975
976

977 Table 4: Scenarios with different combinations of $d^{18}\text{O}$ and Site Preference (SP) endmember values and $\eta_{\text{N}_2\text{O}}$
 978 N_2 values to calculate maximum and minimum estimates of %Bden (minimum, maximum and average values
 979 adopted from Lewicka-Szczabak et al., 2017^{6a}).
 980

	SPOBD	SPOFDN	η_{SP}	$\eta^{18}\text{O}$
model (min endmember plus η)	-11	30	-2	-12
model (max endmember plus η)	0	37	-8	-12
model (max endmember)	0	37	-5.4	-12
model (min endmember)	-11	30	-5.4	-12
model (max η)	-5	33	-8	-12
model (min η)	-5	33	-2	-12

981
 982
 983

984 Table 5. Ratios $N_2O / (N_2O + N_2)$ for all treatments
 985

Days	SAT/sat		HALFSAT/sat		UNSAT/halfsat		UNSAT/sat	
	mean	s.e.	mean	s.e.	mean	s.e.	mean	s.e.
-1	0.276	0.043	0.222	0.009	0.849	0.043	0.408	0.076
0	0.630	0.022	0.538	0.038	0.763	0.053	0.861	0.043
1	0.371	0.025	0.360	0.019	0.622	0.018	0.644	0.031
2	0.096	0.016	0.139	0.015	0.425	0.005	0.296	0.020
3	0.004	0.002	0.015	0.006	0.439	0.052	0.256	0.025
4	0.017	0.002	0.008	0.001	0.475	0.049	0.232	0.012
5	0.019	0.003	0.012	0.001	0.503	0.037	0.174	0.010
6	0.068	0.008	0.020	0.001	0.459	0.052	0.135	0.010
7	0.085	0.008	0.047	0.003	0.333	0.057	0.127	0.003
8	0.106	0.004	0.066	0.002	0.277	0.006	0.122	0.002
9	0.089	0.003	0.053	0.005	0.265	0.006	0.122	0.005
10	0.060	0.003	0.090	0.014	0.428	0.086	0.118	0.006
11	0.063	0.002	0.053	0.002	0.414	0.051	0.125	0.005

986
 987

988
989

Table 6. The temporal trends in $\delta^{15}\text{N}_{\text{bulk}}$, $\delta^{18}\text{O}$, $\delta^{15}\text{N}_{\alpha}$, Site Preference (SP) and %B_{DEN} for all experimental treatments (values in brackets are the standard deviation of the mean)

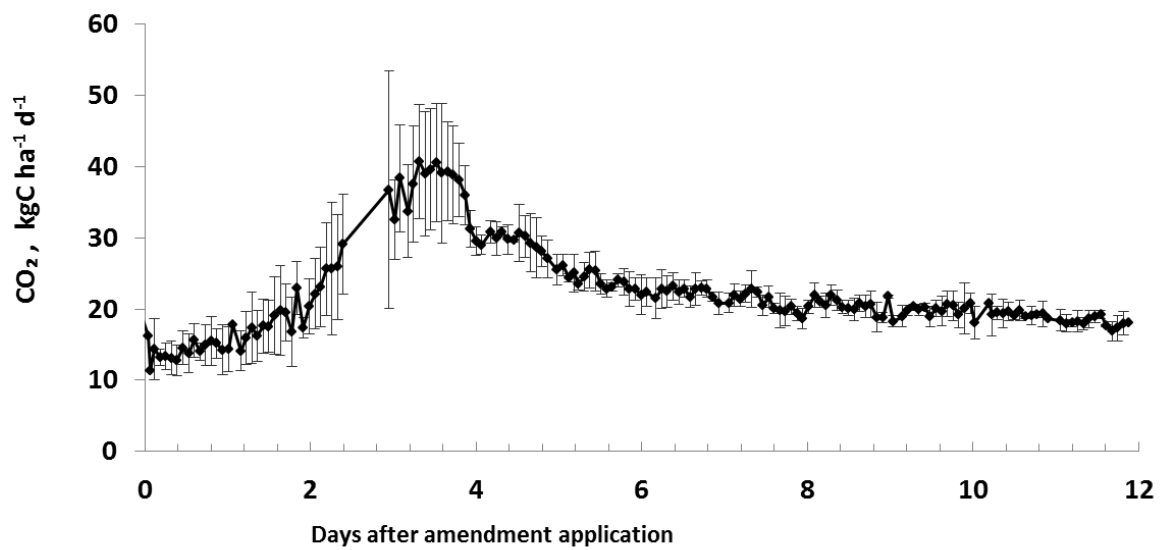
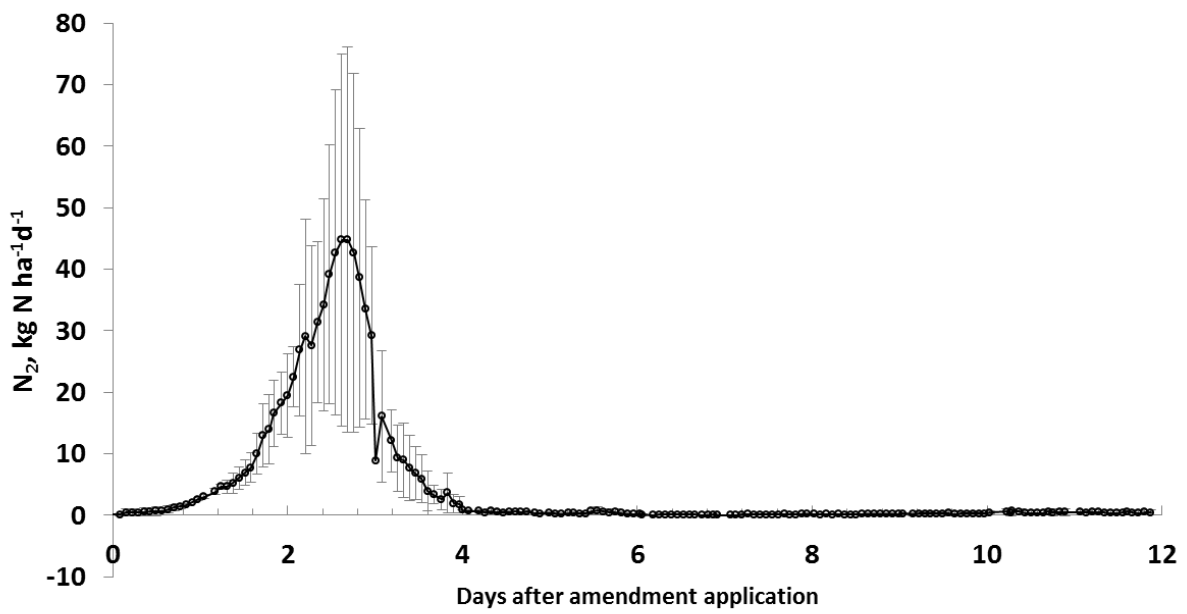
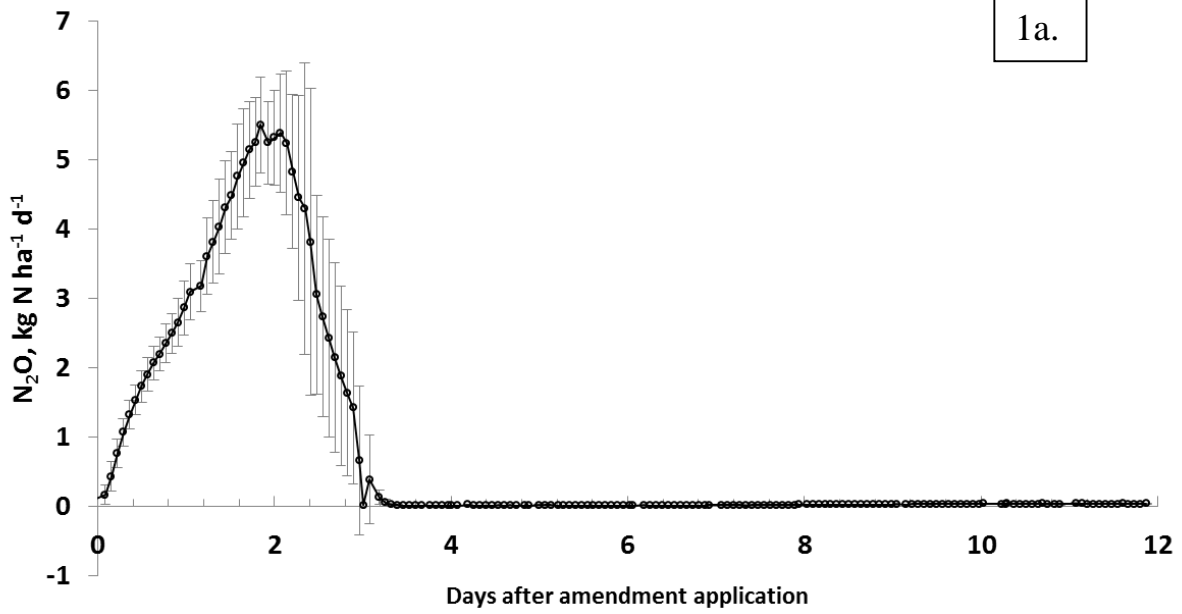
Day	$\delta^{15}\text{N}_{\text{bulkAIR}}$ (‰)			
	SAT/sat	HALFSAT/ <u>Satsat</u>	UNSAT/ <u>Satsat</u>	UNSAT/halfsat
-1	-3.8 (2.1)	-6.2 (1.5)	-14.2 (10.9)	-23.6 (1.1)
1	-18.9 (1.6)	-25.5 (4.6)	-20.3 (2.6)	-20.8 (2.3)
2	-7.7 (4.2)	-12.7 (2.7)	-12.2 (2.0)	-13.9 (5.7)
3	-2.4 (1.8)	14.0 (2.2)	-1.1 (7.6)	-4.4 (3.0)
4	-0.9 (2.2)	-0.3 (3.6)	-7.8 (4.6)	-9.3 (3.7)
5	-6.9 (0.9)	-4.3 (6.1)	-11.3 (3.7)	-8.9 (7.7)
7	-9.6 (1.5)	-10.0 (1.6)	-14.3 (4.7)	-13.4 (13.5)
12	-7.5 (1.2)	-8.6 (0.9)	-11.8 (2.6)	-21.3 (6.9)
	$\delta^{18}\text{O}_{\text{SMOW}}$ (‰)			
	SAT/sat	HALFSAT/ <u>Satsat</u>	UNSAT/ <u>Satsat</u>	UNSAT/halfsat
-1	33.3 (2.6)	32.7 (3.0)	31.4 (9.8)	25.2 (4.9)
1	42.9 (2.4)	37.1 (3.8)	32.3 (3.6)	33.3 (2.1)
2	54.0 (5.7)	48.7 (4.5)	42.7 (5.3)	40.5 (5.0)
3	45.7 (1.5)	59.7 (3.2)	53.4 (5.7)	41.2 (1.0)
4	42.5 (1.4)	42.0 (3.7)	38.1 (4.5)	39.9 (7.7)
5	36.0 (2.9)	34.6 (3.7)	30.4 (2.6)	36.5 (6.9)
7	32.2 (5.5)	31.6 (5.5)	28.4 (4.4)	32.7 (5.4)
12	34.9 (5.6)	34.1 (2.7)	32.4 (2.9)	28.5 (5.0)
	$\delta^{15}\text{N}_{\alpha\text{AIR}}$ (‰)			
	SAT/sat	HALFSAT/ <u>Satsat</u>	UNSAT/ <u>Satsat</u>	UNSAT/halfsat
-1	-0.3 (3.4)	-2.6 (1.8)	-9.5 (12.0)	-19.7 (2.1)
1	-17.4 (1.8)	-24.0 (5.8)	-20.2 (2.0)	-21.1 (2.6)
2	-4.6 (4.2)	-9.5 (3.6)	-11.1 (1.1)	-13.8 (5.9)
3	-0.8 (1.3)	17.2 (4.0)	7.6 (4.7)	-2.7 (3.2)
4	1.0 (2.5)	0.7 (2.2)	-3.5 (3.7)	-2.8 (7.7)
5	-5.9 (0.7)	-2.9 (5.4)	-9.4 (3.9)	-5.2 (7.9)
7	-7.8 (2.3)	-5.3 (4.2)	-12.3 (5.6)	-7.7 (11.5)
12	-3.3 (2.1)	-4.6 (0.6)	-8.1 (4.2)	-15.3 (5.5)
	SP _{AIR}			
	SAT/sat	HALFSAT/ <u>Satsat</u>	UNSAT/ <u>Satsat</u>	UNSAT/halfsat
-1	7.0 (3.9)	7.1 (4.2)	9.4 (2.1)	7.7 (1.9)
1	2.9 (0.6)	3.0 (2.3)	0.1 (1.8)	-0.7 (1.4)
2	6.3 (0.64)	6.4 (1.9)	2.2 (2.0)	0.2 (1.9)
3	3.3 (1.0)	6.4 (6.9)	11.9 (12.4)	5.9 (0.8)
4	3.7 (0.6)	2.0 (6.2)	8.7 (5.9)	5.4 (3.0)
5	2.0 (0.4)	3.0 (2.1)	3.9 (0.5)	7.4 (2.3)
7	5.0 (2.1)	9.2 (5.2)	3.9 (1.8)	11.2 (4.1)
12	8.4 (3.3)	7.9 (0.8)	7.3 (3.7)	11.8 (5.3)
	Estimated range of %B _{DEN}			
	SAT/sat	HALFSAT/sat	UNSAT/sat	UNSAT/halfsat
-1	63-100	60-100	53-85	56-84
1-2	68-100	67-100	73-100	77-100
3-12	78-100	79-100	60-100	54-86

990 Table 7. Equations of fitted functions and correlation coefficients corresponding to Figure 5 for Site
 991 Preference (SP) (Y axis) vs $\delta^{18}\text{O}$ (X axis) in all treatments for three periods. Correlations are
 992 unadjusted, the P value tests if the slope is different from zero.
 993

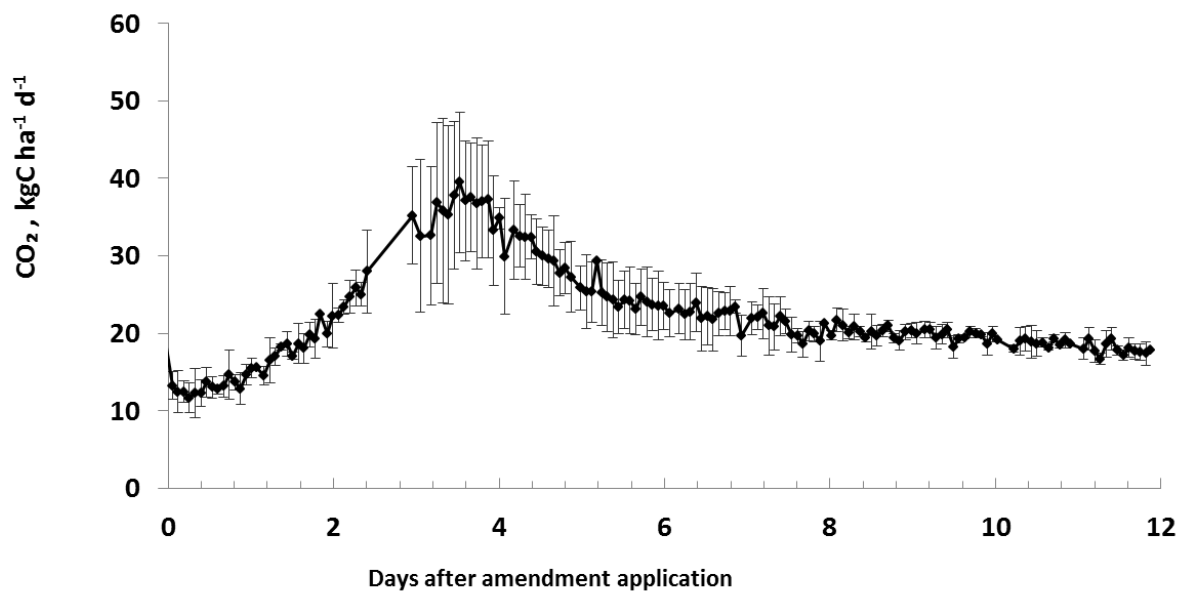
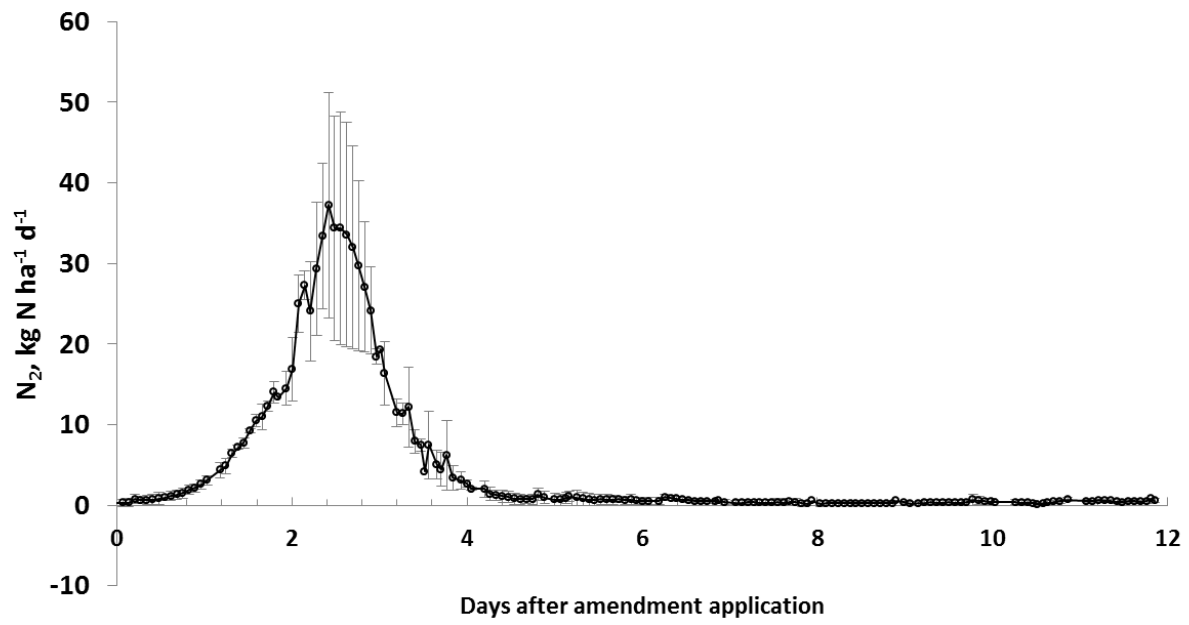
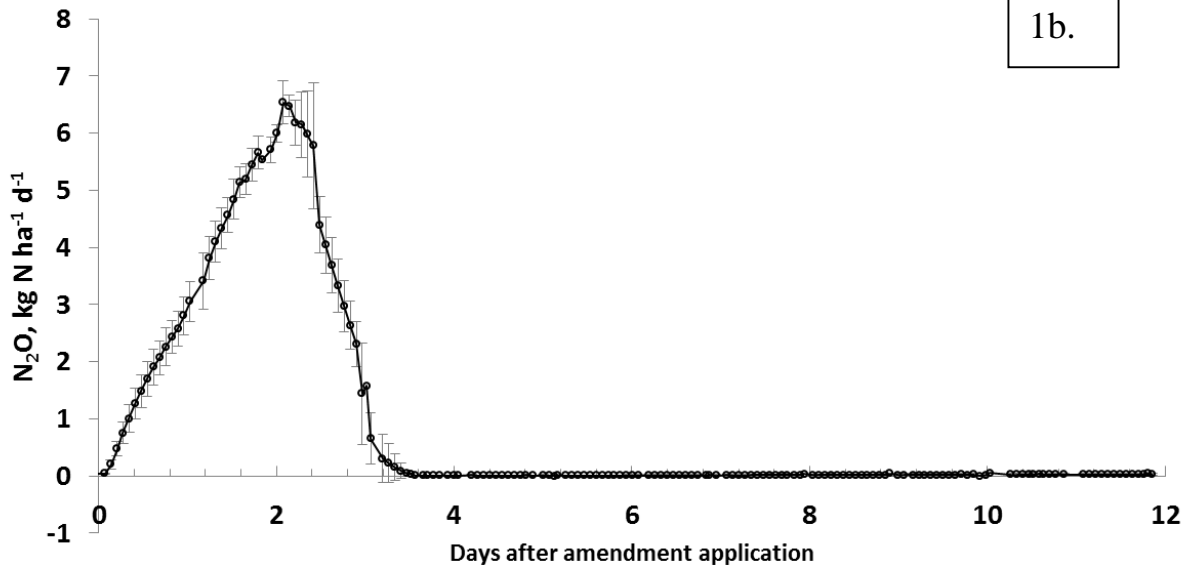
Treatment	Days 1-2	Days 3-5	Days 7-12
SAT/sat	$y = 0.2151x - 5.8386$, $R^2 = 0.6529$ P=0.05	$y = 0.1204x - 1.848$, $R^2 = 0.397$ P=0.129	$y = 0.5872x - 12.223$, $R^2 = 0.985$ P<0.001
HALFSAT/sat	$y = 0.3447x - 10.129$, $R^2 = 0.9048$ P=0.004	$y = 0.18x - 4.5966$, $R^2 = 0.1728$ P=0.266	$y = 0.4063x - 6.2632$, $R^2 = 0.6876$ P=0.171
UNSAT/sat	$y = 0.2709x - 8.9968$, $R^2 = 0.8664$ P=0.007	$y = 0.7248x - 18.874$, $R^2 = 0.507$ P=0.031	$y = 0.6848x - 15.236$, $R^2 = 0.7156$ P=0.034
UNSAT/halfsat	$y = -0.0146x + 0.2506$, $R^2 = 0.0024$ P=0.927	$y = 0.3589x - 7.2194$, $R^2 = 0.4839$ P=0.037	$y = -0.318x + 21.261$, $R^2 = 0.1491$ P=0.450

994
 995

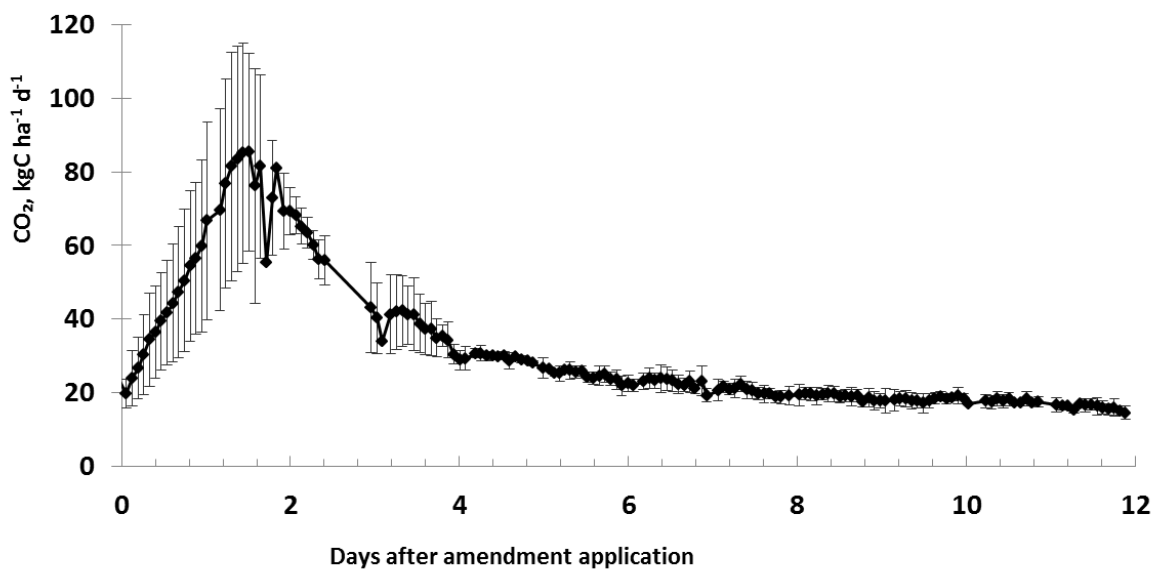
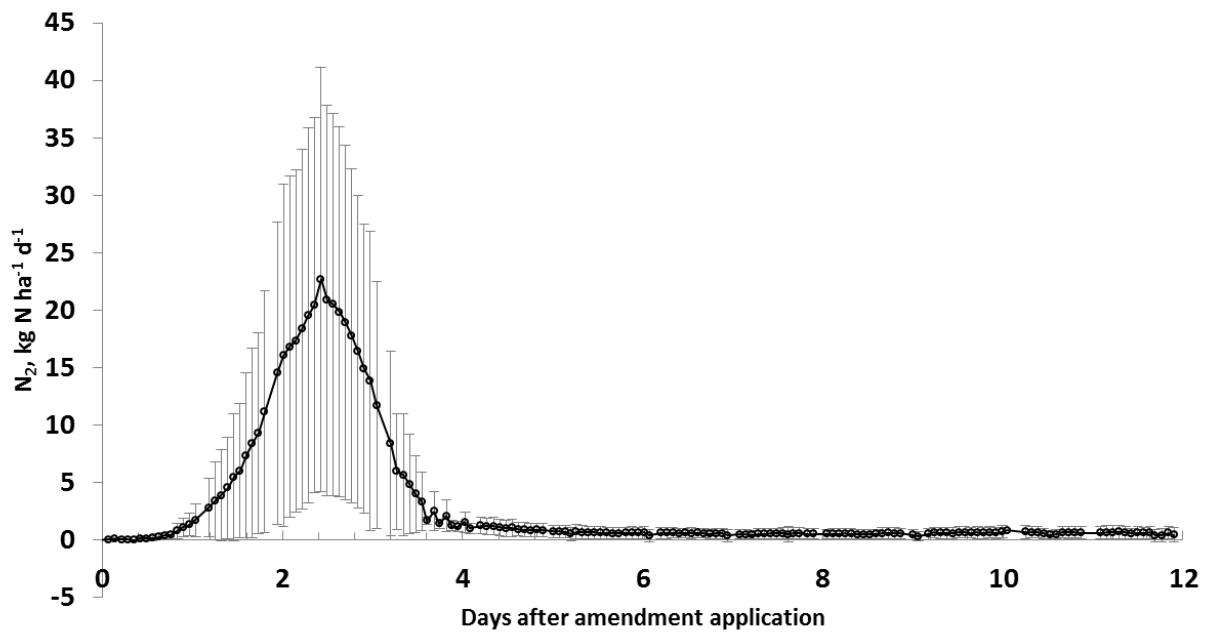
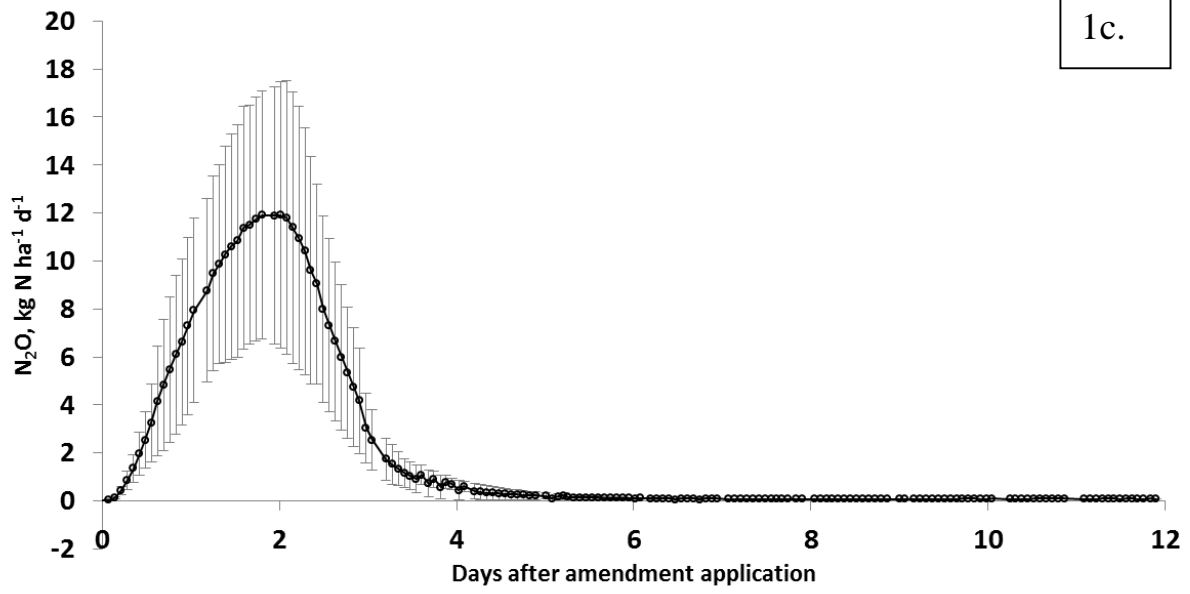
1a.



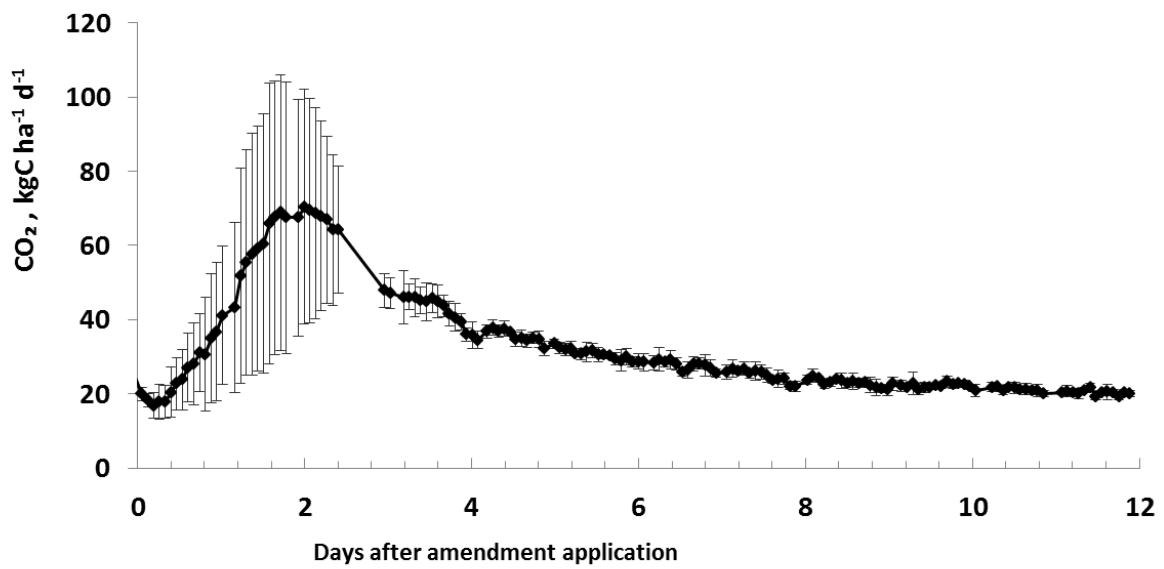
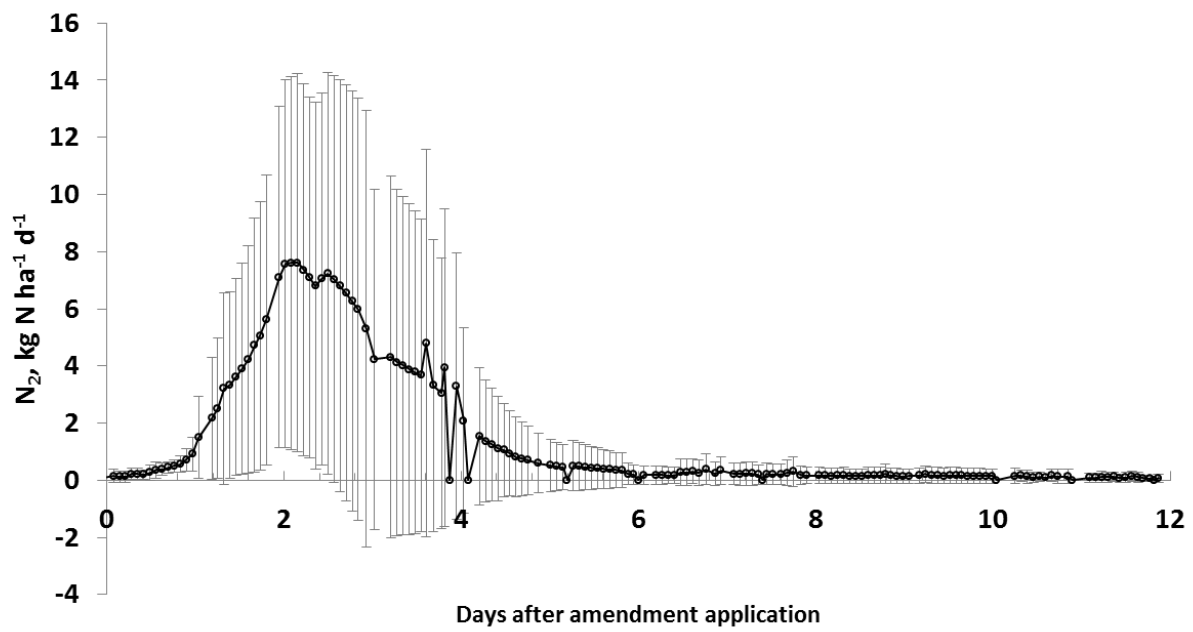
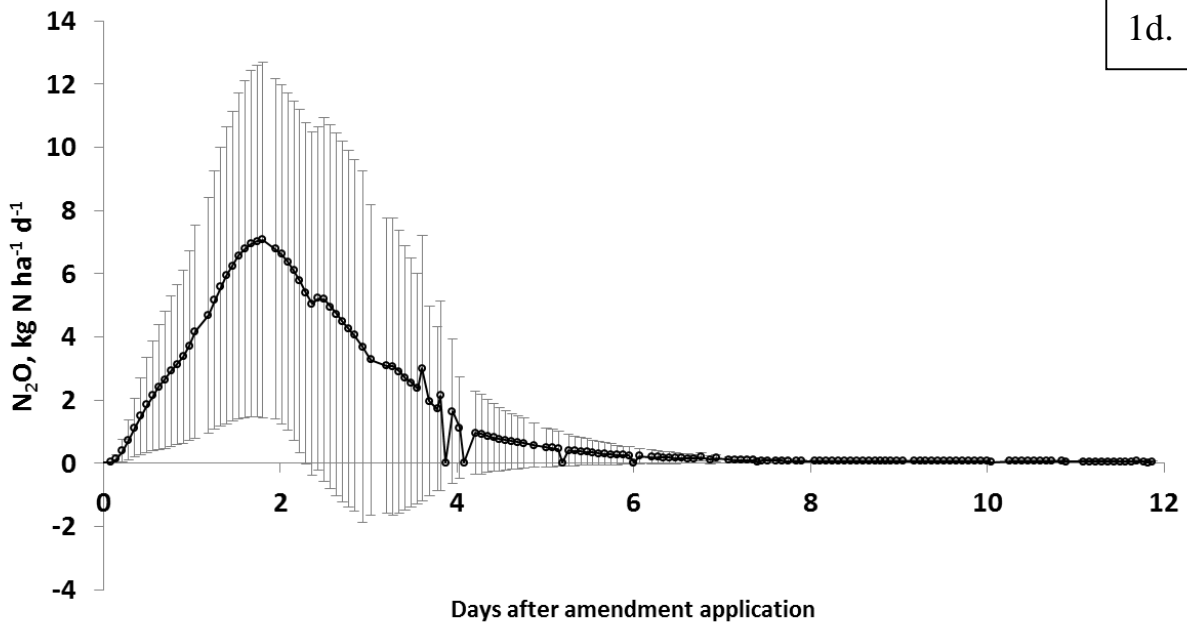
1b.

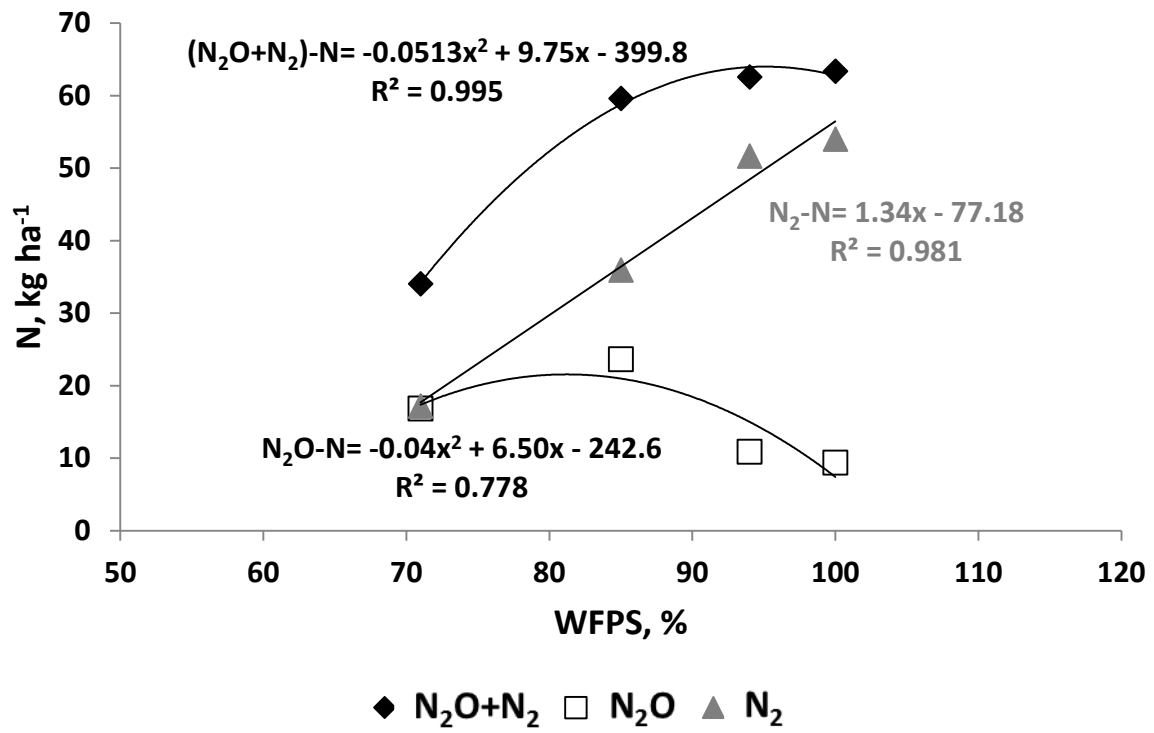


1c.

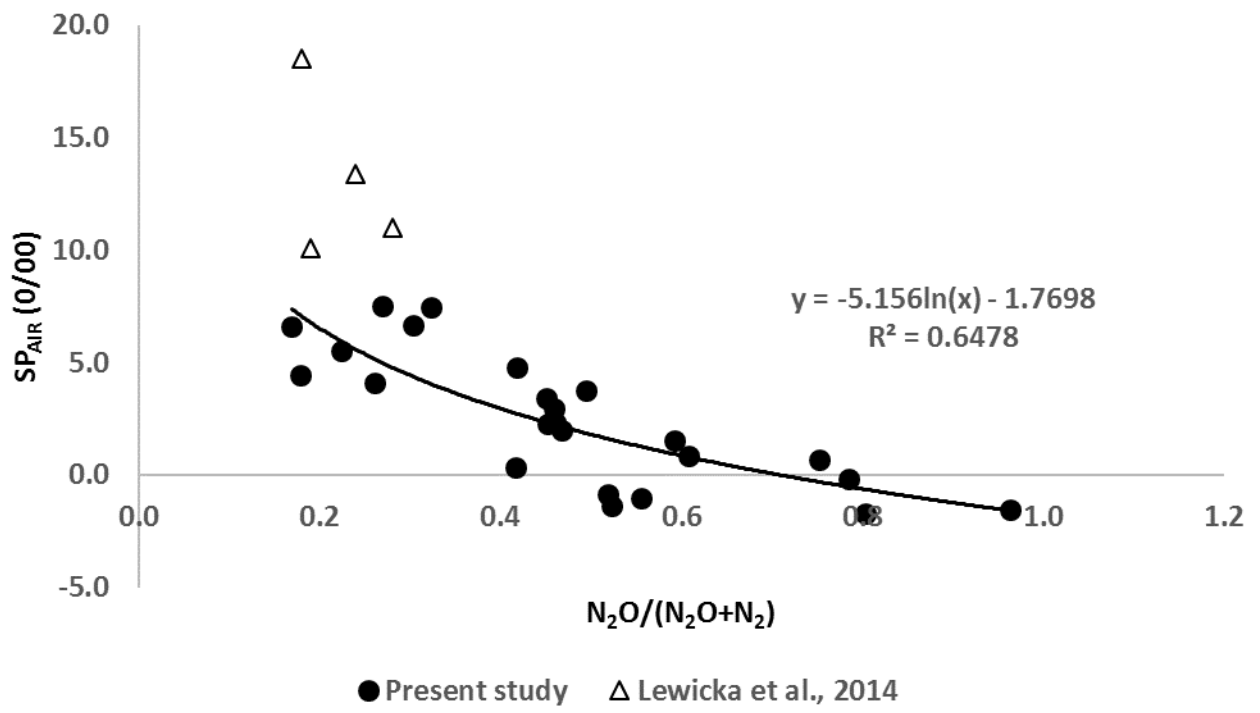


1d.



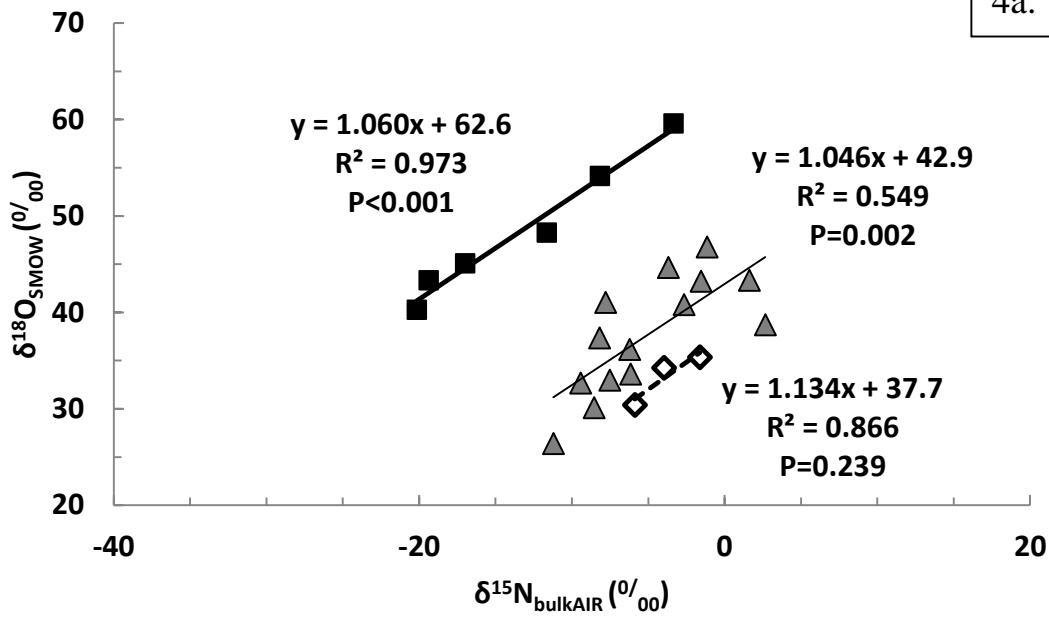


1000
1001 Figure 2
1002
1003



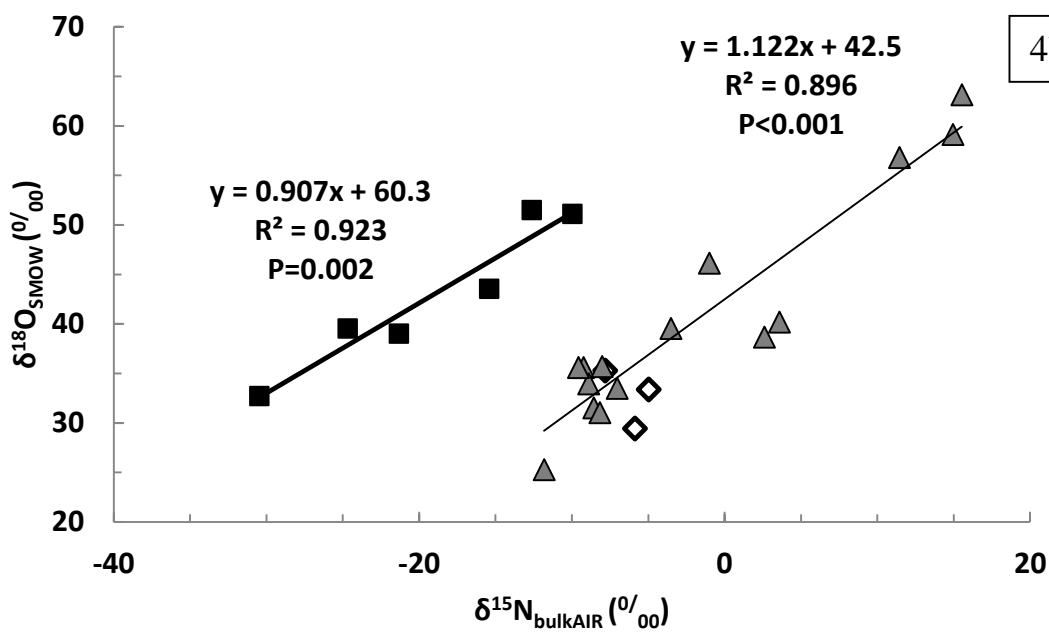
1004
 1005 Figure 3
 1006
 1007

4a.



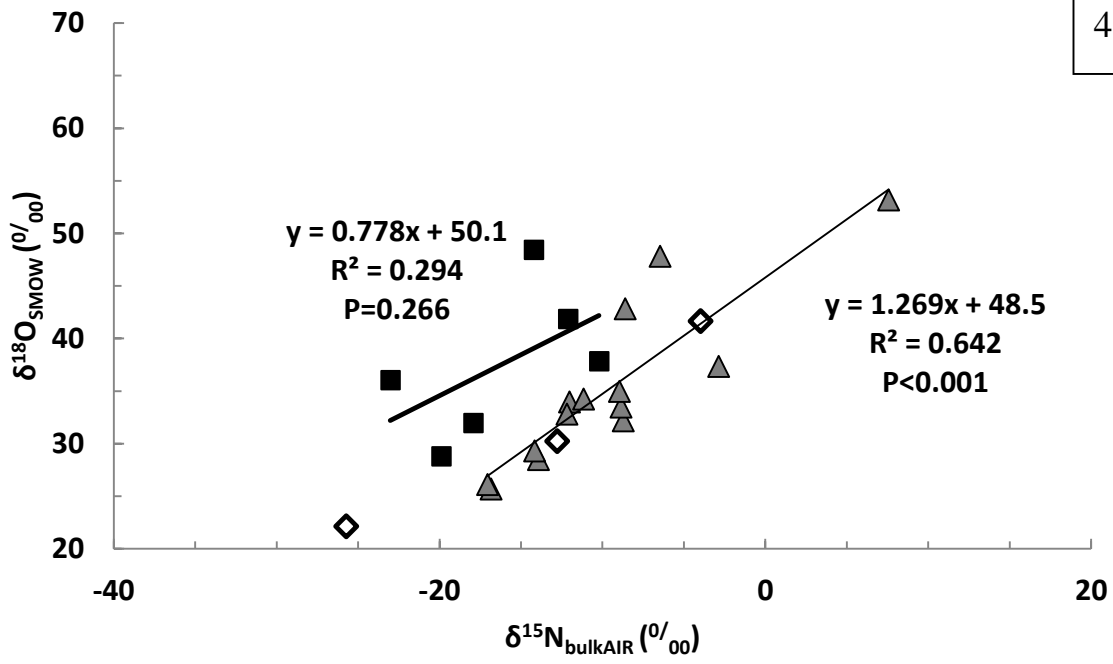
1008
1009
1010
1011

4b.



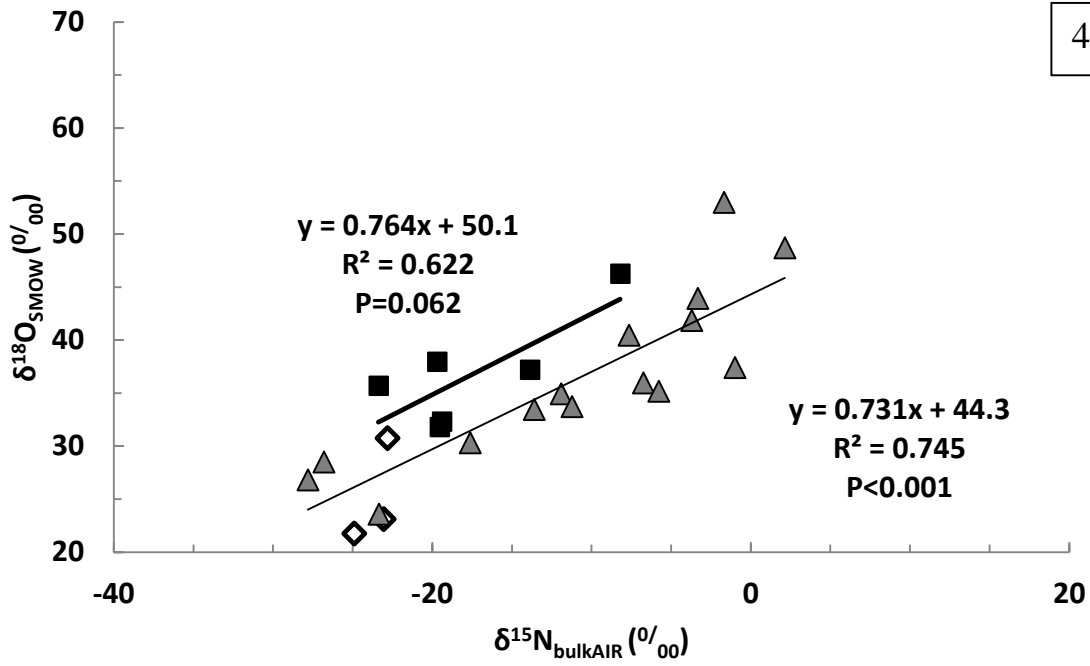
1012
1013

4c

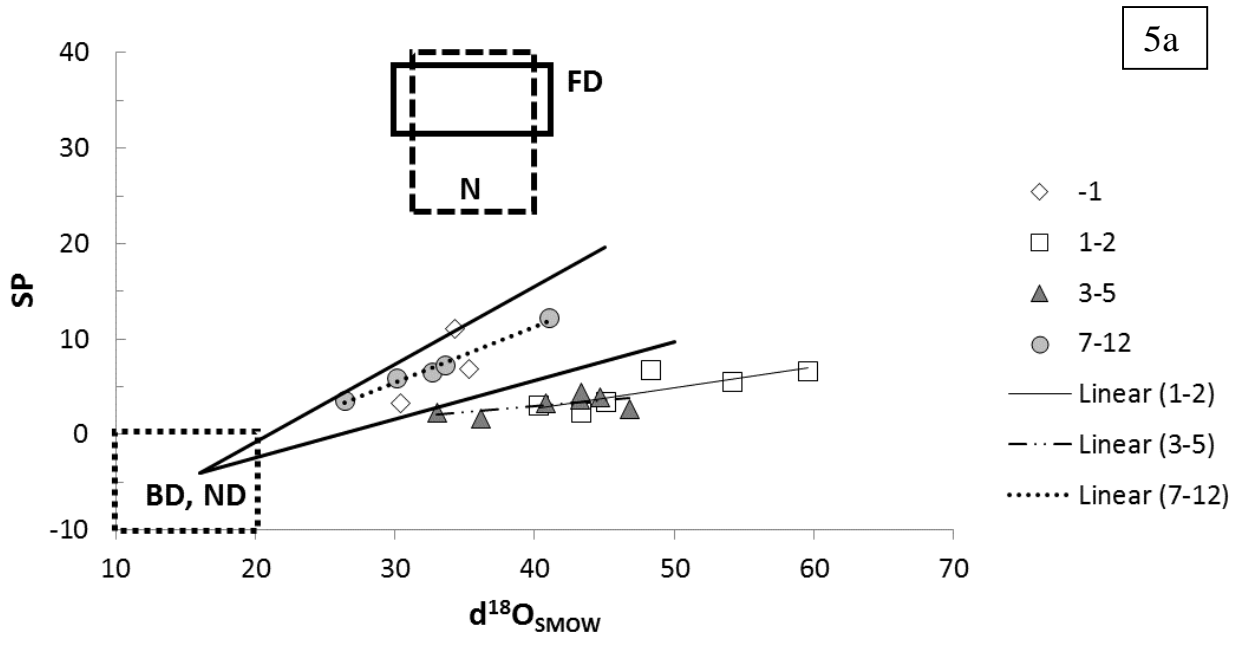


1014
1015
1016

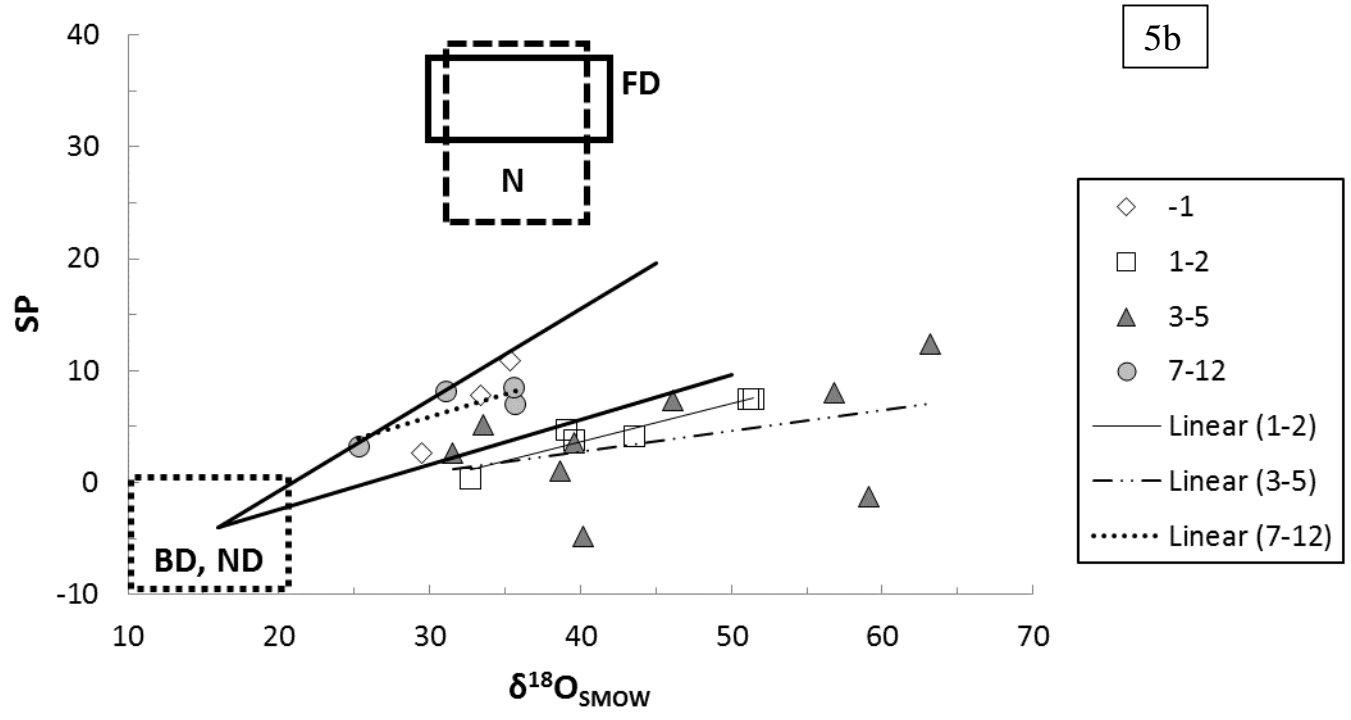
4d



1017
1018
1019

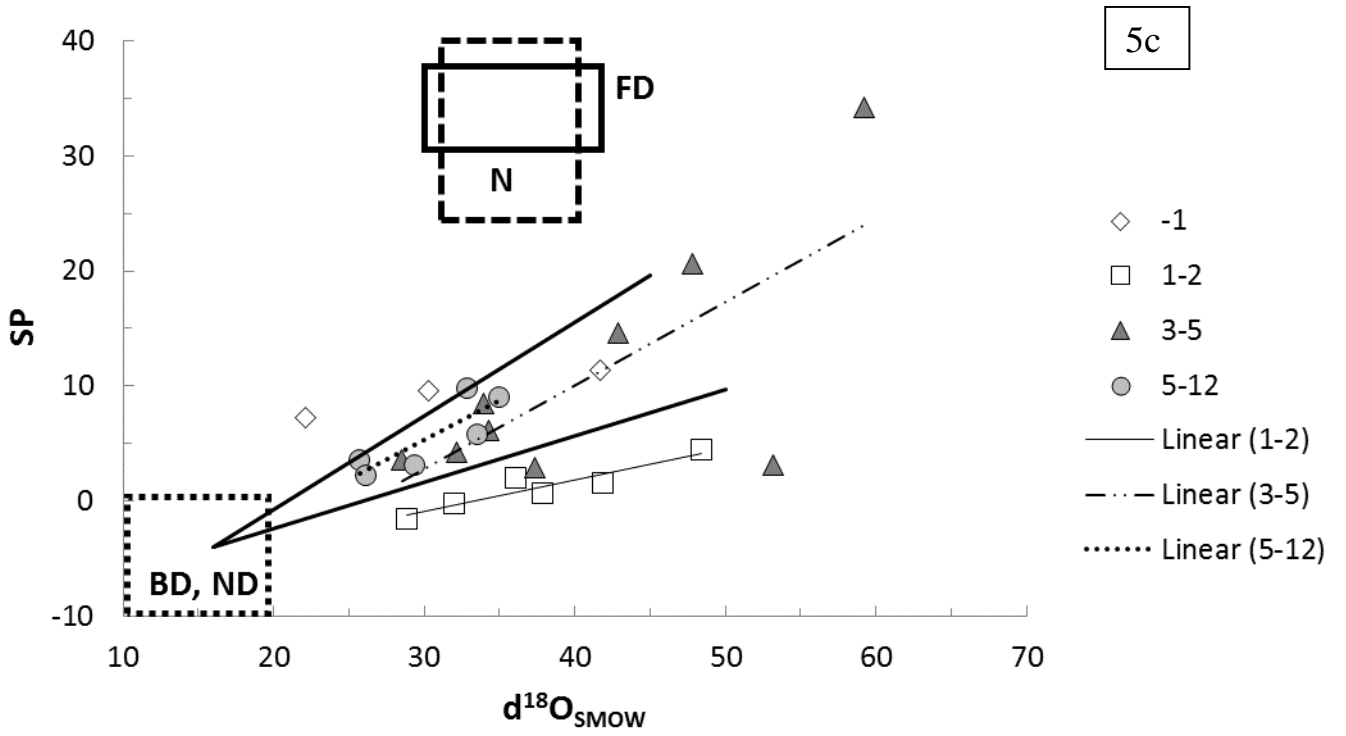


1020
1021
1022

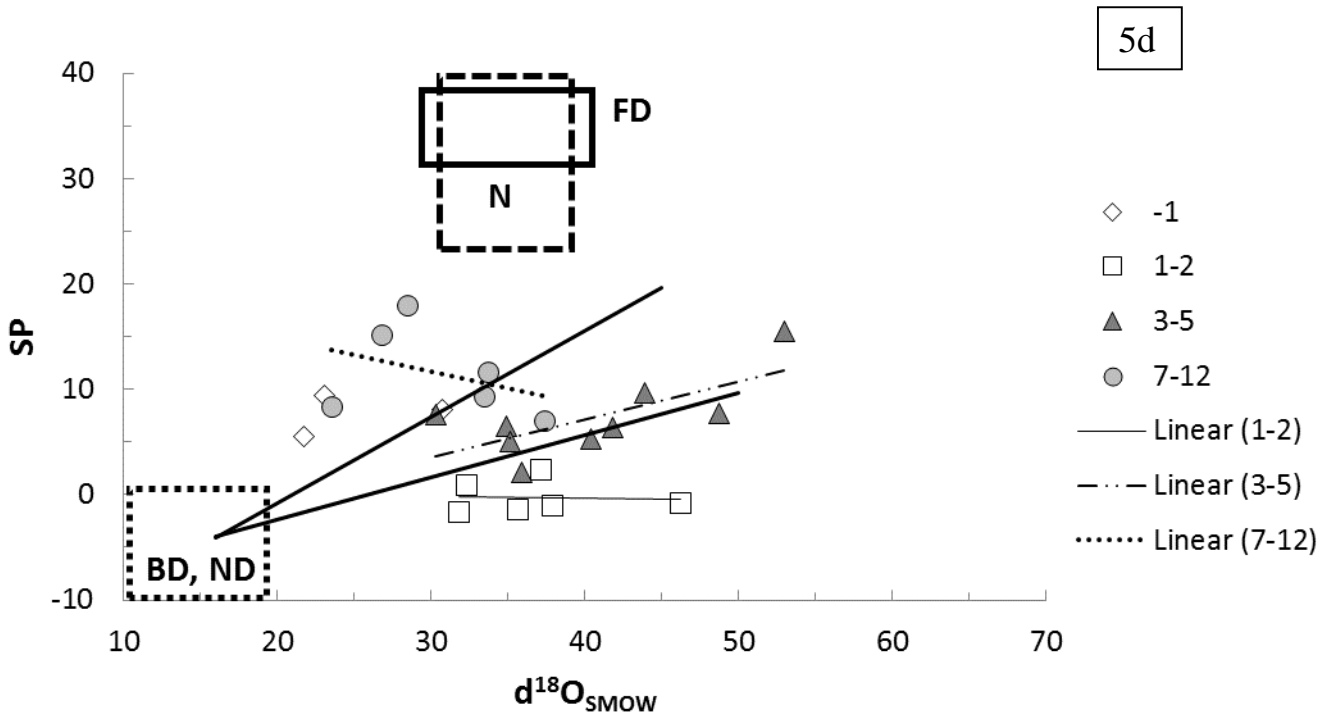


1023
1024
1025
1026
1027
1028
1029
1030
1031

1032



1033
1034
1035



1036
1037

1038 **REFEREE 1**

1039

1040 The paper aims to quantify N₂O and N₂ production process in grassland soils and its dependence
1041 on compaction. N₂O and N₂ emissions and their isotopic signature have been monitored over a
1042 period of 12 days after amendment of KNO₃. The presented laboratory studies simplify the
1043 complex soil pore system into macro and micropores and uses four stages in a rather narrow range
1044 of 70 to 95% “mean” WFPS.

1045 The experimental setup is described in detail. The results agree with the expected values, i.e.
1046 domination of bacterial denitrification processes for the higher water content and an increasing
1047 share of other contribution for when part of the pores is dry. The measurement of the isotopic
1048 signature allows to distinguish different production processes and their dependence on the water
1049 status of the macro and micropores.

1050 I had difficulties to follow the argumentation and get quickly lost in too many in details. I also miss
1051 a discussion of the significance of the presented findings for the characterization of the emissions of
1052 N-species for real grassland systems, although in the introduction (e.g. lines 62 and 63) the study is
1053 set in this context.

1054 The used soil stem from a long-term permanent grassland. But the preparation of the samples (a
1055 necessary step for the laboratory study) destroys the specific characterization of a grassland soil.
1056 Roots and the organization of the aggregates are removed and there is no plant growth that greatly
1057 influence the distribution and availability of N substrate as well as the oxygen supply. It should also
1058 be mentioned that a large share of N-input in agricultural system occurs in reduced N-form
1059 (excrement's, urea or ammonium nitrate). In grazed system, spatial heterogeneity is related to the
1060 urine patches with a very high N-input on a very limited area. Also, compaction (trampling by
1061 animal, tractor tracks) is spatially very heterogeneous and likely uncoupled to N-substrate input.

1062 **R: the authors agree that soil structure is destroyed, but as the referee says himself,**
1063 **this is a laboratory study, so we are not trying to reproduce the field conditions but**
1064 **to understand soil processes. In fact, we are assessing the potential for this soil to**
1065 **emit N₂O and for this reason we have optimised the conditions for denitrification.**
1066 **The plant is not included for the same reason, as we aim to understand the**
1067 **processes in the soil, although we agree that the plant plays a major role in**
1068 **modifying these processes. The soil used in this study is not sourced from a grazed**
1069 **grassland, but a grassland that is cut, so the effect of the animal, via grazing, soil**
1070 **compaction and excreta deposition is not relevant.**

1071 The results from the present study shows for N₂O as well as (N₂O and N₂) emission a remarkably
1072 low variability among the four treatment, much lower as typically experienced in field
1073 measurements.

1074 Below are given specific comments as a guideline to improve the manuscript

1075 Abstract:

1076 Lines 16 and 17: The soil emitted N₂O is predominantly derived from denitrification and to a
1077 smaller extent, nitrification in soils,

1078 This is a too crude generalization. There are many ways to produce N₂O and the share between
1079 them depends in a complex manner from the main driver, such as oxygen content, substrate
1080 availability, etc.

1081 **R: the authors agree with the referee point and in fact the sentence goes on to say:**
1082 **‘both processes controlled by environmental factors and their interactions, and are**
1083 **influenced by agricultural management’. We have however made it clear that it is a**
1084 **generalisation.**

1085 Lines 20 and 21: Soil water content expressed as water filled pore space (WFPS) is a major
1086 controlling factor of emissions and its interaction with compaction, has not been studied at the
1087 micropore scale.

1088 This is slightly misleading as the experimental setup can only measure net fluxes across the surface
1089 of the entire soil samples and naturally does not allow to determine N₂O production/consumption in
1090 and out of the micropores.

1091 **R: yes, the referee is right in that we are not looking at production and consumption**
1092 **separately; but we only claim the control is on emissions (not production and/or**
1093 **consumption) and we are controlling moisture at the micropore scale.**

1094 Introduction

1095 Lines 210 and 211: concentration) for 24 h, or until the system and the soils atmosphere were
1096 emitting low background levels of both N₂ and N₂O (N₂ can get down to levels of 280 ppm much
1097 smaller than atmospheric values).

1098 Please indicate these „background“ values.

1099 **R: the flushing goes on until there is no further decrease in the background signal.**
1100 **This normally occurs within 24 hours. Values can reach a few gN/ha/d (much lower**
1101 **than atmospheric values of 70%).**

1102 Lines 222 and 223: Flushing was carried out with He for half an hour before the solution was
1103 required for application to the soil cores and continued during the application process to avoid
1104 atmospheric N₂ contamination (a total of one and a half hours).

1105 How this affects the oxygen availability?

1106 **R: the flushing is done to the amendment outside the incubation vessel, so we**
1107 **remove N₂ from the liquid before application. The incubation vessel on the other**
1108 **hand continues to receive He/O₂ so it should not affect O₂ availability, in fact the**
1109 **increase in CO₂ in later experiments supports this assumption.**

1110 Lines 304 and 305: We accepted these as unavoidable features of the experimental set-up, but we
1111 suggest that the main response of the gaseous emissions occurred under the initial conditions, prior
1112 to the loss of water over subsequent days.

1113 “We suggest” is a strange formulation, either the time course of the emissions clearly shows this, or
1114 it is an assumption.

1115 **R: this statement came after a comment from a previous reviewer. We have changed**
1116 **the text now to say ‘we assume’.**

1117 Results

1118 Lines 311 UNSAT/halfsat (50-100 N kg- dry soil)

1119 Unit of NO₃⁻ seems incorrect. Also, the header of Table 2 is wrong (twice UNSAT/SAT)

1120 **R: the referee is correct, units and heading have been amended.**

1121 Lines 349 to 351: The results showed that the total N emission (N₂O+N₂) (Table 3) had a
1122 consistent decreasing trend, with decreasing soil moisture i.e. from 63.4 for SAT/sat (100% WFPS)
1123 to 34.1 kg N ha⁻¹ (71% WFPS) for UNSAT/halfsat.

1124 I don't see a consistent decreasing trend. Only the driest treatment shows a lower emission.

1125 **R: we have modified the text to reflect this properly: ‘The results showed that the**
1126 **total N emission (N₂O+N₂) (Table 3) decreased between the highest and the lowest**
1127 **soil moistures i.e. from 63.4 for SAT/sat (100% WFPS) to 34.1 kg N ha⁻¹ (71% WFPS)**
1128 **for UNSAT/halfsat’**

1129 It also would make more sense to use the same reference for the mineral N content as well as the
1130 cumulative gaseous emissions (e.g. per g soil).

1131 **R: we agree this is a good suggestion. So we have included this extra information in**
1132 **table 3.**

1133 Lines 351 and 352: The maximum cumulative N₂O occurred at around 80% WFPS as Fig. 2 shows.
1134 This is an overinterpretation. There are four values and a fit with three unknown is applied.

1135 **R: we agree that there are no many points, but the value of this analysis is that for a**
1136 **narrow soil moisture range (70-100%) there seems to be a linear response for the N₂**
1137 **but not for the N₂O and the total flux. Those shown were the best fits.**

1138 Noticeable emissions of N₂O and N₂ occur in all four treatment only up to day four. Bacterial
1139 denitrification is identified as the main production pathway. This is due to the experimental setup

1140 with a combined amendment of KNO₃ and glucose, a setup that produce good conditions for
1141 denitrification irrespective of the specific treatment.

1142 **R: as mentioned earlier, we optimised conditions for denitrification, except for soil**
1143 **moisture that is the factor we are studying.**

1144
1145 **REFEREE 2**

1146 General remarks

1147 This paper presents results from a sophisticated laboratory experiment in which an
1148 agricultural soil was compacted and adjusted to 4 different moisture conditions. Glucose
1149 and nitrate was added and the formation, isotopic and isoptomeric composition of
1150 gaseous N was measured over a period of 12 days. Using those data the authors try
1151 to determine the contribution of different processes to N gas formation. The paper is a
1152 good example how much information you can get from experimental data if you spend
1153 a lot of energy in calculations and data analysis. However, in my eyes the paper has
1154 three critical weaknesses:

1155
1156 1.) The results are not really new. It is known for a long time that addition of nitrate and
1157 glucose stimulates denitrification in soils and that denitrification is favored under
1158 wetterconditions. All the points in the conclusions are not new. If there is new knowledge
1159 obtained from the study, it has to be elaborated more clearly.

1160 **R: we agree that some of the general points are known, for example the effect of soil**
1161 **moisture on emissions, but this is normally considered in relation to ranges of**
1162 **<60%, 60-75% and >75%. We have looked at a more detailed moisture adjustment,**
1163 **four levels at a relatively high moisture range, between 70 to 100% WFPS. We have**
1164 **also studied the isotopocules of N₂O and found isotopic similarities at similar**
1165 **moisture levels. Moreover, for the first time we have conducted N₂ +N₂O flux**
1166 **measurements at defined saturation of pores size fractions as a prerequisite to**
1167 **model denitrification as a function of water status.**

1168
1169 2.) The paper is lacking a clear story. It is not really clear to me what was the final purpose
1170 of all those detailed analysis. There are some hypothesis mentioned at the end of
1171 the introduction but the rest of the manuscript is not tailored to address those hypotheses.
1172 The hypothesis that wetter conditions reduce heterogeneity could be answered
1173 from just looking at the error bars in figure 1 – you do not need sophisticated analysis
1174 to prove this point. Aiming to understand what is going on in one0s own experiment (as
1175 stated in the last sentence of the introduction) is not a sufficient aim of a paper.

1176 **R: We have done a detailed control of soil moisture in the soil and in order to do this**
1177 **we had to do the detailed analysis the reviewer refers to in terms of the moisture**
1178 **adjustment. In this way we ensured that the four moisture levels above 70% WFPS**
1179 **were as accurate as possible. We also used tools such as the isotopomers to**
1180 **confirm source processes, and this is the result of our research in the last 15 years,**
1181 **when we have built up a large database of isotopomers of N₂O to improve the**
1182 **uncertainty in the determination of the sources. In this particular experiment we**
1183 **have been able to elucidate the effect of saturation on processes at relatively high**
1184 **moisture levels when combined with the measurements of N₂O and N₂ emissions.**

1185
1186 3.) There are some problems with the experimental approach which limit interpretation
1187 of the data. First, moisture conditions were not constant but changed a lot during
1188 the experiment. The second treatment, for example at the end of the experiment had
1189 the same water content as the third treatment in the beginning. They had changing
1190 substrate concentrations in parallel to changing moisture conditions. Thus, the
1191 interpretation

1192 of moisture effects during the course of the experiment is difficult. A way to
1193 minimize that effect would have been to moisten the supplied He/O₂ gas. I would also
1194 expect that water loss was highest in the beginning, when the surface layer was drying.
1195 A way to get some information about temporal changes of water content would have
1196 been to weigh the incubation vessels during the incubation. Second, they measured
1197 gas emission – not gas production. They mention this problem in the paper but somehow
1198 ignore its consequences. The emitted gas probably originates from those sites
1199 which are physically linked to the atmosphere, while gas production, e.g. in the center
1200 of aggregates did probably contribute less to the emitted gas. So, the conclusions
1201 drawn from the analysis could be valid only for a part of the soil volume.

1202 **R: we are aware there are limitations to the experimental approach. In order to moist**
1203 **the gas we would have to have an extra vessel where we flush the gas through.**
1204 **Measuring N₂ is very difficult due to background atmospheric levels and any**
1205 **additions to the experimental system poses a risk of leaks. In addition, adding moist**
1206 **gas will likely block the tubing as these are very narrow (1/8"o.d.). The flow of the**
1207 **gas is very slow (10 ml/min) simulating a low wind speed so normally this would dry**
1208 **the soil in field conditions too. It would represent a rainfall event where the initial**
1209 **moisture differs between treatments but some drying occurs due to the wind flow.**
1210 **We believe the effect of drying will be more relevant (and significant relative to the**
1211 **initial moisture) later in the incubation. We also know that if drying is significantly**
1212 **affecting the microbes, we would see an increase in CO₂ emissions which did not**
1213 **happen later in the incubation. We have introduced changes in the text to make the**
1214 **reader aware of this and have reflected this as ‘the effect of initial soil moisture’.**

1215
1216 Detailed comments

1217
1218 I.17: remove “soils”
1219 **R: removed**

1220
1221 I.40: What do you mean with “benign” for the environment. Do you mean the process is
1222 important because it closes the global N cycle because it reverses N-fixation?
1223 **R: no, it is benign because it does not cause harm to the environment.**

1224
1225 I.64-73: I would move this paragraph to an earlier point, before talking about
1226 compactation.

1227 **R: we have placed this paragraph after the compaction, as it follows from the**
1228 **previous paragraph where we discuss the effect of livestock on compaction. It also**
1229 **leads to the following text on effect of compaction on soil water: ‘reducing the soil**
1230 **air volume and therefore increasing the WFPS’.**

1231
1232 I.72: I would replace “powerful tool” by “basis”.
1233 **R: changed**

1234
1235 I.81: If there are several references for one statement, present them in chronological order.
1236 **R: changed**

1237
1238 I.81-82: Remove sentence
1239 **R: removed**

1240
1241 I.83: “: : under the conditions: : :”
1242 **R: changed**

1243

1244 I.92: Be more specific. What do you mean by “other steps of denitrification”?
1245 **R: we agree that this sentence was not clear enough so we rewrote to:** “Simultaneous
1246 occurrence production and reduction of N₂O as in natural conditions presents a challenge for
1247 isotopic factors determination due to uncertainty on N₂ reduction and the co-existence of different
1248 microbial communities producing N₂O (Lewicka-Szczebak *et al.*, 2014).
1249 I.93: “reported here”.
1250 **R: changed**
1251
1252 I.100: Does that mean that those results are only relevant at elevated C and N?
1253 **R: We have modified the text as follows:** ‘The results generally confirmed the range of
1254 values of η (net isotope effects) and $\eta^{18}\text{O}/\eta^{15}\text{N}$ ratios reported by previous studies for N₂O reduction
1255 for that part of the soil volume where denitrification was enhanced by the N+C amendment. This did
1256 not apply for the other part of the soil volume not reached by the N+C amendment, showing that the
1257 validity of published net isotope effects for soil conditions with low denitrification activity still
1258 needs to be evaluated’.
1259
1260 I.108: Why CO₂?
1261 **R: we have changed the text:** ‘soil to assess the impact of different levels of soil saturation on
1262 N₂O and N₂ emissions after compaction. CO₂ emissions were measured in addition as an estimate of
1263 respiration and thus of O₂ consumption’.
1264 I.112: “controlled laboratory conditions”
1265 **R: changed but this text is now in section 2.4 as recommended by another referee.**
1266
1267 I.119: What do you mean by “heterogeneity in N emissions”?
1268 **R: spatial distribution of emissions, text changed to clarify**
1269
1270 I.120: I am not a soil scientist, but is that really new?
1271 **R: prediction of N₂O emissions is very difficult in part due to their spatial variability.**
1272 **We are trying to understand how this effect occurs in a relatively narrow range of**
1273 **moisture (70-100%). As far as we know there no other studies going to this level of**
1274 **detail. This has been included in the text (end of introduction section).**
1275
1276 I.121: Aiming to understand what is going on in one’s own experiment is not a sufficient
1277 aim of a paper.
1278 **R: we have changed the text:** ‘We aimed to understand changes in the ratio N₂O/(N₂O+N₂) at
1279 the different moisture levels studied in a controlled manner on soil micro and macropores.
1280 Moreover, we used isotopic values of N₂O to evaluate if the contribution of bacterial
1281 denitrification to the total N₂O flux was affected by moisture status’
1282
1283 I.137: Verb missing. “was applied”?
1284 **R: the verb is early on in the paragraph. The paragraph is now split to make it clear.**
1285
1286 I.228: “CO₂ was measured: : :”
1287 **R: changed**
1288
1289 I.230: replace “pulled together in one sample” by “pooled”
1290 **R: changed**
1291
1292 I.232: Remove sentence. There is a similar sentence in the results section.
1293 **R: removed**
1294
1295 I.268: Were the data normal distributed?

1296 **R: yes, all datasets were tested by fitting a Gaussian model resulting in**
1297 **F_{prob}<0.001. this was added in the results section.**
1298
1299 I.275: “mixing model was then used” (use past tense)
1300 **R: changed**
1301
1302 I.283: When did this occur and what is a possible explanation?
1303 Wrong fractionation factors?
1304 **We clarified the variability of endmember values and fractionation factors in the**
1305 **introduction: “The analysis comprised measurements of the N₂O and N₂ fluxes**
1306 **combined with isotopocule data. Net isotope effects (η values) are variable to a**
1307 **certain extent as they result from a combination of several processes causing**
1308 **isotopic fractionation (Well et al., 2012). The results generally confirmed the range**
1309 **of of η values and η¹⁸O/η¹⁵N ratios reported by previous studies for N₂O reduction**
1310 **for the soil volume reached by the N+C amendment. This did not apply for the soil**
1311 **volume not reached by the N+C amendment.”**
1312
1313 I.290: A TCD is an detector – not an analyzer.
1314 **R: changed analysed for determined**
1315
1316 I.303 Why was the gas stream not bubbled through water to saturate it with water?
1317 **R: see our explanation above in point 3.**
1318
1319 I.305: I would expect the highest water loss right in the beginning.
1320 **R: the flowrate is very low so drying will take a while, we are assuming that the**
1321 **significant water loss will affect later in the incubation, later than the peaks appear.**
1322 **However, as explained earlier, we have now referred to the effect of the initial soil**
1323 **moisture in the treatments.**
1324
1325 I.306. But they were similar between treatments in the end although different starting
1326 conditions.
1327 **R: yes**
1328
1329 I.314-316: There was a high variability in the data.
1330 **R: but only for NH₄⁺ it was not significant. A sentence was added**
1331
1332 I.318: Remove “The results showed that”
1333 **R: removed**
1334
1335 I.329: I do not see that in Figure 1. In Unsat/sat the N₂O maximum was at 12 kg N/ha d,
1336 not
1337 around 7.
1338 **R: the referee is correct, we have now amended the text to reflect this: ‘The N₂O**
1339 **maximum in the SAT/sat and HALFSAT/sat treatments was of similar magnitude**
1340 **(means of 5.5 and 6.5 kg N ha⁻¹ d⁻¹, respectively) and but not those of UNSAT/sat and**
1341 **UNSAT/halfsat (means of 7.1 and 11.9 kg N ha⁻¹ d⁻¹, respectively).**
1342
1343 I.348. Right. But what are the consequences of this for your experiment and
1344 its interpretation?
1345 **R: this belongs to the discussion (4.1) so have been moved in there to explain the**
1346 **potential underestimation of the production due to low diffusion.**
1347

1348 I.354: You probably mean “CO₂ fluxes”. Why was CO₂ measured?
1349 **R: yes, added fluxes in the sentence. CO₂ indicates aerobic respiration and as**
1350 **explained above (l.108) is also affected by the soil moisture and level of compaction.**
1351
1352 I.360: The carbon budget is interesting but complicated. Could you calculate recovery
1353 rates for the added glucose? It looks as if there are recoveries higher than 100%. Can
1354 this be interpreted as a priming effect? A problem with using CO₂ for carbon budgeting
1355 is, that depending on pH you also have other IC species in the soil solution. Do you
1356 know the pH in your soils?
1357 **R: pH is 5.63 as shown in Table 1. We did not do a C budget, but it is possible that**
1358 **soil C would have also contributed to the CO₂ emitted but to a lower extent**
1359 **compared to the added glucose.**
1360
1361 I.370: Add article before “period”
1362 **R: added**
1363
1364 I.375: The SP data have a high standard deviation. Are the differencers discussed in this
1365 paragraph real?
1366 **R: we think the larger variation (high SD) of SP around day 3 corresponds to the**
1367 **with highest variation of N₂ and N₂O fluxes (which is evident from Figs**
1368
1369 I.391: You may consider adding these data to the plot.
1370 **R: data added to figure**
1371
1372 I.394: Separate into two sentences.
1373 Start second one with “In our data, maximum : : :.”
1374 **R: changed**
1375
1376 I.404 So what is the message of this paragraph with respect to the first sentence of the
1377 paragraph?
1378 **R: we have rewritten: ‘the question arises to which extent the relationships between**
1379 **the d18O and d15Nbulk and between d18O and SP within the individual treatments**
1380 **denitrification dynamics. We checked this to evaluate the robustness of isotope**
1381 **effects during N₂O reduction as a prerequisite to calculate the percentage of**
1382 **bacterial denitrification in N₂O production.’”**
1383
1384 I.405: Why was this done?
1385 **R: we have found that the isotopologues seem to be potentially more powerful than**
1386 **initially thought. By looking at these relationships we have learnt how the**
1387 **responses relate to the sources of these gases.**
1388
1389 I.428: Why was this plot done?
1390 **R: the same reason as above**
1391
1392 I.441: I do not see data within those areas in the plots.
1393 **R: we have not been so clear, and we refer to the vectors more than the areas. Text**
1394 **has been changed to reflect this.**
1395
1396 I.456: “sat” page 19: It is difficult to detect the storyline on this page.
1397 **R: we are explaining that from our results we are providing a refinement in the soil**
1398 **moisture (WFPS) thresholds previously established as borderline for nitrification-**
1399 **denitrification. We are also proposing that WFPS which was previously established**

1400 **as a normalised parameter for these type of soil moisture thresholds, might actually**
1401 **change with soil type.**
1402

1403 L513: Could it be that there was C limitation in the dryer treatments because glucose was
1404 metabolized aerobically?
1405 **R: if glucose was metabolised we would have expected C to have been less limiting**
1406

1407 I.534-537: The message of the CO₂ paragraph is not really
1408 clear. Are the CO₂ data helpful in this manuscript?
1409 **R: we have deleted the paragraph as suggested.**
1410

1411 I.539: How much is the unaccounted N-loss in comparison to the accounted gaseous
1412 losses?
1413 **R: we added: " unaccounted-for N loss is two to three times the total measured gas**
1414 **loss (Table 3)".**
1415

1416 I.541: NO: What are typical NO fluxes in the literature? Can the NO flux have a significant
1417 magnitude? The same applies to microbial biomass: Is the microbial biomass potentially
1418 formed from the unaccounted N-loss in a realistic order of magnitude?
1419 **R: we are now able to measure NO fluxes in the system. Loick et al reports a ratio**
1420 **N₂O/NO of 0.4 for example, so yes, it can be significant. We did not do microbial**
1421 **biomass in this instance.**
1422

1423 I.567: How should nitrification contribute to BDEN? Do you mean nitrifier-denitrification?
1424 **R: thus large contributions to the total N₂O flux from nitrification were not probable**
1425

1426 I.636: I do not understand the content and purpose of this paragraph.
1427 **R: text changed to: The question arises, if the poor coincidence of Pool 2**
1428 **isotopologue fluxes with previous N₂O reduction studies reflects the variability of**
1429 **isotope effects of N₂O reduction or if the contribution of other processes like fungal**
1430 **denitrification could explain this (Lewicka-Szczabek et al, 2017). The latter**
1431 **explanation is evaluated in section 4.3.**
1432

1433 I.719: Don't you have 4 periods in the figure? Table 3: Unit missing for Total emitted N.
1434 Tables 5 and 6: I wonder whether these data could be presented better in figures.
1435 **R: no, only three. Units included. Yes, figures can illustrate better, but as we**
1436 **explained in the initial review, this data is very useful for models and we think**
1437 **providing the values will be more useful.**
1438

1439 Figure 5: the four sub-graphs are quite similar. Isn't a conclusion that the results were not
1440 much influenced by soil moisture?
1441 Do you really need 4 graphs?
1442 **R: we concluded that there were similarities between the 2 high moisture and 2 low**
1443 **moisture treatments. We believe this is an important finding due to the relatively**
1444 **narrow range of soil moisture we have studied, above 70%, in which we still find**
1445 **differences in fluxes. Davidson stated that the threshold for nitrification-**
1446 **denitrification lies at about 60%, in our case we have managed to refine this.**
1447

1448 **REFEREE 3**
1449

1450 This is an interesting study that addresses the roles of soil compaction and water
1451 saturation

1452 levels on N₂O production and the microbial origins of N₂O. The results are not
1453 terribly profound but this is an important contribution to the literature as the precise
1454 causes of N₂O hot spot production are still unresolved. Overall I found the writing
1455 to suffer from incorrect grammar and English writing style. Further, the manuscript is
1456 much longer than it needs to be. The manuscript would greatly benefit from a major
1457 rewrite and could be re-written as a short concise note rather than a full research paper.
1458 I've identified some issues with the writing below but there are numerous problems
1459 beyond what I have listed.

1460 **R: the majority of the authors consist of native English speakers and the English**
1461 **has been revised by them, so we believe the quality of the English is good. We think**
1462 **that providing the current level of detail in this manuscript as a full research paper**
1463 **is required to give further evidence for the need to use isotopic signatures and**
1464 **modelling approaches of N₂O in order to describe the driving source processes of**
1465 **this gas as emitted from soils.**

1466 Line 26 to 29: As this sentence contains both a colon and a semi-colon it needs to
1467 be broken into at least two sentences. I do not understand the meaning of the portion
1468 after the colon (28-29).

1469 **R: thanks for the suggestion, paragraph has been split.**

1470
1471 Line 73 and 74: Please check with Coplen (2011) regarding the correct usage of
1472 "isotopologues" and "isotopomers".

1473
1474 **R: we have now modified the text according to Coplen's definitions below and used**
1475 **isotopocule always if SP AND d¹⁸O are addressed, isotopomer if ONLY SP is**
1476 **addressed.**

1477 **According to Coplen: 'The molecular species can be an isotopologue, an**
1478 **isotopomer, or neither. For example, the three molecular species ¹⁵N₂ ¹⁶O, ¹⁴N¹⁵N¹⁶O,**
1479 **and ¹⁵N¹⁴N¹⁶O are isotopocules, but they are neither isotopologues (because the**
1480 **latter two do not differ in isotopic composition) nor isotopomers (only the latter two**
1481 **are isotopomers). Isotopolog: Molecular species that differ only in isotopic**
1482 **composition (number of isotopic substitutions) and relative molecular**
1483 **Mass. Isotopomers: Molecular species having the same number of each isotopic**
1484 **atom (thus, the same relative molecular mass) but differing in their positions.'**
1485 **We defined these in the introduction as: 'Isotopologues of N₂O represent the**
1486 **isotopic substitution of the O and/or the two N atoms within the N₂O molecule. The**
1487 **isotopomers of N₂O, are those differing in the peripheral (β) and central N-positions**
1488 **(α) of the linear molecule' which we believe agree with the definition given by**
1489 **Coplen.**

1490
1491 Line 97-98: Why is "soil volume" the key control on the net isotope effect? This seems
1492 more like an experimental condition rather than a governing soil process.

1493 **R: we changed the text for: "The results generally confirmed the range of values of η (net**
1494 **isotope effects) and η¹⁸O/η¹⁵N ratios reported by previous studies for N₂O reduction for that**
1495 **part of the soil volume where denitrification was enhanced by the N+C amendment. This did**
1496 **not apply for the other part of the soil volume not reached by the N+C amendment."**

1497
1498 Line 111-112: Generally avoid one-sentence paragraphs. This statement belongs
1499 more appropriately in the Methods section and could be deleted here.

1500 **R: text has been moved as suggested**

1501
1502 Line 159: This paragraph is much longer and more detailed than it needs to be.

1503 **R: section has been moved to a supplementary material.**

1504
1505 Line 323-324: Use past tense here.
1506 **R: all throughout this section (3.2) there is only past tense. I am not sure where the**
1507 **reviewer refers to.**
1508
1509 Line 338: Delete “already”.
1510 **R: deleted as suggested.**
1511
1512 Line 351: Incorrect word use. SP values don’t “show”; rather they are obtained. Use
1513 past tense to describe trends in the experimental data throughout this paragraph.
1514 **R: text has been amended.**
1515
1516 Line 363: Don’t describe “the plot”; rather simply refer to the trends between the
1517 parameters.
1518 **R: text amended.**
1519
1520 Line 365: Regressions don’t suggest but simply describe a (presumably significant)
1521 relationship between two parameters. You can state that the intercept of the regression
1522 equation relating SP and the $N_2O/(N_2O+N_2)$ was – 2 per mil.
1523 **R: changes have been introduced.**
1524
1525 Line 367-369: The writing is confusing here; I cannot follow the meaning of this sentence.
1526 **R: These are the lines in the submitted pdf: “This is in juxtaposition with the**
1527 **situation when the N emissions are dominated by N_2 or N_2O is low, where the SP**
1528 **values of soil emitted N_2O were much higher (Fig. 3), pointing to an overall product**
1529 **ratio related to an ‘isotopic shift’ of 10 to 12.5o/oo.”**
1530 **We modified to (including previous sentence):**
1531 **“The plot of the $N_2O / (N_2O + N_2)$ ratio vs SP for all treatments in the first two days**
1532 **(when N_2O was increasing and the $N_2O / (N_2O + N_2)$ ratio decreasing) shows a**
1533 **significant negative response of the SP when the ratio increased (Fig. 3). The**
1534 **regression suggests that when the emitted gaseous N is dominated by N_2O (ratio**
1535 **close to 1) the SP values will be slightly negative with values around -2 (Fig. 3), i.e.**
1536 **within the range SP range of bacterial denitrification. With decreasing $N_2O / (N_2O +$**
1537 **$N_2)$ ratio the SP values of soil emitted N_2O were increasing to values up to 8 per**
1538 **mil.”**
1539
1540 Line 370: It is not helpful to refer to data in a figure of another paper. Describe the main
1541 significance to the similarity between these data sets.
1542 **R: I think the reviewer here refers to line 389. We are not referring to a figure**
1543 **necessarily but to the data from Lewicka-Szczebak et al. (2014). The significance**
1544 **was explained in the discussion: ‘These results confirm from 2 independent studies**
1545 **Lewicka-Szczebak et al., 2014) that there is a relationship between the product**
1546 **ratios and isotopic signatures of the N_2O emitted.’**
1547
1548 Line 374: Again, don’t state what is plotted in Figure 4, describe the relationships between
1549 the variables and refer to the figure.
1550 **R: This is in line 406. We have edited the text as suggested.**
1551
1552 Line 383: The r^2 values by themselves are not very relevant. What is relevant is if the
1553 relationships are significant and their associated p values.
1554 **R: R^2 are reported in lines 412 onwards. We have analysed the regressions and**
1555 **introduced the P values as suggested.**

1556

1557 Line 389: See comment for line 374.

1558 **R: I think reviewer refers to line 428. We have stated the new figure was done**
1559 **similarly to the previous one, so we have left the text as it was.**

1560

1561 Tables 1, 4, 5 and 6: These tables could readily be placed in the Supplementary
1562 Documents.

1563 **R: yes, it would be possible, but we would like to have the editor's view before**
1564 **moving them.**

1565

1566 Figure 5: These figures are not well organized. Put a box around the legends so that
1567 we know they are legends. Within the legend, the line should be placed through the
1568 data points rather than defining each line as "Linear". The y-axis title should display
1569 delta not "d".

1570 **R: Legends have now been enclosed by a box. The 'Linear' word in the legend**
1571 **clarifies that a linear function was fitted so we have left this as it was. The reviewer**
1572 **refers to the X axis, delta has been changed.**

1573