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Interactive comment

Interactive comment on "Seagrass as major source of transparent exopolymer particles in the oligotrophic Mediterranean coast" by Francesca luculano et al.

Francesca luculano et al.

fiuculano@imedea.uib-csic.es

Received and published: 11 August 2017

Answer to the general comments:

We thank the reviewer for the positive assessment acknowledging the research reported as original and interesting. We also thank the reviewer for the comments and suggestions for improvements, which will help produce a much improved revised version of the manuscript. We will improve the manuscript following the constructive reviews and adding the references proposed (see specific actions below).

We agree that the discussion on the impact of TEP production by P. oceanica at the

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scale of the Mediterranean suffers from a number of limitations, including a number of uncertainties around the TEP production at the basin scale, which derive from uncertainties in the components of such estimate. We will address these limitations and will, on the light of these limitations, tone down the claims at the scale of the Mediterranean. We will also revise the text to improve clarity.

This text will be changed in the revised version to:

"These observations suggest that P. oceanica meadows, the dominant ecosystem in Mediterranean coastal waters, are an important source of TEP precursors in the Mediterranean Sea. Considering the average leaf production of P. oceanica of 876 g DW m-2 y-1 (Duarte and Chiscano, 1999), the estimated 37,000 Km2 covered by P. oceanica in the Mediterranean Sea (range 31,040 to 43,550 Km2, Marbà et al. 2014), and the average TEP yield from leaf litter experimentally derived here (2344 µg C g DW-1) we calculated that P. oceanica releases about 76 Gg C as TEP annually to the Mediterranean Sea. However, this estimate should be considered a first-order estimate, as it involves considerable uncertainty, compounding that derived from the substantial variability in primary production of P. oceanica (Duarte and Chiscano, 1999), that in the area covered by P. oceanica meadows in the Mediterranean Sea, and variability in TEP yield across meadows and over time, as the estimate used was derived from a single meadow in the fall. Improving this estimate will require narrowing down these sources of uncertainty as well as the capacity to compare it with estimates of other sources of TEP, such as phytoplankton, which are not yet available at the basin scale. The contribution of P. oceanica meadows to TEP release may contribute to explain, along with other processes, the elevated TEP/Chl a ratios characteristic of the Mediterranean Sea (Ortega et al., 2010). The role of P. oceanica as a relevant source of TEP precursors is enhanced by the contrast between the high production of P. oceanica meadows (Duarte and Chiscano, 1999), resulting in a high production of detritus (e.g. Mateo and Romero, 1997; Cebrian and Duarte, 2001) releasing TEP precursors, and the oligotrophic nature of the Mediterranean Sea, leading to low production

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in the pelagic compartment. In fact, both P. oceanica (e.g. Alcoverro et al., 1997) and phytoplankton (e.g. Krom et al., 1991) are likely to be strongly nutrient-limited in the Mediterranean Sea, which has been shown to enhance the release of TEP precursors through carbon overflow during nutrient limiting conditions (Mari et al., 2001; Radić et al., 2005). Despite the limitations acknowledge above, our estimates highlight the important role of P. oceanica litter as source of TEP in the Mediterranean, and suggest that seagrass meadows may play a similarly important role in other regions supporting extensive seagrass meadows, such as the Caribbean, Australia and South East Asia".

We provide below, our response to the specific comments:

We agree that the estimate of the TEP production of 0.10 Tg C y-1 we propose involve significant uncertainties, which should be acknowledged, and represent, therefore, a first-order estimate. We will acknowledge and discuss these limitations in the revised version of the manuscript (see above). We agree that the area covered involves uncertainties, and now report the range of the most robust assessment to date (range 31,040 to 43,550 Km2, Marbà et al. 2014). We will revise the estimate of TEP production, accordingly, downwards to 76 Gg C y-1 (compared to 0.1 Tg C reported originally), and acknowledge that this is a first-order estimate with substantial uncertainty. We will also acknowledge that the yield of TEP may be variable across meadows and over time, and that these variability need be considered (see above). The production reported above of 876 g DW m-2 y-1 (Duarte and Chiscano, 1999), is indeed leaf production, and is a more thorough estimate, the average of 17 estimates, than those provided in Pergent et al. (1994, 1997). We also acknowledge that this estimate carries significant uncertainty (reported in Duarte and Chiscano, 1999).

Line 123-125: We will acknowledge that the leaf litter of P. oceanica supports a complex community of heterotrophic microbes that may contribute to TEP release. The text will be revised to read: "The experimental evidence reported further confirms the role of TEP formed by precursors released by P. oceanica leaf litter, together with the associated microbial heterotrophic community (Peduzzi et al. 1991), in explaining the

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differences between the two sites, as the TEP concentration reached, using a concentration of leaf litter similar to that observed in Es Caragol, is comparable to the maximum values observed in situ."

Answer to the technical corrections:

Line 35: We agree, in the new version of the manuscript we deleted the reference of Thingstad and Rassoulzadegan (1999).

Line 33-40: We agree and the text has been modified accordingly (see above).

Line 41: We agree, the new version of the manuscript will be corrected from "study" to "studying".

The following references will be included in the revised manuscript:

Alcoverro, T., Romero, J., Duarte, C. M. and López, N. I.: Spatial and temporal variations in nutrient limitation of seagrass Posidonia oceanica growth in the NW Mediterranean, Mar. Ecol. Prog. Ser., 146, 155–161, 1997.

Cebrian, J. and Duarte, C. M.: Detrital stocks and dynamics of the seagrass Posidonia oceanica (L.) Delile in the Spanish Mediterranean, Aquat. Bot., 70, 295–309, doi:10.1016/S0304-3770(01)00154-1, 2001.

Krom, M. D., Kress, N., Brenner, S. and Gordon, L. I.: Phosphorus limitation of primary productivity in the eastern Mediterranean Sea, Limnol. Oceanogr., 36(3), 424–432, 1991.

Marbà, N., Díaz-Almela, E. and Duarte, C. M.: Mediterranean seagrass (Posidonia oceanica) loss between 1842 and 2009, Biol. Conserv., 176, 183–190, doi:10.1016/j.biocon.2014.05.024, 2014. Mateo, M. A. and Romero, J.: Detritus dynamics in the seagrass Posidonia oceanica: elements for an ecosystem carbon and nutrient budget, Mar. Ecol. Prog. Ser., 151, 43–53, 1997.

Peduzzi, P. and Herndl, G. J.: Decomposition and significance of seagrass leaf litter

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(Cymodocea nodosa) for the microbial food web in coastal waters (Gulf of Trieste, Northern Adriatic Sea), Mar. Ecol. Prog. Ser., 71, 163–174, 1991.

Please also note the supplement to this comment: https://www.biogeosciences-discuss.net/bg-2016-558/bg-2016-558-AC2-supplement.pdf

Interactive comment on Biogeosciences Discuss., https://doi.org/10.5194/bg-2016-558, 2017.

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Answer to the general comment:

We thank the reviewer for the assessment that our results clearly show the response of TEP by Posidonia oceanica leaf litter release in the coastal area of the oligotrophic Mediterranean Sea and our field observations are confirmed by the experiment conducted in the laboratory. We also agree with the reviewer that, as also pointed out by reviewer 1, the discussion on the role of TEP release by P. oceanica at the scale of the Mediterranean basin needs be improved by acknowledging uncertainties around the estimates provided.

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"These observations suggest that P. oceanica meadows, the dominant ecosystem in Mediterranean coastal waters, are an important source of TEP precursors in the Mediterranean Sea. Considering the average leaf production of P. oceanica of 876 g DW m-2 y-1 (Duarte and Chiscano, 1999), the estimated 37,000 Km2 covered by P. oceanica in the Mediterranean Sea (range 31,040 to 43,550 Km2, Marbá et al. 2014). and the average TEP yield from leaf litter experimentally derived here (2344 µg C g DW-1) we calculated that P. oceanica releases about 76 Gg C as TEP annually to the Mediterranean Sea. However, this estimate should be considered a first-order estimate, as it involves considerable uncertainty, compounding that derived from the substantial variability in primary production of P. oceanica (Duarte and Chiscano, 1999), that in the area covered by P. oceanica meadows in the Mediterranean Sea, and variability in TEP vield across meadows and over time, as the estimate used was derived from a single meadow in the fall. Improving this estimate will require narrowing down these sources of uncertainty as well as the capacity to compare it with estimates of other sources of TEP, such as phytoplankton, which are not yet available at the basin scale. The contribution of P. oceanica meadows to TEP release may contribute to explain, along with other processes, the elevated TEP/Chl a ratios characteristic of the Mediterranean Sea (Ortega et al., 2010). The role of P. oceanica as a relevant source of TEP precursors is enhanced by the contrast between the high production of P. oceanica meadows (Duarte and Chiscano, 1999), resulting in a high production of detritus (e.g. Mateo and Romero 1997, Cebrian and Duarte 2001) releasing TEP precursors, and the oligotrophic nature of the Mediterranean Sea, leading to low production in the pelagic compartment. In fact, both P. oceanica (e.g. Alcoverro et al., 1997) and phytoplankton (e.g. Krom et al., 1991) are likely to be strongly nutrient-limited in the Mediterranean Sea, which has been shown to enhance the release of TEP precursors through carbon overflow during nutrient limiting conditions (Mari et al., 2001; Radic et al., 2005). Despite the limitations acknowledge above, our estimates highlight the important role of P. oceanica litter as source of TEP in the Mediterranean, and suggest

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that seagrass meadows may play a similarly important role in other regions supporting extensive seagrass meadows, such as the Caribbean, Australia and South East Asia."

Answer to the minor comments:

Line 19: We agree. The revised version of the manuscript will correct "deoxyc" with "deoxy".

Line 41: We agree. The revised version of the manuscript will correct "studying" with "study".

Line 51: We agree. The revised version of the manuscript will clarify "for three years since 2006" with "for three years since January 2012". (This study started in January 2012 in Cap Ses Salines and in August 2012 in Es Caragol beach. However, the time series project in Cap Ses Salines started in 2006. We agree that it is not necessary give this detail as it may confound the reader). We will also add in line 54 "for two years since August 2012", when sampling in Es Caragol beach started.

Line 55: We agree. The revised version of the manuscript will correct "in the shore" with "on the shore".

Line 56: We agree. The revised version of the manuscript will correct "on 2 L Nalgene bottles" with "in 2 L Nalgene bottles".

Line 91: "12 hours" listed twice in the revised version of the manuscript will be corrected with "24 hours" as we also sampled at this time interval.

Line 140: We agree, the revised version will be corrected to read: "Despite the limitations acknowledge above, our estimates highlight the important role of P. oceanica litter as source of TEP in the Mediterranean, and suggest that seagrass meadows may play a similarly important role in other regions supporting extensive seagrass meadows, such as the Caribbean, Australia and South East Asia".

Line 143: We agree. The revised version of the manuscript will correct "assess" with

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"assessed".

Line 145: We agree. The revised version of the manuscript will add "for the" particles dynamics in the ocean at the end of this sentence.

Bar Zeev et al., 2011; Duarte and Cebrian, 1996 will be cited in the revised version of the manuscript. Myklestad, 1977 will be changed with Myklestad, 1995 in the text line 20 and in the reference list.

Parsons et al., 1984 yes it is already cited in the text in line 63.

Please also note the supplement to this comment: https://www.biogeosciences-discuss.net/bg-2016-558/bg-2016-558-AC1-supplement.pdf

Interactive comment on Biogeosciences Discuss., https://doi.org/10.5194/bg-2016-558, 2017.

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Seagrass as major source of transparent exopolymer particles in the

2 oligotrophic Mediterranean coast

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9 **Abstract.** The role of seagrass, *Posidonia oceanica*, meadows as a source of transparent exopolymer particles (TEP) to 10 Mediterranean coastal waters was tested by comparing the TEP dynamics in two adjacent coastal waters in the oligotrophic 11 NW Mediterranean Sea, one characterized by oligotrophic open-sea waters and the other accumulating seagrass leaf litter, 12 together with an experimental examination of TEP release by seagrass litter. TEP concentrations ranged from 4.6 μg XG Eq L⁻¹ to 90.6 μg XG Eq L⁻¹, with mean (± SE) values of 38.7 (± 2.02) μg XG Eq L⁻¹ in the site devoid of seagrass litter, whereas the coastal beach site accumulating leaf litter had > 10-fold mean TEP concentrations of 487.02 (± 72.8) μg XG Eq L⁻¹. Experimental evaluation confirmed high rates of TEP production by *P. oceanica* litter, allowing calculations of the associated TEP yield. We demonstrated that *P. oceanica* is an important source of TEP to the Mediterranean Sea,

contributing an estimated 76, Gg C as TEP annually. TEP release by P. oceanica seagrass explains the elevated TEP

Transparent exopolymer particles (TEP) are acidic and sulphated polysaccharides enriched in deoxy sugars and galactose

18 concentration relative to the low chlorophyll a concentration in the Mediterranean Sea.

1 Introduction

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21 (Myklestad, 1995) which are stainable with a cian blue (Alldredge et al., 1993). These organic particles belong to the POC 22 (particulate, organic carbon) pool (Zhou et al., 1998) and are ubiquitous in marine and limnetic ecosystems (Passow, 2002). 23 Their roles in several biogeochemical processes and their importance in sedimentary carbon fluxes has been extensively 24 documented (Engel and Passow, 2001) as, due to its sticky properties, the aggregation of these particles may enhance the sinking flux and export of organic matter (Kiørboe and Hansen, 1993; Simon et al., 2002) with important consecuences for 25 26 the efficiency of the biological carbon pump (Mari et al., 2017 and references therein). Phytoplanktonic cells, mainly 27 diatoms, are believed to be the major sources of TEP in the marine environment (Passow and Alldredge, 1995a), although 28 benthic organisms, such as suspension feeders (Heinonen et al., 2007) and macroalgal detritus (Thornton, 2004) have been 29 also identified as TEP sources. Indeed, marine macrophytes are important sources of dissolved organic carbon (DOC) to coastal waters (Barron et al., 2006), and may therefore release precursors conducive to TEP formation, such as reported by 30 31 Thornton (2004) for macroalgae. However, seagrass meadows are also important sources of DOC to the marine environment (Barrón et al., 2014), but their role as a source of TEP has not yet been assessed. 32 Posidonia oceanica Delile (L.) is the dominant seagrass species of the Mediterranean Sea (Duarte, 2004). P. oceanica 33 meadows are highly productive (Duarte and Chiscano, 1999) and release high amounts of dissolved organic carbon (Barron 34 35 et al., 2014) as well as leaf litter (Cebrian and Duarte, 2001; Gacia et al., 2002). The large production of DOC and detritus by

Zeev et al., 2011) at levels higher than expected, as indicated by high TEP/Chl a and TEP/bacterial abundance ratios compared to other marine systems (Ortega et al., 2010; Ortega et al., 2017). Whereas TEP are often assumed to be of phytoplankton origin, the relatively high levels of TEP (i.e. high TEP/Chl a ratios) in oligotrophic Mediterranean waters

P. oceanica contrasts with the low planktonic primary production in the oligotrophic Mediterranean littoral zone (Duarte et

al., 1999), where TEP are nevertheless present (Mari, 2001; Beauvais et al., 2003; Prieto et al., 2006; Ortega et al., 2010; Bar

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- 53 suggest that DOC release by Posidonia oceanica meadows could be a source of TEP, explaining the relative high TEP
- 54 concentration reported for Mediterranean waters (Ortega et al., 2010). Although macroalgae have been identified as sources
- of TEP, we are not yet aware of any study examining the role of seagrass as source of TEP.
- 56 In this study, we monitored the dynamics of TEP concentrations in two adjacent, but contrasting, oligotrophic littoral sites of
- 57 Majorca Island (NW Mediterranean Sea), an open coastline flushed with open sea waters and an adjacent, 2 Km, beach
- 58 accumulated Posidonia oceanica leaf litter. We tested the hypothesis that seagrass leaf litter of P. oceanica represents an
- 59 important source of TEP to this ecosystem explaining the contrasting TEP concentrations and dynamics observed in these
- 60 coastal sites using a laboratory experiment.

61 2 Materials and methods

2.1 Sampling sites and time series observations

63 The study was carried out at two sites in the coastal NW Mediterranean Sea of Majorca Balearic Island, a) the Faro Cap Ses

64 Salines experimental field station (Lat 39.264724 N; Lon 3.054446 E), where TEP concentrations were monitored

65 fortnightly for three years since January 2012. This is a pristine and oligotrophic rocky shore ecosystem, with an extensive

seagrass of *P. oceanica* meadow extended around 500 m offshore (Álvarez et al., 2015) and flushed with open-sea water

67 (Fig. 1a), and b) Es Caragol beach (Lat 39.276784, N; Lon 3.043779, E), where TEP dynamics were monitored for two

68 years since August 2012. This is a natural sandy beach in a site of community importance (EU directive-red natura2000)

69 where abundant seagrass detritus accumulates on the shore (Fig. 1b), where it plays an important geomorphological role

where abundant seagrass detritus accumulates of the shore (Fig. 10), where it plays an important geomorphological following

70 (Simeone and De Falco, 2012).

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71 Surface water samples at Faro Cap Ses Salines and Es Caragol were collected fortnightly (monthly during winter months) in

72 2 L Nalgene bottles at noon and 3:00 pm, respectively. A total of 76 sampling events were completed at Faro Cap Ses

73 Salines between 09 January 2012 to 23 March 2015, while 45 sampling events were completed at Es Caragol (from 09

74 August 2012 to 24 September 2014). Surface seawater samples of 250 mL from Faro Cap Ses Salines for chlorophyll a

75 determination were filtered through Whatman GF/F filters and stored at -20 °C. Filters were extracted in 6 mL 90 % acetone

76 for 24 hours followed by fluorometric (Trilogy, Turner design) Chl a determination, calibrated with pure Chl a, after Parsons

et al. (1984). Sea-surface temperature was measured in situ using a data logger (HOBO).

78 TEP concentrations were determined following the colorimetric method of Passow and Alldredge (1995b), where TEP are

detected after staining with alcian blue (Sigma), a cationic copper phthalocyanine dye that complexes carboxyl (-COO-) and

half-ester sulphate (OSO₂) reactive groups of acidic polysaccharides. Following each sampling event, triplicate aliquots

81 (Faro Cap Ses Salines: 300-700 mL; Es Caragol: 50-500 mL, depending on the saturation of filters) were filtered onto 0.4

82 μm pore size, 25 mm diameter polycarbonate filters under low and constant pressure (150 mmHg). Filters were subsequently

83 stained with 1000 μ L of a 0.02 % working solution of a cian blue (pre-filtered through 0.2 μ m) in 0.06 % acetic acid (pH =

2.5), allowed to stain for a few seconds, repeated filtering and rinsed twice with MilliQ water, to eliminate excess dye. Dyed

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filters were stored at -80 °C until extraction at IMEDEA laboratory. To perform the extraction, filters were placed in acid-104 clean 10 mL glass tubes, by adding 5 mL of 80 % sulphuric acid, for 2 to 3 hours, shaking 2 to 3 times to enhance extraction. 105 106 Absorbance was read spectrophotometrically (Shimadzu dual beam spectrophotometer) at 787 nm in 1 cm disposable 107 cuvettes. Triplicate blank filters were also analysed for every batch of samples. Blank absorbance values at 787 nm were 108 then subtracted from the total absorbance values of samples, to account for the capacity of alcian blue to stain filters. Four, 109 calibrations of the acian blue solutions were performed by using Xanthan Gum as standard (XG). The calibration factor (F) was calculated as the mean of the eight estimates obtained. TEP concentrations (TEP) were expressed in μg Xanthan Gum 110

(XG) equivalents per litre (µg XG Eq L⁻¹) and calculated following Eq. (1): 112 $TEP = (a_{sample} - a_{blank}) V^{-1} \cdot F,$

113 where a_{sample} and a_{blank} are absorbance values at 787 nm for samples and blank filters, respectively; V is the sampled volume

(in L) and F is the calibration factor. The detection limit of the method was $2.2~\mu g$ XG Eq L⁻¹ and the analytical coefficient 115 of variation was 13 %. TEP concentrations were transformed to carbon units (µg C L⁻¹) by using the conversion factor of

0.75 proposed by Engel and Passow (2001) in order to estimate the total TEP yield of P. oceanica leaf litter. 116

117 2.2 Experimental evaluation of TEP release by P. oceanica leaf litter

118 P. oceanica leaf litter and surface seawater were sampled on 8 September 2014, the period of leaf shedding for P. oceanica, 119 from the seashore of Es Caragol and stored at 4 °C for transport to the laboratory. Six 5L Pyrex glass bottles were filled with 120 seawater, pre-filtered by gravity through a 0.2 µm pore membrane size cartridge filter. Three replicated bottles received 16.6 mg fresh weight L⁻¹ of P. oceanica leaf litter, to obtain a final concentration similar to that measured in the near shore waters 121 122 at Es Caragol, and three replicated bottles, without P. oceanica leaf litter, were used as control. The bottles were gently aerated with an air pump to provide mixing and avoid the development of anoxic conditions. The bottles were incubated at 123 124 the in situ temperature at the time of sampling (26.3 °C) in a temperature controlled chamber, and water samples for TEP determinations were collected at increasing time intervals: time 0 (11 September), 6 hours, 12 hours, 24 hours, 48 hours and 125 264 hours (22 September) after the start of the experiment. The water volume and leaf biomass (fresh weight and dry weight 126 127 following desiccation at 60 °C for 24 hours in a drying oven) in the bottles were measured. Replicated 50 mL to 100 mL volumes, pre-filtered trough a 100 µm mesh to remove leaf litter, were sampled using a 60 mL syringe and immediately 128

3 Results 131

Alldredge, 1995b).

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132 Surface seawater temperature ranged from 12.4 °C to 27.8 °C, registered in February 2012 and September 2014, respectively,

filtered onto 0.4 µm to collect, dye and quantify TEP concentration following the procedure described above (Passow and

along the study (average \pm SE = 19.4 \pm 0.54 °C). Chlorophyll a concentration ranged from 0.02 to 0.54 μ g L⁻¹ in July 2014 133

and March 2013, respectively, along the study (average \pm SE = 0.23 \pm 0.01 μ g L⁻¹).

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TEP concentrations ranged from 4.6 to 90.6 µg XG Eq L⁻¹ in Faro Cap Ses Salines and from 26.8 to 1878.4 µg XG Eq L⁻¹ in 142 Es Caragol, with significantly (paired t-test, p < 0.05) higher mean TEP concentrations at Es Caragol (38.7 \pm 2.02 μ g XG Eq 143

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L⁻¹) compared to Faro Cap Ses Salines (487.02 ± 72.8 μg XG Eq L⁻¹). TEP concentrations changed greatly seasonally, with 144 145 maximum TEP values in waters sampled at the Faro Cap Ses Salines observed in February, likely associated with the

phytoplankton bloom occurring at that time, and June (Fig. 2a). In contrast, TEP dynamics showed a more erratic temporal 146

pattern at Es Caragol, with no clear seasonal patters (Fig. 2b). Mean (± SE) TEP/Chl a ratios were also > 10-fold greater at 147

Es Caragol (3109.9 ± 468.9) than at the Faro Cap Ses Salines (286.3 ± 55.7), with a clear seasonal cycle characterized by 148

149 maximum TEP/Chl a ratios in June and July at the Faro Cap Ses Salines whereas at Es Caragol they remained elevated

throughout the year, except between January and March when values were relatively low (Fig. 3a, b). 150

151 During the experimental evaluation initial TEP concentrations (30.4 µg XG Eq L⁻¹) increased slightly after 6 h incubation, to

remain uniform throughout the rest of the experiment in the absence of P. oceanica leaf litter (Fig. 4). In contrast, TEP 152

153 concentrations increased greatly throughout the experiment in the presence of P. oceanica litter, reaching values of 1551 µg

XG Eq L⁻¹, comparable to maximum values observed at Es Caragol, after 264 h (Fig. 4). The corresponding TEP yield of P. 154

oceanica corresponded to 14.128 ± 11.294 μg XG Eq L⁻¹ or 2344 ± 357.26 μg C g DW⁻¹. The yield of TEP in the presence 155

of P. oceanica litter was 9.77 times greater than that in control bottles $(1.384 \pm 1.582 \,\mu g \, \text{XG Eq L}^{-1})$. 156

157 4 Discussion

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158 The results presented provide, to the best of our knowledge, the first evidence that seagrass leaf litter is a source of TEP to coastal waters. Thornton (2004) demonstrated the formation of TEP from the acidic polysaccharides released by macroalgal 159 160 detritus of different species, but the role of seagrass litter as a source of TEP has not been reported to-date. The role of P. oceanica leaf litter as a source of TEP is demonstrated here through the > 10-fold difference in concentration and TEP/Chl a 161 162 ratios between the two adjacent coastal areas studied, one containing rapidly flushed open-sea water and the other representing an accumulation site for P. oceanica leaf litter. The experimental evidence reported further confirms the role of 163 TEP formed by precursors released by P. oceanica leaf litter, together with the associated microbial heterotrophic 164 165 community (Peduzzi et al., 1991), in explaining the differences between the two sites, as the TEP concentration reached, using a concentration of leaf litter similar to that observed in Es Caragol, is comparable to the maximum values observed in 166 167 P. oceanica, as well as seagrasses in general, exports a large fraction of its net primary production as leaf litter, on average 168 about 24 % of NPP (Duarte and Cebrian, 1996). A fraction of this leaf litter is exported to the shoreline following leaf 169

and re-entrained in the water during storms, resulting in the pulses of TEP observed at Es Caragol. 171

shedding by P. oceanica in the late summer and early autumn (Mateo et al., 2003). Leaf litter is then deposited on the beach

The seasonal variability in TEP/Chl a ratios at Faro Cap Ses Salines, where leaf litter accumulation is precluded by strong 173 currents, shows a maximum in the summer (June and July), likely resulting from TEP precursors released by the nearby

seagrass meadow. Ortega et al. (2010) already reported elevated TEP/Chl a ratios during early summer in the Mediterranean Sea, with values comparable to those we observe at the Faro Cap Ses Salines. These observations suggest that P. oceanica meadows, the dominant ecosystem in Mediterranean coastal waters, are an important source of TEP precursors in the Mediterranean Sea (Ortega et al., 2010). Considering the average leaf production of P. oceanica of 876 g DW m⁻² y-1 (Duarte and Chiscano, 1999), the estimated 37,000 Km² covered by P. oceanica in the Mediterranean Sea (range 31,040 to 43,550 Km², Marbà et al., 2014) and the average TEP yield from leaf litter experimentally derived here (2344 µg C g DW⁻¹) we calculated that P. oceanica releases about 76 Gg C as TEP annually to the Mediterranean Sea. However, this estimate should be considered a first-order estimate, as it involves considerable uncertainty, compounding that derived from the substantial variability in primary production of P. oceanica (Duarte and Chiscano, 1999), that in the area covered by P. oceanica meadows in the Mediterranean Sea, and variability in TEP yield across meadows and over time, as the estimate used was derived from a single meadow in the fall. Improving this estimate will require narrowing down these sources of uncertainty as well as the capacity to compare it with estimates of other sources of TEP, such as phytoplankton, which are not yet available at the basin scale. The contribution of P. oceanica meadows to TEP release may contribute to explain, along with other processes, the elevated TEP/Chl a ratios characteristic of the Mediterranean Sea (Ortega et al., 2010). The role of P. oceanica as a relevant source of TEP precursors is enhanced by the contrast between the high production of P. oceanica meadows (Duarte and Chiscano, 1999), resulting in a high production of detritus (e.g. Mateo and Romero, 1997; Cebrián and Duarte, 2001) releasing TEP precursors, and the oligotrophic nature of the Mediterranean Sea, leading to low production in the pelagic compartment. In fact, both P. oceanica (e.g. Alcoverro et al., 1997) and phytoplankton (e.g. Krom et al. 1991) are likely to be strongly nutrient-limited in the Mediterranean Sea, which has been shown to enhance the release of TEP precursors through carbon overflow during nutrient limiting conditions (Mari et al., 2001; Radić et al., 2005). Despite the limitations acknowledge above, estimates highlight the important role of P. oceanica, litter as source of TEP in the Mediterranean, and suggest that seagrass meadows may play a similarly important role in other regions supporting extensive seagrass meadows, such as the Caribbean, Australia and South East Asia. Seagrass meadows have been recently shown to be globally relevant sources of DOC to the marine ecosystem (Barron et al., 2014) and Mari et al. (2017) have recently assessed that the global TEP production could represent 2.5 to 5 Pg C y⁻¹. Here we provide the first evidence that seagrass meadows can also play a relevant, even locally dominant, role as sources of TEP

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new pathway to be accounted for when considering the fate and fluxes of organic matter in the continuum of DOM-POM

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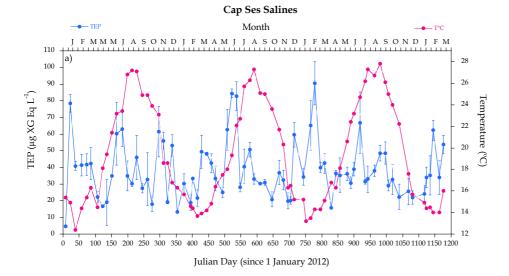
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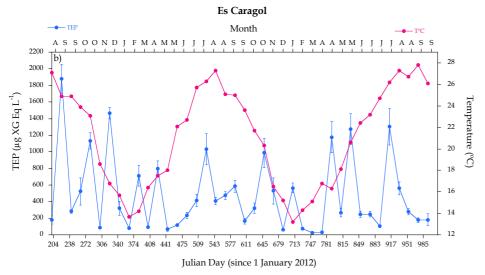
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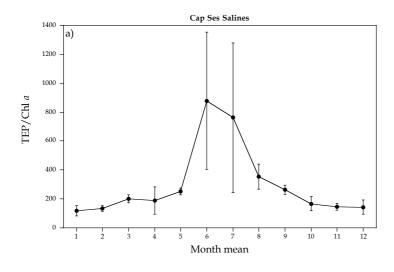
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330 Figure 2: Time series of TEP concentrations (μg XG Eq L⁻¹± SE) and Temperature (°C) at Cap Ses Salines (a) and Es Caragol (b).



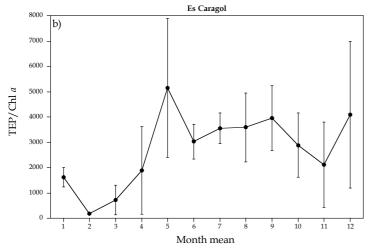


Figure 3: Monthly TEP/Chl a ratios means \pm SE at Cap Ses Salines (a) and Es Caragol (b).

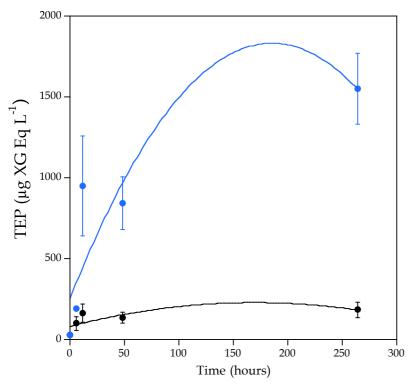


Figure 4: TEP accumulation (mean \pm SE) in the presence (blue line) and absence (dark line) of *P. oceanica* litter. The solid lines show the fitted second order polynomial equations ($R^2 = 0.77$ and 0.53, respectively).

- a)
- b)