

## Response to Associate Editor Comments

An observational constraint on stomatal function in forests: evaluating coupled carbon and water vapor exchange with carbon isotopes in the Community Land Model (CLM 4.5)

Brett Raczka, Henrique F. Duarte, Charles D. Koven, Daniel Ricciuto, Peter E. Thornton, John C. Lin, David R. Bowling

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Our response and associated manuscript changes for the two minor editor comments are described below. We are very pleased with how the manuscript has improved since our first submission. We thank all the reviewers and the editor for their efforts.

Thank you for the opportunity to publish in Biogeosciences.

On behalf of all authors,

Dr. Brett Raczka

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Response to editor comments:

Editor 1: point #2 [from previous comments]: The text explaining eq 11 still need to be modified to state that eq 11 attempts to mimic eq5, but fails to do so. Much of what you are investigating is the unintended consequence of a model implementation, which focusses on N limitation for biogeochemical, but not biogeophysical processes. It needs to be clear to the reader that one could have derived a “correct” implementation by adjusting  $g_s$  in concert with  $A_n$ , and therefore that eq 11 does not satisfy eq 5.

Author 1: We have added to the text that the mismatch between the actual photosynthesis and the intracellular  $CO_2$ : ...” is a result of the carbon water ( $A_n$ - $g_s$ ) coupling (eq. 5) being imposed prior to the effect of nitrogen limitation (eq. 9) and is an artifact of the model implementation. We also test a separate model formulation (described in detail in the next paragraph) specific to this analysis that imposes nitrogen limitation through the  $V_{cmax}$  parameterization and removes the artifact of  $f_{dreg}$ . ” (Page 8. Lines 1-8).

Editor 2: point #25 [from previous comments]: The boundary conditions for the global simulation are not clear. If there is no published reference, then in the main text, include,

[...driven by observed historical climate etc.... , (unpublished, K. Lindsay 20XX)]. Because the global simulation was not introduced in the Methods, I would prefer if you could also state “a global CLM4.5 run with historic climate” the second time you mention this.

Author 2: We have made this change. (page 17, line 13-16, page 17, lines 24-26). We now specify that the climate is driven by CO<sub>2</sub> emissions, and simulated as an emergent property of CAM5 –the atmospheric model within CESM.

Thank You.

# **An observational constraint on stomatal function in forests: evaluating coupled carbon and water vapor exchange with carbon isotopes in the Community Land Model (CLM 4.5)**

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## **Abstract**

Land surface models are useful tools to quantify contemporary and future climate impact on terrestrial carbon cycle processes, provided they can be appropriately constrained and tested with observations. Stable carbon isotopes of CO<sub>2</sub> offer the potential to improve model representation of the coupled carbon and water cycles because they are strongly influenced by stomatal function. Recently, a representation of stable carbon isotope discrimination was incorporated into the Community Land Model component of the Community Earth System Model. Here, we tested the model's capability to simulate whole-forest isotope discrimination in a subalpine conifer forest at Niwot Ridge, Colorado, USA. We distinguished between isotopic behavior in response to a decrease of  $\delta^{13}\text{C}$  within atmospheric CO<sub>2</sub> (Suess effect) vs. photosynthetic discrimination ( $\Delta_{\text{canopy}}$ ), by creating a site-customized atmospheric CO<sub>2</sub> and  $\delta^{13}\text{C}$  of CO<sub>2</sub> time series. We implemented a seasonally-varying  $V_{\text{cmax}}$  model calibration that best matched site observations of net CO<sub>2</sub> carbon exchange, latent heat exchange and biomass. The model accurately simulated observed  $\delta^{13}\text{C}$  of needle and stem tissue, but underestimated the  $\delta^{13}\text{C}$  of bulk soil carbon by 1-2 ‰. The model overestimated the multi-year (2006-2012) average  $\Delta_{\text{canopy}}$  relative to prior data-based estimates by 2-4 ‰. The amplitude of the average seasonal cycle of  $\Delta_{\text{canopy}}$  (i.e. higher in spring/fall as compared to summer) was correctly modeled but only when using a revised, fully-coupled  $A_n-g_s$  (net assimilation rate, stomatal

conductance) version of the model in contrast to the partially-coupled  $A_n$ - $g_s$  version used in the default model. The model attributed most of the seasonal variation in discrimination to  $A_n$ , whereas inter-annual variation in simulated  $\Delta_{\text{canopy}}$  during the summer months was driven by stomatal response to vapor pressure deficit (VPD). The model simulated a 10% increase in both photosynthetic discrimination and water-use efficiency (WUE) since 1850 which is counter to established relationships between discrimination and WUE. The isotope observations used here to constrain CLM suggest 1) the model overestimated stomatal conductance and 2) the default CLM approach to representing nitrogen limitation (partially-coupled model) was not capable of reproducing observed trends in discrimination. These findings demonstrate that isotope observations can provide important information related to stomatal function driven by environmental stress from VPD and nitrogen limitation. Future versions of CLM that incorporate carbon isotope discrimination are likely to benefit from explicit inclusion of mesophyll conductance.

## 1 Introduction

The net uptake of carbon by the terrestrial biosphere currently mitigates the rate of atmospheric CO<sub>2</sub> rise and thus the rate of climate change. Approximately 25% of anthropogenic CO<sub>2</sub> emissions are absorbed by the global land surface (Le Quéré et al., 2015), but it is unclear how projected changes in temperature and precipitation will influence the future of this land carbon sink (Arora et al., 2013; Friedlingstein et al., 2006). A major source of uncertainty in climate model projections results from the disagreement in projected strength of the land carbon sink (Arora et al., 2013). Thus, it is critical to reduce this uncertainty to improve climate predictions, and to better inform mitigation strategies (Yohe et al., 2007).

An effective approach to reduce uncertainties in terrestrial carbon models is to constrain a broad range of processes using distinct and complementary observations. Traditionally, terrestrial carbon models have relied primarily upon observations of land-surface fluxes of carbon, water and energy derived from eddy-covariance flux towers to calibrate model parameters and evaluate model skill. Flux measurements best constrain processes that occur at diurnal and seasonal time scales (Braswell et al., 2005; Ricciuto et al., 2008). Traditional ecological metrics of carbon pools (e.g. leaf area index (LAI), biomass) are also commonly used to provide independent and complementary constraints upon ecosystem processes at

longer time scales (Ricciuto et al., 2011; Richardson et al., 2010). However, neither flux nor carbon pool observations provide suitable constraints for the model formulation of plant stomatal function and the related link between the carbon and water cycles.

Stable carbon isotopes of CO<sub>2</sub> are influenced by stomatal activity in C3 plants (e.g. evergreen trees, deciduous trees), and thus provide a valuable but under-utilized constraint on terrestrial carbon models. Plants assimilate more of the lighter of the two major isotopes of atmospheric carbon (<sup>12</sup>C vs. <sup>13</sup>C). This preference, termed photosynthetic discrimination ( $\Delta_{\text{canopy}}$ ), is primarily a function of two processes, CO<sub>2</sub> diffusion rate through the leaf boundary layer and into the stomata, and the carboxylation of CO<sub>2</sub>. The magnitude of  $\Delta_{\text{canopy}}$  is controlled by CO<sub>2</sub> supply (depending on for instance atmospheric CO<sub>2</sub> concentration and stomatal conductance) and demand (depending on for instance photosynthetic rate; Flanagan et al., 2012). In general, environmental conditions favorable to plant productivity result in higher  $\Delta_{\text{canopy}}$  during carbon assimilation compared to unfavorable conditions. Plants respond to unfavorable conditions by closing stomata and reducing the stomatal conductance which reduces  $\Delta_{\text{canopy}}$ . Most relevant here,  $\Delta_{\text{canopy}}$  responds to atmospheric moisture deficit (Andrews et al., 2012; Wingate et al., 2010), soil water content (McDowell et al., 2010), precipitation (Roden and Ehleringer, 2007) and nutrient availability (Cernusak et al., 2013). After carbon is assimilated, additional post-photosynthetic isotopic changes occur (Bowling et al., 2008; Brüggemann et al., 2011), but these impose a small influence on land-atmosphere isotopic exchange relative to photosynthetic discrimination.

The Niwot Ridge Ameriflux site, located in a sub-alpine conifer forest in the Rocky Mountains of Colorado, U.S.A., has a long legacy of yielding valuable datasets to test carbon and water functionality of land surface models using stable isotopes. Niwot Ridge has a 17-year record of eddy covariance fluxes of carbon, water, and energy, as well as environmental data (Hu et al., 2010; Monson et al., 2002) and a 10-year record of  $\delta^{13}\text{C}$  of CO<sub>2</sub> in forest air (Schaeffer et al., 2008). From a carbon balance perspective, Niwot Ridge is representative of subalpine forests in Western North America that, in general, act as a carbon sink to the atmosphere (Desai et al., 2011). Western forests make up a significant portion of the carbon sink in the United States (Schimel et al., 2002), yet this sink is projected to weaken with projected changes in temperature and precipitation (Boisvenue and Running, 2010).

The Community Land Model (CLM), the land sub-component of the Community Earth System Model (CESM) has a comprehensive representation of biogeochemical cycling (Oleson

et al., 2013) that can be applied across a range of temporal (hours to centuries) and spatial (site to global) scales. A mechanistic representation of photosynthetic discrimination based upon diffusion and enzymatic fractionation (Farquhar et al., 1989) was included in the latest release of CLM 4.5 (Oleson et al., 2013), and is similar to the formulation implemented in other land surface models (Flanagan et al., 2012; Scholze et al., 2003; Wingate et al., 2010; van der Velde et al., 2013). An early version of CLM simulated carbon (but not carbon isotope) dynamics at Niwot Ridge with reasonable skill (Thornton et al., 2002).

Here, we evaluate the performance of the  $^{13}\text{C}/^{12}\text{C}$  isotope discrimination sub-model within CLM 4.5 against a range of isotopic observations at Niwot Ridge, to examine what new insights an isotope-enabled model can bring upon ecosystem function. Specifically, we test whether CLM simulates the expected isotopic response to environmental drivers of  $\text{CO}_2$  fertilization, soil moisture and atmospheric vapor pressure deficit (VPD). A previous analysis at Niwot Ridge showed a seasonal correlation between vapor pressure deficit (VPD) and photosynthetic discrimination (Bowling et al., 2014) suggesting that leaf stomata are responding to changes in VPD, and influencing discrimination. We use CLM to test whether VPD is the primary environmental driver of isotopic discrimination, as compared to soil moisture and net assimilation rate. Next we determine whether site-specific boundary conditions (including for instance  $\delta^{13}\text{C}$  of atmospheric  $\text{CO}_2$ ) combined with the representation of long term (multi-decadal to century) photosynthetic discrimination and simulated carbon pool turnover within the model, can accurately reproduce the measured  $\delta^{13}\text{C}$  in leaf tissue, roots and soil carbon. We then use CLM to determine if the increase in atmospheric  $\text{CO}_2$  since 1850 has led to an increase in water-use efficiency (WUE), and whether net assimilation or stomatal conductance is the primary driver of such a change. Finally, we ask what distinct insights site level isotope observations bring in terms of both model parameterization (i.e. stomatal conductance) and model structure as compared to the traditional observations (e.g. carbon fluxes, biomass).

## 2 Methods

We focus the description of CLM 4.5 (Section 2.1) upon photosynthesis, and its linkage to nitrogen, soil moisture and stomatal conductance (Section 2.1.1). Next we describe the model representation of carbon isotope discrimination by photosynthesis (Section 2.1.2). Because preliminary simulations demonstrated that model results were strongly influenced by nitrogen limitation, we used three separate nitrogen formulations (described in Section 2.1.2)

to better diagnose model performance. Next, to provide context for subsequent descriptions of site-specific model adjustments we describe the field site, Niwot Ridge, including the site level observations (Section 2.2) used to constrain and test model the model.

Patterns in plant growth and  $\delta^{13}\text{C}$  of biomass are strongly influenced by atmospheric  $\text{CO}_2$  and  $\delta^{13}\text{C}$  of atmospheric  $\text{CO}_2$  ( $\delta_{atm}$ ). Therefore we designed a site-specific synthetic atmospheric  $\text{CO}_2$  product (Section 2.3.1) and  $\delta_{atm}$  product (Section 2.3.2) for these simulations. The model setup and initialization procedure, intended to bring the system into steady state, is described in Section (2.3.3). This is followed by an explanation of the model calibration procedure that provided a realistic simulation of carbon and water fluxes (Section 2.4).

## 2.1 Community Land Model, Version 4.5

We used the Community Land Model, CLM 4.5 (Oleson et al., 2013), which is the land component of the Community Earth System Model (CESM) version 1.2 (<https://www2.cesm.ucar.edu/models/current>). Details regarding the Community Land Model can be found in Mao et al., (2016) and Oleson et al., (2013). Here, we emphasize the mechanistic formulation that controls photosynthetic discrimination ( $\Delta_{canopy}$ ) and factors that influence  $\Delta_{canopy}$  including photosynthesis, stomatal conductance, water stress and nitrogen limitation. A list of symbols is provided in Table (1).

### 2.1.1 Net Photosynthetic Assimilation

The leaf-level net carbon assimilation of photosynthesis,  $A_n$  is based on Farquhar et al., (1980) as,

$$A_n = \min(A_c, A_j, A_p) - Resp_d, \quad (1)$$

where  $A_c$ ,  $A_j$  and  $A_p$  are the enzyme (Rubisco)-limited, light-limited, and product-limited rates of carboxylation respectively, and  $Resp_d$  the leaf-level respiration. The enzyme limited rate is defined as

$$A_c = \frac{V_{cmax}(c_i - \Gamma_*)}{c_i + K_c(1 + \frac{o_i}{K_o})}, \quad (2)$$

where  $c_i$  is the intercellular leaf partial pressure of  $\text{CO}_2$ ,  $o_i = 0.209 P_{atm}$ ,  $P_{atm}$  is atmospheric pressure, and  $K_c$ ,  $K_o$  and  $\Gamma_*$  are constants. The maximum rate of carboxylation at  $25^\circ\text{C}$ ,  $V_{cmax25}$ , is defined as

$$V_{cmax25} = N_a F_{LNR} F_{NR} a_{R25} \beta_t, \quad (3)$$

where  $N_a$  is the nitrogen concentration per leaf area,  $F_{LNR}$  the fraction of leaf nitrogen within the Rubisco enzyme,  $F_{NR}$  the ratio of total Rubisco molecular mass to nitrogen mass within Rubisco, and  $a_{R25}$  is the specific activity of Rubisco at 25°C. The  $V_{cmax25}$  is adjusted for leaf temperature to provide  $V_{cmax}$  in Eq. 2, used in the final photosynthetic calculation. Both  $A_j$  and  $A_p$  are functions of  $V_{cmax}$  as well (not shown). The variable  $\beta_t$  represents the level of soil moisture availability, which influences both  $V_{cmax}$  (Sellers et al., 1996), and stomatal conductance (eq. 5). CLM calculates  $\beta_t$  as a factor (0-1, high to low stress) by combining soil moisture, the rooting depth profile, and a plant-dependent response to soil water stress as

$$\beta_t = \sum_i w_i r_i, \quad (4)$$

where  $w_i$  is a plant wilting factor for soil layer  $i$  and  $r_i$  is the fraction of roots in layer  $i$ . The plant wilting factor is scaled according to soil moisture and water potential, depending on plant functional type (PFT). Soil moisture is predicted based upon prescribed precipitation and vertical soil moisture dynamics (Zeng and Decker, 2009). The root fraction in each soil layer depends upon a vertical exponential profile controlled by PFT dependent root distribution parameters adopted from Zeng (2001).

The carbon and water balance are linked through  $c_i$  by the stomatal conductance to  $CO_2$ ,  $g_s$ , following the Ball-Berry model (Ball, 1987) as defined by Collatz et al., (1991),

$$g_s = m \frac{A_n}{c_s/P_{atm}} h_s + b \beta_t, \quad (5)$$

where  $m$  is the stomatal slope,  $c_s$  the partial pressure of  $CO_2$  at the leaf surface,  $h_s$  the relative humidity at the leaf surface and  $b$  the minimum stomatal conductance when the leaf stomata are closed.

The version of CLM used here has a 2-layer (shaded, sunlit) representation of the vegetation (Oleson et al. 2013). Photosynthesis and stomatal conductance are calculated separately for the shaded and sunlit portion and the total canopy photosynthesis is the potential gross primary productivity (GPP),  $CF_{GPPpot}$ :

$$CF_{GPPpot} = [(A_n + Resp_d)_{sunlit} (LAI)_{sunlit} + (A_n + Resp_d)_{shaded} (LAI)_{shaded}] * 12.011^{-6}, \quad (6)$$

where  $LAI$  is the leaf area index and  $12.011^{-6}$  is a unit conversion factor. The total carbon available for new growth allocation ( $CF_{avail\_alloc}$ ) is defined as



$$CF_{avail\_alloc} = CF_{GPPpot} - CF_{GPP,mr} - CF_{GPP,xs}, \quad (7)$$

where the maintenance respiration is derived either from recently assimilated photosynthetic carbon ( $CF_{GPP,mr}$ ), or, if photosynthesis is low or zero (e.g. night), the maintenance respiration is drawn from a carbon storage pool ( $CF_{GPP,xs}$ ). In contrast,  $CF_{alloc}$  is the actual carbon allocated to growth calculated from the available nitrogen and fixed C:N ratios for new growth (e.g. stem, roots, leaves). The downregulation of photosynthesis from nitrogen limitation,  $f_{dreg}$ , is given by

$$f_{dreg} = \frac{CF_{avail\_alloc} - CF_{alloc}}{CF_{GPPpot}}. \quad (8)$$

The actual, nitrogen-limited GPP is defined as:

$$GPP = CF_{GPPpot}(1 - f_{dreg}) \quad (9)$$

11

## 2.1.2 Photosynthetic Carbon Isotope Discrimination

The canopy-level fractionation factor  $\alpha$  is defined as the ratio of  $^{13}\text{C}/^{12}\text{C}$  within atmospheric  $\text{CO}_2$  ( $R_a$ ) and the products of photosynthesis ( $R_{GPP}$ ) as  $\alpha = \frac{R_a}{R_{GPP}}$ . The preference of C3 vegetation to assimilate the lighter  $\text{CO}_2$  molecule during photosynthesis is simulated in CLM with two steps: diffusion of  $\text{CO}_2$  across the leaf boundary layer and into the stomata, followed by enzymatic fixation to give the leaf-level fractionation factor:

$$\alpha = 1 + \frac{4.4 + 22.6 \frac{c_i^*}{c_a}}{1000}. \quad (10)$$

where  $c_i^*$  and  $c_a$  are the intracellular and atmospheric  $\text{CO}_2$  partial pressure respectively. The numbers 4.4 and 22.6 represent the diffusional and enzymatic contributions to isotopic discrimination respectively (Farquhar et al., 1989). The variable  $c_i^*$  (known in CLM as the ‘revised intracellular  $\text{CO}_2$  partial pressure’) is marked with an asterisk to indicate the inclusion of nitrogen downregulation as defined as,

$$c_i^* = c_a - A_n (1 - f_{dreg}) P_{atm} \frac{(1.4g_s) + (1.6g_b)}{g_b g_s} \quad (11)$$

where  $g_b$  is the leaf boundary layer conductance. Equation 11 is a general expression for  $c_i^*$ , where within the model  $c_i^*$  and discrimination are calculated for the sunlit and shaded layer of leaves separately and subject to the local environmental conditions unique to each layer (Oleson

et al. 2013). The inclusion of the nitrogen downregulation factor  $f_{dreg}$  in the above expression reflects the two-stage process in which the potential photosynthesis (Eq. 6) and the actual photosynthesis (Eq. 9) are calculated within CLM and prevents a mismatch between the actual photosynthesis and the intracellular CO<sub>2</sub>. This mismatch is a result of the carbon-water (An- gs) coupling (eq. 5) being imposed prior to the effect of nitrogen limitation (eq. 9), and is an artifact of the model implementation. We also test a separate model formulation (described in detail in the next paragraph) specific to this analysis that imposes nitrogen limitation through the  $V_{cmax}$  parameterization and removes the artifact of  $f_{dreg}$ . Equation 11 is a general expression for  $c_i^*$ , where within the model  $c_i^*$  and discrimination are calculated for the sunlit and shaded layer of leaves separately and subject to the local environmental conditions unique to each layer (Oleson et al. 2013).

The sensitivity of preliminary model results to nitrogen limitation led us to test three distinct discrimination formulations (Figure 1; Table 2). The *limited nitrogen* formulation was based on the default version of CLM 4.5 and included both nitrogen limitation and the nitrogen downregulation factor within the calculation of  $c_i^*$  as given in equation (11). The second, *unlimited nitrogen* formulation, which we created specifically for this analysis, also follows equation (11), however, the vegetation is allowed unlimited access to nitrogen ( $CF_{GPPpot} = GPP, f_{dreg} = 0$ ) which ignores the nitrogen budget within CLM. We account for the increased productivity in the *unlimited nitrogen* model simulations by calibrating  $V_{cmax}$  (Section 2.4). Finally, in the *no downregulation discrimination* formulation, (also created specifically for this analysis) we included nitrogen limitation, but removed the downregulation factor  $f_{dreg}$  within the isotopic discrimination equation (11).

In the *unlimited nitrogen* formulation, we use a different modifier on  $V_{cmax25}$  (Figure 1; described in section 2.4 and Fig. S1, S2) in the calibrated runs to give similar carbon flux, water flux and biomass as in the other two formulations, such that all three formulations have fluxes and biomass that are similar to what is observed at the site, and which presumably reflect nitrogen limitation. Thus the distinction between these three formulations can be viewed entirely as *when* nitrogen limitation is imposed in relation to photosynthesis: (1) *after photosynthesis* via a downregulation between potential and actual GPP (equation 9) that feeds back on the  $c_i^*/c_a$  used for isotopic discrimination but *not on*  $g_s$  or  $A_n$  in the *limited nitrogen* formulation; (2) *before photosynthesis* via  $V_{cmax}$ , which limits photosynthetic capacity affecting both  $c_i^*/c_a$ ,  $g_s$  and  $A_n$  in the *unlimited nitrogen* formulation; and (3) *after photosynthesis* with

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no effect on either the  $c_i/c_a$  for isotopic discrimination or  $g_s$  or  $A_n$  in the *no downregulation* discrimination formulation. The downscaled portion of the carbon during nitrogen limitation ( $CF_{GPPpot} - GPP$ ) is removed from the system and does not appear as a respired flux (Figure 1). In summary, the *limited nitrogen (post-photosynthetic)* formulation adjusts the photosynthetic rate by explicitly tracking N availability, whereas the *unlimited nitrogen (pre-photosynthetic)* formulation takes into account any N-limitation through the  $V_{cmax}$  parameterization. Because the *limited nitrogen* formulation reduces  $A_n$  during the nitrogen downregulation step without explicitly solving for  $g_s$ , the carbon-water cycle is ‘partially-coupled’, whereas the *unlimited nitrogen* formulation is ‘fully-coupled’.

Carbon isotope ratios are expressed by standard delta notation,

$$\delta^{13}C_x = \left( \frac{R_x}{R_{VPDB}} - 1 \right) \times 1000, \quad (12)$$

where  $R_x$  is the isotopic ratio of the sample of interest, and  $R_{VPDB}$  is the isotopic ratio of the Vienna Pee Dee Belemnite standard. The delta notation is dimensionless but expressed in parts per thousand (‰) where a positive (negative) value refers to a sample that is enriched (depleted) in  $^{13}C/^{12}C$  relative to the standard. Because this is the only carbon isotope ratio we are concerned with in this paper, the ‘13’ superscript is omitted for brevity in subsequent definitions using the delta notation. The canopy-integrated photosynthetic discrimination,  $\Delta_{canopy}$ , is defined as the difference between the  $\delta^{13}C$  of the atmospheric and assimilated carbon,

$$\Delta_{canopy} = \delta_{atm} - \delta_{GPP}. \quad (13)$$

The difference between  $\delta^{13}C$  of the total ecosystem respiration (ER) and GPP fluxes, called the isotope disequilibrium (Bowling et al., 2014), is defined as,

$$disequilibrium = \delta_{ER} - \delta_{GPP}. \quad (14)$$

The ecosystem-level water-use efficiency (WUE) is defined as actual carbon assimilated ( $GPP$ ) per unit water transpired ( $E_T$ ) per unit land surface area,

$$WUE = \frac{GPP}{E_T}. \quad (15)$$

The intrinsic water-use efficiency ( $iWUE$ ) from leaf-level physiological ecology is defined as,

$$iWUE = \frac{A_n}{g_s}, \quad (16)$$

1 where  $A$  is the net carbon assimilated per unit leaf area and  $g_s$  is the stomatal conductance.  
2 CLM calculates  $g_s$  (Equation 5) for shaded and sunlit portions of the canopy separately,  
3 therefore an overall conductance was calculated by weighting the conductance by sunlit and  
4 shaded leaf areas.

## 5 2.2 Niwot Ridge and site-level observations

6 Site-level observations and modeling were focused on the Niwot Ridge Ameriflux  
7 tower (US-NR1), a sub-alpine conifer forest located in the Rocky Mountains of Colorado,  
8 U.S.A. The forest is approximately 110 years old and consists of lodgepole pine (*Pinus*  
9 *contorta*), Engelmann spruce (*Picea engelmanni*), and subalpine fir (*Abies lasiocarpa*). The  
10 site is located at an elevation of 3050 m above sea level, with mean annual temperature of  
11 1.5°C and precipitation of 800 mm, in which approximately 60% is snow. More site details  
12 are available elsewhere (Hu et al., 2010; Monson et al., 2002). Flux and meteorological data  
13 were obtained from the Ameriflux archive (<http://ameriflux.lbl.gov/>).

14 Net carbon exchange (NEE) observations were derived from flux tower measurements  
15 based on the eddy covariance method and were partitioned into component fluxes of GPP and  
16 ER according to two separate methods described by Reichstein et al., (2005) and Lasslop et al.,  
17 (2010) using an online tool provided by the Max Planck Institute for Biogeochemistry  
18 (<http://www.bgc-jena.mpg.de/~MDIwork/eddyproc/>). Seasonal patterns in  $\delta_{GPP}$  and  $\delta_{ER}$  were  
19 derived from measurements as described by Bowling et al., (2014). Observations of  $\delta^{13}C$  of  
20 biomass (Schaeffer et al., 2008) and carbon stocks (Bradford et al., 2008; Scott-Denton et al.,  
21 2003) were compared to model simulations. Schaeffer et al., (2008) reported soil, leaf and  
22 root observations specific to each conifer species, however, the observed mean and standard  
23 error for all species were used for comparison because CLM treated all conifer species as a  
24 single PFT.

## 25 2.3 Atmospheric CO<sub>2</sub>, isotope forcing and initial vegetation state

### 26 2.3.1 Site-specific atmospheric CO<sub>2</sub> concentration time series

27 Global average atmospheric CO<sub>2</sub> concentrations increased roughly 40% from 1850 to  
28 2013 (from 280 to 395 ppm). The standard version of CLM 4.5 includes an annually and  
29 globally averaged time series of this CO<sub>2</sub> increase, however, this does not capture the observed

seasonal cycle of ~10 ppm at Niwot Ridge (Troler et al., 1996). Therefore we created a site-specific atmospheric CO<sub>2</sub> time series (Figure 2) to provide a seasonally realistic atmosphere at Niwot Ridge. From 1968-2013 the CO<sub>2</sub> time series was fit to flask observations (Dlugokencky et al., 2015) from Niwot Ridge. Prior to 1968, the CO<sub>2</sub> time series was created by combining the average multi-year seasonal cycle based on the Niwot Ridge flask data to the annual CO<sub>2</sub> product provided by CLM. More details are located in the supplemental material.

### 2.3.2 Customized $\delta^{13}\text{C}$ atmospheric CO<sub>2</sub> time series

As atmospheric CO<sub>2</sub> has increased, the  $\delta^{13}\text{C}$  of atmospheric CO<sub>2</sub> ( $\delta_{atm}$ ) has become more depleted (Francey et al., 1999), and this change continues at Niwot Ridge at -0.25 ‰ per decade (Bowling et al., 2014). The  $\delta_{atm}$  also varies seasonally, and depends on latitude (Troler et al., 1996). However, CLM 4.5 as released assigned a constant  $\delta^{13}\text{C}$  of -6 ‰. We therefore created a synthetic time series of  $\delta_{atm}$  from 1850-2013 (Figure 2). From 1990-2013 the time series was fit to the flask observations (White et al., 2015) as described in Section 2.3.1. Prior to 1990 the inter-annual variation within the  $\delta_{atm}$  time series was fit to the ice core data from Law Dome (Francey et al., 1999; see also Rubino et al., 2013). This annual data product was then combined with the average seasonal cycle at Niwot Ridge as determined by the flask observations to create the synthetic product from 1850-1990. More details are located in the supplemental material.

### 2.3.3 Model Initialization

We performed an initialization to transition the model from near bare-ground conditions to present day carbon stocks and LAI that allowed for proper evaluation of isotopic performance. This was implemented in 4 stages: 1) accelerated decomposition (1000 model years) 2) normal decomposition (1000 model years) 3) parameter calibration (1000 model years) and 4) transient simulation period (1850-2013). The first two stages were pre-set options within CLM with the first stage used to accelerate the equilibration of the soil carbon pools, which require a long period to reach steady state (Thornton and Rosenbloom, 2005). The parameter calibration stage was not a pre-set option but designed specifically for our analysis. For this we introduced a seasonally varying  $V_{cmax}$  that scaled the simulated GPP and ecosystem

respiration fluxes to present day observations (Section 2.4). In the transient phase, we introduced time-varying atmospheric conditions from 1850-2013 including nitrogen deposition (CLM provided), atmospheric CO<sub>2</sub>, and  $\delta_{atm}$  (site-specific as described above). Environmental conditions of temperature, precipitation, relative humidity, radiation, and wind speed were taken from the Niwot Ridge flux tower observations from 1998-2013 and then cycled continuously for the entirety of the initialization process. We used a scripting framework (PTCLM) that automated much of the workflow required to implement several of these stages in a site level simulation (Mao et al., 2016; Oleson et al., 2013).

## 2.4 Specific model details and model calibration

This version of CLM included a fully prognostic representation of carbon and nitrogen within its vegetation, litter and soil biogeochemistry. We used the Century model representation for soil (3 litter and 3 soil organic matter pools) with 15 vertically resolved soil layers (Parton et al. 1987). Nitrification and prognostic fire were turned off. Our initial simulations used prognostic fire, but we found that simulated fire was overactive leading to low simulated biomass compared to observations. Although Niwot Ridge has been subject to disturbance from fire and harvest in the past, ultimately our final simulations did not include either fire or harvest disturbance because the last disturbance occurred over 110 years ago (early 20th century logging; Monson et al., 2005).

Ecosystem parameter values (Table 3) used here were based upon the temperate evergreen needleleaf PFT within CLM. These values were based upon observations reported by White et al., (2000) intended for a wide range of temperate evergreen forests, and by Thornton et al., (2002) for Niwot Ridge. For this analysis two site-specific parameter changes were made. First, the e-folding soil decomposition parameter was increased from 5 to 20 meters. This parameter is a length-scale for attenuation of decomposition rate for the resolved soil depth from 0 to 5 meters where an increased value effectively increases decomposition at depth, thus reducing total soil carbon and more closely matching observations. Second, we performed an empirical photosynthesis scaling (equation 17, below) that reduced the simulated photosynthetic flux, as guided by eddy covariance observations (Figure 3; Figure S1). Consequently, all downstream carbon pools and fluxes including ecosystem respiration, aboveground biomass, and leaf area index which provided a better match to present day

observations. This approach also removed a systematic overestimation of winter photosynthesis. The model simulations without the photosynthetic scaling are referred to within the text and figures as the *uncalibrated* model, whereas model simulations that include the photosynthetic scaling are referred to as the *calibrated* model. We modified CLM for this scaling approach by reducing  $V_{cmax}$  at 25° C,

$$V_{cmax25} = N_a F_{LNR} F_{NR} a_{R25} \beta_t f_{df}, \quad (17)$$

where  $f_{df}$  is the photosynthetic scaling factor, and all other parameters are identical to equation (3). These parameters were constant for the entirety of the simulations except for  $f_{df}$ , an empirically derived time dependent parameter ranging from 0-1. The value was set to zero to force photosynthesis to zero between November 13th and March 23rd, consistent with flux tower observations where outside of this range  $GPP > 0$  was never observed. During the growing season period ( $GPP > 0$ ) within days of year 83-316,  $f_{df}$  was calculated as

$$f_{df} = \frac{\text{observed } GPP(\text{day of year})}{\text{simulated } GPP(\text{day of year})}, \quad 82 < \text{day of year} < 317 \quad (18)$$

where the *observed GPP* was the daily average calculated from the partitioned flux tower observations (Reichstein et al., 2005) from 2006-2013, and the *simulated GPP* was the daily average of the unscaled value during the same time. A polynomial was fit to equation (18) that represented  $f_{df}$  for 1) both the *limited nitrogen* and *no downregulation discrimination formulations* and 2) the *unlimited nitrogen* formulation (Figure S2). Note that CLM already includes a *daylength factor* that also adjusts the magnitude of  $V_{cmax}$  according to time of year, however, that default parameterization alone was not sufficient to match the observations. The light-limited rate and product-limited rate of carboxylation ( $A_j, A_p$ ; eq. 1) and maintenance leaf respiration are functions of  $V_{cmax}$  (not shown) and are therefore subject to the same calibration.

## 1    3    Results & Discussion

### 2    3.1    Calibrated model performance

#### 3    3.1.1    Fluxes & carbon pools

4            The CLM model (*limited nitrogen* simulation) was successful at simulating GPP, ER, and  
5    latent heat fluxes (Fig. 3), leaf area index (LAI), and aboveground biomass (Fig. 4), but only  
6    following site-specific calibration. Similar improvement was observed after calibration for the  
7    *unlimited nitrogen* run (not shown). The calibration also eliminated erroneous winter GPP.  
8    In general, terrestrial carbon models tend to overestimate photosynthesis during cold periods  
9    for temperate/boreal conifer forests (Kolari et al., 2007), including Niwot Ridge (Thornton et  
10    al., 2002). Although our calibration approach forced  $V_{cmax}$  to zero during the winter, it did not  
11    solve the underlying mechanistic shortcoming. A more fundamental approach should address  
12    either cold inhibition (Zarter et al., 2006) of photosynthesis or soil water availability associated  
13    with snowmelt (Monson et al., 2005) to achieve the photosynthetic reduction. Nevertheless,  
14    within the confines of our study area, our calibration approach was sufficient to provide a  
15    skillful representation of photosynthesis and provided a sufficient testbed for evaluating carbon  
16    isotope behavior. We caution that because the  $f_{df}$  parameter (eq. 17) was calibrated specifically  
17    for Niwot Ridge, it would not be applicable outside this study area.

#### 18    3.1.2    $\delta^{13}\text{C}$ of carbon pools

19            The model performed better simulating  $\delta^{13}\text{C}$  biomass of bulk needle tissue, roots and soil  
20    carbon (Figure 5) for the *unlimited nitrogen* and no *downregulation discrimination* cases as  
21    compared to the *limited nitrogen* case. When nitrogen limitation was included the model  
22    underestimated  $\delta^{13}\text{C}$  of sunlit needle tissue (1.8 ‰), bulk roots (1.0 ‰), and organic soil carbon  
23    (0.7‰). All simulations fell within the observed range of  $\delta^{13}\text{C}$  in needles that span from -28.7  
24    ‰ (shaded) to -26.7 (sunlit). This vertical pattern in  $\delta^{13}\text{C}$  of leaves is common (Martinelli et  
25    al., 1998) and results from vertical differences in nitrogen allocation and photosynthetic  
26    capacity. The model results integrated the entire canopy and ideally should be closer to sun  
27    leaves (as in Figure 5) given that the majority of photosynthesis occurs near the top of the  
28    canopy.

29            Model simulations of  $\delta^{13}\text{C}$  of living roots were ~1 ‰ more negative as compared to the  
30    structural roots. This range in  $\delta^{13}\text{C}$  results from decreasing  $\delta_{atm}$  with time (Suess effect, Figure



2). The living roots had a relatively fast turnover time of carbon within the model, whereas the structural roots had a slower turnover time and reflected an older (more enriched  $\delta_{atm}$ ) atmosphere. The *limited nitrogen* simulation was a poor match to observations relative to the others (Figure 5, panel B).

There was an observed vertical gradient in  $\delta^{13}C$  of soil carbon (-24.9 to -26 ‰) with more enriched values at greater depth (Figure 5, panel C). This vertical gradient is commonly observed (Ehleringer et al., 2000). Simulated  $\delta^{13}C$  of soil carbon was most consistent with the organic horizon observations. There are a wide variety of post-photosynthetic fractionation processes in the soil system (Bowling et al., 2008; Brüggemann et al., 2011) that are not considered in the CLM 4.5 model, so the match with observations is perhaps fortuitous.

## 3.2 Photosynthetic discrimination

### 3.2.1 Decadal changes in photosynthetic discrimination and driving factors

All modeled carbon pools showed steady depletion in  $\delta^{13}C$  since 1850 (coinciding with the start of the transient phase of simulations, Figure 5). For the *limited nitrogen* run, there was a decrease in  $\delta^{13}C$  of 2.3 ‰ for needles, 2.3 ‰ for living roots, and 0.1 ‰ for soil carbon. This occurred because of 1) decreased  $\delta_{atm}$  (Suess effect, Figure 2) and 2) increased photosynthetic discrimination. We quantified the contribution of the Suess effect by performing a control run with constant  $\delta_{atm}$ , and kept other factors the same (Figure 6). Approximately 70% of the reduction in  $\delta^{13}C$  of needles occurred due to the Suess effect, and the remaining 30% was caused by increased photosynthetic discrimination. This occurred as plants responded to  $CO_2$  fertilization as illustrated in Figure (7). The model indicated that plants responded to increased atmospheric  $CO_2$  (~40% increase) by decreasing stomatal conductance (Equation 5) by 20% for the *limited nitrogen* run and 30% for the *unlimited nitrogen* run (Figure 7, panel B) with associated change in  $c_i/c_a$  (Figure 7, panel A). Other influences upon stomatal conductance were less significant, including  $A_n$  (+ 10% *limited nitrogen*, -10% *unlimited nitrogen*, Figure 7, panel D), soil moisture availability (2-3%, Figure 7, panel E), and negligible changes in relative humidity (multi-decadal climate change effects are neglected due to methodological cycling of weather data). This finding that stomatal conductance responded to atmospheric  $CO_2$  is consistent with both tree ring studies (Saurer et al., 2014) and site-level experiments (Ward et al., 2012).

The effect of CO<sub>2</sub> fertilization, and associated response of stomatal conductance and net assimilation led to a multi-decadal increase in  $c_i^*/c_a$  for all model formulations (Figure 7, panel A). The  $c_i^*/c_a$  increased from 0.71 to 0.76, 0.67 to 0.71 and 0.66 to 0.68 for the *limited nitrogen*, *unlimited nitrogen* and *no downregulation discrimination* formulations respectively from 1850-2013. All simulations therefore suggested an *increase* in photosynthetic discrimination. This increase in discrimination falls in between two hypotheses posed by Saurer et al., (2004) regarding stomatal response to increased CO<sub>2</sub>: 1) reduction in stomatal conductance causes  $c_i$  to proportionally increase with  $c_a$  keeping  $c_i/c_a$  constant and 2) minimal stomatal conductance response where  $c_i$  increases at the same rate as  $c_a$  (constant  $c_a - c_i$ ) causing  $c_i/c_a$  to increase. Our simulation generally agrees with the observed trend in  $c_i/c_a$  as estimated from tree ring isotope measurements from a network of European forests (Frank et al., 2015). When controlled for trends in climate, Frank et al. (2015) found that  $c_i/c_a$  was approximately constant during the last century. If the Niwot Ridge multi-decadal warming trends in temperature and humidity (Mitton and Ferrenberg, 2012) were included in the CLM simulations (this analysis did not consider multi-decadal climate change) the stomatal response may have been stronger thereby holding  $c_i/c_a$  constant.

The simulated stomatal closure in response to CO<sub>2</sub> fertilization led to an increase in iWUE and WUE of approximately 20% and 10% respectively (Figure 7, panel F) from 1960-2000. This simulated increase in iWUE is consistent with the observation-based studies (Ainsworth and Long, 2005; Franks et al., 2013; Peñuelas et al., 2011) which indicate a 15-20% increase in iWUE for forests during that time. The overall increase in WUE suggests that the vegetation at Niwot Ridge has some ability to maintain net ecosystem productivity when confronted with low soil moisture, low humidity conditions. Ultimately, whether Niwot Ridge maintains the current magnitude of carbon sink (Figure 3; Figure S1) will depend upon the severity of drought conditions, as improvements in WUE, in general, are only likely to negate weak to moderate levels of drought (Frank et al., 2013).

The *limited nitrogen* formulation simulated larger values of  $A_n$  and  $g_s$ , and smaller iWUE as compared to the *unlimited nitrogen* formulation (Figure 7). This is because the *unlimited nitrogen* formulation was fully coupled (i.e. solved simultaneously) between  $A_n$  and  $g_s$  (Eq. 5). The *limited nitrogen* formulation, however, was only partially coupled because  $A_n$  and  $g_s$  were initially solved simultaneously through the potential  $A_n$  (Eq. 1), however, under N-limitation

$A_n$  becomes limited below its potential value (Eq. 9) through  $f_{dreg}$ . Therefore  $g_s$  is calculated through the potential  $A_n$  (Eq. 5) and not the N-limited  $A_n$ .

The simultaneous increase in both simulated photosynthetic discrimination and iWUE conflicts with previous literature where increases in iWUE are typically linked with weakening discrimination (e.g. Saurer et al., 2004) using a linear model. In general, an increase in atmospheric CO<sub>2</sub> alone tends to increase iWUE because of reduced stomatal conductance, however, the impact upon discrimination is close to neutral because the increased supply of CO<sub>2</sub> external to the leaf is offset by reduced stomatal conductance (Saurer et al., 2004). The VPD likely plays an important role in determining the final trends for iWUE and discrimination, where an increasing VPD should further reduce stomatal conductance ( $VPD \propto \frac{1}{h_s}$ , eq. 5) thereby promoting the well-established relationship (increasing iWUE, decreasing discrimination). In contrast, a weak or decreasing trend in VPD should promote the opposite relationship (increasing iWUE, increasing discrimination).

The CLM model at present neglects mesophyll conductance ( $g_m$ ). When Seibt et al. (2008) included  $g_m$  in a model that linked iWUE to discrimination they found there were certain conditions when iWUE and discrimination increased together. This is in part because mesophyll conductance, unlike stomatal conductance, does not respond as strongly to changes in VPD, yet has a significant impact upon  $c_i/c_a$  and discrimination (Flexas et al., 2006). Harvard Forest is an example of a site that was observed to show simultaneous increase in iWUE and discrimination over the last two decades derived from tree rings (Belmecheri et al., 2014). In our model simulation, we do not consider multi-decadal trends in climate or mesophyll conductance, therefore increasing atmospheric CO<sub>2</sub> must be the primary driver for the modeled simultaneous increase in discrimination and iWUE at Niwot Ridge (Figure 7). These trends in iWUE and discrimination have also been found in a fully-coupled, isotope enabled, global CESM1.2 model run with climate simulated by CAM5 (Community Atmospheric Model) simulation driven by CO<sub>2</sub> emissions (unpublished, K. Lindsay; Figure S3). Specifically, a random sample of land model grid cells representing conifer species in British Columbia (lat: 52.3° N, lon: -122.5° W) and Quebec (lat: 49.5° N, lon: -70.0° W) all showed an increase in photosynthetic discrimination and a 10% increase in WUE from 1850-2005. These randomly chosen grid cells are likely better analogs to the site-level simulations described here because they represent boreal conifer forests, whereas the grid cells that are in the Niwot Ridge area

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were heterogeneous in land cover (e.g. tundra, grassland, forest) and a poor representation of conifer forest.

The relationship between iWUE and discrimination in the global [CESM1.2 model runsimulation \(with model-simulated climate\)](#) suggest that the site level trends are not isolated to the specific conditions of Niwot Ridge, but are a function of the model formulation. There is a relationship between iWUE and  $c_i^*/c_a$  (discrimination) as derived from equation (11) within the CLM model,

$$\frac{c_i^*}{c_a} \cong 1 - \frac{1.6}{c_a} iWUE. \quad (19)$$

The full derivation is provided in the supplement. Note that according to equation (19) increasing iWUE can be consistent with weakening discrimination (decreasing  $c_i^*/c_a$  ( $\sim \alpha$ )) and therefore consistent with established understanding between trends in iWUE and discrimination. However, this can be moderated by increasing  $c_a$ . During the course of our simulation (1850-2013) iWUE increased between 15-20% (Figure 7), however,  $c_a$  increased by 40%.

### 3.2.2 Magnitude of photosynthetic discrimination

The simulated photosynthetic discrimination (Fig. 8) was significantly larger than an estimate derived from observations and an isotopic mixing model (Bowling et al., 2014). For brevity we refer to the estimates based on the Bowling et al. (2014) method as ‘observed’ discrimination but highlight that they are derived from observations and not directly measured. On average, the simulated monthly growing season mean canopy discrimination was greater than observed values by 4.0, 2.3, and 1.8‰ for the *limited nitrogen*, *unlimited nitrogen*, and *no downregulation discrimination* formulations respectively. The model-observation mismatch in discrimination, despite model-observation agreement to biomass, carbon and latent heat flux tower observations (Figure 3) highlights the independent, and useful constraint isotopic observations provide for evaluating model performance. Specifically, the overestimation of discrimination may suggest the stomatal slope in the Ball-Berry model ( $m=9$  in Eq. 5) used for these simulations was too high. This is supported by Mao et al., (2016), who found a reduced stomatal slope ( $m=5.6$ ) was necessary for CLM 4.0 to match observed  $\delta^{13}\text{C}$  in an isotope labeling study of loblolly pine forest in Tennessee. The stomatal slope was also important to

match discrimination behavior in the ISOLSM model (Aranibar et al., 2006), a predecessor to CLM. A global analysis of stomatal slope inferred from leaf gas exchange measurements found that evergreen coniferous species, such as those at Niwot Ridge, had near the lowest values compared to other PFTs (Lin et al., 2015). In addition, they found that low stomatal slope values were characteristic of species with low stemwood construction costs per water transpired (high WUE), low soil moisture availability, and cold temperatures.

Alternatively, discrimination may be overestimated because CLM does not consider the resistance to CO<sub>2</sub> diffusion into the leaf chloroplast. The ability of CO<sub>2</sub> to diffuse across the chloroplast boundary layer, cell wall, and liquid interface is collectively known as the mesophyll conductance ( $g_m$ ) (Flexas et al., 2008). Multiple studies suggest that  $g_m$  is comparable in magnitude to  $g_s$ , and responds similarly to environmental conditions (Flexas et al., 2008). CLM does not account for  $g_m$ , and as a result assumes the intracellular CO<sub>2</sub> is the same as intercellular CO<sub>2</sub>, when it can be significantly lower (Di Marco et al., 1990; Sanchez-Rodriguez et al., 1999). The overestimation of  $c_i^*$  could have two important impacts upon our simulation. First, this may lead to unrealistically low values of  $V_{cmax}$  in order to compensate for the overestimation of  $c_i^*$ . In fact, we reduced the default value of  $V_{cmax}$  as much as 50% in our simulation to match the eddy covariance flux tower observations (see Section 4.1). Second, the overestimation of  $c_i^*$  should cause an overestimation of discrimination (Eq. 10), which is also consistent with our simulations (Figure 8). To determine whether the simulated discrimination bias is a model parameter calibration issue ( $g_s$ ) or from excluding  $g_m$ , we recommend a mechanistic representation of mesophyll conductance within CLM.

The mixing model approach estimate of  $\Delta_{canopy}$  (17 ‰) (Bowling et al. 2014), combined with  $\delta_{atm}$  (-8.25 ‰) implies a  $\delta^{13}C$  of biomass between -26 to -25 ‰ (Figure 8). This range is only slightly more enriched than the observed ranges of  $\delta^{13}C$  of needle and root biomass (-27 to -26 ‰). The fact that the different approaches to measure discrimination differ by only 1 ‰, whereas CLM simulates a  $\Delta_{canopy}$  that is 1.8 to 4.0 ‰ greater than the mixing model discrimination, strongly suggests that the model has overestimated discrimination from 2006-2012. Therefore what appeared to be a successful match between the simulated and observed  $\delta^{13}C$  biomass, may in fact have been fortuitous. A multi-decadal time series of discrimination

1 inferred from  $\delta^{13}\text{C}$  of tree rings (Saurer et al., 2014; Frank et al., 2015) would be useful to  
2 investigate this mismatch as a function of time, but these data are not presently available.

3         If the overestimation of modeled discrimination originates from a lack of response of  
4 stomatal conductance to environmental conditions this could be a result of one or several of the  
5 following within the model: 1) the stomatal slope value is too high, 2) multi-decadal trends in  
6 climate (e.g. VPD) have not been included in the simulation 3) the model neglects  $g_m$  or 4) the  
7 Ball-Berry representation of  $g_s$  is not sensitive enough to changes in environmental conditions  
8 (e.g. humidity, soil moisture). It has been shown that VPD may be an improved predictor of  $g_s$   
9 (Katul et al., 2000; Leuning, 1995) and discrimination (Ballantyne et al., 2010, 2011) as  
10 compared to relative humidity, currently used in CLM 4.5. Future work should consider which  
11 of these scenarios is responsible for overestimation of discrimination.

### 12 3.2.3 Seasonal pattern of photosynthetic discrimination

13         The model formulations that did not explicitly consider the influence of nitrogen  
14 limitation upon discrimination (*unlimited nitrogen, no downregulation discrimination*) were  
15 most successful at reproducing the seasonality of discrimination (Figure 8; Figure S4). In  
16 general, the observed discrimination was stronger during the spring and fall and weaker during  
17 summer. This observed  $\Delta_{\text{canopy}}$  seasonal range (excluding November) varied from 16.5 to 18  
18 ‰ using Reichstein partitioning (Figure 8), and was more pronounced using Lasslop  
19 partitioning (16.5 to 23 ‰) (Figure S4). The nitrogen limited simulated  $\Delta_{\text{canopy}}$  had no seasonal  
20 trend whereas the *unlimited nitrogen* and *no downregulation discrimination* simulations ranged  
21 from 18.4 to 21.2 and 17.8 to 20.6 ‰ respectively.

22         The main driver of the seasonality of discrimination was the net assimilation ( $A_n$ ) for  
23 the *unlimited nitrogen* formulation (Figure 9). This was evident given the inversely  
24 proportional relationship between the simulated fractionation factor ( $\alpha$ ) and  $A_n$ , consistent with  
25 equation (11). Stomatal conductance ( $g_s$ ) also influenced the seasonal pattern. The most direct  
26 evidence for this was during the period between days 175-200 (Figure 9), where  $A_n$  descended  
27 from its highest value (favoring higher  $\alpha$ ), and  $g_s$  abruptly ascended to its highest value  
28 (favoring higher  $\alpha$ ). The  $\alpha$  responded to this increase in  $g_s$  with an abrupt increase by  
29 approximately 0.003 (3 ‰). Similarly, the *limited nitrogen* simulation seasonal discrimination  
30 pattern was shaped by both  $A_n$  and  $g_s$ , although the magnitude for both was approximately 30%  
31 higher during the summer months as compared to the unlimited nitrogen simulation. This was

because the calibrated  $V_{cmax}$  value for the *limited nitrogen* simulation was much higher than for the *unlimited nitrogen* simulation (section 4.1). The difference in  $\alpha$  between the two model formulations coincided with the sharp increase in  $f_{dreg}$  between days 125 and 275, providing strong evidence that the downregulation mechanism within the *limited nitrogen* formulation led to increased discrimination during the summer. Therefore, it follows that the nitrogen downregulation mechanism was the root cause of the small range in simulated seasonal cycle discrimination for the *limited nitrogen* formulation, which was inconsistent with the observations.

### 3.2.4 Environmental factors influencing seasonality of discrimination

The simulated  $\Delta_{canopy}$  was driven primarily by net assimilation ( $A_n$ ), followed by vapor pressure deficit (VPD) (Fig. 10). The correlation between VPD and  $\Delta_{canopy}$  was strongest for the *unlimited nitrogen* simulation, where the range in monthly average  $\Delta_{canopy}$  spanned values from 22 to 18 ‰ (Figure 10, panels B, E and H). This resembled the observed range in response based upon a fitted relationship from Bowling et al., (2014) that spanned from roughly 16 to 19 ‰ (Fig. 10, panels A, B and C), although with a consistent discrimination bias. The correlation between VPD and  $\Delta_{canopy}$ , however, does not demonstrate causality. If that were the case, given that  $g_s$  is a function of VPD ( $h_s$  term in Eq. 5) and discrimination is a function of  $g_s$  (Eq. 10; Eq. 11), a similar relationship should have existed between  $g_s$  and  $\Delta_{canopy}$ . This, in fact, was not the case. Overall, the influence of  $g_s$  (responding to VPD) (R-value = -0.50) was secondary to  $A_n$  (R-value = -0.77) in driving changes in discrimination (Figure 10). The model suggested that the range in seasonal discrimination (intra-annual variation) was driven by the magnitude of  $A_n$  based on the inverse relationship between  $A_n$  and  $\Delta_{canopy}$ , (equation 11) illustrated by the separation between months of low photosynthesis (October, May) vs. high photosynthesis (June, July, August). During times of relatively low photosynthesis  $A_n$  also drove the inter-annual variation in  $\Delta_{canopy}$ . On the other hand,  $g_s$  (VPD) was most influential in driving the inter-annual variation of discrimination during the summer months only, judging by the directly proportional relationship during the months of June, July and August. Strictly speaking,  $g_s$  is a function of  $h_s$  (leaf relative humidity) and not atmospheric VPD in CLM. However, the two are closely related and the relationship between either variable (atmospheric VPD or simulated leaf humidity) to  $\Delta_{canopy}$  was similar (Figure S5).

The *limited nitrogen* formulation did not produce as wide a range in discrimination as compared to the observations (Figure 10, panels A, D and G). Part of this result was attributed

to the lack of response between  $A_n$  and  $\Delta_{\text{canopy}}$ . In this case, the discrimination did not decrease with increasing  $A_n$  because the signal was muted by the countering effect of  $f_{\text{dreg}}$ . The *limited nitrogen* formulation was, however, able to reproduce the same discrimination response to  $g_s$  as compared to the other model formulations. The tendency for the limited nitrogen model to simulate discrimination response to  $g_s$  and not to  $A_n$  may negatively impact its ability to simulate multi-decadal trends in discrimination. This may not be a major detriment to sites such as Niwot Ridge which have maintained a consistent level of carbon uptake during the last decade, and is likely more susceptible to environmental impact upon stomatal conductance. However, sites that have shown a significant increase in assimilation rate (e.g. Harvard Forest; (Keenan et al., 2013)) are less likely to be well represented by this model formulation.

Given the dependence of forest productivity at Niwot Ridge on snowmelt (Hu et al., 2010), it was surprising that the model simulated minimal soil moisture stress (Fig. 9, panel E) and therefore minimal discrimination response to soil moisture. However, this finding was consistent with Bowling et al., (2014), who did not find an isotopic response to soil moisture. In addition, lack of response to change in soil moisture may not be indicative of poor performance of the isotopic sub-model performance, but rather an effect of the hydrology sub-model (Duarte et al. (in prep)). However, a comparison of observed soil moisture at various depths at Niwot Ridge generally agrees with the CLM simulated soil moisture (not shown), suggesting the lack of model response to soil moisture was not from biases in the hydrology model.

## 4 Discussion

### 4.1 Discrimination formulations: implications for model development

The *limited* and *unlimited model* formulations tested in this study represented two approaches to account for nitrogen limitation within ecosystem models. The *limited nitrogen* formulation reduced photosynthesis, *after the main photosynthesis calculation*, so that the carbon allocated to growth was accommodated by available nitrogen. This *allocation downscaling* approach is common to a subset of models, for example, CLM (Thornton et al., 2007), DAYCENT (Parton et al., 2010) and ED2.1 (Medvigy et al., 2009). Another class of models limits photosynthesis based upon foliar nitrogen content and adjusts the photosynthetic capacity through nitrogen availability in the leaf through  $V_{\text{cmax}}$  (e.g. CABLE, GDAY, LPJ-GUESS, OCN, SDVGM, TECO, see Zaehle et al., 2014). These *foliar nitrogen* models are



1 similar to the *unlimited nitrogen* formulation of CLM because the scaling of photosynthesis  
2 was taken into account in the  $V_{cmax}$  scaling methodology (see discussion in section 2.1.2 and  
3 2.4), *prior to the photosynthesis calculation*. In general, there were no categorical differences  
4 in behavior between these two classes of models during CO<sub>2</sub> manipulation experiments held at  
5 Duke forest and ORNL (Zachle et al., 2014). However, CLM 4.0 was one of the few models in  
6 that study to consistently underestimate the NPP response to an increase of atmospheric CO<sub>2</sub>  
7 due to nitrogen limitation. This finding was attributed to a lower initial supply of nitrogen and  
8 too strong of a coupling between carbon and nitrogen that limited biomass production. Also  
9 within this experiment, it was found that models that had no or partial coupling (CLM 4.0,  
10 DAYCENT) between  $A_n$  and  $g_s$ , generally predicted lower than observed WUE response to  
11 increases in CO<sub>2</sub> (De Kauwe et al., 2013). Similar to CLM 4.0, the *limited nitrogen* formulation  
12 of CLM 4.5 in this manuscript is partially coupled (see Section 3.2.1). The unlimited nitrogen  
13 formulation of CLM 4.5, on the other hand, is fully coupled and similar to De Kauwe et al.,  
14 (2013) outperformed the partially coupled version of CLM.

15 The *unlimited nitrogen* formulation described in our study has similarities to a foliar  
16 nitrogen model, in that, the influence of nitrogen limitation is parameterized within  $V_{cmax}$ . A  
17 true foliar nitrogen model, however, couples a dynamic nitrogen cycle directly with the  
18 calculation of  $V_{cmax}$ . This capability was recently developed within CLM (Ghimire et al., 2016)  
19 and is scheduled to be included in the next CLM release. Future work should test its  
20 functionality.

21 The performance of the *unlimited nitrogen* formulation was nearly identical to the *no*  
22 *downregulation discrimination* formulation in terms of isotopic behavior despite the  
23 mechanistic differences. The *no downregulation discrimination* formulation included nitrogen  
24 limitation within the bulk carbon behavior but ignored the impact of  $f_{dreg}$  upon discrimination  
25 behavior. The relative high simulation skill with this formulation implied that the ‘potential’  
26 GPP linked to  $A_n$ , was a more effective predictor of discrimination behavior than the  
27 ‘downscaled’ GPP, which is linked to  $A_n * (1 - f_{dreg})$  (equation 11). There are several potential  
28 explanations for an unrealistically large value of  $f_{dreg}$ . First this could indicate that the  $V_{cmax}$   
29 parameter was too large, thereby requiring a large  $f_{dreg}$  to compensate. As noted in Section  
30 (3.1) the default temperate evergreen  $V_{cmax25}$  was  $\sim 62 \mu\text{mol m}^{-2} \text{s}^{-1}$ , much larger than what was  
31 found based on literature reviews (Monson et al., 2005; Tomaszewski and Sievering, 2007).  
32 We found to match the observed GPP we had to impose  $f_{dreg}$  that had the same effect as reducing

$V_{cmax}$  (Figure S2) to values of 51 and 34  $\mu\text{mol m}^{-2} \text{s}^{-1}$  for the *limited nitrogen* and *unlimited nitrogen* formulations respectively. Alternatively, it could be that there are physiological processes that are acting to reduce nitrogen limitation (e.g. nitrogen storage pools or transient carbon storage as non-structural carbohydrates), or that the current measurement techniques are underestimating GPP due to biases within the flux partitioning methods.

#### 4.2 Disequilibrium, possible explanations of mismatch

Carbon cycle models (e.g. Fung et al., 1997) indicate that the steady decrease of  $\delta_{\text{atm}}$  (Suess effect, Fig. 2) should lead to a positive disequilibrium between land surface processes ( $\delta^{13}\text{C}$  difference between GPP and ER, Eq. 14). This is because the  $\delta_{\text{GPP}}$  reflects the most recent ( $\delta^{13}\text{C}$  depleted) state of the atmosphere, whereas the  $\delta_{\text{ER}}$  reflects carbon (e.g. soil carbon) assimilated from an older ( $\delta^{13}\text{C}$  enriched) atmosphere. This positive disequilibrium pattern promoted by the Suess effect was consistent with all CLM formulations for this study with an annual average disequilibrium of 0.8 ‰. In contrast, a negative disequilibrium (-0.6 ‰) was identified at Niwot Ridge based upon observations (Bowling et al. 2014) as well as in other forests (Flanagan et al., 2012; Wehr and Saleska, 2015; Wingate et al., 2010). Bowling et al. (2014) hypothesized several reasons for this: 1) a strong seasonal stomatal response to atmospheric humidity, 2) decreased photosynthetic discrimination associated with  $\text{CO}_2$  fertilization, 3) decreased photosynthetic discrimination associated with multi-decadal warming and increased VPD, and 4) post-photosynthetic discrimination. We evaluated the first three hypotheses within the context of our CLM simulations.

The model results suggest a seasonal variation of discrimination that is a function of both VPD and  $A_n$ . The simulated seasonal range in discrimination (Figure 8; Figure S4) varied by approximately 2 ‰, and this range in seasonal discrimination could contribute to a negative disequilibrium provided specific timing of assimilation, assimilate storage and respiration not currently considered in the model. For example, if a significant portion of photosynthetic assimilation was stored during the spring with relatively high discrimination, and then respired during the summer, the net effect would deplete the  $\delta_{\text{ER}}$  and thereby promote negative disequilibrium during the summer months when discrimination is lower. Theoretically, this could be achieved by explicitly including carbohydrate storage pools within CLM. Isotopic tracer studies have shown assimilated carbon can exist for weeks to months within the vegetation and soil before it is finally respired (Epron et al., 2012; Hogberg et al., 2008).

1 Although carbon storage pools are included in CLM, their allocation is almost always  
2 instantaneous for evergreen systems and could not provide the isotopic effect described above.

3 The CO<sub>2</sub> fertilization effect tends to favor photosynthesis in plants and has been shown to  
4 simultaneously increase WUE and decrease stomatal conductance as inferred from  $\delta^{13}\text{C}$  in tree  
5 rings (Frank et al., 2015; Flanagan et al., 2012; Wingate et al., 2010). In general a decrease in  
6 stomatal conductance and increase in WUE is associated with a decrease in C3 discrimination  
7 (Farquhar et al., 1982), which opposes the disequilibrium trend imposed by the Suess effect.  
8 The model simulation agrees with both these trends in WUE and stomatal conductance, yet  
9 simulates an *increase* in discrimination (Figure 6; Figure 7), which reinforces the Suess effect  
10 pattern upon disequilibrium. Although this appears to be a mismatch between forest processes  
11 and model performance the model is operating within the limits of the discrimination  
12 parameterization (Eq. 17) in which the magnitude of photosynthetic discrimination is inversely  
13 proportional to the iWUE, but is also proportional to atmospheric CO<sub>2</sub> (see section 3.2.1).

14 A multi-decadal decrease in photosynthetic discrimination may also result from change in  
15 climate. Meteorological measurements at Niwot Ridge during the last several decades  
16 generally support conditions of higher VPD based upon a warming trend from an average  
17 annual temperature of 1.1 °C in the 1980's to 2.7 °C in the 2000's (Mitton and Ferrenberg,  
18 2012) and no overall trend in precipitation. It is possible that a multi-decadal trend in increasing  
19 VPD contributed to multi-decadal weakening in photosynthetic discrimination given the  
20 observed (Bowling et al., 2014) and modelled (Figure 10) correlation between  $\Delta_{\text{canopy}}$  and VPD.  
21 The model meteorology only included the years 1998-2013 and did not include the rapid  
22 warming after the 1980's. It is unclear whether, if the full period of warming were to be  
23 included in the simulation, the simulated discrimination response to VPD would be enough to  
24 counter the Suess effect and lead to negative disequilibrium. Still, there is evidence that the  
25 model is overestimating contemporary discrimination (Section 4.2) and the exclusion of the full  
26 multi-decadal shift in VPD could be a significant reason why.

27 Finally, post-photosynthetic discrimination processes are likely to impact the magnitude  
28 and sign of the isotopic disequilibrium (Bowling et al., 2008; Brüggemann et al., 2011) at  
29 multiple temporal scales. None of these isotopic processes are currently modelled within CLM  
30 4.5, so at present the model cannot be used to examine them.

## 5 Conclusions

This study provides a rigorous test of the representation of C isotope discrimination within the mechanistic terrestrial carbon model CLM. CLM was able to accurately simulate  $\delta^{13}\text{C}$  in leaf and stem biomass and the seasonal cycle in  $\Delta_{\text{canopy}}$ , but only when  $V_{\text{cmax}}$  was calibrated to account for nitrogen limitation *prior* to photosynthesis (*unlimited nitrogen* formulation).

Although the *unlimited nitrogen* formulation (fully-coupled carbon water cycle) was able to match observed  $\delta^{13}\text{C}$  of biomass and seasonal patterns in discrimination, it still overestimated the contemporary magnitude of discrimination (2006-2012). Future work should identify whether this overestimation was a result of parameterization (stomatal slope), exclusion of multi-decadal shifts in VPD, limitations in the representation of stomatal conductance (Ball-Berry model) or absence of the representation of mesophyll conductance.

The model attributed most of the range in seasonal discrimination to variation in net assimilation rate ( $A_n$ ) followed by variation in VPD, with little to no impact from soil moisture. The model suggested that  $A_n$  drove the seasonal range in discrimination (across-month variation) whereas VPD drove the inter-annual variation during the summer months. This finding suggests that to simulate multi-decadal trends in photosynthetic discrimination, response to assimilation rate and VPD must be well represented within the model.

The model simulated a positive disequilibrium that was driven by both the Suess effect, and increased photosynthetic discrimination from  $\text{CO}_2$  fertilization. It is possible that the negative disequilibrium that was inferred from observations (Bowling et al., 2014) was driven from the impacts of climate change and/or post-photosynthetic discrimination – not considered in this version of the model.

The model simulated a consistent increase in water-use efficiency as a response to  $\text{CO}_2$  fertilization and decrease in stomatal conductance. The model simulated an increase in WUE despite an increase in discrimination, however C3 plants typically express the opposite trends (increase in WUE, decrease in discrimination). Although CLM includes parameterization that promotes an increase in WUE with a decrease in discrimination, this trend was likely moderated by an increase in  $c_a$ .

Initial indications are that  $\delta^{13}\text{C}$  isotope data can bring additional constraint to model parameterization beyond what traditional flux tower measurements of carbon, water exchange, and biomass measurements. The isotope measurements suggested a stomatal conductance

1 value generally lower than what was consistent with the flux tower measurements.  
2 Unexpectedly, the isotopes also provided guidance upon model formulation related to nitrogen  
3 limitation. The success of our empirical approach to account for nutrient limitation within the  
4  $V_{max}$  parameterization, suggests that additional testing of foliar nitrogen models are  
5 worthwhile.

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7

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19

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1 Table 1. List of symbols used.

Symbol	Description	Unit or Unit Symbol
$\alpha$	Fractionation factor ( $R_a/R_{GPP}$ )	dimensionless
$\beta_t$	Soil water stress parameter (BTRAN)	dimensionless
$\Delta_{canopy}$	photosynthetic carbon isotope discrimination	‰
$\delta^{13}C$	$^{13}C/^{12}C$ isotope composition (relative to VPDB)	‰
$\delta_{atm}$	$\delta^{13}C$ of atmospheric $CO_2$	‰
$\delta_{ER}$	$\delta^{13}C$ of ecosystem respiration	‰
$\delta_{GPP}$	$\delta^{13}C$ of net photosynthetic assimilation	‰
$\Gamma^*$	$CO_2$ compensation point	Pa
$A_c$	Enzyme-limiting rate of photosynthetic assimilation	$\mu mol\ m^{-2}\ s^{-1}$
$A_j$	Light-limiting rate of photosynthetic assimilation	$\mu mol\ m^{-2}\ s^{-1}$
$A_p$	Product-limiting rate of photosynthetic assimilation	$\mu mol\ m^{-2}\ s^{-1}$
$A_n$	net photosynthetic assimilation	$\mu mol\ m^{-2}\ s^{-1}$
$Resp_d$	Leaf-level respiration	$\mu mol\ m^{-2}\ s^{-1}$
$a_{R25}$	Specific activity of Rubisco at 25°C	$\mu mol\ g^{-1}\ Rubisco\ s^{-1}$
$b$	Minimum stomatal conductance	$\mu mol\ m^{-2}\ s^{-1}$
$CF_{alloc}$	Actual carbon allocated to biomass (N-limited)	$gC\ m^{-2}\ s^{-1}$
$CF_{av,alloc}$	Maximum carbon available for allocation to biomass	$gC\ m^{-2}\ s^{-1}$
$CF_{GPPpot}$	Potential gross primary production (non N-limited)	$gC\ m^{-2}\ s^{-1}$
$c_a$	Atmospheric $CO_2$ partial pressure	Pa
$c_i$	Leaf intercellular $CO_2$ partial pressure	Pa
$c_i^*$	Leaf intracellular $CO_2$ partial pressure, (N-limited)	Pa
$c_s$	Leaf surface $CO_2$ partial pressure	Pa
$e_l$	Saturation vapor pressure	Pa
$e_s$	Water vapor pressure at leaf surface	Pa
$E_T$	Ecosystem Transpiration	$\mu mol\ m^{-2}\ s^{-1}$
ER	Ecosystem respiration	$\mu mol\ m^{-2}\ s^{-1}$
GPP	Gross primary productivity (photosynthesis)	$\mu mol\ m^{-2}\ s^{-1}$
$F_{LNR}$	Fraction of leaf nitrogen within Rubisco	$gN\ Rubisco\ g^{-1}\ N$
$F_{NR}$	Total Rubisco mass per nitrogen mass within Rubisco	$g\ Rubisco\ g^{-1}\ N\ Rubisco$
$f_{df}$	$V_{cmax}$ scaling factor	dimensionless
$f_{dreg}$	Nitrogen photosynthetic downregulation factor	dimensionless
$g_b$	Leaf boundary layer conductance	$\mu mol\ m^{-2}\ s^{-1}$
$g_s$	Leaf stomatal conductance	$\mu mol\ m^{-2}\ s^{-1}$
$h_s$	Leaf surface relative humidity	$Pa\ Pa^{-1}$
$K_c$	$CO_2$ Michaelis-Menten constant	Pa
$K_o$	$O_2$ Michaelis-Menten constant	Pa
LE	Latent heat flux	$W\ m^{-2}$
$m$	Stomatal slope (Ball Berry conductance model)	dimensionless
$Na$	Leaf nitrogen concentration	$gN\ m^{-2}\ leaf\ area$
NEE	Net ecosystem exchange	$\mu mol\ m^{-2}\ s^{-1}$
NPP	Net primary production	$\mu mol\ m^{-2}\ s^{-1}$
$o_i$	$O_2$ atmospheric partial pressure	Pa
PFT	Plant functional type	N/A
$P_{atm}$	Atmospheric pressure	Pa
$R_a$	Isotopic ratio of canopy air	$^{13}C/^{12}C$

$R_{GPP}$	Isotopic ratio of net photosynthetic assimilation	$^{13}C/^{12}C$
$R_{VPDB}$	Isotopic ratio of Vienna Pee Dee Belemnite standard	$^{13}C/^{12}C$
$r$	Fraction of roots (for $\beta_i$ )	dimensionless
$V_{cmax25}$	Maximum carboxylation rate at 25°C	$\mu\text{mol m}^{-2} \text{s}^{-1}$
$V_{cmax}$	Maximum carboxylation rate at leaf temperature	$\mu\text{mol m}^{-2} \text{s}^{-1}$
VPD	Vapor pressure deficit	Pa
$w$	Plant wilting factor (for $\beta_i$ )	dimensionless
$WUE$	Water use efficiency, ground area basis	$\mu\text{mol C mol H}_2\text{O}^{-1}$
$iWUE$	Intrinsic water use efficiency, leaf area basis	$\mu\text{mol C mol H}_2\text{O}^{-1}$

1 Table 2. CLM 4.5 model formulation description based upon timing of nitrogen limitation.  
2 Pre-photosynthetic and post-photosynthetic nitrogen limitation are achieved through  $V_{\text{cmax}25}$   
3 calibration (equation 17) and  $f_{dreg}$  (equation 8) respectively.

Formulation	Pre-Photosynthetic Nitrogen Limitation	Post-Photosynthetic Nitrogen Limitation	Impacts $c_i^*/c_a$ & discrimination	$A_n-g_s$ coupling
<i>Limited nitrogen (default)</i>	Yes (weak)	Yes, $f_{dreg} > 0$	Yes	partial
<i>Unlimited nitrogen</i>	Yes (strong)	No, $f_{dreg} = 0$	Yes	full
<i>No downregulation discrimination</i>	Yes (weak)	Yes, $f_{dreg} > 0$	No	partial



1 Table 3. CLM 4.5 key parameter values for all model formulations

Parameter	Description	Value	Units
<i>froot_leaf</i>	new fine root C per new leaf C	0.5	gC gC <sup>-1</sup>
<i>froot_cn</i>	fine root (C:N)	55	gC gN <sup>-1</sup>
<i>leaf_long</i>	leaf longevity	5	years
<i>leaf_cn</i>	leaf (C:N)	50	gC gN <sup>-1</sup>
<i>lflitcn</i>	leaf litter (C:N)	100	gC gN <sup>-1</sup>
<i>slatop</i>	specific leaf area (top canopy)	0.007	m <sup>2</sup> gC <sup>-1</sup>
<i>stem_leaf</i>	new stem C per new leaf C	2	gC gC <sup>-1</sup>
<i>mp</i>	stomatal slope	9	
<i>croot_stem</i>	coarse root: stem allocation	0.3	gC gC <sup>-1</sup>
<i>deadwood_cn</i>	dead wood (C:N)	500	gC gN <sup>-1</sup>
<i>livewood_cn</i>	live wood (C:N)	50	gC gN <sup>-1</sup>
<i>flnr</i>	fraction of leaf nitrogen within Rubisco enzyme	0.0509	gN gN <sup>-1</sup>
<i>decomp_depth_e_folding</i>	controls soil decomposition rate with depth	20	m

2

3

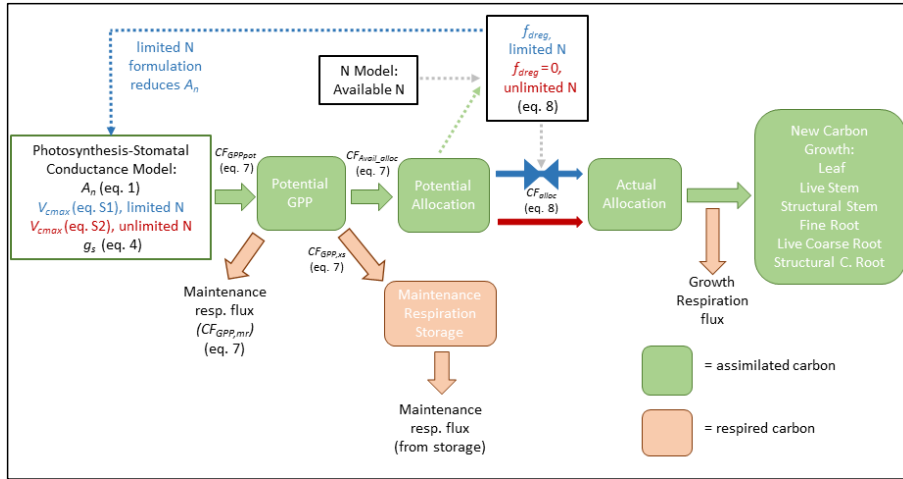


Figure 1: A simplified representation within CLM 4.5 of assimilation and allocation of carbon for conifer species. The colored boxes and solid arrows represent carbon pools and carbon fluxes respectively. The clear background boxes represent CLM sub-models. N-limitation is applied if the available N cannot meet the demand determined by the available carbon for allocation ( $CF_{avail,alloc}$ ) and the C:N biomass ratio. The blue and red text and arrows represent the *limited* and *unlimited* nitrogen formulations respectively. The *no-downregulation discrimination* formulation is exactly the same as the limited N formulation in this schematic.

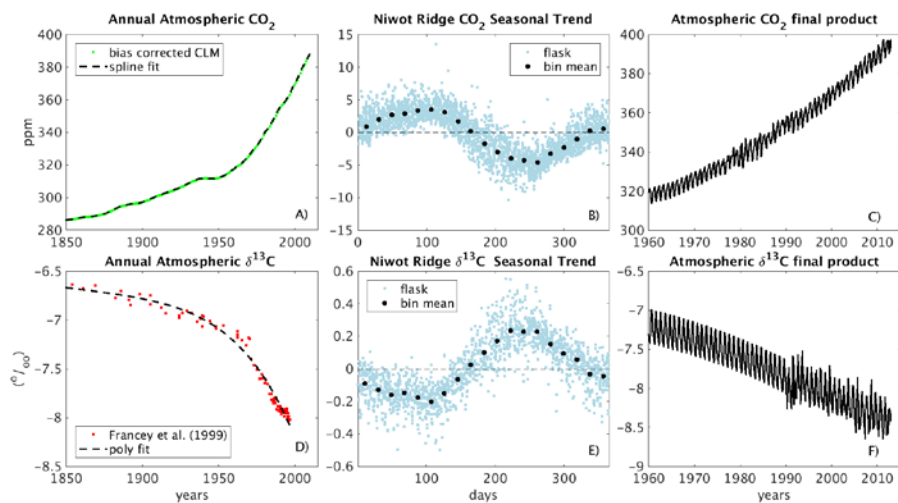
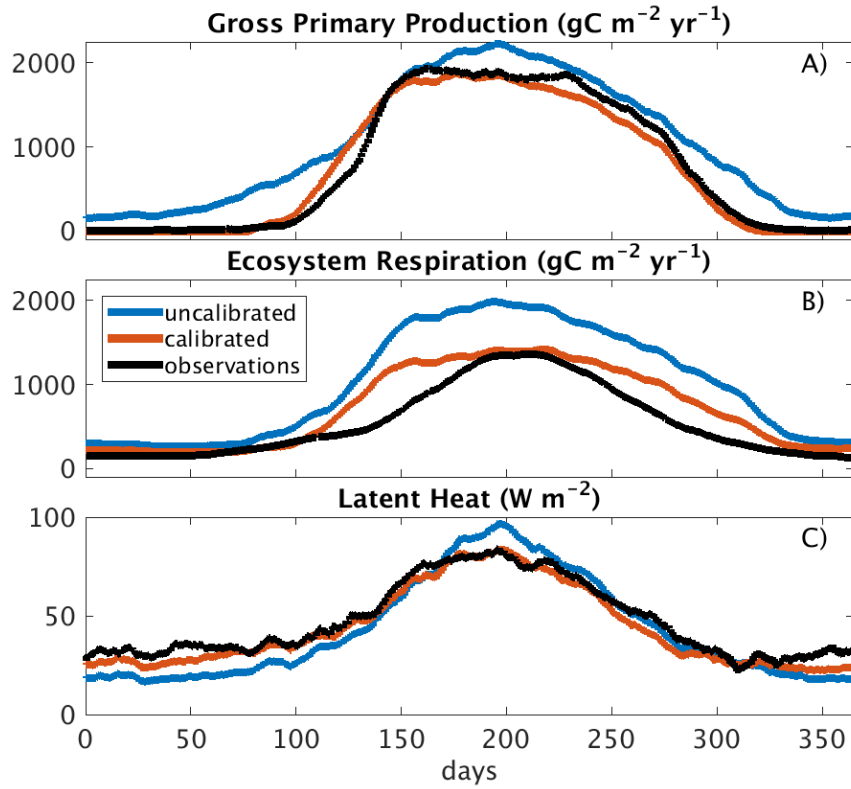
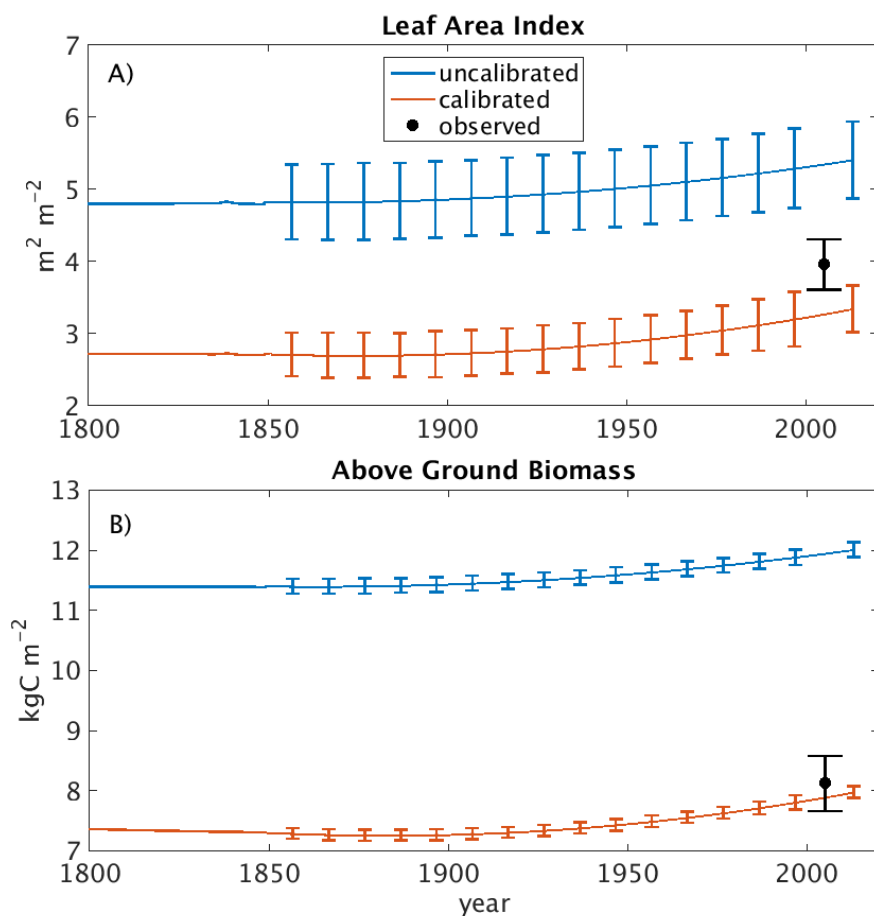


Figure 2. Niwot Ridge synthetic data product for atmospheric  $\text{CO}_2$  concentration ( $c_a$ ) (panels A, B and C) and  $\delta^{13}\text{C}$  of  $\text{CO}_2$  ( $\delta_{\text{atm}}$ ) (panels D, E and F). The final time series (panels C and F) were used as a boundary condition for CLM, and created by combining the annual trends reported by Francey et al. (1999) adjusted for Niwot Ridge (panels A and D) with the mean seasonal cycles measured at Niwot Ridge (panels B and E).

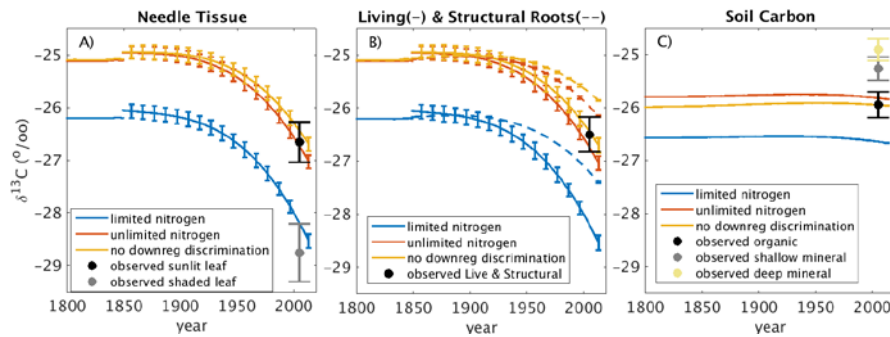


1  
2 Figure 3. Seasonal averages (1999-2013) of simulated and observed land-atmosphere fluxes  
3 for A) gross primary production (GPP) B) ecosystem respiration (ER) and C) latent heat (LE)  
4 for the *limited nitrogen* simulation. The ‘observations’ are taken from the Ameriflux L2  
5 processed eddy covariance flux tower data, partitioned into GPP and ER using the method of  
6 Reichstein et al. (2005). The *uncalibrated* simulation represents the CLM simulation without  
7  $V_{max}$  scaling and the *calibrated* simulation represents the CLM run using the  $V_{max}$  scaling  
8 approach.  
9

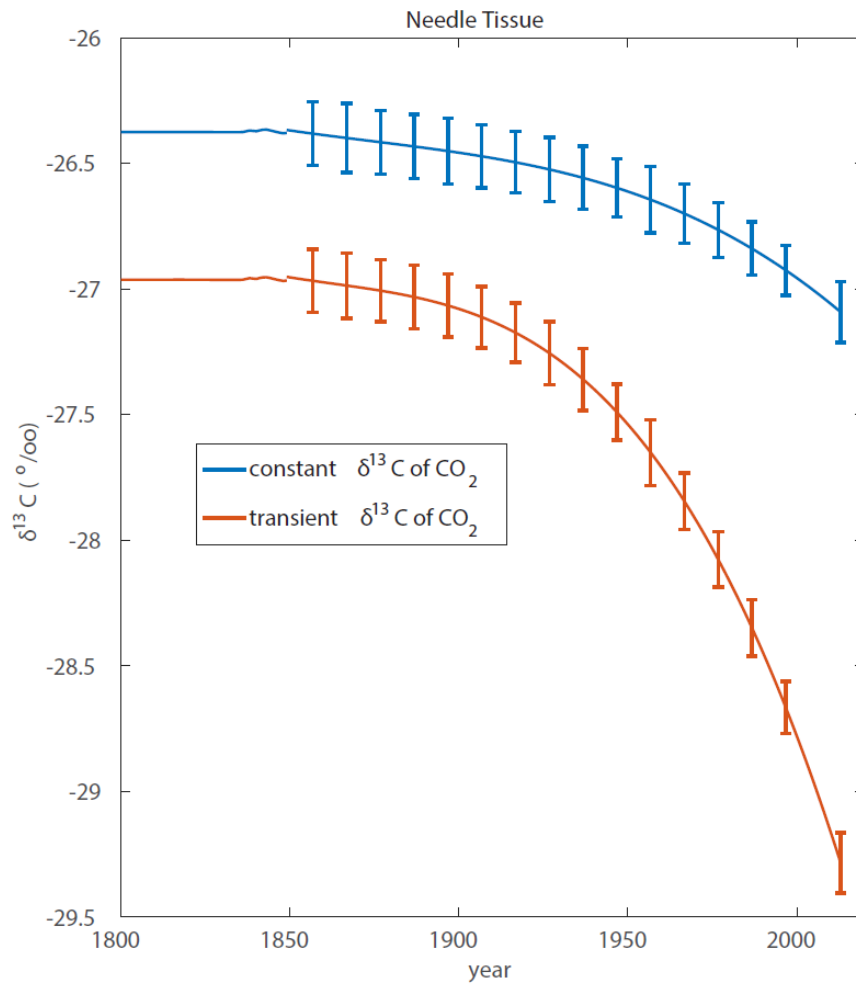


1  
2 Figure 4. Simulation of A) leaf area index and B) above ground biomass for both uncalibrated  
3 and calibrated ( $V_{\text{cmax}}$  downscaled, *limited nitrogen*) simulation. Observations are from  
4 Bradford et al. (2008) with uncertainty bars representing standard error. Uncertainty bars on  
5 simulated runs represent 95% confidence of biomass variation as a result of cycling the site  
6 level meteorology observations.

7



1  
2  
3 Figure 5. Simulation of  $\delta^{13}\text{C}$  of A) bulk needle tissue, B) bulk roots and C) bulk soil carbon.  
4 A description of model formulations are provided in Table (2). Uncertainty bars for simulations  
5 represent 95% confidence intervals of  $\delta^{13}\text{C}$  variation as a result of cycling the site level  
6 meteorology observations. The observed values are from Schaeffer et al. (2008) with  
7 uncertainty bars representing standard error. Solid lines and dashed lines in middle panel  
8 represent living roots and structural roots respectively.



1  
2 Figure 6. Simulation of  $\delta^{13}\text{C}$  of needle tissue using the *limited nitrogen* (default) CLM run. In  
3 the *constant  $\delta^{13}\text{C}$  of  $\text{CO}_2$  ( $\delta_{\text{atm}}$ )* simulation the model boundary condition was -6 ‰, whereas  
4 the *transient  $\delta_{\text{atm}}$*  simulation varied over time (Figure 2).

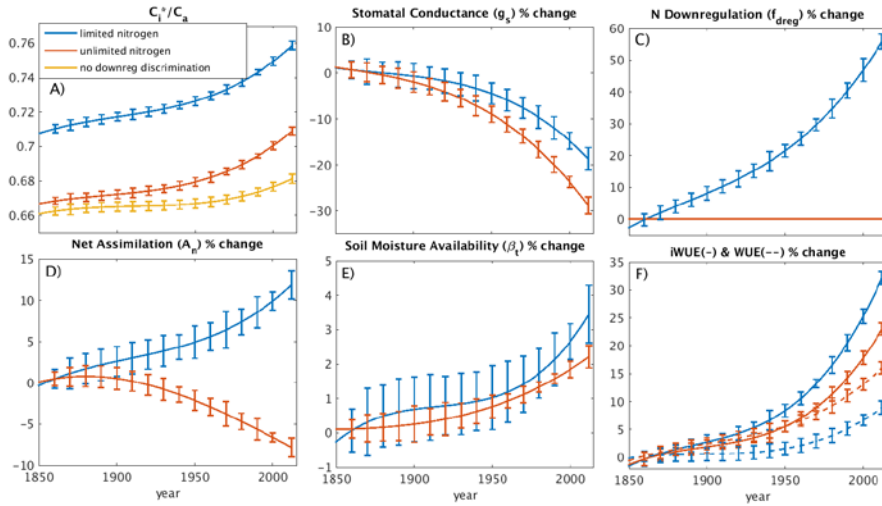


Figure 7. Diagnostic model variables that explain the discrimination trends (Figure 5) for the three model formulations as described in Table (2) for A)  $c_i^*/c_a$ , B)  $g_s$ , C)  $f_{dreg}$ , D)  $A_n$ , E)  $\beta_s$ , and F) the water use efficiency (WUE) and intrinsic water use efficiency (iWUE). Where the *no downregulation discrimination* simulation is not shown, it was identical to the *limited nitrogen* simulation. Uncertainty bars represent 95 % confidence intervals of diagnostic variable variation as a result of cycling the site level meteorology observations. The dashed lines represent WUE and the solid lines represent iWUE in panel F.



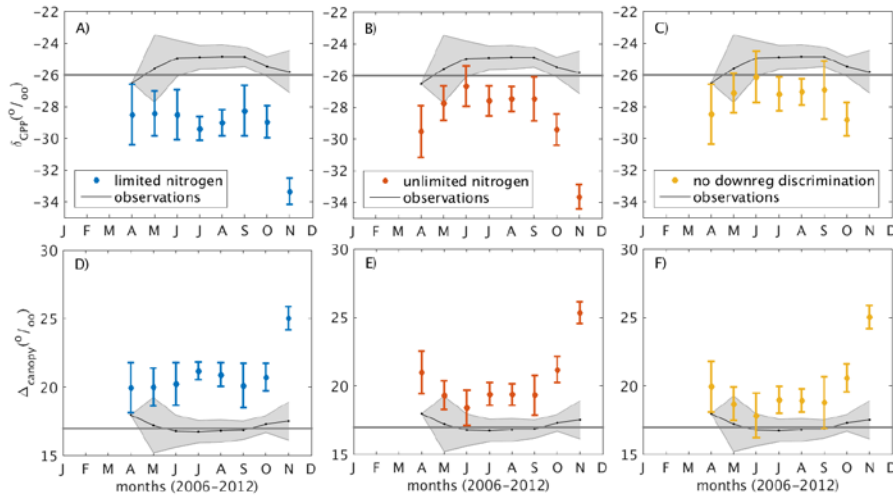


Figure 8. The seasonal pattern of photosynthetic discrimination as shown through  $\delta_{GPP}$  (panels A, B and C) and  $\Delta_{canopy}$  (panels D, E, and F). Uncertainty bars represent 95% confidence bounds of simulated monthly average values from 2006-2012. Gray-shaded observation bounds represent 95% confidence intervals of ‘observed’ monthly average values based upon isotopic mixing model using Reichstein et al. (2005) partitioning of net ecosystem exchange flux described by (Bowling et al. 2014). The horizontal lines at  $\delta^{13}C$  of -26 ‰ (panels A, B and C) and 17 ‰ (panels D, E and F) are included for reference.

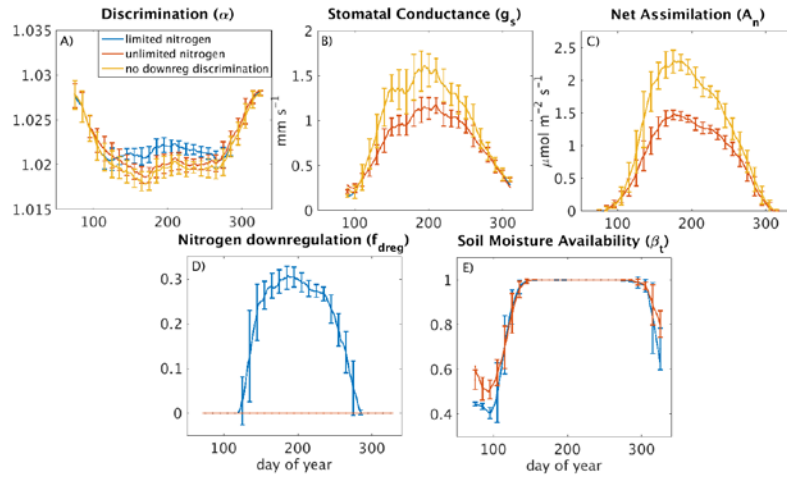


Figure 9. The seasonal pattern of discrimination (panel A) and diagnostic variables that explain the discrimination pattern in Figure (8). The individual tiles provide behavior from days 75-325 for A)  $\alpha$ , B)  $g_s$ , C)  $A_n$ , D)  $f_{dreg}$ , and E)  $\beta_s$ . Where the *no downregulation discrimination* model simulation is not shown, it is identical to the *limited nitrogen* simulation. Uncertainty bars represent 95 % confidence intervals of inter-annual variation from 2006-2012.

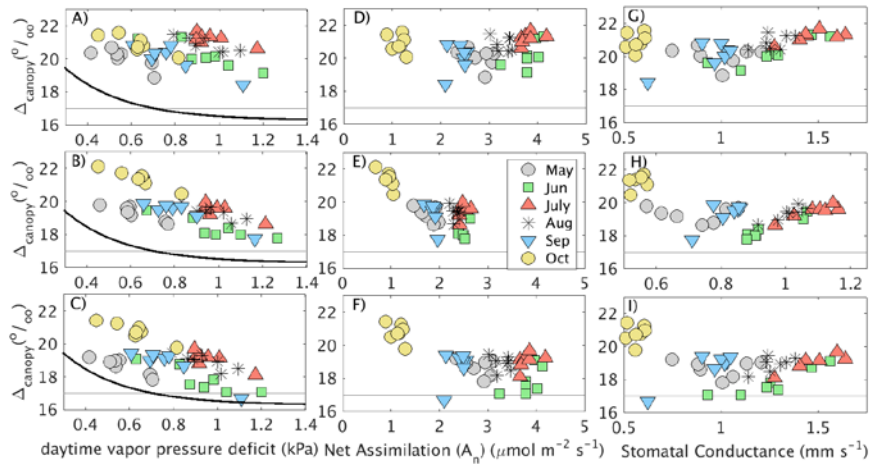


Figure 10. Relationship between monthly average photosynthetic discrimination and monthly average vapor pressure deficit (panels A, B and C),  $A_n$  (panels D, E and F) and  $g_s$  (G, H and I) from 2006-2012. The rows represent the *limited nitrogen* (panels A, D and G), *unlimited nitrogen* (panels B, E and H), and *no downregulation discrimination* (panels C, F and I) simulations. The black lines in panels A, B and C are based on an exponential fitted line from the observed relationship at Niwot Ridge (Bowling et al. 2014). The horizontal lines represent  $\delta^{13}\text{C}$  of 17 ‰ and are included for reference.