- **An observational constraint on stomatal function in forests:**
- 2 evaluating coupled carbon and water vapor exchange with
- 3 carbon isotopes in the Community Land Model (CLM 4.5)
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12 Abstract

13 Land surface models are useful tools to quantify contemporary and future climate impact on 14 terrestrial carbon cycle processes, provided they can be appropriately constrained and tested with observations. Stable carbon isotopes of CO₂ offer the potential to improve model 15 representation of the coupled carbon and water cycles because they are strongly influenced by 16 17 stomatal function. Recently, a representation of stable carbon isotope discrimination was 18 incorporated into the Community Land Model component of the Community Earth System 19 Model. Here, we tested the model's capability to simulate whole-forest isotope discrimination in a subalpine conifer forest at Niwot Ridge, Colorado, USA. We distinguished between 20 isotopic behavior in response to a decrease of δ^{13} C within atmospheric CO₂ (Suess effect) vs. 21 photosynthetic discrimination (Δ_{canopy}), by creating a site-customized atmospheric CO₂ and 22 δ^{13} C of CO₂ time series. We implemented a seasonally-varying V_{cmax} model calibration that 23 24 best matched site observations of net CO₂ carbon exchange, latent heat exchange and biomass. The model accurately simulated observed δ^{13} C of needle and stem tissue, but underestimated 25 26 the δ^{13} C of bulk soil carbon by 1-2 ‰. The model overestimated the multi-year (2006-2012) 27 average Δ_{canopy} relative to prior data-based estimates by 5-6 %. The amplitude of the average seasonal cycle of Δ_{canopy} (i.e. higher in spring/fall as compared to summer) was correctly 28 29 modeled but only when using pre-photosynthetic nitrogen limitation in contrast to the post-

photosynthetic nitrogen limitation used in the default version of the model. The model attributed 1 2 most of the seasonal variation in discrimination to the net assimilation rate (A_n) , whereas interannual variation in simulated Δ_{canopy} during the summer months was driven by stomatal 3 response to vapor pressure deficit. Soil moisture did not influence modeled Δ_{canopy} . The model 4 5 simulated a 10% increase in both photosynthetic discrimination and water use efficiency 6 (WUE) since 1850 as a result of CO₂ fertilization, forced by constant climate conditions. This 7 increasing trend in discrimination is counter to well-established relationships between 8 discrimination and WUE. The isotope observations used here to constrain CLM suggest 1) the 9 model overestimated stomatal conductance and 2) the default CLM approach to representing 10 nitrogen limitation (post-photosynthetic limitation) was not capable of reproducing observed 11 trends in discrimination. These findings demonstrate that isotope observations can provide 12 important information related to stomatal function driven by environmental stress from VPD 13 and nitrogen limitation. Future versions of CLM that incorporate carbon isotope discrimination 14 are likely to benefit from explicit inclusion of mesophyll conductance.

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17 **1** Introduction

18 The net uptake of carbon by the terrestrial biosphere currently mitigates the rate of atmospheric CO_2 rise and thus the rate of climate change. 19 Approximately 25% of 20 anthropogenic CO₂ emissions are absorbed by the global land surface (Le Quéré et al., 2015), 21 but it is unclear how projected changes in temperature and precipitation will influence the future 22 of this land carbon sink (Arora et al., 2013; Friedlingstein et al., 2006). A major source of uncertainty in climate model projections results from the disagreement in projected strength of 23 the land carbon sink (Arora et al., 2013). Thus, it is critical to reduce this uncertainty to improve 24 25 climate predictions, and to better inform mitigation strategies (Yohe et al., 2007).

An effective approach to reduce uncertainties in terrestrial carbon models is to constrain a broad range of processes using distinct and complementary observations. Traditionally, terrestrial carbon models have relied primarily upon observations of land-surface fluxes of carbon, water and energy derived from eddy-covariance flux towers to calibrate model parameters and evaluate model skill. Flux measurements best constrain processes that occur at diurnal and seasonal time scales (Braswell et al., 2005; Ricciuto et al., 2008). Traditional ecological metrics of carbon pools (e.g. leaf area index (LAI), biomass) are also commonly used to provide independent and complementary constraints upon ecosystem processes at
 longer time scales (Ricciuto et al., 2011; Richardson et al., 2010). However, neither flux nor
 carbon pool observations provide suitable constraints for the model formulation of plant
 stomatal function and the related link between the carbon and water cycles.

5 Stable carbon isotopes of CO_2 are influenced by stomatal activity in C3 plants (e.g. 6 evergreen trees, deciduous trees), and thus provide a valuable but under-utilized constraint on 7 terrestrial carbon models. Plants assimilate more of the lighter of the two major isotopes of atmospheric carbon (¹²C vs. ¹³C). This preference, termed photosynthetic discrimination 8 (Δ_{canopv}) , is primarily a function of two processes, CO₂ diffusion rate through the leaf boundary 9 layer and into the stomata, and the carboxylation of CO₂. The magnitude of Δ_{canopy} is controlled 10 by CO₂ supply (atmospheric CO₂ concentration, stomatal conductance) and demand 11 (photosynthetic rate; Flanagan et al., 2012). In general, environmental conditions favorable to 12 13 plant productivity result in higher Δ_{canopy} during carbon assimilation compared to unfavorable 14 conditions. Plants respond to unfavorable conditions by closing stomata and reducing the 15 stomatal conductance which reduces Δ_{canopy} . Most relevant here, Δ_{canopy} responds to atmospheric moisture deficit (Andrews et al., 2012; Wingate et al., 2010), soil water content 16 17 (McDowell et al., 2010), precipitation (Roden and Ehleringer, 2007) and nutrient availability. After carbon is assimilated, additional post-photosynthetic isotopic changes occur (Bowling et 18 19 al., 2008; Brüggemann et al., 2011), but these impose a small influence on land-atmosphere 20 isotopic exchange relative to photosynthetic discrimination.

21 The Niwot Ridge Ameriflux site, located in a sub-alpine conifer forest in the Rocky 22 Mountains of Colorado, U.S.A., has a long legacy of yielding valuable datasets to test carbon 23 and water functionality of land surface models using stable isotopes. Niwot Ridge has a 17-24 year record of eddy covariance fluxes of carbon, water, and energy, as well as environmental data (Hu et al., 2010; Monson et al., 2002) and a 10-year record of δ^{13} C of CO₂ in forest air 25 26 (Schaeffer et al., 2008). From a carbon balance perspective, Niwot Ridge is representative of subalpine forests in Western North America that, in general, act as a carbon sink to the 27 28 atmosphere (Desai et al., 2011). Western forests make up a significant portion of the carbon 29 sink in the United States (Schimel et al., 2002), yet this sink is projected to weaken with projected changes in temperature and precipitation (Boisvenue and Running, 2010). 30

The Community Land Model (CLM), the land sub-component of the Community Earth
 System Model (CESM) has a comprehensive representation of biogeochemical cycling (Oleson

et al., 2013) that can be applied across a range of temporal (hours to centuries) and spatial (site 1 2 to global) scales. A mechanistic representation of photosynthetic discrimination based upon 3 diffusion and enzymatic fractionation (Farguhar et al., 1989) was included in the latest release 4 of CLM 4.5 (Oleson et al., 2013), and is similar to the formulation implemented in other land 5 surface models (Flanagan et al., 2012; Scholze et al., 2003; Wingate et al., 2010; van der Velde et al., 2013). An early version of CLM simulated carbon (but not carbon isotope) dynamics at 6 7 Niwot Ridge with reasonable skill (Thornton et al., 2002). To date, we are not aware of any 8 CLM-based studies that have used CO₂ isotopes at natural abundance to quantify the accuracy 9 of the photosynthetic discrimination sub-model, or to evaluate the utility of CO₂ isotopes to 10 constrain carbon and water cycle coupling.

Here, we evaluate the performance of the ${}^{13}C/{}^{12}C$ isotope discrimination sub-model 11 within CLM 4.5 against a range of isotopic observations at Niwot Ridge, to examine what new 12 13 insights an isotope-enabled model can bring upon ecosystem function. Specifically, we test 14 whether CLM simulates the expected isotopic response to environmental drivers of CO₂ 15 fertilization, soil moisture and atmospheric vapor pressure deficit (VPD). A previous analysis at Niwot Ridge showed a seasonal correlation between vapor pressure deficit (VPD) and 16 photosynthetic discrimination (Bowling et al., 2014) suggesting that leaf stomata are 17 responding to changes in VPD, and influencing discrimination. We use CLM to test whether 18 VPD is the primary environmental driver of isotopic discrimination, as compared to soil 19 moisture and net assimilation rate. Next we determine whether including site-specific δ^{13} C of 20 21 atmospheric CO₂ within the model simulation combined with simulated long term (multi-22 decadal to century) photosynthetic discrimination and simulated carbon pool turnover, accurately reproduces the measured δ^{13} C in leaf tissue, roots and soil carbon. We then use CLM 23 24 to determine if the increase in atmospheric CO₂ since 1850 has led to an increase in water use 25 efficiency (WUE), and whether net assimilation or stomatal conductance is the primary driver 26 of such a change. Finally, we ask what distinct insights site level isotope observations bring in 27 terms of both model parameterization (i.e. stomatal conductance) and model structure as 28 compared to the traditional observations (e.g. carbon fluxes, biomass).

29 2 Methods

We focus the description of CLM 4.5 (Section 2.1) upon photosynthesis, and its linkage to nitrogen, soil moisture and stomatal conductance (Section 2.1.1). Next we describe the model representation of carbon isotope discrimination by photosynthesis (Section 2.1.2). Because preliminary simulations demonstrated that model results were strongly influenced by nitrogen limitation, we used three separate nitrogen formulations (described in Section 2.1.2) to better diagnose model performance. Next, to provide context for subsequent descriptions of site-specific model adjustments we describe the field site, Niwot Ridge, including the site level observations (Section 2.2) used to constrain and test model the model.

6 Patterns in plant growth and δ^{13} C of biomass are strongly influenced by atmospheric CO₂ 7 and δ^{13} C of atmospheric CO₂ (δ_{atm}). Therefore we designed a site-specific synthetic 8 atmospheric CO₂ product (Section 2.3.1) and δ_{atm} product (Section 2.3.2) for these simulations. 9 The model setup and initialization procedure, intended to bring the system into steady state, is 10 described in Section (2.3.3). This is followed by an explanation of the model calibration 11 procedure that provided a realistic simulation of carbon and water fluxes (Section 2.4).

12 **2.1 Community Land Model, Version 4.5**

We used the Community Land Model, CLM 4.5 (Oleson et al., 2013), which is the land 13 14 component of the Community Earth System Model (CESM) version 1.2 15 (https://www2.cesm.ucar.edu/models/current). Details regarding the Community Land Model can be found in Mao et al., (2016) and Oleson et al., (2013). 16 Here, we emphasize the 17 mechanistic formulation that controls photosynthetic discrimination (Δ_{canopy}) and factors that 18 influence Δ_{canopy} including photosynthesis, stomatal conductance, water stress and nitrogen 19 limitation. A list of symbols is provided in Table (1).

20 2.1.1 Net Photosynthetic Assimilation

21 The leaf-level net carbon assimilation of photosynthesis, A_n is based on Farquhar et al., 22 (1980) as,

23
$$A_n = \min(A_c, A_j, A_p) - Resp_d,$$
(1)

where A_c , A_j and A_p are the enzyme (Rubisco)-limited, light-limited, and product-limited rates of carboxylation respectively, and $Resp_d$ the leaf-level respiration. The enzyme limited rate is defined as

27
$$A_{c} = \frac{V_{cmax}(c_{i} - \Gamma_{*})}{c_{i} + K_{c}(1 + \frac{o_{i}}{K_{o}})},$$
(2)

1 where c_i is the intercellular leaf partial pressure of CO₂, $o_i = 0.209 P_{atm}$, P_{atm} is atmospheric 2 pressure, and K_c , K_o and Γ_* are constants. The maximum rate of carboxylation at 25°C, V_{cmax25} ,

3 is defined as

$$4 \quad V_{cmax25} = N_a \ F_{LNR} \ F_{NR} \ a_{R25}, \tag{3}$$

5 where N_a is the nitrogen concentration per leaf area, F_{LNR} the fraction of leaf nitrogen within 6 the Rubisco enzyme, F_{NR} the ratio of total Rubisco molecular mass to nitrogen mass within 7 Rubisco, and a_{R25} is the specific activity of Rubisco at 25°C. The V_{cmax25} is adjusted for leaf 8 temperature to provide V_{cmax} in Eq. 2, used in the final photosynthetic calculation.

9 The carbon and water balance are linked through c_i by the stomatal conductance to CO₂, 10 g_s , following the Ball-Berry model (Ball, 1987) as defined by Collatz et al., (1991),

11
$$g_s = m \frac{A_n}{c_s/P_{atm}} h_s + b\beta_t,$$
(4)

where *m* is the stomatal slope, c_s the partial pressure of CO₂ at the leaf surface, h_s the relative humidity at the leaf surface and *b* the minimum stomatal conductance when the leaf stomata are closed. The variable β_t represents the level of soil moisture availability, which influences stomatal conductance directly, but also indirectly through A_n by multiplying V_{cmax} by β_t (Sellers et al., 1996). CLM calculates β_t as a factor (0-1, high to low stress) by combining soil moisture, the rooting depth profile, and a plant-dependent response to soil water stress as

$$18 \qquad \beta_t = \sum_i w_i r_i, \tag{5}$$

where w_i is a plant wilting factor for soil layer *i* and r_i is the fraction of roots in layer *i*. The plant wilting factor is scaled according to soil moisture and water potential, depending on plant functional type (PFT). Soil moisture is predicted based upon prescribed precipitation and vertical soil moisture dynamics (Zeng and Decker, 2009). The root fraction in each soil layer depends upon a vertical exponential profile controlled by PFT dependent root distribution parameters adopted from Zeng (2001).

The version of CLM used here has a 2-layer (shaded, sunlit) representation of the vegetation. Photosynthesis and stomatal conductance are calculated separately for the shaded and sunlit portion and the total canopy photosynthesis is the potential gross primary productivity (GPP), CF_{GPPpot} :

29
$$CF_{GPPpot} = \left[(A_n + Resp_d)_{sunlit} (LAI)_{sunlit} + (A_n + Resp_d)_{shaded} (LAI)_{shaded} \right] * 12.011^{-6}, \tag{6}$$

1 where *LAI* is the leaf area index and 12.011^{-6} is a unit conversion factor. The total carbon 2 available for new growth allocation (*CF_{avail alloc}*) is defined as

3
$$CF_{avail_alloc} = CF_{GPP,mr} - CF_{GPP,xs},$$
 (7)

where the maintenance respiration is derived either from recently assimilated photosynthetic carbon $(CF_{GPP,mr})$, or, if photosynthesis is low or zero (e.g. night), the maintenance respiration is drawn from a carbon storage pool $(CF_{GPP,xs})$. In contrast, CF_{alloc} , is the actual carbon allocated to growth calculated from the available nitrogen and fixed C:N ratios for new growth (e.g. stem, roots, leaves). The downregulation of photosynthesis from nitrogen limitation, f_{dreg} , is given by

$$10 f_{dreg} = \frac{CF_{avail_alloc} - CF_{alloc}}{CF_{GPPpot}}.$$
(8)

11 The actual, nitrogen-limited GPP is defined as:

$$12 \quad GPP = CF_{GPPpot}(1 - f_{dreg}) \tag{9}$$

13

14 2.1.2 Photosynthetic Carbon Isotope Discrimination

15 The canopy-level fractionation factor α is defined as the ratio of ${}^{13}C/{}^{12}C$ within 16 atmospheric CO₂ (R_a) and the products of photosynthesis (R_{GPP}) as $\alpha = \frac{R_a}{R_{GPP}}$. The preference 17 of C3 vegetation to assimilate the lighter CO₂ molecule during photosynthesis is simulated in 18 CLM with two steps: diffusion of CO₂ across the leaf boundary layer and into the stomata, 19 followed by enzymatic fixation to give the leaf-level fractionation factor:

20
$$\alpha = 1 + \frac{4.4 + 22.6 \frac{c_i^*}{c_a}}{1000}$$
 (10)

where c_i^* and c_a are the intracellular and atmospheric CO₂ partial pressure respectively. The numbers 4.4 and 22.6 represent the diffusional and enzymatic contributions to isotopic discrimination respectively. The variable c_i^* (known in CLM as the 'revised intracellular CO2 partial pressure') is marked with an asterisk to indicate the inclusion of nitrogen downregulation as defined as,

26
$$c_i^* = c_a - A_n \left(1 - f_{dreg}\right) P_{atm} \frac{(1.4g_s) + (1.6g_b)}{g_b g_s}$$
 (11)

1 where g_b is the leaf boundary layer conductance. The inclusion of the nitrogen downregulation 2 factor f_{dreg} in the above expression reflects the two-stage process in which the potential 3 photosynthesis (Eq. 6) and the actual photosynthesis (Eq. 9) are calculated within CLM and 4 prevents a mismatch between the actual photosynthesis and the intracellular CO₂.

5 The sensitivity of preliminary model results to nitrogen limitation led us to test three 6 distinct discrimination formulations (Figure 1; Table 2). The *limited nitrogen* formulation was 7 based on the default version of CLM 4.5 and included both nitrogen limitation and the nitrogen 8 downregulation factor within the calculation of c_i^* as given in equation (11). The second, 9 unlimited nitrogen formulation also follows equation (11), however, the vegetation is allowed unlimited access to nitrogen ($CF_{GPPpot} = GPP$, $f_{dreg} = 0$). Finally, in the no downregulation 10 11 discrimination formulation, we included nitrogen limitation, but removed the downregulation 12 factor f_{drea} within the isotopic discrimination equation (11).

13 In the *unlimited nitrogen* formulation, we use a different modifier on V_{cmax25} (Figure 1; 14 described in section 2.4 and Fig. S1, S2) in the calibrated runs to give similar carbon flux, water 15 flux and biomass as in the other two formulations, such that all three formulations have fluxes and biomass that are similar to what is observed at the site, and which presumably reflect 16 17 nitrogen limitation. Thus the distinction between these three formulations can be viewed 18 entirely as when nitrogen limitation is imposed in relation to photosynthesis: (1) after 19 photosynthesis via a downregulation between potential and actual GPP (equation 9) that feeds back on the c_{i*}/c_a used for isotopic discrimination but not on g_s or A_n in the limited nitrogen 20 formulation; (2) before photosynthesis via V_{cmax}, which limits photosynthetic capacity affecting 21 both c_{i*}/c_a , g_s and A_n in the unlimited nitrogen formulation; and (3) after photosynthesis with 22 23 no effect on either the c_i*/c_a for isotopic discrimination or g_s or A_n in the no downregulation 24 discrimination formulation. The downscaled portion of the carbon during nitrogen limitation $(CF_{GPPpot} - GPP)$ is removed from the system and does not appear as a respired flux (Figure 25 In summary, the limited nitrogen (post-photosynthetic) formulation adjusts the 26 1). 27 photosynthetic rate by explicitly tracking N availability, whereas the unlimited nitrogen (pre*photosynthetic*) formulation takes into account any N-limitation through the V_{cmax} 28 29 parameterization.

31
$$\delta^{13}C_x = \left(\frac{R_x}{R_{VPDB}} - 1\right) \times 1000,$$
 (12)

1 where R_x is the isotopic ratio of the sample of interest, and R_{VPDB} is the isotopic ratio of the 2 Vienna Pee Dee Belemnite standard. The delta notation is dimensionless but expressed in parts 3 per thousand (‰) where a positive (negative) value refers to a sample that is enriched (depleted) 4 in ${}^{13}C/{}^{12}C$ relative to the standard. Because this is the only carbon isotope ratio we are 5 concerned with in this paper, the '13' superscript is omitted for brevity in subsequent definitions 6 using the delta notation. The canopy-integrated photosynthetic discrimination, Δ_{canopy} , is 7 defined as the difference between the $\delta^{13}C$ of the atmospheric and assimilated carbon,

8
$$\Delta_{canopy} = \delta_{atm} - \delta_{GPP}$$
 (13)

9 The difference between δ^{13} C of the total ecosystem respiration (ER) and GPP fluxes, called the 10 isotope disequilibrium (Bowling et al., 2014), is defined as,

11
$$disequilibrium = \delta_{ER} - \delta_{GPP.}$$
 (14)

12 The ecosystem-level water use efficiency (WUE) is defined as actual carbon assimilated 13 (*GPP*) per unit water transpired (E_T) per unit land surface area,

$$14 \quad WUE = \frac{GPP}{E_T}.$$
(15)

15 The intrinsic water use efficiency (*iWUE*) from leaf-level physiological ecology is defined as,

$$16 \qquad iWUE = \frac{A_n}{g_s} , \qquad (16)$$

17 where A is the net carbon assimilated per unit leaf area and g_s is the stomatal conductance. 18 CLM calculates g_s (Equation 4) for shaded and sunlit portions of the canopy separately, 19 therefore an overall conductance was calculated by weighting the conductance by sunlit and 20 shaded leaf areas.

21 **2.2** Niwot Ridge and site-level observations

Site-level observations and modeling were focused on the Niwot Ridge Ameriflux
tower (US-NR1), a sub-alpine conifer forest located in the Rocky Mountains of Colorado,
U.S.A. The forest is approximately 110 years old and consists of lodgepole pine (*Pinus contorta*), Engelmann spruce (*Picea engelmanni*), and subalpine fir (*Abies lasiocarpa*). The
site is located at an elevation of 3050 m above sea level, with mean annual temperature of
1.5°C and precipitation of 800 mm, in which approximately 60% is snow. More site details

are available elsewhere (Hu et al., 2010; Monson et al., 2002). Flux and meteorological data
 were obtained from the Ameriflux archive (http://ameriflux.lbl.gov/).

3 Net carbon exchange (NEE) observations were derived from flux tower measurements based on the eddy covariance method and were partitioned into component fluxes of GPP and 4 ER according to two separate methods described by Reichstein et al., (2005) and Lasslop et al., 5 (2010) using an online tool provided by the Max Planck Institute for Biogeochemistry 6 7 (http://www.bgc-jena.mpg.de/~MDIwork/eddyproc/). Seasonal patterns in δ_{GPP} and δ_{ER} were 8 derived from measurements as described by Bowling et al., (2014). Observations of δ^{13} C of 9 biomass (Schaeffer et al., 2008) and carbon stocks (Bradford et al., 2008; Scott-Denton et al., 2003) were compared to model simulations. 10 Schaeffer et al., (2008) reported soil, leaf and 11 root observations specific to each conifer species, however, the observed mean and standard error for all species were used for comparison because CLM treated all conifer species as a 12 single PFT. 13

14 **2.3** Atmospheric CO₂, isotope forcing and initial vegetation state

15 2.3.1 Site-specific atmospheric CO₂ concentration time series

16 Global average atmospheric CO₂ concentrations increased roughly 40% from 1850 to 17 2013 (from 280 to 395 ppm). The standard version of CLM 4.5 includes an annually and 18 globally averaged time series of this CO₂ increase, however, this does not capture the observed 19 seasonal cycle of ~10 ppm at Niwot Ridge (Trolier et al., 1996). Therefore we created a site-20 specific atmospheric CO₂ time series (Figure 2) to provide a seasonally realistic atmosphere at Niwot Ridge. From 1968-2013 the CO₂ time series was fit to flask observations (Dlugokencky 21 22 et al., 2015) from Niwot Ridge. Prior to 1968, the CO₂ time series was created by combining 23 the average multi-year seasonal cycle based on the Niwot Ridge flask data to the annual CO₂ 24 product provided by CLM. More details are located in the supplemental material.

25

26 2.3.2 Customized δ^{13} C atmospheric CO₂ time series

As atmospheric CO₂ has increased, the δ^{13} C of atmospheric CO₂ (δ_{atm}) has become more depleted (Francey et al., 1999), and this change continues at Niwot Ridge at -0.25 ‰ per decade (Bowling et al., 2014). The δ_{atm} also varies seasonally, and depends on latitude (Trolier

et al., 1996). However, CLM 4.5 as released assigned a constant δ^{13} C of -6 ‰. We therefore 1 2 created a synthetic time series of δ_{atm} from 1850-2013 (Figure 2). From 1990-2013 the time 3 series was fit to the flask observations (White et al., 2015) as described in Section 2.3.1. Prior to 1990 the inter-annual variation within the δ_{atm} time series was fit to the ice core data from 4 5 Law Dome (Francey et al., 1999; see also Rubino et al., 2013). This annual data product was 6 then combined with the average seasonal cycle at Niwot Ridge as determined by the flask 7 observations to create the synthetic product from 1850-1990. More details are located in the 8 supplemental material.

9

10 2.3.3 Model Initialization

We performed an initialization to transition the model from near bare-ground conditions 11 12 to present day carbon stocks and LAI that allowed for proper evaluation of isotopic performance. This was implemented in 4 stages: 1) accelerated decomposition (1000 model 13 14 years) 2) normal decomposition (1000 model years) 3) parameter calibration (1000 model years) and 4) transient simulation period (1850-2013). The first two stages were pre-set options 15 16 within CLM with the first stage used to accelerate the equilibration of the soil carbon pools, 17 which require a long period to reach steady state (Thornton and Rosenbloom, 2005). The 18 parameter calibration stage was not a pre-set option but designed specifically for our analysis. 19 For this we introduced a seasonally varying V_{cmax} that scaled the simulated GPP and ecosystem 20 respiration fluxes to present day observations (Section 2.4). In the transient phase, we 21 introduced time-varying atmospheric conditions from 1850-2013 including nitrogen deposition (CLM provided), atmospheric CO₂, and δ_{atm} (site-specific as described above). Environmental 22 23 conditions of temperature, precipitation, relative humidity, radiation, and wind speed were 24 taken from the Niwot Ridge flux tower observations from 1998-2013 and then cycled 25 continuously for the entirety of the initialization process. We used a scripting framework (PTCLM) that automated much of the workflow required to implement several of these stages 26 in a site level simulation (Mao et al., 2016; Oleson et al., 2013). 27

28

1 **2.4** Specific model details and model calibration

2 This version of CLM included a fully prognostic representation of carbon and nitrogen 3 within its vegetation, litter and soil biogeochemistry. We used the Century model representation 4 for soil (3 litter and 3 soil organic matter pools) with 15 vertically resolved soil layers (Parton 5 et al. 1987). Nitrification and prognostic fire were turned off. Our initial simulations used 6 prognostic fire, but we found that simulated fire was overactive leading to low simulated 7 biomass compared to observations. Although Niwot Ridge has been subject to disturbance from 8 fire and harvest in the past, ultimately our final simulations did not include either fire or harvest 9 disturbance because the last disturbance occurred over 110 years ago (early 20th century 10 logging; Monson et al., 2005).

11 Ecosystem parameter values (Table 3) used here were based upon the temperate 12 evergreen needleleaf PFT within CLM. These values were based upon observations reported by White et al., (2000) intended for a wide range of temperate evergreen forests, and by 13 14 Thornton et al., (2002) for Niwot Ridge. For this analysis two site-specific parameter changes 15 were made. First, the e-folding soil decomposition parameter was increased from 5 to 20 16 meters. This parameter is a length-scale for attenuation of decomposition rate for the resolved 17 soil depth from 0 to 5 meters where an increased value effectively increases decomposition at 18 depth, thus reducing total soil carbon and more closely matching observations. Second, we 19 performed an empirical photosynthesis scaling (equation 17, below) that reduced the simulated 20 photosynthetic flux, as guided by eddy covariance observations (Figure 3; Figure S1). 21 Consequently, all downstream carbon pools and fluxes including ecosystem respiration, 22 aboveground biomass, and leaf area index which provided a better match to present day 23 observations. This approach also removed a systematic overestimation of winter 24 photosynthesis. The model simulations without the photosynthetic scaling are referred to within 25 the text and figures as the *uncalibrated* model, whereas model simulations that include the photosynthetic scaling are referred to as the *calibrated* model. We modified CLM for this 26 scaling approach by reducing V_{cmax} at 25° C, 27

28
$$V_{cmax25} = N_a F_{LNR} F_{NR} a_{R25} f_{df},$$
 (17)

where f_{df} is the photosynthetic scaling factor, and all other parameters are identical to equation (3). These parameters were constant for the entirety of the simulations except for f_{df} , an empirically derived time dependent parameter ranging from 0-1. The value was set to zero to force photosynthesis to zero between November 13th and March 23rd, consistent with flux 1 tower observations where outside of this range GPP > 0 was never observed. During the 2 growing season period (GPP>0) within days of year 83-316, f_{df} was calculated as

3
$$f_{df} = \frac{observed GPP(day of year)}{simulated GPP(day of year)}, 82 < day of year < 317$$
 (18)

where the *observed GPP* was the daily average calculated from the partitioned flux tower observations (Reichstein et al., 2005) from 2006-2013, and the *simulated GPP* was the daily average of the unscaled value during the same time. A polynomial was fit to equation (18) that represented f_{df} for 1) both the *limited nitrogen* and *no downregulation discrimination formulations* and 2) the *unlimited nitrogen* formulation (Figure S2). Note that CLM already includes a *daylength factor* that also adjusts the magnitude of V_{cmax} according to time of year, however, that default parameterization alone was not sufficient to match the observations.

11

12

13 3 Results & Discussion

14 This section is organized into four parts. First the calibrated model performance is 15 evaluated against observed bulk carbon pool and bulk carbon flux behavior (Section 3.1.1), and against the observed δ^{13} C within carbon pools (Section 3.1.2). Second, the simulated 16 17 photosynthetic discrimination is evaluated for multi-decadal trends (Section 3.2.1), magnitude 18 (Section 3.2.2) and seasonal patterns (Section 3.2.3), including the environmental factors that 19 were most responsible for driving the seasonal discrimination (Section 3.2.4). Third, we discuss 20 how isotope observations can be used to guide model development related to nitrogen limitation 21 (Section 3.3). Finally, we evaluate the capability of the model to reproduce the magnitude and 22 trends of disequilibrium (Section 3.4).

23 **3.1 Calibrated model performance**

24 3.1.1 Fluxes & carbon pools

The CLM model was successful at simulating GPP, ER, and latent heat fluxes (Fig. 3), leaf area index (LAI), and aboveground biomass (Fig. 4), but only following site-specific calibration. The *uncalibrated* simulation (*limited nitrogen* formulation) overestimated LAI (39 %), aboveground biomass (48%), average peak warm season GPP (15%), and average peak warm season ER (40%) and overestimated cold-season GPP by 200 g C m⁻² yr⁻¹. The *calibrated* simulation was much closer to the observations for LAI and aboveground biomass
(Figure 4). The calibrated peak warm and cold season GPP, and warm season ER matched
observations. The simulated latent heat fluxes were relatively insensitive to the calibration.
Overall the simulated latent heat during the warm season overestimated the observations by
10% and underestimated by 10% during the cold season. Similar improvement was observed
after calibration for the *unlimited nitrogen* run (not shown).

7 The calibration also eliminated erroneous winter GPP. In general, terrestrial carbon 8 models tend to overestimate photosynthesis during cold periods for temperate/boreal conifer 9 forests (Kolari et al., 2007), including Niwot Ridge (Thornton et al., 2002). One approach to 10 correct for this is to include an acclimatization temperature (e.g. Flanagan et al., 2012) that reduces photosynthetic capacity during the spring and fall. The CLM 4.5 model includes 11 12 functionality to adjust the photosynthetic capacity, including both a temperature acclimatization 13 and a day length factor that reduces V_{cmax} (Bauerle et al., 2012; Oleson et al., 2013). However, 14 this alone was not sufficient to match the observed fluxes. Although our calibration approach 15 forced V_{cmax} to zero during the winter, it did not solve the underlying mechanistic shortcoming. A more fundamental approach should address either cold inhibition (Zarter et al., 2006) of 16 17 photosynthesis or soil water availability associated with snowmelt (Monson et al., 2005) to achieve the photosynthetic reduction. Nevertheless, within the confines of our study area, our 18 19 calibration approach was sufficient to provide a skillful representation of photosynthesis and 20 provided a sufficient testbed for evaluating carbon isotope behavior.

21 3.1.2 δ^{13} C of carbon pools

The model performed better simulating δ^{13} C biomass of bulk needle tissue, roots and soil 22 23 carbon (Figure 5) for the *unlimited nitrogen* and no *downregulation discrimination* cases as compared to the *limited nitrogen* case. When nitrogen limitation was included the model 24 25 underestimated δ^{13} C of sunlit needle tissue (1.8 ‰), bulk roots (1.0 ‰), and organic soil carbon (0.7%). All simulations fell within the observed range of δ^{13} C in needles that span from -28.7 26 % (shaded) to -26.7 (sunlit). This vertical pattern in δ^{13} C of leaves is common (Martinelli et 27 al., 1998) and results from vertical differences in nitrogen allocation and photosynthetic 28 29 capacity. The model results integrated the entire canopy and ideally should be closer to sun 30 leaves (as in Figure 5) given that the majority of photosynthesis occurs near the top of the 31 canopy.

1 Model simulations of δ^{13} C of living roots were ~1 ‰ more negative as compared to the 2 structural roots. This range in δ^{13} C results from decreasing δ_{atm} with time (Suess effect, Figure 3 2). The living roots had a relatively fast turnover time of carbon within the model, whereas the 4 structural roots had a slower turnover time and reflected an older (more enriched δ_{atm}) 5 atmosphere. The *limited nitrogen* simulation was a poor match to observations relative to the 6 others (Figure 5, middle panel).

There was an observed vertical gradient in δ^{13} C of soil carbon (-24.9 to -26 ‰) with more enriched values at greater depth (Figure 5, right panel). This vertical gradient is commonly observed (Ehleringer et al., 2000). Simulated δ^{13} C of soil carbon was most consistent with the organic horizon observations. There are a wide variety of post-photosynthetic fractionation processes in the soil system (Bowling et al., 2008; Brüggemann et al., 2011) that are not considered in the CLM 4.5 model, so the match with observations is perhaps fortuitous.

13 **3**

3.2 Photosynthetic discrimination

14 3.2.1 Decadal changes in photosynthetic discrimination and driving factors

All modeled carbon pools showed steady depletion in δ^{13} C since 1850 (coinciding with 15 16 the start of the transient phase of simulations, Figure 5). For the *limited nitrogen* run, there was a decrease in δ^{13} C of 2.3 ‰ for needles, 2.3 ‰ for living roots, and 0.1 ‰ for soil carbon. This 17 18 occurred because of 1) decreased δ_{atm} (Suess effect, Figure 2) and 2) increased photosynthetic 19 discrimination. We quantified the contribution of the Suess effect by performing a control run with constant δ_{atm} , and kept other factors the same (Figure 6). Approximately 70% of the 20 reduction in δ^{13} C of needles occured due to the Suess effect, and the remaining 30% was caused 21 22 by increased photosynthetic discrimination. This occurred as plants responded to CO_2 23 fertilization as illustrated in Figure (7). The model indicated that plants responded to increased atmospheric CO₂ (~40% increase) by decreasing stomatal conductance (Equation 4) by 20% 24 25 for the *limited nitrogen* run and 30% for the *unlimited nitrogen* run (Figure 7B) with associated 26 change in $c_i * c_a$ (Figure 7A). Other influences upon stomatal conductance were less significant, 27 including A_n (+ 10% limited nitrogen, -10% unlimited nitrogen, Figure 7D), soil moisture 28 availability (2-3%, Figure 7E), and negligible changes in relative humidity (potential climate 29 change effects are neglected due to methodological cycling of weather data). This finding that 30 stomatal conductance responded to atmospheric CO₂ is consistent with both tree ring studies 31 (Saurer et al., 2014) and flux tower measurements (Keenan et al., 2013).

1 The effect of CO_2 fertilization, and associated response of stomatal conductance and net 2 assimilation led to a multi-decadal increase in c_i^*/c_a for all model formulations (Figure 7A). The c_i^*/c_a increased from 0.71 to 0.76, 0.67 to 0.71 and 0.66 to 0.68 for the *limited nitrogen*, 3 4 unlimited nitrogen and no downregulation discrimination formulations respectively from 1850-5 2013. All simulations therefore suggested an *increase* in photosynthetic discrimination. This 6 increase in discrimination falls in between two hypotheses posed by Saurer et al., (2004) 7 regarding stomatal response to increased CO₂: 1) reduction in stomatal conductance causes c_i 8 to proportionally increase with c_a keeping c_i/c_a constant and 2) minimal stomatal conductance 9 response where c_i increases at the same rate as c_a (constant $c_a - c_i$) causing c_i/c_a to increase. Our 10 simulation generally agrees with the observed trend in c_i/c_a as estimated from tree ring isotope 11 measurements from a network of European forests (Frank et al., 2015). When controlled for 12 trends in climate, Frank et al. (2015) found that c_i/c_a was approximately constant during the last 13 century. If the Niwot Ridge multi-decadal warming trends in temperature and humidity (Mitton 14 and Ferrenberg, 2012) were included in the CLM simulations the stomatal response may have 15 been stronger thereby holding c_i/c_a constant.

16 The simulated stomatal closure in response to CO₂ fertilization led to an increase in 17 iWUE and WUE of approximately 20% and 10% respectively (Figure 7F) from 1960-2000. This simulated increase in iWUE is consistent with the observation-based studies (Ainsworth 18 19 and Long, 2005; Franks et al., 2013; Peñuelas et al., 2011) which indicate a 15-20% increase in 20 iWUE for forests during that time. The overall increase in WUE suggests that the vegetation at 21 Niwot Ridge has some ability to maintain net ecosystem productivity when confronted with 22 low soil moisture, low humidity conditions. Ultimately, whether Niwot Ridge maintains the 23 current magnitude of carbon sink (Figure 3; Figure S1) will depend upon the severity of drought 24 conditions, as improvements in WUE, in general, are only likely to negate weak to moderate 25 levels of drought (Frank et al., 2013).

The *limited nitrogen* formulation simulated larger values of A_n and g_s , and smaller iWUE as compared to the *unlimited nitrogen* formulation (Figure 7). This is because the *unlimited nitrogen* formulation was fully coupled (i.e. solved simultaneously) between A_n and g_s (Eq. 4). The *limited nitrogen* formulation, however, was only partially coupled because A_n and g_s were initially solved simultaneously through the potential A_n (Eq. 1), however, under N-limitation A_n becomes limited below its potential value (Eq. 9) through f_{dreg} . Therefore g_s is calculated through the potential A_n (Eq. 4) and not the N-limited A_n .

1 The simultaneous increase in both simulated photosynthetic discrimination and iWUE 2 conflicts with previous literature where increases in iWUE are typically linked with weakening 3 discrimination (e.g. Saurer et al., 2004) using a linear model. In general, an increase in 4 atmospheric CO₂ alone tends to increase iWUE because of reduced stomatal conductance, 5 however, the impact upon discrimination is close to neutral because the increased supply of CO₂ external to the leaf is offset by reduced stomatal conductance (Saurer et al., 2004) The 6 7 VPD likely plays an important role in determining the final trends for iWUE and discrimination, 8 where an increasing VPD should further reduce stomatal conductance thereby promoting the 9 well-established relationship (increasing iWUE, decreasing discrimination). In contrast, a weak 10 or decreasing trend in VPD should promote the opposite relationship (increasing iWUE, 11 increasing discrimination).

12 The CLM model at present neglects mesosphyll conductance (g_m) . When Seibt et al. (2008) included g_m in a model that linked iWUE to discrimination they found there were certain 13 14 conditions when iWUE and discrimination increased together. This is in part because mesophyll 15 conductance, unlike stomatal conductance, does not respond as strongly to changes in VPD, yet has a significant impact upon c_i/c_a and discrimination (Flexas et al., 2006). Harvard Forest is 16 17 an example of a site that was observed to show simultaneous increase in iWUE and 18 discrimination over the last two decades derived from tree rings (Belmecheri et al., 2014). In 19 our model simulation, we do not consider multi-decadal trends in climate or mesophyll 20 conductance, therefore increasing atmospheric CO₂ must be the primary driver for the modeled 21 simultaneous increase in discrimination and iWUE at Niwot Ridge (Figure 7). These trends in 22 iWUE and discrimination have also been found in a fully-coupled, isotope enabled, global 23 CESM simulation (Figure S3). Specifically, a random sample of land model grid cells 24 representing conifer species in British Columbia (lat: 52.3°N, lon: -122.5°W) and Quebec (lat: 25 49.5° N, lon: -70.0° W) all showed an increase in photosynthetic discrimination and a 10% 26 increase in WUE from 1850-2005. These randomly chosen grid cells are likely better analogs 27 to the site-level simulations described here because they represent boreal conifer forests, 28 whereas the grid cells that are in the Niwot Ridge area were heterogeneous in land cover (e.g. 29 tundra, grassland, forest) and a poor representation of conifer forest.

30

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The relationship between iWUE and discrimination in the global simulation suggest that the site level trends are not isolated to the specific conditions of Niwot Ridge, but are a function

1 of the model formulation. There is a relationship between iWUE and c_{i*}/c_a (discrimination) as 2 derived from equation (11) within the CLM model,

3
$$\frac{c_i^*}{c_a} \cong 1 - \frac{1.6}{c_a} iWUE.$$
 (19)

4 The full derivation is provided in the supplement. Note that according to equation (19) 5 increasing iWUE can be consistent with weakening discrimination (decreasing $c_i*/c_a(\sim \alpha)$) and 6 therefore consistent with established understanding between trends in iWUE and 7 discrimination. However, this can be moderated by increasing c_a . During the course of our 8 simulation (1850-2013) iWUE increased between 15-20% (Figure 7), however, c_a increased by 9 40%.

10

11 3.2.2 Magnitude of photosynthetic discrimination

The simulated photosynthetic discrimination (Fig. 8) was significantly larger than an 12 estimate derived from observations and an isotopic mixing model (Bowling et al., 2014). For 13 14 brevity we refer to the estimates based on the Bowling et al. (2014) method as 'observed' 15 discrimination but highlight that they are derived from observations and not directly measured. 16 On average, the simulated monthly growing season mean canopy discrimination was greater 17 than observed values by 6.3, 6.1, and 5.1% for the *limited nitrogen*, *unlimited nitrogen*, and *no* downregulation discrimination formulations respectively. The model-observation mismatch in 18 19 discrimination, despite model-observation agreement to biomass, carbon and latent heat flux 20 tower observations (Figure 3) highlights the independent, and useful constraint isotopic 21 observations provide for evaluating model performance. Specifically, the overestimation of 22 discrimination may suggest the stomatal slope in the Ball-Berry model (m=9 in Eq. 4) used for 23 these simulations was too high. This is supported by Mao et al., (2016), who found a reduced stomatal slope (m=5.6) was necessary for CLM 4.0 to match observed δ^{13} C in an isotope 24 25 labeling study of loblolly pine forest in Tennessee. The stomatal slope was also important to match discrimination behavior in the ISOLSM model (Aranibar et al., 2006), a predecessor to 26 CLM. A global analysis of stomatal slope inferred from leaf gas exchange measurements found 27 that evergreen coniferous species, such as those at Niwot Ridge, had near the lowest values 28 29 compared to other PFTs (Lin et al., 2015). In addition, they found that low stomatal slope values were characteristic of species with low stemwood construction costs per water transpired
 (high WUE), low soil moisture availability, and cold temperatures.

3 Alternatively, discrimination may be overestimated because CLM does not consider the resistance to CO₂ diffusion into the leaf chloroplast. The ability of CO₂ to diffuse across the 4 5 chloroplast boundary layer, cell wall, and liquid interface is collectively known as the 6 mesophyll conductance (g_m) (Flexas et al., 2008). Multiple studies suggest that g_m is 7 comparable in magnitude to g_s , and responds similarly to environmental conditions (Flexas et 8 al., 2008). CLM does not account for g_m , and as a result assumes the intracellular CO₂ is the 9 same as intercellular CO₂, when it can be significantly lower (Di Marco et al., 1990; Sanchez-Rodriguez et al., 1999). The overestimation of c_i^* could have two important impacts upon our 10 simulation. First, this may lead to unrealistically low values of V_{cmax} in order to compensate 11 for the overestimation of c_i^* . In fact, we reduced the default value of V_{cmax} as much as 50% in 12 13 our simulation to match the eddy covariance flux tower observations (see Section 3.3). Second, 14 the overestimation of c_i^* should cause an overestimation of discrimination (Eq. 10), which is 15 also consistent with our simulations (Figure 8). To determine whether the simulated discrimination bias is a model parameter calibration issue (g_s) or from excluding g_m , we 16 17 recommend a mechanistic representation of mesophyll conductance within CLM.

18 The mixing model approach estimate of Δ_{canopy} (17 ‰), combined with δ_{atm} (-8.25 ‰) implies a δ^{13} C of biomass between -26 to -25 ‰ (Figure 8). This range is only slightly more 19 enriched than the observed ranges of δ^{13} C of needle and root biomass (-27 to -26 ‰). The fact 20 21 that the different approaches to measure discrimination differ by only 1 ‰, whereas CLM simulates a Δ_{canopy} that is 5-6 ‰ greater than the mixing model discrimination, strongly suggests 22 23 that the model has overestimated discrimination from 2006-2012. Therefore what appeared to be a successful match between the simulated and observed δ^{13} C biomass, may in fact have been 24 fortuitous. A multi-decadal time series of discrimination inferred from $\delta^{13}C$ of tree rings 25 (Saurer et al., 2014; Frank et al., 2015) would be useful to investigate this mismatch as a 26 27 function of time, but these data are not presently available.

If the overestimation of modeled discrimination originates from a lack of response of stomatal conductance to environmental conditions this could be a result of one or several of the following within the model: 1) the stomatal slope value is too high, 2) multi-decadal trends in climate (e.g. VPD) have not been included in the simulation 3) the model neglects g_m or 4) the Ball-Berry representation of g_s is not sensitive enough to changes in environmental conditions 1 (e.g. humidity, soil moisture). It has been shown that VPD may be an improved predictor of g_s 2 (Katul et al., 2000; Leuning, 1995) and discrimination (Ballantyne et al., 2010, 2011) as 3 compared to relative humidity, currently used in CLM 4.5. Future work should consider which 4 of these scenarios is responsible for overestimation of discrimination.

5 3.2.3 Seasonal pattern of photosynthetic discrimination

6 The model formulations that did not explicitly consider the influence of nitrogen 7 limitation upon discrimination (unlimited nitrogen, no downregulation discrimination) were 8 most successful at reproducing the seasonality of discrimination (Figure 8; Figure S4). In 9 general, the observed discrimination was stronger during the spring and fall and weaker during 10 summer. This observed Δ_{canopy} seasonal range (excluding November) varied from 16.5 to 18 11 ‰ using Reichstein partitioning (Figure 8), and was more pronounced using Lasslop 12 partitioning (16.5 to 23 %) (Figure S4). The nitrogen limited simulated Δ_{canopy} had no seasonal 13 trend whereas the unlimited nitrogen and no downregulation discrimination simulations both 14 ranged from 21 to 23 %.

15 The main driver of the seasonality of discrimination was the net assimilation (A_n) for This was evident given the inversely 16 the *unlimited nitrogen* formulation (Figure 9). 17 proportional relationship between the simulated fractionation factor (α) and A_n , consistent with 18 equation (11). Stomatal conductance (g_s) also influenced the seasonal pattern. The most direct 19 evidence for this was during the period between days 175-200 (Figure 9), where A_n descended from its highest value (favoring higher α), and g_s abruptly ascended to its highest value 20 21 (favoring higher α). The α responded to this increase in g_s with an abrupt increase by 22 approximately 0.003 (3 ‰). Similarly, the *limited nitrogen* simulation seasonal discrimination 23 pattern was shaped by both A_n and g_s although the magnitude for both was approximately 30% 24 higher during the summer months as compared to the unlimited nitrogen simulation. This was 25 because the calibrated V_{cmax} value for the *limited nitrogen* simulation was much higher than for 26 the *unlimited nitrogen* simulation (section 3.3). The difference in α between the two model 27 formulations coincided with the sharp increase in f_{dreg} between days 125 and 275, providing 28 strong evidence that the downregulation mechanism within the *limited nitrogen* formulation led 29 to increased discrimination during the summer. Therefore, it follows that the nitrogen 30 downregulation mechanism was the root cause of the small range in simulated seasonal cycle 31 discrimination for the limited nitrogen formulation, which was inconsistent with the 32 observations.

1 3.2.4 Environmental factors influencing seasonality of discrimination

2 The simulated Δ_{canopy} was driven primarily by net assimilation (A_n), followed by vapor 3 pressure deficit (VPD) (Fig. 10). The correlation between VPD and Δ_{canopy} was strongest for the *unlimited nitrogen* simulation, where the range in monthly average Δ_{canopy} spanned values 4 5 from 22 to 18 ‰ (Figure 10, middle row). This resembled the observed range in response based 6 upon a fitted relationship from Bowling et al., (2014) that spanned from roughly 16 to 19 ‰ 7 (left panels of Fig. 10), although with a consistent discrimination bias. The correlation between 8 VPD and Δ_{canopy} , however, does not demonstrate causality. If that were the case, given that g_s 9 is a function of VPD (h_s term in Eq. 4) and discrimination is a function of g_s (Eq. 10; Eq. 11), a similar relationship should have existed between g_s and Δ_{canopy} . This, in fact, was not the case. 10 11 Overall, the influence of g_s (responding to VPD) (R-value = -0.50) was secondary to A_n (R-12 value = -0.77) in driving changes in discrimination (Figure 10). The model suggested that the 13 range in seasonal discrimination (intra-annual variation) was driven by the magnitude of A_n 14 based on the inverse relationship between A_n and Δ_{canopy} (equation 11) illustrated by the separation between months of low photosynthesis (October, May) vs. high photosynthesis 15 16 (June, July, August). During times of relatively low photosynthesis A_n also drove the interannual variation in Δ_{canopy} . On the other hand, g_s (VPD) was most influential in driving the 17 18 inter-annual variation of discrimination during the summer months only, judging by the directly proportional relationship during the months of June, July and August. Strictly speaking, g_s is a 19 20 function of h_s (leaf relative humidity) and not atmospheric VPD in CLM. However, the two 21 are closely related and the relationship between either variable (atmospheric VPD or simulated 22 leaf humidity) to Δ_{canopy} was similar (Figure S5).

23 The *limited nitrogen* formulation did not produce as wide a range in discrimination as 24 compared to the observations (Figure 10, top row). Part of this result was attributed to the lack 25 of response between A_n and Δ_{canopy} . In this case, the discrimination did not decrease with increasing A_n because the signal was muted by the countering effect of f_{dreg} . The *limited* 26 27 *nitrogen* formulation was, however, able to reproduce the same discrimination response to g_s . 28 as compared to the other model formulations. The tendency for the limited nitrogen model to simulate discrimination response to g_s and not to A_n may negatively impact its ability to simulate 29 multi-decadal trends in discrimination. This may not be a major detriment to sites such as 30 Niwot Ridge which have maintained a consistent level of carbon uptake during the last decade, 31 32 and is likely more susceptible to environmental impact upon stomatal conductance. However, sites that have shown a significant increase in assimilation rate (e.g. Harvard Forest; (Keenan
 et al., 2013)) are less likely to be well represented by this model formulation.

3 Given the dependence of forest productivity at Niwot Ridge on snowmelt (Hu et al., 4 2010), it was surprising that the model simulated minimal soil moisture stress (Fig. 9e) and therefore minimal discrimination response to soil moisture. However, this finding was 5 6 consistent with Bowling et al., (2014), who did not find an isotopic response to soil moisture. 7 In addition, lack of response to change in soil moisture may not be indicative of poor 8 performance of the isotopic sub-model performance, but rather an effect of the hydrology sub-9 model (Duarte et al. (in prep)). However, a comparison of observed soil moisture at various 10 depths at Niwot Ridge generally agrees with the CLM simulated soil moisture (not shown), 11 suggesting the lack of model response to soil moisture was not from biases in the hydrology 12 model.

3.3 Discrimination formulations: implications for model development

14 The limited and unlimited model formulations tested in this study represented two 15 approaches to account for nitrogen limitation within ecosystem models. The *limited nitrogen* formulation reduced photosynthesis, after the main photosynthesis calculation, so that the 16 17 carbon allocated to growth was accommodated by available nitrogen. This *allocation* 18 downscaling approach is common to a subset of models, for example, CLM (Thornton et al., 19 2007), DAYCENT (Parton et al., 2010) and ED2.1 (Medvigy et al., 2009). Another class of 20 models limits photosynthesis based upon foliar nitrogen content and adjusts the photosynthetic 21 capacity through nitrogen availability in the leaf through V_{cmax} (e.g. CABLE, GDAY, LPJ-GUESS, OCN, SDVGM, TECO, see Zaehle et al., 2014). These foliar nitrogen models are 22 23 similar to the *unlimited nitrogen* formulation of CLM because the scaling of photosynthesis was taken into account in the V_{cmax} scaling methodology (see discussion in section 2.1.2 and 24 25 2.4), prior to the photosynthesis calculation. In general, there were no categorical differences 26 in behavior between these two classes of models during CO₂ manipulation experiments held at 27 Duke forest and ORNL (Zaehle et al., 2014). CLM 4.0 was one of the few models in that study to consistently underestimate the NPP response to an increase of atmospheric CO₂ due to 28 29 nitrogen limitation, however this finding was attributed to a lower initial supply of nitrogen. 30 Also within this experiment, it was found that models that had no or partial coupling (CLM 4.0, 31 DAYCENT) between A_n and g_s , generally predicted lower than observed WUE response to 32 increases in CO₂ (De Kauwe et al., 2013). Similar to CLM 4.0, the *limited nitrogen* formulation

of CLM 4.5 in this manuscript is partially coupled (see Section 3.2.1). The unlimited nitrogen
 formulation of CLM 4.5, on the other hand, is fully coupled and similar to De Kauwe et al.,
 (2013) outperformed the partially coupled version of CLM.

The *unlimited nitrogen* formulation described in our study is a simplified foliar nitrogen model, in that, all of the information about nitrogen limitation is incorporated within the V_{cmax} downscaling approach. A more versatile approach would link a dynamic nitrogen cycle directly with the calculation of V_{cmax} . This capability was recently developed within CLM (Ghimire et al., 2016) and is scheduled to be included in the next CLM release. Future work should test its functionality.

10 The performance of the *unlimited nitrogen* formulation was nearly identical to the *no* 11 downregulation discrimination formulation in terms of isotopic behavior despite the 12 mechanistic differences. The no downregulation discrimination formulation included nitrogen limitation within the bulk carbon behavior but ignored the impact of f_{dreg} upon discrimination 13 14 behavior. The relative high simulation skill with this formulation implied that the 'potential' 15 GPP linked to A_n , was a more effective predictor of discrimination behavior than the 16 'downscaled' GPP, which is linked to $A_n * (1-f_{dreg})$ (equation 11). There are several potential 17 explanations for an unrealistically large value of f_{dreg} . First this could indicate that the V_{cmax} 18 parameter was too large, thereby requiring a large f_{dreg} to compensate. As noted in Section (3.1) the default temperate every reen V_{cmax25} was ~62 µmol m⁻² s⁻¹, much larger than what was 19 found based on literature reviews (Monson et al., 2005; Tomaszewski and Sievering, 2007). 20 We found to match the observed GPP we had to impose f_{dreg} that had the same effect as reducing 21 V_{cmax} (Figure S2) to values of 51 and 34 µmol m⁻² s⁻¹ for the *limited nitrogen* and *unlimited* 22 nitrogen formulations respectively. Alternatively, it could be that there are physiological 23 24 processes that are acting to reduce nitrogen limitation (e.g. nitrogen storage pools or transient 25 carbon storage as non-structural carbohydrates), or that the current measurement techniques are underestimating GPP due to biases within the flux partitioning methods. 26

27 **3.4** Disequilibrium, possible explanations of mismatch

Carbon cycle models (e.g. Fung et al., 1997) indicate that the steady decrease of δ_{atm} (Suess effect, Fig. 2) should lead to a positive disequilibrium between land surface processes (δ^{13} C difference between GPP and ER, Eq. 14). This is because the δ_{GPP} reflects the most recent (δ^{13} C depleted) state of the atmosphere, whereas the δ_{ER} reflects carbon (e.g. soil carbon) assimilated

from an older (δ^{13} C enriched) atmosphere. This positive disequilibrium pattern promoted by 1 2 the Suess effect was consistent with all CLM formulations for this study with an annual average 3 disequilibrium of 0.8 %. In contrast, a negative disequilibrium (-0.6 ‰) was identified at 4 Niwot Ridge based upon observations (Bowling et al. 2014) as well as in other forests (Flanagan 5 et al., 2012; Wehr and Saleska, 2015; Wingate et al., 2010). Bowling et al. (2014) hypothesized several reasons for this: 1) a strong seasonal stomatal response to atmospheric humidity, 2) 6 7 decreased photosynthetic discrimination associated with CO₂ fertilization, 3) decreased 8 photosynthetic discrimination associated with multi-decadal warming and increased VPD, and 9 4) post-photosynthetic discrimination. We evaluated the first three hypotheses within the 10 context of our CLM simulations.

11 The model results suggest a seasonal variation of discrimination that is a function of both 12 VPD and A_n . The simulated seasonal range in discrimination (Figure 8; Figure S4) varied by 13 approximately 2 ‰, and this range in seasonal discrimination could contribute to a negative 14 disequilibrium provided specific timing of assimilation, assimilate storage and respiration not currently considered in the model. For example, if a significant portion of photosynthetic 15 assimilation was stored during the spring with relatively high discrimination, and then respired 16 17 during the summer, the net effect would deplete the δ_{ER} and thereby promote negative disequilibrium during the summer months when discrimination is lower. Theoretically, this 18 19 could be achieved by explicitly including carbohydrate storage pools within CLM. Isotopic tracer studies have shown assimilated carbon can exist for weeks to months within the 20 21 vegetation and soil before it is finally respired (Epron et al., 2012; Hogberg et al., 2008). 22 Although carbon storage pools are included in CLM, their allocation is almost always 23 instantaneous for evergreen systems and could not provide the isotopic effect described above.

24 The CO₂ fertilization effect tends to favor photosynthesis in plants and has been shown to simultaneously increase WUE and decrease stomatal conductance as inferred from δ^{13} C in tree 25 rings (Frank et al., 2015; Flanagan et al., 2012; Wingate et al., 2010). In general a decrease in 26 stomatal conductance and increase in WUE is associated with a decrease in C3 discrimination 27 28 (Farquhar et al., 1982), which opposes the disequilibrium trend imposed by the Suess effect. 29 The model simulation agrees with both these trends in WUE and stomatal conductance, yet 30 simulates an *increase* in discrimination (Figure 6; Figure 7), which reinforces the Suess effect pattern upon disequilibrium. Although this appears to be a mismatch between forest processes 31 32 and model performance the model is operating within the limits of the discrimination parameterization (Eq. 17) in which the magnitude of photosynthetic discrimination is inversely
 proportional to the iWUE, but is also proportional to atmospheric CO₂ (see section 3.2.1).

3 A multi-decadal decrease in photosynthetic discrimination may also result from change in 4 climate. Meteorological measurements at Niwot Ridge during the last several decades 5 generally support conditions of higher VPD based upon a warming trend from an average 6 annual temperature of 1.1 °C in the 1980's to 2.7 °C in the 2000's (Mitton and Ferrenberg, 7 2012) and no overall trend in precipitation. It is possible that a multi-decadal trend in increasing 8 VPD contributed to multi-decadal weakening in photosynthetic discrimination given the 9 observed (Bowling et al., 2014) and modelled (Figure 10) correlation between Δ_{canopy} and VPD. The model meteorology only included the years 1998-2013 and did not include the rapid 10 11 warming after the 1980's. It is unclear whether, if the full period of warming were to be 12 included in the simulation, the simulated discrimination response to VPD would be enough to 13 counter the Suess effect and lead to negative disequilibrium. Still, there is evidence that the 14 model is overestimating contemporary discrimination (Section 3.4) and the exclusion of the full 15 multi-decadal shift in VPD could be a significant reason why.

Finally, post-photosynthetic discrimination processes are likely to impact the magnitude and sign of the isotopic disequilibrium (Bowling et al., 2008; Brüggemann et al., 2011) at multiple temporal scales. None of these isotopic processes are currently modelled within CLM 4.5, so at present the model cannot be used to examine them.

20 4 Conclusions

This study provides a rigorous test of the representation of C isotope discrimination within the highly mechanistic terrestrial carbon model CLM. CLM was able to accurately simulate δ^{13} C in leaf and stem biomass and the seasonal cycle in Δ_{canopy} , but only when V_{cmax} was calibrated to mimic the functionality of a foliar nitrogen model by accounting for nitrogen limitation *prior* to photosynthesis.

Although the *unlimited nitrogen* formulation was able to match observed δ^{13} C of biomass and seasonal patterns in discrimination, it still overestimated the contemporary magnitude of discrimination (2006-2012). Future work should identify whether this overestimation was a result of parameterization (stomatal slope), exclusion of multi-decadal shifts in VPD, limitations in the representation of stomatal conductance (Ball-Berry model) or absence of the representation of mesophyll conductance. 1 The model attributed most of the range in seasonal discrimination to variation in net 2 assimilation rate (A_n) followed by variation in VPD, with little to no impact from soil moisture. 3 The model suggested that A_n drove the seasonal range in discrimination (across-month 4 variation) whereas VPD drove the inter-annual variation during the summer months. This 5 finding suggests that to simulate multi-decadal trends in photosynthetic discrimination, 6 response to assimilation rate and VPD must be well represented within the model.

7 The model simulated a positive disequilibrium that was driven by both the Suess effect, 8 and increased photosynthetic discrimination from CO_2 fertilization. It is possible that the 9 negative disequilibrium that was inferred from observations (Bowling et al., 2014) was driven 10 from the impacts of climate change and/or post-photosynthetic discrimination – not considered 11 in this version of the model.

The model simulated a consistent increase in water-use efficiency as a response to CO_2 fertilization and decrease in stomatal conductance. The model simulated an increase in WUE despite an increase in discrimination, however C3 plants typically express the opposite trends (increase in WUE, decrease in discrimination). Although CLM includes parameterization that promotes an increase in WUE with a decrease in discrimination, this trend was likely moderated by an increase in *c_a*.

Initial indications are that δ^{13} C isotope data can bring additional constraint to model 18 19 parameterization beyond what traditional flux tower measurements of carbon, water exchange, 20 and biomass measurements. The isotope measurements suggested a stomatal conductance 21 value generally lower than what was consistent with the flux tower measurements. 22 Unexpectedly, the isotopes also provided guidance upon model formulation related to nitrogen 23 limitation. The success of our empirical approach to account for nutrient limitation within the 24 V_{cmax} parameterization, suggests that additional testing of foliar nitrogen models are worthwhile. 25

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- 27

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1 References

- 2 Ainsworth, E. A. and Long, S. P.: What have we learned from 15 years of free-air CO2 3 enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy 4 properties and plant production to rising CO2, New Phytol., 165(2), 351-372, 5 doi:10.1111/j.1469-8137.2004.01224.x, 2005.
- 6 Andrews, S. F., Flanagan, L. B., Sharp, E. J. and Cai, T.: Variation in water potential, hydraulic 7 characteristics and water source use in montane Douglas-fir and lodgepole pine trees in 8 southwestern Alberta and consequences for seasonal changes in photosynthetic capacity, Tree 9 Physiol., 32, 146–160, doi:10.1093/treephys/tpr136, 2012.
- 10 Aranibar, J. N., Berry, J. A., Riley, W. J., Pataki, D. E., Law, B. E. and Ehleringer, J. R.:
- Combining meteorology, eddy fluxes, isotope measurements, and modeling to understand 11
- 12 environmental controls of carbon isotope discrimination at the canopy scale, Glob. Change
- 13 Biol., 12(4), 710–730, 2006.
- 14 Arora, V. K., Boer, G. J., Friedlingstein, P., Eby, M., Jones, C. D., Christian, J. R., Bonan, G.,
- 15 Bopp, L., Brovkin, V., Cadule, P., Hajima, T., Ilyina, T., Lindsay, K., Tjiputra, J. F. and Wu,
- 16 T.: Carbon-Concentration and Carbon-Climate Feedbacks in CMIP5 Earth System Models, J.
- 17 Clim., 26(15), 5289-5314, doi:10.1175/jcli-d-12-00494.1, 2013.
- 18 Ballantyne, A. P., Miller, J. B. and Tans, P. P.: Apparent seasonal cycle in isotopic 19 discrimination of carbon in the atmosphere and biosphere due to vapor pressure deficit, Glob. 20 Biogeochem. Cycles, 24, GB3018, doi:10.1029/2009GB003623, 2010.
- 21 Ballantyne, A. P., Miller, J. B., Baker, I. T., Tans, P. P. and White, J. W. C.: Novel applications 22 of carbon isotopes in atmospheric CO2: What can atmospheric measurements teach us about 23 processes in the biosphere?, Biogeosciences, 8(10), 3093–3106, 2011.
- 24 Ball, J.T., Woodrow, I.E. & Berry, J. A. Progress in Photosynthesis Research, Martinus Nijhoff 25 Publishers, 1987.
- 26 Bauerle, W. L., Oren, R., Way, D. A., Qian, S. S., Stoy, P. C., Thornton, P. E., Bowden, J. D., 27 Hoffman, F. M. and Reynolds, R. F.: Photoperiodic regulation of the seasonal pattern of 28 photosynthetic capacity and the implications for carbon cycling, Proc. Natl. Acad. Sci. U. S.
- 29 A., 109(22), 8612–8617, 2012.
- 30 Belmecheri, S., Maxwell, R. S., Taylor, A. H., Davis, K. J., Freeman, K. H. and Munger, W. J.:
- Tree-ring δ 13C tracks flux tower ecosystem productivity estimates in a NE temperate forest, 31 32 Environ. Res. Lett., 9(7), 74011, doi:10.1088/1748-9326/9/7/074011, 2014.
- 33 Boisvenue, C. and Running, S. W.: Simulations show decreasing carbon stocks and potential 34 for carbon emissions in Rocky Mountain forests over the next century, Ecol. Appl., 20(5), 35 1302–1319, 2010.
- 36 Bowling, D. R., Pataki, D. E. and Randerson, J. T.: Carbon isotopes in terrestrial ecosystem 37
- pools and CO2 fluxes, New Phytol., 178, 24–40, doi: 10.1111/j.1469–8137.2007.02342.x,
- 38 2008.

- 1 Bowling, D. R., Ballantyne, A. P., Miller, J. B., Burns, S. P., Conway, T. J., Menzer, O.,
- 2 Stephens, B. B. and Vaughn, B. H.: Ecological processes dominate the 13C land disequilibrium
- 3 in a Rocky Mountain subalpine forest, Glob. Biogeochem. Cycles, 28(4), 2013GB004686,
- 4 doi:10.1002/2013GB004686, 2014a.
- 5 Bowling, D. R., Ballantyne, A. P., Miller, J. B., Burns, S. P., Conway, T. J., Menzer, O.,
- 6 Stephens, B. B. and Vaughn, B. H.: Ecological processes dominate the 13C land disequilibrium
- 7 in a Rocky Mountain subalpine forest, Glob. Biogeochem. Cycles, 28(4), 2013GB004686,
- 8 doi:10.1002/2013GB004686, 2014b.
- 9 Bradford, M. A., Fierer, N. and Reynolds, J. F.: Soil carbon stocks in experimental mesocosms
- 10 are dependent on the rate of labile carbon, nitrogen and phosphorus inputs to soils, Funct. Ecol.,
- 11 22(6), 964–974, doi:10.1111/j.1365-2435.2008.01404.x, 2008.
- 12 Braswell, B. H., Sacks, W. J., Linder, E. and Schimel, D. S.: Estimating diurnal to annual
- ecosystem parameters by synthesis of a carbon flux model with eddy covariance net ecosystem
 exchange observations, Glob. Change Biol., 11, 335–355, doi:10.1111/j.13652486 2005 00897 x 2005
- 15 2486.2005.00897.x, 2005.
- 16 Brüggemann, N., Gessler, A., Kayler, Z., Keel, S. G., Badeck, F., Barthel, M., Boeckx, P.,
- 17 Buchmann, N., Brugnoli, E., Esperschütz, J., Gavrichkova, O., Ghashghaie, J., Gomez-
- 18 Casanovas, N., Keitel, C., Knohl, A., Kuptz, D., Palacio, S., Salmon, Y., Uchida, Y. and Bahn,
- 19 M.: Carbon allocation and carbon isotope fluxes in the plant-soil-atmosphere continuum: a
- 20 review, Biogeosciences, 8(11), 3457–3489, doi:10.5194/bg-8-3457-2011, 2011.
- Collatz, G. J., Ball, J. T., Grivet, C. and Berry, J. A.: Regulation of stomatal conductances and
 transpiration a physiological model of canopy processes, Agric. For. Meteorol., 54, 107–136,
 1991.
- 24 De Kauwe, M. G., Medlyn, B. E., Zaehle, S., Walker, A. P., Dietze, M. C., Hickler, T., Jain, A.
- 25 K., Luo, Y., Parton, W. J., Prentice, I. C., Smith, B., Thornton, P. E., Wang, S., Wang, Y.-P.,
- 26 Wårlind, D., Weng, E., Crous, K. Y., Ellsworth, D. S., Hanson, P. J., Seok Kim, H., Warren, J.
- M., Oren, R. and Norby, R. J.: Forest water use and water use efficiency at elevated CO2: a
- 28 model-data intercomparison at two contrasting temperate forest FACE sites, Glob. Change
- 29 Biol., 19(6), 1759–1779, doi:10.1111/gcb.12164, 2013.
- Desai, A. R., Moore, D. J. P., Ahue, W. K. M., Wilkes, P. T. V., De Wekker, S. F. J., Brooks,
 B. G., Campos, T. L., Stephens, B. B., Monson, R. K., Burns, S. P., Quaife, T., Aulenbach, S.
 M. and Schimel, D. S.: Seasonal pattern of regional carbon balance in the central Rocky
 Mountains from surface and airborne measurements, J. Geophys. Res., 116, G04009(4),
 doi:10.1029/2011JG001655, 2011.
- Di Marco, G., Manes, F., Tricoli, D. and Vitale, E.: Fluorescence Parameters Measured
 Concurrently with Net Photosynthesis to Investigate Chloroplastic CO2 Concentration in
 Leaves of Quercus ilex L., J. Plant Physiol., 136(5), 538–543, doi:10.1016/S01761617(11)80210-5, 1990.
- Dlugokencky, E. J., Lang, P. M., Masarie, K. A., Crotwell, A. M. and Crotwell, M. J.:
 Atmospheric Carbon Dioxide Dry Air Mole Fractions from the NOAA ESRL Carbon Cycle

- 1 Cooperative Global Air Sampling network, 1968-2014, Version: 2015-08-03, Path:
- 2 ftp://aftp.cmdl.noaa.gov/data/trace_gases/co2/flask/surface/, 2015.
- Ehleringer, J. R., Buchmann, N. and Flanagan, L. B.: Carbon isotope ratios in belowground
 carbon cycle processes, Ecol. Appl., 10(2), 412–422, 2000.
- 5 Epron, D., Bahn, M., Derrien, D., Lattanzi, F. A., Pumpanen, J., Gessler, A., Högberg, P.,
- 6 Maillard, P., Dannoura, M., Gérant, D. and Buchmann, N.: Pulse-labelling trees to study carbon
- 7 allocation dynamics: a review of methods, current knowledge and future prospects, Tree
- 8 Physiol., 32(6), 776–798, doi:10.1093/treephys/tps057, 2012.
- 9 Farquhar, G. D., von Caemmerer, S. and Berry, J. A.: A Biochemical Model of Photosynthetic
- 10 CO2 Assimilation in Leaves of C3 Species, Planta, 149, 78–90, 1980.
- 11 Farquhar, G. D., O'Leary, M. H. and Berry, J. A.: On the relationship between carbon isotope
- discrimination and the intercellular carbon dioxide concentration in leaves, Aust. J. Plant
- 13 Physiol., 9(2), 121–137, 1982.
- Farquhar, G. D., Ehleringer, J. R. and Hubick, K. T.: Carbon isotope discrimination and
 photosynthesis, Annu. Rev. Plant Physiol. Plant Mol. Biol., 40, 503–537, 1989.
- 16 Flanagan, L. B., Cai, T., Black, T. A., Barr, A. G., McCaughey, J. H. and Margolis, H. A.:
- 17 Measuring and modeling ecosystem photosynthesis and the carbon isotope composition of
- 18 ecosystem-respired CO2 in three boreal coniferous forests, Agric. For. Meteorol., 153, 165-
- 19 176, 2012.
- Flexas, J., Ribas-Carbó, M., Hanson, D. T., Bota, J., Otto, B., Cifre, J., McDowell, N., Medrano,
 H. and Kaldenhoff, R.: Tobacco aquaporin NtAQP1 is involved in mesophyll conductance to
- 22 CO2in vivo, Plant J., 48(3), 427–439, doi:10.1111/j.1365-313X.2006.02879.x, 2006.
- Flexas, J., Ribas-Carbo, M., Diaz-Espej, A., Galmes, J. and Medrano, H.: Mesophyll
 conductance to CO2: current knowledge and future prospects, Plant Cell Environ., 31(5), 602–
 621, 2008.
- Francey, R. J., Allison, C. E., Etheridge, D. M., Trudinger, C. M., Enting, I. G., Leuenberger,
 M., Langenfelds, R. L., Michel, E. and Steele, L. P.: A 1000-year high precision record of d13C
 in atmospheric CO2, Tellus, 51B, 170–193, 1999.
- 29 Frank, D. C., Poulter, B., Saurer, M., Esper, J., Huntingford, C., Helle, G., Treydte, K., 30 Zimmermann, N. E., Schleser, G. H., Ahlström, A., Ciais, P., Friedlingstein, P., Levis, S., 31 Lomas, M., Sitch, S., Viovy, N., Andreu-Hayles, L., Bednarz, Z., Berninger, F., Boettger, T., 32 D'Alessandro, C. M., Daux, V., Filot, M., Grabner, M., Gutierrez, E., Haupt, M., Hilasvuori, 33 E., Jungner, H., Kalela-Brundin, M., Krapiec, M., Leuenberger, M., Loader, N. J., Marah, H., 34 Masson-Delmotte, V., Pazdur, A., Pawelczyk, S., Pierre, M., Planells, O., Pukiene, R., 35 Reynolds-Henne, C. E., Rinne, K. T., Saracino, A., Sonninen, E., Stievenard, M., Switsur, V. 36 R., Szczepanek, M., Szychowska-Krapiec, E., Todaro, L., Waterhouse, J. S. and Weigl, M.: 37 Water-use efficiency and transpiration across European forests during the Anthropocene, Nat. 38 Clim. Change, 5(6), 579–583, doi:10.1038/nclimate2614, 2015.
- Franks, P. J., Adams, M. A., Amthor, J. S., Barbour, M. M., Berry, J. A., Ellsworth, D. S.,
 Farquhar, G. D., Ghannoum, O., Lloyd, J., McDowell, N., Norby, R. J., Tissue, D. T. and von

- 1 Caemmerer, S.: Sensitivity of plants to changing atmospheric CO2 concentration: From the
- 2 geological past to the next century, New Phytol., 197(4), 1077–1094, 2013.
- 3 Friedlingstein, P., Cox, P. M., Betts, R. A., Bopp, L., von Bloh, W., Brovkin, V., Cadule, P.,
- 4 Doney, S. C., Eby, M., Fung, I. Y., Bala, G., John, J., Jones, C. D., Joos, F., Kato, T., Kawamiya,
- 5 M., Knorr, W., Lindsay, K., Matthews, H. D., Raddatz, T., Rayner, P., Reick, C., Roeckner, E.,
- 6 Schnitzler, K.-G., Schnur, R., Strassmann, K., Weaver, A. J., Yoshikawa, C. and Zeng, N.:
- 7 Climate-carbon cycle feedback analysis: Results from the C4MIP model intercomparison, J.
- 8 Clim., 19, 3337–3353, 2006.
- 9 Fung, I. Y., Field, C. B., Berry, J. A., Thompson, M. V., Randerson, J. T., Malmstrom, C. M.,
- 10 Vitousek, P. M., Collatz, G. J., Sellers, P. J., Randall, D. A., Denning, A. S., Badeck, F. and
- 11 John, J.: Carbon 13 exchanges between the atmosphere and biosphere, Glob. Biogeochem.
- 12 Cycles, 11(4), 507–533, 1997.
- 13 Ghimire, B., Riley, W. J., Koven, C. D., Mu, M. and Randerson, J. T.: Representing leaf and
- 14 root physiological traits in CLM improves global carbon and nitrogen cycling predictions, J.
- 15 Adv. Model. Earth Syst., n/a-n/a, doi:10.1002/2015MS000538, 2016.
- Greenland, D.: The Climate of Niwot Ridge, Front Range, Colorado, U.S.A., Arct. Alp. Res.,
 21(4), 380–391, 1989.
- 18 Hogberg, P., Hogberg, M. N., Gottlicher, S. G., Betson, N. R., Keel, S. G., Metcalfe, D. B.,
- Campbell, C., Schindlbacher, A., Hurry, V., Lundmark, T., Linder, S. and Nasholm, T.: High
 temporal resolution tracing of photosynthate carbon from the tree canopy to forest soil
 microorganisms, New Phytol., 177(1), 220–228, 2008.
- Hu, J., Moore, D. J. P., Burns, S. P. and Monson, R. K.: Longer growing seasons lead to less
 carbon sequestration by a subalpine forest, Glob. Change Biol., 16(2), 771–783,
 doi:10.1111/j.1365-2486.2009.01967.x, 2010.
- Katul, G. G., Ellsworth, D. S. and Lai, C.-T.: Modelling assimilation and intercellular CO2
 from measured conductance: a synthesis of approaches, Plant Cell Environ., 23(12), 1313–
 1328, doi:10.1046/j.1365-3040.2000.00641.x, 2000.
- Keenan, T. F., Hollinger, D. Y., Bohrer, G., Dragoni, D., Munger, J. W., Schmid, H. P. and
 Richardson, A. D.: Increase in forest water-use efficiency as atmospheric carbon dioxide
 concentrations rise, Nature, 499(7458), 324–327, doi:10.1038/nature12291, 2013.
- Kolari, P., Lappalainen, H. K., HäNninen, H. and Hari, P.: Relationship between temperature
 and the seasonal course of photosynthesis in Scots pine at northern timberline and in southern
 boreal zone, Tellus B, 59(3), 542–552, doi:10.1111/j.1600-0889.2007.00262.x, 2007.
- Lasslop, G., Reichstein, M., Papale, D., Richardson, A., Arneth, A., Barr, A., Stoy, P. and Wohlfahrt, G.: Separation of net ecosystem exchange into assimilation and respiration using a light response curve approach: critical issues and global evaluation, Glob. Change Biol., 16,
- 37 187–208, 2010.
- 38 Le Quéré, C., Moriarty, R., Andrew, R. M., Peters, G. P., Ciais, P., Friedlingstein, P., Jones, S.
- 39 D., Sitch, S., Tans, P., Arneth, A., Boden, T. A., Bopp, L., Bozec, Y., Canadell, J. G., Chini, L.
- 40 P., Chevallier, F., Cosca, C. E., Harris, I., Hoppema, M., Houghton, R. A., House, J. I., Jain, A.

- 1 K., Johannessen, T., Kato, E., Keeling, R. F., Kitidis, V., Klein Goldewijk, K., Koven, C.,
- 2 Landa, C. S., Landschützer, P., Lenton, A., Lima, I. D., Marland, G., Mathis, J. T., Metzl, N.,
- 3 Nojiri, Y., Olsen, A., Ono, T., Peng, S., Peters, W., Pfeil, B., Poulter, B., Raupach, M. R.,
- 4 Regnier, P., Rödenbeck, C., Saito, S., Salisbury, J. E., Schuster, U., Schwinger, J., Séférian, R.,
- 5 Segschneider, J., Steinhoff, T., Stocker, B. D., Sutton, A. J., Takahashi, T., Tilbrook, B., van
- 6 der Werf, G. R., Viovy, N., Wang, Y.-P., Wanninkhof, R., Wiltshire, A. and Zeng, N.: Global
- 7 carbon budget 2014, Earth Syst. Sci. Data, 7(1), 47–85, doi:10.5194/essd-7-47-2015, 2015.

Leuning, R.: A critical appraisal of a combined stomatal-photosynthesis model for C3 plants,
Plant Cell Environ., 18(4), 339–355, doi:10.1111/j.1365-3040.1995.tb00370.x, 1995.

- 10 Lin, Y.-S., Medlyn, B. E., Duursma, R. A., Prentice, I. C., Wang, H., Baig, S., Eamus, D., de
- 11 Dios, V. R., Mitchell, P., Ellsworth, D. S., de Beeck, M. O., Wallin, G., Uddling, J., Tarvainen,
- L., Linderson, M.-L., Cernusak, L. A., Nippert, J. B., Ocheltree, T. W., Tissue, D. T., Martin-StPaul, N. K., Rogers, A., Warren, J. M., De Angelis, P., Hikosaka, K., Han, Q., Onoda, Y.,
- Gimeno, T. E., Barton, C. V. M., Bennie, J., Bonal, D., Bosc, A., Löw, M., Macinins-Ng, C.,
- Rey, A., Rowland, L., Setterfield, S. A., Tausz-Posch, S., Zaragoza-Castells, J., Broadmeadow,
- 16 M. S. J., Drake, J. E., Freeman, M., Ghannoum, O., Hutley, L. B., Kelly, J. W., Kikuzawa, K.,
- 17 Kolari, P., Koyama, K., Limousin, J.-M., Meir, P., Lola da Costa, A. C., Mikkelsen, T. N.,
- 18 Salinas, N., Sun, W. and Wingate, L.: Optimal stomatal behaviour around the world, Nat. Clim.
- 19 Change, 5(5), 459–464, doi:10.1038/nclimate2550, 2015.
- 20 Mao, J., Ricciuto, D. M., Thornton, P. E., Warren, J. M., King, A. W., Shi, X., Iversen, C. M.
- and Norby, R. J.: Evaluating the Community Land Model in a pine stand with shading manipulations and δ 13CO2 labeling, Biogeosciences, 13(3), 641–657, doi:10.5194/bg-13-641-
- 23 2016, 2016.
- 24 Martinelli, L. A., Almeida, S., Brown, I. F., Moreira, M. Z., Victoria, R. L., Sternberg, L. S. L.,
- Ferreira, C. A. C. and Thomas, W. W.: Stable carbon isotope ratio of tree leaves, boles and fine litter in a tropical forest in Rondonia, Brazil, Oecologia, 114(2), 170–179, 1998.
- 20 inter in a tropical forest in Kondonia, Brazii, Oecologia, 114(2), 170–179, 1998.
- 27 McDowell, N. G., Allen, C. D. and Marshall, L.: Growth, carbon-isotope discrimination, and
- drought-associated mortality across a Pinus ponderosa elevational transect, Glob. Change Biol.,
- 29 16(1), 399–415, 2010.
- 30 Medvigy, D., Wofsy, S. C., Munger, J. W., Hollinger, D. Y. and Moorcroft, P., R.: Mechanistic
- 31 scaling of ecosystem function and dynamics in space and time: Ecosystem Demography model
- 32 version 2, J. Geophys. Res.-Biogeosciences, 114, G01002, doi:10.1029/2008JG000812, 2009.
- Mitton, J. . and Ferrenberg, S. M.: Mountain pine beetle develops an unprecedented summer
 generation in response to climate warming, Am. Nat., 179(5), 1–9, 2012.
- 35 Monson, R. K., Turnipseed, A. A., Sparks, J. P., Harley, P. C., Scott-Denton, L. E., Sparks, K.
- 36 and Huxman, T. E.: Carbon sequestration in a high-elevation, subalpine forest, Glob. Change
- 37 Biol., 8, 459–478, 2002.
- 38 Monson, R. K., Sparks, J. P., Rosentiel, T. N., Scott-Denton, L. E., Huxman, T. E., Harley, P.
- 39 C., Turnipseed, A. A., Burns, S. P., Backlund, B. and Hu, J.: Climatic influences on net
- 40 ecosystem CO2 exchange during the transition from wintertime carbon source to springtime
- 41 carbon sink in a high-elevation, subalpine forest, Oecologia, 146, 130-147-5-169–2, 2005.

1 Oleson et al.: Technical Description of version 4.5 of the Community Land Model (CLM),

Available

- 2 [online]
- 3 http://www.cesm.ucar.edu/models/cesm1.2/clm/CLM45_Tech_Note.pdf, 2013.
- 4 Parton, W. J., Hanson, P. J., Swanston, C., Torn, M., Trumbore, S. E., Riley, W. and Kelly, R.:
- 5 ForCent model development and testing using the Enriched Background Isotope Study
- 6 experiment, J. Geophys. Res. Biogeosciences, 115(G4), G04001, doi:10.1029/2009JG001193,
- 7 2010.
- Parton, W.J., Schimel, D.S., Cole, C.V., Ojima, D.S., 1987. Analysis of factors controlling soil
 organic-matter levels in great-plains grasslands. Soil Sci. Soc. Am. J. 51 (5).
- 10 Peñuelas, J., Canadell, J. G. and Ogaya, R.: Increased water-use efficiency during the 20th
- 11 century did not translate into enhanced tree growth, Glob. Ecol. Biogeogr., 20(4), 597–608,
- 12 doi:10.1111/j.1466-8238.2010.00608.x, 2011.
- 13 Reichstein, M., Falge, E., Baldocchi, D., Papale, D., Aubinet, M., Berbigier, P., Bernhofer, C.,
- 14 Buchmann, N., Gilmanov, T., Granier, A., Grunwald, T., Havrankova, K., Ilvesniemi, H.,
- 15 Janous, D., Knohl, A., Laurila, T., Lohila, A., Loustau, D., Matteucci, G., Meyers, T., Miglietta,
- 16 F., Ourcival, J. M., Pumpanen, J., Rambal, S., Rotenberg, E., Sanz, M., Tenhunen, J., Seufert,
- 17 G., Vaccari, F., Vesala, T., Yakir, D. and Valentini, R.: On the separation of net ecosystem
- 18 exchange into assimilation and ecosystem respiration: review and improved algorithm, Glob.
- 19 Change Biol., 11(9), 1424–1439, 2005.
- 20 Ricciuto, D. M., Davis, K. J. and Keller, K.: A Bayesian calibration of a simple carbon cycle
- 21 model: The role of observations in estimating and reducing uncertainty, Glob. Biogeochem.
- 22 Cycles, 22, GB2030, doi:10.1029/2006GB002908, 2008.
- 23 Ricciuto, D. M., King, A. W., Dragoni, D. and Post, W. M.: Parameter and prediction 24 uncertainty in an optimized terrestrial carbon cycle model: Effects of constraining variables and record 25 data length, J. Geophys. Res. Biogeosciences, 116(G1), G01033. doi:10.1029/2010JG001400, 2011. 26
- Richardson, A. D., Williams, M., Hollinger, D. Y., Moore, D. J. P., Dail, D. B., Davidson, E.
 A., Scott, N. A., Evans, R. S., Hughes, H., Lee, J. T., Rodrigues, C. and Savage, K.: Estimating
 parameters of a forest ecosystem C model with measurements of stocks and fluxes as joint
 constraints, Oecologia, 164(1), 25–40, 2010.
- Roden, J. S. and Ehleringer, J. R.: Summer precipitation influences the stable oxygen and
 carbon isotopic composition of tree-ring cellulose in Pinus ponderosa, Tree Physiol., 27(4),
 491–501, 2007.
- 34 Rubino, M., Etheridge, D. M., Trudinger, C. M., Allison, C. E., Battle, M. O., Langenfelds, R.
- 35 L., Steele, L. P., Curran, M., Bender, M., White, J. W. C., Jenk, T. M., Blunier, T. and Francey,
- 36 R. J.: A revised 1000 year atmospheric δ 13C-CO2 record from Law Dome and South Pole,
- 37 Antarctica, J. Geophys. Res. Atmospheres, 118(15), 8482–8499, doi:10.1002/jgrd.50668, 2013.
- 38 Sanchez-Rodriguez, J., Perez, P. and Martinez-Carrasco, R.: Photosynthesis, carbohydrate
- 39 levels and chlorophyll fluorescence-estimated intercellular CO2 in water-stressed Casuarina
- 40 equisetifolia Forst. & Forst., Plant Cell Environ., 22(7), 867-873, doi:10.1046/j.1365-
- 41 3040.1999.00447.x, 1999.

from:

- 1 Saurer, M., Siegwolf, R. T. W. and Schweingruber, F. H.: Carbon isotope discrimination
- 2 indicates improving water-use efficiency of trees in northern Eurasia over the last 100 years,
- 3 Glob. Change Biol., 10(12), 2109–2120, doi:10.1111/j.1365-2486.2004.00869.x, 2004.

Saurer, M., Spahni, R., Frank, D. C., Joos, F., Leuenberger, M., Loader, N. J., McCarroll, D.,
Gagen, M., Poulter, B., Siegwolf, R. T. W., Andreu-Hayles, L., Boettger, T., Dorado Liñán, I.,
Fairchild, I. J., Friedrich, M., Gutierrez, E., Haupt, M., Hilasvuori, E., Heinrich, I., Helle, G.,
Grudd, H., Jalkanen, R., Levanič, T., Linderholm, H. W., Robertson, I., Sonninen, E., Treydte,
K., Waterhouse, J. S., Woodley, E. J., Wynn, P. M. and Young, G. H. F.: Spatial variability and
temporal trends in water-use efficiency of European forests, Glob. Change Biol., 20(12), 3700–

- 10 3712, doi:10.1111/gcb.12717, 2014.
- 11 Schaeffer, S. M., Miller, J. B., Vaughn, B. H., White, J. W. C. and Bowling, D. R.: Long-term
- 12 field performance of a tunable diode laser absorption spectrometer for analysis of carbon
- 13 isotopes of CO2 in forest air, Atmospheric Chem. Phys., 8, 5263–5277, 2008.
- 14 Schimel, D. T., Kittel, G. F., Running, S., Monson, R., Turnispeed, A. and Anderson, D.:
- 15 Carbon sequestration studied in western U.S. mountains, Eos Trans AGU, 83(40), 445–449,
- 16 2002.
- 17 Scholze, M., Kaplan, J. O., Knorr, W. and Heimann, M.: Climate and interannual variability of
- 18 the atmosphere-biosphere 13CO2 flux, Geophys. Res. Lett., 30(2), 1097,
- 19 doi:10.1029/2002GL015631, 2003.
- 20 Scott-Denton, L. E., Sparks, K. L. and Monson, R. K.: Spatial and temporal controls of soil 21 respiration rate in a high-elevation, subalpine forest, Soil Biol. Biochem., 35, 525–534, 2003.
- Seibt, U., Rajabi, A., Griffiths, H. and Berry, J. A.: Carbon isotopes and water use efficiency:
 sense and sensitivity, Oecologia, 155(3), 441–454, 2008.
- 24 Sellers, P. J., Randall, D. A., Collatz, G. J., Berry, J. A., Field, C. B., Dazlich, D. A., Zhang, C.,
- 25 Collelo, G. D. and Bounoua, L.: A revised land surface parameterization (SiB2) for atmospheric
- 26 GCMs. Part I: Model formulation, J. Clim., 9(4), 676–705, 1996.
- Thornton, P. E. and Rosenbloom, N. A.: Ecosystem model spin-up: Estimating steady state conditions in a coupled terrestrial carbon and nitrogen cycle model, Ecol. Model., 189(1–2), 25–48, doi:10.1016/j.ecolmodel.2005.04.008, 2005.
- 30 Thornton, P. E., Law, B. E., Gholz, H. L., Clark, K. L., Falge, E., Ellsworth, D. S., Golstein, A.
- H., Monson, R. K., Hollinger, D., Falk, M., Chen, J. and Sparks, J. P.: Modeling and measuring
 the effects of disturbance history and climate on carbon and water budgets in evergreen
- needleleaf forests, Agric. For. Meteorol., 113(1–4), 185–222, 2002.
- Thornton, P. E., Lamarque, J.-F., Rosenbloom, N. A. and Mahowald, N. M.: Influence of carbon-nitrogen cycle coupling on land model response to CO2 fertilization and climate variability, Glob. Biogeochem. Cycles, 21, GB4018, doi:10.1029/2006GB002868, 2007.
- Tomaszewski, T. and Sievering, H.: Canopy uptake of atmospheric N deposition at a conifer
 forest: Part II- response of chlorophyll fluorescence and gas exchange parameters, Tellus B,
 59(3) [online] Available from: http://www.tellusb.net/index.php/tellusb/article/view/17021
 (A assessed 28 May 2015) 2007
- 40 (Accessed 28 May 2015), 2007.

- 1 Trolier, M., White, J. W. C., Tans, P. P., Masarie, K. A. and Gemery, P. A.: Monitoring the
- 2 isotopic composition of atmospheric CO2: Measurements from the NOAA Global Air
- 3 Sampling Network, J. Geophys. Res.-Atmospheres, 101(D20), 25,897-25,916, 1996.

4 van der Velde, I. R., Miller, J. B., Schaefer, K., Masarie, K. A., Denning, S., White, J. W. C.,
5 Tans, P. P., Krol, M. C. and Peters, W.: Biosphere model simulations of interannual variability

- 6 in terrestrial 13C/12C exchange, Glob. Biogeochem. Cycles, 27(3), 637–649,
- 7 doi:10.1002/gbc.20048, 2013.

8 Wehr, R. and Saleska, S. R.: An improved isotopic method for partitioning net ecosystem–
9 atmosphere CO2 exchange, Agric. For. Meteorol., 214–215, 515–531,
10 doi:10.1016/j.agrformet.2015.09.009, 2015.

11 White, J. W. C., Vaughn, B. H., Michel, S. E., University of Colorado and Institute of Arctic 12 and Alpine Research (INSTAAR): Stable Isotopic Composition of Atmospheric Carbon

13 Dioxide (13C and 18O) from the NOAA ESRL Carbon Cycle Cooperative Global Air Sampling

- 14 Network, 1990-2014, Version: 2015-10-26, Path:
- 15 ftp://aftp.cmdl.noaa.gov/data/trace_gases/co2c13/flask/, 2015.

White et al.: Parameterization and Sensitivity Analysis of the Biome-BGC Terrestrial
Ecosystem Model: Net Primary Production Controls, [online] Available from:
http://secure.ntsg.umt.edu/publications/2000/WTRN00/White_2000.pdf, 2000.

- 19 Wingate, L., Ogee, J., Burlett, R., Bosc, A., Devaux, M., Grace, J., Loustau, D. and Gessler,
- 20 A.: Photosynthetic carbon isotope discrimination and its relationship to the carbon isotope
- signals of stem, soil and ecosystem respiration, New Phytol., 188(2), 576–589, 2010.
- 22 Yohe, G.W., R.D. Lasco, Q.K. Ahmad, N.W. Arnell, S.J. Cohen, C. Hope, A.C. Janetos and
- R.T. Perez, 2007: Perspectives on climate change and sustainability. Climate Change 2007:
 Impacts, Adaptation and Vulnerability. Contribution of Working Group II to the Fourth
- 25 Assessment Report of the Intergovernmental Panel on Climate Change, M.L. Parry, O.F.
- 26 Canziani, J.P. Palutikof, P.J. van der Linden and C.E. Hanson, Eds., Cambridge University
- 27 Press, Cambridge, UK, Section 20.6, pgs. 821-825
- Zaehle, S., Medlyn, B. E., De Kauwe, M. G., Walker, A. P., Dietze, M. C., Hickler, T., Luo,
 Y., Wang, Y.-P., El-Masri, B., Thornton, P., Jain, A., Wang, S., Warlind, D., Weng, E., Parton,
 W., Iversen, C. M., Gallet-Budynek, A., McCarthy, H., Finzi, A., Hanson, P. J., Prentice, I. C.,
- 31 Oren, R. and Norby, R. J.: Evaluation of 11 terrestrial carbon-nitrogen cycle models against
- 32 observations from two temperate Free-Air CO2 Enrichment studies, New Phytol., 202(3), 803–
- 33 822, doi:10.1111/nph.12697, 2014.

Zarter, C. R., Demmig-Adams, B., Ebbert, V., Adamska, I. and Adams, W. W.: Photosynthetic
capacity and light harvesting efficiency during the winter-to-spring transition in subalpine
conifers, New Phytol., 172(2), 283–292, doi:10.1111/j.1469-8137.2006.01816.x, 2006.

- Zeng, X.: Global Vegetation Root Distribution for Land Modeling, J. Hydrometeorol., 2(5),
 525–530, doi:10.1175/1525-7541(2001)002<0525:GVRDFL>2.0.CO;2, 2001.
- 39 Zeng, X. and Decker, M.: Improving the Numerical Solution of Soil Moisture–Based Richards
- 40 Equation for Land Models with a Deep or Shallow Water Table, J. Hydrometeorol., 10(1), 308–
- 41 319, doi:10.1175/2008JHM1011.1, 2009.

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PFT

Plant functional type

Unit or Unit Symbol Symbol Description dimensionless Fractionation factor (R_{a/} R_{GPP}) α Soil water stress parameter (BTRAN) dimensionless β_t photosynthetic carbon isotope discrimination ‰ Δ_{canopy} $^{13}C/^{12}C$ isotope composition (relative to VPDB) $\delta^{13}C$ ‰ δ^{13} C of atmospheric CO₂ ‰ δ_{atm} δ^{13} C of ecosystem respiration ‰ δ_{ER} δ^{13} C of net photosynthetic assimilation ‰ δ_{GPP} CO₂ compensation point Pa Γ_* Ac Enzyme-limiting rate of photosynthetic assimilation µmol m⁻² s⁻¹ Light-limiting rate of photosynthetic assimilation Ai μ mol m⁻² s⁻¹ μ mol m⁻² s⁻¹ Product-limiting rate of photosynthetic assimilation Ap umol m⁻² s⁻¹ net photosynthetic assimilation An μ mol m⁻² s⁻¹ **Resp**_d Leaf-level respiration Specific activity of Rubisco at 25°C µmol g⁻¹ Rubisco s⁻¹ a_{R25} Minimum stomatal conductance µmol m⁻² s⁻¹ b $gC m^{-2} s^{-1}$ Actual carbon allocated to biomass (N-limited) CF_{alloc} $gC m^{-2} s^{-1}$ Maximum carbon available for allocation to biomass CF_{av_alloc} $gC m^{-2} s^{-1}$ Potential gross primary production (non N-limited) CF_{GPPpot} Atmospheric CO₂ partial pressure Pa C_a Leaf intercellular CO₂ partial pressure Pa C_i c_i^* Leaf intracellular CO₂ partial pressure, (N-limited) Pa Leaf surface CO₂ partial pressure Pa Cs Saturation vapor pressure Pa e_l Water vapor pressure at leaf surface Pa e_s µmol m⁻² s⁻¹ **Ecosystem Transpiration** E_T umol m⁻² s⁻¹ ER Ecosystem respiration µmol m⁻² s⁻¹ GPP Gross primary productivity (photosynthesis) gN Rubisco g⁻¹ N Fraction of leaf nitrogen within Rubisco FLNR g Rubisco g⁻¹ N Rubisco F_{NR} Total Rubisco mass per nitrogen mass within Rubisco fdf V_{cmax} scaling factor dimensionless Nitrogen photosynthetic downregulation factor dimensionless f_{dreg} µmol m⁻² s⁻¹ Leaf boundary layer conductance gb μ mol m⁻² s⁻¹ Leaf stomatal conductance gs Pa Pa⁻¹ Leaf surface relative humidity h_{s} K_c Pa CO₂ Michaelis-Menten constant Pa K_o O2 Michaelis-Menten constant $W m^{-2}$ Latent heat flux LE Stomatal slope (Ball Berry conductance model) dimensionless т gN m⁻² leaf area Leaf nitrogen concentration Na NEE Net ecosystem exchange μ mol m⁻² s⁻¹ NPP umol m⁻² s⁻¹ Net primary production O₂ atmospheric partial pressure Pa O_i

N/A

Patm	Atmospheric pressure	Ра
Ra	Isotopic ratio of canopy air	$^{13}C/^{12}C$
R _{GPP}	Isotopic ratio of net photosynthetic assimilation	$^{13}C/^{12}C$
R _{VPDB}	Isotopic ratio of Vienna Pee Dee Belemnite standard	$^{13}C/^{12}C$
r	Fraction of roots (for β_t)	dimensionless
V _{cmax25}	Maximum carboxylation rate at 25°C	µmol m ⁻² s ⁻¹
V _{cmax}	Maximum carboxylation rate at leaf temperature	μ mol m ⁻² s ⁻¹
VPD	Vapor pressure deficit	Pa
W	Plant wilting factor (for β_t)	dimensionless
WUE	Water use efficiency, ground area basis	µmol C mol H ₂ 0 ⁻¹
iWUE	Intrinsic water use efficiency, leaf area basis	μ mol C mol H ₂ 0 ⁻¹

1 Table 2. CLM 4.5 model formulation description based upon timing of nitrogen limitation.

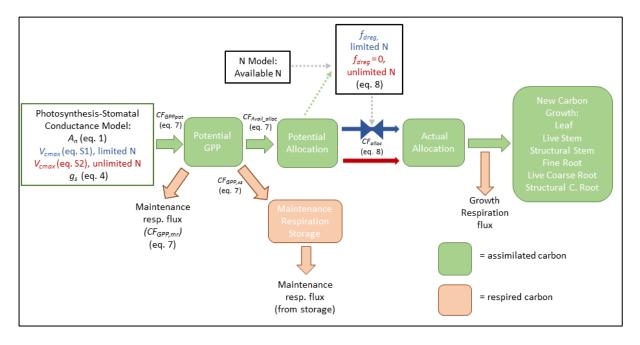
2 3 Pre-photosynthetic and post-photosynthetic nitrogen limitation are achieved through V_{cmax25}

Formulation	Pre-	Post-	Impacts c _{i*} /c _a	Impacts g_s	
	Photosynthetic	Photosynthetic	& discrimination	and A_n	
	Nitrogen	Nitrogen			
	Limitation	Limitation			
Limited	Yes (weak)	Yes, $f_{dreg} > 0$	Yes	No	
nitrogen					
(default)					
Unlimited	Yes (strong)	No, $f_{dreg} = 0$	Yes	Yes	
nitrogen					
No	Yes (weak)	Yes, $f_{dreg} > 0$	No	No	
downregulation					
discrimination					

calibration (equation 17) and f_{dreg} (equation 8) respectively.

Parameter	Description	Value	Units
froot_leaf	new fine root C per new leaf C	0.5	gC gC ⁻¹
froot_cn	fine root (C:N)	55	gC gN ⁻¹
leaf_long	leaf longevity	5	years
leaf_cn	leaf (C:N)	50	gC gN ⁻¹
lflitcn	leaf litter (C:N)	100	gC gN ⁻¹
slatop	specific leaf area (top canopy)	0.007	$m^2 gC^{-1}$
stem_leaf	new stem C per new leaf C	2	gC gC ⁻¹
mp	stomatal slope	9	
croot_stem	coarse root: stem allocation	0.3	gC gC ⁻¹
deadwood_cn	dead wood (C:N)	500	gC gN ⁻¹
livewood_cn	live wood (C:N)	50	gC gN ⁻¹
flnr	fraction of leaf nitrogen within	0.0509	gN gN ⁻¹
	Rubisco enzyme		
decomp_depth_e_folding	controls soil decomposition rate with depth	20	m

1 Table 3. CLM 4.5 key parameter values for all model formulations
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3 Figure 1: A simplified representation within CLM 4.5 of assimilation and allocation of

4 carbon for conifer species. Colored boxes and arrows represent carbon pools and carbon

5 fluxes respectively. Clear background boxes represent CLM sub-models. The N-limitation

6 is determined by required N availability to meet demand from C:N ratio based on CF_{Avail_alloc}.

7 The blue and red text and arrows represent the limited and unlimited nitrogen formulations

8 respectively. The no-downregulation discrimination formulation is exactly the same as the

9 limited N formulation in this schematic.

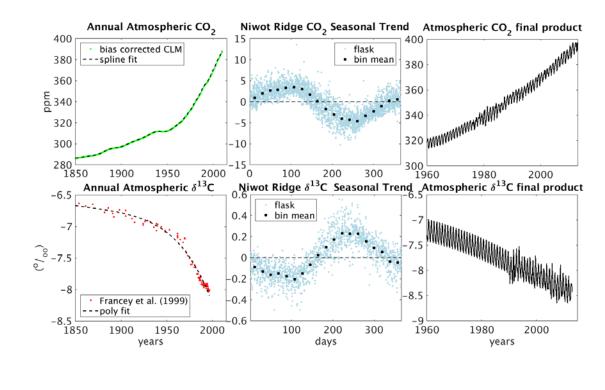
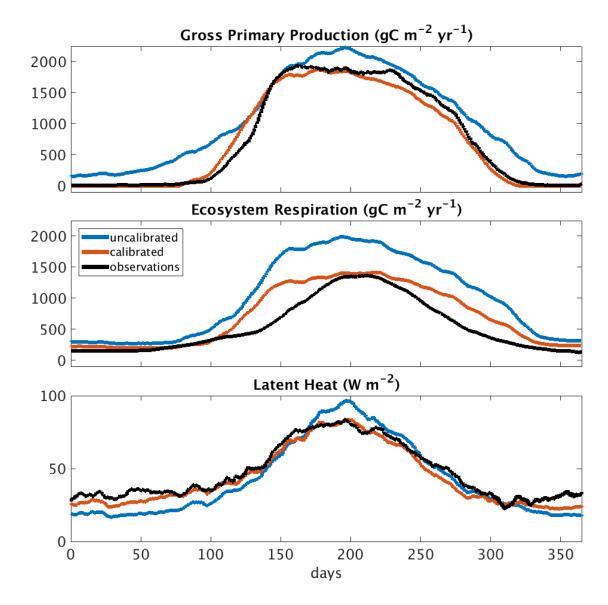


Figure 2. Niwot Ridge synthetic data product for atmospheric CO₂ concentration (c_a) (top row) and δ^{13} C of CO₂ (δ_{atm}) (bottom row). The final time series (right column) was used as a boundary condition for CLM, and created by combining the annual trends reported by Francey et al. (1999) adjusted for Niwot Ridge (left column) with the mean seasonal cycles measured at Niwot Ridge (middle column).

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Figure 3. Seasonal averages (1999-2013) of simulated and observed land-atmosphere fluxes for A) gross primary production (GPP) B) ecosystem respiration (ER) and C) latent heat (LE) for the *limited nitrogen* simulation. The 'observations' are taken from the Ameriflux L2 processed eddy covariance flux tower data, partitioned into GPP and ER using the method of Reichstein et al. (2005). The *uncalibrated* simulation represents the CLM simulation without V_{cmax} scaling and the *calibrated* simulation represents the CLM run using the V_{cmax} scaling approach.

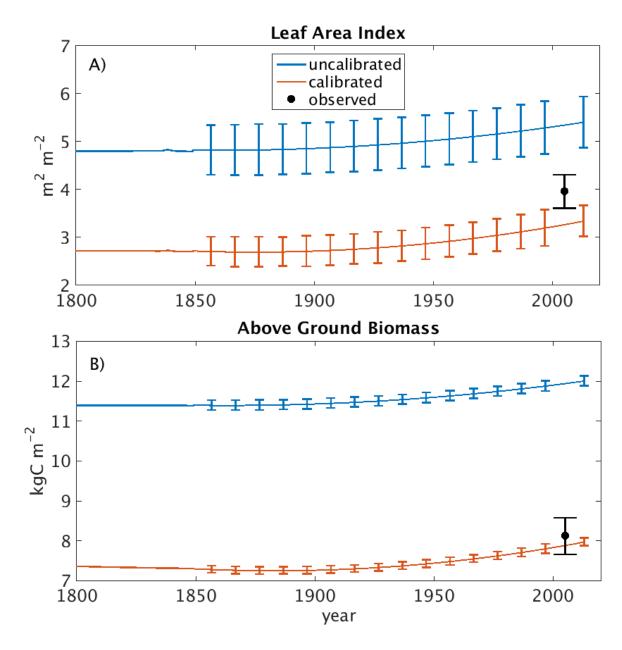


Figure 4. Simulation of A) leaf area index and B) above ground biomass for both uncalibrated
and calibrated (V_{cmax} downscaled, *limited nitrogen*) simulation. Observations are from
Bradford et al. (2008) with uncertainty bars representing standard error. Uncertainty bars on
simulated runs represent 95% confidence of biomass variation as a result of cycling the site
level meteorology observations.

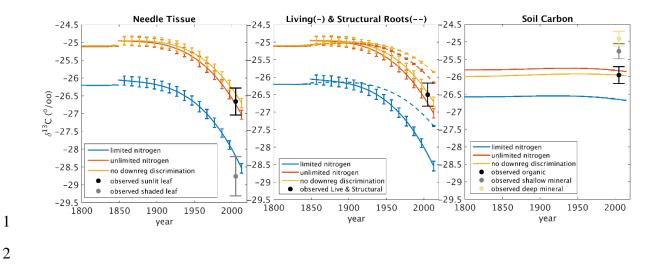


Figure 5. Simulation of δ^{13} C of bulk needle tissue, bulk roots and bulk soil carbon. A description of model formulations are provided in Table (2). Uncertainty bars for simulations represent 95% confidence intervals of δ^{13} C variation as a result of cycling the site level meteorology observations. The observed values are from Schaeffer et al. (2008) with uncertainty bars representing standard error. Solid lines and dashed lines in middle panel represent living roots and structural roots respectively.

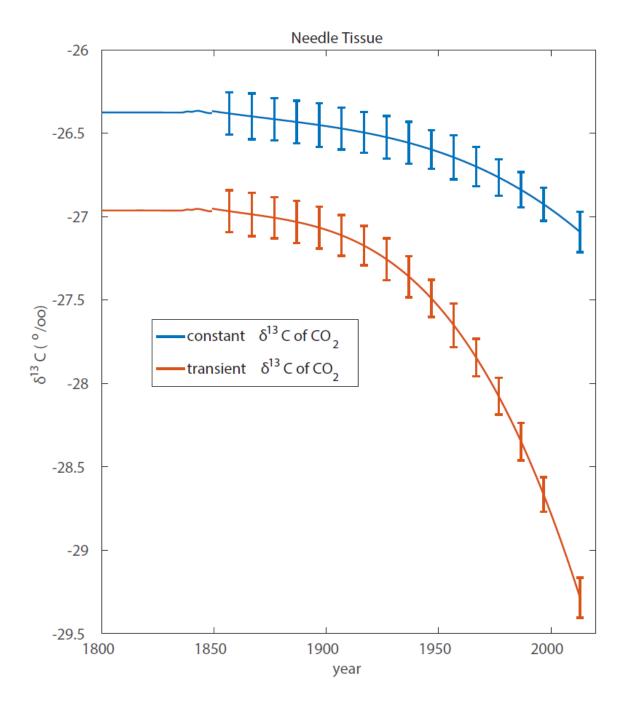


Figure 6. Simulation of δ^{13} C of needle tissue using the *limited nitrogen* (default) CLM run. In the *constant* δ^{13} C of CO2 (δ_{atm}) simulation the model boundary condition was -6 ‰, whereas the *transient* δ_{atm} simulation varied over time (Figure 2).

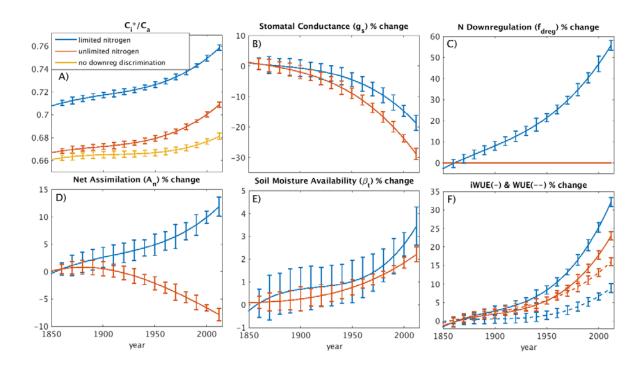


Figure 7. Diagnostic model variables that explain the discrimination trends (Figure 5) for the three model formulations as described in Table (2) for A) c_i^*/c_a , B) g_s , C) f_{dreg} , D) A_n , E) β_t , and F) the water use efficiency (WUE) and intrinsic water use efficiency (iWUE). Where the no 6 downregulation discrimination simulation is not shown, it was identical to the limited nitrogen 7 simulation. Uncertainty bars represent 95 % confidence intervals of diagnostic variable 8 variation as a result of cycling the site level meteorology observations. The dashed lines 9 represent WUE and the solid lines represent iWUE in panel F.

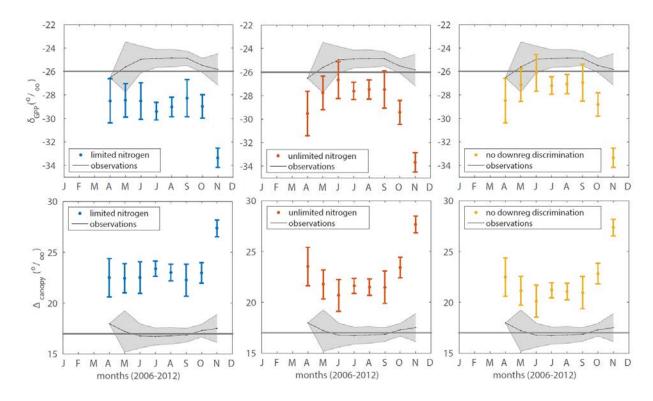


Figure 8. The seasonal pattern of photosynthetic discrimination as shown through δ_{GPP} (top row) and Δ_{canopy} (bottom row). Uncertainty bars represent 95% confidence bounds of simulated monthly average values from 2006-2012. Gray-shaded observation bounds represent 95% confidence intervals of 'observed' monthly average values based upon isotopic mixing model using Reichstein et al. (2005) partitioning of net ecosystem exchange flux described by (Bowling et al. 2014). The horizontal lines at δ^{13} C of -26 ‰ (top row) and 17 ‰ (bottom row) are included for reference.

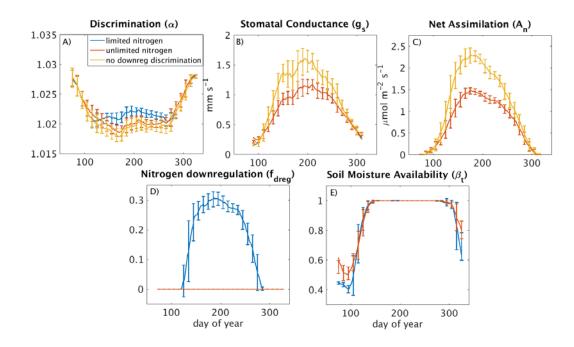
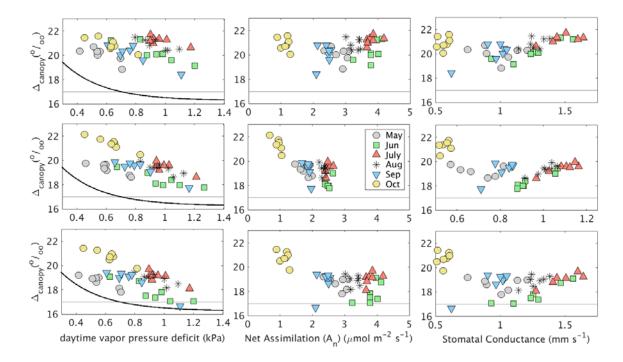


Figure 9. The seasonal pattern of discrimination (panel A) and diagnostic variables that explain the discrimination pattern in Figure (8). The individual tiles provide behavior from days 75-325 for A) α , B) g_s , C) A_n , D) f_{dreg} , and E) β_t . Where the *no downregulation discrimination* model simulation is not shown, it is identical to the *limited nitrogen* simulation. Uncertainty bars represent 95 % confidence intervals of inter-annual variation from 2006-2012.

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- 9



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Figure 10. Relationship between monthly average photosynthetic discrimination and monthly average vapor pressure deficit (1st column), A_n (2nd column) and g_s (3rd column) from 2006-2012. The rows represent the *limited nitrogen* (row 1), *unlimited nitrogen* (row 2), and *no downregulation discrimination* (row 3) simulations. The black line in the 1st column is based on exponential fitted line from observed relationship at Niwot Ridge (Bowling et al. 2014). The horizontal lines represent δ^{13} C of 17 ‰ and are included for reference.