

Interactive comment on “A novel acclimative biogeochemical model and its implementation to the southern North Sea” by Onur Kerimoglu et al.

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We would like to thank all three anonymous referees for their constructive comments and criticism. The referees pointed to the need for a more representative title, improved clarity in text and figures, including a separate discussion section, additional comparison of some model estimates with observations and literature beyond what is already presented, and additional analyses for the justification of some conclusions, in several instances harmoniously. While we agree with most comments as detailed below, we believe some of the suggested extensions require more elaborate analysis than can be included here. To recapitulate, the objectives of the current study are; *i*) gaining insight into the behavior of an acclimative model in a 3D framework for the first time to the best of our knowledge; *ii*) evaluating the skill of the new model system at various spa-

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tial and temporal scales, which requires consideration of an extremely diverse array of observation sets. These objectives lead to a wide scope, and generation of a number of research questions that are better treated separately.

We have a remark relevant to all three referees, therefore placed here: upon further examination of our model simulation presented in our original manuscript, we found out that the performance of the hydrodynamical model could be improved by not specifying the momentum fluxes at the open boundaries, and a re-parameterization of the bottom friction. We also realized that, due to a wrong configuration file, the atmospheric nitrogen deposition was not correctly registered during the model initialization in the simulation presented in the original manuscript. A new simulation run for the entire simulation period with the improvements in hydrodynamical model and inclusion of atmospheric nitrogen deposition results in better model performance overall, although not qualitatively affecting our conclusions based on the original manuscript. In a revised version of the manuscript, we thus would like to present the results obtained with this new simulation run.

Detailed response to Referee #1

Title: I have two problems with the title: 1) as the acclimatisation scheme has been published previously, I would advise against using the word 'novel'; 2) 'implementation' suggests the presentation and discussion of how the acclimatisation method is implemented in the biogeochemical model, which is included in the manuscript, but not related to the application to the SNS. So I would suggest reformulating to, eg., The application of an acclimative biogeochemical model to the southern North Sea.

We would like to thank the referee for this careful observation and thoughtful suggestions. Although the referee is right that the acclimatisation scheme of phytoplankton growth was published previously in a 0D context, the full 3D setup including many model variables is described in this manuscript the first time. However the real 'novelty' of this study is embedding the acclimative phytoplankton growth model in a 3D

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coupled physical-biogeochemical framework, considering that similarly complex models have been previously studied in much simpler contexts. We agree that the previous title was not reflecting this, so we will change it as: 'A novel 3D coupled physical-biogeochemical model resolving phytoplankton acclimation and its application to the southern North Sea'.

Structure: The authors should introduce a separate discussion section.

We will include a separate discussion section as suggested by the referee.

Comparison: In comparing model results with observations, the text is too qualitative, using expressions such as 'compare well', 'reasonable match', and so on, without defining what these are. This should be tightened up and quantified throughout. The same holds for comparison with previous work in the literature: a small subset of earlier biogeochemical modelling work is referenced, and it is suggested that the current model performs better, but without providing the evidence and quantifying the differences. It is also unclear why these studies were selected, and not others.

We will use more precise formulations for the evaluation of the model. When referring to literature, we attempted to refer to all recent work on the modelling of a relevant model domain. We will check the literature again for potentially omitted work. A detailed and precise comparison of the performance of our model with other models is not in the scope of our study: as was already expressed in the manuscript, such a comparison requires a dedicated effort with standardized benchmarking data and tools. We will stress the need for regional model data bases that will facilitate such model intercomparisons .

Logic/interpretation: The logic and interpretation tend to be hand-waving at best, flawed in some cases, and don't always consider multiple options. Examples are listed in the details section below. This needs to be improved. Separating the discussion will help.

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We will try to improve the interpretations along the comments of the referees, and include a separate discussion section.

Is it really 'better'? The authors state at several points in the paper that their acclimative phytoplankton growth method is better what's used in more traditional biogeochemical models. However, unfortunately, they fail to provide any proof of this. In the very least, there sWe will increase the font sizes where necessary.hould be an in-depth, quantified discussion comparing the current results with those of a suitably wide range of 'traditional' models.

As explained above, we did not intend to claim that our model performs better than the others, and we had expressed clearly in the manuscript that such would require some dedicated effort and is out of the scope of the current work. We will nevertheless try to formulate more precise expressions for evaluating the model performance.

I get the impression from the manuscript that the 'novel' biogeochemical model was constructed by stripping an existing 'traditional' biogeochemical model of the relevant parts, and replacing these with the acclimative methods. If this is indeed the case, the authors would strengthen the manuscript immensely by providing and discussing a comparison with a similar run with the earlier model version.

Referee #3 also suggested a comparison with a non-acclimative model version. We did not start from a traditional model and upgrade to an acclimative one, so we do not have such an earlier model version. However, it is possible to turn-off the acclimative features of the model so we will perform a model run and address this relevant issue in the revised version.

The authors will also need to discuss the following in a systematic way. More traditional biogeochemical models may lack (to various extents depending on the model) the full suite of acclimatisation as presented here, but they make up for that at least to some extent by representing several types of phytoplankton. This allows for spatial and temporal changes/patterns in phytoplankton composition. One could argue that the new

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model reflects this with one type with a range of traits, but it presumably has more flexibility in changing these traits over time for the same biomass than could happen in nature (one type of plankton can not change into another).

The plankton functional type (PFT) models might make up for the unrepresented acclimation processes to some extent, but we are not aware of any study which tested this idea rigorously. We do not really see why our model is presumably more flexible than reality: consider the case of the competition of two species, where the first species, dominant at the beginning, is being gradually replaced by the second until it completely vanishes by the end of the experiment. Throughout this plausible experiment, representation of the traits may completely change, possibly without considerable changes in total biomass. A hypothetically perfect simulation of this experiment by our model in terms of biomass and the average trait representation in the system, might seem to suggest that one plankton type changed into another, which is however just an interpretation and not a limitation inherent to our approach. In conclusion, a comparison of the intracellular Chl:C:N:P ratios observed, e.g., in chemostat experiments, and estimations by a PFT model and our acclimation model might provide valuable insights in this direction, however such a comparison would be beyond the scope of the current study. The relevant discussion of the changes in traits in reality vs. as represented by our model provided in the original manuscript (in P.19, L.13-24) is sufficient in our assessment.

Also, the authors are suggesting that they plan the inclusion of additional phytoplankton types. That would require curtailing the ranges of acclimatisation. Would that throw the baby out with the bath water, or have they already done so and would this be an attempt to get it back in?

The plastic response simulated by our model can already be seen at the species level as shown by Wirtz and Kerimoglu (2016). Introduction of further plankton groups for resolving other ecophysiological traits such as silicate limitation and edibility will also allow taxa-specific parameterization of resource utilization traits, which is expected to

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further improve the representation spatio-temporal distribution of the overall cellular composition of the phytoplankton.

Figures: Not all of the figures are clearly readable, and some information is missing. Fig 3: I suspect that the colour scale is truncated, both at the high and low end, resulting in artificial saturation of the figure. This must be addressed. Also this figure would benefit from using a wider range of colours.

The scale was truncated (from the lower range) on purpose, as doing so helps emphasizing the salinity front. We will mention this in the caption of the figure. We will also use the ('viridis') color scheme used in other contour plots and discrete color levels (as well as in other contour plots), as this enables comparing the location of certain value ranges in the measured and simulated data.

Fig 4. S and T are partly obscured by the dots, the cursive eta and n are barely visible on my printout

We will increase the vertical spacing on Taylor diagrams and use larger font size in scatter plots.

Fig 5-7. These are all too small. I can hardly read the axis legends and legends. Names on maps are cluttered.

For the plot size, we used the standard width required by Biogeosciences. However we will improve the plots by: 1) using larger fonts, 2) using less intrusive markers for observations, 3) reworking the maps, such that names do not overlap.

Fig 8. Does ICES store chlorophyll? If so it would help if this were included.

Referee #2 raised the same question. Our choice to exclude chlorophyll was based on the fact that the spatio-temporal representativeness of the chlorophyll measurements was much inferior in comparison to that of DIN and DIP data, and consideration that the performance of the model with regard to chlorophyll is separately evaluated based on the satellite and Scanfish measurements (Fig. 9 and Fig. 13).

Fig 9. Re-plot in colour. I can't work out the route taken from the cruise track figure.

We will re-plot the Figure in color.

Fig 10, 11, 14. The black contours are partially obscured by the dark blue.

We found out that using light-gray contour lines produce better results. We will renew these figures accordingly.

Fig 11. I understand that these are surface values. Please also provide the bottom values.

The winter concentrations of DIN and DIP at the bottom are almost identical to the values at the surface and this will be mentioned in the revised manuscript.

Fig. 12. The colour scale is symmetrical around the centre, making it impossible to distinguish spring and autumn values. Please re-plot.

Objective of coloring in this figure was mainly to distinguish winter, but it is indeed possible that distinguishing spring and autumn values might be of interest, so we will re-plot with an asymmetric color scheme.

Grammar and language: Please check the grammar. There are quite a few anomalies that even a grammar checker would pick up (I'm not going to list them all). Also use past tense to describe the results throughout.

We will go through the text and improve the language.

Further detailed comments

We provide more detailed answers to the following selection of comments. We will address each of the other minor issues in the revised version.

p. 8 - l. 6. Other explanations could be that: 1) the river-runoff is too high, or 2) the set of open boundary conditions used for the hydrodynamics and dissolved components restricts the amount of flushing, leading to an accumulation of fresh water, nutrients,

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etc. Or a combination. Please discuss.

We thank the referee for this insightful comment. As explained at the beginning of this response letter, we found out not specifying the momentum fluxes led to a better representation of the tidal dynamics, hence, the residual currents and as a consequence, spatial distribution of salinity and other transported variables.

p. 9 - Fig 4. There seems to be a 1:1 relationship, but with an anomaly on top. Does the anomaly in T correspond to the low values of S that bend away from the 1:1 line? Does this cluster represent a particular geographic area (front?)? Or a particular event/year (2010?)?

We will include a 1:1 line in the scatter plots. In the new model run, that deviation from the 1:1 relationship is largely resolved, but we will investigate the source of such deviations with respect to certain geographic regions and/or events.

p. 15 - l. 29-34. This seems a ridiculous over-interpretation of a potential contribution by estuarine overturning circulation. There's no evidence of overall higher nutrient concs in bottom waters (fig 8). Providing bottom values in fig 11 will likely support this. What's happening is that the nutrient-rich riverine waters enter/mix with the coastal waters, which are trapped by the coastal density(salinity) front.

The dynamic effect of horizontal density gradients on residual transport of particulate (organic) matter, which we refer to, is a known feature of shallow, tidal seas, as elaborated in detail with observational and modelling approaches in the cited literature (P.15, l. 31-32). The coastal gradients as a result of accumulation of organic matter, is found along the whole SNS coast but the referee is right, that near the estuaries and the regions of freshwater influence (ROFIs), nutrient concentrations are determined more by the riverine nutrient input. The updated model version, which produces weaker stratification than the previous version, hence weaker differences between the surface-bottom waters overall also suggests a more pronounced importance of the estuaries for the nutrient conditions, therefore we will revise the interpretation of gradients in the

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light of the explanation by the referee.

p. 17 - l. 9-12. This is an unfair comparison. The observations in fig 5 are instantaneous, whereas the satellite composites are 3-monthly averaged. It's obvious that the satellite values presented in this way should be lower! This statement requires a proper comparison.

Both other referees raised relevant questions. This sentence was actually based on such a point-to-point comparison of the raw satellite (not-averaged) and station data (Fig. R1), which reveals a bias in the form of low concentrations by the satellite data although we recognize that the sentence referred by the referee was not formulated clear enough. In the new version of the simulation, this overestimation problem is largely resolved, therefore such statements will not be necessary in the revised version of the manuscript.

Detailed response to Referee #2

The time series comparison of chlorophyll results with in-situ data suggest that chlorophyll concentrations are systematically over-predicted in spring at many monitoring stations. The validation plots with in-situ data are only presented for other model variables and not for chlorophyll.

While the chlorophyll concentrations are over-predicted consistently for every year for three out of five stations at the coasts of Netherlands (Fig.6), this is the case in only one out of five stations in the German Bight (Fig.5), while in two others (S. Amrum, Norderelbe) the bias is even negative, so we do not agree that the over-prediction problem was 'systematic'. However, with the updated model run (please see the beginning of the response letter), the simulated chlorophyll concentrations match to the observations better in almost all problematic stations. The Referee #1 also pointed to the need to include the chlorophyll in the validation plots. However, as we stated in our response there, 'our choice to exclude chlorophyll was based on the fact that the

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spatio-temporal representativeness of the chlorophyll measurements was much inferior in comparison to that of DIN and DIP data, and because the performance with regard to chlorophyll is separately evaluated based on the satellite and Scanfish measurements (Fig. 9 and Fig. 13)'.

The validation with satellite data shows also that chlorophyll is systematically over-predicted throughout the model domain during spring. The authors conclude that the satellite data are wrong. This is not supported by any comparison with in-situ data, but the above comparison with time series suggests that the model over-predicts chlorophyll in spring.

This is an issue mentioned by both other referees. As we responded there, a comparison of the raw satellite data with station data clearly reveals a bias (see Fig.R1). On the other hand, with the new model run the overestimation problem during spring is largely resolved, such a critical discussion of the deviations will not be necessary in the new revision.

It is unclear what trait effects are included in the model, which are not included in existing models. On page 4, line 20 a few traits are listed (very brief) but in the discussion at page 15 and 17 other effects are mentioned, such as effects on chlorophyll to carbon ratio and sinking rates.

Such trait effects, along with other process descriptions that are not included in traditional models were described in appendix A1 (e.g., effect of nutrient state of the cells on sinking was described in in Page 21, lines 8-12). We will mention the important processes also in the main text, with necessary links and references.

A critical discussion of the novel aspects of the phytoplankton model is lacking. For example: are the chlorophyll to carbon ratios in spring in a realistic range for spring conditions? How do sinking rates change over the year and how does that relate to observations?

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The agreement between the coastal pattern displayed by the chlorophyll to carbon ratios estimated by the model and that reported by Alvarez-Fernandez and Riegman (2014) was pointed out in the manuscript (page 17, lines 15-16), but a comparison of the actual values was indeed missing. The Chl:C ratios ranging between 0.01-0.1 gChl/gC at the coastal stations and 0.002-0.02 gChl/gC at the off-shore stations reported by Alvarez-Fernandez and Riegman (2014) envelope our estimated seasonal average values of 0.045 and 0.015 within the respective regions. We will mention this in the text. For the sinking rates of phytoplankton in the southern North Sea as well, we will search the literature for relevant information.

There is no validation of the light climate (as K_d) included in the manuscript. This would be helpful in explaining differences between the model and observed data.

The largest source of errors for the representation of the light climate in our study is already known: shading by suspended particulate matter (SPM) was incorporated as a climatological model forcing, which has a rather coarse horizontal resolution (about 20km), and does not represent the vertical heterogeneities as well as inter-annual and sub-daily variations. Therefore, a through evaluation of the light field should encompass various spatio-temporal scales, which is therefore better treated in a separate study, which may be dedicated to improving the representation of the light climate. However, we recognize that the issue is relevant for the spatial distribution of the chlorophyll concentrations and chlorophyll:carbon ratios, which is one of the core findings of this study, therefore we will investigate the possibilities to gain some further insight.

Specific comments:

Page 4, line 20 and equation 1. This part needs to give a complete list of acclimation effects included in the model. It should also describe in words how it works. Like it is written on page 17: “sinking speed of algae in MAECS is inversely related to nutrient quota of cells.”. So there are not only effects of nutrients on growth rate (as suggested by eq 1) but also on other aspects. And there are effects of light on chlorophyll to

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carbon ratio. And does a flexibility constant represent?

A more detailed description of the model was provided in Appendix A1 (e.g., regarding the effects of nutrient quota of cells on the sinking rates), and a full description of the phytoplankton growth model can be found in Wirtz and Kerimoglu (2016, e.g., regarding the effects of light on chlorophyll to carbon ratio). However, as stated above, we will extend the relevant section in the main text with appropriate references and links.

Page 5, caption of Figure 2 mentions Fe-P:P adsorbed in iron-phosphorus complexes. I don't see this in the figure. Or you should refer to bAP in the caption.

Indeed b-AP instead of Fe-P should be referred to in the caption. We will correct this in the revised manuscript.

Page 7: Could you please clarify in more detail the source of the ESA-CCI dataset. Is there a website where these data can be downloaded and where we can find validation reports of this dataset?

Information on the source of this data set was provided in the text (Page 4, lines 5-8) but we will include the requested additional information in the revised version.

Figure 4: the T and S are too small to read and overlap with the dots.

We will use larger fonts and increase the vertical spacing of the labels and the markers.

Page 9, line 10. I don't see that the classical seasonal pattern of phosphorus is entirely reversed in the data. This may be partly due to the small size of the figure. But also it may be that phosphorus concentrations in shallow muddy areas of the Wadden Sea are higher during the summer than during winter due to release of phosphorus from anoxic sediments. This does not reverse the seasonal pattern, because there is a classical drop in phosphorus concentrations during the spring bloom

The observed summer-high instead of the typical winter-high in the S. Amrum station was the basis for the 'reverse seasonal pattern', but we agree that this was overly

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vague. We will clarify this.

Page 9: line 15: “potentially inadequate description of certain processes”. Here a more thorough discussion of model functioning is needed. Now the validation data is more critically discussed than the model. I would expect that at location with a measurement frequency of several weeks to months, there is not much smoothing effect in monthly averages. Anyway such effect cannot explain structural differences between model and in-situ data, as shown in Figure 5a: DIN is consistently underpredicted and DIP overpredicted by the model.

In this sentence, we had already hinted at what might be the relevant processes (‘..such as the grazing formulation of zooplankton and the representation of the light climate’) but we will extend this in the revised version. Moreover, the smoothing effect in this sentence was about Chlorophyll, and not DIN and DIP. On the other hand, the consistent under-prediction of DIN and over-prediction of DIP was found to be caused by the atmospheric deposition not properly registered during model initialization due to a wrong configuration file -we would like to thank the referee for this careful observation. In the new model version which includes atmospheric deposition, this problem is resolved.

Page 10, Figure 5: the Pearson coefficients in the figures are too small to read. It would be clearer to present them in a table.

We will include either a table or a Taylor Diagram for summarizing the model performance against station data in the revised manuscript.

Page 13, Figure 8: Please also include similar figures for chlorophyll. Chlorophyll is the only model variable that is relevant to judge the validity of the novel modeling approach.

As explained above, performance of the model for chlorophyll is separately assessed using other data sources, and the representativeness of the chlorophyll measurements available in the ICES data base is much inferior in comparison to that of the DIN and

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DIP measurements.

Page 15, lines 5 – 13. The reader has no information to judge whether the sinking speeds in MAECS are more realistic than in other models. I would expect that the variability in the physical model underlying the ecosystem model is the main driver of vertical variability in phytoplankton concentrations. I don't see any information to convince me that "intracellular regulation of nutrient storages and pigmentary material" plays any role in this.

We did not intend to claim that the sinking speeds in MAECS are more realistic than other models, and full assessment of such a claim is again beyond the scope of the current study. We argued however, that the formation of thin chlorophyll layers is captured better than some other recent modelling attempts, citing one of those as an example. The reproduction of these structures is sensitive to the dependency of sinking rates on nutrient quotas. Description of this dependency in our model was based on earlier literature as explained in Appendix A1 (page21, lines 8-12, eq. A25). The sentence quoted by the referee is based on the fact that the internal quotas in our model scheme is affected by the allocation of resources to light harvesting (as proxied by pigmentary material) and nutrient acquisition (as proxied by nutrient stores), although we recognize that the sentence was possibly misleading, which we will revise.

Page 17, lines 3-5. This is not an entirely open question. There are some interesting papers about this effect, such as: Burson, Amanda, et al. "Unbalanced reduction of nutrient loads has created an offshore gradient from phosphorus to nitrogen limitation in the North Sea." Limnology and Oceanography (2016) and references therein.

We thank the referee for pointing to this study, which we were not aware of. We will extend this section with the findings of Burson et al. (2016) and references therein.

Page 17, line 10: If you use a data source for validation of the model you cannot conclude that the data are wrong instead of the model. Also the reason that some in-situ measurements in Figure 5 are above 50 is not valid. Figure 5 shows that the

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majority of the in-situ data is well below 50. So to make a fair comparison between in-situ data and satellite data, you should compare the seasonal averages, also at the offshore stations.

Although we agree that our sentence here was misleading, the 'bias' implied in this sentence was based on a pointwise comparison of the in-situ data with the satellite estimates (Fig.R1). Nevertheless, such tedious discussion will not be necessary in the revised version, as the overestimation problem is largely resolved with the updated model.

Page 17: lines 13 – 18. Here you only compare patterns in chlorophyll-c ratios with literature, but not the actual ranges. The numbers in Figure 14 are too small to read so I cannot judge whether the overprediction in chlorophyll in spring (Figure 13) is caused by too much phytoplankton biomass or too high chlorophyll to carbon ratios.

The over-prediction was caused by high phytoplankton concentrations, which is largely resolved in the new model run while the range of chlorophyll to carbon concentrations remains the same, which is between 0.015-0.045 gChl/gC for the seasonal averages as shown in this figure. We will improve the clarity of Figure 14 and add units.

Page 20: Lines 3 – 4. This is an interesting conclusion, but it is not well supported by the results presented in this paper.

We agree with the referee. Based on further elaboration of our model results, we will formulate a more precise statement here.

Technical comments: The text is too small to read in most figures.

We will increase the font sizes where necessary.

Detailed response to Referee #3

1) Model formulation: The authors consider a grazing rate function of prey biomass whatever the phytoplankton species represented. There are potential issues with this

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hypothesis as Phaeocystis colonies (that can dominate the spring bloom in some of the coastal stations of the studied area) is not grazed by copepods. This should be modified or/and discussed.

Our model indeed does not resolve the differences between phytoplankton taxa with respect to grazing defences, as well as other relevant features such as silicate limitation by diatoms. Extension of the model at this stage to resolve such differences is not feasible. We had mentioned (page 19, line24 - page20, line2) that the model could be extended in the future with multiple phytoplankton groups each of which resolve the acclimation processes as described in this study. We will discuss in more detail the implications of having omitted the taxa-specific features, including the grazing resistance of *Phaeocystis*.

2) Model validation: In general, the model reasonably well reproduced available data. However, it is not clear which criteria is used to determine when observed data are realistically represented or not (e.g. p9 L1). This needs to be clarified.

Referee #1 also raised concerns about subjective statements regarding the assessment of the model skill. We will formulate more objective and precise expressions for evaluating the model performance.

3) Model exploitation: The mechanistic description of the regulation of phytoplankton composition is pointed as an important process and an improvement compared to other existing models to correctly describe primary producers but also nutrient cycling. However, this is not directly evidenced in the paper based on model results. A comparison of results obtained with and without taking into account for these processes is needed to support this conclusion.

Referee #1 also suggested a comparison with a non-acclimative version. We will address this indeed relevant issue in the revised version.

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Figure 4: legend 'T' and 'S' on the dots: not clear

We will increase the vertical spacing between the labels and markers and use larger font sizes.

P9 L1: How determine 'realistic' and 'not realistic' results ? (see general comment 2)

Again, we will formulate more precise and objective statements for describing model performance.

P15 L6-8: This is an important result and could be developed and evidenced based on model results (Figure with different parameterization of under-water light climate and sinking rate of phytoplankton for example).

We will perform suggested sensitivity runs and report the results in the revised version.

Figure 12: Why N:P variability of model results is always lower than the one observed?

The variability in the N:P ratios is indeed relevant for a better understanding of this Figure. As mentioned in the text, the high N:P ratio is driven by the riverine fluxes, and actually also high atmospheric N-deposition rates close to the land. With the inclusion of the previously omitted N-deposition rates we indeed obtain a better representation of the high N:P cases within the shallower regions (<10m), although not to a full extent, which might be related with the unrepresented seasonality of atmospheric deposition we use as the model forcing. As also mentioned in the text, the low N:P ratios is consequence of a chain of events: phytoplankton growth, sedimentation, and denitrification and phosphorus release in the benthic layer. The inability of the model to reproduce the low N:P events is therefore presumably due to the coarse representation of the benthic processes by the model. However, with the updated model, we also observed a slight improvement in the reproduction of these low N:P cases, which is probably due to a better representation of the tidal currents in the system with a net effect of a restricted transport of high N:P coastal waters to the off-shore areas. We will extend the discussion of the patterns displayed in Fig. 12 in revised manuscript.

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P17 L10-11: This is not so clear for me: Fig 5 also shows an important overestimation of simulated Chl a compared to observation.

Both other referees raised similar questions about this sentence. Please see Fig. R1, which indeed suggests a systematic underestimation of the higher range of values by the satellite product, although we agree that the sentence there comparing seasonal average values with snapshot values was not fair, but will nevertheless not be needed in the revised manuscript.

P 18 L5: The variability of Chl:C can also partly result from the overestimation of Chl a in the model (see previous comment).

A similar concern was raised by the Referee #2 too. With the updated model results, we obtained lower chlorophyll concentrations overall, although the degree of variability in Chl:C ratios was not affected.

P20 L 3-10: This should be evidenced based on comparison of two simulations (with and without taking account for photoacclimation) (see general comment 3)

As stated above, we will address this relevant issue in the revised manuscript.

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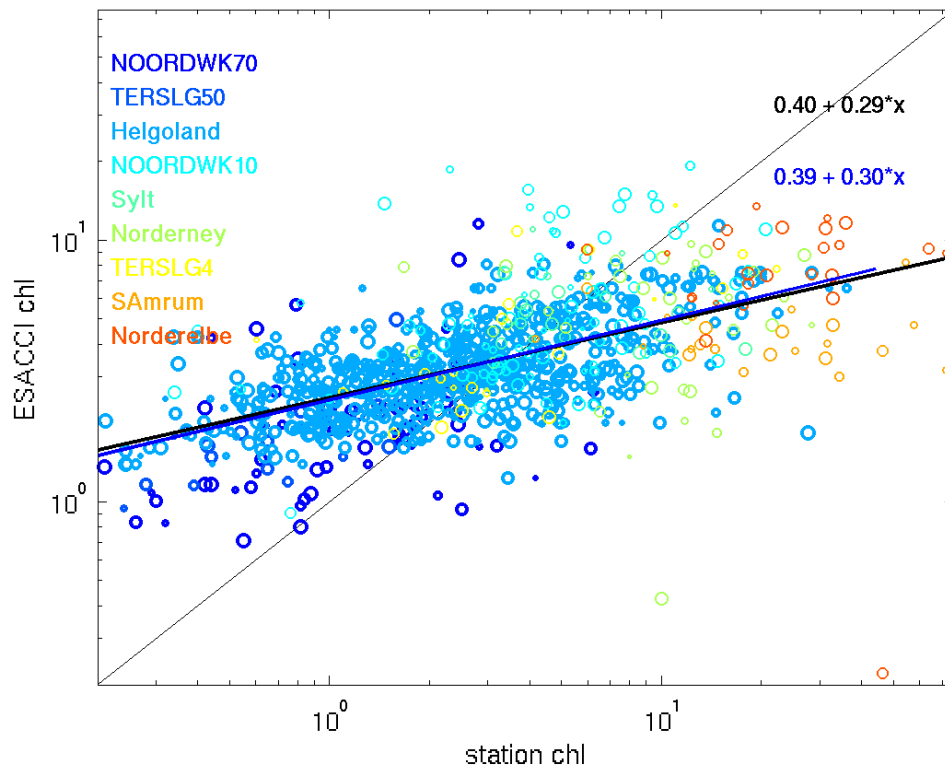


Fig. 1. Figure R1: Chlorophyll concentrations measured at the stations vs. estimated by the ESA-CCI images.

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