Response to reviewer 1#

specific comments

comments: I would move or remove the first paragraph about ozone depletion as it does not flowbonto the rest of the introduction. The importance of CB is enough to justify the study.

Response： We have removed ozone depletion part.

comments: Line 160-163: This movement of P, PA or PAB to "another treatment" – but which? You do not specify and this is very confusing. Then in Figure 2 on Carbon and N2 fixation you also have UVA, UVB and UVR and I have no idea what they represent in terms of your treatments. Lines 307 to 324 that detail the results using P, PA, PAB, P_0, PA_0, PAB_0 and UVA, UVB and UVR are all confusing.

Response: We have added description in Line 166-168: (namely P grown cells divided into P′, PA′, PAB′ treatments; PAB grown cells divided into P′, PA′, PAB′ treatments)

comments: Figure 1: why are there no damage and repair rates for P treatment? Values for all three treatments are given in the text (lines 259 to 262) but not in Figure 1C.

Response: We added damage and repair rates of P treatment in Fig 1C and D.

comments: Figure 2: For both carbon fixation and N2 fixation you calculated inhibition induced by UVA, UVB and UVR and termed this I_P, I_{PA} and I_{PAB}. Why not use this

terminology in the Figures 2B, 2D? – instead you use two different namings –UVA, UVB and UVR– this is confusing.

Response: We changed namings of UVA, UVB and UVR to I_{UVA}, I_{UVB} and $I_{UVA+UVB}$ in Fig 2 and Fig 6.

comments: Line 274: In your UVB treatment, it includes UVA, right so the treatment is actually UVA+UVB?

Response: there actually three treatments in short-term exposure, one is PAR alone, one is PA (which is PAR+UVA), the other is PAB (which is PAR+UVA+UVB) treatment.

comments: Line 282: "other phytoplankton" is referred to here and in Figura 3A. Given that you are comparing with other cultures, you need to specify in the methods how they were grown and give their full names as they are abbreviated in the Figure itself. Some readers may not be aware of these species.

Response: Yes, I have already given detail information about the full names of those species and growth conditions, which were written in Line 204-205 in M&M.

comments: Line 294: "addition of UVR significantly reduced the trichome length by 22% and 11%" How can one treatment (UVR) cause two different reductions (22 and 11%)?

Respones: this experiment was conducted outdoor, light irradiance was different every day, so as the growth rate, the trichome length was measured on the day 11^{th} and day 15^{th}, on those two days, the trichome length of PAR+UVA+UVB treatment was reduced by 22% on day

11^{th} and by 11% on the day 15^{th}, compare to the PAR treatment. I have modified the sentence to make this statement more clearly in Line 304-305.

comments: Lines 366-368: I think you should cite Neale et al 1998 J Phycol here.

One aspect that should be discussed more is that fact that UV absorbing compounds (most likely MAAs) are expensive to make (see Litchman et al 2002) in terms of Nitrogen in particular so this is an interesting aspect that should be discussed given your results. At the end of the paragraph (lines 465-467) would be a good place.

Response: Thanks for your advice, I have cite this reference there. We added the citation in new Line 382-386: A red-tide dinoflagellate *Gymnodinium sanguineum Hirasaka* accumulates about 14-fold more MAAs in high (76 W $\cdot m^{-2}$) than in low (15 W $\cdot m^{-2}$) growth light and the high-light grown ones have lower sensitivity to UV radiation at wavelengths strongly absorbed by the MAAs (Neale et al., 1998).

We also added new lines to discuss N limitation and UV sensitivity, in new Line 485-489: On the other hand, the UV absorbing compounds (most likely MAAs) are expensive to make in terms of nitrogen in particular (Singh et al., 2008). Decreased nitrogen supplied may increase sensitivity of phytoplankton assemblages to UV further (Litchman et al 2002), thus potentially creating a positive feedback between N-limitation and the UV sensitivity.

comments: Technical corrections:

Response: Revised

Response to Reviewer 2#

General Comments

Comments: Also the ecological consequences in a climate change context must be highlighted in the discussion section as well as including future research that would be necessary to confirm and/or deepen the consequences of the studied effects in C and N cycle on the ocean (see Trichodesmium ecological role as C and N source in the ocean, Berger et al., 2012).

Response: we added discussion in Line 514-515:

"future research that would be necessary to confirm and/or deepen the consequences of UV effects in carbon and nitrogen cycle in the ocean."

Specific Comments

Comments:Introduction The general objective of your investigation is not sufficiently justified, it would be better to connect your work with the need to investigate about the topics that are not explored yet (i.e., UVR effects on N2 fixation) and emphasize the importance of your results in the context of climate change. For example: Because of the importance of Trichodesmium in the input of carbon and nitrogen on oligotrophic oceans, and the lack of studies about the impact of enhanced UVR on the C and N fixation, is that we design experiments : : :: : :: : :: : :: : :.. In particular, we evaluated the role of UVR in decreasing: : :: : :: : : The UVR doses we used represent realistic values in a current scenario (or future scenario of climatic change by the year : : :: : :: : :)

Response: we added texts in new line78-80: "Because of the importance of

Trichodesmium in the input of carbon and nitrogen on oligotrophic oceans, and the lack of studies about the impact of enhanced UVR on the C and N fixation, is that we design the experiments."

Comments: Material and Methods

1.-Line 87: I would replace "Estrategy Work" by "Experimental Design", and start explaining the experiments regarding the study0s objective. For example, "The experiments to evaluate: : :: : :: : :: : :.were carried on..... as follows:"

Responses:

We replace "study strategy" by "experimental design".

And added texts in line 88-90: "The experiments to evaluate how UVR affects photosynthesis and N_2 fixation of *Trichodesmium* were carried on indoor and outdoor as follows:"

2.-Line 154: The specific growth rate is only calculated for days 8 to 11 and 12 to 16. What happened from days 1 to 7 is not shown, nor justified the reasons for that. If your study only assesed the exponential growth phase, it is necessary to define it.

Responses:

We added texts to explain it in new line 154-155: "In order to evaluate adaptation responses of *Trichodesmium* to natural solar irradiance, all parameters were obtained

after one week acclimation outdoor."

3.-Line 167: The measurement of effective quantum photochemical yield is not justified. It would be clarifying to include a paragraph explaining what this proxy indicates.

Responses: we added texts to explain Fv'/Fm' in line 173-175: "Effective photochemical quantum yield (F_V'/F_M') is generally considered to be light quantum using efficiency. We use this parameter to indicate Photosystem II activity."

4.-Line 199: Because the procedure for absorption spectra measurement is explained before for Trichodesmium, it's not necessary to repeat the same for the other species.

Responses: we added text "as the same method in *Trichodesmium*" in lin 208 to illustrate the same measurement as Trichodesmium. But in the Trichodesmium part I emphasize the Chlorophyll-specific absorption cross-sections (a*) measurements not the Chl a measurement.

5.-Line 239: Acclimatization conditions of cultures instead of culture conditions is better understood

Responses: revised in new line 247.

Comments: Results

1.-Line 286: Because UVACs values before the 10 hours exposure are not shown, it is not clear if the change is referred to time or to differences among PAB, PA and P. In this latter

case, it would be better if you explained the idea in the following way: “did not present differences between radiation treatments after exposure: : :: : :: : :.”

Response：we added texts : “….not present differences between radiation treatments after exposure to UV for 10 hrs.” in line 295.

2.- Line 312: The paragraph is not clear and/or wrong because you talk about long-term UV-A exposure, and the long term treatments were only PAB and P, there was not PA. I would replace this paragraph with “inhibition induced by UV-A at short exposures in PAB and P acclimated cells. was..... and higher than inhibition induced by UV-B"

Responses: we revised the text to “However, inhibition induced by UV-A at short exposures was about 58% in both P and PAB treatments and significantly higher than inhibition induced by UV-B radiation (Fig. 6B, $p<0.01$).”in line 321-325.

Comments: Discussion

1.- It would be necessary to give a better closure to the discussion adding future research (see General Comments)

Responses: we emphasize the future research in the last paragraph: “….future research that would be necessary to confirm and/or deepen the consequences of UV effects in carbon and nitrogen cycle in the ocean.”

2.- Lines 348, 431: The genus Anabaena for planktic morphotypes was replace by Dolichospermum since 2009 (see Wacklin et al., 2009)

We added the new name in brackets in line 359

3.-Line 412: I would replace “adaptation” with “acclimatization capacity depending on intensity and spectral quality of radiation”. The latter is based on the difference between adaptation and acclimatization terms.

Responses：replaced

4.-Line 429: See Fiorda et al., 2011. It would be very valuable adding their results in the discussion about the change of morfology due to UVR exposure

Responses:We added texts to show their discussion : “….. because UVR may affect calcium signaling then the expression of the key genes responsible for cell differentiation”

Technical corrections

Responses: All revised.

Effects of ultraviolet radiation on photosynthetic performance and N_2 fixation in *Trichodesmium erythraeum* IMS 101

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Abstract

Biological effects of ultraviolet radiation (UVR; 280–400 nm) on marine primary producers are of general concern, as oceanic carbon fixers that contribute to the marine biological CO_2 pump are being exposed to increasing UV irradiance due to global change and ozone depletion. We investigated the effects of UV-B (280-320 nm) and UV-A (320-400 nm) on the biogeochemically-critical filamentous marine N_2-fixing cyanobacterium *Trichodesmium* (strain IMS101) using a solar simulator as well as under natural solar radiation. Short exposure to UV-B, UV-A, or integrated total UVR significantly reduced the effective quantum yield of photosystem II (PSII) and photosynthetic carbon and N_2 fixation rates. Cells acclimated to low light were more sensitive to UV exposure compared to high-light grown ones, which had more UV absorbing compounds, most likely mycosporine-like amino acids (MAAs). After acclimation under natural sunlight, the specific growth rate was lower (by up to 44%), MAAs content was higher, and average trichome length was shorter (by up to 22%) in the full spectrum of solar radiation with UVR, than under a photosynthetically active radiation (PAR) alone treatment (400-700 nm). These results suggest that prior shipboard experiments in UV-opaque containers may have substantially overestimated in-situ nitrogen fixation rates by *Trichodesmium*, and that natural and anthropogenic

elevation of UV radiation intensity could significantly inhibit this vital source of new nitrogen to the current and future oligotrophic oceans.

Introduction

Global warming is inducing shoaling of the upper mixed layer and enhancing a more frequent stratification of the surface layer, thus exposing phytoplankton cells which live in the upper mixed layer to higher depth-integrated irradiance including UV radiation (Häder and Gao, 2015). The increased levels of UV radiation have generated concern about their negative effects on aquatic living organisms, particularly phytoplankton, which require light for energy and biomass production.

Cyanobacteria are the largest and most widely distributed group of photosynthetic prokaryotes on the Earth, and they contribute markedly to global CO_2 and N_2 fixation (Sohm et al., 2011). Fossil evidence suggests that cyanobacteria first appeared during the Precambrian era (2.8 to 3.5 $\times 10^9$ years ago) when the atmospheric ozone shield was absent (Sinha and Häder, 2008). Cyanobacteria have thus often been presumed to have evolved under more elevated UV radiation conditions than any other photosynthetic organisms, possibly making them better equipped to handle UV radiation.

Nevertheless, a number of studies have shown that UV-B not only impairs the DNA, pigmentation and protein structures of cyanobacteria, but also several key metabolic activities, including growth, survival, buoyancy, nitrogen metabolism, CO_2 uptake, and ribulose 1,5-bisphosphate carboxylase activity (Rastogi et al., 2014). To deal with UV stress cyanobacteria have evolved a number of defense strategies, including migration to escape from UV radiation, efficient DNA repair mechanisms, programmed cell death, the production of antioxidants, and the biosynthesis of UV-absorbing compounds, such as MAAs and scytonemin (Rastogi et al., 2014; Häder et al., 2015).

The non-heterocystous cyanobacterium *Trichodesmium* plays a critical role in the marine nitrogen cycle, as it is one of the major contributors to oceanic nitrogen fixation

(Capone et al., 1997) and furthermore is an important primary producer in the tropical and sub-tropical oligotrophic oceans (Carpenter et al., 2004). This global importance of *Trichodesmium* has motivated numerous studies regarding the physiological responses of *Trichodesmium* to environmental factors, including visible light, phosphorus, iron, temperature, and CO_2 (Kranz et al., 2010; Shi et al., 2012; Fu et al., 2014; Spungin et al., 2014; Hutchins et al., 2015). However, to the best of our knowledge, nothing has been documented about how UV exposure may affect *Trichodesmium*.

Trichodesmium spp. have a cosmopolitan distribution throughout much of the oligotrophic tropical and subtropical oceans, where there is a high penetration of solar UV-A and UV-B radiation (Carpenter et al., 2004). It also frequently forms extensive surface blooms (Westberry and Siege, 2006), where it is presumably exposed to very high levels of UV radiation. Moreover, in the ocean, *Trichodesmium* populations may experience continuously changing irradiance intensities as a result of vertical mixing. Cells photoacclimated to reduced irradiance at lower depths might be subject to solar UVR damage when they are vertically delivered close to the sea surface due to mixing. Therefore, this unique cyanobacterium may have developed defensive mechanisms to overcome harmful effects of frequent exposures to intense UV radiation. Understanding how its N_2 fixation and photosynthesis respond to UV irradiance will thus further our knowledge of its ecological and biogeochemical roles in the ocean.

When estimating N_2 fixation using incubation experiments in the field, marine scientists have typically excluded UV radiation by using incubation bottles made of UV-opaque materials like polycarbonate (Capone et al., 1998; Olson et al., 2015). Thus, it seems possible that most shipboard measurements of *Trichodesmium* N_2 fixation rates could be overestimates of actual rates under natural UV exposure conditions in the surface ocean. Because of the importance of *Trichodesmium* in the input of carbon and nitrogen on oligotrophic oceans, and the lack of studies about the impact of enhanced UVR on the C and N fixation, is that we design the experiments. In this study, *Trichodesmium* was exposed to spectrally realistic irradiances of UVR in laboratory

experiments to examine the short-term effects of UVR on photosynthesis and N_2 fixation. In addition, *Trichodesmium* was grown under natural solar irradiance outdoors in order to assess UV impacts on longer timescales, and to test for induction of protective mechanisms to ameliorate chronic UV exposure effects.

Materials and methods

Experimental~~Study strategy~~Experimental design The experiments to evaluate how UVR affects photosynthesis and N_2 fixation of *Trichodesmium* were carried on indoor and outdoor as follows: thist~~This~~ study included two parts: (1) A short-term experiment under a solar stimulator (refer to Fig.S1 for the spectrum) to examine the responses of *Trichodesmium erythraeum* IMS 101 to a range of acute UV radiation exposures, and (2) A long-term UV experiment under natural sunlight to examine acclimated growth and physiology of *Trichodesmium* IMS 101. The first set of experiments was intended to mimic intense but transitory UV exposures, as might occur sporadically during vertical mixing, while the second set was intended to give insights into responses during extended near-surface UV exposures, such as during a surface bloom event.

Short-term UV experiment *Trichodesmium erythraeum* IMS101 strain was isolated from the North Atlantic Ocean (Prufert-Bebout et al., 1993) and maintained in laboratory stock cultures in exponential growth phase in autoclaved artificial seawater enrich with nitrogen free YBCII medium (Chen et al., 1996). For the short-term UV experiment, the cells were grown under low light (LL) 70 μmol photons m^{-2} s^{-1} and hight light (HL) 400 μmol photons m^{-2} s^{-1} (12:12 light: dark) of PAR for at least 50 generations (about 180 days) prior to the UV experiments. These two light levels represent growth sub-saturating and super-saturating levels for *Trichodesmium* (Cai et al., 2015). Cultures were grown in triplicate using a dilute semi-continuous culture method, with medium renewed every 4-5 days at 25°C. The cell concentration was maintained at $< 5 \times 10^4$ cell ml^{-1}.

To determine the short-term responses of *Trichodesmium* IMS101 to UV radiation, subcultures of *Trichodesmium* IMS101 were dispensed at a final cell density of 2-4 × 10^4 cells ml^{-1} into containers that allow transmission of all or part of the UV spectrum, including 35 ml quartz tubes (for measurements of carbon fixation or measurements of fluorescence parameters), 100 ml quartz tubes (for pigment measurements), or 13 ml gas-tight borosilicate glass vials (for N_2 fixation measurements). Three triplicated radiation treatments were implemented: (1) PAB (PAR+UV-A+UV-B) treatment, using tubes covered with Ultraphan film 295 (Digefra, Munich, Germany), thus receiving irradiances >295 nm; (2) PA (PAR+UV-A) treatment, using tubes covered with Folex 320 film (Montagefolie, Folex, Dreieich, Germany), and receiving irradiances >320 nm; and (3) P treatment: tubes covered with Ultraphan film 395 (UV Opak, Digefra), with samples receiving irradiances above 395 nm, representing PAR (400-700 nm). Since the transmission spectrum of the borosilicate glass was similar to that of Ultraphan film 295, the borosilicate glass vials for N_2 fixation measurements of PAB treatment were uncovered. Transmission spectra of these tubes (quartz and borosilicate) and the various cut-off foils used in this study are shown in Fig. S1.

The experimental tubes were placed under a solar simulator (Sol 1200W; Dr. Hönle, Martinsried, Germany) at a distance of 110 cm from the lamp, and maintained in a circulating water bath for temperature control (25ºC) (CTP-3000, Eyela, Japan). Irradiance intensities were measured with a LI-COR 2π PAR sensor (PMA2100, Solar light, USA) that has channels for PAR (400-700 nm), UV-A (320-400 nm) and UV-B (280-320 nm). Measured values at the 110 cm distance were 87 Wm^{-2} (PAR, ca. 400 µmol photons m^{-2} s^{-1}), 28 Wm^{-2} (UV-A) and 1 Wm^{-2} (UV-B), respectively. For the fluorescence measurements, samples were exposed under a solar stimulator for 60 min and measurements of fluorescence parameters were performed during the exposure (see below). Due to analytical sensitivity issues, for the carbon and N_2 incorporation measurements, the exposure duration was 2 hrs, and for the measurements of UVAC (UV-absorbing compounds) contents, the exposure time was 10 hrs.

Long-term UV experiment To assess the long-term effects of solar ultraviolet radiation on *Trichodesmium* IMS101, an outdoor experiment was carried during the winter (Jan 1st to Jan 26th, 2014) in subtropical Xiamen, China. 300-400 ml cell cultures were grown in 500 ml quartz vessels exposed to 100% daytime natural solar irradiance (surface ocean irradiance) (daytime PAR average of ~120W m^{-2}, highest PAR at noon ~300W m^{-2}). All of the quartz vessels were placed in a shallow water bath at 25ºC using a temperature control system (CTP-3000, Eyela, Japan). Two triplicated radiation treatments were implemented: (1) treatment P: PAR alone (400-700 nm), tubes covered with Ultraphan film 395 (UV Opak, Digefra); (2) treatment PAB: PAR+UV-A+UV-B (295-700 nm), unwrapped quartz tubes. Incident solar radiation was continuously monitored with a broadband Eldonet filter radiometer (Eldonet XP, Real Time Computer, Möhrendorf, Germany) that was placed near the water bath. Daily doses of solar PAR, UV-A and UV-B during the experiments are shown in Fig. S2. The photoperiod during the outdoor incubation was 11:13 light:dark (light period from 7:00-18:00 of local time). Cells were maintained in exponential growth phase (cell density < 5×10^4), with dilutions (after sunset) every 4 days. All parameters were measured after acclimation under P or PAB radiation for a week.

In order to evaluate adaptation responses of *Trichodesmium* to natural solar irradiance, all parameters were obtained after one week acclimation outdoor. Specific growth rate (μ, d^{-1}) of *Trichodesmium* IMS101 was determined based on the change in cell concentrations over 4 days during the 8-11th and 12-15th day using microscopic counts (Cai et al., 2015), the corresponding total dose from Day 8 to Day 11 and from Day 12 to Day 15 were 17.03 and 18.51 MJ m^{-2}, respectively. Chl *a* content was measured at the 11th, 15th and 19th day, and Chl *a*-specific absorption spectrum was measured at the 18th day. Carbon and N_2 fixation rate were measured at 11:00-13:00 on the 18th day; the diel solar irradiance record on that day is given in Fig. S3. In order to separate the respective effects of UV-A and UV-B on carbon and N_2 fixation, a shift experiment was carried out: subcultures from either P or PAB treatments were

transferred into another P (PAR), PA (PAR+UV-A), PAB (PAR+UV-A+UV-B) treatment, which were marked as P′, PA′, PAB′ treatments, respectively (namely P grown cells divided into P′, PA′, PAB′ treatments; PAB grown cells also divided into P′, PA′, PAB′ treatments). 35 ml quartz tubes and 13 ml gas-tight borosilicate glass vials were used for carbon and N_2 fixation measurements, respectively, as described below. Triplicate samples were used for each radiation treatment for carbon and N_2 fixation, and the incubations were performed under 100% solar irradiance for 2 hrs.

Measurements and analyses

Effective photochemical quantum yield Effective photochemical quantum yield (F_V'/F_M') is generally considered to be light quantum using efficiency. We use this parameter to indicate Photosystem II activity. During the exposure under the solar stimulator in the short-term experiment, small aliquots of cultures (2 ml) were withdrawn at time intervals of 3-10 min and immediately measured (without any dark adaptation) using a Pulse-Amplitude-Modulated (PAM) fluorometer (Xe-PAM, Walz, Germany). The quantum yield of PSII (F_V'/F_M') was determined by measuring the instant maximum fluorescence (F_M') and the steady state fluorescence (Ft) under the actinic light. The maximum fluorescence (F_M') was determined using a saturating light pulse (4000 μmol photons m^{-2} s^{-1} in 0.8 s) with the actinic light level set at 400 μmol photons m^{-2} s^{-1}, similar to the PAR level during the solar simulator exposure The quantum yield was calculated as: $F_V'/F_M' = (F_M'-Ft)/F_M'$ (Genty et al., 1989).

Chlorophyll-specific absorption spectra and UV-absorbing compounds (UVACs) Chl *a*-specific absorption spectra were measured on the 18th day, after consecutive sunny days. Cellular absorption spectra were measured using the ‘‘quantitative filter technique’’ (Kiefer and SooHoo, 1982; Mitchell 1990). The cells were filtered onto GF/F glass fiber filters and scanned from 300 to 800 nm using a 1-nm slit in a spectrophotometer equipped with an integrating sphere to collect all the transmitted or forward-scattered light (i.e., light diffused by the filter and the quartz diffusing plate).

Filters soaked in culture medium were used as blanks. Chlorophyll-specific absorption cross-sections (a*) were calculated according to Cleveland and Weidemann (1993) and Anning et al., (2000). Content of Chl *a* and UV-absorbing compounds (UVACs) were measured by filtering the samples onto GF/F filters and subsequently extracted in 4 mL of 100% methanol overnight in darkness at 4 ^{o}C. The absorption of the supernatant was measured by a scanning spectrophotometer (Beckman Coulter Inc., Fullerton, CA, USA). The concentration of Chl *a* was calculated according to Ritchie (2006). The main absorption values for UV-absorbing compounds ranged between wavelengths of 310 and 360 nm, and the peak absorption value at 332 nm was used to estimate total absorptivity of UVACs according to Dunlap et al., (1995). The absorptivity of UVACs was finally normalized to the Chl *a* content (μg (μg Chl *a*)$^{-1}$).

Trichodesmium IMS101 UVACs content was compared to that of three other marine phytoplankton species, including *Chlorella*.sp, *Phaeodactylum tricornutum*, and *Synechococcus* WH7803, representing a green alga, a diatom and a unicellular cyanobacterium, respectively. All cultures were maintained under the same conditions (25^{o}C, 150 μmol photons m^{-2} s^{-1}) for several days prior to pigment extraction. The absorption spectra were measured~~–~~ as the same method in *Trichodesmium* by filtering the samples on GF/F filters that were subsequently extracted in 4 mL of 100% methanol overnight at 4 ^{o}C. The absorption spectra of the supernatant were scanned from 250 to 800 nm in a spectrophotometer (Beckman Coulter Inc., Fullerton, CA, USA). The Optical Density (OD) values were then normalized to OD (662 nm), Chl *a* peak.

Carbon fixation rates Carbon fixation rate of both short- and long-term experiments were measured using the ^{14}C method. A total of 20 ml samples were placed in 35 ml quartz tubes and inoculated with 5μCi (0.185 MBq) of labeled sodium bicarbonate (ICN Radiochemicals), and were then maintained under the corresponding radiation treatments for 2 hrs. After incubation, the cells were filtered onto Whatman GF/F filters (Φ 25 mm) and stored at -20^{o}C until analysis. To determine the radioactivity, the filters were thawed and then exposed to HCl fumes overnight and dried at 60^{o}C for 4 hrs

220 before being placed in scintillation cocktail (Hisafe 3, Perkin-Elmer, Shelton, CT, USA),
221 and measured with a scintillation counter (Tri-Carb 2800TR, Perkin-Elmer, Shelton,
222 CT, USA) as previously described (Cai et al., 2015).

223 **N_2 fixation rates** Rates of N_2 fixation for both short- and long-term experiments were
224 measured in parallel with the carbon fixation measurements using the acetylene
225 reduction assay (ARA) (Capone et al., 1993). Samples of 5 ml subcultures were placed
226 in 13 ml gas-tight borosilicate vials (described above), and 1ml acetylene was injected
227 into the headspace before incubating for 2 hrs under the corresponding radiation
228 treatment conditions. A 500 μl headspace sample was then analyzed in a gas
229 chromatograph equipped with a flame-ionization detector and quantified relative to an
230 ethylene standard. The ethylene produced was calculated using the Bunsen gas
231 solubility coefficients according to Breitbarth et al., (2004) and an ethylene production
232 to N_2 fixation conversion factor of 4 was used to derive N_2 fixation rates, which were
233 then normalized to cell number.

234 **Data analysis** The inhibition of ΦPSII, carbon fixation and N_2 fixation due to UVR,
235 UV-A, or UV-B was calculated as:

236 $$\text{UVR-induced inhibition} = (I_P - I_{PAB})/I_P \times 100\%$$

237 $$\text{UV-A-induced inhibition} = (I_P - I_{PA})/I_P \times 100\%$$

238 $$\text{UV-B-induced inhibition} = UVR_{inh} - UVA_{inh}$$

239 where I_P, I_{PA}, I_{PAB} indicate the values of carbon fixation or N_2 fixation in the P, PA
240 and PAB treatments, respectively. Repair (r) and damage (k) rates during the 60 min
241 exposure period in the presence of UV were calculated using the Kok model (Heraud
242 and Beardall, 2000):

243 $$P/P_{initial} = r/(r+k) + k/(r+k) \times \exp(-(r+k) \times t),$$

244 where $P_{initial}$ and P were the yield values at the beginning and at exposure time t.
245 Three replicates for culture conditions or each radiation condition was used in all

experiments, and the data are plotted as mean and standard deviation values. Two way ANOVA tests were used to determine the interaction between ~~culture~~ acclimatization conditions and UVR at a significance level of p=0.05.

Results

Short-term UV experiment The effects of acute UVR exposure on cells grown under LL and HL conditions are shown in Fig.1. For the cells grown under LL condition, the F_V'/F_M' declined sharply within 10 min after first exposure in all radiation treatments, and then leveled off. F_V'/F_M' decreased less in the samples receiving PAR alone (to 43% of the initial value) than those additionally receiving UV-A (to 30% of the initial value) or UV-A+UV-B (to 24% of the initial value) (Fig.1A). The F_V'/F_M' value of PA and PAB treatments were significantly lower compared to the PAR treatment (p=0.03 and p<0.01, respectively). F_V'/F_M' of HL grown cells declined less and more slowly compared to the LL grown cells. The F_V'/F_M' of HL cells under PAR alone remained more or less constant during the exposure, since the PAR level was similar to the growth level of HL (400 μmol photons m^{-2} s^{-1}). In contrast, the F_V'/F_M' decreased to 75% and 65% of its initial value for the PA and PAB treatment, respectively, and were significantly lower than the P~~PAR~~ treatment (p<0.01) (Fig.1B).

The damage and repair rates of the PSII reaction center estimated from the exponential decay in the effective quantum yield showed higher damage and lower repair rates in the LL-grown cells than in the HL-grown ones (Fig.1C,D). The PSII damage rates (k, min^{-1}) of LL grown cells were 0.14, 0.16 and 0.15 min^{-1} in the P, PA and PAB treatments, respectively, about 2 times faster than in the cells grown under HL conditions (Fig.1C). The PSII repair rates (r, min^{-1}) of LL grown cells were 0.1, 0.06 and 0.05 min^{-1} in the P, PA and PAB treatments, which were 83% (p<0.01), 33% (p<0.01) and 54% (p<0.01) lower than in HL grown cells, respectively (Fig.1D). The damage rate was not significantly different among P, PA and PAB treatments within either of

the LL- and HL-grown treatments ($p>0.05$), but the repair rate was much higher in the P treatment without UV than in PA or PAB treatments in the HL-grown cells ($p<0.01$).

The photosynthetic carbon fixation and N_2 fixation rates during the UV exposure are shown in Fig. 2. The HL-grown cells had 17% higher photosynthetic carbon fixation rates than the LL-grown ones under the PA treatment ($p<0.01$), however, the LL and HL-grown cells didn't show significant differences in carbon fixation rates under the P and PAB treatments ($p=0.29$, and $p=0.06$). In the presence of UV radiation, carbon fixation was significantly inhibited in both LL and HL-grown cells (Fig.2A). Carbon fixation inhibition induced by UV-A was about 35-45%, much larger than that induced by UV-B, which caused only about a 10% inhibition of carbon fixation ($p<0.01$). The UV-A exposed carbon fixation rate was significantly higher in the LL- grown cells than in HL grown cells ($p<0.01$), while UV-B did not cause a significant difference in inhibition between the HL~~H~~C~~H~~L- and LL~~L~~C~~L~~L-grown cells ($p=0.88$) (Fig. 2B). N_2 fixation rates were about twofold higher in HL-grown cells in all radiation treatments (Fig.2C, $p<0.01$), but the UV-induced N_2 fixation inhibition showed no significant differences between the LL and HL grown cells regardless of UV-A or UV-B exposures (Fig. 2D, $p=0.80$, 0.62, 0.39 for UVA-, UVB-, and UVR-induced inhibition, respectively).

Compared to other phytoplankton under the same growth conditions, *Trichodesmium* IMS101 had much higher absorbance in the UV region (300-400 nm) (Fig. 3A). In this study, the absorbance at 332 nm of HL-grown cells was about twofold higher compared to LL-grown ones (Fig. 3B). However, the cellular Chl *a* content (data not shown) and UVACs contents of both LL and HL grown cells did not not present differences between radiation treatments after exposure~~change after exposure~~ to UV for 10 hrs (Fig. 3C).

Long-term UV experiment After being acclimated under full natural solar radiation for 7 days, the specific growth rates of cells grown under the PAB treatment were

0.15±0.01 and 0.14±0.06 during the 8-11th day and 12-15th day periods, respectively. These growth rates were significantly lower by 44% and 39% compared to cells grown under the P treatment, respectively (Fig.4A, p=0.014 and p=0.03). The mean trichome lengths of PPAR treatment cells on the 11th and 15th day were 758±56 and 726±19 μm, while addition of UVR significantly reduced the trichome length by 22% (Day 11th, p=0.02) and 11% (Day 15th, p=0.02).

Analysis of the Chl *a* specific absorption spectra, a*(λ), demonstrated that UVR had a major effect on the absorbance of UV regions and phycobilisomes (Fig. 5). The optical absorption spectra revealed a series of peaks in the UV and visible wavelengths corresponding to the absorption peaks of UVACs at 332 nm, Chl *a* at 437 and 664 nm, phycourobilin (PUB) at 495 nm, phycoerythrobilin (PEB) at 545 nm, phycoerythrocyanin (PEC) at 569 nm, and phycocyanin (PC) at 627 nm. In the UV region, the a*(λ) value was higher in the PAB treatment cultures than in the P treatment cultures (Fig. 5). The UVR treatments did not show clear effects on Chl *a* content compared to acclimation to PPAR alone measured on different days (Fig. S3). However, the ratio of UVACs to Chl *a* was increased by 41% in the PAB compared to the P treatment ($p<0.01$).

The cells grown in the long-term P and PAB treatments showed different responses for carbon and N_2 fixation after being transferred to short-term P′, PA′, and PAB′ radiation treatments at noon on the 18th day (Fig. 6). P and PAB acclimated cells did not show significant differences in carbon fixation among all short-term P′, PA′, PAB′ treatments (Fig. 6A, p=0.17, p=0.22, p=0.51, respectively), nor in the UV-induced inhibition of carbon fixation (Fig. 6B, $p>0.05$). However, inhibition induced by UV-A at short exposures was about 58% in both P and PAB treatments and significantly higher than inhibition induced by UV-B ~~long-term UV-A exposure inhibited short-term carbon fixation by about 58% in both the P and the PAB treatments, significantly higher than that induced by UV-B~~ radiation (Fig. 6B, $p<0.01$).

N_2 fixation rates of P acclimated cells were significantly higher than PAB acclimated cells in all P′, PA′, and PAB′ treatments (Fig. 6C, $p<0.01$). The N_2 fixation inhibition induced by UV-A of PAB acclimated cells was 49%, significantly higher by 47% than that of P acclimated cells ($p=0.03$), while there was no significant difference in UVB-induced N_2 fixation inhibition between P and PAB acclimated cells (Fig. 6D, $p=0.62$). The carbon fixation rates measured under P (P~~PAR (PAR~~ treated cells to P') and PAB (PAB treated cells to PAB') conditions were 89.2 and 47.1 fmol C $cell^{-1}$ h^{-1}, respectively, while N_2 fixation rates measured under those conditions were 1.9 and 0.5 fmol N_2 $cell^{-1}$ h^{-1}. UVR exposure lowered estimates of carbon and N_2 fixation rates by 47% and 65%, respectively.

Discussion

Our study shows that growth, photochemistry, photosynthesis and N_2 fixation in *Trichodesmium*.sp are all significantly inhibited by UVR, including both UV-A and UV-B. These effects occur in both short-term, acute exposures, as well as after extended exposures during acclimated growth. These results are ecologically relevant, since this cyanobacterium is routinely exposed to elevated solar irradiances in its tropical habitat either transiently, during vertical mixing, or over longer periods during surface blooms. *Trichodesmium* provides a biogeochemically-critical source of new N to open ocean food webs, so significant UV inhibition of its growth and N_2 fixation rates could have major consequences for ocean biology and carbon cycling.

Short exposure to UVR causes a significant decline in the quantum yield of photosystem II (PSII) fluorescence of *Trichodesmium*, that is consistent with damage to critical PSII proteins such as D1 in a brackish water cyanobacterium *Arthrospira (Spirulina) platensis* (Wu et al., 2011). UV-induced degradation of D1 proteins results in inactivation of PSII, leading to reduction in photosynthetic activity (Campbell et al., 1998). In addition, studies of various microbial mats have shown that Rubisco activity

and supply of ATP and NADPH are inhibited under UV exposure, which might also lead to the reduction in photosynthetic carbon fixation (Cockell and Rothschild, 1999; Sinha et al., 1996, 1997).

Exposure to UVR had an impact on nitrogenase activity in *Trichodesmium*, since both the short- and the long-term UV exposure led to significant reduction of N_2 fixation of up to 30% (short-term) or ~60% (long-term) (Fig. 2D and 6D). Studies on the freshwater cyanobacterium *Anabaena*. sp~~S~~psp (subg. *Dolichospermum*). showed a 57% decline in N_2 fixation rate after 30 min exposure to UVR of 3.65W (Lesser, 2007). Some rice-field cyanobacteria completely lost N_2 fixation activity after 25-40 min exposure to UV-B of 2.5 W (Kumar et al., 2003). In our results, long-term exposure to UV led to higher inhibition of N_2 fixation, implying that accumulated damage to the key N_2-fixing enzyme, nitrogenase, could have occurred during the growth period under solar radiation in the presence of UVR.

Compared to N_2 fixation, UVR induced an even higher degree of inhibition of carbon fixation. The carbon fixation rate decreased by 50% in the presence of UVR. UV-A induced higher inhibition than UV-B, indicating that although UV-B photons (295-320 nm) are in general more energetic and damaging than UV-A (320-400 nm), the greater fluxes of UV-A caused more inhibition of carbon fixation, which was consistent with other studies of spectral dependence of UV effects (Cullen and Neale 1994; Neale 2000). This finding is ecologically significant, since UV-A penetrates much deeper into clear open ocean and coastal seawater than does UV-B.

Compared to low light-grown cells, the high light-grown ones were more resistant to UVR, which was reflected in the lower PSII damage rate and faster recovery rate in the presence of UVR, as well as the significantly lower levels of carbon fixation inhibition caused by UV-A and/or UV-B. Such a reduced sensitivity to UVR coincided well with a significant increase in UV-absorbing compounds in the HL-grown cells compared to the LL-grown ones. Similar dependence of photosynthetic sensitivity to

UV inhibition on growth light levels has been reported in other species of phytoplankton (Litchman and Neale, 2005; Sobrino and Neale, 2007). A red-tide dinoflagellate *Gymnodinium sanguineum Hirasaka* accumulates 14-fold MAAs in high-light grown cells (76 W m^{-2}) than in low-light grown ones (15 W m^{-2}) and the former ones have lower sensitivity to UVRat~~UV~~R ~~radiation at~~ wavelengths strongly absorbed by the MAAs (Neale et al., 1998). The sensitivity of PSII quantum yield to UV exposure in *Synechococcus* WH7803 was also less in high-light-grown versus low-light-grown cells (Garczarek et al., 2008). In addition, it has been observed that phytoplankton from turbid waters or acclimated to low-light conditions are more sensitive to UVR than those from clear waters (Villafane et al., 2004; Litchman and Neale, 2005; Helbing et al., 2015). These observations suggest that *Trichodesmium* sp. may acclimate to growth in the upper mixed layer by producing UV-absorbing compounds, making them more tolerant of UVR than cells living at deeper depths.

Although UVR ~~radiation~~ can clearly cause damage to PSII and inhibit physiological processes in *Trichodesmium* sp., this cyanobacterium has evolved protective biochemical mechanisms to deal with UVRin~~UV~~R ~~radiation in~~ their natural high-UV habitat. One important class of UV-absorbing substances are mycosporine-like amino acids (MAAs) and scytonemin. These compounds strongly absorb in the UV-A and/or UV-B region of the spectrum, and dissipate its energy as heat without forming reactive oxygen species, protecting the cells from UV and from photooxidative stress (Banaszak 2003). The ‘‘mycosporine-like amino acids’’ (MAAs), which have strong UV-absorption maxima between 310 and 362 nm (Sinha and Häder, 2008) as identified by HPLC in other studies, consist of a group of small, water-soluble compounds, including asterina-332 (λmax=332) and shinorine (λmax=334), which are the most abundant, as well as mycosporine-glycine (λmax=310), porphyra-334 (λmax=334), and palythene (λmax=360) (Shick and Dunlap 2002; Subramaniam et al., 1999). As was found previously in *Trichodesmium* spp., high absorbance in the UV region is mainly due to the presence of ‘‘mycosporinelike amino acids’’ (MAAs), with

absorbance maxima between 310~362 nm (Sinha and Häder, 2008).

Our investigation strongly suggests that *Trichodesmium* is able to synthesize MAAs (λmax ~330 nm and 360 nm) in response to elevated PAR and UVR~~ radiation~~. Synthesis of MAAs has been reported to be stimulated by high PAR and UVR~~UV radiation~~R in other phytoplankton (Karsten et al., 1998; Vernet and Whitehead, 1996; Sinha et al., 2001). Our high light-grown cells were more tolerant of UVR, likely at least partly due to their ability to synthesize double the amount of MAAs in comparison to low light-grown ones (Fig.3B). It has been showed that accumulation of MAAs may represent a natural defensive system against exposure to biologically harmful UVR~~UV radiation~~R (Karsten et al., 1998) and cells with high concentrations of MAAs are more resistant to UVR than cells with small amounts of these compounds (Garcia-Pichel and Castenholz, 1993). In fact, MAAs concentrations varying between 0.9 and 8.4 ug mg (dry weight)$^{-1}$ have been measured in cyanobacterial isolates (Garcia-Pichel and Castenholz, 1993), and ratios of MAAs to Chl *a* in the range from 0.04 to 0.19 have been reported in cyanobacterial mats (Quesada et al., 1999). In our study, we found that *Trichodesmium* contained a much higher concentration of MAAs (the highest value in HL-grown cells is 5 pg cell^{-1}) and that the ratio of these compounds to Chl *a* was 5, was consisted with previous reports in regard to *Trichodesmium* (Subramaniam et al., 1999), which is much higher than in other phytoplankton. This acclimatization capacity depending on intensity and spectral quality of radiationradiation~~adaptation~~ could be a major reason for the ability of *Trichodesmium* to grow and form extensive surface blooms under strong irradiation in the oligotrophic oceans.

In our study, no significant changes in the amount of MAAs were observed after 10 h of exposure to UVR under the solar simulator. In contrast, a significant increase of 23% in the concentration of MAAs was observed in full solar spectrum treated cells compared to PAR-treated ones grown outdoors after consecutive sunny days (on the 18th). It seems that the synthesis of MAAs takes a relatively long time. Other studies have shown the time required for induction of MAAs in other cyanobacteria is

dependent on UV doses and species, and shows a circadian rhythm (Sinha et al., 2001; Sinha et al., 2003).

Not only did long-term exposure to high solar UVR~~UV radiation~~R significantly reduce *Trichodesmium*'s growth rate (by 37~44%), but it also significantly shortened its average trichome length (less cell per filament) (Fig. 4). The decreased growth rates correlated with decreased trichome length are consistent with our previous studies under different light levels without UVR (Cai et al., 2015). It has been reported that enhanced UVR is one of the environmental factors that not only inhibit the growth of cyanobacteria, but also change their morphology (Rastogi et al., 2014). Natural solar UVR can suppress formation of heterocysts and shorten the filament length of *Anabaena* sp. PCC7120, because –UVR may affect calcium signaling then the expression of the key genes responsible for cell differentiation (Gao et al., 2007). Natural levels of solar UVR in the Southern China were also found to break the filaments and alter the spiral structure of *Arthrospira* (*Spirulina*) *platensis*, with a compressed helix that lessens UV exposures for the cells (Wu et al., 2005). Cells in the trichomes of the estuarine cyanobacterium *Lyngbya aestuarii* coil and then form small bundles in response to UV-B irradiation (Rath and Adhikari, 2007). However, the shortened trichomes of *Trichodesmium* in this work may be a result of UV-inhibited growth rather than a responsive strategy against UV.

Carbon fixation in the long-term experiment showed similar patterns with the short-term UV experiment, demonstrating that UV-A played a larger role in inhibiting carbon fixation than UV-B. Since the ratio of UV-B to UV-A is lower in natural solar light (1:50) than under our artificial UVR (1:28), the inhibitory effects of UV-B were smaller compared to UV-A in the cultures under sunlight. Carbon fixation and N_2 fixation rates measured outdoors indicated that UV-induced carbon fixation inhibition recovers quickly following transfer to PAR conditions, while the UV-induced N_2 fixation inhibition does not (Fig.6AC). Factors that might be responsible include lower turnover rate of nitrogenase than that of RuBisco; more UV-induced damage to

nitrogenase with lower efficiency of repair (Kumar et al., 2003); and indirect harm caused by ROS (Reactive Oxygen Species) induced by UV (Singh et al., 2014).

The UV effects in our study were measured under conditions that minimized self-shading, namely during growth as single filaments. However, in its natural habitat *Trichodesmium* often grows in a colonial form, with packages of many cells held together by an extracellular sheath (Capone et al., 1998). In such colonial growth forms, the effective cellular pathlengths for UVR~~UV radiation~~R are likely greatly increased, thereby amplifying the overall sunscreen factor for the colony. *Trichodesmium*.spp might use this colony strategy to protect themselves from natural UV damage in the ocean.

Our investigation shows that this cyanobacterium appears to have evolved the ability to produce exceptionally high levels of UV protective compounds, likely mycosporine-like amino acids. However, even this protective mechanism is insufficient to prevent substantial inhibition of nitrogen and carbon fixation in the high-irradiance environment where this genus lives. *Trichodesmium* spp are distributed in the upper layers of the euphotic zone in oligotrophic waters, and its population densities are generally greatest at relatively shallow depths (20 to 40 m) in the upper water column (Capone et al., 1997). It seems likely that UV inhibition therefore significantly reduces the amount of critical new nitrogen supplied by *Trichodesmium* to the N-limited oligotrophic gyre ecosystems, a possibility that has not been generally considered in regional or global models of the marine nitrogen cycle. On the other hand, the UV absorbing compounds (most likely MAAs) are expensive to make in terms of nitrogen in particular (Singh et al., 2008). Decreased nitrogen supplied may increase sensitivity of phytoplankton assemblages to UV further (Litchman et al 2002), thus potentially creating a positive feedback between N-limitation and the UV sensitivity.

Trichodesmium can form dense, extensive blooms in the surface oceans, and a frequently cited estimate of global nitrogen fixation rates by *Trichodesmium* blooms is

~42 Tg N yr^{-1} (Westberry et al., 2006). Previous biogeochemical models of global N_2 fixation have emphasized controls by many environmental factors, including solar PAR ~~radiation~~, temperature, wind speed, and nutrient concentrations (Luo et al., 2014), but have largely neglected the effects of UVR ~~radiation~~. When estimating N_2 fixation using incubation experiments in the field, however, marine scientists have typically excluded UVR~~by~~ ~~UV~~R ~~radiation by~~ using incubation bottles made of UV-opaque materials like polycarbonate (Olson et al., 2015). Our results suggest that under solar radiation at the surface ocean, including realistic levels of UVR inhibition lowers estimates of carbon fixation and N_2 fixation by around 47% and 65%, respectively (Fig.6).

Thus, it seems likely that shipboard measurements and possibly current model projections of *Trichodesmium* N_2 fixation and primary production rates that do not take into account UV inhibition could be substantial overestimates. However, our study was only carried out under full solar radiation, simulating sea surface conditions, so further studies are needed to investigate depth-integrated UV inhibition. Moreover, the response to UVR ~~UV radiation~~R may be taxon-specific. For example, unicellular N_2-fixing cyanobacteria such as the genus *Crocosphaera*, with smaller cell size and thus greater light permeability, may be more vulnerable to UVR ~~UV radiation~~R than *Trichodesmium* (Wu et al., 2015). In the future, as enhanced stratification and decreasing mixed layer depth expose cells to relatively higher UV levels, differential sensitivities to UVR ~~UV radiation~~R may result in changes in diazotroph community composition. Such UV-mediated assemblage shifts could have potentially major consequences for marine productivity, and for the global biogeochemical cycles of nitrogen and carbon, future research that would be necessary to confirm and/or deepen the consequences of UV effects in carbon and nitrogen cycle in the ocean.

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Figures

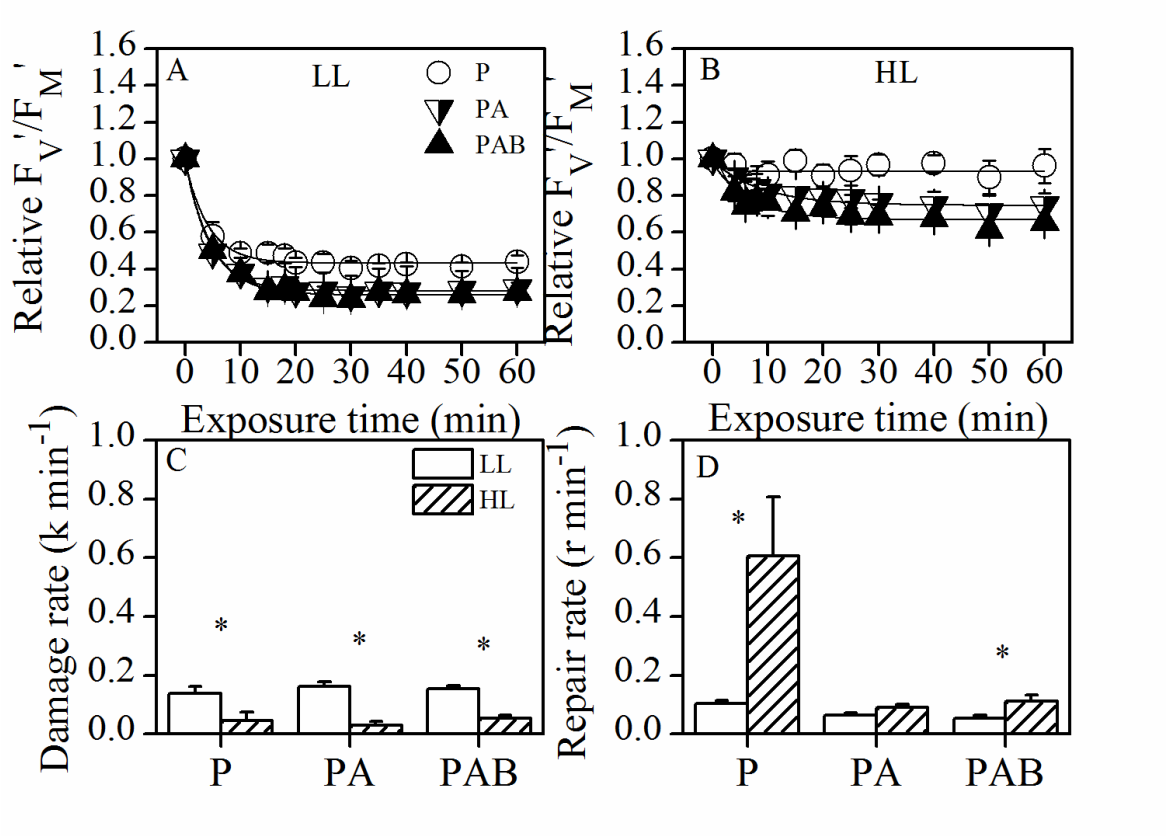
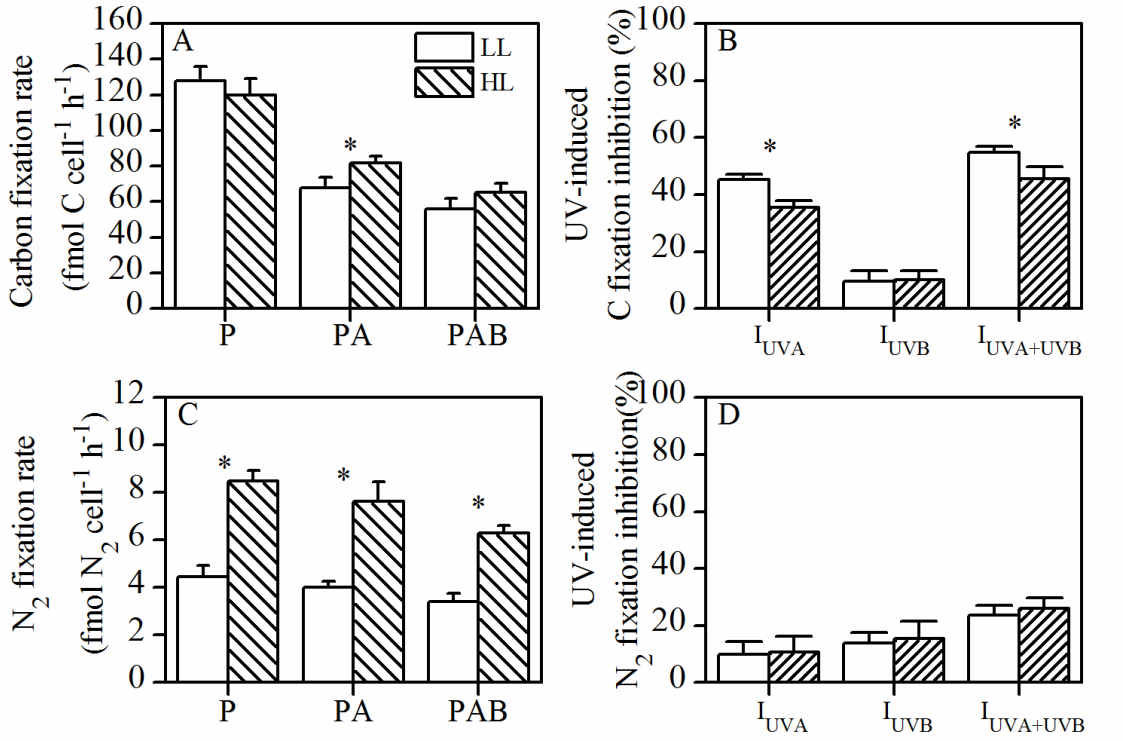


Fig.1 Changes of effective quantum yield (F_V'/F_M') of *Trichodesmium* IMS101 grown under (A) LL and (B) HL conditions while exposed to PAR (P), PAR+UVA (PA) and PAR+UVA+UVB (PAB) under solar stimulator for 60 min. PSII damage (C; k, in min^{-1}) and repair rates (D; r, in min^{-1}) of LL- and HL-grown cells were derived from the yield decline curve in the upper panels. Asterisks above the histogram bars indicate significant differences between LL- and HL-grown cells. Values are the mean ±SD, triplicate incubations.



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564 Fig.2 Photosynthetic carbon fixation rate (A; fmol C $cell^{-1}$ h^{-1}) and UV-induced C
565 fixation inhibition (B), N_2 fixation rate (C; fmol N_2 $cell^{-1}$ h^{-1}) and corresponding UV-
566 induced N_2 fixation inhibition (D) of *Trichodesmium* IMS101 grown under LL and HL
567 conditions. Asterisks above the histogram bars indicate significant differences between
568 LL- and HL-grown cells. Values are the mean ±SD, triplicate incubations.

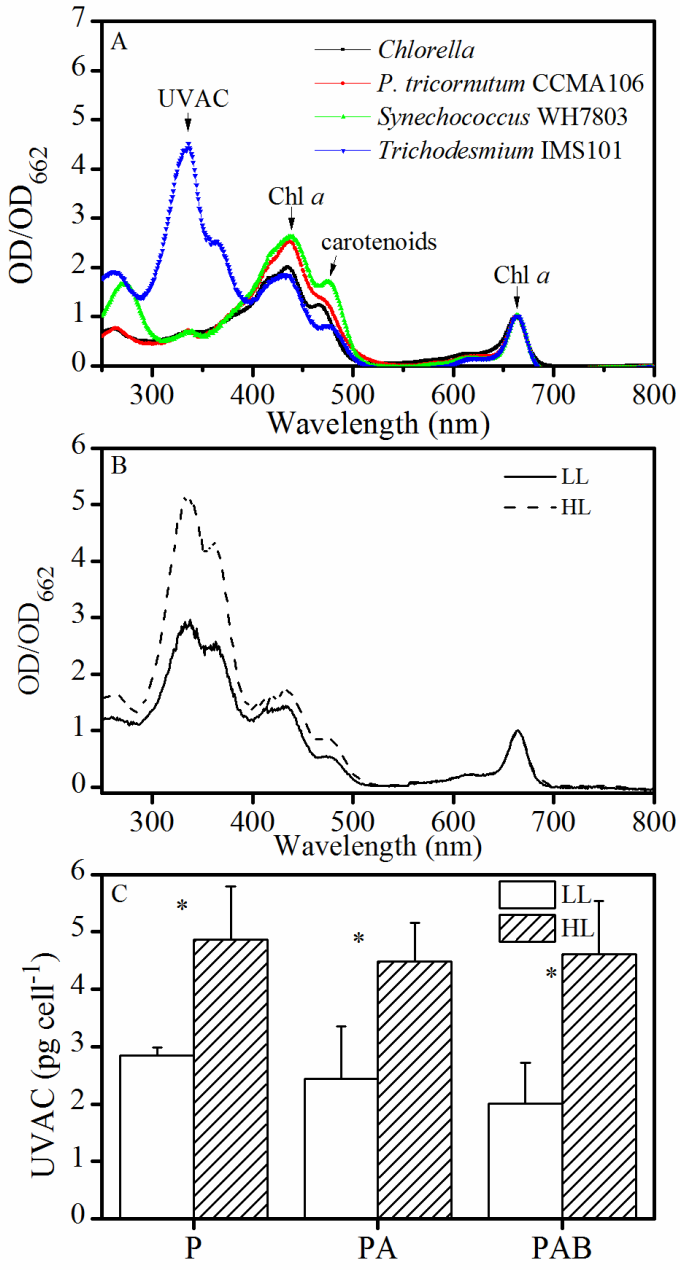

Fig.3 (A) Absorption spectrum of *Trichodesmium* IMS101 compared to other phytoplankton. Pigments were extract by 100% methanol. OD value normalized to OD_{662} (Chl *a*). (B) Absorption spectrum of the *Trichodesmium* IMS101 grown under LL and HL conditions, OD value normalized to OD_{662} (Chl *a*). (C) Cellular contents of UVACs of *Trichodesmium* IMS101 grown under LL and HL conditions after exposure to PAR (P), PAR+UVA (PA), PAR+UVA+UVB (PAB) under solar stimulator for 10 h. Asterisks above the histogram bars indicate significant differences between LL- and HL-grown cells. Values are the mean ±SD, triplicate incubations.

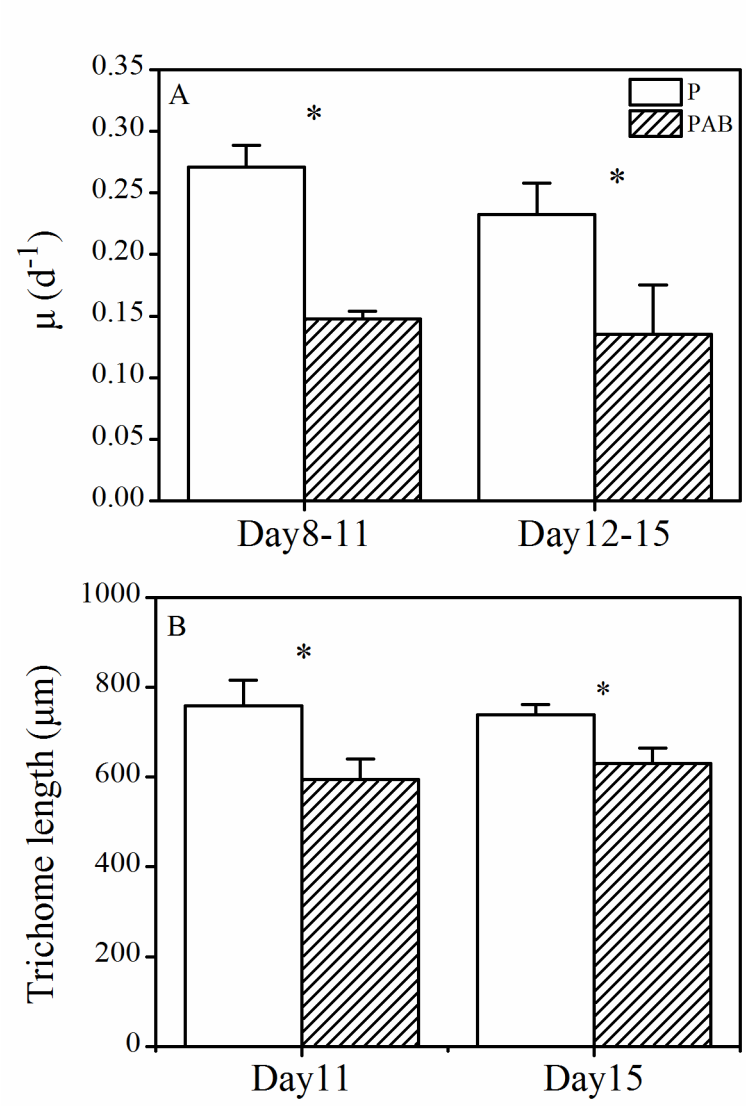

Fig.4 (A) Specific growth rate (measured during 8th-11th and 12th-15th day) of *Trichodesmium* IMS101 grown under solar PAR (P) and PAR+UVA+UVB (PAB). Corresponding total solar doses from Day 8 to Day 11 and from Day 12 to Day 15 were 17.03 and 18.51 MJ, respectively. (B) Trichome length (measured on the 11th and 15th day) of *Trichodesmium* IMS101 grown under solar PAR (P) and PAR+UVA+UVB (PAB). The asterisks indicate significant differences between radiation treatments. Values are the mean ±SD, triplicate cultures.

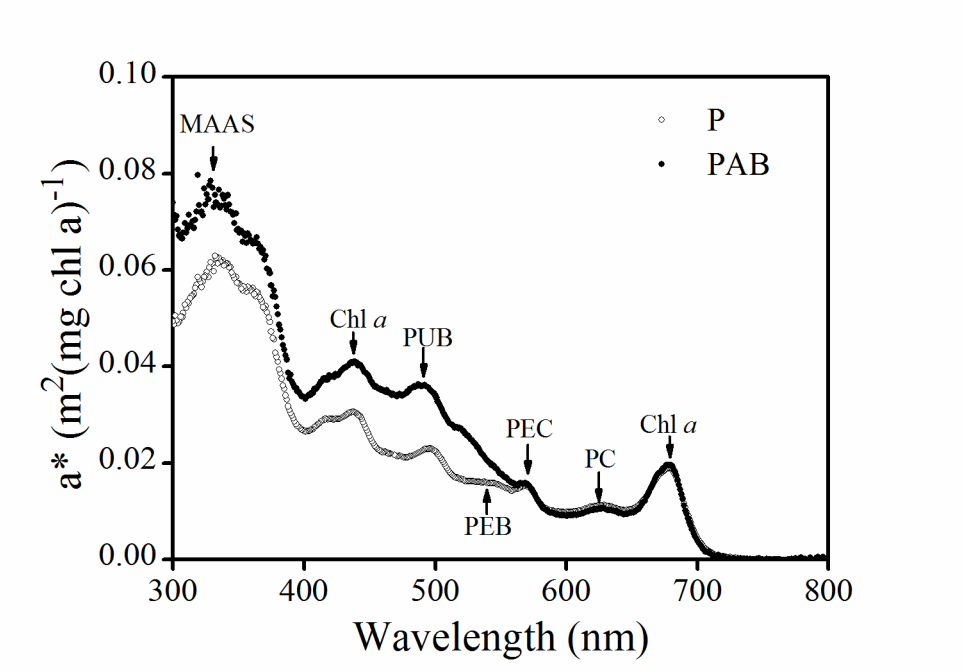

Fig.5 Chl *a* specific absorption spectrum (a^*) of *Trichodesmium* IMS101 grown under solar PAR (P) and PAR+UVA+UVB (PAB). The measurements were taken on the 18th day. The absorption peaks of MAAs (330 nm), PUB (495 nm), PEB (545 nm), PEC (569 nm), PC (625nm) and Chl *a* (438 and 664 nm) are indicated.

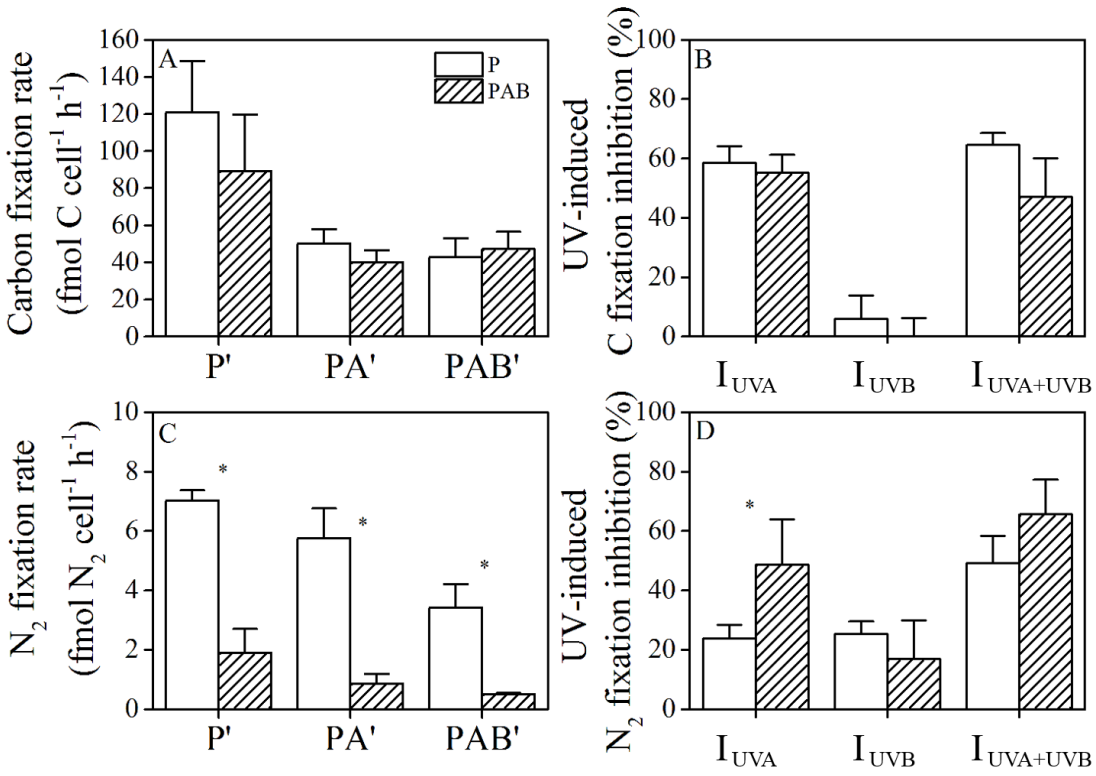

Fig. 6 Photosynthetic carbon fixation rate (A; fmol C $cell^{-1}$ h^{-1}) and UV-induced C fixation inhibition (B), N_2 fixation rate (C; fmol N_2 $cell^{-1}$ h^{-1}) and corresponding UV-induced N_2 fixation inhibition (D) of *Trichodesmium* IMS101 grown under solar PAR (P) and PAR+UVA+UVB (PAB) transferred to another P′, PA′, PAB′ treatments. The measurement was taken on the 18th day at 11:00~13:00. Asterisks above the histogram bars indicate significant differences between P and PAB treatments. Values are the mean ±SD, triplicate incubations.

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