

## Referee 1

*The study presents a new model development and calibration to an interesting horizontally heterogeneous system. It is based on an impressive compiled data set of observations and derivations of relevant inputs and state variables to compare. The main conclusion is that the observed increases in SOM stocks in an agroforest system are due to higher litter input compared to an agricultural control. The modelling exercise is interesting to the soil modelling community, and to the community researching interactions of vegetation components and management. I state several main points followed by more detailed comments.*

**Response:** We thank you for your interest in our work, we really appreciated your comments and suggestions. We tried to take into account all your comments and corrected the manuscript on the requested points.

**NOTE for responses: Pages and lines refer here to the marked-up manuscript version.**

### 1 Main points

*1) One calibration aspect of the study that convinces me (and probably other readers as well) of the validity of the study is currently not well highlighted. The model was calibrated to the control plot only. Despite of simplifying assumptions on similarities in climate and vertical transport between the control and the agroforestry system, the model predicted the differing C-stocks in tree rows and alleys and its depth distribution well. This is a strong validation.*

**Response:** Thanks for this rewarding comment. We better highlighted this result in the abstract “The model was calibrated to the control plot only...The model was strongly validated, describing properly the measured SOC stocks and distribution with depth in agroforestry tree rows and alleys”. (P2L30-34), but also in the discussion part “Despite these simplifying assumptions on similarities in climate but also on vertical transport between the control and the agroforestry system, the model calibrated to the control plot was able to reproduce SOC stocks in tree rows and alleys and its depth distribution well. This strong validation also suggests that OC inputs is the main driver of SOC storage, and that a potential effect of agroforestry microclimate on SOC mineralization is of minor importance” (P47L896-902).

*2) The quantification of the priming effect (PE) seems to be a bit complicated with running the no-PE model variant with a decomposition rate that was calibrated with the PE-model variant. To my opinion there are more straightforward quantifications already in the data (see detailed comments). I suggest highlighting the result that the priming model variant in Fig 4 was able to capture the depth distribution of C-stocks while no-PE model variant did not.*

**Response:** We tried to better describe how the PE intensity was quantified (P29-30L635-657) (see below) and why we chose this calculation method. We also highlighted in the abstract the fact that only the PE model was able to describe SOC profiles “Moreover, only a priming effect variant of the model was able to capture the depth distribution of SOC stocks” (P2L36-37).

3) While the mathematical model is well described, information is missing on the solution of the forward model, i.e. the solution of the presented partial-differential equation given a set of parameters. Which method has been used? What was the spatial grid, the same grid as the measurements? Was this grid sufficient to represent the steep concentration gradient in the top soil? Have different grid sizes been tested?

**Response:** Partial-differential equations were solved using the R package *deSolve* and the *ode.ID* method (Soetaert et al., 2010) (P28L624-625). The spatial grid was as close as possible to the measurements. Due to some field difficulties, the sampling grid is not totally regular but the modelling grid is. We indeed implicitly assumed this resolution to be sufficient to represent the steep concentration but we did not deeply evaluate the effect of different grid size but if really needed we can provide an analysis in the supplementary material

4) To my understanding of the study, the increased C stocks at the walnut tree lines are explained in a big part to increase of the above-ground carbon input by the herbaceous summer vegetation between trees (Fig. 3). I would like to read some discussion on this point. Was there an organic layer?

**Response:** Yes, this is absolutely true, the herbaceous vegetation growing between trees in the tree rows plays an important role on SOC storage. This was very much suggested by previous works on SOC storage in these systems (Cardinael et al., 2015, 2017), but proven here with the quantification of OC inputs. We now discussed this point more into details: “The increased SOC stocks in the tree rows were explained in a big part by an important above-ground carbon input (2.13 t C ha<sup>-1</sup> yr<sup>-1</sup>) by the herbaceous vegetation between trees. This result had already been suggested by Cardinael et al., (2015b) and by Cardinael et al., (2017) who showed that even young agroforestry systems could store SOC in the tree rows while trees are still very small. These “grass strips” indirectly introduced by the tree planting in parallel tree rows have a major impact on SOC stocks of agroforestry systems” (P48L925-931).

As commonly observed on grass strips, there was a very thin organic layer (maximum 0.3 cm thick), but not permanent during the season. Climatic conditions are very favorable for litter decomposition there, and we therefore assumed that this thin organic layer did not significantly change moisture and temperature conditions for the below mineral soil.

## 2 Detailed comments

*L 412: Instead of interpolating parameters of several fits, I suggest fitting a single equation to the entire dataset with an additional variable “distance to tree” and parameters a and b depend on this distance. However, the simplified procedure here seems to work and this point does not affect the conclusions.*

**Response:** Yes, this is indeed another possibility. As we were able to well reproduce root profiles with this simplified method, we think it is not really necessary to look for another equation as it would indeed not change the conclusions.

*L 444: Please specify exactly which observations and which predictions have been used for calibration.*

**Response:** We used SOC stocks measured in 2013 in the control plot (observations) and predicted SOC stocks (predictions) for the calibration. These stocks were considered at equilibrium (P26L568-572).

*Table 7: The prior knowledge in eq. 19 was specified as normal distribution. Table 7 instead reports a range of values instead of a mean and a variance (xb and diagonal of Pb in equation 1). Moreover many ranges span several orders of magnitudes suggesting that the parameters should be log-transformed before estimation. Where does the variance of the posterior come from? And what is the meaning of “prior values” in the posterior column?*

*Table 7: Where did the prior information come from? Are these uninformative priors or does it affect the results if you take different priors?*

**Response:** We acknowledge that this point was not clear enough. The optimization procedure that we used is sensitive to local minima. We therefore performed 30 optimization procedures starting with different parameter prior values to check that the results did not correspond to a local minimum. The prior range presented in Table 7 represents the range in which prior values were sampled for the 30 optimizations, it is therefore normal that they span several orders of magnitudes. The prior values presented in brackets in the posterior column represent the prior values that minimized the  $\mathbf{J}(\mathbf{x})$  value. The variance of the posterior is based on Santaren et al., 2007 (GBC 21, GB2013). The BFGS algorithm does not directly calculate variance of posteriors. To obtain them, we quantified the variance using the curvature cost function at its minimum once it was reached.

We clarified it in the text: “To determine an optimal set of parameters which minimizes  $\mathbf{J}(\mathbf{x})$ , we used the BFGS gradient-based algorithm (Tarantola, 1987). For each model variant, we performed 30 optimizations starting with different parameter prior values to check that the results did not correspond to a local minimum. As the BFGS algorithm does not directly calculate the variance of posteriors, they were quantified using the curvature cost function at its minimum once it was reached (Santaren et al., 2007).” (P27L589-595), and in the Table 7 (now Table 5) footnote: “The prior range represents the range in which prior values were sampled for the 30 optimizations per model variant. The prior values presented in brackets in the posterior column represent the prior values that minimized the  $\mathbf{J}(\mathbf{x})$  value (Eq. (34)).” (P39L824-825).

*Eq 21: Please explain the derivation. Usually the  $BIC = \ln(n)k - 2\log(L)$ , which involves the Likelihood instead of the mean squared deviation. From a Bayesian perspective  $-2\log(L) \propto J_{data}(p)$ , where  $J_{data}(p)$  is the first term of  $\mathbf{J}$  of eq. 19 (excluding the prior term).*

**Response:** Here, we used the MSD to estimate the maximum likelihood. This is indeed not the classical BIC. This approach is similar to Manzoni et al., 2012 (SBB 50, 66-76) who used the residual sum of square to estimate the maximum likelihood. We rephrase to clarify: “where  $N$  is the number of observations,  $MSD$  is the mean squared deviation used to estimate the maximum likelihood, and  $k$  is the number of model parameters” (P28L604-605).

*L 478: Please, clarify terminology of spin-up vs model calibration. To my understanding you calibrated 4 or 5 parameters depending on the three model variants so that equilibrium stocks,*

*i.e. simulations after 5000 years, were close to observed C-stocks (n=?) of the control plot in 2013. I suggest putting this content to the calibration section.*

**Response:** We moved this paragraph to the optimization procedure section and we clarified the terminology of spin-up vs model calibration: “These four or five parameters were calibrated so that equilibrium SOC stocks, i.e. after 5000 years of simulation, equaled SOC stocks of the control plot in 2013. The associated uncertainty was estimated with the 93 soil cores sampled in the control plot (see section 2.2.1). Due to a lack of relevant data, we assumed that the climate and the land use were the same for the last 5000 years, and that SOC stocks in the control plot were at equilibrium at the time of measurement. Therefore, SOC stocks at the end of the 5000 years of simulation equaled SOC stocks in the control plot. Three different calibrations were performed, corresponding to the three different models that were used: one calibration with the two pools model without the priming effect, one calibration with the two pools model with the priming effect, and one calibration with the three pools model” (P26L564-575). “SOC pools were initialized after a spin-up of 5000 years in the control plot. At  $t_0$ , SOC stocks in the agroforestry plot therefore equaled SOC stocks of the control plot” (P28L620-622).

*L 508: This derivation of the effect of priming is hard to grasp. To my opinion its more straightforward is compare predictions of the PE-variant model versus the non- PE variant; each consistently calibrated and applied for prediction:*

- *Effects of litter inputs: predictions of no-priming variant only: agroforestry stocks vs control stocks*
- *Combined effect: prediction of the priming model variant only: at agroforestry plot versus the control plot*
- *Effects of priming only: prediction of the priming model variant versus the predictions of the no-priming variant for the agroforestry system*

*Since the profile was not matched well with the no-priming model one can focus on sums.*

**Response:** We agree that the calculation was not straightforward and we clarified it in the new version (see below). Nevertheless, we consider our calculation as the most correct even though it is a bit complex to understand it. Indeed, we can not directly compare the different versions of the model to calculate priming because the decomposition rate of a classical first order kinetics takes implicitly into account a fixed fraction of decomposition due to priming. In all situations, there are regular inputs inducing priming and when we optimized the decomposition rate parameter in the control plot we implicitly represented this priming but at a fixed rate. Therefore comparing the different versions of the model would not estimate the priming in the agroforestry plots.

“Furthermore, at equilibrium state (i.e. when the input rate is constant) the decomposition rate of a first order equation (Eq. (6)) takes *PE* implicitly into account. Indeed, when FOC enters the system, there is an induced priming, a constant FOC input rate therefore induces a constant priming. This means that when we optimized the decomposition rate parameter in the control plot, we implicitly represented this priming but at a fixed rate. When FOC inputs are modified,

due to the tree growth for instance, the PE intensity is modified and this effect cannot be represented by classical first order kinetics.” (P29L635-642).

“To estimate the change of SOC decomposition rate due to priming when trees are planted, the decomposition fluxes predicted by Eq. (7)  $\left(-k_{HSOC,z} \times (1 - e^{-PE \times FOC_{t,z,d}})\right)$  in the agroforestry plot must be compared to the fluxes in agroforestry plot using the decomposition from the control plot calculated by Eq. (7) with  $FOC_{t,z,d}$  corresponding to the FOC inputs in the control plot. Thus, to calculate the importance of priming on SOC storage when trees are planted, we used the decomposition rates calculated following Eq. (7) in the control plot and we applied this decomposition rate to the agroforestry plot as a classical first order kinetics (without the FOC control, i.e.  $k_{new} = k_{HSOC,z} \times (1 - e^{-PE \times FOC_{t,z,d}})$  with  $FOC_{t,z,d}$  fixed constant)”. (P29L642-653).

*Fig 3: Please, note that the largest above ground input comes from herbaceous vegetation. Is this an important aspect for C-stocks of the agroforestry system?*

**Response:** Yes, this is definitely an important aspect for C-stocks in the agroforestry system. We added the following sentence to the result section: “In the agroforestry plot, the largest aboveground OC input to the soil comes from the herbaceous vegetation, and not from the trees” (P28-29L636-637).

*L698 (3.4.2): Please, remind the reader that C-stocks of the agroforestry plot were not part of model calibration (that used the control plot only) but are used here for validation.*

**Response:** As suggested, we added the following sentence at the beginning of the section: “As a reminder, SOC stocks of the agroforestry plot were not part of model calibration (that used the control plot only) but were used here for validation” (P36L800-801).

*Fig. 4: This is a nice demonstration of priming formulation being able to match the depth-shape. Although uncertainty of the mean (standard error) is low due to the high sample number, you may add the standard deviation across 93 measurements in order to get an impression of the variability.*

*I would like to see a figure, where C-depth profiles can be compared between cases without being dispersed across facets. Maybe zoom in to 5 to 15 stock range.*

**Response:** Yes, the uncertainty of the mean is extremely low for measured SOC stocks, as suggested we instead added the standard deviation of measurements (P43L857-858).

Concerning the C-depth profiles of Fig 4., this was actually our first idea. But SOC profiles are extremely close, especially between the control and the alleys, and the figure was very messy. We would therefore prefer to stick to this presentation, which is much clearer, even if we have to compare different facets.

*Fig. 5: Please, use a color scale with a clear zero.*

**Response:** We changed the color scale as requested. We also added a 2D graph of modeled control and agroforestry SOC stocks (P44).

*Fig. 6 Please, add difference in measured stocks to the “Inputs+PE” column for comparison.*

**Response:** Thanks for this suggestion, it was done (P46).

*L 753: Suggest: “Despite of these simplifying assumptions, the model calibrated to the control plot was able to ...”*

**Response:** This sentence was changed as follows: “Despite these simplifying assumptions on similarities in climate but also on vertical transport between the control and the agroforestry system, the model calibrated to the control plot was able to reproduce SOC stocks in tree rows and alleys and its depth distribution well. This strong validation also suggests that OC inputs is the main driver of SOC storage at this site, and that a potential effect of agroforestry microclimate on SOC mineralization is of minor importance” (P47L896-902).

## **Referee 2**

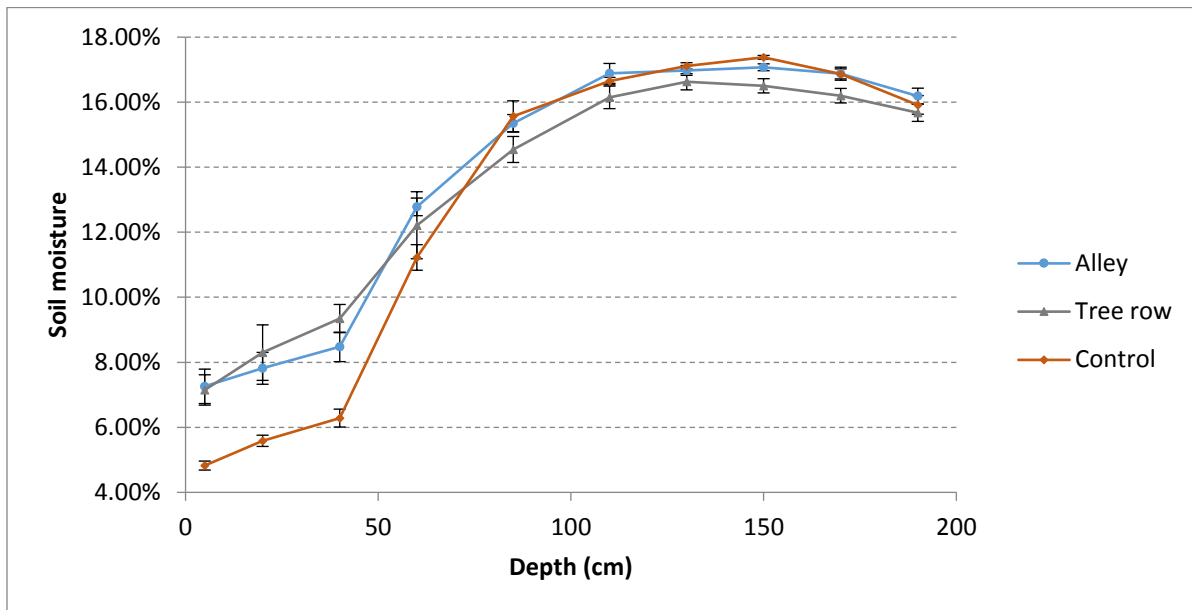
*This is a comprehensive study that uses an impressive set of field data to build a model for exploring agroforestry impacts on soil organic carbon (SOC). The topic is of interest and fits the scope of the journal. The combination of both field and modeling data is a key strength of this paper and provides interesting results regarding the spatial distribution of SOC in an agroforestry system. The modeling further highlights the potential negative impacts of priming on SOC storage. The methodology, results and most of the interpretation is sound. I therefore recommend this manuscript may be published after addressing the concerns and comments outlined below.*

**Response:** We thank you for your interest and you positive comments on our work.

*Major comments: 1) Due to lack of data, the authors assume that ‘soil temperature and soil moisture conditions were the same in the agroforestry tree rows, alleys and in the control plot (L388ff)’. Given the otherwise extensive data collection at this site it is surprising that these key variables have not been measured. As the authors acknowledge at various places, the impact of agroforestry on the SOC is primarily a result of the altered soil abiotic conditions. In my view the lack of these data hamper the understanding of the true controls and mechanisms responsible for change in SOC in the agroforestry system compared to the agricultural control field.*

**Response:** We agree with this comment, it is a pity that soil moisture and soil temperature sensors have not been installed in both fields, and on the long term. But this trial was first established to study crop yield and tree growth in association, and questions on SOC dynamics came very recently. In May 2013 (late Spring, about 15 days after the last rain), we sampled 40 soil cores in the tree rows, 60 in the alleys, and 93 in the control, and we measured soil moisture

on 23 of them. Soil cores were first taken in the agroforestry plot, and then in the control plot, under sunny conditions for both plots. The results showed that soil moisture was lower in the first 40 cm of soil in the control plot, but that there was no difference below:



During the last sampling day in the agroforestry plot, some cores were also taken in the control plot, and the same difference in terms of soil moisture was observed, suggesting that the lowest soil moisture in the control plot were not due to the sampling delay. Trees in the agroforestry plot probably slowed down the soil evaporation due to the shade. Most of the additional SOC storage in the agroforestry plot was observed in the topsoil. The lower topsoil soil moisture observed in the control in May 2013 would induce a reduction of SOC decomposition compared to the agroforestry plot, and then would reduce the observed SOC storage. But we can not conclude with this punctual observation, this phenomenon probably alternates during the season. For instance, we could hypothesize that in summer, deep soil will be drier in the agroforestry plot than in the control due to tree water absorption. Due to these uncertainties, we thought it was wiser to consider a mean annual soil temperature and moisture identical in both fields.

*The sensitivity analysis performed by the authors in an attempt to address this limitation cannot replace the missing information on soil abiotic controls since it merely reflects the model sensitivity to these parameters rather than their actual control on SOC. This shortcoming also limits some of the discussion. In my view, the related conclusions that ‘that OC inputs is the main driver of SOC storage (L752)’, that ‘a decrease of SOC mineralization due to the agroforestry microclimate is not obvious (L753)’ and that ‘soil microclimate in the agroforestry plot are not major drivers of the SOC storage (L766)’ are therefore not justified.*

**Response:** We tried to detail but also nuance our conclusions as suggested: “Despite these simplifying assumptions on similarities in microclimate but also on vertical transport between the control and the agroforestry system, the model calibrated to the control plot was able to reproduce SOC stocks in tree rows and alleys and its depth distribution well. This strong validation also suggests that OC inputs is the main driver of SOC storage at this site, and that a

potential effect of agroforestry microclimate on SOC mineralization is of minor importance.” (P47L896-902).

“A sensitivity analysis performed on these two boundary conditions showed that the model was not very sensitive to soil temperature and soil moisture (Fig. S4), but the real effect of these two parameters on SOC dynamics under agroforestry systems should be better investigated in future studies” (P48L912-915).

*2) The SOC stock is the product of C concentration per unit soil multiplied by the amount of soil per volume (i.e. bulk density). The study however is entirely focused on explaining changes in SOC due to changes in C concentration (as a result of C input/output) whereas changes in bulk density are not reported. It therefore remains unclear what the separate roles of changes in C concentration and bulk density are in controlling the changes in the total SOC stock (L743ff). While the authors acknowledge that the presence of trees (roots) could modify soil structure (L820), the effects of tree planting on such physical soil properties and subsequently SOC stocks are not well addressed in this study.*

**Response:** This is a very relevant point, soil bulk densities were only lower in the topsoil in the tree rows compared to the alleys and to the control plot. Bulk densities were published earlier (Cardinael et al., 2015b) and thus not reported here. In the model, we used the measured soil bulk densities for the control, tree rows and alleys from Cardinael et al., (2015b) (P8L172). We then expressed SOC stocks on an equivalent soil mass basis, and not at fixed depth. Therefore, the change in bulk density was implicitly taken into account in this study.

*3) The authors argue that the two pools model with priming effect was the best one, as shown by the BICs (Fig. 4, Table S1) (L704). However this is not true for the agroforestry alley which had a similar BIC and RMSE than the noPE model in Fig.4. Since the alley covers most of the area in an agroforestry system, this indicates that the priming effect might be overall less significant for this system as proposed by the authors.*

**Response:** In this case, alleys occupied 84% of the agroforestry area. The BIC and RMSE were lower with the PE model than with the noPE model as indicated in Fig. 4, but we acknowledge the difference is small. In the alleys, the first soil layer (0-10 cm) was worse represented by the PE model than by the noPE model. As the BIC is calculated on the whole profile, this bad fit impacts the BIC even if the PE model performs much better for the other soil layers, this is well shown in Table S2. We therefore think that the PE is needed to represent correctly the profile in the alley.

*4) Overall I find that the ms is too long, especially the method section is exhaustive (16 pages incl. Figures and Tables) but also parts of the results could be condensed. Given that the compilation of the C stock data is not a primary study goal (L118ff), I suggest that methods and results related to these data could be considerably shortened and partly moved into the supplementary part or refer to by references. For instance, data shown in Table 4 is already published (Cardinael et al., (2015b) and thus there is no need show this Table once more. Section 3.1 and 3.2, specifically the equations developed here should be moved to the Method*



*or Supplementary section. Details of Section 2.7 could also be moved to the Supplementary part.*

**Response:** We agree that the MS is very long, which is mainly due to the huge amount of data that are compiled here. Moreover, it also includes the differential equations of a new model, which we think are better to be presented in the main manuscript than in the supplementary. We however performed the following changes in order to shorten the description and facilitate comprehension:

Tree fine root biomass data previously shown in Table 4 were moved to the supplementary part (Table S1). Moreover, Section 3.1.1 “Carbon stock in the walnut tree biomass” and Table 3 were deleted as results were already presented in Fig. 3.

Section 3.1.2 “Tree growth” was moved to the Method part and merged with section 2.6.2 “Interpolation of tree growth” (P18L400-403).

Section 3.1.3 “Crop yield” was also moved to the Method part and merged with section 2.6.5 “Aboveground and belowground input from the crop” (P20-24L461-534).

Section 3.2.1 “Leaf litterfall” was moved to the Method part and merged with section 2.6.3 “Change of tree litterfall over time” (P18L407-417).

Section 3.2.2 “Tree fine root C input from mortality” was moved to the Method part and merged with section 2.6.4 “Tree fine root C input from mortality” (P19-20L420-458).

Section 3.2.3 “Aboveground carbon input from the crop” and section 3.2.4 were moved to the Method part and merged with section “Aboveground and belowground input from the crop” (P20-24L461-534).

Section 3.2.5 “Aboveground and belowground carbon inputs from the tree row herbaceous vegetation” was moved to the Method part and merged with section 2.6.6 “Aboveground and belowground input from herbaceous vegetation in the tree rows” (P25L541-557).

Section 3.2.6 “Organic carbon inputs and SOC stocks: a synthesis from field measurements” was however kept in the Results (now Section 3.1).

*Minor comments: Line 658: Here and at other places the authors use the word ‘globally’ which seems inappropriate in the given context.*

**Response:** “Globally” was replaced by “Overall” (P36L798 and P40L847).

*L706: ‘The spatial distribution of SOC storage was also well described (Fig. 5)’ – I disagree, Fig.5 shows the ‘additional’ SOC in the agroforestry system relative to control but not the absolute amount of SOC storage.*

**Response:** We now also added to Figure 5 both SOC stocks in the control and in the agroforestry plot (P44). This sentence was modified to “The spatial distribution of SOC stocks and of additional SOC storage was also well described (Fig. 5), with a very high additional SOC stock storage in the topsoil layer in the tree row” (P40L849-851).

*L725: 'The priming effect increases the decomposition rate when more FOC is available'– provide a reference for this statement or use past tense to indicate that this is a result from this study.*

**Response:** We added the reference of Fontaine et al., (2007) (P46L870-871).

*L772, 797, 873: At the several places the authors refer to 'the model' while several models (or model variations) were used in this study. Please clarify in each case which of the models (model variation) is meant when referring to one specific model.*

**Response:** We now specified it “the two pools model with priming effect” (P48L923, P49L954 and P52L1031).

*Figure 4: It would be helpful to add separate legends to the middle and right column sub-figures in Fig 4; also how is it possible that the model PE follows the measured SOC profile most closely but results in similar BIC than the noPE model?*

**Response:** As suggested, we added a common legend for all sub-figures at the bottom of Fig 4 (P42). In the alleys, The PE model has almost similar BIC than the noPE model only because the first soil layer (0-10 cm) was worse represented by the PE model:  $(\text{Model} - \text{Measures})^2 = 7.71$  compared to 1.28 kg/m<sup>3</sup> for the noPE model. As the BIC is calculated on the whole profile, this bad fit impacts the BIC even if the PE model performs much better for the other soil layers, this is well shown in Table S2.

1 **High organic inputs explain shallow and deep SOC storage in a long-term agroforestry**  
2 **system – Combining experimental and modeling approaches.**

3

4 Rémi Cardinael<sup>a,b,c\*</sup>, Bertrand Guenet<sup>d</sup>, Tiphaine Chevallier<sup>a</sup>, Christian Dupraz<sup>e</sup>, Thomas  
5 Cozzi<sup>b</sup>, Claire Chenu<sup>b</sup>

6

7 <sup>a</sup> IRD, UMR Eco&Sols, Montpellier SupAgro, 2 place Viala, 34060 Montpellier, France

8 <sup>b</sup> AgroParisTech, UMR Ecosys, Avenue Lucien Brétignières, 78850 Thiverval-Grignon, France

9 <sup>c</sup> CIRAD, UPR AIDA, Avenue d'Agropolis, 34398 Montpellier, France (present address)

10 <sup>d</sup> Laboratoire des Sciences du Climat et de l'Environnement, UMR CEA-CNRS-UVSQ, CE  
11 L'Orme des Merisiers, 91191 Gif-Sur-Yvette, France

12 <sup>e</sup> INRA, UMR System, Montpellier SupAgro, 2 place Viala, 34060 Montpellier, France

13 \* Corresponding author. Tel.: +33 04.67.61.53.08. E-mail address: remi.cardinael@cirad.fr

14

15 *Keywords:* priming effect, deep roots, deep soil organic carbon, spatial heterogeneity,  
16 silvoarable system, crop yield, SOC modeling

17

18 **Abstract**

19 Agroforestry is an increasingly popular farming system enabling agricultural diversification  
20 and providing several ecosystem services. In agroforestry systems, soil organic carbon (SOC)  
21 stocks are generally increased, but it is difficult to disentangle the different factors responsible  
22 for this storage. Organic carbon (OC) inputs to the soil may be larger, but SOC decomposition  
23 rates may be modified owing to microclimate, physical protection, or priming effect from roots,  
24 especially at depth. We used an 18-year-old silvoarable system associating hybrid walnut trees  
25 (*Juglans regia* × *nigra*) and durum wheat (*Triticum turgidum* L. subsp. *durum*), and an adjacent

26 agricultural control plot to quantify all OC inputs to the soil - leaf litter, tree fine root  
27 senescence, crop residues, and tree row herbaceous vegetation -, and measure SOC stocks down  
28 2 m depth at varying distances from the trees. We then proposed a model that simulates SOC  
29 dynamics in agroforestry accounting for both the whole soil profile and the lateral spatial  
30 heterogeneity. The model was calibrated to the control plot only.  
31 Measured OC inputs to soil were increased by about 40% (+ 1.11 t C ha<sup>-1</sup> yr<sup>-1</sup>) down to 2 m  
32 depth in the agroforestry plot compared to the control, resulting in an additional SOC stock of  
33 6.3 t C ha<sup>-1</sup> down to 1 m depth. The model was strongly validated, ~~described~~ describing properly  
34 the measured SOC stocks and distribution with depth in agroforestry tree rows and alleys. It  
35 showed that the increased inputs of fresh biomass to soil explained the observed additional SOC  
36 storage in the agroforestry plot. Moreover, only a priming effect variant of the model was able  
37 to capture the depth distribution of SOC stocks. Modeling revealed a strong priming effect that  
38 would reduce the potential SOC storage due to higher organic inputs in the agroforestry system  
39 by 75 to 90%. This result questions the potential of soils to store large amounts of carbon,  
40 especially at depth. Deep-rooted trees modify OC inputs to soil, a process that deserves further  
41 studies given its potential effects on SOC dynamics.

42

## 43 **1 Introduction**

44 Agroforestry systems are complex agroecosystems combining trees and crops or pastures  
45 within the same field (Nair, 1993, 1985; Somarriba, 1992). More precisely, silvoarable systems  
46 associate parallel tree rows with annual crops. Some studies showed that these systems could  
47 be very productive, with a land equivalent ratio (Mead and Willey, 1980) reaching up to 1.3  
48 (Graves et al., 2007). Silvoarable systems may therefore produce up to 30% more marketable  
49 biomass on the same area of land compared to crops and trees grown separately. This  
50 performance can be explained by a better use of water, nutrients and light by the agroecosystem

51 throughout the year. Trees grown in silvoarable systems usually grow faster than the same trees  
52 grown in forest ecosystems, because of their lower density, and because they also benefit from  
53 the crop fertilization (Balandier and Dupraz, 1999; Chaudhry et al., 2003; Chiffot et al., 2006).  
54 In temperate regions, farmers usually grow one crop per year, and this association of trees can  
55 extend the growing period at the field scale, especially when winter crops are intercropped with  
56 trees having a late bud break (Burgess et al., 2004). However, after several years, a decrease of  
57 crop yield can be observed in mature and highly dense plantations, especially close to the trees,  
58 due to competition between crops and trees for light, water, and nutrients (Burgess et al., 2004;  
59 Dufour et al., 2013; Yin and He, 1997).

60 Part of the additional biomass produced in agroforestry is used for economical purposes, such  
61 as timber or fruit production. Leaves, tree fine roots, pruning residues and the herbaceous  
62 vegetation growing in the tree rows will usually return to the soil, contributing to a higher input  
63 of organic carbon (OC) to the soil compared to an agricultural field (Peichl et al., 2006).

64 In such systems, the observed soil organic carbon (SOC) stocks are also generally higher  
65 compared to a cropland (Albrecht and Kandji, 2003; Kim et al., 2016; Lorenz and Lal, 2014).  
66 Cardinael *et al.*, (2017) measured a mean SOC stock accumulation rate of 0.24 (0.09-0.46) t C  
67 ha<sup>-1</sup> yr<sup>-1</sup> at 0-30 cm depth in several silvoarable systems compared to agricultural plots in  
68 France. Higher SOC stocks were also found in Canadian agroforestry systems, but measured  
69 only to 20 cm depth (Bambrick et al., 2010; Oelbermann et al., 2004; Peichl et al., 2006).

70 To our knowledge, we are still not able to disentangle the factors responsible for such a higher  
71 SOC storage. This SOC storage might be due to higher OC inputs but it could also be favored  
72 by a modification of the SOC decomposition owing to a change in SOC physical protection  
73 (Haile et al., 2010), and/or in soil temperature and moisture.

74 The introduction of trees in an agricultural field modifies the amount, but also the distribution  
75 of fresh organic carbon (FOC) input to the soil, both vertically and horizontally (Bambrick et

76 al., 2010; Howlett et al., 2011; Peichl et al., 2006). FOC inputs from the trees decrease with  
77 increasing distance from the trunk and with soil depth (Moreno et al., 2005). On the contrary,  
78 crop yield usually increases with increasing distance from the trees (Dufour et al., 2013; Li et  
79 al., 2008). Therefore, the proportions of FOC coming from both the crop residues and the trees  
80 change with distance from the trees, soil depth, and time.

81 Tree fine roots (diameter  $\leq 2$  mm) are the most active part of root systems (Eissenstat and Yanai,  
82 1997) and play a major role in carbon cycling. In silvoarable systems, tree fine root distribution  
83 within the soil profile is strongly modified due to the competition with the crop, inducing a  
84 deeper rooting compared to trees grown in forest ecosystems (Cardinael et al., 2015a; Mulia  
85 and Dupraz, 2006). Deep soil layers may therefore receive significant OC inputs from fine root  
86 mortality and exudates. Root carbon has a higher mean residence time in the soil compared to  
87 shoot carbon (Kätterer et al., 2011; Rasse et al., 2006), presumably because root residues are  
88 preferentially stabilized within microaggregates or adsorbed to clay particles. Moreover,  
89 temperature and moisture conditions are more buffered in the subsoil than in the topsoil. The  
90 microbial biomass is also smaller at depth (Eilers et al., 2012; Fierer et al., 2003), and the spatial  
91 segregation with organic matter is larger (Salomé et al., 2010) resulting in lower decomposition  
92 rates. Deep root carbon input in the soil could therefore contribute to a SOC storage with high  
93 mean residence times. However, some studies showed that adding FOC – a source of energy  
94 for microorganisms - to the subsoil enhanced decomposition of stabilized carbon, a process  
95 called « priming effect » (Fontaine et al., 2007). The priming effect is stronger when induced  
96 by labile molecules like root exudates than by root litter coming from the decomposition of  
97 dead roots (Shahzad et al., 2015). Therefore, the net effect of deep roots on SOC stocks has to  
98 be assessed, especially in silvoarable systems.

99 Models are crucial as they allow virtual experiments to best design and understand complex  
100 processes in these systems (Luedeling et al., 2016). Several models have been developed to

101 simulate interactions for light, water and nutrients between trees and crops (Charbonnier et al.,  
102 2013; Duursma and Medlyn, 2012; van Noordwijk and Lusiana, 1999; Talbot, 2011) or to  
103 predict tree growth and crop yield in agroforestry systems (Graves et al., 2010; van der Werf et  
104 al., 2007). However, none of these models are designed to simulate SOC dynamics in  
105 agroforestry systems and they are therefore not useful to estimate SOC storage. Oelbermann &  
106 Voroney (2011) evaluated the ability of the CENTURY model (Parton et al., 1987) to predict  
107 SOC stocks in tropical and temperate agroforestry systems, but with a single-layer modeling  
108 approach (0-20 cm). The approach of modeling a single topsoil layer assumes that deep SOC  
109 does not play an active role in carbon cycling, while it was shown that deep soil layers contain  
110 important amounts of SOC (Jobbagy and Jackson, 2000), and that part of this deep SOC could  
111 cycle on decadal timescales due to root inputs or to dissolved organic carbon transport (Baisden  
112 and Parfitt, 2007; Koarashi et al., 2012). The need to take into account deep soil layers when  
113 modeling SOC dynamics is now well recognized in the scientific community (Baisden et al.,  
114 2002; Elzein and Balesdent, 1995), and several models have been proposed (Braakhekke et al.,  
115 2011; Guenet et al., 2013; Koven et al., 2013; Taghizadeh-Toosi et al., 2014; Ahrens et al.,  
116 2015). Using vertically discretized soils is particularly important when modeling the impact of  
117 agroforestry systems on SOC stocks, but to our knowledge, vertically spatialized SOC models  
118 have not yet been tested for these systems.

119

120 The aims of this study were then twofold: (i) to propose a model of soil C dynamics in  
121 agroforestry systems able to account for both vertical and lateral spatial heterogeneities and (ii)  
122 to test whether variations of fresh organic carbon (FOC) input could explain increased SOC  
123 stocks both using experimental data and model runs.

124 For this, we first compiled data on FOC inputs to the soil obtained in a 18-year-old agroforestry  
125 plot and in an agricultural control plot in southern France, in which SOC stocks have been

126 recently quantified to 2 m depth (Cardinael et al., 2015b). FOC inputs comprised tree fine roots,  
127 tree leaf litter, aboveground and belowground biomass of the crop and of the herbaceous  
128 vegetation in the tree rows. We compiled recently published data for FOC inputs (Cardinael et  
129 al., 2015a; Germon et al., 2016), and measured the others (Table 1).

130

131 We then modified a two pools model proposed by Guenet *et al.*, (2013), to create a spatialized  
132 model over depth and distance from the tree, the CARBOSAF model (soil organic CARBOn  
133 dynamics in Silvoarable AgroForestry systems). Based on data acquired since the tree planting  
134 in 1995 (crop yield, tree growth), and on FOC inputs, we modeled SOC dynamics to 2 m depth  
135 in both the silvoarable and agricultural control plot. We evaluated the model against measured  
136 SOC stocks along the profile and used this opportunity to test the importance of priming effect  
137 (*PE*) for deep soil C dynamics in a silvoarable system. The performance of the two pools model  
138 including *PE* was also compared with a model version including three OC pools.

139

## 140 **2 Materials and methods**

### 141 **2.1 Study site**

142 The experimental site is located at the Restinclières farm Estate in Prades-le-Lez, 15 km North  
143 of Montpellier, France (longitude 04°01' E, latitude 43°43' N, elevation 54 m a.s.l.). The  
144 climate is sub-humid Mediterranean with an average temperature of 15.4°C and an average  
145 annual rainfall of 973 mm (years 1995–2013). The soil is a silty and carbonated (pH = 8.2) deep  
146 alluvial Fluvisol (IUSS Working Group WRB, 2007). In February 1995, a 4.6 hectare  
147 silvoarable agroforestry plot was established with the planting of hybrid walnut trees (*Juglans*  
148 *regia* × *nigra* cv. NG23) at a density of 192 trees ha<sup>-1</sup> but later thinned to 110 trees ha<sup>-1</sup>. Trees  
149 were planted at 13 m × 4 m spacing, and tree rows are East–West oriented. The cultivated alleys  
150 are 11 m wide. The remaining part of the plot (1.4 ha) was kept as an agricultural control plot.



151 Since the tree planting, the agroforestry alleys and the control plot were managed in the same  
 152 way. The associated crop is most of the time durum wheat (*Triticum turgidum* L. subsp. *durum*),  
 153 except in 1998, 2001 and 2006, when rapeseed (*Brassica napus* L.) was cultivated, and in 2010  
 154 and 2013, when pea (*Pisum sativum* L.) was cultivated. The soil is ploughed to a depth of 0.2  
 155 m before sowing, and the wheat crop is fertilized with an average of 120 kg N ha<sup>-1</sup> yr<sup>-1</sup>. Crop  
 156 residues (wheat straw) are also exported, but about 25% remain on the soil. Tree rows are  
 157 covered by spontaneous herbaceous vegetation. Two successive herbaceous vegetation types  
 158 occur during the year, one in summer and one in winter. The summer vegetation is mainly  
 159 composed of *Avena fatua* L., and is 1.5 m tall. In winter, the vegetation is a mix of *Achillea*  
 160 *millefolium* L., *Galium aparine* L., *Vicia* L., *Ornithogalum umbellatum* L. and *Avena fatua* L,  
 161 and is 0.2 m tall.

162

163 **Table 1.** Synthesis of the different field and laboratory data available or measured, and their  
 164 sources.

Description of the data	Source
Soil texture, bulk densities, SOC stocks	Cardinael <i>et al.</i> , (2015a)
Soil temperature and soil moisture	Measured
Tree growth (DBH)	Measured
Tree wood density	(Talbot, 2011)
Tree fine root biomass	Cardinael <i>et al.</i> , (2015b)
Tree fine root turnover	Germon <i>et al.</i> , (2016)
Crop yield and crop ABG biomass	Dufour <i>et al.</i> , (2013) and measured
Crop root biomass	Prieto <i>et al.</i> , (2015) and measured
Tree row herbaceous vegetation – ABG biomass	Measured
Tree row herbaceous vegetation – root biomass	Measured
Biomass carbon concentrations	Measured
Potential decomposition rate of roots	Prieto <i>et al.</i> , (2016a)
HSOC potential decomposition rate	Measured

165 DBH: Diameter at Breast Height; ABG: aboveground; OC: organic carbon; HSOC: humified  
 166 soil organic carbon.

167

## 168 2.2 Organic carbon stocks

## 169 **2.2.1 Soil organic carbon stocks**

170 SOC data have been published in Cardinael *et al.*, (2015a2015b). Briefly, soil cores were  
171 sampled down to 2 m depth in May 2013, 100 in the agroforestry plot, and 93 in the agricultural  
172 control plot. SOC concentrations, soil bulk densities, SOC stocks, and soil texture were  
173 measured for ten soil layers (0.0-0.1, 0.1-0.3, 0.3-0.5, 0.5-0.7, 0.7-1.0, 1.0-1.2, 1.2-1.4, 1.4-1.6,  
174 1.6-1.8, and 1.8-2.0 m). In the agroforestry plot, 40 soil cores were taken in the tree rows, while  
175 60 were sampled in the alleys at varying distances from the trees. Soil organic carbon stocks  
176 were quantified on an equivalent soil mass basis (Ellert and Bettany, 1995).

177

## 178 **2.2.2 Tree aboveground and stump carbon stocks**

179 Three hybrid walnuts were chopped down in 2012. The trunk circumference was measured  
180 every meter up to the maximum height of the tree to estimate its volume. The trunk biomass  
181 was estimated by multiplying the trunk volume by the wood density that was measured at 616  
182 kg m<sup>-3</sup> during a previous work at the same site (Talbot, 2011). Then, branches were cut, the  
183 stump was uprooted, and they were weighted separately. Samples were brought to the  
184 laboratory to determine the moisture content, which enabled calculation of the branches and the  
185 stump dry mass.

186

## 187 **2.3 Measurements of organic carbon inputs in the field**

### 188 **2.3.1 Carbon inputs from tree fine root mortality**

189 The tree fine root (diameter  $\leq$  2 mm) biomass was quantified and coupled with an estimate of  
190 the tree fine root turnover in order to predict the carbon input to the soil from the tree fine root  
191 mortality. A detailed description of the methods used to estimate the tree fine root biomass can  
192 be found in Cardinael *et al.*, (2015b2015a). In March 2012, a 5 (length)  $\times$  1.5 (width)  $\times$  4 m  
193 (depth) pit was open in the agroforestry plot, perpendicular to the tree row, at the North of the

194 trees. The tree fine root distribution was mapped down 4 m depth, and the tree fine root biomass  
195 was quantified in the tree row and in the alley. Only results concerning the first two meters of  
196 soil, among those obtained by Cardinael *et al.*, (~~2015b~~2015a) will be ~~presented~~used here.

197 In July 2012, sixteen minirhizotrons were installed in the agroforestry pit, at 0, 1, 2.5 and 4 m  
198 depth, and at two and five meters from the trees. The tree root growth and mortality was  
199 monitored during one year using a scanner (CI-600 Root Growth Monitoring System, CID,  
200 USA), and analyzed using the WinRHIZO Tron software (Régent, Canada). A detailed  
201 description of the methods and of results used to estimate the tree fine root turnover can be  
202 found in Germon *et al.*, (2016).

203

### 204 **2.3.2 Tree litterfall**

205 In 2009, the crowns of two walnut trees were packed with a net in order to collect the leaf  
206 biomass from September to January. The same was done in 2012 with three other walnut trees.  
207 The leaf litter was then dried, weighted and analyzed for C to quantify the leaf carbon input per  
208 tree.

209

### 210 **2.3.3 Aboveground and belowground input from the crop**

211 Since the tree planting in 1995, the crop yield was measured 14 times (in 1995, 2000, 2002,  
212 2003, 2004, 2005, 2007, 2008, 2009, 2010, 2011, 2012, 2013, and 2014), while the wheat straw  
213 biomass and the total aboveground biomass were measured six times (in 2007, 2008, 2009,  
214 2011, 2012, and 2014) in both the control and the agroforestry plot (Dufour *et al.*, 2013), using  
215 sampling subplots of 1 m<sup>2</sup> each. In the control plot, five subplots have been sampled while in  
216 the agroforestry plot five transects have been sampled. Each transect was made of three  
217 subplots, 2 m North from the tree, 2 m South from the tree, and 6.5 m from the tree (middle of  
218 the alley). In March 2012, a 2 m deep pit was opened in the agricultural control plot (Prieto *et*

219 al., 2015), and the root biomass was quantified to the maximum rooting depth (1.5 m). The  
220 root:shoot ratio of durum wheat was measured in the control plot. We assumed that the crop  
221 root biomass turns out once a year, after the crop harvest.

222

#### 223 **2.3.4 Above and belowground input from the tree row herbaceous vegetation**

224 As two types of herbaceous vegetation grow in the tree rows during the year, samples were  
225 taken in summer and winter. In late June 2014, twelve subplots of 1 m<sup>2</sup> each were positioned  
226 in the tree rows, around 4 walnut trees. In January 2015, six subplots of 1 m<sup>2</sup> each were  
227 positioned in the tree rows, around 2 walnut trees. The middle of each subplot was located at 1  
228 m, 2 m and 3 m, respectively, from the selected walnut tree. All the aboveground vegetation  
229 was collected in each square. In the middle of each subplot, root biomass was sampled with a  
230 cylindrical soil corer (inner diameter of 8 cm). Soil was taken at three soil layers, 0.0-0.1, 0.1-  
231 0.3 and 0.3-0.5 m. In the laboratory, soil was gently washed with water through a 2 mm mesh  
232 sieve, and roots were collected. Roots from the herbaceous vegetation were easily separated  
233 manually from walnut roots, as they were soft and yellow compared to walnuts roots that were  
234 black. After being sorted out from the soil and cleaned, the root biomass was dried at 40°C and  
235 measured.

236

#### 237 **2.4 Carbon concentration measurements**

238 All organic carbon measurements were performed with a CHN elemental analyzer (Carlo Erba  
239 NA 2000, Milan, Italy), after samples were oven-dried at 40°C for 48 hours (Table 2). Dry  
240 biomasses (t DM ha<sup>-1</sup>) of each organic matter inputs were multiplied by their respective organic  
241 carbon concentrations (mg C g<sup>-1</sup>) to calculate organic carbon stocks (t C ha<sup>-1</sup>).

242

243 **Table 2.** Organic carbon concentrations and C:N ratio of the different types of biomass.

Type of biomass	Organic C concentration (mg C g <sup>-1</sup> )	C:N	Number of replicates
Walnut trunk	445.7 ± 1.0	159.1 ± 25.2	3
Walnut branches	428.6 ± 1.7	62.2 ± 11.7	3
Wheat straw	433.2 ± 0.7	55.5 ± 2.1	5
Wheat root	351.4 ± 19	24.8 ± 2.1	8
Walnut leaf	449.4 ± 3.7	49.1 ± 0.4	3
Walnut fine root	437.0 ± 3.3	28.6 ± 3.4	8
Summer vegetation (ABG)	448.4 ± 1.9	37.8 ± 2.2	5
Summer vegetation (roots)	314.5 ± 8.3	33.8 ± 1.7	6
Winter vegetation (ABG)	447.7 ± 5.3	11.2 ± 0.4	3
Winter vegetation (roots)	397.4 ± 5.0	24.7 ± 0.7	3

244 The organic matter called “vegetation” stands for the herbaceous vegetation that grows in the  
245 tree row. ABG: aboveground. Errors represent standard errors.

246

## 247 **2.5 General description of the CARBOSAF model**

### 248 **2.5.1 Organic carbon decomposition**

249 We adapted a model developed by Guenet et al. (2013) where total SOC is split in two pools,  
250 the FOC and the humified soil organic carbon (HSOC) for each soil layer (Fig. 1a). Input to the  
251 FOC pool comes from the plant litter and the distribution of this input within the profile is  
252 assumed to depend upon depth from the surface ( $z$ ), distance from the tree ( $d$ ), and time ( $t$ ).  
253 Equations describing inputs to the FOC pool ( $I_{t,z,d}$ ) at a given time, depth, and distance are  
254 fully explained in the Results.

255

256 The FOC mineralisation is assumed to be governed by first order kinetics, being proportional  
257 to the FOC pool, as given by:

$$258 \quad \frac{\partial FOC_{t,z,d}}{\partial t} = -k_{FOC} \times FOC_{t,z,d} \times f_{clay,z} \times f_{moist,z} \times f_{temp,z} \quad (1)$$

259 where  $FOC_{t,z,d}$  is the FOC carbon pool (kg C m<sup>-2</sup>) at a given time ( $t$ , in years), depth ( $z$ , in m)  
260 and distance ( $d$ , in m), and  $k_{FOC}$  is its decomposition rate. The potential decomposition rates of  
261 the different plant materials were assessed with a 16-week incubation experiment during a  
262 companion study at the site (Prieto et al., 2016). The decomposition rate  $k_{FOC}$  was weighted by

263 the respective contribution of each type of plant litter as a function of the tree age, soil depth  
 264 and distance from the tree. The rate modifiers  $f_{clay,z}$ ,  $f_{moist,z}$  and  $f_{temp,z}$  are functions depending  
 265 respectively on the clay content, soil moisture and soil temperature at a given depth  $z$ , and range  
 266 between 0 and 1.

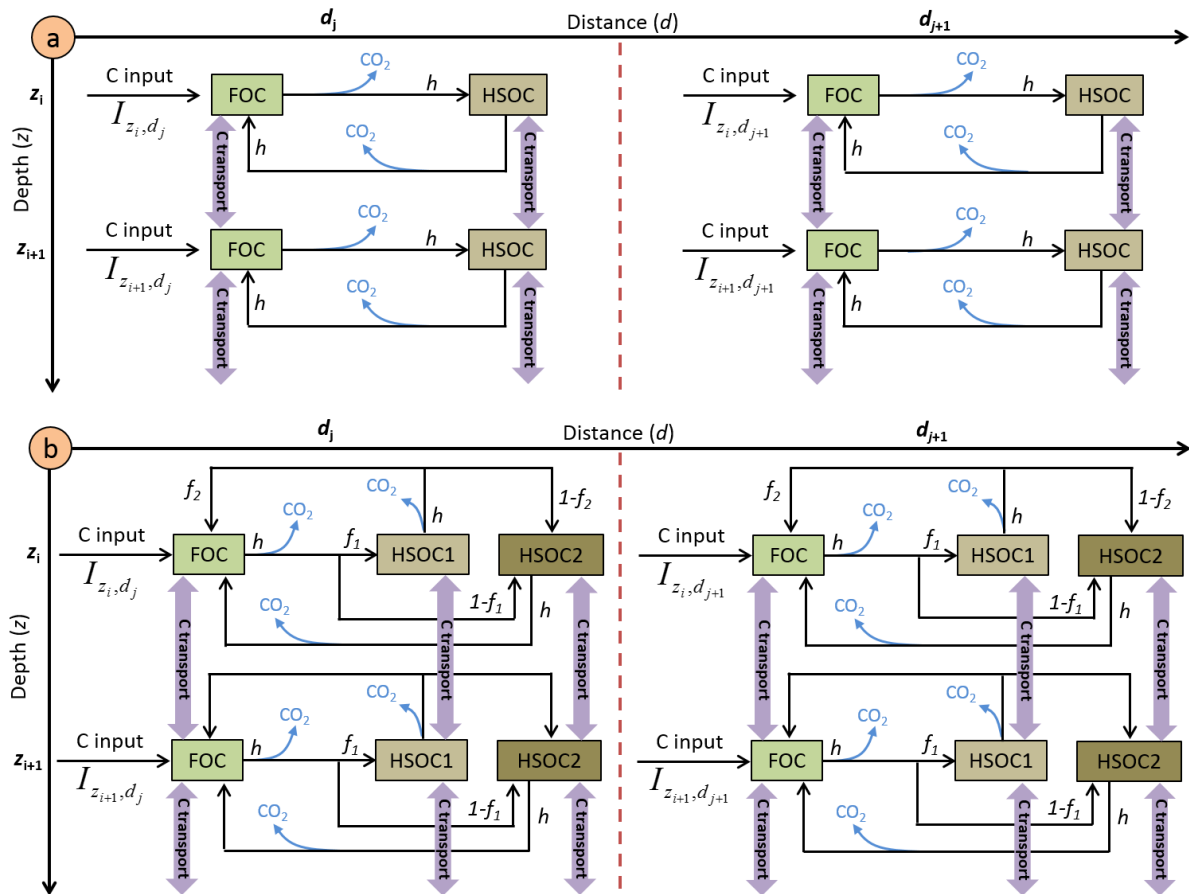
267

268 The  $f_{clay}$  function originated from the CENTURY model (Parton et al., 1987):

269 
$$f_{clay,z} = 1 - 0.75 \times Clay_z \quad (2)$$

270 where  $Clay_z$  is the clay fraction (ranging between 0 and 1) of the soil at a given depth  $z$ .

271



272

273 **Fig. 1.** Schematic representation of the pools and the fluxes of the (a) two pools model and (b)

274 three pools model.

275

276 The  $f_{moist,z}$  function originated from the meta-analysis of Moyano *et al.*, (2012) and is affected  
 277 by soil properties (clay content, SOC content). Briefly, the authors fitted linear models on 310  
 278 soil incubations to describe the effect of soil moisture on decomposition. Then, they normalized  
 279 such linear models between 0 and 1 to apply these functions to classical first order kinetics. All  
 280 details are described in Moyano *et al.*, (2012). To save computing time, we calculated  $f_{moist,z}$   
 281 only once using measured SOC stocks instead of using modelled SOC stocks and repeated the  
 282 calculation at each time step.

283

284 The temperature sensitivity of the soil respiration is expressed as  $Q_{10}$ :

$$285 \quad f_{temp,z} = Q_{10}^{\frac{temp_z - temp_{opt}}{10}} \quad (3)$$

286 with  $temp_z$  being the soil temperature in K at each soil depth  $z$  and  $temp_{opt}$  a parameter fixed to  
 287 304.15 K. The  $Q_{10}$  value was fixed to 2, a classical value used in models (Davidson and  
 288 Janssens, 2006).

289

290 Once the FOC is decomposed, a fraction is humified ( $h$ ) and another is respired as  $CO_2$  ( $1-h$ )  
 291 (Fig. 1a) following equations (4) and (5).

$$292 \quad \text{Humified } FOC_{t,z,d} = h \times \frac{\partial FOC_{t,z,d}}{\partial t} \quad (4)$$

$$293 \quad \text{Respired } FOC_{t,z,d} = (1 - h) \times \frac{\partial FOC_{t,z,d}}{\partial t} \quad (5)$$

294

295 Two mathematical approaches are available in the model to describe the mineralisation of  
 296 HSOC: a first order kinetics, as given by Eq. (6) or an approach developed by Wutzler &  
 297 Reichstein, (2008) and by Guenet *et al.*, (2013) introducing the priming effect, i.e., the  
 298 mineralisation of HSOC depends on FOC availability, and given by Eq. (7):

$$299 \quad \frac{\partial HSOC_{t,z,d}}{\partial t} = -k_{HSOC,z} \times HSOC_{t,z,d} \times f_{moist,z} \times f_{temp,z} \quad (6)$$

300 
$$\frac{\partial HSOC_{t,z,d}}{\partial t} = -k_{HSOC,z} \times HSOC_{t,z,d} \times (1 - e^{-PE \times FOC_{t,z,d}}) \times f_{moist,z} \times f_{temp,z} \quad (7)$$

301 where  $HSOC_{t,z,d}$  is the humified SOC carbon pool at a given time ( $t$ , in years), depth ( $z$ , in m)  
 302 and distance ( $d$ , in m),  $k_{HSOC,z}$  is its decomposition rate ( $\text{yr}^{-1}$ ) at a given depth  $z$ , and  $PE$  is the  
 303 priming effect parameter. The parameters  $f_{moist,z}$  and  $f_{temp,z}$  are functions depending respectively  
 304 on soil moisture and soil temperature at a given depth  $z$ , and affecting the decomposition rate  
 305 of HSOC. They correspond to the moisture equation from Moyano *et al.*, (2012) and to Eq. (3),  
 306 respectively. The decomposition rate  $k_{HSOC,z}$  was an exponential law depending on soil depth  
 307 ( $z$ ) as shown by an incubation study (see paragraph *HSOC decomposition rate* further in the  
 308 M&M):

309 
$$k_{HSOC,z} = a \times e^{-b \times z} \quad (8)$$

310 The  $b$  parameter of this equation represented the ratio of labile C/stable C within the HSOC  
 311 pool. The effect of clay on HSOC decomposition was implicitly taken into account in this  
 312 equation as clay content increased with soil depth.

313 A fraction of decomposed HSOC returns to the FOC assuming that part of the HSOC  
 314 decomposition products is as labile as FOC ( $h$ ) and another is respired as  $\text{CO}_2$  (Fig. 1a) in the  
 315 two pools model.

316

317 Finally, we also developed an alternative version of the model with three pools by splitting the  
 318 HSOC pools into two pools with different turnover rates, HSOC2 being more stabilized than  
 319 HSOC1 (Fig. 1b). The non-respired decomposed FOC is split between HSOC1 and HSOC2  
 320 following a parameter  $f_1$ . The non-respired decomposed HSOC1 is split between HSOC2 and  
 321 FOC following a parameter  $f_2$  whereas non-respired decomposed HSOC2 is only redistributed  
 322 into the FOC pools. The decomposition of HSOC1 and HSOC2 both follow the equation (8)  
 323 but with different parameter values for  $a$ .

324



## 325 **2.5.2 Carbon transport mechanisms**

326 The transport of C between the different soil layers was represented by both advection and  
327 diffusion mechanisms (Elzein and Balesdent, 1995), which have been shown to usually describe  
328 well the C transport in soils (Bruun et al., 2007; Guenet et al., 2013). The advection represents  
329 the C transport due to the water infiltration in the soil, while the diffusion represents the C  
330 transport due to the fauna activity. The same transport coefficients were applied to the two C  
331 pools, FOC and HSOC.

332

333 The advection is defined by:

$$334 \quad F_A = A \times C \quad (9)$$

335 where  $F_A$  is the flux of C transported downwards by advection, and  $A$  is the advection rate (mm  
336  $\text{yr}^{-1}$ ).

337

338 The diffusion is represented by the Fick's law:

$$339 \quad F_D = -D \times \frac{\partial^2 C}{\partial z^2} \quad (10)$$

340 where  $F_D$  is the flux of C transported downwards by diffusion,  $-D$  the diffusion coefficient ( $\text{cm}^2$   
341  $\text{yr}^{-1}$ ) and  $C$  the amount of carbon in the pool subject to transport (FOC or HSOC).

342

343 To represent the effect of soil tillage ( $z \leq 0.2$  m), we added another diffusion term using the  
344 Fick's law but with a value of  $D$  several orders of magnitude higher to represent the mixing due  
345 to tillage. It must be noted that no tillage effect on the decomposition was represented here  
346 because of the large unknowns on these aspects (Dimassi et al., 2013; Virto et al., 2012).

347

348 In this model, the flux of C transported downwards by the advection and diffusion ( $F_{AD}$ ) was  
349 represented as the sum of both mechanisms, following Elzein & Balesdent (1995):

350 
$$F_{AD} = F_A + F_D \quad (11)$$

351

352 The FOC and HSOC pools dynamics in the two pools model correspond to:

353 
$$\frac{\partial FOC}{\partial t} = I_{t,z,d} + \frac{\partial F_{AD}}{\partial z} + h \times \frac{\partial HSOC_{t,z,d}}{\partial t} - \frac{\partial FOC_{t,z,d}}{\partial t} \quad (12)$$

354 
$$\frac{\partial HSOC}{\partial t} = \frac{\partial F_{AD}}{\partial z} + h \times \frac{\partial FOC_{t,z,d}}{\partial t} - \frac{\partial HSOC_{t,z,d}}{\partial t} \quad (13)$$

355

356 Finally, the FOC, HSOC1 and HSOC2 pools dynamics in the three pools model correspond to:

357 
$$\frac{\partial FOC}{\partial t} = I_{t,z,d} + \frac{\partial F_{AD}}{\partial z} + h \times f_2 \times \frac{\partial HSOC1_{t,z,d}}{\partial t} + h \times \frac{\partial HSOC2_{t,z,d}}{\partial t} - \frac{\partial FOC_{t,z,d}}{\partial t} \quad (14)$$

358 
$$\frac{\partial HSOC1}{\partial t} = \frac{\partial F_{AD}}{\partial z} + h \times f_1 \times \frac{\partial FOC_{t,z,d}}{\partial t} - \frac{\partial HSOC1_{t,z,d}}{\partial t} \quad (15)$$

359 
$$\frac{\partial HSOC2}{\partial t} = \frac{\partial F_{AD}}{\partial z} + h \times (1 - f_1) \times \frac{\partial FOC_{t,z,d}}{\partial t} + h \times (1 - f_2) \times \frac{\partial HSOC1_{t,z,d}}{\partial t}$$
  

360 
$$- \frac{\partial HSOC2_{t,z,d}}{\partial t} \quad (16)$$

361

### 362 **2.5.3 Depth dependence of HSOC potential decomposition rates**

363 The shape of the function (i.e. the  $b$  parameter) describing the HSOC potential decomposition  

364 rate (Eq. (8)) was determined by incubating soils from the control, the alley and the tree row,  

365 and from different soil layers (0.0-0.1, 0.1-0.3, 0.7-1.0 and 1.6-1.8 m). Soils were sieved at 5  

366 mm, and incubated during 44 days at 20°C at a water potential of -0.03 MPa. Evolved CO<sub>2</sub> was  

367 measured using a micro-GC at 1, 3, 7, 14, 21, 28, 35, 44 days. The three first measurement  

368 dates corresponded to a pre-incubation period and were not included in the analysis. For a given  

369 depth, the cumulative mineralised SOC was expressed as a percentage of total SOC and was  

370 plotted against the incubation time. The slopes represented the potential SOC mineralisation  

371 rate at a given soil depth and location. The potential SOC mineralisation rates were then plotted

372 against soil depth (Fig. S1). We used the soil incubations to determine only the  $b$  parameter of  
373 the curve: with such short term incubations, the SOC decomposition rate over the soil profile  
374 is overestimated because the  $\text{CO}_2$  measured during the incubations mainly originates from the  
375 labile C pool. The  $a$  parameter was optimized following the procedure described further.

376

## 377 **2.6 Boundary conditions of the CARBOSAF model**

### 378 **2.6.1 Annual aggregates of soil temperature and soil moisture**

379 In April 2013, eight soil temperature and moisture sensors (Campbell CS 616 and Campbell  
380 107, respectively) were installed in the agroforestry plot at 0.3, 1.3, 2.8 and 4.0 m depth, and at  
381 2 and 5 m from the trees. Soil temperature and moisture were measured for 11 months.

382 The mean annual soil temperature in the agroforestry plot was described by the following  
383 equation:

$$384 \quad T = -0.89 \times z + 288.24 \quad (R^2 = 0.99) \quad (17)$$

385 where  $T$  is the soil temperature (K) and  $z$  is the soil depth (m).

386

387 The mean annual soil moisture was described with the following equation:

$$388 \quad \theta = 0.05 \times z + 0.28 \quad (R^2 = 0.99) \quad (18)$$

389 where  $\theta$  is the soil volumetric moisture ( $\text{cm cm}^{-3}$ ) and  $z$  is the soil depth (m).

390 Due to a lack of data in the agricultural plot, we assumed that the soil temperature and the soil  
391 moisture were the same in the agroforestry tree rows, alleys and in the control plot, but we  
392 further performed a sensitivity analysis of the model on these two parameters.

393

### 394 **2.6.2 Interpolation of tree growth**

395 The tree growth has been measured in the field since the establishment of the experiment. We  
396 used the diameter at breast height ( $DBH$ ) as a surrogate of the tree growth preferentially to the

397 tree height as the field measurements were more accurate. Indeed, *DBH* is easier to measure  
398 than height, especially when trees are getting older. To describe the temporal dynamic of *DBH*  
399 since the tree planting, a linear equation was fitted on the data.

400 Tree growth measurements enabled us to fit the following equation that was used in the model:

$$401 \quad DBH_t \begin{cases} 0.01, & t \leq 3 \\ 0.0157 \times t - 0.0391 & (R^2 = 0.997) \quad 3 < t \leq 20 \end{cases} \quad (2219)$$

402 where  $DBH_t$  is the diameter at breast height (m) and  $t$  represents the time since tree planting  
403 (years).

### 405 2.6.3 Change of tree litterfall over time

406 For the five walnut trees where the leaf biomass was quantified, *DBH* was also measured. The  
407 ratio between the leaf biomass and *DBH* was then calculated for the five replicates. ~~A linear~~  
408 ~~relationship between the leaf biomass and *DBH* was then considered to describe the increase of~~  
409 ~~the leaf litter C input with the tree growth.~~ Total leaf biomass was  $8.96 \pm 1.45$  kg DM tree<sup>-1</sup> and  
410 the carbon concentration of walnut leaves was  $449.4 \pm 3.7$  mg C g<sup>-1</sup> (Table 2). With a density  
411 of 110 trees ha<sup>-1</sup>, leaf litterfall was estimated at  $0.73 \pm 0.06$  t C ha<sup>-1</sup> in 2012 and at the plot scale.  
412 The ratio between leaf biomass and *DBH* was  $0.0277 \pm 0.0024$  t C tree<sup>-1</sup> m<sup>-1</sup> or  $3.05$  t C ha<sup>-1</sup> m<sup>-1</sup>  
413 . The following linear relationship was therefore used in the model to describe leaf litter C  
414 input with the tree growth:

$$415 \quad L_t = 3.05 \times DBH_t \quad (260)$$

416 where  $L_t$  is the leaf litter input (t C ha<sup>-1</sup>) at the year  $t$ , and  $DBH_t$  the diameter at breast height  
417 (m) the year  $t$ .

### 419 2.6.4 Tree fine root C input from mortality

In 2012, the measured tree fine root biomass was higher in the tree row than in the alley (Table S1). From 0 to 1 m distance from the tree (in the tree row), the tree fine root biomass was homogeneous and was 1.01 t C ha<sup>-1</sup> down 2 m depth.

~~A decreasing exponential function was fitted on the root biomass data obtained from the pit in 2012 to describe total fine root biomass (TFRB) down to 2 m depth as a function of distance from the tree.~~ In 2012 and in the alley, the tree fine root biomass (TFRB) decreased with increasing distance from the tree and was represented by an exponential function:

$$TFRB = \begin{cases} 1.01, & 0 \leq d \leq 1 \\ 1.29 \times e^{-0.28 \times d} \quad (R^2 = 0.90), & 1 < d \leq 6.5 \end{cases} \quad (271)$$

where  $TFRB$  represents tree fine root biomass down 2 m depth (t C ha<sup>-1</sup>), and  $d$  the distance from the tree (m).

We considered a linear increase of  $TFRB$  with increasing  $DBH$ , and a linear regression was performed between  $TFRB$  in 2012 and  $TFRB$  in 1996, the first year after planting (biomass considered as negligible). The following linear relationship was used to simulate  $TFRB$  as a function of tree growth:

$$TFRB_{t,d} = \begin{cases} 3.69 \times DBH_t, & 0 \leq d \leq 1 \\ 4.70 \times DBH_t \times e^{-0.28 \times d}, & 1 < d \leq 6.5 \end{cases} \quad (282)$$

where  $TFRB_t$  represents the tree fine root biomass to 2 m depth (t C ha<sup>-1</sup>) at the year  $t$ ,  $DBH_t$  the diameter at breast height (m) at the year  $t$ , and  $d$  the distance to the tree (m).

A changing distribution of tree fine roots within the soil profile was taken into account with increasing distance to the tree. For this purpose, exponential functions ( $a \times e^{-b \times z}$ ) were fitted in the alley every 0.5 m distance, and a linear regression was fitted between their coefficients  $a$  and  $b$  and distance from the tree. However, the distribution of  $TFRB$  within the soil profile and with the distance to the tree was considered constant with time.

A decreasing exponential function best represented the changing distribution of tree fine roots within the soil profile with increasing distance to the tree:

$$p_{TFRB,z,d} = \begin{cases} 13.92 \times e^{-1.39 \times z} & (R^2 = 0.68), & 0 \leq d \leq 1 \\ a \times e^{-b \times z}, & & 1 < d \leq 6.5 \end{cases} \quad (293)$$

and

$$a = 10.31 - 1.15 \times d \quad (R^2 = 0.69) \quad (3024)$$

$$b = -1.10 + 0.19 \times d \quad (R^2 = 0.51) \quad (3125)$$

Finally,

$$p_{TFRB,z,d} = \begin{cases} 13.92 \times e^{-1.39 \times z}, & 0 \leq d \leq 1 \\ (10.31 - 1.15 \times d) \times e^{-(-1.10+0.19 \times d) \times z}, & 1 < d \leq 6.5 \end{cases} \quad (326)$$

where  $p_{TFRB,z,d}$  is the proportion (%) of the total tree fine root biomass (TFRB) at a given depth  $z$  (m), and at a distance  $d$  from the tree (m).

To finally estimate the tree fine root input due to the mortality, TFRB was multiplied by the measured root turnover. The tree fine root turnover ranged from 1.7 to 2.8 yr<sup>-1</sup> depending on fine root diameter, with an average turnover of 2.2 yr<sup>-1</sup> for fine roots < 2 mm and to a depth of 2 m (Germon et al., 2016).

### **2.6.5 Aboveground and belowground input from the crop**

As there were more crop yield measurements (14) than straw biomass measurements (6), the effect of agroforestry on the crop yield with time was used as an estimate for change in the aboveground and belowground wheat biomass.

For this, the relative yield ( $Rel Y_{AF}$ ) in the agroforestry system was calculated for each year as the ratio between the agroforestry yield and the control yield ( $Y_C$ ). ~~A linear regression was then fitted between the relative yield and the DBH.~~

467 The average annual crop yield in the control plot was  $Y_C = 3.79 \pm 0.40$  t DM ha<sup>-1</sup> for the 14  
468 studied years. In the agroforestry plot, the average relative yield decreased linearly with time  
469 (increasing  $DBH$ ) and was described using the following linear equation (Fig. 2):

$$470 \quad Rel Y_{AF_t} = -93.33 \times DBH_t + 100 \quad (R^2 = 0.12, \quad p - value = 0.02) \quad (273)$$

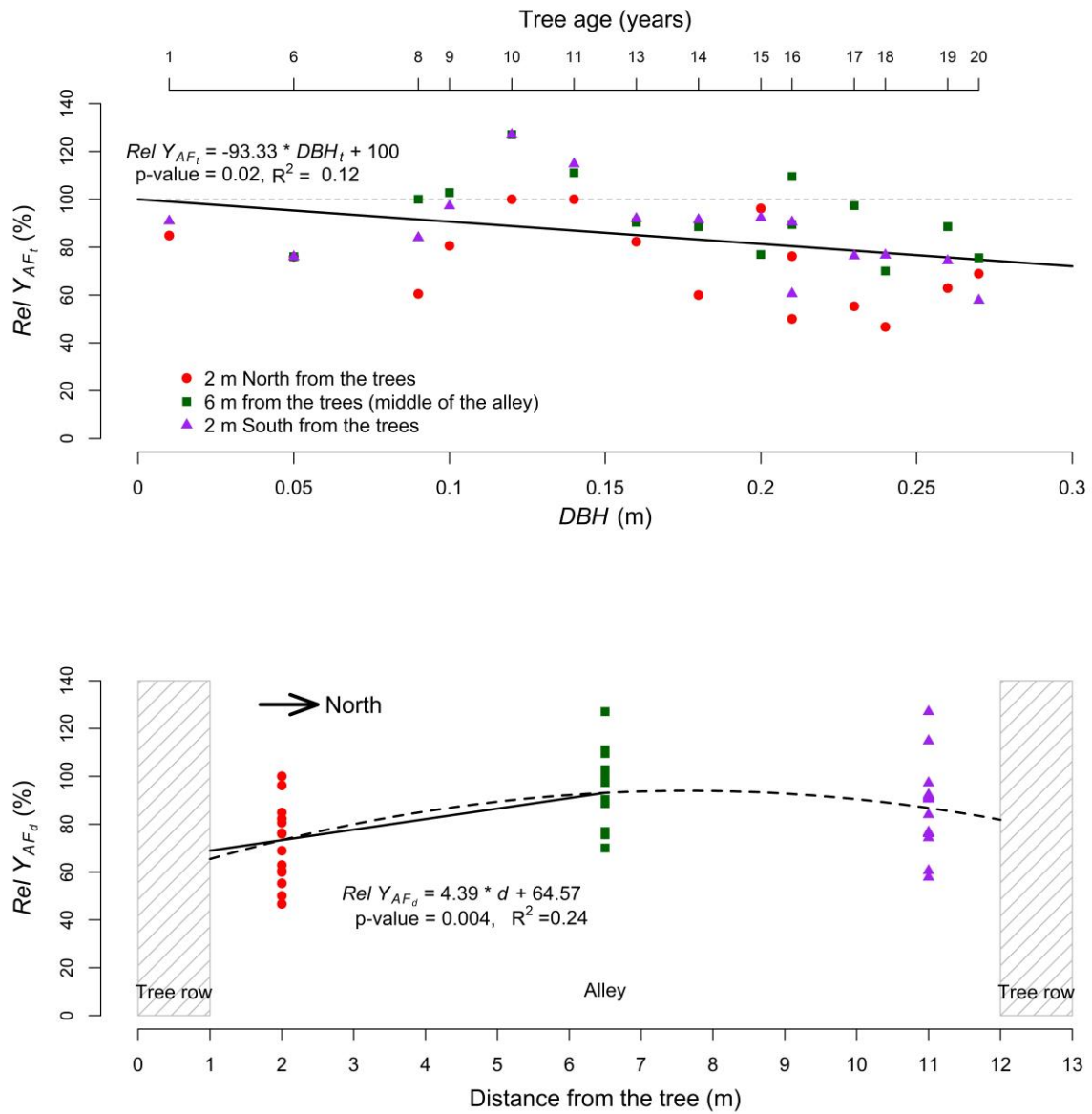
471 where  $Rel Y_{AF_t}$  is the average relative crop yield (%) in the agroforestry plot compared to the  
472 control plot at year  $t$ , and  $DBH_t$  is the diameter at breast height (m) at year  $t$ .

473  
474 The variation of crop yield with distance from the trees was described with a quadratic equation  
475 (Fig. 2). But as we aimed to predict SOC stocks up to 6.5 m distance from the trees (middle of  
476 the alley), a linear increase of crop yield with increasing distance from the tree gave similar  
477 results as the quadratic equation over the 6.5 m distance and was more parsimonious: ~~In the~~  
478 ~~agroforestry plot, a linear relationship was used to describe the relative crop yield increase from~~  
479 ~~the tree to the middle of the alley (Fig. 2):~~

$$480 \quad Rel Y_{AF_d} = 4.39 \times d + 64.57 \quad (R^2 = 0.24), \quad 1 < d \leq 6.5 \quad (284)$$

481 where  $Rel Y_{AF_d}$  is the relative crop yield (%) in the agroforestry plot at a distance  $d$  (m) from  
482 the tree compared to the control plot.

483



484

485 Fig. 2. Top: Relative yield ( $Rel Y_{AF_t}$ ) in the agroforestry plot compared to the control plot as a

486 function of tree growth, represented by the diameter at breast height ( $DBH$ ) at year  $t$ .

487 Bottom: Relative yield ( $Y_{AF_{t,d}}$ ) as a function of the distance from the tree.

488

489

490 Finally, the crop yield in the agroforestry plot was modeled as follows:

491 
$$Y_{AF_{t,d}} = Rel Y_{AF_t} \times Y_C \times Rel Y_{AF_d} \quad (R^2 = 0.19), \quad 1 < d \leq 6.5 \quad (295)$$



492 where  $Y_{AF,t,d}$  is the crop yield (t DM ha<sup>-1</sup>) in the agroforestry plot at the year  $t$  and at a distance  
493  $d$  (m) from the tree. Because three linear equations were used to describe the crop yield in the  
494 agroforestry plot, errors were accumulated and we finally came up with a standard  
495 underestimation of the crop yield in the agroforestry plot that we corrected by multiplying our  
496 equation by 1.2.

497  
498 ~~Finally,  $t$~~ The ratio between the straw biomass and the crop yield was calculated as the average  
499 of the six measurements, and was considered constant with time. This ratio was used to convert  
500 crop yield into straw biomass. In the agroforestry plot, the carbon input to the soil from the  
501 aboveground crop biomass was:

$$502 \quad ABC_{crop,t,d} = Y_{AF,t,d} \times (\text{straw biomass: crop yield}) \times C_{straw} \times (1 - \text{export}) \quad (330)$$

503 where  $ABC_{crop,t,d}$  is the aboveground carbon input from the crop (t C ha<sup>-1</sup>) at the year  $t$  and  
504 distance  $d$  from the tree,  $Y_{AF,t,d}$  is the agroforestry crop yield. The average ratio between the  
505 straw biomass (t DM ha<sup>-1</sup>) and the crop yield (t DM ha<sup>-1</sup>) equaled  $1.03 \pm 0.11$  (n=6). The wheat  
506 straw was exported out of the field after the harvest, but it was estimated that 25% of the straw  
507 biomass was left on the soil, thus  $\text{export}=0.75$ . In the control plot,  $Y_{AF,t,d}$  was replaced by  $Y_{C_s}$

508  
509 To estimate fine root biomass of the crop, we hypothesized that the root:shoot ratio of the durum  
510 wheat was the same in both the agroforestry and agricultural plot, in the absence of any  
511 published data on the matter. In the agroforestry plot, the belowground crop biomass was  
512 represented by:

$$513 \quad BEC_{crop,t,d} = Y_{AF,t,d} \times (\text{shoot: crop yield}) \times (\text{root: shoot}) \times C_{root} \quad (31)$$

514 where  $BEC_{crop,t,d}$  is the belowground crop biomass (t C ha<sup>-1</sup>) at the year  $t$  and at a distance  $d$   
515 from the tree,  $Y_{AF,t,d}$  is the agroforestry crop yield. The average ratio between the total crop

516 aboveground biomass (*shoot*) and the crop yield equaled  $2.45 \pm 0.15$  (n=6). In 2012, total fine  
 517 root biomass was  $2.29 \pm 0.32$  t C ha<sup>-1</sup> in the control (Table 3).

518

519 **Table 3.** Wheat fine root biomass in the agricultural control plot in 2012.

<u>Soil depth (m)</u>	<u>Wheat fine root biomass</u>	
	<u>(kg C m<sup>-3</sup>)</u>	<u>(t C ha<sup>-1</sup>)</u>
<u>0.0-0.1</u>	<u><math>0.48 \pm 0.05</math></u>	<u><math>0.48 \pm 0.05</math></u>
<u>0.1-0.3</u>	<u><math>0.34 \pm 0.04</math></u>	<u><math>0.69 \pm 0.09</math></u>
<u>0.3-0.5</u>	<u><math>0.22 \pm 0.04</math></u>	<u><math>0.44 \pm 0.08</math></u>
<u>0.5-1.0</u>	<u><math>0.10 \pm 0.04</math></u>	<u><math>0.52 \pm 0.20</math></u>
<u>1.0-1.5</u>	<u><math>0.03 \pm 0.04</math></u>	<u><math>0.17 \pm 0.19</math></u>
<u>Total</u>	<u>=</u>	<u><math>2.29 \pm 0.32</math></u>

520 Errors represent standard errors.

521

522 Therefore, the wheat *root:shoot* ratio equaled  $0.79 \pm 0.12$  (n=1). The carbon concentration of  
 523 wheat root was  $C_{root} = 35.14 \pm 1.90$  mg C g<sup>-1</sup>. In the control plot,  $Y_{AF,t,d}$  was replaced by  $Y_C$ .

524

525 In 2012, no wheat roots were observed below 1.5 m, and root biomass decreased exponentially  
 526 with increasing depth (Table 3). The distribution of crop roots within the soil profile was  
 527 described as follows:

$$528 \quad p_{CRBc,z} = \begin{cases} 26.44 \times e^{-2.59 \times z} & (R^2 = 0.99), & z \leq 1.5 \\ 0, & & z > 1.5 \end{cases} \quad (32)$$

529 where  $p_{CRBc,z}$  is the proportion (%) of total crop root biomass in the control plot at a given  
 530 depth  $z$  (m).

531 Since the same maximum rooting depth of the crop was observed in the agroforestry plot and  
 532 in the control plot, we inferred that the wheat root distribution within the soil profile was not  
 533 modified by agroforestry, but only its biomass. The crop root turnover was assumed to be 1 yr<sup>-1</sup>,  
 534 root mortality occurring mainly after crop harvest.

~~The wheat root distribution within the soil profile as a function of total wheat root biomass was described by an exponential fit. Since the same maximum rooting depth of the crop was observed in the agroforestry plot and in the control plot, we inferred that the wheat root distribution within the soil profile was not modified by agroforestry, but only its biomass.~~

## 2.6.6 Aboveground and belowground input from herbaceous vegetation in the tree rows

~~We fitted an exponential function to describe the herbaceous root biomass with depth. We assumed for simplification that the aboveground and belowground biomasses of the herbaceous vegetation in the tree row were constant over time.~~ The distance from the trees had no effect on the above and belowground biomass of the herbaceous vegetation (data not shown), therefore average values are presented. The summer aboveground biomass was almost three times higher than in winter, whereas the belowground biomass was two times higher (Table 4). The total aboveground carbon input was  $2.13 \pm 0.14 \text{ t C ha}^{-1} \text{ yr}^{-1}$  and the total belowground carbon input was  $0.74 \pm 0.05 \text{ t C ha}^{-1} \text{ yr}^{-1}$  to 0.5 m depth.

**Table 4.** Aboveground and belowground biomass of the herbaceous vegetation in the tree rows.

	Soil depth (m)	Herbaceous biomass (t C ha <sup>-1</sup> )	
		Summer	Winter
Aboveground	-	$1.57 \pm 0.11$	$0.56 \pm 0.09$
	0.0-0.1	$0.22 \pm 0.03$	$0.17 \pm 0.01$
Belowground	0.1-0.3	$0.16 \pm 0.02$	$0.06 \pm 0.01$
	0.3-0.5	$0.09 \pm 0.04$	$0.04 \pm 0.01$
	Total	$0.46 \pm 0.04$	$0.27 \pm 0.02$

Errors represent standard errors.

The belowground carbon input from the tree row vegetation ( $BEC_{veg,z}$ , t C ha<sup>-1</sup>) at a given depth  $z$  (m) was described by the following equation:

$$BEC_{veg,z} = \begin{cases} 0.44 \times e^{-3.12 \times z}, & z \leq 1.5 \\ 0, & z > 1.5 \end{cases} \quad (33)$$

556 We assumed for simplification that the aboveground and belowground biomasses of the  
557 herbaceous vegetation in the tree row were constant over time.

558

## 559 **2.7 Optimization procedure**

560 Depending on the model variant, four to Ffive parameters were optimized with a Bayesian  
561 statistical method (Santaren et al., 2007; Tarantola, 1987, 2005) using measured SOC stocks  
562 from the control plot only. These parameters were  $A$ , the advection rate,  $D$ , the diffusion  
563 coefficient,  $h$  the humification yield,  $a$  the coefficient of the  $k_{HSOC}$  rate from Eq. (10), and  $PE$   
564 the priming coefficient. These four or five parameters were calibrated so that equilibrium SOC  
565 stocks, i.e. after 5000 years of simulation, equaled SOC stocks of the control plot in 2013. ~~SOC~~  
566 ~~pools were initialized after a spin-up of 5000 years in the control plot. Measured SOC stocks in~~  
567 ~~2013 in the control plot were used for the spin-up.~~ The associated uncertainty was estimated  
568 with the 93 soil cores sampled in the control plot (see section 2.2.1). Due to a lack of relevant  
569 data, we assumed that the climate and the land use were the same for the last 5000 years, and  
570 that SOC stocks in the control plot were at equilibrium at the time of measurement. Therefore,  
571 SOC stocks at the end of the ~~spin-up~~5000 years of simulation equaled SOC stocks in the control  
572 plot. Three different ~~spin-ups~~calibrations were performed, corresponding to the three different  
573 models that were used: one ~~spin-up~~calibration with the two pools model without the priming  
574 effect, one ~~spin-up~~calibration with the two pools model with the priming effect, and one ~~spin-~~  
575 ~~up~~calibration with the three pools model.

576 TheEach model variant was fitted to the control SOC stocks data using a Bayesian curve fitting  
577 method described in Tarantola (1987), after a conversion from SOC stocks in  $\text{kg C m}^{-2}$  to SOC  
578 stocks in  $\text{kg m}^{-3}$  due to the different soil layers' thickness. We aimed to find a parameter set  
579 that minimizes the distance between model outputs and the corresponding observations,  
580 considering model and data uncertainties, and prior information on parameters. With the

581 assumption of Gaussian errors for both the observations and the prior parameters, the optimal  
 582 parameter set corresponds to the minimum of the cost function  $J(\mathbf{x})$ :

$$583 \quad J(\mathbf{x}) = 0.5 \times [(\mathbf{y} - \mathbf{H}(\mathbf{x}))^t \times \mathbf{R}^{-1} \times (\mathbf{y} - \mathbf{H}(\mathbf{x})) + (\mathbf{x} - \mathbf{x}_b)^t \times \mathbf{P}_b^{-1} \times (\mathbf{x} - \mathbf{x}_b)] \quad (1934)$$

584 that contains both the mismatch between modelled and observed SOC stock and the mismatch  
 585 between a priori and optimized parameters.  $\mathbf{x}$  is the vector of unknown parameters,  $\mathbf{x}_b$  the  
 586 vector of a priori parameter values,  $\mathbf{H}()$  the model and  $\mathbf{y}$  the vector of observations. The  
 587 covariance matrices  $\mathbf{P}_b$  and  $\mathbf{R}$  describe a priori uncertainties on parameters, and observations,  
 588 respectively. Both matrices are diagonal as we suppose the observation uncertainties and the  
 589 parameter uncertainties to be independent. To determine an optimal set of parameters which  
 590 minimizes  $J(\mathbf{x})$ , we used the BFGS gradient-based algorithm (Tarantola, 1987). ~~We For each~~  
 591 ~~model variant, we~~ performed 30 optimizations starting with different parameter prior values to  
 592 check that the results did not correspond to a local minimum. ~~To optimize the parameters we~~  
 593 ~~only used the data coming from the control plot. As the BFGS algorithm does not directly~~  
 594 ~~calculate the variance of posteriors, they were quantified using the curvature cost function at its~~  
 595 ~~minimum once it was reached~~ (Santaren et al., 2007).

596

## 597 2.8 Comparison of models

598 Model predictions with and without priming effect were compared calculating the coefficients  
 599 of determination, root mean square errors (RMSE) and Bayesian information criteria (BIC).

$$600 \quad RMSE = \sqrt{\frac{1}{N} \sum_{i=1}^N (x_i - \bar{x})^2} \quad (2035)$$

601 where  $i$  is the number of observations (1 to N),  $x_i$  is the predicted value and  $\bar{x}$  is the mean  
 602 observed value.

$$603 \quad BIC = N \times \ln(MSD) + k \times \ln(N) \quad (2136)$$

604 where  $N$  is the number of observations,  $MSD$  is the mean squared deviation used to estimate  
605 the maximum likelihood (Manzoni et al., 2012), and  $k$  is the number of model parameters.

606  
607 The model was run at a yearly time step using mean annual soil temperature and moisture and  
608 annual C inputs to the soil. ~~SOC pools were initialized after a spin-up of 5000 years in the~~  
609 ~~control plot. Measured SOC stocks in 2013 in the control plot were used for the spin-up. The~~  
610 ~~associated uncertainty was estimated with the 93 soil cores sampled in the control plot (see~~  
611 ~~section 2.2.1). Due to a lack of relevant data, we assumed that the climate and the land use were~~  
612 ~~the same for the last 5000 years, and that SOC stocks in the control plot were at equilibrium.~~  
613 ~~Therefore, SOC stocks at the end of the spin-up equaled SOC stocks in the control plot. Three~~  
614 ~~different spin-ups were performed, corresponding to the three different models that were used:~~  
615 ~~one spin-up with the two pools model without the priming effect, one spin-up with the two~~  
616 ~~pools model with the priming effect, and one spin-up with the three pools model.~~ In the  
617 agroforestry, the model was run from the ground (0 m) to 2 m depth, and from the tree (0 m) to  
618 6.5 m from the tree (middle of the alley). The model was applied separately across locations of  
619 a tree-distance gradient having varying OC inputs, each soil column was considered  
620 independent from another. SOC pools were initialized after a spin-up of 5000 years in the  
621 control plot. At  $t_0$ , SOC stocks in the agroforestry plot therefore equaled SOC stocks of the  
622 control plot. The model was then run from  $t_0$  to  $t_{18}$  (years) after tree planting. The spatial  
623 resolution was 0.1 m both vertically and horizontally. The model was developed using R 3.1.1  
624 (R Development Core Team, 2013). Partial-differential equations were solved using the R  
625 package *deSolve* and the *ode.ID* method (Soetaert et al., 2010).

## 627 **2.9 Estimation of the priming intensity and its impact on SOC storage**

628 In equation (7), the priming effect (*PE*) is considered as a control of the FOC on the HSOC  
629 decomposition and not as an accelerating factor of the HSOC decomposition. This method  
630 followed the Wutzler & Reichstein, (2008) approach based on the microbial biomass and  
631 adapted to the FOC by Guenet *et al.*, (2013) for models without explicit microbial biomass.  
632 Models able to reproduce priming effect generally need an explicit microbial biomass  
633 controlling the decomposition (Blagodatsky et al., 2010; Perveen et al., 2014). The priming  
634 scheme used here allows some simplifications in the model structure since an explicit  
635 representation of the microbial biomass is not needed. Furthermore, at equilibrium state (i.e.  
636 when the input rate is constant) the decomposition rate of a first order equation (Eq. (6)) takes  
637 *PE* implicitly into account. Indeed, when FOC enters the system, there is an induced priming,  
638 a constant FOC input rate therefore induces a constant priming. This means that when we  
639 optimized the decomposition rate parameter in the control plot, we implicitly represented this  
640 priming but at a fixed rate. When FOC inputs are modified, due to the tree growth for instance,  
641 the *PE* intensity is modified and this effect cannot be represented by classical first order  
642 kinetics. To estimate the importance of priming on SOC storage in the agroforestry plot, the  
643 simulations using first order equations (Eq. (6)) can therefore not be directly compared to the  
644 simulations using the FOC-dependant decomposition rate (Eq. (7)). To estimate the change of  
645 SOC decomposition rate due to priming when trees are planted, the decomposition ~~rate fluxes~~  
646 predicted by Eq. (7)  $\left(-k_{HSOC,z} \times (1 - e^{-PE \times FOC_{t,z,d}})\right)$  in the agroforestry plot ~~has to~~ must be  
647 compared to the fluxes in the agroforestry plot using the decomposition rate from the control  
648 plot calculated by Eq. (7) with  $FOC_{t,z,d}$  corresponding to the FOC inputs in the control  
649 plot, decomposition rate. Thus, to calculate the importance of priming on SOC storage when  
650 trees are planted, we used the decomposition rates calculated following Eq. (7) in the control  
651 plot  $\left(-k_{HSOC,z} \times (1 - e^{-PE \times FOC_{t,z,d}})\right)$  and we applied this decomposition rate to the  
652 agroforestry plot as a classical first order kinetics (without the control FOC ~~from the control~~

653 ~~plot, i.e.  $k_{new} = -k_{HSOC,z} \times (1 - e^{-PE \times FOC_{t,z,d}})$  with  $FOC_{t,z,d}$  fixed constant).~~ This  
 654 simulation corresponded to the absence of priming due to trees in the agroforestry plot (i.e.  
 655 decomposition not controlled by the FOC of the agroforestry plot). By difference with the  
 656 simulation performed with the full two pools model (Eq. (7)), i.e., taking account of FOC input  
 657 and priming, we calculated the priming intensity.

658

### 659 3 Results

#### 660 3.1 Experimental results

##### 661 3.1.1 Carbon stock in the walnut tree biomass

662 ~~The measured aboveground (trunk + branches) and stump carbon stock of 18-year-old walnut~~  
 663 ~~trees are presented in Table 3.~~

664

665 ~~**Table 3.** Carbon stocks in the aboveground biomass and in the stump of 18-year-old walnut~~  
 666 ~~trees (110 trees ha<sup>-1</sup>).~~

667

	Tree biomass carbon stock	
	-(kg C tree <sup>-1</sup> )	-(t C ha <sup>-1</sup> )
Trunk	55.06 ± 4.35	6.06 ± 0.48
Branches	40.98 ± 7.65	4.51 ± 0.84
Stump	21.21 ± 1.07	2.33 ± 0.12
Total	117.25 ± 8.87	12.9 ± 0.98

668

669

670

671 ~~Errors represent standard errors.~~

672

##### 673 3.1.2 Tree growth

674 ~~Tree growth measurements enabled us to fit the following equation that was used in the model:~~

675

$$DBH_t \begin{cases} 0.01, & t \leq 3 \\ 0.0157 \times t - 0.0391 & 3 < t \leq 20 \end{cases} \quad (R^2 = 0.997) \quad (22)$$

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~~where  $DBH_t$  is the diameter at breast height (m) and  $t$  represents the time since tree planting~~  
~~(years).~~



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### 3.1.3 Crop yield

The average annual crop yield in the control plot was  $Y_C = 3.79 \pm 0.40 \text{ t DM ha}^{-1}$  for the 14 studied years. In the agroforestry plot, the average relative yield decreased linearly with time (increasing *DBH*) and was described using the following linear equation (Fig. 2):

$$Rel Y_{AF_t} = -93.33 \times DBH_t + 100 \quad (R^2 = 0.12, \quad p - value = 0.02) \quad (23)$$

where  $Rel Y_{AF_t}$  is the average relative crop yield (%) in the agroforestry plot compared to the control plot at year  $t$ , and  $DBH_t$  is the diameter at breast height (m) at year  $t$ .

In the agroforestry plot, a linear relationship was used to describe the relative crop yield increase from the tree to the middle of the alley (Fig. 2):

$$Rel Y_{AF_d} = 4.39 \times d + 64.57 \quad (R^2 = 0.24), \quad 1 < d \leq 6.5 \quad (24)$$

where  $Rel Y_{AF_d}$  is the relative crop yield (%) in the agroforestry plot at a distance  $d$  (m) from the tree compared to the control plot.

Finally, the crop yield in the agroforestry plot was modeled as follows:

$$Y_{AF_{t,d}} = Rel Y_{AF_t} \times Y_C \times Rel Y_{AF_d} \quad (R^2 = 0.19), \quad 1 < d \leq 6.5 \quad (25)$$

where  $Y_{AF_{t,d}}$  is the crop yield ( $\text{t DM ha}^{-1}$ ) in the agroforestry plot at the year  $t$  and at a distance  $d$  (m) from the tree. Because three linear equations were used to describe the crop yield in the agroforestry plot, errors were accumulated and we finally came up with a standard underestimation of the crop yield in the agroforestry plot that we corrected by multiplying our equation by 1.2.

~~Fig. 2. Top: Relative yield ( $Rel Y_{t,t}$ ) in the agroforestry plot compared to the control plot as a function of tree growth, represented by the diameter at breast height ( $DBH$ ) at year  $t$ . Bottom: Relative yield ( $Y_{t,t}$ ) as a function of the distance from the tree.~~

### 3.2 Carbon inputs to the FOC pool

#### 3.2.1 Leaf litterfall

~~Total leaf biomass was  $8.96 \pm 1.45$  kg DM tree<sup>-1</sup> and the carbon concentration of walnut leaves was  $449.4 \pm 3.7$  mg C g<sup>-1</sup> (Table 2). With a density of 110 trees ha<sup>-1</sup>, leaf litterfall was estimated at  $0.73 \pm 0.06$  t C ha<sup>-1</sup> in 2012 and at the plot scale. The ratio between leaf biomass and  $DBH$  was  $0.0277 \pm 0.0024$  t C tree<sup>-1</sup> m<sup>-1</sup> or  $3.05$  t C ha<sup>-1</sup> m<sup>-1</sup>. The following linear relationship was therefore used in the model to describe leaf litter C input:~~

$$L_t = 3.05 \times DBH_t \quad (26)$$

~~where  $L_t$  is the leaf litter input (t C ha<sup>-1</sup>) at the year  $t$ , and  $DBH_t$  the diameter at breast height (m) the year  $t$ .~~

#### 3.2.2 Tree fine root C input from mortality

~~In 2012, the measured tree fine root biomass was higher in the tree row than in the alley (Table 4). From 0 to 1 m distance from the tree (in the tree row), the tree fine root biomass was homogeneous and was  $1.01$  t C ha<sup>-1</sup> down 2 m depth.~~

~~Table 4. Walnut tree fine root biomass (t C ha<sup>-1</sup>) as a function of depth and distance from the trees (m).~~

Soil depth (m)	Tree fine root biomass (t C ha <sup>-1</sup> )			
	Tree row		Alley	
	[0, 1] m	]1, 2.5] m	]2.5, 4.0] m	]4.0, 5.5] m
0.0-0.1	$0.08 \pm 0.01$	$0.08 \pm 0.01$	$0.01 \pm 0.00$	$0.00 \pm 0.00$
0.1-0.3	$0.14 \pm 0.02$	$0.24 \pm 0.02$	$0.15 \pm 0.02$	$0.05 \pm 0.01$

0.3-0.5	0.22 ± 0.02	0.16 ± 0.02	0.08 ± 0.01	0.05 ± 0.01
0.5-1.0	0.35 ± 0.04	0.14 ± 0.01	0.14 ± 0.01	0.08 ± 0.01
1.0-1.5	0.15 ± 0.02	0.10 ± 0.01	0.08 ± 0.01	0.08 ± 0.01
1.5-2.0	0.07 ± 0.01	0.13 ± 0.01	0.09 ± 0.01	0.07 ± 0.01
Total	1.01 ± 0.06	0.84 ± 0.04	0.55 ± 0.03	0.34 ± 0.02

Data modified from . Errors represent standard errors.

In 2012 and in the alley, the tree fine root biomass decreased with increasing distance from the tree and was represented by an exponential function:

$$TFRB = \begin{cases} 1.01, & 0 \leq d \leq 1 \\ 1.29 \times e^{-0.28 \times d} & (R^2 = 0.90), \quad 1 < d \leq 6.5 \end{cases} \quad (27)$$

where  $TFRB$  represents tree fine root biomass down 2 m depth ( $t \text{ C ha}^{-1}$ ), and  $d$  the distance from the tree (m).

The following linear relationship was used to simulate  $TFRB$  as a function of tree growth:

$$TFRB_{t,d} = \begin{cases} 3.69 \times DBH_t, & 0 \leq d \leq 1 \\ 4.70 \times DBH_t \times e^{-0.28 \times d}, & 1 < d \leq 6.5 \end{cases} \quad (28)$$

where  $TFRB_t$  represents the tree fine root biomass to 2 m depth ( $t \text{ C ha}^{-1}$ ) at the year  $t$ ,  $DBH_t$  the diameter at breast height (m) at the year  $t$ , and  $d$  the distance to the tree (m).

A decreasing exponential function best represented the changing distribution of tree fine roots within the soil profile with increasing distance to the tree:

$$p_{TFRB,z,d} = \begin{cases} 13.92 \times e^{-1.39 \times z} & (R^2 = 0.68), \quad 0 \leq d \leq 1 \\ a \times e^{-b \times z}, & 1 < d \leq 6.5 \end{cases} \quad (29)$$

and

$$a = 10.31 - 1.15 \times d \quad (R^2 = 0.69) \quad (30)$$

$$b = -1.10 + 0.19 \times d \quad (R^2 = 0.51) \quad (31)$$

Finally,

$$p_{TFRB,z,d} = \begin{cases} 13.92 \times e^{-1.39 \times z}, & 0 \leq d \leq 1 \\ (10.31 - 1.15 \times d) \times e^{-(-1.10+0.19 \times d) \times z}, & 1 < d \leq 6.5 \end{cases} \quad (32)$$

where  $p_{TFRB,z,d}$  is the proportion (%) of the total tree fine root biomass ( $TFRB$ ) at a given depth  $z$  (m), and at a distance  $d$  from the tree (m).

The tree fine root turnover ranged from 1.7 to 2.8  $\text{yr}^{-1}$  depending on fine root diameter, with an average turnover of 2.2  $\text{yr}^{-1}$  for fine roots  $\leq 2$  mm and to a depth of 2 m (Germon et al., 2016).

### 3.2.3 Aboveground carbon input from the crop

In the agroforestry plot, the carbon input to the soil from the aboveground crop biomass was:

$$ABC_{crop,t,d} = Y_{AF,t,d} \times (\text{straw biomass: crop yield}) \times C_{straw} \times (1 - \text{export}) \quad (33)$$

where  $ABC_{crop,t,d}$  is the aboveground carbon input from the crop ( $\text{t C ha}^{-1}$ ) at the year  $t$  and distance  $d$  from the tree,  $Y_{AF,t,d}$  is the agroforestry crop yield. The average ratio between the straw biomass ( $\text{t DM ha}^{-1}$ ) and the crop yield ( $\text{t DM ha}^{-1}$ ) equaled  $1.03 \pm 0.11$  ( $n=6$ ). The wheat straw was exported out of the field after the harvest, but it was estimated that 25% of the straw biomass was left on the soil, thus  $\text{export}=0.75$ . In the control plot,  $Y_{AF,t,d}$  was replaced by  $Y_C$ .

### 3.2.4 Belowground carbon input from the crop

In the agroforestry plot, the belowground crop biomass was represented by:

$$BEC_{crop,t,d} = Y_{AF,t,d} \times (\text{shoot: crop yield}) \times (\text{root: shoot}) \times C_{root} \quad (34)$$

where  $BEC_{crop,t,d}$  is the belowground crop biomass ( $\text{t C ha}^{-1}$ ) at the year  $t$  and at a distance  $d$  from the tree,  $Y_{AF,t,d}$  is the agroforestry crop yield. The average ratio between the total crop aboveground biomass ( $\text{shoot}$ ) and the crop yield equaled  $2.45 \pm 0.15$  ( $n=6$ ). In 2012, total fine root biomass was  $2.29 \pm 0.32 \text{ t C ha}^{-1}$  in the control (Table 5).

**Table 5.** Wheat fine root biomass in the agricultural control plot in 2012.

Soil depth (m)	Wheat fine root biomass	
	(kg C m <sup>-3</sup> )	(t C ha <sup>-1</sup> )
0.0-0.1	0.48 ± 0.05	0.48 ± 0.05
0.1-0.3	0.34 ± 0.04	0.69 ± 0.09
0.3-0.5	0.22 ± 0.04	0.44 ± 0.08
0.5-1.0	0.10 ± 0.04	0.52 ± 0.20
1.0-1.5	0.03 ± 0.04	0.17 ± 0.19
Total	-	2.29 ± 0.32

Errors represent standard errors.

Therefore, the wheat *root:shoot* ratio equaled  $0.79 \pm 0.12$  ( $n=1$ ). The carbon concentration of wheat root was  $C_{root} = 35.14 \pm 1.90$  mg C g<sup>-1</sup>. In the control plot,  $V_{AF, \text{tree}}$  was replaced by  $V_C$ . In 2012, no wheat roots were observed below 1.5 m, and root biomass decreased exponentially with increasing depth (Table 5). The distribution of crop roots within the soil profile was described as follows:

$$p_{CRBC, z} = \begin{cases} 26.44 \times e^{-2.59 \times z} & (R^2 = 0.99), & z \leq 1.5 \\ 0, & & z > 1.5 \end{cases} \quad (35)$$

where  $p_{CRBC, z}$  is the proportion (%) of total crop root biomass in the control plot at a given depth  $z$  (m).

The crop root turnover was assumed to be 1 yr<sup>-1</sup>, root mortality occurring mainly after crop harvest.

### 3.2.5 Aboveground and belowground carbon inputs from the tree row herbaceous vegetation

The distance from the trees had no effect on the above and belowground biomass of the herbaceous vegetation (data not shown), therefore average values are presented. The summer aboveground biomass was almost three times higher than in winter, whereas the belowground

biomass was two times higher (Table 6). The total aboveground carbon input was  $2.13 \pm 0.14$  t C ha<sup>-1</sup> yr<sup>-1</sup> and the total belowground carbon input was  $0.74 \pm 0.05$  t C ha<sup>-1</sup> yr<sup>-1</sup> to 0.5 m depth.

**Table 6.** Aboveground and belowground biomass of the herbaceous vegetation in the tree rows.

	Soil depth (m)	Herbaceous biomass (t C ha <sup>-1</sup> )	
		Summer	Winter
Aboveground	-	$1.57 \pm 0.11$	$0.56 \pm 0.09$
	0.0-0.1	$0.22 \pm 0.03$	$0.17 \pm 0.01$
Belowground	0.1-0.3	$0.16 \pm 0.02$	$0.06 \pm 0.01$
	0.3-0.5	$0.09 \pm 0.04$	$0.04 \pm 0.01$
	Total	$0.46 \pm 0.04$	$0.27 \pm 0.02$

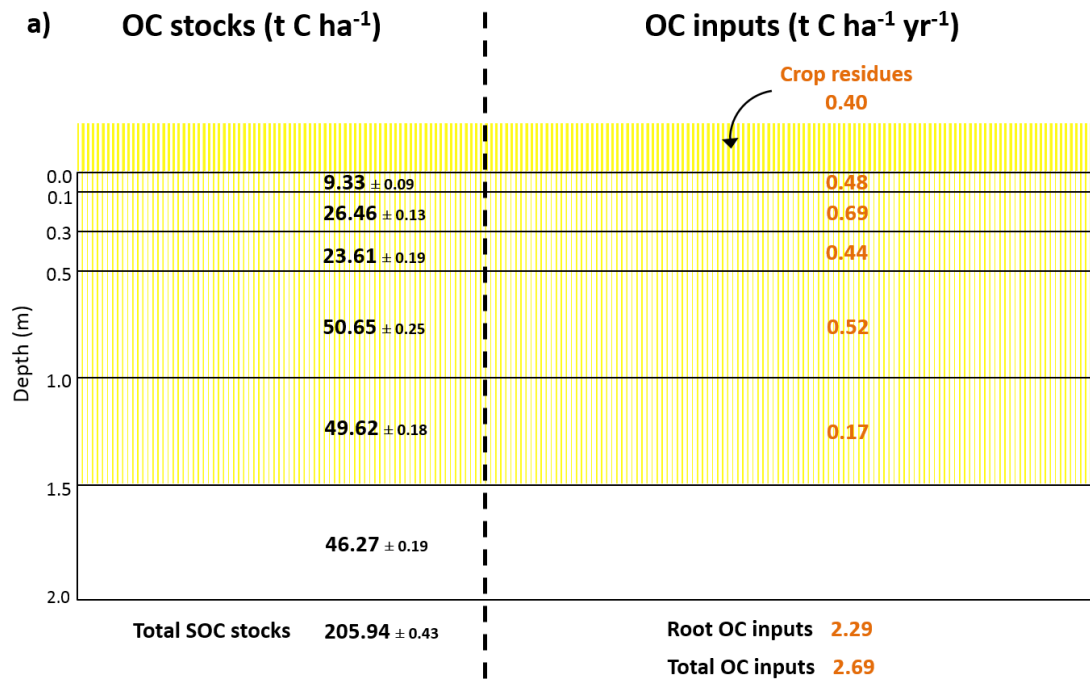
Errors represent standard errors.

The belowground carbon input from the tree row vegetation ( $BEC_{veg,z}$  t C ha<sup>-1</sup>) at a given depth  $z$  (m) was described by the following equation:

$$BEC_{veg,z} = \begin{cases} 0.44 \times e^{-3.12 \times z}, & z \leq 1.5 \\ 0, & z > 1.5 \end{cases} \quad (36)$$

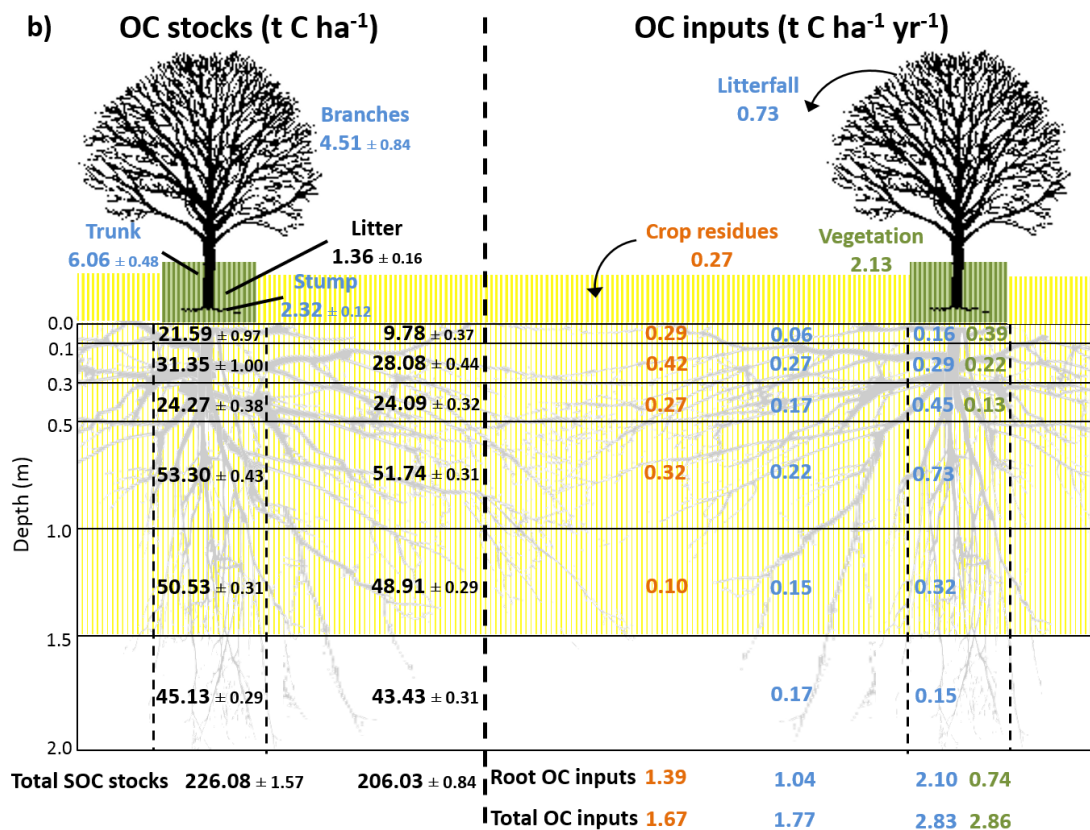
### 3.21.6. Organic carbon inputs and SOC stocks: a synthesis from field measurements

Tree rows in the agroforestry system received two times more organic carbon (OC) inputs compared to the control plot (Fig. 3), and 65% more than alleys. Globally Overall, the agroforestry plot had 41% more OC inputs to the soil than the control plot to 2 m depth ( $3.80$  t C ha<sup>-1</sup> yr<sup>-1</sup> compared to  $2.69$  t C ha<sup>-1</sup> yr<sup>-1</sup>). In the agroforestry plot, the largest aboveground OC input to the soil comes from the herbaceous vegetation, and not from the trees. In the control plot, 85% of OC inputs are wheat root litters. In the agroforestry plot, root inputs represent 71% of OC inputs in the alleys, and 50% in the tree rows.



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**Color legend** Durum wheat Walnut trees Herbaceous vegetation

807 **Fig. 3.** Measured soil organic carbon stocks and organic carbon inputs to the soil a) in the  
808 agricultural control plot, b) in the 18-year-old agroforestry plot. Associated errors are  
809 standard errors. Values are expressed per hectare of land type (control, alley, tree row).  
810 To get the values per hectare of agroforestry, data from alley and tree row have to be  
811 weighted by their respective surface area (i.e., 84% and 16%, respectively) and then  
812 added up. OC: organic carbon; SOC: soil organic carbon. SOC stocks data are issued  
813 from Cardinael *et al.*, (2015a), data of tree root OC inputs are combined from Cardinael  
814 *et al.*, (2015b) and from Germon *et al.*, (2016).

815

### 816 **3.3-2 HSOC decomposition rate**

817 The soil incubation experiment showed that the HSOC mineralization rate decreased  
818 exponentially with depth (Fig. S1) and could be described with:

$$819 \quad k_{HSOC,z} = 6.114 \times e^{-1.37 \times z} \quad (R^2 = 0.76) \quad (34)$$

820 where  $z$  is the soil depth (m), and where the  $a$  ( $\text{yr}^{-1}$ ) coefficient ( $a = 6.114$ ) was further optimized  
821 (Table 75).

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**Table 75.** Summary of optimized model parameters.

Model parameter	Meaning	Prior range	Posterior values $\pm$ variance (prior values)		
			2 pools - without <i>PE</i>	2 pools - with <i>PE</i>	3 pools – without <i>PE</i>
<i>a</i>	coefficient from Eq. (8) of the HSOC decomposition ( $\text{yr}^{-1}$ )	$3.65\text{e}^{-6}$ -3.65	$0.01\text{e}^{-2} \pm <10^{-4}$ ( $0.01\text{e}^{-2}$ )	$0.01\text{e}^{-2} \pm <10^{-4}$ ( $0.01\text{e}^{-2}$ )	-
<i>a</i> <sub>1</sub>	coefficient from Eq. (8) of the HSOC1 decomposition ( $\text{yr}^{-1}$ )	$3.65\text{e}^{-6}$ -3.65	-	-	$0.01\text{e}^{-2} \pm <10^{-4}$ ( $0.01\text{e}^{-2}$ )
<i>a</i> <sub>2</sub>	coefficient from Eq. (8) of the HSOC2 decomposition ( $\text{yr}^{-1}$ )	$3.65\text{e}^{-6}$ -3.65	-	-	$0.83\text{e}^{-2} \pm 0.17\text{e}^{-2}$ ( $0.83\text{e}^{-2}$ )
<i>D</i>	diffusion coefficient ( $\text{cm}^2 \text{yr}^{-1}$ )	$1\text{e}^{-6}$ -1	$4.62\text{e}^{-4} \pm 5.95\text{e}^{-4}$ ( $9.64\text{e}^{-4}$ )	$5.63\text{e}^{-4} \pm 1.42\text{e}^{-4}$ ( $9.01\text{e}^{-4}$ )	$5.24\text{e}^{-4} \pm 7.62\text{e}^{-4}$ ( $9.64\text{e}^{-4}$ )
<i>A</i>	advection rate ( $\text{mm yr}^{-1}$ )	$1\text{e}^{-6}$ -1	$21.25\text{e}^{-4} \pm 5.02\text{e}^{-4}$ ( $8.54\text{e}^{-4}$ )	$6.63\text{e}^{-4} \pm 2.38\text{e}^{-4}$ ( $4.27\text{e}^{-4}$ )	$21.60\text{e}^{-4} \pm 2.24\text{e}^{-4}$ ( $8.54\text{e}^{-4}$ )
<i>h</i>	humification yield	0.01-1	$0.32 \pm <10^{-4}$ (0.34)	$0.25 \pm 1.00\text{e}^{-4}$ (0.13)	$0.34 \pm 0.03$ (0.34)
<i>PE</i>	priming coefficient	0.1-160	-	$9.66 \pm 1.49$ (102.95)	-
<i>f</i> <sub>1</sub>	fraction of decomposed FOC entering the HSOC1 pool	0-1	-	-	$0.99 \pm 0.18$ (0.86)
<i>f</i> <sub>2</sub>	fraction of decomposed HSOC1 entering the FOC pool	0-1	-	-	$0.94 \pm 1.10\text{e}^{-3}$ (0.80)

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The prior range represents the range in which prior values were sampled for the 30 optimizations per model variant. The prior values presented in

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brackets in the posterior column represent the prior values that minimized the  $\mathbf{J}(\mathbf{x})$  value (Eq. (19)).

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### 827 **3.4.3 Modeling results**

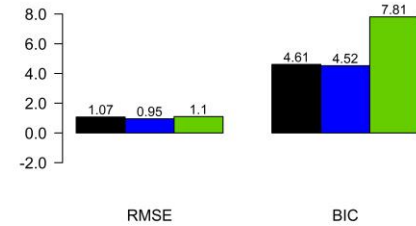
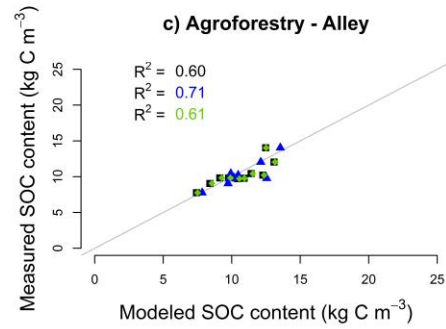
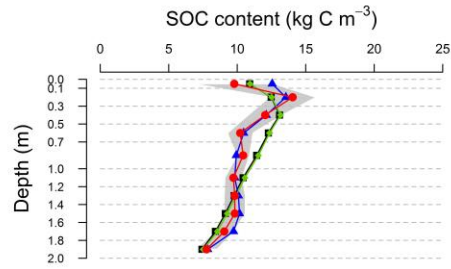
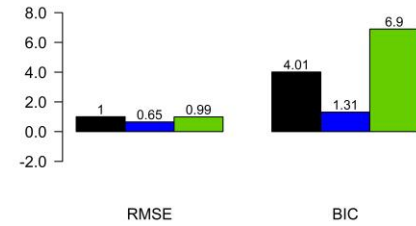
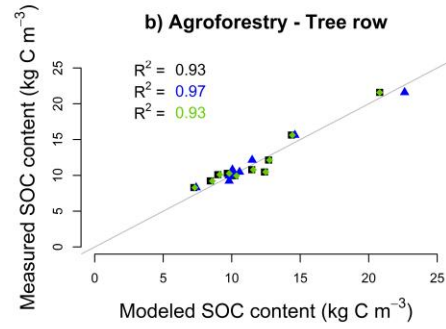
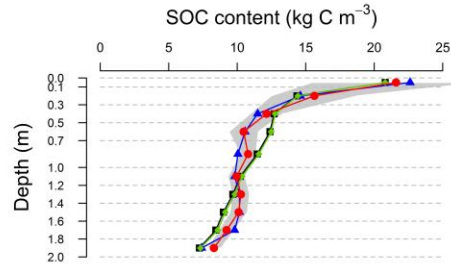
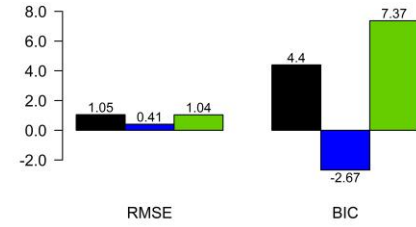
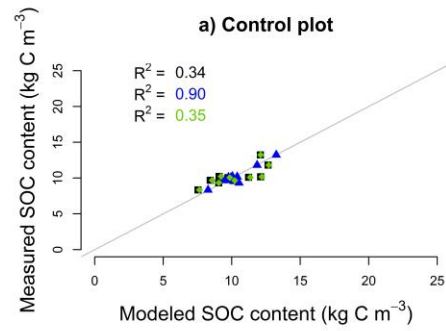
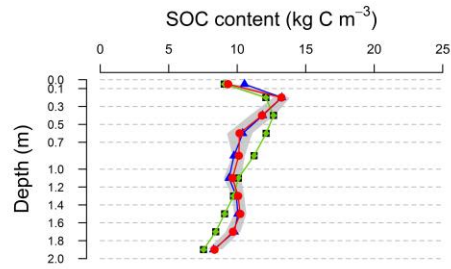
#### 828 **3.4.3.1 Optimized parameters and correlation matrix**

829 The optimized parameters and their prior modes are presented in [Table 75](#). For the two pools  
830 model without priming effect, the most important correlation was observed between  $h$  and  $A$   
831 which control the humification and the transport by advection. Concerning the two pools model  
832 with priming effect, the most important correlations were observed between  $h$  and  $PE$  which  
833 controls the effect of the FOC on HSOC decomposition, and between  $h$  and  $A$ .  $A$  and  $PE$  were  
834 also positively correlated ([Fig. S2](#)). For the three pools model,  $f_1$  and  $f_2$  were by definition  
835 negatively correlated, but  $f_2$  and  $A$  were also correlated. Considering the method used to  
836 optimize the parameters, these important correlation factors hinder the presentation of the  
837 model output within an envelope. Therefore, we presented the model results using the optimized  
838 parameter without any envelope.

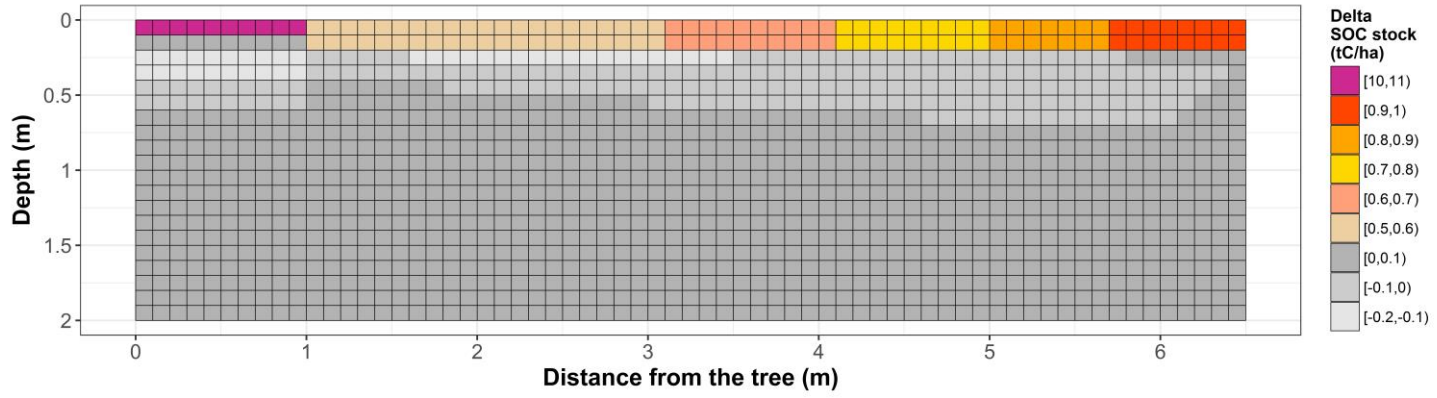
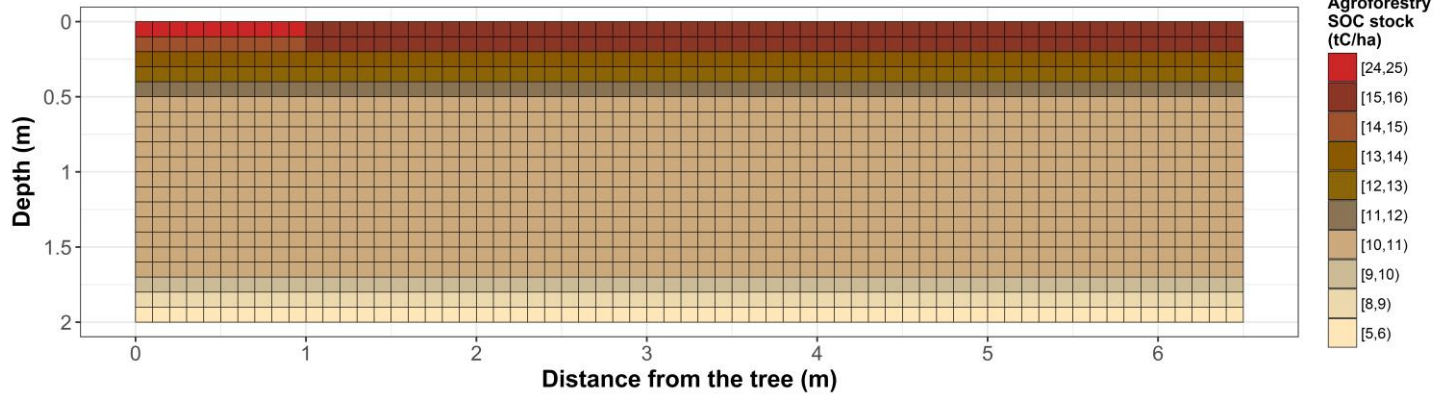
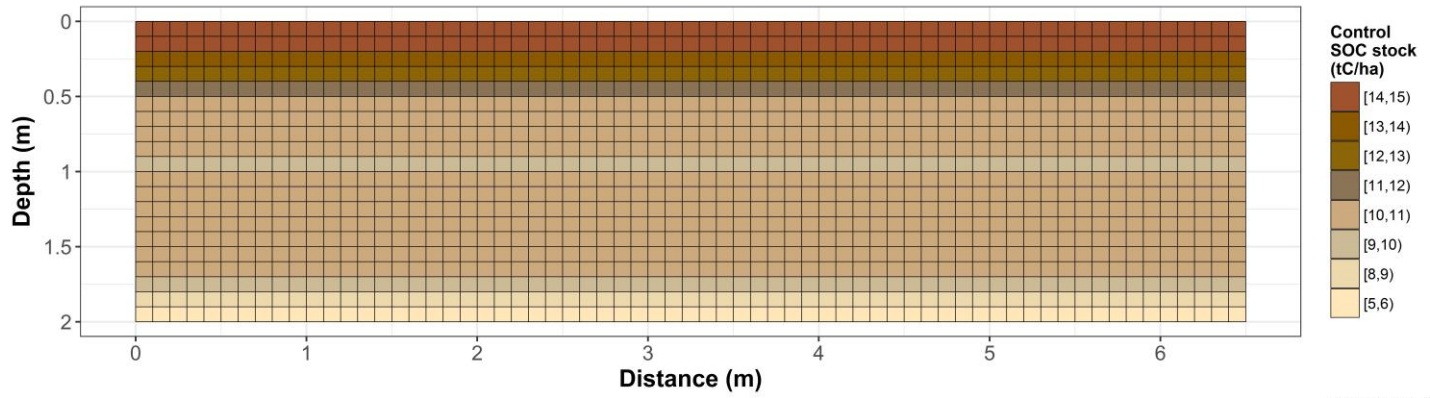
#### 839 840 **3.4.3.2 Modeled SOC stocks**

841 As a reminder, SOC stocks of the agroforestry plot were not part of model calibration (that used  
842 the control plot only) but were used here for validation. Observed SOC stocks were not well  
843 represented by the two pools model without priming effect, with RMSE ranging from 1.00 to  
844 1.07 kg C m<sup>-3</sup> ([Fig. 4, Table S1S2](#)). The model performed better when the priming effect was  
845 taken into account, with RMSE ranging from 0.41 to 0.95 kg C m<sup>-3</sup>, and the SOC profile was  
846 well described. The representation of SOC stocks was not improved by the inclusion of a third  
847 C pool in the model. GloballyOverall, the two pools model with priming effect was the best  
848 one, as shown by the BICs ([Fig. 4, Table S1S2](#)). For all models, SOC stocks below 1 m depth  
849 were better described than above SOC stocks ([Table S1S2](#)). The spatial distribution of SOC  
850 stocks and of additional SOC storage was also well described ([Fig. 5](#)), with a very high  
851 additional SOC ~~stock~~ storage in the topsoil layer in the tree row. Most modeled SOC storage in

852 the agroforestry plot was located in the first 0.2 m depth, ~~and~~but SOC storage was slightly  
853 higher in the middle of the alleys than in the alleys close to the tree rows.



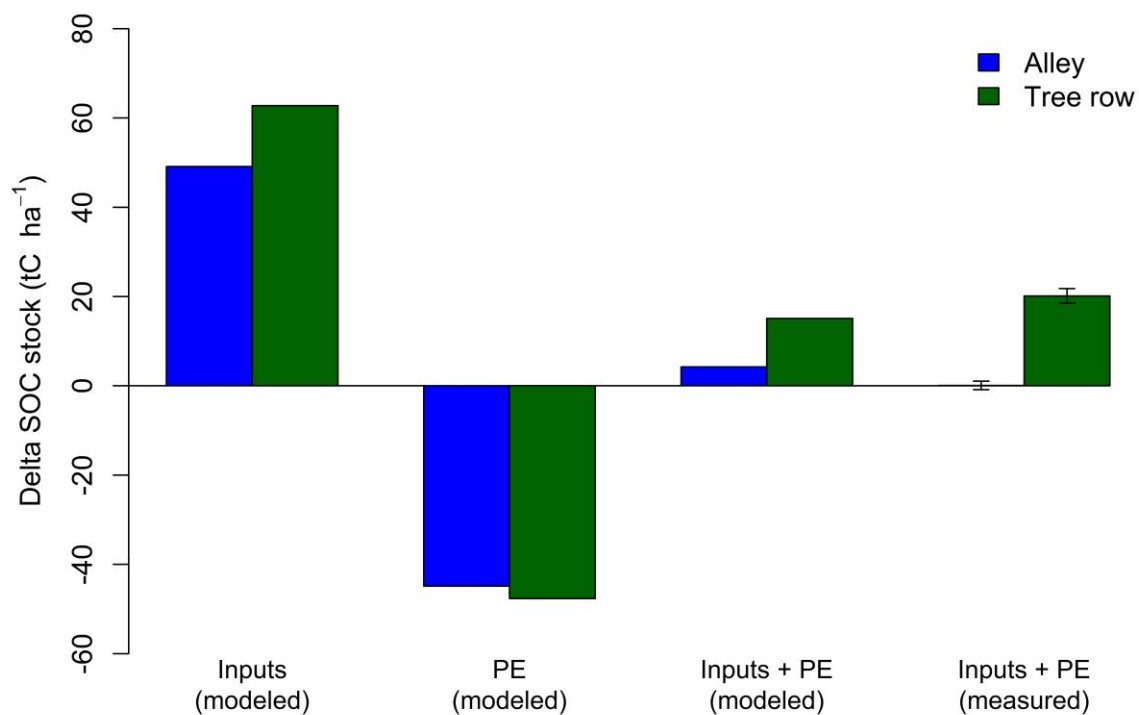
855 **Fig. 4.** Measured and modeled soil organic carbon contents ( $\text{kg C m}^{-3}$ ) in an agricultural control plot and in an 18-year-old silvoarable system with  
856 a two pools model without priming effect (no *PE*), with a two pools model with priming effect (*PE*) and with a three pools model without  
857 *PE*. Gray shaded bands represent standard deviations of measured SOC stocks (n=93 in the control, n=40 in the tree rows, and n=60 in the  
858 alleys).  
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867 **Fig. 5.** Spatial distribution of control SOC stocks (top), agroforestry SOC stocks (middle), and additional SOC storage ( $\text{t C ha}^{-1}$ ) in an 18-year-old  
868 silvoarable system compared to an agricultural control plot and represented by the two pools model with priming effect.

869 **3.43.3 Antagonist effect of priming on SOC storage**

870 The priming effect increases the decomposition rate when more FOC is available (Fontaine et  
871 al., 2007). Therefore, the effect of a C inputs increase on SOC storage in the agroforestry plot  
872 might be counterbalanced by priming. With our model we were able to estimate the contribution  
873 of each driver on SOC storage. The introduction of priming effect in the model reduced the  
874 potential SOC storage due to higher organic inputs in the agroforestry system by 91% in the  
875 alley, and by 76% in the tree rows (Fig. 6). The potential effect of OC inputs alone on SOC  
876 storage was 49.12 to 62.77 t C ha<sup>-1</sup>, but the effect of priming on SOC storage was -44.89 to -  
877 47.67 t C ha<sup>-1</sup>, resulting in a modeled SOC storage of 4.23 t C ha<sup>-1</sup> in the alley and of 15.09 t C  
878 ha<sup>-1</sup> in the tree row down 2 depth (Fig. 6). The negative effect of priming effect on SOC storage  
879 increased with increasing soil depth (Fig. S3).



880

881 **Fig. 6.** Decoupling the role of C inputs and priming effect (*PE*) on SOC storage in an 18-year-  
882 old silvoarable system down 2 m depth. Inputs: only the input effect is modeled; *PE*:



883 only the priming effect is modeled; Inputs + *PE*: model prediction with both processes  
884 taken into account.

885

## 886 **4 Discussion**

### 887 **4.1 OC inputs drive SOC storage in agroforestry systems**

888 Increased SOC stocks in the agroforestry plot compared to the control may be explained either  
889 by increased OC inputs, or decreased OC outputs by SOC mineralization, or both. Measured  
890 organic carbon inputs to soil were increased by 40% down to 2m depth in the 18-year-old  
891 agroforestry plot compared to the control plot. Increased OC inputs in agroforestry systems has  
892 been shown in other studies but they were only quantified in the first 20 cm of soil (Oelbermann  
893 et al., 2006; Peichl et al., 2006). This study is therefore the first one also quantifying deep OC  
894 inputs to soil. In this study and due to a lack of data, soil temperature and soil moisture were  
895 considered the same in both plots so that abiotic factors controlling SOC decomposition were  
896 identical. Despite these simplifying assumptions on similarities in microclimate but also on  
897 vertical transport between the control and the agroforestry system, the model calibrated to the  
898 control plot was able to well-reproduce SOC stocks in tree rows and alleys and its depth  
899 distribution well in the agroforestry plot. This strong validation suggesting also suggests that  
900 OC inputs is the main driver of SOC storage at this site, and that a decrease of SOC  
901 mineralisation due to the agroforestry potential effect of agroforestry microclimate on SOC  
902 mineralisation is not obvious of minor importance. Reduced soil temperature is often observed  
903 in agroforestry systems (Clinch et al., 2009; Dubbert et al., 2014), but effect of agroforestry on  
904 soil moisture is much more complex. The soil evaporation is reduced under the trees, but water  
905 is lost through their transpiration (Ilstedt et al., 2016; Ong and Leakey, 1999), and these effects  
906 vary with the distance from the tree (Odhiambo et al., 2001). Moreover, the water infiltration  
907 and the water storage can be increased under the trees after a rainy event (Anderson et al.,

908 2009). Therefore, the effect of agroforestry on soil moisture is variable in time and space, and  
909 should be investigated more in details. Interactions between soil temperature and soil moisture  
910 on the SOC decomposition are known to be complex (Conant et al., 2011; Moyano et al., 2013;  
911 Sierra et al., 2015) and up to now it is not possible to predict the effect of agroforestry  
912 microclimate on the SOC decomposition rate. A sensitivity analysis performed on these two  
913 boundary conditions showed that the model was not very sensitive to soil temperature and soil  
914 moisture (Fig. S4), but the real effect of these two parameters on SOC dynamics under  
915 agroforestry systems should be better investigated in future studies. suggesting that the potential  
916 changes in soil microclimate in the agroforestry plot are not major drivers of the SOC storage.  
917 Furthermore, the SOC decomposition rate could also be modified due to an absence of soil  
918 tillage in the tree rows (Balesdent et al., 1990) or to an increased aggregate stability (Udawatta  
919 et al., 2008) in the topsoil.

920

#### 921 **4.2 Representation of SOC spatial heterogeneity in agroforestry systems**

922 The lateral spatial heterogeneity of SOC stocks in the agroforestry plot was well described by  
923 the two pools model including priming effect, with higher SOC stocks in the tree rows' topsoil  
924 than in the alleys. Inputs from the herbaceous vegetation had an important impact on SOC  
925 storage in this agroforestry system. The increased SOC stocks in the tree rows were explained  
926 in a big part by an important above-ground carbon input (2.13 t C ha<sup>-1</sup> yr<sup>-1</sup>) by the herbaceous  
927 vegetation between trees. This result had already been suggested by Cardinael et al., (2015b)  
928 and by Cardinael et al., (2017) who showed that even young agroforestry systems could store  
929 SOC in the tree rows while trees are still very small. These “grass strips” indirectly introduced  
930 by the tree planting in parallel tree rows have a major impact on SOC stocks of agroforestry  
931 systems. The model treated the carbon from this herbaceous litter as an input to the upper layer  
932 of the mineral soil, in the same way as inputs by roots. Introduction of nitrogen in the model

933 could be further tested in order to take into account a lower carbon use efficiency due to a lack  
934 of nutrients for microbial growth in this litter. For all models, SOC stocks were better described  
935 in the tree rows than in the alleys. In the alleys, the spatial distribution of organic inputs is more  
936 complex and thus more difficult to model. The tree root system is influenced by the soil tillage  
937 and by the competition with the crop roots, and thus the highest tree fine root density is not  
938 observed in the topsoil but in the 0.3-0.5 m soil layer (Cardinael et al., 2015a). In the model,  
939 we were not able to represent this specific tree root pattern with commonly used mathematical  
940 functions, and tree root profiles were modeled, by default, using a decreasing exponential.  
941 Indeed, piecewise linear functions introduce threshold effects not desirable for transport  
942 mechanisms, especially diffusion. This simplification could partly explain the model  
943 overestimation of SOC stocks in the 0.0-0.1 m layer of the alleys compared to observed data.  
944 This result suggests that it could be useful to couple the CARBOSAF model with a model  
945 describing root architecture and root growth (Dunbabin et al., 2013; Dupuy et al., 2010), using  
946 for instance voxel automata (Mulia et al., 2010). Moreover, the model described a slight  
947 increase of SOC stocks in the middle of the alleys than close to the trees in the alleys. This  
948 could be explained by the linear equation used to describe the crop yield as a function of the  
949 distance from the trees, leading to an overestimation of the crop yield reduction close to the  
950 trees. It could also be explained by the formalism used to model leaf litter distribution in the  
951 plot. We considered a homogeneous distribution of leaf inputs in the agroforestry plot, which  
952 was the case in the last years, but probably not in the first years of the tree growth where leaves  
953 might be more concentrated close to the trees (Thevathasan and Gordon, 1997).

954 The two pools model with priming effect also represented a slight SOC storage in the  
955 agroforestry plot below 1.0 m depth, but it was not observed in the field. This could be linked  
956 to an overestimation of C input from tree fine root mortality. Indeed, a constant root turnover  
957 was considered along the soil profile, but several authors reported a decrease of the root

958 turnover with increasing soil depth (Germon et al., 2016; Hendrick and Pregitzer, 1996; Joslin  
959 et al., 2006). However, the sensitivity analysis showed that the model was not sensitive to this  
960 parameter (Fig. S4).

961

#### 962 **4.3 Vertical representation of SOC profiles in models**

963 The best model to represent SOC profiles considered the priming effect. This process can act  
964 in two different ways on the shape of SOC profiles. It has a direct effect on the SOC  
965 mineralization and it therefore modulates the amount of SOC in each soil layer, creating  
966 different SOC gradients. This indirectly affects the mechanisms of C transport within the soil  
967 profile, as shown by a modification of transport coefficients in the case of priming effect (Table  
968 75). Contrary to what was shown by Cardinael *et al.*, (2015c) in long term bare fallows  
969 receiving contrasted organic amendments, the addition of another SOC pool could not surpass  
970 the inclusion of priming effect in terms of model performance. Together with Wutzler &  
971 Reichstein, (2013) and Guenet *et al.*, (2016), this study therefore suggests that implementing  
972 priming effect into SOC models would improve model performances especially when  
973 modelling deep SOC profiles.

974 We considered here the same transport coefficients for the FOC and HSOC pools, but the  
975 quality and the size of OC particles are different, potentially leading to various movements in  
976 the soil by water fluxes or fauna activity (Lavelle, 1997). Moreover, we considered identical  
977 transport parameters in the agroforestry and in the control plot, but the presence of trees could  
978 modify soil structure, soil water fluxes (Anderson et al., 2009), and the fauna activity (Price  
979 and Gordon, 1999). However, the model was little sensitive to these parameters (Fig. S4).  
980 Further study could investigate the role of different transport coefficients on the description of  
981 SOC profiles.

982

#### 983 **4.4 Higher OC inputs or a different quality of OC?**

984 The introduction of trees in an agricultural field not only modifies the amount of litter residues,  
985 but also their quality. Tree leaves, tree roots, and the herbaceous vegetation from the tree row  
986 have different C:N ratios, lignin and cellulose contents than the crop residues. Recent studies  
987 showed that plant diversity had a positive impact on SOC storage (Lange et al., 2015; Steinbeiss  
988 et al., 2008). One of the hypothesis proposed by the authors is that diverse plant communities  
989 result in more active, more abundant and more diverse microbial communities, increasing  
990 microbial products that can potentially be stabilized. In our model, litter quality is not related  
991 to different SOC pools, but is implicitly taken into account in the FOC decomposition rate,  
992 which is weighted by the respective contribution from the different types of OC inputs. To test  
993 this, we performed a model run considering that all OC inputs in the agroforestry plot were crop  
994 inputs (all FOC decomposition rates equaled wheat decomposition rate), but results were not  
995 significantly different from the one presented here. We then consider that changes in litter  
996 quality in the agroforestry plot did not significantly influence SOC decomposition rates.

997

#### 998 **4.5 Possible limitation of SOC storage by priming effect**

999 Our modelling results showed that the priming effect could considerably reduce the capacity of  
1000 soils to store organic carbon. Our study showed that the increase of SOC stocks was not  
1001 proportional to OC inputs, especially at depth. This result has often been observed in Free Air  
1002 CO<sub>2</sub> Enrichment (FACE) experiments. In these experiments, productivity is usually increased  
1003 due to CO<sub>2</sub> fertilization, but several authors also reported an increase in SOC decomposition  
1004 but not linearly linked to the productivity increase (van Groenigen et al., 2014; Sulman et al.,  
1005 2014). In this study, the estimation of the priming effect intensity was possible because most  
1006 OC inputs to the soil were accurately measured. The modelled intensity of priming effect was  
1007 very strong, offsetting 75 to 90% of potential SOC storage due to OC inputs. In a long-term

1008 FACE experiment, Carney *et al.*, (2007) also found that SOC decreased due to priming effect,  
1009 offsetting 52% of additional carbon accumulated in aboveground and coarse root biomass. The  
1010 priming effect intensity also relies on nutrient availability (Zhang *et al.*, 2013). In agroforestry  
1011 systems, tree roots can intercept leached nitrate below the crop rooting zone (Andrianarisoa *et*  
1012 *al.*, 2016), reducing nutrient availability. This beneficial ecosystem service could indirectly  
1013 increase the priming effect intensity in deep soil layers.

1014 However, this strong intensity could also partially be linked to the formalism used to simulate  
1015 priming effect. This formalism assumes that there is no mineralisation of the SOC in the  
1016 absence of fresh OC inputs (no basal respiration). This is a strong hypothesis, but this situation  
1017 never occurs since the FOC pool is never empty (data not shown). In the alleys and below the  
1018 maximum rooting depth of crops, there are no direct inputs of FOC, but OC is transported in  
1019 these deep layers due to transport mechanisms. However, further studies could study the impact  
1020 of the priming effect formalism on the estimation of its intensity by using explicit microbial  
1021 biomass for instance (Blagodatsky *et al.*, 2010; Perveen *et al.*, 2014).

1022 Finally, root exudates were not quantified in this study. Several authors showed that they could  
1023 induce strong priming effects (Bengtson *et al.*, 2012; Keiluweit *et al.*, 2015), but root exudates  
1024 are also a source of labile carbon, potentially contributing to stable SOC (Cotrufo *et al.*, 2013).  
1025 These opposing effects of root exudates on SOC should be further investigated, especially  
1026 concerning the deep roots in agroforestry systems.

1027

## 1028 **5 Conclusions**

1029 We proposed the first model that simulates soil organic carbon dynamics in agroforestry  
1030 accounting for both the whole soil profile and the lateral spatial heterogeneity in agroforestry  
1031 plots. ~~This~~ The two pools model with priming effect described reasonably well the measured  
1032 SOC stocks after 18 years of agroforestry and SOC distributions with depth. It showed that the

1033 increased inputs of fresh biomass to soil in the agroforestry system explained the observed  
1034 additional SOC storage and suggested priming effect as a process controlling SOC stocks in the  
1035 presence of trees. This study points out at processes that may be modified by deep rooting trees  
1036 and deserve further studies given their potential effects on SOC dynamics, such as additional  
1037 inputs of C as roots exudates, or altered soil structure leading to modified SOC transport rates.

1038

## 1039 **6 Data availability**

1040 The data and the model are freely available upon request and can be obtained by contacting the  
1041 author (remi.cardinael@cirad.fr).

1042

## 1043 **Information about the Supplement**

1044 The Supplement includes the [walnut tree fine root biomass \(Table S1\)](#), the different model  
1045 performances (Table [S21](#)), the potential SOC decomposition rate as a function of soil depth  
1046 (Fig. S1), the correlation matrix of optimized parameters (Fig. [8S2](#)), the decoupling of OC  
1047 inputs and priming effect as a function of soil depth (Fig. S3), and a sensitivity analysis of the  
1048 model (Fig. S4).

1049

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1063

## 1064 **References**

1065 Ahrens, B., Braakhekke, M. C., Guggenberger, G., Schrumpf, M. and Reichstein, M.:  
1066 Contribution of sorption, DOC transport and microbial interactions to the  $^{14}\text{C}$  age of a soil  
1067 organic carbon profile: Insights from a calibrated process model, *Soil Biol. Biochem.*, 88, 390–  
1068 402, 2015.

1069 Albrecht, A. and Kandji, S. T.: Carbon sequestration in tropical agroforestry systems, *Agric.*  
1070 *Ecosyst. Environ.*, 99, 15–27, 2003.

1071 Anderson, S. H., Udawatta, R. P., Seobi, T. and Garrett, H. E.: Soil water content and infiltration  
1072 in agroforestry buffer strips, *Agrofor. Syst.*, 75(1), 5–16, 2009.

1073 Andrianarisoa, K., Dufour, L., Bienaime, S., Zeller, B. and Dupraz, C.: The introduction of  
1074 hybrid walnut trees (*Juglans nigra* x *regia* cv. NG23) into cropland reduces soil mineral N  
1075 content in autumn in southern France, *Agrofor. Syst.*, 90, 193–205, 2016.

1076 Baisden, W. T. and Parfitt, R. L.: Bomb  $^{14}\text{C}$  enrichment indicates decadal C pool in deep soil?,  
1077 *Biogeochemistry*, 85, 59–68, 2007.

1078 Baisden, W. T., Amundson, R., Brenner, D. L., Cook, A. C., Kendall, C. and Harden, J. W.: A  
1079 multiisotope C and N modeling analysis of soil organic matter turnover and transport as a  
1080 function of soil depth in a California annual grassland soil chronosequence, *Global*  
1081 *Biogeochem. Cycles*, 16, 82-1-82–26, 2002.



1082 Balandier, P. and Dupraz, C.: Growth of widely spaced trees. A case study from young  
1083 agroforestry plantations in France, *Agrofor. Syst.*, 43, 151–167, 1999.

1084 Balesdent, J., Mariotti, A. and Boisgontier, D.: Effect of tillage on soil organic carbon  
1085 mineralization estimated from  $^{13}\text{C}$  abundance in maize fields, *J. Soil Sci.*, 41, 587–596, 1990.

1086 Bambrick, A. D., Whalen, J. K., Bradley, R. L., Cogliastro, A., Gordon, A. M., Olivier, A. and  
1087 Thevathasan, N. V.: Spatial heterogeneity of soil organic carbon in tree-based intercropping  
1088 systems in Quebec and Ontario, Canada, *Agrofor. Syst.*, 79, 343–353, 2010.

1089 Bengtson, P., Barker, J. and Grayston, S. J.: Evidence of a strong coupling between root  
1090 exudation, C and N availability, and stimulated SOM decomposition caused by rhizosphere  
1091 priming effects, *Ecol. Evol.*, 2(8), 1843–1852, 2012.

1092 Blagodatsky, S., Blagodatskaya, E., Yuyukina, T. and Kuzyakov, Y.: Model of apparent and  
1093 real priming effects: Linking microbial activity with soil organic matter decomposition, *Soil*  
1094 *Biol. Biochem.*, 42, 1275–1283, 2010.

1095 Braakhekke, M. C., Beer, C., Hoosbeek, M. R., Reichstein, M., Kruijt, B., Schrumphf, M. and  
1096 Kabat, P.: SOMPROF: A vertically explicit soil organic matter model, *Ecol. Modell.*, 222,  
1097 1712–1730, 2011.

1098 Bruun, S., Christensen, B. T., Thomsen, I. K., Jensen, E. S. and Jensen, L. S.: Modeling vertical  
1099 movement of organic matter in a soil incubated for 41 years with  $^{14}\text{C}$  labeled straw, *Soil Biol.*  
1100 *Biochem.*, 39, 368–371, 2007.

1101 Burgess, P. J., Incoll, L. D., Corry, D. T., Beaton, A. and Hart, B. J.: Poplar (*Populus* spp)  
1102 growth and crop yields in a silvoarable experiment at three lowland sites in England, *Agrofor.*  
1103 *Syst.*, 63, 157–169, 2004.

1104 Cardinael, R., Mao, Z., Prieto, I., Stokes, A., Dupraz, C., Kim, J. H. and Jourdan, C.:  
1105 Competition with winter crops induces deeper rooting of walnut trees in a Mediterranean alley

1106 cropping agroforestry system, *Plant Soil*, 391, 219–235, 2015a.

1107 Cardinael, R., Chevallier, T., Barthès, B. G., Saby, N. P. ., Parent, T., Dupraz, C., Bernoux, M.  
1108 and Chenu, C.: Impact of alley cropping agroforestry on stocks, forms and spatial distribution  
1109 of soil organic carbon - A case study in a Mediterranean context, *Geoderma*, 259–260, 288–  
1110 299, 2015b.

1111 Cardinael, R., Eglin, T., Guenet, B., Neill, C., Houot, S. and Chenu, C.: Is priming effect a  
1112 significant process for long-term SOC dynamics? Analysis of a 52-years old experiment,  
1113 *Biogeochemistry*, 123, 203–219, 2015c.

1114 Cardinael, R., Chevallier, T., Cambou, A., Béral, C., Barthès, B. G., Dupraz, C., Durand, C.,  
1115 Kouakoua, E. and Chenu, C.: Increased soil organic carbon stocks under agroforestry: A survey  
1116 of six different sites in France, *Agric. Ecosyst. Environ.*, 236, 243–255, 2017.

1117 Carney, K. M., Hungate, B. A., Drake, B. G. and Megonigal, J. P.: Altered soil microbial  
1118 community at elevated CO<sub>2</sub> leads to loss of soil carbon, *PNAS*, 104, 4990–4995, 2007.

1119 Charbonnier, F., le Maire, G., Dreyer, E., Casanoves, F., Christina, M., Dauzat, J., Eitel, J. U.  
1120 H., Vaast, P., Vierling, L. a. and Roupsard, O.: Competition for light in heterogeneous canopies:  
1121 Application of MAESTRA to a coffee (*Coffea arabica* L.) agroforestry system, *Agric. For.*  
1122 *Meteorol.*, 181, 152–169, 2013.

1123 Chaudhry, A. K., Khan, G. S., Siddiqui, M. T., Akhtar, M. and Aslam, Z.: Effect of arable crops  
1124 on the growth of poplar (*Populus deltoides*) tree in agroforestry system, *Pakistan J. Agric. Sci.*,  
1125 40, 82–85, 2003.

1126 Chiffhot, V., Bertoni, G., Cabanettes, A. and Gavaland, A.: Beneficial effects of intercropping  
1127 on the growth and nitrogen status of young wild cherry and hybrid walnut trees, *Agrofor. Syst.*,  
1128 66, 13–21, 2006.

1129 Clinch, R. L., Thevathasan, N. V., Gordon, A. M., Volk, T. A. and Sidders, D.: Biophysical

1130 interactions in a short rotation willow intercropping system in southern Ontario, Canada, *Agric.*  
1131 *Ecosyst. Environ.*, 131, 61–69, 2009.

1132 Conant, R. T., Ryan, M. G., Ågren, G. I., Birge, H. E., Davidson, E. A., Eliasson, P. E., Evans,  
1133 S. E., Frey, S. D., Giardina, C. P., Hopkins, F. M., Hyvönen, R., Kirschbaum, M. U. F.,  
1134 Lavallee, J. M., Leifeld, J., Parton, W. J., Megan Steinweg, J., Wallenstein, M. D., Martin  
1135 Wetterstedt, J. Å. and Bradford, M. A.: Temperature and soil organic matter decomposition  
1136 rates - synthesis of current knowledge and a way forward, *Glob. Chang. Biol.*, 17, 3392–3404,  
1137 2011.

1138 Cotrufo, M. F., Wallenstein, M. D., Boot, C. M., Deneff, K. and Paul, E.: The Microbial  
1139 Efficiency-Matrix Stabilization (MEMS) framework integrates plant litter decomposition with  
1140 soil organic matter stabilization: do labile plant inputs form stable soil organic matter?, *Glob.*  
1141 *Chang. Biol.*, 19, 988–95, 2013.

1142 Davidson, E. A. and Janssens, I. A.: Temperature sensitivity of soil carbon decomposition and  
1143 feedbacks to climate change, *Nature*, 440, 165–173, 2006.

1144 Dimassi, B., Cohan, J.-P., Labreuche, J. and Mary, B.: Changes in soil carbon and nitrogen  
1145 following tillage conversion in a long-term experiment in Northern France, *Agric. Ecosyst.*  
1146 *Environ.*, 169, 12–20, 2013.

1147 Dubbert, M., Mosena, A., Piayda, A., Cuntz, M., Correia, A. C., Pereira, J. S. and Werner, C.:  
1148 Influence of tree cover on herbaceous layer development and carbon and water fluxes in a  
1149 Portuguese cork-oak woodland, *Acta Oecologica*, 59, 35–45, 2014.

1150 Dufour, L., Metay, A., Talbot, G. and Dupraz, C.: Assessing Light Competition for Cereal  
1151 Production in Temperate Agroforestry Systems using Experimentation and Crop Modelling, *J.*  
1152 *Agron. Crop Sci.*, 199, 217–227, 2013.

1153 Dunbabin, V. M., Postma, J. A., Schnepf, A., Pagès, L., Javaux, M., Wu, L., Leitner, D., Chen,

1154 Y. L., Rengel, Z. and Diggle, A. J.: Modelling root-soil interactions using three-dimensional  
1155 models of root growth, architecture and function, *Plant Soil*, 372, 93–124, 2013.

1156 Dupuy, L., Gregory, P. J. and Bengough, A. G.: Root growth models: Towards a new generation  
1157 of continuous approaches, *J. Exp. Bot.*, 61, 2131–2143, 2010.

1158 Duursma, R. . and Medlyn, B. .: MAESPA: a model to study interactions between water  
1159 limitation, environmental drivers and vegetation function at tree and stand levels, with an  
1160 example application to [CO<sub>2</sub>] × drought interactions, *Geosci. Model Dev.*, 5, 919–940, 2012.

1161 Eilers, K. G., Debenport, S., Anderson, S. and Fierer, N.: Digging deeper to find unique  
1162 microbial communities: The strong effect of depth on the structure of bacterial and archaeal  
1163 communities in soil, *Soil Biol. Biochem.*, 50, 58–65, 2012.

1164 Eissenstat, D. M. and Yanai, R. D.: The Ecology of Root Lifespan, *Adv. Ecol. Res.*, 27, 1–60,  
1165 1997.

1166 Ellert, B. H. and Bettany, J. R.: Calculation of organic matter and nutrients stored in soils under  
1167 contrasting management regimes, *Can. J. Soil Sci.*, 75, 529–538, 1995.

1168 Elzein, A. and Balesdent, J.: Mechanistic simulation of vertical distribution of carbon  
1169 concentrations and residence times in soils, *Soil Sci. Soc. Am. J.*, 59, 1328–1335, 1995.

1170 Fierer, N., Schimel, J. P. and Holden, P. A.: Variations in microbial community composition  
1171 through two soil depth profiles, *Soil Biol. Biochem.*, 35, 167–176, 2003.

1172 Fontaine, S., Barot, S., Barré, P., Bdioui, N., Mary, B. and Rumpel, C.: Stability of organic  
1173 carbon in deep soil layers controlled by fresh carbon supply, *Nature*, 450, 277–281, 2007.

1174 Germon, A., Cardinael, R., Prieto, I., Mao, Z., Kim, J. H., Stokes, A., Dupraz, C., Laclau, J.-P.  
1175 and Jourdan, C.: Unexpected phenology and lifespan of shallow and deep fine roots of walnut  
1176 trees grown in a silvoarable Mediterranean agroforestry system, *Plant Soil*, 401, 409–426, 2016.

1177 Graves, A. R., Burgess, P. J., Palma, J. H. N., Herzog, F., Moreno, G., Bertomeu, M., Dupraz,

1178 C., Liagre, F., Keesman, K., van der Werf, W., de Nooy, a. K. and van den Briel, J. P.:  
1179 Development and application of bio-economic modelling to compare silvoarable, arable, and  
1180 forestry systems in three European countries, *Ecol. Eng.*, 29, 434–449, 2007.

1181 Graves, A. R., Burgess, P. J., Palma, J., Keesman, K. J., van der Werf, W., Dupraz, C., van  
1182 Keulen, H., Herzog, F. and Mayus, M.: Implementation and calibration of the parameter-sparse  
1183 Yield-SAFE model to predict production and land equivalent ratio in mixed tree and crop  
1184 systems under two contrasting production situations in Europe, *Ecol. Modell.*, 221, 1744–1756,  
1185 2010.

1186 van Groenigen, K. J., Qi, X., Osenberg, C. W., Luo, Y. and Hungate, B. A.: Faster  
1187 decomposition under increased atmospheric CO<sub>2</sub> limits soil carbon storage, *Science*, 344, 508–  
1188 509, 2014.

1189 Guenet, B., Eglin, T., Vasilyeva, N., Peylin, P., Ciais, P. and Chenu, C.: The relative importance  
1190 of decomposition and transport mechanisms in accounting for soil organic carbon profiles,  
1191 *Biogeosciences*, 10, 2379–2392, 2013.

1192 Guenet, B., Moyano, F. E., Peylin, P., Ciais, P. and Janssens, I. A.: Towards a representation  
1193 of priming on soil carbon decomposition in the global land biosphere model ORCHIDEE  
1194 (version 1.9.5.2), *Geosci. Model Dev.*, 9, 841–855, 2016.

1195 Haile, S. G., Nair, V. D. and Nair, P. K. R.: Contribution of trees to carbon storage in soils of  
1196 silvopastoral systems in Florida, USA, *Glob. Chang. Biol.*, 16, 427–438, 2010.

1197 Hendrick, R. L. and Pregitzer, K. S.: Temporal and depth-related patterns of fine root dynamics  
1198 in northern hardwood forests, *J. Ecol.*, 84, 167–176, 1996.

1199 Howlett, D. S., Moreno, G., Mosquera Losada, M. R., Nair, P. K. R. and Nair, V. D.: Soil  
1200 carbon storage as influenced by tree cover in the Dehesa cork oak silvopasture of central-  
1201 western Spain, *J. Environ. Monit.*, 13, 1897–904, 2011.

1202 Ilstedt, U., Bargués Tobella, A., Bazié, H. R., Bayala, J., Verbeeten, E., Nyberg, G., Sanou, J.,  
1203 Benegas, L., Murdiyarso, D., Laudon, H., Sheil, D. and Malmer, A.: Intermediate tree cover  
1204 can maximize groundwater recharge in the seasonally dry tropics, *Sci. Rep.*, 6, 21930, 2016.

1205 IUSS Working Group WRB: World Reference Base for Soil Resources 2006, first update 2007.  
1206 World Soil Resources Reports No. 103. FAO, Rome., 2007.

1207 Jobbagy, E. G. and Jackson, R. B.: The vertical distribution of soil organic carbon and its  
1208 relation to climate and vegetation, *Ecol. Appl.*, 10, 423–436, 2000.

1209 Joslin, J. D., Gaudinski, J. B., Torn, M. S., Riley, W. J. and Hanson, P. J.: Fine-root turnover  
1210 patterns and their relationship to root diameter and soil depth in a <sup>14</sup>C-labeled hardwood forest,  
1211 *New Phytol.*, 172, 523–535, 2006.

1212 Kätterer, T., Bolinder, M. A., Andrén, O., Kirchmann, H. and Menichetti, L.: Roots contribute  
1213 more to refractory soil organic matter than above-ground crop residues, as revealed by a long-  
1214 term field experiment, *Agric. Ecosyst. Environ.*, 141, 184–192, 2011.

1215 Keiluweit, M., Bougoure, J. J., Nico, P. S., Pett-Ridge, J., Weber, P. K. and Kleber, M.: Mineral  
1216 protection of soil carbon counteracted by root exudates, *Nat. Clim. Chang.*, 5, 588–595, 2015.

1217 Kim, D.-G., Kirschbaum, M. U. F. and Beedy, T. L.: Carbon sequestration and net emissions  
1218 of CH<sub>4</sub> and N<sub>2</sub>O under agroforestry: Synthesizing available data and suggestions for future  
1219 studies, *Agric. Ecosyst. Environ.*, 226, 65–78, 2016.

1220 Koarashi, J., Hockaday, W. C., Masiello, C. A. and Trumbore, S. E.: Dynamics of decadal  
1221 cycling carbon in subsurface soils, *J. Geophys. Res.*, 117, 1–13, 2012.

1222 Koven, C. D., Riley, W. J., Subin, Z. M., Tang, J. Y., Torn, M. S., Collins, W. D., Bonan, G.  
1223 B., Lawrence, D. M. and Swenson, S. C.: The effect of vertically resolved soil biogeochemistry  
1224 and alternate soil C and N models on C dynamics of CLM4, *Biogeosciences*, 10, 7109–7131,  
1225 2013.

1226 Lange, M., Eisenhauer, N., Sierra, C. A., Bessler, H., Engels, C., Griffiths, R. I., Mellado-  
1227 Vázquez, P. G., Malik, A. A., Roy, J., Scheu, S., Steinbeiss, S., Thomson, B. C., Trumbore, S.  
1228 E. and Gleixner, G.: Plant diversity increases soil microbial activity and soil carbon storage,  
1229 *Nat. Commun.*, 6, 6707, 2015.

1230 Lavelle, P.: Faunal activities and soil processes: adaptative strategy that determine ecosystem  
1231 function., 1997.

1232 Li, F., Meng, P., Fu, D. and Wang, B.: Light distribution, photosynthetic rate and yield in a  
1233 Paulownia-wheat intercropping system in China, *Agrofor. Syst.*, 74, 163–172, 2008.

1234 Lorenz, K. and Lal, R.: Soil organic carbon sequestration in agroforestry systems. A review,  
1235 *Agron. Sustain. Dev.*, 34, 443–454, 2014.

1236 Luedeling, E., Smethurst, P. J., Baudron, F., Bayala, J., Huth, N. I., van Noordwijk, M., Ong,  
1237 C. K., Mulia, R., Lusiana, B., Muthuri, C. and Sinclair, F. L.: Field-scale modeling of tree-crop  
1238 interactions: Challenges and development needs, *Agric. Syst.*, 142, 51–69, 2016.

1239 Manzoni, S., Piñeiro, G., Jackson, R. B., Jobbágy, E. G., Kim, J. H. and Porporato, A.:  
1240 Analytical models of soil and litter decomposition: Solutions for mass loss and time-dependent  
1241 decay rates, *Soil Biol. Biochem.*, 50, 66–76, 2012.

1242 Mead, R. and Willey, R. W.: The concept of a “land equivalent ratio” and advantages in yields  
1243 from intercropping, *Exp. Agric.*, 16, 217–228, 1980.

1244 Moreno, G., Obrador, J. J., Cubera, E. and Dupraz, C.: Fine root distribution in Dehesas of  
1245 central-western Spain, *Plant Soil*, 277, 153–162, 2005.

1246 Moyano, F. E., Vasilyeva, N., Bouckaert, L., Cook, F., Craine, J., Curiel Yuste, J., Don, a.,  
1247 Epron, D., Formanek, P., Franzluebbers, a., Ilstedt, U., Kätterer, T., Orchard, V., Reichstein,  
1248 M., Rey, A., Ruamps, L., Subke, J. A., Thomsen, I. K. and Chenu, C.: The moisture response  
1249 of soil heterotrophic respiration: Interaction with soil properties, *Biogeosciences*, 9, 1173–

1250 1182, 2012.

1251 Moyano, F. E., Manzoni, S. and Chenu, C.: Responses of soil heterotrophic respiration to  
1252 moisture availability: An exploration of processes and models, *Soil Biol. Biochem.*, 59, 72–85,  
1253 2013.

1254 Mulia, R. and Dupraz, C.: Unusual fine root distributions of two deciduous tree species in  
1255 southern France: What consequences for modelling of tree root dynamics?, *Plant Soil*, 281, 71–  
1256 85, 2006.

1257 Mulia, R., Dupraz, C. and van Noordwijk, M.: Reconciling root plasticity and architectural  
1258 ground rules in tree root growth models with voxel automata, *Plant Soil*, 337, 77–92, 2010.

1259 Nair, P. K.: An introduction to agroforestry, Kluwer Academic Publishers, Dordrecht, The  
1260 Netherlands., 1993.

1261 Nair, P. K. R.: Classification of agroforestry systems, *Agrofor. Syst.*, 3, 97–128, 1985.

1262 van Noordwijk, M. and Lusiana, B.: WaNuLCAS, a model of water, nutrient and light capture  
1263 in agroforestry systems, *Agrofor. Syst.*, 43, 217–242, 1999.

1264 Odhiambo, H. O., Ong, C. K., Deans, J. D., Wilson, J., Khan, A. A. H. and Sprent, J. I.: Roots,  
1265 soil water and crop yield: Tree crop interactions in a semi-arid agroforestry system in Kenya,  
1266 *Plant Soil*, 235, 221–233, 2001.

1267 Oelbermann, M. and Voroney, R. P.: An evaluation of the century model to predict soil  
1268 organic carbon: examples from Costa Rica and Canada, *Agrofor. Syst.*, 82, 37–50, 2011.

1269 Oelbermann, M., Voroney, R. P. and Gordon, A. M.: Carbon sequestration in tropical and  
1270 temperate agroforestry systems: a review with examples from Costa Rica and southern Canada,  
1271 *Agric. Ecosyst. Environ.*, 104, 359–377, 2004.

1272 Oelbermann, M., Voroney, R. P., Thevathasan, N. V., Gordon, A. M., Kass, D. C. L. and  
1273 Schlönvoigt, A. M.: Soil carbon dynamics and residue stabilization in a Costa Rican and



1274 southern Canadian alley cropping system, *Agrofor. Syst.*, 68, 27–36, 2006.

1275 Ong, C. K. and Leakey, R. R. B.: Why tree-crop interactions in agroforestry appear at odds with  
1276 tree-grass interactions in tropical savannahs, *Agrofor. Syst.*, 45, 109–129, 1999.

1277 Parton, W. J., Schimel, D. S., Cole, C. V and Ojima, D. S.: Analysis of factors controlling soil  
1278 organic matter levels in great plains grasslands, *Soil Sci. Soc. Am. J.*, 51, 1173–1179, 1987.

1279 Peichl, M., Thevathasan, N. V, Gordon, A. M., Huss, J. and Abohassan, R. A.: Carbon  
1280 sequestration potentials in temperate tree-based intercropping systems, southern Ontario,  
1281 Canada, *Agrofor. Syst.*, 66, 243–257, 2006.

1282 Perveen, N., Barot, S., Alvarez, G., Klumpp, K., Martin, R., Rapaport, A., Herfurth, D.,  
1283 Louault, F. and Fontaine, S.: Priming effect and microbial diversity in ecosystem functioning  
1284 and response to global change: A modeling approach using the SYMPHONY model, *Glob.*  
1285 *Chang. Biol.*, 20, 1174–1190, 2014.

1286 Price, G. W. and Gordon, A. M.: Spatial and temporal distribution of earthworms in a temperate  
1287 intercropping system in southern Ontario, Canada, *Agrofor. Syst.*, 44, 141–149, 1999.

1288 Prieto, I., Roumet, C., Cardinael, R., Kim, J., Maeght, J.-L., Mao, Z., Portillo, N.,  
1289 Thammahacksa, C., Dupraz, C., Jourdan, C., Pierret, A., Roupsard, O. and Stokes, A.: Root  
1290 functional parameters along a land-use gradient: evidence of a community-level economics  
1291 spectrum, *J. Ecol.*, 103, 361–373, 2015.

1292 Prieto, I., Stokes, A. and Roumet, C.: Root functional parameters predict fine root  
1293 decomposability at the community level, *J. Ecol.*, 104, 725–733, 2016.

1294 R Development Core Team: R: A language and environment for statistical computing, 2013.

1295 Rasse, D. P., Mulder, J., Moni, C. and Chenu, C.: Carbon turnover kinetics with depth in a  
1296 French loamy soil, *Soil Sci. Soc. Am. J.*, 70, 2097–2105, 2006.

1297 Salomé, C., Nunan, N., Pouteau, V., Lerch, T. Z. and Chenu, C.: Carbon dynamics in topsoil

1298 and in subsoil may be controlled by different regulatory mechanisms, *Glob. Chang. Biol.*, 16,  
1299 416–426, 2010.

1300 Santaren, D., Peylin, P., Viovy, N. and Ciais, P.: Optimizing a process-based ecosystem model  
1301 with eddy-covariance flux measurements: A pine forest in southern France, *Global*  
1302 *Biogeochem. Cycles*, 21, 1–15, 2007.

1303 Shahzad, T., Chenu, C., Genet, P., Barot, S., Perveen, N., Mougin, C. and Fontaine, S.:  
1304 Contribution of exudates, arbuscular mycorrhizal fungi and litter depositions to the rhizosphere  
1305 priming effect induced by grassland species, *Soil Biol. Biochem.*, 80, 146–155, 2015.

1306 Sierra, C. A., Trumbore, S. E., Davidson, E. A., Vicca, S. and Janssens, I.: Sensitivity of  
1307 decomposition rates of soil organic matter with respect to simultaneous changes in temperature  
1308 and moisture, *J. Adv. Model. Earth Syst.*, 7, 335–356, 2015.

1309 Soetaert, K., Petzoldt, T. and Woodrow Setzer, R.: Solving Differential Equations in R:  
1310 Package deSolve, *J. Stat. Softw.*, 33, 1–25, 2010.

1311 Somarriba, E.: Revisiting the past: an essay on agroforestry definition, *Agrofor. Syst.*, 19(3),  
1312 233–240, 1992.

1313 Steinbeiss, S., Beßler, H., Engels, C., Temperton, V. M., Buchmann, N., Roscher, C.,  
1314 Kreutziger, Y., Baade, J., Habekost, M. and Gleixner, G.: Plant diversity positively affects  
1315 short-term soil carbon storage in experimental grasslands, *Glob. Chang. Biol.*, 14, 2937–2949,  
1316 2008.

1317 Sulman, B. N., Phillips, R. P., Oishi, A. C., Shevliakova, E. and Pacala, S. W.: Microbe-driven  
1318 turnover offsets mineral-mediated storage of soil carbon under elevated CO<sub>2</sub>, *Nat. Clim.*  
1319 *Chang.*, 4, 1099–1102, 2014.

1320 Taghizadeh-Toosi, A., Christensen, B. T., Hutchings, N. J., Vejlin, J., Kätterer, T., Glendining,  
1321 M. and Olesen, J. E.: C-TOOL: A simple model for simulating whole-profile carbon storage in

1322 temperate agricultural soils, *Ecol. Modell.*, 292, 11–25, 2014.

1323 Talbot, G.: L'intégration spatiale et temporelle du partage des ressources dans un système  
1324 agroforestier noyers-céréales: une clef pour en comprendre la productivité ?, PhD Dissertation,  
1325 Université Montpellier II., 2011.

1326 Tarantola, A.: *Inverse problem theory: methods for data fitting and model parameter estimation*,  
1327 edited by Elsevier., 1987.

1328 Tarantola, A.: *Inverse Problem Theory and Methods for Model Parameter Estimation*, edited  
1329 by SIAM., 2005.

1330 Thevathasan, N. V. and Gordon, A. M.: Poplar leaf biomass distribution and nitrogen dynamics  
1331 in a poplar-barley intercropped system in southern Ontario, Canada, *Agrofor. Syst.*, 37, 79–90,  
1332 1997.

1333 Udawatta, R. P., Kremer, R. J., Adamson, B. W. and Anderson, S. H.: Variations in soil  
1334 aggregate stability and enzyme activities in a temperate agroforestry practice, *Appl. Soil Ecol.*,  
1335 39, 153–160, 2008.

1336 Virto, I., Barré, P., Burlot, A. and Chenu, C.: Carbon input differences as the main factor  
1337 explaining the variability in soil organic C storage in no-tilled compared to inversion tilled  
1338 agrosystems, *Biogeochemistry*, 108, 17–26, 2012.

1339 van der Werf, W., Keesman, K., Burgess, P., Graves, A., Pilbeam, D., Incoll, L. D., Metselaar,  
1340 K., Mayus, M., Stappers, R., van Keulen, H., Palma, J. and Dupraz, C.: Yield-SAFE: A  
1341 parameter-sparse, process-based dynamic model for predicting resource capture, growth, and  
1342 production in agroforestry systems, *Ecol. Eng.*, 29, 419–433, 2007.

1343 Wutzler, T. and Reichstein, M.: Colimitation of decomposition by substrate and decomposers  
1344 - a comparison of model formulations, *Biogeosciences*, 5, 749–759, 2008.

1345 Wutzler, T. and Reichstein, M.: Priming and substrate quality interactions in soil organic matter

1346 models, *Biogeosciences*, 10, 2089–2103, 2013.

1347 Yin, R. and He, Q.: The spatial and temporal effects of paulownia intercropping: The case of  
1348 northern China, *Agrofor. Syst.*, 37, 91–109, 1997.

1349 Zhang, W., Wang, X. and Wang, S.: Addition of external organic carbon and native soil organic  
1350 carbon decomposition: a meta-analysis., *PLoS One*, 8, e54779, 2013.

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