

# ***Interactive comment on* “Distribution of planktonic biogenic carbonate organisms in the Southern Ocean south of Australia: a baseline for ocean acidification impact assessment” by Thomas W. Trull et al.**

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Review on: “Distribution of planktonic biogenic carbonate organisms in the Southern Ocean south of Australia: a baseline for ocean acidification impact assessment “ by Trull et al. In this study, Trull et al., investigate Diatom and calcifier distribution patterns in the Southern Ocean. Their analysis is based on BSi, POC and size fractionated PIC data. They compare their ground truth data with satellite data and model predictions and report important discrepancies and consistencies. I think their study is very valuable and their paper contains key information to document climate change effects on

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diC1 BGD Interactive comment Printer-friendly version Discussion paper atoms and calcifiers in the Southern Ocean. I really only have minor comments. Some of these are addressing their methods and some refer to the discussion/conclusion part. Line 26: Are diatoms really the most abundant phytoplankton? I can understand that they might be dominant in terms of biomass but would intuitively assume that smaller groups (e.g. picoeukaryotes such as *Micromonas*) are more abundant than diatoms. (I may be wrong here but just to double check.) Reviewer is correct. Sentence modified as follows: Ancillary measurements of biogenic silica (BSi) and particulate organic carbon (POC) provided context, as estimates of the biomass of diatoms (the highest biomass phytoplankton in polar waters), and total microbial biomass, respectively.

Line 56: I am not sure that the under-saturation is primarily due to low TA. I would assume that it is due to the low temperature that leads to generally low carbonate ion concentration. Reviewer is correct – the dominant effect is temperature. For example, (based on CO2SYS with standard default constants), for seawater Salinity=35, Alkalinity=2320  $\mu\text{mol/kg}$  waters in equilibrium with  $p\text{CO}_2=400$   $\mu\text{atm}$  air, cooling from 15 to 5 C reduces the carbonate anion concentration from 160 to 112  $\mu\text{mol/kg}$ , whereas at 15C dilution of salinity from 35 to 33 and alkalinity proportionally reduces carbonate anion concentrations from 160 to 145  $\mu\text{mol/kg}$ . That is, temperature accounts for ~80% of the total effect. Sentence modified as follows: The low temperature and low alkalinity of Southern Ocean waters make this region particularly susceptible to ocean acidification, . . . .

Line 63: It is a bit weird that you say that their relative importance is poorly known but then in the same sentence say that they will have an influence ecosystem health. The second part of the sentence implicitly contradicts the first part. Furthermore, I did not understand how the “importance” will “influence of the overall impact”. This sentence could perhaps be rephrased. Agreed, and we have reordered and rewritten these sentences to make the issue clearer: Carbonate forming organisms in the Southern Ocean include coccolithophores (the dominant carbonate forming phytoplankton; e.g.

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[Rost and Riebesell, 2004]), foraminifera (the dominant carbonate forming zooplankton; e.g. [Moy et al., 2009; Schiebel, 2002]), and pteropods (a larger carbonate forming zooplankton, which can be an important component of fish diets; e.g. [Doubleday and Hopcroft, 2015; Roberts et al., 2014]). However, the importance of carbonate forming organisms relative to other taxa is unclear in the Southern Ocean [Watson W. Gregg and Casey, 2007b; Holligan et al., 2010].

Line 80: Aren't these results? Perhaps move this sentence to results part. Furthermore, I do not understand the use of the second "suggested" in this sentence. Please check. Agreed. Result sentence removed.

Line 91: In this context it may also be useful to remind the reader that the PIC50 fraction could also contain aggregated coccolithophore calcite (e.g. within fecal pellets). We don't think this is likely, and accordingly we have not added this possibility to the text. The PIC50 fraction is collected at a high flow rate, sufficient to disaggregate most faecal matter. This perspective was corroborated by not seeing faecal pellets during visual inspection of the 50 $\mu$ m mesh screens to remove rare zooplankton. We added text explaining this perspective in the Methods section: Based on visual examination, the high flow rate through the 50  $\mu$ m nylon mesh was sufficient to disaggregate faecal pellets and detrital aggregates.

Line 152: It would be helpful to know whether or not you expect a loss of CaCO<sub>3</sub> by sieving the samples. Are there large quantities of CaCO<sub>3</sub> expected in the >1000  $\mu$ m size fraction? In our experience the >1000 $\mu$ m fraction rarely contains anything, and early attempts to analyse this fraction yielded negligible CaCO<sub>3</sub>. The filter is occasionally useful for preventing krill and other large zooplankton from entering the filtration system. We have not tried to assess what would be "expected" in this large fraction, because we are not aware of data that would make this possible and we consider that the ship intake is unlikely to provide an unbiased sampling of organisms of this size which are often both mobile and rare. We made no changes to the text.

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Line 152: What do you mean by “ship clean”? Please clarify. Text has been added to clarify the meaning: All samples were collected from the ships’ underway “clean” seawater supply lines with intakes at ~4 m depth. These supply lines are separate from the engine intakes, have scheduled maintenance and cleaning, and are only turned on offshore (to avoid possible contamination from coastal waters).

Line 154: Can you provide any information if the 50  $\mu\text{m}$  filter tended to block when such a large volume is filtered? I am asking because it could be that towards the end of the filtration process also smaller particles might have been retained on the filter due to clogging. I know this is difficult to reconstruct, but in case you have any further information it would be useful to provide them. I have personally made bad experiences with sequential filtrations. The short answers are that: 1. We were aware of this potential problem and designed our filtration processes to minimize it. We do not consider that clogging was a problem. 2. If clogging retained PIC on the 50  $\mu\text{m}$  filter, than our PIC01 estimates would too low and our PIC50 estimates would be too high. Because the fraction of total PIC on the PIC50 mesh was generally quite small (10% or less), this possible redistribution does not affect any of our conclusions. The longer answer is that it is challenging to create a filtration system capable of filtering water across the diverse conditions of the entire Southern Ocean. Our filtration system evolved over time, partly to deal with the issue of filter clogging. The first leg (VL1) was a purely sequential filtration system where the volume of water filtered was sometimes limited by either filter clogging, so that insufficient material was obtained on the other filter. For this reason prior to VL2 we added a pressure relief valve between the 50 $\mu\text{m}$  and 1 $\mu\text{m}$  filters which allowed large volumes of water to pass through the 50 $\mu\text{m}$  filter and bypass the 1 $\mu\text{m}$  filter. The second improvement prior to VL6 was the introduction of digital flow meters which recorded instantaneous flow rates. The final improvement prior to VL8 was the introduction of electronically controlled ball valves that stopped filtration when flow rates fell below threshold values (0.5L/min hiflo and 0.05L/min loflo). We believe these stop thresholds are very conservative and the filters are not truly clogged at this stage. Using our final configuration of the system, on VL8 & VL9,

samples from the lower latitudes (approx. 44S - 50S) have reasonably high levels of 1-50 $\mu$ m particles which can reduce flow rates through the 1  $\mu$ m filter below our cut-off threshold of 0.05L/min prior to the 2 hour filtration time limit (Figure 1). However, very large volumes of water pass through the 50  $\mu$ m filter (Figure 1). In mid latitudes (approx. 50S – 58S) flow rates remain high through both filters and most samples filter for 2 hours (Figure 1). At high latitudes (approx. 58S - 68S) flow rates through the 50  $\mu$ m filter often reduced rapidly to the 0.5L/min cut-off threshold because the 50 $\mu$ m filter collects many large chain forming diatoms. In our filtration system this shuts down filtration through both filters which is reflected in the low filtration times and volumes (Figure 1). Ideally we would like to filter larger volumes of water at high latitudes which may require a filtration system capable of switching to a second 50  $\mu$ m filter or something similar. For VL8 & VL9 with their conservative cut-off thresholds we believe that clogging of the 50  $\mu$ m filter and retention of smaller particles is unlikely. The picture is less clear for earlier legs where instantaneous flow rate data was not available and flowrate cut-off thresholds were not used. However, the data across all voyages shows quite consistent trends in PIC concentration and the PIC50/PIC01 ratio. We have added these filtration times and volumes for all PIC samples in Table S1, included this discussion and figures in the Supplementary Material, and added a sentence into the main text pointing to this material: The flow rate and flow volume data also suggests that filter clogging was uncommon (see the Supplementary Information for expanded discussion).

Line 158: I am a bit nervous about the PIC filter cleaning procedure. Omega is 0 in the deionized water and the pH is (probably) low. Does the deionized water have the potential to dissolve CaCO<sub>3</sub>? We also were nervous about this. Accordingly we used degassed de-ionized water (boiling to remove CO<sub>2</sub> and obtain close to neutral pH). The contact time of seconds and no loss of sharp edges on coccolithophores collected in this way and examined by scanning electron microscopy (Cubillos et al., 2007) reassured us. We added the following text: We consider that this rinse did not dissolve PIC, based on the sharp (non-eroded) features of coccolithophores collected

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in this way and examined by scanning electron microscopy (Cubillos et al., 2007).

Line 336: “as resulting” twice. Deleted one “as resulting”

Line 402: I do not understand why mesoscale variability makes the comparison difficult. If you are at a certain location with a ship and sample PIC and you have satellite data for the very same time, you could easily compare these values, couldn't you? Strong mesoscale variability means that the match-up length scale must be very small. This limits the amount of match-ups that can be achieved. The variability length scale can also be smaller than the satellite pixel size. The correlation length scale for chlorophyll in the Southern Ocean degrades at distances > 10-15 km, as recently shown in attempting to match Biogeochemical-Argo fluorescent chlorophyll and satellite ocean colour estimates for a large set of observations (Haëntjens N, Boss E, Talley LD (2017) Revisiting Ocean Color algorithms for chlorophyll a and particulate organic carbon in the Southern Ocean using biogeochemical floats. *Journal of Geophysical Research: Oceans* 122:6583-6593).

Using a somewhat longer match-up length scale of 25 km (i.e. the ship and satellite observations must be within 25 km of each other on the same day), we were able to retain 116 match-ups and we have added these results to the paper. The match-ups tend to occur in clusters of several samples along a transit when the ship encountered cloud-free conditions, so that the amount of independent observations is less than this. Nonetheless the match-up results are valuable and they confirm that the satellite SPIC values are reasonable estimates in Subantarctic waters but very much too high in Antarctic waters.

The new results are described by a new figure and new text: Both cloudy conditions and strong mesoscale variability limit the number of direct comparisons (match-ups) that can be made. Using a match-up length scale of 25 km (i.e. the ship and satellite observations must be within 25 km of each other on the same day), which is somewhat larger than the correlation length scale for chlorophyll in the Southern Ocean of 10-

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15 km [Haëntjens et al., 2017], allowed us to retain 116 match-ups. These results, shown in Figure 6, confirm that the satellite SPIC values are reasonable estimates in Subantarctic waters, within a factor of 2-3 [W M Balch et al., 2011], but very much too high in Antarctic waters.

Line 405: What is “e-folding”? The term has not been introduced. This is a common term to describe exponential behaviour, e.g. from Wikipedia: “In science, e-folding is the time interval in which an exponentially growing quantity increases by a factor of e; it is the base-e analog of doubling time.” No change was made to the text.

Line 469: Dominant in terms of abundance? Dominant in terms of biomass would probably be the more important metric here. Agreed, and we changed “abundance” to “biomass” here and throughout this paragraph.

Lines 470 and 473 : These results imply that diatoms (and to a limited extent coccolithophores) more or less exclusively contribute to the bulk POC in Antarctic waters. I am not so experienced with the plankton communities in the Southern Ocean but would intuitively disagree. Is it really possible that diatoms are so dominant? What about grazers? Did the analysis include e.g. copepod as a POC source or were these not captured on the filters? I think the result of bulk POC = diatom POC in the Antarctic is very interesting.

We have added a qualifying sentence as follows (in italics here but not in text): As shown in Figure 4b, this suggests that diatoms dominate the accumulation of organic carbon throughout the Southern Ocean, with coccolithophores generally contributing less than half that of diatoms in the SAZ and less than a tenth of that in Antarctic waters. This statement is of course limited to POC captured by our small volume, size limited (1-1000  $\mu\text{m}$ ) sampling procedure, and variability in the extent of dominance and the scaling of POC to biogenic minerals still allows significant contributions from other POC sources.

Line 480: Abundance of calcifiers or concentration of  $\text{CaCO}_3$ ? I think you should stick

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to the latter term to be more precise. Agreed, and sentence changed to: Finally, we note that the relatively low levels of PIC across the Southern Ocean as observed here means that POC/PIC ratios are high, greater than 4 in the SAZ and ranging up to 20 in Antarctic waters (Figure 4a).

Line 482: You argue that PIC/POC is low which leads to little influence on the TA mediated reduction of atmospheric CO<sub>2</sub> uptake. I agree with that. However, PIC can induce biogeochemical feedbacks in other ways e.g. through ballasting (as you mention in the paragraph before). So I think that it is not really valid to say that coccolithophores had a limited influence on the uptake capacity of atmospheric CO<sub>2</sub> if you neglect other feedback mechanisms than TA reduction. We agree with the reviewer on the multiple mechanisms of influence of calcification on air-sea CO<sub>2</sub> transfer, and modified this sentence to make clear that only the aspect of alkalinity affects of surface ocean pCO<sub>2</sub> is under consideration: This suggests calcification has a negligible countering impact on the reduction of surface ocean CO<sub>2</sub> partial pressure by phytoplankton uptake, even smaller than the few to ~10% influence identified earlier from deep sediment trap compositions in HNLC [P. W. Boyd and Trull, 2007a] and iron-enriched waters, respectively [Salter et al., 2014].

Section 3.4: In section 3.4 you compare model predictions with field data to test whether they predict meaningful trends. I think this is extremely valuable. I have, however, two comments. 1) You first use the Bach et al., 2015 and Langdon et al. 2000 models. These models only consider carbonate chemistry conditions and no other environmental parameter to predict calcification rates. Your data nicely shows that carbonate chemistry is obviously not the driving factor behind the latitudinal trend in the Southern Ocean because model prediction and latitudinal patterns are inconsistent. The Bach et al., model basically predicts that the carbonate chemistry conditions are close to ideal throughout the Southern Ocean. The Langdon et al., model predicts a decline which reflects the trend in Omega. Both models describe calcification response to carbonate chemistry and not distribution patterns of calcifiers. The reason

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why I mention this is because at the end of this part of the paper you state: “Thus, and unsurprisingly, coccolithophore abundances are clearly not controlled by inorganic carbon chemistry alone” (Lines 603- 604). I could not agree more with this statement. However, the way this is formulated implies to some extent that your finding contradicts what we have concluded in our study. But this is not the case. In Bach et al. (2015) we wrote: “great care must be taken when correlating carbonate chemistry with coccolithophore dispersal because this is by no means the only parameter controlling it. Physical (e.g. temperature), other chemical (e.g. nutrient concentrations), or ecological (e.g. grazing pressure) factors will under many if not most circumstances outweigh the influence of carbonate chemistry conditions, unless differences in the latter are extreme. We will therefore focus the discussion on those cases where differences in carbonate chemistry conditions are rather extreme.” Thus, our valuation is very similar to that of the authors of this manuscript. I would appreciate if you could point out that your main conclusion in this paragraph (that carbonate chemistry is probably not the key factor controlling coccolithophore distribution) is also in line with (and not conflicting with) what we assumed in our studies. 2) We are very happy to do this, and added this information explicitly in the summary of this section: Thus, and unsurprisingly, coccolithophore abundances are clearly not controlled by inorganic carbon chemistry alone. This perspective has been strongly emphasized previously, including by Bach et al., (2015), who noted “ . . .great care must be taken when correlating carbonate chemistry with coccolithophore dispersal because this is by no means the only parameter controlling it. Physical (e.g. temperature), other chemical (e.g. nutrient concentrations), or ecological (e.g. grazing pressure) factors will under many if not most circumstances outweigh the influence of carbonate chemistry conditions. . .”.

2) I am a bit skeptical about the growth rate vs. temperature argument based on the Norberg model. The model predicts a decline of coccolithophore growth rates due to decreasing temperature. This in itself is not convincing because the decrease of growth rate would apply for every other phytoplankton group as well. What you would really have to look at is if the growth rate of coccolithophores decreases over-proportionally

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relative to other phytoplankton species. If this was then case, then you could argue that coccolithophores become less competitive the further South you go. We made no changes in response to this comment, because while it has merit we were already careful to describe at the start of this paragraph that the presentation of the Norberg model was limited to an exploration of the possible response to temperature in a univariate sense: “To provide a brief visualization of the expected univariate response, we fit the “Norberg” thermal optimum envelope model . . . and we already re-emphasize at the end of the paragraph that this exploration was limited in scope: There are of course many other possible explanations (as noted at the start of this section). In addition, in the following paragraphs we were already careful to note again that autotrophic completion was a larger issue - see our response to the next comment.

Accordingly, we made no further changes.

Line 653: In this concluding remark you only consider the bottom-up control on diatom vs. coccolithophore distribution. Have you also considered if top-down mechanisms could have played a role here? Even though there may not be appropriate data available to test this in the present study, it may still be useful to remind the reader that this mechanism exists and could also have played a role. I think the Assmy et al., (2013) study nicely made the case that predators may have an important influence on phytoplankton composition in the Southern Ocean. We had already mentioned this possibility, but have augmented it with a final clause in parentheses to cite the Assmy et al., 2013 study: Importantly, in addition to multivariate environmental control of coccolithophore distributions via their growth rates, there is the possibility of control by resource competition with other autotrophs (presumably mainly for iron) and/or stronger loss terms to grazers in Antarctic than Subantarctic waters ([Assmy et al., 2013] has suggested preferential grazing as a control on community structure; but we have no data to allow us to evaluate this).

Table 1: I think the uppercase 3 also needs to be added to PIC01, POC, and BSi. Agreed and we added upper case 3 to PIC01, POC, BSi

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Figure 1: It would be helpful to add full names and abbreviations of the various fronts to the figure caption. Full names and abbreviations added

Figure 3: One particularly interesting finding presented in Figure 3 is that PIC50 (foraminifera) concentrations are considerably lower than PIC01 (coccolithophores) concentrations except for maybe the most Southern stretch of the transects. Sometimes the discrepancies are orders of magnitude. This implies that coccolithophores are the much more important pelagic calcifiers in the Southern Ocean than foraminifera and pteropods. Is this conclusion valid? If so, I think this finding definitely deserves more attention in your paper. Furthermore, it would be interesting to compare this with the results from Broecker and Clark (2009) who found roughly equal contribution of coccolithophores and foraminifera to the sediment CaCO<sub>3</sub> (although their most southern sample came from 40 South).

We deliberately avoided discussion of the PIC50 distributions in any detail for multiple reasons, as we had stated early in the Introduction. Comparison to sediments would bring in the further complexity of the extent of losses of these organisms after leaving surface waters, and become very speculative. Accordingly, we have not added discussion on this issue, and instead have further strengthened our sentence regarding why we do not discuss these results in any detail in the revised version: We do not discuss the PIC50 results in any detail because of this complexity, because controls on foraminifera distributions appear to involve strongly differing biogeography of several co-dominant taxa, rather than dominance by a single species [Be and Tolderlund, 1971], because the numbers of these organisms collected by our procedures was small, and because assessing these issues is beyond the scope of this paper.

Figure 4: What was the rationale of showing the POC/PIC ratio? I think readers will generally be more familiar with PIC/POC ratios. Both are in common use. We preferred POC/PIC (and BSi/PIC) because these emphasized our key findings that BSi and POC are both much more abundant than PIC. We made no change.

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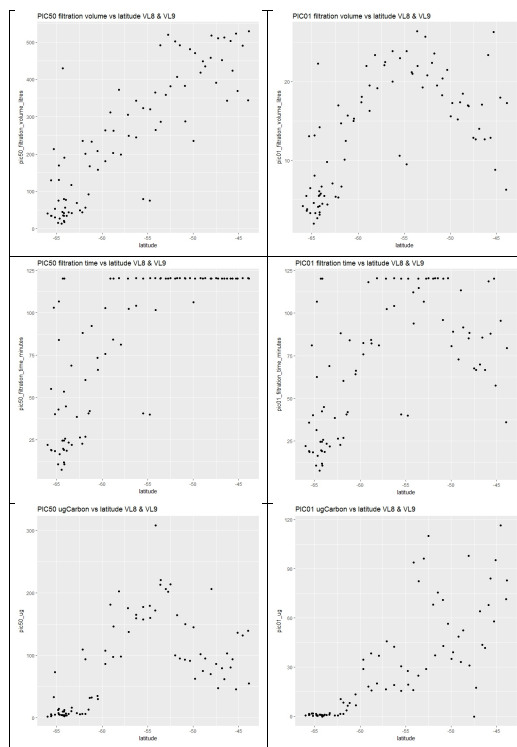


Fig. 1. Fig1 filtration volume and time