Stable isotopes of nitrate reveal different nitrogen processing mechanisms in streams across a land use gradient during wet and dry periods

Response to Reviewer Comments

We thank the reviewers and the associate editor for their constructive comments. We have addressed the reviewers' comments individually (as detailed below) and have revised the manuscript accordingly. Please note that page/line numbers in reviewers' comments refer to the original manuscript while our references to page/line numbers refer to the revised manuscript.

Reviewer #2

GENERAL COMMENTS

Wong and co-authors present a study of stable isotopes of nitrate from five streams within the same catchment area in Southeast Australia sampled during wet and dry periods. The five streams show different degrees of land use intensities. The aim is to reveal different sources and transformation processes of nitrate compared between rainfall patterns through the isotopic composition (d15N-NO3- and d18O-NO3-). Results show that differences between wet and dry periods can be explained by the dominance of different sources on the isotopic composition. During wet periods artificial fertilizer was probably the main source, whereas nitrified organic matter in sediment and nitrified manure dominated the sources during dry periods. The manuscript is well written and presents the results in a logical order. The figures illustrate the findings very well. This novel dataset is suitable for publication in Biogeosciences, however, there are some points that should be addressed by the authors.

MAJOR COMMENTS

(1) To study the impact of rainfall on isotopic composition in a more rigorous way, it would have been interesting to use samplings with different amounts of rainfall in the previous days (instead of only differentiating between wet and dry periods) in order to see whether rainfall and isotopic composition could be correlated. At least the authors should explain why such a study was not carried out.

There was no correlation between the isotope values (both $\delta^{15}N$ and $\delta^{18}O$) of the streams and the total amount of rainfall for 10 days before each sampling event as shown in Fig. 1. A linear relationship between streams nitrate isotopes and rainfall amount might be expected for a pristine environment, however for heavily anthropogenic-affected environments like the Western Port catchment; it is impossible to observe such correlation because of the dominance of other sources of nitrate. In this study, the condition of the soil (i.e. wet versus dry) in the catchment and the residence time of the rivers were affecting the occurrence and the extent of certain biological processes in the catchment and thus the isotope values of the residual nitrate rather than rainfall. We believe this has been shown nicely in the manuscript. The amount of rainfall in this study was used as a direct indicator of soil condition and residence time of the rivers, hence why the isotope data was grouped into wet and dry periods instead of the actual amount of rainfall. Fig. 1, however; is a strong evidence to show that the amount of rainfall had minimal effect on the isotope values in this study; supporting our contention on the insignificant direct contribution of rainfall amount to the overall nitrate dynamic in the catchment. Fig. 1 has now been included in the supplementary information in the revised manuscript.

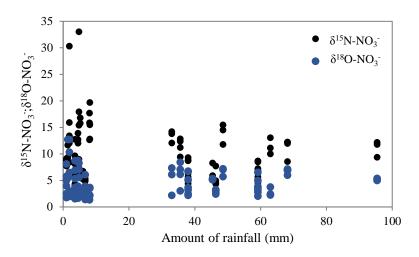


Figure 1: Relationship between (a) δ^{15} N-NO₃⁻; (b) δ^{18} O-NO₃⁻ of the streams and the total amount of rainfall for 10 days before the sampling event.

(2) Could the data not have been explored more thoroughly, e.g. other statistical methods than linear regression in order to identify multiple sources? What about isotope mixing and emission modeling for source identification?

Apportioning the contribution of multiple sources of nitrate using the suggested models requires well-defined isotope values of the end members, accurate fractionation factors of the possible processes (i.e. denitrification and mineralisation) as well as the loads and rates of each possible source and process particularly for an emission model. Unfortunately, determining the fractionation factors of the processes was beyond the scope of this study. These models were not suitable in this study because there was significant overlapping of the end member nitrate isotope values as well as the lack of information on the rates of different types of fertiliser applications. Hence, only a qualitative assessment of the sources was presented in this study.

(3) Significant correlations with very low r2 for isotopic composition and % agriculture (Fig 4) are used as argument for "dominance of anthropogenic nitrogen inputs within the catchment". The discussion should include a more detailed comparison to studies which found a similar but much stronger correlation between d15N of nitrate and land use. We have included more detailed comparisons to studies with similar findings in the revised manuscript.

Page 8 Line 11: Other researchers have also documented similar trends of enriched δ^{15} N-NO₃⁻ with increasing percentage agriculture. For example Harrington et al. 1998, Mayer et al. 2002 and Voss et al. 2006 observed highly significant positive relationships between percentage agriculture land area and δ^{15} N-NO₃⁻ with $r^2 \sim 0.7$. However, these studies showed comparatively narrower and more depleted ranges of δ^{15} N-NO₃⁻ with 2.0 to 7.3‰; 4 to 8‰ and -0.1 to 8.3‰; respectively, suggesting more subtle changes in δ^{15} N-NO₃⁻ over a large span of agriculture land areas in these studies compared to our study.

(4) Is there any information to take away from individual samplings within the same stream? There is no information on river flow rates, for example. Could patterns of isotope data within the streams be explained by mixing of sources or in-stream processing?

Individual samplings within the same stream did not provide sufficient data points for the interpretation on the processes governing the isotope values of the residual nitrate in the streams. For example, at Watson creek only three samples were obtained during each sampling trip. We are not confident to deduce any findings based on that even though the isotope biplot or the keeling plots for some of the sampling events showed significant correlations.

SPECIFIC COMMENTS

Page 2 line 11: Kendall 2007; "et al." is missing Reference has been corrected.

Page 2 line 21-24: please explain in more detail in which way rainfall patterns are different in the southern hemisphere compared to the northern hemisphere.

More detailed explanation has been added to the revised manuscript.

<u>Page 2 Line 30</u>: The southern hemisphere tends to have more sporadic and variable rainfall patterns compared to the northern hemisphere and Australia is an example of this.

Page 2 line 28: please delete space in "samplin g". Format has been corrected.

Page 3 line 21-23: what are the criteria to give the amount of rainfall for 5 days (5-10 days) before sampling of wet periods (dry periods)? Following up, what is the residence time of water in the aquifer and in the river?

There were no specific criteria used to classify the wet and dry periods in this study other than the amount of rainfall prior to the sampling dates. The samplings for wet periods were carried out after a few days of continuous rain. The number of days (5 for wet and 5-10 for dry) were solely to give the readers an idea on the duration of the rain. The same explanation applies for the dry period - the area had received no rain for 5 to 10 days. We also cross-checked the rainfall amount with the discharge of a few streams which were gauged in the area. The discharge of the streams was doubled during the wet periods compared to the dry periods. This information; however, was not discussed in the manuscript as we do not have the complete stream discharge dataset for all the studied streams. We also did not have the information for the residence time of water in both river and aquifer; hence rainfall was used as the qualitative indicator of the residence time of the river.

Page 4 line 10: a figure with some additional water quality parameters would be nice to include as a supplementary material.

This is a good suggestion but all the water quality parameters were relatively consistent throughout the sampling sites and there were no interesting trends or patterns across different sampling sites/periods. All the important data has been presented in the results section.

Page 4 line 16-18: how many samples of fertilizer and cow manure were analysed? Please specify.

A total of 4 fertiliser and 5 cow manure samples were analysed. This has now been specified in the revised manuscript.

<u>Page 4 Line 28</u>: In addition to stream water and sediment, we also collected four samples of artificial/inorganic fertiliser (from the fertiliser distributor in the area) and five cow manure samples from local farmers.

Page 6 line 14/15: what about atmospheric deposition? It is only mentioned on page 7 line 1. Couldn't mixing lead to a depletion of the d18O, with NO3- from atmospheric deposition still contributing partly to the signal?

Contribution from atmospheric deposition although possible was not significant and this has been explained in the manuscript (Page Line). Mixing with atmospheric- NO_3^- could potentially change the $\delta^{18}O$ of the residual NO_3^- but it would only get more enriched rather than more depleted because $\delta^{18}O$ of atmospheric nitrate has been reported to be >60% in the literature.

Page 6 line 27: delete the d of "comprised".

This has been corrected.

Page 7 line 8-10: there is no statistic evidence given by the authors to show that there is an actual trend, so this should be rephrased.

These lines have been rephrased.

Page 8 Line 2: Agricultural land use (i.e. market gardens and cattle rearing) appeared to influence NO_3^- concentrations in our study sites. As shown in Fig. 4(a), during the wet periods, high NO_3^- concentrations (> 40 μM) were particularly observed at sites with more than 70% agricultural land use. During the dry periods, although NO_3^- concentrations were generally lower than 36μM, the outliers were observed at sites with more than 70% agricultural land use.

Page 7 line 10-12: for the dry periods there are at least 6 data points with NO3- conc. $> 50 \mu M$, so "consistently lower" (than 36 μM) is not correct.

The word 'consistently' has been replaced by 'generally'.

Page 7 line 13: replace "entered" by "entering". This has been corrected.

Page 7 line 21/22: please state clearly, that although significant, correlation coefficients r2 are 0.2 and 0.39, respectively, so quite low. From there on it is obvious that the relationship between d15N and % agriculture is not evident at all from this study. This has to be expressed more clearly.

We agree that the correlation between $\delta^{15}N\text{-NO}_3^-$ and percentage agriculture during the dry periods was low and could be marginally significant. However, we are convinced that there was a significant and strong relationship between the two variables during the wet periods. The low r^2 in Fig. 4 was due to 4 data points at Toomuc Creek which have skewed the relationship (as indicated in Fig. 2 in this document). The r^2 value increased to 0.58 (p value <0.01) when the 4 data points were excluded. We have rephrased all the related texts in the manuscript to reflect on this.

Page 8 Line 5: Similarly, enriched δ^{15} N-NO₃⁻ in the streams were mainly found at sites with high percentage agricultural land use (between 75 to 85%) for both dry and wet periods suggesting that enriched δ^{15} N-NO₃⁻ in the stream were originated from agricultural activities. In fact, the most enriched δ^{15} N-NO₃⁻ values (>30‰) were observed at the most downstream site of Watson Creek which has the largest percentage of market gardens (although the total agricultural area is not the highest amongst all the studied sites). We also observed a significant positive relationship between δ^{15} N-NO₃⁻ and percentage agriculture during the wet periods (Fig. 4b). This further supports the contention that agricultural activities were the main control of the δ^{15} N-NO₃⁻ in the streams.

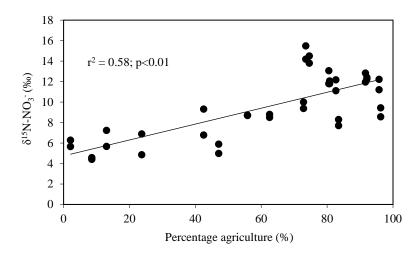


Figure 2: Relationship between δ^{15} N-NO₃ and percentage agriculture during the wet periods

Page 7 line 24-26: the comparison to the other studies has to be made more in detail. For example, Voss et al (2006) observed a significant correlation for 11 streams and weighted monthly means of d15N and % agriculture. If comparable to this data, it should be Figure 7 from this study (if it is correct that it represents average values per stream). And for this representation, there is no significant correlation.

The δ^{15} N-NO₃⁻ values in Figure 7 are not average values but they are the y-intercepts from the Keeling plot for individual stream (Figure 6). These values represent the predicted δ^{15} N-NO₃⁻ of the initial end member. As such, Figure 7 in this study is not comparable to the figure in the study by Voss et al. (2006). This has been made clearer in the figure caption.

Figure 7: Relationship between δ^{15} N-NO₃ of the dominant initial source (indicated by the y-intercept of the Keeling plots; Figure 6) and percentage agriculture during wet periods. Data for Bass-dry period was also presented because only the Keeling plot for Bass-dry period indicates mixing between different sources. The shaded area represents the δ^{15} N-TN of the potential end members.

Page 8 line 18-20 (and following sentences): Give first all the evidence that allows you to conclude that in-stream processing was not the dominant process for regulating the isotopic composition. These arguments could be supported by a more detailed discussion of the relevant literature.

This paragraph has been restructured as suggested by the reviewer.

Page 9 Line 12: In-stream processing of NO_3^- was not evident during the wet periods based on the lack of relationships between $\delta^{18}O\text{-}NO_3^-$ and $[NO_3^-]$ as well as between $\delta^{18}O\text{-}NO_3^-$ and $\delta^{15}N\text{-}NO_3^-$ for the individual streams (shown in Supplementary Fig. 1). If denitrification was dominant, both $\delta^{15}N\text{-}NO_3^-$ and $\delta^{18}O\text{-}NO_3^-$ values are expected to increase at low NO_3^- concentration and there would be systematic increase of both N and O isotopes of NO_3^- (Fry 2008). In addition, high DO in the water column ruled out the possibility of pelagic denitrification.

Careful examination of the Keeling plots for individual streams (Fig. 6) revealed that during the wet periods, NO_3^- concentrations were significantly and linearly correlated with $1/[NO_3^-]$ in all the streams. These relationships strongly suggest mixing between two sources (with distinctive isotopic signatures) as the dominant process regulating the isotopic composition of the residual NO_3^- in the streams during the wet periods.

Page 8 line 20-22: Put figures of d18O vs [NO3-] and d18O vs d15N for individual streams in supplementary material to support your argument.

The relationships between $\delta^{18}O$ vs $[NO_3^-]$ and $\delta^{18}O$ vs. $\delta^{15}N$ have been added as Figure S1 in supplementary material.

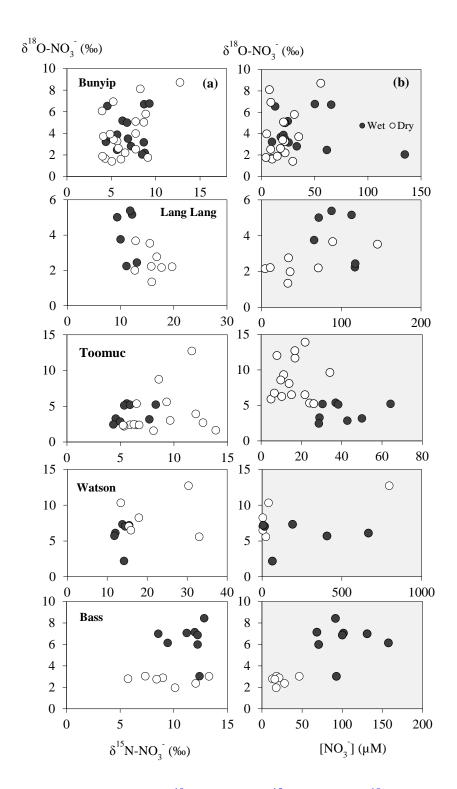


Figure S1: Relationships between (a) $\delta^{18}\text{O-NO}_3^-$ and $\delta^{15}\text{N-NO}_3^-$; (b) $\delta^{18}\text{O-NO}_3^-$ and NO_3^- concentration

Page 9 line 23: add "be" in between "subsequently" and "nitrified". This has been corrected.

Page 10 line 19-21: as stated above, according to figure 4 there is no significant correlation for [NO3-] and percentage agriculture and r2 for the corr. between d15N and percentage agriculture are low, so please rephrase this conclusion. Similarly please rephrase the related sentence in the abstract (page 1 line 17/18).

This has been explained in the earlier comment.

Figures

Fig. 1: indicate percentage agriculture for each sampling site.

Percentage agriculture for each of the sampling site has been added to Fig. 1 as suggested by the reviewer

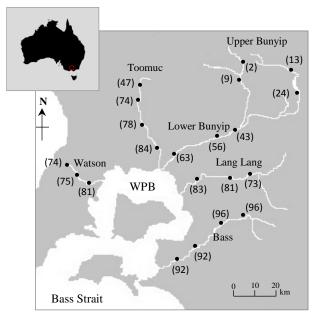


Figure 1: Map of Western Port Bay (WPB) in southern Victoria, Australia and major rivers discharging into WPB. Closed circles represent sampling sites where surface water samples were obtained. Values in parentheses represent the % agriculture area.

Fig. 2: For the Watsons river the "distance from WPB" does not correspond to the values from Fig. 3 (max= 30 km).

The "distance from WPB" has been corrected.

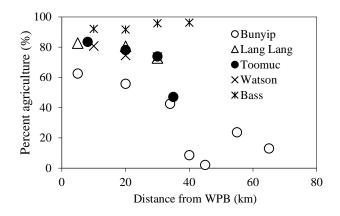


Fig. 4: use A and B for the two panels. In the lower panel indicate which trend curve corresponds to which dataset.

The caption of the figure has been updated according to reviewer's comment.

Figure 4: Relationship between (a) NO_3^- concentration; (b) $\delta^{15}N-NO_3^-$ and the percentage of agricultural land use. In (b) solid line represents the relationship between the variables during dry periods; dotted line represents wet periods.

Fig. 7: Are these average values per site? If so, please indicate the SD. For Bass river (dry period) the value is somewhat high compared to Fig. 6. Please explain.

These are not average values per site but the y-intercept value from the Keeling plots for

individual stream. This has been made clearer in the caption of the figure.

Figure 7: Relationship between δ^{15} N-NO₃ of the dominant initial source (indicated by the y-intercept of the Keeling plots; Figure 6) and percentage agriculture during wet periods. Data for Bass-dry period was also presented because only the Keeling plot for Bass-dry period indicates mixing between different sources. The shaded area represents the δ^{15} N-TN of the potential end members.

Reviewer #3

GENERAL COMMENTS

In the manuscript Stable isotopes of nitrate reveal different nitrogen processing mechanisms in streams across a land use gradient during wet and dry periods, Wong et al present natural abundance nitrate isotopes from five streams across a land use gradient during wet and dry seasons, allowing them to elucidate the controls on sources and transformations of nitrate. This is an interesting dataset and the authors have been resourceful and knowledgeable in their presentation and interpretation of the data. However, prior to publication the manuscript would benefit from a clear and concise definition of terms, and a clearer explanation of the isotope effects and there subsequent implications, as currently it seems hard to follow in places for the none expert reader. An important aspect of the interpretation of this dataset is that there is a tight coupling of mineralisation and nitrification, resulting in no isotope effect being expressed and hence the 15N of organic matter / ammonium and nitrate are similar. Currently this is not fully explained until Page 9 Line 24, making it difficult to understand the authors interpretation of the data prior to this, explaining this earlier on in the discussion will enable the reader to follow your thoughts / interpretation. A good example of this is Page 6 Line 15/16, break this thought down and explain to the reader here the tight coupling between mineralization and nitrification and hence no isotope effect being observed.

We have added a few sentences to explain the minimal isotope effect of the combined reaction of mineralisation and nitrification as suggested by the reviewer.

Page 6 Line 31: Nitrogen isotope of the NO_3^- produced from these end members usually retains the signature of the $\delta^{15}N$ -TN as a result of tight coupling between mineralisation (production of ammonium from organic matter) and nitrification (oxidation of ammonium to NO_3^-) as well as the minimal isotopic fractionation of both processes. It is well documented in the literature that in soil environment, mineralisation causes a small isotopic fractionation ($\pm 1\%$; Kendall et al. 2007) to the produced NH_4^+ . In agricultural areas where NH_4^+ is rapidly consumed or assimilated by crops, nitrification rate is usually low and would also exert a small isotopic fractionation to the produced NO_3^- .

SPECIFIC COMMENTS

Page 2 Line 10: it would be valuable here to state that you are talking about kinetic isotope effects and not equilibrium.

This has been made clearer in the revised manuscript

<u>Page 2 Line 8</u>: Preferential utilisation of lighter isotopes (¹⁴N and ¹⁶O) over heavier isotopes (¹⁵N and ¹⁸O) leads to distinctive isotopic signatures that differentiate the various NO₃⁻ sources/end members (e.g. inorganic and organic fertiliser, animal manure, atmospheric deposition) and the predictable kinetic fractionation effect when NO3- undergoes different biological processes (e.g. nitrogen fixation and denitrification).

Page 2 Line 23 to 25: for the none expert, please explain why rainfall patterns are different in the southern hemisphere and its subsequent effects.

This has been explained in the revised manuscript.

<u>Page 2 Line 30</u>: The southern hemisphere tends to have more sporadic and variable rainfall patterns compared to the northern hemisphere and Australia is an example of this. The variable rainfall patterns can modulate different efficiencies of denitrification in soils and thus different fractionation effects to the residual NO₃⁻ pool.

Page 2 Line 32 to 33: start preparing your reader here, why are denitrification and assimilation more prevalent in wet periods. Study area: throughout this section you refer to the gradient in land use across the catchment, a map of this would be a great addition to the manuscript (or could maybe be added to Figure 1).

Following sentences have been added to the revised manuscript to explain why denitrification can be more prevalent in wet periods. A land use map have been added as supplementary material (Figure S2).

<u>Page 3 Line 7</u>: In some studies (e.g. Riha et al. 2014; Kaushal et al. 2011), denitrification and assimilation by plants and algae have been reported to be more prominent during the dry seasons compared to the wet seasons but in other studies (e.g. Murdiyarso et al. 2010; Enanga et al. 2016) denitrification appeared to be more prevalent during the wet seasons as precipitation induces saturation of soils resulting in oxygen depletion and thereby low redox potentials that favour denitrification.

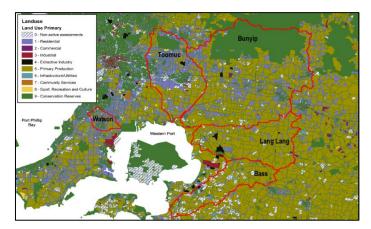


Figure S2: Land use map of Western Port catchment

Page 3 Line 32: how do the authors think using an integrated signal could of biased their interpretation of the results?

We think the integrated signal could potentially bias the interpretation of the results; however, the integrated signal was the best representation of the percentage agriculture area in the catchment.

Page 5 Lines 9 to 10: the term total nitrogen needs to be defined here, as it is important for the mineralization discussion later on. Would particulate organic nitrogen not be a more suitable term? Also, please add in that the values are relative to AIR and the precision of the measurements.

Particulate organic matter is a more suitable term however this was not specifically measured in our study. We used $\delta^{15}N$ of total nitrogen of the soil to directly represent the soil organic portion as most of the nitrogen in soils is generally bound in organic forms. This has been explained more thoroughly in the revised manuscript.

Page 6 Line 30: The average δ^{15} N-TN value of soils is used to directly represent the soil organic portion as most of the nitrogen in soils is generally bound in organic forms.

Page 5 Lines 24: Please add in nitrite concentrations, to confirm for the reader that the values are less than 1% of the nitrate (as stated in the methods).

Nitrite concentrations have been added to the results section of the revised manuscript.

Page 6 Line 6: Nitrite concentrations ranged between 0.1µmol/L and 0.4µmol/L.

Page 6 Line 3: Be clear that you are talking about 15N values here.

This sentence has been corrected to reflect more clearly on the $\delta^{15}N$ values.

<u>Page 6 Line 18</u>: Overall, δ^{15} N of the riverine NO₃ spanned a wide range (+4 to +33%).

Page 6 Line 4: The enriched 15N-nitrate values seem to be constrained to a thin band between 70 to 85% agriculture, but then drop away again at higher percentage agriculture, do the authors have any hypotheses for this?

We hypothesised that the drop off of δ^{15} N-NO₃ at > 85% agriculture was due to recent and possibly over-fertilisation of NH₄+ fertiliser resulting in active nitrification. As a large amount of NH₄+ was available, oxidation of NH₄+ to NO₃- became the rate-determining step resulted in large fractionation (-38% to -14%; Casciotti et al. 2003) and depleted δ^{15} N-NO₃- in the residual NO₃- pool. Unfortunately we could not test this hypothesis as we did not have the information on the rates of fertiliser application and nitrification. This would be a good avenue for future research.

Page 6 Line 5: Surely the same is true for the Bass.

We agree with the reviewer and Bass has been included as exhibiting the same effect as Bunyip in the revised manuscript.

<u>Page 6 Line 20</u>: Among all sites, δ^{15} N-NO₃ values in the Bunyip and Bass were relatively depleted (+4 to +12% for Bunyip and +10 to 12% for Bass), with the lower range found at upper Bunyip (+4 to +8%).

Page 6 Line 16 to 28 and Equation 1: It would be valuable to explain to the readers the value of using both N and O isotopes i.e. N is recycled between fixed N pools and the O atoms are removed and then replaced by nitrification and thereby sensitive to internal processing (this

could come here or in the introduction). The authors need to discuss the more recent literature when introducing and determining the oxygen isotope signal imparted by nitrification, the work of Carly Buchwald is particularly pertinent here.

We have added a few sentences in the introduction to discuss the value of using both N and O isotopes. We have also discussed the calculation used to estimate $\delta^{18}\text{O-NO}_3^-$ imparted by nitrification in more detail. All the texts and figures in the manuscript have been revised to reflect on these changes.

Page 2 Line 7: To date, the most promising tool to investigate the sources and sinks of NO_3^- are the dual isotopic compositions of NO_3^- at natural abundance level (expressed as $\delta^{15}N-NO_3^-$ and $\delta^{18}O-NO_3^-$ in ‰). Preferential utilisation of lighter isotopes (^{14}N and ^{16}O) over heavier isotopes (^{15}N and ^{18}O) leads to distinctive isotopic signatures that differentiate the various NO_3^- sources/end members (e.g. inorganic and organic fertiliser, animal manure, atmospheric deposition) and the predictable kinetic fractionation effect when NO_3^- undergoes different biological processes (e.g. nitrogen fixation and denitrification). For instance, denitrification and phytoplankton assimilation fractionate N and O isotopes in a 1:1 pattern. Simultaneous measurement of $\delta^{15}N-NO_3^-$ and $\delta^{18}O-NO_3^-$ also provides complementary information on the cycling of NO_3^- in the environment. $\delta^{18}O-NO_3^-$ is a more effective proxy of internal cycling of NO_3^- (i.e. assimilation, mineralisation and nitrification) compared to $\delta^{15}N-NO_3^-$. This is because during NO_3^- assimilation and mineralisation, N atoms are recycled between fixed N pools and the O atoms are removed and replaced by nitrification (Sigman et al. 2009; Buchwald et al. 2012).

Page 7 Line 6: The δ^{18} O of NO₃ generated by nitrification of these sources is decoupled from δ^{15} N-NO₃ but relies on the oxygen isotope of water (δ^{18} O-H₂O), oxygen isotope of dissolved oxygen (δ^{18} O-O₂) as well as the kinetic and equilibrium isotope effects during the sequential oxidation of NH₄⁺ to NO₂⁻ then NO₃⁻ (Casciotti et al. 2010; Buchwald et al. 2012). Previous culture studies (Casciotti et al. 2010; Buchwald and Casciotti 2010; Buchwald et al. 2012) and observations in various marine systems (Sigman et al. 2009; Granger et al. 2013; Rafter et al. 2013) have found that δ^{18} O values for nitrified NO₃ were within a few ‰ of the δ^{18} O-H₂O. Hence, -5.3%; the average value of δ^{18} O-H₂O is adopted to represent the lower estimate of δ^{18} O of the nitrified NO₃⁻ in this study. In a system where equilibrium exchange of oxygen between H₂O and NO₂⁻ is negligible but respiration and denitrification are prevalent/cooccurring, $\delta^{18}\text{O-NO}_3^-$ can be much greater than that of $\delta^{18}\text{O-H}_2\text{O}$. In this study, the $\delta^{18}\text{O-NO}_3^$ values were all more enriched than -5.3% suggesting the co-occurrence of a fractionating process, most likely denitrification (this is discussed in the following section). Based on this reason, using -5.3% can potentially underestimate the δ^{18} O of the nitrified NO₃⁻. The conventional 2:1 (δ^{18} O-H₂O: δ^{18} O-O₂) fractional source contribution model (Equation 1) is therefore used to calculate the maximum estimate of δ^{18} O of the nitrified NO₃₋ in our study which is +4.3% by using -5.3% for the average δ^{18} O-H₂O and +23.5% for δ^{18} O-O₂.

Page 6 Line 29: Is this value for cow manure similar to the literature to date? Yes these values are similar to the literature to date.

Page 7 Line 14: 'terrestrial' what are the authors referring to here, fertilizer, manure, leaf litter, please be consistent with the use of terms throughout.

This term refers to a combination of sources and this has been made clearer in the revised manuscript.

<u>Page 8 Line 22</u>:in stream NO₃⁻ comprised mainly of terrestrially derived NO₃⁻ (i.e. inorganic fertiliser, manure and soil organic matter) entered the streams through surface runoff...

Page 7 Lines 15 onwards: a slight restructure here would be beneficial, you are presenting your conclusions before the evidence, discussing your isotope data first in this section would make it easier to follow.

This section has been restructured as suggested by the reviewer.

Page 8 Line 2: Agricultural land use (i.e. market gardens and cattle rearing) appeared to influence NO₃⁻ concentrations in our study sites. As shown in Fig. 4(a), during the wet periods, high NO₃⁻ concentrations (> 40 μM) were particularly observed at sites with more than 70% agricultural land use. During the dry periods, although NO₃- concentrations were generally lower than 36µM, the outliers were observed at sites with more than 70% agricultural land use. Similarly, enriched δ^{15} N-NO₃ in the streams were mainly found at sites with high percentage agricultural land use (between 75 to 85%) for both dry and wet periods suggesting that enriched δ^{15} N-NO₃ in the stream were originated from agricultural activities. In fact, the most enriched δ¹⁵N-NO₃ values (>30‰) were observed at the most downstream site of Watson Creek which has the largest percentage of market gardens (although the total agricultural area is not the highest amongst all the studied sites). We also observed a significant positive relationship between δ¹⁵N-NO₃ and percentage agriculture during the wet periods (Fig. 4b) which further supports the contention that agricultural activities were the main control of the δ^{15} N-NO₃⁻ in the streams. Other researchers have also documented similar trends of enriched δ^{15} N-NO₃⁻ with increasing percentage agriculture. For example Harrington et al. 1998, Mayer et al. 2002 and Voss et al. 2006 observed highly significant positive relationships between percentage agriculture land area and δ^{15} N-NO₃ with $r^2 \sim 0.7$. However, these studies showed comparatively narrower and more depleted ranges of δ^{15} N-NO₃ with 2.0 to 7.3%; 4 to 8% and -0.1 to 8.3%; respectively, suggesting more subtle changes in δ^{15} N-NO₃ over a large span of agriculture land areas in these studies compared to our study.

Given that none of the predicted sources of NO_3^- in the Western Port catchment exhibited an initial $\delta^{15}N-NO_3^-$ of more than +6%, the isotopically-enriched NO_3^- as well as the variability of NO_3^- concentrations observed in this study were consequences of a series of transformation processes. Hence, we propose the following factors to explain the heavy isotopes and the different NO_3^- concentrations across different periods observed in our study:

- (1) During the wet period when surface runoff was conspicuous and residence time of the water column was low, in-stream NO_3^- comprised mainly of terrestrially derived NO_3^- (i.e. fertilisers, manure and soil organic matter) and there was limited in-stream processing of these NO_3^- . The high NO_3^- concentrations and the heavy $\delta^{15}N-NO_3^-$ values reflect the occurrence of mineralisation, nitrification and subsequent preferential denitrification of the isotopically lighter NO_3^- source/s in either the waterlogged soil or in the soil zone underneath the market gardens before transport to the streams through surface runoff.
- (2) During the dry periods when surface runoff was negligible and residence time of the water column was high, there was minimal introduction of terrestrial NO₃⁻ into the streams and in-stream processing of NO₃⁻ was more apparent than during the wet periods. In addition to mineralisation and nitrification, volatilisation and assimilation by plant and algae was highly likely to occur in the stream further reducing the NO₃⁻ concentration and further fractionating the isotopic signature of NO₃⁻.

Page 7 Line 21: I think the authors are referring to Table 2 here. This has been corrected

Page 8 Line 15 and 32: I strongly suggest the authors cite and discuss the implications of the outcomes from the work of Granger and Wankel, 2016 (Isotopic overprinting of nitrification on denitrification as a ubiquitous and unifying feature of environmental nitrogen cycling; PNAS) and how this may influence your interpretation of N turnover in your catchment. The study by Granger and Wankel (2016) has been discussed in the revised manuscript as suggested by the reviewer.

Page 10 Line 5: It is worth noting that although the dual isotopic composition of δ^{18} O- NO_3^- and $\delta^{15}N-NO_3^-$ deviates from a trajectory of 1 (trajectory of 1 indicates denitrification), it is still a salient trend indicating the occurrence of denitrification and is consistent with the δ^{18} O-NO₃⁻:δ¹⁵N-NO₃⁻ recurrently observed in freshwater systems (Kendall et al. 2007). This deviation in our study could be explained by concurrent NO₃- production catalysed by nitrification and/or annamox (Granger and Wankel 2016) although the significance of annamox is still disputable. Based on the multi-process model developed by Granger and Wankel (2016), the two most important factors in the nitrification pathway that govern the δ^{18} O of the newly produced NO₃⁻ are δ^{18} O of the ambient water and the flux of NO₂ oxidation (Granger and Wankel 2016). Deflation of δ^{18} O-NO₃⁻: δ^{15} N:NO₃⁻ trajectory below 1 observed in this study was likely to be associated with the low δ^{18} O-H₂O values which contributed to lower δ^{18} O values for nitrified NO_3^- . Higher NO_3^- reduction rate versus NO_2^- oxidation rate which contributed to the $\delta^{15}N$ enriched pool of nitrified NO_3^- , greater than the denitrified NO_3^- also drives the $\delta^{18}O-NO_3^-$: $\delta^{15}N-$ NO₃ trajectory to values below 1 (see Granger and Wankel 2016 for explanation). All in all, this highlights the significant contribution of nitrification along with denitrification in the WP catchment.

Page 11 Line 5: NO_3^- in group B has variable $\delta^{15}N$ and $\delta^{18}O$ values as shown by Bunyip and Toomuc. This could be attributed to isotopic fractionation either during plant and/or algae uptake or denitrification as substantiated by the parallel increase of $\delta^{18}O\text{-}NO_3^-$ versus $\delta^{15}N\text{-}NO_3^-$ (Fig. 9). Based on Fig. 9, the large uncertainties in the $\delta^{18}O\text{-}NO_3^-$ of the nitrified end members have resulted in overlapping of isotopic signatures of the three major sources (nitrified cow manure, nitrified inorganic fertiliser and nitrified SOM). All three sources appeared to have influenced the $\delta^{15}N$ and $\delta^{18}O$ of the residual NO_3^- in the stream. This scenario reinstates the sensitivity and the importance of accurately determining the $\delta^{18}O\text{-}NO_3^-$ of the initial NO_3^- in the effort to apportion the relative contribution of different sources.

Page 10 section (3): an earlier introduction of the different behaviors of N and O isotopes during internal processing of nitrate will make this section easier to understand for the none expert reader. I would not put denitrification under the heading recycling, if the authors are referring to nitrate reduction, followed by reoxidation please say so.

We have included a paragraph in the introduction to briefly discuss about the different behaviours of nitrate isotopes during internal processing of nitrate. We have also changed the term 'recycling' to 'internal processes'.

<u>Page 2 Line 7</u>: To date, the most promising tool to investigate the sources and sinks of NO_3^- are the dual isotopic compositions of NO_3^- at natural abundance level (expressed as $\delta^{15}N-NO_3^-$ and $\delta^{18}O-NO_3^-$ in ‰). Preferential utilisation of lighter isotopes (^{14}N and ^{16}O) over heavier isotopes (^{15}N and ^{18}O) leads to distinctive isotopic signatures that differentiate the various NO_3^-

sources/end members (e.g. inorganic and organic fertiliser, animal manure, atmospheric deposition) and the predictable kinetic fractionation effect when NO_3^- undergoes different biological processes (e.g. nitrogen fixation and denitrification). For instance, denitrification and phytoplankton assimilation fractionate N and O isotopes in a 1:1 pattern. Simultaneous measurement of $\delta^{15}N$ - NO_3^- and $\delta^{18}O$ - NO_3^- also provides complementary information on the cycling of NO_3^- in the environment. $\delta^{18}O$ - NO_3^- is a more effective proxy of internal cycling of NO_3^- (i.e. assimilation, mineralisation and nitrification) compared to $\delta^{15}N$ - NO_3^- . This is because during NO_3^- assimilation and mineralisation, N atoms are recycled between fixed N pools and the O atoms are removed and replaced by nitrification (Sigman et al. 2009; Buchwald et al. 2012).

Page 10 Line 11: Do the authors know when fertilizer is applied in this catchment, how does this align with your runoff / turnover hypotheses?

Unfortunately we do not have the information on when fertiliser was applied in the catchment hence no further conclusion could be drawn on the relationship of fertiliser application and the runoff processes.

Figure 1: Please mark on the map of Australia where southern Victoria is. The red circle in Figure 1 indicates the location of southern Victoria

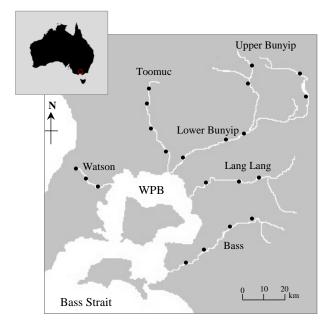


Figure 3: Mark on upper / lower Bunyip.
Figure 3 has been updated in the revised manuscript:

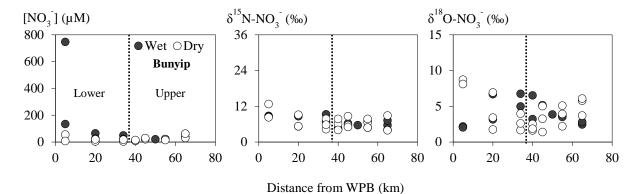


Figure 5: Where have the authors taken these isotope effects from? Please cite the relevant literature in the caption. A positive / inverse isotope effect for nitrification? More details have been provided in the caption of Figure 5:

<u>Figure 5</u>: Conceptual diagram illustrating the sources and processes of NO_3^- during the wet and dry periods in the Western Port catchment. The values of enrichment factor (ϵ) were obtained from the literature (Kendall et al. 2007) to indicate the relative contribution of the transformation processes to the isotopic compositions of the residual NO_3^- .

Figure 7: More details are needed in the figure caption, what do the crosses and dashed line mean? I also assume that it is the y intercept values determined in Figure 6 that have been plotted.

More details have been added to the caption of Figure 7:

Figure 7: Relationship between δ^{15} N-NO₃ of the dominant initial source (indicated by the y-intercept of the Keeling plots; Figure 6) and percentage agriculture during wet periods. Data for Bass-dry period was also presented because only the Keeling plot for Bass-dry period indicates mixing between different sources. The shaded area represents the δ^{15} N-TN of the potential end members.

Figure 9: define what starting values you have used and where they have come from, particularly for the oxygen isotopes.

More details have been added to the caption of Figure 9:

<u>Figure 9</u>: Biplot of δ^{15} N-NO₃- versus δ^{18} O-NO₃⁻ for Bunyip and Toomuc (group B data in Fig. 8b). Shaded areas represent theoretical assimilation trends for cow manure, SOM and inorganic fertiliser. The minimum starting value for δ^{18} O-NO₃⁻ was estimated from the average δ^{18} O-H₂O and the maximum δ^{18} O-NO₃⁻ value was estimated from Equation 1. The starting δ^{15} N-NO₃⁻ is the δ^{15} N-TN value of respective end member. Solid lines represent the assimilation trends for Bunyip (both lower and upper Bunyip) and Toomuc.

Stable isotopes of nitrate reveal different nitrogen processing mechanisms in streams across a land use gradient during wet and dry periods

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Abstract. Understanding the relationship between land use and the dynamics of nitrate (NO₃⁻) is the key to constrain sources of NO₃⁻ export in order to aid effective management of waterways. In this study, isotopic compositions of NO₃⁻ (δ¹⁵N-NO₃⁻ and δ¹⁸O-NO₃⁻) were used to elucidate the effects of land use (agriculture in particular) and rainfall on the major sources and sinks of NO₃⁻ within the Westernport catchment, Victoria, Australia. This study is one of the very few studies carried out in temperate regions with highly stochastic rainfall patterns; enabling a more comprehensive understanding of the applications of NO₃⁻ isotopes in catchment ecosystems with different climatic conditions. Longitudinal samples were collected from five streams with different agriculture land use intensities on five occasions – three during dry periods and two during wet periods. At the catchment scale, we observed significant positive relationships between NO₃⁻ concentrations, δ¹⁵N-NO₃⁻ and percentage agriculture reflecting the dominance of anthropogenic nitrogen inputs within the catchment. Different rainfall conditions appeared to be major controls on the predominance of the sources and transformation processes of NO₃⁻ in our study sites. Artificial fertiliser was the dominant source of NO₃⁻ during the wet periods while nitrified organic matter in sediment and nitrified manure were more apparent sources of NO₃⁻ to the surface water during the dry periods. Denitrification was prevalent during the wet periods while uptake of NO₃⁻ by plants or algae was only observed during the dry periods in two streams. The outcome of this study suggests that effective reduction of NO₃⁻ load to the streams can only be achieved by prioritising management strategies based on different rainfall conditions.

25 1 Introduction

Anthropogenic sources of NO₃⁻ from catchments can pose substantial risk to the quality of freshwater ecosystems (Vitousek et al. 1997; Galloway et al. 2004; Galloway et al. 2005). Over-enrichment of NO₃⁻ in freshwater systems is a major factor in development of algal blooms which often promote bottom water hypoxia and anoxia. Such anoxia intensifies nutrient recycling and can lead to disruption of ecosystem functioning and ultimately loss of biodiversity (Galloway et al. 2004; Carmago and Alonso 2006). Freshwater streams are often sites for enhanced denitrification (Peterson et al. 2001; Barnes and Raymond

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2010). However, when NO₃⁻ loading from the catchment exceeds the removal and retention capacity of the streams, NO₃⁻ is transported to downstream receiving waters including estuaries and coastal embayments, which are often nitrogen-limited, further compounding the problem of eutrophication.

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Understanding the sources, transport and sinks of NO_3^- is critical, particularly in planning and setting guidelines for better management of the waterways (Xue et al. 2009). Establishing the link between land use and the biogeochemistry of NO_3^- provides fundamental information to help develop NO_3^- reduction and watershed restoration strategies (Kaushal et al. 2011). To date, the most promising tool to investigate the sources and sinks of NO_3^- are the dual isotopic compositions of NO_3^- at natural abundance level (expressed as $\delta^{15}N-NO_3^-$ and $\delta^{18}O-NO_3^-$ in ‰). Preferential utilisation of lighter isotopes (^{14}N and ^{16}O) over heavier isotopes (^{15}N and ^{18}O) leads to distinctive isotopic signatures that differentiate the various NO_3^- sources/end members (e.g. inorganic and organic fertiliser, animal manure, atmospheric deposition) and the predictable kinetic fractionation effect when NO_3^- undergoes different biological processes (e.g. nitrogen fixation and denitrification). For instance, denitrification and phytoplankton assimilation fractionate N and O isotopes in a 1:1 pattern. Simultaneous measurement of $\delta^{15}N-NO_3^-$ and $\delta^{18}O-NO_3^-$ also provides complementary information on the cycling of NO_3^- in the environment. $\delta^{18}O-NO_3^-$ is a more effective proxy of internal cycling of NO_3^- (i.e. assimilation, mineralisation and nitrification) compared to $\delta^{15}N-NO_3^-$. This is because during NO_3^- assimilation and mineralisation, N atoms are recycled between fixed N pools and the O atoms are removed and replaced by nitrification (Sigman et al. 2009; Buchwald et al. 2012).

In addition to constraining NO_3^- budget and N cycling in various environmental settings, previous studies have also utilized the dual isotopic signatures of NO_3^- to study the effects of different land uses on the pool of NO_3^- in headwater streams (Barnes and Raymond 2010, Sebilo et al. 2003), creeks (Danielescu and MacQuarrie 2013) and large rivers (Voss et al. 2006; Battaglin et al. 2001). Barnes and Raymond (2010) for example found that both $\delta^{15}N-NO_3^-$ and $\delta^{18}O-NO_3^-$ varied significantly between urban, agricultural and forested areas in the Connecticut River watershed, USA. Several other investigators (Mueller et al. 2016; Mayer et al. 2002) showed positive relationships between $\delta^{15}N-NO_3^-$ and the percent of agricultural land in their study area, indicating the applicability of $\delta^{15}N-NO_3^-$ and $\delta^{18}O-NO_3^-$ to distinguish NO_3^- originating from different land uses. Danielescu and MacQuarrie (2013) and Chang et al. (2002) on the other hand, found no correlations between NO_3^- isotopes and land use intensities in the Trout River catchment and the Mississippi River Basin; respectively. These studies attributed the lack of correlation to catchment size (Danielescu and MacQuarrie, 2013) and the homogeneity of land use (Chang et al. 2002).

Despite the extensive application of NO₃⁻ isotopes to study the transport of terrestrial NO₃⁻ to the tributaries in the catchment; majority of these studies were carried out in the United States and Western Europe where climatic conditions, for example temperature and rainfall patterns are different compared to that in the southern hemisphere. The southern hemisphere tends to have more sporadic and variable rainfall patterns compared to the northern hemisphere and Australia is an example of this. The variable rainfall patterns can modulate different efficiencies of denitrification in soils and thus different fractionation effects to the residual NO₃⁻ pool. However, the lack of NO₃⁻ isotope studies in the southern hemisphere (Ohte et al. 2013) impedes a more thorough understanding of NO₃⁻ dynamics within catchment ecosystems.

Most previous studies investigating the relationship between land use and NO_3^- export using $\delta^{15}N$ - NO_3^- and $\delta^{18}O$ - NO_3^- have either focused on the seasonal or spatial variations in one stream, or used multiple streams with one site per stream (i.e. Mayer et al. 2002; Yevenes et al. 2016). Far fewer studies have incorporated longitudinal sampling of multiple streams over multiple seasons. Nitrate concentrations and concomitant isotopic signatures can change substantially, not only spatially but temporally. Changes in hydrological and physicochemical (notably temperature) conditions of a river can affect the relative contribution of different sources of NO_3^- and the seasonal predominance of a specific source (Kaushal et al. 2011; Panno et al. 2008). In some studies (e.g. Riha et al. 2014; Kaushal et al. 2011), denitrification and assimilation by plants and algae have been reported to be more prominent during the dry seasons compared to the wet seasons but in other studies (e.g. Murdiyarso et al. 2010; Enanga et al. 2016) denitrification appeared to be more prevalent during the wet seasons as precipitation induces saturation of soils resulting in oxygen depletion and thereby low redox potentials that favour denitrification. As such, if spatial and temporal variations of $\delta^{15}N$ - NO_3^- and $\delta^{18}O$ - NO_3^- are not considered thoroughly in a sampling regime, it can lead to misinterpretation of the origin and fate of NO_3^- . Proper consideration of the temporal variability of NO_3^- isotope signatures and transformation are particularly pertinent in catchments with highly stochastic rainfall patterns, such as Australia.

In this study, we examine both spatial and temporal variations of NO_3^- concentrations and isotopic compositions within and between 5 streams in 5 catchments spanning an agricultural land-use gradient, enabling us to evaluate (1) the effects of agriculture land use on the sources and transformation processes of NO_3^- and (2) the effects of rainfall on the predominance of the sources and fate of NO_3^- in the catchments.

2 Materials and methods

2.1 Study area

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This study was undertaken using 5 major streams (Bass River, Lang Lang River, Bunyip River, Watsons Creek and Toomuc Creek) draining into Western Port (Fig. 1) which lies approximately 75km south east of Melbourne, Australia. Western Port is a nitrogen-limited coastal embayment (CSIRO, 1996) recognised as a Ramsar site for migratory birds. The catchments in the Western Port contain three marine national parks, highlighting its environmental and ecological significance. The catchments cover an area of 3,721 km² with land uses ranging from semi-pristine/state forest to high density residential and intense agricultural activities. The area experiences a temperate climate with average annual rainfall ranging from 750mm along the coast to 1200mm in the northern highlands. Mean monthly rainfall was about 20mm and 53mm in 2014 and 2015, respectively (Australian Bureau of Meteorology 2014 - http://www.bom.gov.au/).

The catchment overlies a multi-layered combined aquifer system. The main aquifer consists of Quaternary alluvial and dune deposit (average thickness of <7m) as well as Baxter, Sherwood and Yallock formations (average thickness between 20 and 175m). These aquifers are generally unconfined with radial groundwater flow direction from the basin edge towards Western Port bay. The hydrogeology of Western Port can be found in Carillo-Rivera, 1975.

Five longitudinal surveys were carried out between April 2014 and May 2015, two during wet periods (14/4/2014; 15/5/2015 - the total rainfall for 5 days before sampling was between 45 and 65mm) and three during dry periods (8/4/2014; 22/5/2014; 21/3/2015 - the total rainfall for 5 to 10 days before sampling was <5mm). A total of 21 sampling sites, indicated in Fig. 1 were selected across a gradient of catchment land use intensity. The five streams were selected based on the extent and distribution of land use types between and within each stream sub-catchment, thus enabling comparisons within and between the streams.

In this study, catchment intensive agriculture was used as predictor of land use intensity in the catchment. These data were obtained from the National Environmental Stream Attributes database v1.1 (Stein et al. 2014), Bureau of Rural Sciences' 2005/06 Land Use of Australia V4 maps (www.agriculture.gov.au/abares/aclump) and Victorian Resources Online (VRO). In the context of this study, the catchment intensive agriculture variable is termed as 'percentage agriculture'. This term represents the percentage of the catchment subject to intensive animal production, intensive plant production (horticulture and irrigated cropping) and grazing of modified pastures. This variable also reflects the integrated diffuse sources of nutrients derived from intense agriculture including animal manure and inorganic fertilisers. The percentage agriculture for the sampling sites ranged between 2 to 96% with the Bass River (94±2%) > Lang Lang (79±5%) > Watsons (76±4%) > Toomuc (71±16%) > Bunyip (upper Bunyip: 12±9%; lower Bunyip: 54±10%; Fig. 2). For the purpose of this study, Bunyip is divided into two sectors (upper and lower Bunyip) based on the proximity of the sampling sites (Fig. 1) and the percentage of land use. All the sampling sites in the upper Bunyip are situated in areas with >30% forestation. In general, the percentage agriculture decreases with increased distance from the Western Port Bay (WPB) for all the streams except Bass River. There is an increase of about 2% in percentage agriculture for Bass River with increased distance from WPB. Watsons Creek has the largest percentage of market gardens (~91%).

2.2 Sample collection and preservation

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Water quality parameters (pH, electrical conductivity, turbidity, dissolved oxygen (DO) concentration and water temperature) were measured using a calibrated Horiba U-10 multimeter. Stream samples were collected for the analyses of dissolved inorganic nutrients-DIN (ammonium, NH_4^+ ; NO_3^- and nitrite, NO_2^-), dissolved organic carbon (DOC) and NO_3^- isotopes ($\delta^{15}N-NO_3^-$ and $\delta^{18}O-NO_3^-$). These samples were filtered on site using 0.2μ m Pall Supor® membrane disc filters. Filtered DOC samples were acidified to pH < 2 with concentrated hydrochloric acid. Samples for $\delta^{18}O-H_2O$ were collected directly from the streams without filtering. Sediment samples were collected from the bottom of the rivers and were kept in zip-lock bags. All samples were stored and transported on ice until they were refrigerated (nutrients samples were frozen) in the laboratory. In addition to stream water and sediment, we also collected four samples of artificial/inorganic fertiliser (from the fertiliser distributor in the area) and five cow manure (from local farmers).

2.3 DIN and DOC concentration measurements

All chemical analyses were performed within 1-2 weeks of sample collection except for isotope analyses (within 2 months). The concentrations of NO₃-, NO₂-, and NH₄+ were determined spectrophotometrically using a Lachat QuikChem 8000 Flow Injection Analyzer (FIA) following standard procedures (APHA 2005). DOC concentrations were determined using a Shimadzu TOC-5000 Total Organic Carbon analyser. Analysis of standard reference materials indicated the accuracy of the spectrophotometric analyses and the TOC analyser was always within 2% relative error.

2.4 Isotopic analyses

The samples for δ^{15} N-NO₃⁻ and δ^{18} O-NO₃⁻ were analysed using the chemical azide method based on the procedure outlined in McIlvin et al. (2005). In brief, NO_x (NO₃⁻ + NO₂⁻) was quantitatively converted to NO₂⁻ using cadmium reduction and then to N₂O using sodium azide. The initial NO₂⁻ concentrations were insignificant, typically <1% relative to NO₃⁻. Hence, the influence of δ^{15} N-NO₂⁻ was negligible and the measured δ^{15} N-N₂O represents the signature of δ^{15} N-NO₃⁻. The resultant N₂O was then analysed on a Hydra 20-22 continuous flow isotope ratio mass spectrometer (CF-IRMS; Sercon Ltd., UK) interfaced to a cryoprep system (Sercon Ltd., UK). Nitrogen and oxygen isotope ratios are reported in per mil (‰) relative to atmospheric air (AIR) and Vienna Standard Mean Ocean Water (VSMOW), respectively. The external reproducibility of the isotopic analyses lies within \pm 0.5% for δ^{15} N and \pm 0.3% for δ^{18} O. The international reference materials used were USGS32, USGS 34, USGS 35 and IAEA-NO₃⁻. Lab-internal standards (KNO₃⁻ and NaNO₂⁻) with pre-determined isotopic values were also processed the same way as the samples to check on the efficiency of the analytical method. The δ^{18} O-H₂O values were measured via equilibration with He-CO₂ at 32°C for 24 to 48 hours in a Finnigan MAT Gas Bench and then analysed using CF-IRMS. The δ^{18} O-H₂O values were referenced to internal laboratory standards, which were calibrated using VSMOW and Standard Light Antarctic Precipitation. Measurement of two sets of triplicate samples in every run showed a precision of 0.2‰ for δ^{18} O-H₂O. Sediment samples for the analysis of δ^{15} N of total nitrogen were dried at 60°C before being analysed on the 20-22 CF-IRMS coupled to an elemental analyzer (Sercon Ltd. UK).

2.5 Data Analysis

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The relationships between percentage agriculture and surface water NO₃⁻ concentrations were assessed using linear regression.

25 Percentage agriculture was the predictor variable. NO₃⁻ concentration, and δ¹⁵N-NO₃⁻ were response variables. Relationships between δ¹⁵N-NO₃⁻ and NO₃⁻ concentration as well as δ¹⁸O-NO₃⁻ and δ¹⁵N-NO₃⁻ were assessed using Pearson's correlation. The NO₃⁻ isotopes response variables were assessed at two spatial scales – individual stream and catchment scale. The catchment scale integrates data from all five studied streams. Any graphical patterns or relationships derived from using these scales represent processes that occur somewhere in the catchment either in the streams or prior to entering the streams with data from the individual stream is likely to represent more localised processes to that particular stream.

3 Results

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The streams were oxic throughout the course of our study period with %DO saturation between 70 to 100%. There was no apparent spatial and temporal variation in DO; however, %DO saturation was slightly lower during the dry periods (average of $73\pm20\%$) compared to the wet periods (average of $82\pm12\%$). Temperature was also relatively consistent with an average of $13\pm2^{\circ}$ C. Ammonium concentration was generally low (<4 μ M) except during the wet periods in Bunyip (~7 μ M), Lang Lang (~21 μ M) and Bass (~29 μ M). DOC concentrations were typically 0.8 ± 0.4 mM. Nitrite concentrations were also low in all the streams; ranged between 0.1μ mol/L and 0.4μ mol/L.

The spatial and temporal variations of NO_3^- concentration, $\delta^{15}N$ and $\delta^{18}O$ across the sites are shown in Fig. 3. NO_3^- concentrations ranged between 7 μ M and 790 μ M with averages of 21 ± 15 μ M, 50 ± 130 μ M, 64 ± 43 μ M, 71 ± 43 μ M and 190 ± 280 μ M for Toomuc, Bunyip, Bass, Lang Lang and Watsons, respectively. The lowest NO_3^- concentration was observed in the lower Bunyip (4 μ M) while the highest NO_3^- concentration was observed in Watsons Creek (790 μ M) at the most downstream site. Nitrate concentrations were generally higher during the wet periods compared to the dry periods in all streams (Fig. 3). During the wet periods, NO_3^- concentrations typically followed an increasing trend heading downstream except for the Bass River which exhibited the opposite NO_3^- trend with lower concentrations at downstream sites. During the dry periods, only the Bunyip and Bass Rivers showed apparent longitudinal patterns in NO_3^- concentrations; with decreasing concentrations moving downstream in both. Sites with high percentage agriculture generally also exhibited high NO_3^- concentrations (Fig. 4), particularly during the wet periods.

Overall, $\delta^{15}N$ of the riverine NO_3^- spanned a wide range (+4 to +33‰). Approximately 62% of the obtained $\delta^{15}N$ - NO_3^- values fell below +10‰. More enriched $\delta^{15}N$ - NO_3^- values (> +10‰) were typically observed during the dry periods and were coincident with a high percentage agriculture (Fig. 4). Among all sites, $\delta^{15}N$ - NO_3^- values in the Bunyip and Bass were relatively depleted (+4 to +12‰ for Bunyip and +10 to 12‰ for Bass), with the lower range found at upper Bunyip (+4 to +8‰). There was no discernible pattern spatially or temporally in $\delta^{18}O$ - NO_3^- , except that higher values were found in Lang Lang and Bass during the wet periods with +4 to +6‰ and +5 to +9‰; respectively compared to the dry periods (<+4‰). For other sampling sites, $\delta^{18}O$ - NO_3^- ranged between +2 to +13‰. The isotope compositions of sediment, water, artificial fertiliser and cow manure/organic fertiliser are presented in Table 1. The $\delta^{15}N$ -TN of three potential sources – artificial fertiliser, organic fertiliser and soil organic matter ranged from -0.5 to +0.7‰, +6 to +13‰ and +4 to +5‰, respectively.

4 Discussion

4.1 Potential sources of NO₃-

There are three major potential sources of NO_3^- in the catchments – artificial fertiliser, cow manure/organic fertiliser and soil organic matter (SOM) – see Table 1 for the δ^{15} N-TN values. The average δ^{15} N-TN value of soils is used to directly represent the soil organic portion as most of the nitrogen in soils is generally bound in organic forms. Nitrogen isotope of the NO_3^-

produced from the potential end members usually retains the signature of the δ^{15} N-TN as a result of tight coupling between mineralisation (production of ammonium from organic matter) and nitrification (oxidation of ammonium to NO₃-) as well as the minimal isotopic fractionation of both processes. It is well documented in the literature that in soil environment, mineralisation causes a small isotopic fractionation (±1%; Kendall et al. 2007) to the produced NH₄⁺. In agricultural areas where NH₄⁺ is rapidly consumed or assimilated by crops, nitrification rate is usually low and would also exert a small isotopic fractionation to the produced NO₃. The δ^{18} O of NO₃ generated by nitrification of these sources is decoupled from δ^{15} N-NO₃ but relies on the oxygen isotope of water ($\delta^{18}\text{O-H}_2\text{O}$), oxygen isotope of dissolved oxygen ($\delta^{18}\text{O-O}_2$) as well as the kinetic and equilibrium isotope effects during the sequential oxidation of NH₄⁺ to NO₂⁻ then NO₃⁻ (Casciotti et al. 2010; Buchwald et al. 2012). Previous culture studies (Casciotti et al. 2010; Buchwald and Casciotti 2010; Buchwald et al. 2012) and observations in various marine systems (Sigman et al. 2009; Granger et al. 2013; Rafter et al. 2013) have found that δ^{18} O values for nitrified NO_3 were within a few % of the $\delta^{18}O-H_2O$. Hence, -5.3%; the average value of $\delta^{18}O-H_2O$ is adopted to represent the lower estimate of δ^{18} O of the nitrified NO₃⁻ in this study. In a system where equilibrium exchange of oxygen between H₂O and NO₂⁻ is negligible but respiration and denitrification are prevalent/co-occurring, $\delta^{18}\text{O-NO}_3^-$ can be much greater than that of $\delta^{18}\text{O-NO}_3^ H_2O$. In this study, the $\delta^{18}O$ - NO_3 values were all more enriched than -5.3% suggesting the co-occurrence of a fractionating process, most likely denitrification (this is discussed in the following section). Based on this reason, using -5.3% can potentially underestimate the δ^{18} O of the nitrified NO₃. The conventional 2:1 (δ^{18} O-H₂O; δ^{18} O-O₂) fractional source contribution model (Equation 1) is therefore used to calculate the maximum estimate of δ^{18} O of the nitrified NO₃- in our study which is +4.3% by using -5.3% for the average δ^{18} O-H₂O and +23.5% for δ^{18} O-O₂.

$$\delta^{18}\text{O-NO}_3^- = \frac{2}{3} \delta^{18}\text{O-H}_2\text{O} + \frac{1}{3} \delta^{18}\text{O-O}_2$$
 (1)

As such, we considered the δ^{18} O of nitrified NO₃⁻ to range from -5.3 to +4.3‰.

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The δ^{15} N-TN of cow manure (+6 to +13‰) was most variable compared to other end members. This variation reflects the extent of volatilisation, a highly fractionating process. Volatilisation can cause a fractionation effect of up to 25‰ in the residual NH₄⁺ (Hubner 1986). As such, the lower value of +6‰ indicates a relatively fresh manure sample and is assumed to represent the initial δ^{15} N of the cow manure before undergoing any extensive fractionation.

Atmospheric deposition did not appear to be an important source of NO_3^- in this study based on the relatively depleted $\delta^{18}O\text{-}NO_3^-$ values (ranged from +2 to +8% during the wet periods; +1.5 to +13% during the dry periods) of the riverine samples. The $\delta^{18}O\text{-}NO_3^-$ of atmospheric deposition were reported to range from +60 to +95% in the literature (Kendall 2007; Elliott et al. 2007; Pardo et al. 2004). Similarly, groundwater was not considered as an important source of NO_3^- to the streams based on the low NO_3^- concentrations (~0.7 to 7.0µM) reported in previous studies (Water Information System Online; http://data.water.vic.gov.au/monitoring.htm).

4.2 General characteristics of NO₃ in the streams

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Agricultural land use (i.e. market gardens and cattle rearing) appeared to influence NO_3^- concentrations in our study sites. As shown in Fig. 4(a), during the wet periods, high NO_3^- concentrations (> 40 μM) were particularly observed at sites with more than 70% agricultural land use. During the dry periods, although NO_3^- concentrations were generally lower than 36μM, the outliers were observed at sites with more than 70% agricultural land use. Similarly, enriched $\delta^{15}N-NO_3^-$ in the streams were mainly found at sites with high percentage agricultural land use (between 75 to 85%) for both dry and wet periods suggesting that enriched $\delta^{15}N-NO_3^-$ in the stream were originated from agricultural activities. In fact, the most enriched $\delta^{15}N-NO_3^-$ values (>30%) were observed at the most downstream site of Watson Creek which has the largest percentage of market gardens (although the total agricultural area is not the highest amongst all the studied sites). We also observed a significant positive relationship between $\delta^{15}N-NO_3^-$ and percentage agriculture during the wet periods (Fig. 4b). This further supports the contention that agricultural activities were the main control of the $\delta^{15}N-NO_3^-$ in the streams. Other researchers (e.g. Mayer et al. 2002 and Voss et al. 2006) have also documented similar trends of enriched $\delta^{15}N-NO_3^-$ with increasing percentage agriculture. For example Harrington et al. 1998, Mayer et al. 2002 and Voss et al. 2006 observed highly significant positive relationships between percentage agriculture land area and $\delta^{15}N-NO_3^-$ with $r^2 \sim 0.7$. However, these studies showed comparatively narrower and more depleted ranges of $\delta^{15}N-NO_3^-$ with $r^2 \sim 0.7$. However, these studies showed comparatively narrower and more depleted ranges of $\delta^{15}N-NO_3^-$ with $r^2 \sim 0.7$. However, these studies compared to our study.

Given that none of the predicted sources of NO_3^- in the Western Port catchment exhibited an initial $\delta^{15}N-NO_3^-$ of more than +6‰, the isotopically-enriched NO_3^- as well as the variability of NO_3^- concentrations observed in this study were consequences of a series of transformation processes. Hence, we propose the following factors to explain the heavy isotopes and the different NO_3^- concentrations across different periods observed in our study:

- (1) During the wet period when surface runoff was conspicuous and residence time of the water column was low, instream NO_3^- comprised mainly of terrestrially derived NO_3^- (i.e. fertilisers, manure and soil organic matter) and there was limited in-stream processing of these NO_3^- . The high NO_3^- concentrations and the heavy $\delta^{15}N-NO_3^-$ values reflect the occurrence of mineralisation, nitrification and subsequent preferential denitrification of the isotopically lighter NO_3^- source/s in either the waterlogged soil or in the soil zone underneath the market gardens before transport to the streams through surface runoff.
- (2) During the dry periods when surface runoff was negligible and residence time of the water column was high, there was minimal introduction of terrestrial NO_3^- into the streams and in-stream processing of NO_3^- was more apparent than during the wet periods. In addition to mineralisation and nitrification, volatilisation and assimilation by plant and algae was highly likely to occur in the stream further reducing the NO_3^- concentration and further fractionating the isotopic signature of NO_3^- .

These processes are conceptualised in Fig. 5 and are corroborated in the following discussion using two graphical methods: the Keeling plot and the isotope biplot. In an agricultural watershed, the co-existence of multiple sources and transformation processes can potentially complicate the use of NO_3^- isotopes as tracers of its origin. Keeling plots ($\delta^{15}N-NO_3^-$ versus $1/[NO_3^-]$) are generally very useful to distinguish between mixing and fractionation (i.e. assimilation and bacterial denitrification) processes (Kendall et al. 1998). The latter typically results in progressively increasing $\delta^{15}N-NO_3^-$ values as NO_3^- concentrations decrease and yields a curved Keeling plot. Meanwhile, mixing of NO_3^- from two or more sources can result in concomitant increase of both $\delta^{15}N-NO_3^-$ and NO_3^- concentrations and results in a straight line on the Keeling plot (Kendall et al. 1998). A biplot ($\delta^{18}O-NO_3^-$ versus $\delta^{15}N-NO_3^-$) on the other hand, is an effective method to differentiate between assimilation and denitrification. Nitrate assimilation creates a 1:1 slope on a biplot while the simultaneous increase of $\delta^{18}O-NO_3^-$ and $\delta^{15}N-NO_3^-$ in a 2:1 pattern indicates the presence of denitrification (Fry 2006).

4.3 Key controlling processes of nitrate during the wet periods

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In-stream processing of NO_3^- was not evident during the wet periods based on the lack of relationships between $\delta^{18}O\text{-NO}_3^-$ and $[NO_3^-]$ as well as between $\delta^{18}O\text{-NO}_3^-$ and $\delta^{15}N\text{-NO}_3^-$ for the individual streams (shown in Supplementary Fig. 1). If denitrification was dominant, both $\delta^{15}N\text{-NO}_3^-$ and $\delta^{18}O\text{-NO}_3^-$ values are expected to increase at low NO_3^- concentration and there would be systematic increase of both N and O isotopes of NO_3^- (Fry 2008). In addition, high DO in the water column ruled out the possibility of pelagic denitrification.

Careful examination of the Keeling plots for individual streams (Fig. 6) revealed that during the wet periods, NO₃concentrations were significantly and linearly correlated with $1/[NO_3]$ in all the streams. These relationships strongly suggest mixing between two sources (with distinctive isotopic signatures) as the dominant process regulating the isotopic composition of the residual NO₃ in the streams during the wet periods. The different trends in the Keeling plots (Fig. 6) for individual streams indicate that the isotopic signature of the dominant NO₃ source varied temporally and spatially across the catchments. Negative trends on the Keeling plots for Bunyip, Lang Lang and Toomuc (Fig. 6) clearly show that the dominant NO₃ source was isotopically enriched (above +10% for Bunyip and Toomuc and +14% for Lang Lang) while the positive trends on the Keeling plots for Bass and Watsons show that the dominant NO_3 source was more isotopically depleted (less than +8% for Bass and less than +9% for Watsons). Nevertheless, the isotopic signatures of the dominant source; indicated by the yintercepts of the Keeling plots were a lot more enriched than the initial δ^{15} N-NO₃ of all three pre-identified NO₃ end members. Interestingly, these δ^{15} N-NO₃ values increased with percentage agriculture except for Bass (Fig. 7). The fact that there was a clear fractionation pattern (~2:1) when integrating the isotope values of all the streams (catchment scale) suggests that denitrification was still prevalent during the wet periods (Fig. 8a) but this process was more likely to occur prior to NO₃entering the streams via surface runoff. We explain these observations on the basis that increased rainfall created a 'hot moment' in the soil whereby organic matter mineralisation and nitrification were stimulated followed by denitrification within the waterlogged soil. Waterlogging can result in root anoxia and increased denitrification; leading to significant isotopic enrichment of the residual NO₃ (Chien et al. 1977, Billy et al. 2010) which was then washed into the streams. The extent of this process (mineralisation – nitrification – denitrification) was greatest at Bass and Watsons; sites with the highest agricultural activity (Fig. 8a). Based on Fig. 8a, the isotope enrichments of the riverine NO_3^- followed the denitrification trend of the artificial fertiliser and the NO_3^- isotopes were distributed in between the denitrification ranges of both artificial fertiliser and SOM suggesting the important contribution of these two sources during the wet periods.

It is worth noting that although the dual isotopic composition of $\delta^{18}O\text{-NO}_3^-$ and $\delta^{15}N\text{-NO}_3^-$ deviates from a trajectory of 1 (trajectory of 1 indicates denitrification), it is still a salient trend indicating the occurrence of denitrification and is consistent with the $\delta^{18}O\text{-NO}_3^-$; $\delta^{15}N\text{-NO}_3^-$ recurrently observed in freshwater systems (Kendall et al. 2007). This deviation in our study could be explained by concurrent NO_3^- production catalysed by nitrification and/or annamox (Granger and Wankel 2016) although the significance of annamox is still disputable. Based on the multi-process model developed by Granger and Wankel (2016), the two most important factors in the nitrification pathway that govern the $\delta^{18}O$ of the newly produced NO_3^- are $\delta^{18}O$ of the ambient water and the flux of NO_2^- oxidation (Granger and Wankel 2016). Deflation of $\delta^{18}O\text{-NO}_3^-$; $\delta^{15}N\text{-NO}_3^-$ trajectory below 1 observed in this study was likely to be associated with the low $\delta^{18}O\text{-H}_2O$ values which contributed to lower $\delta^{18}O$ values for nitrified NO_3^- . Higher NO_3^- reduction rate versus NO_2^- oxidation rate which contributed to the $\delta^{15}N\text{-enriched}$ pool of nitrified NO_3^- , greater than the denitrified NO_3^- also drives the $\delta^{18}O\text{-NO}_3^-$; $\delta^{15}N\text{-NO}_3^-$ trajectory to values below 1 (see Granger and Wankel 2016 for explanation). All in all, this highlights the significant contribution of nitrification along with denitrification in the WP catchment.

4.4 Key controlling processes of nitrate during the dry periods

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Unlike the wet periods, only NO_3^- in the Bass River showed an apparent relationship with $\delta^{15}N\text{-NO}_3^-$ (Fig. 6) during the dry periods. There was no obvious relationships between $\delta^{15}N\text{-NO}_3^-$ and $1/[NO_3^-]$ for all other systems during the dry periods limiting the interpretation available from the Keeling plots. This also suggests that mixing between two end members alone is inadequate to explain the variability of $\delta^{15}N\text{-NO}_3^-$ during the dry periods. In general, during the dry periods, none of the samples show a noticeable pattern of denitrification on a biplot of $\delta^{18}O$ vs. $\delta^{15}N$ (Fig. 8b). The isotopic composition of the riverine NO_3^- appeared to be clustered into three groups (A, B and C in Fig 8b):

(1) NO₃⁻ in group A showed consistent $\delta^{18}O$ but variable $\delta^{15}N$. This is demonstrated by the Lang Lang and Bass; coincident with the highest percentage of agriculture. The consistent $\delta^{18}O$ ($\delta^{18}O$ of ~2.5%) shows the importance of nitrification ($\delta^{18}O$ of ~0.3 to 4.3%) and at the same time ruled out the occurrence of denitrification and assimilation. In the absence of the removal processes, the heavy and variable $\delta^{15}N$ -NO₃⁻ values (+6% to +20%) imply that animal manure was an apparent source of NO₃⁻ during the dry periods for Lang Lang and Bass. This is because volatilization of ¹⁴N ammonia from the animal manure over time can lead to enrichment of ¹⁵N in the residual NH₄⁺ to > +20% (Batman and Kelly 2007) which can subsequently be nitrified to produce isotopically-enriched NO₃⁻ without affecting its $\delta^{18}O$ -NO₃⁻. Tight coupling between mineralisation and nitrification results in NO₃⁻ retaining the isotopic signature of the residual NH₄⁺ (Deutsch et al. 2009) in the manure. Hence, it is not surprising that $\delta^{15}N$ -NO₃⁻ > +13% in the group A dataset is indicative of nitrified 'aged' animal manure. Because

of the huge variability in the fractionation effect of ammonia volatilisation, it is difficult to affix an average $\delta^{15}N$ value to represent the isotopic signature of this end member. As such, apportioning the relative contribution of nitrified manure versus other sources (nitrified organic matter in the sediment and inorganic fertiliser) is not possible.

- (2) NO_3^- in group B has variable $\delta^{15}N$ and $\delta^{18}O$ values as shown by Bunyip and Toomuc. This could be attributed to isotopic fractionation either during plant and/or algae uptake or denitrification as substantiated by the parallel increase of $\delta^{18}O-NO_3^-$ versus $\delta^{15}N-NO_3^-$ (Fig. 9). Based on Fig. 9, the large uncertainties in the $\delta^{18}O-NO_3^-$ of the nitrified end members have resulted in overlapping of isotopic signatures of the three major sources (nitrified cow manure, nitrified inorganic fertiliser and nitrified SOM). All three sources appeared to have influenced the $\delta^{15}N$ and $\delta^{18}O$ of the residual NO_3^- in the stream. This scenario reinstates the sensitivity and the importance of accurately determining the $\delta^{18}O-NO_3^-$ of the initial NO_3^- in the effort to apportion the relative contribution of different sources.
- (3) NO_3^- in group C comprised the most enriched $\delta^{15}N$ and $\delta^{18}O$ in the entire dataset (Fig. 8). These isotope values were observed in Watsons Creek which has the highest percentage of market gardens. These samples were collected when the creek was not flowing, hence the enriched $\delta^{15}N$ and $\delta^{18}O$ values could be indications of repeated cycles of internal processes (i.e. volatilisation, nitrification, denitrification and assimilation) in the same pool which enriched the N isotope but had slight effects on the O isotope of NO_3^- .

Although the isotope values during the dry periods appeared to be more likely controlled by manure and SOM, the contribution from artificial fertiliser cannot be excluded. As mentioned in the preceding text, most of the fertiliser-derived NO_3^- was denitrified in the catchment during the wet periods creating an artefact of heavy NO_3^- isotopes in the streams. This NO_3^- could exhibit a similar enriched isotopic composition as the volatilised manure. Overlapping of these isotopic values made it difficult to distinguish between the two sources – a disadvantage of using NO_3^- isotopes in a system where multiple sources and transformation processes coexist. One piece of compelling evidence to show that contribution of artificial fertiliser was also prominent during the dry periods is the deviation of the group B data towards the theoretical assimilation ranges of artificial fertiliser (Fig. 8b). However, calculating the relative contribution of the fertiliser end member is not possible.

25 **5 Conclusions**

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This study highlights the effect of rainfall conditions on the predominance of sources and transformation processes of NO_3^- on both individual stream and catchment scale. The significant positive relationships between percentage agriculture and NO_3^- concentrations as well as $\delta^{15}N-NO_3^-$ showed that enriched NO_3^- concentrations and $\delta^{15}N-NO_3^-$ values resulted from agricultural activities. The dual isotopic compositions of NO_3^- revealed that both mixing of diffuse sources and biogeochemical attenuation controlled the fate of NO_3^- in the streams of the Western Port catchments. During the wet periods, inorganic fertiliser appeared to be the primary source of NO_3^- to the streams while volatilised animal manure and SOM were the dominant sources of NO_3^- during the dry periods. Denitrification in the catchment appeared to be the more active removal process during the wet periods

in contrast to a greater importance of in-stream assimilation during the dry periods. However, these removal processes were insufficient to remove the agricultural-derived NO_3^- inferring that the streams were unreactive conduits of NO_3^- which might pose a potential NO_3^- enrichment threat to downstream ecosystems. To the best of our knowledge, this is the first study in Australia and also one of the very few targeted studies in the southern hemisphere investigating the origin and sink of NO_3^- on a catchment scale using both $\delta^{15}N$ and $\delta^{18}O$ of NO_3^- . The application of NO_3^- isotopes in a region with highly variable and unpredictable rainfall patterns such as the Western Port catchments although challenging; is imperative particularly in setting guidelines for sustainable land use management actions.

References

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- Barnes, R.T. and Raymond, P.A.: Land-use controls on sources and processing of nitrate in small watersheds: insights from dual isotopic analysis, Ecological Applications, 20(7), 1961-1978, doi: 10.1890/08-1328.1, 2010.
- Bateman, A.S. and Kelly S.D.: Fertilizer nitrogen isotope signatures, Isotopes in Environmental and Health Studies, 43(3), 237-247, doi: 10.1080/10256010701550732, 2007.
- Battaglin, W.A., Kendall, C., Chang, C.C.Y., Silva, S.R., and Campbell, D.H.: Chemical and isotopic evidence of nitrogen transformation in the Mississippi River, 1997–98, Hydrological Processes, 15(7), 1285-1300, doi: 10.1002/hyp.214, 2001.
- Billy, C., Billen, G., Sebilo, M., Birgand, F., and Tournebize, J.: Nitrogen isotopic composition of leached nitrate and soil organic matter as an indicator of denitrification in a sloping drained agricultural plot and adjacent uncultivated riparian buffer strips, Soil Biology and Biochemistry, 42(1), 108-117, doi: 10.1016/j.soilbio.2009.09.026, 2010.
- Buchwald, C., Santoro, A.E., McIlvin, M.R., and Casciotti, K.L.: Oxygen isotopic composition of nitrate and nitrite produced by nitrifying cocultures and natural marine assemblages, Limnology and Oceanography, 58(5), 1361-1375, doi: 10.4319/lo.2012.57.5.1361, 2012.
 - Burns, D.A., Boyer, E.W., Elliott, E.M., and Kendall, C.: Sources and transformations of nitrate from streams draining varying land uses: Evidence from dual isotope analysis, Journal of Environmental Quality, 38(3), 1149-59, doi: 10.2134/jeq2008.0371, 2009.
 - Carillo-Rivera, J. J. Hydrogeology of Western Port. Geological Survey of Victoria, 1975.
 - Carmargo, J.A. and Alonso, A.: Ecological and toxicological effects of inorganic nitrogen pollution in aquatic ecosystems: A global assessment, Environment International, 32, 831 849, doi: 10.1016/j.envint.2006.05.002, 2006.

- Casciotti, K.L., Sigman, D.M., Hastings, M.G., Böhlke, J.K., and Hilkert, A.: Measurement of the oxygen isotopic composition of nitrate in seawater and freshwater using the denitrifier method, Analytical Chemistry, 74(19), 4905-4912, doi: 10.1021/ac020113w, 2002.
- Casciotti, K.L., McIlvin, M., and Buchwald, C.: Oxygen isotopic exchange and fractionation during bacterial ammonia oxidation, Limnology and Oceanography, 55(2), 753-762, doi: 10.4319/lo.2010.55.2.0753, 2010.
- Chang, C.C.Y., Kendall, C., Silva, S.R., Battaglin, W.A., and Campbell, D.H.: Nitrate stable isotopes: Tools for determining nitrate sources among different land uses in the Mississippi River Basin, Canadian Journal of Fisheries and Aquatic Sciences, 59(12), 1874-1885, doi: 10.1139/F02-153, 2002.
- Chen, F., Jia, G., and Chen, J.: Nitrate sources and watershed denitrification inferred from nitrate dual isotopes in the Beijiang River, South China, Biogeochemistry, 94(2), 163-174, doi: 10.1007/s10533-009-9316-x, 2009.
- Chien, S.H., Shearer, G., and Kohl, D.H.: The nitrogen isotope effect associated with nitrate and nitrite loss from waterlogged soils, Soil Sci. Soc. Am. J., 41, 63-69, doi:10.2136/sssaj1977.03615995004100010021x, 1977.
- Danielescu, S. and MacQuarrie, K.T.B.: Nitrogen and oxygen isotopes in nitrate in the groundwater and surface water discharge from two rural catchments: implications for nitrogen loading to coastal waters, Biogeochemistry, 115(1), 111-127, doi: 10.1007/s10533-012-9823-z, 2013.
- Deutsch, B., Voss, M., and Fischer, H.: Nitrogen transformation processes in the Elbe River: Distinguishing between assimilation and denitrification by means of stable isotope ratios in nitrate, Aquatic Sciences, 71(2), 228-237, doi: 10.1007/s00027-009-9147-9, 2009.
- Elliott, E.M., Kendall, C., Wankel, S.D., Burns, D.A., Boyer, E.W., Harlin, K., Bain, D.J., and Butler, T.J.:

 Nitrogen isotopes as indicators of NOx source contributions to atmospheric nitrate deposition across the midwestern and northeastern United States, Environ. Sci. Technol, 41, 7661-7667, doi: 10.1021/es070898t, 2007.
- Enanga, E.M., Creed, I.F., Casson, N.J., and Beall, F.D.: Summer storms trigger soil N2O efflux episodes in forested catchments, Journal of Geophysical Research: Biogeosciences, 121(1), 95-108, doi: 10.1002/2015JG003027, 2016.
- Fry, B.: Stable isotope ecology, USA, New York, Springer, 2006.

10

15

20

- Galloway, J.N., Dentener, F.J., Capone, D.G., Boyer, E.W., Howarth, R.W., Seitzinger, S.P., Asner, G.P., Cleveland, C.C., Green, P.A., Holland, E.A., Karl, D.M., Michaels, A.F., Porter, J.H., Townsend, A.R., and Vörösmarty, C.J.: Nitrogen cycles: past, present, and future, Biogeochemistry, 70, 153 226, doi: 10.1007/s10533-004-0370-0, 2004.
 - Galloway J.N.: The global nitrogen cycle: past, present and future. Science in China, Science in China Series C: Life Sciences, 48, 669-677, doi: 10.1007/BF03187108, 2005.
 - Granger, J. and Wankel. S.D.: Isotopic overprinting of nitrification on denitrification as a ubiquitous and unifying feature of environmental nitrogen cycling, PNAS, 113(42), E6391-E6400, doi:10.1073/pnas.1601383113, 2016.

- Harrington, R.R., Kennedy, B.P., Chamberlain, C.P., Blum, J.D., and Folt, C.L.: 15N enrichment in agricultural catchments: field patterns and applications to tracking Atlantic salmon (Salmo salar), Chemical Geology, 147(3-4), 281-294, doi: 10.1016/S0009-2541(98)00018-7, 1998.
- 5 Hübner, H.: Isotope effects of nitrogen in the soil and biosphere, Fritz, P. and Fontes, J.C., Handbook of Environmental Isotope Geochemistry, The Terrestrial Environment. Elsevier, Amsterdam, 2b, 361-425, 1986.
 - Hunter, W.J.: Pilot-scale vadose zone biobarriers removed nitrate leaching from a cattle corral, Journal of Soil and Water Conservation, 68, 52-59, doi: 10.2489/jswc.68.1.52, 2013.
 - Kalff, J.: Limnology, Prentice-Hall, New Jersey, 2001.

- 10 Kaste, Ø., Bechmann, M., and Mørkved, P.T.: Tracing sources of nitrate in agricultural catchments by natural stable isotopes, Norwegian Institute for Water Research, 2006.
 - Kaushal, S.S., Groffman, P.M., Mayer, P.M., Striz, E., and Gold, A.J.: Effects of stream restoration on denitrification in an urbanizing watershed, Ecological Applications, 18(3), 789–804, doi: 10.1890/07-1159.1, 2008.
 - Kaushal, S.S., Groffman, P.M., Band, L.E., Elliott, E.M., Shields, C.A., and Kendall, C.: Tracking nonpoint source nitrogen pollution in human-impacted watersheds, Environmental Science & Technology, 45(19), 8225-8232, doi: 10.1021/es200779e, 2011.
 - Kendall, C. and Caldwell, E.A.: Fundamentals of isotope geochemistry, Isotope tracers in catchment hydrology, Kendall, C. and McDonnell, J.J., Elsevier, Amsterdam, 51-86, 1998.
 - Kendall, C., Elliott, E.M., and Wankel, S.D.: Tracing anthropogenic inputs of nitrogen to ecosystems, Stable isotopes in ecology and environmental science, Michener, R.H. and Lajtha, K., Blackwell Publishing Ltd, Boston, 375–449, 2007.
 - Kroopnick, P. and Craig, H.: Atmospheric oxygen: isotopic composition and solubility fractionation, Science, 175(4017), 54-55, doi: 10.1126/science.175.4017.54, 1972.
- Mayer, B., Boyer, E.W., Goodale, C., Jaworski, N.A., Van Breemen, N., Howarth, R.W., Seitzinger, S., Billen, G.,
 Lajtha, K., Nadelhoffer, K., Van Dam, D., Hetling, L.J., Nosal, M., and Paustian, K.: Sources of nitrate in rivers draining sixteen watersheds in the northeastern U.S.: Isotopic constraints, Biogeochemistry, 57-58, 171-197, doi: 10.1023/A:1015744002496, 2002.
- McIlvin, M.R. and Altabet, M.A.: Chemical conversion of nitrate and nitrite to nitrous oxide for nitrogen and oxygen isotopic analysis in freshwater and seawater, Analytical Chemistry, 77(17), 5589-5595, doi: 10.1021/ac050528s, 2005.

- Mueller, C., Zink, M., Samaniego, L., Krieg, R., Merz, R., Rode, M., and Knöller, K.: Discharge driven nitrogen dynamics in a mesoscale river basin as constrained by stable isotope patterns, Environmental Science and Technology, 17, 9187-9196, doi: 10.1021/acs.est.6b01057, 2016.
- Murdiyarso, D., Hergoualc'h, K., and Verchot, L.V.: Opportunities for reducing greenhouse gas emissions in tropical peatlands, PNAS, 107(46) 19655-19660, doi: 10.1073/pnas.0911966107, 2010.
- Nestler, A., Berglund, M., Accoe, F., Duta, S., Xue, D.M., Boeckx, P., and Taylor, P.: Isotopes for improved management of nitrate pollution in aqueous resources: review of surface water field studies, Environmental Science and Pollution Research International, 18, 519–533, doi: 10.1007/s11356-010-0422-z, 2011.

10

15

- Ohte, N.: Tracing sources and pathways of dissolved nitrate in forest and river ecosystems using high-resolution isotopic techniques: a review, Ecological Research, 28(5), 749-757, doi: 10.1007/s11284-012-0939-3, 2013.
- Ohte, N., Dahlgren, R.A., Silva, S.R., Kendall, C., Kratzer, C.R., and Doctor, D.H.: Sources and transport of algae and nutrients in a Californian river in a semi-arid climate, Freshwater Biology, 52(12), 2476-2493, doi: 10.1111/j.1365-2427.2007.01849.x, 2007.
- Panno, S.V., Kelly, W.R., Hackley, K.C., Hwang, H.H., and Martinsek, A.T.: Sources and fate of nitrate in the Illinois River Basin, Illinois, Journal of Hydrology, 359(1–2), 174-188, doi: 10.1016/j.jhydrol.2008.06.027, 2008.
- Pardo, L.H., Kendall, C., Pett-Ridge, J., and Chang, C.C.Y.: Evaluating the source of streamwater nitrate using $\delta^{15}N$ and $\delta^{18}O$ in nitrate in two watersheds in New Hampshire, USA, Hydrological Processes, 18(14), 2699-2712, doi: 10.1002/hyp.5576, 2004.
- Peterson, B.J., Wollheim, W.M., Mulholland, P.J., Webster, J.R., Meyer, J.L., Tank, J.L., Marti, E., Bowden, W.B., Valett, H.M., Hershey, A.E., McDowell, M.H., Dodds, W.K., Hamilton, S.K., Gregory, S., and Morrall, D.D.: Control of nitrogen export from watersheds by headwater streams, Science, 292(5514), 86-90, doi: 10.1126/science.1056874, 2001.
 - Quay, P.D., Wilbur, D.O., Richey, J.E., Devol, A.H., Benner, R., and Forsberg, B.R.: The δ¹⁸O:δ¹⁶O of dissolved oxygen in rivers and lakes in the Amazon Basin: Determining the ratio of respiration to photosynthesis rates in freshwaters, Limnology and Oceanography, 40(4), 718-729, 1995.
 - Rafter P.A., DiFiore, P.J., and Sigman, D.M.: Coupled nitrate nitrogen and oxygen isotopes and organic matter remineralization in the Southern and Pacific Oceans, Journal of Geophysical Research: Oceans, 118(10), 4781-4794, doi: 10.1002/jgrc.20316, 2013.
- Riha, K.M., Michalski, G., Gallo, E.L., Lohse, K.A., Brooks, P.D., and Meixner, T.: High atmospheric nitrate input and nitrogen turnover in semi-arid urban catchments, Ecosystems, 17(8), 1309-1325, doi: 10.1007/s10021-014-9797-x, 2014

- Sebilo, M., Billen, G., Grably, M., and Mariotti, A.: Isotopic composition of nitrate-nitrogen as a marker of riparian and benthic denitrification at the scale of the whole Seine River system, Biogeochemistry, 63(1), 35–51, doi: 10.1023/A:1023362923881, 2003.
- Sigman, D.M., DiFiore, P.J., Hain, M.P., Deutsch, C., Wang, Y., Karl, D.M., Knapp, A.N., Lehmann, M.F., Pantoja, F.: The dual isotopes of deep nitrate as a constraint on the cycle and budget of oceanic fixed nitrogen, Deep Sea Research Part I: Oceanographic Research Papers, 56(9), 1419-1439, doi: 10.1016/j.dsr.2009.04.007, 2009.

20

25

- Stein, J.L., Hutchinson, M.F., and Stein, J.A.: A new stream and nested catchment framework for Australia, Hydrol Earth Syst Sci, 18, 1917–1933, doi: 10.5194/hess-18-1917-2014, 2014.
- Vitousek, P.M., Aber, J., Howarth, R.W., Likens, G.E., Matson, P.A., Schindler, D.W., Schlesinger, W.H., and
 Tilman, G.D.: Human alteration of the global nitrogen cycle: Causes and consequences, Ecological Applications,
 7(3), 737-750, doi: 10.1890/1051-0761(1997)007[0737:HAOTGN]2.0.CO;2, 1997.
 - Voss, M., Deutsch, B., Elmgren, R., Humborg, C., Kuuppo, P., Pastuszak, M., Rolff, C., Schulte, U.: Source identification of nitrate by means of isotopic tracers in the Baltic Sea catchments, Biogeosciences, 3(4), 663-676, doi: 10.5194/bg-3-663-2006, 2006.
- 15 Xue, D., Botte, J., De Baets, B., Accoe, F., Nestler, A., Taylor, P., Van Cleemput, O., Berglund, M., and Boeckx, P.: Present limitations and future prospects of stable isotope methods for nitrate source identification in surface- and groundwater, Water Research, 43(5), 1159-1170, doi: 10.1016/j.watres.2008.12.048, 2009.
 - Yevenes, M.A., Soetaert, K., and Mannaerts, C.M.: Tracing nitrate-nitrogen sources and modifications in a stream impacted by various land uses, south Portugal, Water, 8(9), 385, doi: 10.3390/w8090385, 2016.

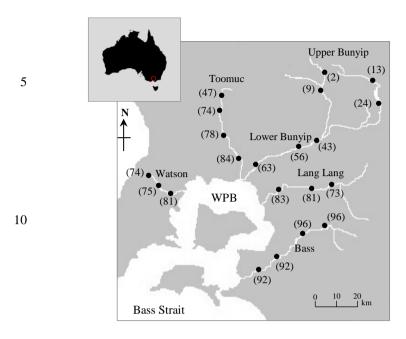
Table 1: The isotopic compositions of potential sources of NO₃⁻ in the catchment

Sample	δ ¹⁵ N-TN (‰)	δ ¹⁸ O-H ₂ O (‰)
Artificial/inorganic fertiliser	-0.5 to +0.7	-
Cow manure/organic fertiliser	+6 to +13	-
Sediment (SOM)	+4 to +5	-
Stream water	-	-5.5 to -4.9

Table 2: Comparison of NO₃⁻ concentrations and isotopes across different systems reported in the literature

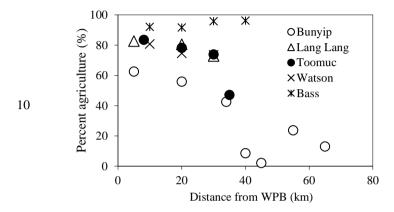
Study area	Percentage agriculture (%)	[NO ₃ -] (μM)	δ ¹⁵ N-NO ₃ - (‰)	δ ¹⁸ O-NO ₃ - (‰)	Reference
Mississippi River Basin, USA	0 to 100	3.6 to 1290	-1.4 to +12.3	+3.1 to +43.3	Chang et al. 2002
Connecticut River Watershed, USA	0.8 to 52	0 to 360	*0 to +14.5	*-2 to +14	Barnes et al. 2010
New York, USA	0 to 72	*5 to 640	*0 to +9	*-8 to +40	Burns et al. 2009
Mid-Atlantic and New England states of the USA	2 to 38	7.9 to 184	+3.6 to +8.4	+11.7 to +18.5	Mayer et al. 2002
Baltic Sea catchment	1 to 81	3 to 216	-1.5 to +14	+10 to +25	Voss et al 2006
Trout River catchment, Atlantic Canada	~39.7	32 to 170	+2.13 to +6.35	+1.51 to +7.07	Danielescu and MacQuarrie 2013
Skuterud catchment, Norway	0 to 100	21 to 1850	+3 to +18	+10 to +24	Kaste et al. 2006
Mørdre catchment, Norway	74 to 100	120 to 2320	+8 to +15	+5 to +20	Kaste et al. 2006
Pearl river drainage basin	~86	41 to 110	+1.9 to +17.6	+5.6 to +17.3	Chen et al. 2009
Westernport catchment, Australia	2 to 96	4 to 790	+5.7 to +33	+1.4 to +12.7	This study

^{*}Values estimated from presented figures, might not accurately represent the actual data



15 Figure 1: Map of Western Port Bay (WPB) in southern Victoria, Australia and major rivers discharging into WPB. Closed circles represent sampling sites where surface water samples were obtained. Values in parentheses represent the % agriculture area in the catchment.





15 Figure 2: The percent agriculture for each of the sampling sites.

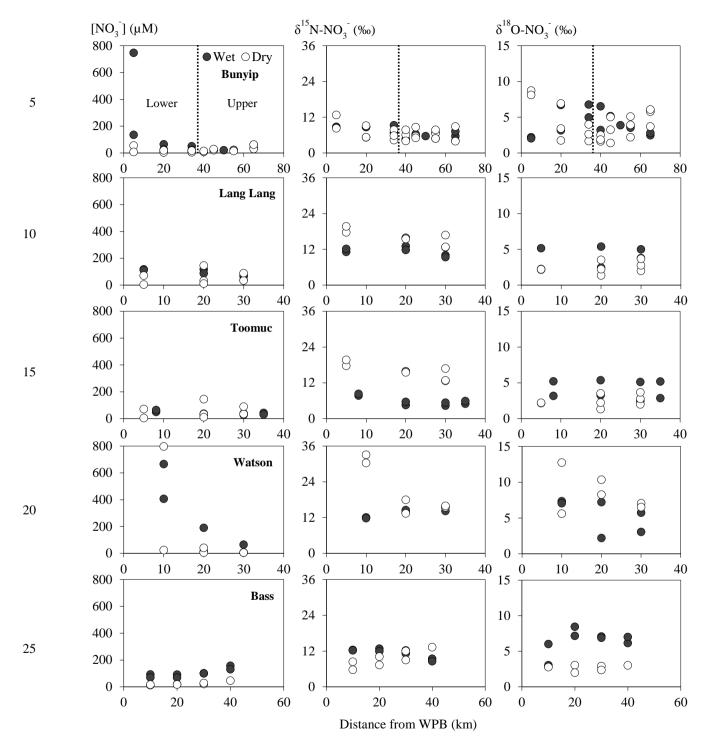


Figure 3: Spatial and temporal variations of nitrate concentrations and isotopes values. Closed circles represent data obtained during the wet periods. Open circles represent data obtained during the dry periods.

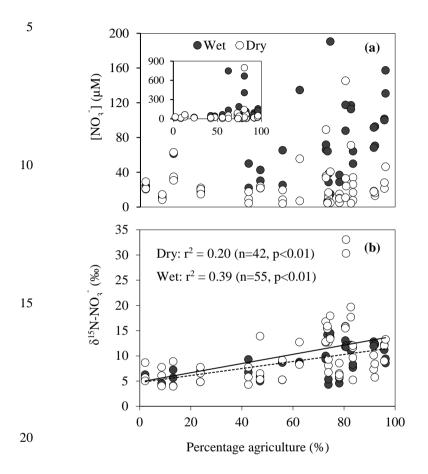
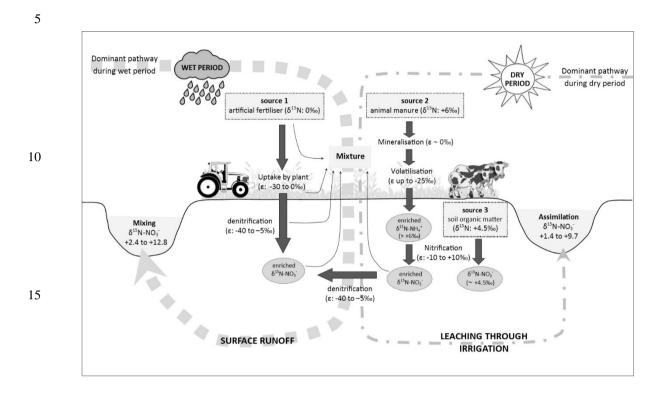


Figure 4: Relationship between (a) NO_3 concentration; (b) $\delta^{15}N$ - NO_3 and the percentage of agricultural land use. In (b) solid line represents the relationship between the variables during dry periods; dotted line represents wet periods.



20 Figure 5: Conceptual diagram illustrating the sources and processes of NO₃ during the wet and dry periods in the Western Port catchment. The values of enrichment factor (ε) were obtained from the literature to indicate the relative contribution of the transformation processes to the isotopic compositions of the residual NO₃.

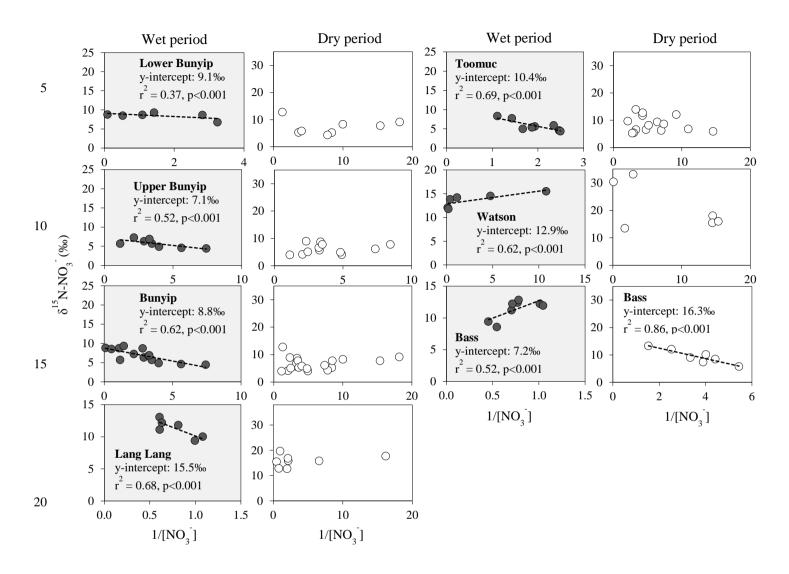
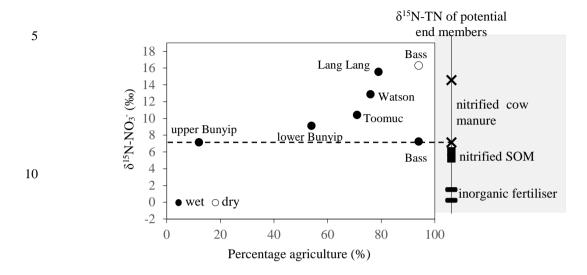


Figure 6: Relationship between $\delta^{15}N$ -NO₃ and 1/[NO₃] for individual streams during the wet and dry periods.



15 Figure 7: Relationship between $\delta^{15}N$ -NO₃ of the dominant initial source (indicated by the y-intercept of the Keeling plots; Figure 6) and percentage agriculture during wet periods. Data for Bass-dry period was also presented because only the Keeling plot for Bass-dry period indicates mixing between different sources. The shaded area represents the $\delta^{15}N$ -TN of the potential end members.

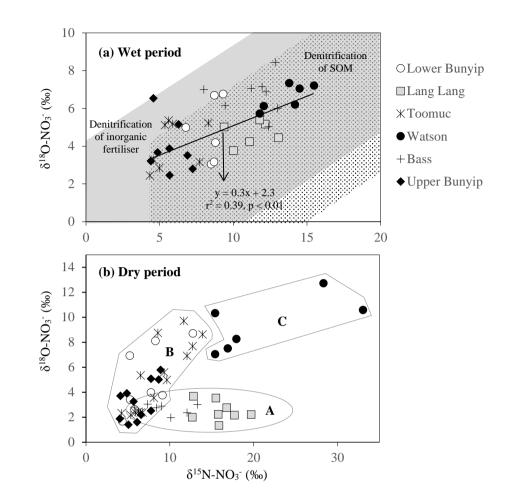


Figure 8: Biplot of $\delta^{15}N\text{-NO}_3$ versus $\delta^{18}O\text{-NO}_3$ for (a) wet and (b) dry periods. Blue shaded area represents possible isotopic compositions of denitrified NO_3 originated from SOM ($\delta^{15}N\text{:}+4.5\%$). Grey shaded area represents the possible isotopic composition of denitrified NO_3 originated from inorganic fertiliser ($\delta^{15}N\text{-NO}_3$: +0.1%). The $\delta^{18}O\text{-NO}_3$ used were -5.3% and +4.3% representing the minimum and maximum estimates of $\delta^{18}O$ of nitrified NO_3 , respectively. The shaded area were plotted based on the theoretical 2:1 denitrification relationship between $\delta^{15}N\text{-NO}_3$ and $\delta^{18}O\text{-NO}_3$ (Kendall et al. 2007).

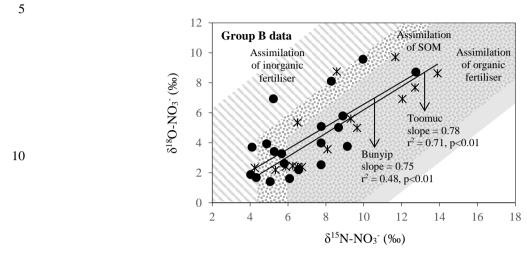


Figure 9: Biplot of δ^{15} N-NO₃ versus δ^{18} O-NO₃ for Bunyip and Toomuc (group B data in Fig. 8b). Shaded areas represent theoretical assimilation trends for cow manure, SOM and inorganic fertiliser. The initial d15N values were Solid lines represent the assimilation trends for Bunyip (both lower and upper Bunyip) and Toomuc.