

BG-2017-255 Editor response to Revised Manuscript

We thank the editor for the constructive comments. We have considered the suggestions and improved the manuscript accordingly.

I would appreciate it if the points below could be addressed in a further revision. I do not believe the changes are especially onerous.

Reviewer #2

Line 216 The reviewer stated ‘ “Temperature had a positive effect on the GI of living maerl. Conversely, GI was significantly reduced under high pCO₂...” The authors fail to mention that in the combined treatment, temperature alleviated the negative effect of pCO₂. This is very important to the story. ‘, and the answer provided was as follows: A: The sentence has been revised due to the change in statistical design. (L. 218-219) “The GI of living maerl was not significantly influenced by increased temperature and pCO₂, regardless of the season” – To my mind this response is not clear. Do you mean that with the new statistical design there were no significant effects of either temperature or CO₂, or that the combined treatment was not significantly different from the control? If the latter, then please at least make the suggestion mentioned by the reviewer that the individual effects of CO₂ and temperature appeared to have cancelled each other out.

Answer: According to the now statistical design, no significant effect of temperature and pCO₂ was detected on G₁ (see table below). We have kept the statement: “The G₁ of living maerl was not significantly influenced by increased temperature and pCO₂, regardless of the season (Table 5, Fig. 2d)” (lines 219-220).

		Light calcification	
		G ₁	
LIVING <i>L. corallioides</i>	WINTER	F	p
	T	3.6	0.08
	pCO ₂	3.2	0.09
	pCO ₂ x T	0.8	0.39
	SUMMER		
	T	3.3	0.07
	pCO ₂	3.6	0.06
pCO ₂ x T	0.2	0.65	

Line 238: The reviewer asks you to state that ‘temperature alone decreased epiphyte biomass in the summer’, however the re-wording that you have made does not make this point clearly (you state ‘while an interactive effect of temperature and pCO₂ was observed in the summer’). I would prefer you to make a clear statement about the direction of this interaction and how the variables concerned were affected. If it is not possible to simply state that elevated temperature led to decreased epiphyte biomass in summer then please state why not (perhaps not supported by the new stats).

A: We have reworded our statement to consider the direction of the interaction for each variable: “In the summer, increased temperature reduced epiphyte biomass under ambient pCO₂ and stimulated epiphyte biomass under elevated pCO₂ (2-way ANOVA, p=0.013, supplementary material e).” (lines 242-243).

Discussion. The reviewer asks you to state more clearly that the combined effects of temperature and CO₂ may be less marked than the individual effects of those variables. In part of your re-worded section you state ‘Under the predicted business-as-usual conditions, epiphyte overgrowth may exacerbate the negative impact of climate change on underlying coralline algae.’ Please check this, as I think you may mean ‘ameliorate’ instead of ‘exacerbate’ (opposite meanings).

A: In the conclusion, we stated that the combined effects of temperature and CO₂ ameliorate the response of assemblages: “In contrast with other studies, [...] we showed here that the effects of pCO₂ and temperature on maerl bed communities were weakened when these factors were combined.” (lines 369 to 372).

However, we also suggest that: “Epiphyte overgrowth is thus likely to adversely affect underlying coralline algae, increasing the shading effect at light and the pH decline in the dark.” (lines 374-375).

Species interactions can shift the response of a maerl bed community to ocean acidification and warming

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Abstract

10 Predicted ocean acidification and warming are likely to have major implications for marine organisms, especially marine calcifiers. However, little information is available on the response of marine benthic communities as a whole to predicted changes. Here, we experimentally examined the combined effects of temperature and partial pressure of carbon dioxide (pCO₂) increases on the response of maerl bed assemblages, composed of living and dead thalli of the free-living coralline alga *Lithothamnion corallioides*, epiphytic fleshy algae, and grazer species. Two three-month experiments were performed

15 in the winter and summer seasons in mesocosms with four different combinations of pCO₂ (ambient and high pCO₂) and temperature (ambient and + 3°C). The response of maerl assemblages was assessed using metabolic measurements at the species and assemblage scales. This study suggests that seasonal variability represent an important driver influencing the magnitude and the direction of species and community response to climate change. Gross primary production and respiration of assemblages were enhanced by high pCO₂ conditions in the summer. This positive effect was attributed to the increase in

20 epiphyte biomass, which benefited from higher CO₂ concentrations for growth and primary production. Conversely, high pCO₂ drastically decreased the calcification rates in assemblages. This response can be attributed to the decline in calcification rates of living *L. corallioides* due to acidification as well as increased dissolution of dead *L. corallioides*. Future changes in pCO₂ and temperature are likely to promote the development of non-calcifying algae to the detriment of the engineer species *L. corallioides*. The development of fleshy algae may be modulated by the ability of grazers to regulate

25 epiphyte growth. However, our results suggest that predicted changes will negatively affect the metabolism of grazers and potentially their ability to control epiphyte abundance. We evidenced here that the effects of pCO₂ and temperature on maerl

bed communities were weakened when these factors were combined. This underlines the importance of examining multi-factorial approaches and community-level processes, which integrate species interactions, to better understand the impact of global change on marine ecosystems.

30 **1. Introduction**

A growing body of literature predicts that ocean acidification and warming will be the main anthropogenic drivers affecting marine species by the end of the century (Kroeker et al., 2013). Due to the increase in atmospheric CO₂, seawater surface temperatures have been predicted to increase by 0.71-2.73°C and pH to decline by 0.07-0.33 units in the surface ocean by the end of the 21st century (Bopp et al., 2013).

35 Species interactions are a key element in ecosystem functioning and are likely to attenuate or amplify the direct effects of climate change on individual species (O'Connor et al., 2011; Hansson et al., 2012; Kroeker et al., 2012). Most research on benthic ecosystems has focused on the impact of ocean acidification and warming on the response of single species (Yang et al., 2016) and despite a growing interest, studies examining the effects of climate change at the community scale are scarce in the literature (Hale et al., 2011; Alsterberg et al., 2013). Understanding the mechanisms and interactions that occur among
40 marine communities that face the predicted changes is necessary for a better overview of marine ecosystem response. Climate change is likely to strongly alter interactions between macroalgae (e.g. calcifying and non-calcifying macroalgae; Olabarria et al., 2013; Short et al., 2014; Short et al., 2015), interactions between grazers and macroalgae (Poore et al., 2016; Sampaio et al., 2017) as well as prey-predator dynamics (Asnaghi et al., 2013; Jellison et al., 2016), inducing drastic consequences on the structure and functioning of marine ecosystems (Widdicombe and Spicer, 2008; Hale et al., 2011).

45 Maerl beds feature high structural and functional diversity arising primarily from the numerous species interactions that occur in this environment — in particular, interactions between fleshy and calcareous macroalgae and grazers and macroalgae (Hily et al., 1992; Guillou et al., 2002; Grall et al., 2006). The accumulation of living and dead thalli of free-living coralline algae (Corallinaceae, Rhodophyta) creates a complex three-dimensional structure that provides habitat for many faunal and floral species (Foster et al., 2007; Amado-Filho et al., 2010; Peña et al., 2014), some of which have high

50 commercial value (Grall and Hall-Spencer, 2003). In some locations, dead maerl can reach high proportions compared with living maerl (Hily et al., 1992), thereby contributing substantially to the local carbonate dynamics (Martin et al., 2007). The main species inhabiting maerl beds may respond differently to ocean acidification and warming. Coralline algae are known to be among the most vulnerable species facing ocean acidification (McCoy and Kamenos, 2015; Martin and Hall-Spencer, 2016), due to their highly soluble Mg-calcite skeleton (Morse et al., 2006). The deleterious consequences of ocean acidification have also been demonstrated for other calcareous marine taxa, such as mollusks (Gazeau et al., 2013; Parker et al., 2013) and echinoderms (Dupont et al., 2010), with reductions in survival, growth, development, and abundance (Kroeker et al., 2013). Conversely, some species can benefit from the increase in CO₂ concentration and temperature. Positive responses, such as increases in primary production and growth, have been found mostly among non-calcifying organisms, such as fleshy algae and seagrasses (Koch et al., 2013; Pajusalu et al., 2013).

60 Here, we experimentally investigated the impact of ocean acidification and warming on the metabolism and the interactions of the main maerl-forming species in Brittany *Lithothamnion coralloides* and the epiphytic fleshy macroalgae and main grazer (gastropods and sea urchins) associated with it. Because the response of species and communities to climate change is likely to vary depending on seasonal changes in environmental factors, such as light intensity, photoperiod and temperature (Godbold and Solan, 2013; Martin et al., 2013; Baggini et al., 2014), it was tested in both winter and summer conditions. The response of marine communities to climate change is likely to be influenced by the direct effects of environmental stressors on individual organisms, and by the indirect effects induced by shifts in interspecific interactions (Harley et al., 2012; Auster et al., 2013). In the present study, we therefore performed metabolic measurements at the species and at the community scale. At the species scale, studying species physiology is useful for understanding how organisms cope with changing climatic conditions and for analyzing the community metabolic response. Community-scale measurements provide information on the potential shifts in species interactions induced by climate change. In particular, we tested the hypothesis that climate change will increase epiphytic fleshy algal growth, exacerbating the deleterious consequences of predicted changes on *L. coralloides* metabolism. We also investigated whether the predicted changes can modify interactions between grazers and macroalgae, and their ability to regulate epiphytic biomass.

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2. Materials and methods

75 2.1. Species collection and assemblages

Organisms were collected from a maerl bed in the Bay of Brest, France (48°18'N 4°23'W) using a naturalist's dredge (width: 1 m, height: 0.2 m, net: 1.5 m long) deployed from the research vessel *Albert Lucas*. In the Bay of Brest, maerl beds are located at depths of between 0.7 and 6.8 m, according to the tide (Dutertre et al., 2015). We deliberately selected thalli of the maerl species *L. corallioides* Crouan and Crouan, 1867 that were devoid of any apparent epiphytes; nonetheless, they were
80 not cleaned so as to retain any epiphyte spores that may have been present on their surface. Medium-sized individuals of the three main species of grazers living in maerl beds were also sampled: two gastropod species (sea snails) *Gibbula magus* Linnaeus, 1758 and *Jujubinus exasperatus* Pennant, 1777 and an urchin species *Psammechinus miliaris* Müller, 1771 (Grall et al., 2006). Samples were collected on 24 January 2015 (winter conditions) and 15 September 2015 (summer conditions). In each season, 1 kg of living thalli of *L. corallioides*, 500 g of dead thalli of *L. corallioides*, 40 individuals of *G. magus*
85 (shell length range 17-29 mm; Table S1), 40 individuals of *P. miliaris* (test diameter range 11-23 mm), and 80 individuals of *J. exasperatus* (shell height range 5-11 mm) were randomly selected and transported in seawater tanks to the Roscoff Marine Station. To mitigate the stress experienced by the species during sampling and transport, they were kept in open-flow aquaria at ambient pH and *in situ* temperature conditions at the time of collection for at least one week before starting the experiments. No mortality was recorded during this period.

90 2.2. Experimental design

Two three-month long experiments were conducted for both winter (March to June 2015) and summer (September to December 2015) conditions. For each season, 20 artificial assemblages were created and randomly assigned to 20 15 L aquaria. Each assemblage was composed of 45 g of living *L. corallioides* thalli, 20 g dead *L. corallioides* thalli, two *G. magus* individuals, two *P. miliaris* individuals and four *J. exasperatus* individuals, according to the proportions observed on
95 maerl beds.

Algae and grazers were acclimated to laboratory conditions for 7 days. Then, the pH was gradually decreased by 0.05 units per day over 7 days and temperature increased by 0.5°C per day. The pH was controlled by modifying pCO₂ through CO₂

bubbling. At each season, two pCO₂ conditions were tested, each with two temperature conditions to examine the interaction between pCO₂ and temperature. These four conditions are presented in table 1.

100 Ambient pCO₂ conditions (A-pCO₂) were determined according to *in situ* winter (7.98) and summer (8.06) mean pH_T (pH on the total scale) monitored above maerl beds in the Bay of Brest (data from Martin, unpublished data). High pCO₂ (H-pCO₂) corresponded to the “business-as-usual” scenario predicted for the end of the century, with a pH decrease of -0.33 units (RCP8.5; Bopp et al., 2013). Ambient temperature (T) corresponded to *in situ* winter (10.0°C) and summer (17.1°C) conditions in the Bay of Brest recorded by SOMLIT (from 2003 to 2014), and high temperature (T + 3°C) was determined
105 according to the business-as-usual scenario predicted for 2100 (Bopp et al., 2013).

The pH and the temperature were controlled in four 100 L tanks, continuously supplied with filtered (5 µm) natural seawater, with a high water flow rate of 150 L h⁻¹ per tank. They were maintained by an off-line feedback system (IKS Aquastar, Karlsbad, Germany) that activated or stopped heaters and solenoid valves, controlling temperature and CO₂ (Air Liquide, France) bubbling in the tanks, respectively. Each 100 L tank provided seawater to five 15 L aquaria for each of the four
110 conditions using pumps. The water flow rate was 15 L h⁻¹ in each aquarium. Temperature was maintained constant in aquaria with water baths. Seawater pH (pH_T, expressed on the total hydrogen ion concentration scale, Dickson et al., 2007) and temperature were monitored every two days in the 20 aquaria, at different times of the day. Seawater pH_T and temperature measurements were carried out using a pH probe associated with a temperature sensor (PHC101, Hach Lange, IntelliCAL). The pH probe was calibrated using Tris/HCl and 2-aminopyridine/HCl buffers (Dickson et al., 2007). The pH values of the
115 off-line feedback system were adjusted from measurements of pH_T carried out every two days in each aquarium. Total alkalinity (A_T) was also monitored during the experiment in each aquarium at different times of the day (n = 28). For A_T analyses, seawater samples (60 mL) were filtered through 0.7 µm Whatman GF/F filters and immediately poisoned with a mercuric chloride solution to prevent further biological activity (Dickson et al., 2007). A_T was determined using open-cell titration on an automatic titrator (Titroline alpha, Schott SI Analytics, Mainz, Germany) according to the method developed
120 by Dickson et al. (2007). A_T was calculated using a Gran function applied to pH values ranging from 3.5 to 3.0 (Dickson et al., 2007) and corrected using standard reference material provided by the Andrew G. Dickson laboratory (CRM Batch 111, accuracy of ± 6 µmol kg⁻¹). Salinity was measured every 2 weeks with a conductivity probe (CDC401, Hach Lange,

IntelliCAL, accuracy of 0.1) and remained constant during experiments (35.2 ± 0.2). From A_T and pH_T measurements, dissolved inorganic carbon (DIC), saturation state of seawater with respect to aragonite (Ω_{Ar}) and saturation state of seawater
125 with respect to calcite (Ω_{Ca}) were calculated with the CO2SYS software. Mean temperature and parameters of the carbonate chemistry are given in table 2.

Irradiance was set to the mean *in situ* daily irradiance at 5 m depth in the Bay of Brest according to Martin et al. (2006a): 30-40 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ in winter and 90-100 $\mu\text{mol photons m}^{-2} \text{ s}^{-1}$ in summer. The light was provided by two or four 80 W fluorescent tubes (JBL Solar Ultra Marin Day, JBL Aquaria, Nelson, New Zealand) above the aquaria under a 10/14 h or
130 14/10 h light/dark photoperiod, for winter or summer conditions, respectively.

2.3. Metabolic measurements

After three months in experimental conditions, metabolic measurements were performed at the species and assemblage level using incubations in acrylic respirometry chambers (Engineering and Design Plastics Ltd, Cambridge, UK). For species-scale measurements, each species was incubated separately. Community-scale measurements were performed on
135 assemblages, incubating all individuals from all species present in each aquarium. The chamber volume was adapted to species size. It was of 80 mL for *J. exasperatus* and epiphytes, 185 mL for *P. miliaris*, *G. magus* and living and dead *L. corallioides*, and 600 mL for the assemblages. After assemblage incubations, epiphytic algae that spontaneously grew on *L. corallioides* during the experiments were carefully removed and incubated separately. Metabolic measurements (net photosynthetic and respiration rates) for the main epiphytic algae *Rhodymenia ardissoni* and *Solieria chordalis* were only
140 examined in the summer, when their biomass was sufficient for measurements. Species were placed on a plastic grid above a stir bar in the chambers to ensure the seawater was well mixed. For *G. magus* and *P. miliaris*, net calcification, respiration and excretion (ammonia release) rates were measured. For *J. exasperatus*, only respiration rates were measured due to its limited size and metabolic rates. For grazers, physiological rates were measured under ambient irradiance. For each grazer species, individuals present in each aquarium were incubated together. For living and dead *L. corallioides* and assemblages,
145 net photosynthetic and light calcification rates were measured under ambient irradiance, and respiration and dark calcification rates were measured in the dark. For light incubations, chambers were placed inside aquaria to control temperature. For dark incubations, chambers were placed in a plastic crate filled with aquaria seawater in an open circuit to

keep the temperature constant. Incubation duration was adjusted to keep oxygen saturation above 80%. Incubations lasted approximately from 1 h for *G. magus* to 2.5 h for dead maerl. For assemblages, the metabolism was measured from the incubations of all species together.

Oxygen concentrations were measured at the beginning and at the end of each incubation, using an optical fiber system (FIBOX 3, PreSens, Regensburg, Germany). Reactive spots were calibrated with 0% and 100% buffer solutions. The 0% buffer solution was prepared by dissolving 1 g of sodium sulfite (Na_2SO_3) in 100 mL of seawater. The 100% buffer solution was prepared by bubbling air into 100 mL of seawater using an air-pump for 20 min to obtain air-saturated seawater. Net primary production (NPP, $\mu\text{mol O}_2 \text{ g DW}^{-1} \text{ h}^{-1}$) or respiration (R, $\mu\text{mol O}_2 \text{ g DW}^{-1} \text{ h}^{-1}$) rates were calculated following Eq. (1):

$$\text{NPP or R} = \frac{\Delta\text{O}_2 \times V}{\Delta t \times \text{DW}} \quad (1)$$

where ΔO_2 is the difference between the initial and final oxygen concentrations ($\mu\text{mol O}_2 \text{ L}^{-1}$), V the volume of the chamber (L), Δt the incubation time (h), and DW the dry weight of the species incubated (g). The dry weight was obtained after 48 h at 60°C. For gastropods, the body was separated from the shell to consider the dry weight of the body only.

For algae and the assemblages, gross primary production (GPP) was calculated following Eq. (2):

$$\text{GPP} = \text{NPP} - \text{R} \quad (2)$$

Control incubations containing only seawater were carried out to correct for oxygen fluxes due to any additional biological activity in seawater. Oxygen fluxes calculated in control chambers were subtracted from oxygen fluxes of chambers containing species.

Seawater samples were taken in the aquaria at the beginning of the incubation and in the chambers at the end of the incubations (except for fleshy algae and *J. exasperatus*) to measure ammonium (NH_4^+) concentration and total alkalinity (A_T). To do so, 45 mL seawater samples for NH_4^+ analyses were fixed with reagent solutions and stored in the dark. NH_4^+ concentrations were determined according to the Solorzano method (Solorzano, 1969). Absorbance was measured by spectrophotometry at a wavelength of 630 nm (spectrophotometer UV-1201V, Shimadzu Corp, Kyoto, Japan). For grazers, ammonia excretion rates (E, $\mu\text{mol NH}_4^+ \text{ g DW}^{-1} \text{ h}^{-1}$) were calculated following Eq. (3):

$$E = \frac{\Delta\text{NH}_4^+ \times V}{\Delta t \times \text{DW}} \quad (3)$$

where ΔNH_4^+ is the difference between the initial and final ammonium concentrations ($\mu\text{mol NH}_4^+ \text{ g DW}^{-1} \text{ h}^{-1}$).

For A_T analyses, 60 mL seawater samples were filtered through 0.7 μm Whatman GF/F filters and were immediately poisoned with a mercuric chloride solution. Total alkalinity was determined according to the method described above. Net calcification rates at light and in the dark (G_l and G_d , respectively; in $\mu\text{mol CaCO}_3 \text{ g DW}^{-1} \text{ h}^{-1}$) were calculated according to
 175 the alkalinity anomaly technique (Smith and Key, 1975) and corrected for NH_4^+ fluxes (Gazeau et al., 2015). This correction was applied to calcareous species and assemblage incubations following Eq. (4):

$$G_l \text{ or } G_d = \frac{(-\Delta A_T + \Delta\text{NH}_4^+) \times V}{2 \times \Delta t \times \text{DW}} \quad (4)$$

where G_l is the net calcification in the light, G_d is the net calcification in the dark, ΔA_T is the difference between the initial and final A_T ($\mu\text{eq L}^{-1}$).

After the three-month experiments, epiphytic algae that spontaneously grew on *L. corallioides* during experiments were
 180 picked off and dried at 60°C for 48 h to determine their dry weight.

2.4. Chlorophyll *a* analysis

At the end of the experiments, thalli of living and dead *L. corallioides* were collected in each aquarium and immediately frozen at -20°C pending analyses. In dead maerl, chlorophyll *a* content was measured in order to check for the presence of associated microflora and potential subsequent metabolism. Then samples were freeze-dried and crushed into a powder using
 185 a mortar, in the dark. An aliquot of 0.15 g of powder was precisely weighed and suspended in 10 mL of 90% acetone and stored in the dark at 4°C for 12 h. Samples were then centrifuged at 4000 rpm. The supernatant was collected and absorbance was measured at 630 (A_{630}), 647 (A_{647}), 664 (A_{664}), and 691 (A_{691}) nm. Chlorophyll *a* (Chl *a*) concentrations ($\mu\text{g g DW}^{-1}$) were calculated from Ritchie (2008) following Eq. (5):

$$\text{Chl } a = \frac{(-0.3319 A_{630} - 1.7485 A_{647} + 11.9442 A_{664} - 1.4306 A_{691}) \times V}{mp} \quad (5)$$

where V is the volume of acetone (mL) and mp the mass of powder (g).

190 2.5. Data analysis

Comparisons in species and assemblage physiological rates between the winter and summer seasons was performed using t-tests, after checking the normality and homogeneity of variances. The influence of temperature and pCO₂ was tested on metabolic rates of grazers (*P. miliaris*, *G. magus* and *J. exasperatus*), living and dead maerl, epiphytic biomass and assemblages. Normality of the data and variance homogeneity were checked for all variables. When assumptions were respected, two-way ANOVA were performed, using temperature and pCO₂ as fixed orthogonal factors. When assumptions were not respected, two-way non-parametric Scheirer-Ray-Hare tests were run. Statistical analyses were conducted separately for winter and summer experiments in order to keep a balanced design. When 2-way ANOVAs showed significant results, post hoc tests (Tukey honest significant difference, HSD) were performed to compare the four treatments. The effects of pCO₂ and temperature on the physiological rates of the epiphytic algae *R. ardissoni* and *S. chodalis* were only tested in the summer. Statistical analyses were carried out using the statistical package R, version 3.2.2.

3. Results

3.1. Metabolic responses of grazers to acidification and warming

In the urchin *P. miliaris* and the gastropod *G. magus*, the metabolism was significantly higher in the summer than in the winter, except for *P. miliaris* R, for which no difference was detected (Table 3). In the gastropod *J. exasperatus*, R did not vary with the season. High temperature (+3°C) reduced *P. miliaris* R in the summer, while pCO₂ had no significant effect on *P. miliaris* R (Fig. 1a; Table 4). *P. miliaris* G₁ was significantly affected by the interaction between temperature and pCO₂ in the summer (Fig. 1b; Supplementary material b), which negated the positive effect of increased temperature or pCO₂ alone. *P. miliaris* E was higher under control conditions in the summer and significantly lower under increased temperature (Table 4; Fig. 1c). *G. magus* R was lower under high pCO₂ in the winter only (Table 4; Fig. 1d). Neither temperature nor pCO₂ increases significantly affected *G. magus* G₁ and E (Table 4; Fig. 1e-f). In *J. exasperatus*, R increased under elevated temperature but in winter conditions only (Table 4; Fig. 1g). *J. exasperatus* R was negatively influenced by the pCO₂ increase in the winter, but positively in the summer.

3.2. Metabolic responses of living *L. corallioides* to acidification and warming

The metabolism of living *L. corallioides* was higher in the summer than in the winter, except for NPP (Table 3). Living
215 maerl NPP did not differ among temperature and pCO₂ conditions regardless of the season, while GPP was lower under high
temperature in the winter (Table 5; Fig. 2a,b). R was significantly reduced by the high temperature condition in the winter,
whereas an increase in R was observed in the summer (Table 5; Fig. 2c). No effect of season was observed on chlorophyll *a*
content (Tables 3; 6). Chlorophyll *a* content was reduced by the high temperature condition in the winter only (Table 5). The
G_l of living maerl was not significantly influenced by increased temperature and pCO₂, regardless of the season (Table 5;
220 Fig. 2d). Conversely, increased temperature enhanced G_d in the winter, but no effect was detected in the summer (Fig. 2e). A
significant decline in G_d occurred under high pCO₂ regardless of the season. Net dissolution, because G_d was negative, was
recorded in the winter under high pCO₂ conditions. This negative effect of increased pCO₂ was alleviated under elevated
temperature.

3.3. Metabolic responses of dead *L. corallioides* to acidification and warming

225 In dead *L. corallioides*, NPP, GPP, R and G_l were significantly higher in the summer, while no effect of season was observed
on G_d (Table 3). The high temperature condition (+3°C) did not affect dead maerl NPP, GPP or R (Table 5; Fig. 2f-h). The
pCO₂ increase did not affect dead maerl NPP and GPP in either season. However, a decrease in R was observed under high
pCO₂ in the summer. Chlorophyll *a* content did not differ between seasons (Tables 3;6) but was significantly affected by the
temperature and pCO₂ interaction (Table 5; Supplementary material a;f). Dead maerl G_l significantly increased under high
230 temperature (Fig. 2i). Conversely, a negative impact of high pCO₂ was on G_l in the winter and summer. In the dark, net
dissolution was observed on dead maerl regardless of the temperature and pCO₂ conditions (Fig. 2j). No temperature effect
was observed on dark dissolution. However, dark dissolution rates were significantly higher under high pCO₂ treatments,
regardless of the season.

3.4. Growth and metabolic responses of epiphytic algae to acidification and warming

235 Mean GPP and R for the two epiphytic algae *R. ardissoni* and *S. chordalis* measured in the summer are presented in Figure
3. *R. ardissoni* NPP and GPP were not affected by high temperature or pCO₂ conditions, and R was reduced under high

pCO₂ (Table 7; Fig. 3a-c). In *S. chordalis*, NPP and GPP were significantly affected by the interaction between temperature and pCO₂ (Table 7; Fig. 3d,e; Supplementary material c;d). R was enhanced by the high temperature and pCO₂ conditions and their combination resulted in a greater R (Fig. 3f).

240 The mean biomass of epiphytic fleshy algae at the end of the experiment was significantly higher in the summer than in the winter (+81%, t-test, p=0.045, Fig. 4). Epiphyte biomass was not affected by increased temperature or pCO₂ in the winter (2-way ANOVA, p=0.95 and 0.67 respectively); In the summer, increased temperature reduced epiphyte biomass under ambient pCO₂ and stimulated epiphyte biomass under elevated pCO₂ while an interactive effect of temperature and pCO₂ was observed in the summer (2-way ANOVA, p=0.013, supplementary material e).

245 3.5. Metabolic responses of assemblages to acidification and warming

Assemblage metabolism was significantly higher in the summer than in the winter regardless of the metabolic parameter tested (Table 3). No temperature effect was observed on NPP, GPP and R in either season (Table 8; Fig. 5a-c). The high pCO₂ condition enhanced NPP in both seasons. High pCO₂ increased GPP in the summer only. Similarly, R significantly increased under high pCO₂ in summer conditions. G₁ increased under high temperature in the summer only (Fig. 5d).

250 Conversely, high pCO₂ reduced G₁ regardless of the season. In the dark, net dissolution was observed in the winter, but net precipitation occurred in summer conditions at high temperature (Fig. 5e). In the winter, high pCO₂ increased net dissolutions rates, while in the summer G_d increased under elevated temperature.

4. Discussion

The response of marine communities to increased temperature and pCO₂ conditions is likely to be a complex function of
255 direct effects of climate variables on species physiology and shifts in species interactions (Lord et al., 2017). Results show that predicted changes may alter interactions among calcifying and fleshy macroalgae via overgrowth of epiphytic algae and an increase in competition for light and nutrients with underlying maerl. Interactions between grazers and macroalgae were also affected because the grazer physiology was adversely affected by acidification and warming with potential consequences on epiphyte biomass regulation. Our results underscore the importance of examining community-level
260 processes to integrate species interactions in the study of the impact of global change on marine ecosystems. This study also

evidences that seasonal variability represent an important driver influencing the magnitude and the direction of species and community response to climate change.

Assemblage exhibited a strong seasonal pattern for all metabolic parameters, which is consistent with the higher metabolism in the summer for most of the species incubated at the specific scale. This higher metabolism in the summer has already been
265 evidenced in urchins (Brockington and Peck, 2001), gastropods (Davies, 1966; Innes and Houlihan, 1985; Martin et al., 2006b) and living maerl (Potin et al., 1990; Martin et al., 2006a) and is strongly related to changes in numerous environmental and biological variables, such as light intensity and photoperiod, temperature and nutrient or food availability (Godbold and Solan, 2013; Thomsen et al., 2013).

Assemblage GPP and R were not affected by the high temperature and pCO₂ conditions in the winter. Conversely, in the
270 summer, GPP and R increased under high pCO₂ conditions. The response of assemblage GPP and R appeared closely related to changes in epiphyte biomass and productivity. For instance, the biomass of maerl epiphytic algae was significantly higher in summer than in winter, which is consistent with other findings in the Bay of Brest (Guillou et al., 2002) and other Atlantic maerl beds (Peña and Barbara, 2010). The high biomass of epiphytic algae in the summer led to high contribution to oxygen fluxes. Under high pCO₂ conditions, the higher availability of CO₂ as substrate for photosynthesis may stimulate epiphyte
275 productivity and growth (Koch et al., 2013). The two main epiphytic algae that grew during the experiments, *R. ardissoni* and *S. chordalis*, are naturally found in maerl beds in Brittany (Peña et al., 2014). The response of the alga *S. chordalis* to increased temperature and pCO₂ differed from that of *R. ardissoni*. This difference suggests that the response is species-specific, even among fleshy algae, as demonstrated by Kram et al. (2016). *R. ardissoni* GPP was not affected by increased temperature and pCO₂, but its R was significantly lower under high pCO₂. Within the same genus, Cook et al. (1986) showed
280 that *Rhodymenia palmata* can potentially use HCO₃⁻ as source of inorganic carbon for photosynthesis. The same process may occur in *R. ardissoni*, suggesting that this alga is not carbon-limited at current oceanic pCO₂ levels. In contrast to *R. ardissoni*, increased pCO₂ stimulated *S. chordalis* GPP under ambient conditions of temperature. In their study, Short et al. (2014) indicate that the overgrowth of filamentous algae occurs synergistically with high pCO₂ levels and decreased photosynthesis in coralline algae. Here, the stimulation of epiphyte productivity and growth under high pCO₂ is likely to
285 increase the competition with underlying maerl, especially through reduction in incident light.

Although assemblages were mainly composed of living and dead maerl, the response of GPP and R of *L. corallioides* to increased temperature and pCO₂ differed from that observed in assemblages. For example, the temperature increase of +3°C reduced living *L. corallioides* R in the winter, but increased R in the summer. Under high pCO₂ conditions, although CO₂ availability for photosynthesis was higher, no difference was observed in *L. corallioides*, probably due to the ability of this
290 species to employ inorganic carbon acquisition mechanisms (Kübler and Dudgeon, 2015). Interestingly, GPP, R and chlorophyll *a* content of dead maerl were of the same magnitude as for living maerl. Although live algae prevent bio-fouling by shedding their surface layers (Keats et al., 1997; Villas Bôas and Figueiredo, 2004), post-mortem colonization by photosynthetic endolithic assemblages may occur within dead crusts (Diaz-Pulido et al., 2012). Moreover, dead thalli may represent a substrate for the settlement of crustose coralline algae that cover small parts of some thalli. Crustose coralline
295 algae colonization may also contribute to the observed GPP and R values. In dead maerl, only R decreased under high pCO₂, while no effect was detected for GPP.

These findings also suggest the importance of dead maerl to assemblage carbonate fluxes during the experiments. For example, endolithic algae appear to play an important role in the dissolution of a crustose coralline alga (CCA) species, *Porolithon onkodes* (Reyes-Nivia et al., 2014). Through their photosynthesis, endolithic algae may elevate interstitial pH
300 within the *P. onkodes* skeleton (Reyes-Nivia et al., 2013), increasing carbonate cement precipitation (Diaz-Pulido et al., 2014). Within dead *L. corallioides*, the presence of endolithic algae combined with the presence of small patches of CCA on the surface of thalli may explain the calcification rates observed in light and dissolution in dark. Considering the high Mg content in the skeleton of *L. corallioides*, increased pCO₂ likely promotes the dissolution of dead thalli. Alternatively, the increase in dissolution observed in the present study may be associated with a reduction of CCA recruitment over the surface
305 of dead thalli under acidified conditions (Jokiel et al., 2008; Ordoñez et al., 2014). These results are consistent with the negative response to increased pCO₂ observed here in assemblage G_l and G_d values, which appeared strongly related to the response of living maerl calcification rates. The high sensitivity of coralline algae to ocean acidification has already been attributed to their high Mg-calcite content (Morse et al., 2006; Hofmann and Bischof, 2014). In the present study, the pCO₂ increase had adverse consequences on assemblage G_d, both in the winter and summer. In the dark, assemblage R reduced
310 seawater pH by releasing CO₂, and hindered the precipitation of CaCO₃ (Cornwall et al., 2013). Under high pCO₂ conditions,

the combined effect of acidification and assemblage R in the dark is likely to increase the sensitivity of living and dead *L. corallioides* to dissolution (Andersson et al., 2009).

Moreover, as discussed above, the overgrowth of epiphytic algae under high pCO₂ increased assemblage R in the dark. Therefore, the negative effect of ocean acidification on *L. corallioides* G_d would be exacerbated by the presence of epiphytic
315 algae, which promote a decline in pH in the dark. In light, several studies have suggested that moderate growth of fleshy macroalgal communities may reduce the impact of ocean acidification on coralline calcification by reducing the CO₂ concentration of seawater through photosynthesis (Semese et al., 2009; Short et al., 2014). Conversely, other studies evidenced that the overgrowth of epiphytic fleshy algae may shade underlying coralline algae and reduce coralline net calcification rates (Garrabou and Ballesteros, 2000; Martin and Gattuso, 2009). The present findings support this idea,
320 because a decline in assemblage G₁ was observed under high pCO₂ and high epiphyte biomass. Under high pCO₂, the overgrowth of epiphytic fleshy algae induced by ocean acidification in the summer may reduce light, oxygen and nutrient availability for underlying maerl, affecting its primary production and calcification (D'Antonio, 1985; Short et al., 2014). Thus, overgrown maerl would be negatively affected by the direct effect of ocean acidification on calcification rates and indirect effects due to shifts in competition dynamics with fleshy epiphytic algae (Kuffner et al., 2008). However, the
325 response of epiphytic algae is likely to be specie-specific and it appears difficult to generalize the impacts of epiphytic algae on coralline algae.

In regard to the present results, the regulation of epiphyte biomass by grazers appears essential to maintain the proper functioning of maerl bed communities (Guillou et al., 2002). The importance of grazers to control epiphytes growth in aquaria has been evidenced by Jokiel et al. (2008). In mollusks and urchins, several studies have demonstrated a link
330 between feeding rates and other metabolic processes, such as respiration, calcification and excretion (Carr and Bruno, 2013; Navarro et al., 2013; Noisette et al., 2016). In mollusks, a wide range of responses to ocean acidification and warming have been revealed (Gazeau et al., 2013; Parker et al., 2013). The differences in sensitivity of mollusks to ocean acidification depend on several parameters, such as the form of CaCO₃ they precipitate during calcification (Ries et al., 2009), as well as their ability to regulate the acid-base balance (Gutowska et al., 2010). Our results corroborate these studies, given that *G. magus* and *J. exasperatus* responded differently to acidification and warming. Increased temperature and pCO₂ had no effect
335

on *G. magus* with regard to the metabolic functions tested. However, despite the apparent resistance of *G. magus* to the applied changes, other physiological parameters that we did not test here may have been affected, such as feeding rates, somatic growth, enzyme activity or immune response (Parker et al., 2013). The respiration rates of *J. exasperatus* showed a decline under high pCO₂ in the winter. The lower growth of epiphytes and biofilm in winter may reduce the energy available
340 to maintain the metabolism under stressful conditions (Thomsen et al., 2013; Pansch et al., 2014). This reduced energy availability may induce changes in energy partitioning and decrease R under high pCO₂. In the summer, the increased R under high pCO₂ can be attributed to higher food supply, which is likely to increase the resistance of *J. exasperatus* to climate change, as reported for several marine taxa (Ramajo et al., 2016).

Given the relatively high resistance of *G. magus* and *J. exasperatus* to predicted changes, the metabolic response of *P.*
345 *miliaris* appears to have stronger implications on assemblage functioning. For example, *P. miliaris* is considered as one of the main macro-epiphytic grazers on maerl beds in the Bay of Brest (Guillou et al., 2002). During the experiments, *P. miliaris* likely played an important role in the regulation of epiphytic biomass. The response of G₁ to temperature and pCO₂ changes was complex. The interaction between temperature and pCO₂ observed in the summer may cause changes in energy partitioning, thereby inducing a trade-off between metabolic processes at the expense of respiration and excretion (Garilli et
350 al., 2015). However, the effect of temperature and pCO₂ on the calcification of *P. miliaris* must be considered carefully. For instance, urchins defecated carbonate pellets following consumption of maerl thalli. These feces are likely to dissolve during incubation, introducing a bias in the measurement of calcification (Gazeau et al., 2015). In the summer, temperature increase by 3°C reduced *P. miliaris* respiration rates. Moreover, the decrease in excretion under high temperature and pCO₂ conditions was modulated by the interaction between these two factors. Temperature is a major factor affecting physiological
355 processes in ectotherms such as metabolic rates and growth (Kordas et al., 2011). In *P. miliaris*, summer temperatures are likely to exceed the physiological thresholds of organisms, inducing a metabolic decline when maintained at 20°C. Although this decline has only been measured for respiration and excretion, the increase in temperature is also likely to affect sea urchin feeding efficiency (Thomas et al., 2000; Carr and Bruno, 2013). Therefore, the ability of *P. miliaris* to regulate epiphyte biomass may be significantly altered under predicted acidification and warming conditions.

360 In addition to the impact of climate change on grazer-fleshy macroalgae interactions, predicted changes may also considerably alter the interaction between grazers and coralline algae. Asnaghi et al. (2013) demonstrated that the grazing activity by urchins may exacerbate pCO₂ effects on coralline algae. Ocean acidification may alter the structural integrity of coralline algae, increasing its sensitivity to grazing (Johnson and Carpenter, 2012; Ragazzola et al., 2012). Coralline algae may thus be more susceptible to grazing by urchins, which also benefit from a higher carbonate uptake from their diet to
365 modulate their response to ocean acidification (Asnaghi et al., 2013). In *L. corallioides*, the decrease in calcification rates may alter its structural integrity and increase its susceptibility to grazing, especially by urchins, which are considered as important bioeroders of coralline algae in marine ecosystems (Ballesteros, 2006; O'Leary and McClanahan, 2010), particularly in maerl beds (Lawrence, 2013).

In conclusion, the community response to climate change does not appear to be only the result of individual species' metabolic responses, but also strongly depends on shifts in species interactions. In contrast with other studies, which
370 evidenced larger impacts of the combination of increased pCO₂ and temperature than that of these factors alone (Reynaud et al., 2003; Anthony et al., 2008; Martin and Gattuso, 2009; Rodolfo-Metalpa et al., 2010), we showed here that the effects of pCO₂ and temperature on maerl bed communities were weakened when these factors were combined. ~~Under the predicted business as usual conditions, epiphyte overgrowth may exacerbate the negative impact of climate change on underlying coralline algae.~~ Here, we also demonstrated that climate change may affect grazer physiology, with major consequences on their ability to regulate epiphyte biomass. Epiphyte overgrowth is thus likely to adversely affect underlying coralline algae, increasing the shading effect at light and the pH decline in the dark. Climate change may also affect other components that we did not assess in the present study, such as algal palatability and potential changes in grazer trophic behavior (Campbell et al., 2014; Duarte et al., 2015; Poore et al., 2013; Poore et al., 2016). Algal palatability to grazers may also be affected by
380 predicted changes through shifts in the composition and the quantity of allelopathic compounds, as suggested by Del Monaco et al. (2017). In order to better understand the consequences of climate change on ecosystem functioning, further work should focus on the response of marine communities and consider more specifically shifts in species interactions, including changes in trophic interactions between algae and grazers.

Authors' Contributions

385 EL SM PR JG JC designed the experiments; EL SM JC collected the data; EL ML analyzed the data; EL SM PR JG prepared the manuscript with contributions from all co-authors.

Competing interests

The authors declare that they have no conflict of interest.

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630 Table 1. Summary of the four experimental treatments. Two pCO₂ (ambient and high pCO₂) and temperature (ambient and high temperature) conditions were tested. High pCO₂ (H-pCO₂) corresponded to a pH decrease of -0.33 units compared to ambient conditions (A-pCO₂). High temperature (T + 3°C) corresponded to a temperature increase of 3°C compared to ambient conditions (T).

	pCO ₂	Temperature	
1 (Control)	Ambient (A-pCO ₂)	Ambient (T)	A-pCO ₂ ; T
2	High (H-pCO ₂)	Ambient (T)	H-pCO ₂ ; T
3	Ambient (A-pCO ₂)	High (T+3°C)	A-pCO ₂ ; T + 3°C
4	High (H-pCO ₂)	High (T+3°C)	H-pCO ₂ ; T + 3°C

635 Table 2. Physicochemical parameters (mean \pm SE) of seawater in each experimental condition (A-pCO₂ = ambient pCO₂; H-pCO₂ = high pCO₂; T = ambient temperature; T+3°C = high temperature) in the winter and the summer. pH_T and temperature were monitored every two days in each aquarium (n = 35). Total alkalinity values (A_T) are means (\pm SE) of 28 samples measured in each aquarium. The CO₂ partial pressure (pCO₂), dissolved inorganic carbon (DIC), and saturation states of seawater with respect to aragonite (Ω_{Ar}) and calcite (Ω_{Ca}) were calculated from pH_T, temperature, salinity, and A_T using CO2SYS.

	Experimental condition	pCO₂ (μatm)	pH_T	Temperature (°C)	A_T (μmol kg⁻¹)	DIC (μmol kg⁻¹)	Ω_{Ar}	Ω_{Ca}
WINTER	A-pCO ₂ ; T	490 (± 5)	7.97 (± 0.04)	10.1 (± 0.3)	2348 (± 6)	2189 (± 6)	1.84 (± 0.02)	2.89 (± 0.02)
	H-pCO ₂ ; T	1183 (± 10)	7.63 (± 0.03)	10.1 (± 0.3)	2342 (± 7)	2306 (± 7)	0.89 (± 0.01)	1.40 (± 0.01)
	A-pCO ₂ ; T+3°C	513 (± 5)	7.97 (± 0.03)	13.7 (± 0.1)	2341 (± 5)	2166 (± 5)	2.01 (± 0.01)	3.14 (± 0.02)
	H-pCO ₂ ; T+3°C	1087 (± 18)	7.64 (± 0.03)	13.6 (± 0.2)	2329 (± 2)	2266 (± 4)	1.09 (± 0.01)	1.70 (± 0.02)
SUMMER	A-pCO ₂ ; T	426 (± 4)	8.03 (± 0.04)	17.1 (± 0.2)	2359 (± 3)	2127 (± 3)	2.60 (± 0.02)	4.03 (± 0.03)
	H-pCO ₂ ; T	948 (± 9)	7.72 (± 0.03)	17.1 (± 0.2)	2382 (± 4)	2279 (± 4)	1.45 (± 0.01)	2.24 (± 0.02)
	A-pCO ₂ ; T+3°C	432 (± 4)	8.01 (± 0.04)	20.0 (± 0.5)	2364 (± 3)	2109 (± 3)	2.88 (± 0.02)	4.43 (± 0.03)
	H-pCO ₂ ; T+3°C	879 (± 7)	7.74 (± 0.02)	20.2 (± 0.3)	2369 (± 2)	2238 (± 2)	1.71 (± 0.01)	2.64 (± 0.02)

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Table 3. Results of mean comparison tests between seasons for the net and gross primary production, respiration, chlorophyll *a* content, light and dark calcification and excretion of the different species and the assemblages. Statistical analyses were performed using t-tests.

	Net production NPP			Gross production GPP			Respiration R			Chlorophyll a			Light calcification G _l			Dark calcification G _d			Excretion E		
	df	t	p	df	t	p	df	t	p	df	t	p	df	t	p	df	t	p	df	t	p
<i>P. miliaris</i>							24	-11.6	<0.001				24	1.5	0.16				38	3.5	0.001
<i>G. magus</i>							27	-20.6	<0.001				19	5.3	<0.001				29	14.1	<0.001
<i>J. exasperatus</i>							38	0.7	0.46												
Living <i>L. corallioides</i>	38	1.4	0.16	38	5.7	<0.001	38	-12.7	<0.001	38	0.4	0.66	25	8.3	<0.001	38	6.6	<0.001			
Dead <i>L. corallioides</i>	26	7.5	<0.001	24	8.4	<0.001	22	-9.8	<0.001	37	0.9	0.35	38	4.2	<0.001	28	0.3	0.80			
Assemblage	31	-4.5	<0.001	31	6.1	<0.001	37	-13.1	<0.001				38	9.0	<0.001	26	3.3	0.003			

Table 4 Results the analysis of variance testing the effects of temperature (T) and pCO₂ on respiration, net calcification and excretion rates in the urchin *Psammechinus miliaris* and the two gastropods *Gibbula magus* and *Jujubinus exasperatus* for winter and summer experiments (n = 5). Statistical analyses were performed using 2-way crossed ANOVAs and Scheirer-Ray-Hare tests when test assumptions were not respected (in italic). Significant p-values are shown in bold ($\alpha = 0.05$). Degrees of freedom = 1

		Respiration		Net Calcification		Excretion	
		R		G		E	
		F	p	F	p	F	p
<i>Psammechinus miliaris</i>	WINTER						
	T	1.8	0.18	0.3	0.59	0.0	0.89
	pCO ₂	0.0	0.87	1.0	0.34	1.2	0.27
	pCO ₂ x T	0.2	0.67	0.2	0.64	1.6	0.22
	SUMMER						
	T	20.8	<0.001 \searrow	4.8	0.045	7.6	0.014 \searrow
	pCO ₂	3.9	0.07	0.1	0.82	2.7	0.12
pCO ₂ x T	1.4	0.25	6.6	0.022	3.1	0.10	
<i>Gibbula magus</i>	WINTER						
	T	1.1	0.30	0.0	0.90	0.4	0.55
	pCO ₂	4.8	0.043 \searrow	0.1	0.79	0.6	0.44
	pCO ₂ x T	0.0	0.88	3.9	0.07	1.3	0.28
	SUMMER						
	T	0.0	0.93	0.8	0.38	2.4	0.15
	pCO ₂	0.2	0.68	0.6	0.45	1.8	0.20
pCO ₂ x T	0.1	0.72	0.4	0.55	0.1	0.75	
<i>Jujubinus exasperatus</i>	WINTER						
	T	8.6	0.010 \nearrow				
	pCO ₂	5.6	0.031 \searrow				
	pCO ₂ x T	0.8	0.39				
	SUMMER						
	T	0.1	0.75				
	pCO ₂	8.9	0.009 \nearrow				
pCO ₂ x T	0.8	0.83					

Table 5. Results the analysis of variance for the effects of temperature (T) and pCO₂ on net and gross primary production, respiration, chlorophyll *a* content and light and dark calcification rates of living and dead *Lithothamnion corallioides* (n = 5). Statistical analyses were performed using 2-way crossed ANOVAs and Scheirer-Ray-Hare tests when test assumptions were not respected (in italic). Significant p-values are shown in bold ($\alpha = 0.05$). Degrees of freedom = 1

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		Net production NPP		Gross production GPP		Respiration R		Chlorophyll a		Light calcification G _l		Dark calcification G _d	
		F	p	F	p	F	p	F	p	F	p	F	p
LIVING <i>L. corallioides</i>	WINTER												
	T	4.4	0.052	8.0	0.012 ⚡	13.1	0.002 ⚡	5.9	0.027 ⚡	3.6	0.08	10.0	0.006 ⚡
	pCO ₂	0.6	0.44	0.6	0.44	0.1	0.73	1.6	0.23	3.2	0.09	153.3	<0.001 ⚡
	pCO ₂ x T	3.2	0.09	1.0	0.33	3.8	0.07	0.0	0.89	0.8	0.39	3.6	0.08
	SUMMER												
	T	2.0	0.18	<i>1.9</i>	<i>0.17</i>	20.9	<0.001 ⚡	<i>1.5</i>	<i>0.23</i>	3.3	<i>0.07</i>	0.0	0.98
pCO ₂	0.8	0.40	<i>0.4</i>	<i>0.55</i>	0.2	0.70	<i>0.7</i>	<i>0.41</i>	3.6	<i>0.06</i>	50.0	<0.001 ⚡	
pCO ₂ x T	1.5	0.24	<i>0.7</i>	<i>0.41</i>	0.4	0.52	<i>0.6</i>	<i>0.45</i>	0.2	<i>0.65</i>	0.0	0.96	
DEAD <i>L. corallioides</i>	WINTER												
	T	<i>0.0</i>	<i>0.91</i>	<i>0.0</i>	<i>0.97</i>	0.0	0.99	0.0	0.99	20.2	<0.001 ⚡	0.1	0.72
	pCO ₂	<i>0.1</i>	<i>0.71</i>	<i>0.1</i>	<i>0.76</i>	0.1	0.73	0.1	0.81	61.1	<0.001 ⚡	99.6	<0.001 ⚡
	pCO ₂ x T	<i>0.3</i>	<i>0.59</i>	<i>0.0</i>	<i>0.93</i>	0.9	0.35	6.3	0.024	0.0	0.88	0.0	0.85
	SUMMER												
	T	3.7	0.07	3.7	0.07	2.0	0.17	0.7	0.41	<i>0.2</i>	<i>0.65</i>	0.2	0.64
pCO ₂	1.1	0.31	1.8	0.20	4.8	0.043 ⚡	9.9	0.006	9.6	0.002 ⚡	17.8	<0.001 ⚡	
pCO ₂ x T	0.9	0.36	0.8	0.39	0.2	0.67	30.3	<0.001	<i>1.9</i>	<i>0.17</i>	0.6	0.44	

660 Table 6. Chlorophyll *a* content (mean \pm SE) of living and dead *L. corallioides* in the different pCO₂ (A-pCO₂ = ambient pCO₂; H-pCO₂ = high pCO₂) and temperature (T = ambient temperature; T+3°C = high temperature) treatments, after being maintained three months in winter and summer conditions, n = 5

	Chlorophyll <i>a</i>			
	µg chlorophyll g DW⁻¹			
	A-pCO ₂ /T	H-pCO ₂ /T	A-pCO ₂ /T+3°C	H-pCO ₂ /T+3°C
Living <i>L. corallioides</i>				
Winter	59.84 (\pm 1.97)	61.66 (\pm 3.83)	52.93 (\pm 3.44)	56.85 (\pm 2.52)
Summer	55.03 (\pm 2.95)	57.63 (\pm 3.99)	60.35 (\pm 0.70)	62.19 (\pm 3.75)
Dead <i>L. corallioides</i>				
Winter	47.09 (\pm 2.72)	39.39 (\pm 5.65)	39.15 (\pm 2.20)	46.36 (\pm 2.19)
Summer	52.21 (\pm 1.92)	36.30 (\pm 1.83)	43.63 (\pm 0.90)	47.96 (\pm 2.54)

Table 7. Summary of the effects of pCO₂ and temperature (T) and their combined effect on gross production and respiration of the two epiphytic algae *R. ardissonaei* and *S. chordalis* in the summer (n = 5). Statistical analyses were performed using a two-way crossed Scheirer-Ray-Hare test. Significant p-values are presented in bold ($\alpha = 0.05$). Degrees of freedom = 1

		Net production NPP		Gross production GPP		Respiration R	
		F	p-value	F	p-value	F	p-value
<i>Rhodymenia ardissonaei</i>	T	0.8	0.37	0.2	0.68	1.3	0.25
	pCO ₂	0.0	0.96	0.8	0.38	8.6	0.003 ↘
	pCO ₂ x T	1.0	0.31	1.0	0.31	0.7	0.42
<i>Solieria chordalis</i>		F	p-value	F	p-value	F	p-value
	T	0.1	0.76	0.1	0.60	5.5	0.019 ↗
	pCO ₂	3.0	0.082	5.5	0.019	3.9	0.049 ↗
	pCO ₂ x T	5.8	0.016	5.5	0.019	0.0	0.88

Table 8. Results the analysis of variance testing the effects of temperature (T) and pCO₂ on net and gross primary production, respiration and light and dark calcification rates, measured on assemblages (n = 5). Significant p-values are presented in bold ($\alpha = 0.05$). Degrees of freedom = 1

		Net production NPP		Gross production GPP		Respiration R		Light calcification G _l		Dark calcification G _d	
		F	p	F	p	F	p	F	p	F	p
Assemblages	WINTER										
	T	0.2	0.70	0.7	0.43	1.7	0.21	0.2	0.71	0.7	0.43
	pCO ₂	8.6	0.011 ↗	0.1	0.72	1.6	0.23	27.3	<0.001 ↘	65.4	<0.001 ↘
	pCO ₂ x T	0.9	0.35	1.1	0.31	1.1	0.31	0.8	0.37	0.2	0.66
	SUMMER										
	T	2.1	0.17	1.6	0.23	0.5	0.51	40.2	<0.001 ↗	6.8	0.020 ↗
	pCO ₂	8.2	0.011 ↗	14.2	0.002 ↗	11.1	0.004 ↗	16.6	<0.001 ↘	3.0	0.10
	pCO ₂ x T	1.9	0.19	1.3	0.27	0.3	0.60	0.8	0.38	3.6	0.08

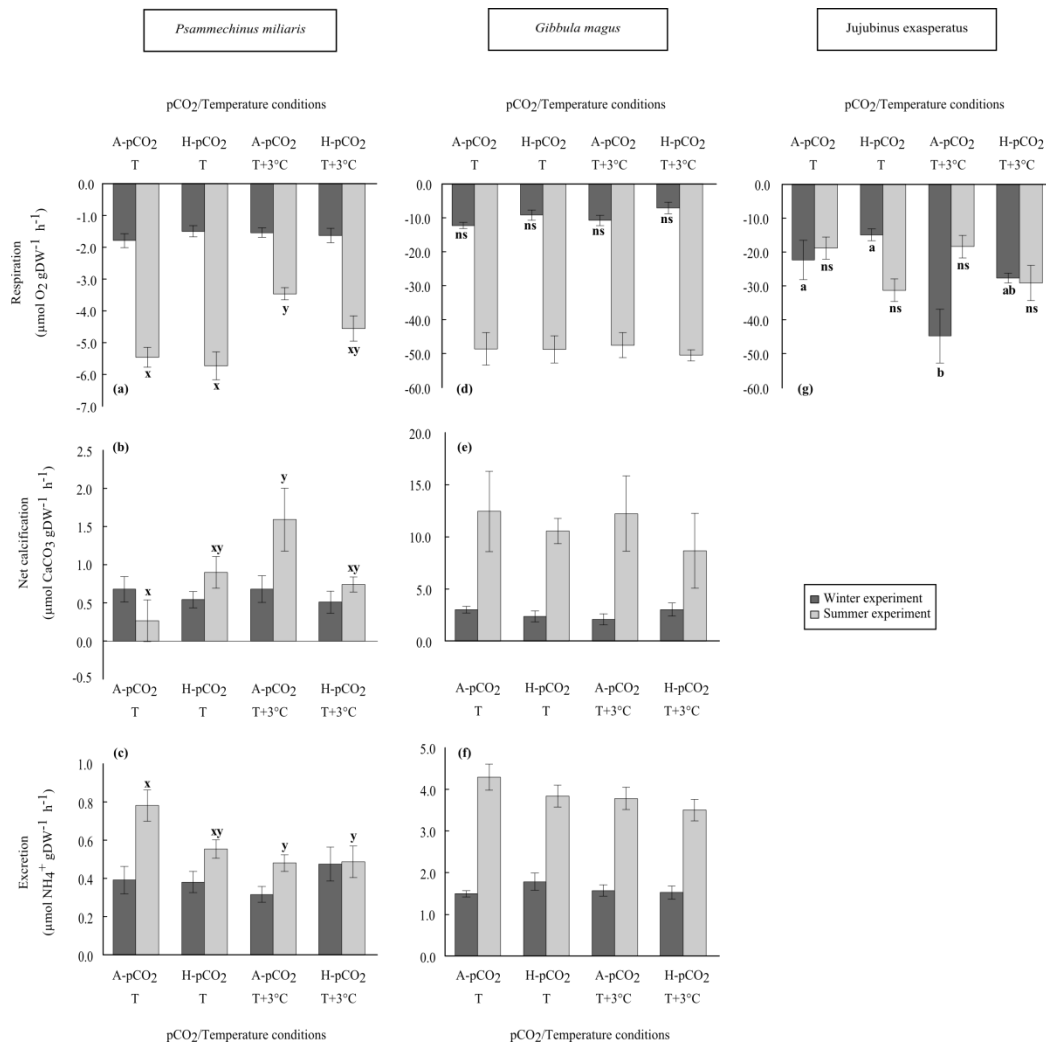


Fig. 1. Respiration, net calcification and excretion rates (mean ± SE, n = 5) of the grazers *P. miliaris* (a to c), *G. magus* (d to f) and respiration of *J. exasperatus* (g) in the different pCO₂ (A-pCO₂ = Ambient pCO₂; H-pCO₂ = High-pCO₂) and temperature (T = Ambient temperature; T+3°C = High temperature) conditions. The species were maintained in assemblages for three months in winter (dark gray) and summer conditions (light gray). Different letters show significant differences (Tukey HSD test) between the four treatments in the winter (letters a and b) and summer (letters x and y). ns = not significant. Tukey tests were performed when a significant effect of temperature or pCO₂ was detected using 2-way ANOVAs.

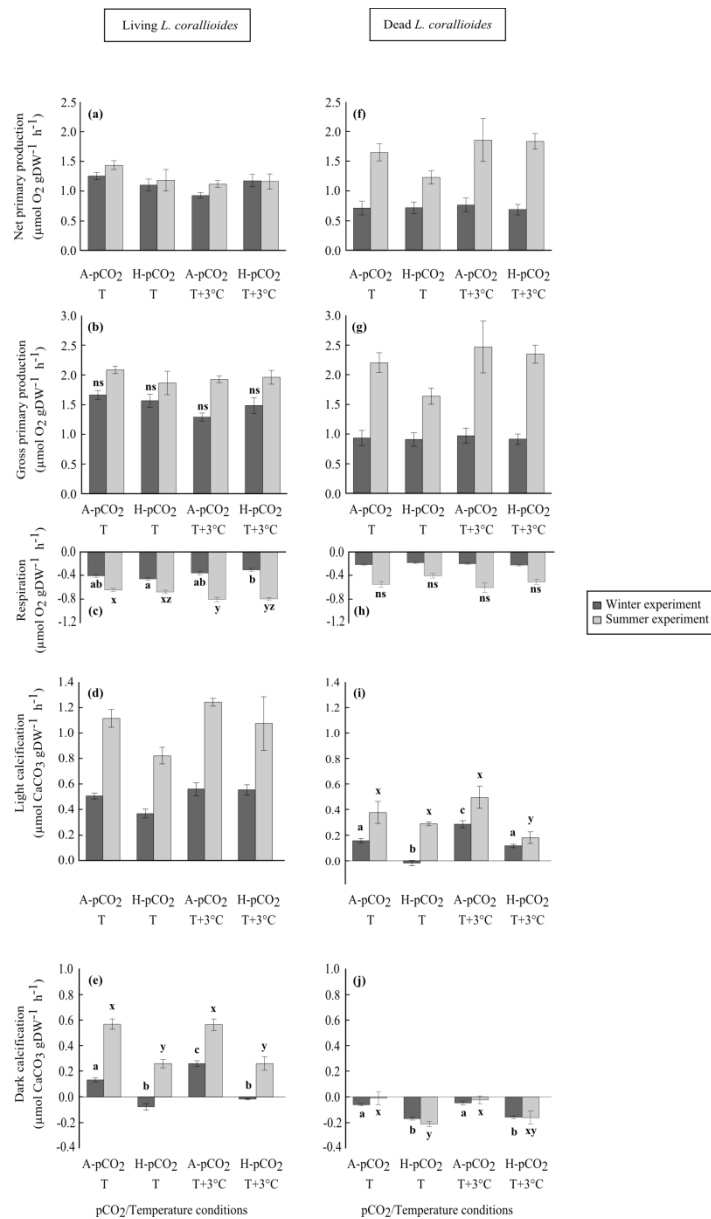


Fig. 2. Net and gross primary production, respiration, light and dark calcification rates (mean \pm SE, n = 5) of living (a to e) and dead thalli (f to j) of *L. corallioides* in the different pCO₂ (A-pCO₂ = Ambient pCO₂; H-pCO₂ = High-pCO₂) and temperature (T = Ambient temperature; T+3°C = High temperature) treatments, after three months in winter (dark gray) and summer conditions (light gray). Letters indicate significant differences between the four treatments in winter (a, b, c) and summer (x, y, z) conditions (Tukey HSD test). ns = not significant. Tukey tests were performed when a significant effect of temperature or pCO₂ was detected using 2-way ANOVAs.

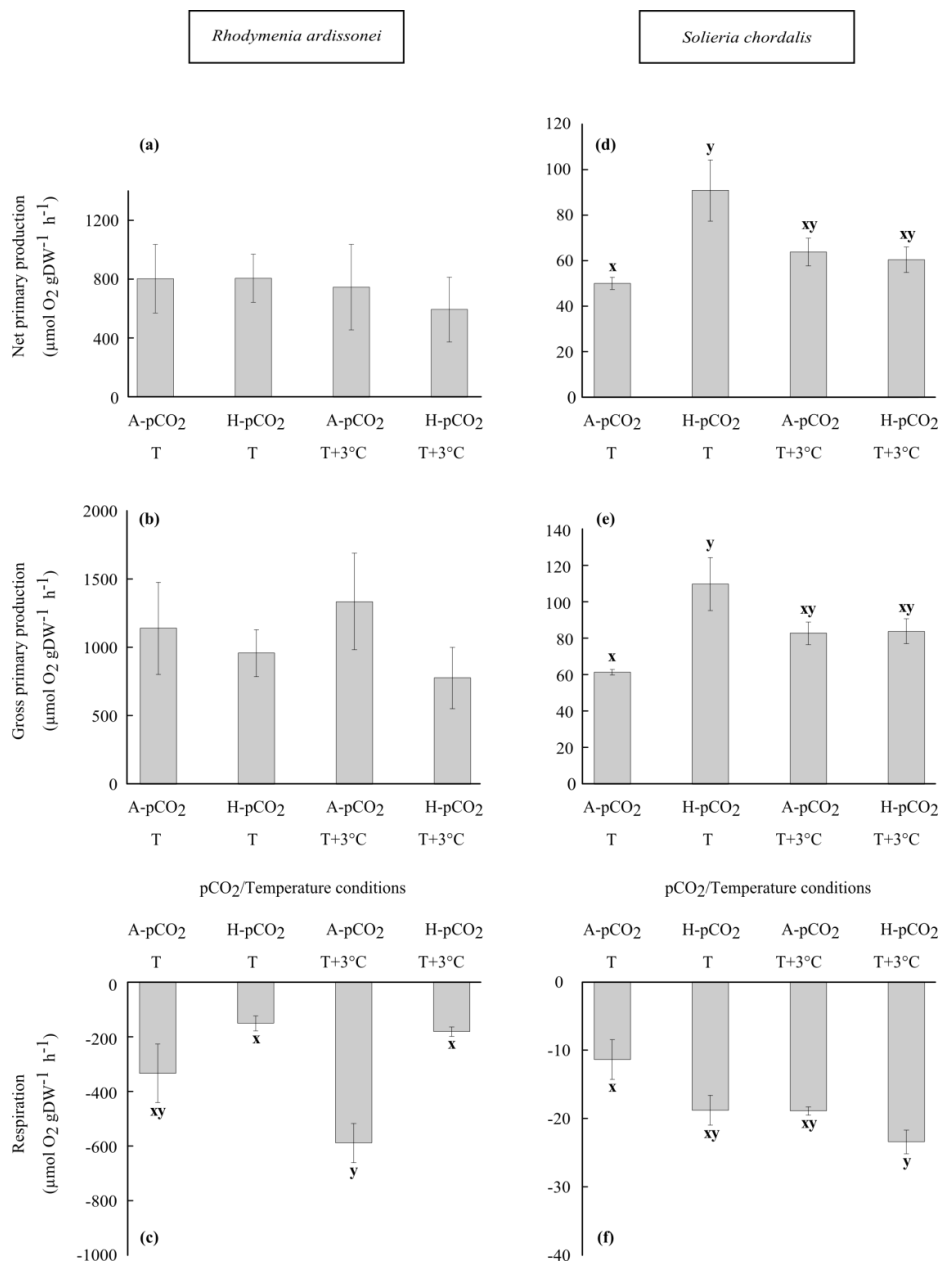


Fig. 3. Summer net and gross primary production and respiration rates (mean \pm SE, n = 5) of the two main epiphytic fleshy algae *Rhodymenia ardissonae* (a to c) and *Solieria chordalis* (d to f), in the different pCO₂ (A-pCO₂ = Ambient pCO₂; H-pCO₂ = High-pCO₂) and temperature (T = Ambient temperature; T+3°C = High temperature) treatments. Letters indicate significant differences between the four treatments in summer (x, y) conditions (Tukey HSD test). Tukey tests were performed when a significant effect of temperature or pCO₂ was detected.

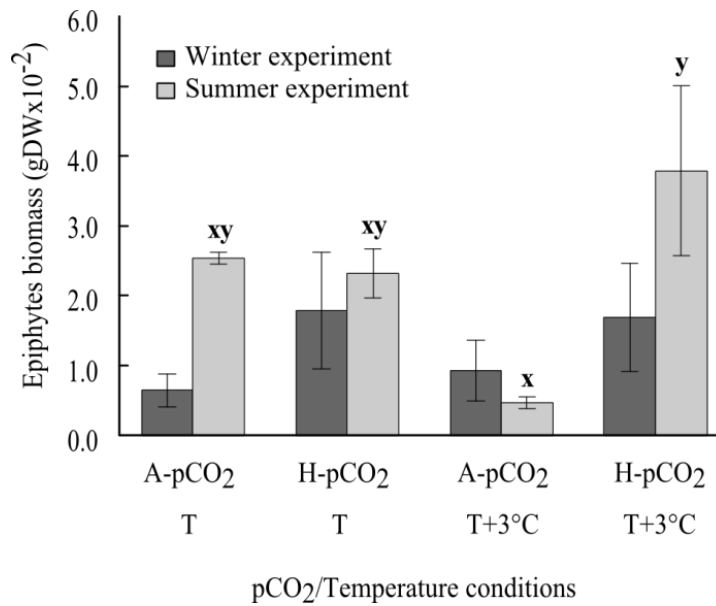


Fig. 4. Biomass of epiphytic fleshy algae (mean \pm SE, n = 5) obtained in the different pCO₂ (A-pCO₂ = Ambient pCO₂; H-pCO₂ = High-pCO₂) and temperature (T = Ambient temperature; T+3°C = High temperature) treatments, after the three-month experiments in winter (dark gray) and summer (light gray) experiments. Letters indicate significant differences between the four treatments (x, y in the summer; Tukey HSD test). Tukey tests were performed when a significant effect of temperature or pCO₂ was detected using 2-way ANOVAs.

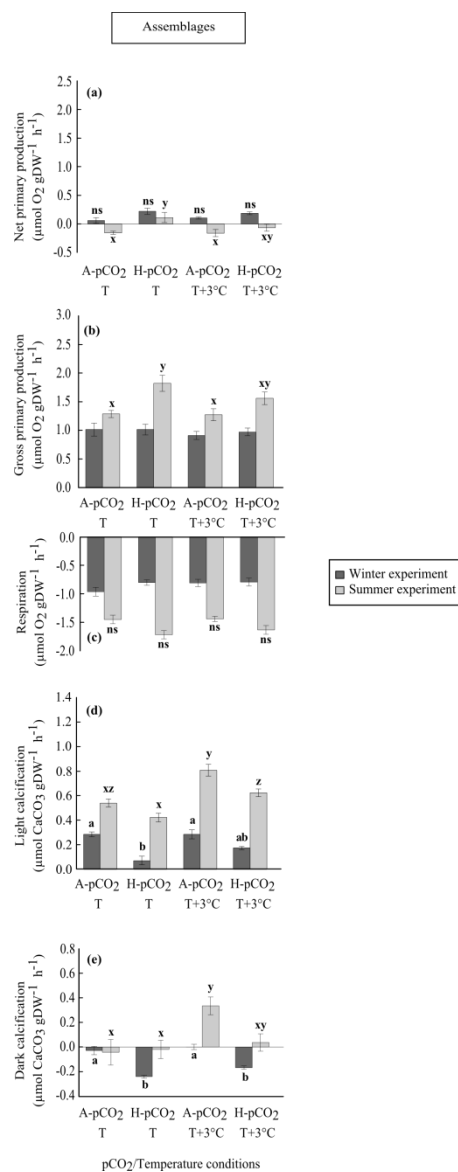


Fig. 5. Net and gross primary production (a and b, respectively), respiration (c) and light and dark calcification rates (d and e, respectively) rates (mean \pm SE, n = 5) of assemblages in the different pCO₂ (A-pCO₂ = Ambient pCO₂; H-pCO₂ = High-pCO₂) and temperature (T = Ambient temperature; T+3°C = High temperature) treatments. The assemblages were maintained during three months in winter (dark gray) and summer conditions (light gray). Letters indicate significant differences between the four treatments in winter (a, b, c) and summer (x, y, z) conditions (Tukey HSD test). ns = not significant. Tukey tests were performed when a significant effect of temperature or pCO₂ was detected using 2-way ANOVAs.