

Dear Emmanuel –

We have revised the manuscript following the reviewers' suggestions. We agree that the clarification based on the reviewer's comments leads to a much improved paper. We attach a version with all changes shown (using track changes in Word) along with the response to reviewers, as well as a version without the track changes.

The responses to the reviewers is almost identical to that already online in the discussion, except that we have now added line number for revisions specific for the track-changes version of the revised manuscript, and corrected a few minor typos and grammatical issues.

In response to the reviews we have added two tables (Table 2 and 3), and added extra panels to Fig 9. We have also redone Fig 4,5,6, 8 and 11 for clarity.

Thank you very much for handling this paper and for earlier comments which also helped in the clarification and suitability to the community.

Yours,

Stephanie Dutkiewicz

We thank the reviewer for this constructive review, and respond to each point below in blue text, and altered text in italics.

This manuscript provides an overview on the how a coupled biogeochemical-ecosystem-optical model can be used to explore ocean colour algorithms, with a focus on Chlorophyll-a. The authors effectively show the kind of interrogation studies that can be done with this type of “virtual laboratory”. They clearly demonstrate how the ocean colour community can explore the bias and uncertainties of algorithms and their products, by investigating the effect of (1) other optically significant materials on derived Chlorophyll-a, and (2) different sized and regionally focused training datasets on robustness of an algorithm. I think this manuscript paves the ground for more detailed studies on the use of a radiative transfer component in a biogeochemical-ecosystem model to investigate ocean colour algorithms. The manuscript is well-written and logically presented, but there are a couple of points where I think a bit more clarity would improve the presentation of the methods & results (see comments).

We thank the reviewer for these positive comments, and especially the understanding of the premise of the paper as a “virtual laboratory”. We hope that this study will allow for others that use such a laboratory for studying ocean colour algorithms and products.

Specific comments:

P2 L20-21: the band-ratio definitely used to be the most commonly used Chl-a algorithm for NASA, but they switched their “default” Chl-a to a merged approach of Hu et al. (2012) and the OCx type algorithms in Reprocessing 2014.0. I am not suggesting you redo your analysis using the band-difference algorithm (because as I understand it, the point in the paper is more to show the kind of analysis you can do with this type of “virtual laboratory”, and dealing with multiple Chl-a algorithms might confuse matters - that being said, it would be an interesting task), but I think it might be worth acknowledging that the OCx algorithms are not the most common for NASA anymore.

Hu et al. (2012), J Geophys. Res., 117(C1). doi: 10.1029/2011jc007395

This is a good point. We had used OC4, because this was what was used on the OC-CCI Chl-a product that we show in Figure 1. We however note that the latest OC-CCI product has switched to a blend of OC3, OC4+CI, OC5. But the improvements in algorithms with new reprocessing of the NASA products is important to acknowledge. We plan to include statements to this effect in the introduction and conclusions of a revised paper. Including a statement that the model might be a good place to explore some of the newer algorithms.

In the introduction (pg 2, line19-30) we now include the following (underlined are added and altered text):

*“There is significant ongoing work to improve algorithms. For instance, the newest National Aeronautics and Space Administration (NASA) reprocessing of Chl-a products has included a merged approach that uses different combination of reflectance bands at low Chl-a (Hu et al., 2012). There have also been many attempts to develop more mechanistically derived algorithms (e.g. using known relationships between absorption, scattering and reflectance). Here we focus on the Chl-a estimated from the blue/green reflectance as it is still the most commonly known product, and until very recently used in products downloaded from both NASA and the European Space Agency (ESA) data portals, as well as merged products such as the Ocean Colour Climate Change Initiative (OC-CCI). However we note that similar techniques used in this paper could help inform on other algorithms. That the satellite-derived*

*products have large errors and specific regional biases is relatively well understood in the ocean colour scientific community (Hu et al., 2000, Moore et al., 2009; Blondeau-Parissier et al., 2014; Szeto et al., 2011). However, there remain many aspects of errors, biases and uncertainties that are poorly quantified, particularly...*

In the conclusion (pg 15, lines 26-29), we finish with:

*“We also hope that the ocean colour community will see the potential of model approaches such as this for deriving sampling strategies, further studies on newer Chl-a algorithms (e.g. NASA Reprocessing 2014.0, and OC-CCI V3 release), other ocean colour products, and will help with algorithm developments for current and future ocean colour measurements.”*

P4 L29-30: While this appears to be true for the January images, it seems to me that the July OC-CCI image (1d) has higher values in the northern high latitudes (around Greenland, Bering Sea, around Scandinavia) than actual July image (1e).

Yes, this is true. The larger values are most notable in the Southern Ocean and in January in the North Pacific. We will make this clearer in the revised version of the paper. We alter this sentence (pg 5, line 7-9) to:

*“As noted (and discussed more fully) in Dutkiewicz et al (2015) there are biases between the model and the observations, in particular larger values in the Southern Ocean and seasonally in the North Pacific than in the real-world satellite-derived Chl-a (Fig 1a,b,d,e).”*

We also are more precise when discussing the model Chl-a (both derived and actual) to OC-CCI at the end of section 3.1 (see below).

P5 L15-27: It is a bit unclear to me which results we are comparing at different points in this paragraph e.g. are the “observations” (L19) the OC-CCI observations? What is the “real world actual Chl-a” (L24)? L19-20: Are you saying the model blue Rrs is too high in the equatorial regions compared to the OC-CCI, coincident with where the model “actual” Chl-a is too low compared to the OC-CCI? Are you meaning OC-CCI is the “real ocean”? Maybe this sentence could be reworded to clarify this.

Indeed this is a difficult section to read (and write). We have tried to rewrite clearer, laying out the terminology first in the introduction. We now refer to the real world OC-CCI Chl-a product, rather suggesting OC-CCI is the “real ocean”. We first have a statement in the introduction (pg 3, line 26-28) to emphasize why we use the term “real-world”:

*“(In this article “real-world” will be used to refer to the real ocean and the real derived ocean colour products that are provide by space agencies. The “real world” is thus different to the numerical biogeochemical/ecosystem/optical model output and the products derived from it.)”*

And have rewritten this Rrs comparison section (pg 5 line 27 to pg 6 line 9) as (underlined is added or altered text):

*“We compare the model output to real world remotely sensed reflectance using the OC-CCI product (Fig 2). We note that the model does not have the exact same wavebands as any of the ocean colour satellites, and as such here we compare to the nearest bands: 450nm model to 443nm for the OC-CCI product, and 550nm model to the 555nm OC-CCI product. The model captures the reversed patterns between blue (443nm/450nm) and green (555nm/550nm)  $R_{RS}$  between gyres and high productive regions. The model blue  $R_{RS}$  (Fig 2a,b,c,d) captures the spatial and seasonal patterns in the real world satellite product. However, the model has lower blue  $R_{RS}$  in the southern Pacific gyre in January. We note though that the model lowest Chl-a in this region is offset from the real-world OC-CCI product (Fig 1a,b). Similarly the model blue  $R_{RS}$  is too high in the equatorial Atlantic and Pacific, but where the model Chl-a is likely too low relative to the real world Chl-a product (see Fig 1). The model has noticeably higher green (550nm)  $R_{RS}$  in the equatorial Atlantic and Indian than the satellite measurements but note that these are regions of high cloud cover where the real world satellite product may be biased. We also find higher green  $R_{RS}$  (Fig 2 e,f,g,h) in the North Pacific, but this might be due to model Chl-a being too high in this region (see Fig 1). In general the differences between model and the real world satellite  $R_{RS}$  appear often to be linked to discrepancies between the model and real world satellite derived Chl-a product (and likely also in situ measurements). The model blue and green  $R_{RS}$  appears to be consistent with the model actual Chl-a fields in a way that is similar to the real world and as such we believe appropriate and useful to use these model remotely sensed reflectance (“model ocean colour”) to construct “satellite-like-derived” Chl-a using the blue to green reflectance ratio algorithm.”*

P7 L4: I think this sentence could be more clearly explained. I think I understand the point you are making: that because the model derived Chl-a compares better with the OC-CCI Chl-a than the model actual Chl-a does, then some of the difference between OC-CCI and model actual Chl-a can be attributed to problems with the band-ratio algorithm (i.e. “product bias”)? Is this what you mean by “product bias” - that there is an intrinsic problem with the band ratio formulation? I think the use of term “model” at the end of this sentence is particularly confusing: often in the ocean colour community, the term “model” is used in terms of a bio-optical proxy/relationship e.g. Chl-a is modelled using the band-ratio. Perhaps use “ecosystem model” (or something similar), to make this distinction clear.


Yes, you understand the point we are trying to make. We have rewritten this section to make this clearer, and in particular added “biogeochemical/ecosystem/optical” to clarify “model”. This difference in use of “model” is important and we try to be careful not to be ambiguous in the paper. In particular we add a sentence to the last paragraph of the introduction (pg 3, lines 29-31) to clarify this:

*“Additionally when we use the word “model” in this article, we refer to the numerical biogeochemical/ecosystem/optical model: In the ocean colour community “model” often refers to bio-optical relationships, we do not use “model” with this definition here.”*

And this section (pg 7, lines 13-31, the last paragraph in Section 3.1) has been rewritten (taking Reviewer 2’s comments into account as well) as (added or altered text underlined):

“Finally in this section, we ask: Which model Chl-a (derived versus actual) best matches real-world OC-CCI product? We do not do this not for model validation purposes (see evaluation in Dutkiewicz et al., 2015), but rather to re-emphasize that the satellite derived Chl-a products are proxies for real world actual Chl-a: The two are not the same thing. We compare climatological monthly model derived Chl-a and model actual Chl-a to OC-CCI monthly climatology regridded to the model configuration (1 degree resolution). We find that the model derived Chl-a has global RMSE of 0.2867mg/m<sup>3</sup>, which is significantly lower than 0.6370mg/m<sup>3</sup> found when comparing model actual Chl-a to OC-CCI. Comparisons are particularly better for the Southern Ocean and North Pacific (Fig 1). Consequently, some (though certainly not all) of the biases noted when comparing model actual Chl-a (Fig 1b,e) to real world satellite derived Chl-a products (Fig 1a,d, section 2 and in the model evaluation done in Dutkiewicz et al. 2015) are due to the real world Chl-a derived product bias and not a deficiency in the biogeochemical/ecosystem/optical model. It follows that a model satellite-like derived products (Fig 1 c,f) might be a better evaluation tool for comparing to ocean colour products derived with the same algorithm (Fig 1a,d) than the model actual Chl-a fields themselves.”

P10 L2-5 (& Appendix B): The exact method is a bit unclear to me here. Did you: take the results of the full run (i.e. those shown in Fig 5a), then take the monthly mean of Rrs output and Chl-a input and derive the 4th order polynomial coefficients on those monthly means? Then, for each of the 3 experiments, did you: do a full run with daily values, take the monthly mean of the model output Rrs and input Chl-a, and use the monthly band-ratio relationship with the monthly Rrs for input? Or did you set up the model with monthly means for the input? Perhaps this could be clarified in the text.

The problem with the sensitivity studies was saving the daily fields for 15 years. So instead we saved off the monthly fields instead. Thus model output for these sensitivity studies was monthly means of Rrs and monthly means of Chl-a. The “default” simulation was rerun saving off the monthly means (rather than the daily means used in Fig 5) so that it would be directly comparable to the sensitivity studies. The 4<sup>th</sup> order polynomials were calculated with these monthly means. We try to explain this clearer in the text now (underlined are altered text). 

In main text we propose to have the following statement (pg 11, lines 20-24) to make this clearer:

“However, given computational and storage constraints we used monthly averaged values of Chl-a and R<sub>RS</sub> to calculate the algorithm coefficients in these experiments rather than daily values (see Appendix B for discussion).”

And for Appendix B (pg 16) we will suggest the following as a change (underlined are the altered text):

“The daily values for 15 years at each grid point creates a very large datafile. Diagnostics with, and storage of, this large dataset becomes extremely computationally expensive. In order to conduct sensitivity studies we found that we needed to reduce this data set. Here we explore only outputting

*monthly means of model  $R_{RS}$  and Chl-a and thus reducing the dataset by 1/30<sup>th</sup>. We determined the algorithm coefficients ( $a_0$  to  $a_4$  in Eq 1) using monthly rather than daily means and subsampling for the GS approach. The resulting function (Fig 4, solid black) is similar at low and intermediate Chl-a, but does deviate at high Chl-a from the algorithm found using daily mean values (light blue line). The  $r^2$  from this algorithm with coefficients defined with monthly means was also not quite as good as that found using daily means (see Table 2 and 3). However we found that the results were similar enough that we could obtain qualitative comparison between sensitivity experiments EXP-1, EXP-2, EXP-3 discussed in Section 5). We also note that the resulting two dimensional histogram (Fig 11) has far lower density when using 4 million relative to 140 million points. Though not perfect, using monthly output does allow us to perform EXP-1 through EXP-3 and still feel confident that the between experiment differences are robust.”*

Fig 11: If I am understanding correctly, Fig 11a is the same as Fig 5a, but 11a uses the monthly coefficients i.e. the black line in Fig 4, whereas 5a uses the light blue line. I think it would be useful to point this out explicitly.

Yes, Fig 11a uses coefficients found using monthly mean Chl-a and  $R_{RS}$ . And yes, the different functions are given by light blue and black lines in Figure 4. We plan to add a further sentence to Fig 11 to make this point. Thus together with the sentences already in the caption we will have the following to clarify this different (underlined are altered text):

*“In these plots, monthly mean output of Chl-a and  $R_{RS}$  were used to calculate the algorithm, and only monthly mean output is shown (4 million versus 140 million points), thus at a great computational savings. The difference in the algorithm is shown in Figure 4 (the light blue line is the algorithm with coefficients found using daily values, versus the solid black line where coefficients were found using monthly values). Differences between 11a and 5a are due to this difference in sampling (discussed in Appendix B). Also notice the difference in values on the colourbars between this figure and Figure 5.”*

P12 L20-21 & L33-P13 L1: I'm not so sure it is quite as simple as this. I agree that the other optically significant materials are contributing to false Chl-a signals: there is a shoulder in all the derived Chl-a time series (Fig 8a), that aligns perfectly with the peak in CDOM and detritus (Fig 8b). But you can see the pattern of the actual Chl-a signal in the derived values, with peaks aligning on around days 60 and 75 - the magnitude of these derived values are just less than the actual values, but I'd say these are the “true peaks” of the spring bloom. After approximately day 75, there is the interference from CDOM and detritus, hence when calculating the initiation of the spring bloom (as described in the appendix), this large “false peak/shoulder” increases the median Chl-a and skews the determined initiation date. So I think what your data could be showing is (1) the Chl-a products do capture the peak of the spring bloom, but the magnitude of that peak is too small, and (2) the CDOM and detritus contribute to a false Chl-a signal, which makes it appear as if the bloom lasts longer and (depending on how you define bloom initiation) makes the initiation date appear to lag compared to the model actual.

We agree that that there is an alignment with the first peak (around day 60), but there is an offset for the maximum peak. Indeed, some of the points we make here are subject to what we define as the “initiation” and “peak”, and we now make this clearer in the revised text: see below.



We use the definitions of Cole et al (2012) and to make our points clearer, we have added extra lines to Fig 8 (see below) to show the timing of the “initiation”. Thus, for this definition of initiation there is indeed a lag between the derived products and the actual Chl-a. (Fig 9)

We add text to clarify this – i.e. acknowledging that the products are capturing the peaks at some times and not others. We also add a figure of the maximum peak offset as an additional panel for Figure 9 (b), as well as for the difference in timings of initiation for CDOM and detrital matter in response to a comment by reviewer 2.

Section 4 (pg 9 and 10) will now read (underlined indicates new or altered text):

“We have noted that in all approaches, though even more obvious in RA, there is a seasonally altering pattern between the derived and actual model Chl-a (Fig 6). The amplitude of the peak of spring blooms is often underestimated in the products derived using global coefficients (GS and GA) in high latitude, especially in the subsampled algorithm (GS) (Figs 6). , Derived Chl-a values were also often higher than model actual Chl-a outside of bloom peaks. We consider the phenology using a single location (in the subpolar North Atlantic) for a single year as illustration (Fig 8a). Though the derived products show similar (though smaller) peaks to the actual Chl-a, and sometimes similar peak timing early in the season (see for instance the first distinct peak in this illustrative location), there are noticeable lags for the maximum peak (shown with a dotted line) and other mismatches later in the season. We also find that the bloom period lasts later into the year. The actual Chl-a also starts its sharp increase in spring (the initiation of the spring bloom, shown with dashed line) considerably before all three derived products (Fig 8a). We follow the approach of Cole et al (2012) for determining the “initiation of the spring bloom” as the time when the Chl-a first increases 5% above the annual median (horizontal dashed line, more description in Appendix A).

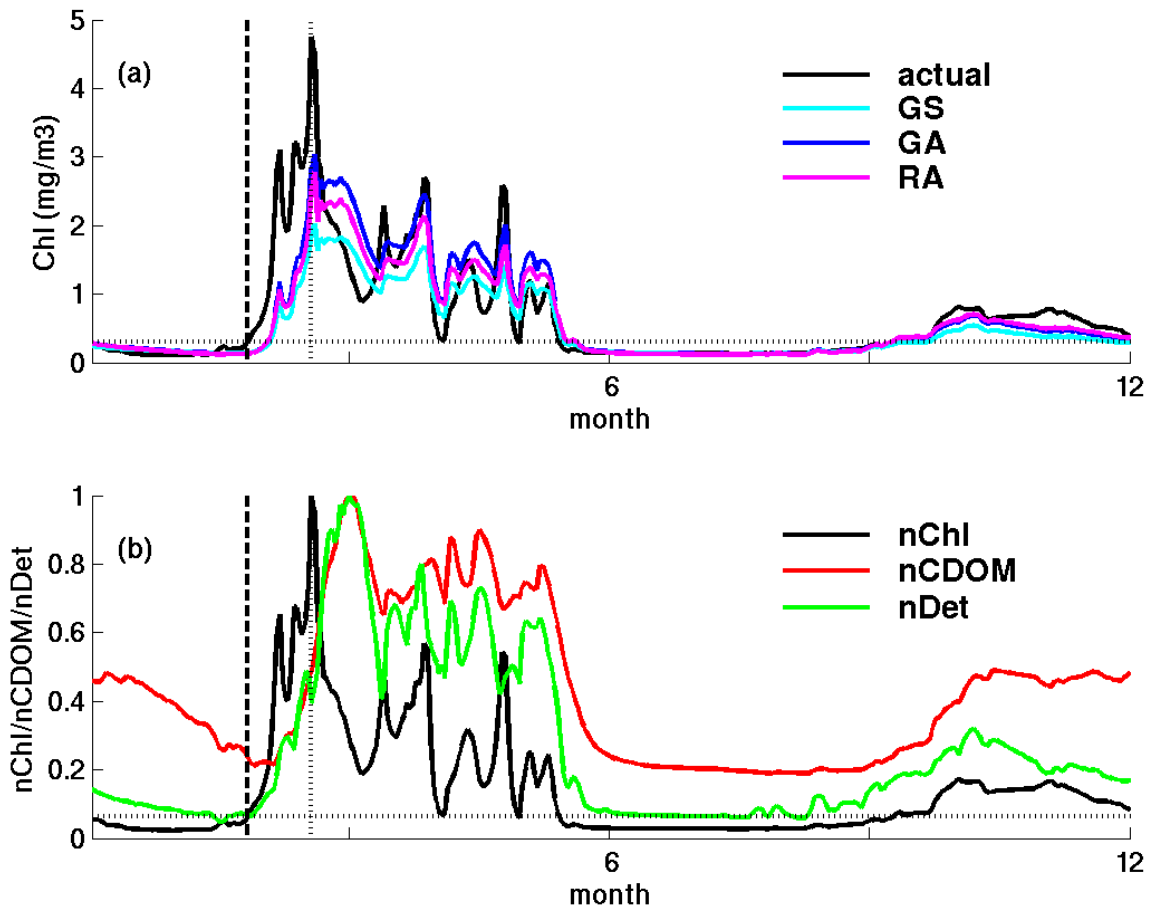
Figure 8 shows just one location for 1 year. To consider the large scale patterns, we determine the lag in the spring initiation (Fig 9a) and maximum bloom timing (Fig 9b) for each location averaged over all years. We find that in almost all locations the derived Chl-a shows the bloom starting later than the model actual Chl-a (Fig 9a). This offset is typically by about 5-10 days but can be as much as 30 days. The maximum Chl-a from the derived product also lags the actual Chl-a in most locations, though by only a few days (Fig 9b). These results indicate that temporal as well as spatial biases occur as a result of deriving Chl-a from X and suggests care should be taken when calculating phenology from satellite products or when evaluating phenology in models using satellite-derived Chl-a. We discuss the reason for the lags in the next section.”

And the following caveats in section 5 (pg 11, lines 5-11):

“We add the caveats that the exact definition of “initiation of bloom” does impact how much of a lag there is in the phenology. For instance, if the first peak in the model actual Chl-a in Figure 8a was defined as “the spring bloom” we would suggest the derived Chl-a does capture the timing better (though not the magnitude). We also note that the model parameterization of CDOM and detrital particle are not necessarily sufficiently well developed to make quantitative statements on the likely real-world lags. Thus, though we do suggest there could be significant lags in phenology in the real world satellite Chl-a product, we do not suggest that the values in Figure 9 are necessarily accurate for the real world. This analysis should instead be seen as a cautionary statement about using satellite-derived products for phenology of the quantities for which they are proxies.”

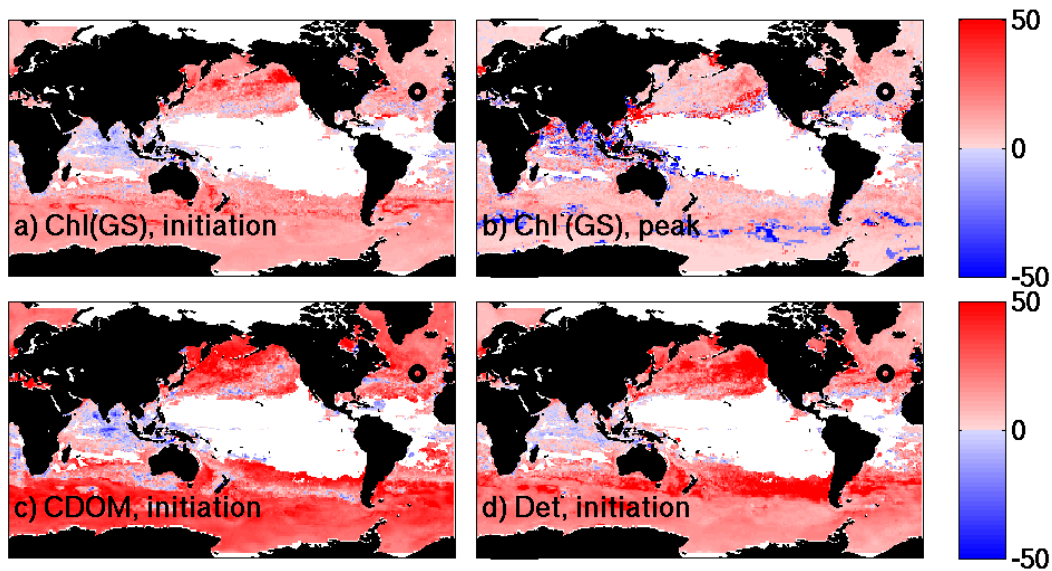
And more-over are careful in the rest of the text to discuss “phenology” rather than “spring bloom” such that the role of definitions of timing are not as relevant.

New versions of Figure 8 and 9 and their captions (altered of added text underline) are provide here. Extra panels in Figure 9 are added at Reviewer 2’s suggestion.



**Figure 8:** Illustrative example timeseries for one year from a single location in the North Atlantic (shown as  $x$  on Fig 9). (a) “actual” Chl-a (black), derived Chl-a using subsampled output (GS, light blue), derived Chl-a using all output (GA, dark blue), and the Chl-a product derived using a regional specific algorithm (RA, purple). (b) actual Chl-a (black), CDOM (red) and detritus (green), all normalized to their peak value. Dashed vertical line indicates the “initiation of the bloom” which is taken to be when Chl-a reaches 5% above the annual median value following Cole et al (2012) and discussed further in Appendix A (dotted horizontal line shows this value for the model actual Chl-a). The vertical dotted line indicates the peak of the bloom. Shown here is only a single year and location, however for larger scale perspective, the difference in initiation and peak timing between model actual and derived Chl-a averaged over all years are shown for the globe in Figure 9.





*Figure 9: Lag in phenology. Number of days between a) the initiation of the spring bloom from model actual Chl-a and that for the model derived Chl-a (GS); b) yearly maximum of model actual Chl-a and that for the derived Chl-a (GS); c) initiation of the spring bloom from model actual Chl-a and the initiation of the CDOM increase; d) initiation of the spring bloom from model actual Chl-a and the initiation of detrital particle increase. Bloom initiation is defined as when Chl-a, CDOM or detrital particles reach 5% above their annual median value (see Appendix A). White areas indicate regions with no significant seasonal cycle or are not resolved by the model (e.g. Arctic Ocean).*

It might be useful to have a table presenting the numerical results (e.g. log/linear RMSE, absolute % bias, etc.) for each approach in Section 3 and 5, to make it easier to compare the different results.

We now plan to add two tables (Table 2 and 3, shown below), one for each section. We split into two tables, to avoid comparison between the daily and the monthly determined algorithms. We also emphasize this difference in the table 3's caption

	Approach 1: GS	Approach 2: GA	Approach 3: RA
$r^2$ (log space)	0.9088	0.9222	0.9466
RMSE (log space)	0.1599	0.1477	0.1215
$r^2$ (linear space)	0.6014	0.7670	0.8301
RMSE (linear space)	0.4816	0.3682	0.3083
absolute % bias	22%	23%	17%

*Table 2: Results of comparison between model “actual” and model “satellite-like” derived Chl-a for the three algorithm approaches discussed in Section 3. Statistics are calculated for each grid and each day*

over 15 years, except for grid cells and times with low light, very low Chl-a and shallow regions (see text).

	Default	EXP-1: uniform $a_{\text{CDOM}}$	EXP-2: uniform $a_{\text{det}}$	EXP-3: phytoplankton optical same
$r^2$ (log space)	0.8999	0.8742	0.8905	0.9493
RMSE (log space)	0.1678	0.1636	0.1663	0.1208
$r^2$ (linear space)	0.5373	0.6298	0.5991	0.7520
RMSE (linear space)	0.4420	0.3811	0.3962	0.2591
absolute % bias	21%	20%	23%	18%

**Table 3: Results of comparison between model “actual” and model “satellite-like” derived Chl-a for the sensitivity experiments discussed in Section 5. All “satellite-like” derived Chl-a was calculated using the GS approach. “Default” is the full experiment discussed in Section 3, but with monthly  $R_{RS}$  used to calculate the algorithm coefficients. Statistics are calculated for each grid cell and each month over 15 years, except for grid cells and times with low light, very low Chl-a and shallow regions (see text).**

Fig 4 and Fig 8: Legends would make these plots easier to read.

Agreed. We add these in the revised paper (see version of Fig 8 above).

Fig 5 and Fig 11: it would be useful to have a title or text on each graph to show which subplot is which e.g. “(a) GS”

Agreed, we add these in the revised paper.

Fig 8: The thick black line on top of the other time series signals masks some of the detail of the derived Chl-a products, particularly after the first 3 months - could this be represented in a different way? Also, check the axis labels, I think Fig 8b is showing days, not months.

We’ve redone the figure with a thinner black line, placed under the other lines. The resulting figures is clearer (see above). And yes, Fig 8b x-axis was days, we now change so that Fig 8a and 8b have the same x-axis. Thank you for catching this.

Technical Corrections:

P1 L15-16: missing the word “to” i.e. sentence should read “...derived Chl-a to the actual...”

Thank you – added in the revised text.

P1 L25: should be either “These results indicate” or “This result indicates”

Thanks –fixed in the revised text as “These results indicate....”

P9 L29: Should this sentence not end with a question mark? i.e. “. . .community structure)?”

Yes, fixed in revised text.

P11 L23: remove the second “like”

Thanks, done.

P12 L24: build should be built

Yes, thank you. Will fix in revised text.

We thank the reviewer for their comments, and respond to each point below in blue text, and proposed altered text in italics.

The paper ‘Modelling Ocean Colour Derived Chlorophyll-a’ by Dutkiewicz et al. uses a modified version of the MIT general circulation model that incorporates scattering and absorption properties of water, detritus, coloured dissolved organic matter (CDOM) and 9 phytoplankton types. ‘Actual’ chlorophyll-a concentration (Chl-a) is determined by summing the variable chlorophyll-a concentration of the 9 phytoplankton types. Remote sensing spectral reflectance, determined from the resulting modelled upwelling and downwelling irradiances are used in a NASA OCx type algorithm to compute satellite-like ‘derived’ chlorophyll-a concentration which is compared to ‘actual’ chlorophyll-a concentration. Firstly, the model output is used to test assumptions used in the derivation of chlorophyll-a concentration using the standard OCx ratio algorithm. Secondly, bloom initiation timings, determined from the ‘derived’ and ‘actual’ chlorophyll concentrations, are compared. Lastly, the impact that other optically important parameters may have on ‘derived’ chlorophyll concentration is explored. The authors conclude that (a) applying a single set of coefficients in the OCx algorithm globally is not as accurate as applying regionally variable coefficients, (b) there is a temporal mismatch between the initiation of the spring bloom defined using ‘actual’ Chl-a compared with that de-fined using ‘derived’ Chl-a and (c) that this mismatch may be caused by the optical influence of other substances such as CDOM and/or detritus.

I found this paper to be generally well-thought out and well-written. For colleagues who are not regular users of these products, this paper provides important caveats for the use of satellite derived data. For those of us who work with ocean colour derived products more often, the results may not be unsurprising, but it serves as a timely reminder of the limitations associated with satellite derived data.

We thank the reviewer for these positive comments and for recognizing that though some of the results are not surprising to those who are experts in ocean colour products, these results offer a reminder of the limitations. We do believe that for non-experts of ocean colour products, these results will be very informative.

Whilst interesting and worthy of publishing in Biogeosciences, I have some concerns with some of the quite sweeping conclusions that appear to be derived from comparison with a single dataset or single datapoint (referred to in more detail below). In addition, I have made other specific comments that I believe should be addressed be-fore this manuscript is suitable for publication.

The reviewer’s comment relates to Fig 8, and the discussion of the results shown in it for one location and one year. This figure was only supposed to be illustrative, not the main basis for our conclusions. We have significantly rewritten this section to make this obvious, and include several more panels in Fig 9 that show that the inferences we gain from that location are relevant for much of the globe. See below.

## SPECIFIC COMMENTS

P 1, L27 – I am unclear whether the term ‘. . .real world Chl-a. . .’ used here refers to actual in-situ chlorophyll-a concentration or satellite derived chlorophyll-a concentration. The term ‘real’ appears to be used interchangeably throughout the manuscript.

We agree that this incident of the use of “real” was confusing and will change this to be “to real world satellite-derived Chl”. We have also gone through the manuscript being careful how we use “real world” in the text. We also add a sentence in the introduction (pg 3, line 27-29) to define “real world” for the rest of the paper:

*“(In this article “real-world” will be used to refer to the real ocean and the real derived ocean colour products that are provide by space agencies. The “real world” is thus different to the numerical biogeochemical/ecosystem/optical model output and the products derived from it.)”*

P5, L15 – Would it make more sense to swap Figures 1 and 2 so that the comparison of model and OC-CCI reflectance appears first as Fig 1 and is followed by the product comparisons as Fig 2. If this were the case, would there then be anything gained by comparing model actual and derived Chl-a to the OC-CCI Chl-a product? As I understand it, this paper is more about using the model output as a test ground to compare model ‘actual’ to ‘derived’ Chl-a rather than testing how well the model replicates the real world’ values. Just a thought.

We like the “thought” – it would make a cleaner paper not getting into the aspect of evaluating the model. However, one of the points we would like to make is that comparing model “actual” or model “derived” Chl-a to “real-world” satellite derived Chl-a leads to different results. There are therefore important implications of our analysis for model validation (i.e. that comparing model chl-a to real-world satellite-derived Chl-a is not comparing like-for-like), so we feel this comparison is an important part of the paper. We have considerably rewritten the relevant paragraph (last one of section 3.1, pg 7, lines 17-31) to make this point clearer and to emphasis that this is not a “evaluation”, but a demonstration (underlined is added or altered text).

*“Finally in this section, we ask: Which model Chl-a (derived versus actual) best matches real-world OC-CCI product? We do not do this not for model validation purposes (see evaluation in Dutkiewicz et al., 2015), but rather to re-emphasis that the satellite derived Chl-a products are proxies for real world actual Chl-a: The two are not the same thing. We compare climatological monthly model derived Chl-a and model actual Chl-a to OC-CCI monthly climatology regrided to the model configuration (1 degree resolution). We find that the model derived Chl-a has global RMSE of 0.2867mg/m<sup>3</sup>, which is significantly lower than 0.6370mg/m<sup>3</sup> found when comparing model actual Chl-a to OC-CCI. Comparisons are particularly better for the Southern Ocean and North Pacific (Fig 1). Consequently, some (though certainly not all) of the biases noted when comparing model actual Chl-a (Fig 1b,e) to real world satellite derived Chl-a products (Fig 1a,d, section 2 and in the model evaluation done in Dutkiewicz et al. 2015) are due to the real world Chl-a derived product bias and not a deficiency in the biogeochemical/ecosystem/optical model. It follows that a model satellite-like derived products (Fig 1 c,f) might be a better evaluation tool for comparing to ocean colour products derived with the same algorithm (Fig 1a,d) than the model actual Chl-a fields themselves.”*

Though we agree that it would be nice to have figure 2 (reflectance) before figure 1 (product: Chl-a) we could not find a way to do this without complicating the paper. So we have left as is.

P6, L13 – The authors talk about comparing ‘. . .locations and dates similar to those in NOMAD.’ What is their definition of similar?

The model has 1 degree resolution – so we are using the degree box within which the actual in situ measurements are taken. This is unnecessarily confusing though and we now state differently (pg 6, line 27):

*“.. at locations and dates nearest in time and space to those in NOMAD”*

P6, L15 – Again they use ‘similar’ to describe the resulting relationship between model ‘actual’ chlorophyll, model X, and real world in situ observations without really defining what similar means.

Here we had meant to reference Figure 4, where we show the OC4 and OC3M-547 functions. We now change this sentence (pg 6, line 29) to reflect this:

*“The resulting relationship between model blue/green reflectance ratio (X) and Chl-a from subsampling the model (Fig 3a) is similar to that found for real-world algorithms (Fig 4).”*

P6, L20 – The authors make no mention of the discrepancy between model reflectance wavebands (blue – 450 nm, 475 nm or 500 nm, green – 550 nm) and those used in the OC4 (blue – 443 nm, 490 nm or 510 nm, green – 555 nm) or OC3M-547 (blue – 443 nm or 488 nm, green – 547 nm) algorithms when comparing coefficients in Table 1. It might make it clearer to those not familiar with the derivation of these algorithms that they are not comparing like with like.

Yes, thank you. This is an important point and in the revised version we will add the following to the text and to the figure caption.

We will add where we compare the model RRS to OC-CCI Rrs (pg 5, line 27-29):

*“We note that the model does not have the exact same wavebands as any of the ocean colour satellites, and as such here we compare to the nearest bands: 450nm model to 443nm for the OC-CCI product, and 550nm model to the 555nm OC-CCI product.”*

Also when we compare the model GS coefficients to OC4 and OC3-457 (Fig 4 and Table 1) we now add the following sentence (pg 6, line 30 to pg 7, line 1):

*“Some of the differences between real-world and model coefficients is likely to come from the use of different exact bands in the blue and green (e.g. 550nm for model green versus 555nm for OC-CCI).”*

We also add the following to the figure caption in Figure 2:

*“We compare the model wavebands against the nearest OC-CCI wavebands, but note that they are not identical.”*

P7, L1-5 – The authors compare the OC-CCI Chl-a product to model derived Chl-a (although see my second comment). Where are the plots to support the statistics? What monthly climatologies are used to generate these statistics (is it a combination of Jan and Jul or all months?) Over what period are the January and July OC-CCI mean values determined? Are the OC-CCI output averaged to 1 degree by 1 degree similar to the model output? What version of the OC-CCI product is used? Are these OC-CCI products just OCx type output or do they include data from the Hu CI algorithm? The OC-CCI output is just one product. The statement at L5 seems to be quite a bold statement to make when only one product has been compared.

The product from OC-CCI was OC4 without the Hu CI component. We discuss the Hu et al adjustments now in the introduction and text (see comments by Reviewer 1).

The RMSE is determined from daily values over the full time period (not just Jan and Jul) averaged globally. We have regridded OC-CCI onto the same grid as the model for these comparisons.

We would prefer not to add any additional figures, especially for this point. However, we do agree with the reviewers' concern on this section and have considerably rewritten it. We add some additional text (also in the figure caption) to explain the statistics better and state what OC-CCI product we are using. Additionally we add text in both introduction and summary to highlight that OC4 is just one product, and that there are newer (and better) products also available. We have stuck here to the OC4 as it was the one that compared to the version of OC-CCI that we downloaded (a new version (V3) has just been released). We do specifically mention the newer OCx+CI versions. We are not convinced that the statement on L5 is too bold. However we do add a caveat in it.

The paragraph questioned here is provided above for the discussion on Fig 1 above. And we add the following sentence to caption of Figure 1:

*"We use version 2 of the OC-CCI, which uses an OC4 algorithm for determining the Chl-a product, and thus comparable algorithm as used in our model derived Chl-a shown in e,f."*

In the introduction (pg 3, line 19-27) we have the following (underlined is added or altered text):

*"There is significant ongoing work to improve algorithms. For instance, the newest National Aeronautics and Space Administration (NASA) reprocessing of Chl-a products has included a merged approach that uses different combination of reflectance bands at low Chl-a (Hu et al., 2012). There have also been many attempts to develop more mechanistically derived algorithms (e.g. using known relationships between absorption, scattering and reflectance). Here we focus on the Chl-a estimated from the blue/green reflectance as it is still the most commonly known product, and until very recently used in products downloaded from both NASA and the European Space Agency (ESA) data portals, as well as merged products such as the Ocean Colour Climate Change Initiative (OC-CCI). However we note that similar techniques used in this paper could help inform on other algorithms."*



The final sentence of the summary (pg 15, lines 27-30) now reads (underlined is new text):

*“We also hope that the ocean colour community will see the potential of model approaches such as this for deriving sampling strategies, further [studies on newer Chl-a algorithms \(e.g. NASA Reprocessing 2014.0, and OC-CCI V3 release\)](#), other ocean colour products, and will help with algorithm developments for current and future ocean colour measurements.”*

P7, L20 – Perhaps it’s my eyesight but I’m not convinced that Figs (b) and (e) show ‘. . .much lower biases at high latitudes. . .’. (I assume you are comparing Fig 6 (b) and (e) to Fig. 6 (a) and (d))

Yes we were comparing b,e to a,d – will make clearer in the revised version. And we agree that this is not obvious from the figure – the improvement is mostly just at the edge of the data before moving into the white areas. We now restate this sentence (pg 8, line 15-16, underlined is new/altered text):

*“Though there is some improvements in some regions in the higher latitudes, there is actually decrease in skill at lower latitudes (Fig 6b,e compared to a,d). There is in fact a slight increase in the mean % absolute bias (23%) between this and the GS estimates: When transformed into percent errors the increased biases at low Chl-a, low latitude regions become more prominent.”*

P8, L3-5 – Are grid cells with depths less than 1000m also excluded?

Yes, will be stated in the revised version.

P8, L22-24 – These statements appear to be derived from data taken from one point in the North Atlantic. Is this a fair representation of the global pattern or is it just representative of this location?

We use the one location as an illustration. The results shown in figure 9 show that same lag in initiation of the bloom occur across much of the high latitudes. We make clear in the text and Figure 8 caption that this is just an illustration. To further strengthen the relevance of this discussion in space and time, we now include other panels in figure 9 to show the mismatch in the peak timing, as well as in the initiation of increase in CDOM and detrital matter relative to actual Chl-a.

Section 4 (pg 9 and 10) now reads (underlined is altered or added text):

*“We have noted that in all approaches, though even more obvious in RA, there is a seasonally altering pattern between the derived and actual model Chl-a (Fig 6). The amplitude of the peak of spring blooms is often underestimated in the products derived using global coefficients (GS and GA) in high latitude, especially in the subsampled algorithm (GS) (Figs 6). , Derived Chl-a values were also often higher than model actual Chl-a outside of bloom peaks. [We consider the phenology using a single location \(in the subpolar North Atlantic\) for a single year as illustration \(Fig 8a\)](#). Though the derived products show similar (though smaller) peaks to the actual Chl-a, and sometimes similar peak timing early in the season*

(see for instance the first distinct peak in this illustrative location), there are noticeable lags for the maximum peak (shown with a dotted line) and other mismatches later in the season. We also find that the bloom period lasts later into the year. The actual Chl-a also starts its sharp increase in spring (the initiation of the spring bloom, shown with dashed line) considerably before all three derived products (Fig 8a). We follow the approach of Cole et al (2012) for determining the “initiation of the spring bloom” as the time when the Chl-a first increases 5% above the annual median (horizontal dashed line, more description in Appendix A).

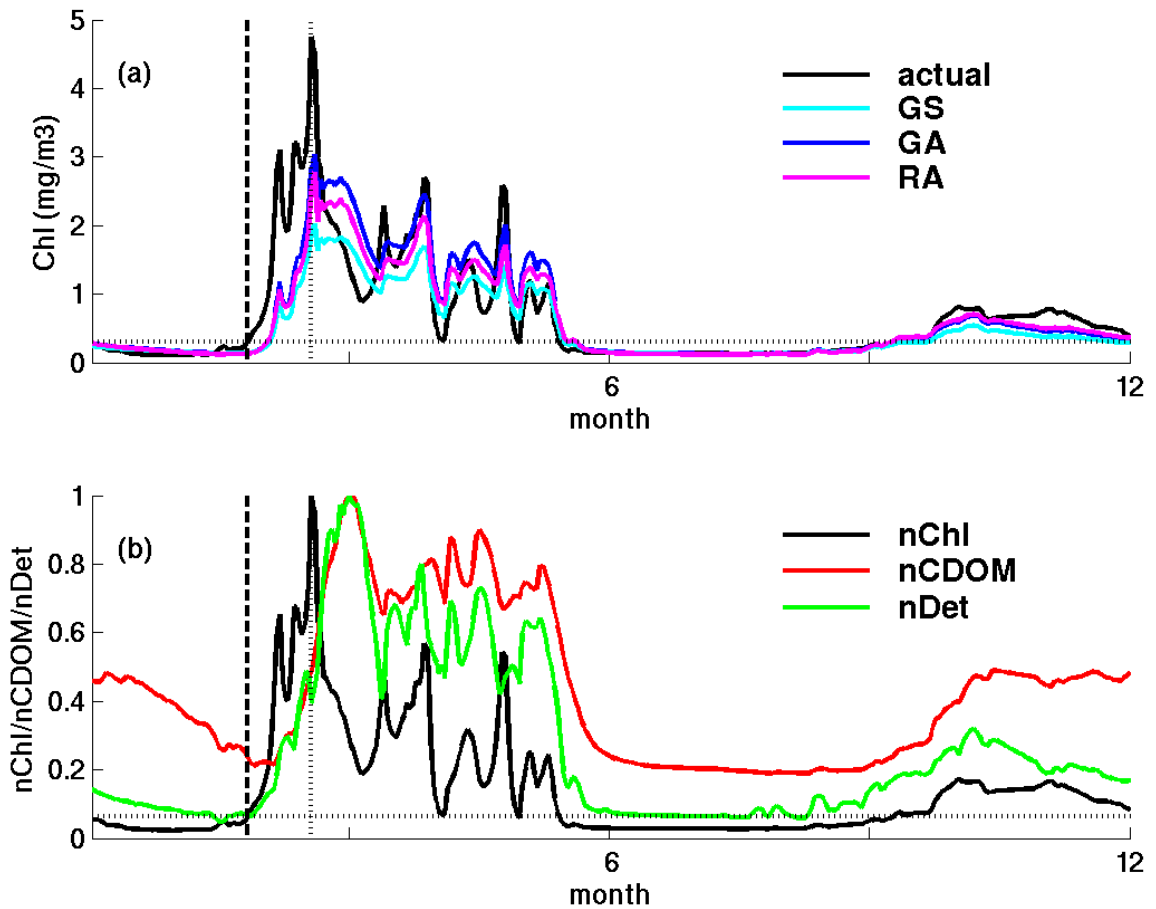
Figure 8 shows just one location for 1 year. To consider the large scale patterns, we determine the lag in the spring initiation (Fig 9a) and maximum bloom timing (Fig 9b) for each location averaged over all years. We find that in almost all locations the derived Chl-a shows the bloom starting later than the model actual Chl-a (Fig 9a). This offset is typically by about 5-10 days but can be as much as 30 days. The maximum Chl-a from the derived product also lags the actual Chl-a in most locations, though by only a few days (Fig 9b). These results indicate that temporal as well as spatial biases occur as a result of deriving Chl-a from X and suggests care should be taken when calculating phenology from satellite products or when evaluating phenology in models using satellite-derived Chl-a. We discuss the reason for the lags in the next section.”

Figure 8 is now more clearly described as just an illustration with the connection to Figure 9 made to make obvious that Fig 9 has the global results. We also add additional lines to Fig 8 to show the initiation of the bloom. We attach the new Fig 8 and 9, and captions below.

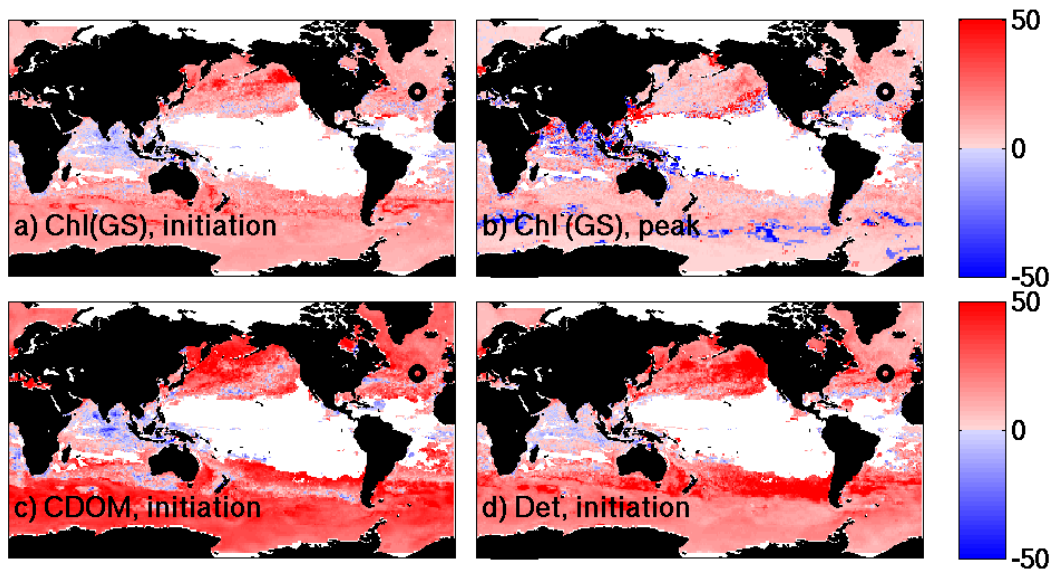
Figure 8: Illustrative example timeseries for one year from a single location in the North Atlantic (shown as x on Fig 9). (a) “actual” Chl-a (black), derived Chl-a using subsampled output (GS, light blue), derived Chl-a using all output (GA, dark blue), and the Chl-a product derived using a regional specific algorithm (RA, purple). (b) actual Chl-a (black), CDOM (red) and detritus (green), all normalized to their peak value. Dashed vertical line indicates the “initiation of the bloom” which is taken to be when Chl-a reaches 5% above the annual median value following Cole et al (2012) and discussed further in Appendix A (dotted horizontal line shows this value for the model actual Chl-a). The vertical dotted line indicates the peak of the bloom. Shown here is only a single year and location, however for larger scale perspective, the difference in initiation and peak timing between model actual and derived Chl-a averaged over all years are shown for the globe in Figure 9.

Figure 8: Illustrative example timeseries for one year from a single location in the North Atlantic (shown as x on Fig 9). (a) “actual” Chl-a (black), derived Chl-a using subsampled output (GS, light blue), derived Chl-a using all output (GA, dark blue), and the Chl-a product derived using a regional specific algorithm (RA, purple). (b) actual Chl-a (black), CDOM (red) and detritus (green), all normalized to their peak value. Dashed vertical line indicates the “initiation of the bloom” which is taken to be when Chl-a reaches 5% above the annual median value following Cole et al (2012) and discussed further in Appendix A (dotted horizontal line shows this value for the model actual Chl-a). The vertical dotted line indicates the peak of the bloom. Shown here is only a single year and location, however for larger scale perspective, the difference in initiation and peak timing between model actual and derived Chl-a averaged over all years are shown for the globe in Figure 9.

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*Figure 8: Illustrative example timeseries for one year from a single location in the North Atlantic (shown as x on Fig 9). (a) “actual” Chl-a (black), derived Chl-a using subsampled output (GS, light blue), derived Chl-a using all output (GA, dark blue), and the Chl-a product derived using a regional specific algorithm (RA, purple). (b) actual Chl-a (black), CDOM (red) and detritus (green), all normalized to their peak value. Dashed vertical line indicates the “initiation of the bloom” which is taken to be when Chl-a reaches 5% above the annual median value following Cole et al (2012) and discussed further in Appendix A (dotted horizontal line shows this value for the model actual Chl-a). The vertical dotted line indicates the peak of the bloom. Shown here is only a single year and location, however for larger scale perspective, the difference in initiation and peak timing between model actual and derived Chl-a averaged over all years are shown for the globe in Figure 9.*



*Figure 9: Lag in phenology. Number of days between a) the initiation of the spring bloom from model actual Chl-a and that for the model derived Chl-a (GS); b) yearly maximum of model actual Chl-a and that for the derived Chl-a (GS); c) initiation of the spring bloom from model actual Chl-a and the initiation of the CDOM increase; d) initiation of the spring bloom from model actual Chl-a and the initiation of detrital particle increase. Bloom initiation is defined as when Chl-a, CDOM or detrital particles reach 5% above their annual median value (see Appendix A). White areas indicate regions with no significant seasonal cycle or are not resolved by the model (e.g. Arctic Ocean).*

P9, L4 – The authors could reference the Dutkiewicz et al. (2015) paper again here.

Good idea, done in the revised version.

P9, L5 – The authors refer to ‘studies’ then reference a single instance.

We add “e.g” to the text and additional references (e.g. Loisel et al., 2010; Brown et al., 2008, Siegel et al., 2005a, 2005b)

Brown, C.A., Huot, Y., Werdell, P.J., Gentili, B., and Claustre, H.: The origin and global distribution of second order variability in satellite ocean color and its potential applications to algorithm development, *Remote Sensing of Environment*, 112, 4186-4203.

Loisel, H., Lebac, B., Dessailly, D., Duforet-Gaurier, L., and Vantrpote, V.: Effect of inherent optical properties variability on chlorophyll retrieval from ocean color remote sensing: an in situ approach. *Optics Express*, 18,

Siegel, D.A., Maritorena, S., Nelson, N.B., and Behrenfeld, M.J.: Independence and interdependencies of global ocean color properties; Reassessing the bio-optical assumption. *J. Geophys. Res.*, 110, C07011, doi:10.1029/2004JC002527, 2005a

Siegel, D.A., Maritorena, S., Nelson, N.B., Behrenfeld, M.J. and McClain, C.R.: Colored dissolved organic matter and its influence on the satellite-based characterization of the ocean biosphere, *Geophys. Res. Letters*, 32, L20605, doi:10.1029/2005GL024310, 2005b.

P9, L17-21 How do the authors support this statement? If it is the timeseries data in Figure 8, then these are data from just one point in the North Atlantic. I don't think that data from one location and for one year is sufficient to warrant these conclusions.

See our comments above for P8, L22-24 above. The results in Figure 9 show the global results averaged over 13 years. However we do see the reviewers point, and now add the extra panels (c,d) to Figure 9. We modify the text in Section 5 (original P9, L17-19) as such (underlined is added or altered text):

*“However, we find that though linked, there are noticeable lags in the sharp increase in accumulation (Fig 8b, Fig 9 c,d) and peak timing and decline(Fig 8b) between CDOM and detrital matter and the model actual Chl-a.”*

Figure 3 – How do you differentiate between zero bias and lack of data? Could I suggest that lack of data is coloured differently to zero bias?

I assume you mean Figure 6? Good idea. We will do have done so in the revised paper.

Figure 4 – Not sure whether the figure order works. The first mention of Fig. 4 that I can find occurs on P11 after reference to all the other figures. Again, if the authors are comparing the polynomials it might make it clearer to the reader if they acknowledge that different wavelengths have been used in the derivation in the legend.

Figure 4 is mentioned first on pg 6 (line 22), but we now also reference it earlier in the newer version of the paper. Since two of lines in Figure 4 are those in Figure 3, it does quite naturally belong here.

We add the acknowledgment of different wavelengths in the caption of Figure 4:

***“Note that the algorithms for the model come from band ratio of 425nm/450nm/475nm and 550nm. For the real world algorithms the band ratios are different and specific for the satellite sensor (SeaWifs or MODIS).”***

Figure 8 – I don't think the x axis matches the label in Fig 8 (b). I assume the vertical dotted line marks the peak in 'actual' Chl-a?

Thank you for catching this – we have altered the axis to match Fig 8a. Yes, the vertical dotted line is the peak – we now mention this in the figure caption. We have also added a line to show the initiation of the spring bloom for clarity. See new figure 8 and caption above.

#### TECHNICAL CORRECTIONS

P1, L16 – Should read '. . .Chl-a to the actual. . .'

Thank you – we fixed this.

P1, L25 – Should read 'This result indicates. . .'

Actually, I think should read "These results indicate...", so will change to this instead. Thank you for catching this inconsistency.

P2, L15 – I think this is the first use of the acronym CDOM and so it should be defined here.

Yes, added.

P2, L18 – Should read 'There have been. . .'

Yes, thank you.

P2, L20 – Should read 'product' instead of 'products'

Yes, fixed

P2, L24 repeats L7

We removed the text at L24 from the revised text.

P3, L28 – In situ is italicised here but nowhere else.

Thanks – we removed the italics to be consistent.

P4, L14, 15, 17, 19 - Repeated uses of 'explicit'.

Yes, have removed excessive use of “explicit”. Thank you.

P5, L15 – ‘Fig.2’ is italicized

We removed the italics. Thank you.

P7, L4 – Missing figure number

Corrected.

P7, L19 - Should read ‘. . .lead to a better. . .’

Yes, thank you.

P9, L3 – Should read ‘lead’ rather than ‘leads’

Yes, thank you

P9, L21 – Should read ‘. . .remains relatively high. . .’

Yes, fixed

P9, L29 – Don’t think there should be a comma after ‘pigments’.

Indeed not, we have removed.

P9, L31 – However, I think there should be one after ‘reflectance’.

Yes, have fixed this in revised version.

P12, L22 Should read ‘. . .by-products. . .’

Yes, thank you.



# Modelling Ocean Colour Derived Chlorophyll-a


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**Abstract.** This article provides a proof-of-concept for using a biogeochemical/ecosystem/optical model with radiative transfer component as a laboratory to explore aspects of ocean colour. We focus here on the satellite ocean colour Chlorophyll-a (Chl-a) product provided by the often-used blue/green reflectance ratio algorithm. The model produces output that can be compared directly to the real world ocean colour remotely sensed reflectance. This model output can then be used to produce an ocean colour satellite-like Chl-a product using an algorithm linking the blue versus green reflectance similar to that used for the real world. Given that the model includes complete knowledge of the (model) water constituents, optics and reflectance, we can explore uncertainties and their causes in this proxy for Chl-a (called “derived Chl-a” in this paper). We compare the derived Chl-a to the “actual” model Chl-a field. In the model we find that the mean absolute bias due to the algorithm is 22% between derived and actual Chl-a. The real world algorithm is found using concurrent in situ measurement of Chl-a and radiometry. We ask whether increased in situ measurements to train the algorithm would improve the algorithm, and find a mixed result. There is a global overall improvement, but at the expense of some regions, especially in lower latitudes where the biases increase. We do find that regional specific algorithms provide a significant  improvement. However, in the model, we find that no matter how the algorithm coefficients are found there can be a temporal mismatch between the derived Chl-a and the actual Chl-a. These mismatches stem from temporal decoupling between Chl-a and other optically important water constituents (such as coloured dissolved organic matter and detrital matter). The degree of decoupling differs regionally and over time. For example, in many highly seasonal regions, the timing of initiation and peak of the spring bloom in the derived Chl-a lags the actual Chl-a by days and sometimes weeks. These results indicate care should also be taken when studying phenology through satellite derived products of Chl-a. This study also re-emphasises that ocean colour derived Chl-a is not the same as the real in situ Chl-a. In fact the model derived Chl-a compares better to real world satellite-derived Chl-a- than the model actual Chl-a. Modellers should keep this in mind when evaluating model output with ocean colour Chl-a and in particular when assimilating this product. Our study spans several disciplines: Our goal is to illustrate the use of numerical laboratory that a) helps users of ocean colour, particularly modellers, gain further understanding of the products they use; and b) the ocean colour community could use to explore other ocean colour products, their biases and uncertainties, as well as to aid in future algorithm development.

## 1 Introduction

Satellite ocean colour measurements have allowed the scientific community an unprecedented ability to study phytoplankton on a global scale and at regular and frequent intervals. In particular, ocean colour products have been used extensively to explore seasonal and interannual variability, trends in ocean surface chlorophyll-a (Chl-a), and in climate, biogeochemical and ecological model evaluation. And yet, there remains a large degree of uncertainty in the satellite-derived Chl-a, with estimates ranging from 30% to >50% (Moore et al., 2009). Uncertainties arise from clouds, patchiness, atmospheric correction, measurement errors, as well as the algorithm used to deduce Chl-a. Here we focus on the uncertainty from one of these algorithms.

10 The most commonly used algorithm for estimating Chl-a from ocean colour uses the fact that phytoplankton absorb more in the blue range of the light spectrum than the green. The ratio of amount of blue to green light reflected at the ocean surface at any location therefore supplies information on the concentration of Chl-a. Using datasets of coincident radiometric observations and in situ Chl-a, a 4<sup>th</sup> order polynomial can be constructed to estimate Chl-a from measured blue/green reflectance ratios (e.g. O'Reilly et al, 2000). This empirical algorithm is then used globally with satellite remotely sensed reflectance. The relationship is typically considered robust in open ocean conditions where the optical effects of phytoplankton co-vary with other optical constituents, ~~(including coloured dissolved organic matter (CDOM) and detritus)~~, so called Case-I conditions (Smith and Baker 1978, Morel 1988, O'Reilly et al, 2000). Though even in these waters the error estimate is about 35% (Moore et al., 2009). Uncertainties that arise from the type of algorithm can be attributed to the potential divergence in the relative role of the optically important water constituents (see e.g. Siegel et al 2005b; Brown et al., 2008). There is significant ongoing work to improve algorithms. For instance, the newest National Aeronautics and Space Administration (NASA) reprocessing of Chl-a products has included a merged approach that uses different combination of reflectance bands at low Chl-a (Hu et al., 2012). There have also been many attempts to develop more mechanistically derived algorithms (e.g. using known relationships between absorption, scattering and reflectance), ~~however~~ Here we focus on the Chl-a estimated from the blue/green reflectance as it is still the most commonly known product, and until very recently used in products downloaded from both ~~National Aeronautics and Space Administration (NASA)~~ and the European Space Agency (ESA) data portals, as well as merged products such as the Ocean Colour Climate Change Initiative (OC-CCI). However we note that similar techniques used in this paper could help inform on other algorithms.

That the satellite-derived products have large errors and specific regional biases is relatively well understood in the ocean colour scientific community (Hu et al., 2000, Moore et al., 2009; Blondeau-Parissier et al., 2014; Szeto et al., 2011).

30 ~~Uncertainties arise from cloud cover, patchiness, atmospheric correction, measurement errors, as well as the algorithm used to determine Chl-a from the reflectance measurements.~~ However, there remain many aspects of errors, biases and uncertainties that are poorly quantified, particularly in regions where there are little or no in situ data to compare to the satellite derived



products. Further, many users of ocean colour products whose main expertise are in other arenas (e.g. numerical modellers) are less aware of these issues. Thus though some of our results may not seem especially exciting to an ocean colour expert at first glance, we note these results could be of much interest in an interdisciplinary context.

5 Ocean colour satellite-derived Chl-a is often used as an evaluation tool for numerical models, and has been used for data assimilation (e.g. Gregg, 2008; Ciavatta et al. 2011, 2014; Rousseaux and Gregg, 2012). The likely biases in the Chl-a estimates are often not appreciated by the modelling community: Modellers sometimes mis-interpret mismatches that are actually potentially due to product biases, or worse have tuned their models or assimilated the products to capture the ocean colour derived Chl-a even where it is likely biased. There is also an inherent disconnect between model output and ocean colour products. Most biogeochemical models have a base currency of carbon, some have a dynamically varying phytoplankton Chl:C, very few resolve spectral irradiance, and even fewer resolve reflectance. However, there are some models that have recently incorporated more thorough treatment of the light field (e.g. Gregg and Casey, 2007; Mobley et al, 2009; Dutkiewicz et al, 2015), and some now include aspects such as reflectance or water leaving irradiances that more directly relate to ocean colour (Dutkiewicz et al 2015; Baird et al, 2015; Gregg and Rousseaux, 2017).

15 By resolving variables that are similar to ocean colour measurements (e.g. reflectance) models can be used to help explore uncertainties in ocean colour products and potentially even to aid in algorithm development. Mouw et al. (2012) used diagnosed optical parameters offline using output from a numerical model to provide ocean colour like products such as reflectance. That study isolated the effects of chlorophyll concentration, phytoplankton cell size and size-varying absorption on remotely sensed reflectance. However, it is only recently that models have directly included the treatment of ocean optics to allow for explicitly including diagnostics such as remotely sensed reflectance (e.g. Dutkiewicz et al., 2015; Baird et al., 2016). Here we use one of these models, a global three-dimensional biogeochemical, ecosystem, and radiative transfer numerical model (Dutkiewicz et al., 2015), that can act as a virtual laboratory to explore the connections between satellite derived products and the ecosystem variability that they are attempting to capture. The model resolves sufficient details of the marine ecosystem, water optical constituents as well as explicit upwelling irradiance.

25 We first briefly describe the numerical model (Section 2), before calculating a "satellite-like" derived Chl-a product from the model spectral reflectance output and explore the potential biases that arise between derived and "actual" model Chl-a (Section 3). Here we focus only on the biases due to the choice of algorithm, and not from other uncertainties that arise in the real world Chl-a products. (In this article "real-world" will be used to refer to the real ocean and the real derived ocean colour products that are provide by space agencies. The "real world" is thus different to the numerical biogeochemical/ecosystem/optical model output and the products derived from it. Additionally, when we use the word "model" in this article, we refer to the numerical biogeochemical/ecosystem/optical model: In the ocean colour community "model" often refers to bio-optical relationships, we do not use "model" with this definition here.); Section 4 examines the temporal mismatches that occur in the derived product. We specifically explore how other optically important constituents, such as coloured dissolved organic matter (CDOM), detrital particles and accessory pigments limit the performance of the algorithm (Section 5).

This paper provides a proof of concept for using numerical model output to explore uncertainties and biases in information derived from surface ocean colour by specifically considering the potential uncertainties in the frequently used blue/green reflectance ratio algorithm for determining Chl-a. Here, using the knowledge of model “actual” Chl-a, other optically important and reflectance at every location and every day allows us to examine these uncertainties and their causes more completely than is possible in the real world with its limited in situ observations.

## 2 The biogeochemical/ecosystem/optical model: Description and Results

We use a biogeochemical/ecosystem/optical numerical model as configured in Dutkiewicz et al (2015). We provide a brief description of the pertinent features here but refer the reader to that paper for more details, equations, parameter values and evaluation. The model resolves the cycling of carbon, phosphorus, nitrogen silica, iron, and oxygen through inorganic, living, dissolved and particulate organic phases (including CDOM). The biogeochemical and biological tracers are transported and mixed by the MIT general circulation model (MITgcm, Marshall *et al.*, 1997) constrained to be consistent with altimetric and hydrographic observations (the ECCO-GODAE state estimates, Wunsch and Heimbach, 2007). This three dimensional configuration has coarse resolution ( $1^\circ \times 1^\circ$  horizontally) and 23 levels ranging from 10m in the surface to 500m at depth. We resolve 9 phytoplankton functional types (diatoms, other large eukaryotes, coccolithophores, pico-eukaryotes, *Synechococcus*, high and low light *Prochlorococcus*, *Trichodesmium* and unicellular diazotrophs) and two grazers. These phytoplankton types differ in the types of nutrients they require (e.g. diatoms require silica), maximum growth rate, nutrient half saturation constants, sinking rates, and palatability to grazers. The phytoplankton also differ in their spectral absorption and scattering (see Figure 1 in Dutkiewicz et al., 2015) and maximum Chl-a:C. The different scattering and absorption spectra for each functional group incorporate the packaging effect (e.g. diatoms have a flatter absorption spectrum than the pico-phytoplankton), but we note that the model does not incorporate changes in the shape of the absorption or scattering spectra due to temporal photo-acclimation. The phytoplankton have dynamic Chl-a:C ratios that change with light availability, temperature and nutrient stress following Geider et al (1998). Thus the model explicitly resolves the Chl-a content of each of the 9 phytoplankton types as model state variables. The sum of this ~~explicit and~~ dynamic Chl-a across all phytoplankton types will be referred to as model “actual” Chl-a in the rest of this manuscript.

This model also explicitly includes ~~explicit~~ radiative transfer of spectral irradiance in 25nm bands between 400 and 700nm. The three stream (downward direct,  $E_d$ , downward diffuse,  $E_s$ , upwelling,  $E_u$ ) model follows Aas (1987), Ackelson et al (1994), and Gregg (2002), though here it is reduced to a tri-diagonal system that is solved explicitly (Dutkiewicz et al., 2015). The model captures the spectral absorption and scattering properties of water molecules, the 9 phytoplankton types, detritus and CDOM. Irradiance just below the surface of the ocean (direct,  $E_{do}$ , and diffuse,  $E_{so}$ , downward) is provided by the Ocean-Atmosphere Spectral Irradiance Model (OASIM, Gregg and Casey, 2009).

The model was run for 10 years for a recurrent "typical" year and then with interannual forcing from 1992 to 2006. Model output compares well to in situ and satellite-derived biogeochemical and ecosystem observations (Dutkiewicz et al., 2015). In particular the magnitudes and patterns of absorption and scattering of different water constituents are captured along the Atlantic Meridional transect cruise (AMT15), as well as the spectral penetration of irradiance and key aspects of the community structure. Model "actual" Chl-a (the sum of the time varying Chl from each of the 9 phytoplankton types resolved) captures the regional patterns seen in a satellite-derived Chl-a. (Here we use the Ocean Colour Climate Change Initiative (OC-CCI) project [V2](#) product). As noted (and discussed more fully) in Dutkiewicz et al (2015) there are biases between the model and the observations, in particular larger values ~~in the high latitudes~~ [in the Southern Ocean and seasonally in the North Pacific](#) than in the [real-world](#) satellite-derived Chl-a (Fig 1a,b,d,e).

10 The numerical model provides spectral surface upwelling irradiance: output that is similar to measurements made by ocean colour satellites. We calculate model subsurface reflectance for each waveband as the upwelling irradiance just below the surface (all diffuse) divided by the total downward (direct and diffuse) irradiance also just below the surface:

$$R(\lambda, 0^-) = \frac{E_{uz}(\lambda)}{E_{d0z}(\lambda) + E_{s0z}(\lambda)},$$

where the  $z$  in the subscript indicates that the irradiance has been re-computed using OASIM code for a zero solar zenith angle to compare more directly to observed normalized reflectance. Satellite sensor measurement (e.g. NASA and ESA products) have been normalized such that they are projected as if there was zero solar zenith angle.

To compare to satellite products, we first convert from irradiance reflectance to remote sensing reflectance using a bidirectional function  $Q$ :


$$R_{RS}(\lambda, 0^-) = R(\lambda, 0^-) / Q.$$

20 The bidirectional function  $Q$  has values between 3 and 5 sr (Morel et al., 2002) and depends on several variables, including inherent optical properties of the water, wavelength, and solar zenith angles (Morel et al., 2002; Voss et al., 2007). Here for simplicity we assume that  $Q = 3$  sr (see Appendix A for discussion of this assumption and for evidence that the choice of  $Q$  makes little difference to model results). We note that Gregg and Rousseaux (2017) make a similar choice of a constant  $Q$ . Secondly we convert to above surface remotely sensed reflectance using the formula of Lee et al (2002):

$$25 \quad R_{RS}(\lambda, 0^+) = 0.52R_{RS}(\lambda, 0^-) / (1 - 1.7R_{RS}(\lambda, 0^-)).$$

Hereafter we will refer to this quantity as  $R_{RS}$ .

We compare the model output to real world remotely sensed reflectance using the OC-CCI product (Fig 2). [We note that the model does not have the exact same wavebands as any of the ocean colour satellites, and as such here we compare to the nearest bands: 450nm model to 443nm for the OC-CCI product, and 550nm model to the 555nm OC-CCI product.](#) The model captures the reversed patterns between blue (443nm/450nm) and green (555nm/550nm)  $R_{RS}$  between gyres and high productive regions. The model blue  $R_{RS}$  (Fig 2a,b,c,d) captures the spatial and seasonal patterns in the [actual-real world](#) satellite product. However, the ~~model has~~ [re-is](#) lower blue  $R_{RS}$  in the southern Pacific gyre in January. We note though that the model lowest Chl-a in this region is offset from ~~the observations~~ [the real-world OC-CCI product](#) (Fig 1a,b). Similarly the model blue  $R_{RS}$  is

too high in the equatorial Atlantic and Pacific, but where the model Chl-a is likely too low relative to the real ~~ocean~~world Chl-a product (see Fig 1). The model has noticeably higher green (550nm)  $R_{RS}$  in the equatorial Atlantic and Indian than the satellite measurements but note that these are regions of high cloud cover where the real world satellite product may be biased. We also find higher green  $R_{RS}$  (Fig 2 e,f,g,h) in the North Pacific, but this might be due to model Chl-a being too high in this region (see Fig 1). In general the differences between model and the real world satellite  $R_{RS}$  appear often to be linked to discrepancies between the model and real world actual-satellite derived Chl-a product (and likely also in situ measurements). The model blue and green  $R_{RS}$  appears to be consistent with the model actual Chl-a fields in a way that is similar to the real world and as such we believe appropriate and useful to use these model remotely sensed reflectance (“model ocean colour”) to construct “satellite-like-derived” Chl-a using the blue to green reflectance ratio algorithm. 

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### 3 Constructing “satellite-like” derived Chl-a

We follow the blue/green reflectance ratio methods used to derive Chl-a from surface reflectance (e.g. O'Reilly et al., 2000). We first determine the log of the blue/green reflectance ratio:  $X = \log(R_{RSB}/R_{RSG})$ , where  $R_{RSB}$  is the largest of the reflectance at 450nm, 475nm, or 500nm at any location and  $R_{RSG}$  is the reflectance at 550nm. We calculate  $X$  using the daily model output from 1992 to 2006. We exclude any grid locations with daily mean PAR is less than  $15 \mu\text{Ein}/\text{m}^2/\text{s}$  (see Appendix A for explanation of this cutoff), with "actual" Chl-a less than  $0.01 \text{ mg Chl}/\text{m}^3$  or with depths less than 1000m since the coarse resolution model does not adequately resolve coastal dynamics.

The blue/green reflectance ratio method uses a 4th order polynomial such that the derived Chl-a ( $chl_d$ ) is:

$$chl_d = 10^{a_0 + a_1 X + a_2 X^2 + a_3 X^3 + a_4 X^4} \quad \text{Eq 1}$$

20 The key here is to determine the best coefficients  $a_0$  to  $a_4$ . We use a least squares fit to find  $a_0$  to  $a_4$  using three different approaches in our model “virtual laboratory”.

#### 3.1 Approach 1: Global Coefficients using Subsampled Fields (GS)

The first approach follows that used in real-world algorithm development (e.g. OC4 for SeaWiFS and OC-CCI, OC3M-547 for MODIS). The NASA bio-Optical Marine Algorithm Data set (NOMAD, Werdell and Bailey, 2005) was constructed from coincident radiometric observations and phytoplankton pigment data and has been extensively used for satellite derived Chl-a (and other) algorithm development. For direct comparison between real-world and emergent within-model relationships we therefore sub-sample the model "actual" Chl-a and reflectance ratio,  $X$ , at locations and dates similar to nearest in time and space those in NOMAD. The resulting relationship between model blue/green reflectance ratio ( $X$ ) and Chl-a from subsampling the model (Fig 3a) is similar to that observed in the real ocean found for real-world algorithms (Fig 4, Table 1 e.g. Werdell and Bailey, 2005). Some of the differences between real-world and model coefficients is likely to come from the use

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of different exact bands in the blue and green (e.g. 550nm for model green versus 555nm for OC-CCI). We note that this subsampling is highly biased to the low latitudes.

We use a least squares fit to find  $a_0$  to  $a_4$  from this subsampled dataset, the corresponding function is shown with a solid line (Fig 3a). We then used these coefficients and  $X$  from every grid cell of the model to produce a model “satellite-like” derived Chl-a (Fig 1c, 5a) for the entire model output (daily from each grid cell, about 140 million data points). This derived Chl-a is analogous to the real-world satellite-derived Chl-a product (e.g. the OC-CCI product). Differences in coefficients relative to those for real-world algorithms (Table 1) are not large and the function looks very similar to those for the real world (Figure 4). We note that while the model world is an idealised system (and hence differences to real world are to be expected) one advantage is that there are no errors on the properties themselves (in contrast to measurement uncertainties on in situ Chl-a and satellite-derived reflectance in the real world) so the model allows for a more precise interrogation of the algorithm biases by themselves.

The root mean square error (RMSE) between the model derived Chl-a and the model actual Chl-a is  $0.48 \text{ mg Chl/m}^3$  ( $0.1599$  for log transformed output) and has an  $r^2$  of  $0.6014$  in linear space ( $0.9088$  in log transformed data, see Table 2). There are substantial errors at higher Chl-a (Fig 5a), which translate to large biases in the high latitudes (Fig 6a,d). The mean value of the absolute bias for all occasions and times where the derived product could be calculated was 22%, though we find that over 35% of the open ocean points (in space and time) had less than 10% absolute error (Fig 7a). We find that the monthly biases have regionally distinct patterns (Fig 6a,d).

Finally in this section, we ask: Which model Chl-a (derived versus actual) best matches real-world OC-CCI product? We do not do this not for model validation purposes (see evaluation in Dutkiewicz et al., 2015), but rather to re-emphasize that the satellite derived Chl-a products are proxies for real world actual Chl-a: The two are not the same thing. The model satellite-derived product compares better to the OC-CCI Chl-a product than it compares to the model “actual” Chl-a (Fig 1), in particular with lower values in the high latitudes. We compare model climatological monthly model derived Chl-a and model actual Chl-a to OC-CCI monthly climatology regridded to the model configuration (1 degree resolution). We find that the model derived Chl-a has a global RMSE of  $0.21 \text{ mg/m}^3$ , which is significantly lower than  $0.48 \text{ mg/m}^3$  found when comparing model actual Chl-a to OC-CCI. Comparisons are particularly better for the Southern Ocean and North Pacific (Fig 1). Consequently, some (though certainly not all) of the biases noted when comparing model actual Chl-a (Fig 1b,e) to real world satellite derived Chl-a products (Fig 1a,d, section 2 and in the model evaluation done in ) (Dutkiewicz et al. 2015) and discussed in Section 2 are due to the real world Chl-a derived product bias and not a deficiency in the biogeochemical/ecosystem/optical model. It follows that a model satellite-like derived products (Fig 1 c,f) might be a better evaluation tools for comparing to ocean colour products derived with the same algorithm (Fig 1a,d) than the model fields actual Chl-a fields themselves.



### 3.2 Approach 2: Global coefficients using output from all locations (GA).

Secondly, we tested whether a lack of data to train the algorithm leads to some of the large errors in the derived Chl-a. We used model output for every surface grid cell and for each day (about 140 million points) to train the algorithm (Fig 3b). We note this is a purely hypothetical exercise: if one knew the ~~real~~ Chl-a at every point and every day, why would one need to derive the Chl-a from a proxy (X)? However, here we are asking rather: Given almost perfect knowledge of Chl-a and X, what is the best that a global set of coefficients for the algorithm given in Eq 1 can do in capturing the actual Chl-a? In other words, even given perfect training dataset (and in an idealised model virtual world), how good could the global OC4-style algorithm possibly be?

In contrast to sub-sampling the model (approach GS), when the full model output is included the relationship between X and actual Chl-a shows considerably more scatter and reveals a distinct cluster below the main body of points at low Chl-a (Fig 3b, the second “tail” below the main cloud). Although this cluster contains only a minority of points (less than 0.003% of the points), the mismatch is of interest and will be discussed in Section 5. Though the coefficients for the algorithm for the full dataset are different (Table 1), the fit is very similar at low Chl-a, but diverges at intermediate and high Chl-a (see solid and dashed line in Fig 3b). When comparing derived and actual Chl-a the GA coefficients lead to a better  $r^2$  (Fig 5b, Table 2) than were achieved using the subsampled algorithm (GS). Though there is improvements in some regions in the higher latitudes, there is actually decrease in skill at lower latitudes (Fig 6b,e compared to a,d). There are much lower biases at high latitudes but there are also larger biases at lower latitudes (Fig 6b,e). ~~TI~~ There is in fact a slight increase in the mean % absolute bias (23%) between this and the GS estimates: When transformed into percent errors the increased biases at low Chl-a, low latitude regions become more prominent.

Not surprisingly our results suggest that a 4<sup>th</sup> order polynomial with one set of global coefficients will not in fact be able to fit both high and low concentrations accurately, no matter how much “data” is available to train the algorithm. Thus, though getting more in situ data in the ocean will still be beneficial for future algorithm development, the use of a single set of coefficients, derived from an improved in situ dataset, used over the whole globe is not likely to significantly improved biases everywhere.

### 3.3. Approach 3. Regional Coefficients (RA)

Recognizing that waters can have distinct optical properties (Moore et al., 2009; Szeto et al, 2011), there have been several projects to produce regionally distinct algorithms (e.g. Szeto et al., 2011, Johnson et al., 2013, latest release (V3) of the OC-CCI project, <https://www.oceancolour.org>). Here we take this concept to the extreme and construct a set of coefficients for each grid cell in the numerical model. We use the algorithm function as provided in Eq 1 and find the coefficients for each location using output from every day over 15 years. Here, we are testing whether X and Chl-a co-vary over time at each location, as opposed to over both time and space as in the previous two approaches (GS, GA). As with the global algorithm

described in Section 3.1 and 3.2 we exclude any grid locations with daily mean PAR less than  $15 \mu\text{Ein}/\text{m}^2/\text{s}$ , ~~and~~ with "model actual" Chl-a less than  $0.01 \text{ mg Chl}/\text{m}^3$ , and where depths are less than 1000m. We also exclude any grid cells where the output falls outside these cut offs for more than half the year.

The regional specific algorithms provide a better Chl-a product with a significant reduction in the bias (Fig 6c,f),  $r^2$ , and RMSE (Fig 5c, [Table 2](#)). The mean absolute bias of all places and occasions where the derived product can be calculated is 17%, lower than the global approaches (GS, GA), and more than 50% of the model output points having less than 10% error (Fig 7c). Average over the full time period, the regional algorithms at every location performs better than either of the global algorithms. There are, however, still significant seasonal biases (Fig 6 c,f) (discussed more below). Note that the biases switch sign between seasons, such that the annual mean bias is extremely low. There are some locations where at some times in the year there is even less accuracy with regional approaches, as seen by the cloud of points at low derived Chl-a (Fig 5c). This indicates that in some regions Chl-a and X do not vary coherently over time and/or that Chl-a and X do not vary in a similar way to the global relationship (noting that the global relationship incorporates both temporal and spatial variability). In these locations the actual Chl-a at some times in the year is closer to the global 4<sup>th</sup> order polynomial (Eq 1) than the local. We note also that there is still a cloud of points below the main cluster (where derived Chl-a is higher than model actual) as was found for approaches GS and GA (Fig 5).

#### 4. Temporal Considerations

We have noted that in all approaches, though even more obvious in RA, there is a seasonally altering pattern between the derived and actual model Chl-a (Fig 6). The amplitude of the peak of spring blooms is often underestimated in the products derived using global coefficients (GS and GA) in high latitude, especially in the subsampled algorithm (GS) (Figs 6, [8a](#)). Derived Chl-a values were also often higher than model actual Chl-a outside of bloom peaks. We consider the phenology, using a single location (in the subpolar North Atlantic) for a single year as illustration (Fig 8a). Though the derived products show similar (though smaller) peaks to the actual Chl-a, and sometimes similar peak timing early in the season (see for instance the first distinct peak in this illustrative location), there are noticeable lags for the maximum peak (shown with a vertical dotted line) and other mismatches later in the season. We also find that the bloom period lasts later into the year. The actual Chl-a also starts its Also noticeable is that the peak of the spring bloom, as well as the sharp increase in Chl-a, spring (the initiation of the spring bloom, shown with dashed line) are offset from the "actual" Chl-a inconsiderably before all three derived products (Fig 8a). We follow the approach of Cole et al (2012) for determining the "initiation of the spring bloom" as the time when the Chl-a first increases 5% above the annual median (horizontal dashed line, more description in Appendix Figure 8 shows just one location for 1 year. To consider the large scale patterns, we determine the lag in the spring initiation (Fig 9a) and maximum bloom timing (Fig 9b) for each location averaged over all years. Since less noisy than bloom peak timing, we find it more informative to consider how the "initiation of the spring bloom" differs between the actual and derived Chl-a. We follow the approach of Cole et al (2012), described in Appendix A. We find

that in almost all locations the derived Chl-a shows the bloom starting later than the model actual Chl-a (Fig 9a). This offset is typically by about 5-10 days but can be as much as 30 days. The maximum Chl-a from the derived product also lags the actual Chl-a in most locations, though by only a few days (Fig 9b). ~~These~~<sup>is</sup> ~~results~~<sup>indicates</sup> that temporal as well as spatial biases occur as a result of deriving Chl-a from X and suggests care should be taken when calculating phenology from satellite products or when evaluating spring bloom timingsphenology in models using satellite-derived Chl-a. We discuss the reason for ~~the~~<sup>is</sup> ~~lags~~ in the next section.

## 5. The Role of other optically important constituents

Chl-a is not the only optically important constituent in seawater. Phytoplankton have a variety of accessory pigments that lead to large difference in their spectral absorption. Additionally different morphologies and structures leads to a variety of scattering spectra (see e.g. Fig 1 in Dutkiewicz et al., 2015). CDOM and detrital particles also absorb more in the blue than the green. How do these other optically important constituents affect the ability of the blue/green ratio algorithm to accurately estimate “in situ” Chl-a? Studies have indeed suggested that second order variability in ocean colour -derived Chl-a can be tracked to the effect of CDOM and non-algal particles (e.g. Loisel et al, 2010; Brown et al., 2008, Siegel et al, 2005a; 2005b). Here, using the knowledge of all constituents in the default experiment (discussed above) in time and space, we can examine the importance of the optically important constituents on model reflectance more thoroughly than is possible in the real world (albeit in the simplified model ocean) and also perform a series of sensitivity experiments targeting individually the other optically important properties.

There is a close connection between CDOM, detrital matter and Chl-a (Fig 10). In general most model data points lie on a linear line: higher Chl-a is closely linked with higher CDOM and detrital matter. The co-variation between CDOM/detrital matter is however not perfect, as has been noted in the real ocean (see e.g. Bricaud et al., 1981; Kitidis et al., 2006; Morel et al., 2010; Siegel et al., 2005b). In the model output there is significant scatter around the core linear relationship (Fig 10). In particular high CDOM can be associated with a wide range of Chl-a concentrations. On the one hand the co-variability between Chl-a, CDOM and detrital matter might help the reflectance ratio algorithm since all absorb more in the blue than the green.

However, we find that though linked, there are noticeable lags in the sharp increase in accumulation (Fig 8b, Fig 9 c,d) and peak timing ~~and decline between all three~~ (Fig 8b). between CDOM and detrital matter and the model actual Chl-a. Since CDOM and detrital matter are a product of primary production, there is a lag in the high latitude spring between the accumulation and peak of CDOM, detrital matter and Chl-a. It is the lag in the accumulation of the different constituents that causes the algorithms to struggle to get the bloom timingsphenology accurate. Moreover detrital particles also lag in their removal, and CDOM (which has relative long remineralization timescales) remains relatively high throughout summer and fall. This leads to the later decrease in derived Chl-a than actual Chl-a seen in Fig 8a. ~~Thus~~<sup>the</sup> the algorithms also all overestimate the background model actual Chl-a. This result may not seem surprising to those in the ocean colour community. In fact, the



role of CDOM as an independent tracer has led to the suggestion that CDOM could be used to track mixing (Nelson et al., 2010). That there is a difference in timing between peaks and CDOM and Chl-a is also known (see e.g. Fig 8 in Nelson et al., 2013). However to our knowledge this is the first time a numerical model has been used to pull apart the differences in timing of the different constituents and that impact on phenology from an algorithm derived Chl-a product.

5 We add the caveats that the exact definition of “initiation of bloom” does impact how much of a lag there is in the phenology. For instance, if the first peak in the model actual Chl-a in Figure 8a was defined as “the spring bloom” we would suggest the derived Chl-a does capture the timing better (though not the magnitude). We also note that the model parameterization of CDOM and detrital particle are not necessarily sufficiently well developed to make quantitative statements on the likely real-world lags. Thus, though we do suggest there could be significant lags in phenology in the real world satellite Chl-a product,

10 we do not suggest that the values in Figure 9 are necessarily accurate for the real world. This analysis should instead be seen as a cautionary statement about using satellite-derived products for phenology of the quantities for which they are proxies.



That the other optically important constituents lead to a mismatch in derived and actual Chl-a leads us to ask the question: Would the algorithms work better if there was not a variation spatially or temporally in detrital matter, CDOM or accessory pigments (and phytoplankton community structure)? The accessory pigments, and the absorption and scattering spectra differ

15 between phytoplankton types and hence the community structure affects the reflectance ratio. However, how the community structure (co-)varies in relation to total Chl-a, and the corresponding combined effects on reflectance, are complex.

To explore how these other constituents affect the algorithm, we perform three sensitivity experiments. Each experiment is performed similar to the “default” run (a 10 year spin-up, 1992-2006 interannually varying component) and we construct 4<sup>th</sup> order polynomials equivalent to Eq 1 using the subsampling approach (GS) for each experiment and derive Chl-a in each case.

20 However, given computational and storage constraints we used monthly averaged values of Chl-a and R<sub>RS</sub> to calculate the algorithm coefficients in these experiments rather than daily values (see Appendix B for discussion). We compare the results from these experiments (Fig 11, Table 3) to the GS results from the default run (i.e. the Chl-a derived product using subset of the data to find the algorithm coefficients, i.e. most like the real satellite product) also using monthly values for consistency.

25 *a) EXP-1 - aCDOM:* This experiment was the same as the default, but  $a_{CDOM}$  was (artificially) set to uniform constant values, specific for each waveband (e.g. 0.016 m<sup>-1</sup> for 450nm, approximately a globally mean). The constant  $a_{CDOM}$  leads to substantial differences in biogeochemistry and community structure (see Dutkiewicz et al., 2015). We construct the satellite-like Chl-a using the approach explained above (Fig. 11b, compare to 11a). The algorithm derived Chl-a compares better to the model actual Chl-a by some metrics (see Fig 11b), but not all, than they did in the original experiment (“default”) (Fig 11a). Thus the

30 correlation between Chl-a and CDOM can enhance the algorithm in some locations (see improvement at high Chl-a), but not at others (e.g. at low Chl-a, see the cloud above the 1-to1 line). However, most noticeable is the lack of points below the main cluster at low Chl-a that was detailed in all other experiments and all types of algorithm approaches (Fig 5 a,b,c, and Fig 11a,c,d). The fact that this cluster of points does not occur in EXP-1 where there is no variability in  $a_{CDOM}$ , helps explain their origin. CDOM is photo-bleached in the surface waters but has long remineralization timescale at depth (Nelson and Siegel,

2013). As such CDOM concentrations (and hence  $a_{CDOM}$ ) tend to be lower in the surface water and higher at mid-depths (see e.g. Nelson and Siegel, 2013); a pattern that is captured in the model (see Fig 3 in Dutkiewicz et al., 2015). When Chl-a is low in the model during autumn, some deep mixing brings high un-bleached CDOM to the surface. This mixing in the highly seasonal regions is what leads to the cluster of points with lower blue/green reflectance ratio than is typical for the actual Chl-a concentration (in Fig 3b).

b) *EXP-2 -Detrital matter*: Similar to EXP-1 experiment, the absorption and scattering by detrital matter was set (artificially) to a constant mean value over the entire globe in this experiment. Detritus itself continued to vary, but the impact of detritus on the optics was as if it constantly had a concentration of  $0.36 \text{ mmolC/m}^3$ . Biogeography and community structure did change as a consequence of the difference in the irradiance. When we calculate the derived Chl-a in a similar manner as in the default with GS approach, we found the  $r^2$  and RMSE are very similar to the default experiment (Fig 11c). However, as with EXP-1, there are some times/places where the algorithm does better than in an ocean with varying optical signature of detritus (e.g. high Chl-a) and some where they were not. This suggest that detritus and Chl-a have a complimentary effect within the algorithm in many, but not all locations.

c) *EXP-3 - Differences in Phytoplankton Absorption and Scattering*: Finally, to explore the role of differing absorption and scattering properties of the different phytoplankton types, we conduct an experiment where all phytoplankton types were assumed to have the same optical properties: the mean of the different absorption and scattering spectra (see black lines in Fig 1 of Dutkiewicz et al., 2015), and same maximum Chl:C. Thus the phytoplankton are optically identical. The main biogeochemistry was similar between this simulation and default experiment, though there is some re-arrangement of the phytoplankton communities as species specific absorption is important to their competitiveness and biogeography (Hickman et al., 2010; Dutkiewicz et al, 2015). We find a substantially higher  $r^2$  and lower RMSE for the derived Chl-a in this experiment. Thus the accessory pigments lead to a large scatter in the relationship between blue/green reflectance ratios and actual Chl-a, making the algorithm approach less accurate. Whether and how these known differences in absorption and scattering make it possible to differentiate species from optical measurements is a promising area of current research (e.g. IOCCG report 2014).

These experiments illustrate how variability in the different optical constituents in time and space lead to significant inaccuracies in deriving Chl-a from the blue/green reflectance ratio, yet on the other hand correlations also enhance the algorithm in many locations. Some of the results and, in particular the statistics are specific to the choices made for the fixed values of  $a_{CDOM}$  and detrital concentration, as well as the mean spectra chosen for EXP-3. On the other, no matter the choice of  $a_{CDOM}$ , EXP-1 is especially useful in elucidating the role of autumn mixing that can bring high CDOM to the surface and impacting the derived Chl-a signature. This is to our knowledge the first time such interactions and their impacts on satellite-derived products have been illustrated using a global biogeochemical model. We believe that similar experiments will be a useful tool in further studies, especially into exploring the impact of different phytoplankton absorption spectra.

## 6. Discussion and Summary

In this study we have used a global three-dimensional biogeochemical, ecosystem, and radiative transfer numerical model to explore how well the magnitudes and seasonal variability of Chl-a can be captured by a product derived from a reflectance ratio algorithm. The model outputs spectral surface upwelling irradiance that includes the effects of the scattering and absorption of optically important water constituents (phytoplankton, water molecules, CDOM and detrital matter), and as such we calculate a remotely sensed reflectance that compares to actual satellite data. We then construct a frequently used algorithm to calculate a "satellite-like" ~~like~~ Chl-a product from the model blue/green reflectance ratio. Given a complete knowledge of all these components and derived products in the model ocean, we can explore the uncertainties and cause more completely than is possible in the real world.

10 When the model algorithm coefficients are calculated from only a subset of data, similar to that which is available in the real world (e.g. NOMAD, Werdell and Bailey, 2005), the resulting function is similar to those used for SeaWiFs and MODIS Chl-a products (Table 1, Fig 4). Using this algorithm, the model derived Chl-a underestimates the actual Chl-a at high latitudes (Fig. 6a,d). Overall the algorithm has a mean absolute bias of 22% in capturing the actual Chl-a, but more than 35% of the model output have less than a 10% absolute error (Fig 7a). However seasonally the errors can be substantially higher and the biases can shift from positive to negative (Fig 6a,d). This study only considers the errors involved in the algorithm development. It does not explore the other potential errors that arise in the real world, for instance from cloud cover, errors in atmospheric correction, instrument drift, and in situ measurement errors and does not resolve all the complexity in the real-world optical constituents. As such, the errors and biases we calculate are underestimates relative to the real world. However, they do suggest that the error of 35% in ocean colour Chl-a estimates that had been desired (e.g. McClain et al 2006) is theoretically possible in many regions of the ocean.

25 We explored the potential to reduce the error by having a much larger dataset (i.e. knowledge of Chl-a at every location every day) for calculating the coefficients of the reflectance ratio algorithm. Although there was improvement in the bias at high Chl-a concentrations, there was also an increase in % bias at low Chl-a concentrations. It is perhaps not surprising that a single set of coefficients for the whole globe will not produce an accurate Chl-a product, even with much improved coverage of data to train the algorithm.

Several studies have explored regional specific algorithms and have indeed found different sets of coefficients work better for different locations (e.g. Szeto et al. 2011, Johnson et al., 2013). To explore this using the model, we again assumed a large dataset (i.e. knowledge of Chl-a at every location and day) and calculated the coefficients for an algorithm unique to each grid cell of the model. The improvement is large, reducing to 17% absolute bias, and almost 50% of the model output points have less than 10% error (Fig. 7c). Though this result is based on the "best case" scenario, it nonetheless suggests that significant improvements in detecting Chl-a from space will be possible with regional specific algorithms. However seasonal biases (Fig 6c,f) can be quite large (though they do cancel out over the course of the year, such that the annual bias is small). Thus temporal variations in Chl-a and reflectance ratio (X) are also not perfectly captured by a 4<sup>th</sup> order polynomial.

Significantly, there is a mismatch between the timing of the spring bloom between any of the algorithm derived products and the actual Chl-a. In almost all seasonal regions the derived Chl-a products suggest the initiation (and peak) of the spring bloom occur later than the actual model Chl-a. We showed how this mismatch could be explained by the role of other optically important water constituents.

5 Because CDOM and detrital matter are also by-products of primary production and subsequent heterotrophic processes, they vary, at least in the surface ocean, in a similar manner to Chl-a (Fig 10) and have a similar effect on the blue/green reflectance ratio. The blue/green ratio algorithm has these co-variations intrinsically built into it (e.g. Morel 1988, 2009). Previous studies have noted that there are however discrepancies with this approach (Bricaud et al., 1998; Siegel et al, 2005a; Siegel et al., 2005b; Brown et al., 2008). In fact, the differences in how CDOM, and detrital matter absorb and scatter light has been used  
10 in algorithm development (e.g. Sathyendranath et al., 1989; Roesler and Perry, 1995; Maritorena et al, 2002). The largest discrepancies between algorithms that explicitly include or exclude the differences in the optical properties is most noticeable at high latitudes (Siegel et al., 2005b), and CDOM and non-algal particles are noted to be especially important (Brown et al., 2008). Additionally it has been found that there are strong seasonal trends in variability of reflectance and reflectance ratios (Brown et al., 2008). Here, we have used the model to show that indeed when the detrital matter and CDOM are distinctly  
15 decoupled from Chl-a there are stronger mismatches between actual and satellite-derived Chl-a. In particular we find that since CDOM and detrital matter in the surface water accumulate later in the spring than Chl-a (Fig 8b), the derived Chl-a increases and peaks later than the actual Chl-a (Fig 8a). Our model suggest that the timing of the spring bloom can be several days to weeks off when using satellite data to determine phenology (Fig 9).

CDOM can also muddy the signal at other times. CDOM is bleached in the surface waters (and is therefore mostly in low  
20 concentration) but is higher at depth where a long remineralization timescale allows it to accumulate. During the fall when in situ Chl-a is low, deep mixing can bring CDOM to the surface. At these times there is an anti-correlation between Chl-a and CDOM, and as such the algorithms tend to overestimate the Chl-a. This leads to the cloud of points where reflectance is lower than anticipated given the in situ Chl-a (Fig 3b), providing higher derived Chl-a than is actually there (Fig 5a,b,c). We suggest that care should be taken when defining a fall bloom from satellite derived products given the effects of CDOM on reflectance  
25 during deep mixing.

It has been recognised that second order variability in reflectance spectra provides a potential method to determine phytoplankton species from space (e.g. Alvain et al, 2005). Using differences in the absorption and scattering spectra by various phytoplankton types to distinguish them optically is an important topic of research (e.g. IOCCG report 2014, see many techniques cited therein; Werdell et al., 2014; Bracher et al., 2017). Here our model results echo this promising direction in  
30 showing that a large amount of the variability in the reflectance ratio versus Chl-a variability is due to the optical differences in phytoplankton (Fig 11d). In our sensitivity study, this appears in fact to have a larger effect than CDOM or detrital particles. The fact that temporal changes in the shape of the Chl-a specific light absorption and scattering spectra for each phytoplankton type does not vary with photo-acclimation in the model formulation means this result is likely under-estimated, though such



within-type variability is likely to have a small effect on sea surface reflectance compared to differences in spectra between types.

Chl-a derived from the model reflectance compares better to the OC-CCI Chl-a than the model actual Chl-a (Fig 1), with a significantly lower RMSE. These differences can particularly be seen in the high latitudes. This finding serves to highlight that Chl-a and reflectance-derived Chl-a are not the same thing and suggests that modellers should be careful in attempting to compare too strongly with satellite derived Chl-a, especially in high latitudes where mismatches between derived and actual Chl-a are shown to be important. Biases are particularly noticeable in the Southern Ocean, which appears to have a very different optical signature (e.g. Szeto et al. 2011; Johnson et al., 2013). Our model results suggest that deep mixer layers (bringing high CDOM to the surface), a potentially different species composition (e.g. Ward, 2015), and high seasonality (leading to mismatches in timing of the peaks in the different optically important constituents) leads these waters to have very different and seasonally varying optical characteristics.

The results presented here provide a novel assessment of the interactions between optical constituents, their effects on reflectance and derived Chl-a that compliment those that are possible for the real world (e.g. Moore et al. 2009; 2014; OCCI uncertainty products, [http://www.esa-oceancolour-cci.org/?q=webfm\\_send/321](http://www.esa-oceancolour-cci.org/?q=webfm_send/321)). In the model all properties are known precisely everywhere allowing details of how the optical constituents and their interactions impacts reflectance and the derived products. However, the results should be taken in context of a modelling approach. The uncertainty estimates reflect only the subset of constituents and mechanisms resolved in the model and the model does not perfectly capture the Chl-a, optics or the reflectance algorithm. Thus the biases presented here should be considered qualitatively, rather than expecting the exact values and statistic to apply to the real ocean. The mismatch in bloom timing should also be interpreted in this way. It is unlikely that the model captures the correct lag between accumulation of CDOM and detrital matter. Thus the exact number of days that the derived Chl-a product lags the actual Chl-a is likely to be different in the real ocean. However the model captures enough of the real world to give insight into the interpretation of the ocean colour product.

A key motivation for this study was to demonstrate that a biogeochemical/ecosystem/optical model with radiative transfer component can be used as a laboratory to explore aspects of ocean colour. As such this study bridges between disciplines: particularly ocean colour and numerical modelling. We believe that our approach could help modellers understand some of the limitations of ocean colour, something that is often lacking when their expertise is not in satellite measurements. We also hope that the ocean colour community will see the potential of model approaches such as this for deriving sampling strategies, further studies on [newer Chl-a algorithms \(e.g. NASA Reprocessing 2014.0, and OC-CCI V3 release\)](#), ~~other different~~ ocean colour products, and will help with algorithm developments for current and future ocean colour measurements.

## Appendix A. Assumptions and Definitions

**Value of bi-directional factor,  $Q$ :** The bidirectional function  $Q$  has values between 3 and 5 sr (Morel et al., 2002) and depends on several variables, including inherent optical properties of the water, wavelength, and solar zenith angles (Morel et al., 2002; Voss et al., 2007). We calculated model reflectance both with a constant value (3 sr) and with time/space/wavelength varying values calculated from the table of Morel et al. (2002). The differences in the relationships between Chl-a and blue/green reflectance ratio with variable and uniform  $Q$  was almost imperceptible (Fig 4). We used the constant/uniform  $Q$  (similar to that used in Gregg and Rousseaux, 2017) in this paper. However we note that the resulting values would only be slightly different if we had used the variable  $Q$ , and, in particular, the choice of  $Q$  would not have changed the interpretation and implications of our results.

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**PAR cutoff:** Satellite measurements of ocean colour cannot be obtained when irradiance fields are too low. These occasions occur during the winter in high latitudes. To compare better to satellite measurements, we choose to not include model data in similar conditions. We examined where and when satellite  $R_{RS}$  (from OC-CCI) lacked data (due to low light) at the high latitudes, and found that the geographic locations and times matched well to when the OASIM input irradiance fields were less than  $15 \mu\text{mol photons m}^{-2}\text{s}^{-1}$  (see Fig 1,2). We thus used this value as a cutoff for calculating derived Chl-a.

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**Determining the Initiation of the Spring Bloom:** We found that the determining the spring bloom peak was quite noisy such that it was more informative to consider “initiation of the spring bloom”. We therefore compared the timing of the bloom initiation between the actual and derived Chl-a. Following the approach of Cole et al (2012), we first reset each “year” at each grid cell by centring to the peak model actual Chl. We then determine when the model actual Chl-a reaches 5% above the annual median value. We define this as the actual “initiation of the spring bloom”. To determine the lag in the initiation (Fig. 9) we calculated the day that the GS derived Chl-a product (that is closest to the real-world satellite-like derived Chl-a. e.g. OC4-like) reaches 5% of its respective median values.

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## 25 Appendix B. Exploring impact of using monthly means to determine algorithm coefficients

The daily values for 15 years, at each grid point creates a very large datafile. Diagnostics with, and storage of, this large dataset becomes extremely computationally expensive. In order to conduct sensitivity studies we found that we needed to reduce this data set. Here we explore using only outputting monthly means of model ~~output~~ ( $R_{RS}$  and Chl-a) and thus reducing the dataset by  $1/30^{\text{th}}$ . We determined the algorithm coefficients ( $a_0$  to  $a_4$  in Eq 1) using monthly rather than daily means and subsampling for the GS approach. The resulting function (Fig 4, solid black) is similar at low and intermediate ~~of~~ Chl-a, but does deviate at high Chl-a from the algorithm found using daily mean values (light blue line). The  $r^2$  from this algorithm with

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coefficients defined with monthly means was also not quite as good as that found using daily means (Fig 11 see Table 2 and 3). However we found that the results were similar enough that we could obtain qualitative comparison between sensitivity experiments EXP-1, EXP-2, EXP-3 discussed in Section 5). We also note that the resulting two dimensional histogram (Fig 11) has far lower density when using 4 million relative to 140 million points. Though not perfect, using monthly output does allow us to perform EXP-1 through EXP-3 and still feel confident that the between experiment differences are robust.

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	$a_0$	$a_1$	$a_2$	$a_3$	$a_4$
model (GS, subsampled)	0.4507	-2.6040	-1.2876	6.5324	-5.1420
model (GA, all output)	0.6588	-3.2742	0.5860	3.2253	-3.0903
OC4 (SeaWiFS)	0.3272	-2.9940	2.7218	-1.2259	-0.5683
OC3M-547 (MODIS)	0.2424	-2.7423	1.8017	0.0015	-1.2280

**Table 1: Coefficients for model global derived algorithms (Eq 1) and for the SeaWiFS and MODIS default algorithms**

10

	Approach 1: GS	Approach 2: GA	Approach 3: RA
$r^2$ (log space)	0.9088	0.9222	0.9466
RMSE (log space)	0.1599	0.1477	0.1215
$r^2$ (linear space)	0.6014	0.7670	0.8301
RMSE (linear space)	0.4816	0.3682	0.3083
absolute % bias	22%	23%	17%

**Table 2: Results of comparison between model “actual” and model “satellite-like” derived Chl-a for the three algorithm approaches discussed in Section 3. Statistics are calculated for each grid and each day over 15 years, except for grid cells and times with low light, very low Chl-a and shallow regions (see text).**

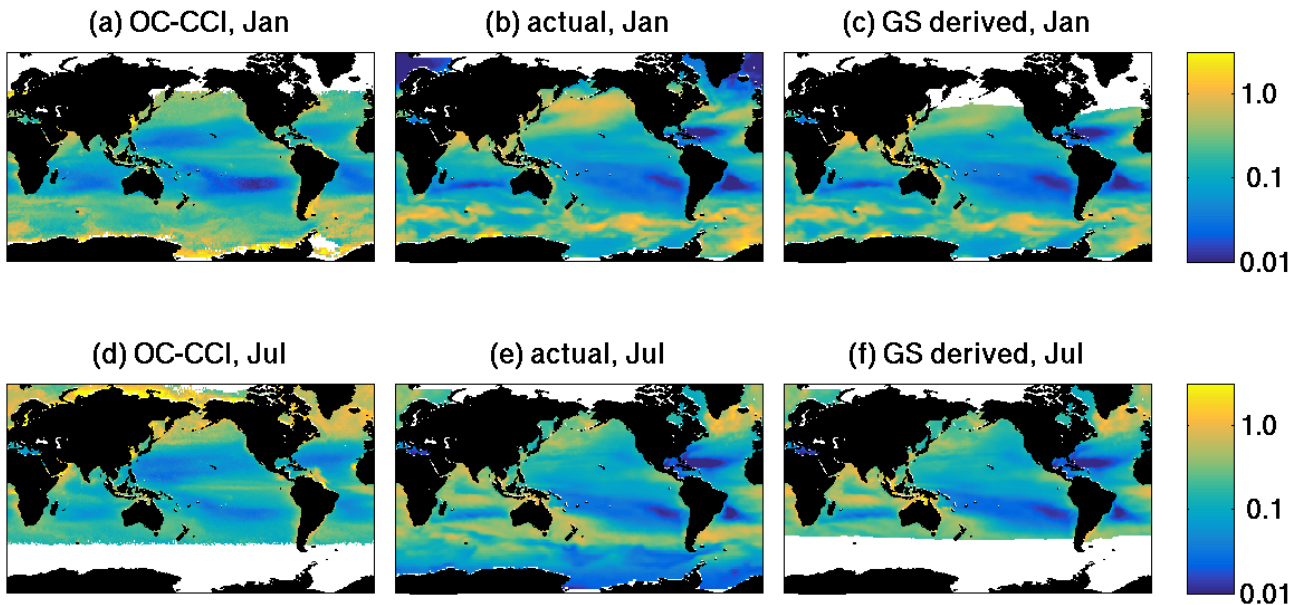
15

	Default	EXP-1: uniform $a_{CDOM}$	EXP-2: uniform $a_{det}$	EXP-3: phytoplankton optical same
$r^2$ (log space)	0.8999	0.8742	0.8905	0.9493
RMSE (log space)	0.1678	0.1636	0.1663	0.1208
$r^2$ (linear space)	0.5373	0.6298	0.5991	0.7520
RMSE (linear space)	0.4420	0.3811	0.3962	0.2591



absolute % bias	21%	20%	23%	18%
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**Table 3: Results of comparison between model “actual” and model “satellite-like” derived Chl-a for the sensitivity experiments discussed in Section 5. All “satellite-like” derived Chl-a was calculated using the GS approach. “Default” is the full experiment discussed in Section 3, but with monthly  $R_{RS}$  used to calculate the algorithm coefficients. Statistics are calculated for each grid cell and each month over 15 years, except for grid cells and times with low light, very low Chl-a and shallow regions (see text).**



**Figure 1: Annual mean Chl-a ( $\text{mg}/\text{m}^3$ ). (a) and (d) OC-CCI-derived; (b) and (e) default model "actual" 0-50m (summed over the 9 phytoplankton types); (c) and (f) default model "derived" (calculated from reflectance ratio and satellite-like algorithm trained with subsampled dataset, GS). Top row are January mean, bottom row are July mean. OC-CCI products (a and d) have no data when irradiances are too low. The model does not resolve the Arctic and thus there is not output here in (b), (c), (e), and (f). Additional lack of output in (c) and (f) indicates regions where PAR is less than  $15 \mu\text{Ein}/\text{m}^2/\text{s}$ . OC-CCI products were downloaded from <https://www.oceancolour.org>. We use version 2 of the OC-CCI, which uses an OC4 algorithm for determining the Chl-a product, and thus comparable algorithm as used in our model derived Chl-a shown in e,f.**

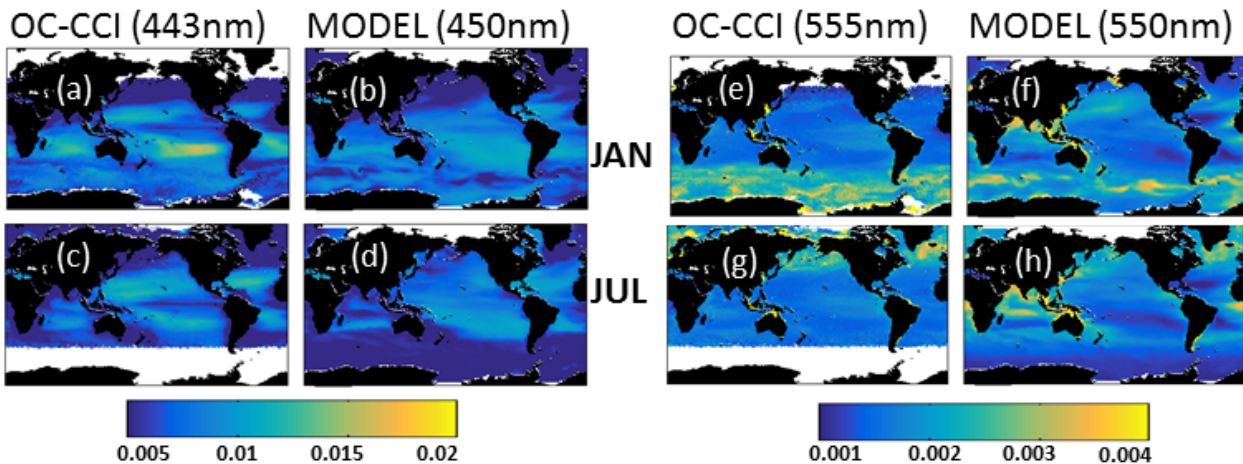


Figure 2: Remotely sensed reflectance (1/Sr) for (a) OC-CCI at 443nm, January; (b) model at 450nm, January; (c) OC-CCI at 443nm, July; (d) model at 450nm, July; (e) OC-CCI at 555nm, January; (f) model at 550nm, January; (g) OC-CCI at 555nm, July; (h) model at 550nm, July. . **We compare the model wavebands against the nearest OC-CCI wavebands, but note that they are not identical.** OC-CCI products (a,c,e,f) have no data when irradiances are too low. For model lack of output indicates regions where PAR is less than  $15 \mu\text{Ein}/\text{m}^2/\text{s}$  or the unresolved Arctic region. OC-CCI products were downloaded from <https://www.oceancolour.org>.

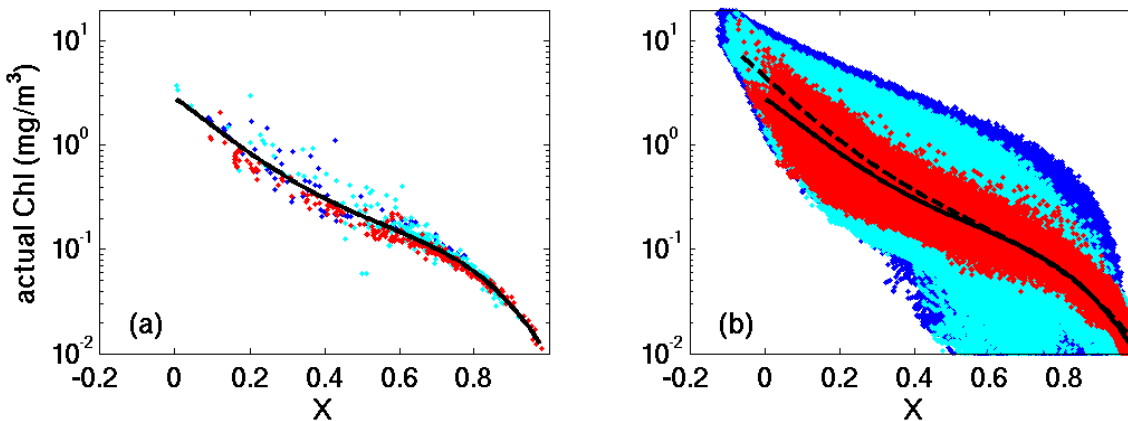


Figure 3: Model "actual" Chl-a and model blue/green reflectance ratio (X) for (a) subset of model output similar to that available from real world in situ observations (e.g NOMAD, Werdell and Bailey, 2005); (b) full model output (every day for 15 years from each grid cell, about 140 million points). Black solid line indicates the algorithm for  $\text{chl}_a$  for where coefficients were determined from the subsampled datasets (GS), and in (b) dashed line is the algorithm where coefficients were calculated using the full dataset (GA). Dots are coloured red for locations equatorward of  $30^\circ$ , light blue for  $30^\circ$  to  $60^\circ$ , and dark blue for poleward of  $60^\circ$ .

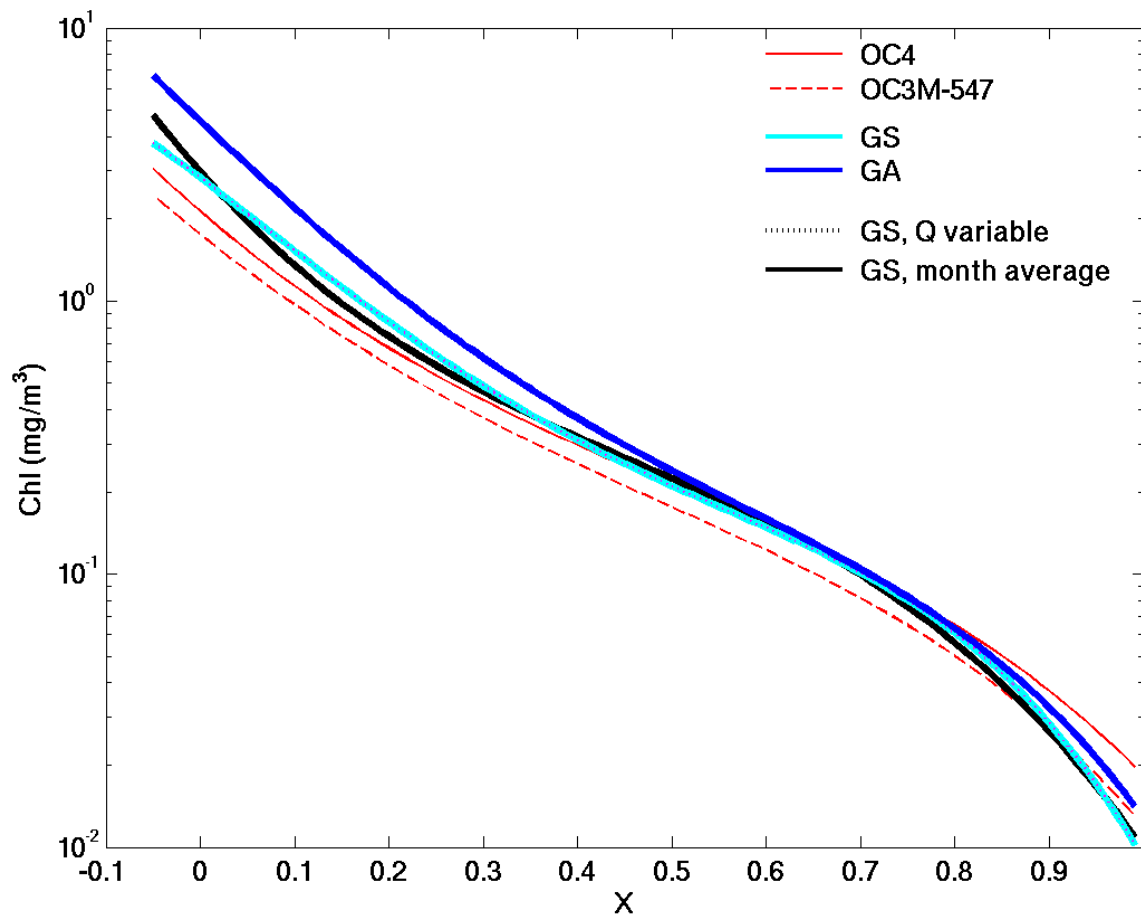
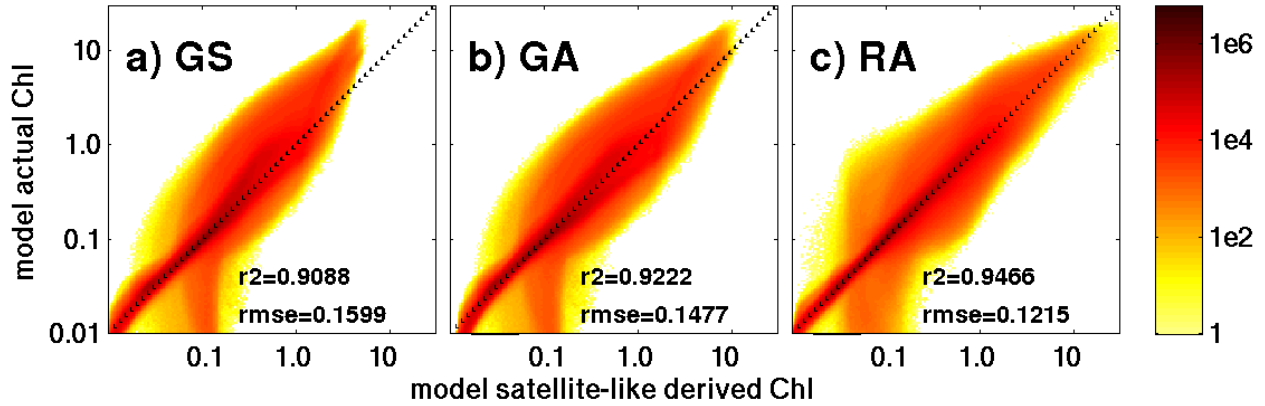
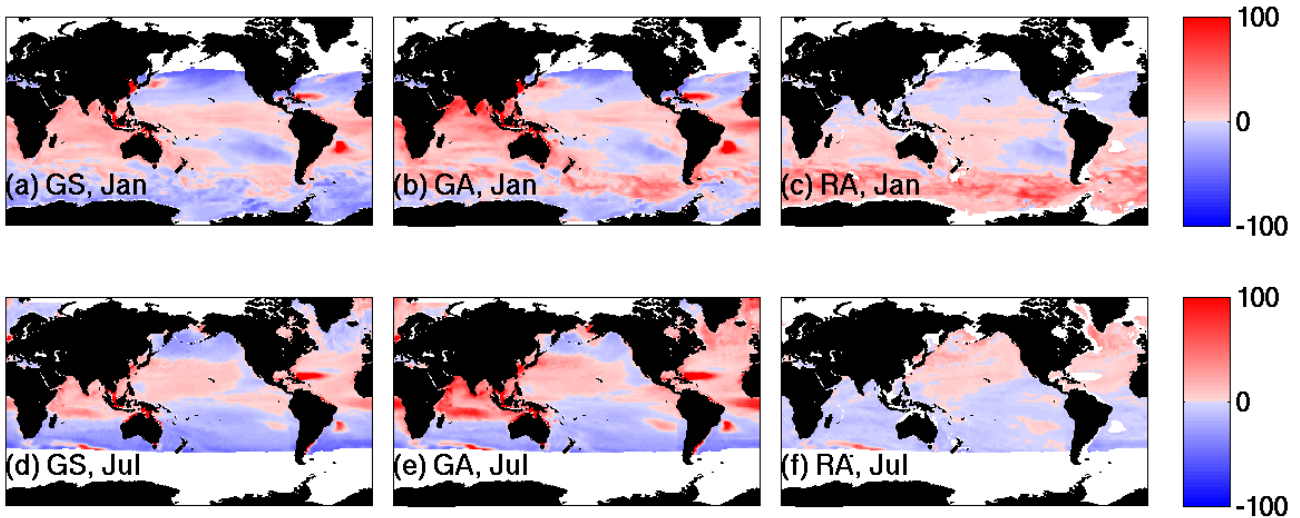


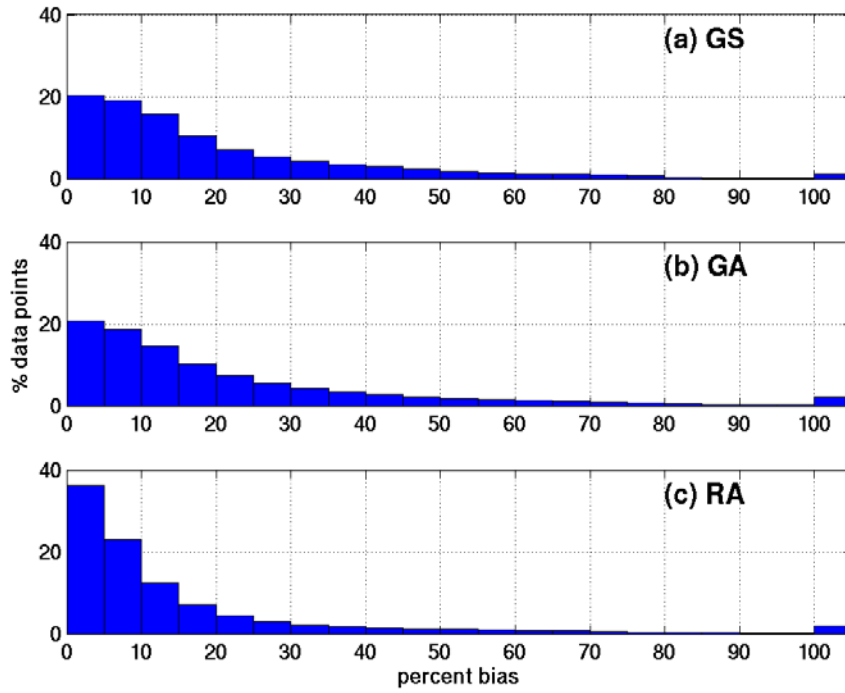
Figure 4: Polynomials for Chl-a algorithm using the blue/green reflectance ratio. Shown are two real world algorithms: NASA OC4 (red solid, used in SeaWiFS and OC-CCI products) and NASA OC3M-547 (red dashed, used for MODIS product). Model algorithms shown are GS (light blue, same as in Fig 3a), where coefficients are found from subset of model output as dictated from real world in situ observations) and GA (dark blue, same as dashed line in Fig 3b), where coefficients are found using the full data set). Also shown are two additional polynomials discussed in the Appendix: one found using a variable bi-directional coefficient, Q, and a subsampling of output as in GS (dotted black line almost exactly on top of the light blue line), and another where coefficients were found from a subsampling as in GS but using monthly average reflectance and Chl-a (black line). Note that the algorithms for the model come from band ratio of 425nm/450nm/475nm and 550nm. For the real world algorithms the band ratios are different and specific for the satellite sensor (SeaWifs or MODIS).



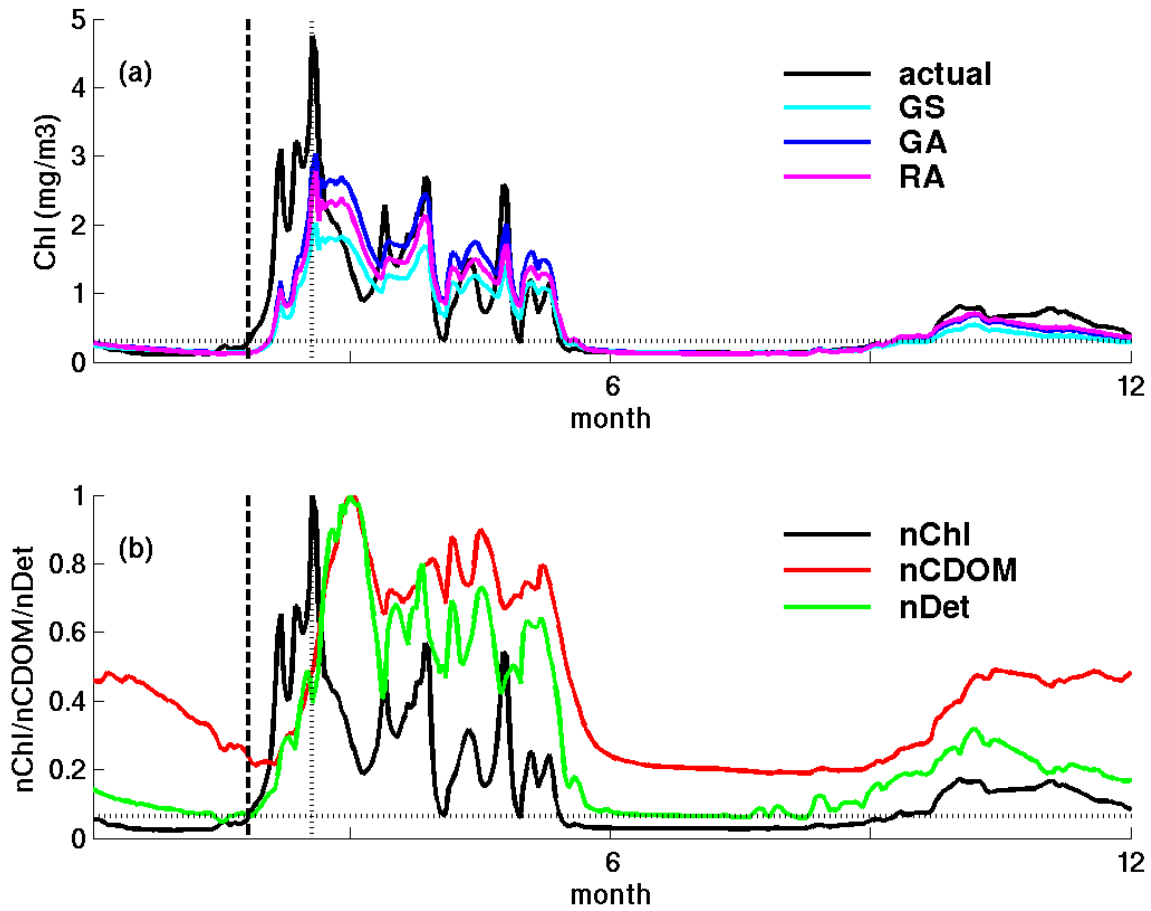
**Figure 5:** Two dimensional density histogram of model "actual" and model "derived" Chl-a using algorithm coefficients found for: (a) default experiment using approach 1 (global, subsampled output, GS), (b) default experiment using approach 2 (global, all output, GA), (c) default experiment using approach 3 (regional, RA), Dashed line indicates 1 to 1. Colour indicate the log of the fraction of all data that occur in phase space (the lightest yellow reflects a single instance in that bin). Statistics noted on the plot are for the log transformed output. ~~In linear space the  $r^2$  is (a) 0.6014, (b) 0.7670, (c) 0.8301 and RMSE is (a) 0.4816, (b) 0.3682, (c) 0.3083 in linear space is provided in Table 2.~~



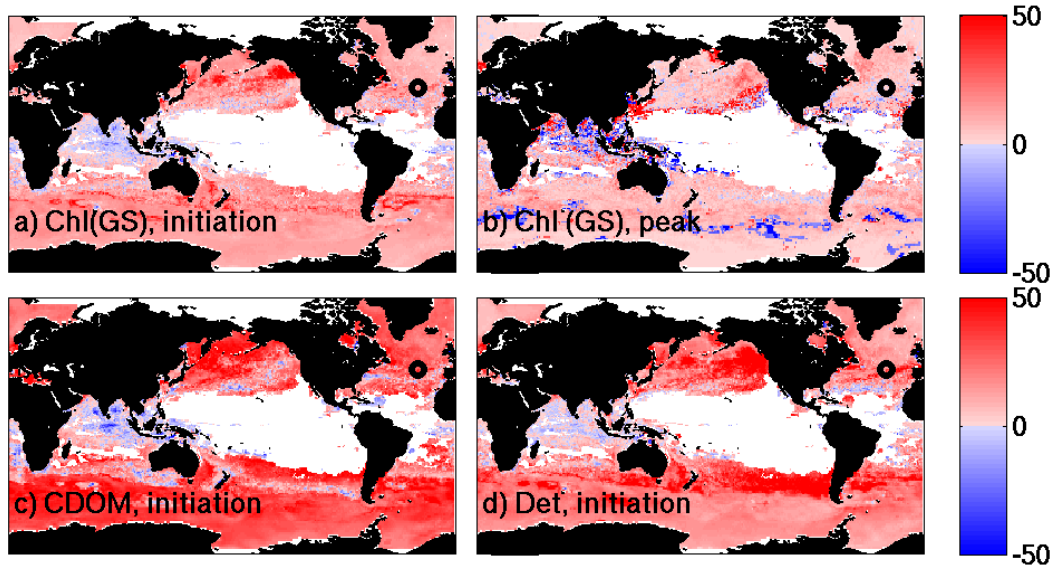
**Figure 6:** Percentage bias between monthly mean model "actual" Chl-a and model "derived" Chl-a ( $chl_a$ ) using algorithm coefficients found for: (a,d) subset of output (GS); (b,e) full model output (GA); and (c,f) each grid cell (regional specific, RA). Top row is for January, Bottom row is for July. ~~White areas Lack of output~~ indicates unresolved Arctic and regions where PAR is less than  $15 \mu\text{Ein}/\text{m}^2/\text{s}$ .



5 **Figure 7:** Distribution of percentage of model output (time and space, about 140 million “data” points) with absolute percent error between model derived Chl-a and model actual Chl-a for (a) global subsampled approach (GS), (b) global all output approach (GA), and (c) regional approach (RA).

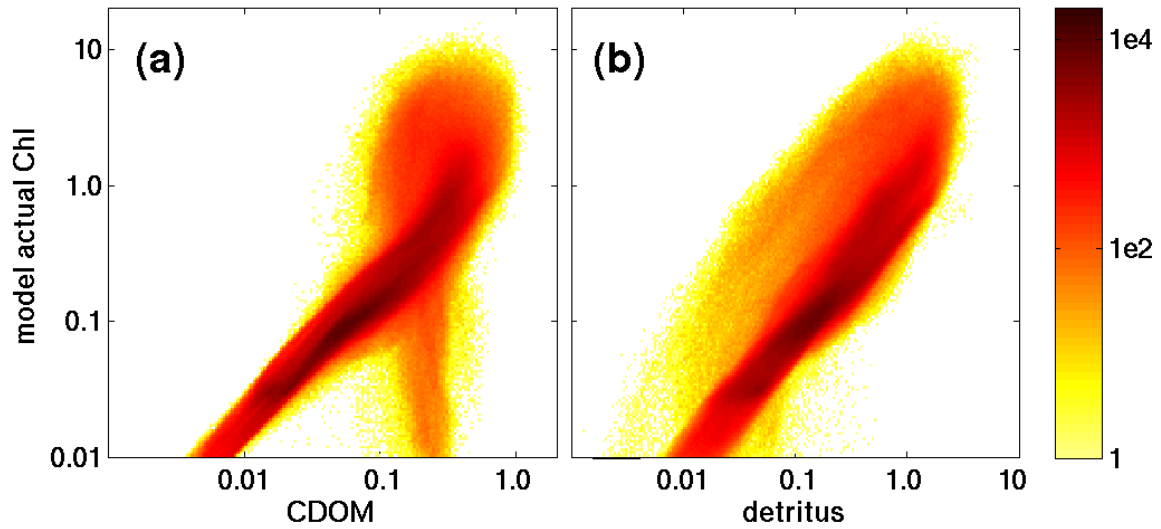


5 **Figure 8: Illustrative example of** timeseries for one year from a single location in the North Atlantic (shown as x on Fig 9). (a) “actual” Chl-a (black), derived Chl-a using subsampled output (GS, light blue), derived Chl-a using all output (GA, dark blue), and the Chl-a product derived using a regional specific algorithm (RA, purple). (b) actual Chl-a (black), CDOM (red) and detritus (green), all normalized to their peak value. Dashed vertical line indicates the “initiation of the bloom” which is taken to be when Chl-a reaches 5% above the annual median value following Cole et al (2012) and discussed further in Appendix A (dotted horizontal line shows this value for the model actual Chl-a). The vertical dotted line indicates the peak of the bloom. Shown here is only a single year and location, however for larger scale perspective, the difference in initiation and peak timing between model actual and derived Chl-a averaged over all years are shown for the globe in Figure 9.

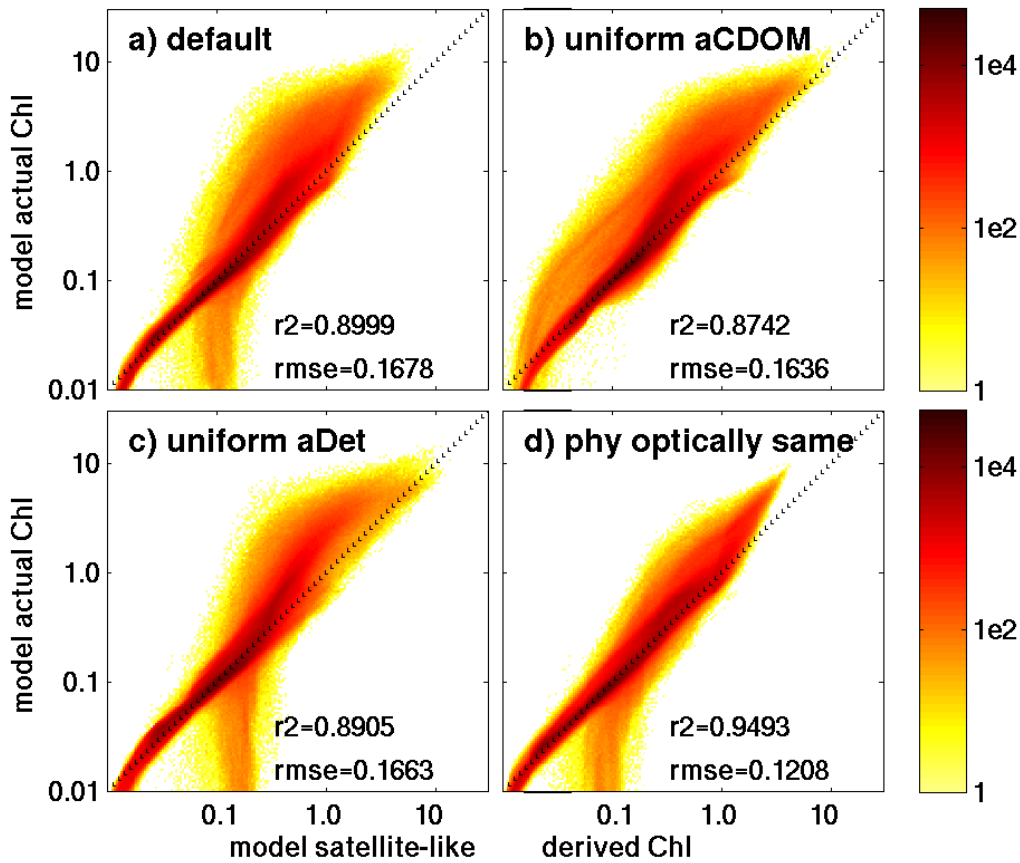


5 **Figure 9: Lag in phenology. Number of days between a) Number of days between the initiation of the spring bloom from model actual Chl-a and that for the model derived Chl-a (GS); Bloom initiation is defined as when Chl-a reaches 5% above the annual median value (see Appendix A); b) yearly maximum of model actual Chl-a and that for the derived Chl-a (GS); c) initiation of the spring bloom from model actual Chl-a and the initiation of the CDOM increase; d) initiation of the spring bloom from model actual Chl-a and the initiation of detrital particle increase. Bloom initiation is defined as when Chl-a, CDOM or detrital particles reaches 5% above their annual median value (see Appendix A). Lack of output White areas indicate did-regions with no significant seasonal cycle or are not resolved by the model (e.g. Arctic Ocean).**





**Figure 10:** Two dimensional histogram of model output for "actual" Chl-a ( $\text{mg}/\text{m}^3$ ) plotted against: (a) model CDOM ( $\text{mmol C}/\text{m}^3$ ), (b) detritus ( $\text{mmol C}/\text{m}^3$ ). Colour indicate the log of the fraction of all data that occur in bins in the phase space (the light yellow reflects a single instance in that bin).



**Figure 11: Sensitivity Experiments.** Two dimensional density histogram of model "actual" and model "derived" Chl-a using algorithm coefficients found for: (a) default experiment using approach 1 (global, subsampled output, GS), (b) EXP-1 (uniform and constant  $a_{CDOM}$ ) approach 1, and (c) EXP-2 (uniform and constant  $a_{det}$  and  $b_{det}$ ) approach 1, (d) EXP-3 (no optical differences between phytoplankton) approach 1. Dashed line indicates 1 to 1. Colour indicate the log of the fraction of all data that occur in phase space (the lightest yellow reflects a single instance in that bin). Statistics noted on the plot are for the log transformed output. In [Table 3](#), [linear space the  \$r^2\$  is \(a\) 0.5373, \(b\) 0.6298, \(c\) 0.5991, \(d\) 0.7520 and RMSE is \(a\) 0.4420, \(b\) 0.3811, \(c\) 0.3962, \(d\) 0.2591.](#) In these plots, monthly mean output of Chl-a and  $R_{RS}$  were used to calculate the algorithm, and only monthly mean output is shown (4 million versus 140 million points), thus at a great computational savings. [The difference in the algorithm is shown in Figure 4 \(the light blue line is the algorithm with coefficients found using daily values, versus the solid black line where coefficients were found using monthly values\).](#) Differences between 11a and 5a are due to this difference in sampling (discussed in Appendix B). Also notice the difference in values on the colourbars between this figure and Figure 5.