



1

## 2 **Reviews and Synthesis: To the bottom of carbon processing at** 3 **the seafloor**

4

5 **Jack J. Middelburg\***

6

7 Earth Sciences, Utrecht University, PO Box 80 021, 3508 TA Utrecht, The

8 Netherlands

9

10 *\* Invited contribution by Jack J. Middelburg, recipient of the EGU Vladimir*  
11 *Vernadsky Medal 2016*

12

### 13 **Abstract**

14

15 Organic carbon processing at the seafloor is studied by geologists to  
16 better understand the sedimentary record, by biogeochemists to quantify burial  
17 and respiration, by organic geochemists to elucidate compositional changes and  
18 by ecologists to follow carbon transfers within food webs. Here I review these  
19 disciplinary approaches and discuss where they agree and disagree. It is shown  
20 that the biogeochemical approach (ignoring the identity of organisms) and the  
21 ecological approach (focussing on growth and biomass of organisms) are  
22 consistent on longer time scales. It is hypothesized that secondary production by  
23 microbes and animals might impact the composition of sedimentary organic  
24 matter eventually buried. Animals impact sediment organic carbon processing  
25 by microbes in multiple ways: by governing organic carbon supply to sediments  
26 and by mixing labile organic matter to deeper layers. An inverted microbial loop  
27 is presented in which microbes profit from bioturbation rather than animals  
28 profiting from microbial processing of otherwise lost dissolved organic  
29 resources. Sediments devoid of fauna therefore function differently and are less  
30 efficient in processing organic matter with the consequence that more organic  
31 matter is buried and transferred from Vernadsky's biosphere to the geosphere.

32

33

### 34 **1 Introduction**

35

36 The seawater-sediment interface represents one of the largest interfaces  
37 on earth and our knowledge of processes at and fluxes through this dynamic and  
38 understudied interface is rather limited. This interface extends a few cm-dm  
39 upwards into the water column, i.e. benthic boundary layer (Boudreau and  
40 Jørgensen, 1992), as well as few cm-dm into the sediments, i.e. the bioturbated,  
41 active surface layer (Berner, 1980; Meysman et al., 2006; Aller, 2013). It serves  
42 as a habitat for organisms, governs the partitioning of material being buried or  
43 recycled and acts as a filter for the paleorecord (Rhoads, 1974). Processes in the  
44 surface sediment layer determine whether carbon is recycled within the  
45 biosphere (short-term cycle) or transferred to the geosphere (long-term cycle)  
46 and as such it functions as a key interface in the System Earth.

47

48 This pivotal role of the seafloor in processing material deposited has been  
49 studied by scientists from various disciplines with their own interests,  
techniques and paradigms (Figure 1). Marine geologists and paleoceanographers



50 study sediments with the primary aim to extract information on past  
51 environmental conditions using down-core measurements of substances  
52 delivered to the seafloor and that have survived the processing at the seafloor  
53 (Burdige, 2006; Bender, 2013). Biogeochemists quantify the fate of material  
54 delivered, in particular how much of the material is eventually buried or  
55 processed, and determine when and in what form the remaining part is recycled  
56 to the water column, because recycling of key nutrients (e.g. N, P, Si, Fe) sustains  
57 primary production (Berner, 1980; Aller, 1980, 2001, 2013; Soetaert et al.,  
58 2000). Organic geochemists investigate how organic matter delivered to the  
59 seafloor is degraded, transformed or preserved using changes in the composition  
60 at the molecular level (Hedges and Keil, 1995; Dauwe et al, 1999; Burdige, 2006;  
61 Bianchi and Canuel, 2011). Ecologists focus on the organisms, i.e. the actors  
62 consuming, producing and transporting the material deposited (Gage and Tyler,  
63 1992; Gray and Elliot, 2009; Herman et al., 1999; Krumins et al., 2013).

64 Although these disciplines often study the same topic, e.g. organic matter  
65 delivered to the seafloor, they focus on different aspects and usually ignore key  
66 concepts, findings and approaches from other disciplines. For example,  
67 ecologists and biogeochemist studying carbon flows at the seafloor normally  
68 ignore detailed molecular information available from organic geochemistry.  
69 Bioturbation, particle transport and mixing by animals at the seafloor, is often  
70 ignored by paleoceanographers, and biogeochemists have developed advanced  
71 transport-reaction models in which the actors, the animals, mix the particles but  
72 do so without consuming organic matter, their food.

73 Here I present the existing views on organic carbon processing at the  
74 seafloor, discuss where they agree and disagree and aim to arrive at an  
75 integrated view of carbon processing at the seafloor that is consistent with  
76 recent views within the organic geochemical, sediment geochemical, ecological  
77 and microbiological research communities. This overview is necessarily and  
78 admittedly incomplete but rather covers personal interests and presents new  
79 concepts on this topic. It is a concise version of the Vernadsky Medal Lecture  
80 presented at the 2017 EGU meeting.

## 82 **2 Geochemists focus on quantification of burial and mineralization.**

84 Organic matter delivered to marine sediments eventually is either  
85 mineralized and the metabolites (carbon dioxide and nutrients) accumulate in  
86 pore-waters and exchange with the overlying water, or it is not consumed and  
87 then buried through the steady accumulation of particles (Fig. 2). This  
88 geochemical view is highly simplified, but for that reason also quantitative and  
89 instructive (Berner, 1980; Boudreau, 1997; Aller, 2013). The percentage of  
90 organic matter buried varies from less than one % to a few tens % and is closely  
91 related with total sediment accumulation rate (Canfield, 1989, 1994; Middelburg  
92 et al., 1993). Since sediment accumulation rates are much higher in vegetated,  
93 coastal, deltaic, shelf and ocean-margin settings, the majority of organic matter  
94 burial occurs there, with organic carbon burial in deep-sea sediments accounting  
95 for <5 % (Berner, 1982; Duarte et al., 2005; Burdige, 2007). In the deep sea and  
96 other low-burial-efficiency settings, almost all organic matter is degraded to  
97 inorganic carbon and organic matter degradation rates provide an excellent  
98 proxy for organic matter deposition (Jørgensen, 1982; Cai and Reimers, 1995;



99 Glud, 2008). Organic matter degradation can be quantified via the consumption  
100 of oxygen, the production of dissolved inorganic carbon and through the use of  
101 pore-water data and diagenetic models (i.e. reaction-transport models for  
102 sediments). At steady state sediment oxygen consumption provides an accurate  
103 measure for total sediment organic matter degradation, independent whether  
104 organic matter is degraded aerobically (i.e. with oxygen) or anaerobically (with  
105 alternative electron acceptors such as nitrate, metal oxides, sulphate), because  
106 almost all reduced metabolites released (ammonium, manganese (II), iron(II),  
107 hydrogen sulfide and methane) are re-oxidized (Jørgensen, 1977, 2006; Berner  
108 and Westrich, 1985; Aller and Rude, 1988; Soetaert et al., 1996; Boetius et al.,  
109 2000; Strous and Jetten, 2004; Raghoebarsing et al., 2006; Middelburg and Levin,  
110 2009).

111 Geochemists have adopted a transport-reaction modeling approach to  
112 accurately quantify organic matter processing (Berner, 1980; Boudreau, 1997;  
113 Burdige, 2006). The basic premise of these diagenetic models is that both  
114 particles and solutes are subject to transport and reaction, making them distinct  
115 from for instance groundwater transport-reaction models in which normally  
116 only solutes and gas phases are mobile. Transport of solutes is due to molecular  
117 diffusion, pore-water advection and biologically mediated processes; enhanced  
118 diffusion due to interstitial fauna (Aller and Aller, 1992) and bio-irrigation due  
119 tube and burrow construction and flushing by macrofauna (Aller, 1980, 1984;  
120 2001). Particle transport is not only due to steady particle deposition but also  
121 due to animal activities (bioturbation, Boudreau, 1997; Aller, 1994, 2013; Rice,  
122 1986; Meysman et al., 2003, 2006, 2010). The reaction terms in these diagenetic  
123 models are normally limited to microbial and chemical reactions and described  
124 using zero, first, second order or Monod/Michaelis-Menten type kinetics  
125 (Bouldin, 1968; Berner, 1980; Soetaert et al., 1996; Boudreau, 1997). There is a  
126 major inconsistency in the basic conceptual model underlying the (numerical)  
127 diagenetic models: animals dominate transport processes via pore-water  
128 irrigation and particle mixing, but without consuming any organic matter. This  
129 has not received much attention because the ruling paradigm within the  
130 microbial ecological and geochemical research communities is that animals  
131 contribute very little to total carbon processing. Multiple recent studies involving  
132 use of  $^{13}\text{C}$  as deliberate tracers show that this premise does not hold on the short  
133 term (days to weeks; Blair et al., 1996; Moodley et al., 2002, 2005a; Woulds et al.,  
134 2009, 2016). Another reason for continuing with the simple approach is that our  
135 understanding of particle mixing due to animals is very limited (Meysman et al.,  
136 2006; 2010), that particle movement may require little energy because of  
137 fracturing (Dorgan et al., 2005) and that diagenetic models can very accurately  
138 reproduce most observations (Soetaert et al., 1996; Berg et al., 2003).

139 These models combined with solid-phase and pore-water concentration  
140 vs. depth profiles, sediment-water exchange fluxes and rate measurement have  
141 resulted in a consistent picture of organic matter degradation pathways in  
142 marine sediments (Berner, 1980; Boudreau, 1997; Aller, 2013). These models  
143 can predict where, when and why organic matter oxidation occurs aerobically or  
144 involves nitrate, metal oxides or sulphate as oxidants (Rabouille and Gaillard,  
145 2001; Boudreau, 1996; Soetaert et al., 1996; Middelburg et al. 1996; van  
146 Cappellen and Wang, 1996; Archer et al., 2002; Meysman et al., 2003; Berg et al.,  
147 2003). They also resolve the re-oxidation of reduced products such as



148 ammonium, manganese(II), iron (II), sulfide and methane (Fig. 2) and as such  
149 define the scope for aerobic and anaerobic organisms and the distribution and  
150 activity of chemoautotrophs. Chemoautotrophs in sediments produce about 0.4  
151  $\text{Tg C y}^{-1}$ , similar to the riverine delivery of organic carbon to the ocean  
152 (Middelburg, 2011). However, these diagenetic models cannot predict organic  
153 carbon burial rates, nor do they provide much insight why organic matter is  
154 buried or why it is labile (reactive) or rather refractory. For this we need to have  
155 a detailed look at the organic geochemistry of sediment organic carbon.

### 157 **3 Organic geochemists focus on the composition of organic matter** 158 **preserved**

159  
160 Organic matter delivered to the seafloor is predominantly produced in the  
161 surface sunlit layer of the ocean (Fig. 3). This organic matter is rich in proteins,  
162 carbohydrates and lipids and follows Redfield stoichiometry (Bianchi and  
163 Canuel, 2011). Organic matter processing leads to preferential degradation of the  
164 more labile components with the result that organic matter becomes less  
165 reactive (Middelburg, 1989; Arndt et al., 2013) and organic matter composition  
166 changes (Fig. 1; Wakeham et al. 1997; Dauwe et al., 1999; Lee et al., 2000). The  
167 proportion of organic matter that can be characterized molecularly decreases  
168 with progressive degradation, i.e. with water depth or depth downcore  
169 (Wakeham et al., 1997; Hedges et al., 2000; Middelburg et al., 1999; Nierop et al.,  
170 2017). This molecularly uncharacterizable material increases to more than 70%  
171 in deep-sea organic matter. The organic geochemical approach to study organic  
172 matter processing is limited not only by our inability to characterize the  
173 majority of the sedimentary organic matter, but also by the simple fact that the  
174 degraded fraction cannot be studied and we have to base our knowledge on the  
175 small fraction of extensively processed organic material remaining.

176 The changes in organic matter composition due to organic matter  
177 processing have been utilized to estimate the lability-digestibility or its reverse  
178 the refractory nature of organic matter with various proxies such as chlorophyll  
179 to bulk organic matter, fraction of nitrogen present as amino acid, and the  
180 contribution of proteins and carbohydrates to total organic matter (Cowie et al.,  
181 1992; Dell'Anno et al., 2000; Danovaro et al., 2001; Koho et al., 2013). The amino  
182 acid based degradation index (Dauwe and Middelburg, 1998) is one of the most  
183 commonly used proxies to quantify the extent of degradation or the quality of  
184 the remaining particulate organic matter (Dauwe et al., 1999; Keil et al., 2000).

185 The compositional changes have also been used to infer transformation of  
186 organic matter by bacteria (production of bacterial transformation products;  
187 accumulation of D-amino acids; Cowie and Hedges, 1994; Dauwe et al., 1999;  
188 Grutters et al., 2001; Vandewiele et al., 2009; Lomstein et al., 2006, 2012), extent  
189 of degradation under oxic and anoxic conditions (Sinninghe Damsté et al., 2002;  
190 Huguet et al., 2008; Nierop et al., 2017) and the relative importance of bacterial  
191 and faunal pathways of organic matter degradation pathways (Sun et al., 1999;  
192 Woulds et al., 2012, 2014). Although some organic geochemical studies hint at  
193 the importance of secondary production (new organic matter produced by  
194 microbes and animals and other heterotrophs; Cowie and Hedges, 1994; Grutters  
195 et al., 2001; Lomstein et al., 2006, 2012), this aspect has received little attention




196 in organic geochemistry, yet is one of the main objectives within the ecological  
197 approach.

198

#### 199 **4 Ecologists focus on the dynamics of organisms using organic matter**

200

201 Benthic communities are usually partitioned into different size classes  
202 (e.g. macrofauna, meiofauna, microbes; Gage and Tyler, 1992; Gray and Elliot,  
203 2009; Herman et al., 1999), which are often studied by different research  
204 communities having distinct objectives, approaches and tools. Organic matter  
205 delivered to the seafloor fuels benthic food webs: i.e. it represents food for the  
206 animals and the energy substrate for heterotrophic microbes. Microbial  
207 ecologists study the growth of microbes on delivered organic matter (e.g.  
208 bacterial production) and subsequent microbial loss processes, including  
209 predation and viral lysis (Kemp, 1988; 1990; Danovaro et al., 2008, 2011, 2016).  
210 Microbial ecologists also study in detail the identities and activities of organisms  
211 involved in (an)aerobic respiration pathways and the re-oxidation of reduced  
212 metabolites produced during anaerobic organic matter degradation (Canfield et  
213 al., 2005). Animal ecologists focus on the response of fauna to food delivery, the  
214 diet and growth of animals and transfer of carbon up the food chain to top  
215 consumers (Krumins et al., 2013; Fig. 3). Interactions among food-web members  
216 are considered key to understand carbon flows (Pimm et al., 1991; van Oevelen  
217 et al., 2010).


218 During the last two decades, isotopically labeled phytodetritus addition  
219 experiments have been performed to identify the organisms involved in the  
220 immediate processing of organic matter delivered to the seafloor (Middelburg,  
221 2014). These studies often covered all size classes (animals and microbes) and  
222 could show that respiration was the major  and that all size classes directly  
223 profited from recently deposited organic matter (Moodley et al., 2002, 2005a;  
224 Woulds et al., 2007, 2009, 2016; Witte et al., 2003). In other words,  
225 heterotrophic microbes and small and big animals compete for the same food.  
226 The relative share of organisms in the processing of organic matter was in some  
227 systems and for some consumers proportional to the biomass of the benthic size  
228 class, but not always (Moodley et al., 2005a; Woulds et al., 2009, 2016). For  
229 instance, foraminifera, amoebid protozoa, sometimes contribute  
230 disproportionately to short-term carbon processing reflecting high turn-over of an  
231 active community (Moodley et al., 2002; Woulds et al., 2007). The spatial  
232 distribution of resources is also a key factor governing the relative use of  
233 phytodetritus by bacteria vs. animals (van Nugteren et al., 2009b).

234

235

#### 236 **5 Towards a synthesis**

237

238 The above discussion on conceptual views within different research disciplines  
239 highlights a few discrepancies and  in our knowledge. Secondary production  
240 by animals and microbes is **not included** in the geochemical view that focuses on  
241 preservation versus mineralization. It is also largely absent from the organic  
242 geochemical literature. Consumption of organic matter is restricted to microbes  
243 in the geochemical view, while the non-fed animals move organic matter,  
244 microbes and particles around by bioturbation and enhance solute transfer by



245 bio-irrigation activities. The consumption of organic matter eventually results in  
246 compositional changes of the organic matter remaining but there is little  
247 information that the identity of the organism processing the organic carbon  
248 matters much. Whole ecosystem labeling experiments revealed direct flow from  
249 detritus to most benthic consumers and to the dissolved inorganic carbon pool,  
250 but these short-term experimental results cannot directly be compared to the  
251 long-term natural processing of deposited organic matter because long-term  
252 transfers in the food web and eventual carbon preservation cannot be resolved  
253 experimentally.

### 254 255 **5.1 On the consistency of food-web carbon processing and the geochemical** 256 **burial-respiration partitioning**

257 Food webs describe the exchange of matter (e.g. carbon or energy) among  
258 different compartments (organisms) within an ecosystem (Pimm et al., 1991; de  
259 Ruiter et al., 1995) and thus formalize the ecological view on carbon processing  
260 (Cole et al., 2006; van Oevelen et al., 2010). The emphasis is on interaction among  
261 organisms and respiration losses are normally lumped into a single carbon  
262 dioxide loss term (Fig. 3 left). Experimental studies using  $^{13}\text{C}$  labelled  
263 phytodetritus as a tracer of sediment carbon processing showed that animals  
264 and microbes both can assimilate labile carbon directly and confirmed that  
265 respiration is the largest sink (Moodley et al., 2005a; Buhning et al., 2006;  
266 Andersson et al. 2008; Woulds et al. 2009, 2016). The geochemical budgeting  
267 approach basically distinguishes only between (refractory) carbon preserved  
268 and buried versus labile organic carbon that is respired to carbon dioxide (Glaser  
269 2008; Aller, 2013; Fig. 3 right). These ecological and geochemical concepts are  
270 consistent when the typical timescale of interest is considered. On the time scale  
271 of days to month deposited carbon is processed by the benthic organisms, a  
272 small part is assimilated and the majority is respired. On longer time scales and  
273 when considering steady-state conditions, i.e. constant faunal and microbial  
274 biomass, there is no net transfer from the detritus pool to the living biomass pool  
275 and all labile carbon is eventually respired because organisms only represent a  
276 (temporary) carbon sink when their biomass is building up or when remains of  
277 secondary producers are buried.

### 278 279 280 **5.2 Secondary production and the formation of molecularly** 281 **uncharacterizable organic matter.**

282 The mere presence of living organisms in sediments clearly indicates that  
283 secondary production is omnipresent. Microbes usually dominate living biomass,  
284 but not always, and living biomass typically contributes a few % at most to the  
285 standing stock of total organic carbon in sediments (Herman et al., 1999).  
286 Various types of experimental evidence have shown that carbon flow through  
287 living compartment is much higher than through the non-living sediment organic  
288 matter pool. Short-term, in situ experiments using  $^{13}\text{C}$  and/or  $^{15}\text{N}$  labelled  
289 organic matter (e.g. phytodetritus) revealed rapid incorporation of  $^{13}\text{C}/^{15}\text{N}$  in  
290 physically separated organisms (macro- and meiofauna and foraminifera) and  
291 microbes, the latter via incorporation of tracers in biomarkers specific for certain  
292 microbial groups (Middelburg et al., 2000; Boschker and Middelburg, 2002;



294 Veuger et al., 2007; Oakes et al., 2012; Woulds et al., 2007, 2016) Similar  
295 ammonium isotope dilution studies have shown that net ammonification is only  
296 a small fraction of the total ammonium regeneration because most of the  
297 ammonium liberated is re-assimilated by the microbial community (Blackburn  
298 and Henriksen, 1983). Clearly the microbes and animals living in sediment  
299 assimilate carbon and synthesize new tissues. How can this be reconciled with  
300 the geochemical and organic geochemical views in which organic matter is either  
301 preferentially degraded to carbon dioxide or selective preserved (Fig. 1, 3).  
302 These two apparently inconsistent views are consistent if all of the newly  
303 produced organic matter is eventually degraded.

304 Detailed investigations of organic matter composition might in principle  
305 resolve this issue as microbial and animal processing of organic matter results in  
306 the formation of distinct compounds (Bradshaw et al., 1990; Sun et al., 1999;  
307 Thomas and Blair, 2002; Woulds et al., 2012, 2014). There are few issues with  
308 this approach: (1) most sedimentary organic matter is molecularly  
309 uncharacterizable and the origin (imported from the water column vs. newly  
310 produced within the sediment) can thus not be investigated, (2) microbes living  
311 within (the guts) of animals may mask the animal signatures (Woulds et al.,  
312 2012, 2014) and (3) different analytical windows (amino acids vs. lipids) may  
313 result in different inferences. On the one hand, the accumulation of bacterial  
314 derived non-protein amino acids and peptidoglycan derived D amino acids are  
315 clear signs that extensively modified organic matter contains a major fraction  
316 that is derived from (heterotrophic) bacteria (Cowie and Hedges, 1994; Dauwe et  
317 al., 1999; Grutters et al., 2001; Lomstein et al., 2006; Keil and Fogel, 2001; Keil et  
318 al., 2000). On the other hand, using a combined lipid-isotope approach Hartgers  
319 et al. (1994) reported only a minor contribution of bacteria to sedimentary  
320 organic carbon pools. However, Gong and Hollander (1997) used fatty acids and  
321 identified a substantial microbial contribution to sedimentary organic matter.

322 Secondary production has potentially major consequences for the  
323 interpretation of sedimentary records. If microbial reworking of deposited  
324 organic matter represents a major carbon processing flow and part of the  
325 material is preserved then one would expect that bulk organic matter properties  
326 such as C, N, P elemental ratios and nitrogen isotopes would reflect this.  
327 Degradation of organic matter usually results in the preferential release of  
328 nitrogen and phosphorus relative to carbon. Microbes normally have lower C/N  
329 ratios than their substrate, implying that secondary production and  
330 accumulation of microbial derived organic matter results in a net decrease of  
331 sediment C/N ratios (Müller, 1977). In contrast, the C/P ratio of heterotrophic  
332 microbes is rather variable because P demands depends on the growth rate  
333 (Sterner and Elser, 2002) and slowly growing benthic microbes may have high  
334 C:P ratios (Steenbergh et al., 2013). Moreover, microbial P storage also depends  
335 on redox conditions with the consequences that sedimentary C:P ratios are  
336 highly variable (Algeo and Ingall, 2007). Sediment  $\delta^{15}\text{N}$  values often show a post-  
337 depositional shift towards heavier values in alternating oxic/anoxic settings  
338 (Moodley et al., 2005b). Such a shift is to be expected because regenerated  
339 ammonium is either transformed into nitrite/nitrate (nitrification) or re-  
340 assimilated by the microbial community. During oxic conditions nitrification  
341 occurs with preference for  $^{14}\text{N}$  and the remaining ammonium available for re-  
342 assimilation by microbes will be relatively rich in  $^{15}\text{N}$ , while during anoxic



343 conditions oxidation of ammonium is less important or absent, and the  
344 ammonium re-assimilated will have similar  $\delta^{15}\text{N}$  values as that regenerated.  
345 Secondary production of microbial biomarkers within sediment may also impact  
346 the interpretation of paleorecords (Schouten et al., 2010).

347 To reconcile the strong experimental evidence for preferential  
348 degradation (Middelburg, 1989), selective preservation (Tegelaar et al., 1989)  
349 and formation of new compounds by secondary producers (Lomstein et al, 2012;  
350 Braun et al, 2017) we present a new integral concept (Fig. 4). Phytodetritus  
351 delivered to sediments is preferentially degraded with the result that new  
352 biomass is formed and that some compounds are selectively preserved. The  
353 newly formed biomass is after death of the organism added to the pool of  
354 degraded detritus and subject to further microbial processing. After multiple  
355 cycles of processing by benthic heterotrophs most of the remaining organic  
356 matter becomes molecularly uncharacterizable. This conceptual model is  
357 consistent with the ruling paradigms of preferential degradation and selective  
358 preservation as well as with the occurrence of secondary production and  
359 formation of molecular uncharacterizable organic matter, but the next step is to  
360 quantify this conceptual view. One approach would be to use proxies for organic  
361 matter degradation state such as fraction of total nitrogen present as amino acid,  
362 non-protein amino acids accumulation and the degradation index (Cowie and  
363 Hedges, 1994; Dauwe and Middelburg, 1998; Dauwe et al., 1999). Lomstein et al.  
364 (2012) and Braun et al. (2017) used amino acid racemization to quantify  
365 turnover of living microbial biomass as well as of bacterially derived organic  
366 matter (necromass) in the deep biosphere. Veuger et al. (2012) executed a  
367  $^{13}\text{C}/^{15}\text{N}$  tracer experiment and followed the isotope labels into carbohydrates,  
368 amino acids and lipids and basically showed that most of the deliberately added  
369 heavy isotopes were recovered from the molecularly uncharacterizable pool  
370 within a few weeks and remained in that pool till the end of the experiment (>  
371 1year). Their study provided direct evidence for rapid formation of new  
372 microbial biomass and subsequent transfer of microbial biomass to the pool of  
373 molecular uncharacterizable organic matter. Moreover, the efficient retention of  
374 label was indicative of recycling of molecules (or parts thereof) by microbes  
375 rather than de novo synthesis, consistent with findings in soil science (Dippold  
376 and Kuzakov, 2016).

377

378

### 379 **5.3 Animals and carbon supply to sediments**

380

381 Marine sediments are often considered donor-controlled systems, i.e. organic  
382 matter is delivered via settling of organic matter produced in the sunlit upper  
383 part of the ocean (Fig. 3) and the consuming sediment communities have no  
384 control on its carbon delivery (5). It is only on the time-scale of ocean  
385 bottom-water renewal (10-100 years) that nutrient regenerated by benthic  
386 organisms may impact primary producer in the sunlit upper part of the ocean  
387 (Soetaert et al., 2000). This is obviously different for sediments in the photic  
388 zone that make up about 1/3 of the coastal ocean (Gattuso et al, 1996) because  
389 animals can directly graze and consume the benthic primary producers at the  
390 sediment surface (Middelburg et al., 2000; Evrard et al., 2010, 2012; Oakes et al.,





391 2012; Fig. 5). Donor and consumer controlled food webs have intrinsic different  
392 dynamics.

393 Animals living in sediments below the photic zone can in multiple ways  
394 impact carbon processing within marine sediments (Fig. 5). Deposit-feeding  
395 animals mix particles (and thus particulate organic carbon) as a consequence of  
396 their activities. In the case of constant organic carbon delivery (donor-control)  
397 bioturbation stimulates organic carbon processing at depth (Herman et al.,  
398 1999). In coastal systems, organic matter delivery is more complex because of  
399 multiple deposition-resuspension events and lateral transport pathways. Rice  
400 and Rhoads (1989) showed that in this case (with constant organic carbon  
401 concentration in the top layer) more bioturbation will increase the organic  
402 matter flux into the sediment. Moreover, organic carbon gradients with depth  
403 are steeper for high-quality than low-quality material and bioturbation thus  
404 results in transfer of high-quality organic matter to (micro-)organisms living at  
405 depth. Animals living in permeable sediment can via surface sediment  
406 topography (bioroughness) induce pore-water flows resulting in the trapping of  
407 phytoplankton (Huettel et al., 2014).

408 Tropical and cold-water corals, coastal and deep-sea sponges, suspension  
409 feeding bivalves and other marine forests communities utilize particulate  
410 organic matter suspended in the water (Herman et al., 1999; Roberts et al., 2006;  
411 Rossi et al., 2017). This organic carbon is used for maintenance respiration and  
412 growth, but part is excreted as faeces or pseudofaeces and becomes then  
413 available for consumers in the sediments. This can result in local hotspots of  
414 biodiversity and microbial activity in the sediments (Herman et al., 1999;  
415 Gutierrez et al., 2003; Cathalot et al., 2015). Moreover, the physical structures  
416 build by these ecosystem engineers impact hydrodynamics with consequences  
417 for local and distant carbon deposition rates. Soetaert et al. (2016) reported  
418 elevated carbon deposition to ocean margin sediments due to cold-water corals  
419 reefs at very large distances.

420 Some sponges have the capability to take up dissolved organic carbon and  
421 transform it into sponge tissue (de Goeij et al., 2013; Fiore et al., 2017; Hoer et  
422 al., 2017). This sponge tissue and in particular its detritus can be consumed by  
423 benthic organisms. This sponge loop (de Goeij et al., 2013; Rix et al., 2016) is  
424 another example how animals can manipulate the transfer of organic carbon  
425 from the water-column to the sediments (Fig. 5).

426

427

#### 428 **5.4 Animal stimulation of microbes: An inverted microbial loop?**

429

430 The microbial loop is a key concept in ocean biogeochemistry (Pomeroy,  
431 1974; Azam et al., 1983). Dissolved organic matter released by phytoplankton,  
432 zooplankton or viral lysis of bacteria, archaea and algae is consumed by  
433 heterotrophic microbes. These heterotrophs are in turn consumed by flagellates,  
434 ciliates and other small consumers that are predated upon by zooplankton  
435 (Azam et al., 1983; Jumars et al. 1989). Energy shunted into the large,  
436 heterogeneous dissolved organic matter pool is in this way made available again  
437 for animals (Fig. 6).

438 After discovery of this loop in the surface ocean water, research has been  
439 executed to identify and quantify it in sediments (Kemp, 1988, 1990). This



440 required substantial investment in developing new methods: these studies  
441 basically revealed that predation on sedimentary bacteria was not that  
442 important (Kemp, 1990; Hondeveld et al., 1992; Hamels et al., 2001; Guilini et al.,  
443 2009). Van Oevelen et al., (2006) made a detailed study on the fate of bacterial  
444 production using in situ <sup>13</sup>C labelling of bacteria. They observed that 8% was lost  
445 by physical processes, 27% was consumed by animal predation, while bacterial  
446 mortality accounted for 65%. Viruses are the most important loss term for  
447 sedimentary microbes (Danovaro et al, 2009, 2011, 2016) and the viral lysis  
448 products (dissolved organic matter) are consumed by microbes. This results in a  
449 dissolved organic carbon-microbes cycle (Fig. 6). This benthic microbial cycle  
450 represents a dead end in terms of food web topology, because there is little  
451 transfer to higher trophic levels and most carbon is eventually respired as  
452 needed for mass-balance closure on the long term (Fig. 3).

453 In fact, the benthic microbial cycle represents more an *inverted microbial*  
454 *loop*: rather than animals profit from the microbial loop sensu Azam et al. (1983),  
455 it appears that benthic microbes profit from animals mixing labile organic matter  
456 downwards into the sediments (Fig. 6). Labile organic matter delivered to the  
457 sediment surface is mixed by animals inhabiting the sediments (Fig. 5). The  
458 transfer of high quality organic matter to deeper sediment layer may prime  
459 sediment microbial communities and in this way stimulate degradation of  
460 indigenous organic matter (van Nugteren et al, 2009a, Bianchi, 2011; Hannadis &  
461 Aller, 2017). This inverted microbial loop is a prime example how animals as  
462 ecosystem engineers impact sediment carbon processing (Meysman et al., 2006).

## 465 5.5 Imagine a world without animals.

467 An ocean floor inhabited solely by microbes and without animals was likely the  
468 reference state during the first four billion of years of Earth's history (Canfield,  
469 2014; Lenton and Watson, 2011). Moreover, in modern systems with anoxic  
470 bottom waters benthic animals are absent (Rhoads and Morse, 1971; Diaz and  
471 Rosenberg, 1995, 2008; Levin, 2003; Levin et al, 2009). In these systems, organic  
472 matter degradation pathways are different not only because of a lack of oxygen  
473 and use of alternative electron acceptors, but also because bio-irrigation and  
474 bioturbation are absent (Aller and Aller, 1998; Levin et al., 2009; Middelburg and  
475 Levin, 2009). Consequently, microbe-fauna interactions (enhanced carbon  
476 delivery, Fig. 5; animal stimulation of microbes, Fig. 6) are impeded.

477 This likely is the reason why more organic carbon is buried in sediments  
478 underlying anoxic bottom waters (Hartnett et al., 1998; Hartnett and Devol,  
479 2003; Middelburg and Levin, 2009; Jessen et al., 2017). Moreover, the organic  
480 matter buried in hypoxic and anoxic settings is usually less degraded (Cowie,  
481 2005, Cowie et al., 2009; Vandewiele et al., 2009; Koho et al., 2013; Jessen et al.,  
482 2017). This presence of animals and all their interactions with organic matter  
483 and microbes has consequences for organic carbon processing in marine  
484 sediments and thus the global carbon cycle. It is obvious for any terrestrial  
485 microbiologist that a world with trees and other macrophytes would be different  
486 than one without. Moreover, biological oceanographers and limnologists agree  
487 that zooplankton and other metazoan consumers contribute to biogeochemical  
488 cycles (Vanni, 2002; Vanni and McIntyre, 2016), and I hope that colleagues



489 studying marine sediments are aware that BIO in sediment biogeochemistry is  
490 more than just microbiology.

491

492 *Acknowledgement*

493 This paper presents my acceptance lecture for the Vladimir Ivanovich Vernadsky  
494 Medal 2017 of the European Geosciences Union. I thank the colleagues that  
495 nominated me for this award and the many colleagues, students and  
496 postdoctoral fellows with which I have had the pleasure to interact and  
497 collaborate over the years. In particular, the late Carlo Heip who was  
498 instrumental for getting the BIO into my biogeochemistry and my long-term  
499 collaborators in modelling benthic ecosystems (Karline Soetaert, Peter Herman,  
500 Filip Meysman and Bernie Boudreau), in food-web studies (Dick van Oevelen,  
501 Leon Moodley), in organic geochemistry (Jaap Sinnighe Damsté and Stefan  
502 Schouten) and in coastal biogeochemistry (Jean-Pierre Gattuso and Carlos  
503 Duarte). Ton Marcus is thanked for graphic support. This is a contribution to the  
504 Netherlands Earth System Science Centre supported by the Dutch ministry of  
505 Education and Science.

506

507



508

509 **References**

510

511 Algeo T.J. and Ingall E.: Sedimentary C<sub>org</sub>:P ratios, paleocean ventilation, and Phanerozoic  
512 atmospheric pO<sub>2</sub>. *Geochim. Cosmochim. Acta* 256: 130-155, 2007.513 Aller, R. C.: Quantifying solute distributions in the bioturbated zone of marine sediments  
514 by defining an average micro environment, *Geochim. Cosmochim. Acta*, 44,  
515 1955-1965, 1980.516 Aller, R. C.: The importance of relict burrow structures and burrow irrigation in  
517 controlling sedimentary solute distributions, *Geochim. Cosmochim. Acta*, 48,  
518 1929-1934, 1984.519 Aller, R. C.: Bioturbation and remineralization of sedimentary organic matter - Effects of  
520 redox oscillation. *Chem. Geol.* 114, 331-345, 1994.521 Aller, R.C.: Transport and reactions in the bioirrigated zone, in: B.P. Boudreau and B.B.  
522 Jørgensen, *The benthic boundary layer: transport processes and biogeochemistry*.  
523 pp. 269-301, 2001.524 Aller, R. C., and Rude, P. D.: Complete oxidation of solid-phase sulphides by manganese  
525 and bacteria in anoxic marine sediments, *Geochim. Cosmochim. Acta*, 52, 751-  
526 765, 1988.527 Aller, R. C., and Aller, J. Y.: Meiofauna and solute exchange in marine muds. *Limnol.*  
528 *Oceanog.* 37, 1018-1033, 1992.529 Aller, R. C., and Aller, J. Y.: The effect of biogenic irrigation intensity and solute exchange  
530 on diagenetic reaction rates in marine sediments, *J. Mar. Res.*, 56, 905-936, 1998.531 Aller, R.C. Sedimentary Diagenesis, Depositional Environments, and Benthic Fluxes  
532 *Treatise on Geochemistry: Second Edition*, 8, pp. 293-33, 2013.533 Andersson, J. H., Woulds, C., Schwartz, M., Cowie, G. L., Levin, L. A., Soetaert, K., and  
534 Middelburg, J. J.: Short-term fate of phytodetritus in sediments across the  
535 arabian sea oxygen minimum zone, *Biogeosciences*, 5, 43-53, 2008.536 Archer, D., Morford, J.L., and Emerson, S.: A model of suboxic diagenesis suitable for  
537 automatic tuning and gridded global domains. *Global Biogeochemical Cycles* 16.  
538 <http://dx.doi.org/10.1029/2000BG001288>, 2002.539 Arndt, S., Jørgensen, B.B., LaRowe, D., Middelburg, J.J., Pancost, R. and Regnier, P., :  
540 Quantification of organic matter degradation in marine sediments: A synthesis  
541 and review, *Earth-Science Reviews*, vol 123. DOI:  
542 <http://dx.doi.org/10.1016/j.earscirev.2013.02.008>, 2013.543 Azam, F., Fenchel, T., Field, J. G., Gray, J. S., Meyer-Reil, L. A., and Thingstad, F.: The  
544 ecological role of water-column microbes in the sea, *Mar. Ecol.-Prog. Ser.*, 10,  
545 257-263, 1983.546 Bender, M.L. *Paleoclimate*. Princeton University Press, 2013.547 Berg, P., Rysgaard, S., and Thamdrup, B.: Dynamic modeling of early diagenesis and  
548 nutrient cycling. A case study in an Arctic marine sediment. *American Journal of*  
549 *Science* 303, 905-955, 2003.550 Berner, R. A.: *Early diagenesis: A theoretical approach*. Princeton Univ. Press, 1980.551 Berner, R. A.: Burial of organic carbon and pyritic sulphur in the modern ocean: Its  
552 geochemical and environmental significance. *American Journal of Science*, 282,  
553 451-473, 1982.554 Berner, R. A., and Westrich, J. T.: Bioturbation and the early diagenesis of carbon and  
555 sulphur, *Am. J. Sci.*, 285, 193-206, 1985.556 Bianchi, T.S.: The role of terrestrially derived organic carbon in the coastal ocean: a  
557 changing paradigm and the priming effect. *PNAS*.  
558 <http://dx.doi.org/10.1073/pnas.1017982108>, 2011559 Bianchi, T.S. and Canuel, E.A. *Chemical biomarkers in aquatic ecosystems*. Princeton  
560 University Press, 2011.



- 561 Blackburn, T.H. and Henriksen, K. Nitrogen cycling in different types of sediments from  
562 Danish waters. *Limnol. Oceanogr.* 28, 477-493, 1983.
- 563 Blair, N. E., Levin, L. A., DeMaster, D. J., and Plaia, G.: The shortterm fate of fresh algal  
564 carbon in continental slope sediments, *Limnol. Oceanogr.*, 41, 1208-1219, 1996.
- 565 Boetius, A., Ravensschlag, K., Schubert, C. J., Rickert, D., Widdel, F., Gieseke, A., Amann, R.,  
566 Jorgensen, B. B., Witte, U., and Pfannkuche, O.: A marine microbial consortium  
567 apparently mediating anaerobic oxidation of methane, *Nature*, 407, 623-626,  
568 2000.
- 569 Boschker, H. T. S. and Middelburg, J. J.: Stable isotopes and biomarkers in microbial  
570 ecology, *FEMS Microbiol. Ecol.*, 40, 85-95, 10.1111/j.1574-6941.2002.tb00940.x,  
571 2002.
- 572 Bouldin, D. R.: Models for describing diffusion of oxygen and other mobile constituents  
573 across mud-water interface, *J. Ecol.*, 56, 77-87, 1968.
- 574 Boudreau, B. P.: A method-of-lines code for carbon and nutrient diagenesis in aquatic  
575 sediments, *Comput. Geosci.*, 22, 479-496, doi:10.1016/0098-3004(95)00115-8,  
576 1996.
- 577 Boudreau, B. P.: Diagenetic models and their implementation, *Modelling transport and*  
578 *reactions in aquatic sediments*, doi:10.1007/978-3-642-60421-8, 1997.
- 579 Boudreau, B. P. and Jorgensen, B. B.: *The Benthic Boundary Layer: Transport Processes*  
580 *and Biogeochemistry*, edited by: Boudreau, B. P. and Jorgensen, B. B., Oxford  
581 University Press, 1st Edn., 2001.
- 582 Bradshaw, S.A. O'hara, S.C.M., Corner, E.D.S. and Eglinton, G: Changes in lipids during  
583 simulated herbivorous feeding by the marine crustacean *Neomysis integer*.  
584 *Journal of the Marine Biological Association of the United Kingdom*, 70, 225-243,  
585 1990.
- 586 Braun S., Mhatre S.S., Jaussi, M. Roy H., Kjeldsen K.U., Pearce C., Seidenkrantz M-S, B.B.  
587 Jorgensen, B. A. Lomstein. Microbial turnover in the deep seabed studied by  
588 amino acid racemization modelling. *Sci. Rep.* 7, Article number: 5680,  
589 doi:10.1038/s41598-017-05972-z, 2017.
- 590 Buhring, S. I., Lampadariou, N., Moodley, L., Tselepides, A. and Witte, U.: Benthic  
591 microbial and whole-community responses to different amounts of <sup>13</sup>C-enriched  
592 algae: In situ experiments in the deep Cretan Sea (Eastern Mediterranean).  
593 *Limnology and Oceanography*, 51, 157-165, 2006.
- 594 Burdige D. J. : Preservation of organic matter in marine sediments: controls,  
595 mechanisms, and an imbalance in sediment organic carbon budgets?. *Chem. Rev.*  
596 107 467-485. doi:10.1002/chin.200720266, 2007.
- 597 Burdige D. : *Geochemistry of Marine Sediments*. Princeton University Press, 2006
- 598 Cai, W. J., and Reimers, C. E.: Benthic oxygen flux, bottom water oxygen concentration  
599 and core top organic carbon content in the deep northeast Pacific Ocean, *Deep-*  
600 *Sea Res. I*, 42, 1681-1699, 1995.
- 601 Canfield, D. E.: Sulphate reduction and oxic respiration in marine sediments -  
602 Implications for organic carbon preservation in euxinic environments, *Deep-Sea*  
603 *Res. I*, 36, 121-138, 1989.
- 604 Canfield, D. E.: Factors Influencing organic carbon preservation in marine sediments.  
605 *Chem. Geol.*, 114, 315-239, 1994.
- 606 Canfield, D. E.: *Oxygen, a four billion year history*, Princeton Univ. Press, 2014.
- 607 Canfield, D. E., Thamdrup, B., and Kristensen, E.: Aquatic geomicrobiology, in: *Advances*  
608 *in Marine Biology*, edited by: Southward, A. J., Tyler, P. A., Young, C. M., and  
609 Fuiman, L. A., Elsevier Academic Press, Amsterdam, vol. 48, 640 pp., 2005.
- 610 Cathalot, C. et al. Cold-water coral reefs and adjacent sponge grounds: Hotspots of  
611 benthic respiration and organic carbon cycling in the deep sea. *Front. Mar. Sci.* 2,  
612 37. doi: 10.3389/fmars.2015.00037, 2015.



- 613 Cole, J. J., Carpenter, S. R., Pace, M. L., Van de Bogert, M. C., Kitchell, J. L. and Hodgson, J.  
614 R.: Differential support of lake food webs by three types of terrestrial organic  
615 carbon. *Ecol. Lett.*, 9, 558-568, 10.1111/j.1461-0248.2006.00898.x, 2006.
- 616 Cowie, G.: The biogeochemistry of Arabian Sea surficial sediments: A review of recent  
617 studies, *Prog. Oceanogr.*, 65, 260-289, 2005.
- 618 Cowie, G.L., Hedges, J.I.: Biochemical indicators of diagenetic alteration in natural  
619 organic-matter mixtures. *Nature* 369 (6478), 304-307, 1994.
- 620 Cowie, G.L, Mowbray S., Lewis M., Matheson H., McKenzie R.: Carbon and nitrogen  
621 elemental and stable isotopic compositions of surficial sediments from the  
622 Pakistan margin of the Arabian Sea Deep-Sea Res. II: 56, 271-282, 2009.
- 623 Cowie, G.L, Hedges J.I., and Calvert S.E.: Sources and relative reactivities of amino acids,  
624 neutral sugars, and lignin in an intermittent anoxic marine environment.  
625 *Geochim. Cosmochim. Acta* 56, 1963-1978, 1992.
- 626 Danovaro R., Dell'Anno A. and Fabiano M.: Bioavailability of organic matter in the  
627 sediments of the Porcupine Abyssal Plain, northeastern Atlantic. *Mar. Ecol. Prog.*  
628 *Ser.*, 220, 2001.
- 629 Danovaro, R., Dell'Anno, A., Corinaldesi, C., Magagnini, M., Noble, R., Tamburini, C., and  
630 Weinbauer, M.: Major viral impact on the functioning of benthic deep-sea  
631 ecosystems, *Nature*, 454, 1084-1088, 2008.
- 632 Danovaro, R., Corinaldesi, C., Dell'Anno, A., Fuhrman, J.A., Middelburg, J.J., Noble, R.T. &  
633 Suttle, C.A.: Marine viruses and global climate change. *FEMS Microbiology*  
634 *Reviews*, 35 (6), 993-1034, 2011.
- 635 Danovaro, R., Dell'Anno, A., Corinaldesi, C., Rastelli, E., Cavicchioli, R., Krupovic, M.,  
636 Noble, R.T., Nunoura, T., and Prangishvili, D.: Virus mediated archaeal hecatomb  
637 in the deep seafloor. *Science Adv.* 2, e1600492, 2016.
- 638 Dauwe B. and Middelburg J. J.: Amino acids and hexosamines as indicators of organic  
639 matter degradation state in North Sea sediments. *Limnol. Oceanogr.* 43, 782-  
640 798, 1998.
- 641 Dauwe, B., Middelburg, J. J., Herman, P. M. J., and Heip, C. H. R.: Linking diagenetic  
642 alteration of amino acids and bulk organic matter reactivity, *Limnol. Oceanogr.*,  
643 44, 1809-1814, 1999.
- 644 Dell'Anno A., Fabiano M., Mei M. L. and Danovaro R.: Enzymatically hydrolysed protein  
645 and carbohydrate pools in deep-sea sediments: estimates of the potentially  
646 bioavailable fraction and methodological considerations. *Mar. Ecol. Prog. Ser.*  
647 196, 15-23, 2000.
- 648 de Goeij, J. M., van Oevelen, D., Vermeij, M. J. A., Osinga, R., Middelburg, J. J., de Goeij, A. F.  
649 P. M. and Admiraal, W.: Surviving in a Marine Desert: The Sponge Loop Retains  
650 Resources Within Coral Reefs, *Science*, 342, 108-110, 10.1126/science.1241981,  
651 2013.
- 652 De Ruiter P.C., Neutel A-M., and Moore J.C.: Energetics, patterns of interaction strengths,  
653 and stability in real ecosystems. *Science* 269: 1257-1260, 1995
- 654 Diaz, R. J., and Rosenberg, R.: Marine benthic hypoxia: A review of its ecological effects  
655 and the behavioural responses of benthic macrofauna. *Ann. Rev. Ocean. Mar.*  
656 *Biol.*, Vol 33, 245-303, London, 1995.
- 657 Diaz, R. J., and Rosenberg, R.: Spreading dead zones and consequences for marine  
658 ecosystems, *Science*, 321, 926-929, 2008.
- 659 Dippold, M.A., and Kuzyakov, Y. Direct incorporation of fatty acids into microbial  
660 phospholipids in soils: position-specific labeling tells the story. *Geochim.*  
661 *Cosmochim. Acta*, 174, 211-221, 2016.
- 662 Dorgan, K.M., Jumars, P.A., Johnson, B.D., Boudreau, B.P., and Landis, E.: Burrowing by  
663 crack propagation: efficient locomotion through muddy sediments. *Nature* 433,  
664 475, 2005.
- 665 Duarte, C. M., Middelburg, J. J. and Caraco, N.: Major role of marine vegetation on the  
666 oceanic carbon cycle. *Biogeosciences*, 2, 1-8, 2005.



- 667 Evrard, V., Soetaert, K., Heip, C. H. R., Huettel, M., Xenopoulos, M. A. and Middelburg, J. J.:  
668 Carbon and nitrogen flows through the benthic food web of a photic subtidal  
669 sandy sediment, *Mar. Ecol. Prog. Ser.*, 416, 1-16, 10.3354/meps08770, 2010.
- 670 Evrard, V., Huettel, M., Cook, P. L. M., Soetaert, K., Heip, C. H. R. and Middelburg, J. J.:  
671 Importance of phytodetritus and microphytobenthos for heterotrophs in a  
672 shallow subtidal sandy sediment, *Mar. Ecol. Prog. Ser.*, 455, 13-31,  
673 10.3354/meps09676, 2012.
- 674 Fiore C.L., Freeman C.J. and Kujawinski E.B.: Sponge exhalant seawater contains a unique  
675 chemical profile of dissolved organic matter. *PeerJ* 5:e2870  
676 DOI10.7717/peerj.2870, 2017
- 677 Gage J.D. and Tyler P.A.: *Deep-sea biology: A natural history of organisms at the deep-sea*  
678 *floor*. Cambridge Univer. Press, 1991.
- 679 Gattuso, J.-P., Gentili, B., Duarte, C. M., Kleyvas, J. A., Middelburg, J. J. and Antoine, D.:  
680 Light availability in the coastal ocean: impact on the distribution of benthic  
681 photosynthetic organisms and their contribution to primary production,  
682 *Biogeosciences*, 3, 489-513, 2006.
- 683 Gray J.S. and Elliot, M.: *Ecology of marine sediments*. 2<sup>nd</sup> edition, Oxford Univer. Press,  
684 2009.
- 685 Gong C. and Hollander D.: Differential contribution of bacteria to sedimentary organic  
686 matter in oxic and anoxic environments, Santa Monica Basin, California.  
687 *Geochim. Cosmochim. Acta* 26, 545-563, 1997.
- 688 Guilini, K., Van Oevelen, D., Soetaert, K., Middelburg, J. J., and Vanreusel, A.: Nutritional  
689 importance of benthic bacteria for deep-sea nematodes from the Arctic ice  
690 margin: Results of an isotope tracer experiment, *Limnol. Oceanogr.*, 55, 1977–  
691 1989, 2010.
- 692 Glud, R. N.: Oxygen dynamics of marine sediments, *Mar. Biol. Res.*, 4, 243-289, 2008.
- 693 Grutters M., van Raaphorst W., Epping E., Helder J., de Leeuw J. W., Glavin D. P. and Bada  
694 J.: Preservation of amino acids from in situ-produced bacterial cell wall  
695 peptidoglycans in northeastern Atlantic continental margin sediments. *Limnol.*  
696 *Oceanogr.* 47, 1521–1524, 2002.
- 697 Gutierrez J.L., Jones C.G., Strayer D.L., Iribarne O.O.: Mollusks as ecosystem engineers: the  
698 role of shell production in aquatic habitats. *Oikos* 101:79–90, 2003.
- 699 Hamels, I., Muylaert, K., Casteleyn, G., and Vyverman, W.: Uncoupling of bacterial  
700 production and flagellate grazing in aquatic sediments: a case study from an  
701 intertidal flat, *Aquat. Microb. Ecol.*, 25, 31–42, 2001.
- 702 Hannides, A. K. and Aller, R.C.: Priming effect of benthic gastropod mucus on  
703 sedimentary organic matter remineralization., *Limnology &*  
704 *Oceanography*, 61, 2016, p. 1640. [doi:10.1002/lno.10325](https://doi.org/10.1002/lno.10325)
- 705 Hartgers W. A., Damste J. S. S., Requejo A. G., Allan J., Hayes J. M. and De Leeuw J. W.:  
706 Evidence for only minor contributions from bacteria sedimentary carbon. *Nature*  
707 369, 224–227, 1994.
- 708 Hartnett, H. E., and Devol, A. H.: Role of a strong oxygen-deficient zone in the  
709 preservation and degradation of organic matter: A carbon budget for the  
710 continental margins of northwest Mexico and Washington State, *Geochim.*  
711 *Cosmochim. Acta*, 67, 247-264, 2003.
- 712 Hartnett, H. E., Keil, R. G., Hedges, J. I., and Devol, A. H.: Influence of oxygen exposure  
713 time on organic carbon preservation in continental margin sediments, *Nature*,  
714 391, 572-574, 1998.
- 715 Hedges, J. I., and Keil, R. G.: Sedimentary Organic-Matter Preservation - an Assessment  
716 and Speculative Synthesis, *Mar. Chem.*, 49, 81-115, 1995.
- 717 Hedges J. I., Eglinton G., Hatcher P. G., Kirchman D. L., Arnosti C., Derenne S., Evershed R.  
718 P., Kögel-Knabner I., de Leeuw J. W., Littke R., Michaelis W. and Rullkötter J.: The  
719 molecularly-uncharacterized component of nonliving organic matter in natural  
720 environments. *Org. Geochem.* 31, 945–958, 2000.



- 721 Herman, P. M. J., Middelburg, J. J., Van de Koppel, J., and Heip, C. H. R.: Ecology of  
722 estuarine macrobenthos. *Advances in Ecological Research*, 29, 195-240, 1999.
- 723 Hoer, D.R., Gibson P.J., Tommerdahl, J.P., Lindquist N.L. and Martens C.S.: Consumption of  
724 dissolved organic carbon by Caribbean reef sponges. *Lim. Ocean.* (2017, first  
725 online).
- 726 Hondeveld, B. J. M., Bak, R. P. M., and Vanduyf, F. C.: Bacterivory by heterotrophic  
727 nanoflagellates in marine sediments measured by uptake of fluorescently  
728 labelled bacteria, *Mar. Ecol.-Prog. Ser.*, 89, 63–71, 1992.
- 729 Huguet, C, de Lange, G.J., Gustafsson, Ö, Middelburg, J.J., Sinninghe Damsté, J.S. and  
730 Schouten, S.: Selective preservation of soil organic matter in oxidized marine  
731 sediments (Madeira Abyssal Plain) *Geochimica et Cosmochimica Acta*, 72, no. 24,  
732 pp. 6061-6068. DOI: 10.1016/j.gca.2008.09.021, 2008.
- 733 Huettel, M., Berg, P., and Kostka J.E.: Benthic exchange and biogeochemical cycling in  
734 permeable sediments. *Ann. Rev. Marine Sciences*, 6, 23-51, 2014.
- 735 Jessen G. L., Lichtschlag A., Ramette A., Pantoja S., Rossel P.E., Schubert C.J., Struck U.,  
736 Boetius A.: Hypoxia causes preservation of labile organic matter and changes  
737 microbial community composition (Black Sea shelf) *Science Advances* 3(2),  
738 e1601897; doi:10.1126/sciadv.1601897, 2017.
- 739 Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate  
740 reduction, *Nature*, 296, 643-645, 1982.
- 741 Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark),  
742 *Limnol. Oceanogr.*, 22, 814-832, 1977.
- 743 Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Schulz and M. Zabel,  
744 *Marine geochemistry*, 169-206, 2006
- 745 Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial  
746 loop: dissolved organic carbon path- way to heterotrophic bacteria from  
747 incomplete ingestion, digestion and absorption in animals, *Deep-Sea Res. Pt. A*,  
748 36, 483– 495, 1989.
- 749 Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact  
750 on sediment bacteria, *Mar. Ecol.-Prog. Ser.*, 49, 163–169, 1988.
- 751 Kemp, P. F.: The fate of benthic bacterial production, *Rev. Aquat. Sci.*, 2, 109–124, 1990.
- 752 Koho, K. A., Nierop, K. G. J., Moodley, L., Middelburg, J. J., Pozzato, L., Soetaert, K., van der  
753 Plicht, J., and Reichart, G.- J.: Microbial bioavailability regulates organic matter  
754 preservation in marine sediments, *Biogeosciences*, 10, 1131–1141,  
755 doi:10.5194/bg-10-1131-2013, 2013.
- 756 Keil R. G. and Fogel M. L.: Reworking of amino acid in marine sediments: stable carbon  
757 isotopic composition of amino acids in sediments along the Washington coast.  
758 *Limnol. Oceanogr.* 46, 14–23, 2001.
- 759 Keil R. G., Tsamakis E. and Hedges J. I.: Early diagenesis of particulate amino acids in  
760 marine systems. In *Perspectives in Amino Acid and Protein Geochemistry* (eds.  
761 G. A. Goodfriend, M. J. Collins, M. L. Fogel, S. E. Macko and J. F. Wehmiller). Oxford  
762 University Press, 2000.
- 763 Krumins, J. A., van Oevelen, D., Bezemer, T. M., De Deyn, G. B., Hol, W. H. G., van Donk, E.,  
764 de Boer, W., de Rooter, P. C., Middelburg, J. J., Monroy, F., Soetaert, K., Thebault, E.,  
765 van de Koppel, J., van Veen, J. A., Viketoft, M. and van der Putten, W. H.: Soil and  
766 Freshwater and Marine Sediment Food Webs: Their Structure and Function,  
767 *Bioscience*, 63, 35-42, 10.1525/bio.2013.63.1.8, 2013.
- 768 Lee C., Wakeham S. G. and Hedges J. I.: Composition and flux of particulate amino acids  
769 and chloropigments in equatorial Pacific seawater and sediments. *Deep Sea Res.*  
770 *Part I* 47, 1535–1568, 2000.
- 771 Lenton T. and Watson A.: *Revolutions that made the earth*. Oxford Univer. Press, 2011.
- 772 Lomstein B. A., Jørgensen B. B., Schubert C. J. and Niggemann J.: Amino acid biogeo- and  
773 stereochemistry in coastal Chilean sediments. *Geochim. Cosmochim. Acta* 70,  
774 2970–2989. doi:10.1016/j.gca.2006.03.015, 2006.





- 775 Lomstein B. Aa., Langerhuus A. T., D'Hondt S., Jørgensen B. B. and Spivack A.: Endospore  
776 abundance, microbial growth and necromass turnover in deep subseafloor  
777 sediment. *Nature* 484, 101–104, 2012.
- 778 Levin, L. A.: Oxygen minimum zone benthos: Adaptation and community response to  
779 hypoxia. *Ann. Rev. Oceanogr. Mar. Biol.*, 41, 1–45, 2003.
- 780 Levin, L.A., Ekau W., Gooday A.J., Jorissen F., Middelburg J.J., Naqvi W., Neira C., Rabalais  
781 N.N., Zhang, J.: Effects of Natural and Human-Induced Hypoxia on Coastal  
782 Benthos. *Biogeosc. Disc.* 6, 3563–3654, 2009.
- 783 Meysman, F.J.R.; Middelburg, J.J.; Herman, P.M.J. and Heip, C.H.R.: Reactive transport in  
784 surface sediments: 2. Media: an object-oriented problem-solving environment  
785 for early diagenesis. *Comput. Geosci.* 29: 301–318, 2003.
- 786 Meysman, F. J. R., Middelburg, J. J., and Heip, C. H. R.: Bioturbation: a fresh look at  
787 Darwin's last idea, *Trends Ecol. Evol.*, 21, 688–695, 2006.
- 788 Meysman, F. J. R., Boudreau, B. P., and Middelburg, J. J.: Relations between local, nonlocal,  
789 discrete and continuous models of bioturbation, *J. Mar. Res.*, 61, 391–410, 2003.
- 790 Meysman, F.J.R., Boudreau, B.P. and Middelburg, J.J.: When and why does bioturbation  
791 lead to diffusive mixing? *Journal of Marine Research* 68, 881–920, 2010.
- 792 Middelburg, J.J., A simple rate model for organic-matter decomposition in  
793 marine-sediments. *Geochimica et Cosmochimica Acta* 53, 1577–1581, 1989.
- 794 Middelburg, J. J.: Chemoautotrophy in the ocean, *Geophys. Res. Lett.*, 38, L24604,  
795 10.1029/2011GL049725, 2011.
- 796 Middelburg, J. J.: Stable isotopes dissect aquatic food webs from the top to the bottom.  
797 *Biogeosciences*, 11, 2357–2371, 2014
- 798 Middelburg, J. J. & Levin, L. A.: Coastal hypoxia and sediment biogeochemistry.  
799 *Biogeosciences*, 6, 1273–1293, 2009.
- 800 Middelburg, J. J., Vlug, T., and Van der Nat, F.: Organic matter mineralization in marine  
801 systems. *Global Planet. Change*, 8, 47–58, 1993.
- 802 Middelburg, J. J., Soetaert, K., Herman, P. M. J., and Heip, C. H. R.: Denitrification in marine  
803 sediments: A model study, *Glob. Biogeochem. Cycle*, 10, 661–673, 1996.
- 804 Middelburg JJ, Nieuwenhuize J, van Breugel P: Black carbon in marine sediments. *Mar*  
805 *Chem* 65:245–252, 1999.
- 806 Middelburg, J. J., Barranguet, C., Boschker, H. T. S., Herman, P. M. J., Moens, T., and Heip, C.  
807 H. R.: The fate of intertidal microphytobenthos carbon: An in situ C-13-labeling  
808 study, *Limnol. Oceanogr.*, 45, 1224–1234, 2000.
- 809 Moodley, L., Middelburg, J. J., Boschker, H. T. S., Duineveld, G. C. A., Pel, R., Herman, P. M. J.  
810 and Heip, C. H. R.: Bacteria and Foraminifera: key players in a short-term deep-  
811 sea benthic response to phytodetritus, *Mar. Ecol. Prog. Ser.*, 236, 23–29,  
812 10.3354/meps236023, 2002.
- 813 Moodley, L., Middelburg, J. J., Soetaert, K., Boschker, H. T. S., Herman, P. M. J. and Heip, C.  
814 H. R.: Similar rapid response to phytodetritus deposition in shallow and deep-  
815 sea sediments, *J. Mar. Res.*, 63, 457–469, 10.1357/0022240053693662, 2005a.
- 816 Moodley, L., Middelburg, J. J., Herman, P. M. J., Soetaert, K., and de Lange, G. J.:  
817 Oxygenation and organic-matter preservation in marine sediments: Direct  
818 experimental evidence from ancient organic carbon-rich deposits, *Geology*, 33,  
819 889–892, 2005b.
- 820 Müller, P.J.: C/N ratios in Pacific deep-sea sediments: effect of inorganic ammonium and  
821 organic nitrogen compounds sorbed by clays. *Geochim. Cosmochim. Acta*, 41,  
822 549–553, 1997.
- 823 Nierop, K.G.J., Reichart, G.-J., Veld, H. and Sinninghe Damsté, J.S. The influence of oxygen  
824 exposure time on the composition of macromolecular organic matter as revealed  
825 by surface sediments on the Murray Ridge (Arabian Sea). *Geochim. Cosmochim.*  
826 *Acta* 206: 40–56. [dx.doi.org/10.1016/j.gca.2017.02.032](https://doi.org/10.1016/j.gca.2017.02.032), 2017.
- 827 Oakes, J. M., Eyre, B. D. and Middelburg, J. J.: Transformation and fate of  
828 microphytobenthos carbon in subtropical shallow subtidal sands: A C-13-



- 829 labeling study, *Limnol. Oceanogr.*, 57, 1846-1856, 10.4319/lo.2012.57.06.1846,  
830 2012.
- 831 Pimm, S.L., Lawton, J.H. and Cohen, J.E.: Food web patterns and their consequences.  
832 *Nature* 350,669-674, 1991.
- 833 Pomeroy, L.: The ocean's food web, a changing paradigm, *Bio-Science*, 24, 499-504,  
834 1974.
- 835 Rabouille, C., Gaillard, J.-F.: Towards the EDGE: early diagenetic global explanation. A  
836 model depicting the early diagenesis of organic matter, O<sub>2</sub>, NO<sub>3</sub>, Mn, and PO<sub>4</sub>.  
837 *Geochimica et Cosmochimica Acta* 55, 2511-2525, 1991.
- 838 Raghoebarsing A.A., Pol A., van de Pas-Schoonen K.T., Smolders A.J.P., Ettwig K.F.,  
839 Rijpstra W.I.C., Schouten S., Sinninghe Damsté J.S., Op den Camp H.J.M., Jetten  
840 M.S.M., Strous M.: A microbial consortium couples anaerobic methane oxidation  
841 to denitrification. *Nature* 440: 918-921, 2006.
- 842 Rhoads, D.C., and Morse, J.W.: Evolutionary and ecological significance of oxygen-  
843 deficient marine basins, *Lethaia*, 4, 413-428, 1971.
- 844 Rhoads, D.C.: Organism-sediment relations on the muddy sea floor. *Oceanogr. Mar. Biol.*  
845 *Ann. Rev.* 12, 263-300, 1974.
- 846 Rice, D.L.: Early diagenesis in bioadvective sediments: relationships between the  
847 diagenesis of beryllium-7, sediment reworking rates, and the abundance of  
848 conveyor-belt deposit-feeders. *Journal of Marine Research* 44, 149-184, 1986.
- 849 Rice, D.L. and Rhoads, D.C.: Early diagenesis of organic matter and the nutritional value  
850 of sediment. In: *Ecology of Marine Deposit Feeders* (Ed. by G. Lopez, G. Taghon  
851 and J. Levinton), pp. 309-317. Springer, Berlin, 1989.
- 852 Rix, L. et al. Coral mucus fuels the sponge loop in warm- and cold-water coral reef  
853 ecosystems. *Sci. Rep.* 6, 18715; doi: 10.1038/srep18715, 2016.
- 854 Roberts, J. M., Wheeler, A. J. and Freiwald A. Reefs of the deep, The biology and geology  
855 of cold-water coral ecosystems. *Science* 312, 543-547, 2006.
- 856 Rossi S., Bramanti, L., Gori, A and O. Covadonga. An overview of the animal forest of the  
857 world. In Rossi (ed). *Marine Animal Forest*. Springer, pp. 1-25, 2017.
- 858 Schouten, S., Middelburg, J.J., Hopmans, E.C. & Sinninghe Damsté, J.S.: Fossilization and  
859 degradation of intact polar lipids in deep subsurface sediments: A theoretical  
860 approach. *Geochimica et Cosmochimica Acta*, 74, 3806-3814, 2010.
- 861 Sinninghe Damsté, J. S., Rijpstra, W. I. C., and Reichart, G. J.: The influence of oxic  
862 degradation on the sedimentary biomarker record II. Evidence from Arabian Sea  
863 sediments, *Geochim. Cosmochim. Acta*, 66, 2737-2754, 2002.
- 864 Soetaert, K., Herman, P. M. J., and Middelburg, J. J.: A model of early diagenetic processes  
865 from the shelf to abyssal depths, *Geochim. Cosmochim. Acta*, 60, 1019-1040,  
866 1996.
- 867 Soetaert, K., Middelburg, J. J., Herman, P. M. J., and Buis, K.: On the coupling of benthic  
868 and pelagic biogeochemical models, *Earth-Sci. Rev.*, 51, 173-201, 2000.
- 869 Soetaert, K., Mohn, C., Rengstorf, A., Grehan, A., and van Oevelen, D.: Ecosystem  
870 engineering creates a direct nutritional link between 600-m deep cold-water  
871 coral mounds and surface productivity, *Sci. Rep.*, 6, 35057,  
872 doi:10.1038/srep35057, 2016.
- 873 Steenbergh A.K., Bodelier P.L.E. Heidal, M., Slomp, C.P., and Laanbroek H.J.: Does  
874 microbial stoichiometry modulate eutrophication of aquatic ecosystems?  
875 *Environ. Microbiol.* 15: 1572-1579, 2013.
- 876 Sterner R.W. and Elser J.J.: *Ecological Stoichiometry*. Princeton University Press, 439 pp,  
877 2002.
- 878 Strous, M., and Jetten, M. S. M.: Anaerobic oxidation of methane and ammonium, *Annu.*  
879 *Rev. Microbiol.*, 58, 99-117, 2004.
- 880 Sun M. Y., Aller R. C., Lee C. and Wakeham S. G.: Enhanced degradation of algal lipids by  
881 benthic macrofaunal activity: effect of *Yoldia limatula*. *J. Mar. Res.* 57, 775-804,  
882 1999.



- 883 Tegelaar, E. W., de Leeuw, J. W., Derenne, S., and Largeau, C.: A reappraisal of kerogen  
884 formation. *Geochim. Cosmochim. Acta* 53, 3103–3106. doi:10.1016/0016-  
885 7037(89)90191-9, 1989.
- 886 Thomas C. J. and Blair N. E.: Transport and digestive alteration of uniformly <sup>13</sup>C-labelled  
887 diatoms in mudflat sediments. *J. Mar. Res.* 60, 517–535, 2002.
- 888 Van Cappellen, P. and Wang, Y. F.: Cycling of iron and manganese in surface sediments:  
889 A general theory for the coupled transport and reaction of carbon, oxygen,  
890 nitrogen, sulfur, iron, and manganese, *Am. J. Sci.*, 296, 197–243,  
891 doi:10.2475/ajs.296.3.197, 1996.
- 892 Vandewiele S., Cowie G., Soetaert K. and Middelburg J. J.: Amino acid biogeochemistry  
893 and organic matter degradation state across the Pakistan margin oxygen  
894 minimum zone. *Deep Sea Res. Part II* 56, 318–334, 2009.
- 895 Vanni, M. J. and McIntyre, P. B.: Predicting nutrient excretion of aquatic animals with  
896 metabolic ecology and ecological stoichiometry: a global synthesis. *Ecology*, 97:  
897 3460–3471. doi:10.1002/ecy.1582, 2016.
- 898 Vanni, M. J.: Nutrient cycling by animals in freshwater ecosystems. *Annual Review of*  
899 *Ecology and Systematics* 33:341–370, 2002.
- 900 Van Nugteren, P., Moodley, L., Brummer, G.-J., Heip, C.H.R., Herman, P.M.J., Middelburg,  
901 J.J.: Seafloor ecosystem functioning: the importance of organic matter priming.  
902 *Marine Biology* 156, 2277–2287, 2009a.
- 903 van Nugteren, P., Herman, P.M., Moodley, L., Middelburg, J.J., Vos, M.: Spatial distribution  
904 of detrital resources determines the outcome of competition between bacteria  
905 and a facultative detritivorous worm. *Limnology and Oceanography* 54, 1413–  
906 1419, 2009b.
- 907 van Oevelen, D., Moodley, L., Soetaert, K. and Middelburg, J. J.: The trophic significance of  
908 bacterial carbon in a marine intertidal sediment: Results of an in situ stable  
909 isotope labeling study, *Limnol. Oceanogr.*, 51, 2349–2359, 2006.
- 910 van Oevelen, D., Van den Meersche, K., Meysman, F. J. R., Soetaert, K., Middelburg, J. J. and  
911 Vezina, A. F.: Quantifying Food Web Flows Using Linear Inverse Models,  
912 *Ecosystems*, 13, 32–45, 10.1007/s10021-009-9297-6, 2010.
- 913 Veuger, B., Eyre, B. D., Maher, D. and Middelburg, J. J.: Nitrogen incorporation and  
914 retention by bacteria, algae, and fauna in a subtropical intertidal sediment: An in  
915 situ N-15-labeling study, *Limnol. Oceanogr.*, 52, 1930–1942,  
916 10.4319/lo.2007.52.5.1930, 2007.
- 917 Veuger B., van Oevelen D. and Middelburg J. J.: Fate of microbial nitrogen, carbon,  
918 hydrolysable amino acids, monosaccharides, and fatty acids in sediment.  
919 *Geochim. Cosmochim. Acta* 83, 217–233, 2012.
- 920 Wakeham S. G., Lee C., Hedges J. I., Hernes P. J. and Peterson M. L.: Molecular indicators  
921 of diagenetic status in marine organic matter. *Geochim. Cosmochim. Acta* 61,  
922 5363–5369, 1997.
- 923 Witte, U., Wenzhofer, F., Sommer, S., Boetius, A., Heinz, P., Aberle, N., Sand, M., Cremer, A.,  
924 Abraham, W. R., Jorgensen, B. B. and Pfannkuche, O.: In situ experimental  
925 evidence of the fate of a phytodetritus pulse at the abyssal sea floor, *Nature*, 424,  
926 763–766, 10.1038/nature01799, 2003.
- 927 Woulds, C., Andersson, J. H., Cowie, G. L., Middelburg, J. J. and Levin, L. A.: The short-term  
928 fate of organic carbon in marine sediments: Comparing the Pakistan margin to  
929 other regions, *Deep-Sea Research Part II-Topical Studies in Oceanography*, 56,  
930 393–402, 10.1016/j.dsr2.2008.10.008, 2009.
- 931 Woulds, C., Cowie, G. L., Levin, L. A., Andersson, J. H., Middelburg, J. J., Vandewiele, S.,  
932 Lamont, P. A., Larkin, K. E., Gooday, A. J., Schumacher, S., Whitcraft, C., Jeffreys, R.  
933 M. and Schwartz, M.: Oxygen as a control on seafloor biological communities and  
934 their roles in sedimentary carbon cycling, *Limnol. Oceanogr.*, 52, 1698–1709,  
935 10.4319/lo.2007.52.4.1698, 2007.



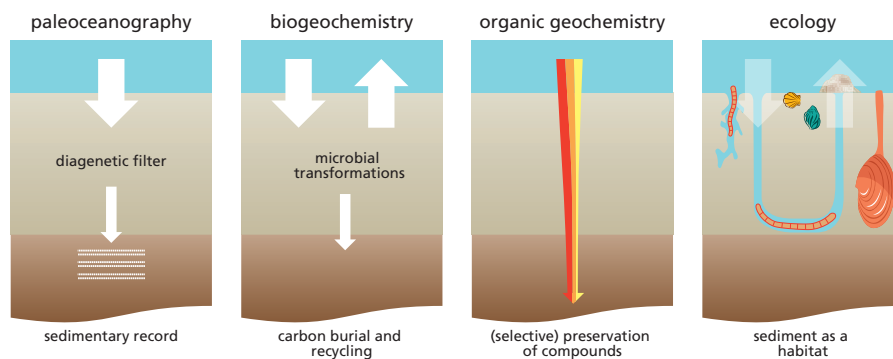
- 936    Woulds, C., Middelburg, J. J. and Cowie, G. L.: Alteration of organic matter during infaunal  
937            polychaete gut passage and links to sediment organic geochemistry. Part I:  
938            Amino acids. *Geochimica Et Cosmochimica Acta*, 77, 396-414, 2012.
- 939    Woulds, C., Middelburg, J. J. & Cowie, G. L.: Alteration of organic matter during infaunal  
940            polychaete gut passage and links to sediment organic geochemistry. Part II: Fatty  
941            acids and aldoses. *Geochimica Et Cosmochimica Acta*, 136, 38-59, 2014.
- 942    Woulds, C., Bouillon, S., Cowie, G. L., Drake, E., Middelburg, J.J. and Witte, U.: Patterns of  
943            carbon processing at the seafloor - the role of faunal and microbial communities  
944            in moderating carbon flows. *Biogeosciences*, 13, 4343-4357, 2016.
- 945



- 946  
947  
948 Figure 1. Different views, approaches and interests on carbon processing in marine  
949 sediments. Paleocceanographers focus on the sedimentary record,  
950 biogeochemists quantify carbon burial and recycling, organic geochemists study  
951 alteration of organic matter and ecologists focus on carbon as food for organisms  
952 living in the sediment  
953  
954 Figure 2. Conceptual model of organic matter (OM) degradation and re-oxidation  
955 pathways (based on Middelburg and Levin, 2009).  
956  
957 Figure 3. Carbon processing in marine sediments on the short-term (left) and the long-  
958 term (right). Organic matter produced in the sunlit layer of the ocean and  
959 delivered to the sediments is either consumed by organisms or buried. The  
960 organic matter consumed by organisms is used to synthesize biomass or  
961 metabolized to carbon dioxide and nutrients. On the long-term or at steady-  
962 state, i.e. the biomass of benthic organisms does not change, the benthic  
963 community can be considered a black box diverting organic matter either into  
964 metabolites or into the geosphere (burial).  
965  
966 Figure 4. Conceptual diagram showing the relationships between molecular  
967 uncharacterizable organics, deposited phytodetritus and secondary production.  
968 Phytodetritus is degraded preferentially and new biomass is formed, which after  
969 death of the organisms is added to the pool of detritus and subject to  
970 degradation. Multiple cycles of organic matter processing eventually results in  
971 the formation of molecular uncharacterizable organic matter.  
972  
973 Figure 5. Organic matter supply to sediments. 1. The traditional view of organic matter  
974 settling passively from the water column (donor control). 2. Sediments in the  
975 photic zone are inhabited by benthic microalgae that produce new organic  
976 matter in situ and grazing animals can impact the growth of these organisms. 3.  
977 Bioturbating animals transfer labile carbon from the sediment surface layer to  
978 deeper layers in the sediments. 4. Suspension feeding organisms enhance the  
979 transfer of suspended particulate matter from the water column to the  
980 sediments (biodeposition). 5. Sponge consume dissolved organic carbon and  
981 produce cellular debris that can be consumed by benthic organisms (i.e. the  
982 sponge loop).  
983  
984 Figure 6. The microbial and inverted microbial loop. In the water column dissolved  
985 organic carbon derived from phytoplankton, zooplankton or microbes (via viral  
986 loop) is consumed by heterotrophic microbes, which in turn are consumed by  
987 protists and small animals with the consequence that carbon flowing through  
988 dissolved organic carbon pools eventually can be used by larger animals  
989 (microbial loop). In sediments, the dissolved organic carbon (from viral lysis  
990 and other sources) is also consumed by heterotrophic microbes but this carbon  
991 is inefficiently transferred to animals. The engineering activities of animals are  
992 key in delivering labile organic matter (phytodetritus) to microbes living in the  
993 subsurface (inverted microbial loop).  
994  
995  
996  
997



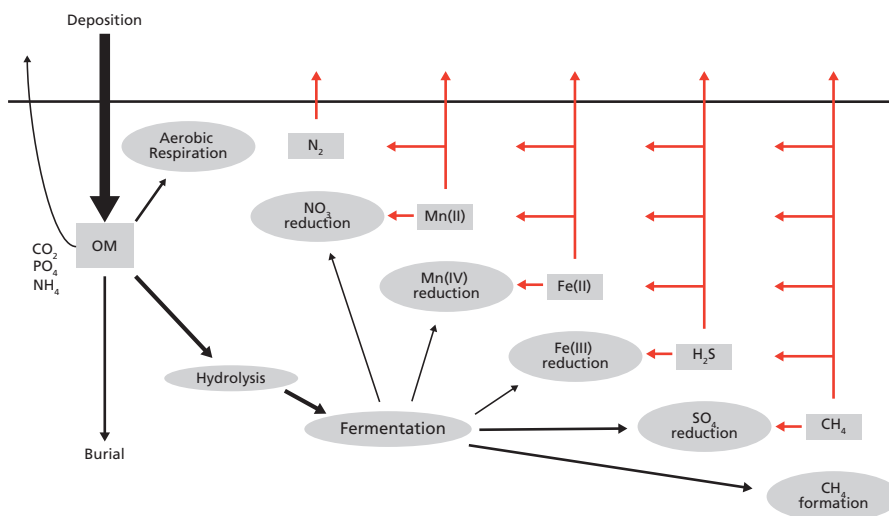
998 Figure 1. Different views, approaches and interests on carbon processing in marine  
999 sediments. Paleocyanographers focus on the sedimentary record,  
1000 biogeochemists quantify carbon burial and recycling, organic geochemists study  
1001 alteration of organic matter and ecologists focus on carbon as food for organisms  
1002 living in the sediment  
1003



1004  
1005



1006 Figure 2. Conceptual model of organic matter (OM) degradation and re-oxidation  
1007 pathways (based on Middelburg and Levin, 2009).  
1008  
1009

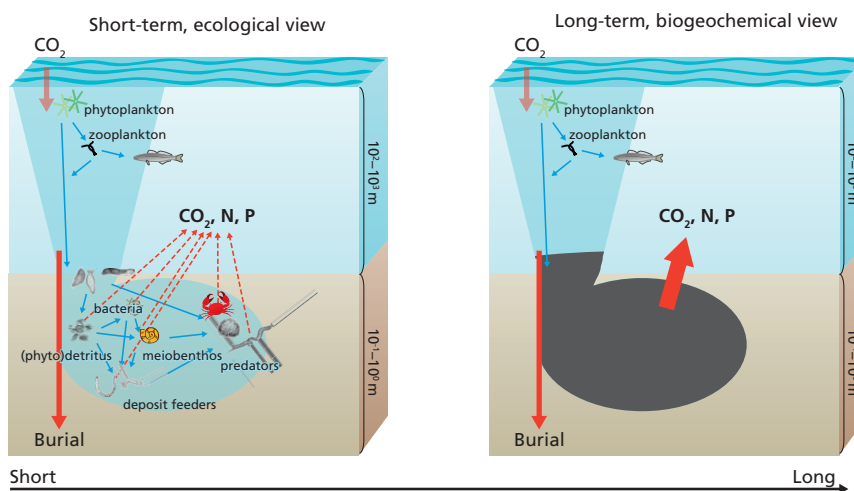


1010  
1011



1012  
1013  
1014  
1015  
1016  
1017  
1018  
1019  
1020  
1021  
1022  
1023

Figure 3. Carbon processing in marine sediments on the short-term (left) and the long-term (right). Organic matter produced in the sunlit layer of the ocean and delivered to the sediments is either consumed by organisms or buried. The organic matter consumed by organisms is used to synthesize biomass or metabolized to carbon dioxide and nutrients. On the long-term or at steady-state, i.e. the biomass of benthic organisms does not change, the benthic community can be considered a black box diverting organic matter either into metabolites or into the geosphere (burial).



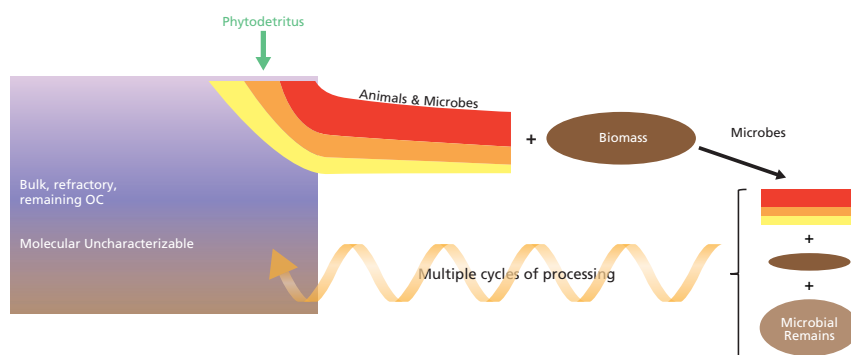
1024





1025  
1026  
1027  
1028  
1029  
1030  
1031  
1032

Figure 4. Conceptual diagram showing the relationships between molecular uncharacterizable organics, deposited phytodetritus and secondary production. Phytodetritus is degraded preferentially and new biomass is formed, which after death of the organisms is added to the pool of detritus and subject to degradation. Multiple cycles of organic matter processing eventually result in the formation of molecular uncharacterizable organic matter.

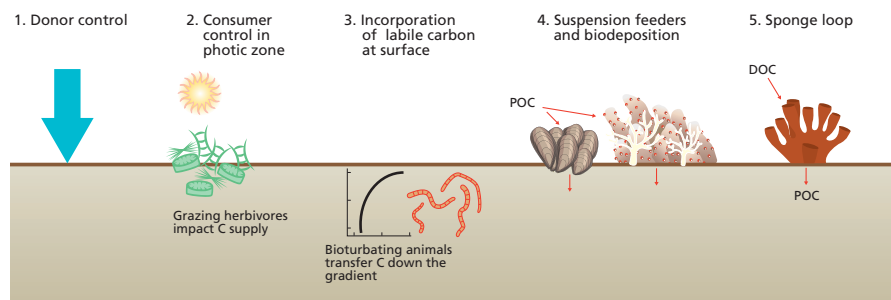


1033  
1034



1035  
1036  
1037  
1038  
1039  
1040  
1041  
1042  
1043  
1044  
1045  
1046  
1047

Figure 5. Organic matter supply to sediments. 1. The traditional view of organic matter settling passively from the water column (donor control). 2. Sediments in the photic zone are inhabited by benthic microalgae that produce new organic matter in situ and grazing animals can impact the growth of these organisms. 3. Bioturbating animals transfer labile carbon from the sediment surface layer to deeper layers in the sediments. 4. Suspension feeding organisms enhance the transfer of suspended particulate matter from the water column to the sediments (biodeposition). 5. Sponges consume dissolved organic carbon and produce cellular debris that can be consumed by benthic organisms (i.e. the sponge loop).



1048  
1049



1050

1051

1052

1053

1054

1055

1056

1057

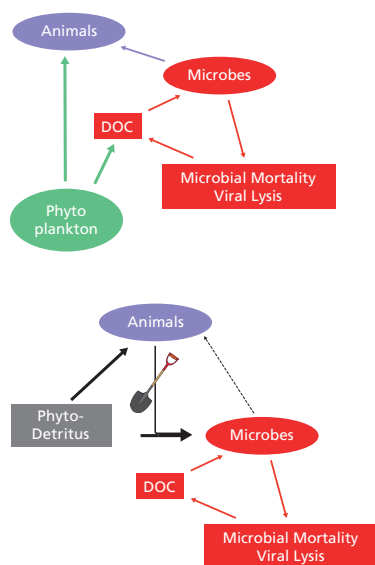
1058

1059

1060

1061

Figure 6. The microbial and inverted microbial loop. In the water column dissolved organic carbon derived from phytoplankton, zooplankton or microbes (via viral lysis) is consumed by heterotrophic microbes, which in turn are consumed by protists and small animals with the consequence that carbon flowing through dissolved organic carbon pools eventually can be used by larger animals (microbial loop). In sediments, the dissolved organic carbon (from viral lysis and other sources) is also consumed by heterotrophic microbes but this carbon is inefficiently transferred to animals. The engineering activities of animals are key in delivering labile organic matter (phytodetritus) to microbes living in the subsurface (inverted microbial loop).



1062

1063

1064

1065