# Reviews and Synthesis: To the bottom of carbon processing at the seafloor 4

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#### 13 Abstract

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15 Organic carbon processing at the seafloor is studied by biogeochemists to quantify burial and respiration, by organic geochemists to elucidate 16 17 compositional changes and by ecologists to follow carbon transfers within food webs. Here I review these disciplinary approaches and discuss where they agree 18 19 and disagree. It will be shown that the biogeochemical approach (ignoring the 20 identity of organisms) and the ecological approach (focussing on growth and 21 biomass of organisms) are consistent on longer time scales. Secondary 22 production by microbes and animals is identified to potentially impact the 23 composition of sedimentary organic matter. Animals impact sediment organic 24 carbon processing by microbes in multiple ways: by governing organic carbon 25 supply to sediments, by aeration via bio-irrigation and by mixing labile organic 26 matter to deeper layers. I will present an inverted microbial loop in which 27 microbes profit from bioturbation rather than animals profiting from microbial 28 processing of otherwise lost dissolved organic resources. Sediments devoid of 29 fauna therefore function differently and are less efficient in processing organic 30 matter with the consequence that more organic matter is buried and transferred 31 from Vernadsky's biosphere to the geosphere.

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### 34 **1 Introduction**

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36 The seawater-sediment interface represents one of the largest interfaces 37 on earth and our knowledge of processes at and fluxes through this dynamic and 38 understudied interface is rather limited. This interface extends a few cm-dm 39 upwards into the water column, i.e. benthic boundary layer (Boudreau and 40 Jørgensen, 1992), as well as few cm-dm into the sediments, i.e. the bioturbated, 41 active surface layer (Berner, 1980; Meysman et al., 2006; Aller, 2013). It serves 42 as a habitat for organisms, governs the partitioning of material being buried or 43 recycled and acts as a filter for the paleorecord (Rhoads, 1974). Processes in the 44 surface sediment layer determine whether remains from organisms (organic 45 matter, biogenic silica) are recycled within the biosphere (short-term cycle) or 46 transferred to the geosphere (long-term cycle) and as such it functions as a key 47 interface in the System Earth.

This pivotal role of the seafloor in processing material deposited has been
studied by scientists from various disciplines with their own interests,

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50 techniques and paradigms (Figure 1). Marine geologists and paleoceanographers study sediments with the primary aim to extract information on past 51 52 environmental conditions using down-core measurements of substances that 53 have survived the processing at the seafloor (Burdige, 2006; Bender, 2013). 54 Biogeochemists quantify the fate of material delivered, in particular how much of 55 that is eventually buried or processed, and determine when and in what form the 56 remaining part is recycled as key nutrients to sustain primary production in the 57 water column (Berner, 1980; Aller, 1980, 2001, 2013; Soetaert et al., 2000). 58 Organic geochemists investigate how organic matter delivered to the seafloor is 59 degraded, transformed or preserved using changes in the composition at the 60 molecular level (Hedges and Keil, 1995; Dauwe et al, 1999; Burdige, 2006; Bianchi and Canuel, 2011). Ecologists focus on the organisms, i.e. the actors 61 62 consuming, producing and transporting the material deposited (Gage and Tyler, 63 1992; Gray and Elliot, 2009; Herman et al., 1999; Krumins et al., 2013).

64 Although these disciplines often study the same topic, e.g. organic matter delivered to the seafloor, they focus on different aspects and usually 65 66 underappreciate or do not incorporate key concepts, findings and approaches 67 from other disciplines. For example, ecologists and biogeochemist studying carbon flows at the seafloor normally ignore detailed molecular information 68 69 available from organic geochemistry (Berner, 1980; Glud, 2008). Bioturbation, 70 biological reworking of sediments (Meysman et al., 2006), is often ignored by 71 paleoceanographers, and biogeochemists (Berner, 1980; Boudreau, 1997) have 72 developed advanced transport-reaction models in which the actors, the animals, 73 mix the sediment but do so without consuming organic matter.

74 Here I present the existing views on organic carbon processing at the 75 seafloor, discuss where they agree and disagree and aim to arrive at an integrated view of carbon processing at the seafloor that is consistent with 76 77 recent views within the organic geochemical, biogeochemical and ecological 78 research communities. This overview is necessarily and admittedly incomplete 79 but rather covers personal interests and presents new concepts on this topic. It 80 is a concise version of the Vernadsky Medal Lecture presented at the 2017 EGU 81 meeting.

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### 2 Biogeochemists focus on quantification of burial and mineralization.

85 Organic matter delivered to marine sediments is either mineralized and 86 the metabolites (carbon dioxide and nutrients) accumulate in pore-waters and 87 exchange with the overlying water, or buried through the steady accumulation of 88 particles (Fig. 2). This biogeochemical view (Berner, 1980; Boudreau, 1997; 89 Aller, 2013) is highly simplified, but for that reason also quantitative and I 90 believe instructive. The percentage of organic matter buried varies from less 91 than one % to a few tens % and is closely and positively related with total 92 sediment accumulation rate (Canfield, 1989, 1994; Middelburg et al., 1993). 93 Since sediment accumulation rates are high in vegetated, coastal, deltaic, shelf 94 and ocean-margin settings, the majority of organic matter burial occurs there, 95 with organic carbon burial in deep-sea sediments accounting for <5 % of the total buried (Berner, 1982; Duarte et al., 2005; Burdige, 2007). In the deep sea 96 97 and other settings with low burial efficiency, almost all organic matter is 98 degraded to inorganic carbon at rates that provide an excellent approximation

99 for organic matter deposition (Jørgensen, 1982; Cai and Reimers, 1995; Glud, 100 2008). Organic matter degradation can be quantified via the consumption of 101 oxygen, the production of dissolved inorganic carbon and through the use of 102 pore-water data and diagenetic models (i.e. reaction-transport models for 103 sediments). At steady state, sediment oxygen consumption provides an accurate 104 measure for total sediment organic matter degradation, independent whether 105 organic matter is degraded aerobically (i.e. with oxygen) or anaerobically (with 106 alternative electron acceptors such as nitrate, metal oxides, sulphate), because 107 almost all reduced metabolites released (ammonium, manganese (II), iron(II), 108 hydrogen sulfide and methane) are re-oxidized (Jørgensen, 1977, 2006; Berner 109 and Westrich, 1985; Aller and Rude, 1988; Soetaert et al., 1996; Boetius et al., 110 2000; Strous and Jetten, 2004; Raghoebarsing et al., 2006; Middelburg and Levin, 111 2009).

112 Biogeochemists have adopted a transport-reaction modeling approach to accurately quantify organic matter processing (Berner, 1980; Boudreau, 1997; 113 114 Burdige, 2006). The basic premise of these diagenetic models is that both 115 particles and solutes are subject to transport and reaction, making them distinct 116 from for instance groundwater transport-reaction models in which normally only solutes and gas phases are mobile (Lichtner, 1996; Appelo, 1996). 117 Transport of solutes is due to molecular diffusion, pore-water advection and 118 119 biologically mediated processes such as enhanced diffusion due to interstitial 120 fauna (Aller and Aller, 1992) and bio-irrigation due to tube and burrow construction and flushing by macrofauna (Aller, 1980, 1984; 2001; Volkenborn 121 et al., 2010; 2016; Kristensen et al., 2012). Particle transport is not only due to 122 123 steady particle deposition but also due to sediment reworking by animals 124 (bioturbation, Boudreau, 1997; Aller, 1994, 2013; Rice, 1986; Meysman et al., 125 2003, 2006, 2010). The reaction terms in these diagenetic models are normally 126 limited to microbial and chemical reactions and are described using zero, first, 127 second order kinetic relationships or Monod/Michaelis-Menten type kinetics 128 (Bouldin, 1968; Berner, 1980; Soetaert et al., 1996; Boudreau, 1997). There is a 129 major inconsistency in the basic conceptual model underlying the (numerical) 130 diagenetic models: animals dominate transport processes via pore-water 131 irrigation and particle mixing, but without consuming any organic matter. This 132 inconsistency has not received much attention because the ruling paradigm 133 within the biogeochemical research community is that animals contribute very 134 little to total carbon processing. Multiple recent studies involving use of <sup>13</sup>C as 135 deliberate tracers show that this premise does not hold on the short term (days 136 to weeks; Blair et al., 1996; Moodley et al., 2002, 2005a; Woulds et al., 2009, 137 2016). Moreover, detailed studies of oxygen consumption have revealed that animals contribute substantially to total sediment oxygen uptake; directly via 138 139 their respiration as well as indirectly via particle and solute mixing (Glud, 2008). 140 Nevertheless, diagenetic models can very accurately reproduce most observations (Soetaert et al., 1996; Berg et al., 2003). 141

Diagenetic models combined with solid-phase and pore-water depth
profiles, sediment-water exchange fluxes and rate measurement have resulted in
a consistent picture of organic matter degradation pathways in marine
sediments (Berner, 1980; Boudreau, 1997; Aller, 2013). These models can
predict where, when and why organic matter oxidation occurs aerobically or
involves nitrate, metal oxides or sulphate as oxidants (Rabouille and Gaillard,

- 148 2001; Boudreau, 1996; Soetaert et al., 1996; Middelburg et al. 1996; van Cappellen and Wang, 1996; Archer et al., 2002; Meysman et al., 2003; Berg et al., 149 2003). They also resolve the re-oxidation of reduced products such as 150 151 ammonium, manganese(II), iron (II), sulfide and methane (Fig. 2) and as such 152 define the scope for aerobic and anaerobic organisms, including the distribution 153 and activity of chemoautotrophs (Middelburg, 2011). Despite these advances, 154 diagenetic models cannot predict organic carbon burial rates, nor do they 155 provide much insight into why organic matter is buried, or why it is either labile 156 (reactive) or refractory. For this we need to have a detailed look at the organic
- 157 geochemistry of sediment organic carbon.
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# 159 **3 Organic geochemists focus on the composition of organic matter** 160 **preserved**

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Organic matter delivered to the seafloor is predominantly produced in the 162 surface sunlit layer of the ocean (Fig. 3). This organic matter is rich in proteins, 163 164 carbohydrates and lipids and generally follows Redfield stoichiometry (Sterner 165 and Elser, 2002; Bianchi and Canuel, 2011). Organic matter processing leads to preferential degradation of the more labile components with the result that 166 organic matter becomes less reactive (Jørgensen, 1979; Westrich and Berner, 167 168 1984; Middelburg, 1989; Arndt et al., 2013) and organic matter composition 169 changes (Fig. 1; Wakeham et al. 1997; Dauwe et al., 1999; Lee et al., 2000). The 170 proportion of organic matter that can be characterized molecularly decreases 171 with progressive degradation, i.e. with water depth or depth downcore 172 (Wakeham et al., 1997; Hedges et al., 2000; Middelburg et al., 1999; Nierop et al., 173 2017). This molecularly uncharacterizable material increases to more than 70% 174 of the total in deep-sea sediment organic matter. The organic geochemical 175 approach to study organic matter processing is limited not only by our inabilities 176 to characterize the majority of the sedimentary organic matter, but also by the 177 simple fact that the degraded fraction cannot be easily studied and we have to 178 base our knowledge on the small fraction of extensively processed organic 179 material remaining.

180 The changes in organic matter composition due to organic matter 181 processing have been utilized to estimate the lability-digestability or the 182 refractory nature of organic matter with various proxies such as chlorophyll to 183 bulk organic matter, fraction of nitrogen present as amino acids, and the 184 contribution of proteins and carbohydrates to total organic matter (Cowie et al., 185 1992; Dell'Anno et al., 2000; Danovaro et al., 2001; Koho et al., 2013). The aminoacid based degradation index (Dauwe and Middelburg, 1998) is one of the most 186 187 commonly used proxies to quantify the extent of degradation or the quality of 188 the remaining particulate organic matter and is based on subtle changes in the 189 amino acid composition due to organic matter processing (Dauwe et al., 1999; 190 Keil et al., 2000).

The compositional changes have also been used to infer transformation of
organic matter by bacteria (Cowie and Hedges, 1994; Dauwe et al., 1999;
Grutters et al., 2001; Vandewiele et al., 2009; Lomstein et al., 2006, 2012), extent
of degradation under oxic and anoxic conditions (Sinninghe Damsté et al., 2002;
Huguet et al, 2008; Nierop et al., 2017) and the relative importance of bacteria
and fauna for organic matter degradation (Sun et al., 1999; Woulds et al., 2012,

197 2014). Although some organic geochemical studies hint at the importance of
198 secondary production (Hayes et al., 1989; Cowie and Hedges, 1994; Grutters et
199 al., 2001; Lomstein et al., 2006, 2012), this aspect has received little attention in
200 organic geochemistry, yet it is one of the main objectives within the ecological
201 approach.

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#### 4 Ecologists focus on the dynamics of organisms using organic matter

205 Benthic communities are usually partitioned into different size classes 206 (e.g. macrofauna, meiofauna and microbes; Gage and Tyler, 1992; Gray and Elliot, 207 2009; Herman et al., 1999), which are often studied by different research 208 communities having distinct objectives, approaches and tools. Organic matter 209 delivered to the seafloor fuels benthic food webs: i.e. it represents food for the 210 animals and the energy substrate for heterotrophic microbes. Microbial ecologists study the growth of microbes on delivered organic matter (e.g. 211 212 bacterial production) and subsequent microbial loss processes, including 213 predation and viral lysis (Kemp, 1988; 1990; Danovaro et al., 2008, 2011, 2016). 214 Microbial ecologists also study in detail the identities and activities or organisms involved in (an)aerobic respiration pathways and the re-oxidation of reduced 215 216 metabolites produced during anaerobic organic matter degradation (Canfield et 217 al., 2005). Animal ecologists focus on the response of fauna to food delivery, the 218 diet and growth of animals and transfer of carbon up the food chain to top consumers (Krumins et al., 2013; Fig. 3). Interactions among food-web members 219 220 are considered the key to understand carbon flows (Pimm et al., 1991; van 221 Oevelen et al., 2010).

222 During the last two decades, <sup>13</sup>C-labeled phytodetritus addition experiments have been performed to identify the organisms involved in the 223 224 immediate processing of organic matter delivered to the seafloor (Middelburg, 225 2014). These studies often covered all size classes (animals and microbes) and 226 could show that respiration was the major fate of added phytodetritus and that 227 all size classes directly profited from recently deposited organic matter (Blair et 228 al., 1996; Moodley et al., 2002, 2005a; Woulds et al., 2007, 2009, 2016; Witte et 229 al., 2003; Nomaki et al., 2005; Sweetman and Witte, 2008). In other words, 230 heterotrophic microbes and small and big animals compete for the same food. 231 Van Nugteren et al. (2009b) have shown that the spatial distribution of resources 232 is a key factor governing the relative use of phytodetritus by bacteria vs. animals. 233 Moreover, the relative share of organisms in the processing of organic matter 234 was in some systems and for some consumers proportional to the biomass of the 235 benthic size class, but not always (Moodley et al., 2005a; Woulds et al, 2009, 236 2016). For instance, foraminifera and amoebid protozoa, sometimes contribute 237 disproportionally to short-term carbon processing reflecting high turn-over of an 238 active community (Moodley et al., 2002; Woulds et al., 2007).

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#### 241 **5 Towards a synthesis**

The above discussion on conceptual views within different research disciplines
highlights a few discrepancies and gaps in our knowledge. Secondary production
by animals and microbes is often not included in the biogeochemical view that

- 246 focuses on preservation versus mineralization. It is also largely absent from the 247 organic geochemical literature. Consumption of organic matter is restricted to 248 microbes in the biogeochemical view, while the non-fed animals move organic 249 matter, microbes and particles around and enhance solute transfer by bioirrigation activities. The consumption of organic matter eventually results in 250 251 compositional changes of the organic matter remaining but there is little 252 information that the identity of the organism matters much. Whole ecosystem 253 labeling experiments revealed direct flow from detritus to most benthic 254 consumers and to the dissolved inorganic carbon pool, but these short-term 255 experimental results cannot directly be compared to the long-term natural 256 processing of deposited organic matter because long-term transfers within the 257 food web and eventual carbon preservation cannot be resolved experimentally.
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#### 5.1 On the consistency of food-web carbon processing and the biogeochemical burial-respiration partitioning 260

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262 Food-web models describe the exchange of matter (e.g. carbon or energy) among 263 different compartments (organisms) within an ecosystem (Pimm et al., 1991; de Ruiter et al., 1995) and usually lump respiration losses (Cole et al., 2006; van 264 Oevelen et al, 2010; Fig. 3 left). Experimental studies using <sup>13</sup>C labelled 265 266 phytodetritus as a tracer of sediment carbon processing showed that animals and microbes both can assimilate labile carbon directly and confirmed that 267 respiration is the largest sink (Moodley et al, 2005a; Buhring et al., 2006; 268 Andersson et al. 2008; Woulds et al. 2009, 2016). The biogeochemical budgeting 269 270 approach basically distinguishes only between (refractory) carbon preserved 271 and buried versus labile organic carbon that is respired to carbon dioxide (Aller, 272 2013; Fig. 3 right). These ecological and biogeochemical concepts can be 273 consistent depending on the timescale considered. On the time scale of days to 274 month deposited carbon is processed by the benthic organisms, a small part is 275 assimilated and the majority is respired. On longer time scales and when 276 considering steady-state conditions, i.e. constant faunal and microbial biomass, 277 there is transfer from the detritus pool to the living biomass pool, but these 278 secondary producers die and their remains are returned to the detritus pool for 279 another cycle with the result that eventually all labile organic matter is respired.

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#### 281 5.2 Secondary production and the formation of molecularly 282 uncharacterizable organic matter.

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284 The mere presence of living organisms in sediments clearly indicates that 285 secondary production is omnipresent. Microbes usually dominate living biomass, 286 but not always, and living biomass typically contributes a few % to the standing 287 stock of total organic carbon in coastal sediments (Herman et al., 1999). The importance of microbial biomass relative to total biomass increases with 288 289 increasing water depth (Rex et al., 2006; Danovaro et al., 2014, 2015), Moreover, 290 living biomass may contribute substantially to total carbon stocks in coarse-291 grained sandy sediments with low background organic carbon contents (Herman 292 et al., 1999; Evrard et al., 2012). Various types of experimental evidence have 293 shown that carbon flow through the living compartment is much higher than 294 through the non-living sediment organic matter pool. Short-term, in situ

experiments using <sup>13</sup>C and/or <sup>15</sup>N labelled organic matter (e.g. phytodetritus) 295 revealed rapid incorporation of  ${}^{13}C/{}^{15}N$  in physically separated organisms 296 (macro- and meiofauna and foraminifera) and microbes, the latter via 297 incorporation of tracers in biomarkers specific for certain microbial groups 298 299 (Middelburg et al., 2000; Boschker and Middelburg, 2002; Veuger et al., 2007; 300 Oakes et al., 2012;, Woulds et al., 2007, 2016) Similarly, ammonium isotope 301 dilution studies have shown that net ammonification (ammonium release) is 302 only a fraction of the total ammonium regeneration because a substantial part of 303 the ammonium liberated is re-assimilated by the microbial community 304 (Blackburn and Henriksen, 1983). Clearly the microbes and animals living in 305 sediment assimilate carbon and synthesize new biomass (Veuger et al., 2012). 306 How can this be reconciled with the biogeochemical and organic geochemical 307 views in which organic matter is either preferentially degraded to carbon dioxide 308 or selectively preserved (Fig. 1, 3). These two apparently inconsistent views are 309 consistent if most of the newly produced organic matter is eventually degraded.

310 Detailed investigations of organic matter composition might in principle 311 resolve this issue as microbial and animal processing of organic matter results in 312 the formation of distinct compounds (Bradshaw et al., 1990; Sun et al., 1999; Thomas and Blair, 2002; Woulds et al., 2012, 2014). There are few issues with 313 314 this approach: (1) most sedimentary organic matter is molecularly 315 uncharacterizable and the origin (imported from the water column vs. newly 316 produced within the sediment) can thus not directly be investigated, (2) microbes living within (the guts) of animals may mask the animal signatures 317 (Woulds et al., 2012, 2014) and (3) different analytical windows (amino acids vs. 318 319 lipids) may result in different inferences. On one hand, the accumulation of 320 bacterial derived non-protein amino acids and peptidoglycan derived D amino 321 acids are clear signs that extensively modified organic matter contains a major 322 fraction that is derived from (heterotophic) bacteria (Cowie and Hedges, 1994; 323 Dauwe et al., 1999; Grutters et al., 2001; Lomstein et al., 2006; Keil and Fogel, 324 2001; Keil et al., 2000). Using fatty acids, Gong and Hollander (1997) also 325 identified a substantial microbial contribution to sedimentary organic matter. On 326 the other hand, using a combined lipid-isotope approach Hartgers et al. (1994) 327 reported only a minor contribution of bacteria to sedimentary organic carbon 328 pools.

329 Secondary production has potentially major consequences for the 330 interpretation of sedimentary records. If microbial reworking of deposited 331 organic matter represents a major carbon flow and part of the material is 332 preserved, then one would expect that bulk organic matter properties such as C. 333 N, P elemental ratios and nitrogen and carbon isotopes would reflect this. 334 Degradation of organic matter initially results in the preferential release of 335 nitrogen and phosphorus relative to carbon. Microbes normally have lower C:N 336 ratios than their substrate (Sterner and Elser, 2002) implying that secondary production and accumulation of microbial derived organic matter should 337 338 eventually result in a net decrease of sediment C:N ratios (Müller, 1977). In 339 contrast, the C:P ratio of heterotrophic microbes is rather variable because P 340 demands depends on the growth rate (Sterner and Elser, 2002) and slowly growing benthic microbes may have high C:P ratios (Steenbergh et al., 2013). 341 Moreover, microbial P storage also depends on redox conditions with the 342 343 consequences that sedimentary C:P ratios are highly variable (Algeo and Ingall,

2007). Sediment  $\delta^{15}$ N values often show a post-depositional shift towards 344 heavier values in alternating oxic/anoxic settings (Moodlev et al., 2005b). Such a 345 346 shift is to be expected because regenerated ammonium is either transformed into nitrite/nitrate (nitrification) or re-assimilated by the microbial community. 347 During oxic conditions nitrification occurs with preference for <sup>14</sup>N and the 348 349 remaining ammonium available for re-assimilation by microbes will be relatively 350 rich in <sup>15</sup>N, while during anoxic conditions oxidation of ammonium is less 351 important or absent, and the ammonium re-assimilated will have similar  $\delta^{15}N$ values as that regenerated. Secondary production within sediment may also 352 353 impact the interpretation of bulk stable carbon isotope records (Haves et al., 1998) and paleorecords of microbial biomarkers (Schouten et al., 2010). 354

355 To reconcile the strong experimental evidence for preferential 356 degradation (Middelburg, 1989), selective preservation (Tegelaar et al., 1989) 357 and formation of new compounds by secondary producers (Lomstein et al, 2012; 358 Braun et al, 2017) I present a new integral concept (Fig. 4). Phytodetritus 359 delivered to sediments is preferentially degraded with the result that new 360 biomass is formed and that some compounds are selectively preserved. The 361 newly formed biomass is after death of the organism added to the pool of degraded detritus and subject to further microbial processing. After multiple 362 cycles of processing by benthic heterotrophs most of the remaining organic 363 364 matter becomes molecularly uncharacterizable. This conceptual model is 365 consistent with the ruling paradigms of preferential degradation and selective preservation as well as with the occurrence of secondary production and 366 formation of molecular uncharacterizable organic matter, but the next step is to 367 368 quantify this conceptual view. One approach would be to use proxies for organic 369 matter degradation state such as fraction of total nitrogen present as amino acid, 370 non-protein amino acids accumulation and the degradation index (Cowie and Hedges, 1994; Dauwe and Middelburg, 1998; Dauwe et al., 1999). Lomstein et al. 371 372 (2012) and Braun et al. (2017) used amino acid racemization to quantify 373 turnover of living microbial biomass as well as of bacterially derived organic 374 matter (necromass) in the deep biosphere. Veuger et al. (2012) executed a 375 <sup>13</sup>C/<sup>15</sup>N tracer experiment and followed the isotope labels into carbohydrates, 376 amino acids and lipids and basically showed that most of the deliberately added 377 heavy isotopes were recovered from the molecularly uncharacterizable pool 378 within a few weeks and remained in that pool till the end of the experiment (> 1 379 year). Their study provided direct evidence for rapid formation of new microbial 380 biomass and subsequent transfer of microbial biomass to the pool of molecular 381 uncharacterizable organic matter. Moreover, the efficient retention of label was 382 indicative of recycling of molecules (or parts thereof) by microbes rather than de 383 novo synthesis, consistent with findings for archaeal lipids in marine sediments (Takano et al., 2010, Lipsewers et al., 2017) and bacteria in soils (Dippold and 384 385 Kuzyakov, 2016).

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#### 5.3 Animals and carbon supply to sediments

Marine sediments are often considered donor-controlled systems, i.e. organic
matter is delivered via settling of organic matter produced in the sunlit upper
part of the ocean (Fig. 3) and the consuming sediment communities have no

393 control on its carbon delivery (Fig. 5). It is only on the time-scale of ocean 394 bottom-water renewal (100-1000 years) that nutrient regenerated by benthic 395 organisms may impact primary producers in the sunlit upper part of the ocean 396 (Soetaert et al., 2000). This is obviously different for sediments in the photic zone that make up about 1/3 of the coastal ocean (Gattuso et al, 1996) because 397 398 animals can directly graze and consume the benthic primary producers at the 399 sediment surface (Middelburg et al., 2000; Evrard et al., 2010, 2012; Oakes et al., 400 2012; Fig. 5). Donor and consumer controlled food webs have intrinsicly 401 different dynamics.

402 Animals living in sediments below the photic zone can in multiple ways 403 impact carbon processing within marine sediments (Fig. 5). Deposit-feeding 404 animals mix particles (and thus particulate organic carbon) as a consequence of 405 their activities. In the case of constant organic carbon delivery (donor-control) 406 bioturbation stimulates organic carbon processing at depth (Herman et al., 407 1999). In coastal systems, organic matter delivery is more complex because of 408 multiple deposition-resuspension events and lateral transport pathways. Rice 409 and Rhoads (1989) showed that in this case (with constant organic carbon 410 concentration in the top layer) more sediment reworking will increase the organic matter flux into the sediment. Moreover, organic carbon gradients with 411 depth are steeper for high-quality than low-quality material and particle mixing 412 413 thus results in transfer of high-quality organic matter to (micro-)organisms 414 living at depth. Animals living in permeable sediment can via surface sediment topography (bioroughness) induce pore-water flows resulting in the trapping of 415 416 phytoplankton (Huettel et al., 2014).

417 Tropical and cold-water corals, coastal and deep-sea sponges, suspension feeding bivalves and other marine forests communities utilize particulate 418 419 organic matter suspended in the water (Herman et al., 1999; Roberts et al., 2006; 420 Rossi et al., 2017). This organic carbon is used for maintenance respiration and 421 growth, but part is excreted as faeces or pseudofaeces and becomes then 422 available for consumers in the sediments. This can result in local hotspots of 423 biodiversity and microbial activity in the sediments (Herman et al., 1999; 424 Gutierrez et al., 2003; Cathalot et al., 2015). Moreover, the physical structures 425 built by these ecosystem engineers impact hydrodynamics with consequences 426 for local and distant carbon deposition rates. Soetaert et al. (2016) reported 427 elevated carbon deposition to ocean margin sediments due to cold-water corals 428 reefs at very large distances.

Some invertebrates have the capability to take up dissolved organic carbon and in particular sponges can assimilate substantial quantities and transform it into biomass (de Goeij et al., 2013; Fiore et al., 2017; Hoer et al., 2017). This sponge tissue and in particular its detritus can be consumed by benthic organisms. This sponge loop (de Goeij et al., 2013; Rix et al., 2016) is another example how animals can manipulate the transfer of organic carbon from the water-column to the sediments (Fig. 5).

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438 **5.4 Animal stimulation of microbes: An inverted microbial loop?** 

440 The microbial loop is a key concept in ocean biogeochemistry (Pomeroy,
441 1974; Azam et al., 1983). Dissolved organic matter released by phytoplankton,

zooplankton or viral lysis of bacteria, archaea and algae is consumed by
heterotrophic microbes. These heterotrophs are in turn consumed by flagellates,
ciliates and other small consumers that are predated upon by zooplankton
(Azam et al., 1983; Jumars et al. 1989). Energy shunted into the large,
heterogeneous dissolved organic matter pool is in this way made available again
for animals (Fig. 6).

After discovery of this loop in the surface ocean water, research has been 448 449 executed to identify and quantify it in sediments (Kemp, 1988, 1990). This 450 required substantial investment in developing new methods: these studies 451 basically revealed that predation on sedimentary bacteria was not that 452 important (Kemp, 1990; Hondeveld et al., 1992; Hamels et al., 2001; Guilini et al., 453 2009). Van Oevelen et al., (2006) made a detailed study on the fate of bacterial 454 production using in situ <sup>13</sup>C labelling of bacteria. They observed that 8% was lost 455 by physical processes, 27% was consumed by animal predation, while bacterial mortality accounted for 65%. Viruses are the most important loss term for 456 sedimentary microbes (Danovaro et al, 2009, 2011, 2016) and the viral lysis 457 458 products (dissolved organic matter) are consumed by microbes. This results in a 459 dissolved organic carbon-microbes cycle (Fig. 6). This benthic microbial cycle represents a dead end in terms of food web topology, because there is little 460 transfer to higher trophic levels and most carbon is eventually respired as 461 462 needed for mass-balance closure on the long term (Fig. 3).

In fact, the benthic microbial cycle represents more an *inverted microbial* 463 *loop*: rather that animals profit from the microbial loop sensu Azam et al. (1983), 464 it appears that benthic microbes profit from animals mixing labile organic matter 465 466 downwards into the sediments (Fig. 6). Labile organic matter delivered to the sediment surface is mixed by animals inhabiting the sediments (Fig. 5). The 467 468 transfer of high quality organic matter to deeper sediment layer may prime 469 sediment microbial communities and in this way stimulate degradation of 470 indigenous organic matter (Canfield, 1994; van Nugteren et al, 2009a, Bianchi, 471 2011; Hannadis & Aller, 2017). Moreover, tube construction and ventilation by 472 animals brings dissolved oxygen to deeper sediment layers and will in this way stimulate organic matter mineralization (Canfield, 1994; Kristensen et al., 1995; 473 474 Dauwe et al., 2001). Bio-irrigation activities may be intermittent (Volkenborn et 475 al., 2016) and cause oscillations in oxygen availability that stimulate organic 476 matter degradation (Aller and Aller, 1998). This inverted microbial loop is a 477 prime example how animals as ecosystem engineers impact sediment carbon 478 processing (Meysman et al., 2006).

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## 481 **5.5 Imagine a world without animals.**

482 An ocean floor inhabited solely by microbes and without animals was likely the 483 reference state during the first four billion of years of Earth's history (Canfield, 484 2014: Lenton and Watson, 2011). Moreover, in modern systems with anoxic 485 486 bottom waters benthic animals are absent (Rhoads and Morse, 1971; Diaz and Rosenberg, 1995, 2008; Levin, 2003; Levin et al, 2009). In these systems, organic 487 matter degradation pathways are different not only because of a lack of oxygen 488 489 and use of alternative electron acceptors, but also because bio-irrigation and 490 sediment reworking are absent (Aller and Aller, 1998; Levin et al., 2009;

491 Middelburg and Levin, 2009). Consequently, microbe-fauna interactions (enhanced carbon delivery, Fig. 5; animal stimulation of microbes, Fig. 6) are 492 493 impeded. 494 While organic matter mineralization in the presence of oxygen may be more 495 efficient than in anoxic environments (Canfield, 1994; Kristensen et al., 1995; 496 Dauwe et al. 2001), the lack of animal stimulation of microbes may also 497 contribute to burying organic carbon more efficiently in sediments underlying 498 anoxic bottom waters (Hartnett et al., 1998; Hartnett and Devol, 2003; Middelburg and Levin, 2009; Jessen et al., 2017). Moreover, the organic matter 499 500 buried in hypoxic and anoxic settings is usually less degraded (Cowie, 2005, Cowie et al., 2009; Vandewiele et al., 2009; Koho et al., 2013; Jessen et al., 2017). 501 502 This presence of animals and all their interactions with organic matter and 503 microbes has consequences for organic carbon processing in marine sediments 504 and thus the global carbon cycle. It is obvious for any terrestrial microbiologist

- that a world with trees and other macrophytes would be different than one
- 506 without. Moreover, biological oceanographers and limnologists agree that
- zooplankton and other metazoan consumers contribute to biogeochemical cycles
  (Vanni, 2002; Vanni and McIntyre, 2016), and I hope that colleagues studying
- marine sediments are aware that BIO in sediment biogeochemistry is more thanjust microbiology.
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530	References
531	
532	Algeo T.J. and Ingall E.: Sedimentary C <sub>org</sub> :P ratios, paleocean ventilation, and Phanerzoic
533	atmospheric pO <sub>2</sub> . Geochim. Cosmochim. Acta 256: 130-155, 2007.
534	Aller, R. C.: Quantifying solute distributions in the bioturbated zone of marine sediments
535	by defining an average micro environment, Geochim. Cosmochim. Acta, 44,
536	1955-1965, 1980.
537	Aller, R. C.: The importance of relict burrow structures and burrow irrigation in
538	controlling sedimentary solute distributions, Geochim. Cosmochim. Acta, 48,
539	1929-1934, 1984.
540 541	Aller, R. C.: Bioturbation and remineralization of sedimentary organic matter - Effects of
541 542	redox oscillation. Chem. Geol. 114, 331-345, 1994.
542 543	Aller, R.C.: Transport and reactions in the bioirrigated zone, in: B.P. Boudreau and B.B. Jørgensen, <i>The benthic boundary layer: transport processes and biogeochemistry.</i>
543 544	pp. 269-301, 2001.
545	Aller, R. C., and Rude, P. D.: Complete oxidation of solid-phase sulphides by manganese
546	and bacteria in anoxic marine sediments, Geochim. Cosmochim. Acta, 52, 751-
547	765, 1988.
548	Aller, R. C., and Aller, J. Y.: Meiofauna and solute exchange in marine muds. Limnol.
549	Oceanog. 37, 1018-1033, 1992.
550	Aller, R. C., and Aller, J. Y.: The effect of biogenic irrigation intensity and solute exchange
551	on diagenetic reaction rates in marine sediments, J. Mar. Res., 56, 905-936, 1998.
552	Aller, R.C. Sedimentary Diagenesis, Depositional Environments, and Benthic Fluxes
553	Treatise on Geochemistry: Second Edition, 8, pp. 293-33, 2013.
554	Andersson, J. H., Woulds, C., Schwartz, M., Cowie, G. L., Levin, L. A., Soetaert, K., and
555 556	Middelburg, J. J.: Short-term fate of phytodetritus in sediments across the
550	arabian sea oxygen minimum zone, Biogeosciences, 5, 43-53, 2008. Appelo C.A.J.: Multicomponent ion exchange and chromatography in natural systems,
558	Rev. Mineralogy 34, 193-228, 1996.
559	Archer, D., Morford, J.L., and Emerson, S.,: A model of suboxic diagenesis suitable for
560	automatic tuning and gridded global domains. Global Biogeochemical Cycles 16.
561	http://dx.doi.org/10.1029/2000BG001288, 2002.
562	Arndt, S., Jorgensen, B.B., LaRowe, D., Middelburg, J.J., Pancost, R. and Regnier, P., :
563	Quantification of organic matter degradation in marine sediments: A synthesis
564	and review, Earth-Science Reviews, vol 123. DOI:
565	http://dx.doi.org/10.1016/j.earscirev.2013.02.008, 2013.
566	Azam, F., Fenchel, T., Field, J. G., Gray, J. S., Meyer-Reil, L. A., and Thingstad, F.: The
567	ecological role of water-column microbes in the sea, Mar. EcolProg. Ser., 10,
568 569	257–263, 1983. Bondon M.L. Balaadimata Dringston University Press, 2012
570	Bender, M.L. Paleoclimate. Princeton University Press, 2013. Berg, P., Rysgaard, S., and Thamdrup, B.: Dynamic modeling of early diagenesis and
570	nutrient cycling. A case study in an Arctic marine sediment. American Journal of
572	Science 303, 905–955, 2003.
573	Berner, R. A.: Early diagenesis: A theoretical approach. Princeton Univ. Press, 1980.
574	Berner, R. A.: Burial of organic carbon and pyritic sulphur in the modern ocean: Its
575	geochemical and environmental significance. American Journal of Science, 282,
576	451-473, 1982.
577	Berner, R. A., and Westrich, J. T.: Bioturbation and the early diagenesis of carbon and
578	sulphur, Am. J. Sci., 285, 193-206, 1985.
579	Bianchi, T.S.: The role of terrestrially derived organic carbon in the coastal ocean: a
580 581	changing paradigm and the priming effect. PNAS.
581	http://dx.doi.org/10.1073/pnas.1017982108, 2011

582	Bianchi, T.S. and Canuel, E.A. Chemical biomarkers in aquatic ecosystems. Princeton
583	University Press, 2011.
584	Blackburn, T.H. and Henriksen, K. Nitrogen cycling in different types of sediments from
585	Danish waters. Limnol. Oceanogr. 28, 477-493, 1983.
586	Blair, N. E., Levin, L. A., DeMaster, D. J., and Plaia, G.: The shortterm fate of fresh algal
587	carbon in continental slope sediments, Limnol. Oceanogr., 41, 1208–1219, 1996.
588	Boetius, A., Ravenschlag, K., Schubert, C. J., Rickert, D., Widdel, F., Gieseke, A., Amann, R.,
589	Jorgensen, B. B., Witte, U., and Pfannkuche, O.: A marine microbial consortium
590	apparently mediating anaerobic oxidation of methane, Nature, 407, 623-626,
591	2000.
592	Boschker, H. T. S. and Middelburg, J. J.: Stable isotopes and biomarkers in microbial
592	5.,, i
	ecology, FEMS Microbiol. Ecol., 40, 85-95, 10.1111/j.1574-6941.2002.tb00940.x,
594	
595	Bouldin, D. R.: Models for describing diffusion of oxygen and other mobile constituents
596	across mud-water interface, J. Ecol., 56, 77-87, 1968.
597	Boudreau, B. P.: A method-of-lines code for carbon and nutrient diagenesis in aquatic
598	sediments, Comput. Geosci., 22, 479–496, doi:10.1016/0098-3004(95)00115-8,
599	1996.
600	Boudreau, B. P.: Diagenetic models and their implementation, Modelling transport and
601	reactions in aquatic sediments, doi:10.1007/978-3-642-60421-8, 1997.
602	Boudreau, B. P. and Jorgensen, B. B.: The Benthic Boundary Layer: Transport Processes
603	and Biogeochemistry, edited by: Boudreau, B. P. and Jorgensen, B. B., Oxford
604	University Press, 1st Edn., 2001.
605	Bradshaw, S.A. O'hara, S.C.M., Corner, E.D.S. and Eglinton, G: Changes in lipids during
606	simulated herbivorous feeding by the marine crustacean <i>Neomysis integer</i> .
607	Journal of the Marine Biological Association of the United Kingdom, 70, 225-243,
608	1990.
609	Braun S., Mhatre S.S., Jaussi, M. Roy H., Kjeldsen K.U., Pearce C., Seidenkrantz M-S, B.B.
610	Jorgensen, B. A. Lomstein. Microbial turnover in the deep seabed studied by
611	amino acid racemization modelling. Sci. Rep. <b>7</b> , Article number: 5680,
612	doi:10.1038/s41598-017-05972-z, 2017.
613	Buhring, S. I., Lampadariou, N., Moodley, L., Tselepides, A. and Witte, U.: Benthic
614	microbial and whole-community responses to different amounts of <sup>13</sup> C-enriched
615	algae: In situ experiments in the deep Cretan Sea (Eastern Mediterranean).
616	Limnology and Oceanography, 51, 157-165, 2006.
617	
618	Burdige D. J. : Preservation of organic matter in marine sediments: controls,
	mechanisms, and an imbalance in sediment organic carbon budgets?. Chem. Rev.
619	107 467–485. doi:10.1002/chin.200720266, 2007.
620	Burdige D. : Geochemistry of Marine Sediments. Princeton University Press, 2006
621	Cai, W. J., and Reimers, C. E.: Benthic oxygen flux, bottom water oxygen concentration
622	and core top organic carbon content in the deep northeast Pacific Ocean, Deep-
623	Sea Res. I, 42, 1681-1699, 1995.
624	Canfield, D. E.: Sulphate reduction and oxic respiration in marine sediments -
625	Implications for organic carbon preservation in euxinic environments, Deep-Sea
626	Res. I, 36, 121-138, 1989.
627	Canfield, D. E.: Factors Influencing organic carbon preservation in marine sediments.
628	Chem. Geol., 114, 315-239, 1994.
629	Canfield, D. E.: Oxygen, a four billion year history, Princeton Univ. Press, 2014.
630	Canfield, D. E., Thamdrup, B., and Kristensen, E.: Aquatic geomicrobiology, in: Advances
631	in Marine Biology, edited by: Southward, A. J., Tyler, P. A., Young, C. M., and
632	Fuiman, L. A., Elsevier Academic Press, Amsterdam, vol. 48, 640 pp., 2005.
633	Cathalot, C. et al. Cold-water coral reefs and adjacent sponge grounds: Hotspots of
634	benthic respiration and organic carbon cycling in the deep sea. Front. Mar. Sci. 2,
635	37. doi: 10.3389/fmars.2015.00037, 2015.
	· · · ·

636 Cole, J. J., Carpenter, S. R., Pace, M. L., Van de Bogert, M. C., Kitchell, J. L. and Hodgson, J. R.: Differential support of lake food webs by three types of terrestrial organic 637 638 carbon, Ecol. Lett., 9, 558-568, 10.1111/j.1461-0248.2006.00898.x, 2006. 639 Cowie, G.: The biogeochemistry of Arabian Sea surficial sediments: A review of recent 640 studies, Prog. Oceanogr., 65, 260-289, 2005. 641 Cowie, G.L., Hedges, J.I.: Biochemical indicators of diagenetic alteration in natural 642 organic-matter mixtures. Nature 369 (6478), 304-307, 1994. 643 Cowie, G.L, Mowbray S., Lewis M., Matheson H., McKenzie R.: Carbon and nitrogen 644 elemental and stable isotopic compositions of surficial sediments from the 645 Pakistan margin of the Arabian Sea Deep-Sea Res. II: 56, 271-282, 2009. 646 Cowie, G.L, Hedges J.I., and Calvert S.E.: Sources and relative reactivities of amino acids, 647 neutral sugars, and lignin in an intermittent anoxic marine environment. 648 Geochim. Cosmochim. Acta 56, 1963-1978, 1992. 649 Danovaro R., Dell'Anno A. and Fabiano M.: Bioavailability of organic matter in the 650 sediments of the Porcupine Abyssal Plain, northeastern Atlantic. Mar. Ecol. Prog. 651 Ser., 220, 2001. 652 Danovaro, R., Dell'Anno, A., Corinaldesi, C., Magagnini, M., Noble, R., Tamburini, C., and 653 Weinbauer, M.: Major viral impact on the functioning of benthic deep-sea 654 ecosystems, Nature, 454, 1084-1088, 2008. 655 Danovaro, R., Corinaldesi, C., Dell'Anno, A., Fuhrman, J.A., Middelburg, J.J., Noble, R.T. & 656 Suttle, C.A.: Marine viruses and global climate change, FEMS Microbiology 657 Reviews, 35 (6), 993-1034, 2011. 658 Danovaro R, Snelgrove PV, and Tyler P, Challenging the paradigms of deep-sea ecology. 659 Trends Ecol Evol 29: 465-475, 2014. 660 Danovaro R, Corinaldesi C, Rastelli E, et al. Towards a better quantitative assessment of 661 the relevance of deep-sea viruses, Bacteria and Archaea in the functioning of the 662 ocean seafloor. Aquat Microb Ecol 75:81-90, 2015. Danovaro, R., Dell'Anno, A., Corinaldesi, C., Rastelli, E., Cavicchioli, R., Krupovic, M., 663 664 Noble, R.T., Nunoura, T., and Prangishvili, D.: Virus mediated archaeal hecatomb 665 in the deep seafloor. Science Adv. 2, e1600492, 2016. 666 Dauwe B. and Middelburg J. J.: Amino acids and hexosamines as indicators of organic 667 matter degradation state in North Sea sediments. Limnol. Oceanogr. 43, 782-668 798.1998. 669 Dauwe, B., Middelburg, J. J., Herman, P. M. J., and Heip, C. H. R.: Linking diagenetic 670 alteration of amino acids and bulk organic matter reactivity, Limnol. Oceanogr., 671 44, 1809-1814, 1999. 672 Dauwe, B., Middelburg, J. J., and Herman, P. M. J.: Effect of oxygen on the degradability of 673 organic matter in subtidal and intertidal sediments of the North Sea area, Mar. 674 Ecol.-Prog. Ser., 215, 13-22, 2001. 675 Dell'Anno A., Fabiano M., Mei M. L. and Danovaro R.: Enzymatically hydrolysed protein 676 and carbohydrate pools in deep-sea sediments: estimates of the potentially 677 bioavailable fraction and methodological considerations. Mar. Ecol. Prog. Ser. 678 196.15-23.2000. 679 de Goeij, J. M., van Oevelen, D., Vermeij, M. J. A., Osinga, R., Middelburg, J. J., de Goeij, A. F. 680 P. M. and Admiraal, W.: Surviving in a Marine Desert: The Sponge Loop Retains 681 Resources Within Coral Reefs, Science, 342, 108-110, 10.1126/science.1241981, 682 2013. 683 De Ruiter P.C., Neutel A-M., and Moore J.C.: Energetics, patterns of interaction strengths, 684 and stability in real ecosystems. Science 269: 1257-1260, 1995 685 Diaz, R. J., and Rosenberg, R.: Marine benthic hypoxia: A review of its ecological effects 686 and the behavioural responses of benthic macrofauna. Ann. Rev. Ocean. Mar. 687 Biol., Vol 33, 245-303, London, 1995. 688 Diaz, R. J., and Rosenberg, R.: Spreading dead zones and consequences for marine 689 ecosystems, Science, 321, 926-929, 2008.

690 Dippold, M.A., and Kuzyakov, Y. Direct incorporation of fatty acids into microbial 691 phospholipids in soils: position-specific labeling tells the story. Geochim. 692 Cosmochim. Acta, 174, 211-221, 2016. 693 Duarte, C. M., Middelburg, J. J. and Caraco, N.: Major role of marine vegetation on the 694 oceanic carbon cycle. *Biogeosciences*, 2, 1-8, 2005. 695 Evrard, V., Soetaert, K., Heip, C. H. R., Huettel, M., Xenopoulos, M. A. and Middelburg, J. J.: 696 Carbon and nitrogen flows through the benthic food web of a photic subtidal 697 sandy sediment, Mar. Ecol. Prog. Ser., 416, 1-16, 10.3354/meps08770, 2010. 698 Evrard, V., Huettel, M., Cook, P. L. M., Soetaert, K., Heip, C. H. R. and Middelburg, J. J.: 699 Importance of phytodetritus and microphytobenthos for heterotrophs in a 700 shallow subtidal sandy sediment, Mar. Ecol. Prog. Ser., 455, 13-31, 701 10.3354/meps09676, 2012. 702 Fiore C.L., Freeman C.J. and Kujawinski E.B.: Sponge exhalent seawater contains a unique 703 chemical profile of dissolved organic matter. PeerJ 5:e2870 704 DOI10.7717/peerj.2870, 2017 705 Gage J.D. and Tyler P.A.: Deep-sea biology: A natural history of organisms at the deep-sea 706 floor. Cambridge Univer. Press, 1991. 707 Gattuso, J.-P., Gentili, B., Duarte, C. M., Kleypas, J. A., Middelburg, J. J. and Antoine, D.: 708 Light availability in the coastal ocean: impact on the distribution of benthic 709 photosynthetic organisms and their contribution to primary production, 710 Biogeosciences, 3, 489-513, 2006. 711 Gray J.S. and Elliot, M.: Ecology of marine sediments. 2<sup>nd</sup> edition, Oxford Univer. Press, 712 2009. 713 Gong C. and Hollander D.: Differential contribution of bacteria to sedimentary organic 714 matter in oxic and anoxic environments, Santa Monica Basin, California. 715 Geochim. Cosmochim. Acta 26, 545-563, 1997. 716 Guilini, K., Van Oevelen, D., Soetaert, K., Middelburg, J. J., and Vanreusel, A.: Nutritional 717 importance of benthic bacteria for deep-sea nematodes from the Arctic ice 718 margin: Results of an isotope tracer experiment, Limnol. Oceanogr., 55, 1977-719 1989, 2010. 720 Glud, R. N.: Oxygen dynamics of marine sediments, Mar. Biol. Res., 4, 243-289, 2008. 721 Grutters M., van Raaphorst W., Epping E., Helder J., de Leeuw J. W., Glavin D. P. and Bada 722 J.: Preservation of amino acids from in situ-produced bacterial cell wall 723 peptidoglycans in northeastern Atlantic continental margin sediments. Limnol. 724 Oceanogr. 47, 1521–1524, 2002. 725 Gutierrez J.L., Jones C.G., Strayer D.L., Iribarne O.O.: Mollusks as ecosystem engineers: the 726 role of shell production in aquatic habitats. Oikos 101:79–90, 2003. 727 Hamels, I., Muylaert, K., Casteleyn, G., and Vyverman, W.: Uncoupling of bacterial 728 production and flagellate grazing in aquatic sediments: a case study from an 729 intertidal flat, Aquat. Microb. Ecol., 25, 31-42, 2001. 730 Hannides, A. K. and Aller, R.C.: Priming effect of benthic gastropod mucus on 731 sedimentary organic matter remineralization., Limnology & 732 Oceanography, 61, 2016, p. 1640. doi:10.1002/lno.10325 733 Hartgers W. A., Damste J. S. S., Requejo A. G., Allan J., Hayes J. M. and De Leeuw J. W.: 734 Evidence for only minor contributions from bacteria sedimentary carbon. Nature 735 369, 224-227, 1994. 736 Hartnett, H. E., and Devol, A. H.: Role of a strong oxygen-deficient zone in the 737 preservation and degradation of organic matter: A carbon budget for the 738 continental margins of northwest Mexico and Washington State, Geochim. 739 Cosmochim. Acta, 67, 247-264, 2003. 740 Hartnett, H. E., Keil, R. G., Hedges, J. I., and Devol, A. H.: Influence of oxygen exposure 741 time on organic carbon preservation in continental margin sediments, Nature, 742 391, 572-574, 1998.

743	Hayes, J.M., Popp, B.N., Takigiku R. and Johnson M.W., An isotopic study of
744	biogeochemical relationships between carbonates and organic carbon in the
745	Greenhorn Formation. Geochim. Cosmochim. Acta 53, 2961-2972, 1998.
746	Hedges, J. I., and Keil, R. G.: Sedimentary Organic-Matter Preservation - an Assessment
747	and Speculative Synthesis, Mar. Chem., 49, 81-115, 1995.
748	Hedges J. I., Eglinton G., Hatcher P. G., Kirchman D. L., Arnosti C., Derenne S., Evershed R.
749	P., Kögel-Knabner I., de Leeuw J. W., Littke R., Michaelis W. and Rullkötter J.: The
750	molecularly-uncharacterized component of nonliving organic matter in natural
751	environments. Org. Geochem. 31, 945–958, 2000.
752	Herman, P. M. J., Middelburg, J. J., Van de Koppel, J., and Heip, C. H. R.: Ecology of
753	estuarine macrobenthos. Advances in Ecological Research, 29, 195-240, 1999.
754	Hoer, D.R., Gibson P.J., Tommerdahl, J.P., Lindquist N.L. and Martens C.S.: Consumption of
755	dissolved organic carbon by Caribbean reef sponges. Lim. Ocean. (2017, first
756	online).
757	Hondeveld, B. J. M., Bak, R. P. M., and Van Duyl, F. C.: Bacterivory by heterotrophic
758	nanoflagellates in marine sediments measured by uptake of fluorescently
759	labelled bacteria, Mar. EcolProg. Ser., 89, 63–71, 1992.
760	
	Huguet, C, de Lange, G.J., Gustafsson, Ö, Middelburg, J.J., Sinninghe Damsté, J.S. and
761	Schouten, S.: Selective preservation of soil organic matter in oxidized marine
762	sediments (Madeira Abyssal Plain) Geochimica et Cosmochimica Acta, 72, no. 24,
763	pp. 6061-6068. DOI: 10.1016/j.gca.2008.09.021, 2008.
764	Huettel, M., Berg, P., and Kostka J.E.: Benthic exchange and biogeochemical cycling in
765	permeable sediments. Ann. Rev. Marine Sciences, 6, 23-51, 2014.
766	Jessen G. L., Lichtschlag A., Ramette A., Pantoja S., Rossel P.E., Schubert C.J., Struck U.,
767	Boetius A.: Hypoxia causes preservation of labile organic matter and changes
768	microbial community composition (Black Sea shelf) Science Advances 3(2),
769	e1601897; doi:10.1126/sciadv.1601897, 2017.
	•
770	Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark),
770 771	Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977
770 771 772	Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977 Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate
770 771 772 773	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models.</li> </ul>
770 771 772 773 774	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> </ul>
770 771 772 773 774 775	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate</li> </ul>
770 771 772 773 774 775 776	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> </ul>
770 771 772 773 774 775 776 777	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel,</li> </ul>
770 771 772 773 774 775 776 777 778	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> </ul>
770 771 772 773 774 775 776 777 778 778 779	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial</li> </ul>
770 771 772 773 774 775 776 777 778 779 780	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A,</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483- 495, 1989.</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A,</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483- 495, 1989.</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483–495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783 784	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483–495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact on sediment bacteria, Mar. EcolProg. Ser., 49, 163–169, 1988.</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783 784 785	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483–495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact on sediment bacteria, Mar. EcolProg. Ser., 49, 163–169, 1988.</li> <li>Kemp, P. F.: The fate of benthic bacterial production, Rev. Aquat. Sci., 2, 109–124, 1990.</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783 784 785 786	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483- 495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact on sediment bacteria, Mar. EcolProg. Ser., 49, 163-169, 1988.</li> <li>Kemp, P. F.: The fate of benthic bacterial production, Rev. Aquat. Sci., 2, 109-124, 1990.</li> <li>Koho, K. A., Nierop, K. G. J., Moodley, L., Middelburg, J. J., Pozzato, L., Soetaert, K., van der</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783 784 785 786 787	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483–495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact on sediment bacteria, Mar. EcolProg. Ser., 49, 163–169, 1988.</li> <li>Kemp, P. F.: The fate of benthic bacterial production, Rev. Aquat. Sci., 2, 109–124, 1990.</li> <li>Koho, K. A., Nierop, K. G. J., Moodley, L., Middelburg, J. J., Pozzato, L., Soetaert, K., van der Plicht, J., and Reichart, G J.: Microbial bioavailability regulates organic matter</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783 784 785 786 785 786 787 788 789 790	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483–495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact on sediment bacteria, Mar. EcolProg. Ser., 49, 163–169, 1988.</li> <li>Kemp, P. F.: The fate of benthic bacterial production, Rev. Aquat. Sci., 2, 109–124, 1990.</li> <li>Koho, K. A., Nierop, K. G. J., Moodley, L., Middelburg, J. J., Pozzato, L., Soetaert, K., van der Plicht, J., and Reichart, G J.: Microbial bioavailability regulates organic matter preservation in marine sediments, Biogeosciences, 10, 1131–1141,</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783 784 785 784 785 786 787 788 787	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483- 495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact on sediment bacteria, Mar. EcolProg. Ser., 49, 163–169, 1988.</li> <li>Kemp, P. F.: The fate of benthic bacterial production, Rev. Aquat. Sci., 2, 109–124, 1990.</li> <li>Koho, K. A., Nierop, K. G. J., Moodley, L., Middelburg, J. J., Pozzato, L., Soetaert, K., van der Plicht, J., and Reichart, G J.: Microbial bioavailability regulates organic matter preservation in marine sediments, Biogeosciences, 10, 1131–1141, doi:10.5194/bg-10-1131-2013, 2013.</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783 784 785 786 785 786 787 788 789 790	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483–495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact on sediment bacteria, Mar. EcolProg. Ser., 49, 163–169, 1988.</li> <li>Kemp, P. F.: The fate of benthic bacterial production, Rev. Aquat. Sci., 2, 109–124, 1990.</li> <li>Koho, K. A., Nierop, K. G. J., Moodley, L., Middelburg, J. J., Pozzato, L., Soetaert, K., van der Plicht, J., and Reichart, G J.: Microbial bioavailability regulates organic matter preservation in marine sediments, Biogeosciences, 10, 1131–1141, doi:10.5194/bg-10-1131-2013, 2013.</li> <li>Keil R. G. and Fogel M. L.: Reworking of amino acid in marine sediments: stable carbon</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783 784 785 786 787 786 787 788 789 790 791	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483–495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact on sediment bacteria, Mar. EcolProg. Ser., 49, 163–169, 1988.</li> <li>Kemp, P. F.: The fate of benthic bacterial production, Rev. Aquat. Sci., 2, 109–124, 1990.</li> <li>Koho, K. A., Nierop, K. G. J., Moodley, L., Middelburg, J. J., Pozzato, L., Soetaert, K., van der Plicht, J., and Reichart, G J.: Microbial bioavailability regulates organic matter preservation in marine sediments, Biogeosciences, 10, 1131–1141, doi:10.5194/bg-10-1131-2013, 2013.</li> <li>Keil R. G. and Fogel M. L.: Reworking of amino acid in marine sediments: stable carbon isotopic composition of amino acids in sediments along the Washington coast.</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783 784 785 786 787 788 787 788 787 788 789 790 791 792	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483- 495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact on sediment bacteria, Mar. EcolProg. Ser., 49, 163-169, 1988.</li> <li>Kemp, P. F.: The fate of benthic bacterial production, Rev. Aquat. Sci., 2, 109-124, 1990.</li> <li>Koho, K. A., Nierop, K. G. J., Moodley, L., Middelburg, J. J., Pozzato, L., Soetaert, K., van der Plicht, J., and Reichart, G J.: Microbial bioavailability regulates organic matter preservation in marine sediments, Biogeosciences, 10, 1131-1141, doi:10.5194/bg-10-1131-2013, 2013.</li> <li>Keil R. G. and Fogel M. L.: Reworking of amino acid in marine sediments: stable carbon isotopic composition of amino acids in sediments along the Washington coast. Limnol. Oceanogr. 46, 14-23, 2001.</li> </ul>
770 771 772 773 774 775 776 777 778 779 780 781 782 783 784 785 786 787 788 785 786 787 788 789 790 791 792 793	<ul> <li>Jørgensen, B. B.: Sulphur cycle of a coastal marine sediment (Limfjorden, Denmark), Limnol. Oceanogr., 22, 814-832, 1977</li> <li>Jørgensen, B. B. A comparison of methods for the quantification of bacterial sulfate reduction in coastal marine sediments. II. Calculation from mathematical models. Geomicrobiol. J. 1,29-47, 1979.</li> <li>Jørgensen, B. B.: Mineralization of organic matter in the sea bed - the role of sulphate reduction, Nature, 296, 643-645, 1982.</li> <li>Jørgensen, B. B.: Bacteria and marine biogeochemistry. In: HD Shulz and M. Zabel, Marine geochemistry, 169-206, 2006</li> <li>Jumars, P. A., Penry, D. L., Baross, J. A., Perry, M. J., and Frost, B.W.: Closing the microbial loop: dissolved organic carbon path- way to heterotrophic bacteria from incomplete ingestion, digestion and absorption in animals, Deep-Sea Res. Pt. A, 36, 483- 495, 1989.</li> <li>Kemp, P. F.: Bacterivory by benthic ciliates: significance as a carbon source and impact on sediment bacteria, Mar. EcolProg. Ser., 49, 163-169, 1988.</li> <li>Kemp, P. F.: The fate of benthic bacterial production, Rev. Aquat. Sci., 2, 109-124, 1990.</li> <li>Koho, K. A., Nierop, K. G. J., Moodley, L., Middelburg, J. J., Pozzato, L., Soetaert, K., van der Plicht, J., and Reichart, G J.: Microbial bioavailability regulates organic matter preservation in marine sediments, Biogeosciences, 10, 1131-1141, doi:10.5194/bg-10-1131-2013, 2013.</li> <li>Keil R. G. and Fogel M. L.: Reworking of amino acid in marine sediments: stable carbon isotopic composition of amino acids in sediments along the Washington coast. Limnol. Oceanogr. 46, 14-23, 2001.</li> <li>Keil R. G., Tsamakis E. and Hedges J. L: Early diagenesis of particulate animo acids in</li> </ul>

797 Kristensen, E., Ahmed, S. I., and Devol, A. H.: Aerobic and anaerobic decomposition of 798 organic matter in marine sediment: Which is fastest?, Limnol. Oceanogr., 40, 799 1430-1437, 1995. 800 Kristensen, E., Penha-Lopes, G., Delefosse, M., Valdemarsne T., Quintana, C.O. and Banta 801 G.T.: What is bioturbation? The need for a precise definition for fauna in aquatic 802 sciences. Mar. Ecol. Prog. Ser. 446, 285–302, doi:10.3354/meps09506, 2012. 803 Krumins, J. A., van Oevelen, D., Bezemer, T. M., De Deyn, G. B., Hol, W. H. G., van Donk, E., 804 de Boer, W., de Ruiter, P. C., Middelburg, J. J., Monroy, F., Soetaert, K., Thebault, E., 805 van de Koppel, J., van Veen, J. A., Viketoft, M. and van der Putten, W. H.: Soil and 806 Freshwater and Marine Sediment Food Webs: Their Structure and Function, 807 Bioscience, 63, 35-42, 10.1525/bio.2013.63.1.8, 2013. 808 Lee C., Wakeham S. G. and Hedges J. I.: Composition and flux of particulate amino acids 809 and chloropigments in equatorial Pacific seawater and sediments. Deep Sea Res. 810 Part I 47, 1535-1568, 2000. 811 Lenton T. and Watson A.: Revolutions that made the earth. Oxford Univer. Press, 2011. 812 Lichtner P. C.: Continuum representation of multi-component-multiphase reactive 813 transport. Rev. Mineralogy 34, 1-82, 1996. Lipsewers, Y.A., Hopmans E.C., Sinnighe Damste J.S. and Villanueva, L.: Potential 814 815 recycling of thaumarchaeotal lipids by DPANN archaea in seasonally hypoxic 816 surface marine sediments. Organic geochemistry (submitted), 2017. 817 Lomstein B. A., Jørgensen B. B., Schubert C. J. and Niggemann J.: Amino acid biogeo- and 818 stereochemistry in coastal Chilean sediments. Geochim. Cosmochim. Acta 70, 819 2970-2989.doi:10.1016/j.gca.2006.03.015, 2006. 820 Lomstein B. Aa., Langerhuus A. T., D'Hondt S., Jørgensen B. B. and Spivack A.: Endospore 821 abundance, microbial growth and necromass turnover in deep subseafloor 822 sediment. Nature 484, 101-104, 2012. 823 Levin, L. A.: Oxygen minimum zone benthos: Adaptation and community response to 824 hypoxia, Ann. Rev. Oceanogr. Mar. Biol., 41, 1–45, 2003. 825 Levin, L.A., Ekau W., Gooday A.J., Jorissen F., Middelburg J.J., Naqvi W., Neira C., Rabalais N.N., Zhang, J.: Effects of Natural and Human-Induced Hypoxia on Coastal 826 827 Benthos. Biogeosc. Disc. 6, 3563-3654, 2009. 828 Meysman, F.J.R.; Middelburg, J.J.; Herman, P.M.J. and Heip, C.H.R.: Reactive transport in 829 surface sediments: 2. Media: an object-oriented problem-solving environment 830 for early diagenesis. Comput. Geosci. 29: 301-318, 2003. 831 Meysman, F. J. R., Middelburg, J. J., and Heip, C. H. R.: Bioturbation: a fresh look at 832 Darwin's last idea, Trends Ecol. Evol., 21, 688-695, 2006. 833 Meysman, F. J. R., Boudreau, B. P., and Middelburg, J. J.: Relations between local, nonlocal, 834 discrete and continuous models of bioturbation, J. Mar. Res., 61, 391-410, 2003. 835 Meysman, F.J.R., Boudreau, B.P. and Middelburg, J.J.: When and why does bioturbation 836 lead to diffusive mixing? Journal of Marine Research 68, 881-920, 2010. 837 Middelburg, J.J., A simple rate model for organic-matter decomposition in 838 marine-sediments. Geochimica et Cosmochimica Acta 53, 1577–1581, 1989. 839 Middelburg, J. J.: Chemoautotrophy in the ocean, Geophys. Res. Lett., 38, L24604, 840 10.1029/2011GL049725, 2011. 841 Middelburg, J. J.: Stable isotopes dissect aquatic food webs from the top to the bottom. 842 Biogeosciences, 11, 2357-2371, 2014 843 Middelburg, J. J. & Levin, L. A.: Coastal hypoxia and sediment biogeochemistry. 844 Biogeosciences, 6, 1273-1293, 2009. 845 Middelburg, J. J., Vlug, T., and Van der Nat, F.: Organic matter mineralization in marine 846 systems. Global Planet. Change, 8, 47-58, 1993. 847 Middelburg, J. J., Soetaert, K., Herman, P. M. J., and Heip, C. H. R.: Denitrification in marine 848 sediments: A model study, Glob. Biogeochem. Cycle, 10, 661-673, 1996. 849 Middelburg JJ, Nieuwenhuize J, van Breugel P: Black carbon in marine sediments. Mar 850 Chem 65:245-252, 1999.

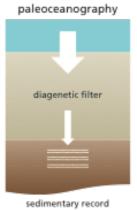
851 852 853	Middelburg, J. J., Barranguet, C., Boschker, H. T. S., Herman, P. M. J., Moens, T., and Heip, C. H. R.: The fate of intertidal microphytobenthos carbon: An in situ C-13-labeling study, Limnol. Oceanogr., 45, 1224–1234, 2000.
854 855	Moodley, L., Middelburg, J. J., Boschker, H. T. S., Duineveld, G. C. A., Pel, R., Herman, P. M. J. and Heip, C. H. R.: Bacteria and Foraminifera: key players in a short-term deep-
856 857	sea benthic response to phytodetritus, Mar. Ecol. Prog. Ser., 236, 23-29, 10.3354/meps236023, 2002.
	, <u>,</u>
858 859	Moodley, L., Middelburg, J. J., Soetaert, K., Boschker, H. T. S., Herman, P. M. J. and Heip, C. H. R.: Similar rapid response to phytodetritus deposition in shallow and deep-
860	sea sediments, J. Mar. Res., 63, 457-469, 10.1357/0022240053693662, 2005a.
861	Moodley, L., Middelburg, J. J., Herman, P. M. J., Soetaert, K., and de Lange, G. J.:
862	Oxygenation and organic-matter preservation in marine sediments: Direct
863	experimental evidence from ancient organic carbon-rich deposits, Geology, 33,
864	889-892, 2005b.
865	Müller, P.J.: C/N ratios in Pacific deep-sea sediments: effect of inorganic ammonium and
866	organic nitrogen compounds sorbed by clays. Geochim. Cosmochim. Actta, 41,
867	549-553, 1997.
868	Nierop, K.G.J., Reichart, GJ., Veld, H. and Sinninghe Damsté, J.S. The influence of oxygen
869	exposure time on the composition of macromolecular organic matter as revealed
870	by surface sediments on the Murray Ridge (Arabian Sea). Geochim. Cosmochim.
871	Acta 206: 40-56. dx.doi.org/10.1016/j.gca.2017.02.032, 2017.
872	Nomaki, H., Heinz, T., Nakatsuka, T., Shimanaga, M., and H. Kitazato, : Species-specific
873	ingestion of organic carbon by deep-sea benthic foraminifera and meiobenthos:
874	In situ tracer experiments. Limnol. Oceanogr. 50: 134–146, 2005.
875	Oakes, J. M., Eyre, B. D. and Middelburg, J. J.: Transformation and fate of
876	microphytobenthos carbon in subtropical shallow subtidal sands: A C-13-
877	labeling study, Limnol. Oceanogr., 57, 1846-1856, 10.4319/lo.2012.57.06.1846,
878	2012.
879 880	Pimm, S.L., Lawton, J.H. and Cohen, J.E.: Food web patterns and their consequences.
881	Nature 350,669-674, 1991. Pomeroy, L.: The ocean's food web, a changing paradigm, Bio-Science, 24, 499–504,
882	1974.
883	Rabouille, C., Gaillard, JF.: Towards the EDGE: early diagenetic global explanation. A
884	model depicting the early diagenesis of organic matter, 02, N03, Mn, and P04.
885	Geochimica et Cosmochimica Acta 55, 2511–2525, 1991.
886	Raghoebarsing A.A., Pol A., van de Pas-Schoonen K.T., Smolders A.J.P., Ettwig K.F.,
887	Rijpstra W.I.C., Schouten S., Sinninghe Damsté J.S., Op den Camp H.J.M., Jetten
888	M.S.M., Strous M.: A microbial consortium couples anaerobic methane oxidation
889	to denitrification. Nature 440: 918-921, 2006.
890	Rex, M. A., Etter, R. J., Morris, J. S., Crouse, J., McClain, C. R., Johnson, N. A., Stuart, C. T.,
891	Deming, J. W., Thies, R., and Avery, R.: Global bathymetric patterns of standing
892	stock and body size in the deep-sea benthos, Mar. EcolProg. Ser., 317, 1–8,
893	2006.
894	Rhoads, D.C., and Morse, J.W.: Evolutionary and ecological significance of oxygen-
895	deficient marine basins, Lethaia, 4, 413-428, 1971.
896	Rhoads, D.C.: Organism-sediment relations on the muddy sea floor. Oceanogr. Mar. Biol.
897	Ann. Rev. 12, 263-300, 1974.
898	Rice, D.L.:. Early diagenesis in bioadvective sediments: relationships between the
899	diagenesis of beryllium-7, sediment reworking rates, and the abundance of
900	conveyor-belt deposit-feeders. Journal of Marine Research 44, 149–184, 1986.
901	Rice, D.L. and Rhoads, D.C.: Early diagenesis of organic matter and the nutritional value
902	of sediment. In: Ecology of Marine Deposit Feeders (Ed. by G. Lopez, G. Taghon
903	and J. Levinton), pp. 309-317. Springer, Berlin, 1989.

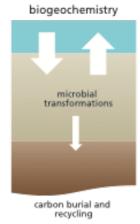
904	Rix, L. et al. Coral mucus fuels the sponge loop in warm- and cold-water coral reef
905	ecosystems. Sci. Rep. <b>6</b> , 18715; doi: 10.1038/srep18715, 2016.
906	Roberts, J. M., Wheeler, A. J. and Freiwald A. Reefs of the deep, The biology and geology
907	of cold-water coral ecosystems. Science 312, 543–547, 2006.
908	Rossi S., Bramanti, L., Gori, A and O. Covadonga. An overview of the animal forest of the
909	world. In Rossi (ed). Marine Animal Forest. Springer, pp. 1-25, 2017.
910	Schouten, S., Middelburg, J.J., Hopmans, E.C. & Sinninghe Damsté, J.S.: Fossilization and
911	degradation of intact polar lipids in deep subsurface sediments: A theoretical
912	approach. Geochmica et Cosmochimica Acta, 74, 3806-3814, 2010.
913	Sinninghe Damsté, J. S., Rijpstra, W. I. C., and Reichart, G. J.: The influence of oxic
914	degradation on the sedimentary biomarker record II. Evidence from Arabian Sea
915	sediments, Geochim. Cosmochim. Acta, 66, 2737-2754, 2002.
916	Soetaert, K., Herman, P. M. J., and Middelburg, J. J.: A model of early diagenetic processes
917 917	from the shelf to abyssal depths, Geochim. Cosmochim. Acta, 60, 1019-1040,
917 918	1996.
919	Soetaert, K., Middelburg, J. J., Herman, P. M. J., and Buis, K.: On the coupling of benthic
920	and pelagic biogeochemical models, Earth-Sci. Rev., 51, 173-201, 2000.
921	Soetaert, K., Mohn, C., Rengstorf, A., Grehan, A., and van Oevelen, D.: Ecosystem
922	engineering creates a direct nutritional link between 600-m deep cold-water
923	coral mounds and surface productivity, Sci. Rep., 6, 35057,
924	doi:10.1038/srep35057, 2016.
925	Steenbergh A.K., Bodelier P.L.E. Heidal, M., Slomp, C.P., and Laanbroek H.J.: Does
926	microbial stoichiometry modulate eutrophication of aquatic ecosystems?
927	Environ. Microbiol. 15: 1572-1579, 2013.
928	Sterner R.W. and Elser J.J.: Ecological Stoichiometry. Princeton University Press, 439 pp,
929	2002.
930	Strous, M., and Jetten, M. S. M.: Anaerobic oxidation of methane and ammonium, Annu.
931	Rev. Microbiol., 58, 99-117, 2004.
932	Sun M. Y., Aller R. C., Lee C. and Wakeham S. G.: Enhanced degradation of algal lipids by
933	benthic macrofaunal activity: effect of Yoldia limatula. J. Mar. Res. 57, 775–804,
934	1999.
935	Sweetman, A. K., and Witte, U: Response of an abyssal macrofaunal community to a
936	phytodetrital pulse. Mar. Ecol. Prog. Ser. 355: 73–84, doi:10.3354/meps07240,
937	2008.
938	Takano, Y., Chikaraishi, Y., Ogawa, N.O., Nomaki, H., Morono, Y., Inagaki, F., Kitazato, H.,
939	Hinrichs, K-U. and Ohkouchi, N.: Sedimentary membrane lipids recycled by deep-
940	sea benthic archaea Nature Geoscience 3, 858–861, doi:10.1038/ngeo983, 2010.
941	Tegelaar, E. W., de Leeuw, J. W., Derenne, S., and Largeau, C.: A reappraisal of kerogen
942	formation. Geochim. Cosmochim. Acta 53, 3103–3106. doi:10.1016/0016-
943	7037(89)90191-9, 1989.
944	Thomas C. J. and Blair N. E.: Transport and digestive alteration of uniformly 13C-labelled
945	diatoms in mudflat sediments. J. Mar. Res. 60, 517–535, 2002.
946	Van Cappellen, P. and Wang, Y. F.: Cycling of iron and manganese in surface sediments:
947	A general theory for the coupled transport and reaction of carbon, oxygen,
948	nitrogen, sulfur, iron, and manganese, Am. J. Sci., 296, 197–243,
949	
	doi:10.2475/ajs.296.3.197, 1996.
950	Vandewiele S., Cowie G., Soetaert K. and Middelburg J. J.: Amino acid biogeochemistry
951	and organic matter degradation state across the Pakistan margin oxygen
952	minimum zone. Deep Sea Res. Part II 56, 318–334, 2009.
953	Vanni, M. J. and McIntyre, P. B.: Predicting nutrient excretion of aquatic animals with
954	metabolic ecology and ecological stoichiometry: a global synthesis. Ecology, 97:
955	3460-3471. doi:10.1002/ecy.1582, 2016.
956	Vanni, M. J.: Nutrient cycling by animals in freshwater ecosystems. Annual Review of
957	Ecology and Systematics 33:341–370, 2002.

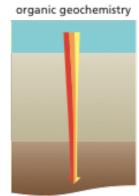
958 Van Nugteren, P., Moodley, L., Brummer, G.-J., Heip, C.H.R., Herman, P.M.J., Middelburg, 959 J.J.: Seafloor ecosystem functioning: the importance of organic matter priming. 960 Marine Biology 156, 2277–2287, 2009a. 961 van Nugteren, P., Herman, P.M., Moodley, L., Middelburg, J.I., Vos, M.: Spatial distribution 962 of detrital resources determines the outcome of competition between bacteria 963 and a facultative detritivorous worm. Limnology and Oceanography 54, 1413-964 1419, 2009b. 965 van Oevelen, D., Moodley, L., Soetaert, K. and Middelburg, J. J.: The trophic significance of 966 bacterial carbon in a marine intertidal sediment: Results of an in situ stable 967 isotope labeling study, Limnol. Oceanogr., 51, 2349-2359, 2006. 968 van Oevelen, D., Van den Meersche, K., Meysman, F. J. R., Soetaert, K., Middelburg, J. J. and 969 Vezina, A. F.: Quantifying Food Web Flows Using Linear Inverse Models, 970 Ecosystems, 13, 32-45, 10.1007/s10021-009-9297-6, 2010. 971 Veuger, B., Eyre, B. D., Maher, D. and Middelburg, J. J.: Nitrogen incorporation and 972 retention by bacteria, algae, and fauna in a subtropical intertidal sediment: An in 973 situ N-15-labeling study, Limnol. Oceanogr., 52, 1930-1942, 974 10.4319/lo.2007.52.5.1930, 2007. 975 Veuger B., van Oevelen D. and Middelburg J. J.: Fate of microbial nitrogen, carbon, 976 hydrolysable amino acids, monosaccharides, and fatty acids in sediment. 977 Geochim. Cosmochim. Acta 83, 217-233, 2012. 978 Volkenborn, N., Polerecky, L., Wethey, D.S., and Woodin, S.A.: Oscillatory porewater 979 bioadvection in marine sediments induced by hydraulic activities of Arenicola 980 marina, Limnology and oceanography 55 (3), 1231-1247, 2010 981 Volkenborn, N., Woodin, S.A., Wethey, D.S. and Polerecky, L.: Bioirrigation in marine 982 sediments, Reference Module in Earth Systems and Environmental Sciences, 983 Elsevier, DOI: 10.1016/B978-0-12-409548-9.09525-7, 2016 984 Wakeham S. G., Lee C., Hedges J. I., Hernes P. J. and Peterson M. L.: Molecular indicators 985 of diagenetic status in marine organic matter. Geochim. Cosmochim. Acta 61, 986 5363-5369, 1997. 987 Westrich, J. T. and Berner, R. A.: The role of sedimentary organic matter in bacterial 988 sulphate reduction – the G model tested, Limnol. Oceanogr., 29, 236–249, 1984. 989 Witte, U., Wenzhofer, F., Sommer, S., Boetius, A., Heinz, P., Aberle, N., Sand, M., Cremer, A., 990 Abraham, W. R., Jorgensen, B. B. and Pfannkuche, O.: In situ experimental 991 evidence of the fate of a phytodetritus pulse at the abyssal sea floor, Nature, 424, 992 763-766, 10.1038/nature01799, 2003. 993 Woulds, C., Andersson, J. H., Cowie, G. L., Middelburg, J. J. and Levin, L. A.: The short-term 994 fate of organic carbon in marine sediments: Comparing the Pakistan margin to 995 other regions, Deep-Sea Research Part Ii-Topical Studies in Oceanography, 56, 996 393-402, 10.1016/j.dsr2.2008.10.008, 2009. 997 Woulds, C., Cowie, G. L., Levin, L. A., Andersson, J. H., Middelburg, J. J., Vandewiele, S., 998 Lamont, P. A., Larkin, K. E., Gooday, A. J., Schumacher, S., Whitcraft, C., Jeffreys, R. 999 M. and Schwartz, M.: Oxygen as a control on seafloor biological communities and 1000 their roles in sedimentary carbon cycling, Limnol. Oceanogr., 52, 1698-1709, 1001 10.4319/lo.2007.52.4.1698, 2007. Woulds, C., Middelburg, J. J. and Cowie, G. L.: Alteration of organic matter during infaunal 1002 1003 polychaete gut passage and links to sediment organic geochemistry. Part I: 1004 Amino acids. Geochimica Et Cosmochimica Acta, 77, 396-414, 2012. 1005 Woulds, C., Middelburg, J. J. & Cowie, G. L.: Alteration of organic matter during infaunal 1006 polychaete gut passage and links to sediment organic geochemistry. Part II: Fatty 1007 acids and aldoses. Geochimica Et Cosmochimica Acta, 136, 38-59, 2014. Woulds, C., Bouillon, S., Cowie, G. L., Drake, E., Middelburg, J.J. and Witte, U.: Patterns of 1008 1009 carbon processing at the seafloor - the role of faunal and microbial communities 1010 in moderating carbon flows. Biogeosciences, 13, 4343-4357, 2016. 1011

1012 1013 1014 1015 1016 1017 1018 1019 1020 1021	<ul> <li>Figure 1. Different views, approaches and interests on carbon processing in marine sediments. Paleoceanographers focus on the sedimentary record, biogeochemists quantify carbon burial and recycling, organic geochemists study alteration of organic matter and ecologists focus on carbon as food for organisms living in the sediment. The red-orange-yellow fractions of organic matter have a different lability.</li> </ul>
1021 1022 1023 1024 1025	Figure 2. Conceptual model of organic matter (OM) degradation and re-oxidation pathways (based on Jørgensen, 2006 and Middelburg and Levin, 2009). The red arrows reflect the fate of (energy-rich) substrates released during anaerobic mineralization.
1026 1027 1028 1029 1030 1031 1032 1033 1034	Figure 3. Carbon processing in marine sediments on the short-term (left) and the long- term (right). Organic matter produced in the sunlit layer of the ocean and delivered to the sediments is either consumed by organisms or buried. The organic matter consumed by organisms is used to synthesize biomass or metabolized to carbon dioxide and nutrients. On the long-term or at steady- state, i.e. the biomass of benthic organisms does not change, the benthic community can be considered a black box diverting organic matter either into metabolites or into the geosphere (burial).
1035 1036 1037 1038 1039 1040 1041 1042	Figure 4. Conceptual diagram showing the relationships between molecular uncharacterizable organics, deposited phytodetritus and secondary production. Phytodetritus is degraded preferentially and new biomass is formed, which after death of the organisms is added to the pool of detritus and subject to degradation. Multiple cycles of organic matter processing eventually results in the formation of molecular uncharacterizable organic matter. The red-orange- yellow fractions of organic matter have a different lability.
1043 1044 1045 1046 1047 1048 1049 1050 1051 1052 1053 1054	Figure 5. Organic matter supply to sediments. 1. The traditional view of organic matter settling passively from the water column (donor control). 2. Sediments in the photic zone are inhabited by benthic microalgae that produce new organic matter in situ and grazing animals can impact the growth of these primary producers. 3. Bioturbating animals transfer labile carbon from the sediment surface layer to deeper layers in the sediments. (Vertical axis is depth, horizontal is concentration) 4. Suspension feeding organisms enhance the transfer of suspended particulate matter from the water column to the sediments (biodeposition). 5. Sponge consume dissolved organic carbon and produce cellular debris that can be consumed by benthic organisms (i.e. the sponge loop).
1055 1056 1057 1058 1059 1060 1061 1062 1063 1064 1065	Figure 6. The microbial (upper panel) and inverted microbial (lower panel) loops. In the water column dissolved organic carbon derived from phytoplankton, zooplankton or microbes (via viral loop) is consumed by heterotrophic microbes, which in turn are consumed by protists and small animals with the consequence that carbon flowing through dissolved organic carbon pools eventually can be used by larger animals (microbial loop). In sediments, the dissolved organic carbon (from viral lysis and other sources) is also consumed by heterotrophic microbes but this carbon is inefficiently transferred to animals. The engineering activities of animals are key in delivering labile organic matter (phytodetritus) to microbes living in the subsurface (inverted microbial loop).

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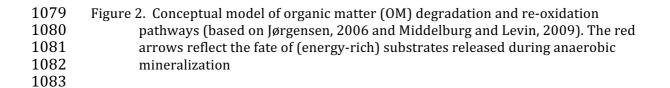


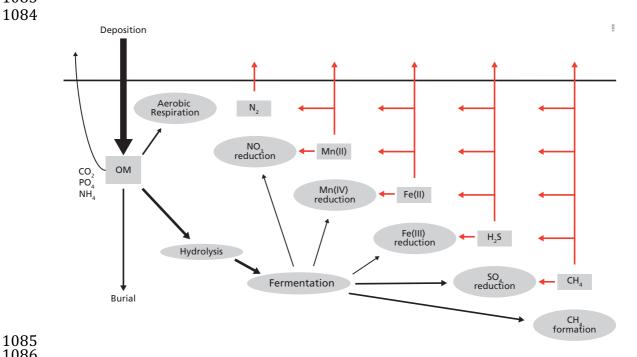


(selective) preservation of compounds



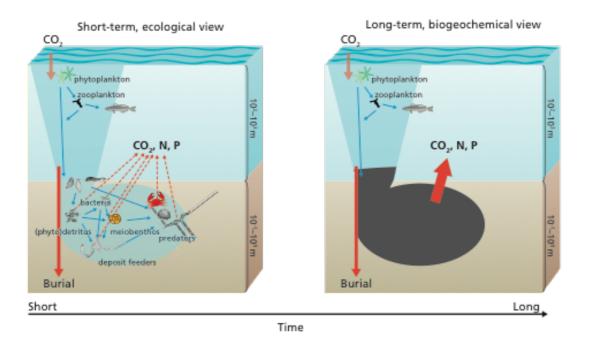
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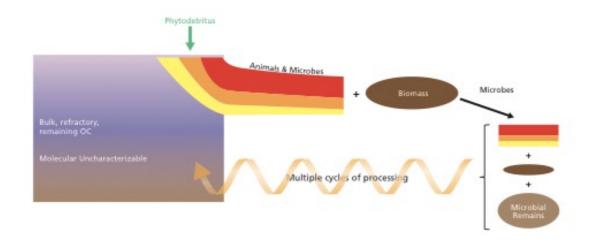


1087 1088 Figure 3. Carbon processing in marine sediments on the short-term (left) and the long-1089 term (right). Organic matter produced in the sunlit layer of the ocean and 1090 delivered to the sediments is either consumed by organisms or buried. The 1091 organic matter consumed by organisms is used to synthesize biomass or 1092 metabolized to carbon dioxide and nutrients. On the long-term or at steady-1093 state, i.e. the biomass of benthic organisms does not change, the benthic 1094 community can be considered a black box diverting organic matter either into 1095 metabolites or into the geosphere (burial). 1096



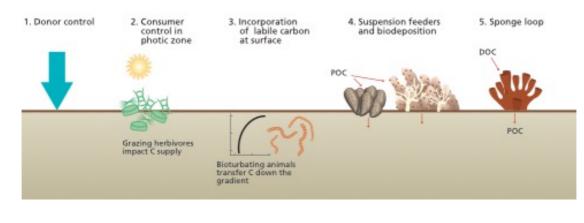


- 1100
- 1101 Figure 4. Conceptual diagram showing the relationships between molecular
- 1102 uncharacterizable organics, deposited phytodetritus and secondary production.
- 1103 Phytodetritus is degraded preferentially and new biomass is formed, which after
- 1104 death of the organisms is added to the pool of detritus and subject to
- 1105 degradation. Multiple cycles of organic matter processing eventually results in
- 1106the formation of molecular uncharacterizable organic matter. The red-orange-1107yellow fractions of organic matter have a different lability.
- 1108

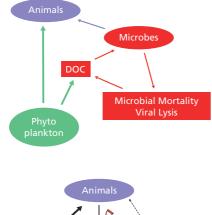


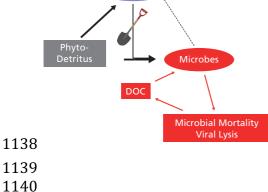
 $\begin{array}{c} 1109\\1110 \end{array}$ 

1112	Figure 5. Organic matter supply to sediments. 1. The traditional view of organic matter
1113	settling passively from the water column (donor control). 2. Sediments in the
1114	photic zone are inhabited by benthic microalgae that produce new organic
1115	matter in situ and grazing animals can impact the growth of these primary
1116	producers. 3. Bioturbating animals transfer labile carbon from the sediment
1117	surface layer to deeper layers in the sediments. (Vertical axis is depth, horizontal
1118	is concentration) 4. Suspension feeding organisms enhance the transfer of
1119	suspended particulate matter from the water column to the sediments
1120	(biodeposition). 5. Sponge consume dissolved organic carbon and produce
1121	cellular debris that can be consumed by benthic organisms (i.e. the sponge loop).
1122	
1123	



- 1126
- 1127 Figure 6. The microbial (upper panel) and inverted microbial (lower panel) loops. In the 1128 water column dissolved organic carbon derived from phytoplankton, 1129 zooplankton or microbes (via viral loop) is consumed by heterotrophic 1130 microbes, which in turn are consumed by protists and small animals with the 1131 consequence that carbon flowing through dissolved organic carbon pools 1132 eventually can be used by larger animals (microbial loop). In sediments, the 1133 dissolved organic carbon (from viral lysis and other sources) is also consumed by heterotrophic microbes but this carbon is inefficiently transferred to animals. 1134 1135 The engineering activities of animals are key in delivering labile organic matter 1136 (phytodetritus) to microbes living in the subsurface (inverted microbial loop). 1137





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