



# Algal diversity of temperate biological soil crusts depends on land use intensity and affects phosphorus biogeochemical cycling

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## Abstract

10 Biological soil crusts (BSCs) form the most productive microbial biomass in many drylands and disturbed areas with a diverse microalgal community as key component. In temperate regions, BSCs are also common, but generally less studied, and they conduct important ecological functions, like stabilization of soil and enrichment of nutrients. Changes in land use and its intensity strongly influence biodiversity per se and it's role for ecosystem processes, particularly in regions which are densely populated like Europe. But systematic studies on land use (i.e. management intensity) gradients in temperate forests on BSCs  
15 are missing up to now. To close this gap of knowledge and enhance the understanding of management effects on BSCs, Cyanobacteria and eukaryotic microalgae as key primary producers of these communities were identified from pine and beech forests under different management regimes. Algae were identified morphologically based on enrichment cultivation and categorized in either coccal taxa, which occur typically in high diversity, or filamentous taxa, which have the potential to initiate BSC formation. In total, 52 algal species were recorded, most from the phylum Chlorophyta, followed by Streptophyta  
20 and Heterokontophyta; Cyanobacteria were much less abundant in temperate forest BSCs. The most abundant crust-initiating filamentous algae were three species of *Klebsormidium* (Streptophyta) a ubiquitous genus often associated with BSCs worldwide and a high tolerance to low pH. Increasing management intensity resulted in a higher richness of algae, especially the proportion of coccal algae rose. Furthermore, the proportion of inorganic phosphorus was positively correlated with the algal richness, indicating that higher diversity of algae results in a more closed P cycle. Thus, management of forests has an  
25 impact on the diversity of phototrophic organisms in BSCs, which in turn affects P cycling in the BSC.

key words: biological soil crusts, forest, management intensity, phosphorus, algae richness, *Klebsormidium*



## Introduction

Biological soil crusts (BSCs) occur on all continents on Earth, predominantly in arid and semi-arid habitats, but also in temperate regions (e.g., Belnap et al., 2001; Weber et al., 2016). In semiarid and arid environments, BSCs were studied, for example, in deserts of Israel and USA or in polar regions (Borchhardt et al., 2017; Flechtner et al., 1998; Kidron et al., 2010).

5 In temperate regions these habitats include dunes with sparse higher vegetation or disturbed areas in open sites (like former mining sites) (Fischer et al., 2010b; Langhans et al., 2009; Lukešová, 2001; Schulz et al., 2016). Although BSCs received raising interest in the past years, reports on BSCs from temperate forests are still missing up to now.

BSCs can be characterized as “ecosystem-engineers” forming water-stable aggregates that have important ecological roles in primary production, nitrogen cycling, mineralization, water retention, and stabilization of soils (Castillo-Monroy et al., 2010; 10 Evans and Johansen, 1999; Lewis, 2007). While the role of BSC in the C- and N-cycle is well documented, less is known about their role in P cycling. However, recent studies indicated that richness of BSC organisms is related to soil P content (Baumann et al., 2017; Schulz et al., 2016). Also, soil texture and carbon content seem to affect the BSC community (Elliott et al., 2014; Steven et al., 2013). But still, only little is known about environmental factors that shape BSC communities and how BSCs in turn affect soil characteristics.

15 Disturbance of BSCs due to land use has been reported to have strong negative effect on BSCs cover, which resulted in higher soil erosion and C and N loss in the top soil (Barger et al., 2006; Belnap, 2003). Studies on the effect of land use on BSCs were up to now exclusively conducted in arid regions. These studies reported, for example, a strong negative influence of intensive livestock grazing due to trampling on BSC cover with a recovery period of up to 27 years (Concostrina-Zubiri et al., 2014; Gomez et al., 2004; Williams et al., 2008). Also ploughing in Australian sand plains reduced the BSCs cover dramatically 20 (Daryanto et al., 2013). In contrast, there are no reports on land use effects in temperate regions or aspects of land use other than grazing on BSCs, like, for example, fertilization of grassland or arable land and silvicultural management.

Cyanobacteria and algae represent the most important phototrophic components of BSCs along with macroscopic lichens and bryophytes (Elbert et al., 2012). Eukaryotic algae are probably the least studied phototrophic component of BSCs, although these organisms are an essential component of BSCs because of their major contribution to C fixation (Weber et al., 2016).

25 BSC algae can be categorized in two functional groups. First, filamentous algae as major BSC forming taxa that stabilize soil particles by gluing them together due to the presence/excretion of mucilage. The filamentous forms occur usually in low diversity but produce high biomass. And second, coccoid algae which are attached to the soil particles or other algae and typically occur in higher diversity but lower biomass (Büdel et al., 2016).

Filamentous cyanobacteria, especially of the genus *Microcoleus*, are often the dominant phototrophic organisms in most BSCs 30 of drylands and in dunes from temperate regions (Garcia-Pichel et al., 2001; Schulz et al., 2016). They are described as important for BSC formation due to their ability to produce mucilage sheds and extracellular polymeric substances forming a network between soil particles (Gundlapally and Garcia-Pichel, 2006). In temperate regions, this key function is often taken



over by filamentous eukaryotic algae, like *Klebsormidium*, *Xanthonema* or *Zygomonium* (Fischer and Subbotina, 2014; Lukešová, 2001; Pluis, 1994).

The aim of the present study was to characterize for the first time the algal community in BSCs collected in temperate forests of different silvicultural management intensities. The data allowed conclusions on land-use effects on BSC algal community  
5 composition, which might result from differences in light regime affecting the phototrophs at these sites. In a previous study we provided for the first time a very detailed picture on the distribution of P content, P pools and P species in temperate BSCs and adhering soil (Baumann et al., 2017). Differences of algal richness in BSCs contributing to P cycling were detected, and the data indicated that BSCs are particularly involved in the transformation of inorganic P to organic P compounds and thus play a key role in the biologically driven P cycling in temperate soils. In addition, BSCs responded differently to management  
10 intensity depending on forest type (beech versus pine) (Baumann et al., 2017). While algal species richness of BSCs was considered as sum parameter, detailed information on the biodiversity is still missing.

Therefore, in the present study we investigated for the first time in detail the influence of forest management intensity on algal  
15 biodiversity in BSCs collected at the same plots as in Baumann et al. (2017), plus additional sampling sites. In addition, the effect of BSC algal biodiversity on C, N and P content, in particular on the different fractions of P was assessed. As a main result we can document that management of forests in temperate regions has an impact on the biodiversity of microalgae and cyanobacteria in BSCs, which in turn affects P cycling in this microecosystem.

## Material and Methods

### Study site

20 BSC samples were collected in June 2014 and 2015 as part of the DFG priority program 1374 Biodiversity Exploratories (Fischer et al., 2010a). Forest plots were sampled in the Schorfheide-Chorin Biosphere Reserve in Northeast Germany. It covers >129,000 hectares and the characteristic postglacial geomorphological structures result in diverse habitats with various  
25 forest types at the plots under study. The plots differed in the dominant tree species: Scots pine (*Pinus sylvestris* L.) or European beech (*Fagus sylvatica* L.). Further, samples were taken from natural, protected forests and from managed forest (age-class forest) (Schall and Ammer, 2013).

The top millimeters of soil, on which BSC had been visually detected as green cover, were collected by pressing a petri dish in the crust and removing gently with a spatula. After transportation to the lab the upper two millimeters of the crust was separated from the adhering soil underneath using a razor blade and stored dry in paper bags prior cultivation. In total, 31 BSCs were collected from 13 pine and 18 beech stands (Table 1).



### Culturing, identification and richness of algae

Solid 3N-Bolds Basal Medium (1.5% agar) with vitamins (Starr and Zeikus, 1993) was used for enrichment cultures in Petri dishes (9.5 cm diameter). Several 7–10 mm BSC pieces were cleaned with forceps to remove all roots and leaves to avoid the additional growth of fungi and bacteria and were placed on the surface of an agar plate under sterile conditions. Plates were 5 incubated at 20°C, 30–35  $\mu\text{mol}$  photons  $\text{m}^{-2} \text{s}^{-1}$  (Osram Lumilux Cool White lamps L36W/840) under a light/dark cycle of 16:8 h L:D. The plates were regularly inspected and colonies were identified four to six weeks after incubation using a light microscope (BX51, Olympus) with Nomarski differential interference optics and 1000x magnification. Light micrographs were taken with an Olympus UC30 camera attached to the microscope and processed with the software cellSens Entry (Olympus). For direct observation of BSC samples, pieces of crust were rewetted with tap water, put on slide and analyzed 10 with the above mentioned microscope with a maximum 400x magnification.

Morphological identification of algae was based on Syllabus (Ettl and Gärtner, 1995) and, more recent publications on certain algae groups (Darienko et al., 2010; Kostikov et al., 2002; Mikhailyuk et al., 2015). Mucilage of algae was stained with an aqueous solution of methylene blue. Algae were identified that belong to Cyanobacteria, Chlorophyta, Streptophyta and some Heterokontophyta (Eustigmatophyceae). Diatoms were regularly observed in direct observation, but excluded from the 15 analyses as the mentioned enrichment cultivation is not suitable for estimation of diatom diversity.

The richness, i.e. the number of algae species in each sample, was used as measurement for diversity rather than diversity indices like evenness, because enrichment cultivation does not allow a clear conclusion about the abundance of each species. Additionally, the community composition of the algae as reflected by the presence or absence of individual species was used as a second parameter for diversity estimation.

### 20 Environmental variables

The forest plots were characterized by differences in the silvicultural management intensity. To evaluate the effect of management, the silvicultural management index (SMI) was used. This index takes into account the tree species, stand age and aboveground living and dead wood biomass (Schall and Ammer, 2013). Thus, the natural forest has a lower SMI than the used forest and a pine stand has a higher SMI than a beech stand.

25 To assess interactions between BSC biodiversity and environmental parameters, the richness, community composition of algae and proportion of filamentous algae was linked to the following environmental parameters: main tree species (pine or beech), silvicultural management intensity (SMI), water content and pH of the bulk soil for all 31 plots (water content and pH provided by I. Schöning, Table 1) and, further, for a subset of the samples (n=19), total C, N and P content, organic and inorganic P proportions, both for labile, moderately labile and stable P. Latter are not shown here but were presented in detail by Baumann 30 et al. (2017).



## Statistical analyses

All statistical analyses were done using the statistical software R version 3.3.0 (R Development Core Team, 2009).

Analysis of Variance (ANOVA) was conducted to reveal the effect of environmental parameters on alga richness and proportion of filamentous algae; their best predictors were selected by backward elimination stepwise regression analysis based  
5 on the BIC (Bayesian information criterion) using ‘step’ command in R. The correlation between environmental parameters were checked by Pearson correlation (cor and cor.test commands in R).

To reveal correlations of single environmental parameters with the community composition of algae, we applied PerManova (with adonis function in R (Anderson, 2001)) using the Bray–Curtis dissimilarity index (Bray and Curtis, 1957), including permutation test with 1000 permutations. The function “adonis” allows applying non-Euclidean distance metrics and handles  
10 both categorical and continuous predictors.

For analysis of co-correlation of environmental factors pearson correlation was used. To test significant differences of environmental factors between tree species, unpaired, two-tailed t-test was performed.

Differences with a p-value below or equal to 0.05 were taken as significant.

## 15 Results

### Algae identification

In total 52 different algae species were detected in enrichment cultures of all 31 BSC samples. *Stichococcus bacillaris* was the most ubiquitous taxon, observed in 27 out of 31 samples; followed by *Coccomyxa simplex* and *Klebsormidium cf. subtile* in 26 out of 23 samples, respectively. All other algal species were detected in less than 50% of the plots; 22 algal species were  
20 observed exclusively at one plot (Figure 1). The total species richness at each plot ranged from three to 14 species, with the mean of eight and a standard deviation of 2.6 (complete species list is provided in the supplemental Table S1).

The phylum Chlorophyta made up 81% of all detected algae species, followed by Streptophyta (11%) and Heterokontophyta (6%) (Figure 2). Cyanobacteria were rare in these BSCs, just one species, *Microcoleus vaginatus*, was observed in only one sample.

25 The identified algae species were differentiated according to their organization form (Figure 3). We found five species with strong filaments (*Klebsormidium cf. flaccidum*, *K. cf. subtile*, *K. cf. nitens*, *Xanthonema cf. exile*, *Microcoleus vaginatus*) and two genera with short or easily disintegrated filaments (*Interflum paradoxum*, *Stichococcus bacillaris*). In each BSC at least two different filamentous species were detected indicating their importance for BSC formation. Especially the genus *Klebsormidium* seemed to be highly important for BSCs in forest: in each BSC at least one of in total three observed  
30 morphospecies was found (Supp. Table S1).



## Land use intensity and soil parameters

The land use intensity was measured by applying the silvicultural management index (SMI), which based on stand density, tree species and stand age. The gravimetric water content of the bulk soil was correlated with the SMI; the pH was independent of the water content, SMI and the main tree species (Table 2).

5 Total C, N and P with a differentiation of P in its fractions were measured and presented by Baumann et al. (2017). Here, we analysed the correlation of this data with the SMI and pH of the forest plots.  
The N content correlated with the C content and both were independent of the SMI and pH. Total P and the proportion of inorganic P were independent of the C and N content, as well as from pH and SMI (Table 2).

## Correlation of algae diversity with plot characteristics and nutrient content

10 The richness of algal species and the proportion of filamentous algae in BSCs only correlated with SMI, water content and proportion of inorganic phosphorus (Table 3). All other tested parameters (C and N content, total P, proportion of organic P, pH, main tree species, and soil horizon) were excluded by stepwise model simplification based on the BIC and thus had no measurable effect on the algal species richness or proportion of filamentous algae. A higher SMI resulted in a higher species richness (Figure 2), especially the proportion of coccoid algae was enhanced.  
15 The community composition of the algae correlated with the main tree species and the water content. The SMI and proportion of inorganic P explained each 5% of the variance, but this was not significant (Table 3).

## Discussion

### Species composition and abundance

20 In total 52 algal species were identified in all BSCs sampled (Figure 1), which is a similar or lower richness compared to other reports on BSCs from temperate regions at open sites (Langhans et al., 2009; Schulz et al., 2016), but similar or higher compared to previous reports on algae from forest bulk soil (Khaybullina et al., 2010; Novakovskaya and Patova, 2008; Starks et al., 1981). Nevertheless, the given number is most probably an underestimation of the real algal biodiversity because our results are based on enrichment cultivation followed by morphological assignment. Enrichment cultivation typically covers  
25 only cultivable algae, which represent only a small part of all algae in the BSCs (Langhans et al., 2009). Furthermore, it is not always possible to distinguish dormant from currently active microalgae. Nevertheless, direct observation of a BSC sample under the microscope gave at least a hint for the dominant active organisms. Using this approach we could prove that all filamentous algae were abundant and always alive in the BSC samples. The morphological identification of algae has known challenges: e.g. sibling species have similar characteristics but are genetically distant (Potter et al., 1997). To overcome these  
30 limitations, researchers proposed to combine molecular and morphological methods, but also molecular techniques sometimes fail to detect some algae (Büdel et al., 2009; Garcia-Pichel et al., 2001).



All observed algal species are known as terrestrial taxa, most of them were also reported in BSCs (Büdel et al., 2016 and references therein; Ettl and Gärtner, 1995). Chlorophyceae were the most abundant phylum, which is typical for temperate regions (Büdel et al., 2016). In contrast, Cyanobacteria were represented by only one single species. Cyanobacteria are often reported as predominant species in BSCs in arid regions such as Israel and drylands of the USA (Garcia-Pichel et al., 2001; 5 Kidron et al., 2010). Nevertheless, Cyanobacteria are less abundant in temperate regions (Gypser et al., 2016; Langhans et al., 2009; Pluis, 1994) and even rare in acidic soils, as in the forest plots of our study site Schorfheide-Chorin (Hoffmann et al., 2007; Lukešová, 2001; Lukešová and Hoffmann, 1996). Absence or presence in small amount in forest soil is concerning with 10 low pH of soil extract which unfavorable for cyanobacteria (Hollerbach & Shtina, 1969; Hoffmann, 1989). It seems that Cyanobacteria play only a minor role in forest ecosystems with consequences for the ecological traits that some Cyanobacteria species occupy. For example, the ability for nitrogen fixation in phototrophic organisms was only reported from Cyanobacteria and never observed in eukaryotic algae. In forest ecosystems litter and other decomposable biomass provides probably sufficient mineral nitrogen compounds, which might lead to the absence of nitrogen-fixing organisms in these systems in contrast to nitrogen-poor habitats such as dunes or deserts (Langhans et al., 2009; Schulz et al., 2016).

The filamentous alga *Klebsormidium* was found in nearly all BSCs of our study, whereas species with similar strong filaments (15 *Microcoleus* and *Xanthonema*) were only found occasionally. Filamentous algae can be regarded as key players in BSCs because of their potential as BSC-initiating organisms by building tight networks among particles (Büdel et al., 2016). Some investigated forest BSC were formed as well by moss protonema, which has filamentous nature and were determined as crust-forming organism (Weber et al. 2016). However, *Klebsormidium* seems to be the most important crust-initiating alga in forest 20 ecosystems of Schorfheide-Chorin. This genus can tolerate a wide range of environmental factors and, thus, has a cosmopolitan distribution in numerous terrestrial and freshwater habitats (Karsten et al., 2016; Rindi et al., 2011 and references therein). Its presence in other terrestrial habitats such as natural rocks in plain and mountainous areas (Mikhailuk et al., 2008), caves (Vinogradova and Mikhailuk, 2009), sand dunes (Schulz et al., 2016), tree barks (Freystein et al., 2008), acidic post-mining sites (Lukešová, 2001), bases of urban walls (Rindi and Guiry, 2004) and building facades (Barberousse et al., 2006) is well 25 documented. As many terrestrial algae, *Klebsormidium* is tolerant to light exposure during dehydration (Gray et al., 2007). This is a typical situation which BSC algae have to cope with because increase of light in the morning is often associated with dehydration (Raanan et al., 2016). A recent study in Central Europe, however, observed that *Klebsormidium* is sensitive to increasing light during cellular water loss (Pierangelini et al., 2017). The distribution of *Klebsormidium* in nearly all samples from Schorfheide-Chorin forest plots may be explained by a lower radiation and also lower evaporation of water in the forest 30 ecosystem compared to open habitats (such as inland dunes), where besides *Klebsormidium* other filamentous algae were dominant (Langhans et al., 2009; Pluis, 1994). Also, the forest soil pH is rather acidic (min: 3.23; max: 3.86, Table 1) which supports a dominance of *Klebsormidium* (Škaloud et al., 2014). Thus, the low light availability, low water evaporation and the acidic soil reaction plausibly explain the presence and dominance of *Klebsormidium* as a potential BSC-initiating algal taxon in nearly all BSCs from Schorfheide-Chorin forest plots.



We identified three morpho-species of the genus *Klebsormidium* in our samples (Figure 2). All three morpho-species were reported from aeroterrestrial habitats in Central Europe (Glaser et al., 2017; Mikhailyuk et al., 2015). *Klebsormidium* has morphological features which can be easily recognized, but the identification down to species level is difficult due to morphological plasticity (Lokhorst, 1996). And still, in times of molecular identification, the debate on species definition in the genus *Klebsormidium* is ongoing (Mikhailyuk et al., 2015; Rindi et al., 2017). Therefore, the definition of clades within *Klebsormidium* was and still is a helpful tool to differentiate between morpho- or genotypes on a species-like level (Rindi et al., 2011). Studies comparing clades at different localities on the one hand observed a global ubiquity, and local endemism on the other hand (Ryšánek et al., 2014). Especially the clade composition seems to differ depending on the habitat. In detail, *Klebsormidium cf. flaccidum* (B/C clade) was abundant in closed as well as in open habitats, whereas *K. cf. nitens* and *K. cf. subtile* (E clade) were predominantly distributed in forest (Glaser et al., 2017; Mikhailyuk et al., 2015). In this study, we also observed in BSCs from forests more often *Klebsormidium cf. subtile* and *K. cf. nitens* than *K. cf. flaccidum*. Nevertheless, in desiccation experiments the recovery rates of these clades were similar (Donner et al., 2016, 2017). It is still an open question, which environmental factors caused the slight habitat preferences of the different clades. Additional ecophysiological experiments combining potential factors, such as light regimes, desiccation frequency and duration and pH, might in future explain this habitat preferences of *Klebsormidium* clades.

Unicellular algae found in investigated soil crusts are represented mostly by Chlorophyta. High biodiversity of soil algae characteristic for humid habitats (genera Chlamydomonas, Chloromonas, Chlorococcum, Tetracystis) are typical for forest soils (Hoffmann 1989).

### Correlation with SMI

The richness of algal species as well as the proportion of coccal algae were positively correlated with the silvicultural management index (SMI) that corresponded with conclusions about high algal biodiversity on disturbed or cultivated soils (Hollerbakh & Shtina, 1969; Hoffmann, 1989). The SMI reflects the main tree species and the stand density as a result of management practice. Most studies in the Biodiversity Exploratories on soil microorganisms in forests observed rather an effect of the main tree species on the community than of the SMI (Goldmann et al., 2015; Kaiser et al., 2016; Purahong et al., 2014); just one study on litter decaying fungi and bacteria observed significant difference between natural and managed beech forests (Purahong et al., 2015). Kaiser et al. (2016) discussed that the different tree species influence the community of soil bacteria by shifting the pH in soil; the pH was described in this study as the main predictor for bacterial community composition. However, the differences in the bulk soil pH between beech and pine forest were not significant in Schorfheide-Chorin (Table 1) and the algae in BSCs were not affected by soil pH. We therefore rejected an effect of the SMI via the pH on the algal species richness in Schorfheide-Chorin.

However, SMI combines other potential factors which may affect BSC microalgae, namely water regime and light availability due to stand density and tree species. The sampled forest plots in the exploratory Schorfheide-Chorin were dominated by either beech or pine trees, both differing in their light regime: in beech forests the canopy shade changes during spring and therefore



radiation on the ground can be higher in winter and spring than in pine forests. Also, the stand density, another parameter of the SMI, could affect the light regime on the ground: higher density would result in less photosynthetically active radiation for photosynthetically active soil organisms. The radiation is often coupled with evaporation of pore water (Raanan et al., 2016) and, hence, the stand density could have an indirect effect on the BSC organisms via an altered water regime. Thus, we expect that

5 the SMI affected the algae diversity in BSC via lower light availability and lower evaporation rates. This is supported by the two-way analysis of water content and SMI, both of which is described as highly important for algal species richness. Nevertheless, it should be noted that the water content was measured in the bulk soil which might differ from that of BSC. For future studies on algal biodiversity it would be important to examine also available light on the ground and the water content in the BSC.

10 Although the SMI positively affected the algal richness, the community composition was correlated with the main tree species but not with the SMI. Broadleaf litter has a higher quality in terms of a more favorable C:N and C:P ratio compared to coniferous litter (Cleveland and Liptzin, 2007; McGroddy et al., 2004). It might be that the community in pine forest is shifted towards algal species, which can cope better with low nutrient availability. But also other, above mentioned factors (light regime and water evaporation) differ between both forest types and could thus cause the observed differences in the algal  
15 community composition.

### Correlation with C/N/P

BSCs have different important ecological functions, for example BSCs enhance the nutrient content in the top soil layer (Baumann et al., 2017; Evans and Johansen, 1999). To assess the relationship between biodiversity and biogeochemical cycling in BSCs, the content of total C, N and P and additionally the different P fractions (organic, inorganic, labile and stable fractions)  
20 were correlated with algal diversity. Baumann et al. (2017) reported that the presence of BSCs leads to the enhanced content of total C, N and P and in particular the proportion of organic P. The present study shows that thereby the richness of algae was independent of the total C, N and P content. Thus, we assume that algal species are functional redundant and a low species richness can conduct the functional role of enhancing C, N and P content in BSCs. A more detailed analysis of the P fractions gave a slightly different picture: the proportion of inorganic P was correlated with the proportion of filamentous algae and  
25 shows a tendency to a correlation with the richness of BSC algae. Soluble inorganic phosphate originates either from P-mineral weathering, desorption of mineral-bound phosphates or from mineralization of organic matter (Mackey and Paytan, 2009) and can be assimilated by organisms. Thus, a low amount of inorganic P could indicate a high take-up rate of BSC organisms and, thus, a more closed P cycle due to higher algae richness (Baumann et al., 2017).



## Conclusion

This study reports for the first time on the algae biodiversity in BSCs from temperate forests with different management intensity. The rather acidic forest soil supports a clear dominance of *Klebsormidium*-morphotypes as the main crust-initiating filamentous algae, while Cyanobacteria play a negligible role in temperate forests. Moss protonema is registered as crust-forming agent in forest ecosystems as well. Higher management intensity resulted in a higher richness of algae, especially the proportion of coccal taxa increased. We expect that the land-use intensity in forests affect the algae biodiversity via changes in the light regime (less light in high stand density and in pine forests), which is positively coupled with water evaporation. As described before, BSCs enhance the content of C, N and P compared to bulk soil (Baumann et al., 2017). In this study, we observed a relation between the proportion of inorganic P with the biodiversity of algae, especially a positive correlation with the proportion of filamentous algae. Thus, the BSC does not only enhance the total amount of P but its algal biodiversity affects the proportion of the inorganic P. Forthcoming studies should include other crust-associated organisms, like fungi and bacteria, to identify key players on the ecological role of BSCs in the P cycle.

*Competing interests.* The authors declare that they have no conflict of interest.

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## References

Anderson, M. J.: A new method for non-parametric multivariate analysis of variance, *Austral Ecol.*, 26(1), 32–46, doi:10.1111/j.1442-9993.2001.01070.pp.x, 2001.

5 Barberousse, H., Tell, G., Yéprémian, C. and Couté, A.: Diversity of algae and cyanobacteria growing on building facades in France, *Algol. Stud.*, 120(1), 81–105, 2006.

Barger, N. N., Herrick, J. E., Van Zee, J. and Belnap, J.: Impacts of Biological Soil Crust Disturbance and Composition on C and N Loss from Water Erosion, *Biogeochemistry*, 77(2), 247–263, doi:10.1007/s10533-005-1424-7, 2006.

10 Baumann, K., Glaser, K., Mutz, J.-E., Karsten, U., MacLennan, A., Hu, Y., Michalik, D., Kruse, J., Eckhardt, K.-U., Schall, P. and Leinweber, P.: Biological soil crusts of temperate forests: Their role in P cycling, *Soil Biol. Biochem.*, 109, 156–166, doi:10.1016/j.soilbio.2017.02.011, 2017.

Belnap, J.: The world at your feet: desert biological soil crusts, *Front. Ecol. Environ.*, 1(4), 181–189, 2003.

15 Belnap, J., Büdel, B. and Lange, O. L.: Biological soil crusts: characteristics and distribution, in *Biological soil crusts: structure, function, and management*, vol. 1, edited by J. Belnap and O. L. Lange, pp. 3–30, Springer-Verlag, Berlin, Heidelberg., 2001.

Borchhardt, N., Schiefelbein, U., Abarca, N., Boy, J., Mikhailyuk, T., Sipman, H. J. M. and Karsten, U.: Diversity of algae and lichens in biological soil crusts of Ardley and King George islands, Antarctica, *Antarct. Sci.*, 1–9, doi:10.1017/S0954102016000638, 2017.

20 Bray, J. R. and Curtis, J. T.: An ordination of the upland forest communities of southern Wisconsin, *Ecol. Monogr.*, 27(4), 325–349, doi:10.2307/1942268, 1957.

Büdel, B., Darienko, T., Deutschwitz, K., Dojani, S., Friedl, T., Mohr, K. I., Salisch, M., Reisser, W. and Weber, B.: Southern african biological soil crusts are ubiquitous and highly diverse in drylands, being restricted by rainfall frequency, *Microb. Ecol.*, 57(2), 229–247, doi:10.1007/s00248-008-9449-9, 2009.

25 Büdel, B., Dulić, T., Darienko, T., Rybalka, N. and Friedl, T.: Cyanobacteria and Algae of Biological Soil Crusts, in *Biological Soil Crusts: An Organizing Principle in Drylands*, edited by B. Weber, B. Büdel, and J. Belnap, pp. 55–80, Springer International Publishing, Cham., 2016.



Castillo-Monroy, A., Maestre, F., Delgado-Baquerizo, M. and Gallardo, A.: Biological soil crusts modulate nitrogen availability in semi-arid ecosystems: insights from a Mediterranean grassland, *Plant Soil*, 333(1–2), 21–34, doi:10.1007/s11104-009-0276-7, 2010.

Cleveland, C. C. and Liptzin, D.: C:N:P stoichiometry in soil: is there a “Redfield ratio” for the microbial biomass?, 5 *Biogeochemistry*, 85(3), 235–252, doi:10.1007/s10533-007-9132-0, 2007.

Concostrina-Zubiri, L., Huber-Sannwald, E., Martínez, I., Flores, J. L., Reyes-Agüero, J. A., Escudero, A. and Belnap, J.: Biological soil crusts across disturbance–recovery scenarios: effect of grazing regime on community dynamics, *Ecol. Appl.*, 24(7), 1863–1877, 2014.

Darienko, T., Gustavs, L., Mudimu, O., Menendez, C. R., Schumann, R., Karsten, U., Friedl, T. and Pröschold, T.: *Chloroidium* 10, a common terrestrial coccoid green alga previously assigned to *Chlorella* (Trebouxiophyceae, Chlorophyta), *Eur. J. Phycol.*, 45(1), 79–95, doi:10.1080/09670260903362820, 2010.

Daryanto, S., Eldridge, D. J. and Wang, L.: Ploughing and grazing alter the spatial patterning of surface soils in a shrub-encroached woodland, *Geoderma*, 200–201, 67–76, doi:10.1016/j.geoderma.2013.02.006, 2013.

Donner, A., Glaser, K., Borchhardt, N. and Karsten, U.: Ecophysiological Response on Dehydration and Temperature in 15 Terrestrial *Klebsormidium* (Streptophyta) Isolated from Biological Soil Crusts in Central European Grasslands and Forests, *Microb. Ecol.*, doi:10.1007/s00248-016-0917-3, 2016.

Donner, A., Ryšánek, D., Mikhailyuk, T. and Karsten, U.: Ecophysiological traits of various genotypes of a green key alga in biological soil crusts from the semi-arid Colorado Plateau, USA, *J. Appl. Phycol.*, doi:10.1007/s10811-017-1158-7, 2017.

Elbert, W., Weber, B., Burrows, S., Steinkamp, J., Büdel, B., Andreae, M. O. and Pöschl, U.: Contribution of cryptogamic 20 covers to the global cycles of carbon and nitrogen, *Nat. Geosci.*, 5(7), 459–462, doi:10.1038/ngeo1486, 2012.

Elliott, D. R., Thomas, A. D., Hoon, S. R. and Sen, R.: Niche partitioning of bacterial communities in biological crusts and soils under grasses, shrubs and trees in the Kalahari, *Biodivers. Conserv.*, 23(7), 1709–1733, doi:10.1007/s10531-014-0684-8, 2014.

Ettl, H. and Gärtner, G.: *Syllabus der Boden-, Luft- und Flechtenalgen*, Spektrum Akademischer Verlag., 1995.

25 Evans, R. D. and Johansen, J. R.: Microbiotic crusts and ecosystem processes, *Crit. Rev. Plant Sci.*, 18(2), 183–225, doi:10.1080/0735268991309199, 1999.



Fischer, M., Bossdorf, O., Gockel, S., Hänsel, F., Hemp, A., Hessenmöller, D., Korte, G., Nieschulze, J., Pfeiffer, S., Prati, D., Renner, S., Schöning, I., Schumacher, U., Wells, K., Buscot, F., Kalko, E. K. V., Linsenmair, K. E., Schulze, E.-D. and Weisser, W. W.: Implementing large-scale and long-term functional biodiversity research: The Biodiversity Exploratories, Basic Appl. Ecol., 11(6), 473–485, doi:10.1016/j.baae.2010.07.009, 2010a.

5 Fischer, T. and Subbotina, M.: Climatic and soil texture threshold values for cryptogamic cover development: a meta analysis, Biologia (Bratisl.), 69(11), doi:10.2478/s11756-014-0464-7, 2014.

Fischer, T., Veste, M., Wiehe, W. and Lange, P.: Water repellency and pore clogging at early successional stages of microbiotic crusts on inland dunes, Brandenburg, NE Germany, CATENA, 80(1), 47–52, doi:10.1016/j.catena.2009.08.009, 2010b.

10 Flechtner, V. R., Johansen, J. R. and William, H. C.: Algal composition of microbiotic crusts from the central desert of Baja California, Mexico, Gt. Basin Nat., 58(4), 295–311, 1998.

Freystein, K., Salisch, M. and Reisser, W.: Algal biofilms on tree bark to monitor airborne pollutants, Biologia (Bratisl.), 63(6), doi:10.2478/s11756-008-0114-z, 2008.

15 Garcia-Pichel, F., Lopez-Cortes, A. and Nubel, U.: Phylogenetic and Morphological Diversity of Cyanobacteria in Soil Desert Crusts from the Colorado Plateau, Appl. Environ. Microbiol., 67(4), 1902–1910, doi:10.1128/AEM.67.4.1902-1910.2001, 2001.

Glaser, K., Donner, A., Albrecht, M., Mikhailyuk, T. and Karsten, U.: Habitat-specific composition of morphotypes with low genetic diversity in the green algal genus *Klebsormidium* (Streptophyta) isolated from biological soil crusts in Central European grasslands and forests, Eur. J. Phycol., 1–12, doi:10.1080/09670262.2016.1235730, 2017.

20 Goldmann, K., Schöning, I., Buscot, F. and Wubet, T.: Forest Management Type Influences Diversity and Community Composition of Soil Fungi across Temperate Forest Ecosystems, Front. Microbiol., 6, doi:10.3389/fmicb.2015.01300, 2015.

Gómez, D. A., Aranibar, J. N., Tabeni, S., Villagra, P. E., Garibotti, I. A. and Atencio, A.: Biological soil crust recovery after long-term grazing exclusion in the Monte Desert (Argentina). Changes in coverage, spatial distribution, and soil nitrogen, Acta Oecologica, 38, 33–40, doi:10.1016/j.actao.2011.09.001, 2012.

25 Gomez, E. D., Garland, J. L. and Roberts, M. S.: Microbial structural diversity estimated by dilution-extinction of phenotypic traits and T-RFLP analysis along a land-use intensification gradient, Fems Microbiol. Ecol., 49(2), 253–259, 2004.



Gray, D. W., Lewis, L. A. and Cardon, Z. G.: Photosynthetic recovery following desiccation of desert green algae (Chlorophyta) and their aquatic relatives, *Plant Cell Environ.*, 30(10), 1240–1255, doi:10.1111/j.1365-3040.2007.01704.x, 2007.

5 Gundlapally, S. R. and Garcia-Pichel, F.: The Community and Phylogenetic Diversity of Biological Soil Crusts in the Colorado Plateau Studied by Molecular Fingerprinting and Intensive Cultivation, *Microb. Ecol.*, 52(2), 345–357, doi:10.1007/s00248-006-9011-6, 2006.

Gypser, S., Herppich, W. B., Fischer, T., Lange, P. and Veste, M.: Photosynthetic characteristics and their spatial variance on biological soil crusts covering initial soils of post-mining sites in Lower Lusatia, NE Germany, *Flora - Morphol. Distrib. Funct. Ecol. Plants*, 220, 103–116, doi:10.1016/j.flora.2016.02.012, 2016.

10 Hoffmann, L. Algae of terrestrial habitats. *Bot. Rev.*, 55(2), 77–105, 1989.

Hoffmann, L., Ector, L. and Kostikov, I.: Algal Flora from Limed and Unlimed Forest Soils in the Ardenne (Belgium), *Syst. Geogr. Plants*, 77(1), 15–90, 2007.

Hollerbach, M.M. & E.A. Shtina. 1969. Soil algae. Nauka Press, Leningrad. [Russian]

15 Kaiser, K., Wemheuer, B., Korolkow, V., Wemheuer, F., Nacke, H., Schöning, I., Schrumpf, M. and Daniel, R.: Driving forces of soil bacterial community structure, diversity, and function in temperate grasslands and forests, *Sci. Rep.*, 6(1), doi:10.1038/srep33696, 2016.

Karsten, U., Herburger, K. and Holzinger, A.: Living in biological soil crust communities of African deserts—Physiological traits of green algal *Klebsormidium* species (Streptophyta) to cope with desiccation, light and temperature gradients, *J. Plant Physiol.*, 194, 2–12, doi:10.1016/j.jplph.2015.09.002, 2016.

20 Khaybullina, L. S., Gaysina, L. A., Johansen, J. R. and Krautová, M.: Examination of the terrestrial algae of the Great Smoky Mountains National Park, USA, *Fottea*, 10(2), 201–215, 2010.

Kidron, G. J., Vonshak, A., Dor, I., Barinova, S. and Abeliovich, A.: Properties and spatial distribution of microbiotic crusts in the Negev Desert, Israel, *CATENA*, 82(2), 92–101, doi:10.1016/j.catena.2010.05.006, 2010.

25 Kostikov, I., Darienko, T., Lukešová, A. and Hoffmann, L.: Revision of the classification system of Radiococcaceae Fott ex Komárek (except the Subfamily Dictyochlorelloideae) (Chlorophyta), *Algol. Stud.*, 104(142), 23–58, 2002.



Langhans, T. M., Storm, C. and Schwabe, A.: Community Assembly of Biological Soil Crusts of Different Successional Stages in a Temperate Sand Ecosystem, as Assessed by Direct Determination and Enrichment Techniques, *Microb. Ecol.*, 58(2), 394–407, doi:10.1007/s00248-009-9532-x, 2009.

5 Lewis, L. A.: Chlorophyta on land: independent lineages of green eukaryotes from arid lands, in *Algae and Cyanobacteria in Extreme Environments*, edited by J. Seckbach, Springer Netherlands, Dordrecht., 2007.

Lokhorst, G. M.: Comparative Taxonomic Studies on the Genus *Klebsormidium* (Charophyceae) in Europe, Gustav Fischer, Stuttgart., 1996.

Lukešová, A.: Soil algae in brown coal and lignite post-mining areas in Central Europe (Czech Republic and Germany), *Restor. Ecol.*, 9(4), 341–350, doi:10.1046/j.1526-100X.2001.94002.x, 2001.

10 Lukešová, A. and Hoffmann, L.: Soil algae from acid rain impacted forest areas of the Krušné hory Mts. 1. Algal communities, *Vegetatio*, 125(2), 123–136, 1996.

Mackey, K. R. M. and Paytan, A.: Phosphorus cycle, in *Encyclopedia of Microbiology*, vol. 3, edited by M. Schaechter, pp. 322–334., 2009.

15 McGroddy, M. E., Daufresne, T. and Hedin, L. O.: Scaling of C: N: P stoichiometry in forests worldwide: Implications of terrestrial redfield-type ratios, *Ecology*, 85(9), 2390–2401, 2004.

Mikhailyuk, T., Sluiman, H. J., Massalski, A., Mudimu, O., Demchenko, E. M., Kondratyuk, S. Y. and Friedl, T.: New streptophyte green algae from terrestrial habitats and an assessment of the genus *Interfilum* (Klebsormidiophyceae, Streptophyta), *J. Phycol.*, 44(6), 1586–1603, doi:10.1111/j.1529-8817.2008.00606.x, 2008.

20 Mikhailyuk, T., Glaser, K., Holzinger, A. and Karsten, U.: Biodiversity of *Klebsormidium* (Streptophyta) from alpine biological soil crusts (Alps, Tyrol, Austria, and Italy), edited by P. Gabrielson, *J. Phycol.*, 51(4), 750–767, doi:10.1111/jpy.12316, 2015.

Novakovskaya, I. and Patova, E.: Green algae in spruce forests in the north-east of European Russia, *Biologia (Bratisl.)*, 63(6), doi:10.2478/s11756-008-0109-9, 2008.

25 Pierangelini, M., Ryšánek, D., Lang, I., Adlassnig, W. and Holzinger, A.: Terrestrial adaptation of green algae Klebsormidium and Zygnema (Charophyta) involves diversity in photosynthetic traits but not in CO<sub>2</sub> acquisition, *Planta*, doi:10.1007/s00425-017-2741-5, 2017.



Pluis, J. L. A.: Algal crust formation in the inland dune area, Laarder Wasmeer, the Netherlands, *Plant Ecol.*, 113(1), 41–51, 1994.

Potter, D., Lajeunesse, T. C., Saunders, G. W. and Anderson, R. A.: Convergent evolution masks extensive biodiversity among marine coccoid picoplankton, *Biodivers. Conserv.*, 6(1), 99–107, 1997.

5 Purahong, W., Hoppe, B., Kahl, T., Schloter, M., Schulze, E.-D., Bauhus, J., Buscot, F. and Krüger, D.: Changes within a single land-use category alter microbial diversity and community structure: molecular evidence from wood-inhabiting fungi in forest ecosystems, *J. Environ. Manage.*, 139, 109–119, 2014.

Purahong, W., Kapturska, D., Pecyna, M. J., Jariyavidyanont, K., Kaunzner, J., Juncheed, K., Uengwetwanit, T., Rudloff, R., Schulz, E., Hofrichter, M., Schloter, M., Krüger, D. and Buscot, F.: Effects of Forest Management Practices in Temperate  
10 Beech Forests on Bacterial and Fungal Communities Involved in Leaf Litter Degradation, *Microb. Ecol.*, 69(4), 905–913, doi:10.1007/s00248-015-0585-8, 2015.

R Development Core Team: R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna Austria [online] Available from: <http://www.R-project.org/>, 2009.

15 Raanan, H., Oren, N., Treves, H., Berkowicz, S. M., Hagemann, M., Pade, N., Keren, N. and Kaplan, A.: Simulated soil crust conditions in a chamber system provide new insights on cyanobacterial acclimation to desiccation: Simulation of BSC conditions and acclimation, *Environ. Microbiol.*, 18(2), 414–426, doi:10.1111/1462-2920.12998, 2016.

Rindi, F. and Guiry, M. D.: Composition and spatial variability of terrestrial algal assemblages occurring at the bases of urban walls in Europe, *Phycologia*, 43(3), 225–235, 2004.

20 Rindi, F., Mikhailyuk, T. I., Sluiman, H. J., Friedl, T. and López-Bautista, J. M.: Phylogenetic relationships in *Interfilum* and *Klebsormidium* (Klebsormidiophyceae, Streptophyta), *Mol. Phylogenet. Evol.*, 58(2), 218–231, doi:10.1016/j.ympev.2010.11.030, 2011.

Rindi, F., Rysanek, D. and Skaloud, P.: Problems of epitypification in morphologically simple green microalgae: a case study of two widespread species of *Klebsormidium* (Klebsormidiophyceae, Streptophyta), *Fottea*, doi:10.5507/fot.2016.017, 2017.

25 Ryšánek, D., Hrčková, K. and Škaloud, P.: Global ubiquity and local endemism of free-living terrestrial protists: phylogeographic assessment of the streptophyte alga *Klebsormidium*: Global biogeography of a terrestrial protist, *Environ. Microbiol.*, 17(3), 689–698, doi:10.1111/1462-2920.12501, 2014.



Schall, P. and Ammer, C.: How to quantify forest management intensity in Central European forests, *Eur. J. For. Res.*, 132(2), 379–396, doi:10.1007/s10342-013-0681-6, 2013.

Schulz, K., Mkhailyuk, T., Dreßler, M., Leinweber, P. and Karsten, U.: Biological Soil Crusts from coastal dunes at the Baltic Sea: cyanobacterial and algal biodiversity and related soil properties, *Microb. Ecol.*, 71(1), 178–193, doi:10.1007/s00248-015-0691-7, 2016.

Škaloud, P., Lukešová, A., Malavasi, V., Ryšánek, D., Hrčková, K. and Rindi, F.: Molecular evidence for the polyphyletic origin of low pH adaptation in the genus *Klebsormidium* (Klebsormidiophyceae, Streptophyta), *Plant Ecol. Evol.*, 147(3), 333–345, doi:10.5091/plecevo.2014.989, 2014.

Starks, T. L., Shubert, L. E. and Trainor, F. R.: Ecology of soil algae: a review, *Phycologia*, 20(1), 65–80, 1981.

10 Starr, R. C. and Zeikus, J. A.: UTEX the culture collection of algae at the University of Texas at Austin, *J. Phycol.*, 29 (Suppl.), 1–106, 1993.

Steven, B., Gallegos-Graves, L. V., Belnap, J. and Kuske, C. R.: Dryland soil microbial communities display spatial biogeographic patterns associated with soil depth and soil parent material, *FEMS Microbiol. Ecol.*, 86(1), 101–113, doi:10.1111/1574-6941.12143, 2013.

15 Vinogradova, O. N. and Mkhailyuk, T. I.: Algal flora of the caves and grottoes of the National Nature Park “Podilsky Tovtry” (Ukraine), *Int. J. Algae*, 11(3), 289–304, doi:10.1615/InterJAlgae.v11.i3.80, 2009.

Weber, B., Büdel, B. and Belnap, J.: Biological soil crusts: an organizing principle in drylands, Springer Nature., 2016.

Williams, W. J., Eldridge, D. J. and Alchin, B. M.: Grazing and drought reduce cyanobacterial soil crusts in an Australian Acacia woodland, *J. Arid Environ.*, 72(6), 1064–1075, doi:10.1016/j.jaridenv.2007.11.017, 2008.



Table 1. Sample location, main tree species, silvicultural management index (SMI), soil horizon material, on which BSC was growing, water content and pH from bulk soil analyses (water content and pH were kindly provided by Ingo Schöning).

Plot	latitude	longitude	main tree species	SMI	Soil Horizon	Water content (weight %)	pH (CaCl <sub>2</sub> )
SW_01	52.9008473	13.846367	pine	0.351	A	12.1	3.64
SW_02	52.9517289	13.7780281	pine	0.329	B	14.4	3.60
SW_03	52.9207074	13.6430016	pine	0.334	A	11.7	3.47
SW_04	52.9173466	13.8473114	pine	0.136	A	13.9	3.50
SW_05	53.0570336	13.8853662	beech	0.211	A	13.9	3.42
SW_06	53.0570336	13.8853662	beech	0.211	A	13.9	3.42
SW_07	52.9074428	13.841688	beech	0.319	A	17.9	3.67
SW_08	52.9074428	13.841688	beech	0.319	C	17.9	3.67
SW_09	53.107348	13.6944189	beech	0.082	C	18.6	3.73
SW_10	53.107348	13.6944189	beech	0.082	A	18.6	3.73
SW_11	53.1917972	13.9303379	beech	0.059	A	20.7	3.38
SW_12	53.1917972	13.9303379	beech	0.059	A	20.7	3.38
SW_13	53.0445869	13.8101029	beech	0.017	B	16.4	3.56
SW_14	53.0445869	13.8101029	beech	0.017	C	16.4	3.56
SW_15	53.0910959	13.6378431	pine	0.381	A	9.9	3.70
SW_16	53.0902937	13.6337038	pine	0.281	A	12.4	3.66
SW_17	52.9179137	13.7521735	pine	0.276	A	15.8	3.38
SW_18	52.9145421	13.7375526	pine	0.330	C	6.1	3.72
SW_19	53.0765832	13.8639856	pine	0.335	A	8.4	3.57
SW_20	53.0886055	13.6353842	pine	0.357	A	9.0	3.66
SW_21	52.9155883	13.7404509	pine	0.218	A	13.0	3.44
SW_22	52.8958259	13.852147	pine	0.217	C	13.3	3.47
SW_23	52.8958259	13.852147	pine	0.217	A	13.3	3.47
SW_24	52.9400216	13.7826121	beech	0.161	A	16.8	3.62
SW_25	52.9400216	13.7826121	beech	0.161	C	16.8	3.62
SW_26	52.9147688	13.8623651	beech	0.250	C	15.7	3.68
SW_27	52.9147688	13.8623651	beech	0.250	A	15.7	3.68
SW_28	52.9009765	13.9283256	beech	0.229	A	18.9	3.72
SW_29	52.9009765	13.9283256	beech	0.229	A	18.9	3.72



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SW_30	53.0512659	13.8449954	beech	0.070	A	14.1	3.71
SW_31	53.0512659	13.8449954	beech	0.070	C	14.1	3.71

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Table 2. Significant Pearson correlation coefficients to reveal correlations between environmental factors, which might affect or be affected by the biodiversity of algae. This co-correlation analysis should support the correct interpretation of potential important factors of the biodiversity. SMI-silvicultural management index; n.s. – not significant

	main tree species	SMI	water content	pH	C content	N content	P content
SMI	-0.6						
water content	0.77	-0.59					
pH	n.s.	n.s.	n.s.				
C content	n.s.	n.s.	n.s.	n.s.			
N content	n.s.	n.s.	n.s.	n.s.	0.94		
P content	n.s.	n.s.	n.s.	n.s.	n.s.	n.s.	
proportion of inorganic P	n.s.	n.s.	n.s.	n.s.	n.s.	-0.78	0.6

5

Table 3. Effect of environmental factors on algae richness, filamentous algae proportion (both estimated by ANOVA) and community composition of algae (estimated by PerMANOVA) quantified by the percentage of explained variance. The significance level is indicated by: \*\*\*-p<0.001, \*\*-p<0.01, \*-p<0.05, °-p<0.1, ns- not significant

10

	algae richness	proportion of filamentous algae	community composition
SMI	30.5 % **	37.7 % ***	5.6 % n.s.
water content	15.7 % *	14.0 % **	9.6 % *
proportion inorganic P	11.0 % °	29.1 % ***	5.8 % n.s.
main tree species	0.9 % n.s.	0.3 % n.s.	14.7 % ***

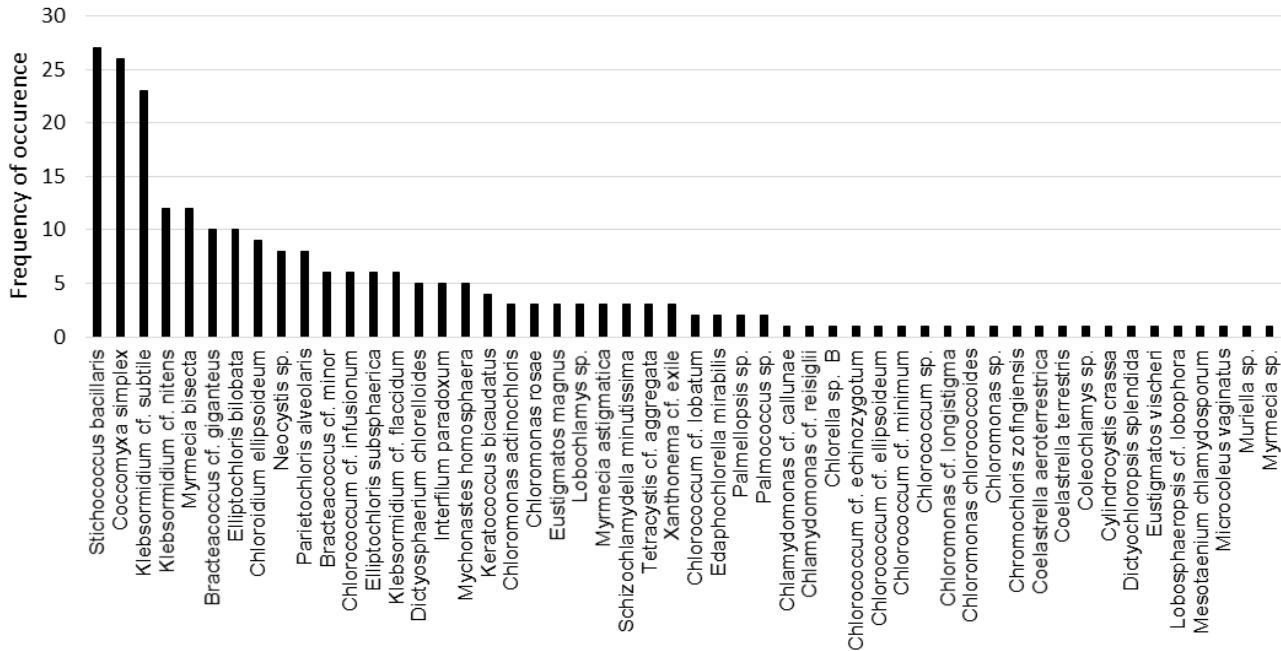


Figure 1. Frequency of occurrence of each algae species in biological soil crust from forest sites (n=31).

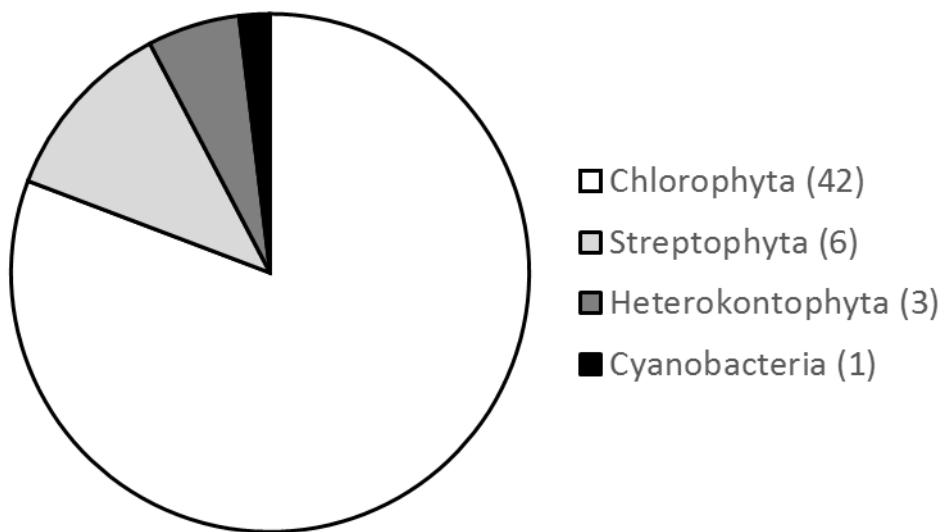


Figure 2. Distribution of algae phyla in BSCs from 31 forest plots based on enrichment cultivation and morphological identification; total number of species for each phylum is given in brackets.

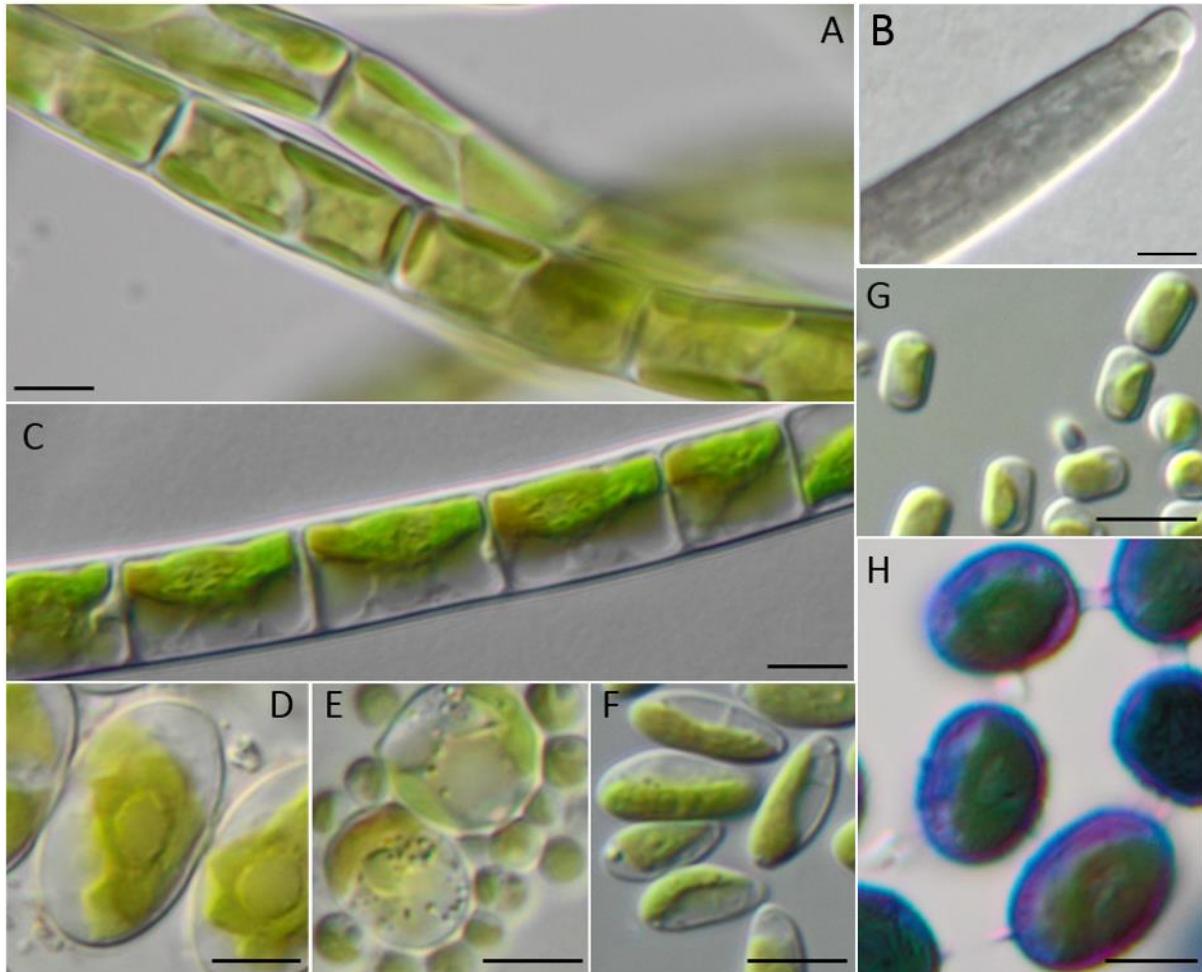


Figure 3. Filamentous and some examples of coccal algae from forest BSCs: algae with strong filaments: A-*Xanthonema cf. exile*, B-*Microcoleus vaginatus*, C-*Klebsormidium cf. flaccidum*; coccal algae: D-*Chloroidium ellipsoideum*, E-*Eustigmatos magnus*, F-*Coccomyxa simplex*; algae with short or easily disintegrated filaments: G-*Stichococcus bacillaris*, H-*Interfilum paradoxum* (mucilage bridges were coloured with methylene blue); scale bar = 5µm

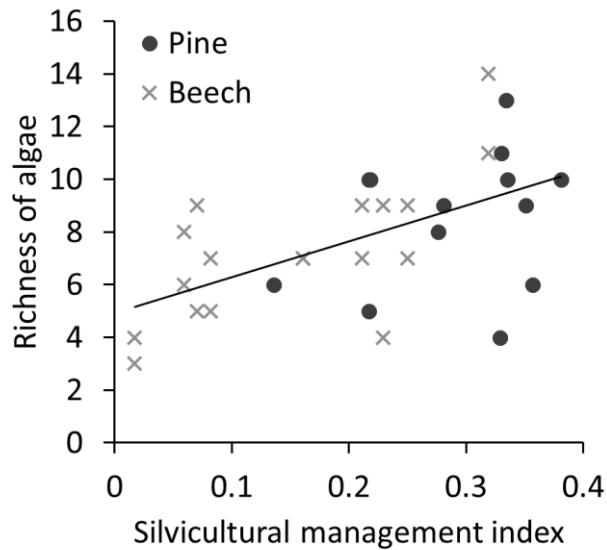


Figure 4. Plot of algae richness over the silvicultural management index (SMI), the line indicates the best linear fit (slope: 13.6,  $p<0.001$ )