Interactive Comment on **"Tracing the origin of the oxygen-consuming organic matter in the hypoxic zone in a large eutrophic estuary: the lower reach of the Pearl River Estuary, China" by Su et al.**

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Referee #1

Authors responded to all my original comments, but I feel that authors do not take into account main points of my comments.

- Uncertainties and difference between the CJE and PRE (similar issue was raised by reviewer #1): Authors re-estimated 13C value of organic carbon remineralized in the hypoxic zone to be -23.2 +/- 1.1, which was originally -21.8 +/- 0.6. Such a large change seems to be simply due to slight changes in end-member values. Authors may have to allow seasonal and spatial variations of end-members. Indeed, the large scattering and outlier evaluation in Figure 8 will result in much larger uncertainties in 13C value of OC remineralized. Authors should explain how the scattering and outlier are included in the uncertainty (+/- 1.1) of OC remineralized values calculated. In addition, I still do not see the visible difference between CJE and PRE in Figure 8 considering the scattering and natural variations. I think that authors try to obtain the accuracy that the method cannot provide.

[Response]: We have to clarify that we did not change the end-member values of our three end-member mixing model in either θ, S, DIC or $\delta^{13}C_{DIC}$. These end-member values have been well justified by taking into account the uncertainties as explicitly explained in our revised submission. The uncertainties of end-member values mostly come from averaging values of several stations at certain salinity, thus including spatial variations to some extent.

The reviewer might be right about the seasonality of the end-member values but here we are examining the summer bottom hypoxia where the mixing scheme of the water masses is rather clear as demonstrated in the T-S diagram shown in Fig. 7a.

A broader range of data at oxygen deficiency (n=38) (instead of those associated with severe oxygen deficiency at AOU> 100 μmol kg^{-1} (n=9) in our original submission) is now being used to derive $\delta^{13}C_{OCx}$ considering on the main comments from Reviewer #1. This re-estimation is justified because the oxygen depletion generating from organic matter remineralization distributes as a continuum in the water column and the accumulative oxygen reduction ultimately results in hypoxia formation.

As shown in Fig. 8, we plotted Δ ($\delta^{13}C_{DIC} \times$ DIC) against ΔDIC including all the scattering points in the subsurface water in the PRE, and obtained a slope of -23.2 ±1.1 ‰ with a high correlation coefficient (R^2=0.86, P< 0.001). Thus, the original $\delta^{13}C$ signature of the remineralized organic matter is -23.2 ‰ with an uncertainty of ±1.1 ‰.

The propagation error is shown in the following figure (Fig. S1). We are not sure how the reviewer concluded that there was no visible difference between PRE and CJE. The linear regression slope of the PRE is -23.2 ±1.1 ‰, which is statistically different from that of the CJE (-18.5 ±1.0 ‰, R^2=0.6, P< 0.001). Our statistic has a confidence level of 95 %.

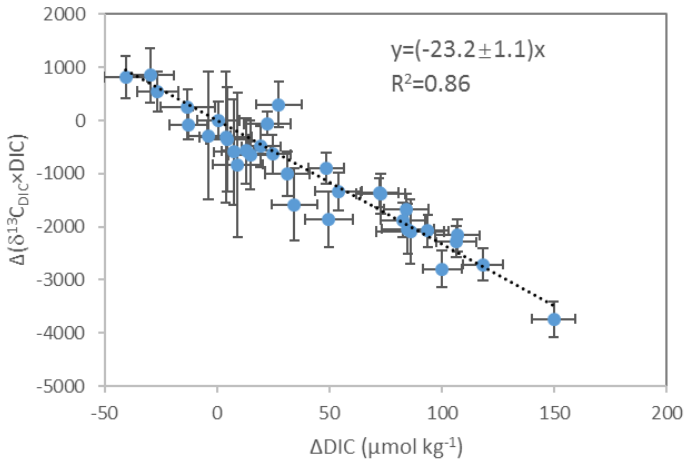

Figure S1. Propagation errors for Δ ($\delta^{13}C_{DIC} \times$ DIC) and ΔDIC in Fig. 8.

- Use of Ca2+ as a conservative tracer: I am not convinced why authors use Ca, instead of salinity, as a conservative tracer although Ca is less conservative.

[Response]: We perfectly agreed that Ca^{2+} is a less conservative tracer than salinity. The fact is that salinity has been applied as a conservative tracer in our three end-member mixing model, and thus we ought to find alternative independent tracers to validate the model. Ca^{2+} is obviously such an alternative. As we explained in our previous responses, Ca^{2+} is conservative in our system as supported by a strong linear relationship between surface water Ca^{2+} and salinity, and aragonite oversaturation (Ω_{arag}=2.6±0.7) in the subsurface water.

- Relative contribution of bottom sediment OM: Authors may compare the water column DO consumption rates relative to the bottom sediment DO consumption rates in order to show that the contribution of bottom sediments are negligible. Figure 4 shows stronger DO consumption in the bottom layer intuitively.

[Response]: In the present study, we used bottom water taken from Station A10 on 27 July and conducted on-deck incubation experiments to estimate total oxygen consumption rate, which was 9.8 μmol L^{-1} d^{-1}. Such a water column oxygen consumption rate could well support the oxygen decline rate observed in situ at Station A10 in the hypoxic zone between 20 July and 27 July (Fig. 6), which was 7.7 μmol L^{-1} d^{-1}. This first order comparison strongly suggests that water column oxygen consumption may be predominate in the formation of the hypoxia in the present case.

Note that we cannot distinguish between the contribution of resuspended sediments vs. sinking POC to oxygen depletion, because both of these isotopic signals would have been reflected in oxidation product (DIC) and the derived $\delta^{13}C$ value of the remineralized OC ($\delta^{13}C_{OCx}$). Please see details in our previous response to comment (3) of Referee #2. Quantifying the relative contribution of bottom sediment OM is beyond the scope of our present study. Moreover, the bottom sediment DO consumption rate is still controversial due to its high spatial variability.

In responding to the concerns of the reviewer, we have added in the further revised manuscript our water column total oxygen consumption rate determined by on-deck

incubation experiments using bottom water taken from Station A10 (Table S1). "As a first order comparison, the water column total oxygen consumption rate of 9.8 μmol L^{-1} d^{-1} could well support the oxygen decline rate observed at Station A10 in the hypoxic zone between 20 July and 27 July (Fig. 6), which was 7.7 μmol L^{-1} d^{-1}. This comparison along with the stoichiometry between ΔDIC and AOU strongly suggests that water column aerobic respiration may be predominate in the formation of the hypoxia in the present case." Please see P4 L14-20 and P10 L12-18 in the further revised manuscript.

Table S1. Comparison of oxygen consumption rate between in situ observations and on-deck incubation experiments at Stations A10 in the hypoxic zone.

Station	$DO_{\_ini}$ (μmol L^{-1})	$DO_{\_end}$ (μmol L^{-1})	Interval (d)	ΔDO (μmol L^{-1})	DO decline rate (μmol L^{-1} d^{-1})	Total oxygen consumption rate (μmol L^{-1} d^{-1})
A10	156	101	7.1	55	7.7	9.8

Note that the subscripts "ini" means the initial sampling and "end" means the end sampling. The Interval shows the days between these two repeated sampling. DO decline rate equals to the amount of DO reduction dividing by interval.

- Significant figures: Authors show that DO and DIC precisions are +/- 1 and 2 respectively. But, they say that "we kept one decimal place for DO/DIC…". Authors should explain why they keep one decimal place although the values are meaningless.
[Response]: Accepted. Do not keep one decimal place of DO/DIC in the main text.

Referee #2

The revisions made in the paper by Su et al. have substantially improved the quality of the paper which, to my opinion, is now ready for publication. You have answered the main points raised in my previous review, especially the one dealing with end-members which is crucial in quantifying the uncertainty of the message provided by the paper. I can see that you have changed the value and expanded the uncertainty of your proportion of terrigeneous OM in the mix generating hypoxia.

I particularly appreciated the long and detailed response letter which made the revisions

easy to check and evaluate.

I have just a few technical points which should further improve the clarity of your paper:

- You use very often "pre" for predicted (by the mixing model) and "PRE" for Pearl River Estuary. It is confusing in the equations and Figures when endmembers are quoted (Eq. 7 page 10, or Figure caption and axis labels of Figure 7). In this Figure (7) especially, "PRE" and "pre" co-exist in the axis title which should be changed. You should choose one of them (e.g. Pearl River Estuary) and replace "pre" by "mod" for model.

[Response]: Accepted. Use “con” (conservative) instead of “pre” (predicted) throughout the manuscript.

- You use very often "PRE" for Pearl River Estuary and CJE for "ChangJiang Estuary". At least for the Figure captions, I would recommand to replace CJE by ChangJiang Estuary, as it is not so obvious for most readers and it would make the Figure easier to read.

[Response]: Accepted. Please see captions in Figs. 5, 7 and 8.

Tracing the origin of the oxygen-consuming organic matter in the hypoxic zone in a large eutrophic estuary: the lower reach of the Pearl River Estuary, China

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Abstract. We assess the relative contributions of different sources of organic matter, marine vs. terrestrial, to oxygen consumption in an emerging hypoxic zone in the lower Pearl River Estuary (PRE), a large eutrophic estuary located in Southern China. Our cruise, conducted in July 2014, consisted of two legs before and after the passing of Typhoon Rammasun, which completely de-stratified the water column. The stratification recovered rapidly, within one day after the typhoon. We observed algal blooms in the upper layer of the water column and hypoxia underneath in bottom water during both legs. Repeat sampling at the initial hypoxic station showed severe oxygen depletion down to 30 μmol kg^{-1} before the typhoon and a clear drawdown of dissolved oxygen after the typhoon. Based on a three end-member mixing model and the mass balance of dissolved inorganic carbon and its isotopic composition, the $\delta^{13}C$ of organic carbon remineralized in the hypoxic zone was -23.2 ±1.1 ‰. We estimated that 65 ±16 % of the oxygen-consuming organic matter was derived from marine sources, and the rest (35 ±16 %) was derived from the continent. In contrast to a recently studied hypoxic zone in the East China Sea off the Changjiang Estuary where marine organic matter stimulated by eutrophication dominated oxygen consumption, here terrestrial organic matter significantly contributed to the formation and maintenance of hypoxia. How varying amounts of these organic matter sources drive oxygen consumption has important implications for better understanding hypoxia and its mitigation in bottom waters.

1 Introduction

The occurrence of hypoxia has been exacerbated worldwide (Nixon, 1995; Diaz and Rosenberg, 2008; Rabalais et al., 2010; Zhang et al., 2013). In recent decades, more than 400 coastal hypoxic systems have been reported with an exponential growth rate of 5.5 ± 0.23 % yr^{-1}, demonstrating their persistence and complexity with respect to both science and management (Diaz and Rosenberg, 2008; Vaquer-Sunyer and Duarte, 2008). Hypoxia may not only reduce biodiversity and endanger aquatic and benthic habitats, but also alter the redox chemistry in both the water column and the underlying sediments, triggering the release of secondary pollutants (Breitburg, 2002; Rutger et al., 2002). Moreover, the management and recovery of these systems is complicated due to the hysteresis of hypoxic conditions, and the varying timescales of biological loss (within hours to weeks) and recovery from hypoxia (from months to years) (Steckbauer et al., 2011).

Coastal hypoxia usually occurs in stratified water columns where the downward mixing of oxygen from the surface is impeded (Kemp et al., 2009). Below the pycnocline, aerobic respiration is usually the predominant sink of oxygen. Organic matter, which consumes dissolved oxygen (DO) as it becomes oxidized, is thus the ultimate cause of hypoxia under favourable physical settings (Rabouille et al., 2008; Rabalais et al., 2014; Qian et al., 2016). The organic carbon (OC) that fuels respiration-driven reduction of oxygen in these systems could originate from either eutrophication-induced primary production (marine OC; OC_{mar}), or naturally and/or anthropogenically driven delivery from terrestrial environments (terrestrial OC; OC_{terr}) (Paerl, 2006; Rabalais et al., 2010).

The question of how much OC in hypoxic zones is supplied from on-site primary production versus the quantity derived from terrestrial sources has been an issue of debate (Wang et al., 2016). Much of the phytoplankton-centric hypoxia literature suggests that OC_{mar} dominates oxygen consumption in hypoxic zones, owing to its higher microbial availability than OC_{terr} (Zimmerman and Canuel, 2000; Boesch et al., 2009; Carstensen et al., 2014). Wang et al. (2016) quantified for the first time the relative contributions of particulate OC_{mar} (POC_{mar}) and particulate OC_{terr} (POC_{terr}) in consuming DO in the bottom waters of the East China Sea (ECS) off the Changjiang Estuary (CJE), and found that POC_{mar} dominated DO consumption. However, other studies suggest that POC_{terr} may also play an important role (Swarzenski et al., 2008;

Bianchi, 2011a; Bianchi et al., 2011b). It is thus very important to quantify the relative contributions of organic matter (OC_{mar} vs. OC_{terr}) driving the onset and maintenance of hypoxia in coastal systems, since reducing organic matter vs. nutrient inputs requires a different set of management strategies.

The Pearl River Estuary (PRE, 21.2 °N–23.1 °N, 113.0 °E–114.5 °E) is surrounded by several large cities including Hong Kong, Shenzhen and Guangzhou and has received very high loads of nutrients from the drainage basin in the last three decades. As such, eutrophication has increasingly become an issue of concern (Huang et al., 2003; Ye et al., 2012). Dissolved inorganic nitrogen (DIN) concentrations in the PRE have increased approximately 4-fold from 1986 (19.3 μmol L^{-1}) to 2002 (76.1 μmol L^{-1}) (He and Yuan, 2007). This DIN increase has been attributed to increased inputs of domestic sewage, industrial wastewater, agricultural runoff and aquaculture in the watershed (Huang et al., 2003).

Recent observations based on monthly surveys between April 2010 and March 2011 and long term monitoring data from 1990 to 2014, have suggested that the lower PRE has emerged as a seasonal hypoxic zone (Qian et al., 2017). This is supported by our current study, as two relatively large hypoxic zones (> 300 km^2) were observed in the lower PRE with DO < 2 mg L^{-1}. However, the origin of the organic matter driving hypoxia in the lower PRE has not previously been examined. Here, we quantified the relative proportions of OC_{mar} and OC_{terr} contributing to DO drawdown in bottom waters of the lower PRE, an economically important coastal region. This study has important biological, societal and managerial implications for the region, particularly relating to water quality in the vicinity of Hong Kong in the lower PRE. For example, the government of Hong Kong is examining the efficacy of its costly Harbour Area Treatment Scheme project and if additional treatment should be implemented (http://www.gov.hk/en/residents/environment/water/harbourarea.htm).

2 Materials and Methods

2.1 Sampling and analysis

Interrupted by Typhoon Rammasun on 17-18 July 2014, our cruise was divided into two legs (Fig. 1). During Leg 1 on 13–16 July, we sampled Transects F4, F5 and Stations A08–A18. During Leg 2 on 19–27 July, we sampled Stations A01–A10, Transects F3 and F4, Stations A11–A17 and Transects F5, F6, F1, and F2, in sequence.

In order to monitor the development of hypoxia before and after the passage of the typhoon, we revisited Station A10 three more times (13, 20 and 27 July).

According to the gauge in the upper Pearl River, water discharge peaked in June and July. Typhoon Rammasun increased discharge during 15-18 July, with daily average values of 19480, 26115, 22981 and 17540 m^3 s^{-1}, respectively. Nevertheless, the freshwater discharge was 18908 m^3 s^{-1} in leg 1 and 15698 m^3 s^{-1} in leg 2, comparable to the long-term (2000–2011) monthly average.

Temperature and salinity were determined with a SBE 25 Conductivity-Temperature-Depth/Pressure unit (Sea-Bird Co.). Water samples were collected using 4 L Go-Flo bottles (General Oceanics). DIC and DO was measured at all stations with depth profiles. Samples for $\delta^{13}C_{DIC}$ were collected primarily along Transect A as well as at depth in low oxygen layers.

The DO concentrations in discrete water samples were measured on board within 8 h using the classic Winkler titration method (Dai et al., 2006). In addition, we conducted on-deck incubation experiments using unfiltered water taken from the hypoxic zone on 27 July, 2014 following He et al. (2014). Bottom water from ~2 m above the sediment surface was collected and incubated for 24 hours in 65 mL BOD bottles in dark at ambient temperature controlled by the flowing surface water. Total oxygen consumption rate was determined by comparing the DO concentration at the initial and end point of the experiment.

DIC was measured with an infrared detector after acidifying 0.5–0.7 mL of water sample with a precision of 0.1 % for estuarine and sea waters (Cai et al., 2004). Dissolved calcium concentrations (Ca^{2+}) were determined using an EGTA titration with a Metrohm 809 TITRANDO potentiometer, which has a precision better than ± 5 μmol kg^{-1} (Cao et al., 2011).

For $\delta^{13}C_{DIC}$ analysis, an ~20 mL DIC sample was converted into gaseous CO_2 and progressively purified through a vacuum line. The pure CO_2 sample was analyzed with an isotope ratio mass spectrometer (IRMS, Finnigan MAT 252, Bremen, Germany). The analytical precision was better than 0.1 ‰.

Water samples for TSM (total suspended matter), POC and $\delta^{13}C_{POC}$ analysis were concentrated onto preweighed and pre-combusted 0.7 μm Whatman GF/F filters after filtering 0.2–1.0 L of water under a mild vacuum (~ 25 kPa). Filters were washed with distilled water and stored at -20 ℃. Prior to analysis, all filters were freeze-dried. TSM

was determined using the net weight increment on the filter and the filtration volume. Filters were decarbonated with 1.0 mol L^{-1} HCl and dried at 40 ℃ for 48 h (Kao et al., 2012) and analyzed for POC and $\delta^{13}C_{POC}$ on an elemental analyzer coupled with an IRMS (EA-IRMS). The analytical precision for $\delta^{13}C_{POC}$ was better than 0.1 ‰. Chl-a was measured with a Turner fluorometer after extracting filters with 90 % acetone (He et al., 2010b). Calibrations were performed using a Sigma Chl-a standard.

2.2 Three end-member mixing model

We adopted a three end-member mixing model to construct the conservative mixing scheme among different water masses (Cao et al., 2011; Han et al., 2012):

$$F_{RI}+F_{SW}+F_{SUB}=1 \quad (1)$$

$$\theta_{RI}\times F_{RI}+\theta_{SW}\times F_{SW}+\theta_{SUB}\times F_{SUB}=\theta \quad (2)$$

$$S_{RI}\times F_{RI}+S_{SW}\times F_{SW}+S_{SUB}\times F_{SUB}=S \quad (3)$$

where θ and S represent potential temperature and salinity; the subscripts RI, SW, and SUB denote the three different water masses (Pearl River plume water, offshore surface seawater and upwelled subsurface water); and F_{RI}, F_{SW}, and F_{SUB} represent the fractions that each end-member contributes to the in situ samples. These fractions were applied to predict conservative concentrations of DIC (DIC_{con}) and its isotopic composition ($\delta^{13}C_{DICcon}$) resulting solely from conservative mixing.

$$DIC_{RI}\times F_{RI}+DIC_{SW}\times F_{SW}+DIC_{SUB}\times F_{SUB}=DIC_{con} \quad (4)$$

$$\frac{\delta^{13}C_{DICRI}\times DIC_{RI}\times F_{RI}+\delta^{13}C_{DICSW}\times DIC_{SW}\times F_{SW}+\delta^{13}C_{DICSUB}\times DIC_{SUB}\times F_{SUB}}{DIC_{con}}=\delta^{13}C_{DICcon} \quad (5)$$

The difference (Δ) between measured and conservative DIC values represents the magnitude of the biological alteration of DIC (Wang et al., 2016).

3 Results

3.1 Horizontal distribution

Although the average freshwater discharge rate during our sampling period (16369 m^3 s^{-1}) was slightly higher than the multi-year (2000–2011) monthly average (15671 m^3 s^{-1}), typhoon Rammasun modified the system to some extent as shown from the evolution of chemical species at Station A10 before and after the typhoon (See Sect. 3.4). The interruption of Leg 1 due to the typhoon (July 17-18) led to a smaller survey area, covering only outside Lingdingyang Bay (traditionally regarded as the PRE),

while Leg 2 covered Lingdingyang Bay from the Humen Outlet to the adjacent coastal sea.

As depicted in Fig. 2, the sea surface temperature (SST) during Leg 1 (28.9-32.2 ℃) was slightly higher than during Leg 2 (28.9-31.0 ℃). Sea surface salinity (SSS) measurements showed that plume water was restricted more landward during Leg 2 than Leg 1. However, a steeper gradient to higher SST offshore during Leg 1 was likely induced by the upwelling of bottom water, featuring relatively high SSS (18.6), high DIC (1789 μmol kg^{-1}) and low DO saturation (DO%, 86 %). During Leg 1, the region with the most productivity was found east of the Wanshan Islands, characterized by high concentrations of Chl-a (8.0 μg kg^{-1}), low concentrations of DIC (1607 μmol kg^{-1}), and DO supersaturation, with the highest DO% greater than 160 % at Station F503. During Leg 2, there were three patches of high productivity, south of Huangmaohai, at the PRE entrance, and off Hong Kong. The central region of high productivity had the highest DO%, greater than 140% at Station A14, and was characterized by relatively high concentrations of Chl-a (7.8 μg kg^{-1}) and low concentrations of DIC (1737 μmol kg^{-1}).

As shown in Fig. 3, bottom water hypoxia during Leg 1 was located more centrally in the study area relative to the surface phytoplankton bloom. The center of the hypoxic zone was found at Station A10, characterized by the lowest observed DO concentrations (as low as 30 μmol kg^{-1}) and a relatively high concentration of DIC (2075 μmol kg^{-1}). During Leg 2, hypoxic conditions were no longer found at Station A10, and instead the largest hypoxic zone was discovered to the southwest of the Wanshan Islands, where the lowest DO values were observed (as low as 7 μmol kg^{-1} at F304), and once again coincided with relatively high concentrations of DIC (2146 μmol kg^{-1}). We were unable to precisely constrain the areas of the regions impacted by bottom water hypoxia due to the limited spatial coverage, but our results suggest it covered an area of > 280 km^2 during Leg 1 and > 290 km^2 during Leg 2 according to the definition of hypoxia as DO < 2 mg L^{-1} or 63 μM, or an area of > 900 km^2 during Leg 1 and > 800 km^2 during Leg 2 assuming the threshold of the oxygen-deficit zone was < 3 mg L^{-1} or 95 μM (Rabalais et al., 2010; Zhao et al., 2017).

3.2 Vertical distribution

During Leg 1, plume water reached 50 km offshore from the entrance of the PRE, forming a 5–10 m thick surface layer (Fig. 4b). Both the thermocline and halocline contributed to the stability of the water column structure, which favored the formation of bottom water hypoxia. The thickness of the bottom water hypoxic layer was ~ 5 m. The region of highest productivity, however, was not observed in the same location as the hypoxic zone, but further offshore.

During Leg 2, although the passing of the typhoon would be expected to absorb large amounts of potential heat and cause extensive mixing of the water column, the enhanced freshwater discharge could rapidly re-stratify the water column and facilitate the re-formation of hypoxia. This time, the primary region of hypoxia was observed directly below the bloom, with a thickness of 3 m (Fig. 4i). Additionally, near the Humen Outlet we observed low DIC (1466 μmol kg^{-1}) and moderately low DO (89 μmol kg^{-1}), which reflected the input of the low DO water mass from upstream as reported previously (Dai et al., 2006; Dai et al., 2008a; He et al., 2014).

3.3 Isotopic composition of DIC and POC

The $\delta^{13}C$ values of DIC became progressively heavier from stations dominated by freshwater (~ -11.4 ‰) to off-shore seawater (~ -0.6 ‰), with a relatively wide range of values beyond a salinity of 13 (Fig. 5). Owing to a malfunction of the instrument, $\delta^{13}C_{POC}$ data from our cruise were not available. Instead, we reported a valid $\delta^{13}C_{POC}$ dataset from a 2015 summer cruise in approximately the same region. $\delta^{13}C_{POC}$ values showed a similar trend with $\delta^{13}C_{DIC}$, i.e. ^{13}C enriched seaward, from ~ -28 ‰ to ~ -20 ‰. In the bloom, where the DO% was above 125 %, the mean $\delta^{13}C$ value for POC was -19.4±0.8 ‰ (n=8), which was within the typical range of marine phytoplankton (Peterson and Fry, 1987). As shown in Fig. 5, there was a large $\delta^{13}C_{POC}$ decrease near a salinity of 15. Geographically, it was located at the mixing dominated zone in inner Lingdingyang Bay, where intense resuspension of ^{13}C depleted sediments may occur (Guo et al., 2009).

3.4 Reinstatement of the hypoxic station after Typhoon Rammasun

Typhoon Rammasun made landfall at Zhanjiang, located 400 km to the southwest of the PRE, at 20:00 LT (Local Time) on 18 July, and was dissipated by 05:00 LT on 20

July. The typhoon completely de-stratified the water column during its passing. However, the associated heavy precipitation and runoff appeared to re-establish stratification rather quickly, within one day, as suggested by the salinity gradient (18–30) from 0–10 m depth during Leg 2 at 15:20 LT on 20 July (Fig. 6b). In order to capture the evolution of DO between the disruption and reinstatement of stratification, we resumed our cruise and revisited Station A10 (Fig. 6). On 13 July, the bottom water at Station A10 was the hypoxic core, with the lowest observed DO (30 μmol kg^{-1}) and highest DIC (2075 μmol kg^{-1}) concentrations. On 20 July, the results showed that the temperature homogeneous layer in the bottom water (9–13 m) might reflect the remnants of typhoon-induced mixing (Fig. 6a), while the reduction in salinity at <9 m depicted the rapid re-establishment of stratification as a result of enhanced freshwater discharge (Fig. 6b). Bottom water DO increased to 153 μmol kg^{-1} and DIC decreased to 1901 μmol kg^{-1} as a result of the typhoon-induced water column mixing and aeration. In addition, TSM increased sharply from 20.2 before the typhoon to 36.6 mg kg^{-1}, suggesting large volumes of sediment had been resuspensed during its passing. On 27 July, one week after the typhoon, strong thermohaline stratification was re-established in the whole water column. Along with the intensifying stratification, bottom water DO decreased to 99 μmol kg^{-1} indicating continuous DO depletion and the potential for hypoxia formation. Meanwhile, bottom water DIC concentrations increased to 2000 μmol kg^{-1} and dissolved inorganic phosphate (DIP) rose from 0.28 to 0.57 μmol kg^{-1}. Moreover, bottom-water TSM returned to pre-typhoon (13 July) levels.

4 Discussion

4.1 Selection of end-members and model validation

The potential temperature-salinity plot displayed a three end-member mixing scheme over the PRE and adjacent coastal waters (Fig. 7a), consisting of Pearl River plume water, offshore surface seawater and upwelled subsurface water. During the summer, a DIC concentration of ~1917 μmol kg^{-1} was observed at S=33.7, which can be regarded as the offshore surface seawater end-member (Guo and Wong, 2015). Here, by choosing S=34.6 as the offshore subsurface water salinity end-member, we obtained a DIC value of ~2023 μmol kg^{-1}, similar to the value at ~100 m depth adopted by Guo and Wong (2015). For the plume end-member, it was difficult to directly select from the

field data, because biological alteration might lead to altered values within the plume influenced area. Therefore, we first assumed that the plume water observed on the shelf consisted of a mixture of freshwater and offshore surface seawater. Then, we compiled 3 years of surface data from the summer (August 2012, July 2014 and July 2015) to extrapolate the relatively stable freshwater end-member and examine the biological effect on DIC-salinity relationships. By constraining DIC end-members (freshwater and offshore surface seawater), we observed that DIC remained overall conservative when salinity was <10.8 but showed removal when salinity was >10.8 (Han et al., 2012). Thus, we derived plume end-member values (1670±50 μmol kg^{-1}) from the DIC-salinity conservative mixing curve at S=10.8. Furthermore, S=10.8 was observed at the innermost station (A08) during Leg 1, which agreed well with the spatial and temporal scale of the actual water mass mixing in our survey. To confirm our results, we also used a freshwater end-member (S=0), but the output of the model showed little difference from that based on the plume end-member at S=10.8.

The $\delta^{13}C_{DIC}$ value was 0.6±0.2 ‰ in the offshore surface seawater at S=~33.7, where nutrient (NO_3^-+NO_2^- and DIP) concentrations were close to their detection limits and DO was nearly saturated, indicating little biological activity. As DIC remained overall conservative when salinity was < 10.8, the $\delta^{13}C_{DIC}$ value of -11.4±0.2 ‰ at S < 0.4 is representative of the freshwater source. Assuming the plume water is a mixture of freshwater and offshore surface seawater, the initial plume end-member of $\delta^{13}C_{DIC}$ at S=10.8 can be calculated via an isotopic mass balance (-7.0±0.8 ‰). A summary of the end-member values used in this study is listed in Table 1.

We calculated the fractions of the three water masses based on potential temperature and salinity equations, so as to predict conservative DIC (DIC_{con}) and its isotopic composition ($\delta^{13}C_{DICcon}$) solely from conservative mixing. We chose the concentration of Ca^{2+} as a conservative tracer to validate our model prediction, assuming $CaCO_3$ precipitation or dissolution is not significant. This assumption is supported by a strong linear relationship between surface water Ca^{2+} and salinity, and aragonite oversaturation (Ω_{arag}=2.6±0.7) in the subsurface water. Our model derived values were in good accordance with the field-observed values (Fig. 7b), which strongly supported our model prediction.

As shown in Fig. 7c, most of the observed DIC concentrations in the subsurface water were higher than the conservative values, as a result of DIC production via OC

oxidation. This coincided with lighter $\delta^{13}C_{DIC}$ values than conservative, owing to the accumulation of isotopically lighter carbon entering the DIC pool from remineralized organic matter (Fig. 7d). Based on the differences between the observed and conservative values of DIC and $\delta^{13}C_{DIC}$, the carbon isotopic composition of the oxygen-consuming organic matter could be traced precisely (see details in Sect. 4.2).

In the subsurface water, the bulk of ΔDIC values varied from 0 to 132 μmol kg^{-1}, coupled with a range of apparent oxygen utilization (AOU) values from 0 to 179 μmol kg^{-1}. ΔDIC values positively correlated with AOU (Fig. 7e), corresponding to the fact that the additional DIC was supplied by organic matter remineralization via aerobic respiration. The slope of ΔDIC vs. AOU in the subsurface water was 0.71 ±0.03, which agrees well with classic Redfield stoichiometry (i.e., 106/138=0.77), providing further evidence for aerobic respiration as the source of added DIC. As a first order comparison, the water column total oxygen consumption rate of 9.8 μmol L^{-1} d^{-1} could well support the oxygen decline rate observed at Station A10 in the hypoxic zone between 20 July and 27 July (Fig. 6), which was 7.7 μmol L^{-1} d^{-1}. This comparison along with the stoichiometry between ΔDIC and AOU strongly suggests that water column aerobic respiration may be predominate in the formation of the hypoxia in the present case.

4.2 Isotopic composition of the oxygen-consuming OC

The DIC isotopic mass balance is shown in Eq. (6) (Wang et al., 2016):

$$\delta^{13}C_{DICobs}\times DIC_{obs}=\delta^{13}C_{DICcon}\times DIC_{con}+\delta^{13}C_{DICbio}\times DIC_{bio} \quad (6)$$

where the subscripts obs, con and bio refer to the field-observed, conservative and biologically altered values.

Degradation of OC typically produces DIC with minor isotopic fractionation from the OC substrate (Hullar et al., 1996; Breteler et al., 2002). Thus, the isotopic composition of DIC_{bio} (i.e., $\delta^{13}C_{DICbio}$) should be identical to the $\delta^{13}C$ of the OC ($\delta^{13}C_{OCx}$), which consumed oxygen and produced DIC_{bio}. $\delta^{13}C_{OCx}$ was derived from the mass balance equations of both DIC and its stable isotope:

$$\delta^{13}C_{OCx}=\frac{\delta^{13}C_{obs}\times DIC_{obs}-\delta^{13}C_{con}\times DIC_{con}}{DIC_{obs}-DIC_{con}} \quad (7)$$

Equation (7) can be rearranged into Eq. (8):

$$\Delta(\delta^{13}C_{DIC}\times DIC)=\delta^{13}C_{OCx}\times\Delta DIC \quad (8)$$

As shown in Fig. 8, the slope of the linear regression represents $\delta^{13}C_{OCx}$ or $\delta^{13}C_{DICbio}$, which here is equal to -23.2±1.1 ‰. This value reflects the original $\delta^{13}C$ signature of the remineralized organic matter contributing to the observed addition of DIC.

Although studies have shown selective diagenesis of isotopically heavy or light pools of organic matter (Marthur et al., 1992; Lehmann et al., 2002), these effects are small compared to the isotopic differences among different sources of organic matter (Meyers, 1997). It is thus reasonable to assume that the isotopic ratios are conservative and that physical mixing of the end-member sources determine the isotopic composition of organic matter in natural systems (Gearing et al., 1984; Cifuentes et al., 1988; Thornton and McManus, 1994). The relative contributions of marine and terrestrial sources to oxygen-consuming organic matter in our study area could be estimated based on the following equation (Shultz and Calder, 1976; Hu et al., 2006):

$$f(\%)=\frac{\delta^{13}C_{mar}-\delta^{13}C_{OCx}}{\delta^{13}C_{mar}-\delta^{13}C_{terr}}\times 100 \quad (9)$$

Here, for the terrestrial end-member ($\delta^{13}C_{terr}$), we adopted the average $\delta^{13}C$ value of POC sampled near the Humen Outlet (S<4), which represents the predominant source of riverine material entering the estuary (He et al., 2010b). The mean $\delta^{13}C_{POC}$ value, -28.3±0.7 ‰ (n=7), is very similar to the freshwater $\delta^{13}C_{POC}$ value of -28.7 ‰ reported by Yu et al. (2010), which reflected a terrigenous mixture of C3 plant fragments and forest soils. For the marine end-member ($\delta^{13}C_{mar}$), we calculated the mean surface water $\delta^{13}C_{POC}$ value (-19.4±0.8 ‰, n=8) from stations with S>26 where significant phytoplankton blooms were observed, as indicated by DO supersaturation (DO% > 125 %) and relatively high pH values (> 8.3) and POC contents (5.3±2.4 %). This value is similar, although slightly heavier than the marine end-member used by Chen et al. (2008), who measured a $\delta^{13}C$ value of -20.9 ‰ in tow-net phytoplankton samples from outer Lingdingyang Bay, in the same region as this study. Additionally, He et al. (2010a) reported a $\delta^{13}C$ value of -20.8±0.4 ‰ in phytoplankton collected from the northern South China Sea. These values are consistent enough for us to compile and use an average $\delta^{13}C_{mar}$ value of -20.5±0.9 ‰. This value agrees well with the reported stable carbon isotopic signature of marine organic matter in other coastal regions. For example, mean isotopic values of phytoplankton were reported as -20.3±0.6 ‰ in Narragansett Bay (Gearing et al., 1984), -20.3±0.9 ‰ in Auke Bay

and Fritz Cove (Goering et al., 1990), and -20.1±0.8 ‰ in the Gulf of Lions (Harmelin-Vivien et al., 2008).

Our model results suggest that marine organic matter contributed to 65±16 % of the observed oxygen consumption, while terrestrial organic matter accounted for the remaining 35±16 %. It is thus clear that marine organic matter from eutrophication-induced primary production dominated oxygen consumption in the hypoxic zone; however, terrestrial organic matter also contributed significantly to the formation and maintenance of hypoxia in the lower PRE and adjacent coastal waters.

4.3 Comparison with hypoxia in the East China Sea off the Changjiang Estuary

As one of the largest rivers in the world, the Changjiang has been suffering from eutrophication for the past few decades (Zhang et al., 1999; Wang et al., 2014). In summer, sharp density gradients with frequent algal blooms and subsequent organic matter decomposition cause seasonal hypoxia in the bottom water of the ECS off the CJE. Wang et al. (2016) revealed that the remineralization of marine organic matter (OC_{mar}) overwhelmingly (nearly 100 %) contributed to DO consumption in the ECS off the CJE. However, our present study showed that less OC_{mar} contributed to the oxygen depletion (65±16 %) in the hypoxic zone of the lower PRE.

As shown in Fig. 5, there is little difference between $\delta^{13}C_{DIC}$ and $\delta^{13}C_{POC}$ values of the marine end-member. However, the $\delta^{13}C_{DIC}$ and $\delta^{13}C_{POC}$ values of the freshwater end-member showed some dissimilarity, with lighter values in the PRE (-11.4±0.2 ‰, -28.3±0.7 ‰) than in the CJE (-8.8 ‰, -24.4±0.2 ‰). In Fig. 7e, the amplitude of ΔDIC and AOU values suggest a similar intensity of OM biodegradation, and the slope of ΔDIC vs. AOU (0.71±0.03 vs. 0.65±0.04) indicates a predominance of aerobic respiration in the two systems. As seen from Table 2, there is no significant difference between the $\delta^{13}C$ values of surface sediments within the hypoxic zones of the PRE and CJE. However, data in Fig. 7a show generally higher water temperatures in the PRE than in the CJE. For instance, the temperature of surface and subsurface seawater end-members in the PRE is 2-3 ℃ higher than in the CJE. From a spatial point of view, the distance from the river mouth to the hypoxic zone in the CJE is 2-3 times longer than in the PRE, possibly resulting in a longer travel time of OC_{terr}. Therefore, we contend that the difference in the predicted distributions of marine and terrestrial sources of organic matter contributing to oxygen consumption in and off the

PRE and CJE is likely related to differences in the bioavailability of OC_{terr} and OC_{mar}, the microbial community structures and the physical settings between these two hypoxic systems.

Although C3 plants dominate and C4 plants are minor in both the Pearl River and Changjiang drainage basins (Hu et al., 2006; Zhu et al., 2011a), the OC_{terr} delivered from these two watersheds experiences varying degrees of degradation within the estuaries before being transported into the coastal hypoxic zones. In the CJE, approximately 50 % of OC_{terr} becomes remineralized during transport through the estuary, likely due to efficient OM unloading from mineral surfaces (Zhu et al., 2011a) and longer residence times within the estuary, facilitating microbial transformation and degradation. In contrast, the PRE appears to be a somewhat intermediate site with the export of OC_{terr} being closely associated with sedimentary regimes and not characterized by extensive degradative loss (Strong et al., 2012). Thus, the bioavailability of OC_{terr} that reached the hypoxic zone is likely higher in the PRE than in the CJE. Moreover, the increased precipitation and runoff during the typhoon may have mobilized additional fresh anthropogenic OM from surrounding megacities (e.g. Guangzhou, Shenzhen and Zhuhai) deposited in the river channel, which could lead to more labile OC_{terr} in the PRE. Additionally, the difference in bacterial community structure between the two systems may have played a role. Recent studies have demonstrated that the bacterial community in the PRE is characterized by higher relative abundances of Actinobacteria and lower relative abundances of Cytophaga-Flavobacteria-Bacteroides (CFB) than in the CJE (Liu et al., 2012; Zhang et al., 2016). Whether such differences would promote the degradation of OC_{terr} in the PRE relative to the CJE remains unknown. Finally, the temperature of the bottom water in the PRE hypoxic zone (27–29 ℃) was higher than in the CJE hypoxic zone (21.5–24.0 ℃), which may have accelerated the rates of bacterial growth and OM decomposition (Brown et al., 2004).

5 Conclusions

Based on a three end-member mixing model and the mass balance of DIC and its isotopic composition, we demonstrated that the organic matter decomposed via aerobic respiration in the stratified subsurface waters of the lower PRE and adjacent coastal waters was predominantly OC_{mar} (49-81 %, mean 65 %), with a significant portion of

OC_{terr} also decomposed (19-51 %, mean 35 %). The relative distribution of organic matter sources contributing to oxygen drawdown differs in the hypoxic zone off the CJE, where it is caused almost entirely by OC_{mar}. These differences have important implications for better understanding the controls on hypoxia and its mitigation. Nevertheless, with respect to increasing coastal nutrient levels, a significant implication of the present study is that reducing and managing nutrients is critical to control eutrophication and, subsequently, to mitigate hypoxia (Conley et al., 2009; Paerl, 2009; Mercedes et al., 2015; Stefan et al., 2016). Given that OC_{terr} also contributes to the consumption of oxygen in the lower PRE hypoxic zone, it is crucial to characterize the source of this oxygen-consuming terrestrial organic matter, whether from natural soil leaching and/or anthropogenic wastewater discharge, so as to make proper policies for hypoxia remediation.

The processes involved in the partitioning of organic matter sources, their isotopic signals and their subsequent biogeochemical transformations in the PRE hypoxic zone are illustrated in the conceptual diagram in Fig. 9. The river delivers a significant amount of nutrients and terrestrial organic matter to the estuary, stimulating phytoplankton blooms in the surface water at the lower reaches of the estuary where turbidity is relatively low and conditions are favourable for phytoplankton growth (Gaston et al., 2006; Dai et al., 2008b; Guo et al., 2009). The subsequent sinking of this biomass along with terrestrial organic matter below the pycnocline consumes oxygen and adds respired DIC to subsurface waters, resulting in coastal hypoxia. Therefore, we conclude that within the PRE and adjacent coastal areas, the most important biological process with respect to forming and maintaining hypoxic conditions is aerobic respiration.

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Table 1. Summary of end-member values and their uncertainties adopted in the three end-member mixing model.

Water Mass	θ(°C)	Salinity	DIC (μmol kg^{-1})	$\delta^{13}C_{DIC}$ (‰)	Ca^{2+} (μmol kg^{-1})
Plume	30.6±1.0	10.8	1670±50[a]	-7.0±0.8[b]	3670±16[c]
Surface	31.0±1.0	33.7±0.2	1917±3	0.6±0.2	9776±132[c]
Subsurface	21.8±1.0	34.6±0.1	2023±6	0.1±0.1	10053

[a] In order to derive a proper plume end-member value, we took advantage of 3 years of surface dataset from summer cruises (see Sect. 4.1). For DIC, the data is from cruises during August 2012, July 2014 and July 2015.

[b] See details in Sect. 4.1.

[c] The Ca^{2+} values of the plume and surface seawater end-member are derived from a conservative mixing calculation (Ca^{2+} vs. S) based on 3 years of surface data during the summer (August 2012, July 2014 and July 2015).

Table 2. Comparison of $\delta^{13}C$ values in surface sediments within the hypoxic zone[a] between the PRE and CJE.

$\delta^{13}C$ (‰)	Mean±SD	Stations involved	References
		Pearl River Estuary	
-23.4 ~ -22.1	-22.9±0.5	A4, A5, C1-C4, D1	Hu et al. 2006
-23.2 ~ -22.3	-22.7±0.5	28, 29, 30	Zong et al. 2006
-23.6 ~ -21.5	-22.5±1.1	E8-1, E7A, S7-1, S7-2	He et al. 2010a
-[b]	-23.1±0.6	Clustering groups G6 and G7	Yu et al. 2010
Average	-22.8±0.6		
		Changjiang Estuary	
-22.9 ~ -20.9	-21.8±0.6	-[c]	Tan et al. 1991
-22.4 ~ -19.9	-21.2±1.0	32, 37, 38, 42, 48, 49, 54, 56, 64	Kao et al. 2003
-22.7 ~ -20.8	-22.0±0.8	H1-12, H2-10, H2-11, S1-2, S2-4	Xing et al. 2011
-23.5 ~ -20.4	-22.6±1.0	3, 12, 13, 20-25	Yao et al. 2014
Average	-21.9±1.0		

[a]In the PRE, the data is from similar sites to our present study, which is in the northeast (Leg 1) and southwest (Leg 2) of the Wanshan Islands. While in the CJE, the hypoxic zone is located around 30.0 °N–32.0 °N, 122.7 °E–123.2 °E, which is frequently reported in previous studies (Li et al., 2002; Zhu et al., 2011b; Wang et al., 2016).

[b]The authors provide an average value of clustering groups instead of individual data from each site.

[c]In Fig. 7 of Tan et al. (1991), the sampling sites are shown without numbers.

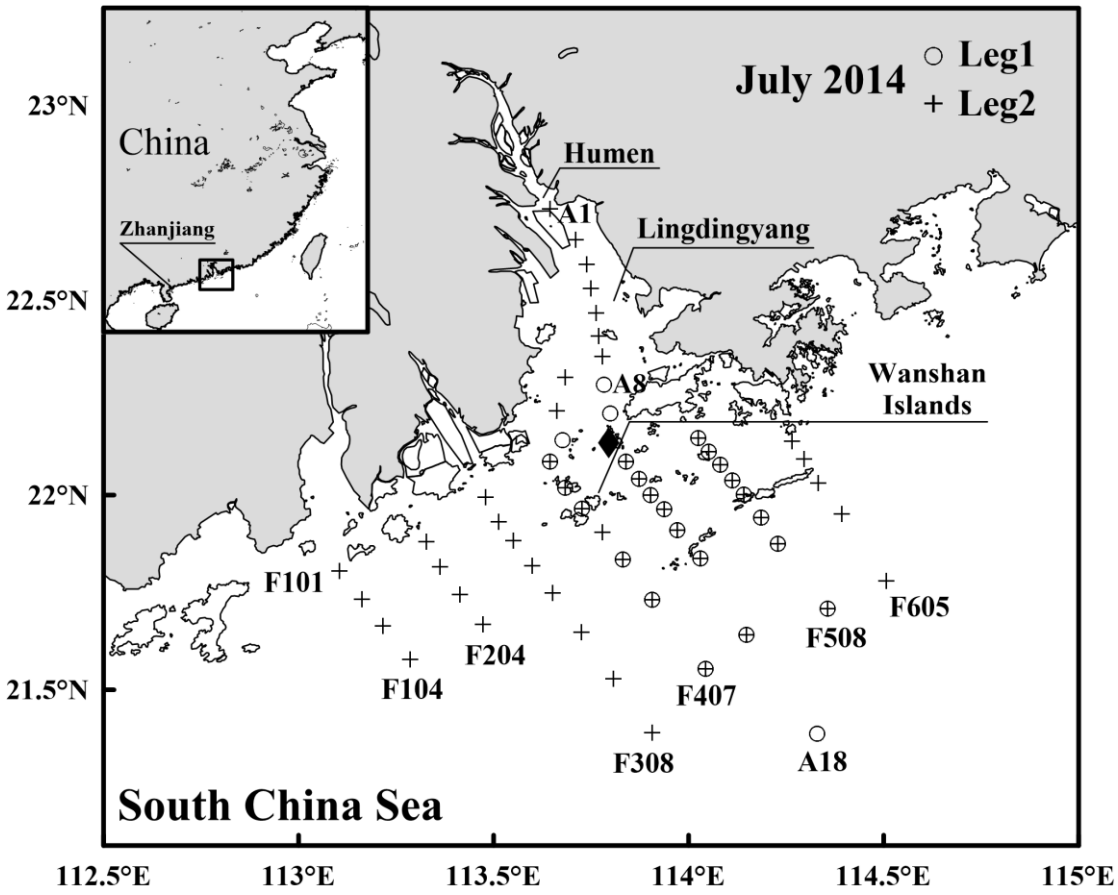

Figure 1. Map of the Pearl River Estuary and adjacent coastal waters. The open circles denote Leg 1 stations visited on 13–16 July 2014, and the crosses represent Leg 2 stations visited on 19–27 July 2014. Note that the filled diamond is the location of Station A10.

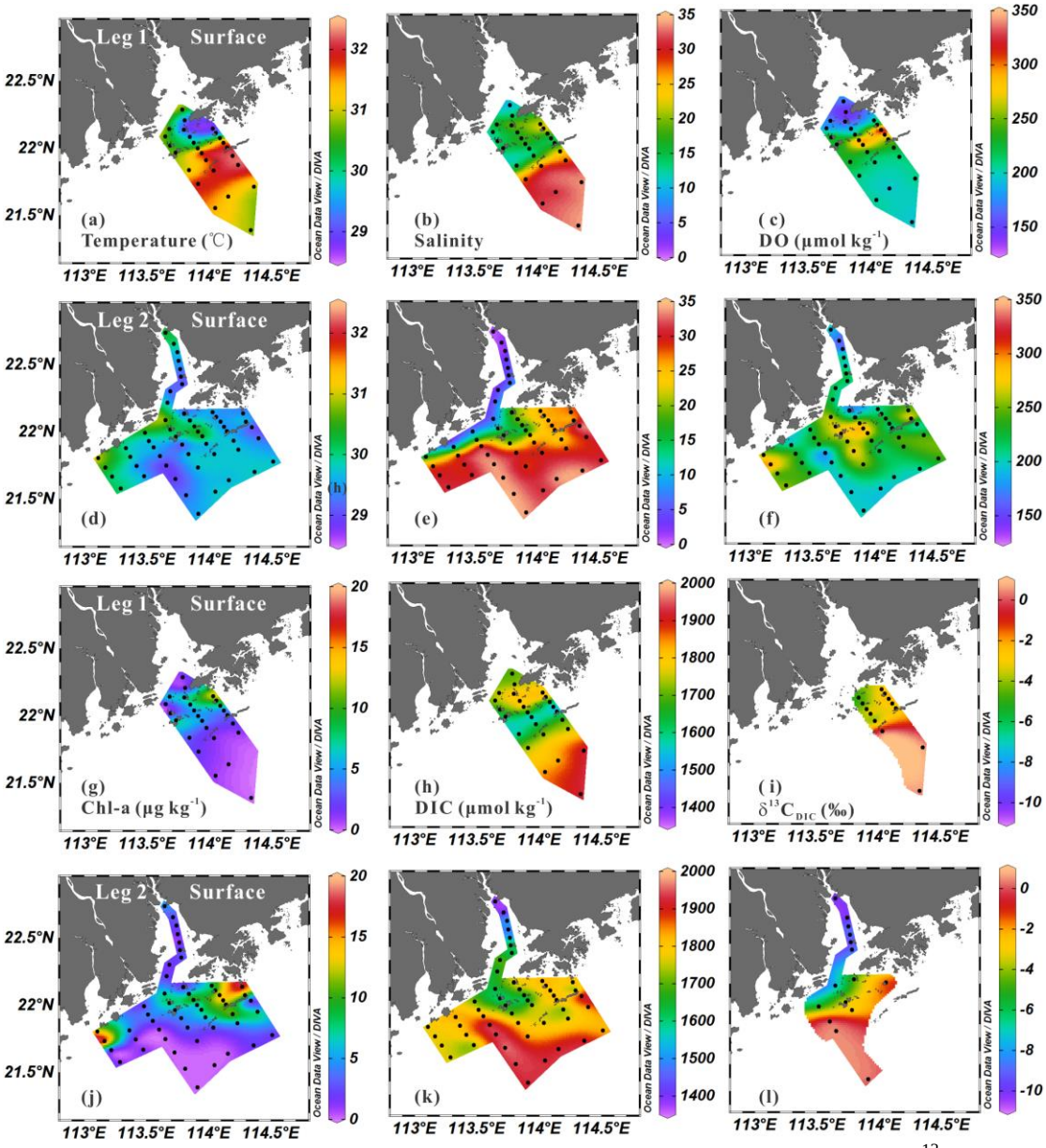

Figure 2. Surface water distribution of temperature, salinity, DO, Chl-a, DIC and $\delta^{13}C_{DIC}$ during Leg 1 (a–c, g–i) and Leg 2 (d–f, j–l).

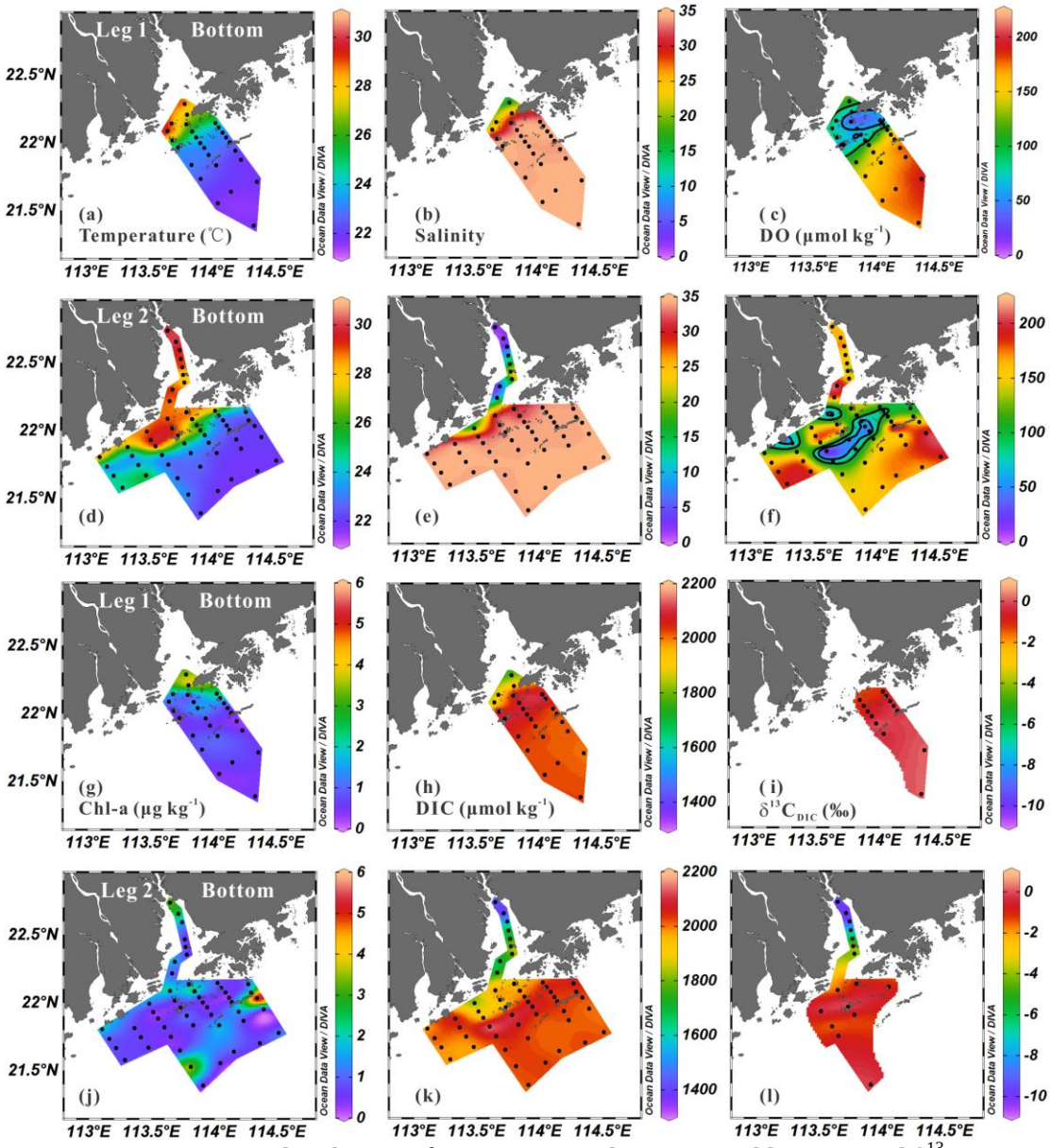

Figure 3. Bottom water distribution of temperature, salinity, DO, Chl-a, DIC and $\delta^{13}C_{DIC}$ during Leg 1 (a–c, g–i) and Leg 2 (d–f, j–l). Note that the black lines in (c) and (f) indicate DO contours of 63 μM and 95 μM.

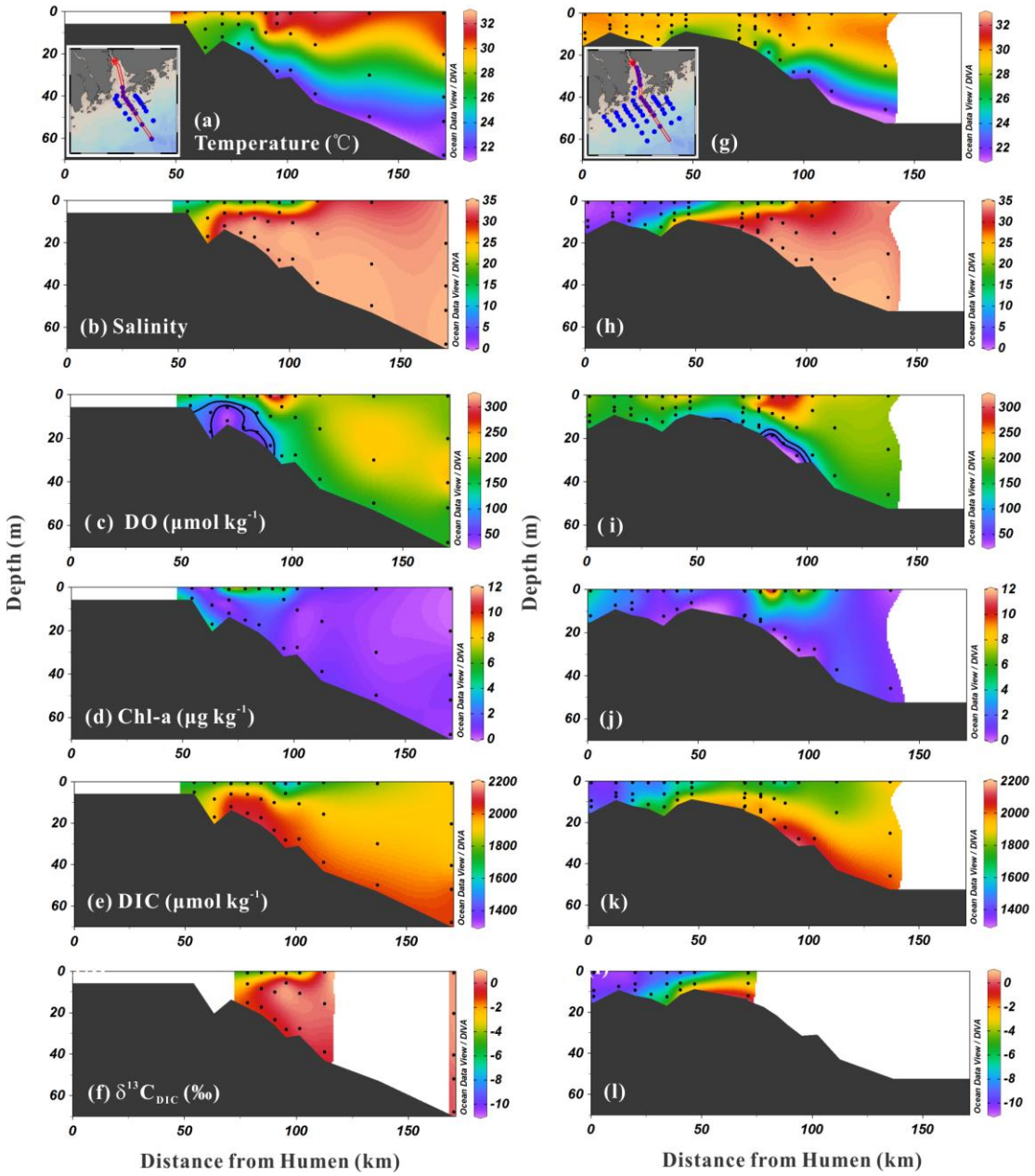

Figure 4. Profiles of temperature, salinity, DO, Chl-a, DIC and $\delta^{13}C_{DIC}$ along Transect A during Leg 1 (a–f) and Leg 2 (g–l). Note that the black lines in (c) and (i) indicate DO contours of 63 μM and 95 μM.

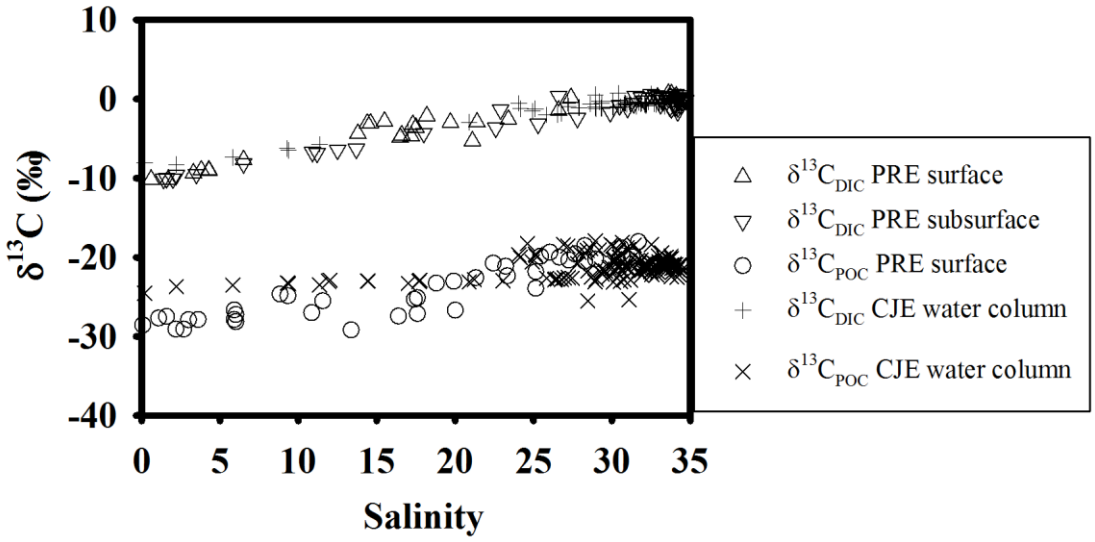

Figure 5. Distribution of $\delta^{13}C_{DIC}$ and $\delta^{13}C_{POC}$ with respect to salinity in the PRE. The up-facing and down-facing triangles denote surface and subsurface $\delta^{13}C_{DIC}$ data, respectively, from July 2014, while the open circles represent $\delta^{13}C_{POC}$ values in surface water from July 2015. Additionally, the plus signs and crosses show the $\delta^{13}C_{DIC}$ and $\delta^{13}C_{POC}$ data, respectively, from the Changjiang Estuary (CJE) in Wang et al. (2016).

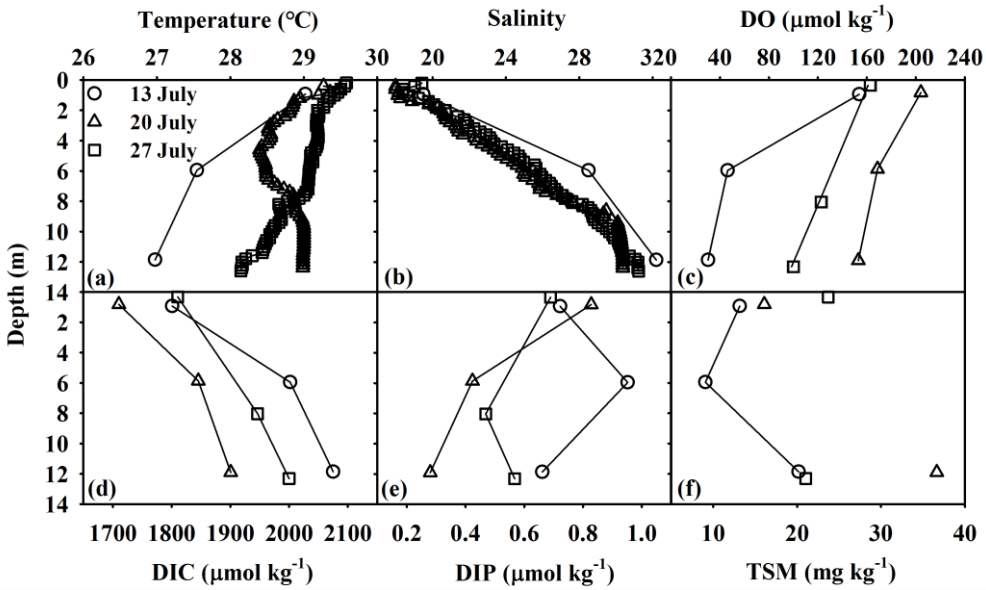

Figure 6. Profiles of (a) temperature, (b) salinity, (c) DO, (d) DIC, (e) DIP, (f) TSM and their evolution during repeated sampling at Station A10.

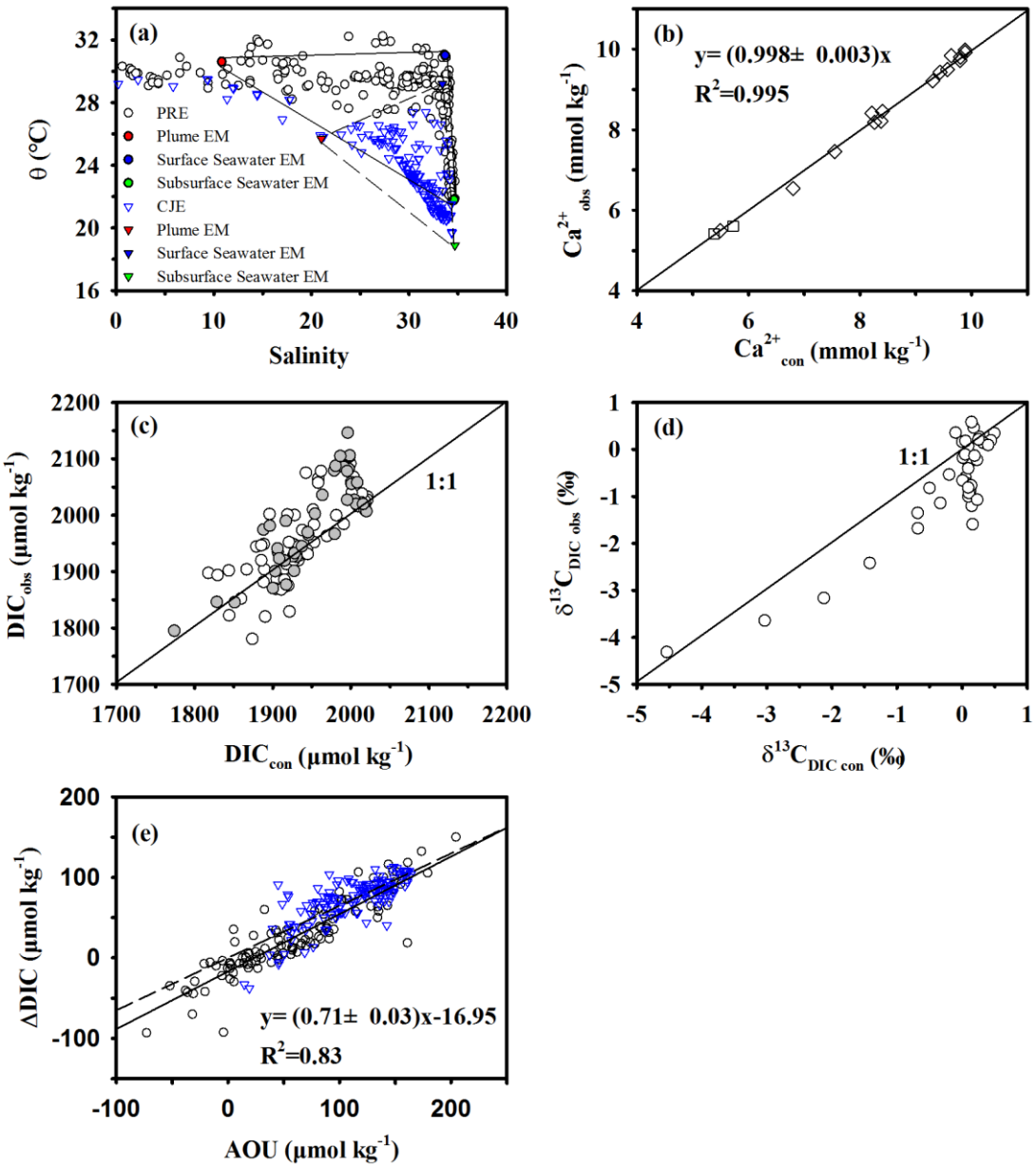

Figure 7. (a) Potential temperature (θ) (℃) vs. salinity in the PRE and adjacent coastal waters (open circles) based on data collected during the July 2014 cruise. The three end-members are shown as different coloured symbols. The blue triangles represent data collected during the August 2011 cruise in the Changjiang Estuary (CJE) (Wang et al., 2016); (b) Correlation between the field-observed Ca^{2+} (Ca^{2+}_{obs}) and conservative Ca^{2+} (Ca^{2+}_{con}). The straight line denotes a linear regression line of both surface (square) and subsurface (diamond) data; (c), (d) Relationship between observed and conservative DIC and $\delta^{13}C_{DIC}$ values. The straight line represents a 1:1 reference line. Note that the grey dots in Fig. 7c identify data also in Fig. 7d; and (e) Correlation of ΔDIC vs. AOU for all subsurface water data. ΔDIC is the difference between the field-observed and conservative DIC concentrations. Also shown is the data from

Wang et al. (2016). The straight and dashed lines indicate linear regressions of data from the PRE and CJE, respectively.

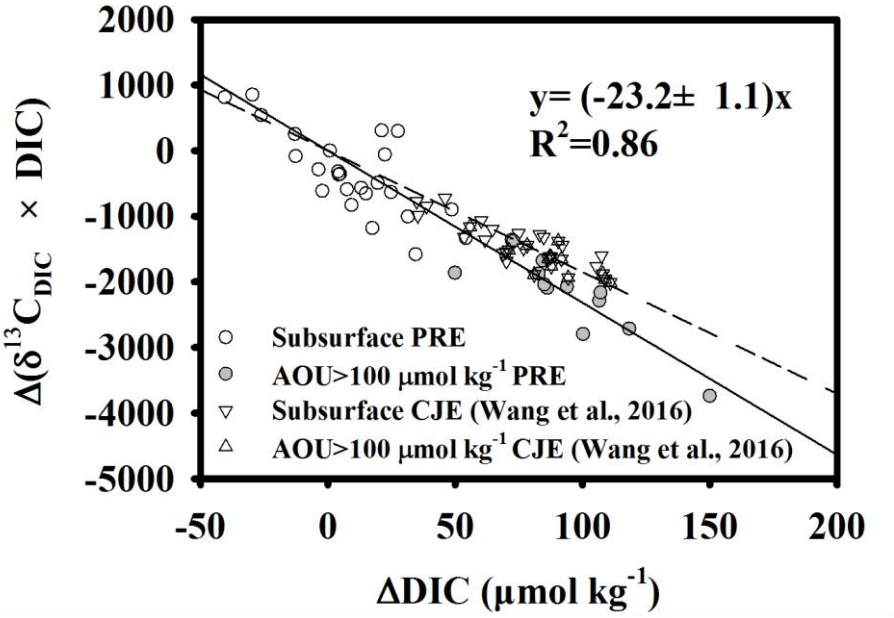

Figure 8. Δ ($\delta^{13}C_{DIC}\times$DIC) vs. ΔDIC in the PRE. Samples were collected from subsurface water (> 5 m). The grey circles represent samples with AOU > 100 μmol kg^{-1}. Δ is the difference between the field-observed and conservative values. Also shown is data from the Changjiang Estuary (CJE) reported by Wang et al. (2016). The straight and dashed lines indicate linear regression lines of data from the PRE and CJE, respectively.

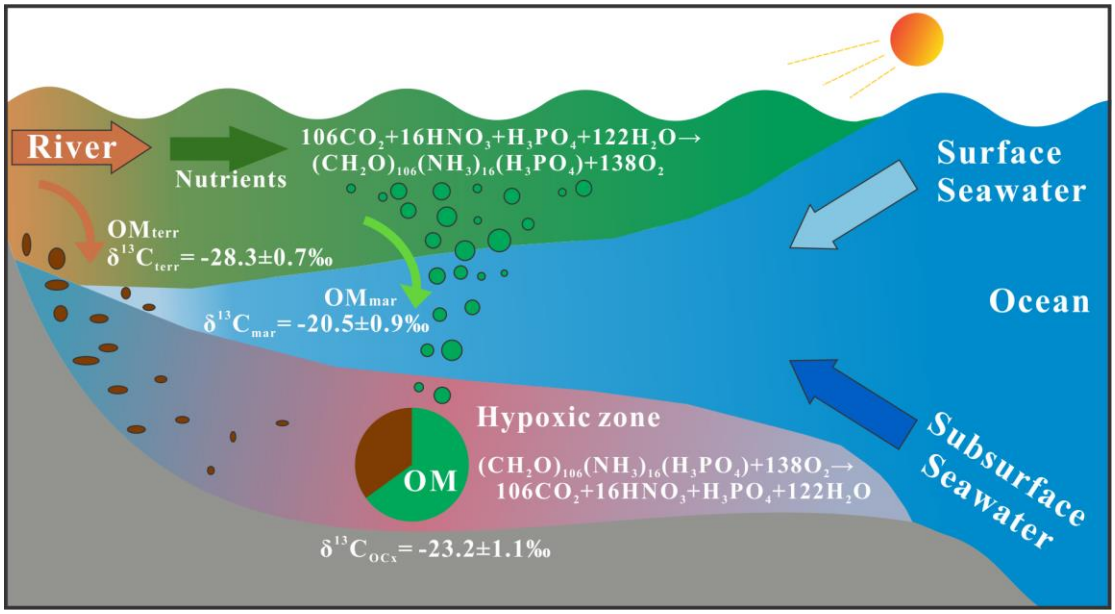

Figure 9. A conceptual diagram illustrating the partitioning of oxygen-consuming organic matter (OC_{mar} vs. OC_{terr}) within the hypoxic zone in the lower PRE and the adjacent coastal area. See Sect. 5 for explanations.