Tracing the origin of the oxygen-consuming organic matter in the hypoxic zone in a large eutrophic estuary: the lower reach of the Pearl River Estuary, China

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Abstract. We assess the relative contributions of different sources of organic matter, 13 marine vs. terrestrial, to oxygen consumption in an emerging hypoxic zone in the lower 14 Pearl River Estuary (PRE), a large eutrophic estuary located in Southern China. Our 15 cruise, conducted in July 2014, consisted of two legs before and after the passing of 16 Typhoon Rammasun, which completely de-stratified the water column. The 17 stratification recovered rapidly, within one day after the typhoon. We observed algal 18 blooms in the upper layer of the water column and hypoxia underneath in bottom water 19 during both legs. Repeat sampling at the initial hypoxic station showed severe oxygen 20 depletion down to 30.3 μ mol kg⁻¹ before the typhoon and a clear drawdown of 21 dissolved oxygen after the typhoon. Based on a three end-member mixing model and 22 the mass balance of dissolved inorganic carbon and its isotopic composition, the δ^{13} C of 23 organic carbon remineralized in the hypoxic zone was -23.2 ± 1.1 %. We estimated that 24 65 ± 16 % of the oxygen-consuming organic matter was derived from marine sources, 25 and the rest (35±16%) was derived from the continent. In contrast to a recently studied 26 hypoxic zone in the East China Sea off the Changjiang Estuary where marine organic 27 matter stimulated by eutrophication dominated oxygen consumption, here terrestrial 28 organic matter significantly contributed to the formation and maintenance of hypoxia. 29 30 How varying amounts of these organic matter sources drive oxygen consumption has important implications for better understanding hypoxia and its mitigation in bottom 31 32 waters.

1 1 Introduction

2 The occurrence of hypoxia has been exacerbated worldwide (Nixon, 1995; Diaz and Rosenberg, 2008; Rabalais et al., 2010; Zhang et al., 2013). In recent decades, more 3 than 400 coastal hypoxic systems have been reported with an exponential growth rate 4 of $5.5 \pm 0.23 \%$ yr⁻¹, demonstrating their persistence and complexity with respect to both 5 science and management (Diaz and Rosenberg, 2008; Vaquer-Sunyer and Duarte, 6 2008). Hypoxia may not only reduce biodiversity and endanger aquatic and benthic 7 habitats, but also alter the redox chemistry in both the water column and the underlying 8 sediments, triggering the release of secondary pollutants (Breitburg, 2002; Rutger et 9 10 al., 2002). Moreover, the management and recovery of these systems is complicated due to the hysteresis of hypoxic conditions, and the varying timescales of biological 11 12 loss (within hours to weeks) and recovery from hypoxia (from months to years) 13 (Steckbauer et al., 2011).

Coastal hypoxia usually occurs in stratified water columns where the downward 14 mixing of oxygen from the surface is impeded (Kemp et al., 2009). Below the 15 pycnocline, aerobic respiration is usually the predominant sink of oxygen. Organic 16 matter, which consumes dissolved oxygen (DO) as it becomes oxidized, is thus the 17 ultimate cause of hypoxia under favourable physical settings (Rabouille et al., 2008; 18 Rabalais et al., 2014; Qian et al., 2016). The organic carbon (OC) that fuels 19 20 respiration-driven reduction of oxygen in these systems could originate from either eutrophication-induced primary production (marine OC; OC_{mar}), or naturally and/or 21 anthropogenically driven delivery from terrestrial environments (terrestrial OC; OC_{terr}) 22 23 (Paerl, 2006; Rabalais et al., 2010).

The question of how much OC in hypoxic zones is supplied from on-site primary 24 25 production versus the quantity derived from terrestrial sources has been an issue of debate (Wang et al., 2016). Much of the phytoplankton-centric hypoxia literature 26 suggests that OC_{mar} dominates oxygen consumption in hypoxic zones, owing to its 27 higher microbial availability than OCterr (Zimmerman and Canuel, 2000; Boesch et al., 28 29 2009; Carstensen et al., 2014). Wang et al. (2016) quantified for the first time the relative contributions of particulate OC_{mar} (POC_{mar}) and particulate OC_{terr} (POC_{terr}) in 30 31 consuming DO in the bottom waters of the East China Sea (ECS) off the Changjiang Estuary (CJE), and found that POC_{mar} dominated DO consumption. However, other 32 studies suggest that POC_{terr} may also play an important role (Swarzenski et al., 2008; 33

Bianchi, 2011a; Bianchi et al., 2011b). It is thus very important to quantify the relative contributions of organic matter (OC_{mar} vs. OC_{terr}) driving the onset and maintenance of hypoxia in coastal systems, since reducing organic matter vs. nutrient inputs requires a different set of management strategies.

The Pearl River Estuary (PRE, 21.2 °N–23.1 °N, 113.0 °E–114.5 °E) is surrounded 5 by several large cities including Hong Kong, Shenzhen and Guangzhou and has 6 received very high loads of nutrients from the drainage basin in the last three decades. 7 As such, eutrophication has increasingly become an issue of concern (Huang et al., 8 9 2003; Ye et al., 2012). Dissolved inorganic nitrogen (DIN) concentrations in the PRE have increased approximately 4-fold from 1986 (19.3 µmol L⁻¹) to 2002 (76.1 µmol 10 L^{-1}) (He and Yuan, 2007). This DIN increase has been attributed to increased inputs of 11 domestic sewage, industrial wastewater, agricultural runoff and aquaculture in the 12 watershed (Huang et al., 2003). 13

Recent observations based on monthly surveys between April 2010 and March 2011 14 and long term monitoring data from 1990 to 2014, have suggested that the lower PRE 15 has emerged as a seasonal hypoxic zone (Qian et al., 2017). This is supported by our 16 current study, as two relatively large hypoxic zones (> 300 km^2) were observed in the 17 lower PRE with $DO < 2 \text{ mg L}^{-1}$. However, the origin of the organic matter driving 18 19 hypoxia in the lower PRE has not previously been examined. Here, we quantified the relative proportions of OC_{mar} and OC_{terr} contributing to DO drawdown in bottom waters 20 of the lower PRE, an economically important coastal region. This study has important 21 biological, societal and managerial implications for the region, particularly relating to 22 23 water quality in the vicinity of Hong Kong in the lower PRE. For example, the government of Hong Kong is examining the efficacy of its costly Harbour Area 24 25 Treatment Scheme project and if additional treatment should be implemented (http://www.gov.hk/en/residents/environment/water/harbourarea.htm). 26

27 2 Materials and Methods

28 **2.1 Sampling and analysis**

Interrupted by Typhoon Rammasun on 17-18 July 2014, our cruise was divided into
two legs (Fig. 1). During Leg 1 on 13–16 July, we sampled Transects F4, F5 and
Stations A08–A18. During Leg 2 on 19–27 July, we sampled Stations A01–A10,
Transects F3 and F4, Stations A11–A17 and Transects F5, F6, F1, and F2, in sequence.

In order to monitor the development of hypoxia before and after the passage of the
 typhoon, we revisited Station A10 three more times (13, 20 and 27 July).

According to the gauge in the upper Pearl River, water discharge peaked in June and July. Typhoon Rammasun increased discharge during 15-18 July, with daily average values of 19480, 26115, 22981 and 17540 m³ s⁻¹, respectively. Nevertheless, the freshwater discharge was 18908 m³ s⁻¹ in leg 1 and 15698 m³ s⁻¹ in leg 2, comparable to the long-term (2000–2011) monthly average.

determined SBE 25 Temperature and salinity were with 8 a Conductivity-Temperature-Depth/Pressure unit (Sea-Bird Co.). Water samples were 9 collected using 4 L Go-Flo bottles (General Oceanics). DIC and DO was measured at 10 all stations with depth profiles. Samples for $\delta^{13}C_{DIC}$ were collected primarily along 11 Transect A as well as at depth in low oxygen layers. 12

The DO concentrations in discrete water samples were measured on board within 8 h using the classic Winkler titration method (Dai et al., 2006). DIC was measured with an infrared detector after acidifying 0.5–0.7 mL of water sample with a precision of 0.1 % for estuarine and sea waters (Cai et al., 2004). Dissolved calcium concentrations (Ca²⁺) were determined using an EGTA titration with a Metrohm 809 TITRANDO potentiometer, which has a precision better than $\pm 5 \ \mu mol \ kg^{-1}$ (Cao et al., 2011).

For $\delta^{13}C_{DIC}$ analysis, an ~20 mL DIC sample was converted into gaseous CO₂ and progressively purified through a vacuum line. The pure CO₂ sample was analyzed with an isotope ratio mass spectrometer (IRMS, Finnigan MAT 252, Bremen, Germany). The analytical precision was better than 0.1 ‰.

Water samples for TSM (total suspended matter), POC and $\delta^{13}C_{POC}$ analysis were 23 concentrated onto preweighed and pre-combusted 0.7 µm Whatman GF/F filters after 24 filtering 0.2–1.0 L of water under a mild vacuum (~ 25 kPa). Filters were washed with 25 26 distilled water and stored at -20 °C. Prior to analysis, all filters were freeze-dried. TSM was determined using the net weight increment on the filter and the filtration volume. 27 Filters were decarbonated with 1.0 mol L⁻¹ HCl and dried at 40 $^{\circ}$ C for 48 h (Kao et al., 28 2012) and analyzed for POC and $\delta^{13}C_{POC}$ on an elemental analyzer coupled with an 29 IRMS (EA-IRMS). The analytical precision for $\delta^{13}C_{POC}$ was better than 0.1 ‰. Chl-a 30 was measured with a Turner fluorometer after extracting filters with 90 % acetone (He 31 et al., 2010b). Calibrations were performed using a Sigma Chl-a standard. 32

1 **2.2 Three end-member mixing model**

We adopted a three end-member mixing model to construct the conservative mixing
scheme among different water masses (Cao et al., 2011; Han et al., 2012):

$$4 \qquad F_{\rm RI} + F_{\rm SW} + F_{\rm SUB} = 1 \tag{1}$$

5
$$\theta_{\rm RI} \times F_{\rm RI} + \theta_{\rm SW} \times F_{\rm SW} + \theta_{\rm SUB} \times F_{\rm SUB} = \theta$$
 (2)

$$6 \qquad S_{\rm RI} \times F_{\rm RI} + S_{\rm SW} \times F_{\rm SW} + S_{\rm SUB} \times F_{\rm SUB} = S \tag{3}$$

where θ and *S* represent potential temperature and salinity; the subscripts RI, SW, and SUB denote the three different water masses (Pearl River plume water, offshore surface seawater and upwelled subsurface water); and F_{RI} , F_{SW} , and F_{SUB} represent the fractions that each end-member contributes to the in situ samples. These fractions were applied to predict concentrations of DIC (DIC_{pre}) and its isotopic composition ($\delta^{13}C_{\text{DICpre}}$) resulting solely from conservative mixing.

13
$$DIC_{\rm RI} \times F_{\rm RI} + DIC_{\rm SW} \times F_{\rm SW} + DIC_{\rm SUB} \times F_{\rm SUB} = DIC_{\rm pre}$$
 (4)

14
$$\frac{\delta^{I3}C_{\text{DICRI}} \times DIC_{\text{RI}} \times F_{\text{RI}} + \delta^{I3}C_{\text{DICSW}} \times DIC_{\text{SW}} \times F_{\text{SW}} + \delta^{I3}C_{\text{DICSUB}} \times DIC_{\text{SUB}} \times F_{\text{SUB}}}{DIC_{\text{pre}}} = \delta^{I3}C_{\text{DICpre}}$$
(5)

15 The difference (Δ) between measured and predicted DIC values represents the 16 magnitude of the biological alteration of DIC (Wang et al., 2016).

17 3 Results

18 **3.1 Horizontal distribution**

Although the average freshwater discharge rate during our sampling period (16369 m³ 19 s^{-1}) was slightly higher than the multi-year (2000–2011) monthly average (15671 m³ 20 s⁻¹), typhoon Rammasun modified the system to some extent as shown from the 21 evolution of chemical species at Station A10 before and after the typhoon (See Sect. 22 23 3.4). The interruption of Leg 1 due to the typhoon (July 17-18) led to a smaller survey area, covering only outside Lingdingyang Bay (traditionally regarded as the PRE), 24 while Leg 2 covered Lingdingyang Bay from the Humen Outlet to the adjacent coastal 25 26 sea.

As depicted in Fig. 2, the sea surface temperature (SST) during Leg 1 (28.9-32.2 °C) was slightly higher than during Leg 2 (28.9-31.0 °C). Sea surface salinity (SSS) measurements showed that plume water was restricted more landward during Leg 2 than Leg 1. However, a steeper gradient to higher SST offshore during Leg 1 was

likely induced by the upwelling of bottom water, featuring relatively high SSS (18.6), 1 high DIC (1788.7 µmol kg⁻¹) and low DO saturation (DO%, 86 %). During Leg 1, the 2 region with the most productivity was found east of the Wanshan Islands, characterized 3 by high concentrations of Chl-a (8.0 μ g kg⁻¹), low concentrations of DIC (1606.8 μ mol 4 kg⁻¹), and DO supersaturation, with the highest DO% greater than 160 % at Station 5 F503. During Leg 2, there were three patches of high productivity, south of 6 Huangmaohai, at the PRE entrance, and off Hong Kong. The central region of high 7 productivity had the highest DO%, greater than 140% at Station A14, and was 8 characterized by relatively high concentrations of Chl-a (7.8 μ g kg⁻¹) and low 9 concentrations of DIC (1737.3 µmol kg⁻¹). 10

As shown in Fig. 3, bottom water hypoxia during Leg 1 was located more centrally in 11 the study area relative to the surface phytoplankton bloom. The center of the hypoxic 12 zone was found at Station A10, characterized by the lowest observed DO 13 concentrations (as low as 30.3 µmol kg⁻¹) and a relatively high concentration of DIC 14 (2074.9 µmol kg⁻¹). During Leg 2, hypoxic conditions were no longer found at Station 15 A10, and instead the largest hypoxic zone was discovered to the southwest of the 16 Wanshan Islands, where the lowest DO values were observed (as low as 7.2 μ mol kg⁻¹ 17 at F304), and once again coincided with relatively high concentrations of DIC (2146.1 18 μ mol kg⁻¹). We were unable to precisely constrain the areas of the regions impacted by 19 bottom water hypoxia due to the limited spatial coverage, but our results suggest it 20 covered an area of $> 280 \text{ km}^2$ during Leg 1 and $> 290 \text{ km}^2$ during Leg 2 according to 21 the definition of hypoxia as $DO < 2 \text{ mg L}^{-1}$ or 63 μ M, or an area of > 900 km² during 22 Leg 1 and $> 800 \text{ km}^2$ during Leg 2 assuming the threshold of the oxygen-deficit zone 23 was $< 3 \text{ mg L}^{-1}$ or 95 μ M (Rabalais et al., 2010; Zhao et al., 2017). 24

25 **3.2 Vertical distribution**

During Leg 1, plume water reached 50 km offshore from the entrance of the PRE, forming a 5–10 m thick surface layer (Fig. 4b). Both the thermocline and halocline contributed to the stability of the water column structure, which favored the formation of bottom water hypoxia. The thickness of the bottom water hypoxic layer was ~ 5 m. The region of highest productivity, however, was not observed in the same location as the hypoxic zone, but further offshore. 1 During Leg 2, although the passing of the typhoon would be expected to absorb large amounts of potential heat and cause extensive mixing of the water column, the 2 enhanced freshwater discharge could rapidly re-stratify the water column and facilitate 3 the re-formation of hypoxia. This time, the primary region of hypoxia was observed 4 directly below the bloom, with a thickness of 3 m (Fig. 4i). Additionally, near the 5 Humen Outlet we observed low DIC (1466.3 µmol kg⁻¹) and moderately low DO (89.1 6 µmol kg⁻¹), which reflected the input of the low DO water mass from upstream as 7 reported previously (Dai et al., 2006; Dai et al., 2008a; He et al., 2014). 8

9 **3.3 Isotopic composition of DIC and POC**

The δ^{13} C values of DIC became progressively heavier from stations dominated by 10 freshwater (~ -11.4 %) to off-shore seawater (~ -0.6 %), with a relatively wide range of 11 values beyond a salinity of 13 (Fig. 5). Owing to a malfunction of the instrument, 12 $\delta^{13}C_{POC}$ data from our cruise were not available. Instead, we reported a valid $\delta^{13}C_{POC}$ 13 dataset from a 2015 summer cruise in approximately the same region. $\delta^{13}C_{POC}$ values 14 showed a similar trend with $\delta^{13}C_{DIC}$, i.e. ¹³C enriched seaward, from ~ -28 ‰ to ~ -20 15 %. In the bloom, where the DO% was above 125 %, the mean δ^{13} C value for POC 16 was -19.4 ± 0.8 % (n=8), which was within the typical range of marine phytoplankton 17 (Peterson and Fry, 1987). As shown in Fig. 5, there was a large $\delta^{13}C_{POC}$ decrease near a 18 salinity of 15. Geographically, it was located at the mixing dominated zone in inner 19 Lingdingyang Bay, where intense resuspension of ¹³C depleted sediments may occur 20 (Guo et al., 2009). 21

3.4 Reinstatement of the hypoxic station after Typhoon Rammasun

Typhoon Rammasun made landfall at Zhanjiang, located 400 km to the southwest of 23 24 the PRE, at 20:00 LT (Local Time) on 18 July, and was dissipated by 05:00 LT on 20 July. The typhoon completely de-stratified the water column during its passing. 25 However, the associated heavy precipitation and runoff appeared to re-establish 26 stratification rather quickly, within one day, as suggested by the salinity gradient (18– 27 28 30) from 0–10 m depth during Leg 2 at 15:20 LT on 20 July (Fig. 6b). In order to capture the evolution of DO between the disruption and reinstatement of stratification, 29 30 we resumed our cruise and revisited Station A10 (Fig. 6). On 13 July, the bottom water at Station A10 was the hypoxic core, with the lowest observed DO (30.3 µmol kg⁻¹) and 31

highest DIC (2074.9 µmol kg⁻¹) concentrations. On 20 July, the results showed that the 1 temperature homogeneous layer in the bottom water (9-13 m) might reflect the 2 remnants of typhoon-induced mixing (Fig. 6a), while the reduction in salinity at <9 m 3 depicted the rapid re-establishment of stratification as a result of enhanced freshwater 4 discharge (Fig. 6b). Bottom water DO increased to 153.1 µmol kg⁻¹ and DIC decreased 5 to 1900.7 μ mol kg⁻¹ as a result of the typhoon-induced water column mixing and 6 aeration. In addition, TSM increased sharply from 20.2 before the typhoon to 36.6 mg 7 kg⁻¹, suggesting large volumes of sediment had been resuspensed during its passing. On 8 27 July, one week after the typhoon, strong thermohaline stratification was 9 re-established in the whole water column. Along with the intensifying stratification, 10 bottom water DO decreased to 98.6 µmol kg⁻¹ indicating continuous DO depletion and 11 the potential for hypoxia formation. Meanwhile, bottom water DIC concentrations 12 increased to 2000.1 µmol kg⁻¹ and dissolved inorganic phosphate (DIP) rose from 0.28 13 to 0.57 µmol kg⁻¹. Moreover, bottom-water TSM returned to pre-typhoon (13 July) 14 levels. 15

16 4 Discussion

17 **4.1 Selection of end-members and model validation**

The potential temperature-salinity plot displayed a three end-member mixing scheme 18 over the PRE and adjacent coastal waters (Fig. 7a), consisting of Pearl River plume 19 water, offshore surface seawater and upwelled subsurface water. During the summer, a 20 DIC concentration of ~1917 μ mol kg⁻¹ was observed at S=33.7, which can be regarded 21 as the offshore surface seawater end-member (Guo and Wong, 2015). Here, by 22 choosing S=34.6 as the offshore subsurface water salinity end-member, we obtained a 23 DIC value of ~2023 μ mol kg⁻¹, similar to the value at ~100 m depth adopted by Guo 24 and Wong (2015). For the plume end-member, it was difficult to directly select from the 25 field data, because biological alteration might lead to altered values within the plume 26 influenced area. Therefore, we first assumed that the plume water observed on the shelf 27 consisted of a mixture of freshwater and offshore surface seawater. Then, we compiled 28 3 years of surface data from the summer (August 2012, July 2014 and July 2015) to 29 extrapolate the relatively stable freshwater end-member and examine the biological 30 effect on DIC-salinity relationships. By constraining DIC end-members (freshwater 31

and offshore surface seawater), we observed that DIC remained overall conservative 1 when salinity was <10.8 but showed removal when salinity was >10.8 (Han et al., 2 2012). Thus, we derived plume end-member values ($1670\pm50 \text{ }\mu\text{mol }kg^{-1}$) from the 3 DIC-salinity conservative mixing curve at S=10.8. Furthermore, S=10.8 was observed 4 at the innermost station (A08) during Leg 1, which agreed well with the spatial and 5 temporal scale of the actual water mass mixing in our survey. To confirm our results, 6 we also used a freshwater end-member (S=0), but the output of the model showed little 7 8 difference from that based on the plume end-member at S=10.8.

The $\delta^{13}C_{DIC}$ value was 0.6±0.2 ‰ in the offshore surface seawater at S=~33.7, where 9 nutrient (NO₃⁻+NO₂⁻ and DIP) concentrations were close to their detection limits and 10 DO was nearly saturated, indicating little biological activity. As DIC remained overall 11 conservative when salinity was < 10.8, the $\delta^{13}C_{DIC}$ value of -11.4±0.2 ‰ at S < 0.4 is 12 representative of the freshwater source. Assuming the plume water is a mixture of 13 freshwater and offshore surface seawater, the initial plume end-member of $\delta^{13}C_{\text{DIC}}$ at 14 S=10.8 can be calculated via an isotopic mass balance (-7.0 \pm 0.8 ‰). A summary of the 15 end-member values used in this study is listed in Table 1. 16

17 We calculated the fractions of the three water masses based on potential temperature and salinity equations, so as to predict DIC (DIC_{pre}) and its isotopic composition 18 $(\delta^{13}C_{DICpre})$ solely from conservative mixing. We chose the concentration of Ca²⁺ as a 19 conservative tracer to validate our model prediction, assuming CaCO₃ precipitation or 20 dissolution is not significant. This assumption is supported by a strong linear 21 relationship between surface water Ca²⁺ and salinity, and aragonite oversaturation 22 ($\Omega_{arag}{=}2.6{\pm}0.7)$ in the subsurface water. Our model derived values were in good 23 accordance with the field-observed values (Fig. 7b), which strongly supported our 24 model prediction. 25

As shown in Fig. 7c, most of the observed DIC concentrations in the subsurface water were higher than the predicted values, as a result of DIC production via OC oxidation. This coincided with lighter $\delta^{13}C_{\text{DIC}}$ values than predicted, owing to the accumulation of isotopically lighter carbon entering the DIC pool from remineralized organic matter (Fig. 7d). Based on the differences between the observed and predicted values of DIC and $\delta^{13}C_{\text{DIC}}$, the carbon isotopic composition of the oxygen-consuming organic matter could be traced precisely (see details in Sect. 4.2). In the subsurface water, the bulk of Δ DIC values varied from 0 to 132.3 µmol kg⁻¹, coupled with a range of apparent oxygen utilization (AOU) values from 0 to 179.1 µmol kg⁻¹. Δ DIC values positively correlated with AOU (Fig. 7e), corresponding to the fact that the additional DIC was supplied by organic matter remineralization via aerobic respiration. The slope of Δ DIC vs. AOU in the subsurface water was 0.71±0.03, which agrees well with classic Redfield stoichiometry (i.e., 106/138=0.77), providing further evidence for aerobic respiration as the source of added DIC.

8 4.2 Isotopic composition of the oxygen-consuming OC

9 The DIC isotopic mass balance is shown in Eq. (6) (Wang et al., 2016):

10
$$\delta^{13}C_{\text{DICobs}} \times DIC_{\text{obs}} = \delta^{13}C_{\text{DICpre}} \times DIC_{\text{pre}} + \delta^{13}C_{\text{DICbio}} \times DIC_{\text{bio}}$$
 (6)

where the subscripts obs, pre and bio refer to the field-observed, model-predicted and
biologically altered values.

13 Degradation of OC typically produces DIC with minor isotopic fractionation from 14 the OC substrate (Hullar et al., 1996; Breteler et al., 2002). Thus, the isotopic 15 composition of DIC_{bio} (i.e., $\delta^{13}C_{DICbio}$) should be identical to the $\delta^{13}C$ of the OC 16 ($\delta^{13}C_{OCx}$), which consumed oxygen and produced DIC_{bio}. $\delta^{13}C_{OCx}$ was derived from the 17 mass balance equations of both DIC and its stable isotope:

$$18 \qquad \delta^{13} C_{OCx} = \frac{\delta^{13} C_{obs} \times DIC_{obs} - \delta^{13} C_{pre} \times DIC_{pre}}{DIC_{obs} - DIC_{pre}}$$
(7)

20
$$\Delta(\delta^{13}C_{DIC} \times DIC) = \delta^{13}C_{OCx} \times \Delta DIC$$
 (8)

As shown in Fig. 8, the slope of the linear regression represents $\delta^{13}C_{OCx}$ or $\delta^{13}C_{DICbio}$, which here is equal to -23.2±1.1 ‰. This value reflects the original $\delta^{13}C$ signature of the remineralized organic matter contributing to the observed addition of DIC.

Although studies have shown selective diagenesis of isotopically heavy or light pools of organic matter (Marthur et al., 1992; Lehmann et al., 2002), these effects are small compared to the isotopic differences among different sources of organic matter (Meyers, 1997). It is thus reasonable to assume that the isotopic ratios are conservative and that physical mixing of the end-member sources determine the isotopic composition of organic matter in natural systems (Gearing et al., 1984; Cifuentes et al., 1988; Thornton and McManus, 1994). The relative contributions of marine and terrestrial sources to oxygen-consuming organic matter in our study area could be
estimated based on the following equation (Shultz and Calder, 1976; Hu et al., 2006):

3
$$f(\%) = \frac{\delta^{13} C_{mar} - \delta^{13} C_{OCx}}{\delta^{13} C_{mar} - \delta^{13} C_{terr}} \times 100$$
 (9)

Here, for the terrestrial end-member ($\delta^{13}C_{terr}$), we adopted the average $\delta^{13}C$ value of 4 POC sampled near the Humen Outlet (S<4), which represents the predominant source 5 of riverine material entering the estuary (He et al., 2010b). The mean $\delta^{13}C_{POC}$ value, 6 -28.3±0.7 ‰ (n=7), is very similar to the freshwater $\delta^{13}C_{POC}$ value of -28.7 ‰ 7 reported by Yu et al. (2010), which reflected a terrigenous mixture of C3 plant 8 fragments and forest soils. For the marine end-member ($\delta^{13}C_{mar}$), we calculated the 9 mean surface water $\delta^{13}C_{POC}$ value (-19.4±0.8 ‰, n=8) from stations with S>26 where 10 significant phytoplankton blooms were observed, as indicated by DO supersaturation 11 (DO% > 125%) and relatively high pH values (> 8.3) and POC contents $(5.3\pm2.4\%)$. 12 This value is similar, although slightly heavier than the marine end-member used by 13 Chen et al. (2008), who measured a δ^{13} C value of -20.9 ‰ in tow-net phytoplankton 14 samples from outer Lingdingyang high Bay, in the same region as this study. 15 Additionally, He et al. (2010a) reported a δ^{13} C value of -20.8±0.4 ‰ in phytoplankton 16 collected from the northern South China Sea. These values are consistent enough for 17 us to compile and use an average $\delta^{13}C_{mar}$ value of -20.5±0.9 %. This value agrees 18 well with the reported stable carbon isotopic signature of marine organic matter in 19 other coastal regions. For example, mean isotopic values of phytoplankton were 20 reported as -20.3±0.6 ‰ in Narragansett Bay (Gearing et al., 1984), -20.3±0.9 ‰ in 21 Auke Bay and Fritz Cove (Goering et al., 1990), and -20.1±0.8 ‰ in the Gulf of Lions 22 (Harmelin-Vivien et al., 2008). 23

Our model results suggest that marine organic matter contributed to 65 ± 16 % of the observed oxygen consumption, while terrestrial organic matter accounted for the remaining 35 ± 16 %. It is thus clear that marine organic matter from eutrophication-induced primary production dominated oxygen consumption in the hypoxic zone; however, terrestrial organic matter also contributed significantly to the formation and maintenance of hypoxia in the lower PRE and adjacent coastal waters.

4.3 Comparison with hypoxia in the East China Sea off the Changjiang Estuary

As one of the largest rivers in the world, the Changjiang has been suffering from 2 eutrophication for the past few decades (Zhang et al., 1999; Wang et al., 2014). In 3 4 summer, sharp density gradients with frequent algal blooms and subsequent organic matter decomposition cause seasonal hypoxia in the bottom water of the ECS off the 5 CJE. Wang et al. (2016) revealed that the remineralization of marine organic matter 6 (OC_{mar}) overwhelmingly (nearly 100 %) contributed to DO consumption in the ECS off 7 8 the CJE. However, our present study showed that less OC_{mar} contributed to the oxygen depletion $(65\pm16\%)$ in the hypoxic zone of the lower PRE. 9

As shown in Fig. 5, there is little difference between $\delta^{13}C_{DIC}$ and $\delta^{13}C_{POC}$ values of 10 the marine end-member. However, the $\delta^{13}C_{DIC}$ and $\delta^{13}C_{POC}$ values of the freshwater 11 end-member showed some dissimilarity, with lighter values in the PRE (-11.4±0.2 ‰, 12 -28.3±0.7 ‰) than in the CJE (-8.8 ‰, -24.4±0.2 ‰). In Fig. 7e, the amplitude of 13 Δ DIC and AOU values suggest a similar intensity of OM biodegradation, and the 14 slope of ΔDIC vs. AOU (0.71 \pm 0.03 vs. 0.65 \pm 0.04) indicates a predominance of 15 aerobic respiration in the two systems. As seen from Table 2, there is no significant 16 difference between the δ^{13} C values of surface sediments within the hypoxic zones of 17 the PRE and CJE. However, data in Fig. 7a show generally higher water temperatures 18 in the PRE than in the CJE. For instance, the temperature of surface and subsurface 19 seawater end-members in the PRE is 2-3 °C higher than in the CJE. From a spatial 20 point of view, the distance from the river mouth to the hypoxic zone in the CJE is 2-3 21 times longer than in the PRE, possibly resulting in a longer travel time of OC_{terr}. 22 Therefore, we contend that the difference in the predicted distributions of marine and 23 terrestrial sources of organic matter contributing to oxygen-consumption in and off 24 the PRE and CJE is likely related to differences in the bioavailability of OCterr and 25 OC_{mar}, the microbial community structures and the physical settings between these two 26 27 hypoxic systems.

Although C3 plants dominate and C4 plants are minor in both the Pearl River and Changjiang drainage basins (Hu et al., 2006; Zhu et al., 2011a), the OC_{terr} delivered from these two watersheds experiences varying degrees of degradation within the estuaries before being transported into the coastal hypoxic zones. In the CJE, approximately 50 % of OC_{terr} becomes remineralized during transport through the estuary, likely due to efficient OM unloading from mineral surfaces (Zhu et al., 2011a)

and longer residence times within the estuary, facilitating microbial transformation and 1 degradation. In contrast, the PRE appears to be a somewhat intermediate site with the 2 export of OC_{terr} being closely associated with sedimentary regimes and not 3 characterized by extensive degradative loss (Strong et al., 2012). Thus, the 4 bioavailability of OC_{terr} that reached the hypoxic zone is likely higher in the PRE than 5 in the CJE. Moreover, the increased precipitation and runoff during the typhoon may 6 have mobilized additional fresh anthropogenic OM from surrounding megacities (e.g. 7 Guangzhou, Shenzhen and Zhuhai) deposited in the river channel, which could lead to 8 9 more labile OC_{terr} in the PRE. Additionally, the difference in bacterial community structure between the two systems may have played a role. Recent studies have 10 demonstrated that the bacterial community in the PRE is characterized by higher 11 relative abundances of Actinobacteria and lower relative abundances of 12 Cytophaga-Flavobacteria-Bacteroides (CFB) than in the CJE (Liu et al., 2012; Zhang et 13 al., 2016). Whether such differences would promote the degradation of OC_{terr} in the 14 15 PRE relative to the CJE remains unknown. Finally, the temperature of the bottom water in the PRE hypoxic zone (27–29 $^{\circ}$ C) was higher than in the CJE hypoxic zone (21.5– 16 24.0 $^{\circ}$ C), which may have accelerated the rates of bacterial growth and OM 17 18 decomposition (Brown et al., 2004).

19 5 Conclusions

20 Based on a three end-member mixing model and the mass balance of DIC and its isotopic composition, we demonstrated that the organic matter decomposed via aerobic 21 22 respiration in the stratified subsurface waters of the lower PRE and adjacent coastal waters was predominantly OC_{mar} (49-81 %, mean 65 %), with a significant portion of 23 OC_{terr} also decomposed (19-51 %, mean 35 %). The relative distribution of organic 24 matter sources contributing to oxygen drawdown differs in the hypoxic zone off the 25 CJE, where it is caused almost entirely by OC_{mar}. These differences have important 26 implications for better understanding the controls on hypoxia and its mitigation. 27 Nevertheless, with respect to increasing coastal nutrient levels, a significant implication 28 of the present study is that reducing and managing nutrients is critical to control 29 30 eutrophication and, subsequently, to mitigate hypoxia (Conley et al., 2009; Paerl, 2009; Mercedes et al., 2015; Stefan et al., 2016). Given that OC_{terr} also contributes to the 31 32 consumption of oxygen in the lower PRE hypoxic zone, it is crucial to characterize the

source of this oxygen-consuming terrestrial organic matter, whether from natural soil
 leaching and/or anthropogenic wastewater discharge, so as to make proper policies for
 hypoxia remediation.

The processes involved in the partitioning of organic matter sources, their isotopic 4 signals and their subsequent biogeochemical transformations in the PRE hypoxic zone 5 are illustrated in the conceptual diagram in Fig. 9. The river delivers a significant 6 amount of nutrients and terrestrial organic matter to the estuary, stimulating 7 phytoplankton blooms in the surface water at the lower reaches of the estuary where 8 9 turbidity is relatively low and conditions are favourable for phytoplankton growth (Gaston et al., 2006; Dai et al., 2008b; Guo et al., 2009). The subsequent sinking of this 10 biomass along with terrestrial organic matter below the pycnocline consumes oxygen 11 and adds respired DIC to subsurface waters, resulting in coastal hypoxia. Therefore, we 12 conclude that within the PRE and adjacent coastal areas, the most important biological 13 process with respect to forming and maintaining hypoxic conditions is aerobic 14 15 respiration.

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1 References

- 2 Bianchi, T. S.: The role of terrestrially derived organic carbon in the coastal ocean: A
- 3 changing paradigm and the priming effect, Proc. Natl. Acad. Sci. U.S.A., 108,
- 4 19473-19481, doi:10.1073/pnas.1017982108, 2011a.
- Bianchi, T. S., Wysocki, L. A., Schreiner, K. M., Filley, T. R., Corbett, D. R., and
 Kolker, A. S.: Sources of terrestrial organic carbon in the Mississippi plume region:
- vidence for the importance of coastal marsh inputs, Aquat. Geochem., 17, 431-456,
- 8 doi:10.1007/s10498-010-9110-3, 2011b.
- 9 Boesch, D. F., Boynton, W. R., Crowder, L. B., Diaz, R. J., Howarth, R. W., Mee, L. D.,

10 Nixon, S. W., Rabalais, N. N., Rosenberg, R., Sanders, J. G., Scavia, D., and Turner,

11 R. E.: Nutrient Enrichment Drives Gulf of Mexico Hypoxia, Eos, Trans. Amer.

12 Geophys. Union, 90, 117-118, doi:10.1029/2009EO140001, 2009.

- 13 Breitburg, D.: Effects of hypoxia, and the balance between hypoxia and enrichment, on
- 14 coastal fishes and fisheries, Estuaries, 25, 767-781, doi:10.1007/BF02804904, 2002.
- 15 Breteler, W. C. K., Grice, K., Schouten, S., Kloosterhuis, H. T., and Damst é, J. S. S.:
- 16 Stable carbon isotope fractionation in the marine copepod Temora longicornis:
- 17 unexpectedly low δ^{13} C value of faecal pellets, Mar. Ecol. Prog. Ser., 240, 195-204, 18 doi:10.3354/meps240195, 2002.
- Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M., and West, G. B.: Toward a
- 20 metabolic theory of ecology, Ecology, 85, 1771-1789, doi:10.1890/03-9000, 2004.
- 21 Cai, W.-J., Dai, M., Wang, Y., Zhai, W., Huang, T., Chen, S., Zhang, F., Chen, Z., and
- 22 Wang, Z.: The biogeochemistry of inorganic carbon and nutrients in the Pearl River
- estuary and the adjacent Northern South China Sea, Cont. Shelf Res., 24, 1301-1319,
 doi:10.1016/j.csr.2004.04.005, 2004.
- Cao, Z., Dai, M., Zheng, N., Wang, D., Li, Q., Zhai, W., Meng, F., and Gan, J.:
 Dynamics of the carbonate system in a large continental shelf system under the
 influence of both a river plume and coastal upwelling, J. Geophys. Res. Biogeosci.,
- 28 116, G02010, doi:10.1029/2010JG001596, 2011.
- 29 Carstensen, J., Andersen, J. H., Gustafsson, B. G., and Conley, D. J.: Deoxygenation of
- the Baltic Sea during the last century, Proc. Natl. Acad. Sci. U.S.A., 111, 5628-5633,
- doi:10.1073/pnas.1323156111, 2014.

- Cifuentes, L., Sharp, J., and Fogel, M. L.: Stable carbon and nitrogen isotope
 biogeochemistry in the Delaware estuary, Limnol. Oceanogr., 33, 1102-1115,
 doi:10.4319/lo.1988.33.5.1102, 1988.
- Conley, D. J., Paerl, H. W., Howarth, R. W., Boesch, D. F., Seitzinger, S. P., Karl, E.,
 Karl, E., Lancelot, C., Gene, E., and Gene, E.: Controlling eutrophication: nitrogen
 and phosphorus, Science, 123, 1014-1015, doi:10.1126/science.1167755, 2009.
- 7 Dai, M., Guo, X., Zhai, W., Yuan, L., Wang, B., Wang, L., Cai, P., Tang, T., and Cai,
- 8 W.-J.: Oxygen depletion in the upper reach of the Pearl River estuary during a winter
- 9 drought, Mar. Chem., 102, 159-169, doi:10.1016/j.marchem.2005.09.020, 2006.
- 10 Dai, M., Wang, L., Guo, X., Zhai, W., Li, Q., He, B., and Kao, S.-J.: Nitrification and
- 11 inorganic nitrogen distribution in a large perturbed river/estuarine system: the Pearl
- 12 River Estuary, China, Biogeosciences, 5, 1227-1244, doi:10.5194/bg-5-1227-2008,
- 13 2008a.
- 14 Dai, M., Zhai, W., Cai, W.-J., Callahan, J., Huang, B., Shang, S., Huang, T., Li, X., Lu,
- Z., Chen, W., and Chen, Z.: Effects of an estuarine plume-associated bloom on the
 carbonate system in the lower reaches of the Pearl River estuary and the coastal zone
 of the northern South China Sea, Cont. Shelf Res., 28, 1416-1423, doi:
- 18 10.1016/j.csr.2007.04.018, 2008b.
- Diaz, R. J. and Rosenberg, R.: Spreading dead zones and consequences for marine
 ecosystems, Science, 321, 926-929, doi:10.1126/science.1156401, 2008.
- 21 Gaston, T. F., Schlacher, T. A., and Connolly, R. M.: Flood discharges of a small river
- into open coastal waters: Plume traits and material fate, Estuar. Coast. Shelf Sci., 69,
 4-9, doi:10.1016/j.ecss.2006.03.015, 2006.
- Gearing, J. N., Gearing, P. J., Rudnick, D. T., Requejo, A. G., and Hutchins, M. J.:
- 25 Isotopic variability of organic carbon in a phytoplankton-based, temperate estuary,
- Geochim. Cosmochim. Acta, 48, 1089-1098, doi:10.1016/0016-7037(84)90199-6,
 1984.
- Goering, J., Alexander, V., and Haubenstock, N.: Seasonal variability of stable carbon
 and nitrogen isotope ratios of organisms in a North Pacific Bay, Estuar. Coast. Shelf
 Sci., 30, 239-260, doi:10.1016/0272-7714(90)90050-2, 1990.
- Guo, X., Dai, M., Zhai, W., Cai, W.-J., and Chen, B.: CO₂ flux and seasonal variability
- in a large subtropical estuarine system, the Pearl River Estuary, China, J. Geophys.
- 33 Res. Biogeosci., 114, G03013, doi:10.1029/2008JG000905, 2009.

Guo, X. and Wong, G. T.: Carbonate chemistry in the northern South China Sea
 shelf-sea in June 2010, Deep-Sea Res. II, 117, 119-130,
 doi:10.1016/j.dsr2.2015.02.024, 2015.

Han, A., Dai, M., Kao, S.-J., Gan, J., Li, Q., Wang, L., Zhai, W., and Wang, L.: Nutrient
dynamics and biological consumption in a large continental shelf system under the
influence of both a river plume and coastal upwelling, Limnol. Oceanogr., 57,
486-502, doi:10.4319/lo.2012.57.2.0486, 2012.

- Harmelin-Vivien, M., Loizeau, V., Mellon, C., Beker, B., Arlhac, D., Bodiguel, X.,
 Ferraton, F., Hermand, R., Philippon, X., and Salen-Picard, C.: Comparison of C and
 N stable isotope ratios between surface particulate organic matter and
 microphytoplankton in the Gulf of Lions (NW Mediterranean), Cont. Shelf Res., 28,
 1911-1919, 2008.
- 13 He, B., Dai, M., Huang, W., Liu, Q., Chen, H., and Xu, L.: Sources and accumulation of

organic carbon in the Pearl River Estuary surface sediment as indicated by elemental,
 stable carbon isotopic, and carbohydrate compositions, Biogeosciences, 7,

16 3343-3362, doi:10.5194/bg-7-3343-2010, 2010a.

He, B., Dai, M., Zhai, W., Wang, L., Wang, K., Chen, J., Lin, J., Han, A., and Xu, Y.:
Distribution, degradation and dynamics of dissolved organic carbon and its major
compound classes in the Pearl River estuary, China, Mar. Chem., 119, 52-64,
doi:10.1016/j.marchem.2009.12.006, 2010b.

- He, B., Dai, M., Zhai, W., Guo, X., and Wang, L.: Hypoxia in the upper reaches of the
 Pearl River Estuary and its maintenance mechanisms: A synthesis based on multiple
 year observations during 2000–2008, Mar. Chem., 167, 13-24,
 doi:10.1016/j.marchem.2014.07.003, 2014.
- He, G.-f. and Yuan, G.-m.: Assessment of the water quality by fuzzy mathematics for
 last 20 years in Zhujiang Estuary, Mar. Environ. Sci., 26, 53-57, 2007.
- Hu, J., Peng, P. a., Jia, G., Mai, B., and Zhang, G.: Distribution and sources of organic
- carbon, nitrogen and their isotopes in sediments of the subtropical Pearl River estuary
 and adjacent shelf, Southern China, Mar. Chem., 98, 274-285,
- 30
 doi:10.1016/j.marchem.2005.03.008, 2006.
- Huang, X., Huang, L., and Yue, W.: The characteristics of nutrients and eutrophication
 in the Pearl River estuary, South China, Mar. Pollut. Bull., 47, 30-36,
 doi:10.1016/S0025-326X(02)00474-5, 2003.

- 1 Hullar, M., Fry, B., Peterson, B., and Wright, R.: Microbial utilization of estuarine
- dissolved organic carbon: a stable isotope tracer approach tested by mass balance,
 Appl. Environ. Microbiol., 62, 2489-2493, 1996.

Kao, S.-J., Terence Yang, J.-Y., Liu, K.-K., Dai, M., Chou, W.-C., Lin, H.-L., and Ren,
H.: Isotope constraints on particulate nitrogen source and dynamics in the upper water

- 6 column of the oligotrophic South China Sea, Global Biogeochem. Cycles, 26,
- 7 GB2033, doi:10.1029/2011GB004091, 2012.
- 8 Kemp, W., Testa, J., Conley, D., Gilbert, D., and Hagy, J.: Temporal responses of
- 9 coastal hypoxia to nutrient loading and physical controls, Biogeosciences, 6,
- 10 2985-3008, doi:10.5194/bg-6-2985-2009, 2009.
- Lehmann, M. F., Bernasconi, S. M., Barbieri, A., and McKenzie, J. A.: Preservation of
 organic matter and alteration of its carbon and nitrogen isotope composition during
 simulated and in situ early sedimentary diagenesis, Geochim. Cosmochim. Acta, 66,
 3573-3584, doi:10.1016/S0016-7037(02)00968-7, 2002.
- Li, D., Zhang, J., Huang, D., Wu, Y., and Liang, J.: Oxygen depletion off the
 Changjiang (Yangtze River) estuary, Sci. China Ser. D-Earth Sci., 45, 1137-1146,
 doi:10.1360/02yd9110, 2002.
- Liu, M., Xiao, T., Wu, Y., Zhou, F., Huang, H., Bao, S., and Zhang, W.: Temporal
 distribution of bacterial community structure in the Changjiang Estuary hypoxia area
 and the adjacent East China Sea, Environ. Res. Lett., 7, 025001,
 doi:10.1088/1748-9326/7/2/025001, 2012.
- Marthur, J. M., Tyson, R. V., Thomson, J., and Mattey, D.: Early diagenesis of marine
 organic matter: Alteration of the carbon isotopic composition, Mar. Geol., 105,
 51-61, doi:10.1016/0025-3227(92)90181-G, 1992.
- 25 Mercedes, M. C. B., Luiz Antonio, M., Tibisay, P., Rafael, R., Jean Pierre, H. B. O.,
- 26 Felipe Siqueira, P., Silvia Rafaela Machado, L., and Sorena, M.: Nitrogen
- 27 management challenges in major watersheds of South America, Environ. Res. Lett.,
- 28 10, 065007, doi:10.1088/1748-9326/10/6/065007, 2015.
- 29 Meyers, P. A.: Organic geochemical proxies of paleoceanographic, paleolimnologic,
- and paleoclimatic processes, Org. Geochem., 27, 213-250,
- doi:10.1016/S0146-6380(97)00049-1, 1997.
- 32 Nixon, S. W.: Coastal marine eutrophication: A definition, social causes, and future
- 33 concerns, Ophelia, 41, 199-219, doi:10.1080/00785236.1995.10422044, 1995.

- 1 Paerl, H. W.: Assessing and managing nutrient-enhanced eutrophication in estuarine
- 2 and coastal waters: Interactive effects of human and climatic perturbations, Ecol.
- 3 Eng., 26, 40-54, doi:10.1016/j.ecoleng.2005.09.006, 2006.
- 4 Paerl, H. W.: Controlling Eutrophication along the Freshwater–Marine Continuum:
 5 Dual Nutrient (N and P) Reductions are Essential, Estuar. Coast., 32, 593-601,
- 6 doi:10.1007/s12237-009-9158-8, 2009.
- Peterson, B. J. and Fry, B.: Stable Isotopes in Ecosystem Studies, Annu. Rev. Ecol.
 Syst., 18, 293-320, 1987.
- 9 Qian, W., Dai, M., Xu, M., Kao, S.-j., Du, C., Liu, J., Wang, H., Guo, L., and Wang, L.:
- Non-local drivers of the summer hypoxia in the East China Sea off the Changjiang
 Estuary, Estuar. Coast. Shelf Sci., doi:10.1016/j.ecss.2016.08.032, 2016.
- 12 Qian, W., Gan, J., Liu, J., He, B., Lu, Z., Guo, X., Wang, D., Guo, L., Huang, T., and
- 13 Dai, M.: Current status of emerging hypoxia in a large eutrophic estuary: the lower
- reach of Pearl River estuary, China, Submitted to Limnol. Oceanogr., 2017.
- 15 Rabalais, N., Cai, W.-J., Carstensen, J., Conley, D., Fry, B., Hu, X., Quiñones-Rivera,
- Z., Rosenberg, R., Slomp, C., Turner, E., Voss, M., Wissel, B., and Zhang, J.:
 Eutrophication-Driven Deoxygenation in the Coastal Ocean, Oceanography, 27,
- 18 172-183, doi:10.5670/oceanog.2014.21, 2014.
- 19 Rabalais, N. N., D áz, R. J., Levin, L. A., Turner, R. E., Gilbert, D., and Zhang, J.:
- 20 Dynamics and distribution of natural and human-caused hypoxia, Biogeosciences, 7,
- 21 585-619, doi:10.5194/bg-7-585-2010, 2010.
- 22 Rabouille, C., Conley, D. J., Dai, M. H., Cai, W. J., Chen, C. T. A., Lansard, B., Green,
- 23 R., Yin, K., Harrison, P. J., Dagg, M., and McKee, B.: Comparison of hypoxia among
- four river-dominated ocean margins: The Changjiang (Yangtze), Mississippi, Pearl,
- and Rhône rivers, Cont. Shelf Res., 28, 1527-1537, doi:10.1016/j.csr.2008.01.020,
 2008.
- 27 Rutger, R., Stefan, A., Birthe, H., Hans, C. N., and Karl, N.: Recovery of marine
- 28 benthic habitats and fauna in a Swedish fjord following improved oxygen conditions,
- 29 Mar. Ecol. Prog. Ser., 234, 43-53, doi:10.3354/meps234043, 2002.
- Shultz, D. J. and Calder, J. A.: Organic carbon ${}^{13}C/{}^{12}C$ variations in estuarine sediments, Geochim. Cosmochim. Acta, 40, 381-385, doi:10.1016/0016-7037(76)90002-8, 1976.

- 1 Steckbauer, A., Duarte, C. M., Carstensen, J., Vaquer-Sunyer, R., and Conley, D. J.:
- 2 Ecosystem impacts of hypoxia: thresholds of hypoxia and pathways to recovery,
- 3 Environ. Res. Lett., 6, 025003, doi:10.1088/1748-9326/6/2/025003, 2011.
- Stefan, R., Mateete, B., Clare, M. H., Nancy, K., Wilfried, W., Xiaoyuan, Y., Albert,
 B., and Mark, A. S.: Synthesis and review: Tackling the nitrogen management
 challenge: from global to local scales, Environ. Res. Lett., 11, 120205,
 doi:10.1088/1748-9326/11/12/120205, 2016.
- 8 Strong, D. J., Flecker, R., Valdes, P. J., Wilkinson, I. P., Rees, J. G., Zong, Y. Q., Lloyd,
- J. M., Garrett, E., and Pancost, R. D.: Organic matter distribution in the modern
 sediments of the Pearl River Estuary, Org. Geochem., 49, 68-82,
 doi:10.1016/j.orggeochem.2012.04.011, 2012.
- Swarzenski, P., Campbell, P., Osterman, L., and Poore, R.: A 1000-year sediment
 record of recurring hypoxia off the Mississippi River: The potential role of
 terrestrially-derived organic matter inputs, Mar. Chem., 109, 130-142,
 doi:10.1016/j.marchem.2008.01.003, 2008.
- Thornton, S. F. and McManus, J.: Application of Organic Carbon and Nitrogen Stable
 Isotope and C/N Ratios as Source Indicators of Organic Matter Provenance in
 Estuarine Systems: Evidence from the Tay Estuary, Scotland, Estuar. Coast. Shelf
 Sci., 38, 219-233, doi:10.1006/ecss.1994.1015, 1994.
- 20 Vaquer-Sunyer, R. and Duarte, C. M.: Thresholds of hypoxia for marine biodiversity,
- Proc. Natl. Acad. Sci. U.S.A., 105, 15452-15457, doi:10.1073/pnas.0803833105,
 2008.
- 23 Wang, H., Dai, M., Liu, J., Kao, S.-J., Zhang, C., Cai, W.-J., Wang, G., Qian, W., Zhao,
- M., and Sun, Z.: Eutrophication-Driven Hypoxia in the East China Sea off the Changjiang Estuary, Environ. Sci. Technol., 50, 2255-2263, doi:10.1021/acs.est.5b06211, 2016.
- 27 Wang, Q., Koshikawa, H., Liu, C., and Otsubo, K.: 30-year changes in the nitrogen
- inputs to the Yangtze River Basin, Environ. Res. Lett., 9, 115005,
 doi:10.1088/1748-9326/9/11/115005, 2014.
- 30 Ye, F., Huang, X., Zhang, X., Zhang, D., Zeng, Y., and Tian, L.: Recent oxygen
- depletion in the Pearl River Estuary, South China: geochemical and microfaunal
- 32 evidence, J. Oceanogr., 68, 387-400, doi:10.1007/s10872-012-0104-1, 2012.

Yu, F., Zong, Y., Lloyd, J. M., Huang, G., Leng, M. J., Kendrick, C., Lamb, A. L., and 1 Yim, W. W.-S.: Bulk organic δ^{13} C and C/N as indicators for sediment sources in the 2 Pearl River delta and estuary, southern China, Estuar. Coast. Shelf Sci., 87, 618-630, 3 doi:10.1016/j.ecss.2010.02.018, 2010. 4 Zhang, J., Zhang, Z. F., Liu, S. M., Wu, Y., Xiong, H., and Chen, H. T.: Human impacts 5 on the large world rivers: Would the Changjiang (Yangtze River) be an illustration?, 6 Global Biogeochem. Cycles, 13, 1099-1105, doi:10.1029/1999GB900044, 1999. 7 8 Zhang, J., Cowie, G., and Naqvi, S. W. A.: Hypoxia in the changing marine environment, Environ. Res. Lett., 8, 015025, doi:10.1088/1748-9326/8/1/015025, 9 2013. 10 Zhang, Y., Xiao, W., and Jiao, N.: Linking biochemical properties of particles to 11 particle-attached and free-living bacterial community structure along the particle 12 density gradient from freshwater to open ocean, J. Geophys. Res. Biogeosci., 121, 13 2261-2274, doi:10.1002/2016JG003390, 2016. 14 Zhao, H.-D., Kao, S.-J., Zhai, W.-D., Zang, K.-P., Zheng, N., Xu, X.-M., Huo, C., and 15 Wang, J.-Y.: Effects of stratification, organic matter remineralization and bathymetry 16 on summertime oxygen distribution in the Bohai Sea, China, Cont. Shelf Res., 134, 17 15-25, doi:10.1016/j.csr.2016.12.004, 2017. 18 19 Zhu, C., Wagner, T., Pan, J.-M., and Pancost, R. D.: Multiple sources and extensive degradation of terrestrial sedimentary organic matter across an energetic, wide 20 21 continental shelf. Geochem. Geophys. Geosyst., 12, Q08011, doi:10.1029/2011GC003506, 2011a. 22 Zhu, Z.-Y., Zhang, J., Wu, Y., Zhang, Y.-Y., Lin, J., and Liu, S.-M.: Hypoxia off the 23 Changjiang (Yangtze River) Estuary: oxygen depletion and organic matter 24 decomposition, Mar. Chem., 125, 108-116, doi:10.1016/j.marchem.2011.03.005, 25 2011b. 26 Zimmerman, A. R. and Canuel, E. A.: A geochemical record of eutrophication and 27 anoxia in Chesapeake Bay sediments: anthropogenic influence on organic matter 28 composition, Mar. Chem., 69, 117-137, doi:10.1016/S0304-4203(99)00100-0, 2000. 29 30 31

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1 Table 1. Summary of end-member values and their uncertainties adopted in the three 2 end-member mixing model.

Water		Salinity	DIC	$\delta^{13}C_{DIC}$	Ca ²⁺
Mass	0(C)		(µmol kg ⁻¹)	(‰)	(µmol kg ⁻¹)
Plume	30.6±1.0	10.8	1670 ± 50^{a}	-7.0±0.8 ^b	3670±16 ^c
Surface	31.0±1.0	33.7±0.2	1917±3	0.6±0.2	$9776 \pm 132^{\circ}$
Subsurface	21.8±1.0	34.6±0.1	2023±6	0.1±0.1	10053

^a In order to derive a proper plume end-member value, we took advantage of 3 years of surface
dataset from summer cruises (see Sect. 4.1). For DIC, the data is from cruises during August
2012, July 2014 and July 2015.
^b See details in Sect. 4.1.

^c The Ca^{2+} values of the plume and surface seawater end-member are derived from a conservative mixing calculation (Ca^{2+} vs. S) based on 3 years of surface data during the summer (August 2012, July 2014 and July 2015).

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1 **Table 2.** Comparison of δ^{13} C values in surface sediments within the hypoxic zone^a between

δ ¹³ C (‰)	Mean±SD	Stations involved	References				
Pearl River Estuary							
-23.4 ~ -22.1	-22.9 ± 0.5	A4, A5, C1-C4, D1	Hu et al. 2006				
-23.2 ~ -22.3	-22.7 ± 0.5	28, 29, 30	Zong et al. 2006				
-23.6 ~ -21.5	-22.5 ± 1.1	E8-1, E7A, S7-1, S7-2	He et al. 2010a				
_b	-23.1±0.6	Clustering groups G6 and G7	Yu et al. 2010				
Average	-22.8±0.6						
Changjiang Estuary							
-22.9 ~ -20.9	-21.8±0.6	_c	Tan et al. 1991				
-22.4 ~ -19.9	-21.2 ± 1.0	32, 37, 38, 42, 48, 49, 54, 56, 64	Kao et al. 2003				
-22.7 ~ -20.8	-22.0 ± 0.8	H1-12, H2-10, H2-11, S1-2, S2-4	Xing et al. 2011				
-23.5 ~ -20.4	-22.6±1.0	3, 12, 13, 20-25	Yao et al. 2014				
Average	-21.9±1.0						

2 the PRE and CJE.

^aIn the PRE, the data is from similar sites to our present study, which is in the northeast (Leg 1)

4 and southwest (Leg 2) of the Wanshan Islands. While in the CJE, the hypoxic zone is located

5 around 30.0 °N-32.0 °N, 122.7 °E-123.2 °E, which is frequently reported in previous studies

6 (Li et al., 2002; Zhu et al., 2011b; Wang et al., 2016).

7 ^bThe authors provide an average value of clustering groups instead of individual data from each

8 site.

⁹ ^cIn Fig. 7 of Tan et al. (1991), the sampling sites are shown without numbers.

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Figure 1. Map of the Pearl River Estuary and adjacent coastal waters. The open circles denote
Leg 1 stations visited on 13–16 July 2014, and the crosses represent Leg 2 stations visited on
19–27 July 2014. Note that the filled diamond is the location of Station A10.



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Figure 4. Profiles of temperature, salinity, DO, Chl-a, DIC and $\delta^{13}C_{DIC}$ along Transect A during Leg 1 (a–f) and Leg 2 (g–l). Note that the black lines in (c) and (i) indicate DO contours of 63 μ M and 95 μ M.



Figure 5. Distribution of $\delta^{13}C_{DIC}$ and $\delta^{13}C_{POC}$ with respect to salinity in the PRE. The up-facing and down-facing triangles denote surface and subsurface $\delta^{13}C_{DIC}$ data, respectively, from July 2014, while the open circles represent $\delta^{13}C_{POC}$ values in surface water from July 2015. Additionally, the plus signs and crosses show the $\delta^{13}C_{DIC}$ and $\delta^{13}C_{POC}$ data, respectively, from the CJE in Wang et al. (2016).





3 Figure 6. Profiles of (a) temperature, (b) salinity, (c) DO, (d) DIC, (e) DIP, (f) TSM and their

4 evolution during repeated sampling at Station A10.



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Figure 7. (a) Potential temperature (θ) (∞) vs. salinity in the PRE and adjacent coastal waters 2 3 (open circles) based on data collected during the July 2014 cruise. The three end-members are 4 shown as different coloured symbols. The blue triangles represent data collected during the August 2011 cruise in the CJE (Wang et al., 2016); (b) Correlation between the field-observed 5 $Ca^{2+} (Ca^{2+}_{obs})$ and model-predicted $Ca^{2+} (Ca^{2+}_{pre})$. The straight line denotes a linear regression 6 line of both surface (square) and subsurface (diamond) data; (c), (d) Relationship between 7 observed and model-predicted DIC and $\delta^{13}C_{DIC}$ values. The straight line represents a 1:1 8 reference line. Note that the grey dots in Fig. 7c identify data also in Fig. 7d; and (e) Correlation 9 10 of ΔDIC vs. AOU for all subsurface water data. ΔDIC is the difference between the 11 field-observed and model-predicted DIC concentrations. Also shown is the data from Wang et

1 al. (2016). The straight and dashed lines indicate linear regressions of data from the PRE and

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2 CJE, respectively.
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Figure 8. Δ ($\delta^{13}C_{DIC} \times DIC$) vs. ΔDIC in the PRE. Samples were collected from subsurface water (> 5 m). The grey circles represent samples with AOU > 100 µmol kg⁻¹. Δ is the difference between the field-observed and model-predicted values. Also shown is data from the CJE reported by Wang et al. (2016). The straight and dashed lines indicate linear regression lines of data from the PRE and CJE, respectively.



2 Figure 9. A conceptual diagram illustrating the partitioning of oxygen-consuming organic

- 3 matter (OC_{mar} vs. OC_{terr}) within the hypoxic zone in the lower PRE and the adjacent coastal
- 4 area. See Sect. 5 for explanations.