



1 **Tracing the origin of the oxygen-consuming organic
2 matter in the hypoxic zone in a large eutrophic estuary:
3 the lower reach of the Pearl River Estuary, China**

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11 **Abstract.** We assess the relative contributions of differently sourced organic matter, marine vs.
12 terrestrial, to oxygen consumption in an emerging hypoxic zone in the lower Pearl River Estuary (PRE),
13 a large eutrophic estuary located in Southern China. Our cruise, conducted in July 2014, contained two
14 legs before and after a passing through of Typhoon Rammasun, which destroyed the water stratification.
15 The stratification recovered rather quickly within one day after the typhoon. We observed, in both legs,
16 algal blooms in the surface water with hypoxias underneath in the bottom water. The repeated samplings
17 at the initial hypoxic station showed severe oxygen depletion down to 30.3 $\mu\text{mol kg}^{-1}$ before the typhoon
18 and a clear drawdown of dissolved oxygen after the typhoon. Based on a three end-member mixing
19 model and the mass balance of dissolved inorganic carbon and its isotopic composition, we derived the
20 $\delta^{13}\text{C}$ of the remineralized organic carbon as $-21.8 \pm 0.6\text{‰}$ in the hypoxia. We estimated that $73 \pm 10\text{ \%}$ of
21 the oxygen-consuming organic matter was marine sourced and, the rest $27 \pm 10\text{ \%}$ was terrestrially
22 sourced. Contrasting to the hypoxic site in the East China Sea off the Changjiang Estuary where
23 eutrophication-stimulated marine sourced organic matter prevailed the oxygen consumption, the
24 terrestrially sourced organic matter significantly contributed to the formation and maintenance of the
25 hypoxia in the present study site. The difference in oxygen sinks in different systems has important
26 implications in better understanding to the controls of hypoxias and their mitigations.



1 1 Introduction

2 The occurrence of hypoxias has been exacerbated worldwide (Nixon, 1995; Rabalais et al., 2010; Zhang
3 et al., 2013). In recent decades, more than 400 coastal hypoxic systems are reported with an exponential
4 growth rate of $5.5 \pm 0.23\% \text{ yr}^{-1}$, demonstrating the persistence and complexity in both science and
5 management (Vaquer-Sunyer and Duarte, 2008; Diaz and Rosenberg, 2008). Hypoxia may not only
6 reduce the biodiversity, endanger aquatic and benthic habitats, but also alter the redox chemistry in both
7 the water column and the underlying sediments, triggering secondary pollutants in the water body
8 (Breitburg, 2002; Rutger et al., 2002). Moreover, the hysteresis-like behavior involved in the recovery
9 from hypoxia, due to the differential timescales of biological loss (within hours to weeks) and recovery
10 from hypoxia (from months to years), indicates the difficulties in the restoring process and urgency for
11 proper managerial actions (Steckbauer et al., 2011).

12 Coastal hypoxia usually occurs in stratified water columns where the downward mixing of oxygen from
13 the surface is impeded (Kemp et al., 2009). Below the pycnocline, the aerobic respiratory process is
14 usually the predominant sink of oxygen. Organic matter that consumes dissolved oxygen (DO) is thus
15 the ultimate cause of hypoxia under favourable physical settings (Rabouille et al., 2008; Rabalais et al.,
16 2014; Qian et al., 2016a). The organic carbon (OC) that fuels the respiratory reduction of oxygen could
17 originate from the eutrophication-induced primary production (marine sourced OC, OC_{mar}) and
18 terrestrially natural leaching and anthropogenic drainage (terrestrially sourced OC, OC_{terr}) (Paerl, 2006;
19 Rabalais et al., 2010).

20 The question of how much OC in the bottom hypoxic zone is imported from the on-site primary
21 production versus how much OC is derived from terrestrial sources has been an issue of debate (Wang et
22 al., 2016). Much of the phytoplankton-centric hypoxia literature suggests that the OC_{mar} dominates the
23 oxygen consumption in hypoxic zones owing to the higher microbial availability (Zimmerman and
24 Canuel, 2000; Boesch et al., 2009; Carstensen et al., 2014). Wang et al. (2016) quantified for the first
25 time the relative contributions of particulate OC_{mar} (POC_{mar}) and particulate OC_{terr} (POC_{terr}) in
26 consuming DO in the bottom water of the East China Sea (ECS) off the Changjiang Estuary (CJE), and
27 found that POC_{mar} prevailed in DO consumption. There existed other studies showing that terrestrially
28 sourced organic matter may also be important (Swarzenski et al., 2008; Bianchi, 2011a; Bianchi et al.,



1 2011b). It is thus highly significant to quantify the relative contributions of organic matter (OC_{mar} vs.
2 OC_{terr}) in different systems, since reducing organic matter and nutrient loadings would require different
3 management strategies.

4 The Pearl River Estuary (PRE, 21.2°N – 23.1°N , 113.0°E – 114.5°E) is surrounded by large cities
5 including Hong Kong, Shenzhen and Guangzhou and has received very high loads of nutrients from the
6 drainage basin in the last three decades, and eutrophication has increasingly become an issue of concern
7 (Ye et al., 2012; Huang et al., 2003). The dissolved inorganic nitrogen concentration in the PRE has
8 increased about 4-fold in 2002 ($76.1 \mu\text{mol L}^{-1}$) compared with 1986 ($19.3 \mu\text{mol L}^{-1}$) (He and Yuan,
9 2007). Such an increase arises mainly from the intensifying domestic sewage, industrial wastewater,
10 agricultural runoff and aquaculture in the watershed (Huang et al., 2003).

11 Recent observations by Qian et al. (Submitted) based on their monthly surveys between April 2010 and
12 March 2011 together with the long term monitoring data from 1990 to 2014 have suggested that the
13 lower PRE has emerged as a seasonal hypoxic zone (Qian et al., 2016b). Moreover, during our cruise,
14 two relatively large hypoxic zones ($\sim 1000 \text{ km}^2$) occurred in the lower PRE with $\text{DO} < 3 \text{ mg L}^{-1}$. However,
15 the origin of the organic matter that induced the emerging hypoxia in the lower PRE has not been
16 examined. The present study thus sought to assess in a quantitative way the origin of the
17 oxygen-consuming organic matter in the lower PRE, an economically important region. We point out
18 that such a study has additionally significant societal relevance, such as the water quality in the vicinity of
19 Hong Kong located at the lower PRE. The government of Hong Kong is examining the efficacy of its
20 costly Harbour Area Treatment Scheme project and if additional treatment should be implemented
21 (<http://www.gov.hk/en/residents/environment/water/harbourarea.htm>).

22 **2 Materials and Methods**

23 **2.1 Sampling and analysis**

24 Interrupted by Typhoon Rammasun on 17–18 July 2014, our cruise was divided into two legs (Fig. 1). In
25 Leg 1 on 13–16 July, we sampled Sections F4, F5 and Stations A08–A18. During Leg 2 on 19–27 July,
26 we sampled Stations A01–A10, Sections F3, F4, Stations A11–A17 and Sections F5, F6, F1, F2 in



1 sequence. In order to monitor the development of the hypoxia before and after the typhoon event, we
2 revisited Station A10 three more times (13, 20 and 27 July).

3 According to the gauge records in the upper Pearl River, water discharge peaked in June and July.
4 Typhoon Rammasun brought about a relatively high discharge ($26115 \text{ m}^3 \text{ s}^{-1}$) on 16 July. Nevertheless,
5 the freshwater discharge during our cruise was rather comparable to the long-term (2000–2011) monthly
6 average, suggesting that this study represented a typical situation of the area in terms of terrestrial
7 material discharge.

8 Temperature and salinity were determined with a SBE 25 Conductivity-Temperature-Depth/Pressure
9 unit (Sea-Bird Co.). Water samples were collected using 4 L Go-Flo bottles (General Oceanics). The DO
10 concentrations in discrete samples were measured on board using the classic Winkler procedure within 8
11 h (Dai et al., 2006). DIC was measured with an infrared detector after acidifying 0.5–0.7 mL of water
12 sample with a precision of 0.1 % for estuarine and sea waters (Cai et al., 2004). Dissolved calcium
13 concentration (Ca^{2+}) was determined using an EGTA titration with a Metrohm 809 TITRANDO
14 potentiometer, which has a precision better than $\pm 5 \mu\text{mol kg}^{-1}$ (Cao et al., 2011).

15 For the analysis of $\delta^{13}\text{C}_{\text{DIC}}$, an ~20 mL DIC sample was converted into gaseous CO_2 and progressively
16 purified through a vacuum line. The pure CO_2 sample was analyzed with an isotope ratio mass
17 spectrometer (IRMS, Finnigan MAT 252, Bremen, Germany). The analytical precision was better than
18 0.1 ‰.

19 TSM (total suspended matter), POC and $\delta^{13}\text{C}_{\text{POC}}$ samples were concentrated onto a preweighed and
20 pre-combusted 0.7 μm Whatman GF/F filters after filtering 0.2–1.0 L water under a mild vacuum (~ 25
21 kPa). Filters were washed with distilled water and stored under a -20 °C environment. Before analysis,
22 the filters were freeze-dried. The TSM was determined using the net weight increment on the filter and
23 the filtration volume. The filters were decarbonated with 1.0 mol L^{-1} HCl and dried at 40 °C for 48 h (Kao
24 et al., 2012) and analyzed for POC and $\delta^{13}\text{C}_{\text{POC}}$ on an elemental analyzer coupled with an isotope-ratio
25 mass spectrometer (IRMS). The analytical precision for $\delta^{13}\text{C}_{\text{POC}}$ was better than 0.1 ‰. Chl-a was
26 measured using a Turner fluorometer after extraction of the filter samples with 90 % acetone (He et al.,
27 2010b). Calibrations were performed using Sigma Chl-a standard.



1 2.2 Three end-member mixing model

2 We adopted a three end-member mixing model to construct the conservative mixing scheme among
3 different water masses (Cao et al., 2011; Han et al., 2012):

$$4 \quad F_{\text{RI}} + F_{\text{SW}} + F_{\text{SUB}} = 1 \quad (1)$$

$$5 \quad \theta_{\text{RI}} \times F_{\text{RI}} + \theta_{\text{SW}} \times F_{\text{SW}} + \theta_{\text{SUB}} \times F_{\text{SUB}} = \theta \quad (2)$$

$$6 \quad S_{\text{RI}} \times F_{\text{RI}} + S_{\text{SW}} \times F_{\text{SW}} + S_{\text{SUB}} \times F_{\text{SUB}} = S \quad (3)$$

7 where θ and S represent the potential temperature and salinity; the subscripts RI, SW, SUB denote the
 8 three different water masses (the Pearl River plume water, the offshore surface seawater and the
 9 upwelled subsurface water); and F_{RI} , F_{SW} , F_{SUB} represent the fractions that each end-member contributes
 10 to the in situ samples. Then, these fractions were applied to predict the DIC and its isotopic composition
 11 ($\delta^{13}\text{C}_{\text{DIC}}$) solely from conservative mixing. The difference (Δ) between measurement and prediction
 12 values could be acquired, which represented the magnitude of the biological alteration (Wang et al.,
 13 2016).

14 **3 Results**

15 3.1 Horizontal distribution

16 The average freshwater discharge rate during our cruises ($16369 \text{ m}^3 \text{ s}^{-1}$) showed a minor difference (<5
17 %) with the multi-years (2000–2011) monthly mean discharge ($15671 \text{ m}^3 \text{ s}^{-1}$). We noted that the survey
18 area in Leg 1 was smaller, covering only outside Lingdingyang Bay, traditionally regarded as PRE.
19 While, in Leg 2, the survey area covered the Lingdingyang Bay from the Humen Outlet to the adjacent
20 coastal sea.

21 In each leg, we found an algal bloom zone at the surface and a hypoxic zone at the bottom (Fig. 2). In
22 Leg 1, the bloom zone, located to the east of the Wanshan Islands, was oxygen supersaturated with the
23 highest DO saturation (DO%) greater than 160 % at Station F503, coupled with higher Chl-a (8.0 μg
24 kg^{-1}) and lower surface DIC (1606.8 $\mu\text{mol kg}^{-1}$). Meanwhile, the bottom hypoxia lay more landward and
25 the center's DO was as low as 30.3 $\mu\text{mol kg}^{-1}$ at Station A10, along with higher DIC (2074.9 $\mu\text{mol kg}^{-1}$).
26 However, in Leg 2, the bloom zone was more westward and larger than the former one, with the highest
27 DO% greater than 140 % at Station A14. In spite of the disappearance of the previous hypoxia near



1 Station A10, another hypoxic zone was discovered to the southeast of the Wanshan Islands, with a lowest
2 DO value of $7.2 \mu\text{mol kg}^{-1}$ and a higher DIC ($2146.1 \mu\text{mol kg}^{-1}$) in F304. We were unable to precisely
3 constrain the areas of the bottom hypoxia due to the limited spatial coverage, but they covered at least
4 $\sim 900 \text{ km}^2$ and $\sim 800 \text{ km}^2$ for Legs 1 and 2.

5 **3.2 Vertical distribution**

6 In Leg 1, the plume water reached 50 km away from the entrance of the PRE, and formed a 5–10 m thick
7 plume (Fig. 3a–e). Both the thermocline and halocline contributed to the stability of the water structure,
8 which favored the formation of the bottom hypoxia. The thickness of the bottom hypoxia was $\sim 10 \text{ m}$. The
9 algal bloom zone was not right above the hypoxia, but more seaward.

10 In Leg 2, although the typhoon attack would absorb enormous potential heat and aggravate water
11 mixing, the enhanced freshwater discharge could soon make the stratification strong enough for the
12 formation of hypoxia. At this time, the hypoxia was right below the bloom zone, with a thickness of 7 m
13 (Fig. 3f–j). Also, near the Humen Outlet, we found low DIC ($1466.3 \mu\text{mol kg}^{-1}$) and moderately low DO
14 water ($89.1 \mu\text{mol kg}^{-1}$), which reflected the input of the low DO water mass from the upstream as
15 reported previously (Dai et al., 2006; Dai et al., 2008a; He et al., 2014).

16 **3.3 Isotopic composition of DIC and POC**

17 The $\delta^{13}\text{C}$ of DIC became heavier from freshwater ($\sim -11.4 \text{ ‰}$) to off-shore seawater ($\sim -0.6 \text{ ‰}$), with a
18 relatively wide range beyond salinity=13 (Fig. 4). Owing to a malfunction of the machine, the $\delta^{13}\text{C}_{\text{POC}}$
19 data from our cruise were not available. Instead, we reported a valid $\delta^{13}\text{C}_{\text{POC}}$ dataset from a 2015 summer
20 cruise in nearly the same region. The $\delta^{13}\text{C}$ of POC showed a similar trend with $\delta^{13}\text{C}_{\text{DIC}}$, i.e. ^{13}C enriched
21 seaward, from $\sim -28 \text{ ‰}$ to $\sim -20 \text{ ‰}$. In the algal bloom zone, where the DO% was beyond 125 %, the
22 mean $\delta^{13}\text{C}$ value for POC was $-19.4 \pm 0.8 \text{ ‰}$ ($n=8$), which was within the typical range of the marine
23 phytoplankton (Peterson and Fry, 1987). As shown in Fig. 4, there was a $\delta^{13}\text{C}_{\text{POC}}$ trough near a salinity of
24 15. Geologically, it was located at the mixing dominated zone in the inner Lingdingyang Bay, where
25 intense resuspension of ^{13}C depleted sediment may occur (Guo et al., 2009).



1 **3.4 Reinstatement of the hypoxic station after Typhoon Rammasun**

2 Typhoon Rammasun landed on Zhanjiang located 400 km to the southwest of the PRE at 20:00 LT on 18
3 July and was dissipated at 05:00 LT on 20 July. This typhoon destroyed the water stratification.
4 However, the associated heavy precipitation and rising runoff appeared to reestablish the stratification
5 rather quickly, within one day, as suggested by the saline gradients (18–30) within the 0–10 m depth
6 during our revisit at 15:20 LT on 20 July (Fig. 5b). In order to capture the evolution of oxygen between
7 stratification disruption and reinstatement, we resumed our cruise and revisited Station A10 on 13, 20
8 and 27 July (Fig. 5). On 13 July, the bottom water of Station A10 was the hypoxic core, with the lowest
9 DO (30.3 $\mu\text{mol kg}^{-1}$) and highest DIC (2074.9 $\mu\text{mol kg}^{-1}$). On 20 July, the results showed that the
10 temperature homogeneous layer in the bottom water (9–13 m) might be the trail of the typhoon induced
11 mixing (Fig. 5a), while the saline stratification (<9 m) depicted the rapid recuperation as a result of
12 enhanced freshwater discharge (Fig. 5b). The bottom DO increased to 153.1 $\mu\text{mol kg}^{-1}$ and the DIC
13 decreased to 1900.7 $\mu\text{mol kg}^{-1}$ because of the water mixing and aeration. In addition, the TSM increased
14 sharply from 20.2 to 36.6 mg kg^{-1} , suggesting strong sediment resuspension. On 27 July, one week after
15 the typhoon, the homogenous mixing layer in the bottom disappeared and strong thermohaline
16 stratification occupied the whole water column again. Along with the intensifying stratification, the
17 bottom DO decreased to 98.6 $\mu\text{mol kg}^{-1}$ indicating continuous DO depletion and the potential trend for
18 hypoxia formation. Meanwhile, the bottom DIC increased to 2000.1 $\mu\text{mol kg}^{-1}$ and the dissolved
19 inorganic phosphate (DIP) rose from 0.28 to 0.57 $\mu\text{mol kg}^{-1}$. Moreover, the bottom-water TSM returned
20 to the same level as prior to the typhoon (13 July).

21 **4 Discussion**

22 **4.1 Selection of end-members and model validation**

23 The potential temperature-salinity plot displayed a three end-member mixing scheme over the PRE and
24 the adjacent coastal water (Fig. 6a), consisting of the Pearl River plume water, offshore surface seawater
25 and the upwelled subsurface water. In summer, the DIC was \sim 1917 $\mu\text{mol kg}^{-1}$ at S=33.7, which can be
26 regarded as the off-shore surface seawater end-member (Guo and Wong, 2015). When we chose S=34.6
27 as the off-shore subsurface water salinity end-member, we obtained a DIC of \sim 2023 $\mu\text{mol kg}^{-1}$. For the



1 plume end-member, it was difficult to directly select from the field data, because biological alteration
2 within the plume influenced area might lead to an improper calculation result. Therefore, we first
3 assumed that the plume water was the mixture between the freshwater water and off-shore surface
4 seawater. Then, we compiled a 3 years' surface dataset in summer (August 2012, July 2014 and July
5 2015) to extrapolate the relatively stable freshwater end-member so as to examine the biological effect
6 along with salinity. From the constraint of the DIC end-members (freshwater and off-shore surface
7 seawater), we observed that DIC remained overall conservative when salinity was <10.8 but showed
8 removal when salinity >10.8 (Han et al., 2012). Thus, we derived plume end-member values from the
9 property-salinity conservative mixing curve at salinity=10.8. Furthermore, S=10.8 was located on the
10 innermost station (A08) in Leg 1, which coincided with the spatial and temporal scale of the actual water
11 mass mixing in our survey. As a guarantee, we also used a freshwater end-member (S=0), but the outputs
12 of the model showed little difference from those based on the plume end-member at S=10.8. The
13 summary of the end-member values in this study is listed in Table 1.

14 We calculated the water fractions of the three water parcels based on potential temperature and salinity
15 equations, so as to predict DIC and its isotopic composition ($\delta^{13}\text{C}_{\text{DIC}}$) solely from conservative mixing.
16 We chose the concentration of Ca^{2+} as a conservative tracer to validate our model prediction, and the
17 model derived values were in good accordance with the field-observed values (Fig. 6b), which strongly
18 supported our model prediction.

19 As shown in Fig. 6c, most of the observed DIC in the subsurface water was higher than the predicted
20 values, as a result of the DIC production via OC oxidation. On the other hand, the majority of observed
21 $\delta^{13}\text{C}_{\text{DIC}}$ values became lighter than the predicted ones, owing to the accumulation of lighter carbon
22 remineralized from the organic matter (Fig. 6d). Based on the difference between the observed and
23 predicted values of DIC and $\delta^{13}\text{C}_{\text{DIC}}$, the carbon isotopic composition of the oxygen-consuming organic
24 matter could be traced precisely (see details in Sect. 4.2).

25 In the subsurface water, most of the ΔDIC varied from 0 to $132.3 \mu\text{mol kg}^{-1}$, coupled with an apparent
26 oxygen utilization (AOU) increment from 0 to $179.1 \mu\text{mol kg}^{-1}$. ΔDIC had a positive correlation with the
27 AOU (Fig. 6e), corresponding with the fact that the additional DIC was primarily produced by organic
28 matter remineralization via aerobic respiration. The slope of ΔDIC vs. AOU in the subsurface water was



1 0.75 \pm 0.03, which matched well with the classic Redfield stoichiometry (i.e., 106/138=0.77), providing
2 further evidence for the occurrence of aerobic respiration.

3 **4.2 Isotopic composition of the oxygen-consuming OC**

4 The mass balance of DIC isotopic composition is shown by Eq. (4) (Wang et al., 2016):

$$5 \quad \delta^{13}\text{C}_{\text{DICobs}} \times \text{DIC}_{\text{obs}} = \delta^{13}\text{C}_{\text{DICpre}} \times \text{DIC}_{\text{pre}} + \delta^{13}\text{C}_{\text{DICbio}} \times \text{DIC}_{\text{bio}} \quad (4)$$

6 where the subscripts obs, pre and bio refer to the field-observed, model-predicted and biologically altered
7 values.

8 Degradation of OC generally produces DIC with minor isotopic fractionation relative to the OC
9 substrate (Hullar et al., 1996; Breteler et al., 2002). Thus, the isotopic composition of DIC_{bio} (i.e.,
10 $\delta^{13}\text{C}_{\text{DICbio}}$) should be identical to the $\delta^{13}\text{C}_{\text{OCx}}$ of the OC, which consumed oxygen and produced DIC_{bio} ,
11 and the $\delta^{13}\text{C}_{\text{OCx}}$ could be derived from the mass balance equations of both DIC and its stable isotope:

$$12 \quad \delta^{13}\text{C}_{\text{OCx}} = \frac{\delta^{13}\text{C}_{\text{obs}} \times \text{DIC}_{\text{obs}} - \delta^{13}\text{C}_{\text{pre}} \times \text{DIC}_{\text{pre}}}{\text{DIC}_{\text{obs}} - \text{DIC}_{\text{pre}}} \quad (5)$$

13 Equation (5) can be rearranged into Eq. (6):

$$14 \quad \Delta(\delta^{13}\text{C}_{\text{DIC}} \times \text{DIC}) = \delta^{13}\text{C}_{\text{OCx}} \times \Delta\text{DIC} \quad (6)$$

15 As shown in Fig. 7, the slope of the linear regression represents $\delta^{13}\text{C}_{\text{OCx}}$ or $\delta^{13}\text{C}_{\text{DICbio}}$, which equals
16 -21.8 \pm 0.6 ‰, reflecting a general $\delta^{13}\text{C}$ of the remineralized organic matter (Wang et al., 2016). The
17 average $\delta^{13}\text{C}$ of POC samples near the Humen Outlet (S<4) was -28.3 \pm 0.7 ‰ (n=7), representing
18 terrestrial origin. Previous studies adopt the $\delta^{13}\text{C}$ values of the terrigenous OC end-member in the PRE
19 from -28.7 to -24.9 ‰ (He et al., 2010a; Yu et al., 2010; Hu et al., 2006). Likewise, Yu et al. (2010) report
20 that the terrigenous freshwater POC has $\delta^{13}\text{C}$ around -28.7 ‰, reflecting a mixed nature of the C3 plant
21 fragments and forest soils. For the surface water data (n=8) at stations with high salinity (S>26) and high
22 DO saturation (DO%>125 %), the average $\delta^{13}\text{C}_{\text{POC}}$ value was -19.4 \pm 0.8 ‰, reflecting a stable carbon
23 isotopic signature mainly from the marine phytoplankton biomass. Since previous studies choose a
24 marine OC end-member from -21.2 to -20.5 ‰ (He et al., 2010a; Yu et al., 2010; Hu et al., 2006; Chen et
25 al., 2008), it was reasonable to assign -19.4 \pm 0.8 ‰ as the $\delta^{13}\text{C}$ value of OC_{mar} in the PRE and the adjacent
26 coastal water.



1 Although studies have shown the effects of selective diagenesis of organic matter fractions that are
2 isotopically heavy or light (Marthur et al., 1992; Lehmann et al., 2002), these effects are small compared
3 to the isotopic differences among differently sourced organic matter (Meyers, 1997). It is thus reasonable
4 to assume that the isotopic ratios are conservative and that the end-member sources determine the
5 isotopic distributions of organic matter in natural systems (Cifuentes et al., 1988). The relative
6 contributions from marine and terrestrially sourced organic matter to oxygen-consuming organic matter
7 could be estimated based on the following equation (Shultz and Calder, 1976; Hu et al., 2006):

$$8 \quad f(\%) = \frac{\delta^{13}\text{C}_{\text{mar}} - \delta^{13}\text{C}_{\text{OCx}}}{\delta^{13}\text{C}_{\text{mar}} - \delta^{13}\text{C}_{\text{terr}}} \times 100 \quad (7)$$

9 The result showed that the marine sourced organic matter contributed $73 \pm 10\%$ to the
10 oxygen-consuming organic matter pool, while the terrestrially sourced organic matter accounted for the
11 remaining $27 \pm 10\%$. It is thus clear that the marine organic matter from eutrophication-induced primary
12 production dominated oxygen consumption in the hypoxic zone while the terrestrially sourced organic
13 matter also significantly contributed to the formation and maintenance of hypoxia in the lower PRE and
14 the adjacent coastal water.

15 The processes involved in the source partitioning of organic matter, their isotopic signals and their
16 subsequent biogeochemical processes in the hypoxic zone are illustrated in the conceptual diagram in Fig.
17 8. River delivers a significant amount of nutrients and terrestrial organic matter into the estuary,
18 stimulating phytoplankton blooms in the surface water at the lower reach of the estuary where turbidity is
19 relatively low and conditions are favorable for phytoplankton growth (Gaston et al., 2006; Dai et al.,
20 2008b; Guo et al., 2009). The subsequent sinking of the biomasses and the delivered terrestrial organic
21 matter below the pycnocline consume oxygen and add respired DIC into subsurface water, thereby
22 forming a coastal hypoxia. Within the hypoxic zone, the most important biological process is aerobic
23 respiration.

24 As one of the largest rivers in the world, the Changjiang has been suffering from eutrophication for the
25 past few decades (Zhang et al., 1999; Wang et al., 2014). In summer, sharp density gradients with
26 frequent algal blooms and subsequent organic matter decomposition cause seasonal hypoxia in the
27 bottom water of the ECS off the CJE. Wang *et al* (2016) revealed that the remineralization of marine
28 sourced organic matter, OC_{mar} , overwhelmingly (nearly 100 %) contributed to DO consumption in the



1 ECS off the CJE. However, our present study showed that less OC_{mar} contributed to the oxygen depletion
2 ($73 \pm 10\%$) in the hypoxic zone of the lower PRE. The exact reasons causing such a difference between
3 two systems are unclear. However, it might be related to the differences in the bioavailability of OC_{terr} ,
4 microbial community and physical settings between these two hypoxic systems. First of all, although C3
5 plants dominate and C4 plants are minor in both the Pearl River and Changjiang drainage basins (Hu et
6 al., 2006; Zhu et al., 2011), the OC_{terr} has gone through different extent of degradation within the
7 estuaries before being transported into the hypoxic zones. In the CJE, approximate 50 % of the OC_{terr} has
8 been mineralized down the estuary likely due to efficient OM unloading from mineral surfaces (Zhu et
9 al., 2011). On the contrary, the PRE appears to be a somewhat intermediate site with respect to the export
10 of OC_{terr} , being closely associated with sedimentary regimes and not characterized by extensive
11 degradative loss (Strong et al., 2012). Thus, the bioavailability of OC_{terr} is likely higher in the PRE than in
12 the CJE. Secondly, the difference in bacterial community structure between two systems may also play a
13 role. Recent studies have demonstrated that the bacterial community was characterized by higher relative
14 abundances of Actinobacteria and lower relative abundances of Cytophaga-Flavobacteria-Bacteroides
15 (CFB) in the PRE than in the CJE (Zhang et al., 2016; Liu et al., 2012). Whether such differences would
16 promote the degradation of OC_{terr} in the PRE remains to be known. Finally, the temperature of the bottom
17 water in the PRE hypoxia (27–29 °C) was higher than in the CJE hypoxia (21.5–24.0 °C), which may
18 have accelerated the rates of bacterial decomposition and growth (Brown et al., 2004).

19 **5 Conclusions**

20 Based on a three end-member mixing model and the mass balance of dissolved inorganic carbon and its
21 isotopic composition, we demonstrated that the decomposed organic matter via aerobic respiration
22 consisted of a major portion of OC_{mar} ($73 \pm 10\%$) and a less but significant portion of OC_{terr} ($27 \pm 10\%$), in
23 the stratified subsurface waters of the lower PRE and the adjacent coastal water. Such source partitioning
24 of organic matter in oxygen sinks differed from the hypoxic zone off the CJE. The difference in oxygen
25 sinks in different systems has important implications in better understanding to the controls of hypoxias
26 and their mitigations. Nevertheless, corresponding to the increasing nutrient levels, a significant
27 implication of the present study is that reducing and managing nutrients is critical to control



1 eutrophication and to subsequently mitigate hypoxias (Conley et al., 2009; Paerl, 2009; Stefan et al.,
2 2016; Mercedes et al., 2015). Given the fact that, the terrestrially sourced organic carbon also contributed
3 to the consumption of oxygen in the lower PRE hypoxic zone, it is also crucial to characterize these
4 oxygen-consuming terrestrial organic matter from natural soil leaching and/or anthropogenic wastewater
5 discharge so as to make proper policies for hypoxia remediation.

6

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13

14 **References**

15 Bianchi, T. S.: The role of terrestrially derived organic carbon in the coastal ocean: A changing paradigm
16 and the priming effect, Proc. Natl. Acad. Sci. U.S.A., 108, 19473-19481,
17 doi:10.1073/pnas.1017982108, 2011a.

18 Bianchi, T. S., Wysocki, L. A., Schreiner, K. M., Filley, T. R., Corbett, D. R., and Kolker, A. S.: Sources
19 of terrestrial organic carbon in the Mississippi plume region: evidence for the importance of coastal
20 marsh inputs, Aquat. Geochem., 17, 431-456, doi:10.1007/s10498-010-9110-3, 2011b.

21 Boesch, D. F., Boynton, W. R., Crowder, L. B., Diaz, R. J., Howarth, R. W., Mee, L. D., Nixon, S. W.,
22 Rabalais, N. N., Rosenberg, R., Sanders, J. G., Scavia, D., and Turner, R. E.: Nutrient Enrichment
23 Drives Gulf of Mexico Hypoxia, Eos, Trans. Amer. Geophys. Union, 90, 117-118,
24 doi:10.1029/2009EO140001, 2009.

25 Breitburg, D.: Effects of hypoxia, and the balance between hypoxia and enrichment, on coastal fishes and
26 fisheries, Estuaries, 25, 767-781, doi:10.1007/BF02804904, 2002.



1 Breteler, W. C. K., Grice, K., Schouten, S., Kloosterhuis, H. T., and Damsté, J. S. S.: Stable carbon
2 isotope fractionation in the marine copepod *Temora longicornis*: unexpectedly low $\delta^{13}\text{C}$ value of
3 faecal pellets, *Mar. Ecol. Prog. Ser.*, 240, 195-204, doi:10.3354/meps240195, 2002.

4 Brown, J. H., Gillooly, J. F., Allen, A. P., Savage, V. M., and West, G. B.: Toward a metabolic theory of
5 ecology, *Ecology*, 85, 1771-1789, doi:10.1890/03-9000, 2004.

6 Cai, W.-J., Dai, M., Wang, Y., Zhai, W., Huang, T., Chen, S., Zhang, F., Chen, Z., and Wang, Z.: The
7 biogeochemistry of inorganic carbon and nutrients in the Pearl River estuary and the adjacent Northern
8 South China Sea, *Cont. Shelf Res.*, 24, 1301-1319, doi:10.1016/j.csr.2004.04.005, 2004.

9 Cao, Z., Dai, M., Zheng, N., Wang, D., Li, Q., Zhai, W., Meng, F., and Gan, J.: Dynamics of the
10 carbonate system in a large continental shelf system under the influence of both a river plume and
11 coastal upwelling, *J. Geophys. Res. Biogeosci.*, 116, G02010, doi:10.1029/2010JG001596, 2011.

12 Carstensen, J., Andersen, J. H., Gustafsson, B. G., and Conley, D. J.: Deoxygenation of the Baltic Sea
13 during the last century, *Proc. Natl. Acad. Sci. U.S.A.*, 111, 5628-5633, doi:10.1073/pnas.1323156111,
14 2014.

15 Chen, F., Zhang, L., Yang, Y., and Zhang, D.: Chemical and isotopic alteration of organic matter during
16 early diagenesis: Evidence from the coastal area off-shore the Pearl River estuary, south China, *J. Mar.*
17 *Syst.*, 74, 372-380, doi:10.1016/j.jmarsys.2008.02.004, 2008.

18 Cifuentes, L., Sharp, J., and Fogel, M. L.: Stable carbon and nitrogen isotope biogeochemistry in the
19 Delaware estuary, *Limnol. Oceanogr.*, 33, 1102-1115, doi:10.4319/lo.1988.33.5.1102, 1988.

20 Conley, D. J., Paerl, H. W., Howarth, R. W., Boesch, D. F., Seitzinger, S. P., Karl, E., Karl, E., Lancelot,
21 C., Gene, E., and Gene, E.: Controlling eutrophication: nitrogen and phosphorus, *Science*, 123,
22 1014-1015, doi:10.1126/science.1167755, 2009.

23 Dai, M., Guo, X., Zhai, W., Yuan, L., Wang, B., Wang, L., Cai, P., Tang, T., and Cai, W.-J.: Oxygen
24 depletion in the upper reach of the Pearl River estuary during a winter drought, *Mar. Chem.*, 102,
25 159-169, doi:10.1016/j.marchem.2005.09.020, 2006.

26 Dai, M., Wang, L., Guo, X., Zhai, W., Li, Q., He, B., and Kao, S.-J.: Nitrification and inorganic nitrogen
27 distribution in a large perturbed river/estuarine system: the Pearl River Estuary, China,
28 *Biogeosciences*, 5, 1227-1244, doi:10.5194/bg-5-1227-2008, 2008a.



1 Dai, M., Zhai, W., Cai, W.-J., Callahan, J., Huang, B., Shang, S., Huang, T., Li, X., Lu, Z., Chen, W., and
2 Chen, Z.: Effects of an estuarine plume-associated bloom on the carbonate system in the lower reaches
3 of the Pearl River estuary and the coastal zone of the northern South China Sea, *Cont. Shelf Res.*, 28,
4 1416-1423, doi: 10.1016/j.csr.2007.04.018, 2008b.

5 Diaz, R. J. and Rosenberg, R.: Spreading dead zones and consequences for marine ecosystems, *Science*,
6 321, 926-929, doi:10.1126/science.1156401, 2008.

7 Gaston, T. F., Schlacher, T. A., and Connolly, R. M.: Flood discharges of a small river into open coastal
8 waters: Plume traits and material fate, *Estuar. Coast. Shelf Sci.*, 69, 4-9,
9 doi:10.1016/j.ecss.2006.03.015, 2006.

10 Guo, X., Dai, M., Zhai, W., Cai, W.-J., and Chen, B.: CO₂ flux and seasonal variability in a large
11 subtropical estuarine system, the Pearl River Estuary, China, *J. Geophys. Res. Biogeosci.*, 114,
12 G03013, doi:10.1029/2008JG000905, 2009.

13 Guo, X. and Wong, G. T.: Carbonate chemistry in the northern South China Sea shelf-sea in June 2010,
14 *Deep-Sea Res. II*, 117, 119-130, doi:10.1016/j.dsr2.2015.02.024, 2015.

15 Han, A., Dai, M., Kao, S.-J., Gan, J., Li, Q., Wang, L., Zhai, W., and Wang, L.: Nutrient dynamics and
16 biological consumption in a large continental shelf system under the influence of both a river plume
17 and coastal upwelling, *Limnol. Oceanogr.*, 57, 486-502, doi:10.4319/lo.2012.57.2.0486, 2012.

18 He, B., Dai, M., Huang, W., Liu, Q., Chen, H., and Xu, L.: Sources and accumulation of organic carbon
19 in the Pearl River Estuary surface sediment as indicated by elemental, stable carbon isotopic, and
20 carbohydrate compositions, *Biogeosciences*, 7, 3343-3362, doi:10.5194/bg-7-3343-2010, 2010a.

21 He, B., Dai, M., Zhai, W., Wang, L., Wang, K., Chen, J., Lin, J., Han, A., and Xu, Y.: Distribution,
22 degradation and dynamics of dissolved organic carbon and its major compound classes in the Pearl
23 River estuary, China, *Mar. Chem.*, 119, 52-64, doi:10.1016/j.marchem.2009.12.006, 2010b.

24 He, B., Dai, M., Zhai, W., Guo, X., and Wang, L.: Hypoxia in the upper reaches of the Pearl River
25 Estuary and its maintenance mechanisms: A synthesis based on multiple year observations during
26 2000–2008, *Mar. Chem.*, 167, 13-24, doi:10.1016/j.marchem.2014.07.003, 2014.

27 He, G.-f. and Yuan, G.-m.: Assessment of the water quality by fuzzy mathematics for last 20 years in
28 Zhujiang Estuary, *Mar. Environ. Sci.*, 26, 53-57, 2007.



1 Hu, J., Peng, P. a., Jia, G., Mai, B., and Zhang, G.: Distribution and sources of organic carbon, nitrogen
2 and their isotopes in sediments of the subtropical Pearl River estuary and adjacent shelf, Southern
3 China, *Mar. Chem.*, 98, 274-285, doi:10.1016/j.marchem.2005.03.008, 2006.

4 Huang, X., Huang, L., and Yue, W.: The characteristics of nutrients and eutrophication in the Pearl River
5 estuary, *South China Mar. Pollut. Bull.*, 47, 30-36, doi:10.1016/S0025-326X(02)00474-5, 2003.

6 Hullar, M., Fry, B., Peterson, B., and Wright, R.: Microbial utilization of estuarine dissolved organic
7 carbon: a stable isotope tracer approach tested by mass balance, *Appl. Environ. Microbiol.*, 62,
8 2489-2493, 1996.

9 Kao, S.-J., Terence Yang, J.-Y., Liu, K.-K., Dai, M., Chou, W.-C., Lin, H.-L., and Ren, H.: Isotope
10 constraints on particulate nitrogen source and dynamics in the upper water column of the oligotrophic
11 South China Sea, *Global Biogeochem. Cycles*, 26, GB2033, doi:10.1029/2011GB004091, 2012.

12 Kemp, W., Testa, J., Conley, D., Gilbert, D., and Hagy, J.: Temporal responses of coastal hypoxia to
13 nutrient loading and physical controls, *Biogeosciences*, 6, 2985-3008, doi:10.5194/bg-6-2985-2009,
14 2009.

15 Lehmann, M. F., Bernasconi, S. M., Barbieri, A., and McKenzie, J. A.: Preservation of organic matter
16 and alteration of its carbon and nitrogen isotope composition during simulated and in situ early
17 sedimentary diagenesis, *Geochim. Cosmochim. Acta*, 66, 3573-3584,
18 doi:10.1016/S0016-7037(02)00968-7, 2002.

19 Liu, M., Xiao, T., Wu, Y., Zhou, F., Huang, H., Bao, S., and Zhang, W.: Temporal distribution of
20 bacterial community structure in the Changjiang Estuary hypoxia area and the adjacent East China Sea,
21 *Environ. Res. Lett.*, 7, 025001, doi:10.1088/1748-9326/7/2/025001, 2012.

22 Marthur, J. M., Tyson, R. V., Thomson, J., and Matthey, D.: Early diagenesis of marine organic matter:
23 Alteration of the carbon isotopic composition, *Mar. Geol.*, 105, 51-61,
24 doi:10.1016/0025-3227(92)90181-G, 1992.

25 Mercedes, M. C. B., Luiz Antonio, M., Tibisay, P., Rafael, R., Jean Pierre, H. B. O., Felipe Siqueira, P.,
26 Silvia Rafaela Machado, L., and Sorena, M.: Nitrogen management challenges in major watersheds of
27 South America, *Environ. Res. Lett.*, 10, 065007, doi:10.1088/1748-9326/10/6/065007, 2015.



1 Meyers, P. A.: Organic geochemical proxies of paleoceanographic, paleolimnologic, and paleoclimatic
2 processes, *Org. Geochem.*, 27, 213-250, doi:10.1016/S0146-6380(97)00049-1, 1997.

3 Nixon, S. W.: Coastal marine eutrophication: A definition, social causes, and future concerns, *Ophelia*,
4 41, 199-219, doi:10.1080/00785236.1995.10422044, 1995.

5 Paerl, H. W.: Assessing and managing nutrient-enhanced eutrophication in estuarine and coastal waters:
6 Interactive effects of human and climatic perturbations, *Ecol. Eng.*, 26, 40-54,
7 doi:10.1016/j.ecoleng.2005.09.006, 2006.

8 Paerl, H. W.: Controlling Eutrophication along the Freshwater–Marine Continuum: Dual Nutrient (N and
9 P) Reductions are Essential, *Estuar. Coast.*, 32, 593-601, doi:10.1007/s12237-009-9158-8, 2009.

10 Peterson, B. J. and Fry, B.: Stable Isotopes in Ecosystem Studies, *Annu. Rev. Ecol. Syst.*, 18, 293-320,
11 1987.

12 Qian, W., Dai, M., Xu, M., Kao, S.-j., Du, C., Liu, J., Wang, H., Guo, L., and Wang, L.: Non-local drivers
13 of the summer hypoxia in the East China Sea off the Changjiang Estuary, *Estuar. Coast. Shelf Sci.*,
14 doi:10.1016/j.ecss.2016.08.032, 2016a.

15 Qian, W., Gan, J., Liu, J., He, B., Lu, Z., Guo, X., Wang, D., Guo, L., Huang, T., and Dai, M.: Current
16 status of emerging hypoxia in a large eutrophic estuary: the lower reach of Pearl River estuary, China,
17 Submitted to *Limnol. Oceanogr.*, 2016b.

18 Rabalais, N., Cai, W.-J., Carstensen, J., Conley, D., Fry, B., Hu, X., Quiñones-Rivera, Z., Rosenberg, R.,
19 Slomp, C., Turner, E., Voss, M., Wissel, B., and Zhang, J.: Eutrophication-Driven Deoxygenation in
20 the Coastal Ocean, *Oceanography*, 27, 172-183, doi:10.5670/oceanog.2014.21, 2014.

21 Rabalais, N. N., Díaz, R. J., Levin, L. A., Turner, R. E., Gilbert, D., and Zhang, J.: Dynamics and
22 distribution of natural and human-caused hypoxia, *Biogeosciences*, 7, 585-619,
23 doi:10.5194/bg-7-585-2010, 2010.

24 Rabouille, C., Conley, D. J., Dai, M. H., Cai, W. J., Chen, C. T. A., Lansard, B., Green, R., Yin, K.,
25 Harrison, P. J., Dagg, M., and McKee, B.: Comparison of hypoxia among four river-dominated ocean
26 margins: The Changjiang (Yangtze), Mississippi, Pearl, and Rhône rivers, *Cont. Shelf Res.*, 28,
27 1527-1537, doi:10.1016/j.csr.2008.01.020, 2008.



1 Rutger, R., Stefan, A., Birthe, H., Hans, C. N., and Karl, N.: Recovery of marine benthic habitats and
2 fauna in a Swedish fjord following improved oxygen conditions, *Mar. Ecol. Prog. Ser.*, 234, 43-53,
3 doi:10.3354/meps234043, 2002.

4 Shultz, D. J. and Calder, J. A.: Organic carbon $^{13}\text{C}/^{12}\text{C}$ variations in estuarine sediments, *Geochim.*
5 *Cosmochim. Acta*, 40, 381-385, doi:10.1016/0016-7037(76)90002-8, 1976.

6 Steckbauer, A., Duarte, C. M., Carstensen, J., Vaquer-Sunyer, R., and Conley, D. J.: Ecosystem impacts
7 of hypoxia: thresholds of hypoxia and pathways to recovery, *Environ. Res. Lett.*, 6, 025003,
8 doi:10.1088/1748-9326/6/2/025003, 2011.

9 Stefan, R., Mateete, B., Clare, M. H., Nancy, K., Wilfried, W., Xiaoyuan, Y., Albert, B., and Mark, A. S.:
10 Synthesis and review: Tackling the nitrogen management challenge: from global to local scales,
11 *Environ. Res. Lett.*, 11, 120205, doi:10.1088/1748-9326/11/12/120205, 2016.

12 Strong, D. J., Flecker, R., Valdes, P. J., Wilkinson, I. P., Rees, J. G., Zong, Y. Q., Lloyd, J. M., Garrett,
13 E., and Pancost, R. D.: Organic matter distribution in the modern sediments of the Pearl River Estuary,
14 *Org. Geochem.*, 49, 68-82, doi:10.1016/j.orggeochem.2012.04.011, 2012.

15 Swarzenski, P., Campbell, P., Osterman, L., and Poore, R.: A 1000-year sediment record of recurring
16 hypoxia off the Mississippi River: The potential role of terrestrially-derived organic matter inputs,
17 *Mar. Chem.*, 109, 130-142, doi:10.1016/j.marchem.2008.01.003, 2008.

18 Vaquer-Sunyer, R. and Duarte, C. M.: Thresholds of hypoxia for marine biodiversity, *Proc. Natl. Acad.*
19 *Sci. U.S.A.*, 105, 15452-15457, doi:10.1073/pnas.0803833105, 2008.

20 Wang, H., Dai, M., Liu, J., Kao, S.-J., Zhang, C., Cai, W.-J., Wang, G., Qian, W., Zhao, M., and Sun, Z.:
21 Eutrophication-Driven Hypoxia in the East China Sea off the Changjiang Estuary, *Environ. Sci.*
22 *Technol.*, 50, 2255-2263, doi:10.1021/acs.est.5b06211, 2016.

23 Wang, Q., Koshikawa, H., Liu, C., and Otsubo, K.: 30-year changes in the nitrogen inputs to the Yangtze
24 River Basin, *Environ. Res. Lett.*, 9, 115005, doi:10.1088/1748-9326/9/11/115005, 2014.

25 Ye, F., Huang, X., Zhang, X., Zhang, D., Zeng, Y., and Tian, L.: Recent oxygen depletion in the Pearl
26 River Estuary, South China: geochemical and microfaunal evidence, *J. Oceanogr.*, 68, 387-400,
27 doi:10.1007/s10872-012-0104-1, 2012.



1 Yu, F., Zong, Y., Lloyd, J. M., Huang, G., Leng, M. J., Kendrick, C., Lamb, A. L., and Yim, W. W.-S.:
2 Bulk organic $\delta^{13}\text{C}$ and C/N as indicators for sediment sources in the Pearl River delta and estuary,
3 southern China, *Estuar. Coast. Shelf Sci.*, 87, 618-630, doi:10.1016/j.ecss.2010.02.018, 2010.

4 Zhang, J., Zhang, Z. F., Liu, S. M., Wu, Y., Xiong, H., and Chen, H. T.: Human impacts on the large
5 world rivers: Would the Changjiang (Yangtze River) be an illustration?, *Global Biogeochem. Cycles*,
6 13, 1099-1105, doi:10.1029/1999GB900044, 1999.

7 Zhang, J., Cowie, G., and Naqvi, S. W. A.: Hypoxia in the changing marine environment, *Environ. Res.*
8 *Lett.*, 8, 015025, doi:10.1088/1748-9326/8/1/015025, 2013.

9 Zhang, Y., Xiao, W., and Jiao, N.: Linking biochemical properties of particles to particle-attached and
10 free-living bacterial community structure along the particle density gradient from freshwater to open
11 ocean, *J. Geophys. Res. Biogeosci.*, 121, 2261-2274, doi:10.1002/2016JG003390, 2016.

12 Zhu, C., Wagner, T., Pan, J.-M., and Pancost, R. D.: Multiple sources and extensive degradation of
13 terrestrial sedimentary organic matter across an energetic, wide continental shelf, *Geochem. Geophys.*
14 *Geosyst.*, 12, Q08011, doi:10.1029/2011GC003506, 2011.

15 Zimmerman, A. R. and Canuel, E. A.: A geochemical record of eutrophication and anoxia in Chesapeake
16 Bay sediments: anthropogenic influence on organic matter composition, *Mar. Chem.*, 69, 117-137,
17 doi:10.1016/S0304-4203(99)00100-0, 2000.

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1 **Table 1.** Summary of the end-member values and their uncertainties adopted in the three end-member mixing
 2 model.

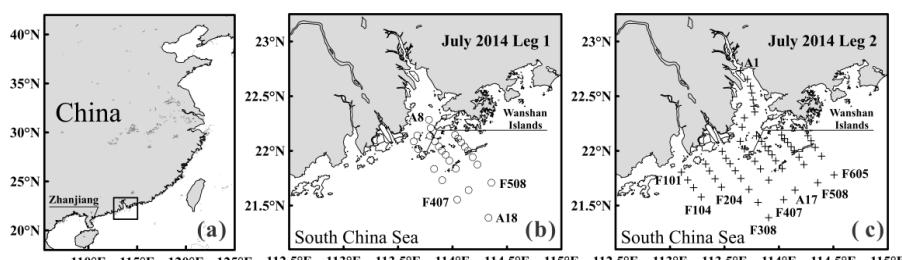
Water Mass	θ (°C)	Salinity	DIC ($\mu\text{mol kg}^{-1}$)	$\delta^{13}\text{C}_{\text{DIC}}$ (‰)	Ca^{2+} ($\mu\text{mol kg}^{-1}$)
Plume	30.6 ± 1.0	10.8	$1670 \pm 50^{\text{a}}$	-7.0 ^b	3620 ^c
Surface	31.0 ± 1.0	33.7 ± 0.2	1917 ± 3	0.6 ± 0.2	9778 ^c
Subsurface	21.8 ± 1.0	34.6 ± 0.1	2023 ± 6	0.1 ± 0.1	10053

3 ^aIn order to derive a proper plume end-member value, we took advantage of a 3 years' surface dataset in summer
 4 cruises (see Sect. 4.1). For DIC, the data is from cruises in August 2012, July 2014 and July 2015.

5 ^bThe $\delta^{13}\text{C}_{\text{DIC}}$ value was 0.6 ± 0.2 ‰ in the off-shore surface seawater at $S \sim 33.7$, and -11.4 ± 0.2 ‰ at $S < 0.4$ near the
 6 freshwater source in this cruise. Assuming the plume water is a mixture between freshwater and the off-shore surface
 7 seawater, the $\delta^{13}\text{C}_{\text{DIC}}$ of plume water end-member ($S=10.8$) can be calculated via the isotope mass balance, which
 8 equals -7.0 ‰.

9 ^cThe Ca^{2+} values of the plume and surface seawater EM are derived from a conservative mixing calculation (Ca^{2+} vs.
 10 S) based on the cruise data in August 2012.

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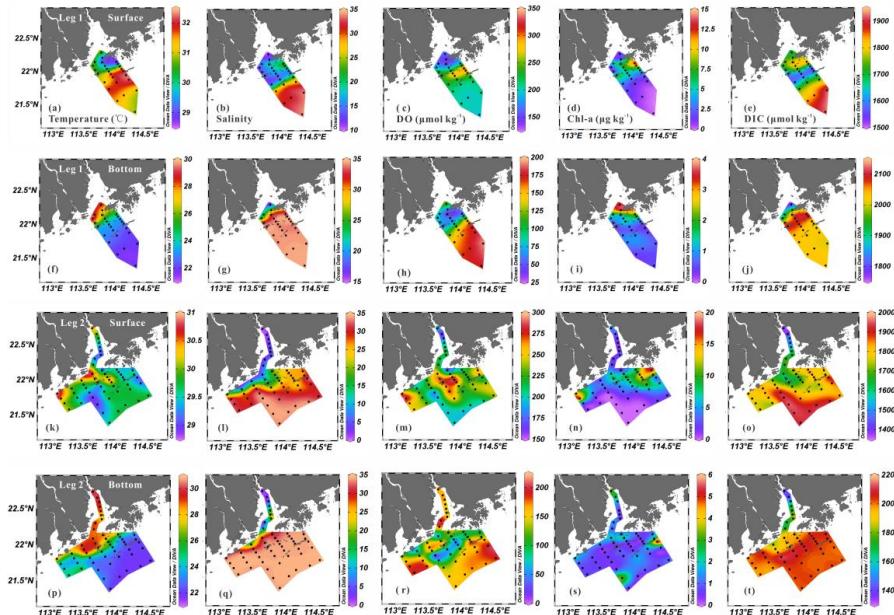


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 14 **Figure 1.** Map of the Pearl River Estuary and the adjacent coastal water (a). The open circles denote the Leg 1
 15 stations on 13–16 July 2014 (b) and the crosses represent the Leg 2 stations on 19–27 July 2014 (c).

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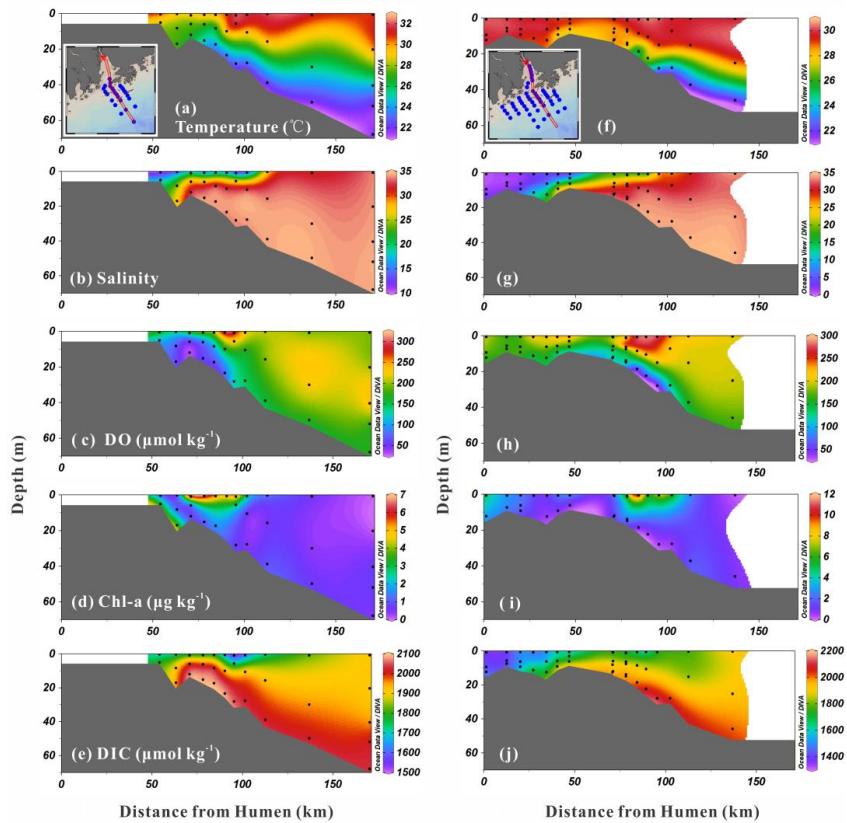
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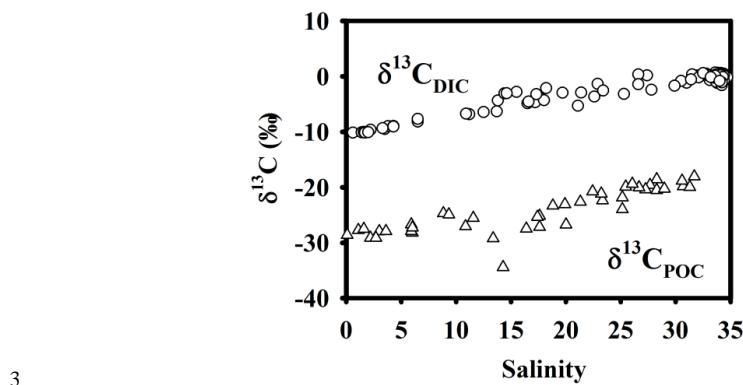
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3 **Figure 2.** Surface and bottom distributions of temperature, salinity, DO, Chl-a and DIC belonging to Leg 1 (a–e, f–
4 j) and Leg 2 (k–o, p–t).

5



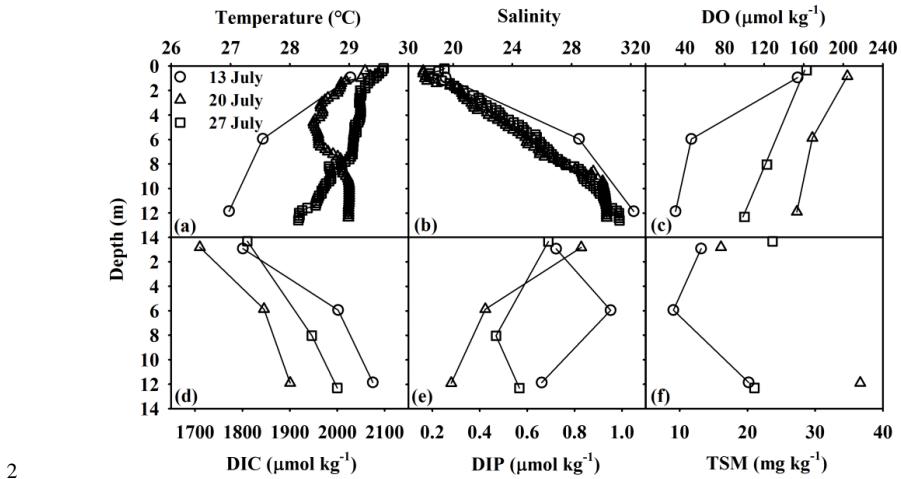
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 2 **Figure 3.** Profiles of temperature, salinity, DO, Chl-a and DIC along Transect A in Leg 1 (a–e) and Leg 2 (f–j).



3
 4 **Figure 4.** Distribution of $\delta^{13}\text{C}_{\text{DIC}}$ and $\delta^{13}\text{C}_{\text{POC}}$ along with salinity in the PRE and the adjacent coastal water. The
 5 open circles denote the $\delta^{13}\text{C}_{\text{DIC}}$ data in July 2014, while the open triangles represent the $\delta^{13}\text{C}_{\text{POC}}$ in July 2015.



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3 **Figure 5.** Profiles of (a) temperature, (b) salinity, (c) DO, (d) DIC, (e) DIP, (f) TSM and their evolution during
4 repeated sampling at Station A10.

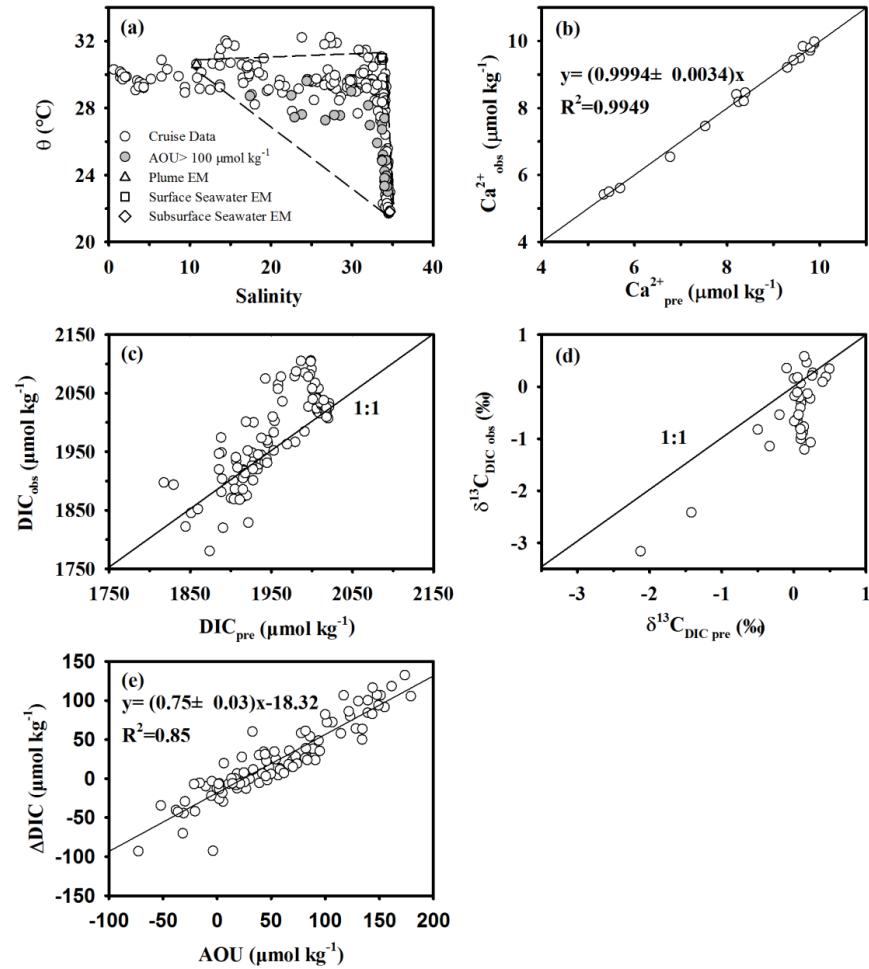
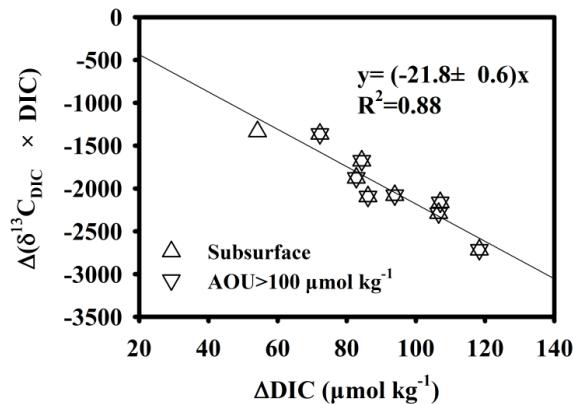


Figure 6. (a) The potential temperature (θ) vs. salinity scheme in the PRE and adjacent coastal water (open circles) based on the data collected in the July 2014 cruise. The grey filled circles show the low oxygen sites, where $\text{AOU} > 100 \mu\text{mol kg}^{-1}$. The three end-members, including plume water (open triangle), off-shore surface seawater (open square) and subsurface seawater (open diamond) are also displayed; (b) Relationships between the field-observed Ca^{2+} ($\text{Ca}^{2+}_{\text{obs}}$) and model-predicted Ca^{2+} ($\text{Ca}^{2+}_{\text{pre}}$). The straight line denotes a linear regression line; (c), (d) Relationships between the observed DIC, $\delta^{13}\text{C}_{\text{DIC}}$ and model-predicted DIC, $\delta^{13}\text{C}_{\text{DIC}}$. The straight lines act as a 1:1 reference line; and (e) Relationships of ΔDIC vs. AOU for all the data belonging to the subsurface water. ΔDIC is the difference between the field-observed DIC and model-predicted DIC.

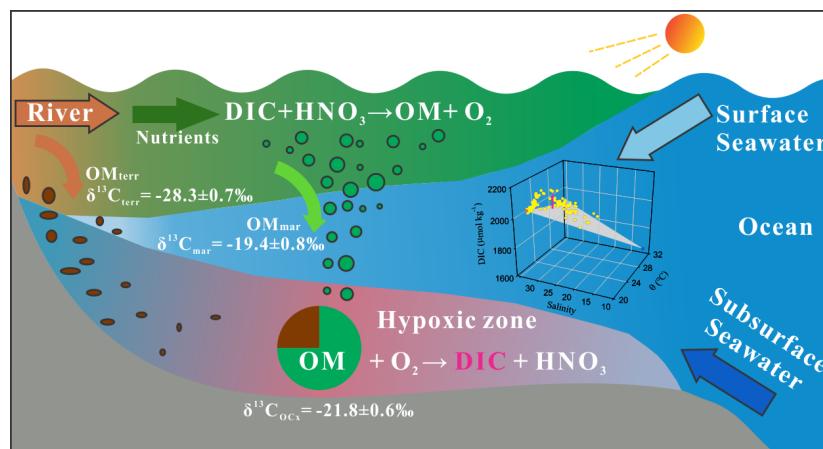
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 2 **Figure 7.** Plot of $\Delta (\delta^{13}\text{C}_{\text{DIC}} \times \text{DIC})$ vs. ΔDIC for samples collected in the subsurface water (>5 m, up triangles). The
 3 down triangles represent the samples with $\text{AOU} > 100 \mu\text{mol kg}^{-1}$. Δ is the difference between the field-observed and
 4 model-predicted values.

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 8 **Figure 8.** A conceptual diagram illustrating the source partitioning of oxygen-consuming organic matter (OC_{mar} vs.
 9 OC_{terr}) within the hypoxic zone in the lower PRE and the adjacent coastal area. See Sect. 4.2 for explanations.

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