1 **1. TITLE PAGE**

Seasonal variability in methane and nitrous oxide fluxes from tropical peatlands in the
 Western Amazon basin

- ⁵ Teh, Yit Arn^{1*}, Murphy, Wayne A.², Berrio, Juan-Carlos², Boom, Arnound², and Page, Susan E.²
- ⁶ ¹ Institute of Biological and Environmental Sciences, University of Aberdeen
- 7 ² Department of Geography, University of Leicester
- 8 * Author to whom all correspondence should be addressed; email: <u>yateh@abdn.ac.uk</u>

9 2. ABSTRACT

The Amazon plays a critical role in global atmospheric budgets of methane (CH₄) and nitrous 10 oxide (N₂O). However, while we have a relatively good understanding of the continental-scale 11 flux of these greenhouse gases (GHGs), one of the key gaps in knowledge is the specific 12 contribution of peatland ecosystems to the regional budgets of these GHGs. Here we report 13 CH₄ and N₂O fluxes from lowland tropical peatlands in the Pastaza-Marañón foreland basin 14 (PMFB) in Peru, one of the largest peatland complexes in the Amazon basin. The goal of this 15 research was to: quantify the range and magnitude of CH₄ and N₂O fluxes from this region; 16 assess seasonal trends in trace gas exchange; and determine the role of different 17 18 environmental variables in driving GHG flux. Trace gas fluxes were determined from the most numerically-dominant peatland vegetation types in the region: forested vegetation, forested 19 20 (short pole) vegetation, Mauritia flexuosa-dominated palm swamp, and mixed palm swamp. Data were collected in both wet and dry seasons over the course of four field campaigns from 21 2012 to 2014. Diffusive CH₄ emissions averaged 36.05 \pm 3.09 mg CH₄-C m⁻² d⁻¹ across the 22 entire dataset, with diffusive CH₄ flux varying significantly among vegetation types and 23 between seasons. Net ebullition of CH₄ averaged 973.3 \pm 161.4 mg CH₄-C m⁻² d⁻¹, and did not 24 vary significantly among vegetation types nor between seasons. Diffusive CH₄ flux was 25 greatest for mixed palm swamp (52.0 \pm 16.0 mg CH₄-C m⁻² d⁻¹), followed by *M. flexuosa* palm 26 swamp (36.7 \pm 3.9 mg CH₄-Cm⁻² d⁻¹), forested (short pole) vegetation (31.6 \pm 6.6 mg CH₄-Cm⁻² 27 d^{-1}), and forested vegetation (29.8 ± 10.0 mg CH₄-C m⁻² d⁻¹). Diffusive CH₄ flux also showed 28 marked seasonality, with divergent seasonal patterns among ecosystems. Forested 29 30 vegetation and mixed palm swamp showed significantly higher dry season (47.2 ± 5.4 mg CH₄-C m^{$^{-2}$} d^{$^{-1}$} and 85.5 ± 26.4 mg CH₄-C m^{$^{-2}$} d^{$^{-1}$}, respectively) compared to wet season emissions 31

32	(6.8 ± 1.0 mg CH ₄ -C m ⁻² d ⁻¹ and 5.2 ± 2.7 mg CH ₄ -C m ⁻² d ⁻¹ , respectively). In contrast, forested
33	(short pole) vegetation and <i>M. flexuosa</i> palm swamp showed the opposite trend, with dry
34	season flux of 9.6 \pm 2.6 and 25.5 \pm 2.9 mg CH ₄ -C m ⁻² d ⁻¹ , respectively, versus wet season flux
35	of 103.4 \pm 13.6 and 53.4 \pm 9.8 mg CH ₄ -C m ⁻² d ⁻¹ , respectively. These divergent seasonal trends
36	may be linked to very high water tables (>1 m) in forested vegetation and mixed palm swamp
37	during the wet season, which may have constrained CH_4 transport across the soil-atmosphere
38	interface. Diffusive N_2O flux was very low (0.70 \pm 0.34 μg $N_2O\text{-}N$ $m^{-2}d^{-1}$), and did not vary
39	significantly among ecosystems nor between seasons. We conclude that peatlands in the
40	PMFB are large and regionally significant sources of atmospheric CH ₄ , that need to be better
41	accounted for in regional emissions inventories. In contrast, N_2O flux was negligible,
42	suggesting that this region does not make a significant contribution to regional atmospheric
43	budgets of N_2O . The divergent seasonal pattern in CH_4 flux among vegetation types challenges
44	our underlying assumptions of the controls on CH_4 flux in tropical peatlands, and emphasizes
45	the need for more process-based measurements during high water table periods.

KEYWORDS

49 methane, nitrous oxide, peat, tropical peatland, Amazonia, Peru

53 **3. INTRODUCTION**

The Amazon basin plays a critical role in the global atmospheric budgets of carbon (C) and 54 greenhouse gases (GHGs) such as methane (CH₄) and nitrous oxide (N₂O). Recent basin-wide 55 studies suggest that the Amazon as a whole accounts for approximately 7 % of global 56 atmospheric CH₄ emissions (Wilson et al., 2016). N₂O emissions are of a similar magnitude, 57 with emissions ranging from 2-3 Tg $N_2 O\text{-}N$ year $^{\text{-}1}$ (or, approximately 12-18 % of global 58 atmospheric emissions) (Huang et al., 2008;Saikawa et al., 2014;Saikawa et al., 2013). While 59 we have a relatively strong understanding of the role that the Amazon plays in regional and 60 global atmospheric budgets of these gases, one of the key gaps in knowledge is the 61 62 contribution of specific ecosystem types to regional fluxes of GHGs (Huang et al., 2008;Saikawa et al., 2014;Saikawa et al., 2013). In particular, our understanding of the 63 64 contribution of Amazonian wetlands to regional C and GHG budgets is weak, as the majority 65 of past ecosystem-scale studies have focused on terra firme forests and savannas (D'Amelio et al., 2009;Saikawa et al., 2013;Wilson et al., 2016;Kirschke et al., 2013;Nisbet et al., 2014). 66 Empirical studies of GHG fluxes from Amazonian wetlands are more limited in geographic 67 scope and have focused on three major areas: wetlands in the state of Amazonas near the 68 city of Manaus (Devol et al., 1990;Bartlett et al., 1990;Bartlett et al., 1988;Keller et al., 1986), 69 70 the Pantanal region (Melack et al., 2004; Marani and Alvalá, 2007; Liengaard et al., 2013), and 71 the Orinoco River basin (Smith et al., 2000;Lavelle et al., 2014). Critically, none of the ecosystems sampled in the past were peat-forming ones; rather, the habitats investigated 72 73 were non-peat forming (i.e. mineral or organo-mineral soils), seasonally-inundated floodplain 74 forests (i.e. varzea), rivers or lakes.

Peatlands are one of the major wetland habitats absent from current bottom-up GHG 76 77 inventories for the Amazon basin, and are often grouped together with non-peat forming 78 wetlands in regional atmospheric budgets (Wilson et al., 2016). Unlike their Southeast Asian 79 counterparts, most peatlands in the Amazon basin are unaffected by human activity at the 80 current time (Lahteenoja et al., 2009a; Lahateenoja et al. 2009b; Lahteenoja and Page 2011), except for ecosystems in the Madre de Dios region in southeastern Peru, which are impacted 81 82 by gold mining (Householder et al., 2012). Because we have little or no data on ecosystem-83 level land-atmosphere fluxes from Amazonian peatlands (Lahteenoja et al., 2012;Lahteenoja 84 et al., 2009b;Kirschke et al., 2013;Nisbet et al., 2014), it is difficult to ascertain if rates of GHG flux from these ecosystems are similar to or different from mineral soil wetlands (e.g. varzea). 85 Given that underlying differences in plant community composition and soil properties are 86 known to modulate the cycling and flux of GHGs in wetlands (Limpens et al., 2008;Melton et 87 al., 2013; Belyea and Baird, 2006; Sjögersten et al., 2014), expanding our observations to 88 include a wider range of wetland habitats is critical in order to improve our understanding of 89 90 regional trace gas exchange, and also to determine if aggregating peat and mineral soil wetlands together in bottom-up emissions inventories are appropriate for regional budget 91 calculations. Moreover, Amazonian peatlands are thought to account for a substantial land 92 area (i.e. up to 150,000 km²) (Schulman et al., 1999;Lahteenoja et al., 2012), and any 93 differences in biogeochemistry among peat and mineral/organo-mineral soil wetlands may 94 95 therefore have important implications for understanding and modelling the biogeochemical functioning of the Amazon basin as a whole. 96

98 Since the identification of extensive peat forming wetlands in the north (Lahteenoja et al., 99 2009a; Lahateenoja et al. 2009b; Lahteenoja and Page 2011) and south (Householder et al., 100 2012) of the Peruvian Amazon, several studies have been undertaken to better characterize 101 these habitats, investigating vegetation composition and habitat diversity (Draper et al., 2014; 102 Kelly et al., 2014; Householder et al., 2012; Lahteenoja and Page, 2011), vegetation history 103 (Lahteenoja and Roucoux et al., 2010), C stocks (Lahteenoja et al., 2012; Draper et al., 2014), 104 hydrology (Kelly et al., 2014), and peat chemistry (Lahteenoja et al., 2009a; Lahteenoja et al., 2009b). Most of the studies have focused on the Pastaza-Marañón foreland basin (PMFB), 105 106 where one of the largest stretches of contiguous peatlands have been found (Lahteenoja et al 2009a; Lahteenoja and Page, 2011; Kelly et al, 2014), covering an estimated area of 35,600 107 \pm 2,133 km² (Draper et al., 2014). Up to 90% of the peatlands in the PMFB lie in flooded 108 backwater river margins on floodplains and are influenced by large, annual fluctuations in 109 water table caused by the Amazonian flood pulse (Householder et al., 2012;Lahteenoja et al., 110 111 2009a). These floodplain systems are dominated by peat deposits that range in depth from 112 ~3.9 m (Lahteenoja et al., 2009a) to ~12.9 m (Householder et al., 2012). The remaining 10% of these peatlands are not directly influenced by river flow and form domed (i.e. raised) 113 nutrient-poor bogs that likely only receive water and nutrients from rainfall (Lahteenoja et 114 al., 2009b). These nutrient-poor bogs are dominated by large, C-rich forests (termed "pole 115 116 forests"), that represent a very high density C store (total pool size of 1391 \pm 710 Mg C ha⁻¹, 117 which includes both above- and belowground stocks); exceeding in fact the C density of nearby floodplain systems (Draper et al., 2014). Even though the peats in these nutrient-poor 118 bogs have a relatively high hydraulic conductivity, they act as natural stores of water because 119 of high rainwater inputs (>3000 mm per annum), which help to maintain high water tables, 120 121 even during parts of the dry season (Kelly et al., 2014).

CH₄ flux in tropical soils are regulated by the complex interplay among multiple factors that 123 124 regulate CH₄ production, oxidation, and transport. Key factors include: redox/water table depth (Couwenberg et al., 2010;Couwenberg et al., 2011;Silver et al., 1999;Teh et al., 125 2005; von Fischer and Hedin, 2007), plant productivity (von Fischer and Hedin, 2007; Whiting 126 and Chanton, 1993), soil organic matter lability (Wright et al., 2011), competition for C 127 substrates among anaerobes (Teh et al., 2008;Teh and Silver, 2006;von Fischer and Hedin, 128 2007), and presence of plants capable of facilitating atmospheric egress (Pangala et al., 2013). 129 Of all these factors, fluctuation in soil redox conditions, as mediated by variations in water 130 131 table depth, is perhaps most critical in regulating CH₄ dynamics (Couwenberg et al., 2010;Couwenberg et al., 2011), because of the underlying physiology of the microbes that 132 produce and consume CH₄. Methanogenic archaea are obligate anaerobes that only produce 133 134 CH₄ under anoxic conditions (Conrad, 1996); as a consequence, they are only active in stably anoxic soil microsites or soil layers, where they are protected from the effects of strong 135 136 oxidants such as oxygen or where competition for reducing equivalents (e.g. acetate, H₂) from other anaerobic microorganisms is eliminated (Teh et al., 2008;Teh and Silver, 2006;Teh et 137 al., 2005;von Fischer and Hedin, 2002;von Fischer and Hedin, 2007). CH₄ oxidation, on the 138 139 other hand, is thought to be driven primarily by aerobic methanotrophic bacteria in tropical soils (Hanson and Hanson, 1996; Teh et al., 2005; Teh et al., 2006; von Fischer and Hedin, 140 2002;von Fischer and Hedin, 2007), with anaerobic CH₄ oxidation playing a quantitatively 141 smaller role (Blazewicz et al., 2012). Thus, fluctuations in redox or water table depth play a 142 fundamental role in directing the flow of C among different anaerobic pathways (Teh et al., 143 2008; Teh and Silver, 2006; von Fischer and Hedin, 2007), and shifting the balance between 144

production and consumption of CH₄ (Teh et al., 2005;von Fischer and Hedin, 2002). Moreover,
water table or soil moisture fluctuations are also thought to profoundly influence CH₄
transport dynamics throughout the soil profile, changing the relative partitioning of CH₄
among different transport pathways such as diffusion, ebullition, and plant-facilitated
transport (Whalen, 2005;Jungkunst and Fiedler, 2007).

150

151 Controls on N₂O flux are also highly complex (Groffman et al., 2009), with N₂O originating from as many as four separate sources (e.g. bacterial ammonia oxidation, archaeal ammonia 152 153 oxidation, denitrification, dissimilatory nitrate reduction to ammonium), each with different environmental controls (Baggs, 2008; Morley and Baggs, 2010; Firestone and Davidson, 154 155 1989; Firestone et al., 1980; Pett-Ridge et al., 2013; Silver et al., 2001; Prosser and Nicol, 2008). 156 Key factors regulating soil N₂O flux include: redox, soil moisture content or water table depth, 157 temperature, pH, labile C availability, and labile N availability (Groffman et al., 2009). As is the case for CH₄, variations in redox/water table depth plays an especially prominent role in 158 regulating N₂O flux in tropical peatland ecosystems, because all of the processes that produce 159 160 N₂O are redox-sensitive, with bacterial or archaeal ammonia oxidation occurring under 161 aerobic conditions (Prosser and Nicol, 2008; Firestone and Davidson, 1989; Firestone et al., 162 1980) whereas nitrate-reducing processes (i.e. denitrification, dissimilatory nitrate reduction to ammonium) are anaerobic ones (Firestone and Davidson, 1989;Firestone et al., 163 1980; Morley and Baggs, 2010; Silver et al., 2001). Moreover, for nitrate reducing processes, 164 which are believed to be the dominant source of N₂O in wet systems, the extent of 165 166 anaerobiosis also controls the relative proportion of N₂O or N₂ produced during dissimilatory

167 metabolism (Firestone and Davidson, 1989;Firestone et al., 1980;Morley and Baggs,
168 2010;Silver et al., 2001).

169

170	In order to improve our understanding of the biogeochemistry and rates of GHG exchange
171	from Amazonian peatlands, we conducted a preliminary study of CH_4 and N_2O fluxes from
172	forested peatlands in the PMFB. The main objectives of this are to:

- 173 1. Quantify the magnitude and range of soil CH_4 and N_2O fluxes from a sub-set of
- 174 peatlands in the PMFB that represent dominant vegetation types
- 175 2. Determine seasonal patterns of trace gas exchange
- 176 3. Establish the relationship between trace gas fluxes and environmental variables

Sampling was concentrated on the four most dominant vegetation types in the area, based on prior work by the investigators (Lahteenoja and Page, 2011). Trace gas fluxes were captured from both floodplain systems and nutrient-poor bogs in order to account for underlying differences in biogeochemistry that may arise from variations in hydrology. Sampling was conducted during four field campaigns (two wet season, two dry season) over a 27-month period, extending from February 2012 to May 2014.

183

184

185 4. MATERIALS AND METHODS

186 **4.1 Study site and sampling design**

187 The study was carried out in the lowland tropical peatland forests of the PMFB, between 2 188 and 35 km south of the city of Iquitos, Peru (Lahteenoja et al., 2009a; Lahteenoja et al., 2009b) 189 (Figure 1, Table 1). The mean annual temperature is 26 °C, annual precipitation is c. 3,100 mm, relative humidity ranges from 80-90 %, and altitude ranges from c. 90 to 130 m above 190 191 sea level (Marengo 1998). The northwestern Amazon basin near Iquitos experiences 192 pronounced seasonality, which is characterized by consistently high annual temperatures, but 193 marked seasonal variation in precipitation (Tian et al., 1998), and an annual river flood pulse 194 linked to seasonal discharge from the Andes (Junk et al., 1989). Precipitation events are 195 frequent, intense and of significant duration during the wet season (November to May) and infrequent, intense and of short duration during the dry season (June to August). September 196 197 and October represent a transitional period between dry and wet seasons, where rainfall patterns are less predictable. Catchments in this region receive no less than 100 mm of rain 198 per month (Espinoza Villar et al., 2009a; Espinoza Villar et al., 2009b) and >3000 mm of rain 199 200 per year. River discharge varies by season, with the lowest discharge between the dry season 201 months of August and September. Peak discharge from the wet season flood pulse occurs 202 between April and May, as recorded at the Tamshiyaku River gauging station (Espinoza Villar et al., 2009b). 203

204

Histosols form the dominant soil type for peatlands in this region (Andriesse, 1988;Lahteenoja
and Page, 2011). Study sites are broadly classified as nutrient-rich, intermediate, or nutrientpoor (Lahteenoja and Page, 2011), with pH ranging from 3.5 to 7.2 (Lahteenoja and Page,
2011;Lahteenoja et al., 2009a;Lahteenoja et al., 2009b). More specific data on pH for our plots
are presented in Table 3. Nutrient-rich (i.e. minerotrophic) sites tend to occur on floodplains

210 and river margins, and account for at least 60 % of the peatland cover in the PMFB (Lahteenoja 211 and Page, 2011; Draper et al., 2014). They receive water, sediment, and nutrient inputs from 212 the annual Amazon river flood pulse (Householder et al., 2012;Lahteenoja and Page, 2011), 213 leading to higher inorganic nutrient content, of which Ca and other base cations form major 214 constituents (Lahteenoja and Page, 2011). Many of the soils in these nutrient-rich areas are 215 fluvaquentic Tropofibrists (Andriesse, 1988), and contain thick mineral layers or minerogenic 216 intrusions, reflective of episodic sedimentation events in the past (Lahteenoja and Page, 217 2011). In contrast, nutrient-poor (i.e. oligotrophic) sites tend to occur further in-land 218 (Lahteenoja and Page, 2011;Draper et al., 2014). They are almost entirely rain-fed, and receive low or infrequent inputs of water and nutrients from streams and rivers (Lahteenoja 219 220 and Page, 2011). These ecosystems account for 10 to 40 % of peatland cover in the PMFB, though precise estimates vary depending on the land classification scheme employed 221 222 (Lahteenoja and Page, 2011; Draper et al., 2014). Soil Ca and base cation concentrations are significantly lower in these sites compared to nutrient-rich ones, with similar concentrations 223 224 to that of rainwater (Lahteenoja and Page, 2011). Soils are classified as typic or hydric 225 Tropofibrists (Andriesse, 1988). Even though Ca and base cations themselves play no direct 226 role in modulating CH₄ and N₂O fluxes, underlying differences in soil fertility may indirectly influence CH₄ and N₂O flux by influencing the rate of labile C input to the soil, the 227 decomposability of organic matter, and the overall throughput of C and nutrients through the 228 229 plant-soil system (Firestone and Davidson, 1989;Groffman et al., 2009;von Fischer and Hedin, 230 2007; Whiting and Chanton, 1993).

We established 239 sampling plots (~30 m² per plot) within five tropical peatland sites that 232 captured four of the dominant vegetation types in the region (Draper et al., 233 234 2014;Householder et al., 2012;Kelly et al., 2014;Lahteenoja and Page, 2011), and which 235 encompassed a range of nutrient availabilities (Figure 1, Table 1) (Lahteenoja and Page, 2011;Lahteenoja et al., 2009a). These four dominant vegetation types included: forested 236 237 vegetation (nutrient-rich; n= 21 plots), forested (short pole) vegetation (nutrient-poor; n= 47 238 plots), Mauritia flexuosa-dominated palm swamp (intermediate fertility, n= 153 plots), and 239 mixed palm swamp (nutrient-rich; n=18 plots) (Table 1). Four of the study sites (Buena Vista, 240 Charo, Miraflores, and Quistococha) were dominated by only one vegetation type, whereas San Jorge contained a mixture of *M. flexuosa* palm swamp and forested (short pole) 241 242 vegetation (Table 1). As a consequence, both vegetation types were sampled in San Jorge to develop a more representative picture of GHG fluxes from this location. Sampling efforts were 243 partially constrained by issues of site access; some locations were difficult to access (e.g. 244 245 centre of the San Jorge peatland) due to water table height and navigability of river channels; 246 as a consequence, sampling patterns were somewhat uneven, with higher sampling densities in some peatlands than in others (Table 1). 247

248

In each peatland site, transects were established from the edge of the peatland to its centre.
Each transect varied in length from 2 to 5 km, depending on the relative size of the peatland.
Randomly located sampling plots (~30 m² per plot) were established at 50 or 200 m intervals
along each transect, from which GHG fluxes and environmental variables were measured
concomitantly. The sampling interval (i.e. 50 or 200 m) was determined by the length of the
transect or size of the peatland, with shorter sampling intervals (50 m) for shorter transects

(i.e. smaller peatlands) and longer sampling intervals (200 m) for longer transects (i.e. largerpeatlands).

257

258 **4.2 Quantifying soil-atmosphere exchange**

259 Soil-atmosphere fluxes (CH₄, N₂O) were determined in four campaigns over a two-year annual water cycle: February 2012 (wet season), June-August 2012 (dry season), June-July 2013 (dry 260 261 season), and May-June 2014 (wet season). The duration of the campaign for each study site varied depending on its size. Each study site was generally sampled only once for each 262 263 campaign, except for a sub-set of plots within each vegetation type where diurnal studies 264 were conducted to determine if CH₄ and N₂O fluxes varied over daily time steps. Gas exchange was quantified using a floating static chamber approach (Livingston and Hutchinson, 1995; 265 Teh et al., 2011). Static flux measurements were made by enclosing a 0.225 m² area with a 266 dark, single component, vented 10 L flux chamber. No chamber bases (collars) were used due 267 268 to the highly saturated nature of the soils. In most cases, a standing water table was present 269 at the soil surface, so chambers were placed directly onto the water. In the absence of a standing water table, a weighted skirt was applied to create an airtight seal. Under these drier 270 271 conditions, chambers were placed carefully on the soil surface. In order to reduce the risk of 272 pressure-induced ebullition or disruption to soil gas concentration profiles caused by the investigators' footfall, flux chambers were lowered from a distance of 2-m away using a 2-m 273 274 long pole. Gas samples were collected with syringes using >2 m lengths of Tygon[®] tubing, 275 after thoroughly purging the dead volumes in the sample lines. To promote even mixing within the headspace, chambers were fitted with small computer fans (Pumpanen et al., 276 277 2004). Headspace samples were collected from each flux chamber at five intervals over a 25

278 minute enclosure period using a gas tight syringe. Gas samples were stored in evacuated 279 Exetainers[®] (Labco Ltd., Lampeter UK), shipped to the UK, and subsequently analysed for CH₄, 280 CO₂ and N₂O concentrations using Thermo TRACE GC Ultra (Thermo Fischer Scientific Inc., 281 Waltham, Massachusetts, USA) at the University of St. Andrews. Chromatographic separation was achieved using a Porapak-Q column, and gas concentrations determined using a flame 282 ionization detector (FID) for CH₄, a methanizer-FID for CO₂, and an electron capture detector 283 284 (ECD) for N₂O. Instrumental precision, determined from repeated analysis of standards, was < 5% for all detectors. 285

286

Diffusive fluxes were determined by using the JMP IN version 11 (SAS Institute, Inc., Cary, 287 288 North Carolina, USA) statistical package to plot best-fit lines to the data for headspace 289 concentration against time for individual flux chambers, with fluxes calculated from linear or 290 non-linear regressions depending on the individual concentration trend against time (Teh et al., 2014). Gas mixing ratios (ppm) were converted to areal fluxes by using the Ideal Gas Law 291 292 to solve for the quantity of gas in the headspace (on a mole or mass basis) and normalized by 293 the surface area of each static flux chamber (Livingston and Hutchinson, 1995). Ebullition-294 derived CH₄ fluxes were also quantified in our chambers where evidence of ebullition was 295 found. This evidence consisted of either: (i) rapid, non-linear increases in CH₄ concentration 296 over time; (ii) abrupt, stochastic increases in CH₄ concentration over time; or (iii) an abrupt stochastic increase in CH₄ concentration, followed by a linear decline in concentration. For 297 observations following pattern (i), flux was calculated by fitting a quadratic regression 298 equation to the data (P < 0.05), and CH₄ flux determined from the initial steep rise in CH₄ 299 300 concentration. For data following pattern (ii), the ebullition rate was determined by 301 calculating the total CH₄ production over the course of the bubble event, in-line with prior 302 work conducted by the investigators (Teh et al., 2011). Last, for data following pattern (iii), a 303 best-fit line was plotted to the CH₄ concentration data after the bubble event, and a net rate 304 of CH₄ uptake calculated from the gradient of the line. While observations (i) – (iii) all reflect 305 the effects of ebullition, only observations following patterns (i) and (ii) indicate net emission to the atmosphere, whereas observations following pattern (iii) indicate emission followed by 306 307 net uptake. As a consequence, patterns (i) and (ii) were categorized as "net ebullition" (i.e. net efflux) whereas observations following pattern (iii) were categorized as "ebullition-driven 308 309 CH₄ uptake" (i.e. net influx).

310

311 4.3 Environmental variables

To investigate the effects of environmental variables on trace gas fluxes, we determined air 312 temperature, soil temperature, chamber headspace temperature, soil pH, soil electrical 313 conductivity (EC; μ Scm⁻²), dissolved oxygen concentration of the soil pore water (DO; 314 315 measured as percent saturation, %) in the top 15 cm of the peat column, and water table position concomitant with gas sampling. Air temperature (measured 1.3 m above the soil) 316 and chamber headspace temperature were measured using a Checktemp® probe and meter 317 (Hanna Instruments LTD, Leighton Buzzard, UK). Peat temperature, pH, DO and EC were 318 measured at a depth of 15 cm below the peat surface and recorded in situ with each gas 319 sample using a HACH[®] rugged outdoor HQ30D multi meter and pH, LDO or EC probe. At sites 320 321 where the water level was above the peat surface, the water depth was measured using a meter rule. Where the water table was at or below the peat surface, the water level was 322

measured by auguring a hole to 1 m depth and measuring water table depth using a meterrule.

325

326 4.4 Statistical Analyses

Statistical analyses were performed using JMP IN version 11 (SAS Institute, Inc., Cary, North 327 Carolina, USA). Box-Cox transformations were applied where the data failed to meet the 328 assumptions of analysis of variance (ANOVA); otherwise, non-parametric tests were applied 329 330 (e.g. Wilcoxon signed-rank test). ANOVA and analysis of co-variance (ANCOVA) were used to 331 test for relationships between gas fluxes and vegetation type, season, and environmental variables. When determining the effect of vegetation type on gas flux, data from different 332 study sites (e.g. San Jorge and Miraflores) were pooled together. Means comparisons were 333 tested using a Fisher's Least Significant Difference (LSD) test. 334

335

336

337 **5. RESULTS**

5.1 Differences in gas fluxes and environmental variables among vegetation types

339	All vegetation types were net sources of CH_4 , with an overall mean (± standard error) diffusive
340	flux of 36.1 \pm 3.1 mg CH ₄ -C m $^{-2}$ d $^{-1}$ and a mean net ebullition flux of 973.3 \pm 161.4 mg CH ₄ -C
341	$m^{-2} d^{-1}$ (Figure 2, Table 2). We also saw examples of ebullition-driven CH ₄ uptake (i.e. a sudden
342	or stochastic increase in CH_4 concentration, followed immediately by a rapid linear decline in
343	concentration), with a mean rate of -504.1 \pm 84.4 mg CH ₄ -C m ⁻² d ⁻¹ (Table 2). Diffusive fluxes

of CH_4 accounted for the majority of observations (83.3 to 93.1 %), while ebullition fluxes accounted for a much smaller proportion of observations (6.9 to 16.7 %; Table 2).

346

347 Diffusive CH₄ flux varied significantly among the four vegetation types sampled in this study (two-way ANOVA with vegetation, season and their interaction, $F_{7, 979}$ = 13.2, P<0.0001; Fig. 348 2a). However, the effect of vegetation was relatively weak (see ANCOVA results in the section 349 'Relationships between gas fluxes and environmental variables'), and a means comparison 350 test on the pooled data was unable to determine which means differed significantly from the 351 others (Fisher's LSD, P > 0.05). For the pooled data, the overall numerical trend was that mixed 352 palm swamp showed the highest mean flux (52.0 ± 16.0 mg CH₄-C m⁻² d⁻¹), followed by M. 353 *flexuosa* palm swamp (36.7 \pm 3.9 mg CH₄-Cm⁻² d⁻¹), forested (short pole) vegetation (31.6 \pm 354 6.6 mg CH₄-Cm^{$^{-2}$} d^{$^{-1}$}), and forested vegetation (29.8 ± 10.0 mg CH₄-C m^{$^{-2}$} d^{$^{-1}$}). CH₄ ebullition 355 (i.e. net ebullition and ebullition-driven uptake) did not vary significant among vegetation 356 types nor between seasons (Table 2). Broadly speaking, however, we saw a greater frequency 357 of ebullition in the *M. flexuosa* palm swamp, followed by mixed palm swamp, forested 358 359 vegetation, and forested (short pole vegetation) (Table 2).

360

These study sites were also a weak net source of N₂O, with a mean diffusive flux of 0.70 ± 0.34 $\mu g N_2 O-N m^{-2} d^{-1}$. We saw only limited evidence of ebullition of N₂O, with only three chambers out of 1181 (0.3 % of observations) showing evidence of N₂O ebullition. These data were omitted from the analysis of diffusive flux of N₂O. Because of the high variance in diffusive N₂O flux among plots, analysis of variance indicated that mean diffusive N₂O flux did not differ significantly among vegetation types (two-way ANOVA, P > 0.5, Fig. 2b). However, when the N₂O flux data were grouped by vegetation type, we see that some vegetation types tended to function as net atmospheric sources, while others acted as atmospheric sinks (Fig. 2b, Table 3). For example, the highest N₂O emissions were observed from *M. flexuosa* palm swamp (1.11 ± 0.44 µg N₂O-N m⁻² d⁻¹) and forested vegetation (0.20 ± 0.95 µg N₂O-N m⁻² d⁻¹). In contrast, forested (short pole) vegetation and mixed palm swamp were weak sinks for N₂O, with a mean flux of -0.01 ± 0.84 and -0.21 ± 0.70 µg N₂O-N m⁻² d⁻¹, respectively.

373

Soil pH varied significantly among vegetation types (data pooled across all seasons; ANOVA, P < 0.0001, Table 3). Multiple comparisons tests indicated that mean soil pH was significantly different for each of the vegetation types (Fisher's LSD, P < 0.0001, Table 3), with the lowest pH in forested (short pole) vegetation (4.10 ± 0.04), followed by *M. flexuosa* palm swamp (5.32 ± 0.02), forested vegetation (6.15 ± 0.06), and the mixed palm swamp (6.58 ± 0.04).

379

Soil dissolved oxygen (DO) content varied significantly among vegetation types (data pooled across all seasons; Kruskal-Wallis, P < 0.0001, Table 3). Multiple comparisons tests indicated that mean DO was significantly different for each of the vegetation types (Fisher's LSD, P <0.05, Table 3), with the highest DO in the forested (short pole) vegetation (25.2 ± 2.1 %), followed by the *M. flexuosa* palm swamp (18.1 ± 1.0 %), forested vegetation (11.8 ± 2.8 %), and the mixed palm swamp (0.0 ± 0.0 %).

Electrical conductivity (EC) varied significantly among vegetation types (data pooled across all seasons; Kruskal-Wallis, P < 0.0001, Table 3). Multiple comparison tests indicated that mean EC was significantly different for each of the vegetation types (Fisher's LSD, P < 0.05; Table 3), with the highest EC in the mixed palm swamp (170.9 ± 6.0 µs m⁻²), followed by forested vegetation (77.1 ± 4.2 µs m⁻²), *M. flexuosa* palm swamp (49.7 ± 1.4 µs m⁻²) and the forested (short pole) vegetation (40.9 ± 3.5 µs m⁻²).

393

Soil temperature varied significantly among vegetation types (data pooled across all seasons; ANOVA, P < 0.0001, Table 3). Multiple comparisons tests indicated that soil temperature in forested (short pole) vegetation was significantly lower than in the other vegetation types (Table 3); whereas the other vegetation types did not differ in temperature amongst themselves (Fisher's LSD, P < 0.05, Table 3).

399

Air temperature varied significantly among vegetation types (data pooled across all seasons; ANOVA, P < 0.0001, Table 3). Multiple comparisons tests indicated that air temperature in M. *flexuosa* palm swamp was significantly lower than in the other vegetation types; whereas the other vegetation types did not differ in temperature amongst themselves (Fisher's LSD, P <0.05, Table 3).

405

Water table depths varied significantly among vegetation types (data pooled across all seasons; ANOVA, P < 0.0001, Table 3). The highest mean water tables were observed in mixed palm swamp (59.6 ± 9.3 cm), followed by forested vegetation (34.0 ± 6.9 cm), *M. flexuosa*

palm swamp (17.4 ± 1.2 cm), and forested (short pole) vegetation (3.5 ± 1.0 cm) (Fisher's LSD,
P < 0.0005).

411

412 **5.2** Temporal variations in gas fluxes and environmental variables

The peatlands sampled in this study showed pronounced seasonal variability in diffusive CH₄ flux (two-way ANOVA, $F_{7, 979} = 13.2$, P < 0.0001; Table 4). For ebullition of CH₄ and ebullitiondriven uptake of CH₄, mean fluxes varied between seasons, but high variability meant that these differences were not statistically significant ((two-way ANOVA, P > 0.8; Table 2). Diffusive N₂O flux showed no seasonal trends (two-way ANOVA, P > 0.5), and therefore will not be discussed further here. Diurnal studies suggest that neither diffusive fluxes of CH₄ nor N₂O varied over the course of a 24-hour period.

420

For diffusive CH₄ flux, the overall trend was towards significantly higher wet season (51.1 \pm 421 7.0 mg CH₄-C m^{$^{-2}$} d^{$^{-1}$}) compared to dry season (27.3 ± 2.7 mg CH₄-C m^{$^{-2}$} d^{$^{-1}$}) flux (data pooled 422 across all vegetation types; t-Test, P < 0.001, Table 4). However, when diffusive CH₄ flux was 423 424 disaggregated by vegetation type, very different seasonal trends emerged. For example, both forested vegetation and mixed palm swamp showed significantly greater diffusive CH₄ flux 425 during the *dry season* with net fluxes of 47.2 \pm 5.4 mg CH₄-C m⁻² d⁻¹ and 64.2 \pm 12.1 mg CH₄-426 C m^{$^{-2}$} d^{$^{-1}$}, respectively (Fisher's LSD, P < 0.05, Table 3). In contrast, wet season flux was 7-16 427 times lower, with net fluxes of 6.7 \pm 1.0 mg CH₄-C m⁻² d⁻¹ and 6.1 \pm 1.3 mg CH₄-C m⁻² d⁻¹, 428 429 respectively (Fisher's LSD, P < 0.05, Table 3). In contrast, forested (short pole) vegetation and *M. flexuosa* palm swamp showed seasonal trends consistent with the pooled data set; i.e. 430

significantly higher flux during the wet season (46.7 ± 8.4 and 60.4 ± 9.1 mg CH₄-C m⁻² d⁻¹, respectively) compared to the dry season (28.3 ± 2.6 and 18.8 ± 2.6 mg CH₄-C m⁻² d⁻¹, respectively) (Fisher's LSD, P < 0.05, Table 3).

434

Even though seasonal trends in CH₄ ebullition were not statistically significant, we will briefly 435 436 describe the overall patterns for the different vegetation types as they varied among ecosystems (Table 2). Forested vegetation only showed evidence of ebullition during the dry 437 season, where ebullition-driven uptake was observed. For forested (short pole) vegetation, 438 net ebullition was generally greater during the wet season, while ebullition-driven uptake was 439 higher during the dry season. For *M. flexuosa* palm swamp, both net ebullition and ebullition-440 441 driven uptake were greater during the wet season. Lastly, for mixed palm swamp, both net 442 ebullition and ebullition-driven uptake were greater during the dry season.

443

For the environmental variables, soil pH, DO, EC, water table depth, and soil temperature 444 varied significantly between seasons, whereas air temperature did not. Thus, for sake of 445 brevity, air temperature is not discussed further here. Mean soil pH was significantly lower 446 447 during the wet season (5.18 \pm 0.03) than during the dry season (5.31 \pm 0.04) (data pooled 448 across all vegetation types; t-Test, P < 0.05, Table 2). When disaggregated by vegetation type, 449 the overall trend was found to hold true for all vegetation types except forested (short pole) vegetation, which displayed higher pH during the wet season compared to the dry season 450 (Table 2). A two-way ANOVA on Box-Cox transformed data using vegetation type, season and 451 452 their interaction as explanatory variables indicated that vegetation type was the best 453 predictor of pH, with season and vegetation type by season playing a lesser role ($F_{7, 1166}$ = 454 348.9, P < 0.0001).

455

456 For DO, the overall trend was towards significantly lower DO during the wet season (13.9 \pm 1.0 %) compared to the dry season (19.3 ± 1.2 %) (data pooled across all vegetation types; 457 Wilcoxon test, P < 0.0001, Table 2). However, when the data were disaggregated by 458 vegetation type, we found that individual vegetation types showed distinct seasonal trends 459 from each other. Forested vegetation and mixed palm swamp were consistent with the 460 overall trend (i.e. lower wet season compared to dry season DO), whereas forested (short 461 pole) vegetation and *M. flexuosa* palm swamp displayed the reverse trend (i.e. higher wet 462 463 season compared to dry season DO) (Table 2). A two-way ANOVA on Box Cox transformed 464 data using vegetation type, season and their interaction as explanatory variables indicated 465 that vegetation type was the best predictor of DO, followed by a strong vegetation by season interaction; season itself played a lesser role than either of the other two explanatory 466 variables (*F*_{7, 1166} = 57.0, *P* < 0.0001). 467

468

For EC, the overall trend was towards lower EC in the wet season $(49.4 \pm 1.8 \ \mu s \ m^{-2})$ compared to the dry season $(65.5 \pm 2.2 \ \mu s \ m^{-2})$ (data pooled across all vegetation types; Wilcoxon test, P < 0.05, Table 2). When the data were disaggregated by vegetation type, this trend was consistent for all the vegetation types except for forested vegetation, where differences between wet and dry season were not statistically significant (Wilcoxon, P > 0.05, Table 2).

Water table depths varied significantly between seasons (data pooled across all vegetation 475 476 types; Wilcoxon test, *P* < 0.0001, Table 2). Mean water table level was significantly higher in 477 the wet $(54.1 \pm 2.7 \text{ cm})$ than the dry $(1.3 \pm 0.8 \text{ cm})$ season. When disaggregated by vegetation 478 type, the trend held true for individual vegetation types (Table 2). All vegetation types had negative dry season water tables (i.e. below the soil surface) and positive wet season water 479 tables (i.e. water table above the soil surface), except for *M. flexuosa* palm swamp that had 480 481 positive water tables in both seasons. Two-way ANOVA on Box-Cox transformed data using 482 vegetation type, season and their interaction as explanatory variables indicated that all three 483 factors explained water table depth, but that season accounted for the largest proportion of the variance in the model, followed by vegetation by season, and lastly by vegetation type (F_{7} . 484 $_{1157} = 440.1, P < 0.0001).$ 485

486

487 For soil temperature, the overall trend was towards slightly higher temperatures in the wet season (25.6 ± 0.0 °C) compared to the dry season (25.1 ± 0.0 °C) (t-Test, P < 0.0001). Analysis 488 489 of the disaggregated data indicates this trend was consistent for individual vegetation types 490 (Table 2). Two-way ANOVA on Box-Cox transformed data using vegetation type, season and 491 their interaction as explanatory variables indicated that all three variables played a significant 492 role in modulating soil temperature, although season accounted for the largest proportion of the variance whereas the other two factors accounted for a similar proportion of the variance 493 494 $(F_{7, 1166} = 21.3, P < 0.0001).$

495

496 **5.3 Relationships between gas fluxes and environmental variables**

To explore the relationships between environmental variables and diffusive gas fluxes, we conducted an analysis of covariance (ANCOVA) on Box-Cox transformed gas flux data, using vegetation type, season, vegetation by season, and environmental variables as explanatory variables. We did not analyze trends between ebullition and environmental variables because of the limitations in the sampling methodology and the limited number of observations.

502

503 For diffusive CH₄ flux, ANCOVA revealed that vegetation by season was the strongest predictor of CH₄ flux, followed by a strong season effect ($F_{13, 917}$ = 9.2, P<0.0001). Other 504 significant drivers included soil temperature, water table depth, and a borderline-significant 505 effect of vegetation type (P < 0.06). However, it is important to note that each of these 506 507 environmental variables were only weakly correlated with CH₄ flux even if the relationships were statistically significant; for example, when individual bivariate regressions were 508 calculated, the r^2 values were less than 0.01 for each plot (see Supplementary Online 509 510 Materials, Figures S1 and S2).

511

For diffusive N₂O flux, ANCOVA indicated that the best predictors of flux rates were dissolved oxygen and electrical conductivity ($F_{13, 1014}$ = 2.2, P < 0.0082). As was the case for CH₄, when the relationships between these environmental variables and N₂O flux were explored using individual bivariate regressions, r^2 values were found to be very low (e.g. less than $r^2 < 0.0007$) or not statistically significant (see Supplementary Online Materials, Figures S3 and S4).

517

6.1 Large and asynchronous CH₄ fluxes from peatlands in the Pastaza-Marañón foreland basin

The ecosystems sampled in this study were strong atmospheric sources of CH₄. Diffusive CH₄ 522 flux, averaged across all vegetation types, was $36.1 \pm 3.1 \text{ mg CH}_4$ -C m⁻² d⁻¹, spanning a range 523 from -100 to 1,510 mg CH₄-C m^{$^{-2}$} d^{$^{-1}$}. This mean falls within the range of other diffusive fluxes 524 observed in Indonesian peatlands (3.7-87.8 mg CH_4 -C m⁻² d⁻¹) (Couwenberg et al., 2010) and 525 other Amazonian wetlands (7.1-390.0 mg CH_4 -C m⁻² d⁻¹) (Bartlett et al., 1990;Bartlett et al., 526 1988; Devol et al., 1990; Devol et al., 1988). Although the ebullition data must be treated with 527 caution because of the sampling methodology (see below), we observed a mean net ebullition 528 flux of 973.3 \pm 161.4 mg CH₄-C m⁻² d⁻¹, spanning a range of 27 to 8,082 mg CH₄-C m⁻² d⁻¹. 529 While data on ebullition from Amazonian wetlands are sparse, these values are broadly in-530 line with riverine and lake ecosystems sampled elsewhere (Bastviken et al., 2010;Smith et al., 531 2000;Sawakuchi et al., 2014). Ebullition-driven CH₄ uptake is not a commonly reported 532 phenomena in other peatland studies because it is likely an artefact of chamber sampling 533 534 methods; as a consequence, we do not discuss these data further here. To summarize, these data on diffusive CH₄ flux and ebullition suggest that peatlands in the Pastaza-Marañón 535 foreland basin are strong contributors to the regional atmospheric budget of CH₄, given that 536 the four vegetation types sampled here represent the dominant cover types in the PMFB 537 538 (Draper et al., 2014; Householder et al., 2012; Kelly et al., 2014; Lahteenoja and Page, 2011)

The overall trend in the diffusive flux data was towards greater temporal (i.e. seasonal) 540 541 variability in diffusive CH₄ flux rather than strong spatial (i.e. inter-site) variability. For the 542 pooled dataset, diffusive CH₄ emissions were significantly greater during the wet season than 543 the dry season, with emissions falling by approximately half from one season to the other (i.e. 51.1 ± 7.0 to 27.3 ± 2.7 mg CH₄-C m^{$^{-2}$} d^{$^{-1}$}). This is in contrast to the data on diffusive CH₄ flux 544 among study sites, where statistical analyses indicate that there was a weak effect of 545 546 vegetation type on CH₄ flux, that was only on the edge of statistical significance (i.e. ANCOVA; 547 P < 0.06 for the vegetation effect term). For the ebullition data, while there was no significant 548 difference among vegetation types nor between seasons, it is interesting to note that ebullition was more common for the two vegetation types – Mixed Palm Swamp and M. 549 550 *flexuosa* palm swamp – that showed the highest rates of diffusive CH_4 flux (Figure 2, Table 2). In contrast, forested (short pole) and forested vegetation, which showed the lowest rates of 551 diffusive CH₄ flux, also showed the lowest occurrence of ebullition (Figure 2, Table 2). This is 552 broadly consistent with the notion that Mixed Palm Swamp and *M. flexuosa* palm swamp may 553 554 produce more CH₄ or possess lower gross CH₄ oxidation rates than the other vegetation types.

555

On face value, these data on diffusive CH_4 flux suggest two findings: first, the relatively weak effect of vegetation type on diffusive CH_4 flux implies that patterns of CH_4 cycling are broadly similar among study sites. Second, the strong *overall* seasonal pattern suggests that – on the whole – these systems conform to our normative expectations of how peatlands function with respect to seasonal variations in hydrology and redox potential; i.e. enhanced CH_4 emissions during a more anoxic wet season (i.e. when water tables rise), and reduced CH_4 emissions during a more oxic dry season (i.e. when water tables fall). However, closer

inspection of the data reveals that different vegetation types showed contrasting seasonal 563 564 emission patterns (Table 3), challenging our basic assumptions about how these ecosystems 565 function. For example, while forested (short pole) vegetation and *M. flexuosa* palm swamp 566 conformed to expected seasonal trends for methanogenic wetlands (i.e. higher wet season compared to dry season emissions), forested vegetation and mixed palm swamp showed the 567 opposite pattern, with significantly greater CH_4 emissions during the dry season. The 568 569 disaggregated data thus imply that the process-based controls on CH₄ fluxes may vary significantly among these different ecosystems, rather than being similar, leading to a 570 571 divergence in seasonal flux patterns.

572

573 What may explain this pattern of seasonal divergence in CH₄ flux? One explanation is that CH₄ 574 emissions from forested vegetation and mixed palm swamp, compared to the other two 575 ecosystems, may be more strongly transport-limited during the wet season than the dry season. This interpretation is supported by the field data; forested vegetation and mixed palm 576 577 swamp had the highest wet season water table levels, measuring 110.8 ± 9.3 and 183.7 ± 1.7 578 cm, respectively (Table 2). In contrast, water table levels for forested (short pole) vegetation 579 and *M. flexuosa* palm swamp in the wet season were 3-7 times lower, measuring only 26.9 ± 580 0.5 and 37.2 \pm 1.7 cm, respectively (Table 2). Moreover, a scatter plot of diffusive CH₄ flux 581 against water table depth shows a peak in diffusive CH₄ emissions when water tables are between 30 to 40 cm above the surface, after which CH₄ emissions decline precipitously 582 (Supplementary Online Materials Figure S2). Thus, the greater depth of overlying water in 583 584 forested vegetation and mixed palm swamp may have exerted a much greater physical constraint on gas transport compared to the other two ecosystems. This interpretation is 585

broadly consistent with studies from other ecosystems, which indicate that high or positive
water tables may suppress CH₄ emissions from wetlands above a system-specific threshold
(Couwenberg et al., 2010;Couwenberg et al., 2011).

589

590 However, transport limitation alone does not fully explain the difference in dry season CH₄ emissions among vegetation types. Forested vegetation and mixed palm swamp showed 591 substantially higher dry season CH₄ emissions (47.2 \pm 5.4 and 85.5 \pm 26.4 mg CH₄-C m⁻² d⁻¹, 592 respectively) compared to forested (short pole) vegetation and *M. flexuosa* palm swamp (9.6 593 \pm 2.6 and 25.5 \pm 2.9 mg CH₄-C m⁻² d⁻¹, respectively), pointing to underlying differences in CH₄ 594 production and oxidation among these ecosystems. One possibility is that dry season 595 596 methanogenesis in forested vegetation and mixed palm swamp was greater than in the other 597 two ecosystems, potentially driven by higher rates of C flow (Whiting and Chanton, 1993). 598 This is plausible given that forested vegetation and mixed palm swamp tend to occur in more nutrient-rich parts of the Pastaza-Marañón foreland basin, whereas forested (short pole) 599 vegetation and *M. flexuosa* palm swamp tend to dominate in more nutrient-poor areas 600 601 (Lahteenoja et al., 2009a), leading to potential differences in rates of plant productivity and 602 belowground C flow. Moreover, it is possible that the nutrient-rich vegetation may be able to utilize the higher concentration of nutrients, deposited during the flood pulse, during the 603 604 Amazonian dry season (Morton et al., 2014; Saleska et al., 2016), with implications for overall 605 ecosystem C throughput and CH₄ emissions. Of course, this interpretation does not preclude other explanations, such as differences in CH₄ transport rates among ecosystems (e.g. due to 606 plant-facilitated transport or ebullition) (Panagala et al., 2013), or varying rates of CH₄ 607 oxidation (Teh et al., 2005). However, these other possibilities cannot be explored further 608

609 without recourse to more detailed process-level experiments. Forthcoming studies on the 610 regulation of GHG fluxes at finer spatial scales (e.g. investigation of environmental gradients 611 within individual study sites) or detailed diurnal studies of GHG exchange (Murphy *et al.*, in 612 prep.) will further deepen our understanding of the process controls on soil GHG flux from 613 these peatlands, and shed light on these questions.

614

Finally, while the trends described here are intriguing, it is important to acknowledge some 615 of the potential limitations of our data. First, given the uneven sampling pattern, it is possible 616 617 that the values reported here do not fully represent the entire range of diffusive flux rates, 618 especially for the more sparsely sampled habitats. However, given the large and statistically 619 significant differences in CH₄ emissions between seasons, it is likely that the main trends that 620 we have identified will hold true with more spatially-extensive sampling. Second, the data are 621 a conservative underestimate of CH₄ emissions, because the low frequency, static chamber sampling approach that we utilized was unable to fully capture erratic ebullition events 622 623 representatively (McClain et al., 2003). Although we attempted to quantify CH_4 ebullition 624 within our static flux chambers, the sampling approach that we utilized was not the best-625 suited for representatively quantifying ebullition. Given the erratic or stochastic nature of 626 ebullition, automated chamber measurements or an inverted "flux funnel" approach would 627 have provided better estimates of ebullition (Strack et al., 2005). However, we lacked the resources to apply these techniques here. We also did not measure CH₄ emissions from the 628 629 stems of woody plants, even though woody plants have been recently identified as an 630 important point of atmospheric egress (Pangala et al., 2013). We did not have enough data on floristic composition or individual plant identities within our plots to develop a sampling 631

design that would adequately represent plant-mediated fluxes from our study sites, nor the 632 633 resources to implement a separate study of stem fluxes. Third and last, our data probably underestimate net CH₄ fluxes for the PMFB because we chose to include fluxes with strong 634 negative values (i.e. more than -10 mg CH_4 -C m⁻² d⁻¹) in our calculation of mean diffusive flux 635 rates. These observations are more negative than other values typically reported elsewhere 636 in the tropical wetland literature (Bartlett et al., 1990;Bartlett et al., 1988;Devol et al., 637 638 1990; Devol et al., 1988; Couwenberg et al., 2010). However, they represent only a small proportion of our dataset (i.e. 7 %, or only 68 out of 980 measurements), and inspection of 639 640 our field notes and the data itself did not produce convincing reasons to exclude these observations (e.g. we found no evidence of irregularities during field sampling, and any 641 642 chambers that showed statistically insignificant changes in concentration over time were removed during our quality control procedures). While headspace concentrations for these 643 644 measurements were often elevated above mean tropospheric levels (>2 ppm), this in itself is not unusual in reducing environments that contain strong local sources of CH₄ (Baldocchi et 645 646 al., 2012). We did not see this as a reason to omit these values as local concentrations of CH₄ are likely to vary naturally in methanogenic forest environments due to poor mixing in the 647 understory and episodic ebullition events. Importantly, exclusion of these data did not alter 648 the overall statistical trends reported above, and only produced slightly higher estimates of 649 diffusive CH₄ flux (41.6 ± 3.2 mg CH₄-C m⁻² d⁻¹ versus 36.1 ± 3.1 mg CH₄-C m⁻² d⁻¹). 650

651

652 6.2 Western Amazonian peatlands as weak atmospheric sources of nitrous oxide

The ecosystems sampled in this study were negligible atmospheric sources of N₂O, emitting only $0.70 \pm 0.34 \ \mu g \ N_2O-N \ m^{-2} \ d^{-1}$, suggesting that peatlands in the Pastaza-Marañón foreland

basin make little or no contribution to regional atmospheric budgets of N₂O. This is consistent 655 656 with N_2O flux measurements from other forested tropical peatlands, where N_2O emissions 657 were also found to be relatively low (Inubushi et al., 2003; Couwenberg et al., 2010). No 658 statistically significant differences in N₂O flux were observed among study sites or between 659 seasons, suggesting that these different peatlands may have similar patterns of N₂O cycling. Interestingly, differences in N₂O fluxes were not associated with the nutrient status of the 660 661 peatland; i.e. more nutrient-rich ecosystems, such as forested vegetation and mixed palm 662 swamp, did not show higher N_2O fluxes than their nutrient-poor counterparts, such as 663 forested (short pole) vegetation and *M. flexuosa* palm swamp. This may imply that N availability, one of the principal drivers of nitrification, denitrification, and N₂O production 664 665 (Groffman et al., 2009; Werner et al., 2007), may not be greater in nutrient-rich versus nutrient-poor ecosystems in this part of the Western Amazon. Alternatively, it is possible that 666 even though N availability and N fluxes may differ between nutrient-rich and nutrient-poor 667 systems, N₂O yield may also vary such that net N₂O emissions are not significantly different 668 669 among study sites (Teh et al., 2014).

670

One potential source of concern are the negative N₂O fluxes that we documented here. While 671 some investigators have attributed negative fluxes to instrumental error (Cowan et al., 672 2014;Chapuis-Lardy et al., 2007), others have demonstrated that N₂O consumption -673 674 particularly in wetland soils – is not an experimental artifact, but occurs due to the complex effects of redox, organic carbon content, nitrate availability, and soil transport processes on 675 denitrification (Ye and Horwath, 2016; Yang et al., 2011; Wen et al., 2016; Schlesinger, 676 2013;Teh et al., 2014;Chapuis-Lardy et al., 2007). Given the low redox potential and high 677 678 carbon content of these soils, it is plausible that microbial N_2O consumption is occurring,

- because these types of conditions have been found to be conducive for N₂O uptake elsewhere
 (Ye and Horwath, 2016;Teh et al., 2014;Yang et al., 2011).
- 681
- 682

683 **7. CONCLUSIONS**

684 Our data suggest that peatlands in the Pastaza-Marañón foreland basin are strong sources of atmospheric CH₄ at a regional scale, and need to be better accounted for in CH₄ emissions 685 inventories for the Amazon basin as a whole. In contrast, N₂O fluxes were negligible, 686 687 suggesting that these ecosystems are weak regional sources at best. Divergent or 688 asynchronous seasonal emissions pattern for CH₄ among different vegetation types was 689 intriguing, and challenges our underlying expectations of how tropical peatlands function. 690 These data highlight the need for greater wet season sampling, particularly from ecosystems 691 near river margins that may experience very high water tables (i.e. >40 cm). Moreover, these 692 data also emphasize the need for more spatially-extensive sampling across both the Pastaza-Marañón foreland basin and the wider Amazon region as a whole, in order to establish if these 693 694 asynchronous seasonal emission patterns are commonplace or specific to peatlands in the 695 PMFB region. If CH₄ emission patterns for different peatlands in the Amazon are in fact 696 asynchronous and decoupled from rainfall seasonality, then this may partially explain some of the heterogeneity in CH₄ source and sinks observed at the basin-wide scale (Wilson et al., 697 2016). 698

699

701 8. AUTHOR CONTRIBUTION

YAT secured the funding for this research, assisted in the planning and design of the experiment, and took the principal role in the analysis of the data and preparation of the manuscript. WAM planned and designed the experiment, collected the field data, analyzed the samples, and took a secondary role in data preparation, data analysis, and manuscript preparation. JCB, AB, and SEP supported the planning and design of the experiment, and provided substantive input into the writing of the manuscript. AB in particular took a lead role in developing the maps of our study sites in the PMFB.

709

710

711 9. ACKNOWLEDGEMENTS

712 The authors would like to acknowledge the UK Natural Environment Research Council for funding this research (NERC award number NE/I015469). We would like to thank MINAG and 713 the Ministerio de Turismo in Iquitos for permits to conduct this research, the Instituto de 714 Investigaciones de la Amazonía Peruana (IIAP) for logistical support, Peruvian rainforest 715 716 villagers for their warm welcome and acceptance, Hugo Vasquez, Pierro Vasquez, Gian Carlo Padilla Tenazoa and Yully Rojas Reátegui for fieldwork assistance, Dr Outi Lahteenoja and Dr 717 718 Ethan Householder for fieldwork planning, and Dr Paul Beaver of Amazonia Expeditions for lodging and logistical support. Our gratitude also goes to Alex Cumming for fieldwork support 719 720 and laboratory assistance, Bill Hickin, Gemma Black, Adam Cox, Charlotte Langley, Kerry Allen, 721 and Lisa Barber of the University of Leicester for all of their continued support. Thanks are 722 also owed to Graham Hambley (St Andrews), Angus Calder (St Andrews), Viktoria Oliver

(Aberdeen), Torsten Diem (Aberdeen), Tom Kelly (Leeds), and Freddie Draper Leeds) for their
help in the laboratory and with fieldwork planning. TD, VO, and two anonymous referees
provided very helpful and constructive comments on earlier drafts of this manuscript. This
publication is a contribution from the Scottish Alliance for Geoscience, Environment and
Society (<u>http://www.sages.ac.uk</u>) and the UK Tropical Peatland Working Group
(<u>https://tropicalpeat.wordpress.com</u>).

- 729
- 730

731 **10. REFERENCES**

- Andriesse, J.: Nature and management of tropical peat soils, 59, Food & Agriculture Org.,
 1988.
- 734 Baggs, E. M.: A review of stable isotope techniques for N2O source partitioning in soils:
- 735 Recent progress, remaining challenges and future considerations. , Rapid Communications
- in Mass Spectrometry 22, 1664-1672, 2008.
- 737 Baldocchi, D., Detto, M., Sonnentag, O., Verfaillie, J., Teh, Y. A., Silver, W., and Kelly, N. M.:
- The challenges of measuring methane fluxes and concentrations over a peatland pasture,
- 739 Agric. For. Meteorol., 153, 177-187, <u>http://dx.doi.org/10.1016/j.agrformet.2011.04.013</u>, 2012.
- 740 Bartlett, K. B., Crill, P. M., Sebacher, D. I., Harriss, R. C., Wilson, J. O., and Melack, J. M.:
- 741 METHANE FLUX FROM THE CENTRAL AMAZONIAN FLOODPLAIN, J. Geophys. Res.-Atmos.,
- 742 93, 1571-1582, 1988.
- 743 Bartlett, K. B., Crill, P. M., Bonassi, J. A., Richey, J. E., and Harriss, R. C.: METHANE FLUX
- 744 FROM THE AMAZON RIVER FLOODPLAIN EMISSIONS DURING RISING WATER, J. Geophys.
- 745 Res.-Atmos., 95, 16773-16788, 10.1029/JD095iD10p16773, 1990.

746	Bastviken, D., Santo	ro, A. L., Marotta	, H., Pinho, L. Q.	., Calheiros, D. F	., Crill, P., and Enrich-
-----	----------------------	--------------------	--------------------	--------------------	---------------------------

- 747 Prast, A.: Methane Emissions from Pantanal, South America, during the Low Water Season:
- 748 Toward More Comprehensive Sampling, Environ. Sci. Technol., 44, 5450-5455,
- 749 10.1021/es1005048, 2010.
- 750 Belyea, L. R., and Baird, A. J.: Beyond "The limits to peat bog growth": Cross-scale feedback
- in peatland development, Ecological Monographs, 76, 299-322, 2006.
- 752 Blazewicz, S. J., Petersen, D. G., Waldrop, M. P., and Firestone, M. K.: Anaerobic oxidation of
- 753 methane in tropical and boreal soils: Ecological significance in terrestrial methane cycling,
- Journal of Geophysical Research: Biogeosciences, 117, n/a-n/a, 10.1029/2011JG001864,
- 755 2012.
- 756 Chapuis-Lardy, L., Wrage, N., Metay, A., Chotte, J.-L., and Bernoux, M.: Soils, a sink for N2O?
- 757 A review, Global Change Biology, 13, 1-17, 10.1111/j.1365-2486.2006.01280.x, 2007.
- 758 Conrad, R.: Soil Microorganisms as Controllers of Atmospheric Trace Gases., Microbiological
- 759 Reviews, 60, 609-640, 1996.
- Couwenberg, J., Dommain, R., and Joosten, H.: Greenhouse gas fluxes from tropical
- 761 peatlands in south-east Asia, Global Change Biology, 16, 1715-1732, 10.1111/j.1365-
- 762 2486.2009.02016.x, 2010.
- 763 Couwenberg, J., Thiele, A., Tanneberger, F., Augustin, J., Bärisch, S., Dubovik, D.,
- Liashchynskaya, N., Michaelis, D., Minke, M., Skuratovich, A., and Joosten, H.: Assessing
- 765 greenhouse gas emissions from peatlands using vegetation as a proxy, Hydrobiologia, 674,
- 766 67-89, 10.1007/s10750-011-0729-x, 2011.
- 767 Cowan, N. J., Famulari, D., Levy, P. E., Anderson, M., Reay, D. S., and Skiba, U. M.:
- 768 Investigating uptake of N₂O in agricultural soils using a high-precision dynamic
- 769 chamber method, Atmos. Meas. Tech., 7, 4455-4462, 10.5194/amt-7-4455-2014, 2014.

- 770 D'Amelio, M. T. S., Gatti, L. V., Miller, J. B., and Tans, P.: Regional N2O fluxes in Amazonia
- derived from aircraft vertical profiles, Atmospheric Chemistry and Physics, 9, 8785-8797,

772 2009.

- 773 Devol, A. H., Richey, J. E., Clark, W. A., King, S. L., and Martinelli, L. A.: Methane emissions to
- the troposphere from the Amazon floodplain, Journal of Geophysical Research:
- 775 Atmospheres, 93, 1583-1592, 10.1029/JD093iD02p01583, 1988.
- 776 Devol, A. H., Richey, J. E., Forsberg, B. R., and Martinelli, L. A.: SEASONAL DYNAMICS IN
- 777 METHANE EMISSIONS FROM THE AMAZON RIVER FLOODPLAIN TO THE TROPOSPHERE, J.
- 778 Geophys. Res.-Atmos., 95, 16417-16426, 10.1029/JD095iD10p16417, 1990.
- 779 Draper, F. C., Roucoux, K. H., Lawson, I. T., Mitchard, E. T. A., Coronado, E. N. H., Lahteenoja,
- 780 O., Montenegro, L. T., Sandoval, E. V., Zarate, R., and Baker, T. R.: The distribution and
- amount of carbon in the largest peatland complex in Amazonia, Environmental Research
- 782 Letters, 9, 12, 10.1088/1748-9326/9/12/124017, 2014.
- 783 Firestone, M. K., Firestone, R. B., and Tiedge, J. M.: Nitrous oxide from soil denitrification:
- Factors controlling its biological production., Science, 208, 749-751, 1980.
- 785 Firestone, M. K., and Davidson, E. A.: Microbiological basis of NO and N2O production and
- 786 consumption in soil, in: Exchange of Trace Gases Between Terrestrial Ecosystems and the
- 787 Atmosphere, edited by: Andrae, M. O., and Schimel, D. S., John Wiley and Sons Ltd., New
 788 York, 7-21, 1989.
- 789 Groffman, P. M., Butterbach-Bahl, K., Fulweiler, R. W., Gold, A. J., Morse, J. L., Stander, E. K.,
- 790 Tague, C., Tonitto, C., and Vidon, P.: Challenges to incorporating spatially and temporally
- 791 explicit phenomena (hotspots and hot moments) in denitrification models, Biogeochemistry,
- 792 93, 49-77, 10.1007/s10533-008-9277-5, 2009.

- Hanson, R. S., and Hanson, T. E.: Methanotrophic Bacteria., Microbiological Reviews, 60,
 439-471, 1996.
- Householder, J. E., Janovec, J., Tobler, M., Page, S., and Lähteenoja, O.: Peatlands of the
- 796 Madre de Dios River of Peru: Distribution, Geomorphology, and Habitat Diversity, Wetlands,
- 797 32, 359-368, 10.1007/s13157-012-0271-2, 2012.
- Huang, J., Golombek, A., Prinn, R., Weiss, R., Fraser, P., Simmonds, P., Dlugokencky, E. J.,
- Hall, B., Elkins, J., Steele, P., Langenfelds, R., Krummel, P., Dutton, G., and Porter, L.:
- 800 Estimation of regional emissions of nitrous oxide from 1997 to 2005 using multinetwork
- 801 measurements, a chemical transport model, and an inverse method, J. Geophys. Res.-
- 802 Atmos., 113, 1-19, D17313
- 803 10.1029/2007jd009381, 2008.
- Jungkunst, H. F., and Fiedler, S.: Latitudinal differentiated water table control of carbon
- dioxide, methane and nitrous oxide fluxes from hydromorphic soils: feedbacks to climate
- change, Global Change Biology, 13, 2668-2683, 10.1111/j.1365-2486.2007.01459.x, 2007.
- 807 Keller, M., Kaplan, W. A., and Wofsy, S. C.: EMISSIONS OF N2O, CH4 AND CO2 FROM
- 808 TROPICAL FOREST SOILS, J. Geophys. Res.-Atmos., 91, 1791-1802,
- 809 10.1029/JD091iD11p11791, 1986.
- 810 Kelly, T. J., Baird, A. J., Roucoux, K. H., Baker, T. R., Honorio Coronado, E. N., Ríos, M., and
- Lawson, I. T.: The high hydraulic conductivity of three wooded tropical peat swamps in
- 812 northeast Peru: measurements and implications for hydrological function, Hydrological
- 813 Processes, 28, 3373-3387, 10.1002/hyp.9884, 2014.
- 814 Kirschke, S., Bousquet, P., Ciais, P., Saunois, M., Canadell, J. G., Dlugokencky, E. J.,
- 815 Bergamaschi, P., Bergmann, D., Blake, D. R., Bruhwiler, L., Cameron-Smith, P., Castaldi, S.,

- 816 Chevallier, F., Feng, L., Fraser, A., Heimann, M., Hodson, E. L., Houweling, S., Josse, B.,
- 817 Fraser, P. J., Krummel, P. B., Lamarque, J. F., Langenfelds, R. L., Le Quere, C., Naik, V.,
- O'Doherty, S., Palmer, P. I., Pison, I., Plummer, D., Poulter, B., Prinn, R. G., Rigby, M.,
- 819 Ringeval, B., Santini, M., Schmidt, M., Shindell, D. T., Simpson, I. J., Spahni, R., Steele, L. P.,
- 820 Strode, S. A., Sudo, K., Szopa, S., van der Werf, G. R., Voulgarakis, A., van Weele, M., Weiss,
- 821 R. F., Williams, J. E., and Zeng, G.: Three decades of global methane sources and sinks,
- Nature Geoscience, 6, 813-823, 10.1038/ngeo1955, 2013.
- Lahteenoja, O., Ruokolainen, K., Schulman, L., and Alvarez, J.: Amazonian floodplains
- harbour minerotrophic and ombrotrophic peatlands, Catena, 79, 140-145,
- 825 10.1016/j.catena.2009.06.006, 2009a.
- Lahteenoja, O., Ruokolainen, K., Schulman, L., and Oinonen, M.: Amazonian peatlands: an
- ignored C sink and potential source, Global Change Biology, 15, 2311-2320, 10.1111/j.1365-
- 828 2486.2009.01920.x, 2009b.
- Lahteenoja, O., and Page, S.: High diversity of tropical peatland ecosystem types in the
- 830 Pastaza-Maranon basin, Peruvian Amazonia, Journal of Geophysical Research-
- Biogeosciences, 116, 14, 10.1029/2010jg001508, 2011.
- Lahteenoja, O., Reategui, Y. R., Rasanen, M., Torres, D. D., Oinonen, M., and Page, S.: The
- 833 large Amazonian peatland carbon sink in the subsiding Pastaza-Maranon foreland basin,
- 834 Peru, Global Change Biology, 18, 164-178, 10.1111/j.1365-2486.2011.02504.x, 2012.
- Lavelle, P., Rodriguez, N., Arguello, O., Bernal, J., Botero, C., Chaparro, P., Gomez, Y.,
- 836 Gutierrez, A., Hurtado, M. D., Loaiza, S., Pullido, S. X., Rodriguez, E., Sanabria, C., Velasquez,
- 837 E., and Fonte, S. J.: Soil ecosystem services and land use in the rapidly changing Orinoco
- 838 River Basin of Colombia, Agriculture Ecosystems & Environment, 185, 106-117,
- 839 10.1016/j.agee.2013.12.020, 2014.

- Liengaard, L., Nielsen, L. P., Revsbech, N. P., Priem, A., Elberling, B., Enrich-Prast, A., and
- 841 Kuhl, M.: Extreme emission of N2O from tropical wetland soil (Pantanal, South America),
- 842 Frontiers in Microbiology, 3, 13, 10.3389/fmicb.2012.00433, 2013.
- Limpens, J., Berendse, F., Blodau, C., Canadell, J. G., Freeman, C., Holden, J., Roulet, N.,
- 844 Rydin, H., and Schaepman-Strub, G.: Peatlands and the carbon cycle: from local processes to
- global implications a synthesis, Biogeosciences, 5, 1475–1491, 2008.
- 846 Marani, L., and Alvalá, P. C.: Methane emissions from lakes and floodplains in Pantanal,
- 847 Brazil, Atmospheric Environment, 41, 1627-1633,
- 848 <u>http://dx.doi.org/10.1016/j.atmosenv.2006.10.046</u>, 2007.
- McClain, M. E., Boyer, E. W., Dent, C. L., Gergel, S. E., Grimm, N. B., Groffman, P. M., Hart, S.
- 850 C., Harvey, J. W., Johnston, C. A., Mayorga, E., McDowell, W. H., and Pinay, G.:
- 851 Biogeochemical hot spots and hot moments at the interface of terrestrial and aquatic
- ecosystems, Ecosystems, 6, 301-312, 10.1007/s10021-003-0161-9, 2003.
- Melack, J. M., Hess, L. L., Gastil, M., Forsberg, B. R., Hamilton, S. K., Lima, I. B. T., and Novo,
- 854 E.: Regionalization of methane emissions in the Amazon Basin with microwave remote
- sensing, Global Change Biology, 10, 530-544, 10.1111/j.1529-8817.2003.00763.x, 2004.
- Melton, J. R., Wania, R., Hodson, E. L., Poulter, B., Ringeval, B., Spahni, R., Bohn, T., Avis, C.
- A., Beerling, D. J., Chen, G., Eliseev, A. V., Denisov, S. N., Hopcroft, P. O., Lettenmaier, D. P.,
- 858 Riley, W. J., Singarayer, J. S., Subin, Z. M., Tian, H., Zurcher, S., Brovkin, V., van Bodegom, P.
- 859 M., Kleinen, T., Yu, Z. C., and Kaplan, J. O.: Present state of global wetland extent and
- 860 wetland methane modelling: conclusions from a model inter-comparison project
- 861 (WETCHIMP), Biogeosciences, 10, 753-788, 10.5194/bg-10-753-2013, 2013.
- Morley, N., and Baggs, E. M.: Carbon and oxygen controls on N2O and N-2 production during
- nitrate reduction, Soil Biol. Biochem., 42, 1864-1871, 10.1016/j.soilbio.2010.07.008, 2010.

- Morton, D. C., Nagol, J., Carabajal, C. C., Rosette, J., Palace, M., Cook, B. D., Vermote, E. F.,
- 865 Harding, D. J., and North, P. R. J.: Amazon forests maintain consistent canopy structure and
- greenness during the dry season, Nature, 506, 221-224, 10.1038/nature13006
- 867 http://www.nature.com/nature/journal/v506/n7487/abs/nature13006.html supplementary-
- 868 <u>information</u>, 2014.
- Nisbet, E. G., Dlugokencky, E. J., and Bousquet, P.: Methane on the Rise—Again, Science,
- 870 343, 493-495, 10.1126/science.1247828, 2014.
- Pangala, S. R., Moore, S., Hornibrook, E. R. C., and Gauci, V.: Trees are major conduits for
- methane egress from tropical forested wetlands, New Phytologist, 197, 524-531,
- 873 10.1111/nph.12031, 2013.
- 874 Pett-Ridge, J., Petersen, D. G., Nuccio, E., and Firestone, M. K.: Influence of oxic/anoxic
- 875 fluctuations on ammonia oxidizers and nitrification potential in a wet tropical soil, FEMS

876 Microbiol. Ecol., 85, 179-194, 10.1111/1574-6941.12111, 2013.

- 877 Prosser, J. I., and Nicol, G. W.: Relative contributions of archaea and bacteria to aerobic
- ammonia oxidation in the environment, Environ. Microbiol., 10, 2931-2941, 10.1111/j.1462-
- 879 2920.2008.01775.x, 2008.
- 880 Saikawa, E., Schlosser, C. A., and Prinn, R. G.: Global modeling of soil nitrous oxide emissions
- from natural processes, Global Biogeochemical Cycles, 27, 972-989, 10.1002/gbc.20087,
- 882 2013.
- 883 Saikawa, E., Prinn, R. G., Dlugokencky, E., Ishijima, K., Dutton, G. S., Hall, B. D., Langenfelds,
- 884 R., Tohjima, Y., Machida, T., Manizza, M., Rigby, M., O'Doherty, S., Patra, P. K., Harth, C. M.,
- Weiss, R. F., Krummel, P. B., van der Schoot, M., Fraser, P. J., Steele, L. P., Aoki, S.,
- Nakazawa, T., and Elkins, J. W.: Global and regional emissions estimates for N2O,
- Atmospheric Chemistry and Physics, 14, 4617-4641, 10.5194/acp-14-4617-2014, 2014.

- Saleska, S. R., Wu, J., Guan, K., Araujo, A. C., Huete, A., Nobre, A. D., and Restrepo-Coupe,
- N.: Dry-season greening of Amazon forests, Nature, 531, E4-E5, 10.1038/nature16457, 2016.
- 890 Sawakuchi, H. O., Bastviken, D., Sawakuchi, A. O., Krusche, A. V., Ballester, M. V. R., and
- 891 Richey, J. E.: Methane emissions from Amazonian Rivers and their contribution to the global
- methane budget, Global Change Biology, 20, 2829-2840, 10.1111/gcb.12646, 2014.
- 893 Schlesinger, W. H.: An estimate of the global sink for nitrous oxide in soils, Global Change
- Biology, 19, 2929-2931, 10.1111/gcb.12239, 2013.
- Schulman, L., Ruokolainen, K., and Tuomisto, H.: Parameters for global ecosystem models,
 Nature, 399, 535-536, 1999.
- 897 Silver, W., Lugo, A., and Keller, M.: Soil oxygen availability and biogeochemistry along
- rainfall and topographic gradients in upland wet tropical forest soils., Biogeochemistry, 44,
 301-328, 1999.
- 900 Silver, W. L., Herman, D. J., and Firestone, M. K. S.: Dissimilatory Nitrate Reduction to
- Ammonium in Upland Tropical Forest Soils., Ecology, 82, 2410-2416, 2001.
- 902 Sjögersten, S., Black, C. R., Evers, S., Hoyos-Santillan, J., Wright, E. L., and Turner, B. L.:
- 903 Tropical wetlands: A missing link in the global carbon cycle?, Global Biogeochemical Cycles,
- 904 28, 1371-1386, 10.1002/2014GB004844, 2014.
- 905 Smith, L. K., Lewis, W. M., Chanton, J. P., Cronin, G., and Hamilton, S. K.: Methane emissions
- 906 from the Orinoco River floodplain, Venezuela., Biogeochemistry, 51, 113-140, 2000.
- 907 Strack, M., Kellner, E., and Waddington, J. M.: Dynamics of biogenic gas bubbles in peat and
- their effects on peatland biogeochemistry, Global Biogeochemical Cycles, 19, n/a-n/a,
- 909 10.1029/2004GB002330, 2005.

- 910 Teh, Y. A., Silver, W. L., and Conrad, M. E.: Oxygen effects on methane production and
- oxidation in humid tropical forest soils, Global Change Biology, 11, 1283-1297,
- 912 10.1111/j.1365-2486.2005.00983.x, 2005.
- 913 Teh, Y. A., and Silver, W. L.: Effects of soil structure destruction on methane production and
- carbon partitioning between methanogenic pathways in tropical rain forest soils, Journal of
- 915 Geophysical Research: Biogeosciences, 111, n/a-n/a, 10.1029/2005JG000020, 2006.
- 916 Teh, Y. A., Silver, W. L., Conrad, M. E., Borglin, S. E., and Carlson, C. M.: Carbon isotope
- 917 fractionation by methane-oxidizing bacteria in tropical rain forest soils, Journal of
- 918 Geophysical Research-Biogeosciences, 111, 10.1029/2005jg000053, 2006.
- 919 Teh, Y. A., Dubinsky, E. A., Silver, W. L., and Carlson, C. M.: Suppression of methanogenesis
- 920 by dissimilatory Fe(III)-reducing bacteria in tropical rain forest soils: implications for
- 921 ecosystem methane flux, Global Change Biology, 14, 413-422, 10.1111/j.1365-
- 922 2486.2007.01487.x, 2008.
- 923 Teh, Y. A., Silver, W. L., Sonnentag, O., Detto, M., Kelly, M., and Baldocchi, D. D.: Large
- 924 Greenhouse Gas Emissions from a Temperate Peatland Pasture, Ecosystems, 14, 311-325,
- 925 10.1007/s10021-011-9411-4, 2011.
- 926 Teh, Y. A., Diem, T., Jones, S., Huaraca Quispe, L. P., Baggs, E., Morley, N., Richards, M.,
- 927 Smith, P., and Meir, P.: Methane and nitrous oxide fluxes across an elevation gradient in the
- tropical Peruvian Andes, Biogeosciences, 11, 2325-2339, 10.5194/bg-11-2325-2014, 2014.
- von Fischer, J., and Hedin, L.: Separating methane production and consumption with a field-
- 930 based isotope dilution technique., Global Biogeochemical Cycles, 16, 1-13,
- 931 10.1029/2001GB001448, 2002.
- 932 von Fischer, J. C., and Hedin, L. O.: Controls on soil methane fluxes: Tests of biophysical
- 933 mechanisms using stable isotope tracers, Global Biogeochemical Cycles, 21, 9, Gb2007

- 934 10.1029/2006gb002687, 2007.
- 935 Wen, Y., Chen, Z., Dannenmann, M., Carminati, A., Willibald, G., Kiese, R., Wolf, B.,
- 936 Veldkamp, E., Butterbach-Bahl, K., and Corre, M. D.: Disentangling gross N2O production
- 937 and consumption in soil, Sci Rep, 6, 8, 10.1038/srep36517, 2016.
- 938 Werner, C., Butterbach-Bahl, K., Haas, E., Hickler, T., and Kiese, R.: A global inventory of N2O
- 939 emissions from tropical rainforest soils using a detailed biogeochemical model, Global
- Biogeochemical Cycles, 21, 1-18, Gb3010
- 941 10.1029/2006gb002909, 2007.
- 942 Whalen, S. C.: Biogeochemistry of methane exchange between natural wetlands and the
- atmosphere, Environ. Eng. Sci., 22, 73-94, 10.1089/ees.2005.22.73, 2005.
- Whiting, G. J., and Chanton, J. P.: Primary production control of methane emission from
 wetlands., Nature, 364, 794-795, 1993.
- 946 Wilson, C., Gloor, M., Gatti, L. V., Miller, J. B., Monks, S. A., McNorton, J., Bloom, A. A.,
- 947 Basso, L. S., and Chipperfield, M. P.: Contribution of regional sources to atmospheric
- methane over the Amazon Basin in 2010 and 2011, Global Biogeochem. Cycles, 30, 400–420,
- 949 10.1002/2015GB005300, 2016.
- 950 Wright, E. L., Black, C. R., Cheesman, A. W., Drage, T., Large, D., Turner, B. L., and
- 951 SjÖGersten, S.: Contribution of subsurface peat to CO2 and CH4 fluxes in a neotropical
- 952 peatland, Global Change Biology, 17, 2867-2881, 10.1111/j.1365-2486.2011.02448.x, 2011.
- Yang, W. H., Teh, Y. A., and Silver, W. L.: A test of a field-based N-15-nitrous oxide pool
- dilution technique to measure gross N2O production in soil, Global Change Biology, 17,
- 955 3577-3588, 10.1111/j.1365-2486.2011.02481.x, 2011.

- 956 Ye, R., and Horwath, W. R.: Nitrous oxide uptake in rewetted wetlands with contrasting soil
- 957 organic carbon contents, Soil Biology and Biochemistry, 100, 110-117,
- 958 <u>http://dx.doi.org/10.1016/j.soilbio.2016.06.009</u>, 2016.

11. TABLES AND FIGURES

Table 1. Site characteristics including field site location, nutrient status, plot and flux chamber

962 replication

Vegetation type	Site name	Nutrient	Latitude (S)	Longitude (W)	Plots	Flux
		status*				chambers
Forested	Buena Vista	Rich	4°14'45.60"S	73°12'0.20"W	21	105
Forested (short pole)	San Jorge (centre)	Poor	4°03'35.95"S	73*12'01.13"W	6	28
Forested (short pole)	Miraflores	Poor	4°28'16.59"S	74° 4'39.95"W	41	204
M. flexuosa Palm Swamp	Quistococha	Intermediate	3°49'57.61"S	73*12'01.13"W	135	668
M. flexuosa Palm Swamp	San Jorge (edge)	Intermediate	4*03'18.83"S	73*10'16.80"W	18	86
Mixed palm swamp	Charo	Rich	4*16'21.80"S	73*15'27.80"W	18	90

963 *After Householder et al. 2012, Lahteenoja et al. 2009a, and Lahteenoja et al. 2009b

- 965 **Table 2.** Proportion of observations for each vegetation type that showed evidence of
- 966 ebullition, mean rates of ebullition and ebullition-driven CH₄ uptake. Values represent

967 means and standard errors.

Vegetation Type	Percentage of Net Ebullition observations (mg CH ₄ -C m		1)	Ebullition-driven uptake (mg CH ₄ -C m ⁻² d ⁻¹)		
	(%)	Wet Season	Dry Season	Wet Season	Dry Season	
Forested	10.5	0	0	0	-136.4 ± 0.1	
Forested (short pole)	6.9	994.6 ± 293.2	512.5 ± 153.0	-95.8 ± 0.0	-245.5 ± 48.9	
<i>M. flexuosa</i> Palm Swamp	16.7	1192.0 ± 305.7	994.3 ± 237.3	-869.4 ± 264.8	-401.4 ± 59.9	
Mixed Palm Swamp	12.2	0	733.6 ± 313.1	0	-464.4 ± 565.9	

- 969 **Table 3.** Environmental variables for each vegetation type for the wet and dry season.
- 970 Values reported here are means and standard errors. Lower case letters indicate significant
- 971 differences among vegetation types within the wet or dry season (Fisher's LSD, *P* < 0.05).

Vegetation Type	Peat Ter	nperature	Air Tem	perature	Conduct	ivity	Dissolve	d Oxygen	Water Ta	able Level	pН	
	(°C)		(°C)		(µS m ⁻²)		(%)		(cm)			
	Wet	Dry	Wet	Dry	Wet	Dry	Wet	Dry	Wet	Dry	Wet	Dry
	Season	Season	Season	Season	Season	Season	Season	Season	Season	Season	Season	Season
Forested	26.1 ±	24.7 ±	28.8 ±	26.4 ±	79.0 ±	75.9 ±	0.2 ±	18.9 ±	110.8 ±	-13.2 ±	5.88 ±	6.31±
	0.1a	0.0a	0.7a	0.3a	5.9a	5.7a	0.1a	4.4a	9.3a	0.7a	0.15a	0.04a
Forested	25.2 ±	24.8 ±	27.6 ±	27.5 ±	21.0 ±	48.5 ±	4.4 ±	33.1 ±	26.9 ±	-4.7 ±	4.88 ±	3.8 ±
(short pole)	0.0b	0.1a	0.1b	0.1b	0.0b	4.8b	0.0a	2.6b	0.5b	0.4b	0.01b	0.03b
M. flexuosa	25.6 ±	25.3 ±	26.3 ±	26.4 ±	45.9 ±	51.9 ±	19.4 ±	17.3 ±	37.2 ±	6.1 ±	5.04 ±	5.49 ±
Palm Swamp	0.6c	0.1b	0.1c	0.1a	2.1c	1.8b	1.3b	1.5a	1.7c	1.3c	0.03c	0.03c
Mixed Palm	26.0 ±	25.0 ±	26.1 ±	28.2 ±	$100.0 \pm$	206.4 ±	0.0 ±	0.0 ±	183.7 ±	-2.4 ±	6.1 ±	6.82 ±
Swamp	0.0a	0.1ab	0.1c	0.3b	0.2d	4.2c	0.0a	0.0c	1.7d	0.3b	0.03a	0.02d

Table 4. Trace gas fluxes for each vegetation type for the wet and dry season. Values reported
here are means and standard errors. Upper case letters indicate significant differences in gas
flux between seasons with a vegetation type, while lower case letters indicate significant
differences among vegetation types within a season (Fisher's LSD, *P* < 0.05).

Vegetation Type	Methane Flux (mg CH ₄ -C m ⁻² c	l ⁻¹)	Nitrous Oxide Flux (μg N ₂ O-N m ⁻² d ⁻¹)		
	Wet Season	Dry Season	Wet Season	Dry Season	
Forested	6.7 ± 1.0Aa	47.2 ± 5.4Ba	2.54 ± 1.48	-1.16 ± 1.20	
Forested (short pole)	60.4 ± 9.1Ab	18.8 ± 2.6Bb	1.16 ± 0.54	-0.42 ± 0.90	
<i>M. flexuosa</i> Palm Swamp	46.7 ± 8.4Ac	28.3 ± 2.6Bc	1.14 ± 0.35	0.92 ± 0.61	
Mixed Palm Swamp	6.1±1.3Aa	64.2 ± 12.1Ba	1.45 ± 0.79	-0.80 ± 0.79	

978 Figure Captions

- 979 **Figure 1.** Map of the study region and field sites. The colour scale to the right of the map
- 980 denotes elevation in meters above sea level (m a.s.l.). Tan and brown tones indicate areas in
- 981 which peatlands are found; however, not all of these areas are peatland-dominated.

- **Figure 2.** Net diffusive (a) methane (CH₄) and (b) nitrous oxide (N₂O) fluxes by vegetation type.
- 984 Error bars denote standard errors.





