



1 Coccolithophore populations and their contribution to carbonate export during an

- 2 annual cycle in the Australian sector of the Antarctic Zone
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13 Abstract

The Southern Ocean is experiencing rapid and relentless change in its physical and 14 15 biogeochemical properties. The rate of warming of the Antarctic Circumpolar Current exceeds that of the global ocean, and the enhanced uptake of carbon dioxide is causing 16 17 basin-wide ocean acidification. Observational data suggest that these changes are influencing the distribution and composition of pelagic plankton communities. Long-term 18 19 and annual field observations on key environmental variables and organisms are a critical 20 basis for predicting changes in Southern Ocean ecosystems. These observations are 21 particularly needed, since high-latitude systems have been projected to experience the most severe impacts of ocean acidification and invasions of allochthonous species. 22

Coccolithophores are the most prolific calcium carbonate producing phytoplankton 23 24 group, playing an important role in Southern Ocean biogeochemical cycles. Satellite imagery has revealed elevated particulate inorganic carbon concentrations near the major 25 26 circumpolar fronts of the Southern Ocean, that can be attributed to the coccolithophore Emiliania huxleyi. Recent studies have suggested changes during the last decades in the 27 28 distribution and abundance of Southern Ocean coccolithophores. However, due to limited field observations, the distribution, diversity and state of coccolithophore populations in 29 30 the Southern Ocean remain poorly characterized.





We report here on seasonal variations in the abundance and composition of 31 coccolithophore assemblages collected by two moored sediment traps deployed in the 32 33 deep ocean (~2000 and 3700 m) in the Australian sector of the Antarctic Zone for one year in 2001-02. Additionally, seasonal changes in coccolith weights of E. huxleyi 34 populations were estimated using circularly polarized micrographs analysed with C-35 36 Calcita software. Our findings indicate that (1) coccolithophore sinking assemblages were nearly monospecific for Emiliania huxleyi morphotype B/C in the Antarctic Zone 37 waters in 2001-2002; (2) coccolith assemblages experienced weight and length reduction 38 39 during the summer months; (3) the estimated annual coccolith weight of E. huxleyi at both sediment trap depths (2.11 \pm 0.96 and 2.13 \pm 0.90 pg at 2000 m and 3700 m) was 40 consistent with previous studies for morphotype B/C in other Southern Ocean settings; 41 (4) coccolithophores accounted for approximately 2-5% of the annual, deep-ocean CaCO₃ 42 43 flux. Our results are the first annual record of coccolithophore abundance, composition 44 and degree of calcification in the Antarctic Zone. They provide a baseline against which to monitor coccolithophorid responses to changes in environmental conditions expected 45 46 for this region in coming decades. 47

48 Key words: Southern Ocean, Antarctic Zone, coccolithophores, coccolith weight,49 sediment traps.

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51 Introduction

The rapid increase in atmospheric CO_2 levels since the onset of the industrial 52 53 revolution is modifying the environmental conditions of marine ecosystems in a variety of ways. The enhanced greenhouse effect, mainly driven by increased atmospheric CO₂ 54 55 levels, is causing ocean warming (Barnett et al., 2005), shallowing of mixed layer depths (Levitus et al., 2000) and changes in light penetration and nutrient supply (Bopp et al., 56 57 2001; Rost and Riebesell, 2004; Sarmiento et al., 2004b; Deppeler and Davidson, 2017). 58 Moreover, the enhanced accumulation of CO₂ in the ocean is giving rise to changes in the ocean carbonate system, including reduction of carbonate ion concentrations and 59 lowering of seawater pH. Most evidence suggests that the ability of many marine 60 calcifying organisms to form carbonate skeletons and shells may be reduced with 61 62 increasing seawater acidification including some species (but not all) of coccolithophores, corals, pteropods and foraminifera (e.g. Orr et al., 2005; Moy et al., 2009; Lombard et al., 63





2010; Beaufort et al., 2011; Andersson and Gledhill, 2013). Since phytoplankton are
extremely sensitive to global environmental change (Litchman et al., 2012) all predicted
changes in marine environmental conditions are likely to modify the abundance,
composition and distribution of phytoplankton communities.

Changes in the relative abundances of major phytoplankton functional groups are 68 likely to influence ocean biogeochemistry and ocean carbon storage, with feedbacks to 69 the rate of climate change (e.g. Boyd and Newton, 1995; Boyd et al., 1999; Falkowski et 70 al., 2004; Cermeño et al., 2008). For example, diatoms can play a prominent role in export 71 72 of organic matter from the surface ocean, because of their heavy siliceous frustules and 73 capacity for aggregation and rapid sinking facilitates efficient transport of organic carbon (Buesseler, 1998; Smetacek, 1999), although this silica-mediated carbon export may not 74 reach the ocean interior efficiently (Francois et al., 2002; Lam and Bishop, 2007). The 75 76 precipitation and sinking of CaCO3 by coccolithophores also has the potential for 77 complex contributions to carbon cycling. Carbonate precipitation removes more alkalinity than dissolved inorganic carbon from surface waters, thereby acting to increase 78 79 pCO_2 in surface waters (the so-called carbonate counter pump, e.g. (Zeebe, 2012), but ballasting by carbonates appears to increase transfer of organic carbon to the ocean 80 81 interior (Armstrong et al., 2002; Klaas and Archer, 2002). On seasonal timescales the 82 counter pump contribution dominates (Boyd and Trull, 2007), but more complex 83 interactions can occur over longer timescales as a result of changing extents of carbonate dissolution in sediments, including the possibility that enhanced calcite dissolution in the 84 Southern Ocean contributed to lower atmospheric CO₂ levels during glacial maxima 85 (Archer and Maier-Reimer, 1994; Sigman and Boyle, 2000; Ridgwell and Zeebe, 2005). 86

87 The Southern Ocean is a critical component of the Earth's ocean-climate system and plays a pivotal role in the global biogeochemical cycles of carbon and nutrients 88 (Sarmiento et al., 2004a; Anderson et al., 2009). Despite its relatively small area (~25% 89 90 of the global ocean), the Southern Ocean contains ~40% of the global ocean inventory of 91 anthropogenic CO₂ (Khatiwala et al., 2009; Takahashi et al., 2009; Frölicher et al., 2015), and it exports nutrients to more northern latitudes ultimately supporting ~ 75% of the 92 ocean primary production north of 30°S (Sarmiento et al., 2004a). Model projections 93 94 suggest that the reduction in the saturation state of CaCO₃ will reach critical thresholds 95 sooner in cold, high-latitude ecosystems such as the Southern Ocean (Orr et al., 2005; McNeil and Matear, 2008; Feely et al., 2009). Therefore, calcifying organisms living in 96 97 these regions will be the first to face the most severe impacts of ocean acidification.





98 In view of the rapid changes in climate and other environmental stressors presently 99 occurring in the Southern Ocean, a major challenge facing the scientific community is to 100 predict how phytoplankton communities will reorganize in response to global change. In 101 this regard, two main aspects of the distributions of coccolithophores are emerging. Firstly, coccolithophores are dominantly present in the Subantarctic Southern Ocean, a 102 feature termed by Balch et al. (2011) as the "Great Calcite Belt" based on satellite 103 104 reflectance estimates of PIC abundances. Although importantly the PIC accumulations 105 are significantly less than those that arise in the North Atlantic, and the satellite algorithm 106 is not reliable in Antarctic waters, where it badly overestimates PIC abundances (Balch et al., 2016; Trull et al., 2017). Secondly, recent studies suggest that the magnitude and 107 geographical distribution of E. huxleyi blooms may be experiencing significant and rapid 108 109 changes. Cubillos et al. (2008) and Winter et al. (2014) postulated that E. huxleyi has 110 expanded its ecological niche south of the Polar Front in the recent decades. 111 Contrastingly, Freeman and Lovenduski (2015) suggested an overall decline in Southern Ocean PIC concentrations using satellite records between 1998 and 2014. The 112 113 explanation of these contrasting results may lie in the methodologies applied. While shipboard surface water observations provide a highly detailed picture of a given 114 115 ecosystem, they are very spares, only represent a snapshot in time, and can easily miss 116 blooms of any given species. The satellite PIC signal has the great advantage of largescale 117 and repeated coverage, but can miss subsurface populations (e.g. Winter et al., 2014) and be mimicked by the spectral characteristics of other scattering sources, such as 118 microbubbles (Zhang et al., 2002), glacial flour (Balch et al., 2011) and noncalcifying 119 organisms such as *Phaeocystis antarctica* (Winter et al., 2014), a colonial 120 121 prymnesiophyte algae very abundant in high latitude systems of the Southern Ocean (e.g. 122 Arrigo et al., 1999; Arrigo et al., 2000). Notably the PIC algorithm performs particularly 123 poorly in Antarctic waters (Balch et al., 2016; Trull et al., 2017)

124 For these reasons, year-round field observations of areas representative of key 125 Southern Ocean regions are essential to determine the current state of coccolithophore 126 communities and to develop baselines against which long-term trends can be detected. 127 Moreover, a better understanding of coccolithophore distribution, ecology and seasonal 128 dynamics is required to improve our interpretations of the sedimentary record and our 129 models of biogeochemistry. Sediment traps are a direct method to collect data about calcareous and siliceous micro and nanoplankton. Traps allow the monitoring of seasonal 130 131 and annual variability of plankton export, document species successions, and help to





determine the specific role of microplankton species in the biological and carbonate
pumps. The autonomous collection capacity of sediment traps is particularly useful in the
remote Southern Ocean, where inaccessibility and harsh working conditions prevent yearround ship-based sampling.

We present here the first record of composition, abundance, and seasonality of 136 137 coccolithophore assemblages in the Antarctic Zone of the Southern Ocean, a record 138 inferred from one-year records from two deep ocean sediment traps deployed on a single 139 mooring south of Australia at the site of the SOIREE ocean iron fertilisation experiment 140 near 61°S, 140°E (Boyd et al., 2000a). Moreover, we report weight and length measurements on E. huxleyi coccoliths, assessing the impact of seasonally varying 141 142 environmental parameters on E. huxleyi coccoliths. That provides a baseline of coccolith 143 dimensions for the populations living in this region. All the above information is needed 144 for monitoring coccolithophore responses, if any, to changing environmental conditions 145 in the Antarctic Zone south of Australia during coming decades.

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147 **2. Material and Methods**

148 2.1 Regional setting and oceanography

149 The southern Antarctic Zone (AZ-S; Parslow et al., 2001) is delimited at the north by the southern branch of the Polar Front (PF) and at the south by the southern front of 150 the Antarctic Circumpolar Current (SAACF). Trull et al. (2001b) summarized the 151 152 seasonal evolution of water column properties in the study region. The intense heat loss 153 of surface waters during winter decreases Sea Surface Temperature (SST) to values < 1°C, resulting in strong vertical convection. Winter mixing extends to depths of about 154 120 m, replenishing the upper water column with nutrients. Chlorophyll-a levels during 155 winter are negligible throughout the region due to the reduced solar radiation and the 156 157 deep, continuous vertical mixing. During summer, increasing solar radiation warms the surface ocean and a seasonal thermocline forms (Fig. 2). By late summer (March) SST 158 ranges between 2 and 3 °C. Considerable nutrient depletion associated with a moderate 159 increase in algal biomass occurs within the mixed layer. Nonetheless, due to the limited 160 sampling of the study region, the timing of the summer nutrient minimum is not well 161 constrained by the available data (Trull et al., 2001b). Silicate exhibits the strongest 162 163 summer draw-down of all the macronutrients, reaching ~30% of its winter values (Fig. 2; 164 Trull et al., 2001), mainly due to diatom growth and subsequent biogenic silica export to





165 the deep sea (Rigual-Hernández et al., 2015a). The low algal biomass accumulation in the region is attributed to the very low iron levels (0.1-0.2 nM; Boyd et al., 2000a; Sohrin et 166 167 al., 2000). Mesozooplankton analysis during the SOIREE experiment by Zeldis (2001) 168 indicates that zooplankton community in the study region is dominated by copepods, mainly large calanoid copepodites. Grazing pressure was low (<1% of the phytoplankton 169 standing stock removed per day) and, therefore, is thought not to play an important role 170 171 in the control of the micro-phytoplankton (primarily diatom) stocks, but nanoflagellate 172 grazer abundances were significant and were likely to have regulated smaller 173 phytoplankton abundances (Hall and Safi, 2001).



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Figure 1. Annual mean sea surface temperature map (World Ocean Atlas; Locarnini et al., 2013) of the Australian sector of the Southern Ocean, showing the position of the main frontal and zonal systems (adapted from Orsi et al., 1995) and the location of the 61°S, 54°S and 47°S sediment trap stations (inverted triangles). Abbreviations: STF Subtropical Front, SAZ - Subantarctic Zone, SAF - Subantarctic Front, PFZ - Polar Frontal Zone, PF - Polar Front, AZ - Antarctic Zone, SACCF - Southern ACC Front and SB - Southern Boundary.







Figure 2: (a) Summary of seasonal evolution of macronutrient concentrations (silicate and nitrate) at the 61°S site taken from the WOCE SR3 transects between 1993 and 1996 (modified from Trull et al., 2001b) (b) Seasonal variation in the vertical structure of temperature (°C) between June 2001 and April 2002 for the 61°S site from the World Ocean Atlas 2009 (Locarnini et al., 2010).

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189 **2.2 Water carbonate chemistry**

Calcite solubility increases at higher pressures and lower temperatures, so that 190 191 dissolution increases with depth in the water column. Based on downward changes in the 192 calcite dissolution rate, two critical depth horizons can be distinguished: the calcite saturation horizon (CSH) that can be defined as the depth at which the water becomes 193 194 undersaturated with respect to calcite (i.e. where $\Omega_{\text{calcite}} = 1$); and the CaCO₃ compensation depth (CCD), the depth at which the rate of calcite rain from the upper 195 196 water column equals the dissolution rate. Figure 3 shows carbonate concentrations $[CO_3^{2-}]$] and calcite saturation ($\Omega_{calcite}$) for the WOCE SR03 2001 transect between Antarctica 197 and Tasmania along the 140°E meridian as estimated by Bostock et al. (2011). In the AZ-198 199 S waters south of Tasmania, the CSH and CCD occur at 3000 and 3700 m, respectively 200 (Fig. 3). Therefore, the location of sediment traps at the 61°S site allows for the assessment of dissolution changes, if any, of coccolithophore assemblages between the 201 202 two critical dissolution depth horizons: the CSH and CCD. Notably, both progressive uptake of anthropogenic CO_2 and increased upwelling of naturally CO_2 rich deep waters 203 204 over the past 20 years is leading to shallowing of these features (Pardo et al., 2017) 205







Figure 3. Cross section of the mooring location (only 2000m and 3700m sediment traps are represented) in comparison to regional seafloor bathymetry, carbonate concentrations $[CO_3^{2-}]$ and calcite saturation ($\Omega_{calcite}$) for WOCE transect SR03 2001 from Bostock et al. (2011), who calculated them from the DIC and alkalinity in the CARINA database (Tanhua et al., 2008). The location of the transects is shown on the map on the right top. $\Omega_{calcite} = 1$ contour is highlighted with a red line to show the approximate depth of the CSH across the transect.

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215 **2.3 Sediment trap experiment**

As part of the SAZ collaborative research program (Trull et al., 2001c), a sediment 216 trap experiment was carried out at the 61°S site (60° 44.43'S; 139° 53.97'E) in the 217 218 Australian sector of the southern Antarctic Zone within the region where the Southern 219 Ocean Iron Release Experiment (SOIREE) was conducted (Boyd et al., 2000a). The 61°S 220 site is characterized by weak currents with a mean eastward geostrophic surface velocity of approximately 0.03 ± 0.02 m s⁻¹ (Trull et al., 2001b). The site is north of the Seasonal 221 Sea-Ice Zone (Massom et al., 2013; Rigual-Hernández et al., 2015a) and remote from any 222 223 known iceberg pathway (Gladstone et al., 2001).

The 61°S mooring was equipped with three McLane Parflux time series sediment traps (Honjo and Doherty, 1988) for approximately one year (November 30, 2001 to September 29, 2002, 317 days). The traps were located at approximately 1000, 2000 and





227 3700 m below the surface in a water column of 4393 m; Figure 3b). Each trap was provided with 21 cups. Sampling intervals were synchronized between traps and in order 228 229 to resolve the seasonal flux cycle ranged from 8 days (in austral summer) to 55 days in 230 austral winter. No samples were recovered from the shallowest trap owing to equipment malfunction and, therefore, only results for the 2000 and 3800 m traps are presented here. 231 Each trap was paired with an Aanderaa current meter and temperature sensors. The 250 232 ml collection cups were filled with a buffered solution of sodium tetraborate (1 g L^{-1}), 233 sodium chloride (5 g L⁻¹), strontium chloride (0.22 g L⁻¹), and mercury chloride (3 g L⁻¹) 234 235 in unfiltered, deep seawater from the region. The two deeper traps completed their collection sequence as programmed, providing continuous time-series for a year. Due to 236 237 the low particle fluxes during the winter, insufficient material remained for phytoplankton 238 analysis of cup 1 from the 2000 m trap and cups 1, 2, 19, 20 and 21 from the 3700 m trap 239 (Table 1).

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241 **2.4 Sample processing and coccolithophore counting procedure**

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243 The sediment trap cup contents were washed through a 1 mm sieve after recovery 244 and then divided into 10 aliquots using a rotary splitter (McLane, Inc.). A description of the analytical procedures for estimation of geochemical fluxes is provided in Trull et al. 245 (2001a) and Rigual-Hernández et al. (2015a). One aliquot was used for siliceous and 246 calcareous micro- and nano-plankton analyses. Each fraction for plankton analysis was 247 refilled with distilled water to 40 ml, from which 10 ml was subsampled and buffered 248 with a solution of sodium carbonate and sodium hydrogen carbonate (pH 8) and kept 249 250 refrigerated for calcareous nannoplankton analysis. Samples for coccolithophore analysis 251 were prepared following the methodology of Flores and Sierro (1997). In short, 300 µl 252 were extracted with a micropipette and dropped onto a glass Petri dish previously filled 253 with a buffered solution and with a cover slip on its bottom. After settling 12 hours, the 254 buffer solution was removed using short strips of filter paper placed at the edge of the dish. Then, the cover slip was left to dry completely and mounted on a glass slide using 255 256 Canada balsam. Coccoliths were identified and counted using a Nikon Eclipse 80i 257 polarized light microscope at 1000× magnification. A minimum of 400 coccoliths were 258 counted in each sample. Coccospheres occurred in much lower numbers than loose 259 coccoliths in these preparations. The coccolith counts were transformed into daily fluxes 260 of specimens m⁻² d⁻¹ following the formula:





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$$F = \frac{N \times \frac{A}{n \times a} \times V \times S}{d \times T}$$

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where "*F*" is the daily coccolith flux, "*N*" the number of coccoliths, "*A*" the total area of a Petri dish, "*n*" the number of fields-of-view analysed, "*a*" the area of a single field of view, "*V*" the dilution volume, "*S*" the split of the cup, "*d*" the number of days of collection and "*T*" the aperture area of the sediment trap.

268 Since the sediment trap collection period was shorter than a full calendar year, an estimate of the annual coccolith flux of the 2000 m trap was calculated. This estimate 269 270 takes into account the fact that the unsampled days occurred in winter when particle fluxes were low, and were obtained by using the flux for the last winter cup (#21 in 2002) to 271 represent mean daily fluxes during the unobserved interval. Due to the lack of samples 272 corresponding to the winter 2002 for the 3700 m sediment trap record, the annualization 273 of the coccolith fluxes for this trap was made based only on the samples with available 274 data. Therefore, the annualized and annual flux data for the 3700 m trap presented in 275 Table 1 should be used with caution. 276

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61_2000	Sampling period	Length	I otal Mass Patx	CaC	03	POC		Diatoms	Cocconnophore mix	Relative abundance		
Cup	mid point	days	$mg m^{2} d^{1}$	$\mathrm{mgm^{'2}d^{'1}}$	%	$mg m^{-2} d^{-1}$	%	10 ⁶ valves m ⁻² d ⁻¹	10 ⁸ coccoliths m ⁻² d ⁻¹	E. huxleyi	C. leptoporus	Other
1	nov. 30, 2001	8	48	14	30	0.7	1.5	-	-	-		
2	dic. 08, 2001	8	78	17	22	1.7	2.2	9	2.5	98.8	1.2	0.0
3	dic. 16, 2001	8	326	62	19	6.9	2.1	82	2.7	98.5	1.3	0.2
4	dic. 24, 2001	8	509	140	28	6.4	1.3	85	8.2	99.5	0.5	0.0
5	ene. 01, 2002	8	1151	44	4	26.9	2.3	408	12.3	99.8	0.2	0.0
6	ene. 09, 2002	8	1069	170	16	14.8	1.4	200	22.3	99.8	0.2	0.0
7	ene. 17, 2002	8	656	60	9	11.3	1.7	159	9.2	99.3	0.7	0.0
8	ene. 25, 2002	8	702	38	5	11.0	1.6	296	8.4	99.3	0.7	0.0
9	feb. 02, 2002	8	666	39	6	12.0	1.8	184	5.4	98.8	1.2	0.0
10	feb. 10, 2002	8	595	24	4	8.2	1.4	295	6.0	99.5	0.5	0.0
11	feb. 18, 2002	8	534	20	4	6.2	1.2	149	9.8	99.0	0.5	0.5
12	feb. 26, 2002	8	524	19	4	4.7	0.9	152	5.0	100.0	0.0	0.0
13	mar. 06, 2002	8	586	15	3	6.9	1.2	120	6.4	99.8	0.2	0.0
14	mar. 14, 2002	8	285	11	4	3.2	1.1	71	2.0	99.8	0.2	0.0
15	mar. 22, 2002	8	290	7	3	3.2	1.1	66	2.0	97.6	1.0	1.5
16	mar. 30, 2002	8	263	8	3	2.6	1.0	87	0.9	99.2	0.8	0.0
17	abr. 08, 2002	10	264	7	3	2.2	0.8	97	1.3	98.1	1.9	0.0
18	may. 08, 2002	50	130	5	4	1.2	1.0	47	0.8	99.8	0.2	0.0
19	jun. 29, 2002	54	65	2	4	0.7	1.0	10	0.7	98.8	0.8	0.4
20	ago. 22, 2002	55	56	2	4	0.8	1.5	19	0.9	99.5	0.2	0.2
21	sep. 29, 2002	20	42	2	4	0.5	1.3	6	0.9	98.0	2.0	0.0
Annualised values			232	17	7.4	3.3	1.4	67	2.8			
Annual flux			85 g m ⁻² y ⁻¹	6 g m ⁻² y ⁻¹		1.2 g m ⁻² y ⁻¹		24 10 ⁹ valves m ⁻² y ⁻¹	$1.03 \ 10^{11}$ coccoliths m $^{-2}$ y $^{-1}$	99.4	0.5	0.1
61_3700	Sampling period	Length	Total Mass Flux	CaCO ₃		POC		Diatoms	Coccolithophore flux	Relative abundance		ce
Cup	mid point	days	$mg m^{-2} d^{-1}$	$mg m^{-2} d^{-1}$	%	$mg m^{-2} d^{-1}$	%	10 ⁶ valves m ⁻² d ⁻¹	10 ⁷ coccoliths m ⁻² d ⁻¹	E. huxleyi	C. leptoporus	Other
1	nov. 30, 2001	8	38	9	23	0.4	1.1	-	-	-	-	-
2	dic. 08, 2001	8	31	9	28	0.4	1.2	-	-	-	-	-
3	dic. 16, 2001	8	99	29	30	1.4	1.4	4	1.3	99.0	0.7	0.2
4	dic. 24, 2001	8	231	59	26	1.4	0.6	12	5.5	99.3	0.5	0.2
5	ene. 01, 2002	8	873	87	10	17.3	2.0	118	11.6	99.8	0.2	0.0
6	ene. 09, 2002	8	1157	154	13	19.8	1.7	479	15.9	100.0	0.0	0.0
7	ene. 17, 2002	8	828	166	20	9.4	1.1	354	20.0	100.0	0.0	0.0
8	ene. 25, 2002	8	490	34	7	6.4	1.3	169	11.0	99.8	0.2	0.0
9	feb. 02, 2002	8	491	32	б	6.5	1.3	385	4.6	100.0	0.0	0.0
10	feb. 10, 2002	8	419	19	4	6.0	1.4	281	4.2	99.8	0.2	0.0
11	feb. 18, 2002	8	584	36	6	6.2	1.1	254	15.9	99.1	0.7	0.2
12	feb. 26, 2002	8	581	31	5	5.2	0.9	238	12.2	100.0	0.0	0.0
13	mar. 06, 2002	8	849	23	3	7.6	0.9	326	15.0	99.8	0.2	0.0
14	mar. 14, 2002	8	369	18	5	3.3	0.9	44	6.6	99.2	0.8	0.0
15	mar. 22, 2002	8	218	8	4	2.6	1.2	32	6.6	99.5	0.2	0.2
16	mar. 30, 2002	8	258	10	4	2.5	1.0	43	6.8	99.3	0.7	0.0
17	abr. 08, 2002	10	257	9	3	2.3	0.9	32	4.8	99.5	0.2	0.2
18	may. 08, 2002	50	118	5	4	1.2	1.0	8	1.2	99.8	0.0	0.2
19	jun. 29, 2002	54	0	0	4	0.0	1.0	-		-	-	
20	ago. 22, 2002	55	0	0	4	0.0	1.0	-		-	-	-
21	sep. 29, 2002	20	0	0	4	0.0	1.0	-		-	-	-
Annualised values			188	17	9	2.3	1.2	62	3.3			
Annual flux			69 g m * y *	6 g m ^{-*} y ⁻¹		0.9 g m ⁻⁺ y ⁻¹		23 10° valves m ⁻² y ⁻¹	1.20 10" coccoliths m ⁻² y ⁻¹	99.7	0.2	0.1

Table 1: Daily export fluxes of total mass flux, calcium carbonate (CaCO₃), particulate
organic carbon (POC), diatom valves and coccoliths registered at the 61°S site from
November 2001 through October 2002. Mass fluxes listed as zero were too small to
measure (<1 mg).

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282 2.5 SEM analysis

As the resolution of the light microscope is insufficient to differentiate Emiliania 283 284 huxleyi morphotypes, the samples of the 2000 m trap record were analysed using Scanning Electron Microscopy. Glass cover-slips were prepared following the 285 286 decantation method outlined by Flores and Sierro (1997). The dried cover-slips were 287 mounted on aluminium stubs and coated in gold. A EVO HD25 SEM (Carl Zeiss) was 288 used to determine the morphotype of Emiliania huxleyi coccoliths found in the samples. 289 Due to the large abundance of diatom valves and the scarcity of coccoliths in the samples, a compromise between number of identified coccoliths and time spent had to be reached. 290 291 Therefore, a target minimum of thirty Emiliania huxleyi coccoliths per sample were 292 identified. The taxonomic concepts of Young and Westbroek (1991), Young et al. (2003),





293 Cubillos et al. (2007) and Hagino et al. (2011) were followed to classify the *Emiliania*

- *huxleyi* coccoliths into morphotypes.
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296 **2.6 C-Calcita analyses**

The glass slides used for coccolith counts were also analysed for coccolith mass 297 298 and size measurements using a with a Nikon Eclipse LV100 POL polarized light 299 microscope equipped with circular polarization and a Nikon DS-Fi1 8-bit colour digital 300 camera. Calibration images were performed on an apical rhabdolith of the genus 301 Acanthoica collected by a sediment trap at the 47°S site (46°48'S, 142°6'E), located in the Australian sector of the Subantarctic Zone. Camera parameters and microscope light 302 303 settings were maintained constant throughout the imaging session. Depending on 304 coccolith concentration, between 13-28 random fields of view per sample were 305 photographed. The images were then analysed by the image processing software C-306 Calcita (Fuertes et al., 2014). The output files for single E. huxleyi coccoliths were visually selected. Length and weight measurements were automatically performed by C-307 308 Calcita software. A total of 2328 coccoliths were analysed with a minimum of 50 309 coccoliths per sample. For more methodological details see Fuertes et al. (2014).

An estimated range of annual contributions of coccoliths to total CaCO₃ export was calculated for the 2000 m trap record by multiplying the coccolith flux of each sampling interval by the maximum and minimum standard deviations of coccolith weight values measured on each sample. Then, the minimum and maximum estimates of coccolith-CaCO₃ fluxes for each sampling interval (i.e. cup) were used to estimate the minimum and maximum annual contribution of coccoliths to total carbonate following the same procedure as for the annual coccolith fluxes.

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318 2.7 Satellite imagery, meteorological and oceanographic data

Weekly mean sea surface temperatures (SST) for the 2001-2002 interval were obtained from the NOAA Optimum Interpolation Sea Surface Temperature Analysis database (Reynolds et al., 2002). Seasonal SST variation range was low, with maximum SSTs of 2.94 °C observed during March 2002 and minimum of 0.12 °C, in early October 2002. SST variations mirrored changes in the vertical structure of the water column temperature profile (Fig. 4) that displayed vertical homogeneity of the water column in autumn and winter and a seasonal thermocline during the austral summer (Fig. 2b).





326 Photosynthetically active radiation (PAR), monthly chlorophyll-a concentration and particulate inorganic carbon (PIC) concentration estimates were obtained from 327 NASA's Giovanni program (Acker and Leptoukh, 2007) (Fig. 4) for the region: 130°E, 328 329 62.5°S, 150°E, 59.5°S. Chlorophyll-a concentration was low throughout the year (ranging from 0.07 to 0.30 mg m⁻³) and in line with previous observations in the study region (Trull 330 et al., 2001b). Algal biomass responded rapidly to the solar radiation increase in 331 332 September 2001 and reached its highest levels in November 2001 (Fig. 4). Chlorophyll-333 a concentration declined throughout the summer, reaching negligible values in autumn and winter (i.e. from March to August 2002). Satellite-derived PIC concentration 334 335 exhibited a clear seasonal pattern similar to that of the chlorophyll-a with peak concentrations in November (up to 0.003 mol m⁻³) and values below detection limit in 336 337 winter (Fig. 4).







Figure 4: (a) Satellite-derived SST (°C), photosynthetically active radiation (Einstein m 340 ² d⁻¹), chlorophyll-a concentration (mg m⁻³) and particulate inorganic carbon (PIC) 341 342 concentration (mol m⁻³) for the period November 2001 to September 2002. It is important to note that satellite PIC concentration estimates have been reported to be biased for high 343 344 latitudes systems of the Southern Ocean where the satellite algorithm is thought to produce overestimates (Balch et al., 2016; Trull et al., 2017). Therefore PIC data 345 346 presented here should be looked with caution. (b) Temporal variability of the total mass, calcium carbonate (CaCO₃) and particulate organic carbon (POC) the < 1mm fraction at 347 2000 and 3700 m water depth from November 2001 through to November 2002 at the 348 61°S site (Rigual-Hernández et al., 2015a). Grey strips represent summer. 349





351	3. Results
352	
353	3.1 Seasonal dynamics of coccolith export fluxes
354	Coccolith fluxes showed a pronounced seasonal pattern at both sediment trap
355	depths, roughly following the chlorophyll- a dynamics in the surface layer with maximum
356	fluxes during the austral summer and minima during winter (Fig. 4 and 5). The summer
357	coccolith particle bloom exhibited a bimodal distribution with a major peak registered in
358	early January (2.2 10^9 coccoliths $m^{\text{-2}}d^{\text{-1}}$ at 2000 m) and a secondary maximum recorded
359	in mid-February (9.8 10^8 coccoliths $m^{\text{-2}}\text{d}^{\text{-1}}$). Coccolith flux was low in autumn and winter
360	(down to 7.5 10^7 coccoliths $m^{\text{-}2}d^{\text{-}1}$). Coccolith fluxes in the deeper trap (3700 m) followed
361	a similar pattern to that in the 2000 m trap with a delay of about one sampling interval.
362	The fluxes of all biogeochemical components were closely correlated (Table 2 in
363	Rigual-Hernández et al., 2015a). Coccolith fluxes at both traps were broadly in line with
364	biogenic particle fluxes estimated by Rigual-Hernández et al. (2015a) showing strongest
365	correlations with Biogenic silica (R 2 = 0.86 at 2000 m and R 2 = 0.71 at 3700 m), followed
366	by PIC (R ² = 0.62 at 2000 m and R ² = 0.47 at 3700 m) and POC (R ² = $~0.56$ at 2000 m
367	and $R^2 = 0.41$ at 3700 m).





368 Coccolithophore sinking assemblages at the 61°S site were nearly monospecific,

- 369 with an overwhelming dominance of *E. huxleyi* that represented >99% of the annual
- 370 coccolith sinking assemblage at both trap depths. Background concentrations of the
- 371 species Calcidiscus leptoporus, Gephyrocapsa spp. and Helicosphaera spp. were also
- registered, together representing 0.6% and 0.3% at 2000 and 3700 m, respectively, of the



total annual coccolith fluxes (Table 1). The numbers of coccospheres found in the samples

374 were negligible in both sediment trap records.

Figure 5: Seasonal variation of total coccolith and diatom valve flux at the 2000 and 3700

376 m sediment traps at the 61° S site. Grey strips represent summer.

377

378 3.2 SEM analyses

379

380 Emiliania huxleyi coccoliths correspond to morphotype B/C, having proximal 381 shields slightly wider than the distal ones and with a central area usually filled by several (usually 5 to 11) flat, wide and thin tiles (see Plate 1, image a). Several coccoliths present 382 383 distal shields partially missing, may be due to the slender and delicate structure of the 384 laths. Distal shield measures ranged between 2 to 4,35 µm in the samples recovered from 385 the 2000 m sediment trap. It is conspicuous that most of the coccoliths display a 386 morphology which is compatible with a secondary recrystallization. Small spherules like recrystallizations are present on these coccoliths, especially on the laths (Plate 1, images 387 388 c-f). However, some coccoliths, mostly from cup 10 (February) have no spherules





covering them (Plate 1, images a and b). These coccoliths present very thin slender laths(usually from 20 to 26) and wider central areas than the coccoliths having spherules.

391

392 **3.3** Coccolith weight and length changes

393 Average coccolith weight at both sediment trap depths exhibited a clear seasonal 394 pattern with high values (2.3 and 2.1 pg/coccolith at 2000 m and 3700 m, respectively) at 395 the onset of the coccolithophorid bloom in early spring, followed by a pronounced decrease (down to 1.6 and 1.9 pg at 2000 m and 3700 m, respectively) in approximately 396 late January - early February. Average coccolith weight followed a gradual increasing 397 trend from approximately mid-February into winter, reaching values up to 2.7 pg in 398 399 August 2002 at 2000 m and up to 2.43 in May at 3700 m, respectively. Average annual 400 coccolith weight at the 61°S traps was 2.11 ± 0.96 and 2.13 ± 0.90 pg at 2000 m and 3700 m, respectively. The annual amplitude of coccolith weight was approximately 1 pg at 401 2000 m and and 0.5 pg at 3700 m. The lower annual amplitude exhibited by the coccolith 402 assemblages captured at the 3700 trap is attributed to the lower sampling duration at that 403 404 depth over the winter season.

Mean coccolith length was greatest in early spring 2001 (3.1 and 3.2 μm at 2000 and 3700 m, respectively), followed by a decrease in early summer (down to 2.8 and 2.9 μm at 2000 and 3700 m, respectively) (Fig. 6). From late February coccolith length increased again reaching the highest values of the record in winter 2002 (up to 3.2 and 3.3 μm at 2000 and 3700 m, respectively).

410 Seasonal variations of coccolith length and weight exhibited a strong correlation 411 at both depths ($R^2 = 0.84$, n = 20 at 2000 m; $R^2 = 0.61$, n = 16 at 3700m), indicating a 412 clear, dependable relationship between the two variables.







Figure 6: Mean and standard deviation of coccolith weight and length over the sediment
trap deployment period at 2000 m and 3700 m at the 61°S site. The red solid line
represents a 3-point running average. Grey strips represent summer.

418

419 4. Discussion

420 4.1 Origin, magnitude and composition of the coccolithophores

Since there is a current debate about the potential expansion of *E. huxleyi* populations south of the Polar Front during recent decades (Cubillos et al., 2007; Winter et al., 2014), it is important to evaluate the likely origins of the sinking coccolith assemblages collected at station 61°S. This assessment is particularly needed in the case of deep–moored, sediment-trap experiment because the source area of the particles collected by the traps can be as wide as hundreds of square kilometres (Buesseler et al., 2007).

Several lines of evidence strongly suggest that the coccolithophore fluxes registered by the traps were produced in waters of the Antarctic Zone. Firstly, the mooring was deployed in a quiescent area of the AZ-S (Trull et al., 2001b), between the stronger flows associated with the southern branch of the PF and the SACCF (Fig. 1). The relatively weak currents around the sediment trap location greatly reduce the area of likely origins of the particles intercepted by the traps, i.e. the statistical funnel (Siegel and





434 Deuser, 1997; Siegel et al., 2008). Moreover, the large magnitude of the coccolith export fluxes at both depths, plus the long duration of the "coccolith particle bloom" (about 3 435 436 months), rule out the likelihood of a transient lateral transport event (e.g., transport by mesoscale eddies) of a coccolithophore bloom produced in more northerly latitudes. 437 Lastly, the composition of the biogeochemical fluxes and diatom assemblages collected 438 439 by the traps are characteristic of AZ waters (Rigual-Hernández et al., 2015a), further supporting the idea that the coccolithophores captured by the traps were produced close 440 to the site. All this clearly indicates that in 2001 E. huxleyi was an established member of 441 the phytoplankton communities of the Antarctic Zone to the south of Australia. 442

The annual coccolith export to the deep ocean at the 61° S site (1.03 x 10^{11} 443 coccoliths $m^{-2} yr^{-1}$ is one sixth that registered by Wilks et al. (2017) (6.5 x 10¹¹ coccolith 444 m⁻² yr⁻¹) in the SAZ waters (station 47°S; Fig. 1) north of the study site. The lower 445 abundance of coccolithophores at the 61°S site is most likely due to the negative effects 446 447 of low temperature and low light levels on coccolithophore growth (Paasche, 2002; Boyd et al., 2010), but important also is the competitive advantage of diatoms over 448 449 coccolithophores in the silicate-rich waters of the AZ. The lower coccolithophore production in the AZ is also reflected in the lower carbonate export at this site, i.e. 6 g m⁻ 450 2 y⁻¹ versus 10-13 g m⁻² y⁻¹ at the 47°S site (Rigual-Hernández et al., 2015b; Wilks et al., 451 452 2017). The non-proportional latitudinal change in coccolith and carbonate fluxes (i.e. 453 sixfold versus twofold changes, respectively) is most likely due to variations in the contribution of heterotrophic calcifiers (i.e. foraminifers and pteropods) to total carbonate 454 export. There are also differences in the carbonate content per coccolith of the 455 456 coccolithophore species and the morphotypes of *E. huxleyi* dwelling in each zonal system. 457 Indeed, mean coccolith weight can vary up to two orders of magnitude between small 458 species such as E. huxleyi (2-3.5 pg) and large and heavily calcified taxa such as 459 Coccolithus pelagicus (~150 pg) (Giraudeau and Beaufort, 2007). Intraspecific size 460 variability is also common in most coccolithophore species, mainly due to growth 461 variations driven by different environmental factors and by genotypic variability (e.g. 462 Knappertsbusch et al., 1997; Poulton et al., 2011).

Recognizing the significant genetical variability they found between Southern Ocean populations of morphotypes A and B/C, Cook et al. (2011) classified these morphotypes as *E. huxleyi* var. *huxleyi* and *E. huxleyi* var. *aurorae*, respectively. Since only morphotype B/C had been reported at and south of the Antarctic Polar Front, Cook et al. (2013) concluded that the rapid drop in water temperature occurring at the Antarctic





Polar Front may act as an open-ocean barrier to gene flow between these the two Southern
Ocean *E. huxleyi* morphotypes/varieties. The monospecific coccolith assemblages of *E. huxleyi* morphotype B/C collected by the 61°S site traps (Plate 1) are consistent with those
studies and supports the idea that the physiological differences in light-harvesting
pigments of morphotype B/C compared to other *E. huxleyi* varieties (Cook et al., 2011)
may represent a critical ecological advantage in the cold and low-light waters of the AZ
south of Australia.

475



476

Plate 1: SEM photos showcasing the different morphologies of *Emiliania huxleyi*morphotype B/C coccoliths found in the sediment traps of the 61°S site. Scale bars =1

479

μm.

480

481 4.2 Seasonal dynamics of the calcareous and siliceous phytoplankton fluxes

The eight-day sampling resolution during spring and summer enabled us to monitor the detailed temporal dynamics of phytoplankton fluxes at the 61°S site. Comparison of satellite-derived PIC and Chl-*a* concentrations for the study region with coccolith fluxes registered by the sediment trap suggests a time lag of about two months between their surface maxima and peak coccolith fluxes registered by the shallower trap (Fig. 4). Therefore, the growth phase of the *E. huxleyi* bloom probably took place between





October and December 2001, a period characterized by very low SSTs (0.1-0.9 °C). It
was before development of any significant stratification in the upper water column (Fig.
2b and 4a). These observations indicate that the very cold temperatures (near 0°C) and
strong mixing of the water column in the Antarctic waters during spring are not an
impediment for the development of an *E. huxleyi* bloom.

493 The onset of seasonal increase in coccolithophore arrivals in the traps occurred at the same time as that of diatoms, suggesting a rapid response of both phytoplankton 494 groups to enhanced light levels. Although both coccolith and diatom fluxes exhibited a 495 pronounced and nearly parallel increase throughout December (Fig. 5), coccolith fluxes 496 peaked one week later than those of diatoms. A similar succession was observed in late 497 summer, when coccoliths displayed a secondary flux maximum, one sampling interval 498 later (8 days) than that of diatoms (Fig. 5). These observations agree with the bloom-499 500 dynamics scheme proposed by Barber and Hiscock (2006) (the so-called coexistence 501 theory), in that neither phytoplankton group seems to outcompete the other during the development of the bloom. Interestingly, diatoms seem to decline earlier than 502 503 coccolithophores, a feature often (but not always) observed in other parts of the world ocean (e.g. Margalef, 1978; Holligan et al., 1983; Lochte et al., 1993; Sieracki et al., 1993; 504 505 Thunell et al., 1996; Balch, 2004). Indeed, a recent study of the phenological characteristics of coccolithophore blooms by Hopkins et al. (2015) concluded that they 506 often follow those of diatoms in many regions, the sequencing driven by increasing 507 stabilization and/or nutrient depletion (mainly silicate and/or Fe, and possibly also 508 509 favoured by associated increase of carbonate saturation; Merico et al, 2004) of the surface 510 layer.

511 Lack of nutrient and mixed-layer-depth measurements during the sediment trap 512 deployment precludes us from establishing robust links between changes in physical and chemical parameters in the upper water column and the observed phytoplankton 513 514 succession. Nonetheless, some shipboard observations of mixed-layer properties from 515 years previous to the sediment trap deployment (Fig. 2; Trull et al., 2001b) can provide 516 some insight about the mechanisms driving the phytoplankton succession. Macronutrient measurements indicate that, although considerable nutrient draw-down often occurs by 517 518 mid-summer, the AZ-S waters never reach potentially limiting concentrations (i.e. below 519 10 µM) of silicate, nitrate or phosphate (Fig. 2a; Trull et al., 2001b). Thus, macronutrient limitation was not a likely driver of the observed phytoplankton succession at the 61°S 520 site traps. Iron levels in the AZ-S, on the other hand, are low year-round (0.1-0.2 nM; 521





Boyd et al., 2000b; Sohrin et al., 2000) and exhibit clear seasonality in the AZ (Tagliabue
et al., 2014). So, iron availability does represent a potential driver for the observed
phytoplankton succession. Indeed, laboratory experiments have shown that *E. huxleyi* has
lower minimum Fe requirements for growth than oceanic diatoms (Brand et al., 1983;
Muggli and Harrison, 1997). This physiology likely provides an ecological advantage
over diatoms in the later stages of the spring-summer bloom, when most iron has been
stripped from the mixed layer.

529 In regard to the mechanism underlying the second diatom-coccolith succession 530 observed at both depths in February (Fig. 5), it is possible that a vertical mixing event as frequently reported in the AZ (e.g. Brzezinski et al., 2001) - supplied waters rich in 531 532 iron and macronutrients to the euphotic zone, resetting the phytoplankton succession. Alternatively, the part of the *Emiliania huxleyi* populations accumulated at or just above 533 534 the nutricline may have increased using the iron moved by diapycnal diffusion through 535 the pycnocline (Tagliabue et al., 2014). Their deposition in February could have been triggered by a drop of the light levels (Fig. 4). This second hypothesis is also consistent 536 537 with the following observations: (1) the presence of a sub-surface chlorophyll-a 538 maximum in the study region during spring and summer (Parslow et al., 2001; Trull et 539 al., 2001b); (2) reports of high E. huxleyi cell accumulations associated with the nutricline 540 in other settings of the world ocean (Beaufort et al., 2008; Henderiks et al., 2012) and (3) 541 peak annual sedimentation in late February of the diatom Thalassiothrix antarctica (Rigual-Hernández et al., 2015a), a typical component of the "shade flora" (Kemp et al., 542 2000; Quéguiner, 2013). Further sampling and taxonomic analysis of the vertical 543 544 distributions of phytoplankton in the AZ south of Australia are required to asses these 545 hypotheses.

546

547 4.3 Seasonal variability in coccolith calcification

548 Two main factors have been proposed as driving seasonal changes in coccolith 549 weights of *E. huxleyi*: a seasonal shift in the dominant morphotypes/ecotypes — each with a different degrees of calcification (Poulton et al., 2011) — and the physiological 550 response of a given morphotype to the seasonal variation of environmental parameters 551 552 (e.g. Smith et al., 2012; Meier et al., 2014). SEM analysis of the 61°S sediment trap samples revealed that only morphotype B/C, sensu Young et al. (2003), thrives in the AZ-553 S waters south of Tasmania. That is consistent with a report by Cubillos et al. (2007) of 554 dominance of B/C south 50°S. Therefore, a seasonal shift in the dominant morphotype 555





can be ruled out in respect to changing coccolith weight. The observed decrease in 556 coccolith weight could have been caused by a change in coccolith calcification or 557 558 reduction in coccolith dimensions. Young and Ziveri (2000) showed that coccolith weight is approximately linearly correlated to the cube of coccolith length. Applying that, the 559 decrease in length by 7.5% (a reduction to 92.5%) observed from the pre-bloom to the 560 summer bloom in the 2000 m traps (i.e. difference in minimum coccolith lengths in cups 561 5 and 8) corresponds to a coccolith weight loss of 21% ($0.925^3 \approx 0.79$). That is similar to 562 563 the observed weight reduction in the 2000 m trap between the pre-bloom and summer 564 bloom coccolith assemblages (16.2 - 27.6%, respectively Fig. 6). When the linear correlation between coccolith length and weight proposed by Young and Ziveri (2000) is 565 566 also applied to the 3700 m trap coccoliths, the predicted reduction of coccolith weight 567 between the pre-bloom and bloom assemblages is 12%. That is again very similar to the 568 reduction in coccolith weight observed in the E. huxleyi coccoliths intercepted by the 569 3700 trap (10%). It is strongly suggested that the seasonal changes in coccolith weight at the 61°S site were mainly driven by changes in coccolith length and were not due to 570 571 significant changes in their degrees of calcification.

572 Laboratory, mesocosm and field studies have shown that multiple environmental 573 factors including irradiance, temperature, macronutrient concentrations and iron 574 availability affect coccolith formation by E. huxleyi cells (e.g. Paasche, 2002; Zondervan, 2007; Langer and Benner, 2009; Feng et al., 2017). Since calcification in E. huxleyi is a 575 light-dependant process (Paasche, 1999, 2002), the observed decrease in coccolith weight 576 577 during summer in both traps was somewhat unexpected. Some field experiments have 578 shown that calcification in coccolithophores can occur at low light levels, or even in the 579 absence of light (e.g. van der Wal et al., 1994). However, it is often reduced compared to 580 that at higher irradiance levels (Zondervan, 2007).

581 In terms of temperature effects, Feng et al. (2017) showed that optimal 582 temperature for calcification on E. huxleyi cultures (morphotype A, strain NIWA1108) 583 was ~20°C, while temperatures below 10° C resulted in a dramatic reduction of calcification rates and severe malformations of coccoliths, such as incomplete distal 584 585 shield elements. Although E. huxleyi morphotype B/C found at the $61^{\circ}S$ site likely 586 represents an ecotype more tolerant to low temperatures than morphotype A (Cubillos et 587 al., 2007; Cook et al., 2013), the frequent variations in the structure of the coccoliths (e.g. incomplete distal shield elements; Plate 1) captured by the traps suggest some degree of 588 589 low-temperature stress at the 61°S site. Despite the important role of temperature in





coccolithophore growth (Paasche, 2002), enhanced summer SSTs may lead to an increase
in coccolith calcification, a response opposite to that observed at both traps. Therefore,
it is unlikely that seasonal SST variations at the 61°S are behind the observed variability
in coccolithophore weight.

In regard to the possible impact of macronutrient concentrations on coccolith weight, both nitrate and phosphate are known to have a pronounced effect on coccolith calcite content and morphology (Zondervan, 2007). However, as mentioned previously, none of these macronutrients reach limiting concentrations throughout the annual cycle in the AZ (Fig. 2; Trull et al., 2001). and, therefore, their influence in the calcification of coccolithophores is likely to be low or negligible.

600 On the other hand, low iron levels have been reported to have a pronounced 601 negative effect on CaCO₃ production by *E. huxleyi* cells (Schulz et al., 2004), so it 602 represents a candidate driver of seasonal changes in coccolith weight. During winter, deep 603 water mixing re-stocks the mixed layer with iron (Tagliabue et al., 2014). As soon as light levels become sufficient for photosynthesis in early spring, phytoplankton rapidly 604 605 develops under non-limiting concentrations of macro- and micronutrients. These favourable conditions for coccolithophore growth could explain the heavier and larger 606 607 coccoliths registered in early December (Fig. 6). As the phytoplankton bloom develops, 608 the dissolved iron stock is rapidly depleted in the photic zone possibly resulting in a size 609 and weight reduction of coccoliths of the already substantial E. huxleyi populations. From late summer throughout autumn, some recycling of iron in the upper water column by 610 611 increasing summer populations of zooplankton feeding on the bloom (Tagliabue et al., 612 2014), coupled with increasing light levels and the continued shallowing of the mixed 613 layer, would allow coccolithophores to produce again longer and heavier coccoliths (Fig. 614 6).

Iron-limitation, therefore, represents the most likely environmental driving factor for the seasonal variability in coccolith weight and length of *E. huxleyi* assemblages at the 61°S site. However, we note again that the absence accompanying *in situ* measurements of chemical and physical parameters of the water column, means that control of coccolith weight by varying iron availability in the AZ remains an hypothesis needing validationby future studies.

621

622 4.4 Effects of calcite dissolution on the sinking coccolith assemblages





623 The similar average annual coccolith weight registered at both traps (2.5 pg at 624 2000 m to 2.6 pg at 3700 m) indicates that negligible coccolith dissolution occurs at meso-625 and bathypelagic depths in the AZ south of Australia. That is despite the fact that 626 coccolith sinking assemblages captured by the deeper trap were exposed to potentially intense dissolution after crossing the CSH (located at 3000 m in the study region; Fig. 2). 627 The similar coccolith values observed at both depths can be attributed to the formation of 628 629 algal and faecal aggregates in the mixed layer that include fine mineral particles (Passow and De La Rocha, 2006) and provide protection against dissolution. They also facilitate 630 rapid transport of the coccoliths down through the water column. The aggregate-631 formation hypothesis is supported by the findings of Closset et al. (2015) who estimated 632 633 that sinking rates at the 61° S site were, at least 213 m d⁻¹ during the productive period, a value consistent with the sinking rates of algal and/or faecal aggregates (Turner, 2002; 634 635 Turner, 2015).

636 Despite not finding increased dissolution with water depth between 2000 and 3700 m, it is possible that coccoliths experienced some carbonate dissolution before reaching 637 638 the traps. Milliman et al. (1999) suggested that the same biological processes that 639 facilitate aggregate formation and flocculation, such as ingestion, digestion and egestion 640 by grazers, may be responsible for significant carbonate dissolution at epipelagic depths 641 (i.e. depths shallower than 800-1000 m. Indeed, the negligible amounts of coccospheres 642 found in both traps, together with the high sinking velocities, suggest that grazing could have been an important influence on export. That supported by findings of Ebersbach et 643 al. (2011) in the PFZ north of 61°S. They documented that an important fraction of the 644 645 particles sinks from the mixed layer as faecal aggregates. However, the available data are 646 insufficient to evaluate the impact of carbonate dissolution in the upper water column.

647

648 4.5 Calcium carbonate content of *Emiliania huxleyi* coccoliths

649 A broad range of calcite contents for E. huxleyi coccoliths (1.4 - 7.0 pg) has been 650 proposed in the literature (e.g. Young and Ziveri, 2000; Beaufort, 2005; Holligan et al., 651 2010; Poulton et al., 2011). The differences in these estimates are most likely due to 652 variability in the amount of coccolith calcite between morphotypes and to the varied 653 methodological biases associated with the three main approaches for estimating coccolith 654 mass: morphometrics, regression and birefringence. Since E. huxleyi morphotype B/C is more weakly calcified than other morphotypes (Young and Ziveri, 2000) and considered 655 to be geographically restricted to the Southern Ocean (Cubillos et al., 2007; Cook et al., 656





657 2013) we limit the comparison of our results to studies conducted only in the Southern658 Ocean.

659 Average annual coccolith quotas at both trap depths at the 61°S site (2.11 ± 0.96) and 2.13 ± 0.90 pg per coccolith at 2000 m and 3700 m, respectively) are almost identical 660 to that estimated by Holligan et al. (2010) $(2.20 \pm 0.60 \text{ pg}; \text{morphotype B/C})$ in the Scotia 661 662 Sea using a regression line between the number of coccoliths against PIC. Moreover, our 663 estimates are slightly higher, but with a considerable overlap in the ranges of coccolith 664 weight, than those estimated by Poulton et al (2011) for the E. huxleyi morphotype B/C populations found in Patagonian shelf waters (1.40 \pm 0.6 pg). The greater standard 665 deviation of our data is most likely due to the time periods compared. While the average 666 coccolith weight estimated for our traps reflects an integration of the annual variability in 667 coccolith weight, the shipboard observations by Poulton et al. (2011) provide a snapshot 668 669 of the summer coccolithophore populations, that likely exhibit lower coccolith size and, 670 thus, variability.

Because our coccolith weight estimates are similar to those of Poulton et al. (2011) 671 672 and Holligan et al. (2010), we can estimate the fractional contribution of coccolithophores 673 to total carbonate production in the AZ south of Australia. Coccolithophores account for 674 approximately 2-5% of the annual deep-ocean CaCO₃ fluxes at mesopelagic depths at the 61°S site. The contribution of coccolithophores to the annual CaCO₃ budget in the AZ 675 676 south of Australia is similar to the estimate by Salter et al. (2014) for the macronutrientrich, but iron deficient M6 site in the Indian sector of the AZ (12%) and remarkably lower 677 than an estimate for the iron-fertilized station A3 over the central Kerguelen Plateau 678 (85%; Rembauville et al., 2016). Due to the different methodologies for estimating 679 680 coccolithophore contributions to carbonate production, comparison of our results with 681 these other studies should be treated with caution. While only whole coccoliths were 682 counted for our calculation, therefore providing a conservative estimate, Salter et al. 683 (2014) and Rembauville et al. (2016) estimated the weight of the $< 20 \,\mu m$ fraction using 684 inductively coupled plasma-atomic emission spectrometry. That approach often results in 685 overestimates of the coccolith contribution to bulk carbonate content. There can be nonnegligible contributions of non-coccolith fragments to the fine fraction (Giraudeau and 686 687 Beaufort, 2007). Despite the biases associated with both methodologies, the general trend 688 appears clear: the fractional contributions of coccolithophores to bulk carbonate export are lower in the iron-limited waters of the AZ compared to those in naturally iron-689 fertilized settings of the Southern Ocean. These findings underscore the secondary role 690





- of this phytoplankton group in the biological carbon pumps (both the in organic carbon
 and carbonate counter pumps) south of the PF where non-calcifying phytoplankton mainly diatoms and *Phaeocystis* largely control the biologically-mediated CO₂
 exchange between the ocean and the atmosphere.
- 695

696 Conclusions

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Analysis of the sediment trap materials captured at the 61°S site allowed for the characterization and quantification of coccolith assemblages in Australian sector of the Antarctic Zone, providing a baseline of the state of coccolithophore populations in this region against which future changes can be assessed. More specifically, our study has shown the following:

703 Coccolithophores were a consistent member of the phytoplankton communities of the 704 Antarctic Zone south of Australia in year 2001. Coccolithophore assemblages in this 705 region are monospecific being composed almost entirely of Emiliania huxleyi 706 morphotype B/C. This observation supports the hypothesis that the physiological 707 differences in light-harvesting pigments of morphotype B/C (or E. huxleyi var. 708 aurorae), compared to other Southern Ocean E. huxleyi varieties (Cook et al., 2011), 709 may represent an ecological advantage in the cold, low-light and iron-limited 710 environment of the Antarctic Zone.

The onset of the coccolithophore productive period took place at the same time as that
of diatoms, indicating that neither phytoplankton group outcompetes the other during
the development of the bloom. We speculate that the diatom-coccolithophore
succession observed during the peak phase of the productive period could result from
the lower minimum iron requirements for growth of *E. huxleyi*, a feature that may
confer a competitive advantage over diatoms.

 A decrease in coccolith weight and size during the summer months was observed at both sediment trap depths. After assessing the potential influence of several environmental parameters, increasing iron limitation seems to be the most likely candidate to drive this change. This hypothesis, however, will need to be validated in future field and laboratory culture experiments with morphotype B/C.

The similar weight of *E. huxleyi* coccolith assemblages captured by the 2000 and 3700
 m sediment traps indicates that negligible coccolith dissolution occurs during transit





- through meso- and bathypelagic depths in the study region. This is most likely due toa rapid transport of the coccoliths in algal and/or faecal aggregates.
- Coccolith weight values calculated for both sediment trap records using a birefringence-based approach were similar to previous estimates of *E. huxleyi* morphotype B/C in other Southern Ocean settings using regression and morphometric methods (Holligan et al., 2010; Poulton et al., 2011, respectively).
- Coccolithophore fluxes at the 61°S site account for only 2-5% of the annual deep ocean CaCO₃ fluxes, suggesting that heterotrophic calcifiers must represent the main
 biogenic carbonate producer in the AZ south of Australia.
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