

## ***Interactive comment on “Aphotic N<sub>2</sub> fixation along an oligotrophic to ultraoligotrophic transect in the Western Tropical South Pacific Ocean” by Mar Benavides et al.***

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In this study, our objective was to sample throughout the mesopelagic zone, not necessarily targeting any specific water masses. The depths sampled (200, 500, 650 and 800 db) were "arbitrarily" chosen according to water volume availability in deep casts during the OUTPACE cruise (note that we needed as much as 40 L per depth to perform all our analyses). Very interestingly, when examining the *nifH* sequencing results, it turned out that a specific phylotype was predominant in a given water mass (subcluster 1G in the SAMW).

Unfortunately, the coverage of our samples throughout the mesopelagic zone is not

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enough to represent all the different water masses present and to identify patterns in N<sub>2</sub> fixation activity or diversity of diazotrophs according to water mass distribution. This can be clearly seen in the T-S diagrams shown in Figure 1 (from the response to reviewers file). On the left, we present a T-S diagram of the water masses sampled during the OUTPACE cruise (as displayed in Fig. 4a in Fumenia et al., this issue). According to this T-S diagram, our N<sub>2</sub> fixation and *nifH* gene measurements (central and right figures) correspond to the lower part of the upper thermocline ( $t=24.7-25.4$ ), lower part of the lower thermocline ( $t=26.5-26.7$ ), and SAMW/AAIW ( $t=26.7-27.3$ ). No measurements are available in the two water masses of the central thermocline.

2. I am not sure what the high resolution analysis of DOM by FTICRMS adds to this manuscript. As stated in the abstract and on page 9, line 10, the n<sub>2</sub> fixation rates were not related to DOM compounds analysed by FTICRMS. The application of such techniques may have been more suitable in an incubation-type experiment, e.g. adding compounds and detecting their uptake and/or incorporation.

Our group investigated aphotic N<sub>2</sub> fixation and its relationship with DOM during two cruises in the Solomon Sea (Benavides et al., 2015) and in the Mediterranean Sea (Benavides et al., 2016), in the frame of the project DIADOM [https://cordis.europa.eu/project/rcn/187917\\_en.html](https://cordis.europa.eu/project/rcn/187917_en.html) In both cases we found positive correlations between labile compounds and N<sub>2</sub> fixation. In the OUTPACE cruise we basically followed the same sampling strategy, but did not find significant relationships between DOM composition and aphotic N<sub>2</sub> fixation. Although the FTICRMS data may itself not add much to the present study, we decided to keep it for comparison with our previous studies and to reinforce the need for a mechanistic understanding of how non-cyanobacterial diazotrophs interact with DOM in the ocean.

Benavides, M., H. Moisaner, P. Berthelot, H. Dittmar, T. Grosso, O. and Bonnet, S.: Mesopelagic N<sub>2</sub> fixation related to organic matter composition in the Solomon and Bismarck Seas (Southwest Pacific), PLoS One, 10(12), 1-19, doi:10.1371/journal.pone.0143775, 2015.

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Benavides, M., Bonnet, S., Hernández, N., Martínez-Pérez, A. M., Nieto-Cid, M., Álvarez-Salgado, X. A., Baños, I., Montero, M. F., Mazuecos, I. P., Gasol, J. M., Osterholz, H., Dittmar, T., Berman-Frank, I. and Arístegui, J.: Basin-wide N<sub>2</sub> fixation in the deep waters of the Mediterranean Sea, *Glob. Biogeochem. Cycles*, 30, 1-19, doi:10.1002/2015GB005326. Received, 2016.

3. Why would fixed N inputs add to this area only if diazotrophy is related to water masses which are moving around the ocean? Is this really only a locally important processes add N to this area only?

It is difficult to speculate here, but in principle fixed N<sub>2</sub> (into ammonium or DON) would be consumed in a short time by the in situ bacterial community. In a recent opinion paper (now in review in *Frontiers in Marine Science*), we estimate that N<sub>2</sub> fixed and eventually remineralized to nitrate in the mesopelagic zone would turn over in 4 to 43 years. Please see our response to this reviewer's comment on the same issue below.

4. Unclear why the depth is reported as dbar here. I suggest the authors change dbar to meters.

It was a general agreement among all the scientists involved in the OUTPACE cruise to use dbar in all of our publications for consistency with CTD files and easy exchange of data among groups.

Figure 1. I suggest that oxygen is reported as  $\mu\text{mol L}^{-1}$  or  $\mu\text{mol kg}^{-1}$  and not mL L<sup>-1</sup> which is an unconventional unit for oxygen on oceanography. This figures is not clear because it is not possible to see the specific rates of nitrogen fixation here. I suggest this is replotted to show the actual values for nitrogen fixation, which would be more useful considering the uniqueness of this data set.

In our previous aphotic N<sub>2</sub> fixation studies we plotted rates as sized dots with oxygen concentrations (color scale) in the background. Since no significant relationships were found between aphotic N<sub>2</sub> fixation and oxygen concentrations during the OUTPACE

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cruise, we agree with this reviewer that it may be more reasonable to plot rates in a different way. We now provide aphotic N<sub>2</sub> fixation rates as sized dots (as we find it very visual and easy to spot where activity is higher), but with actual rates superimposed in coloured numbers (see Figure 2 in the response to reviewers file).

Figure 5. This is not clear due to words in blue overlapping as well as SD5 to SD15 overlapping. Can this be replotted, e.g as colour codes?

We agree that the relative positioning of overlapping samples within the gray box was not clear, however, assigning distinct color or shape codes to the samples would not resolve this problem as some samples are directly over one another. Thus, one of the conclusions from this figure is that these samples are very similar to each other, driven by the relationship to PC1 of the DOM analysis (which they fall under). The variation among samples within the gray box is extremely small when compared to the distance to other samples, which differ based on the various factors shown. We have addressed the issue of overlapping text by removing the text from the grey box, and are now showing the sample names only in the zoom-out box on the left. Other text in the figure was also made smaller, which improved overall readability.

Figure S1. The DIN and phosphate around station 7 look odd? There is no DIN and phosphate between 400 and 1000m. Please check.

Indeed, this was an error. DIN, DIP and several variables are considered core parameters shared among all researchers participating in the OUTPACE cruise special issue in *Biogeosciences*. We have therefore decided to refer the reader to figures 5a-b in Fumenia et al.'s paper (same issue) <https://www.biogeosciences-discuss.net/bg-2017-557/bg-2017-557.pdf>, where nitrate and phosphate concentrations are shown.

Table S1 has fallen off the bottom of the page. Please explain in the legend how to interpret the numbers. Are these p values or is a high value good, i.e. means a strong relationship. What do the stars mean?

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We apologize for this. In the current version we have re-dimensioned the table so that it does not fall off the page. One asterisk means significant correlation at the 0.05 level, two asterisks mean significant correlation at the 0.01 level. This information has been provided in the table caption.

Minor details/comments: Abstract, line 29: remove 'here'. Change of tense, suggest 'we measured....and identified...'

Corrected as suggested.

This sentence is awkward 'Because non-cyanobacterial diazotrophs presumably need external dissolved organic matter (DOM) sources for their nutrition, we also identified DOM compounds using Fourier Transform Ion Cyclotron Mass Spectrometry (FTI-CRMS)' - suggest change to 'DOM sources were identified.....because non-cyans...

We agree that this sentence was rather incomplete. We have rewritten it as follows: "Because non-cyanobacterial diazotrophs presumably need external dissolved organic matter (DOM) sources for their nutrition, we also identified DOM compounds using Fourier Transform Ion Cyclotron Resonance Mass Spectrometry (FTICRMS) with the aim of searching for relationships between the composition of DOM and non-cyanobacterial N<sub>2</sub> fixation in the aphotic ocean."

Page 2, line 1: remove majorly

We have replaced it with "mostly", in order to conserve the meaning of the sentence.

Page 2, line 8: '....that aphotic N<sub>2</sub> fixation may contribute significantly to fixed nitrogen inputs in this area.' As above....Why just this area? Considering the deep ocean consists of water masses moving water and its properties around the ocean, what would the nitrogen fixation here contribute to the N budget here only?

It is difficult to speculate here, but in principle fixed N<sub>2</sub> (into ammonium or DON) would be consumed in a short time by the in situ bacterial community. In a recent opinion paper (now in review in *Frontiers in Marine Science*), we estimate that N<sub>2</sub> fixed and

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eventually remineralized to nitrate in the mesopelagic zone would turn over in 4 to 43 years. We have however rephrased it to: "While the data available is still too scarce to elucidate the distribution and controls of mesopelagic non-cyanobacterial diazotrophs in the WTSP, their prevalence in the mesopelagic layer and the consistent detection of active N<sub>2</sub> fixation activity at all depths sampled during our study suggest that aphotic N<sub>2</sub> fixation may contribute significantly to fixed nitrogen inputs in this area and/or areas downstream of water mass circulation."

Page 3: Line 17: the N<sub>2</sub> fixation rate should be removed as a volumetric rate rather than integrated rate. For example, it may only be high because it is integrated over a thick layer of the ocean?

With this sentence we intended to highlight the importance of the WTSP as a hotspot of photic N<sub>2</sub> fixation worldwide. As noted in the text: "The WTSP has been recently recognized as a global hotspot of photic N<sub>2</sub> fixation, harboring among the highest N<sub>2</sub> fixation rates ever recorded (~600  $\mu\text{mol N m}^{-2} \text{d}^{-1}$ ; Bonnet et al., 2017), mostly attributed to *Trichodesmium* and *UCYN-B* (Berthelot et al., 2017; Bonnet et al., 2015; 2009; Stenegren et al., 2017)." These photic measurements correspond to the integration of rates obtained at 5 to 7 levels in the sunlit layer, and therefore we believe it is legitimate to present it as integrated rates.

Page 5: Line 5: 'measured the initial  $\delta^{15}\text{N}$  of N<sub>2</sub> in the incubation on each incubation bottle by membrane inlet mass spectrometry analyses (MIMS; Kana et al., 1994)' - do you mean after the addition of  $^{15}\text{N}_2$ ? Then this needs to be clearer here. But range of enrichments were you achieving here? In light of the newness of this approach, it would be appropriate to include some detail here.

MIMS samples were taken at the end of incubations. The  $^{15}\text{N}$  at% values obtained were  $7.548 \pm 0.557$  at% (Bonnet et al., 2018). We have rewritten this part of the M&M as follows: "To obtain accurate N<sub>2</sub> fixation rates we (1) measured the  $\delta^{15}\text{N}$  of background N<sub>2</sub> in the incubation on each incubation bottle by membrane inlet mass

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spectrometry analyses (MIMS; Kana et al., 1994) -the values obtained were  $7.548 \pm 0.557$  at% (Bonnet et al., 2018)-,"

Bonnet, S., Caffin, M., Berthelot, H., Grosso, O., Benavides, M., Helias-Nunige, S., Guieu, C., Stenegren, M. and Foster, R. A.: In depth characterization of diazotroph activity across the Western Tropical South Pacific hot spot of N<sub>2</sub> fixation, *Biogeosciences*, (January), 1-30, doi:10.5194/bg-2017-567, 2018.

Page 11: Note that Tricho colonies have been detected in sediment traps elsewhere, e.g. Pabortsava et al 2017 in *Nature Geosciences*

We have cited Pabortsava et al. as well.

Interactive comment on *Biogeosciences Discuss.*, <https://doi.org/10.5194/bg-2017-542>, 2018.

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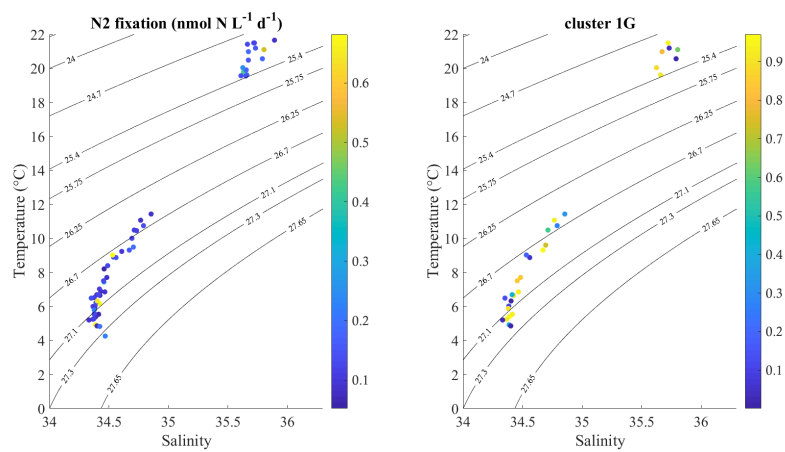


Fig. 1.

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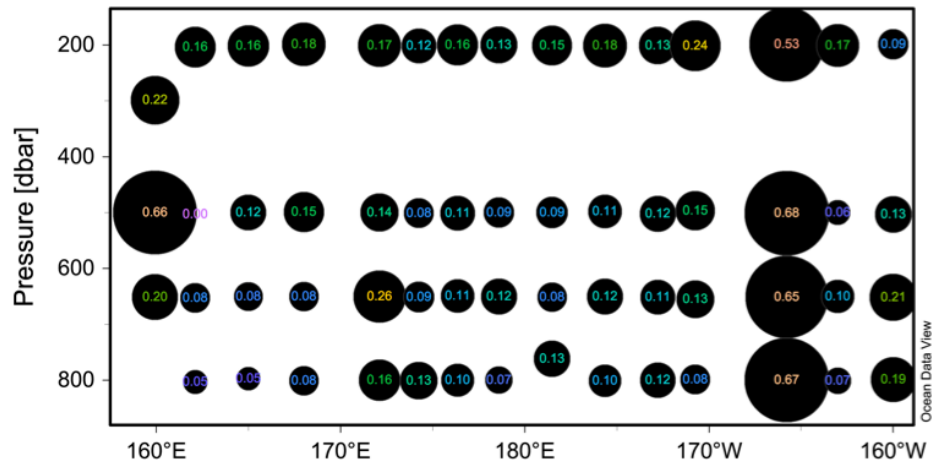


Fig. 2.