



1 **Bryophyte-dominated biological soil crusts mitigate soil**
2 **erosion in an early successional Chinese subtropical forest**

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29 **Abstract.** This study investigated the development of biological soil crust (biocrust) covers in an early
30 successional subtropical forest ecosystem and their impact on soil erosion. Within a biodiversity and ecosystem
31 functioning experiment in Southeast China (BEF China), sediment discharge and runoff measurements were
32 conducted with micro-scale runoff plots under natural rainfall and biocrust covers were surveyed over a five-year
33 period.

34 Results showed that biocrusts occurred widely in our experimental forest ecosystem and developed from initial
35 light cyanobacteria- and algae-dominated crusts to later-stage bryophyte-dominated crusts in only three years.
36 Biocrust covers were still increasing after six years of tree growth. Within later stage crusts, 25 bryophyte species
37 were determined. The development of biocrusts was significantly influenced by the surrounding vegetation cover
38 and terrain attributes. Besides high crown cover and leaf area index, the development of biocrusts was favoured
39 by low slope gradients, slope orientations towards the incident sunlight and the altitude of the research plots. Our
40 measurements showed, that bryophyte-dominated biocrusts were importantly decreasing soil erosion and more
41 effective in erosion reduction than abiotic soil surface covers. Hence, their significant role to mitigate sediment
42 discharge and runoff generation in mesic forest environments and their ability to quickly colonize gaps in higher
43 vegetation layers are of particular interest for soil erosion control in early stage forest plantations. A detailed record
44 of different biocrust species and their functional influence on soil erosion processes as well as a thorough
45 monitoring of biocrust covers under closing tree canopy in subtropical forests is required in further studies.

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61 1 Introduction

62 Biological soil crusts (hereinafter referred to as biocrusts) are a living soil cover, which plays significant functional
63 roles in many environments (Weber et al., 2016). In initial ecosystems, communities of cyanobacteria, algae, fungi,
64 lichens, bryophytes and bacteria in varying combinations are the first to colonize the substrate (Evans and
65 Johansen, 1999). Those highly specialized communities form a biological crust immediately on top or within the
66 first millimetres of the soil surface (Büdel, 2005). Biocrusts generally occur under harsh conditions of temperature
67 or light and when the cover of vascular vegetation is sparse (Allen, 2010). Therefore, biocrusts are generally
68 widespread under dryland conditions (Berkeley et al., 2005; Belnap, 2006; Büdel et al., 2009), whereas under
69 mesic conditions they mostly occur as a successional stage after disturbance or in environments under regularly
70 disturbed regimes (Büdel et al., 2014).

71 Biocrusts are generally less capable of competing with phanerogamic plants and thus the crust development is
72 limited when closed plant canopies or litter layers come into play (Belnap et al., 2003a). This limitation is due to
73 the competition for light (Malam Issa et al., 1999) and nutrients (Harper and Belnap, 2001). Nevertheless, a high
74 number of studies could not clarify the relation between biocrust cover and vascular plant cover and some studies
75 even showed a positive correlation (Belnap et al., 2003b). This coherence was explained with enhanced nutrient
76 levels provided for vascular plants growing on crusted compared to non-cruste soil surfaces (Kleiner and Harper,
77 1977; Belnap, 2002). Therefore, the improvement of soil fertility by biocrusts has been shown to be fundamental
78 for the development of vascular plant communities in some regions (St. Clair and Johansen, 1993; Harper and
79 Belnap, 2001). Biocrusts are able to quickly colonize gaps in higher vegetation layers (Belnap et al., 2003a) or
80 gaps appearing after disturbance (Dojani et al., 2011; Chiquoine et al., 2016). Such disturbances can occur e.g. in
81 forest ecosystems by treefall or after clear-cutting (Barnes and Spurr, 1998). The complete removal of a forest
82 signifies a harsh cut in vegetation development and creates a starting point for new vascular plant as well as
83 biocrust communities (Bormann et al., 1968; Keenan and Kimmins, 1993; Beck et al., 2008). Nevertheless, the
84 development of biocrusts in early successional forest ecosystems under a closing tree canopy has not been in focus
85 of research so far (Su et al., 2007; Zhang et al., 2016). Furthermore, evidence for different biocrust types in mesic
86 vegetation zones and especially from southeast Asia are rare (Büdel, 2003a; Bowker et al., 2016).

87 Functions of biocrusts have been investigated for decades, but less attention has been paid to their spatial
88 distribution and characteristics (Allen, 2010). Biocrust cover varies across different scales (from centimetres to
89 kilometres) and depends not only on the surrounding vascular vegetation cover, but also on geomorphology and
90 (micro-)topography or terrain (Evans and Johansen, 1999; Ullmann and Büdel, 2003; Kidron et al., 2009; Bowker
91 et al., 2016). Different biocrust distributions could be related to elevation and terrain-influenced microclimatic
92 gradients (Kutiel et al., 1998), different geomorphic zones (Eldridge, 1999), varying aspects (George et al., 2000)
93 or soil types (Bu et al., 2016). To our knowledge, investigations on the influence of small-scale (centimetres to
94 metres) topographic variations on biocrust development are rare and further studies will assist in the understanding
95 of their abundance and distribution (Garcia-Pichel and Belnap, 2003; Bu et al., 2016; Bowker et al., 2016).
96 Furthermore, as the development of biocrusts is characterized by a high complexity and spatial heterogeneity with
97 many micro-climatic and micro-environmental factors, it is of great significance to conduct comparative studies
98 on the spatial distribution of biocrusts (Bu et al., 2013b). This is particularly true for initial forest ecosystems
99 (Weber et al., 2016).



100 Moreover, biocrusts have been recognized as having major influences on terrestrial ecosystems (Buscot and
101 Varma, 2005), as they protect soil surfaces against erosive forces by both wind and water (Bowker et al., 2008;
102 Zhao et al., 2014), enhance soil stability (Malam Issa et al., 2001; Warren, 2003) and influence the hydrological
103 cycle (Belnap, 2006). Nevertheless, impacts of biocrusts on soil stability and soil hydrology may differ with regard
104 to soil texture, surface roughness, water repellency and finally different crust species and developmental stages
105 (Warren, 2003; Belnap and Büdel, 2016). Furthermore, studies that directly relate biocrust cover to rates of soil
106 erosion are few (Allen, 2010) and the influence of biocrusts on sediment discharge and runoff in humid climates
107 has been largely disregarded (Belnap and Lange, 2003; Weber et al., 2016).

108 This study aims to investigate the development of biocrust cover in an early successional subtropical forest
109 ecosystem and the impact of those biocrusts on soil erosion. Therefore, interrill erosion was measured with runoff
110 plots and the occurrence, distribution and development of biocrusts was recorded. The study was conducted in an
111 experimental forest plantation (BEF China), which aims to study biodiversity and ecosystem functioning
112 relationships in southeast China (Yang et al., 2013; Bruelheide et al., 2014). The following hypotheses were
113 addressed:

114 (1) Biocrusts widely develop in early successional subtropical forest ecosystems, but crust cover decreases with
115 ongoing tree growth.

116 (2) The development of biocrusts is influenced by the surrounding vegetation cover, but also by soil and terrain
117 attributes.

118 (3) Biocrusts mitigate interrill soil erosion in early successional subtropical forest ecosystems.

119

120 **2 Material and methods**

121 2.1 Study site and experimental design

122 The study was carried out within the BEF China experiment (Bruelheide et al., 2014) in Xingangshan, Jiangxi
123 Province, PR China (29°06.450' N and 117°55.450' E). The experimental area is located in a mountainous
124 landscape at an elevation of 100 m a.s.l. to 265 m a.s.l. with slopes from 15° to 41° (Scholten et al., 2017). The
125 bedrock is non-calcareous slates weathered to saprolite and predominant soil types are Cambisols with Anthrosols
126 in downslope positions and Gleysols in valleys (Scholten et al., 2017). The mean annual temperature is 17.4 °C
127 and the annual precipitation is 1635 mm with about 50 % falling during May to August (Goebes et al., 2015). The
128 climate is typical for summer monsoon subtropical regions. The potential natural vegetation of this region is a
129 subtropical broadleaved forest with dominating evergreen species. The experimental area is structured in 566
130 research plots (25.8 m × 25.8 m each) at two sites (A and B) which were clear-cut and replanted with 400 tree
131 saplings per plot in 2009 and 2010, respectively (Yang et al., 2013). A selection of 34 research plots (VIPs, Very
132 Intensively studied Plots) was used for this study.

133 2.2 Field methods



134 Biocrust cover was determined photogrammetrically in 70 selected runoff plots (ROPs, 0.4 m × 0.4 m; Seitz et al.,
135 2015) at five timesteps (November 2011, May 2012, May 2013, May 2014 and May 2015) and general biocrust
136 types were described in the field. During the rainy season in summer 2013, an extended survey linked to soil
137 erosion measurements was conducted in 170 ROPs (see below and Table 1). At each ROP, perpendicular images
138 were taken with a single lens reflex camera system (Canon 350D, Tokio, Japan) and processed with the grid
139 quadrat method in GIMP 2.8 using a digital grid overlay with 100 subdivisions (cf. Belnap et al., 2001). Stone
140 cover and biocrust cover were separated by hue distinction. A continuous leaf litter cover, which may impede
141 analyses, was not present during measurements. Biocrusts were collected in 2013 and samples were dried at 40 °C
142 (Dörrex drying unit, Netstal, Switzerland). The identification of species was carried out by morphological
143 characteristics using a stereomicroscope (Leitz TS, Wetzlar, Germany), a transmitted-light microscope (Leitz
144 Laborlux S, Wetzlar, Germany) and ultraviolet light. Bryophytes (dominating taxa in 2013) were determined to
145 the species level, wherever possible and separated into mosses (Bischler-Causse, 1989; Moos flora of China: Gao
146 et al., 1999; 2001; 2002; 2003; 2005; 2007; 2008; 2011) and liverworts (Zhu, 2006; Söderström et al., 2016 and
147 Alfons Schäfer-Verwimp, personal communication). Comparisons were conducted with specimen hosted in the
148 herbarium of the State Museum of Natural History in Stuttgart, Germany (Herbarium STU).

149 Sediment discharge and surface runoff were measured within 34 research plots on five micro-scale ROPs each in
150 summer 2013 (n=170, Table 1). After four timesteps, 334 valid ROP measurements entered the analysis (for
151 detailed information see Seitz et al., 2016). Sediment discharge was sampled, dried at 40 °C and weighed, whereas
152 surface runoff and rainfall amount were measured in situ. At every ROP, crown cover and leaf area index (LAI)
153 were measured with a fish-eye camera system (Nikon D100 with Nikon AF G DX 180°, Tokio, Japan) and
154 calculated with HemiView V.8 (Delta-T devices, Cambridge, UK). Measurements of tree height and crown width
155 were provided by Li et al. (2014) at research plot scale (n=34). Tree species richness and tree composition resulted
156 from the experimental setup of BEF China (Bruehlheide et al., 2014).

157 Soil attributes (Table 1) were determined for every research plot (n=34) using pooled samples from nine point
158 measurements each. Soil pH was measured in KCl (WTW pH-meter with Sentix electrodes, Weilheim, Germany),
159 bulk soil density was determined by the mass-per-volume method and total organic carbon (TOC) was measured
160 using heat combustion (Elementar Vario EL III, Hanau, Germany). Soil organic matter (SOM) was calculated by
161 multiplying TOC with the factor 2 (Pribyl, 2010).

162 2.3 Digital terrain analysis

163 Terrain attributes (Table 1) were derived from a digital elevation model (DEM, Scholten et al., 2017) at research
164 plot scale (n=34). Attributes were the terrain ruggedness index (TRI, Riley et al., 1999) to describe the
165 heterogeneity of the terrain, the Monte-Carlo based flow accumulation (MCCA, Behrens et al., 2008) to diagnose
166 terrain driven water availability, altitude above sea level to address elevation effects and the eastness and the
167 northness (Roberts, 1986) to describe plant related climatic conditions. Those terrain attributes cover major
168 landscape features of the experimental area and were not correlated. Slope was additionally measured with an
169 inclinometer at every ROP (n=170, see Seitz et al., 2016).

170



171 [Table 1]

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173 2.4 Statistical methods

174 The temporal development of biocrust covers (1) from 2011 to 2015 was assessed at five timesteps within 70 ROPs
175 (see above) by an analysis of variance (ANOVA) and Tukey's Honestly Significant Difference (HSD) test (n=350).

176 The influences of vegetation, soil and topographic attributes on biocrust cover (2) in 170 ROPs (see above) were
177 assessed by linear mixed effects (LME) models (n=334). Crown cover, bulk soil density, SOM, pH, altitude, slope,
178 MCCA, TRI, eastness, northness and tree species richness were fitted as fixed effects and biocrust cover as
179 response variable. The attributes were tested with Pearson's correlation coefficient before fitting. LAI was fitted
180 individually in exchange to crown cover due to multi-collinearity. Experimental site and research plot were fitted
181 as random effects and hypotheses were tested with an ANOVA type 1 with Satterthwaite approximation for
182 degrees of freedom.

183 The influences on soil erosion (3) were assessed by LME models with restricted maximum likelihood (n=334) and
184 sediment discharge and surface runoff as response variables, respectively. Crown cover, slope, surface cover,
185 SOM, rainfall amount and tree species richness were fitted as fixed effects. Surface cover was than split into
186 surface cover by biocrusts and by stones, which entered the analysis as fixed conjoined factors. Precipitation events
187 nested in plot, tree species composition, experimental site and ROP nested in plot were fitted as random effects.
188 Attributes were not correlated. The hypothesis was tested with an ANOVA type 1 with Satterthwaite
189 approximation for degrees of freedom. Moreover, the Wilcoxon rank sum test was applied to test for differences
190 between biocrust cover and stone cover on sediment discharge and surface runoff. Therefore, the dataset was split
191 into data points where biocrust cover exceeds stone cover (n=281) and data points where stone cover exceeds
192 biocrust cover (n=53).

193 All response variables were log-transformed before modelling and analyses were performed with R 3.1.2 (R Core
194 Team, 2014). LME modelling was conducted with "lmerTest" (Kuznetsova et al., 2014) and rank sum tests with
195 "exactRankTests" (Hothorn and Hornik, 2015). Figures were designed with "ggplot2" (Wickham, 2009).

196

197 **3 Results**

198 3.1 Temporal development of biocrust cover

199 Biocrusts were detected in 94 % of all ROPs and their cover within ROPs ranged between 1 % and 88 % over all
200 five years. The mean biocrust cover of all ROPs more than tripled from their installation in 2011 to the last
201 measurement in 2015 (Fig. 1). This increase was significant from 2011 to 2015 and from 2012 to 2013, 2013 to
202 2014 and 2014 to 2015 (p<0.001).

203

204 [Figure 1]



205 Whereas a clear bryophyte-dominance of biocrusts was evident at the time of sampling in 2013, different
206 successional stages were identified in the field and on ROP photos from 2011 to 2015 (Fig. 2). In 2011, a smooth,
207 light cyanobacteria- and algae-dominated crust with traces of lichens and few bryophytes indicated a pioneer stage
208 of biocrust development (cf. Colesie et al., 2016). In 2013, 25 moss and liverwort species were classified (Table
209 2) and formed a bryophyte-dominated crust with cyanobacteria, algae, lichens and micro-fungi still observed in
210 minor numbers within ROPs. The same was true in 2015, but first evidence of vascular plants (*Selaginella* and
211 *Poaceae*) indicated a further advanced biocrust stage.

212

213 [**Figure 2**]

214

215 [**Table 2**]

216

217 3.2 The influence of vegetation, soil and terrain on biocrust cover

218 The development of biocrust cover in 2013 was positively influenced by crown cover and LAI as attributes of the
219 surrounding vegetation (Table 3). Furthermore, it was negatively affected by slope and northness and slightly
220 positively affected by the altitude of the research plots (Table 3). Further soil or terrain attributes did not affect
221 biocrust cover.

222

223 [**Table 3**]

224

225 3.3 The impact of biocrust cover on soil erosion

226 Both biocrust and stone cover, as well as soil surface cover (comprising both biocrust and stone cover) negatively
227 affected sediment discharge ($p < 0.001$, Table 4). In addition, soil surface cover negatively affected surface runoff
228 ($p = 0.003$). However, only biocrust but not stone cover mediated the effect of runoff. Furthermore, crown cover,
229 SOM and rainfall amount affected sediment discharge, whereas runoff was affected by crown cover and rainfall
230 amount. ROPs primarily influenced by stone cover showed higher sediment discharge and surface runoff than
231 those primarily influenced by biocrust cover (Fig. 3).

232

233 [**Table 4**]

234



235 [Figure 3]

236

237 4 Discussion

238 4.1 Temporal development of biocrust cover

239 Biocrusts were detected widely within the experiment and occupied a considerable area in the interspaces of the
240 growing tree community. Thus, the first part of hypothesis 1 can be confirmed, as biocrusts colonized the newly
241 created habitats originating from the disturbance by forest clear-cutting and weeding (Bruehlheide et al., 2014). The
242 deforestation provided a local arid micro-environment which initiated early biocrust development (Büdel, 2003b).
243 At this early stage of the ecosystem, biocrusts were highly competitive and formed a pioneer vegetation (Langhans
244 et al., 2009), which then prepared the upper soil layer for further growth of vascular plants by the input of carbon
245 and nitrogen (West, 1990; Evans and Johansen, 1999). Biocrusts generally facilitate the succession of vascular
246 plants to more advanced stages (Bowker, 2007). Accordingly, tree growth provide shade and protection from wind,
247 which then leads to advancement in biocrust development. The bryophyte-dominance of biocrusts in 2013
248 documented this development into a later and somewhat moister successional stage (Williams and Büdel, 2012).
249 Biocrusts are often dominated by one organism group, with cyanobacterial crusts being indicators for early stage
250 crusts and drier conditions (Malam Issa et al., 1999; Malam Issa et al., 2007). The successional development of
251 biocrusts within the BEF China experiment seemed to be faster than e.g. reported by Zhao et al. (2010) for the
252 Chinese Loess Plateau, who claimed biocrusts from a 3-year old site as early successional dominated by
253 cyanobacteria. Bryophytes in biocrusts have received comparatively little attention and in Asia only 23 different
254 species have been reported up to now (Seppelt et al., 2016). Thus, this study with 25 recorded moss and liverwort
255 species, most of them being new records within Asian biocrusts (Burkhard Büdel, personal communication)
256 substantially increases the knowledge on biocrusts of this region.

257 Nevertheless, the extent of biocrusts was strongly increasing since 2012 i.e. three years after tree replantation and
258 still gaining coverage in the sixth year after our experimental setup. Thus, the second part of hypothesis 1 has to
259 be rejected. Even if single trees were already up to 7.4 m high (Li et al., 2014) and LAI was up to 5.35 in 2013,
260 biocrusts still remained competitive within the forest ecosystem. Moreover, increasing crown cover and LAI
261 seemed to foster the development of bryophyte-dominated biocrusts at this ecological stage. It is assumed that
262 with continuing tree growth the biocrust communities will adapt and the composition of moss and liverwort species
263 will further change (Eldridge and Tozer, 1997). Thus, bryophytes will likely switch from species favouring sunny
264 habitats to more shade-tolerant species (Zhao et al., 2010; Müller et al., 2016). In addition, there might also be a
265 reduction in bryophyte diversity due to shady conditions, where only smaller number of species could prevail.
266 Nevertheless, adapting biocrusts seem to be able to coexist widely with vascular plants under a nearly closed tree
267 canopy, even if it is assumed that biocrust cover will decrease in later years with an increasing leaf litter layer
268 (Belnap and Lange, 2003). In this context, the ecological roles of biocrusts in succession models and plant
269 restoration are of interest (Hawkes, 2004; Bowker, 2007). Restoration of biocrusts in disturbed ecosystems could
270 be a practical approach to improve and accelerate plant community rehabilitation after disturbance and there are
271 several projects under way to establish successful restoration techniques (Rosentreter et al., 2003; Bowker, 2007;
272 Chiquoine et al., 2016).



273 4.2 The influence of vegetation, soil and terrain on biocrust cover

274 The development of biocrusts was influenced by vegetation and terrain attributes, but not by soil attributes. Thus,
275 hypothesis 2 can be partly confirmed. As already shown before, high crown cover and LAI affected the
276 development of biocrust cover in 2013. This finding is due to the successional alteration of biocrusts towards
277 bryophyte-dominance. Mosses and liverworts profit from humid conditions and a higher protection from light
278 compared to cyanobacteria- or lichen-dominated crusts (Ponzetti and McCune, 2001; Marsh et al., 2006; Williams
279 et al., 2013). Environmental factors such as water content, light intensity and temperature influence e.g.
280 photosynthesis and respiration (Zhao et al., 2010; Weber et al., 2012).

281 Furthermore, several terrain attributes affected biocrust cover. Slope was the most prominent of those factors,
282 causing a considerable decline in biocrust cover with increasing slope, being explained by their decreasing ability
283 to fasten themselves on the soil surface at high slope angles, especially when large surface water flows occurred
284 during rainfall events (Chamizo et al., 2013; Bu et al., 2016). Northness showed a positive impact on biocrust
285 covers and indicated that slope orientations towards the incident sunlight directly influence the biocrust
286 development (Bowker et al., 2002; Zaady et al., 2007). Furthermore, biocrust development depended on the
287 altitude, probably by affecting microclimatic conditions (Kutiel et al., 1998; Chamizo et al., 2016; Bu et al., 2016).

288 Interestingly, SOM and pH did not affect biocrust cover in this study. Increased organic matter contents and acidic
289 conditions, as they were found at the experimental area (Scholten et al., 2017), generally favour the development
290 of bryophyte-dominated biocrusts (Eldridge and Tozer, 1997; Seppelt et al., 2016). Nevertheless, discrepancies
291 between the research plots were small and apparently not large enough to cause prominent differences in biocrust
292 development.

293 4.3 The impact of biocrust cover on soil erosion

294 Biocrust cover clearly mitigated interrill soil erosion in this early stage ecosystem and thus hypothesis 3 was
295 confirmed. Biocrusts attenuate the impact of raindrops on the soil surface and greatly improve its resistance against
296 soil erosion (Eldridge and Greene, 1994; Goebes et al., 2014; Zhao et al., 2014). Moreover, they have the ability
297 to glue loose soil particles by polysaccharides extruded by cyanobacteria and green algae (Buscot and Varma,
298 2005). In the current study, protonemata and rhizoids of mosses and liverworts were observed to be most effective
299 by weaving and thus fixing the first millimetres of the top soil (Bowker et al., 2008). *Pogonatum inflexum* and
300 *Atrichum subseriatum* are well known to have a positive effect on erosion control due to their sustained protonema
301 system. Furthermore, bryophytes increase the formation of humus, which in turn assists to bind primary particles
302 into aggregates (Scheffer et al., 2010; Zhang et al., 2016). Thus, biocrusts contribute to the aggregation of soil
303 particles and stabilize the upper soil surface. Furthermore, biocrusts are known to influence hydrological processes
304 such as surface runoff and infiltration rates (Belnap, 2006; Cantón et al., 2011; Chamizo et al., 2012). Just recently,
305 Chamizo et al. (2016) showed that runoff and infiltration also depend on the investigation scale. Whereas point
306 based measurements showed both increasing and decreasing runoff through biocrusts depending on the study site,
307 studies of larger scale (>2 m²) revealed that biocrusts decrease runoff generation (Chamizo et al., 2016). Moreover,
308 reducing effects on runoff are related to the biocrusts species composition (Belnap and Lange, 2003). Especially
309 bryophyte-dominated crusts appear to enhance infiltration and reduce runoff due to their rhizome system, while
310 soil erosion rates stay low (Warren, 2003; Yair et al., 2011). Field studies in Utah, USA, revealed that later stage



311 biocrusts, containing both lichens and mosses, offer more protection against soil erosion than cyanobacterial crusts
312 (Belnap and Gillette, 1997). They provide higher infiltration than biocrusts dominated by cyanobacteria (Kidron,
313 1995) and decrease the aggradation of soil pores by reducing the kinetic energy of raindrops (Eldridge and Greene,
314 1994). Moreover, biocrusts dominated by bryophytes increase surface roughness and thus slow down runoff
315 (Kidron et al., 1999; Rodríguez-Caballero et al., 2012). Furthermore, they absorb water and provide a certain water
316 storage capacity (Warren, 2003; Belnap, 2006). Especially *Leucobryum juniperoideum* is known for its water
317 absorbing capacity. Whereas a partial stone cover does not enhance infiltration, bryophyte-dominated biocrusts
318 positively influence the hydrological processes in the top soil layer regarding erosion control. Thus, they actively
319 mitigate initial soil erosion compared to abiotic components such as stones. This study showed, that biocrust covers
320 play an important role in the avoidance of severe soil erosion in early successional forest plantations. This effect
321 should be considered for the replantation of forests in regions endangered by soil erosion. Furthermore, the
322 artificial cultivation of mosses in such initial forest ecosystems could improve erosion control (Bu et al., 2013a;
323 Zhao et al., 2016). At this point, the importance of biocrusts in the rehabilitation of disturbed ecosystems comes
324 into focus again (Bowker, 2007).

325

326 5 Conclusion

327 This study investigated the development and distribution of biocrusts in an early stage subtropical forest ecosystem
328 as well as their impact on interrill soil erosion. The following conclusions were obtained:

329 (1) Biocrusts occurred widely in this early successional forest ecosystem in subtropical China. They developed
330 quickly to later-stages in this mesic environment and were already dominated by bryophytes after three years of
331 tree growth (25 bryophyte species classified). After six years of continuing canopy closure, biocrust cover was
332 still increasing. Further monitoring under closing tree canopy is of importance to detect changes in biocrust cover
333 and species composition in subtropical environments.

334 (2) The surrounding vegetation and underlying terrain affected biocrust development, whereas soil attributes did
335 not have an effect. Besides high crown cover and LAI, the development of biocrusts was favoured by low slope
336 gradients, slope orientations towards the incident sunlight and altitude. Further research appears to be necessary
337 to explain effects of terrain attributes such as aspect or elevation and effects of underlying soils and substrates.

338 (3) Soil surface cover of biocrusts largely affected soil erosion control in this early stage of the forest ecosystem.
339 Bryophyte-dominated crusts had erosion-reducing characteristics regarding both sediment discharge and surface
340 runoff. Furthermore, they were more effective to decrease soil loss compared to abiotic soil surface covers. These
341 functional properties of bryophytes with regard to soil erosion need to be considered for management practices in
342 early stage forest plantations. Further research is required on functional mechanisms of different biocrust species
343 and their impact on soil erosion processes.

344

345 Data availability

346 Data are publicly accessible and archived at the BEF China data portal (<http://china.befdata.biow.uni-leipzig.de>).



347

348 **Author contribution**

349 Steffen Seitz and Thomas Scholten designed the experiment and Steffen Seitz, Zhengshan Song, Kathrin Käppeler
350 and Carla L. Webber carried it out. Martin Nebel and Kathrin Käppeler classified biocrust types and determined
351 bryophyte species. Steffen Seitz, Philipp Goebes and Karsten Schmidt performed the statistical models. Steffen
352 Seitz and Bettina Weber prepared the manuscript with contributions from all co-authors. The authors declare that
353 they have no conflict of interest.

354

355 **Acknowledgements**

356 We are grateful to the BEF China research group and especially to our students Mario Ahner, Milan Daus, Marlana
357 Hall, Madeleine Janker, Paula Kersten, Vera Müller and Andrea Wadenstorfer for assistance in fieldwork. We also
358 thank Alfons Schäfer-Verwimp for assistance in determination of bryophytes, Karl Forchhammer for giving us
359 first insights into the world of cyanobacteria and the participants of BioCrust3 for helpful comments on the results.

360 This work was funded by the German Research Foundation (DFG FOR 891/2 and 891/3). We also benefitted from
361 travel grants by the Sino-German Centre for Research Promotion (GZ 699 and GZ 785).

362

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676 **Tables**

677 **Table 1: Erosion, soil, soil cover, vegetation and terrain attributes in runoff plots (ROPs, n=170) and on research plots**
 678 **(n=34) in Xingangshan, Jiangxi Province, PR China in 2013.**

	<i>Min</i>	<i>Mean</i>	<i>Max</i>
<i>Runoff plots (ROPs, n=170)</i>			
Sediment discharge [g]	21.6	195.5	989.0
Surface runoff [ml]	3.1	40.3	111.8
Rainfall amount [ml]	25	94	178
Slope [°]	5	29	60
Soil cover [%]	0	19	62
- Biological soil crust cover [%]	0	24	62
- Stone cover [%]	0	4	42
Crown cover [%]	0.00	0.32	1.00
Leaf area index (LAI)	0.00	0.73	5.35
<i>Research plots (n=34)</i>			
Bulk soil density [g cm ⁻²]	0.83	0.98	1.12
Soil organic matter [%]	4.2	6.5	9.7
pH (KCl)	3.24	3.66	4.00
Altitude [m]	119	167	244
MCCA	0.98	2.07	3.81
TRI	0.72	2.39	3.86
Eastness	-0.86	0.09	0.99
Northness	-0.87	0.23	0.99
Tree height [m]	1.0	2.2	7.4
Crown width [m]	0.4	1.2	3.0

679 **Soil cover: proportion of soil surface area covered by biocrusts or stones, crown cover: proportion of soil surface area**
 680 **covered by crowns of live trees, leaf area index: one-sided green leaf area per unit soil surface area, MCCA: Monte-**
 681 **Carlo based flow accumulation (Behrens), TRI: terrain ruggedness index (Riley), Eastness and Northness: state of**
 682 **being east or north (Roberts), tree height: distance from stem base to apical meristem, crown width: length of longest**
 683 **spread from edge to edge across the crown**

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685 **Table 2: Liverwort and moss species sampled in the BEF China experiment in Xingangshan, Jiangxi Province, PR**
 686 **China in 2013.**

Family	Species	Author
<u>Liverworts</u>		
<i>Calypogeiaceae</i>	<i>Calypogeia fissa</i>	(L.) Raddi
<i>Conocephalaceae</i>	<i>Conocephallum salebrosum</i>	Szweyk., Buczk. et Odrzyk.
<i>Lophocoleaceae</i>	<i>Heteroscyphus zollingeri</i>	(Gottsche) Schiffn.
<i>Marchantiaceae</i>	<i>Marchantia emarginata</i>	Reinw., Blume et Nees
<i>Acrobolbaceae</i>	<i>Notoscyphus lutescens</i>	(Lehm. et Lindenb.) Mitt.
<u>Mosses</u>		
<i>Polytrichaceae</i>	<i>Atrichum subserratum</i>	(Harv. et Hook. f.) Mitt.
<i>Pottiaceae</i>	<i>Barbula unguiculata</i>	Hedw.
<i>Bryaceae</i>	<i>Bryum argenteum</i>	Hedw.
<i>Leucobryaceae</i>	<i>Campylopus atrovirens</i>	De Not.
<i>Dicranellaceae</i>	<i>Dicranella heteromalla</i>	(Hedw.) Schimp.
<i>Pottiaceae</i>	<i>Didymodon constrictus</i>	(Mitt.) K. Saito
<i>Pottiaceae</i>	<i>Didymodon ditrichoides</i>	(Broth.) X.J. Li et S. He
<i>Ditrichaceae</i>	<i>Ditrichum pallidum</i>	(Hedw.) Hampe
<i>Entodontaceae</i>	<i>Entodon spec.</i>	sterile
<i>Hypnaceae</i>	<i>Hypnum cupressiforme</i>	Hedw.
<i>Hypnaceae</i>	<i>Hypnum macrogynum</i>	Besch.
<i>Leucobryaceae</i>	<i>Leucobryum juniperoideum</i>	(Brid.) Müll. Hal.
<i>Bartramiaceae</i>	<i>Philonotis marchica</i>	(Hedw.) Brid.
<i>Bartramiaceae</i>	<i>Philonotis mollis</i>	(Dozy et Molck.) Mitt.
<i>Bartramiaceae</i>	<i>Philonotis roylei</i>	(Hook. f.) Mitt.
<i>Mniaceae</i>	<i>Plagiommium acutum</i>	(Lindb.) T.J. Kop.
<i>Polytrichaceae</i>	<i>Pogonatum inflexum</i>	(Lindb.) Sande Lac.
<i>Thuidiaceae</i>	<i>Thuidium glaucinoides</i>	Broth.
<i>Mniaceae</i>	<i>Trachycystis microphylla</i>	(Dozy et Molck.) Lindb.
<i>Pottiaceae</i>	<i>Trichostomum crispulum</i>	Bruch

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694 **Table 3: Results of the final linear mixed effects (LME) model for vegetation, soil and terrain attributes on biological**
 695 **soil crust cover in Xingangshan, Jiangxi Province, PR China in 2013 (***: $p < 0.001$; **: $p < 0.01$; *: $p < 0.05$; .: $p <$**
 696 **0.1; ns: not significant; n=215).**

Biological soil crust cover				
	denDF	F	Pr	estim.
<i>Fixed effects</i>				
Crown cover	136	12.9	***	10.8
Bulk soil density	37	0.03	ns	3.65
SOM	39	1.11	ns	(-)0.95
pH (KCl)	38	2.47	ns	(-)16.7
Altitude	37	3.69	.	0.80
Slope	191	7.53	**	(-)2.72
MCCA	39	0.02	ns	0.33
TRI	38	0.04	ns	(-)0.40
Eastness	37	2.73	ns	(-)4.23
Northness	37	9.14	**	5.99
Tree species richness	38	1.22	ns	(-)0.27
<i>Random effects</i>				
		<i>SD</i>	<i>Variance</i>	
Site		<0.01	<0.01	
Plot		<0.01	<0.01	
<i>Vegetation attribute fitted in exchange to crown cover</i>				
Leaf area index	107	42.8	***	5.98

697 **SOM: soil organic matter; MCCA: monte carlo based flow accumulation; TRI: topographic roughness index; denDF:**
 698 **denominator degrees of freedom; F: F value; Pr: significance; estim.: estimates**

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705 **Table 4: Results of the final linear mixed effects (LME) models for sediment discharge and surface runoff with**
 706 **surface cover split into biological soil crust cover and stone cover in Xingangshan, Jiangxi Province, PR China in 2013**
 707 **(***: $p < 0.001$; **: $p < 0.01$; *: $p < 0.05$; .: $p < 0.1$; ns: not significant; $n=334$).**

	Sediment discharge				Surface runoff			
	den DF	F	Pr	estim.	den DF	F	Pr	estim.
<i>Fixed effects</i>								
Crown cover	130	6.53	*	(-)0.15	173	9.11	**	(-)0.14
Slope	151	1.23	ns	0.06	168	2.25	ns	(-)0.06
Surface cover								
- Biocrust	151	50.2	***	(-)0.38	159	8.11	**	(-)0.12
- Stone	136	10.3	**	(-)0.19	188	1.66	ns	(-)0.06
SOM	44	5.71	*	(-)0.08	72	2.43	ns	0.12
Rainfall	95	5.46	*	(-)0.08	302	13.2	***	0.14
Tree species richness	22	0.46	ns	0.05	68	0.11	ns	(-)0.03
<i>SD</i>								
<i>Random effects</i>								
		<i>SD</i>	<i>Variance</i>			<i>SD</i>	<i>Variance</i>	
Precip. event : plot		0.199	0.040			0.537	0.288	
Tree composition		0.292	0.085			0.000	0.000	
Site		0.466	0.217			0.443	0.196	
Plot : ROP		0.441	0.195			0.269	0.073	

708 **SOM: soil organic matter; denDF: denominator degrees of freedom; F: F value; Pr: significance; estim.: estimates**

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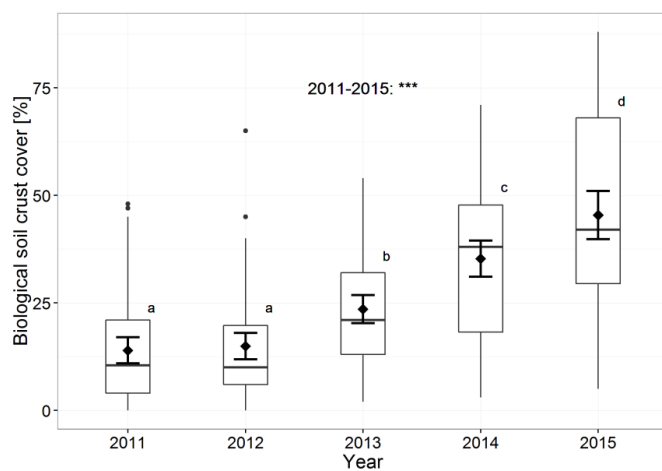
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718 **Figures**



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720 **Figure 1: The development of biological soil crust cover in runoff plots of the BEF China experiment from 2011 to**
721 **2015 in Xingangshan, Jiangxi Province, PR China (n=350). Horizontal lines within boxplot represent medians and**
722 **diamonds represent means with standard error bars. Points signify outliers and small letters significant differences**
723 **($p < 0.001$).**

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732 **Figure 2: Successional stages of biological soil crusts in two exemplary runoff plots (top row and bottom row, 0.4 m ×**
733 **0.4 m each) in 2011, 2013 and 2015 (from left to right) at the BEF China experiment in Xingangshan, Jiangxi**
734 **Province, PR China.**

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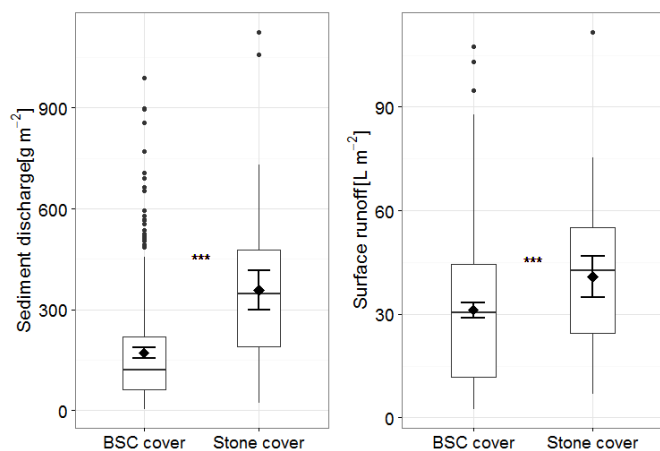
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743 **Figure 3: The influence of runoff plots dominated by biological soil crust cover (BSC, n=281) and stone cover (n=53)**

744 **on sediment discharge and surface runoff in Xingangshan, Jiangxi Province, PR China in 2013. Horizontal lines**

745 **within box plots represent median and diamonds represent mean with standard error bars.**

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