Response to reviewers

"Comparison of CO_2 and O_2 fluxes demonstrate retention of respired CO_2 in tree stems from a range of tree species"

Hilman et al. reply

We thank the reviewers for their comments. Detailed responses to the comments appear below, numbered and in **bold**. The main changes were: (1) Re-organization of the methods section and Table 1; (2) adding information about PEPC and the possible fates of the refixed carbon to the introduction and discussion; (3) Re-organization of the discussion, especially its second half, and including a short discussion on the possible effect of corticular photosynthesis (L435-441).

Reviewer 1

The biggest potential issue with this manuscript is the choice of ANOVA as an analytical approach, particularly in the use of ARQ as a response variable. Since ARQ is not measured directly, but is the ratio of the two measurements, it does not necessarily have the correct statistical properties for ANOVA. In particular, if measurement error scales with the component fluxes of ARQ, then the relative error of ARQ increases as the component fluxes decrease. This is of particular concern in light of the admission that in multiple cases, samples were excluded for having very low flux measurements, presumably approaching detection limits. I would recommend the authors consider another approach that preserves the original scaling of the measurements, such as ANCOVA or multiple regression. I would not object to summarizing some of the findings in terms of ARQ, however, as it is a useful tool for explanation.

1. **The perception that relative measurement errors increase in smaller signals is correct. However, we used ANOVA for compare measurements in which the gas fluxes were similar in magnitude, thus the relative analytical errors are of similar magnitude. In addition, our instrumental design minimizes analytical errors. Most of the ARQ results in the paper, including those ANOVA was applied with, were measured in the *Hampadah,* a closed-system that contains both CO_2 and O_2 analyzers. The inclusion of both analyzers in the same system means that temperature and pressure effects will occur simultaneously, and will be canceled while dividing the CO_2/O_2 measurements. Thus, no systematic bias in the ARQ estimation is expected. This was demonstrated in very good precision for duplicate ARQ samples (±0.01) (Hilman and Angert, 2016a). The O_2 measurement with the *Hampadah* is validated by measurement of O_2 by mass spectrometry; Licor and optode measurements are also calibrated to ensure accuracy (Hilman and Angert, 2016a). Yet, the precision in the concentrations of the gases is poorer than for ARQ and the errors for the individual gas fluxes are higher than for the ARQ. Therefore, the ARQ measurement can be regarded as a direct measurement of the ratio and thus compatible with ANOVA. As results are measured for multiple trees, the most important expression of uncertainty is how reproducible the measurement is — and how does that standard deviation across measurements compare to the expressed error for an individual measurement.**

Another potential issue is the combination of analyses across so many species and sites. While, on one hand, this a strength of the paper, there is an implicit assumption that the mechanisms are consistent across species and sites. This is not necessarily the case, especially in regard to the transport of DIC in sap, which could be effected greatly by species or wood anatomy. One may expect that this process and the contribution of transport to observed ARQ values would depend greatly on the depth of active sapwood, vessel size and other anatomical characteristics, such as medullary rays.

2. **The text was corrected according to this comment. In the revised discussion we linked wood anatomy to the contradicting literature regarding the contribution of in-stem CO_2 to surface efflux (L401-L411). Later in the discussion we further highlight that the ARQ value is probably the sum of numerous mechanisms, which might vary between species and wood anatomies (L432-435 and L441-443).**

One possible spot for improvement is connecting the putative mechanism of PEPC fixation of CO_2 with transport and canopy-level measurements of CO_2 and O_2 exchange. In particular how this could result in similar decoupling of the component fluxes in time rather than or in addition to spatial decoupling from processes such as the transport of DIC in sap.

3. **We added discussion in this regard. In L422-431 we offer two sinks for PEPC products: export of malate to the canopy where 'C4-like photosynthesis' might happen, and/or export of organic acids to the soil as root exudates. In L449-454 we predict the CO_2 and O_2 exchange expected for each of those possible sinks: ARQ >1.0 in the rhizosphere as result of organic acid catabolism, and increase of the photosynthetic oxidative ratio (O_2 produced/CO_2 consumed) as the internally transported C replaces the atmospheric CO_2 when assimilation is measured.**

Reviewer 2

This paper has an important result, which is that ARQ values lower than 1 are widespread across biomes, and these low values cannot be explained by dissolution and transport of respired CO2 in the xylem stream. Overall, while well-written overall, this paper has significant issues with organization and clarity. The introduction is compelling and reads smoothly, but the methods section in particular is difficult to follow. Additionally, the discussion section introduces a new concept to explain the results and dwells on concepts that the introduction stated were not important. There are different methods used and experiments performed at each study site, and this information is not presented logically. Table 1 was very difficult to read due to spacing within the table and it did not contain easily obtainable information about which experiments were performed at which site. I recommend reorganization of these sections.

4. **The methods section was revised as well as Table 1 according to the reviewer's suggestions. We edited section 2.2 (L164-220) by providing a list of the conducted experiments, and which questions those experiments test. The list is linked now to Table 1, which was re-designed.**

Additionally, I was unsatisfied with the PEPC explanation for the low ARQ values that was only introduced at the very end of the paper. This new concept was introduced without sufficient context, such as 'what is a reasonable value for PEPC fixation?' – there is only 1 true value presented in the table.

This concept should be introduced earlier in the paper with proper setup, because as is it feels like a surprise.

5. **An introduction to PEPC was added (L70-74), mirroring the discussion. Unfortunately, PEPC measurements in stems are extremely rare. We were able to find one more paper that enables calculation of PEPC activity that is comparable to our measurements (Ivanov et al., 2005). The calculation is included in Table 2 and results in activity similar to that in the first cited paper (Berveiller and Damesin, 2008).**

Finally, I am unconvinced by the authors' assertion that ARQ values from "instantaneous" vs. "steady state" sampling are equivalent because the regression was forced through zero and R2 was not reported, masking bias that could be present based on previous studies. The authors achieve their original objective to obtain estimates of ARQ in different biomes across seasons, but this important result was obfuscated by a complicated methods section and a scattered discussion section.

6. **The aim of Fig. 2 is to demonstrate the overall validity of this comparison. Our box-model allows comparison of ARQ values measured shortly after the beginning of chamber incubation (instantaneous) and after steady-state is reached (>day). Considering the reviewers' comment, we agree that linear regression between the ARQ determined each way is not the best way to demonstrate the adequacy of the steady-state and the instantaneous measurements, because they are not dependent-independent variables. The figure was slightly changed, and the 1:1 line (which is what we expect if the methods agree) was plotted instead of the linear fit. We additionally report the mean difference and RMSD from the 1:1 line, which are respectively 0.02 and 0.15. As we discussed in the paper (L349-358) and previously (Hilman and Angert, 2016a), the considerable scatter around the 1:1 line and the large RMSD could partly be attributed to temporal differences in ARQ during the time between the instantaneous and steady-state measurements, while the model assumes constant ARQ with time. Additionally, the precision for "instantaneous" ARQ was lower than for "steady state" values, due to smaller changes in O_2 over the shorter time periods. This may also contribute to the scatter in Fig. 2 (Hilman and Angert, 2016b).**

I recommend significant revisions to this paper that include: reorganization of methods and results sections for clarity, making the discussion mirror the introduction, and improving overall cohesiveness. The authors lay out clear objectives in the introduction, but the discussion has a lot of information in it that isn't set up in the introduction. To make this story more cohesive, the authors should keep their main objectives in mind in the revisions, introduce important concepts earlier in the paper, and make sure the discussion section follows logically from the results presented.

7. **Please see answers 4-6.**

Major comments: I recommend reorganization and attention to consistency in referring to species vs. sites. For example, Figure 4 refers to species, but when presented in the results (lines 260-263), the "Bartlett" and "Harvard" are referenced. The reader should not have to go to Table 1 for reference to understand to which panel in figure 4 the text refers. Sometimes the authors mix species and site names

in the same sites, for example in lines 317-318 when they refer to trees in Jerusalem. The reader should again, not have to refer to Table 1 to figure out which species were in Jerusalem. Site names should be consistent throughout the manuscript. Sometimes sites are referred to by name (e.g. “Hebrew University” or “Carmel Ridge”), sometimes by location (e.g. “Jerusalem” or “Brazil”, and sometimes by a more general name like “Israel”). In this example, there is no “Hebrew University” referred to in Table 1 so the reader cannot even be certain which site is being discussed when this term is used. Please be consistent throughout the manuscript with your names for each site.

8. **The study sites names are consistent now throughout the manuscript.**

You assert that ARQ values from “instantaneous” vs. “steady state” sampling are equivalent – however, the regression was forced through zero, and authors only report the slope. Forcing the regression through the origin will mask any main effect bias. There is no reason to assume the regression will go through zero, conversely, Angert et al. (2012) showed a large difference in ARQ between these two approaches (Figure 2). You state several times that the methods are equivalent and that there are no measurement effects, but the test used to support the statement is insufficient.

9. **Please see answer 6.**

I suggest reorganizing the “measurements made in the site” column of table 1 so it is easier to understand. It would be better if the reader could look at the numbered list of experiments (lines 182-191) and know which experiment was performed at which site. Please have the measurements in the table use the same wording as the subsections in the methods section. For example, the reader should be able to read the section heading 3.3 “Tissue Incubations” and easily discern from Table 1 where these measurements were performed.

10. **Please see answer 4. In addition, we reorganize the column, which is named now “Experiments in the site” and contains information about the experiments done in the site (with reference to the list in section 2.2) and the dates the experiments were performed. The “tissue incubations” (sections 2.6 and 3.3) are referred in the Table according to the list in section 2.2 (G) and specifically by the measured tissue (stem cores/leaves).**

The methods section (in particular sections 2.2 – 2.6) is difficult to follow and should be reorganized for clarity. For example, you could try organizing section 2.2 by site, which might make it easier to keep track of which experiment was performed at which site. Or, you could try incorporating the numbered list of experiments (lines 182-191) in the following sections. As is, the section is difficult to follow.

11. **Was corrected, please see answers 4, 10. We organized the experiments in section 2.2 by experiments and not by site.**

Minor Comments: I found the lengthy calculation in the discussion section (lines 383- 393) to be strange. Again, it is nearly the end of the paper, and a new two pool model is introduced and a calculation is performed. Please clarify the purpose of this calculation to answering your overall objectives, and consider how to shorten and make it more conceptual.

12. **The calculation using the two-pool mixing model was removed from the text.**

The distinction in greenness between Harvard and Bartlett forest is not discernable from Figure 4 (discussed lines 320-322), please clarify.

13. **Figure 4 presents trees measured in the Jerusalem site. The results from Harvard and Bartlett forests are not presented visually.**

I was surprised that pre-dawn water potentials were not referred to as a measure of water stress in lines 190-192, especially since stress is stated as a potential explanation for lower ARQ values in the discussion (line 307-308).

14. **Stress was mentioned in these lines as potential explanation for lipid respiration that can cause a decrease in ARQ. As natural trees in Israel are acclimated to the long dry Mediterranean summer, we don't think the term "stress" describes exactly the water status of these trees. To eliminate misunderstanding we removed the reference to "stress" in the text.**

Similarly, I was surprised to see the lengthy discussion of lipid storage in stems in lines 304 to 312, when the authors discount this as a reasonable explanation for lower ARQ values in the introduction in lines 39-41. The introduction made it seem like this was an unlikely explanation anyway, as only a few genera of tree species store lipids in tree stems.

15. **We revised the description in the introduction (L40-41) and edited the discussion about lipid storage (L329-333). In brief, lipids are believed to have small importance in storage and respiration in most trees. However, this assumption is based on very few lipid measurements in the literature, and we could not confidently refute the possibility that lipids potentially play an important role in tree stems.**

Why does ARQ plateau at 0.7 in the model presented in Figure 1? Shouldn't it plateau closer to 1 if that's what the theory suggests?

16. **The theory dictates the ratio DCO2/DO2 in the beginning of stem incubation should equal 0.76*DCO2/DO2 at the plateau stage. In this model run, ARQ=0.5, hence the DCO2/DO2 plateaus at ~0.65.**

Figure 3 is graph is good and easy to read, but the two yellow colors on this graph are difficult to distinguish

17. **Was corrected.**

Figure 4: It is difficult to distinguish colors and symbols on these graphs as many points overlap. Perhaps different symbols for the different heights would help?

18. **The symbols of the stem-base chambers are smaller now. In the software used to design the plots it is impossible in this type of plot to change the symbol style.**

Post-hoc comparisons – your methods (lines 249-251) state you did Tukey's post hoc comparisons, but I don't see letters corresponding to the post hoc tests like I'd expect to see on Figures 8 and 5 in particular. Were these tests performed for the corresponding analyses, and if so, why aren't the results on the figure?

19. **We performed post-hoc comparison only in the experiment presented in figure 9, where the letters indicate significant differences are shown.**

Line by line comments: Line 275 – what is the duplicates error?

20. **Single ARQ measurement is the average of duplicate flasks taken from the stem chamber, and the error is the standard deviation (SD). This is now clarified it in the text (L140-141).**

Please be consistent with the placement of the ARQ measurement type in-text, for example in line 188, "continuous" should be right after "ARQ" instead of at the end of the sentence. Line 342 should read "must" not "much".

21. **Was corrected.**

Berveiller, D., and Damesin, C.: Carbon assimilation by tree stems: potential involvement of phospho*enol*pyruvate carboxylase, Trees - Structure and Function, 22, 149-157, doi:10.1007/s00468-007-0193-4, 2008.
Hilman, B., and Angert, A.: Measuring the ratio of CO_2 efflux to O_2 influx in tree stem respiration, Tree Physiology, 36, 1422-1431, 10.1093/treephys/tpw057, 2016a.
Hilman, B., and Angert, A.: Measuring the ratio of CO_2 efflux to O_2 influx in tree stem respiration, Tree Physiol, 36, 1422, 2016b.
Ivanov, A. G., Krol, M., Sveshnikov, D., Malmberg, G., Gardeström, P., Hurry, V., Öquist, G., and Huner, N. P. A.: Characterization of the photosynthetic apparatus in cortical bark chlorenchyma of Scots pine, Planta, 223, 1165, 10.1007/s00425-005-0164-1, 2005.

Comparison of CO_2 and O_2 fluxes demonstrate retention of respired CO_2 in tree stems from a range of tree species

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Abstract. The ratio of CO_2 efflux to O_2 influx (ARQ, apparent respiratory quotient) in tree stems is expected to be 1.0 for carbohydrates, the main substrate supporting stem respiration. In previous studies of stem fluxes, ARQ values below 1.0 were observed and hypothesized to indicate retention of respired carbon within the stem. Here, we demonstrate that stem ARQ <1.0 values are common across 85 tropical, temperate, and Mediterranean forest trees from 9 different species. Mean ARQ values per species per site ranged from 0.39 to 0.78, with an overall mean of 0.59. Assuming that O_2 uptake provides a measure of *in situ* stem respiration (due to the low solubility of O_2), the overall mean indicates that on average 41% of CO_2 respired in stems is not emitted from the local stem surface. The instantaneous ARQ did not vary with sap flow. ARQ values of incubated stem cores were similar to those measured in stem chambers on intact trees. We therefore conclude that dissolution of CO_2 in the xylem sap and transport away from the site of respiration cannot explain the low ARQ values. We suggest ~~to examine~~ refixation of respired CO_2 in biosynthesis reactions as possible mechanism for low ARQ values.

1 Introduction

The global annual CO_2 efflux from tree stems to the atmosphere is estimated at 6.7 ±1.1 Pg C yr^{-1} ~~(Yang et al., 2016)~~(Yang et al., 2016), but the drivers of stem CO_2 efflux are not well understood ~~(Trumbore et al., 2013)~~(Trumbore et al., 2013). CO_2 in tree stems originates primarily from aerobic respiration, which consumes oxygen (O_2). ~~Respiratory~~ The respiratory quotient (RQ) is defined as the ratio between CO_2 produced ~~CO_2~~ and O_2 consumed ~~O_2~~, and its value is derived from the stoichiometry of the metabolized substrate. Carbohydrates are believed to be the main respiratory substrate in tree stems ~~(Hoch et al., 2003; Plaxton and Podestá, 2006)~~(Hoch et al., 2003; Plaxton and Podestá, 2006)~~, and their metabolism results in an RQ of ~1.0. Metabolism that relies~~

~~entirely on lipids yields an RQ value of ~0.7, but significant storage of lipids in stems is uncommon and limited to several tree genera called 'fat trees' (Sinnott, 1918). RQ values greater than 1.0 are associated with organic acid catabolism~~, and their metabolism results in an RQ of ~1.0. Respiration that relies entirely on lipids predicts RQ values of ~0.7, but it is not clear to what extent lipids are stored and used in trees as they are rarely measured (Hartmann and Trumbore, 2016). Current understanding suggests that significant storage of lipids in stems is uncommon and limited to several tree genera, the so-called 'fat-trees' (Sinnott, 1918). RQ values greater than 1.0 are associated with organic acids catabolism, due to the greater O content of the molecules being oxidized.

Initial measurements of the ratio of CO_2 efflux to O_2 influx from the stem surface for six tree species found values mostly below 1.0 (the expected value for RQ from carbohydrate metabolism) ~~(Angert and Sherer, 2011; Angert et al., 2012)~~(Angert and Sherer, 2011; Angert et al., 2012a). The flux ratio is referred to in those studies, and here, as the "apparent" RQ (ARQ), because it potentially includes ~~processes that incorporate~~additional sources or sinks of CO_2 and/or O_2 in the stem in addition to the respiration taking place in tissue beneath a chamber placed on the stem surface. Processes that can potentially reduce the emission of CO_2 and thereby decrease ARQ below 1.0 include: (1) dissolution and transport of CO_2 in the xylem sap ~~(Teskey et al., 2008)~~(Teskey et al., 2008), and (2) carboxylating reactions during biosynthesis of compounds more oxidized than carbohydrates that involve refixation of CO_2 by the enzyme phosphoenolpyruvate carboxylase (PEPC) ~~(Lambers et al., 2008)~~(Lambers et al., 2008). Alternatively, it may be hypothesized that ARQ below 1.0 is the result of non-respiratory O_2 uptake, e.g. by oxidases and hydroxylases that are O_2 consuming enzymes.

Carbon dioxide is ~30 times more soluble in water than O_2, and dissolved CO_2 reacts with water to form bicarbonate (HCO_3^-) and carbonate (CO_3^{2-}) ions, further increasing the amount of dissolved inorganic carbon (DIC). The rate of O_2 uptake is thus assumed to provide a better measure of stem respiration than CO_2 efflux, which can be complicated by dissolution and transport within the xylem sap ~~(Teskey et al., 2008)~~(Teskey et al., 2008)~~, potentially contributing to low ARQ values.~~, potentially contributing to low ARQ values. There is evidence from studies with an isotopically labeled stem CO_2 pool that a significant portion of C is transported as DIC to photosynthetic tissues where it might be refixed to organic C (Bloemen et al., 2013; McGuire et al., 2009; Powers and Marshall, 2011). If transport of CO_2 within the stem is important, ARQ measured at the stem surface is expected to be inversely related to sap velocity. As the difference in solubility between CO_2 and O_2 decreases with increasing temperature ~~(Gevantman, 2018)~~(Gevantman, 2018), ARQ also might be expected to increase with temperature if all other factors remain constant. In addition, variations of ARQ with stem height are to be expected. A model of CO_2 diffusion and advection in the xylem sap by Hölttä and Kolari ~~(2009)~~(2009) predicted that the accumulation of dissolved CO_2 in the ascending xylem sap, together with a reduction in stem diameter with height, induces faster CO_2 diffusive loss to the atmosphere in the upper parts of the stem. Thus, an increase in ARQ (higher CO_2 loss per mole of O_2 uptake) with stem height is expected. ~~However, there is evidence from studies with an isotopically labeled stem CO_2 pool that a significant portion of C is transported as DIC to photosynthetic tissues where it might be refixed to organic C (Bloemen et al., 2013; McGuire et al., 2009; Powers and Marshall, 2011). To date, studies of these processes in large trees are scarce, and it is not clear which process are responsible for low ARQ.~~

The second possible explanation for low ARQ is local dark refixation in the stem by PEPC (Angert et al., 2012b). PEPC is present in tree stems (Berveiller and Damesin, 2008; Höll, 1974; Ivanov et al., 2005), and its

activity was suggested to be sufficient to have a measureable impact on respired CO_2 in *Ricinus communis* (Gessler et al., 2009). Stem ARQ values would remain below unity as long as the products of PEPC fixation (e.g. malate and citrate) are not inhibiting further fixation. To date, studies of these processes in large trees are scarce, and it is not clear which processes are responsible for low ARQ. If lower than unity ARQ values are prevalent and result from processes that retain CO_2 in the stem, estimates of tree stem respiration based on CO_2 efflux measurements must be reconsidered. Thus, the first objective of this work is to determine whether ARQ values lower than 1.0 is observed in a variety of trees from different biomes and across seasons. A secondary objective of this study was to test whether ARQ varies with xylem stream characteristics or with tree height.

2 Materials and Methods

2.1 Methods for evaluating ARQ

We report tree stem ARQ results based on measurement methods described in ~~(Hilman and Angert, 2016).~~Hilman and Angert (2016a). These methods overcome the difficulty of measuring small changes in O_2 against the high atmospheric background by using a static stem chamber, in which the O_2 changes are considerably larger than in an open flow chamber.

We used three different approaches to measuring ARQ: two are based on discrete gas samples of headspace air, and one based on direct measurement of ~~instantaneous fluxes~~the headspace air using gas sensors ~~in the first hour after chamber sealing.~~("continuous" sampling). Discrete gas samples are either taken within 30 minutes to few hours after chamber sealing ("instantaneous" sampling) or after the chamber has been sealed to the stem for more than 24 hours, once steady state conditions have been achieved ("steady state"). These ~~timings~~methods and the time required for achieving steady state were confirmed by comparing with "continuous" measurements ~~with sensors (Hilman and Angert, 2016).~~ (Hilman and Angert, 2016a). For each site and experiment described in section 2.2, we identify the method used to estimate ARQ as "instantaneous", "steady state" (for flask samples) or "continuous" (Table 1).

2.1.1 ARQ measurement from discrete samples

The evaluation of ARQ from discrete gas measurements is based on a one-box model that describes gas dynamics in the headspace of a static chamber sealed to the surface of a tree stem ~~(Angert and Sherer, 2011; Angert et al., 2012; Hilman and Angert, 2016)~~(Angert and Sherer, 2011; Angert et al., 2012a; Hilman and Angert, 2016a). In the model, the gas in the chamber headspace has initial mean atmospheric values (20.95% O_2, 0.04% CO_2), ensured by flushing the chamber with ambient air before measurement. Once the chamber is closed and the headspace above the stem surface is isolated, metabolic reactions in the stem control the chamber's air composition. For the first few hours, headspace concentrations of CO_2 increase and O_2 decrease at rates that are roughly linear with time ("instantaneous" incubation, Fig. 1, S1). During this linear stage, ARQ is calculated by:

$$\mathrm{ARQ} = \frac{\mathrm{CO_2\ efflux}}{\mathrm{O_2\ influx}} = \frac{\Delta \mathrm{CO_2}}{\Delta \mathrm{O_2}} \quad (1)$$

where ΔCO_2 and ΔO_2 are the changes in $[CO_2]$ and $[O_2]$ during the initial period after the chamber was sealed, and for discrete samples can also be determined from the difference in concentrations between the chamber air sampled at a specific time and the initial atmosphere. "Instantaneous" fluxes of CO_2 and O_2 reported here are obtained either by monitoring concentration change during the first hour following chamber closure with sensors directly in the field or by sampling headspace air with glass flasks within 30 minutes to a few hours of closing the chamber. The flasks were transported to the laboratory for measurement of CO_2 and O_2.

After the first hours, the initially linear rates of change in headspace gas concentration with time decline, and concentrations eventually remain constant (Fig. 1, S1). In this phase the gases in the chamber and the outer part of the stem, where most of the metabolism takes place, are assumed to be in equilibrium. This "steady state" occurs when the rates of addition of CO_2 and loss of O_2 from the stem to the chamber headspace are balanced by diffusive (assuming no strong wind) exchange of headspace air with outside air through porous portions of the outer stem. For "steady state" samples, the chamber is sealed to the surface of the stem and left for a period longer than 24 hours, after which the headspace air is sampled using glass flasks. The CO_2 and O_2 concentrations must be corrected for differences in diffusivity between CO_2 and O_2, as detailed in ~~(Angert and Sherer, 2011; Angert et al., 2012; Hilman and Angert, 2016)~~(Angert and Sherer, 2011; Angert et al., 2012a; Hilman and Angert, 2016b) in order to estimate the ratio of the gas fluxes from the concentrations in the static chamber:

$$\mathrm{ARQ} = \frac{gCO_2 \times \Delta CO_2}{gO_2 \times \Delta O_2}$$

~~(2)~~(2)

where gCO_2 and gO_2 are the CO_2 and O_2 conductance values in the outer layer of the stem between the chamber and the atmosphere. The structure of the path along which diffusion occurs is the same for CO_2 and O_2 and hence the conductance ratio gCO_2/gO_2 depends solely on the ratio of diffusivities of the gases in air, which is 0.76 ~~(Massman, 1998)~~(Massman, 1998). As a result, at steady state:

$$\mathrm{ARQ} = 0.76 \times \frac{\Delta CO_2}{\Delta O_2} \quad (3)$$

~~Hilman and Angert (2016) demonstrated excellent agreement for direct comparisons of the "instantaneous" and "steady state" measurement methods, and the results are further compared here.~~

~~The data we report here were collected in different sites and over different years, and chamber designs and methods applied varied from site to site, as described in Sect. 2.2 and in Table 1. In all cases, a chamber is attached to the surface of the stem with an air-tight seal (using a sealant in most cases – see Table 1 for details). Ports (to which sampling flasks can later be attached) or a separate lid with ports allow the chamber to remain open to the atmosphere when not in use; openings are covered with screen to prevent insect damage inside the chambers. For a measurement, the chamber is first flushed with ambient air using a syringe, then all openings are closed, and CO_2 is allowed to accumulate (and O_2 to be consumed) in the headspace trapped within the chamber. The chambers contain sampling ports to which glass flasks equipped with O-ring valves (LouwersHanique, Hapert, The Netherlands) are attached. Initially the valves are open. Air from the chambers is sampled passively by closing the valves. For "steady state" field measurements, two glass flasks are connected to a stem chamber and closed after at least one day of incubation. For "instantaneous" ARQ, the valves are closed after shorter incubation periods (30 minutes to a few hours).~~

~~The flasks were analyzed in the laboratory at the Hebrew University in Jerusalem in a closed system [The *Hampadah* (Hilman and Angert, 2016)]. Two analyzers are included in the *Hampadah* system; an infra-red gas analyzer (IRGA) for CO_2 measurement (LI 840A LI-COR; Lincoln, NE, USA) and a fuel-cell based analyzer (FC-10; Sable Systems International, Las Vegas, NV, USA) for measuring O_2. The principle of operation of the *Hampadah* is measurement of the change in CO_2 and O_2 concentrations in the system's air after flask opening, and calculation of the concentration in the flask that would yield such change.~~

~~2.1.2~~Assuming constant CO_2/O_2 fluxes over time, samples taken either by "instantaneous" or "steady state" methods will yield the same ARQ values. Indeed, Hilman and Angert (2016a) demonstrated excellent agreement for direct comparisons of the "instantaneous" and "steady state" measurement methods, and the results are further compared here.

2.1.2 Stem chambers and gas measurements

All data reported here was collected by using chambers attached to the stem surface to create a gas-tight incubation headspace. Chamber designs and sampling details differed between sites (see section 2.2 and Table 1), but generally all chambers were equipped with sampling ports for attaching glass flasks equipped with O-ring valves (LouwersHanique, Hapert, The Netherlands). Outside incubations, permanently installed chambers were protected against insect infestation using screens. Incubations were always started at ambient concentration, and flasks were allowed to equilibrate with the headspace by opening the flasks' valves during incubation. Incubation time varied from between 30 minutes to a few hours for "instantaneous" ARQ samples to more than 24h for "steady state" samples. At the end of the incubation period, the flask valve was closed and the gas sample was shipped to the laboratory for analysis. Each reported ARQ measurement is the average of duplicate flasks taken from the stem chamber, and the error is the standard deviation.

The CO_2/O_2 ratios in the flasks were analyzed in the laboratory at the Hebrew University in Jerusalem in a closed system (The *Hampadah* (Hilman and Angert, 2016b)). Two analyzers are included in the *Hampadah* system; an infra-red gas analyzer (IRGA) for CO_2 measurement (LI 840A LI-COR; Lincoln, NE, USA) and a fuel-cell based analyzer (FC-10; Sable Systems International, Las Vegas, NV, USA) for measuring O_2. The principle of operation of the *Hampadah* is measurement of the change in CO_2 and O_2 concentrations in the system's air after the addition of the air from a given sample flask of known volume, and calculation of the concentration in the flask that would yield that overall concentration change (Hilman and Angert, 2016b).

2.1.3 Continuous ARQ measurements

Sensitive detection of small changes in O_2 is difficult in the field, which is why we used the flask samples and long chamber closure times ("steady state") in most field sites. However, to measure diurnal changes in stem ARQ values of *Malus domestica*, we were able to make continuous measurements with a small IRGA CO_2 sensor (COZIR Wide Range 0-20% CO_2 Sensor, CO2Meter, Inc~~.) and a quenching based optode (Fibox 3, PreSens Precision Sensing) for O_2 measurement (Hilman and Angert, 2016).~~., Ormond Beach FL, USA) and a quenching based optode (Fibox 3, PreSens-Precision Sensing, Regensburg, Germany) for O_2 measurement (Hilman and Angert, 2016a). The sensors' reading was extracted every 30 seconds. A temperature sensor was placed next to the optode sensor for temperature and water vapor corrections. ~~-~~The inlet of a small diaphragm pump (KNF micro-pump) and a non-return valve (SMC AKH 12mm, RS, UK) were connected to the chamber

headspace~~, for periodic automatic venting of~~ and used to automatically vent the chamber headspace every 4 hours. The CO_2 efflux and the O_2 influx were calculated using a linear fit over ~120 gas concentration measurements during the first hour of incubation, the chamber volume, and the stem surface area under the chamber. We used the data from this experiment to examine the sensitivity of ARQ to temperature, which affects the gas solubility constants. The strongest effects are expected during the night, when daytime influences on stem fluxes associated with sap flow and low turgor pressure (Salomón et al., 2018) are minimized.

~~For each site and experiment described below, we identify the method used to estimate ARQ as "instantaneous", "steady state" (for flask samples) or "continuous".~~

2.2 Study sites and experimental design

For addressing ~~our first goal of determining~~ the variation in stem ARQ values across a range of tree species and environments, ~~we measured ARQ in~~our study included trees located in tropical forests ~~in the Republic of~~ (Panama and ~~in~~ Brazil~~,~~), in temperate forests ~~in the northeast US~~ (Bartlett and Harvard ~~forests),~~, USA), and in a Mediterranean savanna ~~in (~~Spain~~, and in~~) and a Mediterranean shrubland (Carmel Ridge, Israel ~~where we sampled various species~~). We also included five trees located on the Hebrew University campus in Jerusalem (Israel) and in the adjacent Botanical Gardens. The trees in Panama were part of a fertilization and litter manipulation projects (Wright et al., 2011; Sayer and Tanner, 2010). No treatment effects were found (Fig. ~~, and in natural Mediterranean shrubland that is located on the Carmel Ridge (~~S2, this topic is not in the scope of this paper). Details about the sites, tree species, stem chambers, stem dimensions, and experiments conducted in each of the sites are presented in Table 1~~, Fig.~~ and Figure 3~~).~~ . The list below summarizes what data was available from the different sites and what questions in particular we addressed with this data (the numbering of the experiments matches Table 1):

A. Seasonal and/or phenological measurements of stem ARQ were performed in Jerusalem, US, and Brazil sites. In Jerusalem, five individual trees from five different species (first five species in Table 1) were measured ~~every 2-3 months between December-February 2011 and July 2014, except for the *M. domestica*, which was measured at monthly intervals between July 2011 and July 2013 ("steady~~. The phenological state~~"). Phenology~~ of ~~the~~ deciduous trees (all except *Quercus calliprinos*) was ~~classified~~separated into four ~~groups~~classes (Fig. 4). In ~~addition, in the same site, we sampled four *Quercus ilex*~~the US sites, trees ~~in July 2016 ("steady state"). Five individuals of *Acer rubrum* were~~ measured at ~~each of the sites in the US in September 2012 ("steady state"). Trees at~~ the northern site (Bartlett Experimental Forest) had fall color development, while leaves at Harvard Forest (southern site) were still green. ~~We questioned if ARQ would vary with the phenological differences.~~ After analysis of flasks, we excluded results from three trees because of suspected air leakage from the chamber (O_2 >20% after six days of stem incubation). In Brazil, six ~~*Scleronema micranthum* trees were measured in five campaigns between March 2012 and March 2014. In the two first campaigns "instantaneous" ARQ was measured, while "steady state" ARQ was measured in the three later campaigns.~~trees were measured. After analysis we excluded results from four out of twelve "instantaneous" measurements because of a weak signal (O_2 >20.7% and SD >0.1 after 3 h of incubation). ~~In Panama we sampled 42 *Tetragastris panamensis* trees ("steady state") in three campaigns: September 2012, September-October 2013, March-April 2014. Some individuals were~~

~~sampled more than once. The trees grew in plots that were part of a fertilization and litter manipulation projects (Wright et al., 2011; Sayer and Tanner, 2010). No treatment effects were found (Fig. S2). In Spain we sampled 16 *Q. ilex* trees during May 2015 ("steady state"). On Carmel Ridge we sampled ARQ of four *Q. calliprinos* trees ("steady state") during April 2012, September 2012, and January 2013.~~

For our second objective, to explore the potential for low ARQ values to reflect dissolution and transport of CO_2 in the xylem sap, we measured instantaneous ARQ at varying sap flow velocities and ~~in~~at different times of a day. Transport of CO_2 was previously reported to be correlated with sap flow ~~(McGuire and Teskey, 2004; Bowman et al., 2005; McGuire et al., 2007)~~(McGuire and Teskey, 2004; Bowman et al., 2005; McGuire et al., 2007). Thus, anti-correlation of ARQ with sap flux, expressed ~~in~~via maximal ARQ values during the night ~~in diel course~~when transport is at a minimum, would provide evidence ~~to support the transport~~ that low ARQ can be explained by export of locally respired ~~C~~ CO_2 (as DIC~~. We also~~) out of the stem region being measured ~~vertical transects of ARQ including in-stem measurements, using chambers~~(experiments B, C, and ~~probes placed at different heights on a single~~E, below). If transport of dissolved CO_2 is the main driver of low ARQ values, we would also expect that: (D) higher ARQ values will be observed at higher temperatures (due to differential temperature dependences of CO_2/O_2 solubility coefficients); (F) ARQ values will increase with stem~~,~~ height due to DIC accumulation and ~~in incubations from~~ stem tapering that induce stronger CO_2 diffusive loss; (F) ARQ values will decrease with depth in the stem (due to the greater proximity to the water conducting vessel elements); and (G) ARQ values in incubated stem cores~~.~~ will be higher than measured values at the stem surface (due to the detachment from the transport system).We performed a number of experiments~~:~~ to test each of these predictions (additional details in Table 1):

B. ~~1)~~ ARQ ("instantaneous") was measured simultaneously with sap flux density measurements in nine ~~*Q.*~~*Quercus ilex* trees with similar diameter (0.35 to 0.49 m at breast height) ~~in~~at the site in Spain.

C. ~~2) At sites where we could not measure sap flux, we measured day-night variation in ARQ.~~ ARQ ("instantaneous") was measured during daytime, at pre-dawn when the transpiration stream should reach its minimum, and again during the next day. We conducted two day-night campaigns on the trees at the site in Jerusalem~~, during July 2012 and April 2013.~~. Additionally, during ~~24-28 April 2013~~4 days, ARQ ("continuous") values were measured every 4 h from the *M. domestica* tree in Jerusalem ~~("continuous").~~

D. ~~3)~~ Nighttime results of the "continuous" ARQ measurements on the *M. domestica* enabled us to examine the relationship between temperature and ARQ. During the night, when sap flux is minimal, the temperature effect on the gases solubility should have its maximum effect on ARQ values.

E. ARQ ("steady state") was measured over spring, summer and winter ~~in the *Q.*~~for *Quercus calliprinos* trees on Carmel Ridge site, simultaneously with pre-dawn shoot water potential (Ψ_{pd}). Ψ_{pd} is a measure for available soil water and therefore is also a rough proxy for seasonal differences in transpiration rates (~~Aranda et al., 2005~~Aranda et al., 2005; Bucci et al., 2005).

F. ~~4)~~ ARQ was measured at different heights on the same tree ~~stem~~stems, while simultaneously ARQ was determined from ~~the stem surface using stem chambers, and also from~~ air sampled inside the stem. ~~In~~During the ~~ARQ~~ seasonal measurements in Jerusalem, ~~the *Q.*~~ARQ ("steady state") was measured at the stem base of the *Q. calliprinos* and the *Platanus occidentalis* trees ~~were measured at their stem~~

base, in addition to the as well as at breast height measurement ("steady state"). In Brazil, we measured "ARQ ("instantaneous" ARQ ") from stem chambers and in-stem probes to sample in-stem gases from stem chambers installedthe tree base up to 11 m above the ground on a single *S.Scleronema micranthum* tree. To evaluate the influence of the internal ARQ on the surface two separate days.

ARQ, we measured in the same tree in-stem gas concentrations and ARQ.

G. 5) "Steady ("steady state" ARQ") measured from stem chambers was compared with ARQ measurements through incubation of stem cores. Measurement of stem tissues should provide better estimation for the stem outer layers' RQ by excluding dissolution and advection in the xylem stream. Incubations were performed on cores taken from four species in four different sites (Table 1). In Jerusalem, we compared repeated stem incubation ARQ with that of leaf incubation.

2.3 Sap flux density

Sap flux density was monitored in 9 trees at the site in Majadas de Tietar (Spain) using heat ratio method (HRM) sensors (SFM1 Sap Flow Meter, ICT International). A description of the installation and measurement is presented in Methods S1. The detailed procedures for sap flux corrections and calculations are described in (Perez-Priego et al., 2017)(Perez-Priego et al., 2017). We tested, whether the daily maximum sap flux density (i.e. average of measurements between 10:00 and 17:00 during the day of the ARQ measurement), which correlated with CO_2 dissolution fluxes (Bowman et al., 2005), would explain variability in "instantaneous" ARQ.

. We tested whether the daily maximum sap flux density (i.e. average of measurements between 10:00 and 17:00 during the day of the ARQ measurement), which correlated with CO_2 dissolution fluxes (Bowman et al., 2005) could explain variability in ARQ ("instantaneous").

2.4 Shoot water potential

Pre-dawn shoot water potential (Ψ_{pd}) on Carmel Ridge was measured using a pressure chamber (PMS Instrument Company, Corvallis, Oregon, USA). At each sampling time, we sampled 2-3 terminal twigs containing 5-10 leaves from each *Q. calliprinos* tree. The samples were wrapped in plastic, placed on ice and measured within an hour of sampling using the pressure chamber technique (Scholander et al., 1965)(Scholander et al., 1965).

2.5 In-stem measurements

For sampling gas from inside the stem, stainless-steel tubes (1.3 cm diameter) were installed 4, 8, and 12 cm deep into the stem, in various stem heights on the same *S. micranthum* tree in Brazil where the vertical ARQ transects were measured. Installation procedure was according to Muhr et al. (2013)Muhr et al. (2013) and tubes were sealed between sampling dates. Using rubber tubing we connected the sampling flasks to the tubes for incubation of 4 days. The flasks were then analyzed for CO_2 and O_2 in the *Hampadah*. Assuming steady state, ARQ was calculated using Eq (3) (Angert et al., 20122012a).

2.6 Measuring ARQ of incubated tissues

Stem cores were extracted immediately after the chamber incubation experiment in Panama, Spain, and Jerusalem using a 1.2 cm diameter cork borer~~, right after the chamber incubation experiment~~. The outer bark and green tissues, as well as sapwood sieves (with paler color than the phloem tissues), were removed from the cores. The cores were cut ~~to fit~~ into ~~the incubation system~~longitudinal halves, wrapped with moist gauze cloth to avoid desiccation~~,~~ and ~~inserted into gas~~placed in an air-tight ~~set of~~incubation system to which glass flasks (two or three) were connected by Swagelok Ultra-Torr fittings (Swagelok, Solon, OH, USA, Fig. ~~S3~~S4). At the end of the incubation period, the flasks were closed and analyzed in the *Hampadah*. Since the incubations took place in a closed system~~,~~ (no diffusive exchange with outside air), the change with time in $[CO_2]$ and $[O_2]$ are assumed to be linear, and ARQ can be calculated using ~~Eq~~equation (1).

In Panama and Spain the incubations were started immediately upon core extraction, at ambient temperature, and lasted 8 h and 3 h, respectively. In ~~Israel,~~Jerusalem the ~~*Q. ilex*~~ cores were kept on moist gauze cloth for 2 h before being sealed in the incubation system ~~that were~~and kept at 25°C in an environmental chamber. Repeated incubations were performed in series, with the incubation systems flushed in between with ambient air. Simultaneously, from each tree, four leaves from an understory branch were cut and inserted into the same incubation systems, for the same incubation durations. The O_2 uptake rate (nmol O_2 g.FW^{-1} s^{-1}) was calculated as follows ~~:~~(adopted from Pruyn et al. ~~(2002a)]:~~(2002)):

$$O_2 \text{ uptake rate} = \frac{\Delta O_2}{100} \times \frac{V_{H}}{T \times M_{FW} \times V_{m}} \times \frac{V_H}{T \times M_{FW} \times V_m} \times 10^9 \quad (4)$$

where ΔO_2 is the decrease in $[O_2]$ during the incubation, V_H is volume of headspace (ml), T is incubation period (s), M_{FW} is fresh weight (g), V_m is the molar volume, and 10^9 converts units to nmol. We dried the samples in an oven at 60°C for two days for the dry weight.

In Brazil, stem cores were extracted by using a 5.15 mm diameter increment corer. After bark was removed the cores were cut to a length of 6 cm each and then allowed to equilibrate with the atmosphere for 6-8 hours, while continually being kept moist. After equilibration, each core was transferred to an incubation chamber equipped with flasks. Prior to starting the incubation, a few ml of water were added to keep the core tissue moist. In this case, incubations were left at room temperature (~25°C) for 24 h before flasks were closed and removed.

2.7 Statistical analysis

All statistical analysis was done using JMP (JMP®, JMP Pro 13, SAS Institute Inc., Cary, NC, USA). Repeated measures analysis of variance was used to evaluate how the interaction of tissue (stem core/leaves) with ARQ and O_2 uptake varies with time in the repeated incubations of the ~~*Q. ilex*~~ tissues from the trees in Jerusalem. Mauchly's test indicated violation of sphericity in the ARQ response in the repeated incubations experiment (χ^2 =18.132, P =0.021), therefore the Greenhouse-Geisser adjusted F test was chosen. One-way analysis of variance (ANOVA) followed by Tukey-Kramer HSD was used to perform comparisons among time points in every tissue. Student's t-test was used for comparisons between stem cores and leaves at each time point.

3 Results

The ARQ estimated from "instantaneous" and "steady state" measurements were in good agreement over a large range of ARQ (Fig. 2). The mean difference between the two assessments is 0.02, and RMSD is 0.15. The average ~~"~~ARQ ("steady state~~" ARQ")~~ value across all species and sites, including results from ~~(Angert et al., 2012)~~(Angert et al., 2012a), was 0.59 (n =229) and the average ARQ of species in the different sites ranged between 0.39 and 0.78 (Fig. 3). For individual measurements, a minimum ARQ value of 0.27 was recorded for *Q. ilex* in Spain and for ~~T.~~*Tetragastris panamensis* in Panama. The highest value was 0.99 for *M. domestica* and *Populus deltoids* in Jerusalem.

Phenology or seasonality had some effect on ARQ ~~(Figure 4). In Brazil, ARQ varied between 0.41 ±0.15 in the wetter season and 0.82 ±0.12) in the drier season.~~. In Jerusalem, the ARQ of *Q. calliprinos* and *Pistacia atlantica* ~~had lowest values~~was lower during spring and ~~highest values~~higher in fall and winter (Fig. ~~4).~~4). In Brazil, ARQ varied between 0.41 ±0.15 in the wet season (March) and 0.82 ±0.12 in the dry season (October, Fig. S3). The average ARQ of the ~~A.~~*Acer rubrum* trees at Harvard Forest, where all leaves were green, was significantly ~~greater~~higher than the average ARQ of the trees at Bartlett Experimental Forest, where the leaves had autumn color development (0.69 vs. 0.57, P <0.05 in a Student's t test).

3.1 ARQ values under varying xylem stream flow and temperature

ARQ ("Instantaneous ~~ARQ"~~) values of nine *Q. ilex* trees in Spain were invariable (mean ±SD of 0.42 ±0.04) in comparison with the larger variation in maximum daily sap flux density among these trees (0.15 ±0.05 m^3 H_2O m^{-2} h^{-1}), and no correlation was found between the ARQ and sap flux density (r^2 =0, P =0.9891).

Mean ARQ ±SD values ("steady state") of the ~~oaks~~trees at the Carmel Ridge site were 0.62 ±0.06, 0.68 ±0.07 and 0.69 ±0.08 for spring, summer and winter, respectively. Repeated-measures analysis of variance found no significant difference between seasons ($F_{2,2}$ =2.52, P =0.28), while Ψ_{pd} varied significantly with seasons ($F_{2,2}$ =207.85, P =0.0048). During summer, Ψ_{pd} was -2.65 MPa, much lower than the spring and winter values (-0.64 and -0.86 MPa, respectively).

In the Jerusalem day-night campaigns ~~done at Hebrew University and the adjacent arboretum~~, ARQ ("instantaneous") values ranged between 0.52 and 1.05, across all trees, seasons, and ~~samples~~sampling times (Fig. 5). Pre-dawn ARQ values ~~higher~~exceeding than daylight values (~~beyond the~~by amounts larger than the differences between duplicates ~~error~~) were observed during the summer in *M. domestica* and in the upper chamber on *Q. calliprinos*. No significant diurnal effect was found in repeated-measures analysis of variance of the breast height chambers, neither when results of all the trees was grouped by season, nor when results were grouped by stem chamber. In "continuous" ~~measurement~~measurements of *M. domestica*, with ARQ ~~value~~values obtained every 4 hours~~.~~. ARQ during the night (0.70; n=12) was not significantly ~~higher than the day time ARQ value [~~(P >0.76 in a ~~student's~~student's t test~~, 0.70 (n =12) vs~~) greater than in the day (0.71 ~~(~~; n =11~~)~~ ~~respectively,~~; Fig. 6~~].~~). The variations among the nighttime values were best ~~fitted with~~ explained using temperatures measured 235 minutes before the ARQ measurement (r^2 =0.84, P =0.0001, ARQ =0.01 × Temperature (C^o) + 0.54). With the same time lag, the coefficient of determination for the daytime values is r^2 =0.44 (P =0.0266).

3.2 Stem surface and in-stem ARQ vertical transects

In *Q. calliprinos*, measured over three years in Jerusalem, ARQ did not differ significantly (P >0.33 in student's t test) between breast height and stem base (ARQ of 0.56 vs. 0.59 respectively, n =14, Fig. 4). For *P. occidentalis* measured for the same period the ARQ measured at breast height was significantly higher than ARQ measured at the stem base (0.74 vs. 0.64 respectively, n =12, P =0.003 in student's t test, Fig. 4). For a single *S. micranthum* tree in Brazil, ARQ values measured at heights of 6.5 m and 11 m above the ground were similar to ARQ measured at breast height (Fig. 7), but also show differences with the stem base. In this tree, ARQ measured in March (0.46 ±0.11; wet season) was lower than in October (0.89 ±0.16; dry season). The in-stem ARQ values ranged between 0.25 and 0.56, with average ±SD of 0.46 ±0.07 in both seasons and at all stem positions and depths. The in-stem ARQ, as well as [CO_2] values, had no clear vertical trend (Fig. 7~~,S4~~; S5).

3.3 Tissue incubations

The average ARQ values of the stem core incubations were similar to the stem chamber ARQ for the four sites/trees where these comparisons were made (Fig. 8). ~~In the time series~~When incubations ~~of~~were repeated over time for *Q. ilex* stem cores and leaves, significant effects of time, tissue (leaves, stem cores), and their interactions (time × tissue) on ARQ and O_2 uptake rates were observed. ARQ of the stem cores increased from 0.44 ±0.08 (mean ±SD, n =4) after 3 h to 0.94 ±0.03 at the end of the experiment (32 h; Fig. 9). The ARQ of incubated leaves of the same trees showed higher initial ARQ of 0.80 ±0.02, with an increase over time to 0.92 ±0.02.

4 Discussion

4.1 ARQ is lower than 1.0 for a wide range of tree species

The ARQ measured in stem chambers installed on 85 individual trees of 9 species including tropical, temperate and Mediterranean forest trees was considerably and almost universally lower than 1.0. ARQ values as low as 0.7 could indicate that lipids were used exclusively as substrates for respiration~~. Lipids respiration is often associated with environmental stresses, for example, initial RQ of ~1 measured in branches of *Pinus sylvestris* L. declined in response to 11 days of shading and drought treatments to values of 0.77–0.75, reflecting mixture of substrates (Hanf et al., 2015). However, many ARQ values are~~, but current understanding suggests this scenario is implausible. However, this understanding relies on low and constant lipid concentrations over seasonal sampling (Hoch et al., 2003); daily changes in lipid concentrations and RQ were measured in response to shading and drought treatments, indicating this substrate might be more important than commonly thought (Fischer et al., 2015; Hanf et al., 2015). Nevertheless, many of the measured ARQ values were below 0.7, so substrate use alone cannot explain them. Additionally, as ARQ values above 1.0 are expected when lipids are produced ~~(De Vries et al., 1974)~~(De Vries et al., 1974), ARQ <1.0 resulting from lipid metabolism must be mirrored with ARQ >1.0 at a different time (assuming the lipids are produced locally). However, ARQ almost never exceeded 1.0. The results demonstrate that O_2 influx to the stems usually exceeded the CO_2 efflux, regardless of tree species, site, season, and time of day. Assuming O_2 uptake provides a measure of *in situ* respiration (due to the low solubility of O_2~~),~~) and carbohydrates are the main substrate, values of ARQ

averaging 0.59 indicate that on average 41% of the CO_2 produced by respiration was not locally emitted to the atmosphere, but apparently retained in the stem.

For sites where we have time series data for the same individuals, considerable ~~variation~~variations in ARQ values ~~was~~were observed over two years in Brazil (Fig. S3) and over three years in ~~Israel.~~Jerusalem (Fig. 4). A decrease in ARQ values was often observed during entrance to dormancy for the deciduous trees in Jerusalem, and an apparent minimum in ARQ for *P. atlantica* and *Q. calliprinos* in spring (Fig. 4). ~~This~~The autumn decrease seems to be in agreement with the ~~findings~~finding of significantly lower ARQ for Bartlett Experimental forest, where leaves were beginning to senesce, compared to the more southerly Harvard forest, where leaves were still green.

The possibility of measurement artifacts as the source for the low ARQ values seems unlikely, as ~~Hilman and Angert (2016)~~Hilman and Angert (2016a) previously demonstrated the validity of the measurement methods and the box-model approach. Further support comes from the ~~slope (1.006) of~~ small mean difference (0.02) between the ~~linear regression of~~ "instantaneous" ~~ARQ vs.~~and "steady state" ARQ measured for the same tree, which ~~is extremely close to 1 (Fig. 2).~~reflects overall agreement between the measures. The considerable ~~scattering~~scatter around perfect agreement (Fig. 2), expressed also in ~~the regression may~~RMSD of 0.15 could be taken as an indication that the measurement methods differ significantly. However, since the model assumes constant ARQ with time, and temporal changes in ARQ are obviously present as shown in Fig. 5 and 6, the scatter could also be attributed to temporal differences in the time integrated by the two types of measurement: the "instantaneous" sampling was typically conducted few days before the "steady state" sampling on the same tree~~, and to lower precision in "instantaneous" samples due to smaller changes in O_2 over the shorter time periods (Hilman and Angert, 2016). We also found strong similarities between the ARQ measured for intact stems with chambers and by incubating cores (Fig. 8), which~~. Additionally, the precision for "instantaneous" ARQ was lower than for “steady state” values, due to smaller changes in O_2 over the shorter time periods. This may also contribute to the scatter in Fig. 2 (Hilman and Angert, 2016a). We also found strong similarities between ARQ measured on intact stems using chambers and ARQ determined by incubating stem cores (Fig. 8). This provides another, indirect, confirmation that the low ARQ values obtained with the stem chamber measurement approaches are measuring something that is occurring in the stem tissues.

4.2 Dissolution and transport of respired CO_2 in xylem stream cannot explain the low ARQ values

Given the low solubility of O_2, stem flux ARQ values <1.0 (or potentially ≤0.7 for ‘fat’ trees) are ~~either~~the result of respired CO_2 ~~that is~~either being exported from the site of respiration before it can be emitted to the atmosphere or being refixed ~~in~~during biosynthesis processes within the stem. As noted earlier, a second possibility is non-respiratory O_2 uptake, e.g. by oxidases and hydroxylases that are O_2 consuming enzymes, most notably used in lignin biosynthesis. However, stoichiometric analysis of this pathway shows that the CO_2 produced from the sucrose that is the lignin's substrate usually exceeds the O_2 consumption, so that the net effect of lignin biosynthesis should be a local increase in ARQ (Amthor, 2003). To the best of our knowledge, there are no other significant O_2 consuming processes in tree stems that might affect the ARQ value.

We conclude that the low stem ARQ ~~much~~must be the result of CO_2 being locally fixed or transported away from the site of respiration. If CO_2 dissolution and DIC transport is the main export mechanism, we would expect ARQ to ~~be related to~~increase with temperature increase (i.e. according to solubility changes with

temperature), be anti-correlated with sap flow ~~(McGuire and Teskey, 2004; McGuire et al., 2007; Bowman et al., 2005)~~(McGuire and Teskey, 2004; McGuire et al., 2007; Bowman et al., 2005), and ~~further that ARQ should~~to increase with height in the stem ~~(Hölttä and Kolari, 2009).~~(Hölttä and Kolari, 2009). Three observations support the idea that this export mechanism controls some of the variability in ARQ. First, nighttime ARQ in ~~the diurnal measurements of the~~ *M. domestica*~~, the nighttime ARQ results were~~ was indeed correlated with temperature, an expected trend given the greater temperature sensitivity of the CO_2 solubility in comparison with O_2 ~~(Gevantman, 2018)~~(Gevantman, 2018). Second, the *P. occidentalis* had higher ARQ values in the upper stem position, especially during the growing ~~seasons~~season (Fig. 4). Third, relatively high ARQ values were observed at 0.2 m above the ground in the *S. micranthum* tree (Fig. 7), which may reflect a burst of in-stem CO_2 that originated from belowground respiration ~~(McGuire and Teskey, 2004; Levy et al., 1999)~~(McGuire and Teskey, 2004; Levy et al., 1999). However, in most of our observations ARQ did not vary as expected if CO_2 dissolution and transport were the main CO_2 export mechanism.

When sap flux density was measured directly, it did not explain the variation in ARQ among *Q. ilex* trees in Spain. Mean ARQ values were fairly stable over spring, summer and winter (0.62-0.69) for *Q. calliprinos* in the Carmel Ridge site, while the transpiration stream probably varied greatly between seasons if related to Ψ_{pd}. Additionally, during dormancy when no leaves were in place to force the transpiration stream, we found ARQ values <1.0 in four deciduous trees (black markers in Fig. 4). ~~Transpiration streams~~Sap flow rates are ~~also~~ assumed to decline during the night, but ARQ values <1.0 during nighttime were measured in five species, and in most cases no nocturnal increase of ARQ in comparison to daytime values was observed (Fig. 5, 6). Thus, the temperature dependency observed for the *M. domestica* tree during the night, which explained variability in ARQ values between 0.65-0.75, must be a second order control on ARQ variability and cannot explain the big deviation from unity (according to the linear fit, an ARQ of 1.0 is expected at the unreasonable temperature of 63^{o}C). Also, the vertical transects of ARQ for *Q. calliprinos* and *S. micranthum*, including in-stem ARQ for the later (Fig. 4, 7, ~~S4~~ S5), showed no consistent pattern of ARQ increasing with stem height, unlike the ARQ increase with height measured in the *P. occidentalis* (Fig. 4). ~~Likewise, no trend of in-stem [CO_2] increase with stem height was observed, suggesting no CO_2 accumulation in the ascending xylem sap (Fig. S4).~~

~~The in-stem ARQ measured in the *S. micranthum* ranged between 0.25 and 0.56. Even lower values, with typical ARQ of 0.13-0.18, have been reported before~~ ARQ values measured in the stem core incubations, where tissues are isolated from the influence of transport in the xylem stream, were well below 1.0 and similar to the chambers' values (Fig. 8, 9). The in-stem ARQ measured in the *S. micranthum* was likewise <1.0, but although the proximity to the xylem was greater, the values were not necessarily lower than the surface ARQ (Fig. 7). It is likely that in-stem ARQ values are influenced by dissolution in the xylem water, but the question is what is the contribution of in-stem CO_2 to the CO_2 efflux from the stem surface? There are contradicting assessments, and the influence likely is related to wood-anatomy. For example, studies of ring- and diffuse-porous species observed tight covariations of in-stem CO_2 and surface efflux and have interpreted this as evidence of strong influence of in-stem CO_2 concentrations ~~(Angert et al., 2012)~~(Teskey and McGuire, 2007; Steppe et al., 2007; Teskey and McGuire, 2002)~~. These low values are consistent with in-stem measurements of small CO_2 increases and large O_2 reductions in comparison to atmospheric concentrations~~, while other studies conducted on conifers with tracheid anatomy inferred only marginal influence of in-stem processes on surface efflux ~~(Pruyn et al., 2002b; Eklund, 1990; Eklund, 1993)~~(Ubierna et al., 2009; Maier and Clinton, 2006)~~. Thus,~~

~~one can speculate that the low ARQ values at the trunk surface are the reflection of the low ARQ in the sapwood itself. However, there are contradicting assessments about the influence of in-stem CO_2 on the CO_2 efflux from the stem surface; while some studies interpreted tight covariations of in-stem CO_2 and surface efflux as strong in-stem influence (Teskey and McGuire, 2007; Steppe et al., 2007; Teskey and McGuire, 2002), other studies inferred only marginal influence of in-stem processes on surface efflux (Ubierna et al., 2009; Maier and Clinton, 2006). Unlike covariation observations, which do not necessarily represent cause and effect relationships (Maier and Clinton, 2006), Muhr et al. (2013) utilized the difference in ^{14}C signature of in-stem CO_2 (5 cm deep) and surface efflux to estimate that <20% of total emitted CO_2 originates from the inner stem. To assess the potential influence on ARQ measured at the surface, we used a two-pool model for sources of the surface CO_2 efflux: (1) an in-stem CO_2 pool that is affected by sap flow transport, and (2) CO_2 produced locally by stem tissues (mostly close to the stem surface) fully released to the atmosphere. Using the results of the *S. micranthum*, we can evaluate the contribution of the in-stem ARQ to the stem surface ARQ. The mean ARQ values for the 4 cm deep probes and for the surface were ~0.5 and 0.67, respectively. Assuming 20% of the CO_2 efflux comes from the in-stem due the diffusive gradient between the sapwood and the atmosphere, the other 80% comes from metabolism in the stem tissues close to the stem surface. With an in-stem ARQ of 0.5, which means that the O_2 influx induced by concentration gradient is twice the CO_2 flux after correcting for relative diffusivity, the O_2 influx between the atmosphere and the inner stem would be equivalent to 40% of the CO_2 efflux. To explain the overall ARQ (measured at the stem surface) of 0.67, the O_2 influx to the stem tissues must therefore be the equivalent of 110% of the CO_2 efflux, i.e. the total ARQ representing the sum of fluxes from diffusion and metabolism would be (20+80)/(40+110) =0.67. The ARQ of the fluxes stem tissue metabolism alone would be 80/110 =0.73, which is still lower than unity.~~

~~ARQ values <1.0 observed in stem core incubations, where tissues are isolated from the influence of transport (Fig. 8,9) further support our conclusion that ARQ resulting from local metabolism of the stem tissue near the surface is <1.0. In light of that, the apparent decoupling between ARQ, sap flux density, and Ψ_{pd} presented above might be derived from strong diffusion barriers that restrict gas exchange between the sapwood and the external cambium, phloem, and bark tissues (Ubierna et al., 2009). A major contribution from respiratory activity concentrated in the outer stem tissues to overall stem respiration would further reduce sap flow effects on surface fluxes (Hölttä and Kolari, 2009; Maier and Clinton, 2006; Ubierna et al., 2009).~~ Nevertheless, observations of covariation in in-stem [CO_2] and CO_2 efflux do not necessarily represent cause-and-effect relationships (Maier and Clinton, 2006). Muhr et al. (2013) utilized the difference in ^{14}C signature of in-stem CO_2 (5 cm deep) and surface efflux, to estimate that <20% of total emitted CO_2 originates from the inner stem in three tropical non-coniferous tree species. Small contribution of in-stem CO_2 to the surface efflux can be easily explained by the slow diffusion through wood of all three anatomical groups (Sorz and Hietz, 2006). The woody diffusional barrier can explain the apparent decoupling between ARQ, sap flux density, and Ψ_{pd} presented above. A major contribution from respiratory activity concentrated in the outer stem tissues to overall stem respiration would further reduce sap flow effects on surface fluxes (Hölttä and Kolari, 2009; Maier and Clinton, 2006; Ubierna et al., 2009).

An alternative~~,~~

~~Overall, our results suggest that CO_2 dissolution and removal in the xylem stream are not the main cause of the low ARQ values that are common to the trees we measured.~~ ~~At the same time, observed ARQ values may be~~

~~influenced by the cumulative effects of some dissolution and transport, partial lipid metabolism, and non-respiratory O_2 consumption. One potential~~ explanation for low ARQ values could be the fixation of CO_2 by biosynthesis with engagement of the enzyme ~~phosphoenolpyruvate carboxylase (~~PEPC~~),~~, which is able to fix respired CO_2. Indirect evidence for PEPC activity can be found in the increase of the ARQ values with time in our repeated incubations, while cellular activity was retained as reflected in O_2 uptake rates (Fig. 9). Such a pattern may reflect a biochemical process, e.g. CO_2 fixation by the enzyme PEPC, which decreases with time due to self-inhibition by the accumulation of the products (Kai et al., 1999; Huber and Edwards, 1975). PEPC fixation rates can easily explain the retained CO_2, according to mass balance calculation for the stem cores incubations and published PEPC fixation rates in young tree stems (Table 2). Assuming refixation is important, the fact that ARQ measured from intact stems is almost always lower than unity indicates that the fixation products, organic acids like malate and citrate or amino acids, are not inhibiting the fixation or being oxidized locally, and are further metabolized or allocated elsewhere in the stem. The malate can be transported in the xylem stream as indicated by an upwards concentration increase in *Acer platanoides* stems (Schill et al., 1996). A possible fate of the malate might be similar to 'C4-like photosynthesis' observed in tobacco, where xylem-transported malate contributes carbon to photosynthesis in leaves (Hibberd and Quick, 2002). Alternatively, the fixation products might be exported via the phloem. One possible sink is excretion of organic acids to the rhizosphere as root exudates, which can account for ample fraction of overall GPP in forests (Abramoff and Finzi, 2016; Finzi et al., 2015). Indications for the transport of organic acids from upper parts of the plant to the roots have already been reported (Hoffland et al., 1992; Shane et al., 2004).

Overall, our results suggest that CO_2 dissolution and removal in the xylem stream are not the main cause of the low ARQ values that are common to the trees we measured. ~~C fixation by the enzyme PEPC, that decreases with time due to self inhibition by the accumulation of the products (Kai et al., 1999; Huber and Edwards, 1975). Based on mass balance calculation for the stem cores incubations and published PEPC fixation rates in tree stems (Table 2), PEPC fixation rates can easily explain the retained CO_2. The fixation products, organic and amino acids, may be exported via the xylem stream and/or via the phloem (Hoffland et al., 1992; Schill et al., 1996). Further investigation into~~We speculate the observed ARQ values resulted by PEPC refixation, with possible cumulative effects of some dissolution and transport, partial lipid metabolism, and some non-respiratory O_2 consumption. Corticular photosynthesis may theoretically influence ARQ, but in complex manner; with assumed O_2/CO_2 exchange ratio of 1.0 and given all other processes yield ARQ <1.0, the photosynthesis will reduce the CO_2 and O_2 concentration gradients between stem-atmosphere in the same absolute numbers, which will cause to further ARQ decrease. However, in our measurements photosynthesis was prevented by shading the measured stem surface. Additionally, most evidence for significant corticualr photosynthesis come from twigs and young stems (Pfanz et al., 2002; Ávila et al., 2014), while stems in current study were mature. Wood anatomy may further impact ARQ by modifying the contribution of internal stem processes on surface fluxes. The numerous mechanisms potentially responsible to ARQ probably varied with the broad range of species and wood anatomies we investigated. Further research to pursue the potential role of PEPC, including direct measurement of PEPC activity, would be needed to assess whether PEPC plays a role in lowering ARQ values to the levels observed. To complete the stem carbon balance, additional evaluation of the relations between the in-stem and the stem surface fluxes are also needed, as well as analysis of how organic and amino acids vary in the stem.

4.3 Implications of low ARQ

From a whole ecosystem perspective, if respired CO_2 in the stem returns to the atmosphere elsewhere (e.g. in the soil, canopy), the overall ecosystem-atmosphere carbon fluxes will not be affected, and high ARQ associated with the release of the transported CO_2 will balance the low ARQ in the stem. ~~However~~Such ARQ >1.0 values are expected in the rhizosphere where organic acids are decomposed. In the canopy, greater re-fixation of internal C is expected to increase the photosynthetically oxidative ratio (O_2 produced/CO_2 consumed), as the internally transported C replaces the atmospheric CO_2 when assimilation is measured. Additionally, such internal transport can cause a discrepancy between the measured above-ground and below-ground CO_2 effluxes and the locations where respiration is actually occurring ~~(Aubrey and Teskey, 2009)~~(Aubrey and Teskey, 2009), and lead to false attribution of respiration responses to environmental conditions. Moreover, the different long-term temperature sensitivity of CO_2 efflux and O_2 influx is of interest, and might explain part of the gap between modeled and observed Q_{10} values of tree respiration ~~(Griffin and Prager, 2017)~~(Griffin and Prager, 2017). For example, decrease in ARQ with rising temperature (due to higher PEPC activity for example) might result in a slow increase in CO_2 efflux, whereas the respiration rate (O_2 uptake) is actually increasing sharply, together with the internal carbon flux. Future studies should determine how temperature and nutrients control long term changes in ARQ, and aim to identify the biochemical process that control the low ARQ reported by the current study.

Author Contribution. B.H and A.A planned and designed the research. B.H performed the ARQ analysis and led the writing of the manuscript. J.M, N.K and S.T carried out the field work in Brazil, and M.S.C carried out the field work in USA. P.Y measured shoot water potential. S.J.W designed the long term experiment in the Republic of Panama. G.M, O.P, M.M, and A.C contributed to the campaign in Spain. O.P measured the sap flux density. J.M.G and Y.O contributed to the campaigns in the Carmel Ridge. T.W contributed to the campaigns in Spain and in Givat Ram campus. J.M, S.T, S.J.W, G.M, O.P, M.M, J.M.G and A.A contributed to the discussion and writing.

Data availability. Data used in this study can be found in figures, tables and in the Supplement.

Competing interests. The authors declare that they have no conflict of interest.

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Table 1 Study sites, tree species sampled at each site, stem chambers ~~and stems dimensions~~, stems diameters, and experiments done in the site.

Site and coordinates	Species	Chamber type, sealant	Diameter at the base of the chamber (cm)[a]			Measurements made in the site	ARQ measurement method
			Mean ± SD	minimum	maximum		
Givat Ram campus, Jerusalem, Israel (31.77°N, 35.20°E)	*Populus deltoids* Bartr. Ex Marsh *Platanus occidentalis* L. *Pistacia atlantica* Desf. *Quercus calliprinos* Webb. *Malus domestica* Borkh. *Quercus ilex* L.	Perspex®[b], hot glue	60.2 43.4 21.2 24.3 16.3			ARQ seasonal comparison Day-night ARQ variation ARQ vs. temperature	Steady state, Instantaneous, Continuous
		Perspex®, vacuum grease	20 ± 8	12	29	ARQ vertical transects ARQ of incubated stem cores and leaves	
Ramat Hanadiv Nature Park, Carmel Ridge, Israel (32.55°N, 34.94°E)	*Quercus calliprinos* Webb.	Perspex®, hot glue	11.2 ± 1.2	9.8	12.7	Simultaneous measurements of ARQ and shoot water potential	Steady state
Bartlett Experimental forest, NH, USA (44.06°N, 71.29°W)	*Acer rubrum* L.	Polypropylene[c], caulking	20 ± 10	10	34	ARQ seasonal comparison	Steady state
Harvard forest, MA, USA (42.53°N, 72.17°W)	*Acer rubrum* L.	Polypropylene, caulking	18 ± 9	9	26	ARQ seasonal comparison	Steady state
Majadas de Tiétar, Caceres, Spain (39°56'25" N, 5°46'28" W)	*Quercus ilex* L.	Perspex®, vacuum grease	45 ± 7	35	64	ARQ survey of 16 trees Simultaneous measurements of ARQ and sap flux density ARQ of incubated stem cores	Steady state, Instantaneous
Gigante peninsula, Barro Colorado Nature Monument, Republic of Panama (9°06'31" N, 79°50'37" W)	*Tetragastris panamensis* (Engl.) Kuntze	Perspex®, vacuum grease	30.0 ± 12.5	12.6	66.8	ARQ survey of 42 trees ARQ of incubated stem cores	Steady state, Instantaneous
A station of the Brazilian National Institute for Research in the Amazon (INPA), north west of Manaus, Brazil (2°38'23" S, 60°09'51")	*Scleronema micranthum* (Ducke) Ducke	Polypropylene[d]	41.2 ± 13.3	27.0	57.8	Seasonal comparison ARQ vertical transects In-stem ARQ and gases concentrations ARQ of incubated stem cores	Steady state, Instantaneous

[a]All chambers were installed at ~1.3 m above the ground, except for the *Q. calliprinos* on Carmel Ridge that were placed near to the ground due to the shrubby canopy, the low branching of the trunk and the constraint of the size of the chamber.

[b] ~~Chambers~~[b]Chambers were made of 10 cm × 12 cm Perspex® plate with four connectors to allow attachment of sampling flasks. The chamber on the *M. domestica* was slightly larger, 12 cm × 19 cm, with six flasks connectors. Chambers were placed on top of a closed cell foam frame that allowed an air-tight seal between the rigid chamber and the uneven surface of the tree stem. We used nylon straps to compress the foam, while the sealant was applied between the foam and stem for ensuring the seal ~~(Hilman and Angert, 2016).~~(Hilman and Angert, 2016a). Sealants were silicone based vacuum grease (Silicaid®1010 manufactured by Aidchim ltd., Raanana, Israel) or hot glue applied by a hot-glue gun.

[e] ~~The~~[c]The chambers are described in ~~(Muhr et al., 2013; Carbone et al., 2013)~~(Muhr et al., 2013; Carbone et al., 2013). Briefly, the chambers were made from an opaque plastic polypropylene pipe T-fitting with fittings for sampling flasks. Sealants were caulking (Nautiflex; OASE GmbH, Oerel- Barchel, Germany) or hot glue applied by a hot-glue gun.

[d] ~~Chambers~~[d]Chambers were built from a 15 cm long piece of polypropylene (PP) tubing (6.5 cm OD) that was welded shut on both sides with a PP disc (6.7 cm diameter). By cutting off a segment (height 2 cm) the tube was turned into an incubation chamber. Opposite the chamber opening, three fittings (Sprint ESKV 20, Wiska, Germany) were installed and sealed around the edges with liquid rubber (Dichtfix, Bindulin, Fürth, Germany). For sampling, chambers were attached to the trees with 4 lashing straps. To achieve a gas tight seal, a frame (25 mm thick) made from closed-porous cellular rubber (EPDM-quality, REIFF Technische Produkte GmbH, Reutlingen, Germany) was placed between the chamber and the stem.

Table 2 Comparison between the calculated PEPC fixation rates required to explain measured ARQ in stem cores incubations and reported PEPC fixation ~~rate.~~rates for young stems.

	ARQ[a] (CO_2 efflux/O_2 uptake)	O_2 uptake[b] (nmol g.DW^{-1} s^{-1})	PEPC fixation rate required to explain the observed ARQ[c] (nmol CO_2 g.DW^{-1} s^{-1})	PEPC fixation rate[d] (nmol C g.DW^{-1} s^{-1})
Quercus ilex (n =4)	0.44 ±0.08[c]	3.84 ±0.30	2.15	
Tetragastris panamensis (n =11)	0.33 ±0.07	1.40 ±0.69	0.93	
Fagus sylvatica L.				12.6
~~[a] Values~~ *Pinus sylvestrys L.*				16.74

[a]Values are mean ±SD

[b] ~~Dry~~Dry weight (DW) was determined after drying in an oven at 60°C for two days.

[c] ~~Calculated~~Calculated as O_2 uptake × (1-ARQ), which is an estimation of the flux of respired CO_2 that didn't diffused out from the core. Based on the assumption that carbohydrates with ARQ =1 are the respiratory substrate.

~~[d] We calculated PEPC fixation rate of *Fagus sylvatica* L. with data from Berveiller and Damesin (2008) as follow:~~

[d]We calculated PEPC fixation rate of *Fagus sylvatica* L. with data from Berveiller and Damesin (2008) as follow:

PEPC activity (nmol C mg^{-1} chl s^{-1}) × total chl (mg g.DW^{-1}) ~~=~~= ~30 × 0.42 = 12.6 nmol C g.DW^{-1} s^{-1}

The chosen PEPC activity was the lowest among seasonal measurements.

We calculated PEPC fixation rate of *Pinus sylvestrys* L. with data from Ivanov et al. (2005) as follow:

PEPC activity (μmol C mg^{-1} chl min^{-1}) × total chl (μg $g.FW^{-1}$) × g.FW/g.DW (using assumed water content of 0.5) × conversion to seconds = 1.04 × 483.02 × 2 × 1/60 = 16.74 nmol C $g.DW^{-1}$ s^{-1}

PEPC activity was measured during winter

Figure captions

Figure 1: ~~Modeled~~Modelled changes in a tree stem chamber of the concentrations of CO_2, O_2, and the ratio between ΔCO_2 and ΔO_2, which are the changes in the gases concentrations from their initial values, and are also the difference in concentrations between the chamber and the atmosphere. The gas dynamics are based on a one-box model with arbitrary fluxes and ARQ ~~=~~(ratio of CO2 efflux/O2 influx for tree stems) = 0.5. The two time frames in which ARQ~~, the ratio of CO_2 efflux/O_2 influx,~~ can be measured from the ratio $\Delta CO_2/\Delta O_2$ are indicated in the figure.

Figure 2: ~~Relation between~~Scatter plot of "instantaneous" ARQ (ratio of CO_2 efflux/O_2 influx for tree stems) measured in stem chambers after incubation of 30 minutes to a few hours and "steady state" ARQ measured in the same experiment with typically two days of incubation (n = 139). ~~The regression forced to go through 0. The P value is the significance of the slope estimation.~~

Figure 3: Summary of "steady state" ~~apparent respiratory quotient (~~ARQ~~) measurements~~ (ratio of CO_2 efflux/O_2 influx for tree stems) for 12 species (n measurements, n individuals). Gases were sampled from chambers at breast height (~1.3 m above soil surface), except for the *Q. calliprinos* in the Mediterranean shrubland, in which chambers were placed near the stem base due to branching stems. Vertical lines are mean values, error bars represent one standard deviation, and colored bars represent the range of measured ARQ values. ~~The Peru data is after Angert et al. (2012).~~The Peru data is after Angert et al. (2012a). The horizontal bars were ordered according to increasing mean ARQ.

Figure 4: Seasonal dynamics of "steady state" ~~apparent respiratory quotient (ARQ, the~~ ARQ (ratio of CO_2 efflux/O_2 influx for tree stems) of five individual trees from five different species. Phenology stage index determined according to: "Defoliation"- from beginning of autumn color development to the end of the fall,

"Winter dormancy"- when the tree was bare from leaves, "Leaf regeneration"- from bud burst to early leaf development stage. The *Q. calliprinos* is evergreen. Markers are mean values and error bars are SD of duplicate samples from the same stem chamber. Markers connected with solid lines represent measurements with chambers at breast height (~1.3 m above soil surface). ~~Markers~~Smaller markers connected with dashed lines represent measurements with chambers positioned at the stem base. The trees grew on Hebrew University campus in Jerusalem, Israel.

Figure 5: Instantaneous ~~apparent respiratory quotient (ARQ,~~ ARQ (ratio CO_2 efflux/O_2 influx of a stem, ± SD of duplicates) values measured over a day-night-day transition ~~in Jerusalem, Israel in~~during July 2012 (a) and April 2013 (b) from different trees growing on Hebrew University campus in Jerusalem, Israel. ~~*Q.*~~*Quercus calliprinos* was measured at two different heights on the stem. First sampling was taken during daylight (day 1), next sampling before dawn (pre-dawn) and last sampling during daylight of the successive day (day 2).

Figure 6: Diurnal patterns of (a) O_2 influx to the stem and CO_2 efflux from the stem, (b) chamber temperature, and (c) instantaneous ~~apparent respiratory quotient (~~ARQ~~,~~ (ratio CO_2 efflux/O_2 influx for tree stems). Shaded areas ~~ndicate~~indicate night periods. Error bars are 95% confidence bounds. All data were obtained from a single *M. domestica* tree during 24-28 April 2013 on Hebrew University campus in Jerusalem, Israel.

Figure 7: Instantaneous ~~apparent respiratory quotient (ARQ,~~ ARQ (ratio of CO_2 efflux/O_2 influx for tree stems) measured from stem chambers installed at different heights above the ground on a *S. micranthum* tree in Brazil. At the same heights ARQ was measured from 4 cm in-stem probes. The measurements were conducted during 30 March and 18 October 2012. Error bars represents SD of duplicate samples from the same stem chamber.

Figure 8: Comparisons of stem chamber ~~apparent respiratory quotient (~~steady state ARQ~~,~~ (ratio CO_2 efflux/O_2 influx for tree stems~~, "steady state"))~~ to ARQ measured from incubations of stem cores (ratio CO_2 increase/O_2 decrease), by species (n individuals) in different sites. Values are means ±SD.

Figure 9: (a) O_2 uptake rate (nmol $g.FW^{-1}$ s^{-1}) and (b) ~~apparent respiratory quotient (~~ARQ~~,~~ (ratio CO_2 increase/O_2 decrease) of *Q. ilex* leaves and stem cores incubated in a closed system (n =4). Values are means ± SD. Asterisks indicate significant difference between tissues at each time step (* $P < 0.05$, ** $P < 0.01$, *** $P < 0.0001$ in Student's t-test). Different letters indicate significant difference in Tukey-Cramer HSD analysis that followed one-way analysis of variance (ANOVA) within tissue type, between time steps.

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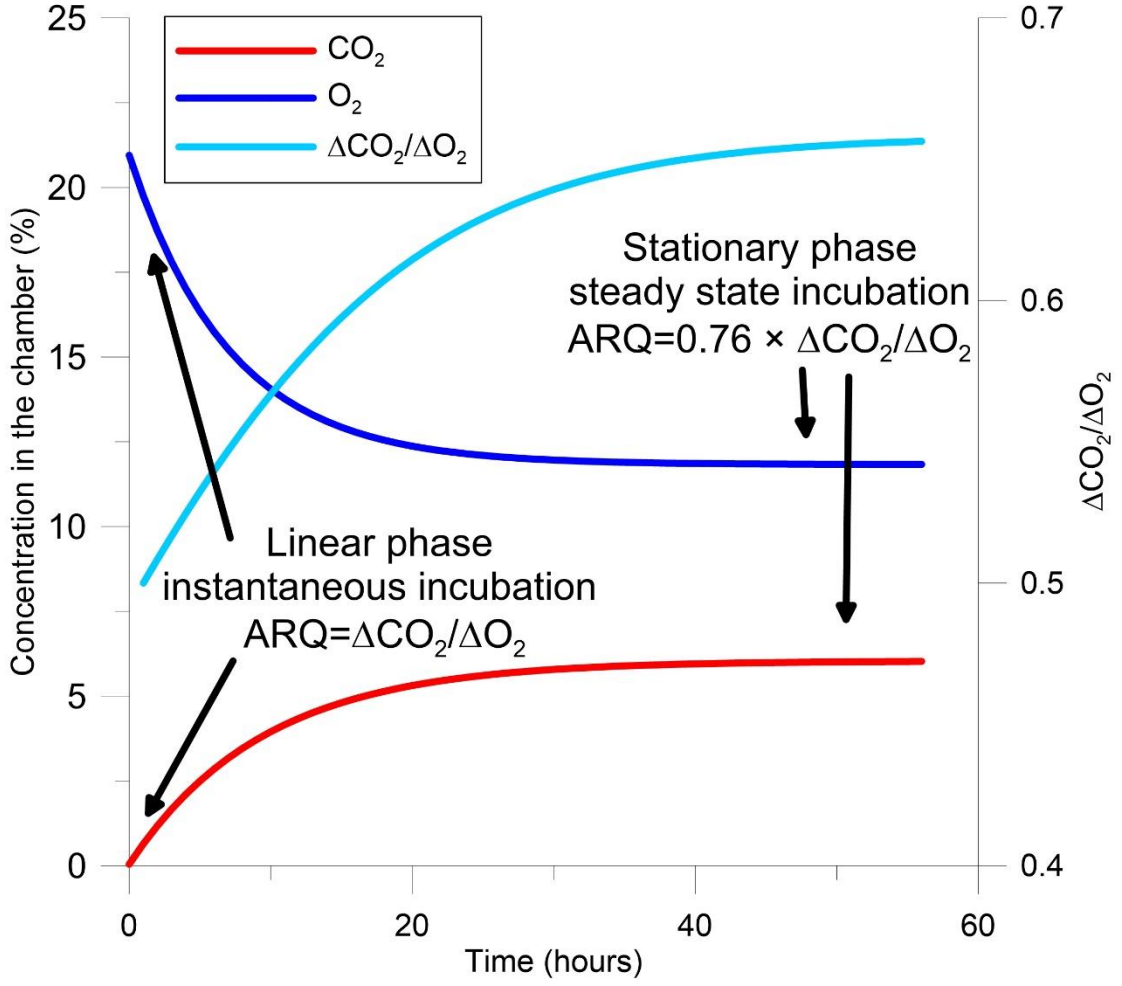
25
20
15
10
5
0
0.7
0.6
0.5
0.4
0
20
40
60
Time (hours)
Concentration in the chamber (%)
ΔCO2/ΔO2
CO2
O2
ΔCO2/ΔO2
Stationary phase
steady state incubation
ARQ=0.76 × ΔCO2/ΔO2
Linear phase
instantaneous incubation
ARQ=ΔCO2/ΔO2

Figure 1

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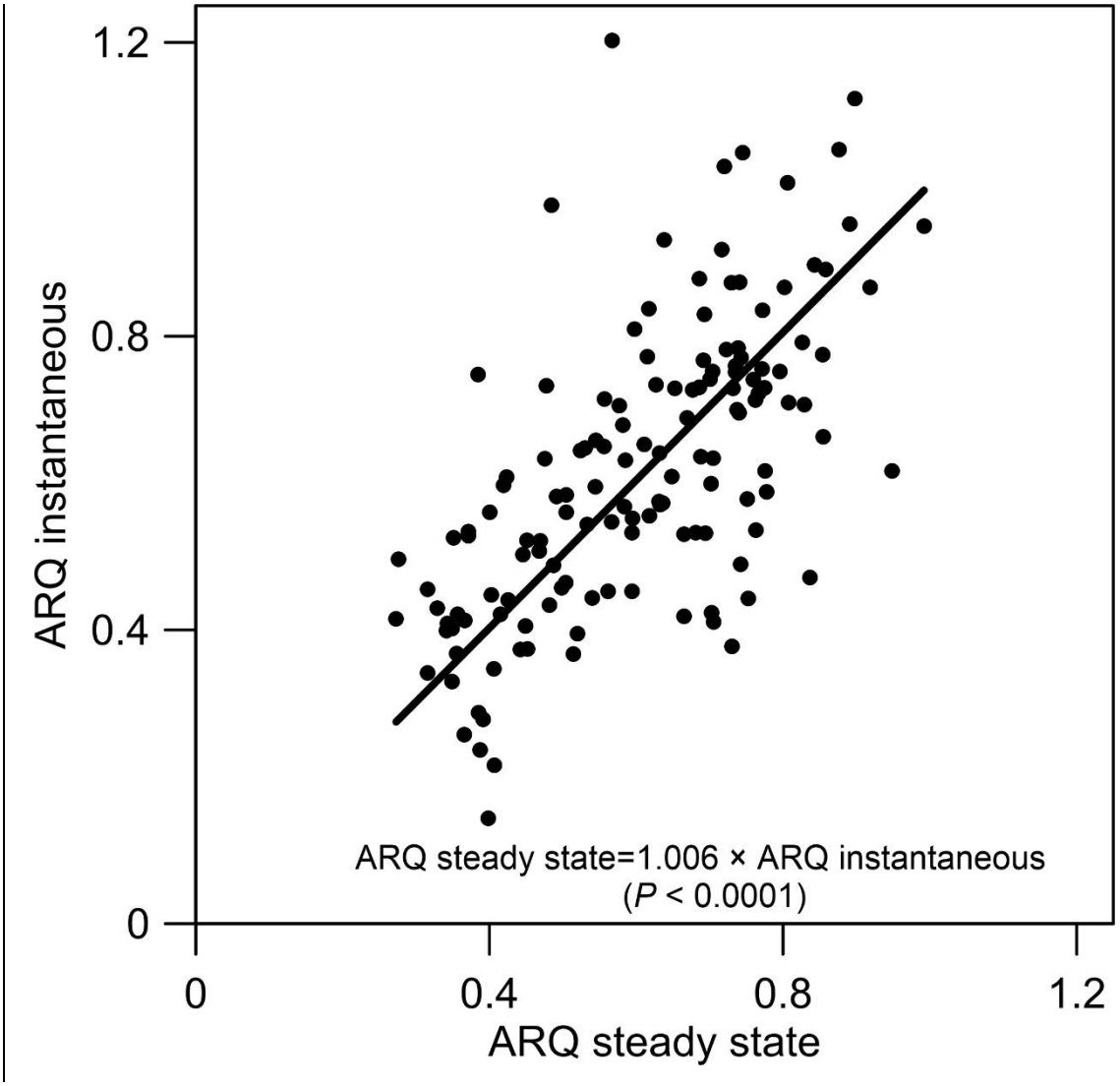

1.2
0.8
0.4
0
ARQ instantaneous
0
0.4
0.8
1.2
ARQ steady state
ARQ steady state=1.006 × ARQ instantaneous
(P < 0.0001)

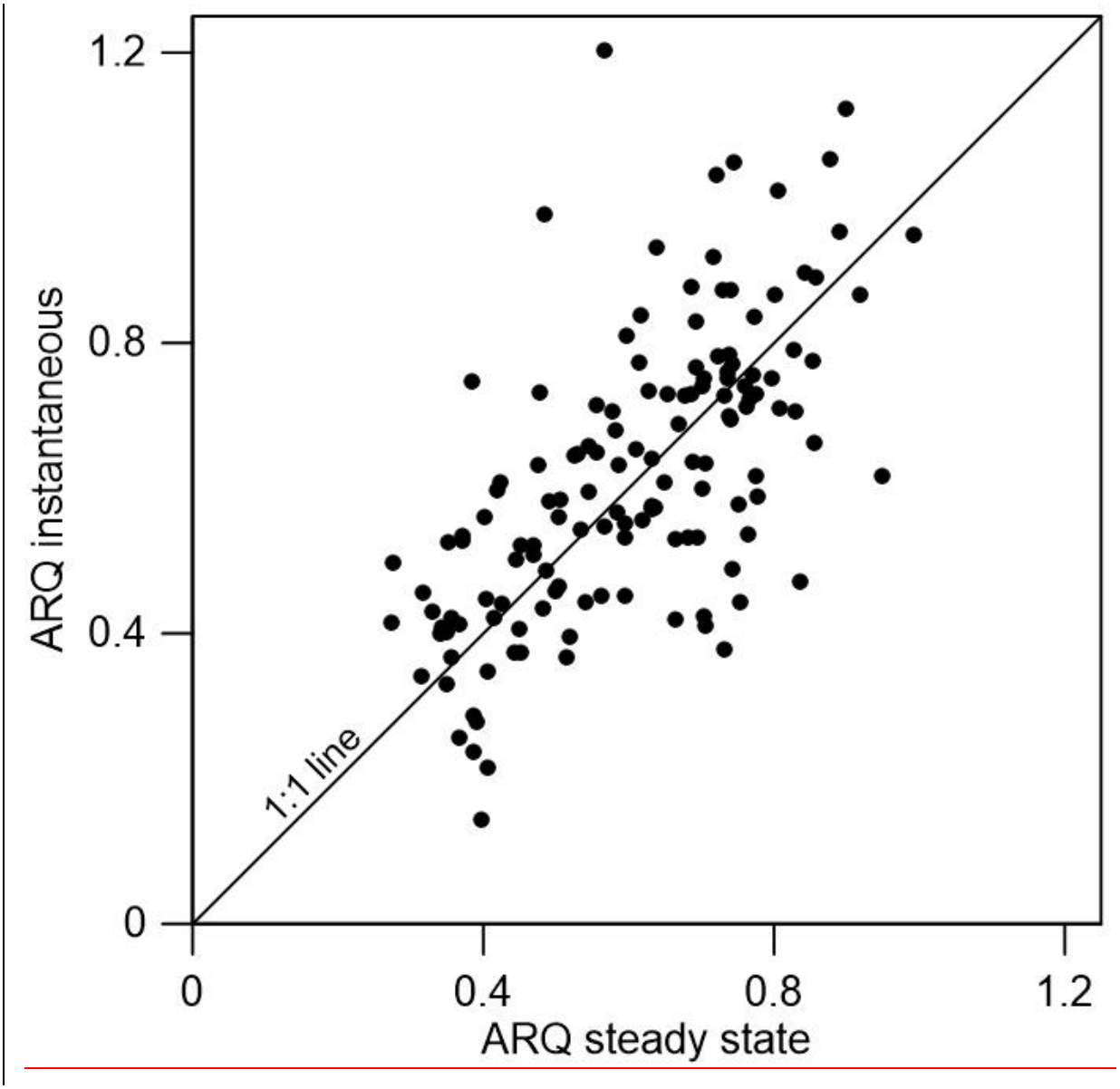
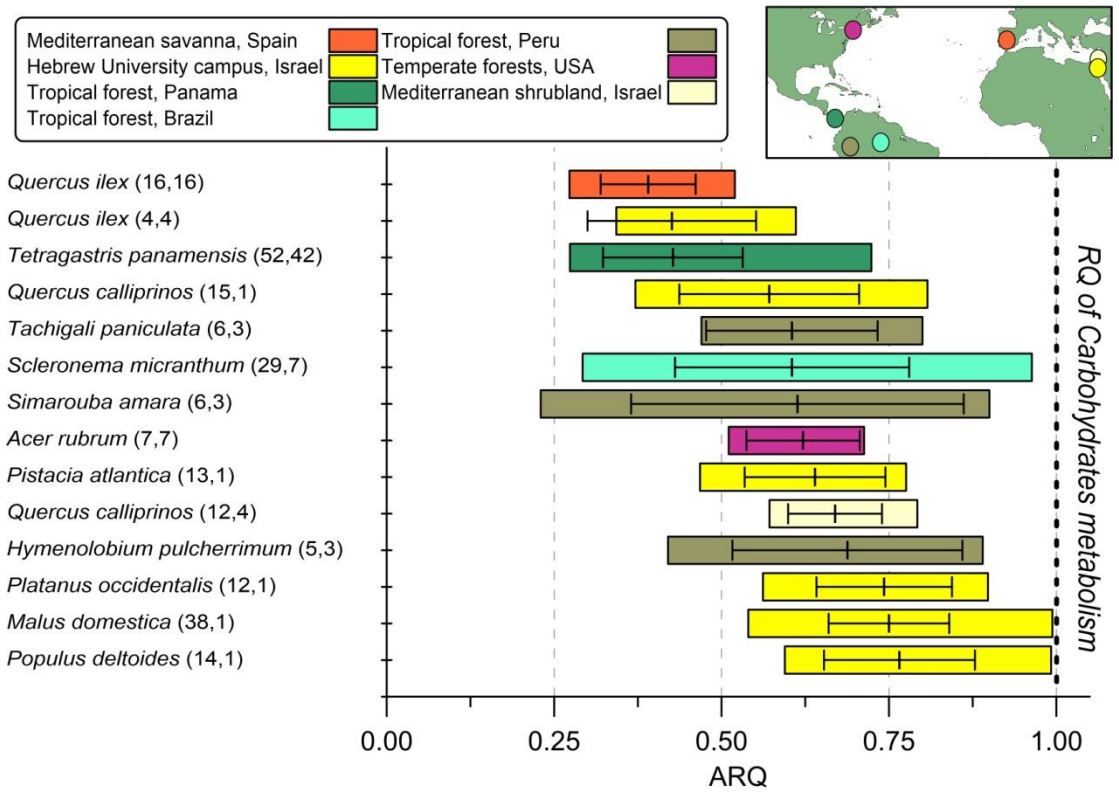

1.2
0.8
0.4
0
ARQ instantaneous
1:1 line
0
0.4
0.8
1.2
ARQ steady state

Figure2

5

5


Mediterranean savanna, Spain
Hebrew University campus, Israel
Tropical forest, Panama
Tropical forest, Brazil
Tropical forest, Peru
Temperate forests, USA
Mediterranean shrubland, Israel
Quercus ilex (16,16)
Quercus ilex (4,4)
Tetragastris panamensis (52,42)
Quercus calliprinos (15,1)
Tachigali paniculata (6,3)
Scleronema micranthum (29,7)
Simarouba amara (6,3)
Acer rubrum (7,7)
Pistacia atlantica (13,1)
Quercus calliprinos (12,4)
Hymenolobium pulcherrimum (5,3)
Platanus occidentalis (12,1)
Malus domestica (38,1)
Populus deltoides (14,1)
0.00
0.25
0.50
0.75
1.00
ARQ
RQ of Carbohydrates metabolism

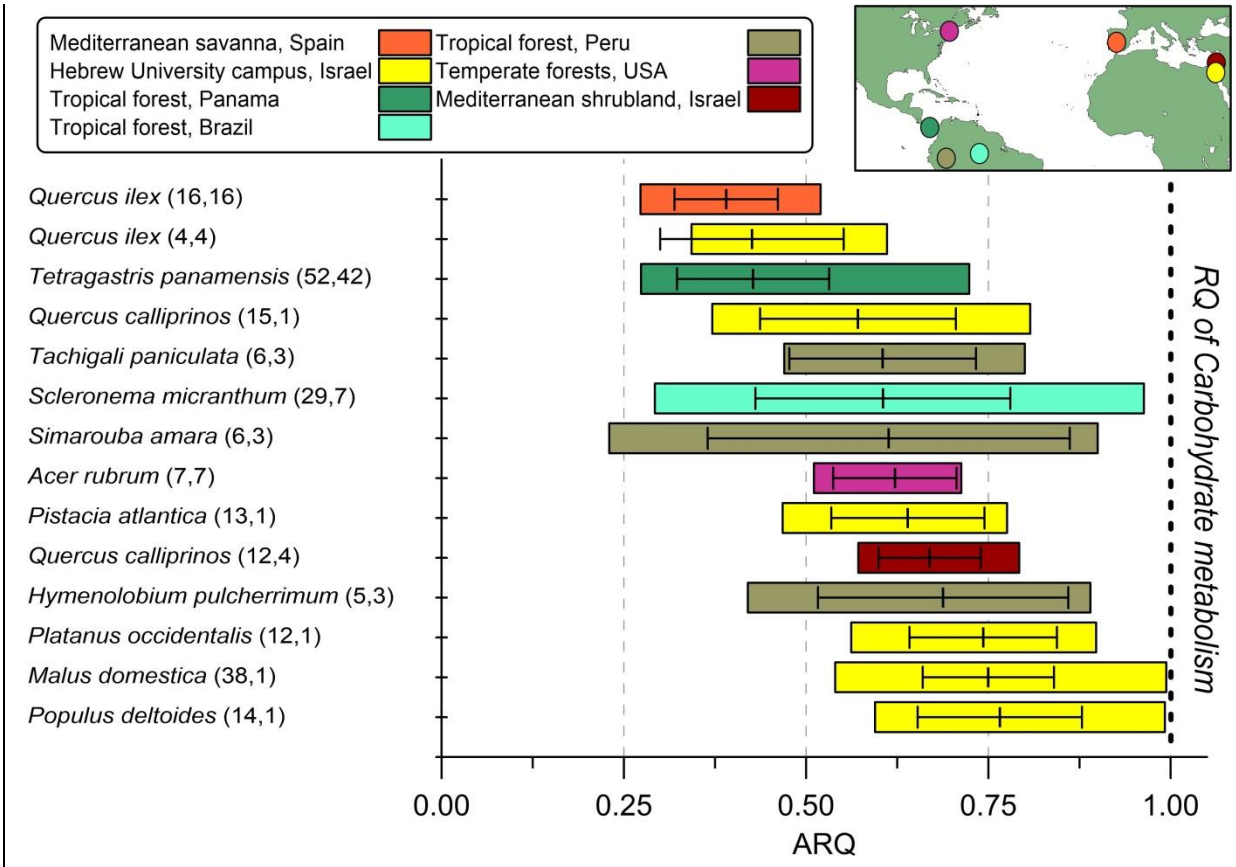
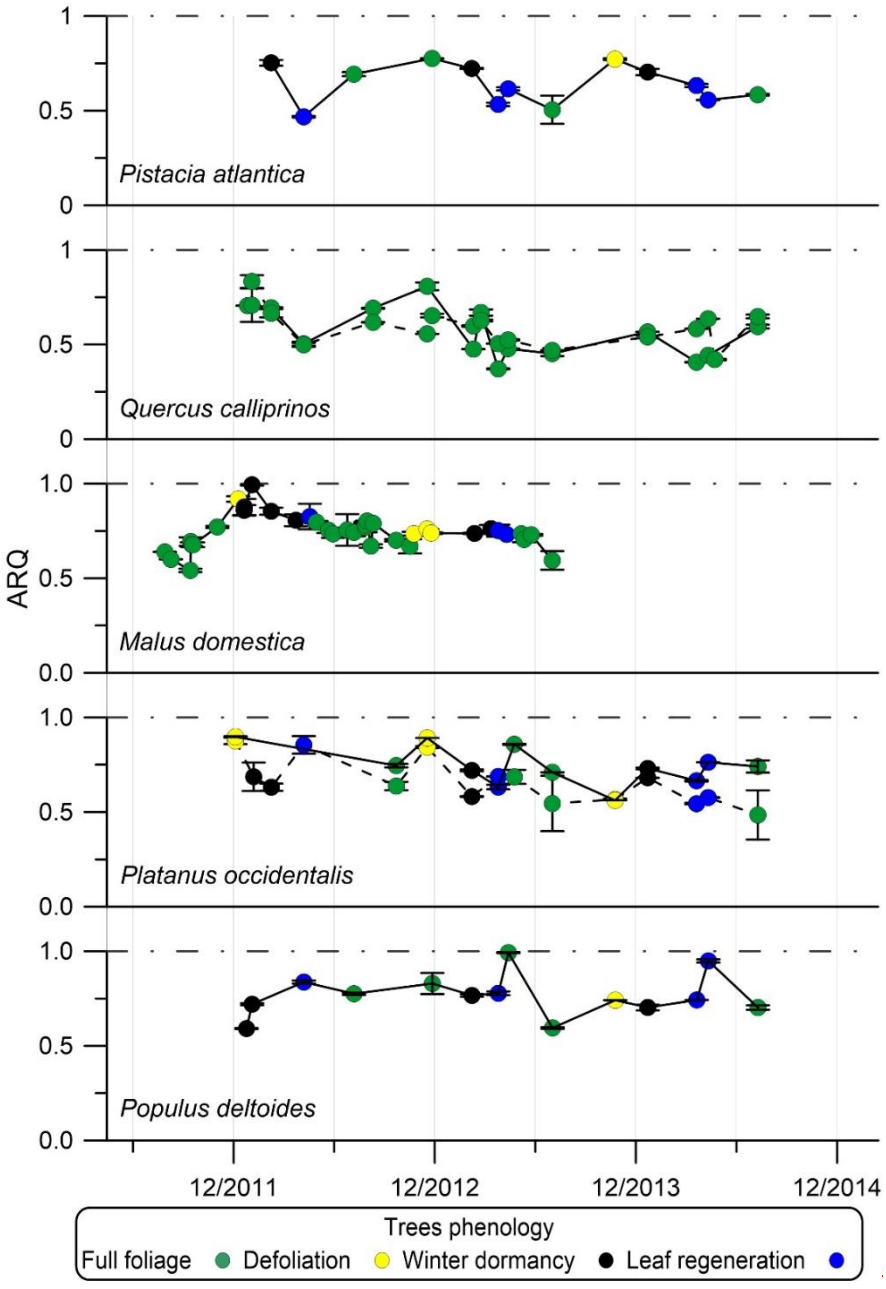

Mediterranean savanna, Spain
Hebrew University campus, Israel
Tropical forest, Panama
Tropical forest, Brazil
Tropical forest, Peru
Temperate forests, USA
Mediterranean shrubland, Israel
Quercus ilex (16,16)
Quercus ilex (4,4)
Tetragastris panamensis (52,42)
Quercus calliprinos (15,1)
Tachigali paniculata (6,3)
Scleronema micranthum (29,7)
Simarouba amara (6,3)
Acer rubrum (7,7)
Pistacia atlantica (13,1)
Quercus calliprinos (12,4)
Hymenolobium pulcherrimum (5,3)
Platanus occidentalis (12,1)
Malus domestica (38,1)
Populus deltoides (14,1)
0.00
0.25
0.50
0.75
1.00
ARQ
RQ of Carbohydrate metabolism

Figure 3


ARQ
Pistacia atlantica
Quercus calliprinos
Malus domestica
Platanus occidentalis
Populus deltoides
12/2011
12/2012
12/2013
12/2014
Trees phenology
Full foliage
Defoliation
Winter dormancy
Leaf regeneration

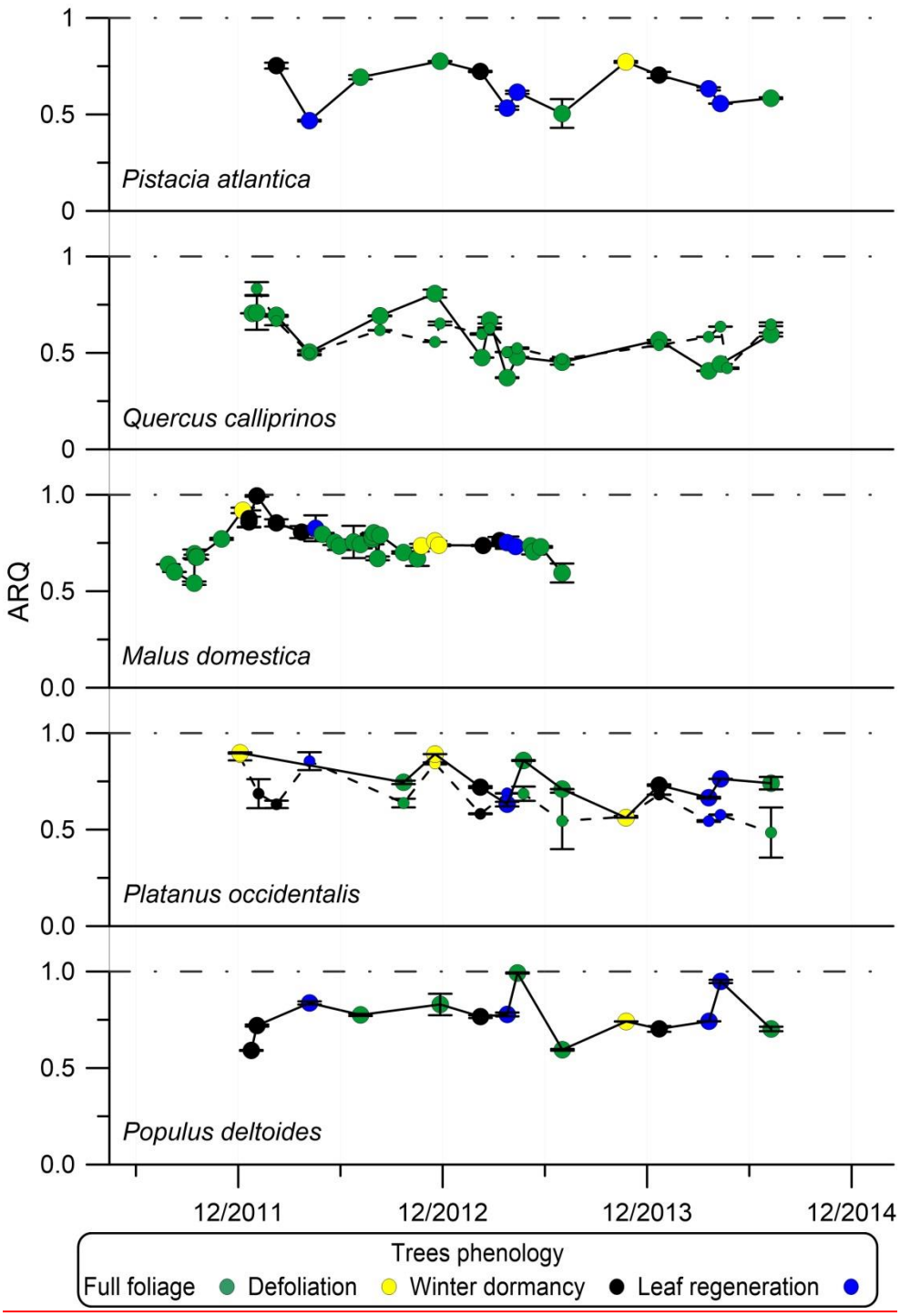

ARQ
Pistacia atlantica
Quercus calliprinos
Malus domestica
Platanus occidentalis
Populus deltoides
12/2011
12/2012
12/2013
12/2014
Trees phenology
Full foliage
Defoliation
Winter dormancy
Leaf regeneration

Figure 4

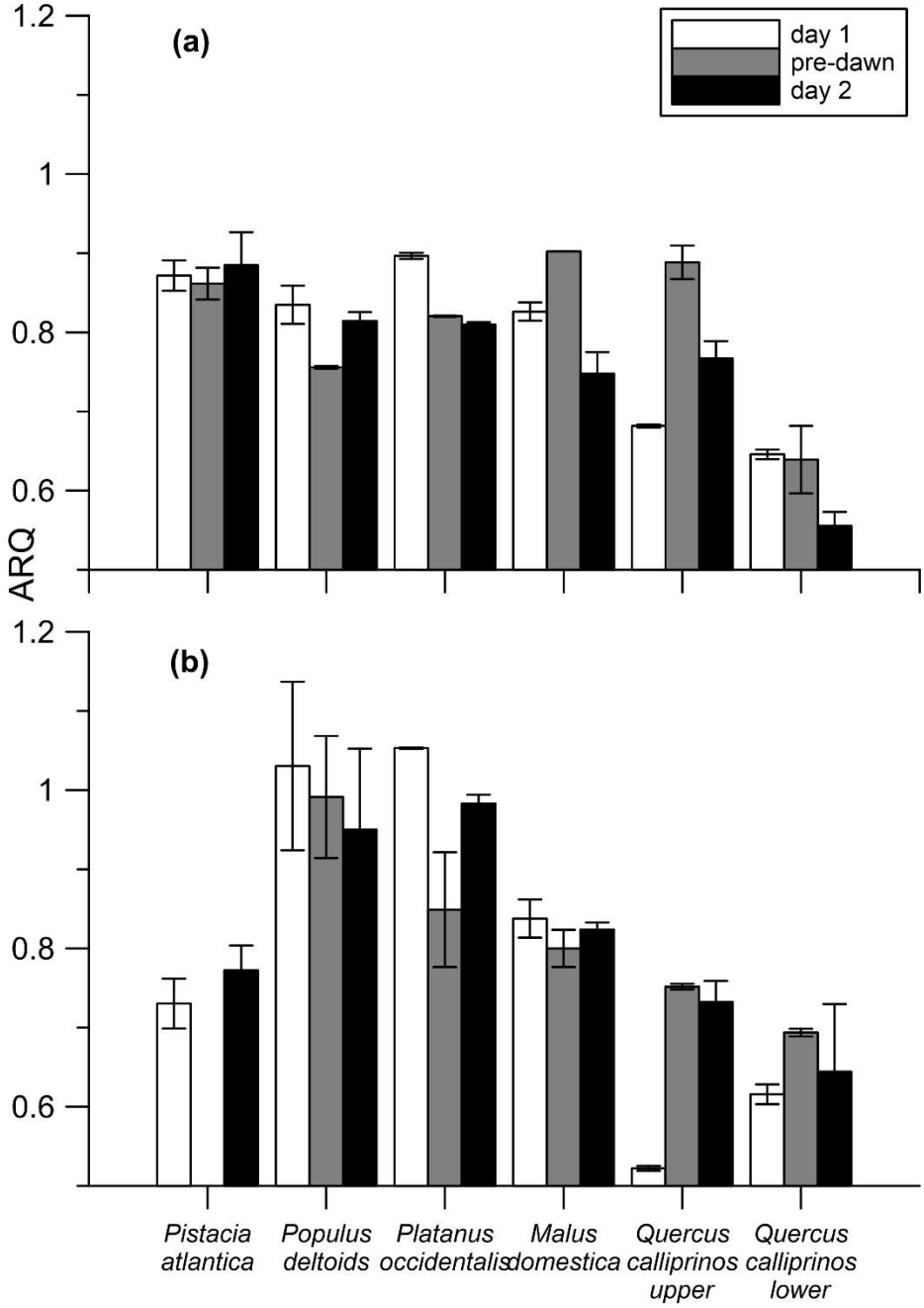

(a)
(b)
day 1
pre-dawn
day 2
ARQ
1.2
1
0.8
0.6
Pistacia atlantica
Populus deltoids
Platanus occidentalis
Malus domestica
Quercus calliprinos upper
Quercus calliprinos lower

Figure 5

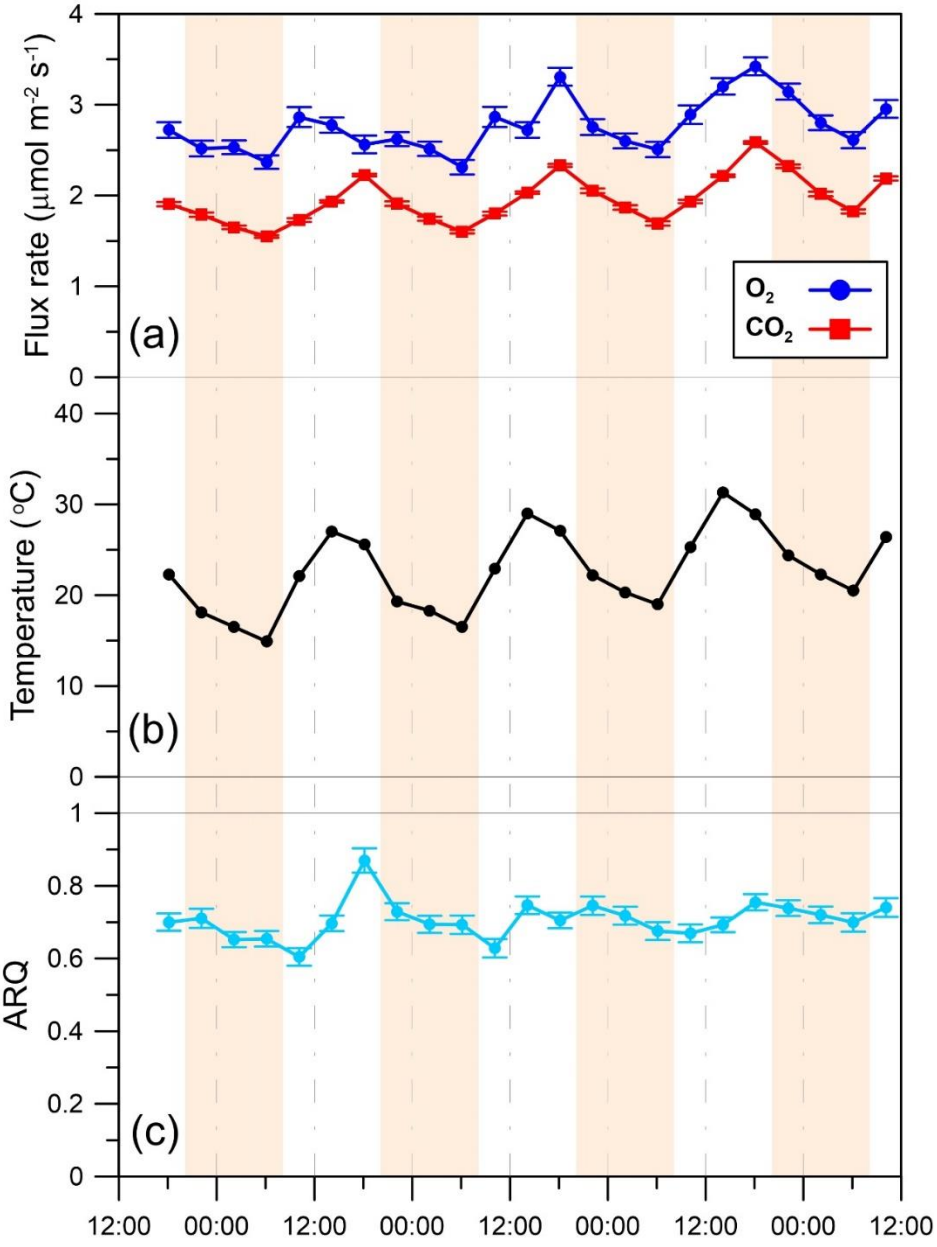

Flux rate (μmol m-2 s-1)
4
3
2
1
0
(a)
O2
CO2
Temperature (°C)
40
30
20
10
0
(b)
ARQ
1
0.8
0.6
0.4
0.2
0
(c)
12:00
00:00
12:00
00:00
12:00
00:00
12:00
00:00
12:00

Figure 6

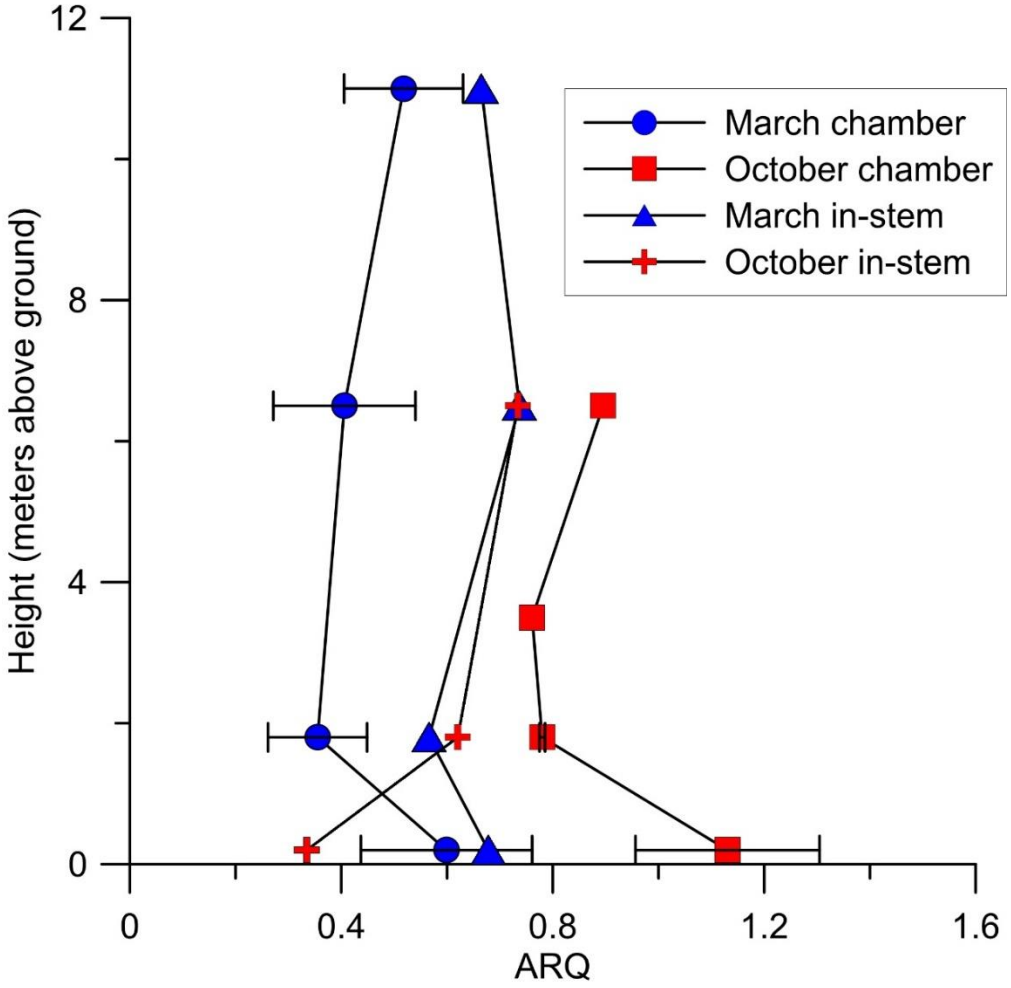


Figure 7

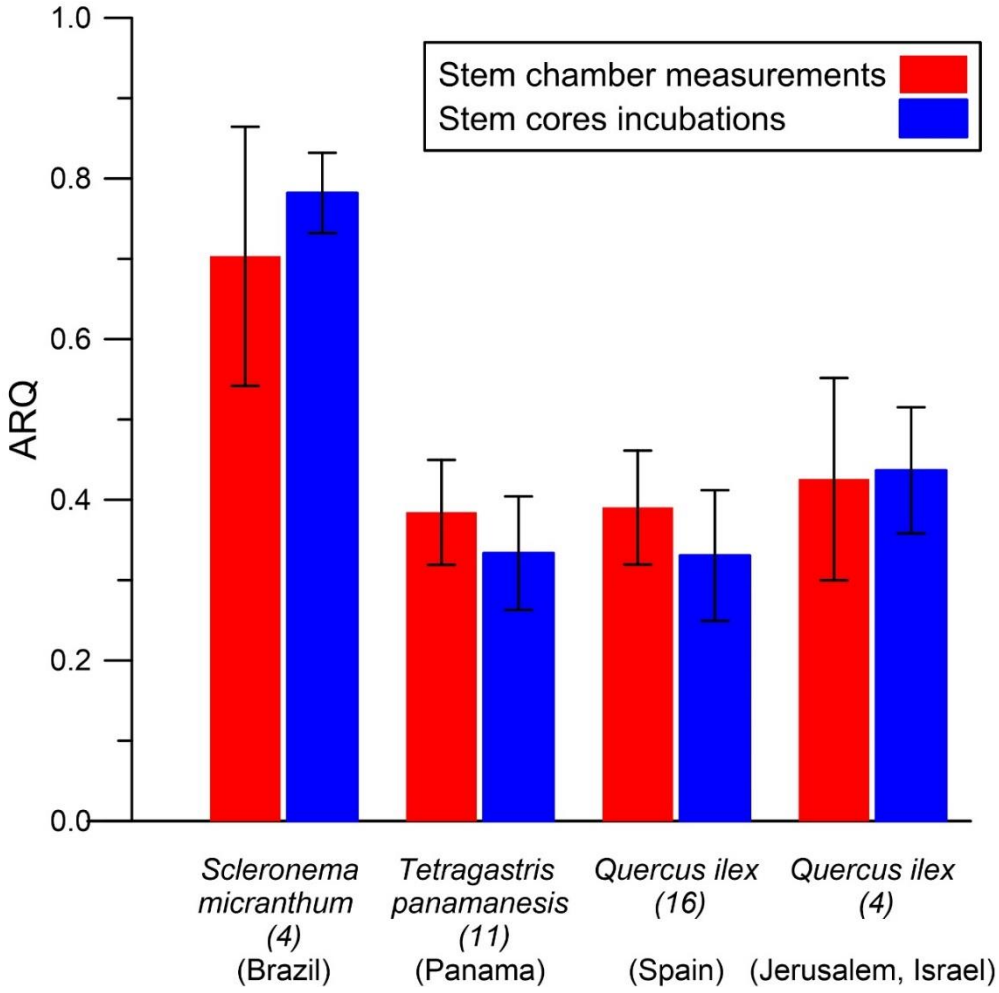

1.0
0.8
0.6
0.4
0.2
0.0
ARQ
Stem chamber measurements
Stem cores incubations
Scleronema micranthum (4) (Brazil)
Tetragastris panamanesis (11) (Panama)
Quercus ilex (16) (Spain)
Quercus ilex (4) (Jerusalem, Israel)

Figure 8

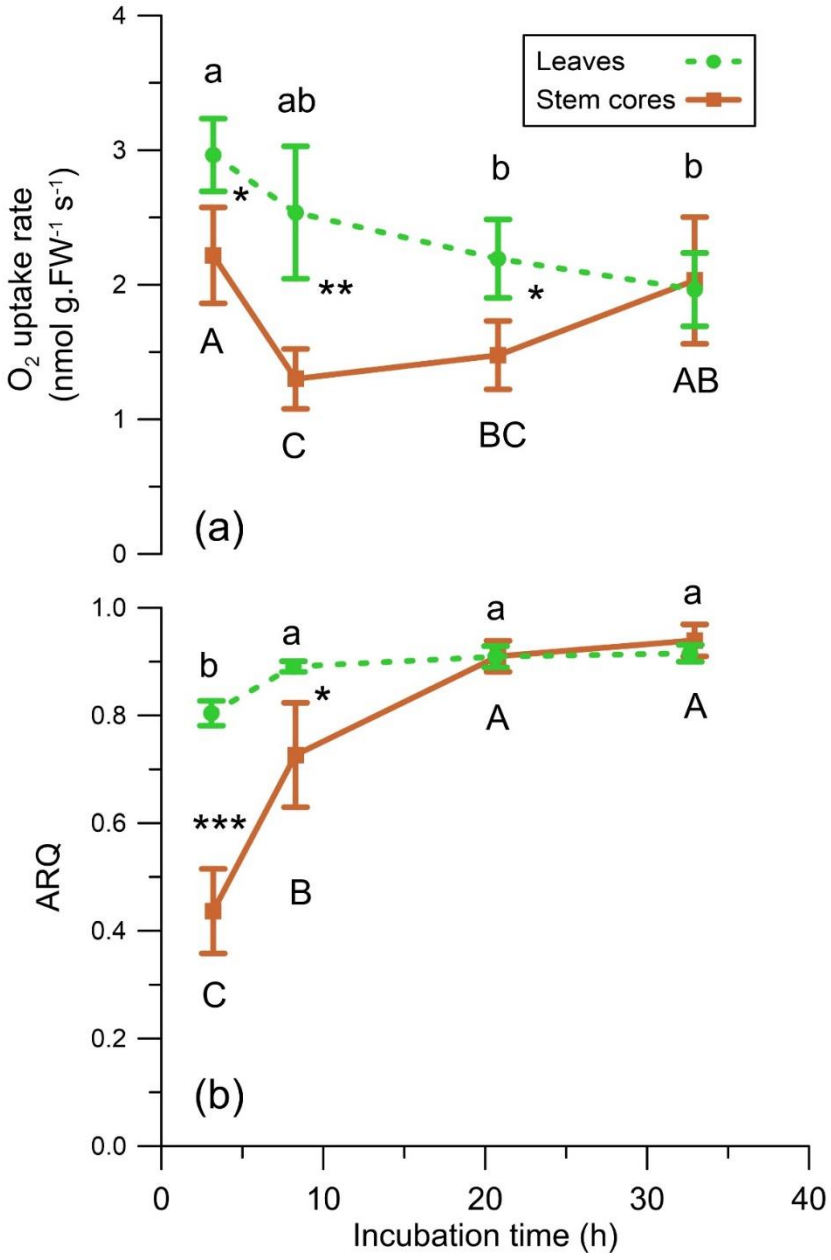

Leaves
Stem cores
O_2 uptake rate (nmol g.FW^{-1} s^{-1})
a
ab
b
b
A
C
BC
AB
(a)
ARQ
b
a
a
a
A
A
C
B
(b)
Incubation time (h)

Figure 9