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Alteration of nitrous oxide emissions from floodplain soils by aggregate size, litter accumulation and plant soil interactions

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Abstract. Semi–terrestrial soils such as floodplain soils are considered potential hotspots of nitrous oxide (N_2O) emissions. Microhabitats in the soil, such as within and outside of aggregates, in the detritusphere, and/or in the rhizosphere, are considered to promote and preserve specific redox conditions. Yet, our understanding of the relative effects of such microhabitats and their interactions on N_2O production and consumption in soils is still incomplete. Therefore, we assessed the effect of aggregate size, buried organic matter, and rhizosphere processes on the occurrence of enhanced N_2O emissions under simulated flooding/drying conditions in a mesocosm experiment. We used two model soils with equivalent structure and texture, comprising macroaggregates (4000–250 µm) or microaggregates (< 250 µm) from a N-rich floodplain soil. These model soils were either planted with basket willow (*Salix viminalis* L.), mixed with leaf litter, or left unamended. After 48 hours of flooding, a period of enhanced N_2O emissions occurred in all treatments. The unamended model soils with macroaggregates emitted significantly more N_2O during this period than those with microaggregates. Litter addition modulated the temporal pattern of the N_2O emission, leading to short-term peaks of high N_2O fluxes at the beginning of the period of enhanced N_2O emissions. The presence of *S. viminalis* strongly suppressed the N_2O emission from the macroaggregated model soil, masking any aggregate size effect. Integration of the flux data with data on soil bulk density, moisture, redox potential and soil solution composition suggest that macroaggregates provided more favorable conditions for spatially coupled nitrification–denitrification, which are particularly conducive to net N_2O production, than microaggregates. The local increase in organic carbon in the detritusphere appears to first stimulate N_2O emissions, but ultimately, respiration of the surplus organic matter shifts the system towards redox conditions where N_2O reduction to N_2 dominates. Similarly, the low emission rates in the planted soils can be best explained by root exudation of low-molecular weight organic substances supporting complete denitrification in the anoxic zones, but also by the inhibition of denitrification in the zone above, where rhizosphere aeration takes place. Together, our experiments highlight the importance of microhabitat formation in regulating O_2 content and the completeness of denitrification in soils during drying after saturation. Moreover, they will help to better predict the conditions under which hotspots and moments of enhanced N_2O emissions are most likely to occur in hydrologically dynamic soil systems like floodplain soils.

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1. Introduction

Nitrous oxide (N_2O) is a potent greenhouse gas with a global warming potential over a 100 year time horizon 298 times higher than the one of carbon dioxide (Forster et al., 2007). Given its role as climate-relevant gas and in the depletion of stratospheric ozone (Ravishankara et al., 2009), the steady increase of its average atmospheric concentration of 0.75ppb yr^{-1} (Hartmann et al., 2013) asks for a quantitative understanding of its sources and the factors that control its production. On a global scale, vegetated soils are the main natural terrestrial sources of N_2O. Agriculture is the main anthropogenic source and the main driver of increasing atmosphere N_2O concentrations (Ciais et al., 2013).

In soils, several biological nitrogen (N) transformation processes produce N_2O either as a mandatory intermediate or as a by-product (Spott et al., 2011). Under oxic conditions, the most important process is obligate aerobic nitrification that yields N_2O as by-product when hydroxylamine decomposes (Zhu et al., 2013). Under low oxygen (O_2) availability, nitrifier denitrification and heterotrophic denitrification with N_2O as intermediate become more relevant (Philippot et al., 2009). At stably anoxic conditions and low concentrations of nitrate, complete denitrification consumes substantial amounts of previously produced N_2O by further reduction to N_2 (Baggs, 2008; Vieten et al., 2009). In environments that do not sustain, stable anoxia but undergo sporadic transitions between oxic and anoxic conditions, the activity of certain N_2O reductases can be suppressed by transiently elevated O_2 concentration and thus can lead to the accumulation of N_2O (Morley et al., 2008).

Nitrous oxide emissions from soils depend on the availability of carbon (C) and N substrates that fuel the involved microbial processes. On the other hand, given its dependency on O_2, N_2O production is also governed by the diffusive supply of O_2 through soils. Similarly, soil N_2O emissions are modulated by diffusive N_2O transport from the site of production to the soil surface (e.g. Böttcher et al., 2011; Heincke and Kaupenjohann, 1999). Substrate availability, gas diffusivity, and the distribution of soil organisms are highly heterogeneous in soils at a small scale, with micro-niches in particular within soil aggregates, within the detritusphere, and within the rhizosphere. These can result in “hot spots” with high denitrification activity (Kuzyakov and Blagodatskaya, 2015).

Soil aggregate formation is a key process in building soil structure and pore space. Soil aggregates undergo different stages in their development, depending on the degradability of the main binding agent (Tisdall and Oades, 1982). Initially, highly persistent primary organo–mineral clusters (20–250 µm) are held together by root hairs and hyphae, thus forming macroaggegates (> 250 µm). Upon decomposition of these temporary binding agents and the subsequent disruption of the macroaggregates, microaggregates (< 250 µm) are released (Elliott and Coleman, 1988; Oades, 1984; Six et al., 2004). These consist of clay-encrusted fragments of organic debris coated with polysaccharides and proteins. This multi-stage development leads to a complex relationship between aggregate size, intra-aggregate structure and soil structure (Ball, 2013; Totsche et al., 2017), which influences soil aeration, substrate distribution and pore water dynamics (Six et al., 2004). Often, micro-site heterogeneity increases with aggregate size, thus fostering the simultaneous activity of different N_2O producing microbial communities with distinct functional traits (Bateman and Baggs, 2005). Aggregate size effects on N_2O production and consumption have generally been studied in static batch incubation experiments with a comparatively small number of isolated aggregates of uniform size, at constant levels of water saturation (Diba et al., 2011; Drury et al., 2004; Jahangir et al., 2011; Khalil et al., 2005; Sey et al., 2008), and through modelling approaches (Renault and Stengel, 1994; Stolk et al., 2011). Previous work provided partially inconsistent results,

which led to an ongoing discourse about the interplay of physicochemical properties and different aggregate sizes in controlling N_2O emission. For example, ostensible inconsistencies may be attributed to the use of different aggregate size classes, other methodological constraints (water saturation, redox potential), and differences in microbial communities. The effects of aggregate size, in combination with fluctuating water saturation, on soil N_2O emissions have, to our knowledge, not been addressed specifically.

Similar to soil aggregates, the detritusphere and the rhizosphere (the zone of the soil that is affected by root activity (Baggs, 2011; Luster et al., 2009), can be considered biogeochemical hot spots (Kuzyakov and Blagodatskaya, 2015; Myrold et al., 2011). Here, carbon availability is much higher than in the bulk soil and thus rarely limiting microbial process rates. The detritusphere consists of dead organic material, which spans a wide range of recalcitrance to microbial decomposition. Spatially confined accumulations of variably labile soil litter form microhabitats that are often colonized by highly active microbial communities (Parkin, 1987). Aggregation of litter particles has been shown to affect N_2O emissions (Loecke and Robertson, 2009). Hill (2011) identified buried organic-rich litter horizons in a stream riparian zone as hot spots of N cycling. Similarly, in the rhizosphere, root exudates and exfoliated root cells provide ample degradable organic substrate for soil microbes (Robertson and Groffman, 2015). Yet, plant growth may also affect soil microbial communities through competition for water and nutrients (e.g., fixed N) (Bender et al., 2014; Myrold et al., 2011). The combined effects of these plant-soil interactions on N_2O production have been reviewed by Philippot et al. (2009). Root-derived bioavailable organic compounds can stimulate heterotrophic microbial activity, specifically N mineralization and denitrification. Nitrification in turn can be enhanced by the elevated N turnover and mineralization rates, but may also be negatively affected by specific inhibitors released from the root or through plant-driven ammonium depletion. The ability of some plants adapted to water-saturated conditions to „pump“ air into the rhizosphere via aerenchyma (gas conductive channels in the root) leads to an improved oxygenation of the rhizosphere and a stimulation of nitrification (Philippot et al., 2009). Surrounded by otherwise anoxic sediments, such aerated micro-environments may create optimal conditions for coupled nitrification–denitrification (Baldwin and Mitchell, 2000; Koschorreck and Darwich, 1998). On the other hand, transport of N_2O produced in the soil to the atmosphere is may be facilitated via these plant-internal channels, bypassing diffusive transport barriers and enhancing soil–atmosphere gas fluxes (Jørgensen et al., 2012).

The dynamics of N_2O emissions are strongly coupled to the dynamics of pore water. Re-wetting of previously dried soil can lead to strong N_2O emissions (Goldberg et al., 2010; Ruser et al., 2006), likely fostered by a wetting-induced flush in N mineralization (Baldwin and Mitchell, 2000). On the other hand, the drying-phase after water saturation of sediments and soils can lead to a period of enhanced N_2O emissions (e.g. Baldwin and Mitchell, 2000; Groffman and Tiedje, 1988; Rabot et al., 2014; Shrestha et al., 2012) when water-filled pore space (WFPS) exceeds 60% (Beare et al., 2009; Rabot et al., 2014). The increased N_2O production has been attributed to enhanced coupled nitrification–denitrification (Baldwin and Mitchell, 2000). Depending on the spatial distribution of water films around soil particles and tortuosity (which is a function of aggregate size and soil structure), the uneven drying of the soil after full saturation may generate conditions that are conducive to the formation of anaerobic zones in otherwise oxic environments (Young and Ritz, 2000). Pore water thereby acts as a diffusion barrier for gas exchange, limiting the O_2 availability in the soil pore space (Butterbach-Bahl et al., 2013). Moreover, pore water serves as a medium for the diffusive dispersal of dissolved C and N substrates, e.g. from the site of litter decomposition to spatially separated N_2O producing microbial communities (Hu et al., 2015). Therefore, fluctuations in water saturation efficiently promote the development of hot spots and hot

moments of N_2O emissions in floodplain soils and other semi-terrestrial soils (Hefting et al., 2004; Shrestha et al., 2012).

The main objective of the present experimental study was to contribute to a better understanding of the factors governing the formation and emission of N_2O in floodplain soils during hot moments after flooding events. Towards this objective, we performed a mesocosm flooding simulation experiment under controlled conditions, with model soils of largely similar structure, but differing in the size distribution of original soil aggregates. We included two additional factorial treatments: a willow-litter addition treatment to assess whether aggregate size effects are modified by such a detritusphere, and a willow cuttings treatment to test whether aggregate effects change in the presence of plants, as result of root soil interactions.

We demonstrate that the level of soil aggregation affects N_2O emission rates from floodplain soils through its modulating control on the model soils physicochemical properties. We further show that these effects are modified by the presence of a detritusphere and by root–soil interactions, through effects on carbon and N substrate availability and redox conditions.

2. Material and methods

2.1 Model soils

In February 2014, material from the uppermost 20 cm of a N-rich gleyic Fluvisol (calcaric, humic siltic) with 20% sand and 18% clay (Samaritani et al., 2011) was collected in the restored Thur River floodplain near Niederneunforn (NE Switzerland 47°35’ N, 8°46’ E, 453 m.a.s.l.; MAT 9.1 °C; MAP 1015 mm). After removing plant residues such as roots, twigs and leaves, the soil was mixed and air-dried to a gravimetric water content of 24.7 ± 0.4 %. In the next step, the floodplain soil material was separated into a macroaggregate fraction (250–4000 µm) and a microaggregate fraction (< 250 µm) by dry sieving. The threshold of 250 µm between macroaggregates and microaggregates was chosen based on Tisdall and Oades (1982). Soil aggregate fractions were then used to re-compose model soils. In order to preserve the original soil structure, the remaining aggregate size fractions were complemented with an inert matrix replacing the removed aggregate size fraction of the original soil. Model Soil 1 (LA) was composed of soil macroaggregates mixed in a 1:1 (w/w) ratio with glass beads of 150–250 µm size serving as inert matrix material replacing the microaggregates of the original soil. Similarly, Model Soil 2 (SA) was composed of soil microaggregates mixed at the same ratio with fine quartz gravel of 2000–3200 µm size. To generate an even mixture of original soil aggregates and the respective inert matrix a Turbula mixer (Willy A. Bachofen AG, Muttenz, Switzerland) was used. The physicochemical properties of the two soils were determined by analysing three random samples of each model soil. Texture of the complete model soils was determined using the pipette method (Gee and Bauder, 1986) and pH was measured potentiometrically in a stirred slurry of 10 g soil in 20 ml of 0.01 M $CaCl_2$. Additionaly C_{org} and N_{tot} were analysed in both aggregate size fractions without the inert material, using the method described by Walthert et al. (2010). The two model soils displayed very similar physicochemical properties (Table 1), except for the C:N ratio that was lower in macroaggregates than in microaggregates. The latter was due to the slightly lower organic carbon content in concert with slightly higher N_{tot} values in the macroaggregates. The high calcium carbonate ($CaCO_3$) content of the source material of our model soils (390 ± 3 g $CaCO_3$ kg^{-1}; Samaritani et al., 2011) buffered the systems at an alkaline pH of 8.00 ± 0.02 for LA and 7.56 ± 0.01 for SA respectively (Table 1), ensuring that the activity of key N-transforming enzymes was not hampered by too low pH, and that the potential

for simultaneous production and consumption of N_2O in our experiment was fully intact (Blum et al., 2018; Frame et al., 2017).

2.2 Mesocosms

For the mesocosm experiments, transparent polyvinyl chloride (PVC) cylinders with polymethyl methacrylate (PMMA) couplings were used. A mesocosm comprised a bottom column section, containing the soil material and a drainage layer as described below, and the upper headspace section with a detachable headspace chamber (Fig. 1). Each column section was equipped with two suction cups (Rhizon MOM Soil Moisture Samplers, Rhizosphere Research Products, Netherlands; pore size 0.15 µm) for soil solution sampling. The suction cups were horizontally inserted at 5 cm and 20 cm below soil surface. For redox potential measurements, two custom-made Pt electrodes (tip with diameter of 1 mm and contact length of 5 mm) were placed horizontally at a 90° angle to the suction cups at the same depths, with the sensor tip being located 5 cm from the column wall. A Ag/AgCl reference electrode (B 2820, SI Analytics, Germany) was installed as shown in Fig. 1. A volumetric water content (VWC) sensor (EC-5, Decagon, USA) was installed 15 cm below the soil surface. To avoid undesired waterlogging, each column section contained a 5 cm thick drainage layer composed of quartz sand with the grain size decreasing with depth from 1 mm to 5.6 mm (Fig. 1). The upper cylinder section was equipped with three way valves for gas sampling, and an additional vent for pressure compensation.

2.3. Experimental setup

The mesocosm experiment had a two factorial design where factor 1 (model soil) had two levels (macroaggregates or microaggregates) and factor 2 (treatment) had three levels (unamened, litter addition and plant presence) resulting in six treatments, each replicated six times (Table 2). As basic material, each mesocosm contained 8.5 kg of either of the two model soils. Unamended model soils were used to investigate exclusively the effect of aggregate size, abbreviated as LAU (large aggregates unamended) and SAU (small aggregates unamended), respectively. In order to assess detritusphere effects, two sets of mesocosms were amended with freshly collected leaves of Basket Willow (*Salix viminalis* L.). Those leaves were cut into small pieces, autoclaved, and then added to the model soil components (8 g kg^{-1} model soil) during the mixing procedure to create treatments LAL (large aggregates litter) and SAL (small aggregates litter), respectively. A third set of mesocosms was planted with cuttings collected from the same basket willow (*Salix viminalis* L.) to evaluate the effects of root–soil interactions in the respective model soils. For each mesocosm (treatments LAP/large aggregate plants and SAP/small aggregates plants, respectively) one cutting was inserted 10 cm into the soil, protruding from the surface about 3 cm.

The addition of leaf litter to the model soils led to an increase of C_{org} and total nitrogen (TN) in LAL relative to LAU by 41 % and 35 %, respectively, and in SAL relative to SAU by 58 % and 44 % respectively. The bulk density of the unamended model soil SAU (1.27 ± 0.01 g cm^{-3}) was slightly higher than the one of LAU (1.22 ± 0.01 g cm^{-3}; adj. *P*: < 0.0001). Regarding the litter addition treatments, the bulk density of LAL (1.13 ± 0.01 g cm^{-3}) was significantly smaller than the one of LAU (adj. *P*: < 0.0001), whereas the bulk density of SAL (1.27 ± 0.02 g cm^{-3}) did not differ significantly from the one of SAU. The soils in the treatments with plants exhibited a similar bulk density (LAP: 1.23 ± 0.02 g cm^{-3}; SAP: 1.24 ± 0.01 g cm^{-3}) as in the respective unamended treatments.

The experiments were conducted inside a climate chamber set to constant temperature (20 ± 1 °C) and relative air humidity (60 ± 10%), with a light/dark cycle of 14/10 h (PAR 116.2 ± 13.7 µmol m^{-2} s^{-1}). The experimental period was divided into four consecutive phases: The conditioning phase (phase 1) lasted for 15 weeks and allowed the model soils to equilibrate and the plants to develop a root system. This was followed by the first experimental phase of nine days (phase 2), serving as a reference period under steady-state conditions. During phases 1 and 2, the soils were continuously irrigated with artificial river water (Na^+: 0.43 µM; K^+: 0.06 µM; Ca^{2+}: 1.72 µM; Mg^{2+}: 0.49 µM; Cl^-: 4.04 µM; NO_3^-: 0.16 µM; HCO_3^-: 0.5 µM; SO_4^{2-}: 0.11 µM; pH: 7.92) via suction cups, to maintain a volumetric water content of 35 ± 5 %. In phase 3, the mesocosms were flooded by pumping artificial river water through the drainage vent at the bottom into the cylinder (10 mL min^{-1}, using a peristaltic pump; IPC-N-24, Ismatec, Germany) until the water level was 1 cm above the soil surface. After 48 h of flooding, the water was allowed to drain and the soil to dry for 18 days without further irrigation (phase 4).

2.4 Sampling and analyses

During the entire experiment, water content and redox potential were automatically logged every 5 minutes (EM5b, Decagon, USA and CR1000, Campbell scientific, USA, respectively).

At selected time points during the experiment, soil-emitted gas and soil solution were sampled. For N_2O flux measurements, 20, 40 and 60 minutes after closing the mesocosms, headspace gas samples (20 mL) were collected using a syringe and transferred to pre-evacuated exetainers. The samples were analyzed for their N_2O concentration using a gas chromatograph (Agilent 6890, Santa Clara, USA; Porapak Q column, Ar/CH_4 carrier gas, micro-ECD detector). Measured headspace N_2O concentrations were converted to moles using the ideal gas law and headspace volume. The N_2O efflux rates were calculated as the slope of the linear regression of the N_2O amounts at the three sampling times, relative to the exposed soil surface area (Fig. 1, Shrestha et al., 2012).

For soil water sampling, 20 mL of soil solution were collected using the suction cups. Water samples were analyzed for dissolved organic carbon (DOC) and TN concentrations with an elemental analyzer (Formacs$^{HT/TN}$, Skalar, The Netherlands). Nitrate and ammonium concentrations were measured by ion chromatography (IC 940, Metrohm, Switzerland), and nitrite concentrations were determined photometrically (DR 3900, Hach Lange, Germany).

2.5 Data analyses

Differences among the six treatments on individual sampling dates in N_2O fluxes, DOC and N-species concentrations in soil solution were tested for significance using the non-parametric Kruskal–Wallis test followed by Dunn's post hoc test. To estimate the total amount of N_2O emitted during the period of enhanced N_2O fluxes in phase 4, Q_{tot}, the N_2O fluxes between day 11 and 25 of the experiment were integrated as follows:

$$Q_{tot} = \frac{1}{2}\sum_{n=1}^{n_{max}}[\Delta_n \times (q_n + q_{n+1})], \quad (1)$$

where Δ_n is the time period between the n^{th} and the $n+1^{th}$ measurement, and q_n and q_{n+1} the mean flux on the n^{th} and $n+1^{th}$ measurement day, respectively. "n=1" refers to day 11, and n_{max} to day 25 of phase 4. The integrated N_2O flux data were tested for differences between treatments and model soils by performing a two way ANOVA and the Tukey's honestly significant difference (HSD) post hoc test. No data transformation was necessary, since the inspection of residuals of the ANOVA model and the application of the Shapiro–Wilk normality test revealed

that the values follow a Gaussian distribution. Significance and confidence levels were set at $\alpha < 0.05$. For the statistical analyses we used Graphpad Prism (GraphPad Software Inc., 2017) and R (R Core Team, 2018).

3. Results

3.1 Soil moisture and redox potential

During phase 1 and 2, saturation levels stabilized at 53.0 ± 2.1% WFPS (water filled pore space) in the treatments with LA soils, and were slightly higher in SA treatments (57.8 ± 2.0%) (Fig. 2). The flooding of the mesocosms for 48 h with artificial river water raised the WFPS for all LA soils to 87.8 ± 0.1%, significantly exceeding the increase of WFPS in SA soils (80.6 ± 0.1%). The water release from the system after the simulated flood resulted in an immediate drop of the WFPS, except for the LAU treatment (Fig. 2). This was followed by slow drying for 1 week, and a more marked decrease in WFPS during the second week after the flood. During the latter period, the plant treatments dried faster than the other treatments. As a result, at the end of the experiment, WFPS was still above pre-flood values in unamended and litter treatments, while WFPS levels in the treatments with plants were lower than before the flooding.

The time course of the redox potential measured in 5 cm and 20 cm depth exhibited distinct patterns depending on the respective model soil (Fig. 3). In all treatments, flooding induced a rapid decrease of the redox potential to values below 250 mV within 36 hours. Upon water release, the redox potential returned rapidly to pre-flood values at both measurement depths only in SA soils. In the LA treatments (most pronounced in LAL), soils at 20 cm depth underwent a prolonged phase of continued reduced redox condition, returning to the initial redox levels only towards the end of the experiment.

3.2 Hydrochemistry of soil solutions

Considering individual treatments, DOC concentrations varied only little with time. Yet, the DOC concentrations were generally much higher in treatments with LA than with SA soils. Nitrate was the most abundant dissolved reactive N species in the soil solution, with pre-flood concentrations of 1 to 5 mM (Fig. 4d–f). In the unamended and plant treatments, NO_3^- concentrations were markedly higher in SA than in LA soils, whereas they were similar in both litter addition treatments. Two distinct temporal patterns in the evolution of NO_3^- concentration could be discerned. In the unamended and litter-addition treatments, NO_3^- concentrations decreased after the flooding, consistently reaching a minimum on day 19, in the case of the litter treatments below the detection limit of 0.2 µM, before increasing again during the later drying phase (Fig 4d,e). In contrast, in the treatments with plants, NO_3^- concentrations steadily declined from concentrations of 1–2 mM to around 0.5 mM at the end of the experiment (Fig. 4f). Nitrite was found at significant concentrations only in LA soils, with highest concentrations in the LAU treatment right after the flooding (33.6 µM) and decreasing concentrations throughout the remainder of the experiment (Fig. 4g–i). In SA soils NO_2^- concentration was always < 5 µM, without much variation. Similarly, in most treatments except SAL, ammonium (NH_4^+) concentrations were < 10 µM, and particularly towards the end of the experiment very close to the detection limit (Fig. 4j, 4l). In the SAL treatment, NH_4^+ concentrations peaked 5 d after the flood with concentrations of around 70 µM (Fig. 4k).

3.3 Nitrous oxide emissions

During phase 2 (i.e., before the flooding), N_2O fluxes were generally low (< 1 µmol m^{-2} h^{-1}; Fig. 2), however, fluxes in the LAL treatment were significantly higher than in the other treatments (adj. P = 0.002–0.039; Fig. 2). The flooding triggered the onset of a "hot moment", defined here as period with strongly increased N_2O emissions, which lasted for about one week independent of the treatment (Fig. 2). The maximum efflux was observed immediately after the flood. The subsequent decline in N_2O emission rates followed different patterns among the various treatments. Normalizing the N_2O flux to the maximum measured efflux for each replicated treatment revealed a slower decrease with time for the unamended soils than for the litter and plant treatments (not shown). The strongest peak emissions were observed in the LAL treatment (91.6 ± 14.0 µmol m^{-2} h^{-1}; mean ± SD). Throughout most of the drying phase, the LAU and LAL treatments exhibited higher N_2O emissions than the corresponding SAU and SAL experiments. In contrast, there was no such difference in the treatments with plant cuttings, and peak N_2O emissions were overall lower than in the other treatments. The integrated N_2O fluxes during the hot moment (days 10 to 25 of the experiment) were significantly higher for the LAU and LAL than for all other treatments (Fig. 5). Again, there was a significant aggregate size effect in the unamended (adj. P = 0.045) and litter-addition treatments (adj. P = 0.008). The integrated N_2O emissions in the two plant treatments did not differ significantly from each other, but were significantly smaller than in the LAU (adj. P = 0.001), and the LAL (adj. P = 0.005) treatments.

4. Discussion

In our experiment, we could confirm the occurrence of periods of enhanced N_2O emissions in the drying phase shortly after flooding, as expected based on previous research (Baldwin and Mitchell, 2000; Groffman and Tiedje, 1988; Rabot et al., 2014; Shrestha et al., 2012). We observed that the six treatments had a substantial effect on the magnitude and temporal pattern of N_2O emissions that could only be captured by observations at relatively high temporal resolution. The fast occurrence of strong N_2O fluxes over a comparatively short period in the litter-amended treatment on the one side, and the relatively weak response to the flooding in the plant treatment on the other, suggests complex interactive mechanisms related to distinct microhabitat effects leading to characteristic periods of enhanced N_2O emission. Rabot et al. (2014) explained N_2O emission peaks during the desaturation phase with the release of previously produced and entrapped N_2O. Such a mechanism may partly contribute to high N_2O emissions in our experiment initially, but the continuing depletion of NO_3^- and NO_2^- during the phase of high N_2O emissions indicates that the flooding and drying has strong effects on N transformations mediated by microorganisms in the soil (e.g., the balance and overall rates of nitrification, nitrifier–denitrification, and denitrification). Hence, physical controls alone clearly do not explain the observed timing and extent of hot moments with regard to N_2O emission. In the following sections we will discuss how the effect of flooding on microbial N_2O production is modulated by differential microhabitat formation (and hence redox conditions) in the various treatments.

4.1 Effect of aggregate size on N_2O emissions

Our results indicate that aggregate size is a major factor in modulating soil N_2O emissions. In the unamended and litter addition treatments, LA model soils exhibited both higher peak and total N_2O emissions during the hot moment in the drying phase than SA model soils (Figs. 2 and 5). By contrast, in the presence of a growing willow, there was no detectable effect of aggregate size on the overall N_2O emission (further discussion below).

The aggregate size effects observed in the unamended and litter treatments can be explained by factors controlling (i) gas diffusion (e.g. water film distribution, tortuosity of the intra-aggregate pore space) and (ii) decomposition of encapsulated SOM regulating the extent of N_2O formation (Neira et al., 2015). In order to isolate the effect of aggregate size (i.e., to minimize the effect of other factors that are likely to influence gas diffusion), we created model soils of similar soil structure (see Materials and Methods). The results of the particle size analysis confirmed a nearly identical texture of the two model soils, and at least in the unamended treatments a similar bulk density was achieved. The effect of soil texture and structure should therefore be similar for both model soils. The same applies to bulk soil chemical properties of the two aggregate size fractions such as C_{org} content and pH. Therefore, we assume in the following that the differences in N_2O emissions among the treatments can mainly be attributed to size-related aggregate properties and their interactions with litter addition or rhizosphere effects.

During phase 3 with near-saturated conditions, no aggregate size effect was observed. High WFPS seem to have limited the gas diffusion (O_2 and N_2O) independent of the aggregate size, limiting soil–atmosphere gas exchange in both model soils equally (Neira et al., 2015; Thorbjørn et al., 2008). As a consequence of inhibited gas exchange/soil aeration, a sharp drop in the redox potential was observed in all treatments, indicating a rapid decline in O_2 availability to suboxic/anoxic conditions. Together with an incipient decrease in soil solution NO_3^-, this indicates that N_2O production is primarily driven by denitrification in this phase.

The aggregate size effects on the formation of moments of enhanced N_2O emission became evident during the subsequent drying period. During the initial drying phase, when a heterogeneous distribution of water films around soil particles/aggregates develops (Young and Ritz, 2000), the macroaggregates in the LA model soils appear to foster micro-environmental conditions that are more beneficial to N_2O production. This could be related to the longer diffusive distances for re-entering O_2 caused by the higher tortuosity of the intra-aggregate pore space of macroaggregates, as reported by Ebrahimi and Or (2016). This may have helped to maintain, or even extend, reducing conditions due to microbial activity inside the core of macroaggregates during drying. Thus, on the one hand, large aggregates favor the emergence of anoxic microhabitats expanding the zones where denitrification occurs. On the other hand, the overall higher porosity of the LA soils supports a better aeration in drained parts of the soil (Sey et al., 2008), and aerobic processes (e.g., nitrification) are supported. As a result, ideal conditions for spatially coupled nitrification–denitrification are created (Baldwin and Mitchell, 2000; Koschorreck and Darwich, 1998). Indeed, the emergence of heterogeneously distributed, spatially confined oxygen minimum zones during soil drying may be reflected by the high variability of the redox conditions observed in replicate mesocosms and, on average, the tendency towards lower redox potentials for a prolonged period of time in the subsoils of the LA model soils (Fig. 3 d–f). In this context, the relevance of water films for the emergence of periods of enhanced N_2O emissions is further highlighted by the fact that elevated flux rates were only observed as long as the WFPS was above 65%. This is consistent with work by Rabot et al. (2014) and Balaine et al. (2013), who found similar soil water saturation thresholds for elevated N_2O emissions from soils, attributing this phenomenon to suboptimal environmental conditions for both nitrification and denitrification at lower saturation levels.

Given the arguments above, we assume that N_2O emissions during the drying phase originate to a large degree from heterotrophic denitrification, and that they are governed mainly by the aggregate size dependent redox conditions within the semi-saturated soils. This conclusion stands in good agreement with findings from Drury et al. (2004) who found higher production of N_2O due to enhanced denitrification with increasing size of intact

arable soil aggregates in a laboratory incubation study. In contrast, the much lower emissions from the SA treatments can best be explained by a rapid return to pre-flood, i.e. oxic, conditions in most of the pore space, under which N_2O production driven by denitrification is inhibited. According to Manucharova et al. (2001) and Renault and Stengel (1994), aggregates smaller than 200 µm are simply not large (and reactive) enough (i.e., molecular diffusive distances for oxygen are too short) to develop suboxic or anoxic conditions in the center, let alone denitrifying zones. Hence, only a relatively small fraction of the total number of microaggregates in the SA soils would have been large enough (between 200 and 250 µm) to host denitrification and act as site of anaerobic N_2O production.

4.2 Litter effect on N_2O emissions

We expected that litter addition would increase N_2O emissions from model soils with both small and large aggregates, as was found earlier (e.g. Loecke and Robertson, 2009; Parkin, 1987). The addition of litter to the model soils changed the temporal dynamics of the N_2O emission substantially, but its effect on the net integrated N_2O emission was rather minor (Fig. 5). More precisely, highest peak emission rates of all treatments were observed in the LAL treatment, but peak emission rates were followed by a faster return to low pre-flood emission rates in the LAL and the SAL treatments relative to the unamended treatments (Fig 2). This confirms that surplus organic carbon can, on short-term, boost N_2O emissions, particularly in the large-aggregate treatment. The fast mid-term return to low N_2O emission suggests that N_2O production by heterotrophic denitrification either becomes limited by substrates other than carbon, and/or that the carbon added to the soils affects the redox-biogeochemistry in a way that shifts the balance between N_2O production and consumption in favor of consumption. Loecke and Robertson (2009) reported similar temporal N_2O emission patterns in field experiments with litter-amended soil, and attributed the observed dynamic of a rapid decline after peak emission to an increased demand for terminal electron acceptors during denitrification shortly after the carbon addition. Nitrate/nitrite limitation leads, under stable anoxic conditions, ultimately to the complete reduction of produced N_2O to N_2 decreasing net N_2O emission. Indeed, the rapid decrease in N_2O emissions after the emission rate peak in the litter addition treatments was accompanied by the complete depletion of NO_3^- in the soil solution at low redox potential, suggesting nitrate limitation. The increased demand for electron acceptors can be attributed to the increased availability of labile C compounds and nutrients provided by the mineralization of litter, and the concomitant stimulation of aggregate-associated microbial communities during the flooding (Li et al., 2016). At the same time, the litter-stimulated soil respiration increases the soil's oxygen demand, maintaining stable low redox conditions for a longer period of time during the drying phase. Since high activity of N_2O reductase requires very low O_2 concentrations (Morley et al., 2008), such conditions may be particularly favorable for complete denitrification to N_2, an additional, or alternative, explanation for the low N_2O emission rates shortly after the N_2O emission peak.

4.3 Effects of *Salix viminalis*

Planted willow cuttings resulted in relatively low maximum N_2O emission rates (LAP: 19.75 ± 9.31 µmol m^{-2} h^{-1}; SAP: 15.07 ± 12.07 µmol m^{-2} h^{-1}; mean ± SD), independent of aggregate size. The high values for WFPS throughout the hot moment, and a low redox potential in the subsoil, imply optimal conditions for denitrification or nitrifier denitrification, but compared to unamended and litter-addition treatments, only little N_2O was emitted

(both during peak N_2O emission rates and with regards to the integrated N_2O flux). *S. viminalis* suppressed peak N_2O emissions, overriding the positive effect of large aggregates on N_2O emissions observed otherwise. The specific mechanisms involved are uncertain. Fender et al. (2013) found in laboratory experiments with soil from a temperate broad-leaved forest planted with ash saplings N_2O fluxes and plant effects very similar to the ones observed in our study. They attributed reduced N_2O emissions in presence of ash partly to plant uptake of nutrients that reduced NO_3^- availability to denitrifiers. Fast-growing plant species like *Salix* are particularly effective in removing soil inorganic N (Kowalik and Randerson, 1994). Such a causal link between reduced N_2O emissions and plant growth is, however, not supported by our data. More precisely, the NO_3^- concentrations during the hot moment of N_2O emissions were always relatively high (> 0.5 mM) and above the levels observed in the litter treatments.

An alternative explanation for the reduced N_2O emissions in the plant treatments could be rhizosphere aeration by aerenchyma, a physiological trait of *Salix viminalis* roots, which prevents the formation of anoxia in their close vicinity (Blom et al., 1990; Randerson et al., 2011). Thus, while aerenchyma in general aid in the gas exchange between the soil and the atmosphere, and would per se accelerate transport of N_2O from soils to the atmosphere, they also inhibit anaerobic N_2O production by aerating the rhizosphere. Indeed, redox potentials in the topsoil were higher in SAP and LAP compared to the other treatments. By contrast, the redox potential in the saturated subsoil below was even lower than observed for the unamended soils. This indicates that the aeration effect by aerenchyma is constrained to the upper soil, or is, in the deeper soil portions, compensated by respiratory rhizosphere processes. According to Fender et al. (2013), in vegetated soils, microbial respiration is stimulated by deposition of root exudates, which in concert with root respiration in a highly saturated pore space, leads to severe and ongoing oxygen depletion. Again, N_2 and not N_2O is the dominant final product of denitrification,under the stable anoxic condition produced this way, and N_2O emissions will be low.

5. Conclusions

In this study, we investigated the distinct effects of aggregate size, surplus organic carbon from litter and vegetation on N_2O emission from model soils after flooding. .Flooding and drying were always associated with hot moments of N_2O production, most likely due to heterotrophic denitrification as result of suboxic O_2 levels at high WFPS. Our results demonstrate that aggregate size is a very important factor in modulating N_2O emission from soils under changing pore space water saturation. Aggregates of a diameter > 250 µm appear to foster suboxic microhabitats that favor denitrification and associated N_2O emission. This soil aggregate size effect may be amplified in the presence of excess carbon substrate, as long as heterotrophic denitrification as the main N_2O producing process is not electron acceptor limited, and extremely reducing conditions in organic rich soils do not promote complete denitrification leading to a further reduction of N_2O to N_2. On the other hand, the higher porosity of the soils with macroaggregates may aid in the formation of microsites at the surface of aggregates where nitrification is re-initialized during drying, supporting favorable conditions for spatially coupled nitrification–denitrification. The mechanisms by which processes in the rhizosphere of *Salix viminalis* effectively suppress N_2O emissions, and thus mask any aggregate size effect, remain ambiguous. Distinct physiological features of *Salix viminalis*, its root metabolism, in combination with microbial respiration can lead to the simultaneous aeration of some parts of the rhizosphere, and the formation of strongly reducing zones in others. In both cases, redox conditions seem to be impedimental for extensive net N_2O production.

Our results demonstrate the importance and complexity of the interplay between soil aggregate size, labile organic C availability, respiratory processes in the rhizosphere, and plant-induced aeration of soils under changing soil water content. Those interactions emerged as modulators of N_2O emissions by controlling the O_2 distribution in the soil matrix. Indeed, O_2 appears as the unifying master variable that ultimately sets the boundary conditions for N_2O production and/or consumption.

The main scope of this work was to expand our knowledge on the controls on net N_2O emissions from floodplain soils. The systematic relationships observed in this study are likely to help anticipating where and when hotspots and hot moments of N_2O emissions are most likely to occur in hydrologically dynamic soil systems like floodplain soils. Further understanding of the complex interaction between plants and soil microorganisms, the detritusphere, and soil aggregation, as well as their influence on N turnover and N_2O accumulation in soils, should focus on how the parameters tested affect the actual activity of the nitrifying and denitrifying communities, with an in-depth investigation into the biogeochemical pathways involved.

Data availability. Data will be openly available at https://datadryad.org/

Competing interests. The authors declare that they have no conflict of interest.

Authors contributions. The initial concept of the experiment was developed by JL, MFL and PAN. ML planned the experiment in detail, set it up and performed it. PAN supervised the measurement of N_2O gas concentrations, whereas ML conducted all other measurements and data analyses. ML wrote the manuscript with major contributions by JL, MFL and PAN.

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References

Baggs, E. M.: A review of stable isotope techniques for N2O source partitioning in soils: recent progress, remaining challenges and future considerations, Rapid Commun. Mass Spectrom., 22(11), 1664–1672, doi:10.1002/rcm.3456, 2008.

Baggs, E. M.: Soil microbial sources of nitrous oxide: Recent advances in knowledge, emerging challenges and future direction, Curr. Opin. Environ. Sustain., 3(5), 321–327, doi:10.1016/j.cosust.2011.08.011, 2011.

Balaine, N., Clough, T. J., Beare, M. H., Thomas, S. M., Meenken, E. D. and Ross, J. G.: Changes in Relative Gas Diffusivity Explain Soil Nitrous Oxide Flux Dynamics, Soil Sci. Soc. Am. J., 77(5), 1496–1505, doi:DOI

10.2136/sssaj2013.04.0141, 2013.

Baldwin, D. S. and Mitchell, A. M.: The effects of drying and re- flooding on the sediment and soil nutrient dynamics of lowland river–floodplain systems: a synthesis, Regul. Rivers Res. Manag., 16(5), 457–467, doi:10.1002/1099-1646(200009/10)16:5<457::AID-RRR597>3.3.CO;2-2, 2000.

Ball, B. C.: Soil structure and greenhouse gas emissions: A synthesis of 20 years of experimentation, Eur. J. Soil Sci., 64(3), 357–373, doi:10.1111/ejss.12013, 2013.

Bateman, E. J. and Baggs, E. M.: Contributions of nitrification and denitrification to N2O emissions from soils at different water-filled pore space, Biol. Fertil. Soils, 41(6), 379–388, doi:10.1007/s00374-005-0858-3, 2005.

Beare, M. H., Gregorich, E. G. and St-Georges, P.: Compaction effects on CO2 and N2O production during drying and rewetting of soil, Soil Biol. Biochem., 41(3), 611–621, doi:10.1016/j.soilbio.2008.12.024, 2009.

Bender, S. F., Plantenga, F., Neftel, A., Jocher, M., Oberholzer, H.-R., Köhl, L., Giles, M., Daniell, T. J. and van der Heijden, M. G.: Symbiotic relationships between soil fungi and plants reduce N2O emissions from soil, ISME J., 8(6), 1336–1345, doi:10.1038/ismej.2013.224, 2014.

Blom, C. W. P. M., Bögemann, G. M., Laan, P., van der Sman, A. J. M., van de Steeg, H. M. and Voesenek, L. A. C. J.: Adaptations to flooding in plants from river areas, Aquat. Bot., 38(1), 29–47, doi:10.1016/0304-3770(90)90097-5, 1990.

Blum, J. M., Su, Q., Ma, Y., Valverde-Pérez, B., Domingo-Félez, C., Jensen, M. M. and Smets, B. F.: The pH dependency of N-converting enzymatic processes, pathways and microbes: Effect on net N2O production, Environ. Microbiol., doi:10.1111/1462-2920.14063, 2018.

Böttcher, J., Weymann, D., Well, R., Von Der Heide, C., Schwen, A., Flessa, H. and Duijnisveld, W. H. M.: Emission of groundwater-derived nitrous oxide into the atmosphere: model simulations based on a 15N field experiment, Eur. J. Soil Sci., 62(2), 216–225, doi:10.1111/j.1365-2389.2010.01311.x, 2011.

Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R. and Zechmeister-Boltenstern, S.: Nitrous oxide emissions from soils: how well do we understand the processes and their controls?, Philos. Trans. R. Soc. Lond. B. Biol. Sci., 368(1621), 20130122, doi:10.1098/rstb.2013.0122, 2013.

Ciais, P., Sabine, C., Bala, G., Bopp, L., Brovkin, V., Canadell, J., Chhabra, A., DeFries, R., Galloway, J., Heimann, M., Jones, C., Quéré, C. Le, Myneni, R. B., Piao, S. and Thornton, P.: Carbon and Other Biogeochemical Cycles, in Climate Change 2013 - The Physical Science Basis, edited by Intergovernmental Panel on Climate Change, pp. 465–570, Cambridge University Press, Cambridge., 2013.

Diba, F., Shimizu, M. and Hatano, R.: Effects of soil aggregate size, moisture content and fertilizer management on nitrous oxide production in a volcanic ash soil, Soil Sci. Plant Nutr., 57(5), 733–747, doi:10.1080/00380768.2011.604767, 2011.

Drury, C. ., Yang, X. ., Reynolds, W. . and Tan, C. .: Influence of crop rotation and aggregate size on carbon dioxide production and denitrification, Soil Tillage Res., 79(1), 87–100, doi:10.1016/j.still.2004.03.020, 2004.

Ebrahimi, A. and Or, D.: Microbial community dynamics in soil aggregates shape biogeochemical gas fluxes from soil profiles – upscaling an aggregate biophysical model, Glob. Chang. Biol., 22(9), 3141–3156, doi:10.1111/gcb.13345, 2016.

Elliott, A. E. T. and Coleman, D. C.: Oikos Editorial Office Let the Soil Work for Us Proceedings of the 4th European Ecology Symposium 7-12 September 1986 , Wageningen Published by : Oikos Editorial Office Stable URL : http://www.jstor.org/stable/20112982 Let the soil work for us, Ecol. Bull., (39), 22–32, 1988.

Fender, A.-C., Leuschner, C., Schützenmeister, K., Gansert, D. and Jungkunst, H. F.: Rhizosphere effects of tree

species – Large reduction of N2O emission by saplings of ash, but not of beech, in temperate forest soil, Eur. J. Soil Biol., 54, 7–15, doi:10.1016/j.ejsobi.2012.10.010, 2013.

Forster, P., V. Ramaswamy, P. Artaxo, T. Berntsen, R. Betts, D.W. Fahey, J. Haywood, J. Lean, D.C. Lowe, G. Myhre, J. Nganga, R. Prinn, G. Raga, M. S. and R. V. D.: Changes in Atmospheric Constituents and in Radiative Forcing., 2007.

Frame, C. H., Lau, E., Joseph Nolan, E., Goepfert, T. J. and Lehmann, M. F.: Acidification enhances hybrid N2O production associated with aquatic ammonia-oxidizing microorganisms, Front. Microbiol., 7(JAN), 1–23, doi:10.3389/fmicb.2016.02104, 2017.

Gee, G. W. and Bauder, J. W.: Particle-size Analysis, in Physical and Mineralogical Methods-Agronomy Monograph no. 9, edited by A. Klute, pp. 383–411, American Society of Agronomy-Soil Science Society of America, Madison, WI., 1986.

Goldberg, S. D., Knorr, K. H., Blodau, C., Lischeid, G. and Gebauer, G.: Impact of altering the water table height of an acidic fen on N2O and NO fluxes and soil concentrations, Glob. Chang. Biol., 16(1), 220–233, doi:10.1111/j.1365-2486.2009.02015.x, 2010.

GraphPad Software Inc.: GraphPad Prism 7.04, La Jolla, CA, www.graphpad.com, 2017.

Groffman, P. M. and Tiedje, J. M.: Denitrification Hysteresis During Wetting and Drying Cycles in Soil, Soil Sci. Soc. Am. J., 52(6), 1626, doi:10.2136/sssaj1988.03615995005200060022x, 1988.

Hartmann, D. J., Klein Tank, A. M. G., Rusticucci, M., Alexander, L. V, Brönnimann, S., Charabi, Y. A.-R., Dentener, F. J., Dlugokencky, E. J., Easterling, D. R., Kaplan, A., Soden, B. J., Thorne, P. W., Wild, M. and Zhai, P.: Observations: Atmosphere and Surface, in Climate Change 2013 - The Physical Science Basis, edited by Intergovernmental Panel on Climate Change, pp. 159–254, Cambridge University Press, Cambridge., 2013.

Hefting, M., Clément, J.-C., Dowrick, D., Cosandey, A. C., Bernal, S., Cimpian, C., Tatur, A., Burt, T. P. and Pinay, G.: Water table elevation controls on soil nitrogen cycling in riparian wetlands along a European climatic gradient, Biogeochemistry, 67(1), 113–134, doi:10.1023/B:BIOG.0000015320.69868.33, 2004.

Heincke, M. and Kaupenjohann, M.: Effects of soil solution on the dynamics of N2O emissions: a review, Nutr. Cycl. Agroecosystems, 55(2), 133–157, doi:10.1023/A:1009842011599, 1999.

Hill, A. R.: Buried organic-rich horizons: their role as nitrogen sources in stream riparian zones, Biogeochemistry, 104(1–3), 347–363, doi:10.1007/s10533-010-9507-5, 2011.

Hu, H.-W., Macdonald, C. A., Trivedi, P., Holmes, B., Bodrossy, L., He, J.-Z. and Singh, B. K.: Water addition regulates the metabolic activity of ammonia oxidizers responding to environmental perturbations in dry subhumid ecosystems, Environ. Microbiol., 17(2), 444–461, doi:10.1111/1462-2920.12481, 2015.

Jahangir, M. M. R., Roobroeck, D., Van Cleemput, O. and Boeckx, P.: Spatial variability and biophysicochemical controls on N2O emissions from differently tilled arable soils, Biol. Fertil. Soils, 47(7), 753–766, doi:10.1007/s00374-011-0580-2, 2011.

Jørgensen, C. J., Struwe, S. and Elberling, B.: Temporal trends in N2O flux dynamics in a Danish wetland - effects of plant-mediated gas transport of N2O and O2 following changes in water level and soil mineral-N availability, Glob. Chang. Biol., 18(1), 210–222, doi:10.1111/j.1365-2486.2011.02485.x, 2012.

Khalil, K., Renault, P. and Mary, B.: Effects of transient anaerobic conditions in the presence of acetylene on subsequent aerobic respiration and N2O emission by soil aggregates, Soil Biol. Biochem., 37(7), 1333–1342, doi:10.1016/j.soilbio.2004.11.029, 2005.

Koschorreck, M. and Darwich, A.: Nitrogen dynamics in sesonally flooded soils in the Amazon floodplain, Wetl.

Ecol. Manag., 11, 317–330, 1998.

Kowalik, P. J. and Randerson, P. F.: Nitrogen and phosphorus removal by willow stands irrigated with municipal waste water?A review of the Polish experience, Biomass and Bioenergy, 6(1–2), 133–139, doi:10.1016/0961-9534(94)90092-2, 1994.

Kuzyakov, Y. and Blagodatskaya, E.: Microbial hotspots and hot moments in soil: Concept & review, Soil Biol. Biochem., 83, 184–199, doi:10.1016/j.soilbio.2015.01.025, 2015.

Li, X., Sørensen, P., Olesen, J. E. and Petersen, S. O.: Evidence for denitrification as main source of N 2 O emission from residue-amended soil, Soil Biol. Biochem., 92, 153–160, doi:10.1016/j.soilbio.2015.10.008, 2016.

Loecke, T. D. and Robertson, G. P.: Soil resource heterogeneity in terms of litter aggregation promotes nitrous oxide fluxes and slows decomposition, Soil Biol. Biochem., 41(2), 228–235, doi:10.1016/j.soilbio.2008.10.017, 2009.

Luster, J., Göttlein, A., Nowack, B. and Sarret, G.: Sampling, defining, characterising and modeling the rhizosphere—the soil science tool box, Plant Soil, 321(1–2), 457–482, doi:10.1007/s11104-008-9781-3, 2009.

Manucharova, N. A., Stepanov, A. L. and Umarov, M. M.: Microbial transformation of nitrogen in water-stable aggregates of various soil types, EURASIAN SOIL Sci., 34(10), 1125–1131, 2001.

Morley, N., Baggs, E. M., Dörsch, P. and Bakken, L.: Production of NO, N2O and N2 by extracted soil bacteria, regulation by NO2- and O2 concentrations, FEMS Microbiol. Ecol., 65(1), 102–112, doi:10.1111/j.1574-6941.2008.00495.x, 2008.

Myrold, D. D., Pett-Ridge, J. and Bottomley, P. J.: Nitrogen Mineralization and Assimilation at Millimeter Scales, in Methods in Enzymology, vol. 496, pp. 91–114, Elsevier Inc., 2011.

Neira, J., Ortiz, M., Morales, L. and Acevedo, E.: Oxygen diffusion in soils: Understanding the factors and processes needed for modeling, Chil. J. Agric. Res., 75(August), 35–44, doi:10.4067/S0718-58392015000300005, 2015.

Oades, J. M.: Soil organic matter and structural stability: mechanisms and implications for management, Plant Soil, 76(1–3), 319–337, doi:10.1007/BF02205590, 1984.

Parkin, T. B.: Soil Microsites as a Source of Denitrification Variability, Soil Sci. Soc. Am. J., 51, 1194–1199, 1987.

Philippot, L., Hallin, S., Börjesson, G. and Baggs, E. M.: Biochemical cycling in the rhizosphere having an impact on global change, Plant Soil, 321(1–2), 61–81, doi:10.1007/s11104-008-9796-9, 2009.

R Core Team: R: A Language and Environment for Statistical Computing, R Found. Stat. Comput., Vienna, https://www.R-project.org/, 2018.

Rabot, E., Hénault, C. and Cousin, I.: Temporal Variability of Nitrous Oxide Emissionsby Soils as Affected by Hydric History, Soil Sci. Soc. Am. J., 78(2), 434, doi:10.2136/sssaj2013.07.0311, 2014.

Randerson, P. F., Moran, C. and Bialowiec, A.: Oxygen transfer capacity of willow (Salix viminalis L.), Biomass and Bioenergy, 35(5), 2306–2309, doi:10.1016/j.biombioe.2011.02.018, 2011.

Ravishankara, A. R., Daniel, J. S. and Portmann, R. W.: Nitrous Oxide (N2O): The Dominant Ozone-Depleting Substance Emitted in the 21st Century, Science (80-.)., 326(5949), 123–125, doi:10.1126/science.1176985, 2009.

Renault, P. and Stengel, P.: Modeling Oxygen Diffusion in Aggregated Soils: I. Anaerobiosis inside the Aggregates, Soil Sci. Soc. Am. J., 58(4), 1017, doi:10.2136/sssaj1994.03615995005800040004x, 1994.

Robertson, G. P. and Groffman, P. M.: Nitrogen Transformations, in Soil Microbiology, Ecology and

Biochemistry, pp. 421–446, Elsevier., 2015.

Ruser, R., Flessa, H., Russow, R., Schmidt, G., Buegger, F. and Munch, J. C.: Emission of N2O, N2 and CO2 from soil fertilized with nitrate: Effect of compaction, soil moisture and rewetting, Soil Biol. Biochem., 38(2), 263–274, doi:10.1016/j.soilbio.2005.05.005, 2006.

Samaritani, E., Shrestha, J., Fournier, B., Frossard, E., Gillet, F., Guenat, C., Niklaus, P. A., Pasquale, N., Tockner, K., Mitchell, E. A. D. and Luster, J.: Heterogeneity of soil carbon pools and fluxes in a channelized and a restored floodplain section (Thur River, Switzerland), Hydrol. Earth Syst. Sci., 15(6), 1757–1769, doi:10.5194/hess-15-1757-2011, 2011.

Sey, B. K., Manceur, A. M., Whalen, J. K., Gregorich, E. G. and Rochette, P.: Small-scale heterogeneity in carbon dioxide, nitrous oxide and methane production from aggregates of a cultivated sandy-loam soil, Soil Biol. Biochem., 40(9), 2468–2473, doi:10.1016/j.soilbio.2008.05.012, 2008.

Shrestha, J., Niklaus, P. a, Frossard, E., Samaritani, E., Huber, B., Barnard, R. L., Schleppi, P., Tockner, K. and Luster, J.: Soil nitrogen dynamics in a river floodplain mosaic., J. Environ. Qual., 41(6), 2033–45, doi:10.2134/jeq2012.0059, 2012.

Six, J., Bossuyt, H., Degryze, S. and Denef, K.: A history of research on the link between (micro)aggregates, soil biota, and soil organic matter dynamics, Soil Tillage Res., 79(1), 7–31, doi:10.1016/j.still.2004.03.008, 2004.

Spott, O., Russow, R. and Stange, C. F.: Formation of hybrid N2O and hybrid N2 due to codenitrification: First review of a barely considered process of microbially mediated N-nitrosation, Soil Biol. Biochem., 43(10), 1995–2011, doi:10.1016/j.soilbio.2011.06.014, 2011.

Stolk, P. C., Hendriks, R. F. A., Jacobs, C. M. J., Moors, E. J. and Kabat, P.: Modelling the effect of aggregates on N 2O emission from denitrification in an agricultural peat soil, Biogeosciences, 8(9), 2649–2663, doi:10.5194/bg-8-2649-2011, 2011.

Thorbjørn, A., Moldrup, P., Blendstrup, H., Komatsu, T. and Rolston, D. E.: A Gas Diffusivity Model Based on Air-, Solid-, and Water-Phase Resistance in Variably Saturated Soil, Vadose Zo. J., 7(4), 1276, doi:10.2136/vzj2008.0023, 2008.

Tisdall, J. M. and Oades, J. M.: Organic matter and water-stable aggregates in soils, J. Soil Sci., 33(2), 141–163, doi:10.1111/j.1365-2389.1982.tb01755.x, 1982.

Totsche, K. U., Amelung, W., Gerzabek, M. H., Guggenberger, G., Klumpp, E., Knief, C., Lehndorff, E., Mikutta, R., Peth, S., Prechtel, A., Ray, N. and Kögel-Knabner, I.: Microaggregates in soils, J. Plant Nutr. Soil Sci., 1–33, doi:10.1002/jpln.201600451, 2017.

Vieten, B., Conen, F., Neftel, A. and Alewell, C.: Respiration of nitrous oxide in suboxic soil, Eur. J. Soil Sci., 60(3), 332–337, doi:10.1111/j.1365-2389.2009.01125.x, 2009.

Walthert, L., Graf, U., Kammer, A., Luster, J., Pezzotta, D., Zimmermann, S. and Hagedorn, F.: Determination of organic and inorganic carbon, δ 13 C, and nitrogen in soils containing carbonates after acid fumigation with HCl, J. Plant Nutr. Soil Sci., 173(2), 207–216, doi:10.1002/jpln.200900158, 2010.

Young, I. . and Ritz, K.: Tillage, habitat space and function of soil microbes, Soil Tillage Res., 53(3–4), 201–213, doi:10.1016/S0167-1987(99)00106-3, 2000.

Zhu, X., Burger, M., Doane, T. a and Horwath, W. R.: Ammonia oxidation pathways and nitrifier denitrification are significant sources of N2O and NO under low oxygen availability, Pnas, 110(16), 6328–6333, doi:10.1073/pnas.1219993110/-/DCSupplemental.www.pnas.org/cgi/doi/10.1073/pnas.1219993110, 2013.

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622 **Table 1: Physicochemical properties of the two aggregate size fractions (macroaggregates and microaggregates) and**
623 **added leaf litter. C_{org} and N_{tot} of the aggregates was measured in triplicates. The leaf litter was analyzed in four**
624 **replicates. Final pH and texture of model soil 1 and 2 was measured in duplicates (means ± SD)**

		Macroaggregates	Microaggregates	Litter (*Salix v.* L.)
C_{org}	g kg^{-1}	19.22 ± 0.55	21.56 ± 2.39	459.9 ± 2.55
Total N	g kg^{-1}	1.58 ± 0.02	1.35 ± 0.14	27.39 ± 0.15
C:N ratio		12.16 ± 0.22	15.99 ± 0.71	16.79 ± 0.06
		Model soil 1	Model soil 2	
pH ($CaCl_2$)		8 ± 0.02	7.56 ± 0.01	
sand	%	71.25 ± 0.05	70.7 ± 0.50	
silt	%	20 ± 0.30	21.1 ± 0.60	
clay	%	8.75 ± 0.25	8.2 ± 0.10	

625

626 **Table 2: Overview of treatments in the flooding–drying experiment**

	LAU	SAU	LAL	SAL	LAP	SAP
Model Soil 1 (LA)	+	-	+	-	+	-
Model Soil 2 (SA)	-	+	-	+	-	+
Leaf litter (*Salix v.*)	-	-	+	+	-	-
Salix v.	-	-	-	-	+	+

627

Figure Captions

Figure 1: Schematic of a mesocosm with gas sampling valves (1), Ag/AgCl reference electrode (2), Pt redox electrodes (3), suction cups (4), volumetric water content sensors (5), vent (6), and water inlet/outlet (7). The top part is only attached during gas sampling.

Figure 2: Mean N_2O emission during the flooding–drying experiment from large-aggregate model soil (LA; filled circles) and small-aggregate model soil (SA, open circles), and corresponding water-filled pore space (WFPS) in LA (filled triangles) and SA (open triangles). Unamended soils (A), litter addition (B) and plant treatment (C). Flooding phase indicated by the grey area. Symbols indicate means; error bars are SE; n= 6.

Figure 3: Redox potential relative to standard hydrogen electrode during the flooding–drying experiment in 5 cm and 20 cm depth (mean ± SE; n=6). Unamended soils (a and d, respectively), litter addition (b and e, respectively), plant treatment (c and f, respectively). LA (filled circles) and SA (open circles); the dotted line at 250 mV marks the threshold, below which denitrification is expected to occur.

Figure 4: DOC (circles), nitrate (squares), nitrite (diamonds) and ammonium (triangles) concentrations in pore water during the flooding–drying experiment. LA (filled symbols) and SA (empty symbols). Unamended soils (a, d, g and j, respectively), litter addition (b, e, h and k, respectively) and plant treatment (c, f, j and l, respectively).; (mean ± SE; n=6).

Figure 5: Integrated N_2O fluxes over the 14 days period of elevated N_2O emissions in the drying phase of the flooding–drying experiment (mean ± SE; n= 6). Black bars represent model soil 1 (macroaggregates 250-4000µm) whereas model soil 2 (microaggregates < 250µm) is depicted as white bars. Significant differences among the six treatments are denoted by different lower case letters at adj. $P < 0.05$.

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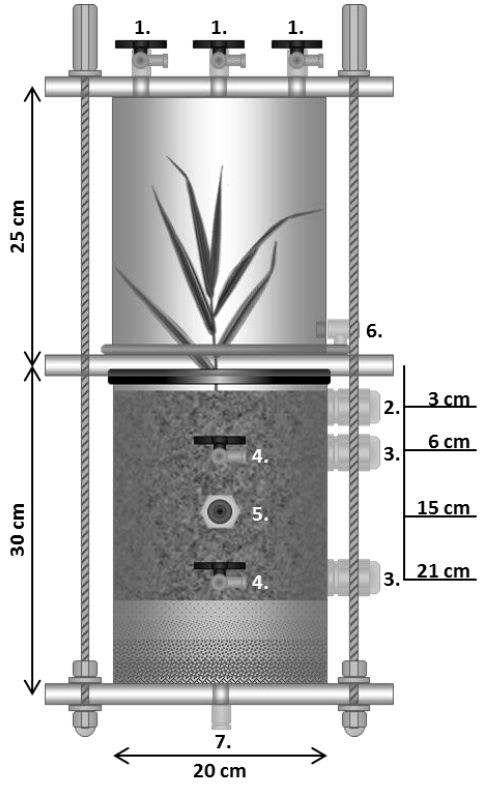

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650 Figure 1

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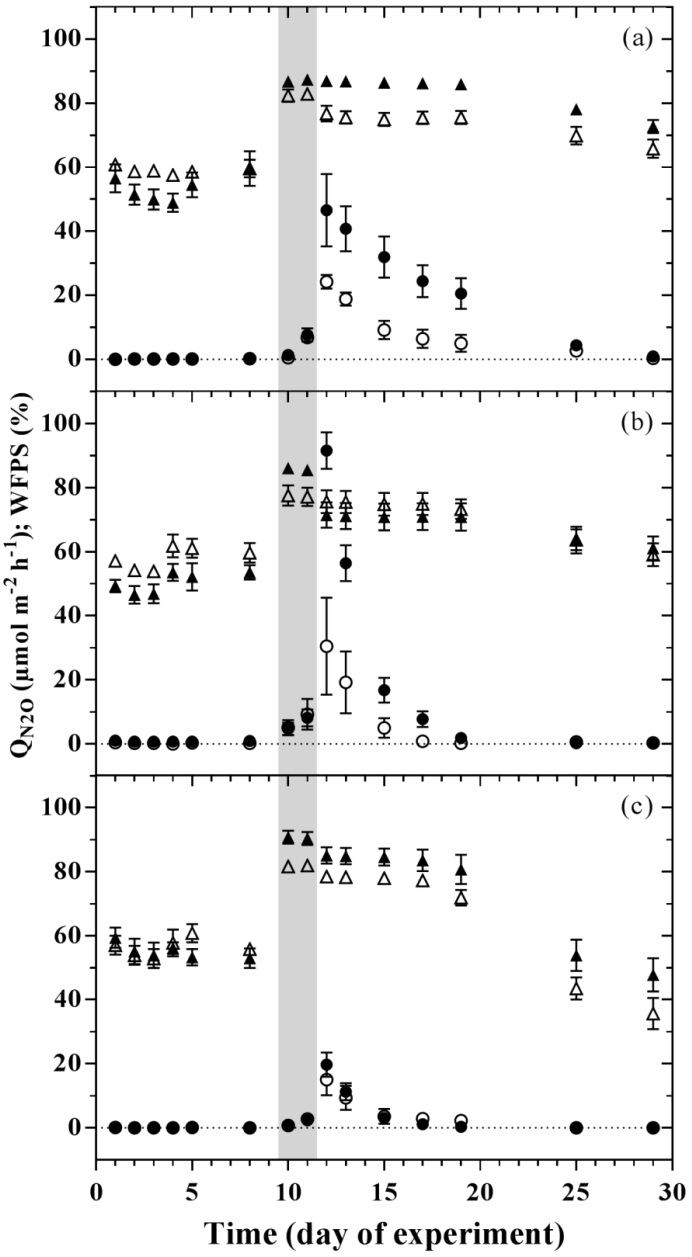

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653 Figure 2

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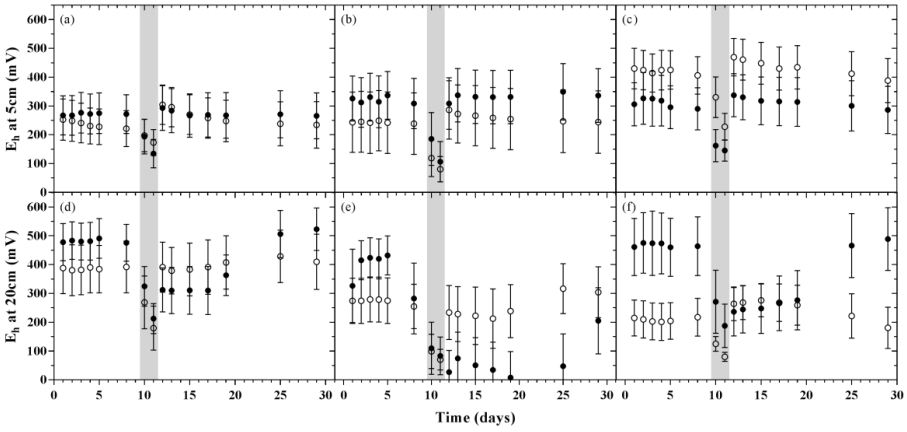

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656 Figure 3

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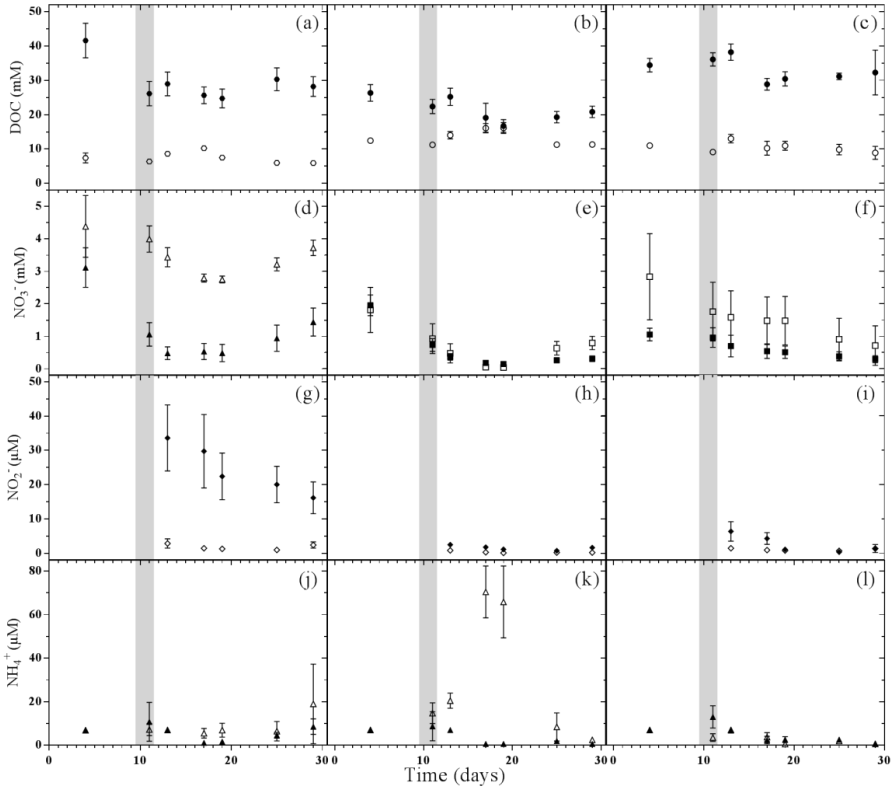

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659 Figure 4

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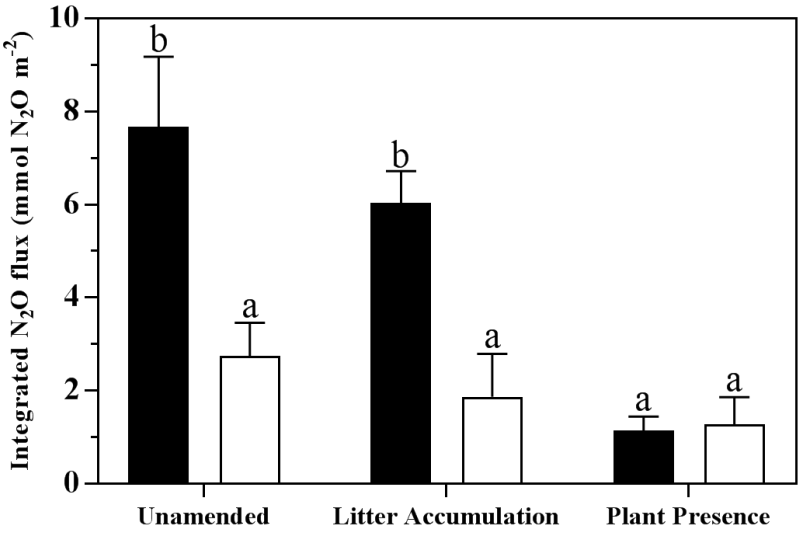

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662 Figure 5