Response to reviewers' comments (bg-2018-290)

Response to interactive comments of Reviewer 1

5 We thank reviewer 1 for helpful comments and corrections. Our responses to specific comments (reprinted in bold) are given below.

The authors tested experimentally whether the slow degradability of boreal forest mosses is caused primarily by the chemically complexity of their tissues or the physical structure of the moss cell wall biochemical matrix
10 **inhibiting decomposition. The authors used various methods to study the decay rate of mosses, and changes in moss tissue C and N composition and physical structure during the 2.5-year laboratory incubation at two different temperatures. The results suggested 1) the moss cell wall matrix protected labile C from microbial decomposition and 2) the N and C cycles were uncoupled. I find the manuscript very interesting and topical in terms of assessing the role of boreal forest soils as sinks and sources of C. Below comments to the aspects**
15 **listed by BG: 1. Does the paper address relevant scientific C1 BGD Interactive comment Printer-friendly version Discussion paper questions within the scope of BG? YES. 2. Does the paper present novel concepts, ideas, tools, or data? YES. 3. Are substantial conclusions reached? YES. 4. Are the scientific methods and assumptions valid and clearly outlined? YES. 5. Are the results sufficient to support the interpretations and conclusions? YES. 6. Is the description of experiments and calculations sufficiently complete and precise to**
20 **allow their reproduction by fellow scientists (traceability of results)? YES. 7. Do the authors give proper credit to related work and clearly indicate their own new/original contribution? YES. 8. Does the title clearly reflect the contents of the paper? YES. 9. Does the abstract provide a concise and complete summary? YES. 10. Is the overall presentation well structured and clear? YES. 11. Is the language fluent and precise? YES. 12. Are mathematical formulae, symbols, abbreviations, and units correctly defined and used? YES. 13.**
25 **Should any parts of the paper (text, formulae, figures, tables) be clarified, reduced, combined, or eliminated? YES, see specific comments. 14. Are the number and quality of references appropriate? I CANNOT ASSES THIS BECAUSE AT LEAST 14 REFERENCES GIVEN IN THE TEXT ARE MISSING FROM THE LIST OF REFERENCES. THE REFERENCES IN TEXT AND IN THE LIST SHOULD ALSO BE CROSS-CHECKED BECAUSE THERE ARE DIFFERENCES IN THE PUBLICATION YEAR OR NAME OF THE**
30 **FIRST AUTHOR IN SOME CASES.**

Discrepancies between the reference list and the references cited in the text have been corrected in the revised manuscript.

35 **15. Is the amount and quality of supplementary material appropriate? YES.**

Page 3, line 13: Tell whether you only sampled green living (fresh?) parts of mosses or was the material a mixture of green and older brown parts.

The collected mosses were separated into green and brown fractions, and the green tissues were used in the incubations. This has been clarified in the revised manuscript. Please see pg. 3 lines 23-29 of the methods section.

Page 9, lines 33-35: Uncoupling of the N and C cycles has also been reported as a result of in situ incubations - see Manninen et al. 2016, Science of the Total Environment 571, 314- 322. Add reference.

This reference has been added in the revised manuscript (Page 10 line 9)

Page 3, lines 18-19 and Table 1: Correct the names of the moss species, i.e. should be Rhytidiadelphus spp., Pleurozium spp. and Ptilium crista-castrensis.

Has been corrected in the revised manuscript

Page 6, line 27: I think the authors should refer to Table 2 (not Table 3).

Changed to Table 2

Page 7, line 15: Replace 'Figure 2' with Fig. 2.

Corrected

Page10, lines 25-33 (and page 11, lines 27-28): Discussion on fungi is very important, given that fungi are important decomposers in acid forest soils. If the authors have data on soil pH at the two sites, it should be added in Table 1.

The soil pH of the two sites is quite low (<4.5 in all cases) and has been added to both the methods section (Page 3, lines 19-20) and Table 1

Table 3: Replace '%Carbon' and '%Nitrogen' with %C and %N, respectively. Replace 'Nitrate' and 'Ammonia' with nitrate+nitrite and ammonium. Use nitrate+nitrite also on page 7, lines 21-22.

Corrected

Fig. 8: Add a, b, c and d to indicate Figs. 8a-8d.

Will be added in the revised manuscript

Cross-check the list of references with references in the text and revise when needed.

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Corrected

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Response to interactive comments of Reviewer 2

We thank reviewer 2 for helpful comments. Our responses to specific comments (reprinted in bold) are given below.

15 **This is an interesting and novel paper that tries to dig into the reasons behind the relatively well documented low decomposition rates of bryophytes that has a huge impact on biogeochemical cycles in the boreal, which as the authors point out, is frequently not taken into account. I think the question that this paper is addressing is important and novel. I have a few concerns about the paper however that in my opinion would have to be addressed before it should be published.**

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1) The bryophyte species were not included as a variable in this test, but they did vary between regions. Bryophyte species, even beyond the true moss/sphagnum split, are far from being homogeneous. I suspect that many of the differences found between the two regions has to do with the different species that were included in the mesocosmes. This point is not addressed anywhere in the text. There is considerable
25 **litterature showing that the nature of the decomposing matter is one if not the most important factor in determining decomposition rate (e.g. Lang et al. 2009 Journal of Ecology). Unfortunately the latin names of almost all the species are mispelled. I feel that including acknowldging this factor and including the associated litterature will strengthen this paper considerably.**

30 We acknowledge that the study prevents the separation of effects of different moss species vs. regional effects due to the differences in moss species between sites. However, the main conclusions of the paper (low decomposition and $Q10$, little change in chemical composition or physical structure) arise from similarities between the two sites. The observation of these similarities despite contrasting climate, moss species, and N availability strengthens these conclusions. This is clarified in the discussion of the revised manuscript.

35

The main difference we observe between the sites was in N dynamics, including changes in %N remaining, C:N, and amino acids. We maintain that these differences most likely arise from higher moss N concentrations at GC than

SR because the changes are consistent with differences in N availability. The differences in moss N concentrations are likely due to site differences in N availability rather than species-specific differences because N concentrations of balsam fir needles follow the same pattern as the moss tissues (Ziegler et al. 2017). We expanded the section on differences in N dynamics between the site to acknowledge the possibility of moss species effects, including additional citations. Please see pg. 9 lines 17-25 and pg. 10 lines 4-16.

The spelling of the Latin names is corrected in the revised manuscript

2) The methods are not clearly enough described. In the annotated manuscript I have highlighted several places where more details are needed to clearly understand the methodology - mostly in the field aspects. The details highlighted in the annotated manuscript will be clarified in the revision. Similarly, I am uncertain about the use of the Philben et al. 2006 approach as "green moss" from a stream is taken as equivalent as a variety of mosses from boreal forests. Can more justification be provided?

Clarification of methodological details, particularly the field collections, are now included as requested. Please see p. 3 lines 20-29 and p.4 lines 6-20.

"Green mosses" in Philben et al. 2016 refer to the green portion of upland boreal forest moss tissues, separated from the underlying brown portion which was reported separately. The mosses in Philben et al. 2016 were collected from the same two forest sites as the present study and the same set of dominant species are represented. This is clarified in the revised manuscript

3) The results could be more clearly presented. I am uncomfortable with a table made up only of p values. It would be much better to have F values and N for the different tests. There also seems to be a contredriction between the table (effect of temperature on mass remaining), the figures (not really) and the texte (there was none).

Table 2 has been revised to include F values and degrees of freedom for each test.

The effect of temperature on mass remaining is significant, as indicated by table 2. The text in the results section (page 6 line 28 – page 7 line 2) also indicates that mass loss and Q10 were significantly higher in the 18°C incubations. Statistics and a reference to Table 2 have been added to these statements for clarity.

Discussion of a small temperature effect is based on low Q10 compared to vascular plant decomposition (page 9, lines 6-16), despite a significant difference in mass loss between temperature treatments.

Also the figures were not always clear as information was lacking from the legends. I do wonder if all of the figures are required, perhaps Fig 6 could be an annexe?

We prefer to keep Fig. 6 in the main text because it clearly illustrates the lack of change in bulk C composition during incubation, which is an important conclusion but not demonstrated in the other figures. A legend has been added to Figure 6 and Figure 8 for clarity.

In conclusion I think this is an interesting paper with a lot of potential. With a little refinement I think it could have a lot of impact.

Biochemical and structural controls on the decomposition dynamics of boreal upland forest moss tissues

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Abstract. Mosses contribute an average of 20% of upland boreal forest net primary productivity and are frequently observed to degrade slowly compared to vascular plants. If this is caused primarily by the chemically complexity of their tissues, moss decomposition could exhibit high temperature sensitivity (measured as Q_{10}) due to high activation energy, which would imply soil organic carbon (SOC) stocks derived from moss remains are especially vulnerable to decomposition with warming. Alternatively, the physical structure of the moss cell wall biochemical matrix could inhibit decomposition, resulting in low decay rates and low temperature sensitivity. We tested these hypotheses by incubating mosses collected from two boreal forests in Newfoundland, Canada, for 959 days at 5 and 18°C, while monitoring changes in the moss tissue composition using total hydrolysable amino acid (THAA) analysis and ^{13}C NMR spectroscopy. Less than 40% of C was respired in all incubations, revealing a large pool of apparently recalcitrant C. The decay rate of the labile fraction increased in the warmer treatment, but the total amount of C loss increased only slightly, resulting in low Q_{10} values (1.23-1.33) compared to L horizon soils collected from the same forests. NMR spectra were dominated by O-alkyl C throughout the experiment, indicating the persistence of potentially labile C. Accumulation of hydroxyproline (derived primarily from plant cell wall proteins) and aromatic C indicates selective preservation of biochemicals associated with the moss cell wall. This was supported by scanning electron microscope (SEM) images of the moss tissues, which revealed few changes in the physical structure of the cell wall after incubation. This suggests the moss cell wall matrix protected labile C from microbial decomposition, accounting for the low temperature sensitivity of moss decomposition despite low decay rates. Climate drivers of moss biomass and productivity, therefore, represent a potentially important regulator of boreal forest SOC responses to climate change that needs to be assessed to improve our understanding of carbon-climate feedbacks.

1 Introduction

Boreal forests account for over half of global forest soil carbon (C) stocks, with areal soil C densities 2-3 times higher than temperate or tropical forests (Malhi et al., 1999). Many factors contribute to high C stocks including low temperatures, high soil moisture in many regions, and the high relative abundance of plants with relatively slow-to-decay litter such as mosses (Coûteaux et al., 2002; Hobbie, 1996; Hobbie, 2013; Hobbie et al., 2000; Wetterstedt et al., 2010). However, the boreal region is warming more rapidly than the global mean (IPCC, 2014), which could lead to C losses both from the direct effects of warming and drying (Kane et al. 2005) and from the indirect effects of changing C sources due to vegetation change (Gornall et al. 2007; Kohl et al. 2018; Turetsky et al. 2012).

~~The low temperatures and, in many regions, high soil moisture likely cause slow rates of decomposition, contributing to the retention of these substantial stores of soil C (Coûteaux et al., 2002; Hobbie, 2013; Hobbie et al., 2000; Wetterstedt et al., 2010). However, these northern areas are expected to be vulnerable to climate change (IPCC, 2014). Warming impacts on carbon cycling and storage within boreal forests are likely due to changes in both C emissions from soils due to decomposition (Kane et al. 2005; Norris et al. 2010) and changes in soil C sources even prior to major vegetation shifts associated with long term climate change (Kohl et al. 2017; Ziegler et al. 2017).~~

Mosses contribute an average of about 20% of the total NPP in upland boreal forests (Turetsky et al., 2010) and can locally exceed vascular plant NPP (Frolking et al. 1996; Gower et al. 2001). Despite this, the unique dynamics of moss biogeochemistry are not typically included in models of boreal forest C cycling, which could result in considerable biases. For example, Bona et al. (2013) used a literature review of the rates of primary production and decomposition of upland mosses to estimate the range of moss C that could be stored in the soils of black spruce-dominated boreal forests, and the result (31-49% of total SOC) was comparable to the difference between modeled and observed C stocks for those forests. This demonstrates that accurately representing moss production and decomposition is essential for modeling the C cycle in moss-rich boreal forests. While many studies estimate moss primary production, only three estimates of degradation rates were available for this meta-analysis, and none estimated the temperature sensitivity of moss decomposition (Bona et al. 2013). This is significant because the limited numbers of studies of upland mosses show slower decomposition than vascular plant litter incubated under similar conditions (Harden et al., 1997; Moore and Basiliko, 2006). Omitting moss-specific dynamics could therefore overestimate decomposition and underestimate C storage in moss-rich soils.

Predicting the temperature sensitivity of moss decomposition is ~~also~~ difficult because it is not clear if the apparent recalcitrance of moss tissues is due to chemical complexity, physical and structural characteristics impeding microbial decomposition, or a combination of the two. Most studies of moss biochemistry have focused on the peat-forming *Sphagnum* genus, and it is not clear to what extent these insights apply to non-*Sphagnum* upland species. While mosses do not contain lignin (Maksimova et al., 2013), they do produce a variety of lignin-like phenols, which have been hypothesized to inhibit decomposition (Tsuneda et al., 2001). In addition, *Sphagnum* mosses produce structural carbohydrates that also appear to impede decomposition (Hájek et al., 2011; Turetsky et al., 2008). If the decomposition of upland mosses is limited by chemical properties, we might expect higher temperature sensitivity of moss decomposition compared to vascular plant litter due to higher activation energy of

more chemically complex compounds (Bosatta and Ågren, 1999; Davidson et al., 2006). This result would suggest the moss C pool is not only under-represented in size due to slow decomposition, but is also particularly vulnerable to decomposition with warming.

Distinct from moss chemical characteristics, the physiochemical matrix of moss cell walls also could play a role in limiting microbial access to moss tissues otherwise useful as microbial resources. Indeed, scanning electron micrographs of a slowly decomposing *Sphagnum* species (*S. fuscum*) revealed little change in the structure of the cell wall after three years of decomposition, suggesting that something inherent about moss cell wall structure presented a barrier to microbial access, even long after cell death (Turetsky et al. 2008). If so, moss tissue chemistry *per se* may not require a higher activation energy than vascular plant tissues for decay to proceed, but decay instead would be limited by poor ~~microbial~~ physical accessibility of microbes to usable resources. However, it is not clear if the observed decay resistance is widespread, as the cell wall structure of another *Sphagnum* species (*S. riparium*) collapsed after one year of decomposition (Turetsky et al. 2008). The degree to which upland moss cell walls retain their physical structure after death and impede microbial decomposition remains unknown.

We observed the decay of upland boreal forest mosses collected from the Newfoundland and Labrador Boreal Ecosystem Latitudinal Transect (NL-BELT) for more than 2.5 y to investigate (1) the temperature sensitivity of moss tissue decomposition; and (2) the relationship between moss chemical composition, cell wall structure, and its decomposition. We combined chemical characterization with scanning electron microscopy (SEM) to determine both chemical and physical changes in the moss tissues during decomposition. In doing so we investigate the relative importance of these factors contributing to the slow turnover of moss tissues in these forests.

2 Methods

2.1 Sample sites

Moss samples were collected in July 2011 from two balsam fir dominated forest sites from within the Newfoundland and Labrador Boreal Ecosystem Latitudinal Transect (NL-BELT). One site was located in the Salmon River watershed near Main Brook on Newfoundland's northern peninsula (hereafter "northern forest~~SR~~"), and one in the Grand Codroy watershed in southwestern Newfoundland ("southern forest~~GC~~"). The mean annual temperature (MAT) ~~at SR~~at the northern forest is 3.2°C lower than ~~GC~~ in the southern forest (2.0 vs. 5.2°C; Table 1), while mean annual precipitation is higher ~~at GC~~at the southern forest (1224 vs. 1505 mm; Environment Canada Normals 1981-2010). Both sites contain humo-ferric podzol soils. Soil pH was measured on water extracts from F and H horizon soils and was low at both sites (4.39±0.26 at the northern forest and 4.03±0.22 at the southern forest; Table 1). The dominant moss species were *Dicranum spp.* a~~A~~nd *Rhytidiadelphus triquetrus* ~~Rhytidiadelfus spp.~~ in ~~SR~~the northern forest, and *Pleurozium schreberi*~~Pluerozium spp.~~, *Hylocomium splendens*, and *Ptilium crista-castrensis* ~~Ptillium crista cristensis~~ in ~~GC~~the southern forest, which had lower overall moss coverage based on examination of 15 1 m^2 plots at each site (Table 1). A total combined sample of over 50 g dry weight of moss tissue (green and brown parts) was sampled from across each site. To obtain a sample representative of each site, subsamples (10-20 gdw each) were collected over ~ 3 m^2 area in each of five locations around the entire edge and

just outside of each site. This was done to avoid destructive sampling of the moss layer within the study site. ~~Representative samples of the dominant moss species were collected from both sites,~~ Moss samples were then separated into green and brown portions and rinsed with DI water. The green tissues of different species were mixed together to create a homogenised sample for each site.~~, and homogenized by site prior to incubation. We sampled mosses from multiple patches in both forests.~~ The collection was focused on accurately reflecting the moss cover at each site and no effort was made to ensure all species were represented.

2.2 Incubation experiment

Incubations were designed to include four destructive-sampling time points which occurred at 69, 283, 648, and 959 days from the beginning of the experiment, starting in October 2011. The green portion of moss tissues~~es~~ from both regions were incubated in the dark within sealed glass jars at 5°C and 18°C. For each site, 24 ~~replicate~~ jars (three replicates per site/temperature/time point combination) were established that contained 1, 1.5, 2.5, or 2.75 g dry weight of moss tissue; jars containing greater weights were sampled at later time points. A microbial inoculum derived from each site's organic horizon was added to their respective jars at the start of the incubation experiment. ~~A soil slurry~~The inoculum was created by saturating the soil sample from each site with nanoUV water (~120 gdw soil L^{-1}). The slurry was then filtered (GF/C; 1.2 μm nominal pore size) to exclude most soil organic matter while retaining many soil microorganisms and pipetted onto the moss sample in each jar at a volume required to achieve 60% water holding capacity (6-21 mL for the 1-2.75 g dry weight moss samples). Moss water holding capacity was predetermined on five replicate subsamples of each integrated moss sample. Whatman filter papers suspended on filter racks were saturated with water and once these filters stopped dripping, five pre-weighed, 1 g (fresh weight) replicates of each site's moss sample was placed onto separate moistened filter papers and water was slowly added until saturated and dripping. When dripping stopped, moss tissue samples were scraped into preweighed oven tins and weighed. Following this each sample was oven-dried in the same tin at 60°C for >48 h. Upon drying, each was weighed again and the amount of water per g dry weight moss was calculated as the water holding capacity for that material (ml g^{-1}).

Following inoculation, jars were sealed and incubated, with half the jars at 5 and half at 18°C (12 jars for each site and temperature combination). We opened all jars biweekly to allow for gas exchange and to add sufficient water to ensure moss was kept at approximately 60% water holding capacity. At each sampling time point all moss was removed from the jars and dried at 40°C to constant mass and weighed. Once weighed, moss was ground using a Wiley mill (Thomas Scientific, Swedesboro) with a 60 mesh (0.25 mm) screen and stored in glass vials in the dark for subsequent analyses.

2.3 Chemical analyses

To determine how decomposition affected organic matter composition, the elemental and stable carbon and nitrogen isotopic composition of the moss samples were analyzed on a Carlo Erba NA 1500 Series elemental analyzer interfaced to a Delta V Plus isotope ratio mass spectrometer via a ConfloIII interface (ThermoFisher

Scientific). In total 48 samples were analyzed, plus 6 initial samples taken as random triplicates from the homogenized prepared moss tissue from each site.

Sub-samples of the initial and final (959 days) moss samples were analyzed using solid state CPMAS ^{13}C-NMR to determine the proportions of carbon functional groups and how they changed with decomposition. Samples

5   were analyzed using a Bruker AVANCE II 600 MHz using a MASHCCND probe. All samples were run at 150.96 MHz for ^{13}C and spun at 20 kHz at a constant temperature of 298 K. Carbonyl and amide (190-165 ppm) were separated from each other by subtracting N:C (multiplied by 100%) from the total peak area ratio to determine the maximum amount carbonyl. The remaining provides an approximation of the maximum amide C proportion.

Water soluble inorganic nitrogen, measured as nitrate plus nitrite and ammonium, was determined using a

10   Lachat 8500 flow injection analyzer. All samples were first extracted using NanoUV water. Briefly, 300 mg of the ground moss material were shaken with 10 ml of water for 2 minutes at room temperature then centrifuged and filtered using a glass fiber filter (GF/F; nominal pore size of 0.45 μm) to remove particulate matter. The filtrate was then analyzed for NO_3^- and NH_4^+ concentration to determine the total water soluble inorganic nitrogen content of each moss tissue sample.

15   Total hydrolysable amino acids (THAA) were analyzed following the method outlined in Philben et al. (2016) using the EZ:Faast kit for amino acid analysis (Phenomenex, USA). Briefly, 20mg subsample of each moss sample was mixed with 1 ml of 6M HCl acid in a 1 ml ampule, which was sealed, shaken, and heated at 110°C for 20 hours. Samples were then transferred to 2 ml vials, and centrifuged. An aliquot of the resulting hydrosylate was transferred into a new vial and evaporated using N_2 gas. 200 μl of 0.01 M HCl was added to each hydrosylate along

20   with norvaline which was added as an internal standard. Amino acids were derivatized with propyl chloroformate using the EZ:Faast kit. Samples were analyzed on an Agilent 6890 gas chromatograph with a ZB-AAA column using a single step oven program of 110-320°C at 30° min^{-1} and quantified using a flame ionization detector. Fifteen amino acids were determined; alanine, glycine, valine, leucine, isoleucine, threonine, serine, proline, aspartic acid, hydroxyproline, glutamic acid, phenylalanine, lysine, histidine and tyrosine.

25   **2.4 Scanning electron microscopy**

We performed scanning electron microscopy (SEM) on prepared fragments of the initial and final moss tissues from this experiment using a JEOL JSM 7100F Field Emission SEM equipment with a Thermo EDS. Three random subsamples where taken from the homogenized whole dried (40˚C) initial moss samples used to establish the incubation mesocosms. ~~These were pooled to provide representative samples of the initial moss tissues.~~

30   Subsamples of each of the final 18˚C incubation samples were used to provide the most degraded samples to compare with the initial sample images. All samples where dried at 50˚C, mounted on aluminum stubs, and coated with 300 angstroms of gold using a SPI-Module Sputter Coater (Structure Probe, Inc.; West Chester, PA, USA).

2.5 Data analysis

Percent mass remaining was calculated using the dry weights of the initial and final mass at the four time points of

35   the incubation. Mass remaining was then fit to the exponential decay equation to calculate the rate of decay:

$$y = L\left(e^{-\mathrm{kt}}\right) + R. \tag{1}$$

L represents the labile fraction of mass that was decomposed by the end of the experiment, R was the residual fraction was left undecomposed, t represents time in days and k is the exponential decay constant (day^{-1}). Carbon and nitrogen remaining was also fit to equation (1) where possible.

5   Q_{10} values were calculated to measure the temperature sensitivity of moss decomposition. This was performed by comparing C loss over the full experiment (equation 2), or using the k-values calculated in equation (1) to estimate the temperature sensitivity of decomposition of the labile fraction (equation 3).

$$Q_{10,Total} = \left(\frac{CL_{T2}}{CL_{T1}}\right)^{\frac{10}{T2-T1}} \tag{2}$$

$$Q_{10,Labile} = \left(\frac{k_{T2}}{k_{T1}}\right)^{\frac{10}{T2-T1}} \tag{3}$$

10   CL indicates the percentage C loss, T2 indicates the warmer temperature (18°C), and T1 indicates the cooler temperature (5°C).

THAA data were analyzed to determine the percentages of total C or N as amino acids, and their change over incubation time, using the equation from Philben et al. (2016):

THAA (%C or N) = Σ[Yield$_{AA}$/(C or N)] x [Wt % (C or N)]$_{AA}$ (4)

15   where Yield$_{AA}$/(C or N) is the normalized yield of each amino acid (15 in total) in mg amino acid per 100mg C or N and [Wt % (C or N)]$_{AA}$ is the weight % of C or N in the amino acid. This equation was used for each individual amino acid, which was then summed for each sample. The THAA results were also used to determine the degradation index, commonly used for interpreting the extent organic matter diagenesis in aquatic environments (Dauwe et al., 1999; Dauwe and Middelburg, 1998; Menzel et al., 2015). The index was modified for use at the sites

20   used in the current study to permit assessment of the degradation state of the sites' soil organic matter pools (Philben et al. 2016). In that work, a principle component analysis (PCA) of a data set including the green ~~mosses~~portion of moss tissues, pooled needle litterfall, and L, F, H and B soil horizons from three regions within the NL-BELT was conducted to identify the most significant differences in the overall THAA composition. Scores of the first principle component declined with increasing decomposition. This approach allows for the comparison of the moss tissues in

25   the current incubation study and the soil profiles at each site. The equation taken from Philben et al. (2016) was:

$$\text{Degradation Index} = \left[\Sigma_i\left(\frac{Mol_i - Avg_i}{SD_i}\right) \times PCI_i\right] \tag{5}$$

where Avg_i and SD_i are the average and standard deviation of the individual amino acid (mol%) determined for the data set described above, Mol_i is the mol% of each amino acid analyzed from the mosses, and PCl_i is the loading of the amino acids on the first principle component of the PCA performed on the dataset of all litter, soil horizons, and

30   moss from all regions across the NL-BELT transect. See Philben et al. (2016) for more detail.

To test for effects of incubation temperature, site, and their interaction on all quantified variables, we applied two-way ANOVA using a mixed effects model ~~to account for the pseudoreplication of repeated measures~~. Additional 2-way ANOVA tests were conducted within each time point to determine if any observed ~~the~~ treatment effects change~~ds~~d over the course of the incubation. A two-way ANOVA was conducted to test the effects of site and

35   temperature on Alkyl: O-Alkyl, using the initial moss samples and final (day 959) moss samples at 5 and 18°C, and

the effects of site and time (before or after incubation) on mol% hydroxyproline, N speciation, and the degradation index. All ~~other~~ analyses were conducted in R 3.3.2 (R core team, 2016).

3 Results

3.1 Moss tissue decay rates and mass and elemental loss.

5 Both C and mass loss measured in this experiment were fit to an exponential equation as described in the methods (Fig 1). When incubated at 18°C, between 34 and 38% of the initial moss was lost, based on both the mass and C loss results. The rate of loss in the 18˚C incubation declined over time, with rapid loss from 0-283 days, less change from 283-648 days, and no change from 648 to 959 days. In the 5°C incubations, between 28 and 30% of mass or C was lost, with mass and C losses continuing throughout the incubation. Regardless of incubation

10 temperature, the percentage of mass or C remaining at the end of the 959 days was not different between sites (Table 2). Decay constants (k) associated with moss tissue mass or C remaining did not differ between sites and were greater in the 18°C incubation than at 5 °C (-0.0130±0.0015 vs. -0.00380±0.0007 respectively; average ± standard deviation). The Q_{10} values of mass loss during the full incubation ($Q_{10, total}$) averaged 1.24 and did not vary by site. The Q_{10} of the labile pool calculated from the fitted k-values were 3.56 and 2.08 for ~~SR and GC~~the northern and

15 southern forests, respectively.

The N remaining did not follow the same trend as mass and C, and could not be fitted to an exponential curve but rather exhibited both increases and decreases over the course of the incubation (Fig. 2). One treatment (~~GC~~ the southern forest at 18°C) experienced a net gain in total N during the incubation, suggesting N fixation occurred. Temperature did not have an effect on N remaining at most time points (Table 2 ~~3~~). However, collection

20 site had an effect on N remaining during decomposition at all time points except on day 69 due to greater N loss during the 18°C incubation of ~~GC mosses~~moss from the southern forest (up to 46%; Table 2).

3.2 Elemental and stable isotope composition of decaying moss tissues.

Mosses collected from the two sites differed in initial N concentration (0.92±0.05% and 0.59±0.03% in the ~~warmer~~southern (GC) and ~~cooler~~northern (SR) forest site, respectively; ~~t-test,~~ p~~<~~=0.00~~1~~3) but were not different in

25 C concentration (43.9±0.1% and 43.6±0.5, respectively~~;~~. ~~p=0.373; Table 2,~~ Table 3). This variation in N was responsible for the lower C:N ratio of 55.6±3 at the southern forest~~GC~~ relative to the higher value of 86.3±3 at ~~SR~~ the northern forest (p<0.001). However, C:N declined with decomposition to a greater extent in ~~SR~~ the northern forest than ~~GC~~in the southern forest, and the site difference in C:N decreased with incubation time (Fig. 3). Though the effect of site on C:N of the moss tissue was significant throughout the experiment, the differences between sites

30 became less pronounced with time, especially in the 18°C incubations, and the p-value increased over the course of the experiment (Table 3).

The initial δ^{13}C and δ^{15}N values of moss did not differ between sites. δ^{13}C was -31.8±0.5‰ and -32.0±0.2‰ for ~~SR and GC~~the northern and southern forests (p=0.3179), while δ^{15}N was -3.19±0.2‰ and -3.72±0.4‰, respectively ~~(p=0.1679)~~. Moss δ^{13}C increased over the course of the 18°C incubation for both sites

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(p=0.010) and increased during the 5°C only for ~~GC~~the southern forest. The increase in $\delta^{13}C$ was greatest in the 18°C incubation of ~~GC~~ southern forest mosses (1.5‰; Fig. 1). In all other incubations $\delta^{13}C$ increased by less than 1‰, with most of that change occurring within the first 400 days when the rate of C and mass loss was highest. Therefore, although $\delta^{13}C$ of moss tissues was not different for the initial values collected, the $\delta^{13}C$ values differed by

5   site throughout the decomposition experiment with the significance of that difference increasing with decomposition (Table 2). There was no effect of temperature on the $\delta^{13}C$, until the final time point when site, temperature, and their interaction had a significant effect due to an increase of $\delta^{13}C$ values in the ~~GC~~ southern forest 18°C incubation to - 30.6±0.3‰. Temperature, but not site, had an effect on $\delta^{15}N$ (p=0.035; Table 2). $\delta^{15}N$ increased during incubation in all treatments except for ~~SR~~ the northern forest at 18°C, by an average of 2.3±1.5‰ (Fig.~~ure~~ 2).

10   **3.3 Molecular composition of C and N and SEM images of decaying moss tissues.**

Nitrogen was further characterized into four compound classes: nitrate+nitrite, ammonium, total hydrolysable amino acids, and molecularly unidentified N (MUN) and expressed as a fraction of total N content (Fig. 4). Water-extractable ammonium increased slightly with decomposition from initial values of 2-2.5% to 4-7% in the 18°C treatment (p<0.001) and 5-11% in the 5°C treatment (p<0.001). Extractable nitrate+nitrite was less than

15   1% of total N in most samples, with the exception of the final time point when some nitrate+nitrite values reached up to 20% of total N. Relatively high nitrate concentrations at the final time point were associated with elevated N remaining values (> 100%) and more negative $\delta^{15}N$ values (Fig. 2 and 4). THAA declined from 51.2±3 and 50.9±5% to 26.4±7 and 21.2±3% of the total N in ~~SR and GC~~the northern and southern forests, respectively (p≤>0.001), and the effect of site was not significant on the decline ~~(p=0.238)~~. The decline in the %N as THAA over

20   the incubation coincided with changing amino acid composition, as the degradation index also declined as expected with decomposition (p≤>0.001). The change was larger in ~~SR~~the northern forest, declining from 1.4±0.7 and 2.6±0.5 to -1.6±0.5 and -1.3±0.1 in ~~GC and SR~~the southern and northern forest, respectively (p=0.0388). Mole % hydroxyproline increased over the first 69 days in both sites (from 0.9 to 1.5% in ~~GC~~ the southern forest and 1.1 to 1.3% in ~~SR~~the northern forest, Fig. 5). It remained elevated for the remainder of the incubation in ~~GC~~ the southern

25   forest and was significantly higher than the initial value (p=0.0496), but declined back to 1.1% in ~~SR~~ the northern forest after 959 days. Because %N as THAA declined with little change in the inorganic N pools, MUN increased in relative abundance with decomposition in both regions.

The CPMAS ^{13}C-NMR results indicate that the moss tissues from both forest sites were similar and dominated by O-alkyl and di-O-alkyl C (70%) with relatively little alkyl, carbonyl or aromatic C (Fig. 6; Table 3).

30   The NMR spectra of the moss tissues before and after incubation were broadly similar and exhibited little change in the relative proportions of the major C groups. As a result, the alkyl:O-alkyl ratio also exhibited no change with decomposition regardless of collection site and despite up to 50% mass loss. The largest change observed was in the calculated maximum amide value from the carbonyl-C and amide-C resolved at chemical shift 190-165 ppm. Calculated maximum amide-C increased from 1.85±0.1% to 2.30±0.04% and 1.16±0.06% to 2.18±0.1% for mosses

35   incubated at 18°C from ~~SR and GC~~northern and southern forests, respectively. Amino acids are likely a major

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source of amide C; however, the %C as THAA declined with decomposition, indicating the increasing relative abundance of amide C was due to MUN compounds.

SEM imaging revealed few apparent differences between the physical structure of moss tissues before and after incubation (Fig. 7, S1, and S2). Intact moss cell walls were visible in both sets of images with few signs of structural change. There was no evidence of warping, gouging, or pitting from microbial degradation of the cell wall following the incubation of mosses from either region.

4 Discussion

4.1 Decomposition of mosses is slower and less temperature sensitive than vascular plants

The low decay rates observed are consistent with previous studies of moss decomposition (Fyles and McGill, 1986; Hobbie et al. 2000; Hogg, 1993; Hagemann and Moroni 2015). Decay rates of moss tissues are typically lower than rates for vascular plant decay under similar conditions (Fyles and McGill, 1987; Hagemann and Moroni, 2015; Hobbie, 1996). Mass loss during one-year litter bag decomposition of balsam fir needles averaged 27% at ~~SR~~ the northern forest and 35% at ~~GC~~ the southern forest (K. Edwards, unpublished data). The needles therefore experienced similar mass loss in one year compared to the mosses after 959 days, indicating more rapid decomposition. The moss decay rates can also be compared to incubations of L horizon soils collected from these sites, which were comprised of ~66% partially decomposed balsam fir litter in ER and ~83% in GC according to visual inspection. Despite their more advanced state of degradation compared to the fresh moss litter, the L horizon soils also experienced C losses similar to those experienced by the moss samples (30-45%) in a shorter time period (68 weeks). The low decay rates observed are unlikely to be an artefact of the laboratory approach, as Bengstton et al. (2016) compared field-based litter bag and laboratory incubation approaches using a common set of *Sphagnum* mosses and found the laboratory approach generally resulted in greater mass loss.

The ~~SR~~ northern forest mosses, but not the ~~GC~~ southern forest mosses, exhibited higher Q_{10} than the bulk L horizon soil (Laganière et al., 2015; Podrebarac et al., 2016) and previous findings for vascular plant tissue decomposition (e.g. Fierer et al. 2005) based on the decay rate of the labile C fraction. However, the higher temperature only slightly increased the total C degraded after 959 days. The Q_{10} value based on total mass loss was therefore lower than the L horizon soils. This indicates that the additional energy in the warmer treatment was not sufficient to induce additional decomposition, suggesting that decomposition was not limited by activation energy, contrary to the C-quality temperature hypothesis (Bosatta and Ågren, 1999; Davidson et al., 2006). Warming and drying trends in the boreal regions that inhibit moss growth (Gower et al., 2001; Turetsky, 2003) could result in the formation of SOM comprised of a greater relative abundance of vascular plant tissues, and thus of SOM that is both more decomposable and more temperature sensitive. This is consistent with our observations of increasing temperature sensitivity of soil respiration at lower latitudes along this boreal forest transect where moss inputs are reduced (Podrebarac et al. 2016).

We did not observe significant differences between sites' total C loss or Q_{10}, despite differences in the moss species incubated and their C:N. These results contrast with previous reports of the importance of species

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composition and N content in determining litter decay rates (Bengtsson et al. 2018; Bragazza et al. 2007; Hobbie, 1996; Lang et al. 2009; Limpens and Berendse, 2003). Decomposition studies utilizing a range of *Sphagnum* species have also found significant species effects. (Bengtsson et al. 2018; Bragazza et al. 2007; Johnson and Damman, 1991; Turetsky et al. 2008). Our study utilized the mixture of moss species naturally occurring at each site, and we therefore cannot isolate the effect of species from that of regional differences in C:N. However, the lack of difference in moss decomposition temperature responses across regions despite differences in moss species represented suggests that slow decomposition and low Q_{10} are more widely distributed among upland mosses in these forest ecosystems.

4.2 The cell wall matrix governs low decomposition rates and temperature sensitivities of decay

The low Q_{10} values for total mass loss suggest the low decay rates are not caused by chemical complexity or recalcitrance of bulk moss tissues, properties associated with high activation energies and correspondingly high temperature sensitivities of decay. This idea is supported by the ^{13}C NMR data, which indicate that the moss OM is rich in carbohydrates with little aromatic or alkyl C. The proportions of O-alkyl C in mosses were higher than vascular plant litter collected from the same study sites (55.3% vs. 37.0%; Kohl et al. 2018). Further, the OM composition did not change significantly following incubation, unlike the decomposition of vascular plant tissues and SOM in which O-alkyl C is often preferentially degraded and alkyl C increases in relative abundance (Baldock et al., 1997; Kögel-Knabner, 1997; Preston et al., 2009). The lack of change in the moss alkyl:O-alkyl ratio during decomposition does not appear to result from low mass loss in these incubations, given the increase in this ratio after a similar amount of mass loss (approximately 40%) in the foliage of each of 10 tree species in Canadian boreal forests (Preston et al., 2009). In conjunction with the relatively low temperature sensitivities of decay, the high proportion of O-alkyl C and lack of change in the alkyl:O-alkyl ratio following decomposition suggests that something other than bulk chemical composition governs the relatively low decomposition rates observed during the moss incubations.

There is also no evidence that decay-inducing microorganisms were limited by N availability. Indeed, varying concentrations of moss tissue N and C:N ratios across sites were not related to changing decay rates. Lower C:N ratios are ~~typically~~ often correlated with faster decay in both mosses (Aerts et al., 2001; Bragazza et al., 2007; Limpens and Berendse, 2003) and vascular plant tissues (Nadelhoffer et al. 199~~2~~1; Hobbie et al. 1996) in boreal soils, suggesting N limitation of decomposition. However, in another field study N fertilization altered the N cycle but did not affect C losses from moss tissues (Manninen et al. 2016). The N content was significantly lower and C:N higher in SR compared to GC (86.3±1.8 and 53.0±3.1, respectively), which would exacerbate N limitation in SR moss tissues. The difference in N content appears to be related to site N availability rather than differences in moss species because the C:N of balsam fir needles are also higher in the northern forest than in the southern forest (Ziegler et al. 2017; Kohl et al. 2018). Limpens et al. (2003) demonstrated that *Sphagnum* decay resulted in net N mineralization below the threshold C:N of 67. Our data are consistent with these results, as net N loss was observed for the ~~GC~~ southern forest moss with C:N < 67 but not from the ~~SR~~ northern forest mosses with higher C:N, although species-specific differences between sites cannot be ruled out. ~~However,~~Regardless of the causes for the

higher C:N in the ~~SR~~northern forest mosses the lack of difference in decomposition ~~despite greater N availability in GC~~ suggests that N availability was not the limiting factor for moss decay rates.

The changing composition of N during the incubations suggests that rapid turnover of the N pool could have reduced microbial N limitation. The %N as THAA declined from ~50% to ~20% in the first 69 days, while the amino acid degradation index declined from ~2 to ~-1 over this period. Total N declined by <15% and increases in inorganic N were <5% over this period, indicating the degraded organic N was mostly transformed to MUN. Previous studies indicate the accumulation of amino sugars derived from microbial residues could contribute to the MUN pool (Tremblay and Benner, 2006). The decline in %N as THAA and the degradation index are similar in magnitude to the difference in composition between the L and the H horizon in these soils (Philben et al. 2016). This indicates extensive degradation and turnover of the relatively small moss N pool, perhaps explaining the apparent lack of microbial N limitation despite high C:N ratios. The rapid turnover of the N pool indicates uncoupling of the N and C cycles in these incubations, and suggests the recalcitrant cell wall matrix does not protect most of the moss protein and peptide N pool, which appear to remain labile.

Despite the rapid turnover of the bulk amino acid pool, changing AA composition during the incubation indicates the cell wall matrix protected some proteins as well. Hydroxyproline is found in glycoproteins in the plant cell wall, but lacks major microbial sources (Philben et al. 2013). The increase in Hyp relative to other amino acids in the incubations of the GC mosses therefore indicates selective preservation of these cell wall proteins. The ~~SR~~ northern forest incubations exhibit an increase in mol% Hyp, followed by a decline to near the pre-incubation value. This regional difference is likely due to the difference in bulk N dynamics: the decline in C:N during the ~~SR~~ northern forest incubations indicates microbial N immobilization, which would produce amino acids but not Hyp and lower its mol%. Dilution of Hyp by microbial protein synthesis appears to have counteracted selective preservation of cell wall glycoproteins at ~~SR~~the northern forest. These dynamics further support the importance of the cell wall matrix in organic matter stabilization during the incubations.

The persistence of the physical structure of the cell wall likely explains the observed combination of slow decomposition and its low temperature sensitivity. The SEM images indicate microbial access to chemically labile C is inhibited by the biochemistry and physical matrix of the moss cell wall. The lack of microbial access to otherwise reactive microbial resources appears to be a bottleneck to decomposition that is not alleviated by increasing temperature, explaining our observation of low Q_{10} and a large O-alkyl C pool despite apparently recalcitrant litter.

Although they share a backbone of cellulose microfibrils, there are important chemical and structural differences between moss and vascular plant cell walls which could contribute to the persistence of the moss cell wall during the incubation (Roberts et al. 2012). Uronic acids (glucuronic acid and galacturonic acid) are more abundant in mosses than vascular plants (Popper and Fry 2003). The NMR spectra indicated O- and di-O alkyl functional groups account for ~70% of the moss C, but molecular analysis of *Sphagnum* found about half of that total in five aldoses (glucose, galactose, mannose, rhamnose, and fucose) (Philben et al. 2014), leaving a large pool of uncharacterized moss carbohydrates. The low ratio of O-alkyl to di-O-alkyl C (3.7 vs. >4 in vascular plants) suggests that uronic acids contribute to this pool. *Sphagnum* also produces a group of pectin-like carbohydrates (termed "sphagnan") composed primarily of rhamnose, mannose, and galacturonic acid (Ballance et al., 2007). This

mixture appears to be resistant to decomposition and/or possess antimicrobial properties (Hájek et al., 2011; Stalheim et al., 2009). While this fraction has only been identified in *Sphagnum*, uronic acid-enriched carbohydrates in these upland mosses could play a similar role.

The NMR spectra also indicate a substantial contribution of phenolic and aromatic C (7% of the total C), despite the lack of lignin. The relative abundance of this fraction increased during decomposition, indicating selective preservation. Tsunida et al. (2001) identified an amorphous phenolic coating on the outside of the moss cell wall, and proposed that a specialized fungal consortium is required to break it down. This is analogous to the accelerated degradation of lignin in wood by white rot fungi (Rice et al. 2006), and is a plausible explanation of the lack of cell wall degradation observed in the SEM images as well as the accumulation of phenolic and aromatic C. If key fungal species were excluded from the soil slurry used as a microbial inoculum in the incubations, or if incubation conditions were not favorable for their growth, then their absence could lead to the persistence of the cell wall's structural integrity.

Overall, our data suggest that some combination of inherent molecular resistance to decomposition and the molecular architecture of the cell wall matrix make it difficult for microbes or their exoenzymes to access and catabolize. While these analyses can identify biochemical differences between moss and vascular plant cell walls, we cannot identify which of these differences specifically contribute to their apparent recalcitrance. This presents an intriguing avenue for future research, as the persistence of the physio-chemical integrity of the moss cell wall matrix is likely important for maintaining the globally significant pools of moss-derived C in both peatlands and forested uplands.

4.3 Implications for the bioavailability of boreal forest SOM

The contrasting responses of the bulk C composition and the THAA-based indices during moss decomposition could complicate interpretation of decay-focusedsuch data sets in moss-rich boreal forest soils. Decomposition of vascular plant tissues is typically characterized by selective loss of carbohydrates. Indices such as the carbohydrate yield and the O-alkyl to alkyl ratio of ^{13}C NMR spectra have therefore become useful and widely used indicators of the degree of SOM degradation, albeit with some caution (Baldock et al., 1997). However, our results indicate the O-alkyl C of moss tissues is not selectively degraded, and O-alkyl:alkyl C did not change following incubation (Fig. 8). The contribution of relatively recalcitrant moss-derived structural carbohydrates could cause the O-alkyl:alkyl C ratio to underestimate diagenesis in moss-rich boreal forest soils (Fig. 8d). In contrast, the change in the THAA-based indices (%N as THAA and the degradation index) following incubation was similar to the change with depth in the organic horizon of these soils (Fig. 8a,c) and the change during incubation of the L horizon (Philben et al. 2016). These results demonstrate that variations in biochemical composition among plant types can confound conventional geochemical interpretation of diagenetic indices, and underscore the value of using multiple independent indices for a holistic understanding of the degradation of different SOM components (Baldock et al., 1997).

Our measurements of the decay rates and Q_{10} of mosses contrast with latitudinal trends of bulk SOM bioavailability observed along the NL-BELT. Despite having the highest contribution of mosses to the organic

horizon, incubation experiments indicated that the ~~coolest region~~northern forest ~~in the transect~~ had the highest ~~specific~~ soil respiration rate (relative to initial soil C) ~~(R10)~~ and similar or lower Q_{10} compared to the other regions (Laganière et al., 2015; Podrebarac et al., 2016). The slow decomposition of moss tissues therefore does not appear to result in slow~~er~~ decomposition in of more moss-rich organic soils. However, the trend in organic matter composition is consistent with moss influence, as the coolest region is enriched in O-alkyl C and depleted in methoxy and aromatic C compared to the other regions (Kohl et al. 201~~7~~8). The larger proportion of labile C compounds in the cooler region soils is correlated with the bioreactivity of the bulk soil (indicated by R_{10}; Kohl et al. 201~~7~~8), suggesting moss contributions of labile C override the effects of slow decomposition of moss tissues themselves. This implies that the physical protection afforded by the cell wall declined in importance over time within the soil profile. It is possible that a fungal species or consortium absent from the soil inoculum are able to ~~can efficiently~~ degrade the cell wall phenolics *in situ*, enabling the broader microbial community to utilize the labile moss C. Physical processes associated with the burial of moss litter and incorporation into SOM could also contribute to a loss of cell wall integrity over time. This study points to the need to better understand the physical and biochemical mechanisms controlling moss tissue decay and its variations among upland forest species and microhabitats, as these factors and their interaction impose important controls on boreal forest soil C stocks and their response to changing climate.

Competing interests

The authors declare they have no conflict of interest.

Data availability

All data are included in the manuscript tables and the supplement file.

Acknowledgements

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Table 1. Study site characteristics including mean annual temperature (MAT) and mean annual precipitation (MAP).

	Salmon River "Northern Forest"	Grand Codroy "Southern Forest"
Climate station	Main Brook	Doyles
MAT(°C)[a]	2.0±2.4	5.2±2.0
MAP(mm)[a]	1223.9	1504.6
rainfall (mm)[a]	708.8	1110.4
Snowfall (cm)[a]	515.0	394.2
GSST[b]	14.00	16.0
Latitude	51°15'21.28"N	47°53'36.34"N
Longitude	56° 8'17.76"W	59°10'28.31"W
Elevation (m)	14	13.10
Soil pH	4.39±0.26	~~4.03±0.21~~4.03±0.21
Dominant moss Species[c]	*Pluerozium spp., Hylocomium splendens, Ptillium crista-cristensis*	*Dicranum spp., Rhytidiadelfus spp.*

[a] Canadian climate normals, 1981–2010, http://www.climate.weatheroffice.gc.ca/climate_normals/index_ e.html. MAT, mean annual temperature; MAP, mean annual precipitation.

[b] Growing season soil temperature (GSST) averaged over July and August in 2010 based on data from Canadian Forest Service, Atlantic Forestry Centre, in Corner Brook, Newfoundland, Canada.

[c] Identified by Kate Buckeridge, KJ Min, Kate Edwards, Andrea Skinner, Amanda Baker, Danny Pink

12　Table 2. Results from 2-way ANOVA tests of the effects of collection site and incubation temperature on mass, C and N remaining, molar C to N ratio (C:N), and stable
13　C (δ^{13}C) and N (δ^{15}N) isotope of the moss tissues. Results provided for both the full experiment (using a repeated measures ANOVA) and for each incubation time
14　(using 2-way ANOVA). Bolded values are significant according (α= 0.05).

		p values		
Parameter	Days	Temperature	Region	Interaction
%Mass Remaining	Full experiment	**<0.001**	0.742	0.083
(log transformed)	0	na	na	na
	69	**<0.001**	**0.028**	0.463
	283	**<0.001**	0.283	**0.009**
	648	**<0.001**	**0.011**	0.088
-	959	**0.007**	0.191	0.476
%C Remaining	Full experiment	**<0.001**	0.908	0.291
(log transformed)	0	na	na	na
	69	**<0.001**	0.375	0.562
	283	**<0.001**	0.879	**0.014**
	648	**<0.001**	0.429	0.310
-	959	**0.001**	0.388	0.568
%N Remaining	Full experiment	**0.007**	**0.009**	0.090
(log transformed)	0	na	na	na
	69	0.053	0.238	0.506
	283	0.157	**0.031**	0.729
	648	**0.005**	**<0.001**	**0.010**
-	959	0.716	**0.012**	0.207
C:N	Full experiment	0.869	**<0.001**	**0.008**
(log transformed)	0	na	**<0.001**	na
	69	0.383	**<0.001**	0.715
	283	0.051	**<0.001**	**0.026**
	648	0.232	**0.008**	**0.015**
-	959	**0.008**	**0.006**	0.124

		Temperature	Region	Interaction
$\delta^{13}C$	Full experiment	**0.010**	**<0.001**	0.235
	0	na	0.113	na
	69	0.818	**0.012**	0.788
	283	0.103	**0.002**	0.240
	648	0.052	**<0.001**	0.423
	959	**0.002**	**<0.001**	**0.025**
$\delta^{15}N$	Full experiment	**0.035**	0.417	0.160
	0	na	**0.028**	na
	69	0.762	0.225	0.797
	283	**<0.001**	0.164	0.309
	648	0.742	**0.032**	0.189
	959	0.841	0.485	0.286

		Temperature				Region			Interaction	
		df	F	p-value	n	F	p-value	n	F	p-value
%Mass Remaining (log transformed)	Full experiment	54	8.7106	**0.005**	54	0.034	0.855	54	0.9643	0.331
	0	na	na	na	na	na	na	na	na	na
	69	1	58.034	**<0.001**	1	7.158	**0.028**	1	0.595	0.463
	283	1	177.465	**<0.001**	1	1.327	0.283	1	11.79	**0.009**
	648	1	192.503	**<0.001**	1	10.984	**0.011**	1	3.782	0.088
	959	1	13.216	**0.007**	1	2.036	0.191	1	0.559	0.476
%C Remaining (log transformed)	Full experiment	54	12.547	**<0.001**	54	0.004	0.948	54	0.3615	0.550
	0	na	na	na	54	na	na	na	na	na
	69	1	43.420	**<0.001**	na	0.881	0.375	1	0.365	0.562
	283	1	245.256	**<0.001**	1	0.025	0.879	1	9.914	**0.014**
	648	1	189.016	**<0.001**	1	0.694	0.429	1	1.175	0.310
	959	1	22.573	**0.001**	1	0.834	0.388	1	0.355	0.568
%N Remaining (log transformed)	Full experiment	54	3.749	0.058	1	3.496	0.067	54	1.4262	0.238
	0	na	na	na	na	na	na	na	na	na
	69	1	5.131	0.053	1	1.629	0.238	1	0.484	0.506

		df	F	p	df	F	p	df	F	p
	283	1	2.436	0.157	1	6.859	**0.031**	1	0.129	0.729
	648	1	15.150	**0.005**	1	61.010	**<0.001**	1	11.32	**0.010**
	959	1	0.142	0.716	1	10.603	**0.012**	1	1.884	0.207
C:N (log transformed)	Full experiment	54	0.018	0.893	54	69.954	**<0.001**	54	4.984	**0.030**
	0	1	na	na	1	248.500	**<0.001**	1	na	na
	69	1	0.850	0.383	1	164.055	**<0.001**	1	0.143	0.715
	283	1	5.245	0.051	1	51.785	**<0.001**	1	7.452	**0.026**
	648	1	1.669	0.232	1	12.045	**0.008**	1	9.447	**0.015**
	959	1	12.035	**0.009**	1	14.156	**0.006**	1	2.948	0.124
$\delta^{13}C$	Full experiment	54	4.342	**0.042**	54	6.342	**0.015**	54	0.869	0.356
	0	1	na	na	1	3.175	0.113	1	na	na
	69	1	0.057	0.818	1	10.418	**0.012**	1	0.077	0.788
	283	1	3.389	0.103	1	20.498	**0.002**	1	1.610	0.240
	648	1	5.213	0.052	1	32.657	**<0.001**	1	0.712	0.423
	959	1	21.328	**0.002**	1	57.429	**<0.001**	1	7.587	**0.025**
$\delta^{15}N$	Full experiment	54	3.197	**0.081**	54	0.834	0.365	54	1.174	0.245
	0	1	na	na	1	7.148	**0.028**	1	na	na
	69	1	0.098	0.762	1	1.730	0.225	1	0.071	0.797
	283	1	120.174	**<0.001**	1	2.349	0.164	1	1.179	0.309
	648	1	0.117	0.742	1	6.704	**0.032**	1	0.117	0.742
	959	1	0.043	0.841	1	0.537	0.485	1	1.306	0.286

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Table 3. The CPMAS ^{13}C-NMR results given as percentages of total C resolved for each chemical shift ~~range, site provided~~, molar C to N ratio (C:N), and weight % C and N of the initial and final moss tissues from both sites and incubated at 5°C and 18°C top. All values are provided as the mean ± on standard deviation of three replicates.

	~~Salmon River (cooler forest)~~ Northern Forest			~~Grand Codroy (warmer forest)~~ Southern Forest		
	Initial	Final (5°C)	Final (18°C)	Initial	Final (5°C)	Final (18°C)
Alkyl (50-0)	9.08 ± 0.2	8.23 ± 0.6	8.57 ± 2	9.88 ± 0.5	8.66 ± 0.2	7.57 ± 0.7
O-alkyl (90-65)	55.31 ± 0.3	54.01 ± 1.0	55.36 ± 3	54.75 ± 1.6	54.98 ± 0.5	57.09 ± 0.3
Di-O-alkyl (110-90)	15.57 ± 0.1	15.32 ± 0.1	15.19 ± 1	15.11 ± 0.1	15.06 ± 0.2	15.49 ± 0.1
Aromatic (145-110)	4.44 ± 0.2	7.18 ± 0.1	6.55 ± 0.02	5.05 ± 1.9	6.05 ± 0.004	5.64 ± 0.9
Phenolic (165-145)	2.35 ± 0.2	2.07 ± 0.1	1.74 ± 0.2	1.68 ± 0.4	1.57 ± 0.1	1.24 ± 0.01
Carbonyl and amide (190-165)	3.40 ± 0.2	4.76 ± 0.2	4.61 ± 1	4.15 ± 0.1	5.12 ± 0.2	4.69 ± 0.2
Aromatic and Phenolic (145-110)	6.79 ± 0.4	9.25 ± 0.1	8.29 ± 0.2	6.74 ± 2.3	7.62 ± 0.1	6.87 ± 0.9
O-alkyl:Di-O-Alkyl	3.55 ± 0.03	3.53 ± 0.04	3.64 ± 0.1	3.62 ± 0.1	3.65 ± 0.01	3.69 ± 0.002
Alkyl:O-Alkyl	0.16 ± 0.003	0.15 ± 0.01	0.16 ± 0.04	0.18 ± 0.004	0.16 ± 0.002	0.13 ± 0.01
Carboxyl/ester *	2.24 ± 0.1	3.14 ± 0.2	2.43 ± 1	2.30 ± 0.1	2.92 ± 0.5	2.39 ± 0.1
Amide*	1.16 ± 0.06	1.62 ± 0.4	2.18 ± 0.1	1.85 ± 0.1	2.20 ± 0.2	2.30 ± 0.04
% C ~~Carbon~~	43.6 ± 0.5	44.4 ± 0.4	43.3 ± 0.3	43.9 ± 0.1	44.1 ± 0.2	42.3 ± 0.7
% N ~~Nitrogen~~	0.590 ± 0.03	0.823 ± 0.1	1.08 ± 0.07	0.920 ± 0.05	1.12 ± 0.09	1.19 ± 0.08
C:N	86.3 3	64.0 9.7	46.9 2.7	55.6 3.2	46.1 3.7	41.6 3.4
δ^{15}N	-3.19 ± 0.2	-1.25 ± 0.8	-2.42 ± 0.4	-3.72 ± 0.4	-1.61 ± 2	-0.8 ± 2
δ^{13}C	-31.8 ± 0.5	-32.1 ± 0.2	-31.8 ± 0.2	-32.0 ± 0.2	-31.5 ± 0.03	-30.6 ± 0.3
% N as ~~n~~Nitrate ~~+nitrite~~	0.058 ± 0.006	6.5 ± 9.0	10.4 ± 9	0.034 ± 0.003	7.27 ± 11	6.21 ± 10.0
% N as ~~a~~Ammoni~~um~~a	2.41 ± 0.1	8.41 ± 2.0	5.24 ± 2	1.59 ± 0.06	8.14 ± 3	6.99 ± 1.0
%C as amino acids	2.54 ± 0.03	na	2.31 ± 0.7	3.83 ± 0.03	na	2.12 ± 0.7
%N as amino acids	52.1 ± 1.1	na	26.4 6.6	50.9 ± 5	na	21.2 ± 3.0
Total amino acids (nmol/mg)	207.2 ± 3	na	193.0 ± 60	313.0 ± 30	na	171.0 ± 20.0

20 *maximum estimated value for amide-C and carboxyl/ester-C determined from the carbonyl-C and C:N ratio, see methods for details.
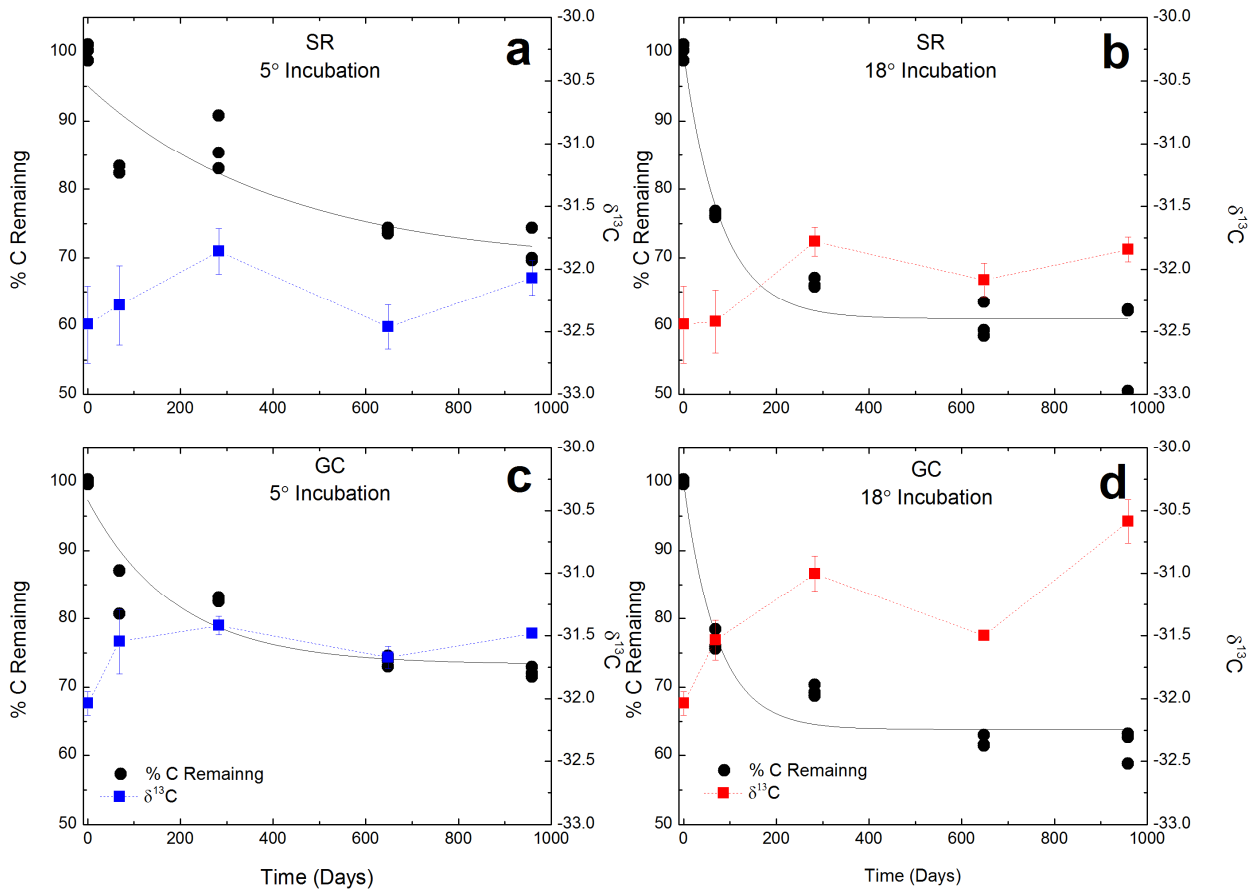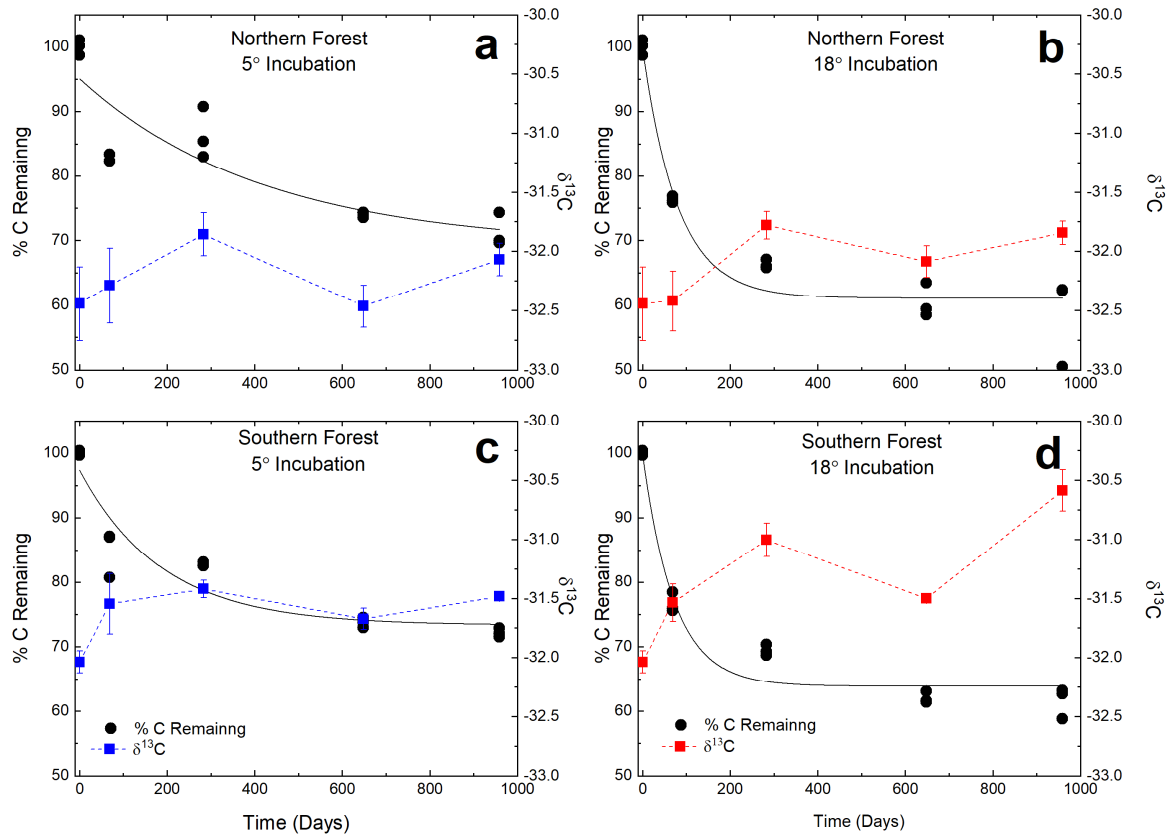
Figure 1. Moss tissue C remaining normalized to the initial tissue C (black circles). The solid black line represents the exponential fit used to assess the decay constant (k). The δ^{13}C values for the moss tissue at each time are indicated by squares and dashed lines, reported as the mean (n=3) with the standard error depicted by error bars. Panels a and c (blue lines and symbols) depict results of the 5°C incubation, and panels b and d (red lines and symbols) depict the 18°C incubation.
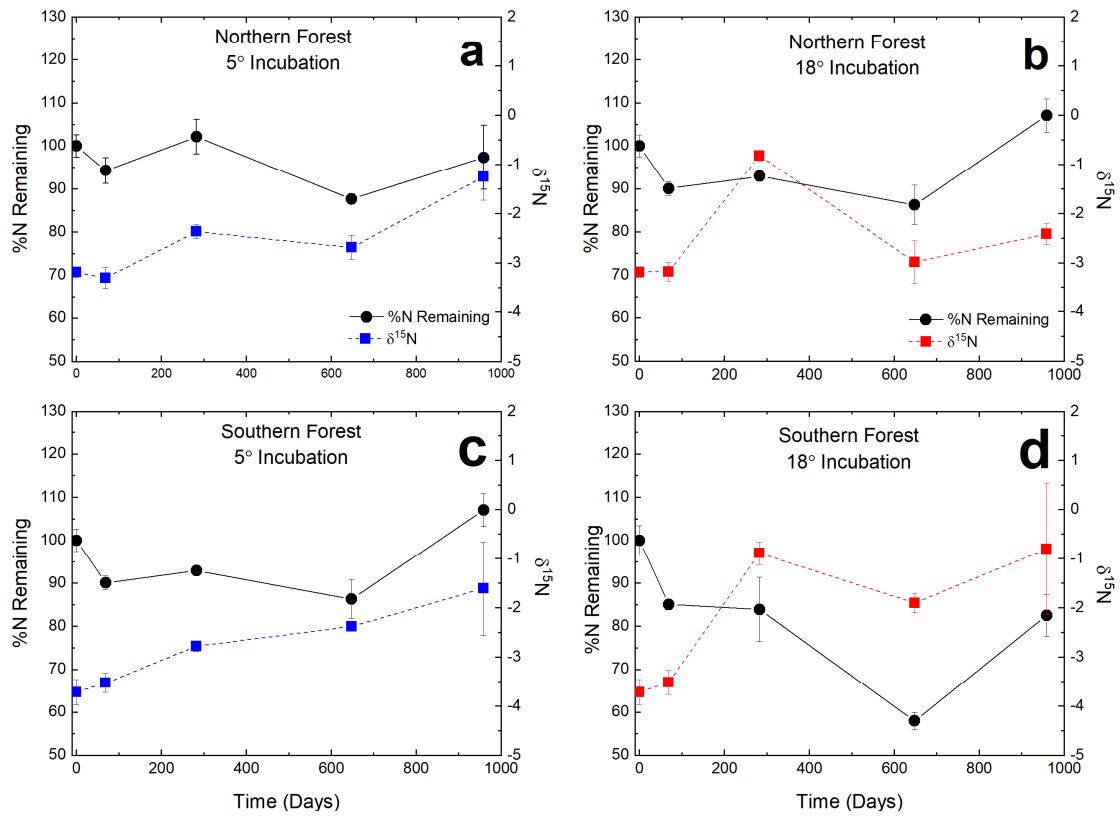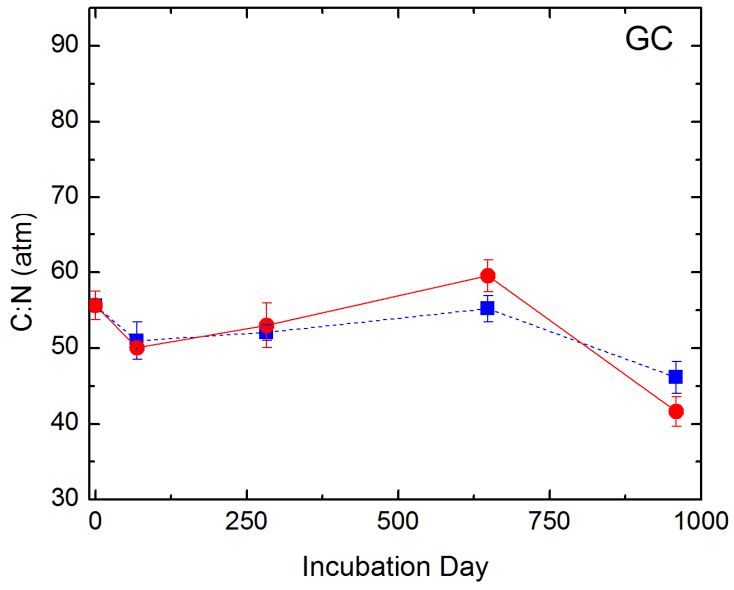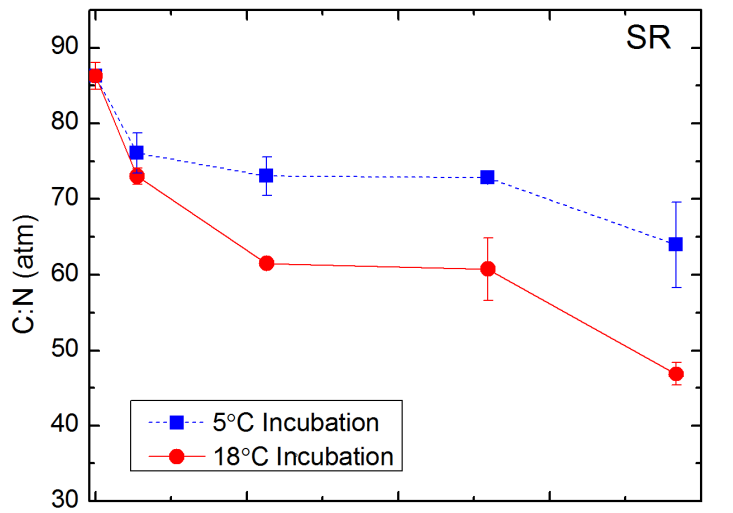
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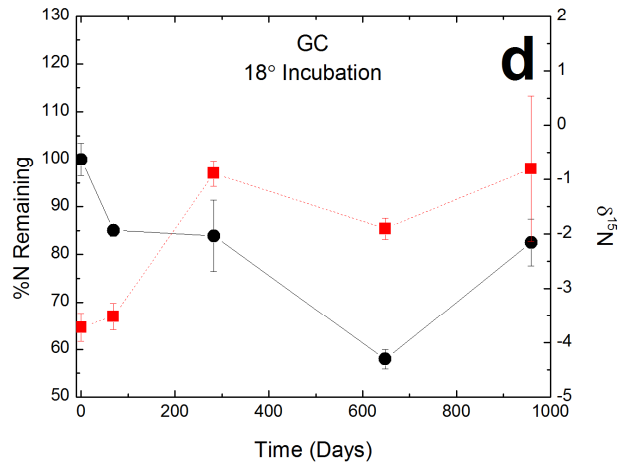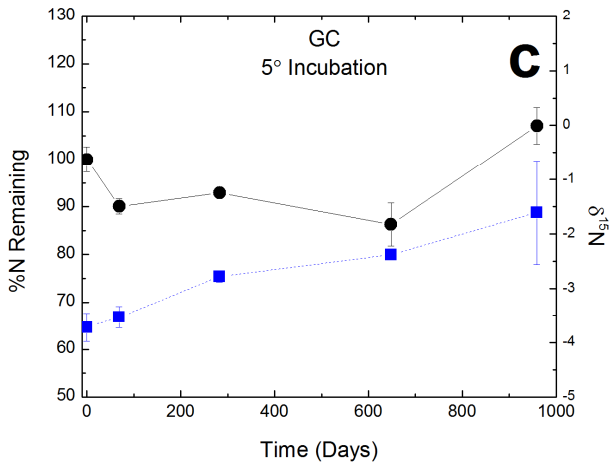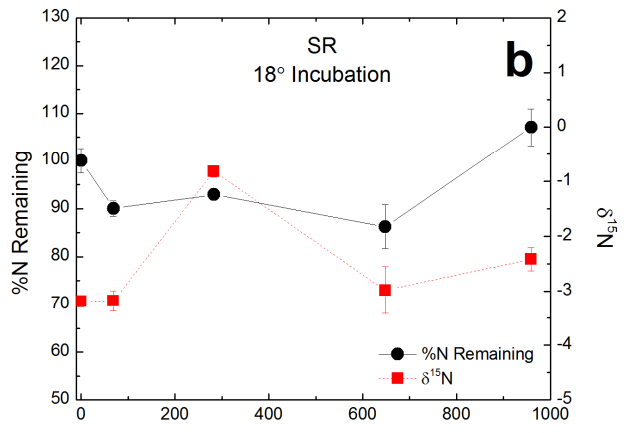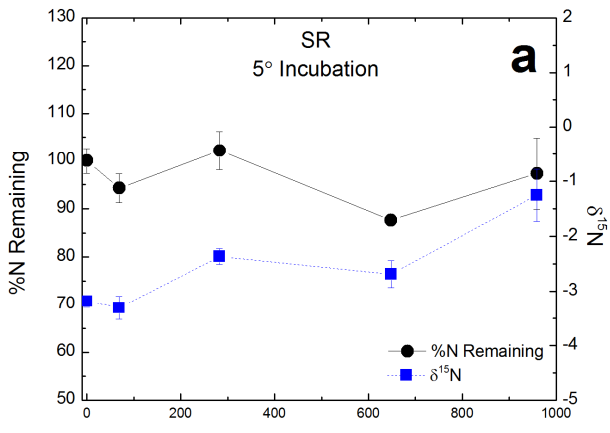
Figure 2. Percentage of initial nitrogen remaining (circles and solid lines) and δ¹⁵N (squares and dashed lines) at each time point. Points indicate the mean (n=3) with standard error depicted by error bars. Values in blue (panels a and c) are results from the 5˙C incubation and those in red (panels b and d) are from the 18˙C incubation.

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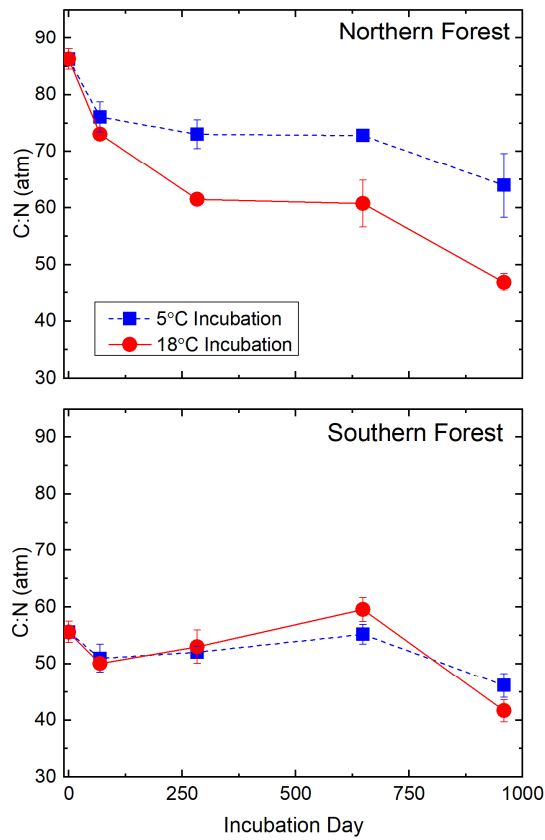
Figure 3. Molar carbon to nitrogen ratio of moss tissues from the cooler (upper panel) and warmer (lower panel) forest sites plotted against incubation time. Values for the 5˚C and 18˚C incubations temperatures are given by the blue squares and red circles, respectively, and reported as the mean (n=3) with standard error depicted by error bars.
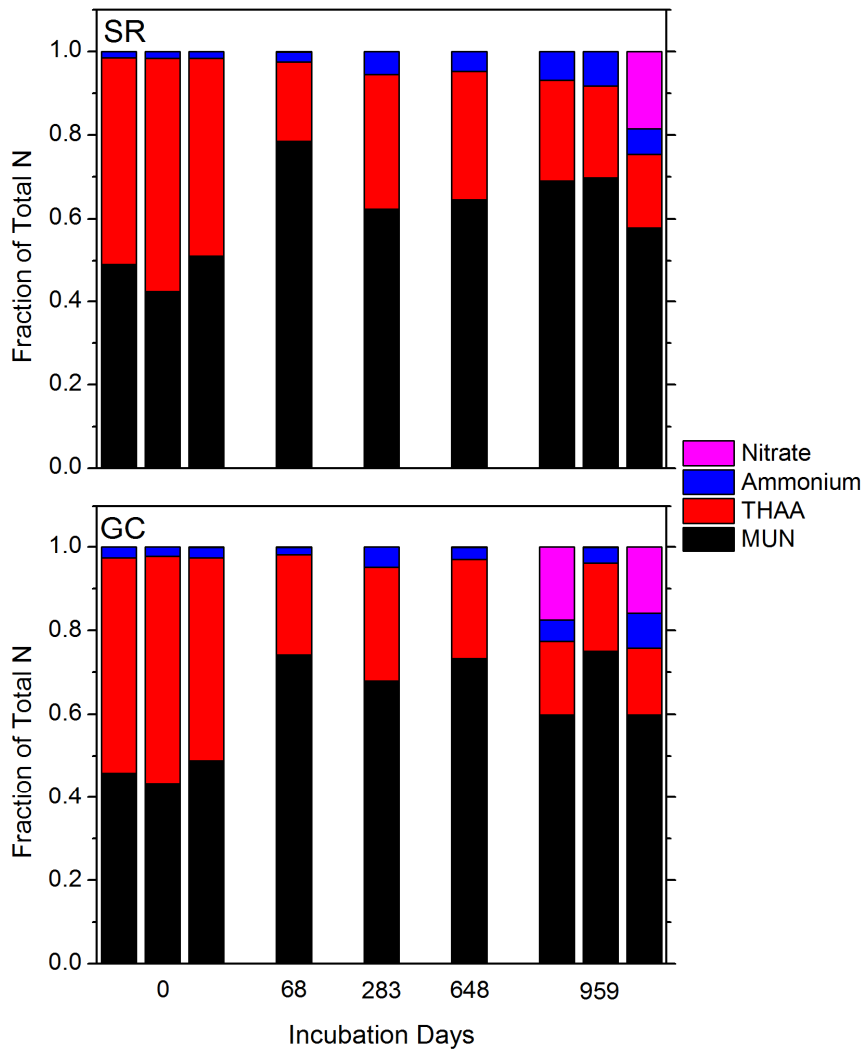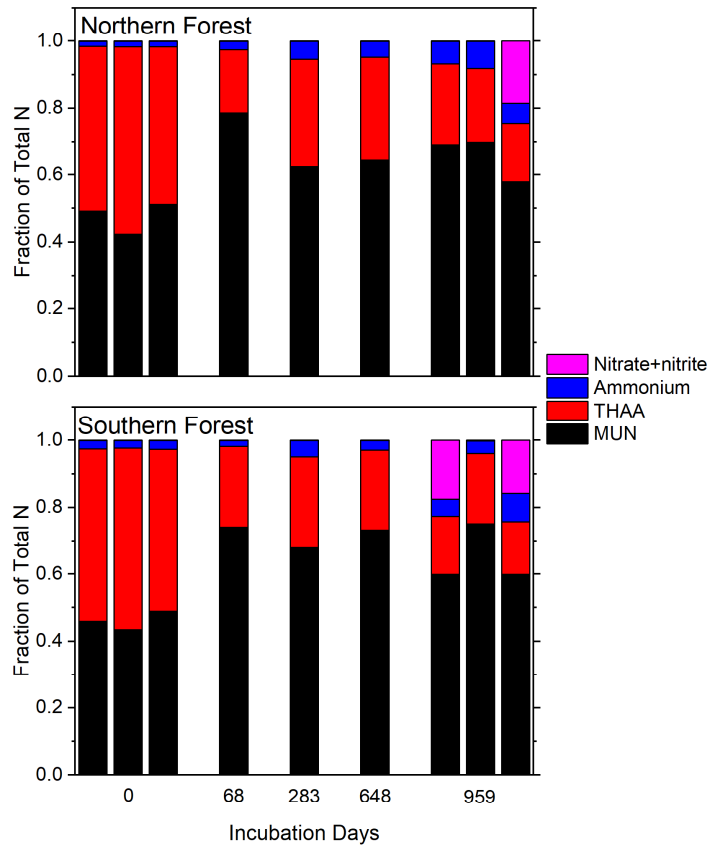
Figure 4. Fraction of the total moss tissue N content as nitrate, ammonium, total hydrolysable amino acids (THAA), and molecularly unidentified N (MUN) in each sample. Total hydrolyzable amino acids were only reported for one triplicate of the 18˚C incubations for the middle time points (69, 283, 648 days) though all samples were tested for the inorganic N species.
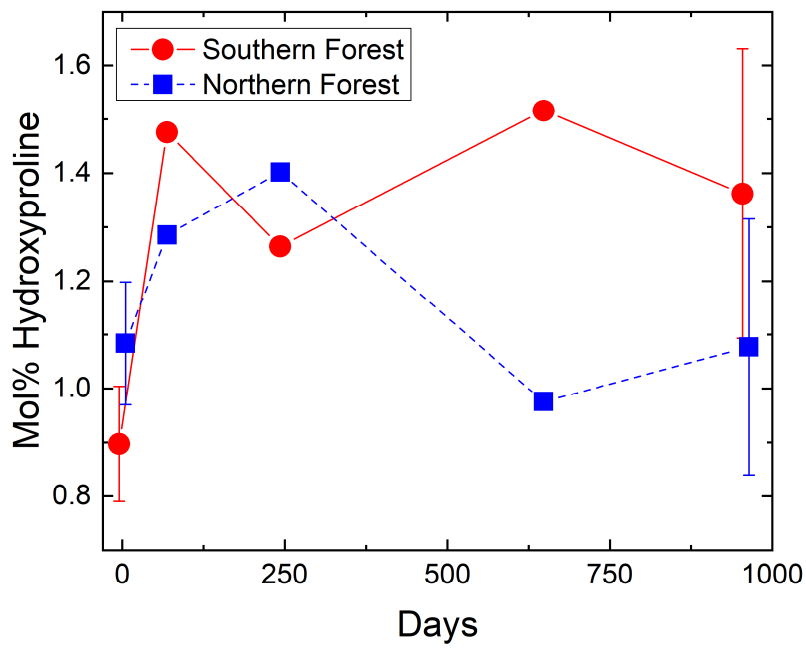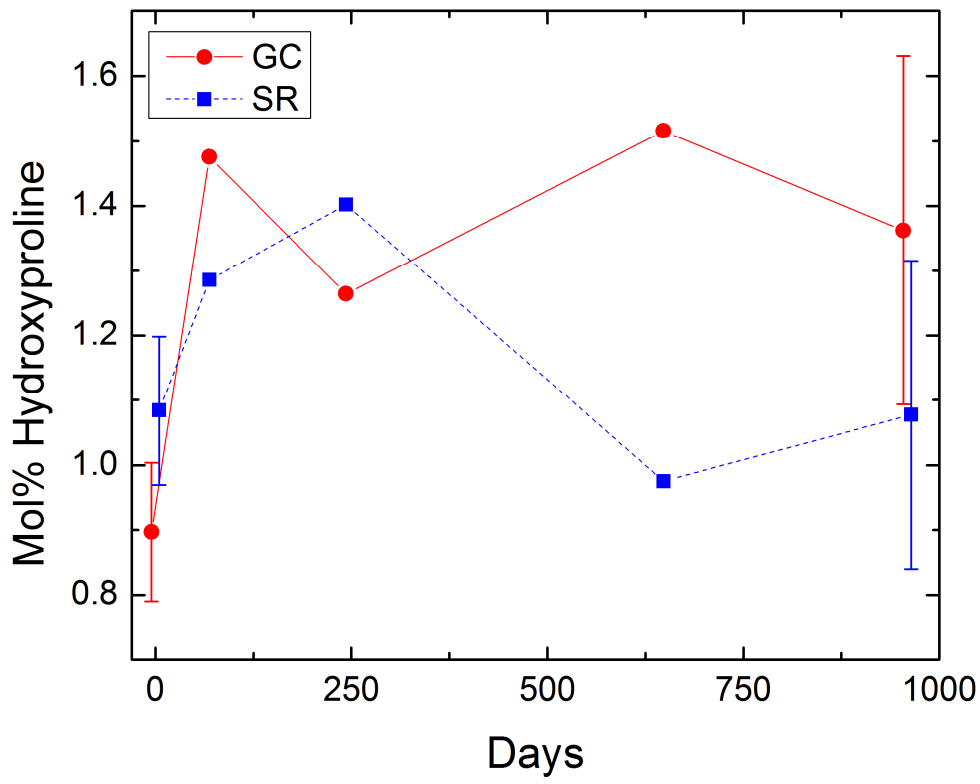
Figure 5: Hydroxyproline yields in the moss tissues as a percentage of total hydrolysable amino acids. Error bars for the initial and final time points (0 and 959 days) indicate standard deviation (n=3). The initial and final time points are jittered (± 5 days) to display error bars without overplotting.
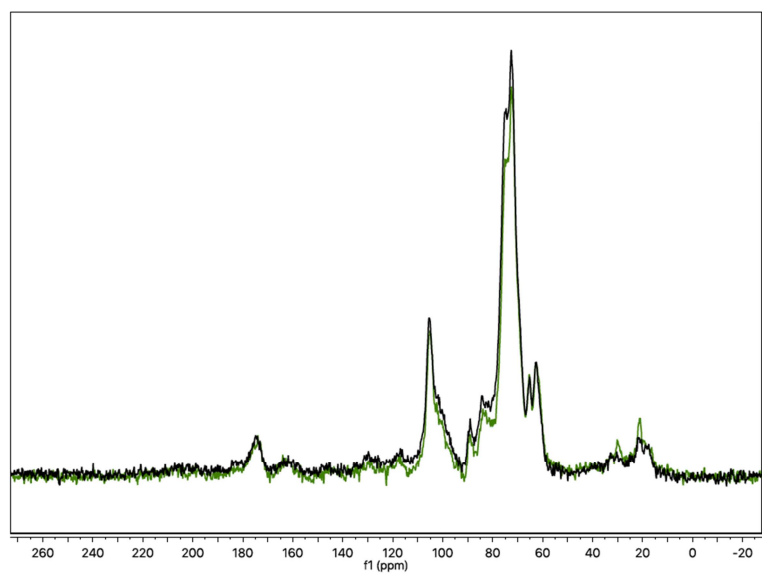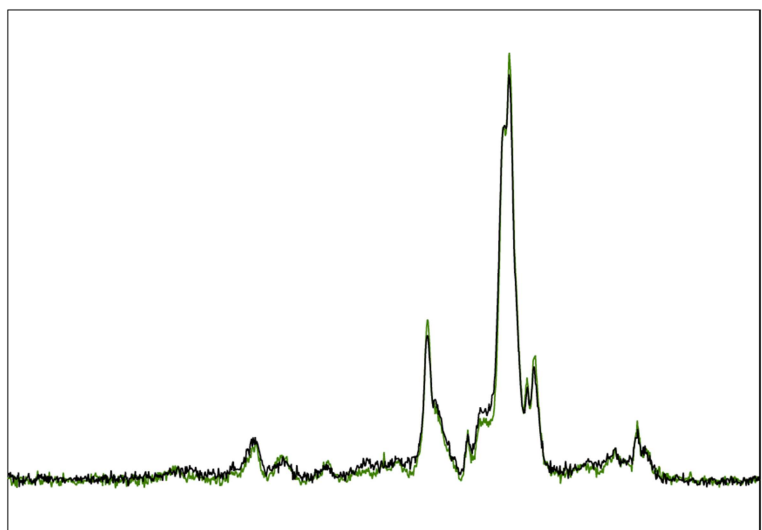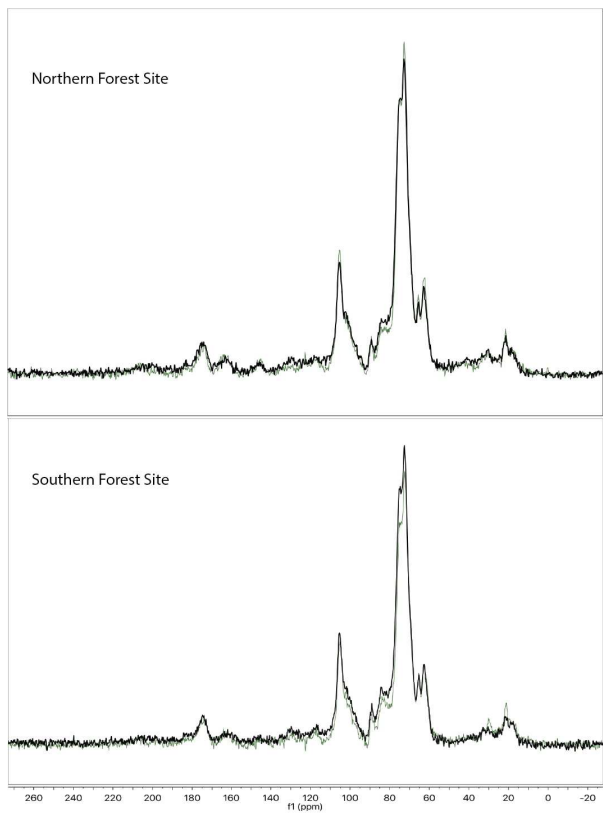
Figure 6. Examples of the CPMAS ^{13}C-NMR results for four out of twelve samples analyzed. The four chosen here are from each of the two forest sites with one undecomposed initial moss tissue replicate and the corresponding most decomposed replicate from the final time point of the 18°C incubation. Top panel provides examples from the cooler forest site (SR) and bottom panel the warmer forest site (GC). The initial samples are in green and a final sample from the 18°C incubation in black. The C types and their ppm range are; alkyl (50-0), O-alkyl (90-65), Di-O-alkyl (110-90), aromatic (165-110), carbonyl and amide (190-165).
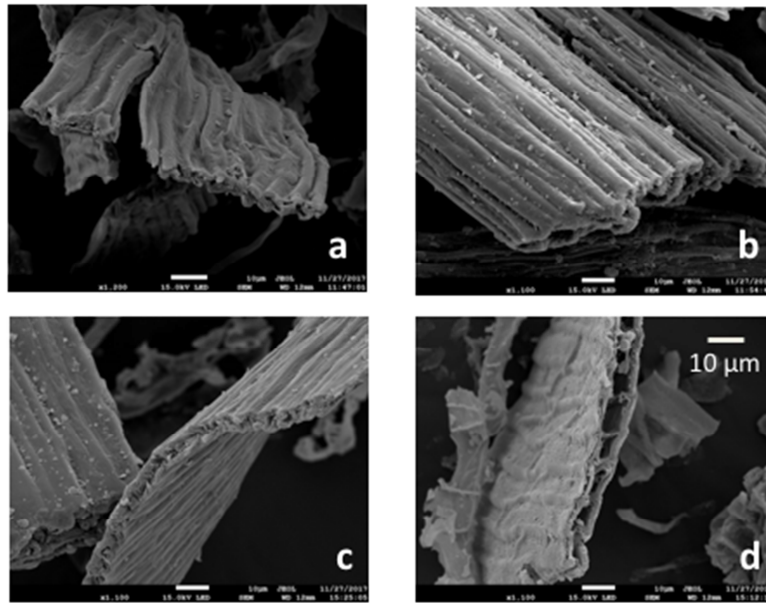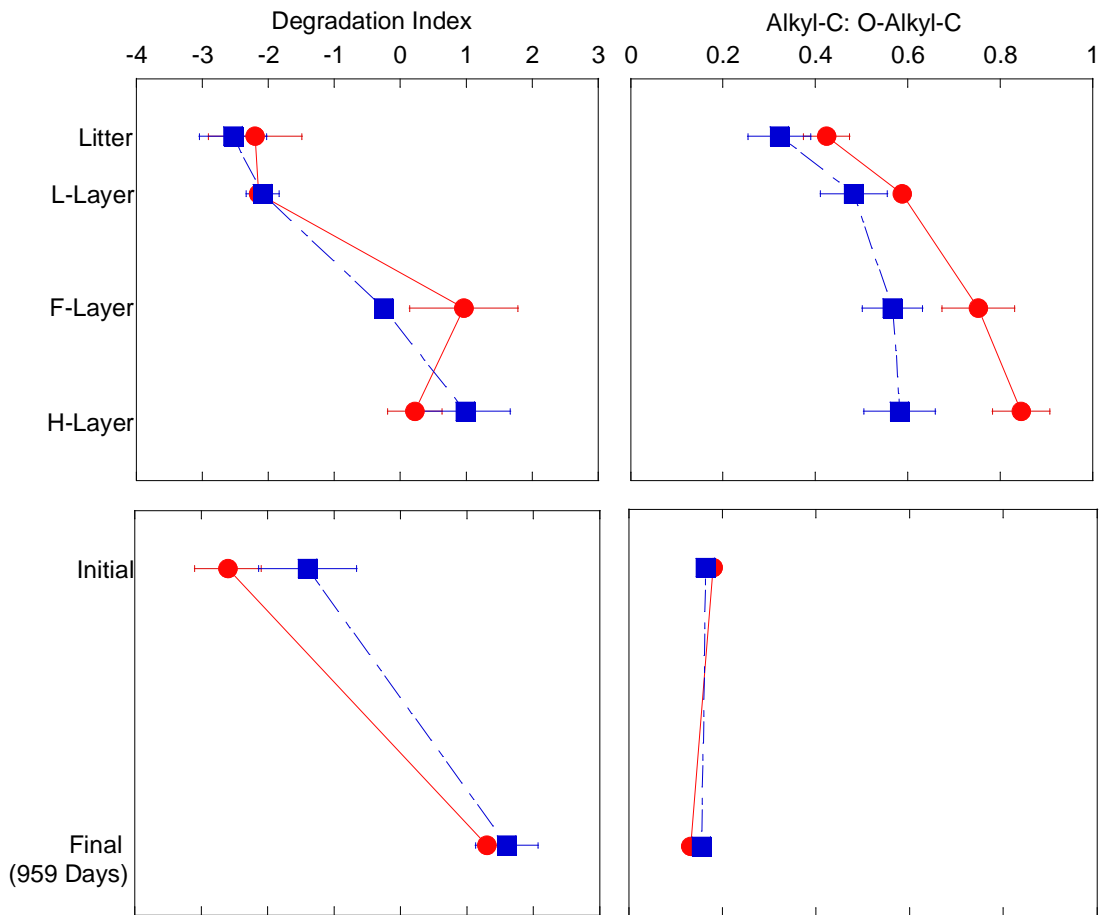
Figure 7. Representative scanning electron micrographs of moss tissues before (a, c) and after (b, d) incubation. The top panels (a and b) depict mosses from the cooler forest, and the bottom panels (c and d) depict mosses from the warmer forest.
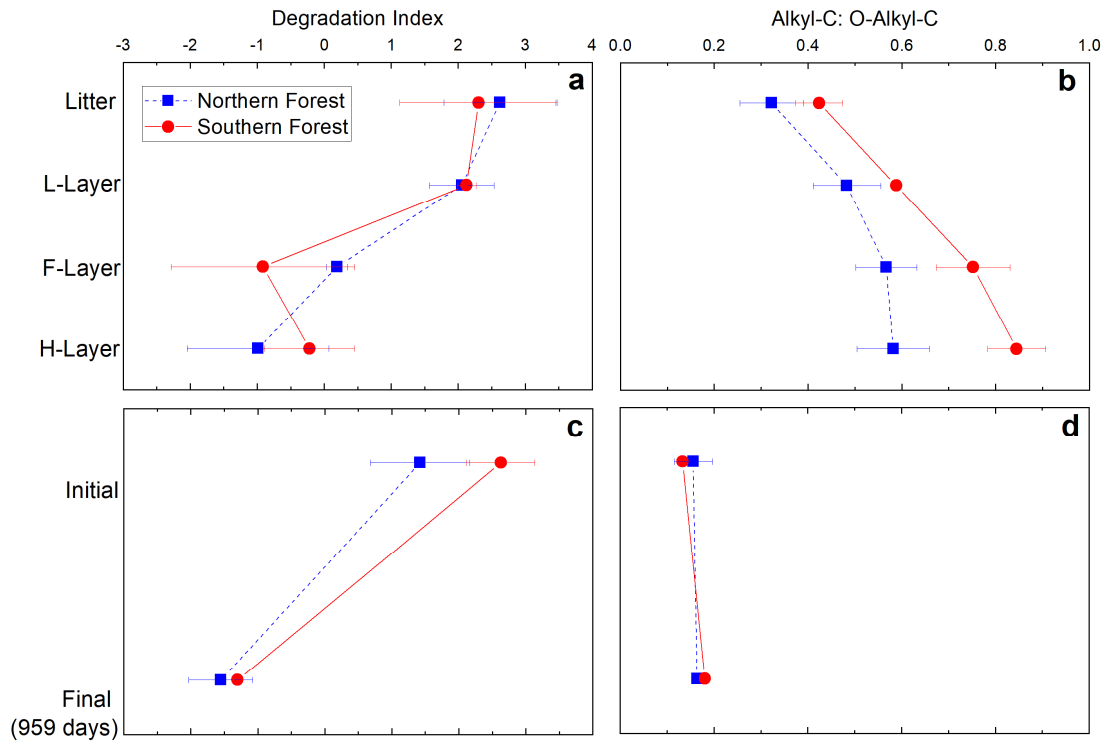
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Figure 8. ~~Top two panels~~Panels a and b show the degradation index and Alkyl-C:OAlkyl-C ratio for the soil profile in the cooler region (blue squares) and warmer region (red circles). Degradation index data from Philben et al (2016). ~~Bottom two panels~~Panels c and d are calculated from this incubation data to contrast two methods for determining level of degradation for mosses. Data points are given as the mean with standard deviation displayed as error bars.

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