

1 **Biogeography and community structure of**
2 **abyssal scavenging Amphipoda (Crustacea) in**
3 **the Pacific Ocean.**

4
5 Patel, Tasnim.^{1,2}, Robert, Henri.¹, D'Udekem D'Acoz, Cedric.³, Martens,
6 Koen.^{1,2}, De Mesel, Ilse.¹, Degraer, Steven.^{1,2} & Schön, Isa.^{1,4}

7
8 ¹ *Royal Belgian Institute of Natural Sciences, Operational Directorate Natural Environment,*
9 *Aquatic and Terrestrial Ecology, Vautierstraat 29, B-1000 Brussels, Gulledele 100, 1000*
10 *Brussels and 3e en 23e linieregimentsplein, 8400 Oostende, Belgium.*

11 ² *University of Ghent, Dept Biology, K.L. Ledeganckstraat 35, B-9000 Ghent, Belgium*

12 ³ *Royal Belgian Institute of Natural Sciences, Operational Directorate Taxonomy &*
13 *Phylogeny, Vautierstraat 29, B-1000 Brussels, Belgium.*

14 ⁴ *University of Hasselt, Research Group Zoology, Agoralaan Building D, B-3590*
15 *Diepenbeek, Belgium.*

16
17 Corresponding author: tpatel@naturalsciences.be

28 **Abstract**

29

30 In 2015, we have collected more than 60,000 scavenging amphipod specimens during two
31 expeditions to the Clarion-Clipperton fracture Zone (CCZ), in the Northeast (NE) Pacific and
32 to the DISturbance and re-COLONisation (DisCOL) Experimental Area (DEA), a simulated
33 mining impact disturbance proxy in the Peru basin, Southeast (SE) Pacific. Here, we compare
34 biodiversity patterns of the larger specimens (> 15 mm) within and between these two
35 oceanic basins. Nine scavenging amphipod species are shared between these two areas, thus
36 indicating connectivity. We further provide evidence that disturbance proxies seem to
37 negatively affect scavenging amphipod biodiversity, as illustrated by a reduced alpha
38 biodiversity in the DEA (Simpson Index (D) = 0.62), when compared to the CCZ (D = 0.73)
39 and particularly of the disturbance site in the DEA and the site geographically closest to it.
40 Community compositions of the two basins differs, as evidenced by a Non-Metric
41 Dimensional Scaling (NMDS) analysis of beta biodiversity. The NMDS also shows a further
42 separation of the disturbance site (D1) from its neighbouring, undisturbed reference areas
43 (D2, D3, D4 and D5) in the DEA. A single species, *Abyssorchomene gerulicorbis*, dominates
44 the DEA with 60% of all individuals.

45 **Keywords**

46

47 JPIO Ecological Aspects of Deep-sea mining, Clarion Clipperton Fracture Zone, CCZ, DisCOL
48 Experimental Area (DEA), Amphipoda.

49

50

51

52

53

54

55

56

57

58

59 **Introduction**

60

61 **The abyssal deep sea** (3000 - 6000 m) represents the largest ecosystem on the planet, with the
62 abyssal seafloor covering approximately 54% of the Earth's solid surface (Rex et al. 1993;
63 Gage & Tyler, 1991). Since it is one of the least investigated ecosystems, there are still
64 extensive gaps in our knowledge of deep-sea fauna (German et al. 2011). Marine research has
65 thus far focused on coastal areas, hydrothermal vents or chemosynthetic habitats, whereas
66 open-ocean abyssal plains have been less extensively investigated (Ramirez-Llodra et al.
67 2010). This is unsurprising given the challenges of sampling this remote environment, which
68 is impeded by several confounding factors. For example, deep-sea sampling is both
69 financially expensive and labour intensive, and furthermore, constrained by the challenge of
70 deploying equipment at low temperatures (0.01 - 4.0°C) and at high hydrostatic pressures
71 (Sweetman et al. 2017). Therefore, to date very little of the deep sea has been sampled, and
72 the oversampling in the North Atlantic basin has created a biased knowledge base (McClain
73 & Hardy, 2010). Consequently, and owing to the low availability of data on deep-sea
74 biodiversity, and with the inherent risk of under-sampling, it is difficult to estimate species
75 richness in the deep sea.

76

77 In the traditional view of the deep sea, the abyss was considered to be homogeneous and
78 many species were thought to have large biogeographical ranges, their dispersal aided by an
79 apparent lack of barriers (Sanders, 1968). This hypothesis was challenged by the discovery of
80 chemosynthetic habitats e.g. hydrothermal vents (Lonsdale, 1977), cold seeps (Paull et al.
81 1984), seasonal fluctuations in primary productivity (Billett et al. 1983) and erratic whale-
82 falls (Smith et al. 1989). All of this research has demonstrated that the deep sea is an
83 extremely heterogeneous environment and is controlled by many factors, including:
84 Particulate Organic Carbon (POC) flux, water depth, flow regime, current circulation,
85 seafloor topography (Laver et al. 1985) and also historical factors e.g. the opening of ocean
86 basins (i.e. rifting), sea-level rise and fall, and periods of deep-sea anoxia (Smith et al. 2006).
87 All of these can result in a mosaic of different communities (Levin et al. 2001), many of
88 which do not follow a latitudinal gradient (Brandt et al. 2007).

89

90 It has also been established that dispersal ability of species on the one hand, and their actual
91 geographic and bathymetric distribution range on the other, are not always linked, and are
92 often dependent on habitat suitability, fragmentation, and ecological flexibility (Lester et al.
93 2007; Liow 2007). Therefore, although the deep-seafloor includes some of the largest
94 contiguous features on the planet, the populations of many deep-sea species are spatially
95 fragmented, and may become increasingly so with continued human disturbance (Hilario et
96 al. 2015).

97

98 In the last decade, there has been an increased demand for exploitation of deep-sea resources
99 e.g. hydrocarbon/rare earth element (REE) extraction (such as those concentrated in
100 manganese nodule provinces) (Ramirez-Llodra et al. 2011). As a result, ecologists are
101 increasingly asked to assess the ecological risks of these mining activities and to provide
102 sustainable solutions for its mitigation, in order to prevent adverse changes to the deep-sea
103 ecosystem (ISA, 2017).

104

105 Glover et al. (2001) showed that abyssal sediments can contain high biodiversity with more
106 than 100 species of meiofaunal invertebrates (e.g. nematodes, copepods) and protists (e.g.
107 foraminifers) found every square meter. In spite of this, our knowledge on the deep-sea
108 ecosystem structure and functioning is still limited, and there is a paucity of data on the
109 distribution, drivers and origins of deep-sea communities at global scales. This is especially
110 true for the biogeography of deep-sea Amphipoda (Barnard 1961; Thurston 1990) and other
111 invertebrates.

112

113 Although recent morphological and molecular studies have shed new light on the distribution
114 and habitat niches of certain benthic-pelagic amphipods (e.g. *Eurythenes*) (Havermans, 2016;
115 Narahara-Nakano et al. 2017), there is little published so far on how widespread other
116 amphipod species may be. This lack of information on species richness and ecological
117 uniqueness hampers the answering of crucial questions on recoverability of anthropogenic
118 impacts. Ultimately this impedes ecologists from providing advice on sustainable deep-sea
119 mining practices, thus, underpinning the need for this dedicated deep-sea ecosystem research.

120

121 Here, we present distribution patterns of scavenging deep-sea amphipod communities, with
122 the first comparisons of their biogeography and community structures in two oceanic basins.

123 These two basins are the research areas for simulating/studying the anthropogenic impacts of
124 deep-sea nodule mining.

125

126 We are investigating whether there are differences and similarities in the species
127 compositions of the two basins (e.g. richness, abundances), and further exploit a disturbance
128 experiment to compare the biodiversity of this mining impact proxy to the undisturbed
129 reference areas. We discuss the possible implications of our findings; aiming to use them to
130 formulate recommendations regarding the pending deep-sea mining of manganese nodule
131 activities in the NE Pacific ecosystem.

132 **Material and Methods**

133

134 *Study area*

135

136 We investigated the amphipod communities of two oceanic basins (Figure 1); (i) the Clarion-
137 Clipperton Fracture Zone (CCZ, six million km², 7000 km wide), an economically important
138 manganese nodule field in the NE Pacific, comprising several different contractor claim
139 areas, (who to date, have exploration licences only), and nine designated Areas of Particular
140 Ecological Interest (APEIs) as designated by the International Seabed Authority (ISA)
141 (Lodge et al, 2014) and (ii) the DISturbance and re-COLONisation (DisCOL) Experimental
142 Area (DEA, 11 km², 4 km wide), a mining disturbance proxy (also containing manganese
143 nodules) in the Peru Basin in the SE Pacific. In 1989, the DEA sediment bed was artificially
144 disturbed using a plough-harrow to create 78 track marks. These are supposed to simulate the
145 type of disruption which would be caused by a commercial mining operation (Appendix 1)
146 (Thiel, 1992). This baseline study was a new approach in deep-sea risk assessment and is still
147 ongoing today, providing us with crucial data from this long-term ecological experiment.

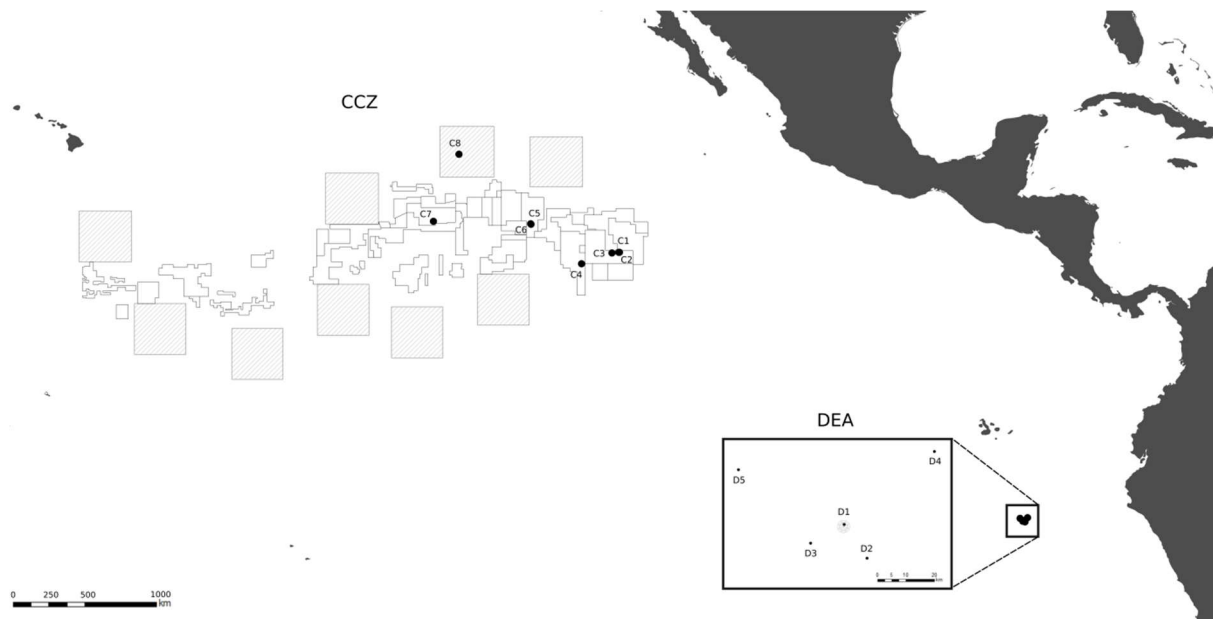
148

149

150

151

152



153

154 **Figure 1: Geographic locations of the two study areas, the Clarion-Clipperton fracture**
 155 **Zone (CCZ) (Northeast Pacific) and the DISTurbance and re-COLonisation (DisCOL)**
 156 **Experimental Area (DEA) (Peru Basin, Southeast Pacific).** There are nine Areas of
 157 Particular Ecological Interest (APEIs) in the CCZ region, which are illustrated by 400 x 400
 158 km² white boxes. Grey boxes indicate the various contractor claim areas in the CCZ. We
 159 deployed eight amphipod traps across the CCZ, which is 16,000 km² and 7000 km wide, and
 160 five in the DEA, which encompasses 11 km² with a width of 4 km.

161

162 *Sampling*

163

164 In 2015 (26 years after the first impact in the DEA in 1989), two research expeditions with
 165 the “RV Sonne” visited the CCZ (cruise SO239) and revisited the DEA (cruise SO242-1 &
 166 SO242-2), to assess if and how the deep-sea faunal communities had recovered within the
 167 DEA, and to attempt to quantify their recolonization potential.

168

169 Amphipod samples were taken from the CCZ and DEA using a free-fall lander (120 x 120 x
 170 120 cm), to which four plastic traps were attached (two 20 x 25 x 40 cm traps with four cm
 171 openings and two 25 x 40 x 60 cm traps with eight cm openings), **baited for each station with**
 172 **an 800 g mixture of mackerel, squid and shrimp.** Using this specially designed deep-sea
 173 sampling equipment, more than 60,000 specimens of scavenging amphipods were collected
 174 from the CCZ and the DEA sites.

175

176 The baited trap was deployed eight times across the CCZ at a depth range of 4116 - 4932 m
 177 (samples C1 - C8), and five times in the DEA at a depth range of 4078 – 4307 m (samples D1

178 - D5; Figure 1, Table 1). In the CCZ, we sampled within three different contractor claim
179 areas (Table 1) to obtain a pre-disturbance baseline, and to then compare it with one of the
180 nine protected APEIs around the CCZ. In contrast, in the DEA, sampling was conducted once
181 within the disturbed area (D1), twice 10 km away (D2, D3) and twice 40 km away (D4, D5)
182 from D1 in four surrounding reference areas (see Figure 1).

183
184
185
186
187
188
189
190
191
192
193
194
195
196
197
198
199
200
201
202
203
204
205

206 **Table 1: Station overview.**

207 Codes refer to the codes used in this paper in figures 1, 3, 4, 6 and Table 3. The original
 208 station code represents the cruise codes from (SO239 and SO232-1). Depth refers to water
 209 depth (m) on deployment. Nodule presence/absence information is known only for stations
 210 D3 and D4.

Deployment Code	Original Station Code	Depth (m)	Known geological features	Remarks	Residence Time (h:m:s)
C1	SO239-33	4122	Plains	German claim	100:20:00
C2	SO239-37	4116	Plains	German claim	125:49:00
C3	SO239-63	4354	Plains	German claim	66:07:00
C4	SO239-96	4388	Seamount	Inter Ocean Metals (IOM) claim	63:02:00
C5	SO239-123	4529	Plains	Belgian claim	26:12:00
C6	SO239-139	4516	Plains North/South + seamount to west	Belgian claim	56:33:00
C7	SO239-173	4934	Plains	French claim	79:40:00
C8	SO239-205	4855	Plains	Area of Particular Ecological Interest (APEI)	55:59:00
D1	SO242/1-8	4146	Plains	Disturbed	44:26:33
D2	SO242/1-30	4307	Plains	Undisturbed Reference	51:11:18
D3	SO242/1-55	4043	Seamount No nodules	Undisturbed Reference	25:09:09
D4	SO242/1-68	4078	Seamount No nodules	Undisturbed Reference	65:20:46
D5	SO242/1-106	4269	Plains	Undisturbed Reference	47:00:50

211
 212
 213
 214
 215
 216
 217
 218

219 ***Processing***

220

221 On recovery of the lander, all traps were disconnected and placed in pre-cooled (4°C) buckets
222 of filtered seawater. All specimens were washed on board in a cool-climate laboratory (4°C),
223 morphologically pre-sorted and fixed in molecular grade (95%) ethanol, before being stored
224 at -20°C.

225

226 Detailed sorting and identification was performed using the morphological species concept
227 (Futuyama, 1998) and the keys of Lowry & Killagen (2014) and Schulenberger & Barnard
228 (1976), to separate the samples into taxonomic “morphotypes”. The larger fraction (> 15 mm
229 length) has been identified to the lowest possible taxonomic resolution. Species not assigned
230 with certainty are denoted here by as affiliated species (e.g. *genus* aff. *species*) or conferred
231 species (e.g. *genus* cf. *species*).

232

233 Specimens with a size of less than 15 mm length were excluded from the analysis, primarily
234 because these were mostly juveniles, and their morphological differences were not
235 sufficiently pronounced to allow an accurate identification to the species or even genus level.
236 Also, all pelagic amphipod specimens were omitted which were accidentally caught when the
237 sampling equipment was retrieved to the sea surface. Genera containing multiple and as yet
238 unidentified species have been summarised as “spp.”

239

240 ***Statistical analyses***

241

242 Our null hypothesis (H_0) here is that there are no differences in the amphipod biodiversity of
243 the two basins. To test this hypothesis, we firstly calculated the alpha biodiversity of the two
244 basins was using the Simpson Index (D) (Simpson, 1949) (Appendix 4). Individual-based
245 rarefaction curves were generated using the rarefy function of the vegan package in R 2.3.0
246 (R Core Team, 2013; Gotelli, 2001) to compare species richness across all thirteen sampling
247 stations and to test for the completeness of sampling.

248

249 Secondly, to compare the beta biodiversity, we estimated the variability of the community
250 compositions between sites. The Bray-Curtis dissimilarity metric (Bray & Curtis, 1957), was
251 used to calculate differences between community compositions based on species densities,
252 and the results were then visualised in 2D using a Non-Metric Dimensional Scaling (NMDS)

253 plot. The ANOSIM function in the vegan package of R (R Core Team, 2013; Taguchi & Ono,
254 2005) was used to test the statistical significance of the differences in species compositions
255 between the two study areas.

256

257

258 **Results**

259

260 ***Basin biodiversity***

261

262 In total, 6916 scavenging amphipods (> 15 mm) were collected from the thirteen trap
263 deployments in the two study areas, representing nineteen morphotypes (Figure 2). In the
264 CCZ, we collected 3932 individuals, which represent ten morphotypes. Five of these have
265 been identified to the species level: *Abyssorhomene distinctus*, *A. gerulicorbis*, *Eurythenes*
266 *sigmiferus*, *Paralicella caperesca* and *Valettietta tenuipes*. A further two are affiliated to a
267 species: *Paracallisoma* aff. *alberti* and *Valettietta* cf. *gracilis*, and the remaining three are at
268 least affiliated to a genus (Tables 2a and 2b). The 2984 individuals from the DEA represent
269 eighteen morphotypes. Six of these have been identified to the species level: *Abyssorhomene*
270 *distinctus*, *A. gerulicorbis*, *Eurythenes sigmiferus*, *Paralicella caperesca*, *Parandaniexis*
271 *mirabilis* and *Tectovallopsiis regelatus*. A further five which have been affiliated to a species:
272 *Eurythenes* sp. 2. aff. *gryllus*, *Eurythenes* sp. 4. aff. *magellanicus*, *Paracallisoma* aff. *alberti*,
273 *Stephonyx* sp. nov. aff. *arabiensis* and *Valettietta* cf. *gracilis* and the remaining seven
274 identified to at least an affiliated genus (Tables 2a and 2b).

275

276 There are nine morphotypes shared between the basins: *Abyssorhomene distinctus*, *A.*
277 *gerulicorbis*, *Abyssorhomene* spp., *Eurythenes sigmiferus*, *Eurythenes* spp. nov.,
278 *Paracallisoma* aff. *alberti*, *Paralicella caperesca*, *Parandania* sp. and *Valettietta* cf. *gracilis*
279 (Figure 2).

280

281 Two morphotypes were found only in the CCZ (*Hirondellea* sp. & *Valettietta tenuipes*), and
282 eight morphotypes were found only in the DEA (*Eurythenes* sp. 1-4, gen. aff. *Cleonardo*,
283 *Parandaniexis mirabilis*, *Stephonyx* sp. nov. aff. *arabiensis*, and *Tectovallopsiis regelatus*)
284 (Table 2).

285

286

287

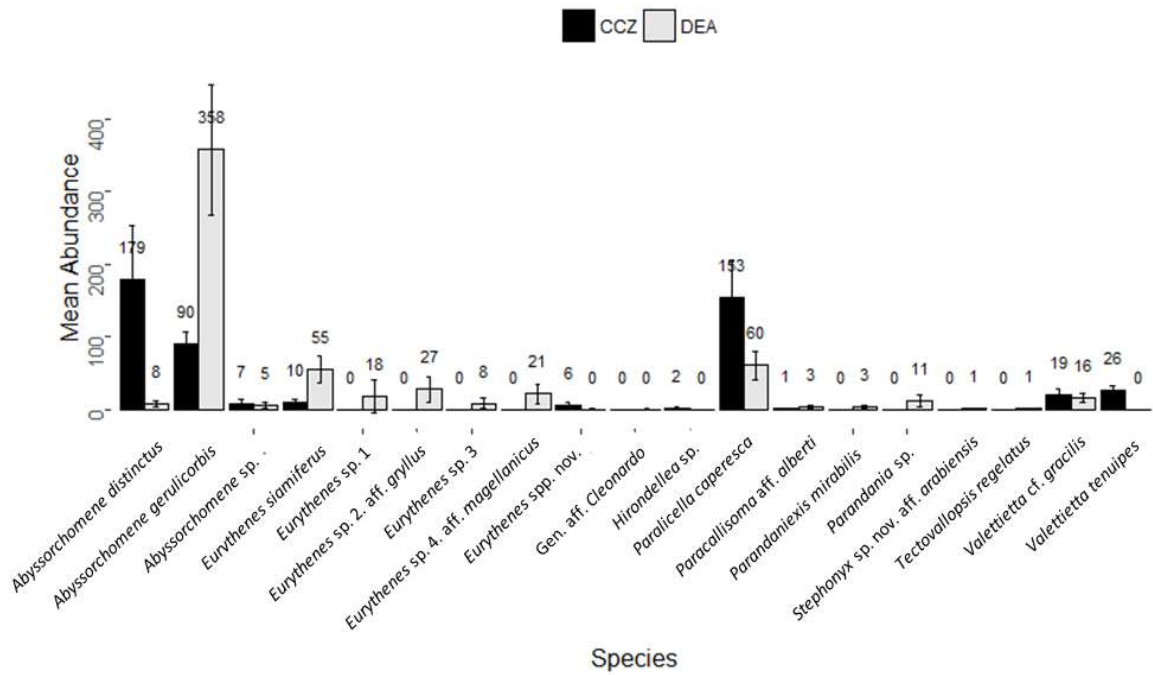
288

289

290

291

Amphipod Trap Biodiversity in the CCZ & DEA



292

293

294 **Figure 2: Histogram showing the species assemblage for the scavenging community in**
295 **the Clarion-Clipperton fracture Zone (CCZ) (black) and the DisCOL Experimental**
296 **Area (DEA) (grey). The abundances of 19 morphotypes are shown.**

297

298

299

300

301

302

303

304

305

306

307

308

309

310

311 **Table 2a: Overview of morphotypes across the Clarion-Clipperton fracture Zone (CCZ)**
 312 **and DisCOL Experimental Area (DEA).**

	Clarion-Clipperton fracture Zone	DisCOL Experimental Area
Total unique morphotypes collected	19 (10 found in the CCZ, 18 found in the DEA)	
Species possibly unique to this basin	2	8
Shared species between basins	9	

313
314

315 **Table 2b: Distribution and abundances of morphotypes across the Clarion-Clipperton**
 316 **fracture Zone (CCZ) and DisCOL Experimental Area (DEA). Green = shared, Yellow =**
 317 **DEA only, Blue = CCZ only.**

318

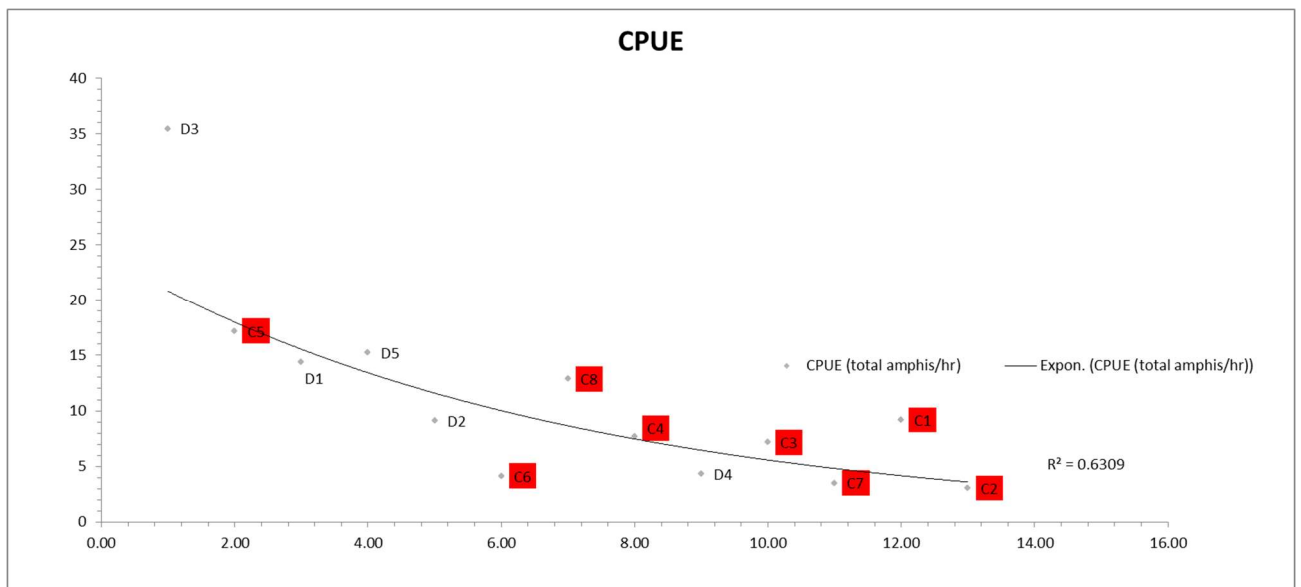
Species	C1	C2	C3	C4	C5	C6	C7	C8	D1	D2	D3	D4	D5
<i>Abyssochomene distinctus</i>	629	312	180	170	64	47	2	25	9	3	25	0	5
<i>Abyssochomene gerulicorbis</i>	73	47	48	107	71	65	184	121	351	143	522	178	595
<i>Abyssochomene spp.</i>	0	0	50	0	0	3	0	0	5	20	0	0	0
<i>Eurythenes sigmiferus</i>	9	3	35	11	12	5	0	6	30	61	127	36	22
<i>Eurythenes</i> sp. 1									0	90	0	0	1
<i>Eurythenes</i> sp. 2. aff. <i>gryllus</i>									119	0	9	0	9
<i>Eurythenes</i> sp. 3									0	0	3	39	0
<i>Eurythenes</i> sp. 4 aff. <i>magellanicus</i>									0	0	59	0	47
<i>Eurythenes</i> .spp.nov.	6	3	2	0	0	20	1	12	0	1	0	0	0
gen. aff. <i>Cleonardo</i>									1	0	0	0	0
<i>Hirondellea</i> sp.	0	2	0	0	0	0	5	10					
<i>Paracallisoma</i> aff. <i>alberti</i>	0	0	0	0	2	1	6	10	4	0	1	1	1
<i>Parallicella caperesca</i>	104	4	114	152	255	75	63	460	86	108	80	21	7
<i>Parandania</i> sp.									5	2	42	5	1
<i>Parandaniexis mirabilis</i>									11	0	3	0	0
<i>Stephonyx</i> sp. nov. aff. <i>arabiensis</i>									0	4	0	0	0
<i>Tectovallopsis regelatus</i>									5	0	0	0	0
<i>Valettieta</i> cf. <i>gracilis</i>	75	11	29	3	2	5	1	23	2	29	17	1	29
<i>Valettieta tenuipes</i>	22	0	14	42	43	9	19	58					

319
320
321
322
323
324
325

326
327
328
329
330
331
332
333
334
335
336
337

Sampling completeness

Due to differences in allocated ship-times (CCZ cruise being 52 days and the DEA cruise being 29 days), the trap deployments were not identical, making it necessary to normalise deployment times. The resulting Catch Per Unit Effort (CPUE) plot (Figure 3,) shows that, with the exception of C5, all stations in the DEA yielded higher abundances/hr than the CCZ. The highest numbers of individuals/hr were collected at station D3. Overall, there is a moderate negative exponential correlation with increasing deployment times ($R = 0.67, p = 0.01$).

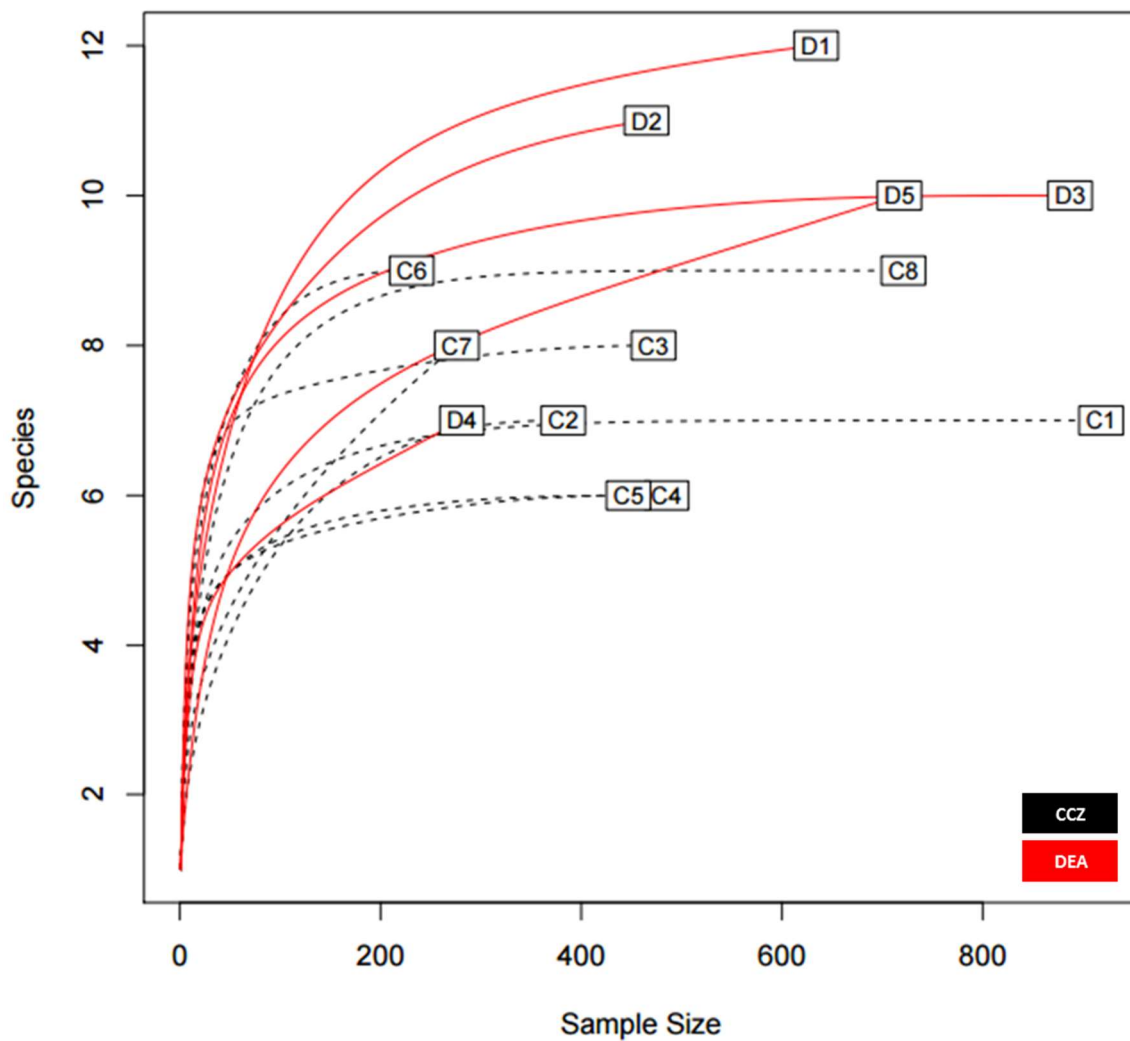


338
339
340
341
342
343
344
345
346
347
348
349

Figure 3: Catch Per Unit Effort (CPUE) illustrating the correlation between sampling time and number of individuals collected. Only the > 15mm fraction was included here to estimate number of collected individuals.

350
351
352
353
354
355
356

The rarefaction results (Figure 4) show that the curves for nine stations reach a plateau, indicating that sampling effort was sufficient to assess diversity levels. These include all CCZ stations except C7. In contrast, four of the five curves for the DEA (stations D1, D2, D4 and D5) are unsaturated.



357
358
359
360
361
362
363

Figure 4: Species rarefaction curves for each of the 13 trap stations across both areas, the Clarion-Clipperton fracture Zone and the DisCOL Experimental Area. Only individuals greater than 15 mm were considered here.

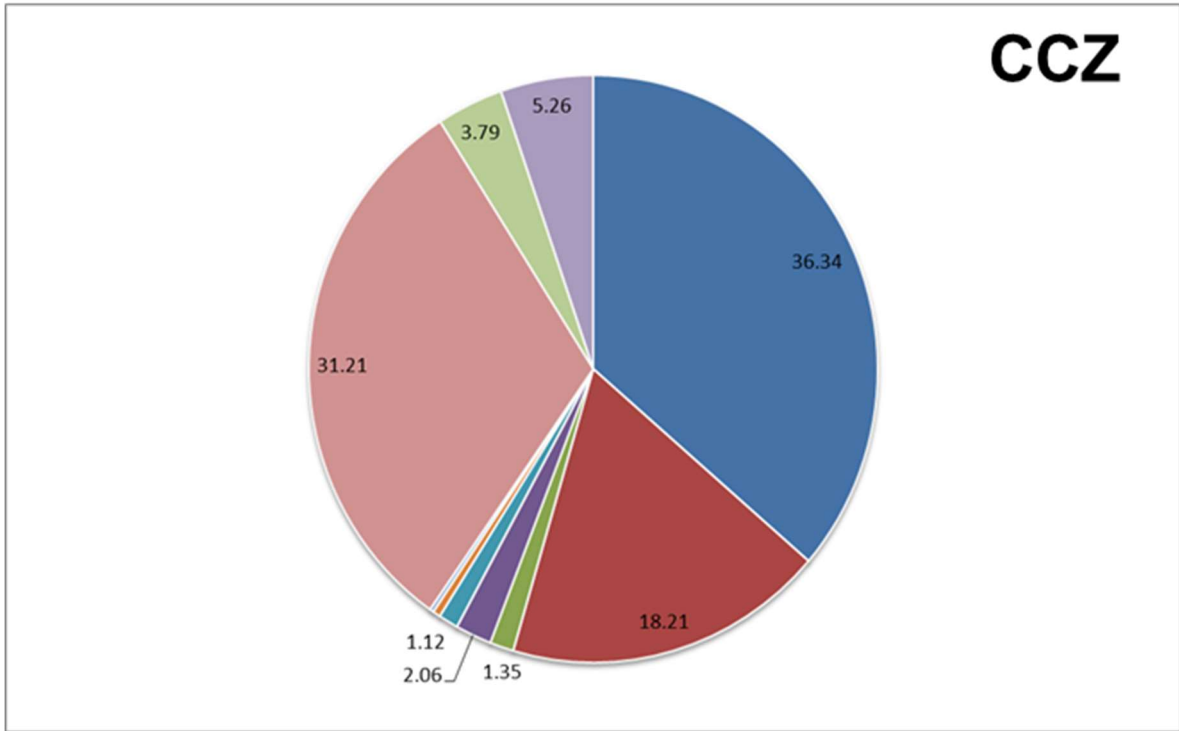
364
365
366
367
368
369
370
371
372
373
374
375
376
377
378
379
380
381
382
383
384
385
386
387
388
389
390
391
392
393
394
395
396
397

Biodiversity

Figures 5a and b show that the scavenging community in the CCZ is dominated by three species, *A. distinctus* (16%), *A. gerulicorbis* (18%) and *Paralicella caperesca* (31%), whereas, in contrast, the DEA scavenging community is dominated by a single species, *A. gerulicorbis*, accounting for almost 60% of all specimens. The Simpson Index (D) for the entire CCZ area is (with 0.73), higher than the 0.616 that was calculated for the whole of the DEA area (Table 3). The biodiversity of each individual station was further explored (Table 3). In the CCZ, the lowest biodiversity was found at C3 and C6 ($D = 0.23$) and the highest at C2 ($D = 0.67$), respectively. In the DEA, the lowest biodiversity of $D = 0.36$ was found at station D1 (the site of the actual disturbance) and just south of the disturbance site at D2 (0.21), while the highest biodiversity was observed at D5 ($D = 0.61$) (Table 3).

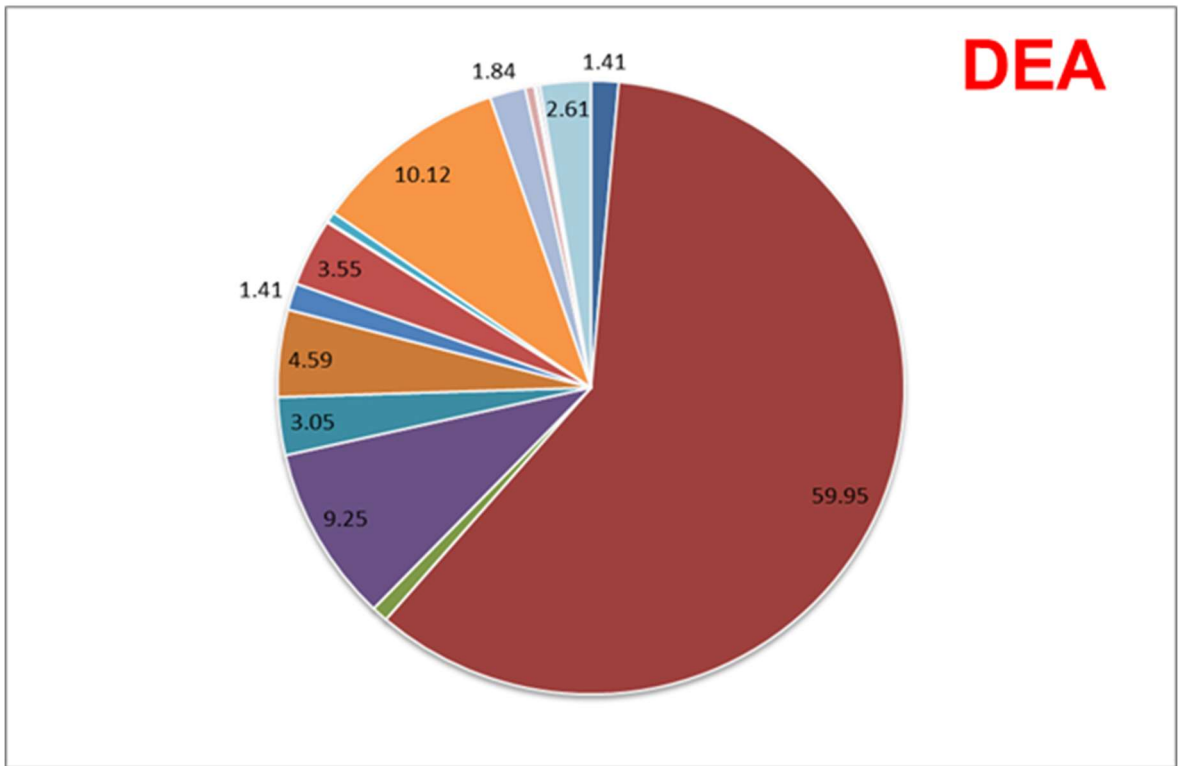
398

399



400

401



402

403

404

- *Abyssorchomene distinctus*
- *Abyssorchomene gerulicorbis*
- *Abyssorchomene* sp.
- *Eurythenes sigmiferus*
- *Eurythenes* sp. 1
- *Eurythenes* sp. 2 (aff. *gryllus*)
- *Eurythenes* sp. 3
- *Eurythenes* sp. 4 (aff. *magellanicus*)
- *Eurythenes* spp. nov.
- gen. aff. *Cleonardo*
- *Hirondellea* sp.
- *Paracallisoma* aff. *alberti*
- *Paralicella caperesca*
- *Parandania* sp.
- *Parandaniexis mirabilis*
- *Stephonyx* sp. nov. aff. *arabiensis*
- *Tectovallopsiopsis regelatus*
- *Valettietta gracilis*
- *Valettietta tenuipes*

405

406

407 **Figures 5a and b: Relative species abundances in the Clarion-Clipperton fracture Zone**

408 **and the DisCOL Experimental Area.** These abundances represent the greater than 15mm

409 fraction of the scavenging amphipod community only.

410

411

412

413

414

415

416

417

418

419

420

421

422 **Table 3: Comparison of biodiversity calculated using the Simpson Index (*D*), for the**
 423 **Clarion-Clipperton fracture Zone and DisCOL Experimental Area, and *D* for each**
 424 **station is shown for further comparisons within these areas.**

Simpson Index (<i>D</i>) whole Clarion-Clipperton Fracture Zone	0.73
Simpson Index (<i>D</i>) whole DisCOL Experimental Area	0.62
C1 (<i>D</i>)	0.41
C2 (<i>D</i>)	0.68
C3 (<i>D</i>)	0.23
C4 (<i>D</i>)	0.27
C5 (<i>D</i>)	0.38
C6 (<i>D</i>)	0.23
C7 (<i>D</i>)	0.45
C8 (<i>D</i>)	0.44
D1 disturbed (<i>D</i>)	0.36
D2 ref 1 (<i>D</i>)	0.21
D3 ref 2 (<i>D</i>)	0.38
D4 ref 3 (<i>D</i>)	0.44
D5 ref 4 (<i>D</i>)	0.61

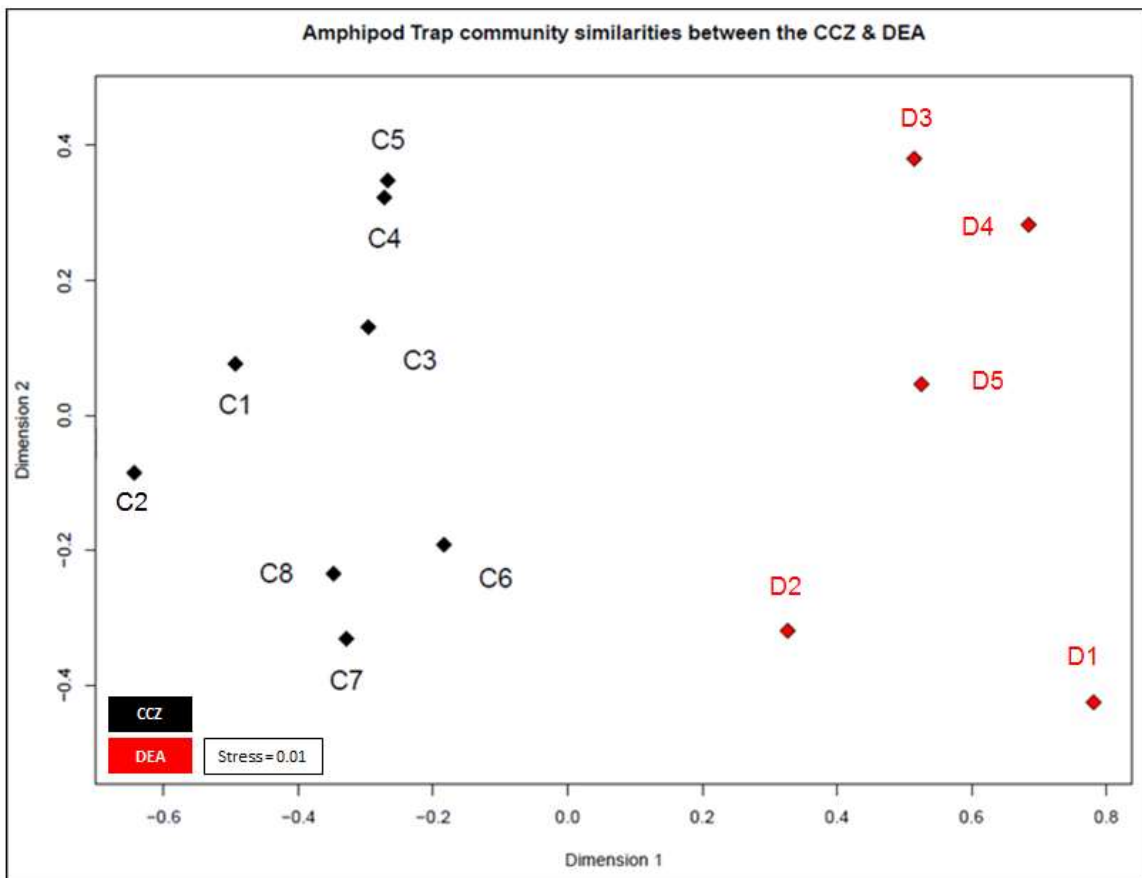
425
 426
 427
 428
 429
 430
 431
 432
 433

434 **Species composition**

435

436 The NMDS shows that the communities of the two basins are clearly separated (ANOSIM: p
437 = 0.002); Figure 6). The disturbed area in the DEA (D1) is showing a clear difference to the
438 four reference areas (D2 - 5). When the communities between the two basins are compared,
439 D2 appears to be most similar to the CCZ community, and more specifically to C6, C7 and
440 C8. The reliability of the data ranking is supported by a low stress value of 0.01.

441



442

443 **Figure 6** : NMDS plot showing the beta biodiversity (dis/similarities) for each of the thirteen
444 amphipod trap sampling stations associated with the two basins, Clarion-Clipperton fracture
445 Zone (CCZ) (black) and the DisCOL Experimental Area (DEA) (red). Data are supported by
446 a low stress value of 0.01.

447

448

449 Discussion

450

451 *An unexplored ecosystem*

452 Although the most recent and comprehensive analysis of the Animal diversity of the World's
453 oceans estimates a total of less than a million species over all depths (Appeltans et al. 2012),
454 it is not currently known how many species actually inhabit the deep-sea. Regarding
455 amphipods, only 328 benthic, demersal and benthopelagic species, belonging to 144 genera
456 and 39 families. Over 7000 marine amphipod species have been found below 2000 m. These
457 numbers are reduced to 173 known species, 87 genera and 37 families at depths below 3000
458 m, and 100 known species, 66 genera and 31 families are known to occur below 4000 m
459 (Vader 2005; Brandt et al, 2012).

460

461 *Lysianassoidea and their biogeography*

462

463 The superfamily Lysianassoidea constitutes an important part of the abyssal amphipod fauna.
464 Also, in our sampling, lysianassoid amphipods were collected in large numbers (99% of the
465 samples taken in both basins). As a superfamily, they comprise 23% of all the species found
466 below 2000 m, 35% of the species found below 3000 m and 31% of the species found below
467 4000 m (Brandt et al. 2012).

468

469 Many species in the Lysianassoidea occur in multiple abyssal basins, and some even have
470 worldwide distributions (Thurston 1990). Despite the Ocean Biogeographic Information
471 System (OBIS) database containing 615,650 records of Amphipoda, many of these are shelf
472 or pelagic species, with very few records from the CCZ and DEA (OBIS, 2017). Here, we
473 provide new data for the known bathymetric range of the seven amphipods which we have
474 identified to species level (*Abyssorchomene distinctus*, *Abyssorchomene gerulicorbis*,
475 *Eurythenes sigmiferus*, , *Paralicella caperesca*, *Parandaniexis mirabilis*, *Tectovallopsis*
476 *regelatus* & *Valettietta tenuipes*) (Table 2b). In addition, we have found two possibly new
477 species of *Eurythenes*, previously not known from these basins, which we plan to analyse
478 further in the future.

479 While we only sampled *Hirondellea* sp. and *Valettietta tenuipes* in the CCZ, and *Eurythenes*
480 sp. 1- 4, gen. aff. *Cleonardo*, *Parandaniexis mirabilis*, *Stephonyx* sp. nov. aff. *arabiensis*, and

481 *Tectovallopsis regelatus* only in the DEA, we cannot conclude based on the current data only
482 if these species are unique to their respective basins without confirming these distribution
483 patterns with additional sampling campaigns.

484

485

486 ***Catch Per Unit Effort***

487

488 Despite the sampling campaign in the CCZ being twice as long as the DEA, the number of
489 individuals/species collected does not correlate positively with deployment effort. We assume
490 that this is rather an effect of abiotic and organic factors, such as the productivity-driven
491 gradients in the CCZ, which decrease from East-West and from North-South (Hannides &
492 Smith, 2003). This lack of correlation is supported by our findings for station C2 (with the
493 shortest deployment time), which shows the highest Simpson Index of all thirteen stations (D
494 = 0.67). Further evidence comes from the patterns visualised in Figure 3, which shows a
495 correlation of $R = 0.67$ for Catch Per Unit Effort (CPUE) and deployment times.

496

497 ***Biodiversity within basins***

498

499 Figures 5a & b show clearly that the DEA scavenging community has reduced abundances of
500 all species including *A. distinctus* (1%) and *P. capresca* (7%), and is now dominated by a
501 single species, *A. gerulicorbis*, accounting for 60% of the DEA community. This indicates an
502 interesting resilience and flexibility in the latter species. Similar patterns have been observed
503 in Potter Cove (Seefeldt et al. 2017), where following glacial retreat, a change in
504 sedimentation rates led to the dominance of a single amphipod scavenging species,
505 *Cheirimedon femoratus*.

506

507 Although the assemblage of the two basins has some overlap in its amphipod diversity (as is
508 exemplified by the nine shared morphotypes), the sampling stations (and the two basins) are
509 heterogeneous concerning species compositions of the subdominant and rarer species

510

511 Thus, we can observe some negative influence (possibly attributed to the disturbance in the
512 DEA) on the scavenging amphipod community. This reduced biodiversity is reflected in the
513 higher Simpson Index (D) for the CCZ (0.73) as compared to the DEA ($D = 0.62$; Table 3).

514

515 To explore whether this reduced diversity in the DEA was truly a result of the simulated
516 disturbance, D was also calculated for each sampling station within each basin (Table 3).
517 In the CCZ, the APEI (C8) shows a moderate level of biodiversity ($D = 0.44$), indicating that
518 it is not optimally-placed for representing the biodiversity of the scavenging amphipod
519 community of the CCZ. Additionally, this pre-existing lower biodiversity (in comparison to
520 the contractor claim areas), indicates that the APEI may not serve well as a refugium for
521 amphipods post-disturbance. However, due to the fact that only one of the eight APEIs have
522 been investigated thus far, this APEI along with the remaining eight APEIs would need to be
523 (re-) sampled.

524

525 Within the DEA, the lowest biodiversities are observed at the site of the disturbance (D1) and
526 south of it (D2; Table 3), indicating that the reduced biodiversity in the DEA could indeed be
527 caused by the simulated disturbance in 1989 (Thiel, 1992).

528

529 The highest abundances in the DEA were collected from station D5 ($n = 1242$); this station
530 also has the highest Simpson Index within the DEA ($D = 0.61$) (Table 3). Side-scan sonar
531 imaging shows a seamount range to the North West (NW) of the disturbed area (D1)
532 (Appendix 3). Although the relief change is only 150m, the range extends laterally for several
533 kilometres (SO242-1 Cruise report, 2016) hampering dispersal across barriers such as sills,
534 canyons and ridges (Smith, et al. 2006; Blankenship et al. 2006; Etter et al. 2011). However,
535 recent studies have shown that due to their mobile nature, geographic isolation alone would
536 not pose a true barrier to benthic-pelagic amphipod species (Havermans, 2016; Ritchie et al.
537 2017), and thus, cannot explain why such a high number of large scavenging individuals was
538 collected at station D5.

539

540

541 *Community similarities*

542

543 Scavenging amphipods are resilient and dispersive, but most importantly, they are highly
544 mobile (Ingram and Hessler 1983; Lörz et al. 2018). Often driven by their search for erratic
545 deposited feeding opportunities (Smith et al. 1989), they are probably less constrained by
546 local environmental abiotic conditions and seafloor topography.

547

548 Beta diversity can be regarded as the dissimilarities in species composition between spatially
549 different communities. As an indication for beta biodiversity, the NMDS (Figure 6) shows a
550 significant separation in the similarity index between the two basins (ANOSIM $p = 0.002$).
551 However, despite the dispersive and resilient nature of scavenging amphipods, their
552 biodiversity appears to have been affected by the disturbance experiment as evidenced by the
553 NMDS (Figure 6), where the disturbed area (D1) and the area closest to it (D2) are separated
554 from the remaining three reference sites (D3, D4 and D5).

555

556 In the CCZ, stations C1, C2, C3, C4 and C5 form one cluster in the NMDS (Figure 6), and
557 stations C6, C7 and C8 a second cluster. The CCZ is a geomorphologically very
558 heterogeneous region, with seamounts of 200 m altitude running from north-south. A barrier
559 of this height would be expected to affect sedimentation rates, nodule presence and currents.
560 Furthermore, the difference in depth from the eastern edge (3950m) and the western edge
561 (5150 m) is more than 1200 m. These combined factors very likely give rise to different
562 trends in species composition (Glover, et al. 2015). However, since it has been established
563 that benthic-pelagic amphipods are less sensitive to such barriers (Havermans, 2016), at this
564 stage, other biotic (e.g. the productivity gradient) and abiotic factors causing this separation
565 cannot be excluded as alternative explanations.

566

567 *Dispersal and connectivity*

568

569 Whilst the NMDS (Figure 6) illustrates a visual separation of the two basins, there is also
570 some similarity in the amphipod fauna between the two areas, (as is obvious by the nine
571 shared species), indicating that the dispersal extent for these nine species might range up to at
572 least 3000 km. However, this hypothesis will need to be confirmed with subsequent
573 molecular analyses.

574

575

576 Abyssal amphipods have been shown to be able to travel actively at speeds of almost 4
577 cm/sec (Laver, 1985), even at temperatures as low as 3°C (Kankaanpää et al. 1995). It is
578 obvious that they are sufficiently strong to swim up weak currents since they can be found
579 several hundred meters above the seafloor searching pelagically for mates (e.g. *Eurythenes*
580 *gryllus* occurring up to 1800 m above the seafloor) (Thurston 1990; Havermans et al. 2013)
581 or following food-falls (Baldwin and Smith 1987).

582

583 However, it is apparent that the dispersal of abyssal amphipods is not always contingent on
584 current direction, but also on passive dispersal. Amphipods can also be carried passively over
585 long distances by stronger currents e.g. (the circumpolar current of the Southern Ocean)
586 (Laver et al. 1985), but even weaker deep-sea currents have been suggested as a mechanism
587 for deep-sea dispersal of amphipods (e.g. *Eurythenes gryllus* (Schüller and Ebbe 2007)). This
588 coupled with their ability to follow odour-plumes (Ide et al. 2006; Premke, 2003),
589 significantly increases the probability and extent of their dispersal (Conlan 1991; Highsmith,
590 1985).

591

592 The lack of a clear dispersal pattern is obvious from Figure 6, where station D2 is the station
593 clustering closest with the CCZ basin in terms of species composition despite the fact that
594 station D5 is geographically the shortest distance away from the CCZ.

595

596 Recent research on *Eurythenes gryllus* has demonstrated that it thrives in every ocean with a
597 wide (pelagic – hadal) depth range. However, assumptions that individual morphotypes of
598 this species belong to the same genetic lineage have been challenged (Havermans et al. 2013
599 & Havermans, 2016). Ritchie et al. (2016) demonstrated with microsatellite markers
600 heterozygote deficiency in *Paralicella tenuipes*, which they attributed to cryptic diversity. It
601 is likely that the connectivity of the nine species we observed as being shared between the
602 CCZ and DEA (based on traditional morphological methods), can be explained by the
603 presence of one or more cryptic amphipod species, which will have to be tested by future
604 molecular research.

605

606 Unfortunately, in the absence of data on deep-sea currents in the study area, especially
607 between the CCZ and DEA, it is not yet possible to fully explain the drivers and mechanisms
608 of amphipod dispersal between these particular basins.

609

610 *The DisCOL Experimental Area as a proxy*

611

612 Higher abundances of scavenging amphipods were collected from the CCZ (3932
613 individuals) as opposed to the DEA (2984 individuals). Yet, we have identified more
614 morphotypes in the DEA (18) than in the CCZ (10), indicating that the DEA is more
615 speciose, and thus, more biodiverse.

616

617 Although the DEA is more speciose, many of its morphotypes were collected in low
618 abundances, with several of these being singletons or doubletons (collected from one or two
619 sampling stations only). This is reflected in the rarefaction curves (Figure 4), which indicate
620 thorough sampling in the CCZ with all but station C7 reaching asymptotes. In contrast, four
621 stations in the DEA (D1, D2, D4 and D5) are **unsaturated. This pattern suggests firstly** that
622 the less abundant species which are present at fewer stations only may not necessarily be rare
623 species and secondly, that there could be as yet undetected biodiversity in the DEA.

624 Therefore, the effects of mining impact could be even more pronounced than we observed in
625 this study. However, as the seafloor environment is subject to seasonal fluctuations (Billett et
626 al. 1983), it is hard to predict exactly what the effects will be at this stage.

627

628 Our preliminary (basin-scale) comparison of the scavenging communities of the two study
629 areas shows that even if the DEA is a small-scale disturbance experiment, it is a very diverse
630 area. Thus, the DEA is a well-chosen site for monitoring the impacts of disturbance and
631 instrumental in its role as a proxy to assess impending mining activities in the CCZ.

632

633

634

635 ***Future research***

636

637 At several stations in both basins, we collected amphipods in very high abundances (C1, C8,
638 D3 & D5) (Table 2b). Since biotic production is contingent on the sinking flux of particles
639 from the euphotic zone (Sweetman, 2017), the biodiversity differences at each of the thirteen
640 stations could be driven by Particulate Organic Carbon (POC) or erratic whale-falls (Smith et
641 al. 1989). **However, not all feeding behaviour of scavenging amphipods is based on**
642 **opportunistic or erratic availability of nutrients (Havermans & Smetacek, 2018). During**
643 **future sampling campaigns, the POC of these areas should be monitored, along with**
644 **experiments on different types of food-fall in addition to obtaining side-scan sonar and**
645 **abiotic data. This will provide a more comprehensive view of the food types required for**
646 **these species to thrive in the deep sea.**

647

648 It is not clear from our results whether substrate type (i.e. nodule/non-nodule) has any effect
649 on the amphipod communities (Smith and Demopoulos, 2003) since this kind of data is only

650 available for stations D3 and D4. To answer this question, resampling of the study areas in
651 combination with an Ocean Floor Observation System (OFOBS) (video/camera) is required.

652

653 Although our study only addresses the scavenging amphipod species longer than 15 mm, we
654 already find indications for a disturbance effect in the DEA. It is obvious that scavenging
655 amphipods are only one of several **bentho-pelagic indicator groups. Other truly** benthic
656 groups such as sponges or less dispersive amphipods (e.g. collected by epibenthic sledge
657 (EBS)) may demonstrate an even more pronounced impact of mining activities and should be
658 investigated in future studies.

659

660 With the application of molecular techniques to identify cryptic species (Delić et al, 2017),
661 more realistic estimates of biodiversity can be obtained (Schön et al. 2012), improving our
662 current knowledge of the biodiversity of this area. If these improved estimates of biodiversity
663 also include cryptic species, it is possible that the biological impact of manganese nodule
664 mining on amphipod and other deep-sea faunal communities may turn out to be even higher.

665 **Conclusions**

666

667 In summary, this study on the scavenging amphipod community of two abyssal oceanic
668 basins has demonstrated that amphipods are present in high abundances across the CCZ and
669 DEA, with nine shared species and some species possibly being unique to their respective
670 basin.

671 Our results have indicated that the simulated mining experiment probably had an impact on
672 the biodiversity of these scavenging amphipods, as demonstrated by the low *D* of the DEA
673 overall, at the disturbance site itself (D1), and the 60% dominance of *A. gerulicorbis* in this
674 region.

675 Given the scarcity of sampling and industry experience of marine habitats at these depths, the
676 formulation of effective regulations is challenging (International Seabed Authority, 2017).
677 Nonetheless, our study provides the first results on possible effects of disturbance activities
678 on the abyssal amphipod biodiversity of deep-sea basins.

679 **Sample and data availability**

680 Biological samples pertaining to this manuscript are stored at the Royal Belgian Institute of Natural
681 Sciences, and the data discussed in the manuscript are submitted to PANGEA.

682 **Acknowledgements**

683

684 The authors would like to acknowledge the Belgian Science and Policy Office (BELSPO) and the
685 German Federal Ministry of Research and Education for funding. We would also like to thank the
686 crew of the Research Vessel “Sonne”, the University of Ghent (Ghent, Belgium) and Zohra
687 Elouaazizi (Brussels, Belgium) for help in the lab. This study would not have been possible without
688 their support.

689

690

691

692

693

694

695

696

697

698

699

700 **References**

701

702 Appeltans, W., Boxshall, G., Bouchet, P., Vanhoorneb., Decock, W., Warren, A., Collinsa.,
703 Kroh A., Schmidt–Rhaesa, A., Berta, A., Barber, A., Todaro, A., Gittenberger, A.,
704 Hoeksema, B., Swalla, B., Neuhaus, B., Hayward, B., Self–Sullivan, C., Fransen, C.,
705 Messing, C., Erséus, C., Emig, C., Boyko, C., Mah, C., Millsc., Nielsen, C., Jaume,
706 D., Fautin, D., Domning, D. P., Gibson, D., Lazarus, D., Gordon, D., Opresko, D.,
707 Schwabe, E., Mac–Pherson E., Thuesen, E., Dahdouh–Guebas, F., Anderson, G.,
708 Poore, G., Williams, G., Walker–Smith, G., Read, G., Lambert, G., Paulay, G.,
709 Segers, H., Furuya, H., Bartsch, I., Van Der Land, J., Reimer, J. D., Vanaverbeke, J.,
710 Saiz–aiz, Saike, J., Sair, J. D., Pilger, J., Norenburg, J., Kolb, J., Schnabel, K. E.,
711 Meland, K., Fauchald, K., Cheng, L., Van Ofwe–Gen L., Błażewicz–Paszkowycz,
712 M., Rius, M., Curini–Galletti, M., Schotte, M., Tasker, M. L., Angel, M. V., Osawa,
713 M., Longshaw M., Guiry, M., Bailly, N., De Voogd, N., Bruce N., Shenkar, N.,
714 Garcia–Alvarez, O., Mclaughlin, P., Kirk, P., Davie, P., Ng, P. K. L., Schuchert P.,
715 Uetz P., Bock P., Pugh P., Lemaitrer., Kristensen R., Van Soestr., Bray, R., Bamber,
716 R. N., Da Rocha, R. M., Hopcroft R., Stöhr, S., De Grave, S., Gerken, S., Gofas, S.,
717 Tyler, S., Ahyong, S., Wilson, S., Brandao S. N., Koenemann, S., Feist, S., Cairns, S.,
718 Timm, T., Cribb, T., Molodtsova, T., Chan, T.–Y., Iseto, T., Artois, T., Scarabino, V.,
719 Siegel, V., Eschmeyer, W., Hummon, W., Perrin W., Sterrer, W., Hernandez, F.,
720 Mees J., Costello, M. J.: The Magnitude of Global Marine Biodiversity, *Curr. Biol.*,
721 22, 2189–2202, 2012.

722

723 Baldwin, R. J. & Smith, K. L. Jr.: Temporal variation in the catch rate, length, color, and sex
724 of the necrophagus amphipod, *Eurythenes gryllus*, from the central and eastern North
725 Pacific, *Deep Sea Res. (part 1 Oceanogr. Res. Pap.)*, 34, 425–439, 1987.

726

727 Barnard, J. L.: Gammaridean Amphipoda from depth of 400 to 6000 m, *Galathea. Rep.*, 5,
728 23–128, 1961.

729

730 Barnard, J. L. & Karaman, G.: The Families and Genera of Marine Gammaridean Amphipoda
731 (Except Marine Gammaroids), *Rec. Aust. Mus. Suppl.* 13., 1–866, 1991.

732

733 Belyaev, G. M.: Deep-Sea Ocean Trenches and Their Fauna. Nauka Publishing House,
734 Moscow, 385 pp, 1989.
735

736 Billett, D. S. M., Lampitt, R. S., Rice, A. L. & Mantoura, R. F. C.: Seasonal sedimentation of
737 phytoplankton to the deep-sea benthos, *Nature*, 302, 520–522. 1983.

738 Blankenship, L. E. & Levin, L. A.: Extreme food webs: Foraging strategies and diets of
739 scavenging amphipods from the ocean's deepest 5 kilometers, *Limnol. Oceanogr.*, 52,
740 1685–1697, 2007.
741

742 Blankenship, L. E., Yayanos, A. A., Cadien, D. B., and Levin, L. A.: Vertical zonation
743 patterns of scavenging amphipods from the Hadal zone of the Tonga and Kermadec
744 Trenches, *Deep-Sea Res. Pt. I*, 53, 48–61, doi:10.1016/j.dsr.2005.09.006, 2006.
745

746 Brandt, A., Gooday A. J., Brix S. B., Brökeland, W., Cedhagen, T., Choudhury, M.,
747 Cornelius, N., Danis, B., De Mesel, I., Diaz R. J., Gillan D. C., Ebbe B., Howe J.,
748 Janussen, D., Kaiser, S., Linse, K., Malyutina, M., Brandao, S., Pawlowski, J. &
749 Raupach, M.: The So Deep Sea: First Insights Into Biodiversity And Biogeography.
750 *Nature*, 447, 307–311, 2007.
751

752 Bray, J. R. & J. T. Curtis.: An ordination of upland forest communities of southern
753 Wisconsin, *Ecol. Mono.*, 27, 325–349, 1957.
754

755 Brenke, N.: An epibenthic sledge for operations on marine soft bottom and bedrock, *Mar.*
756 *Technol. Soc. J.*, 39, 10–21, 2005. hdl.handle.net/10.4031/002533205787444015.
757

758 Carmona, L., Malaquias, M. A. E., Gosliner, T. M., Pola, M. & Cervera, J. L. Amphi-Atlantic
759 distributions and cryptic species in Sacoglossan sea slugs, *J. Molluscan Stud.*, 77,
760 401–412, 2011.
761

762 Conlan K. E.: Precopulatory mating behavior and sexual dimorphism in the amphipod
763 Crustacea, *Hydrobiologia*, 223, 255–282, 1991.
764

765 Delić, T., Trontelj, P., Rendoš, M. & Fišer., C.: The importance of naming cryptic species
766 and the conservation of endemic subterranean amphipods. *Sci. Rep.*, 7, 3391, 2017.
767 <https://doi.org/10.1038/s41598-017-02938-z>
768
769

770 Etter, R. J., Boyle, E. E., Glazier, A., Jennings, R. M., Dutra, E. & Chase. M. R.
771 “Phylogeography of a Pan-Atlantic Abyssal Protobranch Bivalve: Implications for
772 Evolution in the Deep Atlantic.” *Molecular Ecology* 20 (4): 829–843, 2011.
773 doi:<http://dx.doi.org/10.1111/j.365-294X.2010.04978.x>.
774

775 France, D. & Kocher, T.: Geographic and bathymetric patterns of mitochondrial 16S rRNA
776 sequence divergence among deep-sea amphipods, *Eurythenes gryllus*. *Mar. Biol.*, 126,
777 633–643, 1996.
778

779 Futuyma, D. J.: *Evolutionary Biology*, ed. 3. Sinauer Associates, Sunderland MA, 751 pp.,
780 1998.
781

782 Gage, J. D. & Tyler, P. A.: *Deep-Sea Biology: a natural history of organisms at the deep-sea*
783 *floor*, Vol. Cambridge University press, Cambridge, 1991.
784

785 German C. R., Ramirez-Llodra, E., Baker, M. C., Tyler, P. A., and the Chess Scientific
786 Steering Committee.: *Deep-Water Chemosynthetic Ecosystem Research during the*
787 *Census of Marine Life Decade and Beyond: A Proposed Deep Ocean Road Map*,
788 *PLoS ONE*, 6, 2011. doi:10.1371/journal.pone.0023259.
789

790 Glover, A., Paterson, G., Bett, B., Gage, J., Sibuet, M., Sheader, M., Hawkins, L.: Patterns in
791 polychaete abundance and diversity from the Madeira Abyssal Plain, northeast
792 Atlantic. *Deep Sea Res. (part 1 Oceanogr. Res. Pap.)*, 48, 217–236, 2001.
793

794 Glover, A., Smith, C. R., Paterson, G. J. L., Wilson, G. D. F., Hawkins, L. & Sheader, M.:
795 Polychaete species diversity in the central Pacific abyss: local and regional patterns,
796 and relationships with productivity, *Mar. Ecol. Prog. Ser.*, 240, 157–170, 2002.
797

798 Gotelli, N. J. & Colwell, R., K.: Quantifying Biodiversity: Procedures and Pitfalls in the
799 Measurement and Comparison of Species Richness.”, *Ecol. Lett.*, 4, 379–91, 2001.
800 doi:10.1046/j.1461-0248.2001.00230.
801

802 Hannides, A., Smith, C. R.: The northeast abyssal Pacific plain. In: *Biogeochemistry of*
803 *Marine Systems*, K. B. Black and G. B Shimmield, eds., CRC Press, Boca Raton,
804 Florida, 208-237, 2003.
805

806 Havermans, C.: Have we so far only seen the tip of the iceberg? Exploring species diversity
807 and distribution of the giant amphipod *Eurythenes*, *Biodiversity*, 2016.
808 DOI: 10.1080/14888386.2016.1172257
809

810 Havermans, C & Smetacek, V. Bottom-up and top-down triggers of diversification: A new
811 look at the evolutionary ecology of scavenging amphipods in the deep sea. *Progress in*
812 *Oceanography*, 164, 37-51, 2018.
813

814 Havermans, C., Sonet, G., d'Udekem d'Acoz, C., Nagy, Z. T., Martin, P., Brix, S., Riehl, T.,
815 Agrawal, S. & Held, C.: Genetic and morphological divergences in the cosmopolitan
816 deep-sea amphipod *Eurythenes gryllus* reveal a diverse abyss and a bipolar species.
817 *PLoS ONE*, 8, e74218, 2013.
818

819 Held, C. & Wägele, J. W.: Cryptic speciation in the giant Antarctic isopod *Glyptonotus*
820 *antarcticus* (Isopoda: Valvirfera: Chaetiliidae), *Sci. Mar.* 69 (Suppl. 2), 175–181,
821 2005.
822

823 Highsmith, R.: Floating and algal rafting as potential dispersal mechanisms in brooding
824 invertebrates, *Mar. Ecol. Prog. Ser.*, 25, 169–179, 1985.
825

826 Hilário, A., Metaxas, A., Gaudron, S. M. ,Howell, K. L., Mercier, A., Mestre, N. C., Ross, R.
827 E. ,Thurnherr, A. M. & Young, C.: Estimating dispersal distance in the deep
828 sea:challenges and applications to marine reserves, *Front. Mar. Sci.*, 2, 2015.
829 doi:10.3389/ fmars.2015.00006.

830 Ide, K., Takahashi, K., Nakano, T., & Minoru, S. & Omori, M. Chemoreceptive foraging in a
831 shallow-water scavenging lysianassid amphipod: Role of amino acids in the location
832 of carrion in *Scopelocheirus onagawae*. Marine Ecology-progress Series, 2006.
833 MAR ECOL-PROGR SER. 317. 193-202. 10.3354/meps317193.
834
835 International Seabed Authority. A Discussion Paper on Developing a Regulatory Framework
836 for Mineral Exploitation in the Area (Env. Matt.), 1–102, 2017.
837
838 Kankaanpää, H., Laurén, M., Mattson, M. & Lindström, M.: Effects of bleached kraft mill
839 effluents on the swimming activity of *Monoporeia affinis* (Crustacea, Amphipoda)
840 Lindström, Chemosphere, 31, 4455–4473, 1995.
841
842 Laver, M. B., Olsson, M. S., Edelman, J. L. & Smith K. L. Jr.: Swimming rates of scavenging
843 deep-sea amphipods recorded with a free-vehicle video camera. Deep-Sea Res. II,
844 32, 1135–1142, 1985.
845
846 Lester, S. E., Ruttenberg, B. I., Gaines, S. D. & Kinlan B. P.: The relationship between
847 dispersal ability and geographic range size. Ecol. Lett, 10, 745–758, 2007.
848
849 Levin, L. A., Etter, R.J., Rex, M. A., Gooday, A. J., Smith, C. R., Pineda, J., Stuart, C. T.,
850 Hessler, R. R. & Pawson, D.: Environmental influences on regional deep-sea species
851 diversity, Annu. Rev. Ecol. Evol. Syst., 32, 51–93, 2001.
852
853 Liow, L. H.: Does versatility as measured by geographic range, bathymetric range and
854 morphological variability contribute to taxon longevity?, Glob. Ecol. Bio., 16, 117–
855 128, 2007.
856
857 Lodge, M., Johnson, D., Le Gurun, G., Wengler, M., Weaver, P. & Gunn, V.: Seabed mining:
858 International Seabed Authority environmental management plan for the Clarion–
859 Clipperton Zone. A partnership approach, Mar. Pol., 49, 66–72, 2014.
860
861 Lonsdale, P.: Clustering of suspension-feeding macrobenthos near abyssal hydrothermal
862 vents at oceanic spreading centers, Deep-Sea Res., 24, 857–863, 1977.

863 Lörz A-N, Jażdżewska AM, Brandt A. A new predator connecting the abyssal with the hadal
864 in the Kuril-Kamchatka Trench, NW Pacific. *Costello M, ed. PeerJ*. 2018;6:e4887.
865 doi:10.7717/peerj.4887.
866
867
868 Managing Impacts of Deep Sea Resource Exploitation (MIDAS) Summary Report:
869 Biodiversity in the Clarion-Clipperton Zone, 1–2, 2016.
870
871 McClain, C. R. & Hardy, S. M.: The dynamics of biogeographic ranges in the deep sea, *Proc.*
872 *Roy. Soc. B: Bio. Sci.*, 277, 3533–3546, 2010.
873
874 Narahara-Nakano, Y., Nakano, T. & Tomikawa, K. Deep-sea amphipod genus *Eurythenes*
875 from Japan, with a description of a new *Eurythenes* species from off Hokkaido
876 (Crustacea: Amphipoda: Lysianassoidea), *Marine Biodiversity*, 1867-1616, 2017. *Mar*
877 *Biodiv* DOI 10.1007/s12526-017-0758-4
878
879 Paull, C. K., Hecker, B., Commeau, R., Freeman-Lynde, R. P., Neumann, C., Corso, W. P.,
880 Golubic, S., Hook, J. E., Sikes, E. & Curray, J.: Biological communities at the Florida
881 escarpment resemble hydrothermal vent taxa, *Science*, 226, 965–967, 1984.
882 Premke K, Muyakshin S, Klages M & Wegner J. Evidence for long-range chemoreceptive
883 tracking of food odour in deep-sea scavengers by scanning sonar data. *J Exp Mar Biol*
884 *Ecol*, 285:283–294, 2003.
885
886 R Core Team. R: A language and environment for statistical computing. R Foundation for
887 Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, 2013.
888 URL <http://www.R-project.org/>.
889
890 Ramirez-Llodra, E., Brandt, A., Danovaro, R., De Mol, B., Escobar, E., German, C. R.,
891 Levin, L. A., Martinez Arbizu, P., Menot, L., Buhl-Mortensen, P., Narayanaswamy,
892 B. E., Smith, C. R., Tittensor, D. P., Tyler, P. A., Vanreusel, A. & Vecchione, M.:
893 Deep, diverse and definitely different: unique attributes of the world's largest
894 ecosystem, *Biogeosciences*, 7, 2851–2899, 2010.
895

896 Ramirez-Llodra, E., Tyler, P. A., Baker, M. C., Bergstad, O. A., Clark, M. R. & Escobar, E.:
897 Man and the last great wilderness: human impact on the Deep Sea, PLoS ONE, 6,
898 2011. doi:10.1371/journal.pone.0022588.
899

900 Rex, M. A., Stuart, C. T., Hessler, R., R., Allen, J. A., Sanders, H. L. & Wilson, G. D. F.:
901 Global-scale latitudinal patterns of species diversity in the deep-sea benthos, Nature,
902 365, 636–639, 1993.

903 Ritchie, H., Jamieson, A. J. & Piortney, S. B. Isolation and Characterization of Microsatellite
904 DNA Markers in the Deep-Sea Amphipod *Paralicella tenuipes* by Illumina MiSeq
905 Sequencing, Journal of Heredity, 367 – 371, 2016.
906

907 Sanders H. L.: Marine benthic diversity: a comparative study, Am. Nat., 102, 243–282, 1968.
908

909 Schön I, Pinto R. L., Halse, S., Smith, A. J. & Martens, K.: Cryptic Species in Putative
910 Ancient Asexual Darwinulids (Crustacea, Ostracoda), PLoS ONE, 7, e39844.
911 doi:10.1371/journal.pone.0039844. 2012.
912

913 Schulenberger, E. & Barnard, J. L.: Clarification of the Abyssal Amphipod, *Paralicella*
914 *tenuipes* Chevreux, Crustaceana, 31, 267–274, 1976.
915

916 Schüller, M. & Ebbe, B.: Global distributional patterns of selected deep-sea Polychaeta
917 (Annelida) from the Southern Ocean, Deep-Sea Res. II, 54, 1737–1751, 2007.
918

919 Simpson, E. H.: Measurement of diversity, Nature, 163, 688, 1949.
920

921 Smith, C. R., Kukert, H., Wheatcroft, R. A., Jumars, P. A. & Deming, J. W.: Vent fauna on
922 whale remains, Nature, 341, 27–28, 1989.
923

924 Smith, C. R. & A.W. J. Demopoulos.: Ecology of the deep Pacific Ocean floor. In:
925 Ecosystems of the World Volume 28: Ecosystems of the Deep Ocean, P. A. Tyler,
926 ed., Elsevier, Amsterdam, pp. 179–218, 2003.
927

- 928 Smith, C. R., Drazen J. & Mincks, S. L.: Deep-sea Biodiversity and Biogeography:
929 Perspectives from the Abyss. International Seabed Authority Seamount Biodiversity
930 Symposium, 1–13, 2006.
- 931
- 932 Somero, G. N.: Adaptations to high hydrostatic pressure, *Ann. rev. physiol.*, 54, 57–577.
933 1992.
- 934
- 935 Stuart, C., Rex, M. & Etter, R.: Large scale spatial and temporal patterns of deep-sea
936 biodiversity. *Ecosystems of the World Volume 28: Ecosys. Deep Oc.*, P. A. Tyler, ed.,
937 Elsevier, Amsterdam, 295–311, 2003.
- 938
- 939 Taguchi Y. H. & Oono, Y.: Relational patterns of gene expression via non-metric
940 multidimensional scaling analysis, *Bioinformatics*, 21, 730–740, 2005.
- 941
- 942 Thiel, H.: Deep-sea Environmental Disturbance and Recovery Potential, *Int. Revue ges.*
943 *Hydrobiol. Hydrogr.*, 77, 331–339, 1992. doi:10.1002/iroh.19920770213.
- 944
- 945 Thurston, M. H.: Abyssal necrophagous amphipods (Crustacea: Amphipoda) in the northeast
946 and tropical Atlantic Ocean, *Prog. Oceanogr.*, 24, 257–274, 1990. doi:10.1016/0079-
947 6611(90)90036-2.
- 948
- 949 Vader, W.: How many amphipods species? 6th International Crustacean Congress, Glasgow,
950 Scotland, 18–22 July 2005, 143, 2005.
- 951
- 952 Wilson, G. D. F., Hessler R.: Speciation in the deep sea, *Ann. Rev. Ecol. Syst.*, 18, 185–207,
953 1987.
- 954
- 955 Zardus, J. D., Etter, R. J., Chase, M. R., Rex, M. A. & Boyle, E. E. Bathymetric and
956 geographic population structure in the pan-Atlantic deep-sea bivalve *Deminucula*
957 *atacellana* (Schenck, 1939), *Mol. Ecol.*, 15, 639–651, 2006.
- 958
- 959

960 **Figure captions**

961

962 **Figure 1: Geographic locations of the two study areas, the Clarion-Clipperton**
963 **fracture Zone (CCZ) (Northeast Pacific) and the DISTurbance and re-COLONisation**
964 **(DisCOL) Experimental Area (DEA) (Peru Basin, Southeast Pacific).** There are nine
965 Areas of Particular Ecological Interest (APEIs) in the CCZ region, which are illustrated
966 by 400 x 400 km² white boxes. Grey boxes indicate the various contractor claim areas in
967 the CCZ. We deployed eight amphipod traps across the CCZ, which is 16,000 km² and
968 7000 km wide, and five in the DEA, which encompasses 11 km² with a width of 4 km.

969

970 **Figure 2: Histogram showing the species assemblage for the scavenging community**
971 **in the Clarion-Clipperton fracture Zone (CCZ) (black) and the DisCOL**
972 **Experimental Area (DEA) (grey).** The abundances of 19 morphotypes are shown.

973

974 **Figure 3: Catch Per Unit Effort (CPUE) illustrating the correlation between**
975 **sampling time and number of individuals collected.** Only the > 15 mm fraction was
976 included here to estimate number of collected individuals.

977

978 **Figure 4: Species rarefaction curves for each of the 13 trap stations across both**
979 **areas, the Clarion-Clipperton fracture Zone and the DisCOL Experimental Area.**
980 Only individuals longer than 15 mm were considered here.

981

982 **Figures 5a and b: Relative species abundances in the Clarion-Clipperton fracture**
983 **Zone and the DisCOL Experimental Area.** These abundances represent the greater than
984 15mm fraction of the scavenging amphipod community only.

985

986 **Figure 6: NMDS plot showing the beta biodiversity (dis/similarities) for each of the**
987 **thirteen amphipod trap sampling stations associated with the two basins, Clarion-**
988 **Clipperton fracture Zone (CCZ) (black) and the DisCOL Experimental Area (DEA) (red).**
989 Data are supported by a low stress value of 0.01.

990

991

992 **Table captions**

993

994 **Table 1: Station overview.** Codes refer to the codes used in this paper in figures 1, 3, 4, 6
995 and Table 3. The original station code represents the cruise codes from (SO239 and SO232-
996 1). Depth refers to water depth (m) on deployment. Nodule presence/absence information is
997 known only for stations D3 and D4.

998 **Table 2a:** Overview of morphotypes across the Clarion-Clipperton fracture Zone (CCZ) and
999 DisCOL Experimental Area (DEA).

1000 **Table 2b:** Overview of morphotypes across the Clarion-Clipperton fracture Zone (CCZ) and
1001 DisCOL Experimental Area (DEA).

1002 **Table 3:** Comparison of biodiversity calculated using the Simpson Index (D), for the
1003 Clarion-Clipperton fracture Zone and DisCOL Experimental Area, and D for each station is
1004 shown for further comparisons within these areas.

1005
1006
1007
1008

1009

1010

1011

1012

1013

1014

1015 **Appendix/Electronic Supplementary Information (ESM) captions**

1016

1017 **Appendix 1: Multibeam scan** - Showing the location of the 78 track marks created by the
1018 plough harrow in the DisCOL Experimental Area to simulate manganese nodule extraction
1019 activity (D1)

1020 **Appendix 2** – Photograph showing the baited free-fall lander trap designed and deployed by
1021 RBINS.

1022 **Appendix 3** - Side-scan sonar image of site D5 showing possible seamount barriers. View
1023 from NW (top) to SE (bottom). Contours are every 25 m. (Source: GEOMAR, 2015).

1024 **Appendix 4** – Calculation of alpha biodiversity used in this manuscript.

1025

1026

1027

1028

1029

1030

1031

1032

1033

1034

1035

1036

1037

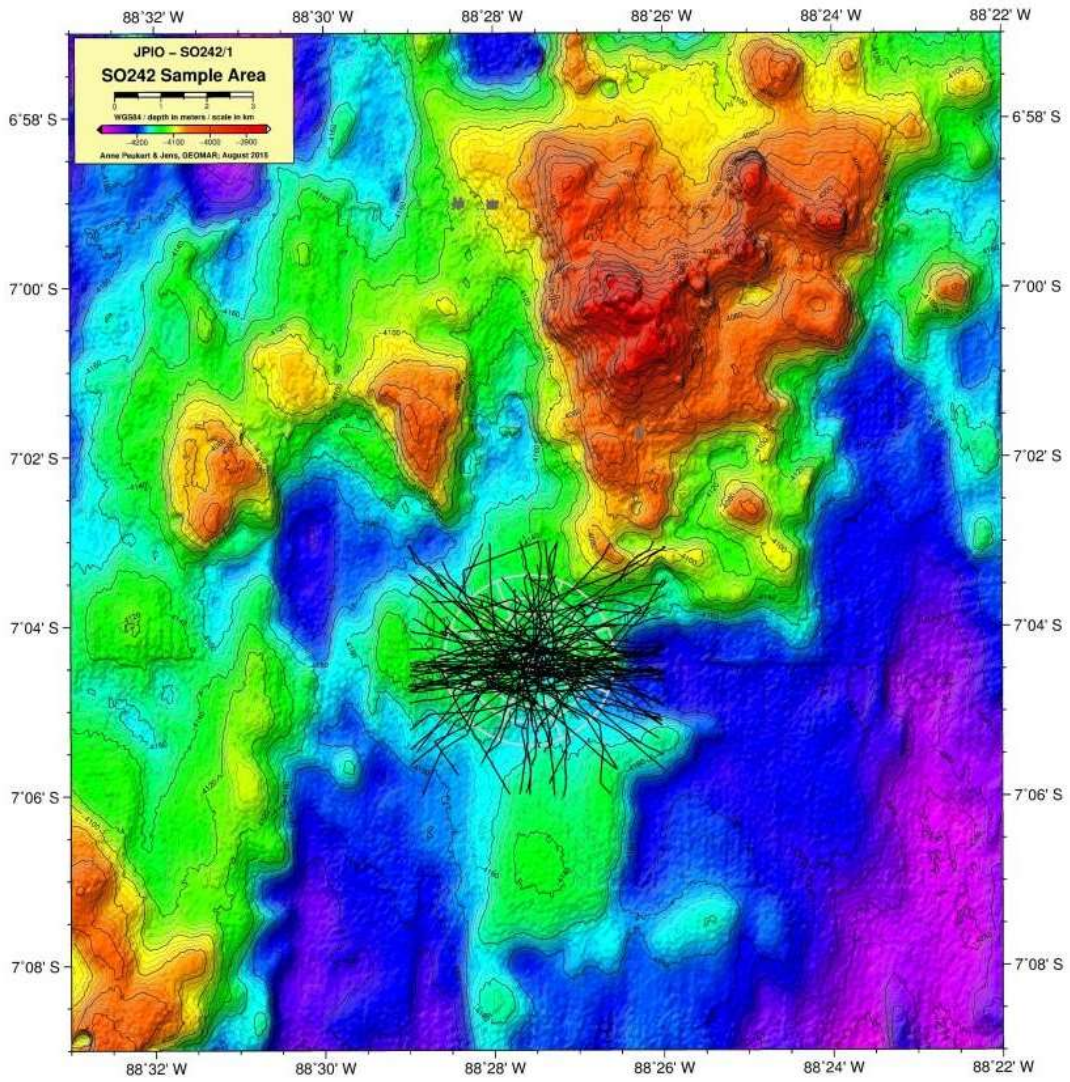
1038

1039

1040

1041 **Appendix 1: Multibeam scan** - Showing the location of the 78 track marks created by the
1042 plough harrow in the DisCOL Experimental Area to simulate manganese nodule extraction
1043 activity.

1044



1045

1046

1047

1048

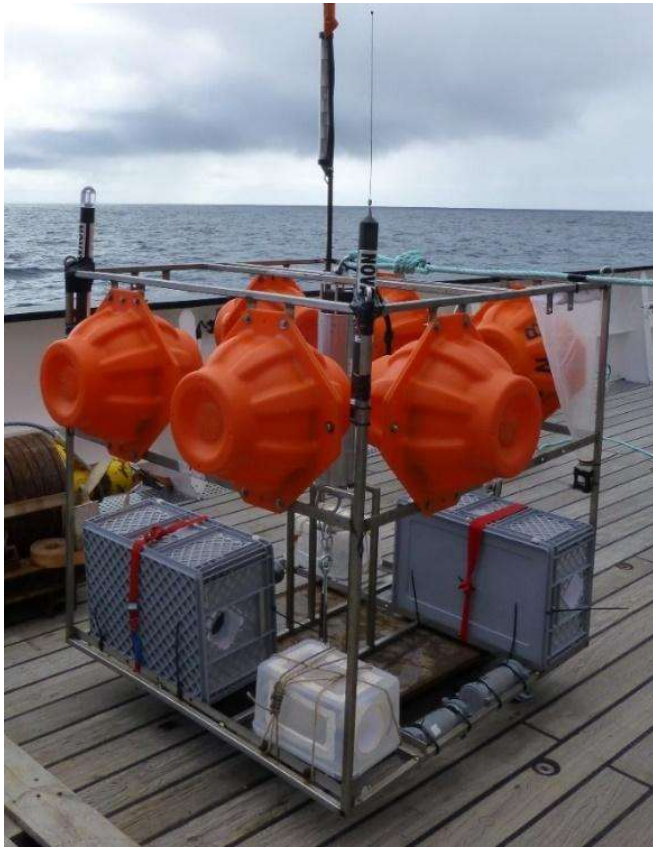
1049

1050

1051

1052

1053 **Appendix 2** – Photograph showing the baited free-fall lander trap designed and deployed by
1054 RBINS.



1055

1056

1057

1058

1059

1060

1061

1062

1063

1064

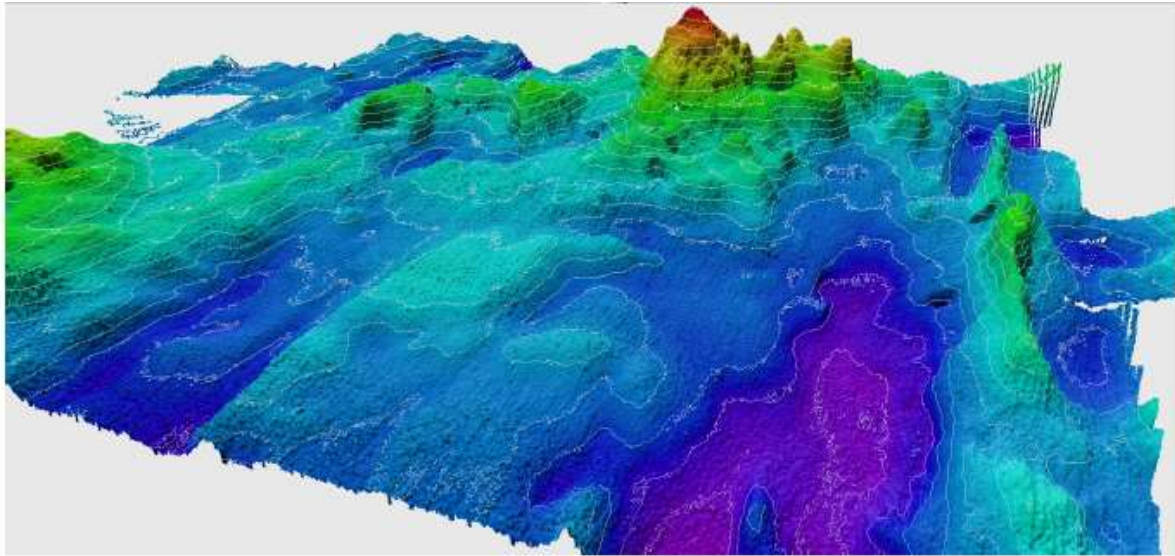
1065

1066

1067

1068 **Appendix 3** - Side-scan sonar image of site D5 showing possible seamount barriers. View
1069 from NW (top) to SE (bottom). Contours are every 25 m. (Source: GEOMAR, 2015).

1070



1071

1072

1073

1074

1075

1076

1077

1078

1079

1080

1081

1082 **Appendix 4** - Calculation of alpha biodiversity used in this manuscript.

1083 Simpson Index

1084 **Simpson Diversity Index (D) = $1 - \frac{\sum n(n-1)}{N(N-1)}$ =**

1085 D = Diversity Index

1086 n = number of individuals in each particular species

1087 N = Total number in community

1088 A high value of D indicates a high species diversity.

1089