File contents

5

- Response to Reviewer 1
- Response to Reviewer 2
- Brief list of changes in manuscript and supplement

- Marked up edited manuscript
 - Marked up edited supplement

Response to Reviewer 1

Balancing the number of acronyms used in a manuscript is always challenging. In response to this comment, we have reduced the number of acronyms in the conclusions section. We find that using the acronyms for each process (e.g. NAR and NIR) in the rest of the text is less confusing than spelling out NO₃⁻ reduction and NO₂⁻ reduction, since it is easier to mistake the 3 and

- 5 2 in the subscripts than the "A" and "P" in the acronyms. There is no perfect solution to this, and we hope that removing the process acronyms in the conclusions will be helpful in this regard. In addition, we have opted to retain widely used three-letter acronyms for chemical species (DIN, DON, PON, etc.), but have revised the less common two-letter acronym 'ON' to 'organic N' throughout. We believe this strikes a balance between both clarity and brevity.
- 10 A brief comment on the importance of improving N cycle models by adding NO2⁻ as a tracer has been added to the abstract.

Specific comments

- Page 2, line 16: Rephrased as requested.
- Page 2, line 17: Rephrased as requested.
- Page 3, line 24: Rephrased as requested.
- Page 3, line 31: The optimization itself was not conducted using the sensitivity analysis, but the optimization procedure is discussed in Section 2.6. Sensitivity analysis was performed in order to narrow the set of parameters that was optimized, since there are too many parameters in the model to optimize them all concurrently due to computational limits.
 - Page 4, line 4: Rephrased as requested.
- Page 4, line 30: Additional comments added to reference NH₄⁺ accumulation in ODZs.
 - Added text: "Though NH4⁺ has been observed to accumulate to micromolar concentrations in ODZs (Bristow et al., 2016; Hu et al., 2016), this occurs largely in shallow, coastal shelf regions, which are not resolved by the model."
 - Page 6, line 15: References added for rair (Mariotti, 1983).
 - Page 7, line 16: The references to the pe ratio have been changed to *P_e* in both equations and the text to improve clarity.
- 25
- Page 7, line 22: Sentence has been restructured to avoid starting with a symbol
- Section 2.3.2: A brief discussion of the exclusion of riverine N inputs from the model has been added to the beginning of the section.
- 30
- Added text: "In our model, atmospheric deposition and N₂ fixation are the only sources of bioavailable N in the model. These are the two largest sources of N to the ocean (Gruber and Galloway, 2008). We do not consider the third largest source of N, riverine fluxes, in the model due to lack of coastal resolution and the expectation that much of the river-derived N is denitrified in the shelf sediments (Nixon et al., 1996;
 - 2

Seitzinger and Giblin, 1996). This would potentially impact the surface NO_3^- and $\delta^{15}N_{NO3}$ but the overall contribution to the N budget would be negligible, especially considering the coarse resolution of the model."

- Page 8, line 4: DON and NO₃⁻/NO₂⁻ must be considered separately in order to introduce dependence on two N species for NO₃⁻ reduction and NO₂⁻ reduction, since these heterotrophic processes require both organic N and either NO₃⁻ or
- NO₂⁻. In order to use the simple linear model setup, DON and NO₃⁻/NO₂⁻ cannot be variables/unknown at the same time. Additional explanation can be found in Section 2.3.1, and some additional discussion has been added to the line in question.
 - Added text: "The assimilation rates for DON and PON must be calculated using observed surface [NO₃⁻], rather than modeled [NO₃⁻], in order for the heterotrophic processes of NAR and NIR to be dependent on both organic N and NO₃⁻ or NO₂⁻ availability, respectively."
- Page 9, line 1: This sentence has been clarified to indicate that having a variable ¹⁴k_{sol} that accurately represents slower ON remineralization under low O₂ conditions would be a refinement that could be incorporated in future model versions.
 - Added text: "A spatially variable ¹⁴k_{sol} that accounts for lower apparent values in ODZs is a refinement that could be introduced in future model versions."
- Page 11, line 1-3: A mention of oxygen as a field that is not explicitly modeled has been added to the end of Section 2.1 (the N cycle model overview).
- Page 12, line 3: Cutoff points discussion has been rephrased to clarify that they are the transition points between the
 piecewise segments of the ([O₂] [NO₅⁻]) vs. sedimentary denitrification rate relationship.
- Added text: "In order for sedimentary denitrification to be properly implemented in our linear model, we broke the original non-linear relationship into three roughly linear segments to create a piecewise relationship between ([O₂] [NO₃⁻]) and sedimentary denitrification rate. We obtained three linear relationships between ([O₂] [NO₃⁻]) and sedimentary denitrification rate, each applicable across a given range of ([O₂] [NO₃⁻]) values (Figure S1). Due to the nature of our linear model, we needed to express the interval cutoff points that define the transition between the piecewise relationship segments in terms of O₂ rather than ([O₂] [NO₃⁻])."
- Page 13, line 21: Sentence has been rephrased to indicate that there are 200,160 total ocean boxes in the model.
 - o Added text: "All model ocean boxes (200,160 in total) are accounted for in the matrices."
- Page 17, line 27: Rephrased as requested.
- Page 17, lines 21-31: This section of text refers to model nitrate δ¹⁵N that is lower than observations on a transect extending westward from the ETSP oxygen deficient zone, which we propose to be due to an underestimate of NO₃⁻ reduction in the model ETSP. We interpret the reviewer's question to be about the potential role of fractionation during NH₄⁺ production and consumption in driving NO₃⁻ isotope variations in the real ocean, which are not fully
 - 3

10

15

5

20

represented in the model. We agree that there typically are large fractionation factors associated with usage of NH4⁺ via uptake or oxidation, and that these fractionation factors should be expressed if NH4+ accumulates in the water column, or if there is a large difference in fractionation factors between competing pathways. The reviewer rightly points out that NH4⁺ is not explicitly included in the model. We feel justified in doing this since accumulation of NH4⁺ in the modern ocean, even in oxygen deficient zones, is typically very low. We agree that ignoring NH4⁺ in euxinic conditions, where it constitutes a large fraction of dissolved inorganic N, would not be advisable. In the current model, while not explicitly representing NH₄⁺ as a DIN species, we do attempt to represent the partitioning of NH₄⁺ generated during DON degradation (ammonification) between ammonia oxidation and anammox below the euphotic zone. From an isotope balance perspective, we also assume that these competing fates have similar fractionation factors, which, while fairly uncertain, is generally supported by available data. Here we apply similarly low fractionation factors (a = 1), which is not likely the case, but turns out to not impact nitrate $\delta^{15}N$ in the circumstances applied here, where NH_4^+ does not accumulate, and both consumption processes have the same isotope effect. As applied here, the $NO_3^$ generated has the same δ^{15} N as remineralized DON. If an isotope effect were applied to ammonia oxidation, this would most likely lower the $\delta^{15}N$ of nitrate, not raise it. We believe that if anything our model results are biased towards he upper limit of nitrate δ^{15} N produced during nitrification. One scenario of concern for our application would be if anammox has a much larger fractionation factor towards NH4⁺ than ammonia oxidation, leaving ¹⁵N-enriched NH_4^+ to be oxidized to NO_3^- in the water column. While theoretically possible, there are no observations that we are aware of that would support this scenario in the modern ocean. Secondly, if partly consumed NH4+ is released from the sediments and oxidized to NO3- in the water column this could contribute to 15N-enrichment of NO3- that is not represented by the model. There is observational support for this effect in the modern ocean, as mentioned by reviewer #2. In response to these two questions about the underrepresentation of NH_4^+ in the model, we have added a few additional comments regarding potential errors in the model associated with our representation.

Added text:

- p. 13: "This is a conservative estimate of the effects of benthic N loss on water column NO₃⁻ isotopes, as several studies suggest that benthic N processes may contribute to water column nitrate ¹⁵N-enrichment (Lehmann et al., 2007; Granger et al., 2011; Somes et al., 2015; Brown et al., 2015). However, our current model parameterization does not require additional benthic fractionation to fit deep ocean δ¹⁵N_{NO3}. Also, our spatial resolution does not well represent regions where this effect might be important."
- 30
- p. 18: "The simplification of NH₄⁺ dynamics in the model could contribute to underestimation of $\delta^{15}N_{NO3}$ values if there were a large flux of ¹⁵N-enriched NH₄⁺ from sediments (Granger et al., 2011), or if ¹⁵N-depleted NH₄⁺ was preferentially transferred to the N₂ pool via anammox. While the isotope effect on NH₄⁺ during anammox (Brunner et al., 2013) is higher than that applied here, we chose to balance this with a low isotope effect during aerobic NH₄⁺ oxidation (Table 1)."
 - 4

10

15

20

25

- Page 19, line 25: These sentences have been reworked to indicate that low NO₂⁻ consumption rates (via NIR, AMX, and NXR) in conjunction with modest NO₂⁻ production rates (via NAR and likely AMO) result in an accumulation of NO₂⁻.
 - Added text: "The accumulation of NO₂" here in the model is likely due to O₂ concentrations falling below the set threshold for NAR but above the threshold for NIR, so NO₂" can accumulate via NAR but cannot be consumed via NIR. Although AMX and NXR occur there, the modeled rates of their NO₂" consumption are rather low, which in combination with high rates of NAR and no NIR leads to more NO₂" being produced than consumed."
- 10 Page 20, line 14: Acronyms have been replaced with full process names in the conclusion.

5

• Figure 4: The figure has been amended to include a color legend for clarity (see attached image).

Response to Reviewer 2

15

20

- Page 3, lines 2-5: It is likely that the isotope effects measurements are subject to both environmental conditions and the microbial community composition. Though the strains chosen for culture by Buchwald and Casciotti (2010) and Casciotti (2009) are thought to be representative of the marine nitrite oxidizing community, that is not necessarily the
- 5 case, and the pure culture studies in the laboratory are missing potential feedbacks from other *in situ* microbial processes or changing environmental conditions on an extremely local scale. There has been evidence for changing isotope effect expression under different environmental conditions (e.g. Krittee et al., 2012), so that could play a role in the different estimates. It is also possible that nitrite oxidation could be occurring with an alternative electron acceptor, but would be difficult to incorporate into our model due to our lack of thorough understanding about what these alternative pathways might be.
 - Page 4, line 20: It would be a considerable computational challenge to increase the resolution of the physical circulation model. The data assimilation process used to construct the data-constrained tracer-transport model used in the present study took more than 1 month on a dedicated computer. With an increase in the horizontal resolution to ~1/2* (the minimum resolution needed to start resolving the coastal ocean) the computational cost per iteration would increase by a factor of 16 and a larger number of iterations would likely be needed for the optimization to converge. Furthermore, the direct matrix inversions which we use to solve the physical and biogeochemical models would no longer be possible. The memory requirements would be too high. As a result, the fast direct solvers would have to be replaced by slower iterative solvers. Thus a global inverse model that resolves the coastal oceans is not presently feasible. It should however be feasible to embed our biogeochemistry model into a regional physical circulation model that resolves the coastal circulation.
 - Page 4, lines 30-32: Additional comments added to reference NH4⁺ accumulation in ODZs.
 - Added text: "Though NH4⁺ has been observed to accumulate to micromolar concentrations in ODZs (Bristow et al., 2016; Hu et al., 2016), this occurs largely in shallow, coastal shelf regions, which are not resolved by the model."
- Page 5, line 23: A definition for the World Ocean Atlas acronym has been added to its first usage.
 - Page 5, lines 28-30 and page 6, lines 1-2: DON in the model is not differentiated between labile and recalcitrant pools. The concentrations at the surface are largely between 0-20 µM, with some higher concentrations in equatorial regions with high surface production. The DON concentration decays rapidly with depth, as its distribution is controlled by PON solubilization (driven by a Martin curve) and remineralization with a first order rate constant. Though the concentration in the surface box is higher than it should be, averaging the concentration over the top two or three boxes (representative of the mixed layer) yields concentrations closer to 10 µM. The representation of ON in the model and the surface processes that affect it is relatively simplified, and could be improved and expanded in subsequent versions of the model.
 - 6

- Page 6, line 9: The modeled N deposition values here match observations fairly well (see Dentener et al., 2006) though
 there is some spatial variation in the goodness of the prediction. Many of the regions with lower prediction accuracy
 are also those with lower fluxes, so the errors do not lead to large under- or over-predictions for global N deposition.
 There are some newer estimates for N deposition (as well as future projections) that will be explored in a subsequent
 paper.
- Page 6, lines 25-28 and page 7, lines 1-2: Though our N₂ fixation parameterization is by no means perfect, we do not exclude non-*Trichodesmium* N₂ fixers. Explicitly adding additional types of N₂ fixers would require different parameters for maximum N₂ fixation rate, temperature constraints, and Fe and PO₄³⁻ limitation, which we do not have good literature values for and are limited computationally by the number of parameters we can optimize. Additional investigations using this model that focus more on N₂ fixation rates could certainly implement different classes of N₂ fixers, but the generally good spatial patterns of N₂ fixation in the model and appropriate global rates (see below) are adequate for the ODZ analyses of interest here.
- Page 7, lines 12-13: The spatial patterns of N₂ fixation produced by the model are very similar to the observationallyconstrained estimates of Luo et al. (2014). The modeled rates of N₂ fixation are appropriately low in the ETSP. Another paper (submitted to Global Biogeochemical Cycles) focuses on assessment of the N cycle process rates and their comparison to both other models and observational data.
- Page 8, lines 6-7: We believe that this is a reasonable assumption within the coarse framework of our model. Assimilation is only represented in the top two boxes of the model, which extend from 0-36 m (box 1) and 36-73 m (box 2). These two boxes are shallower than the characteristic high accumulation of NO₂⁻ in the oxygen deficient zone, and accumulation of NO₂⁻ in the primary NO₂⁻ maximum usually only reaches a maximum of 1-2 μM, which would be negligible compared to NO₃⁻.
- Page 8, lines 30-31: Though carbon burial is higher under anoxic conditions, there have been other work that has
 shown preferential mineralization of N relative to C under anoxic conditions (e.g. Van Mooy et al., 2002; Roberts et
 al., 2012). Since variable C:N ratios are beyond the scope of this model, and in order to keep things simple, we assume
 that the preferential mineralization of N relative to C balances the slower mineralization rate under anoxic conditions.
- Page 10, lines 14-15: We have revised this section to note that our estimate of denitrification does not include chemoautotrophic denitrification, citing Lavik et al., 2009. As our model does not include cycling of reduced sulfur species, it is not possible for us to include chemoautotrophic denitrification at this time, leading to our estimates potentially underestimating overall rates of denitrification. However, since our model is optimized to fit observations of NO₃⁻ and NO₂⁻ concentration and isotopes, the overall rate of denitrification is constrained. If chemolithotrophic denitrification can be assumed to have a similar N isotope effect as heterotrophic denitrification (Frey et al., 2014), it may just be part of the mix of our signal.
 - Added text: "When NAR occurs chemoautotrophically, it would be dependent primarily on the presence of NO₃- and an electron donor, such as hydrogen sulfide (Lavik et al., 2009). Since we do not model the
 - 7

10

5

15

- 20
- 25

production of reduced sulfur species in our model, our estimates of denitrification would not explicitly include the effects of this process. However, chemolithotrophic denitrification could be tacitly accounted for in the optimization process, since the rate constants that control the rates of NAR and NIR are optimized in order to best fit the observations, and the isotope effect for chemolithotrophic denitrification is thought to be similar to that of heterotrophic denitrification (Frey et al., 2014)."

- Page 11, line 1: If the thresholds were set to nanomolar levels as suggested by Dalsgaard et al. (2014), there would be virtually no NAR, NIR, or AMX occurring in the model at all. The coarse resolution of the model results in depth-averaged [O₂] in many ODZ boxes that are above zero, even after applying the empirical correction put forward by Bianchi et al. (2012) to improve WOA O₂ data fit in ODZs. It is not possible in this coarse, time-independent scheme to model any truly anoxic microenvironments. As a result, the area of the model ocean which is truly anoxic is very small. The area in which NAR, NIR, and AMX have been measured, observed, inferred, and modeled in previous
- ODZ studies is much greater than the extent of the anoxic model boxes, and thus we have chosen to be more flexible in our O₂ thresholds for anoxic processes in order to best reflect current knowledge of what is occurring within ODZs. As discussed in Section 4.2, the model dependency on this input O₂ has its downsides and does not adequately represent all ODZs despite our accommodating thresholds. The O₂ thresholds and the effect of changing the thresholds on process rates is discussed further in another paper (submitted to Global Biogeochemical Cycles).
 - Page 11, line 13: Previous studies on the biomechanics of anammox have included the nitrate oxidoreductase enzyme, as the oxidation of nitrite via this enzyme produced electrons that are needed for autotrophic CO₂ fixation under normal anammox conditions (de Almeida et al., 2011; Kartal and Keltjens, 2016). Culture studies have also measured the stoichiometric production of NO₃⁻, presumably by this enzyme (Strous et al., 1999; Brunner et al., 2013).
 - Page 12, lines 10-13: This is an excellent suggestion, which we appreciate. We have thought about how to address
 this in the paper, as unfortunately it is not possible in the scope of the current model to incorporate this process in the
 model. The quoted text below represents our revisions to the paper in accordance.
- 25

30

20

5

10

• Added text:

- p. 13: "This is a conservative estimate of the effects of benthic N loss on water column NO₃⁻¹ isotopes, as several studies suggest that benthic N processes may contribute to water column nitrate ¹⁵N-enrichment (Lehmann et al., 2007; Granger et al., 2011; Somes et al., 2015; Brown et al., 2015). However, our current model parameterization does not require additional benthic fractionation to fit deep ocean δ¹⁵N_{NO3}. Additionally, our spatial resolution does not well represent regions where this effect might be significant on bottom water δ¹⁵N_{NO3}, such as the shallow shelves."
- p. 18: "The simplification of NH4⁺ dynamics in the model could contribute to underestimation of δ¹⁵N_{N03} values if there were a large flux of ¹⁵N-enriched NH4⁺ from sediments (Granger et al., 2011), or if ¹⁵N-depleted NH4⁺ was preferentially transferred to the N₂ pool via anammox. While
 - 8

the isotope effect on NH₄⁺ during anammox (Brunner et al., 2013) is higher than that applied here, we chose to balance this with a low isotope effect during aerobic NH₄⁺ oxidation (Table 1)."

- Page 14, lines 9-10: This paper has been submitted to Biogeosciences Discussions and will be available for proper citation shortly.
- Page 15, lines 22-25: The significance of these relationships was not calculated, and the lines shown are simply 1:1 lines to aid visual comparison between the modeled and observed data in the test set. It is likely that calculating such values would result in strange regression data and low significance. In many cases (particularly for NO₂⁻), the magnitude of the model is incorrect, but observing [NO₂⁻] and $\delta^{15}N_{NO2}$ in a profile or section view reveals that the overall patterns are correct.
- Page 16, lines 17-18: The model cannot resolve episodic events, since it is a steady state, time-independent model that relies on a generalized circulation matrix. Though in the ETSP there is likely some upwelling that is incorporated into the circulation matrix, the more nuanced, time-dependent changes that occur in N cycling as a result of upwelling cannot be accounted for in this model.
 - Added text: "This could be due in part to the time-independent nature of this steady state inverse model, which does not capture the effects of upwelling events in the ETSP on N supply and cycling."
 - Page 16, lines 20-23: An additional line accounting for the impact of sedimentary PO₄³ release on N* has been added, though attempting to account for said release is beyond the capabilities of this model. This is particularly true since the shallow shelves where low O₂ waters come into contact with sediments are not resolved in the model.
- 20

25

30

- Added text: "Negative N* values are associated with N loss due to AMX or NIR or release of PO4³⁻ from anoxic sediments (Noffke et al., 2012), while positive N* values are associated with input of new N through N₂ fixation (Gruber and Sarmiento, 1997)."
- Page 17, line 2: It is not surprising that the model does not reproduce the high δ¹⁵N_{NO3} values observed in the eddy for two reasons. First, the ETSP processes are not well-resolved by the model due to the aforementioned O₂ threshold issues. Second, the time-independent and steady-state nature of the model would not be able to capture transient features such as eddies, where the δ¹⁵N_{NO3} increases over time due to lack of NO₃⁻ resupply within the closed system of the eddy.
- Page 19, lines 2-3: This section has been rephrased. The original intent was to indicate the small fractional offset between the DIN and organic N model runs, both locally and globally.
 - Added text: "However, the majority of DIN assimilation estimates were within 10 μM/yr of the organic N production estimates, with an average offset of approximately 3.5% compared to DIN assimilation. The total global assimilation rates were within 0.4%, with some spatially variable differences due to offset between surface [NO₃⁻] and modeled [NO₃⁻]. However we find that the WOA surface NO₃⁻ values are fairly well represented by our modeled surface NO₃⁻ (Figure S4)."
 - 9

- Page 19, lines 8-21: The O₂ data product available through WOA is imperfect and coarse, which is why O₂ thresholds have been adjusted to accommodate the values. The empirical correction calculated by Bianchi et al. (2012) that corrects the WOA using the GLODAP data attempts to tackle this issue, but the O₂ values in ODZs are still too high. It is likely that if we had more thorough spatial coverage of O₂ data from STOX sensors (rather than Seabird), we
- could use the model to more accurately and thoroughly probe some of the questions about O_2 limitation for these processes. However, using a time-independent, steady-state model would still be unable to capture small, dynamic changes in O_2 that may be important for driving some N cycling within regions that are close to the O_2 detection limit or the O_2 limit for these processes.
 - Page 20, lines 12-13: Additional reference to Bourbonnais et al. (2015) has been added.
- 10

5

- Figure 3: The lines shown are not regressions, they are 1:1 lines and do not have associated r² or significance values. See above for further discussion of the comparison between model output and test set data.
- Figure 4: These are indeed offshore profiles. Due to low model resolution, there are very few coastal or on-shelf boxes, especially in ODZs. A note has been added to the figure legend and description to indicate that the profiles are offshore.
- Figure 5-6: Section profiles comparing the modeled and observed [NO₂⁻] and δ¹⁵N_{NO2} for the GP16 transect have been added to the supplement (and are attached to this comment). NO₂⁻ does not accumulate along the GA03 transect and no δ¹⁵N_{NO2} measurements were made, so those profiles are not included.
 - Page 9, line 15: This font error and a few others were corrected.
 - Page 21, line 8: This grammatical error was corrected.

Brief list of changes in manuscript

5

- Title has been updated to include nitrate
- Minor textual changes for clarity, including reducing the number of acronyms used
- Some discussion of N2 fixation has been added, with an additional figure and table included in the supplement
- Supplemental figure with GP16 NO2⁻ concentration and isotopes has been added
- Discussion of the treatment of NH4⁺ has been added throughout the manuscript, as it is one of the major model caveats
- Figure 4 has been updated to include a legend
- · References have been added to further justify model assumptions and address shortcomings with this model setup

Modeling oceanic<u>nitrate and</u> nitrite concentrations and isotopes using a 3D inverse N cycle model

Taylor S. Martin¹, François Primeau², Karen L. Casciotti^{1*}

¹Stanford University, Department of Earth System Science ²University of California, Irvine, Department of Earth System Science

*Corresponding author: kcasciotti@stanford.edu; 650-721-5545

Abstract. Nitrite (NO_2^{-}) is a key intermediate in the marine nitrogen (N) cycle and a substrate in nitrification, which produces nitrate (NO_3^{-}) , as well as water column N loss processes, denitrification and anammox. In models of the marine N cycle, NO_2^{-} is often not considered as a separate state variable, since NO_3^{-} occurs in much higher concentrations in the ocean. In oxygen

- 10 deficient zones (ODZs), however, NO2⁻ represents a substantial fraction of the bioavailable N, and modeling its production and consumption is important to understanding the N cycle processes occurring there, especially those where bioavailable N is lost from or retained within the water column. <u>Improving N cycle models by including NO2⁻</u> is important in order to better <u>quantify N cycling rates in ODZs</u>, <u>particularly N loss rates</u>. Here we present the expansion of a global 3D inverse N cycle model to include NO2⁻ as a reactive intermediate as well as the processes that produce and consume NO2⁻ in marine ODZs.
- 15 NO₂⁻ accumulation in ODZs is accurately represented by the model involving NO₃⁻ reduction, NO₂⁻ reduction, NO₂⁻ oxidation, and anammox. We model both ¹⁴N and ¹⁵N and use a compilation of oceanographic measurements of NO₃⁻ and NO₂⁻ concentrations and isotopes to place a better constraint on the N cycle processes occurring. The model is optimized using a range of isotope effects for denitrification and NO₂⁻ oxidation, and we find that the larger (more negative) inverse isotope effects for NO₂⁻ oxidation along with relatively high rates of NO₂⁻ oxidation give a better simulation of NO₃⁻ and NO₂⁻
- 20 concentrations and isotopes in marine ODZs.

1 Introduction

5

Nitrogen (N) is an important nutrient to consider when assessing the biogeochemical cycling in the ocean. The N cycle is intrinsically tied to the carbon (C) cycle, whereby N can be the limiting nutrient for primary production and carbon dioxide
uptake (Moore et al., 2004; Codispoti, 1989). Understanding the distribution and speciation of bioavailable N in the ocean allows us to make inferences about the effects on other nutrient cycles and potential roles that N may play in a regime of climate change (Gruber, 2008).

There are several chemical species in which N can be found in the ocean. The largest pool of bioavailable N is nitrate (NO₃⁻), 30 a dissolved inorganic species, which can be taken up by microbes for use in assimilatory or dissimilatory processes. Another Deleted: nitrogen

dissolved inorganic species, nitrite (NO₂⁻), accumulates in much lower concentrations but is a key intermediate in many N cycling processes. Models of the marine N cycle often include NO₃⁻ and NO₂⁻ together as a single dissolved inorganic N (DIN) pool, or exclude NO₂⁻ entirely (DeVries et al., 2013; Deutsch et al., 2007; Brandes and Devol, 2002). However, NO₂⁻ does accumulate significantly in oxygen deficient zones (ODZs) in features known as secondary NO₂⁻ maxima_k and it is an intermediate or substrate in many important N cycle processes occurring there.

5

ODZs are hotspots for marine N loss (Codispoti et al., 2001; Deutsch et al., 2007), which is driven by processes that result in conversion of bioavailable DIN to dinitrogen gas (N₂). The two main water column N loss processes, denitrification and anammox, use NO_2^- as a substrate. Denitrification involves the stepwise reduction of NO_3^- to NO_2^- and then to gaseous <u>pitric</u>

10 oxide (NO), nitrous oxide (N₂O), and N₂. Anammox consists of the anaerobic oxidation of ammonium (NH₄⁺) to N₂ using NO₂⁻ as the electron acceptor. NO₂⁻ is also oxidized to NO₃⁻ during anammox, representing an alternative fate for NO₂⁻ in ODZs. Indeed, NO₂⁻ oxidation appears to be prevalent in ODZs, with more NO₂⁻ oxidation occurring than can be explained by anammox alone (Gaye et al., 2013; Peters et al., 2016; Peters et al., 2018; Babbin et al., 2017; Buchwald et al., 2015; Casciotti et al., 2013; Martin and Casciotti, 2017). NO₂⁻ oxidation results in the regeneration of NO₃⁻ that would otherwise be converted

15 to N₂ and lost from the system. The close coupling between NO₃⁻ reduction to NO₂⁻ and NO₂⁻ oxidation back to NO₃⁻ represents a control valve on the marine N budget (Penn et al., 2016; Bristow et al., 2016). Where NO₂⁻ oxidation can outcompete NO₂⁻ reduction via denitrification and anammox, bioavailable N is retained. Water column N losses only occur where NO₂⁻ oxidation <u>rates are</u> limited by oxygen availability. Thus, understanding the NO₂⁻ dynamics in ODZ waters is critical to assess the N loss occurring there.

20

The observed NO₃⁻ and NO₂⁻ concentrations alone do not allow us to fully characterize the N cycling processes occurring in a given region. Stable isotope measurements of NO₃⁻ and NO₂⁻ provide additional insight and constraints on marine N cycle processes. There are two stable isotopes of N, ¹⁴N and ¹⁵N. The isotopic ratios for a given N species, usually expressed in delta notation as $\delta^{15}N$ (‰) = ($^{15}N/^{14}N_{sample} \div {}^{15}N/^{14}N_{standard} -1$)×1000, is an integrated measure of the processes that have produced and consumed that N species. Each process imparts a unique isotope effect $\frac{42}{45} = {}^{14}k/{}^{15}k - 1 \times 1000$, where ${}^{14}k$ and ${}^{15}N$ containing molecules, respectively) that impacts the isotopic composition of the substrate and the product (Mariotti et al., 1980). In particular, NO₂⁻ cycling processes have distinct isotope effects for NO₂⁻ reduction, which occurs with normal isotopic fractionation (Bryan et al., 1983; Martin and Casciotti, 2016; Brunner et al.,

2013) and NO^{2⁻} oxidation, which occurs with an unusual inverse kinetic isotope effect (Casciotti, 2009; Buchwald and
 Casciotti, 2010; Brunner et al., 2013). Thus, the isotopes of NO²⁻ are very sensitive to the relative importance of NO²⁻ oxidation and NO²⁻ reduction in NO²⁻ consumption (Casciotti, 2009; Casciotti et al., 2013).

Models of the marine N cycle have employed isotopes and isotope effects in conjunction with N concentrations to elucidate N cycle processes (Brandes and Devol, 2002; Sigman et al., 2009; Somes et al., 2010; DeVries et al., 2013; Casciotti et al., 2013;

13

Deleted: (SNMs),

Deleted: products

(Deleted: kind of
(Deleted: Where
(Deleted: is
(Deleted: , then N loss can occur.
\mathcal{A}	Deleted: assessing

Deleted: (ɛ

Formatted: Font: Times New Roman

Formatted: Font: Times New Roman

Buchwald et al., 2015; Peters et al., 2016). A model can either assume a set of processes and infer the underlying isotope effects, or assume isotope effects and infer a set of processes. These isotope models are highly dependent on the chosen isotope effects used for given processes. Though there are estimates of isotope effects for processes based on both environmental measurements and <u>Jaboratory</u> studies, there is not always agreement between them. For example, laboratory cultures of NO²⁻

5 oxidizers indicate an N isotope effect of ¹⁵ε = <u>10 to -20</u>‰ (Casciotti, 2009; <u>Buchwald and Casciotti, 2010</u>), while measured concentrations and isotopes of NO₃⁻ and NO₂⁻ in ODZs indicate that <u>isotope effects closer to -30‰ are more consistent with</u> the observations (Buchwald et al., 2015; Casciotti et al., 2013; <u>Peters et al., 2016</u>).

 Here we present an expansion of an existing global ocean 3D inverse isotope-resolving N cycling model (DeVries et al., 2013)
 to investigate the isotopic constraints on N cycling in ODZs and the impact of these regions on global ocean N isotope patterns. <u>An important step was to</u> include NO₂⁻ and its isotopes as tracers. The addition of NO₂⁻ allows us to include additional internal N cycling processes, as well as a more nuanced and realistic version of the processes occurring in ODZs. We used a database of NO₃⁻ and NO₂⁻ observations in order to assess the performance of the model as well as optimize the model N cycle parameters for which we do not have good prior estimates. In the model we employ a variety of isotope effect estimates for

15 three important ODZ processes—NO₃⁻ reduction, NO₂⁻ reduction, and NO₂⁻ oxidation—to discern what isotope effect estimates result in the best fit to the observations

2 Methods

2.1 Inverse nitrogen cycle model overview

- The model used here is a steady-state inverse model that solves for the concentration and δ¹⁵N of NO₅⁻, NO₂⁻, particulate organic N (PON), and dissolved organic N (DON) using a set of linear equations. Because the model assumes that the system is in steady state, it is not able to capture time_z dependent properties of the system such as seasonality and anthropogenic change. However, on inter-annual timescales the N cycle is thought to be approximately in balance (Gruber, 2004; Bianchi et al., 2012). The residence time of N in the ocean, which is thought to be on the order of 2000-3000 years (Gruber, 2008), is sufficiently long to preclude any detectable changes in the global N inventory to date on timescales commensurate with the
- 25 global overturning circulation. An important advantage of the steady-state assumption for our linear model is that it is possible to find solutions by direct matrix inversion without the need for a spin-up period as required by forward models. The solution to the system provides ¹⁴N and ¹⁵N concentrations of the N species of interest at every grid point in the model system. Working with a linear system imposes some restrictions on how complicated the rate equations can be, but there are improvements in model performance and ease of use, allowing us to test hypotheses about the processes that govern the marine N cycle and
- 30 budget, particularly those occurring in and around oceanic oxygen deficient zones. We aimed to produce a realistic N cycle model that represented ODZ processes accurately while limiting the number of free parameters. The description below outlines the dependencies and simplifications employed in this version of the model,

Deleted: in-lab	
Deleted: 13	
Deleted: an	
Deleted: effect	
Deleted: is	

Deleted: many

Deleted: This should result in a reasonably accurate global representation of NO₃⁻ and NO₂⁻ concentrations and isotopes.

Formatted: Font: Times New Roman

Oeleted:

Deleted: is		

Deleted:

Deleted:

The model's uncertain biological parameters were determined through an optimization process that minimizes the difference between the modeled and observed NO_3 ⁻ and NO_2 ⁻ concentration and isotope data. Computational time limits the number of

parameters that we were able to optimize. We therefore <u>focused</u> our investigation on parameters that are poorly constrained
by literature values and to which the model solution is most sensitive. In order to determine the parameters for optimization, a sensitivity analysis was performed on each parameter, varying them individually by ±10% and computing the change in the modeled ¹⁴N and ¹⁵N. <u>Those parameters that resulted in modeled ¹⁴N and ¹⁵N</u> variability of >5% were chosen for optimization in the model. The sensitivity analysis and the optimal values of the parameters contribute to an improved understanding of the cycling of nitrogen in the ocean in general and in the ODZs in particular. <u>The optimization process is discussed in further</u>
detail in Section 2.6.

The sensitivity analysis revealed that the modeled distribution of ${}^{15}N$ was very sensitive to isotope effects, parameters that control the relative rates of ${}^{15}N$ and ${}^{14}N$ in chemical and biological processes. There are literature estimates <u>for</u> each of the isotope effects of interest in this work, although there is often a discrepancy between isotope effects estimated in laboratory

- 15 studies and those expressed in oceanographic measurements (<u>Kritee et al., 2012;</u> Casciotti et al., 2013; <u>Bourbonnais et al.,</u> 2015; Martin and Casciotti, 2017; Fuchsman et al., <u>2017; Marconi et al.,</u> 2017; Peters et al., 2018b). Rather than optimizing the isotope effect values, we have chosen to use multiple cases with different combinations of previously estimated isotope effects in order to assess which values best fit the observations.
- 20 In addition to the optimized parameters and isotope effects, there were some non-sensitive parameters that were fixed prior to the optimization and whose values were chosen using literature estimates (Table 1). Some N cycle processes are also dependent on prescribed input fields that are not explicitly modeled, such as temperature, phosphate, <u>oxygen</u>, and net primary production. These external input fields will be discussed in detail in the relevant sections for each N cycle process.

2.2 Model grid and transport

- 25 The model uses a uniform 2°x2° grid with 24 depth levels. The thickness of each model layer increases with depth, from 36 m at the top of the water column to 633 m near the bottom. Bottom topography was determined using 2-minute gridded bathymetry (ETOPO2v2) that was then interpolated to the model grid._Our linear N cycle model relies on the transport of dissolved N species (NO₃⁻, NO₂⁻, and DON) in the ocean. For this we use the annual averaged circulation as captured by a tracer transport operator that governs the rate of transport of dissolved N species (NO₃⁻, NO₂⁻, and DON) between boxes. The
- 30 original version of the tracer data-assimilation procedure used to generate the transport operator for dissolved species (T_i) is described by DeVries and Primeau (2011), and the higher resolution version used here is described by DeVries et al. (2013).

15

Deleted: focus

Deleted: Those exhibiting

(Deleted: Buchwald et al., 2015;

2.3 N cycle

In the N cycling portion of the model, we track four different N species, (Figure 1). There are two organic N (ON) pools: <u>dissolved (DON)</u> and <u>particulate (PON)</u>. There are also two dissolved inorganic N (DIN) pools: NO₃⁻ and NO₂⁻. We <u>did not</u> <u>explicitly model</u> ammonium (NH₄⁺) because it typically <u>occurs</u> in low concentrations throughout the ocean, and scarcity of

5 data (especially ¹⁵N data) would make model validation difficult. <u>Though NH4+ has been observed to accumulate to micromolar</u> <u>concentrations in some ODZs</u> (Bristow et al., 2016; Hu et al., 2016), this occurs largely in shallow, coastal shelf regions that <u>are not resolved by the model</u>.

Because we <u>used the concentrations of both ¹⁴N and ¹⁵N of each N species to constrain the rate parameters, two sets of</u> governing equations were employed: one that depends on ¹⁴N and another that depends on ¹⁵N. Generally, the rate for ¹⁵N processes was dependent on the rate of ¹⁴N processes and an isotopic fractionation factor (a) that is specific to each process and substrate. By solving for steady-state solutions to both ¹⁴N and ¹⁵N concentrations, we were able to model global distributions of [NO₅⁻], [NO₂⁻], and their corresponding δ ¹⁵N values.

2.3.1 N cycle parameterization

30

15 We will first address the ¹⁴N equations and the general format of the N cycle in the model. Each equation is then broken down into its component parts for further explanation of the biological processes and their parameterization. The ¹⁵N equations and isotope implementation will be discussed in a later section.

The governing equations for the ¹⁴N-containing DIN (NO₁²⁻ and NO_2^{-}) and organic N (DON and PON) state variables can be 20 written as follows:

 $1. \quad \left[\frac{\partial}{\partial t} + T_{f}\right]^{14} NO_{3}^{-} = J_{14}^{dep} - J_{14}^{assim,NO3} - J_{14}^{NAR} + J_{14}^{NXR} + 0.3 J_{14}^{AMX} - J_{14}^{sed}$

$$\left[\frac{\partial}{\partial t} + T_{f}\right]^{14}NO_{2}^{-} = J_{14}^{AMO} - J_{14}^{assim,NO2} + J_{14}^{NAR} - J_{14}^{NXR} - J_{14}^{NIR} - 1.3J_{14}^{AMX}$$

- 3. $\left[\frac{\partial}{\partial t} + T_f\right] DO^{14}N = \sigma(J_{14}^{fix} + J_{14}^{assim,WOA}) + J_{14}^{sol} J_{14}^{remin}$
- 25 4. $\left[\frac{\partial}{\partial t} + T_p\right] PO^{14}N = (1 \sigma)(J_{14}^{fix} + J_{14}^{assim,WOA}) J_{14}^{sol}$

The model is designed to represent a steady state, thus the $\frac{\partial}{\partial t}$ term is 0. The J terms represent the source and sink processes for each state variable, expressed in units of mmol/m³/yr and will be described in more detail below. Briefly, J_{14}^{dep} is the spatiallyvariable deposition of NO₃⁻ from the atmosphere to the sea surface. In the DIN model equations, $J_{14}^{assim,NO3}$ and $J_{14}^{assim,NO3}$ represent the assimilation of NO₃⁻ and NO₂⁻, respectively, by phytoplankton in the upper two box levels. This assimilated NO₃⁻

Deleted:	are	interested	iı

Deleted: from the model

Deleted: only accumulates

shown in Figure 1

Deleted: A schematic diagram of the N cycle in the model is

Deleted:

Deleted: . Deleted: excluded

Deleted: concentrations

Deleted: ON Deleted: DIN

Deleted: assumed Deleted: be in

produces DON and PON, with proportions set by a spatially-variable $\underline{term}_{,\underline{\sigma}}$. Assimilation in the DON and PON equations is represented by $J_{14}^{assim,WOA}$ and is dependent on 2013 World Ocean Atlas (WOA) [NO₃⁻] <u>interpolated to the model grid. N2</u> fixation (\int_{14}^{15}) is split between DON and PON with the same $\underline{\sigma}$ term. NO₃⁻ reduction (\int_{14}^{NAR}), NO₂⁻ reduction (\int_{14}^{NIR}), and anammox (\int_{14}^{AMX}) act on the NO₃⁻ and NO₂⁻ pools. J^{sed} represents the removal of NO₃⁻ via benthic denitrification. J^{sed} represents the dissolution of PON into DON. J^{semin} represents the degradation of DON, which feeds into

ammonia oxidation (J_{14}^{AMO}) and J_{14}^{AMX} as described below.

Through the use of these J terms, the governing equations are all linear with respect to the state variables. <u>However</u>, in order to introduce dependence of rates on the concentrations of multiple state variables, for example allowing heterotrophic NO_{3}^{2}

10 reduction to be dependent on organic N as well as NO₃⁻, we run the <u>organic N</u> equations first and the DIN equations second. When [DON] is found in the [DIN] governing equations, that [DON] value has already been determined for each grid box from the <u>organic N</u> model. When [NO₃⁻] is found in the DON governing equations, it is drawn from 2013 World Ocean Atlas, annual data interpolated to the model grid.

2.3.2 N source processes

15 Atmospheric deposition and N₂ fixation are the only sources of new bioavailable N in the model. These are the two largest sources of N to the ocean (Gruber and Galloway, 2008). We do not consider the third largest source of N, riverine fluxes, in the model due to lack of coastal resolution and the expectation that much of the river-derived N is denitrified in the shelf sediments (Nixon et al., 1996; Seitzinger and Giblin, 1996). Representing these processes may be possible in a future version of the model, but is beyond the scope of the current model, given its coarse resolution near the coasts.

20

Atmospheric deposition

N deposition is assumed to only occur in the top box of the model, and we assume that most of the N deposited is as NO_3^- , and that the other species would be rapidly oxidized to NO_3^- in the oxic surface waters.

25
$$\underline{5.} J_{14}^{dep} = r_{14}^{dep} S^{dep}$$

To calculate I_{14}^{dep} , the atmospheric deposition rate of ¹⁴N, we use modeled total inorganic N deposition for 1993, S^{dep} (Galloway et al., 2004; Dentener al., 2006; data available et online at https://daac.ornl.gov/CLIMATE/guides/global N deposition maps.html), which was interpolated to our model grid. This term, S^{dep} , is then multiplied by a prescribed fractional abundance of ¹⁴N in the deposited N (r_{14}^{dep}), which is calculated from 30 the isotopic composition of deposited N ($\delta^{15}N_{dep}$, 4‰; Equation 6), to yield the deposition of ¹⁴N to the sea surface in each

ns is	· Deleted: σ
L N2	Deleted: as described below.
NO ₂ -	Deleted: o
nthic	
into	Deleted: is related to
order	Deleted: In
<u>NO3</u> =	
ond.	Deleted: ON
box	
Atlas	Deleted: ON
	Deleted: (WOA13)
	Formatted

Deposition of N to the ocean from the atmosphere is one of two

Formatted: Indent: Left: 0.5", No bullets or numbering

sources of bioavailable N supplied to the ocean in this model.

Deleted: Atmospheric deposition

box (I_{14}^{dep}) . To calculate r_{14}^{dep} from $\delta^{15}N_{dep}$, we first calculate r_{15}^{dep} using r_{15}^{air} , a standard with a value of 0.003676 (Equation 6: Mariotti, 1983).

6.
$$r_{15}^{dep} = \left(\frac{\delta^{15}N_{dep}}{1000} + 1\right) \times r_{15}^{aii}$$

Then, assuming that $[^{14}N] \sim [^{14}N] + [^{15}N]$, we calculate r_{14}^{dep} as $(1 - r_{15}^{dep})$. The units of S^{dep} are given in mg N/m²/yr, which we convert to mmol NO₃⁻/m³/yr via dimensional analysis and by dividing by the depth of the surface box. This source term of N to the model is independent of the modeled N terms.

10 N₂ fixation

5

20

 N_2 fixation is the other source of new N to the model, and is assumed to only occur in the top box of the model. It is parameterized similarly to N_2 fixation in the model of DeVries et al. (2013), with partial inhibition by NO₃⁻ (Holl and Montoya, 2005) and dependence on iron (Fe) and phosphate (PO4³⁻) availability (Monteiro et al., 2011).

15 7.
$$J_{14}^{\text{fix}} = r_{14}^{\text{fix}} F_0 e^{-N0_{3,0bs}/\lambda} e^{\frac{T_{obs}-T_{max}}{T_0}} \frac{Fe}{Fe+K_{Fe}} \frac{PO_4}{PO_4+K_P}$$

Fo is the maximum rate of N₂ fixation (1.5 mmol/m³/yr; Table 1) and is calculated from the estimated areal rate of N₂ fixation in the western tropical Atlantic (Capone et al., 2005) divided by the depth of the top model box. NO_{3,obs} is the 2013 World Ocean Atlas, annually averaged surface NO₃⁻ interpolated to the model grid (Garcia et al., 2014). λ is an inhibition constant for N₂ fixation in the presence of NO₃⁻ (Table 1).

The temperature (T) terms scale the rate of N_2 fixation based on the observed temperature (T_{obs}), maximum observed sea surface temperature (T_{max}), and the minimum preferred growth temperature for *Trichodesmium* (T_0 ; Capone et al., 2005). The temperature data were taken from 2013 <u>World Ocean Atlas</u> annually averaged temperature interpolated to the model grid

25 (Locarnini et al., 2013). We recognize that this will likely provide a conservative estimate of N₂ fixation, given the growing recognition of N₂ fixation outside of the tropical and subtropical ocean by organisms other than *Trichodesmium* (Shiozaki et al., 2017; Harding et al., 2018; Landolfi et al., 2018).

Fe is the modeled deposition of soluble Fe interpolated to the model grid (mmol Fe/m²/yr; Chien et al., 2016) divided by the 30 depth of the top model grid box to give units of mmol Fe/m³/yr. Fe and PO4³⁻ are assumed to limit N₂ fixation at low concentrations via Michaelis-Menten kinetics. K_{Fe} and K_P are their respective half-saturation constants. Additionally, there is a term that allows us to set the isotopic ratio of newly fixed N, r_{14}^{fix} , which is the fractional abundance of ¹⁴N in newly fixed N

18

Deleted: equation 6).

Formatted: Indent: Left: 0.5", No bullets or numbering

Deleted: (WOA)
Deleted: `.

Deleted: WOA

and is calculated as in Equation 6 from $\delta^{15}N_{fix}$ (-1‰; Table 1). All of the N₂ fixation parameters are fixed rather than optimized (Table 1). Due to the use of non-optimized parameters and an input NO₃⁻ field rather than modeled NO₃⁻, N₂ fixation serves as an independent check that our modeled N cycle produces reasonable N concentrations and overall N loss rates. However, N₂ fixation is not explicitly modeled here and is instead taken as a fixed, though spatially variable, input field₄(Figure S1). The

5 global rate of N₂ fixation produced by this parameterization is 131 Tg N/yr, which is in line with several current estimates (Table S1).

In the model, N₂ fixation and NO₃⁻ assimilation (Section 2.3.3) are assumed to be the two processes that create exportable ρ rganic N. A fraction, σ , of this ρ rganic N is portioned into DON rather than PON (Equations 3-4). In order to create spatial variability in this constant, we assumed (1- σ), the fraction of assimilated N partitioned to PON, is equal to the particle export (P_{c}) ratio. This P_{c} ratio is the ratio of particle export to primary production, and is equivalent to the fraction of ρ rganic N that is exported from the euphotic zone as particulate matter rather than recycled or solubilized into DON. The P_{c} ratio is calculated for each model grid square from the mixed layer temperature (T_{ml}) and net primary production (NPP) as described by Dunne et al. (2005):

15

8. $P_e = \phi T_{ml} + 0.582 \log(\text{NPP}) + 0.419$

The constant φ has a value of -0.0101 °C¹ as determined by Dunne et al. (2005). Net primary production estimates in units of mmol carbon/m²/yr were taken from a satellite-derived productivity model (Westberry et al., 2008), annually averaged, and
 interpolated onto the model grid. T_{ml} is calculated from the 2013 World Ocean Atlas annual average (Locarnini et al., 2013), which has been interpolated to the model grid. The temperature of the top two model boxes were averaged to give T_{ml}. As temperature increases, the P_e ratio decreases and less PON is exported, resulting in more DON recycling in the surface with several possible explanatory mechanisms discussed in greater detail by Dunne et al. (2005). As anterprimary production increases, the P_e ratio increases and more PON is exported; and primary production explains 74% of the observed variance in

25 particle export (Dunne et al., 2005).

2.3.3 Internal N cycling processes

Assimilation of nitrate and nitrite

Assimilation accounts for the uptake of DIN and its incorporation into <u>organic matter in the shallowest two layers of the global</u> model. Since assimilation affects both the <u>organic and inorganic N</u> pools, we must account for it in both sets of model runs.
 We will first address assimilation in the organic N model (Equations 9 and 10).

9.
$$J_{14}^{assim,WOA} = {}^{14}k_{assim}[NO_3^-]_{ob}$$

Deleted: OM	
Deleted: OM	
Deleted: pe	
Deleted: pe	
Deleted: OM	
Deleted: pe	
Deleted: pe =	
Deleted: ϕ has a	
Deleted: o has a	
Deleted: . NPP	
Deleted: WOA	
Deleted: pe	
Deleted: NPP	
Deleted: pe	
Deleted: NPP	

Deleted:

Deleted: OM.	
Deleted: ON	
Deleted: DIN	

$$\underline{10.}^{14}k_{assim} = \frac{_{NPP}}{_{r_{C:N}[NO_3^-]_{obs}}}$$

Since the organic N model is run first and the assimilation rates are dependent on DIN concentrations, assumptions must be made about the DIN field in order to account for assimilation prior to the DIN model runs. Here we used observed [NO3] from 5 the 2013 World Ocean Atlas annual product interpolated to the model grid [NO3⁻]obs (Garcia et al., 2014) to calculate the

- assimilation rates for DON and PON production ($\int_{14}^{assim,WOA}$). For this assumption to be valid, our modeled surface [NO₃⁻] must be close to the observed values, which we will test in Section 3.1. The rate constant for assimilation, $^{14}k_{assim}$, varies spatially and is determined using observations of surface [NO3-] and satellite derived net primary production estimates (NPP: Westberry et al., 2008). The rate constant is converted to N units using the ratio of carbon (C) to N in organic matter (rc:N)
- 10 which we assume to be 106:16 (Redfield et al., 1963). The value of the rate constant is only non-zero in the top two boxes of the model, where we assume primary production to be occurring. The same rate constant is used in both the organic N and DIN assimilation equations. We also assume from the perspective of organic N that only NO3- is being assimilated, since NO2is present at relatively low concentrations in the surface ocean, and it may be characterized as recycled production. Assimilated N is partitioned between PON and DON using the Pe ratio as previously described and shown in Equations 3 and 4.

15

20

The setup for assimilation in the DIN model (Equations 11 and 12) is similar, but can use modeled [NO3⁻] and [NO2⁻] rather than the World Ocean Atlas values. In order to appropriately reflect surface NO3 and NO2 concentrations, both NO3 and NO2 are assimilated. ¹⁴k_{assim} is calculated as described above and is assumed to be the same for both NO3⁻ and NO2⁻. We justify using only [NO₃⁻] to parameterize ${}^{14}k_{assim}$ because NO₃⁻ generally makes up the bulk of DIN available for assimilation at the surface, but this assumption will be discussed in more detail below.

11.
$$J_{14}^{assim,NO3} = {}^{14}k_{assim}[{}^{14}NO_3^-]$$

12. $J_{14}^{assim,NO2} = {}^{14}k_{assim}[{}^{14}NO_7^-]$

25 Solubilization

Solubilization is the transformation of PON to DON, and is dependent only on [PON] and a solubilization rate constant (14ksol), which is optimized (Table 2).

13.
$$J_{14}^{sol} = {}^{14}k_{sol}[PO^{14}N]$$

30

The solubilization of PON, together with the particle transport operator (T_p), produces a particle flux attenuation curve similar to a Martin curve with exponent b = -0.858 (Table 1) (Martin et al., 1987). While in the real world, the length scale for particle flux attenuation is somewhat longer in ODZs compared to oxygenated portions of the water column, and also varies regionally

20

Deleted: ON

Deleted: We will first address assimilation in

Deleted: ON equations.

Deleted: must be calculated using observed surface [NO3"], rather than modeled [NO3"].

Deleted: We also assume from the perspective of ON that only NO3" is being assimilated, since NO2" is present at relatively low concentrations in the surface ocean and it may be characterized as recycled production. Assimilated N is partitioned between PON and DON using the pe ratio as previously described and shown in Equations 3 and 4.¶ $J_{14}^{assim} = {}^{14}k_{assim}[NO_3^-]_{obs}$ ¶

 $\frac{V_{14}}{1^4 k_{assim}} = \frac{\frac{NPP}{r_{cN} |N\sigma_1^-|os|}}{\frac{NPP}{r_{cN} |N\sigma_1^-|os|}}$ interpolated to the model grid (Garcia et al., 2014).

Deleted: (NPP)

Formatted: Font: Italic

Deleted: ON and DIN assimilation equations

Deleted: equations Deleted: WOA

(Berelson et al., 2002; Buesseler et al., 2008; Buesseler and Boyd, 2009), our model uses a spatially-invariant ${}^{14}k_{sol}$. A spatially variable ${}^{14}k_{sol}$ that accounts for lower apparent values in ODZs is a refinement that could be introduced in future model versions.

Remineralization

5 Remineralization, or ammonification, is the release of DON into the DIN pool. This is determined using the concentration of DON and a remineralization rate constant (¹⁴k_{remin}), which is optimized (Table 2).

$$14. J_{14}^{remin} = {}^{14}k_{remin} [DO^{14}N]$$

10 The removal of this remineralized DON, since it does not accumulate as NH4⁺, is either through ammonia oxidation (AMO) or anammox (AMX), depending on [O₂] as described below and in Section 2.3.4. We use the same remineralization rate constant regardless of the utilized electron acceptor (e.g. O₂, NO₃⁻). Since particle flux attenuation is observed to be somewhat weaker in oxygen deficient zones compared with oxygenated water (Van Mooy et al., 2002), this may slightly overestimate the rates of heterotrophic remineralization occurring in ODZs.

15

20

Ammonia oxidation

Ammonia oxidation (AMO) uses ammonia (NH_3) as a substrate. Since we do not include NH_3 or NH_4^+ in the model system, we treat remineralized DON as the substrate for AMO. In order to maintain consistency between the <u>organic N and DIN model</u> runs, remineralized DON is routed either to AMO or AMX (lost from the system) based on the O₂ dependencies of AMO and AMX. Rather than using a strict O₂ cutoff for AMO, it is limited by O₂ using Michaelis-Menten kinetics. The half-saturation

constant for O_2 , K_m^{AMO} (Table 1), sets the O_2 concentration at which AMO reaches half of its maximal value.

15.
$$J_{14}^{AMO} = (1 - \eta_{AMX}) J_{14}^{rem} + \eta_{AMX} \frac{[O_2]}{[O_2] + K_m^{AMO}} J_{14}^{remin}$$

25 Recent studies have shown that AMO and NO₂⁻ oxidation (NXR), both O₂-requiring processes, have very low O₂ half saturation constants and can occur down to nM levels of [O₂] (Peng et al., 2015; Bristow et al., <u>2016</u>). In contrast, O₂-inhibited processes such as AMX are only allowed to occur at O₂ concentrations below a given threshold. The handling of O₂ thresholds for anaerobic processes is discussed in more detail below (Section 2.3.4), though we describe it briefly here due to the interplay between AMO and AMX in the model. Briefly, the O₂ dependence of AMX is represented by the parameter η_{AMX}, which has
30 a value between 0 and 1 for a given grid box depending on the average number of months in a year its <u>2013</u> World Ocean Atlas [O₂] threshold for anammox (O₂^{AMX}, Table 1). If, for example, [O₂] in a given grid box is always

above the threshold for AMX, $\eta_{AMX} = 0$ and all of the remineralized DON (represented by J_{14}^{rem}) will be oxidized via AMO. If [O₂] is less than O₂^{AMX}, η_{AMX} will be non-zero and a smaller fraction of the remineralized DON will be oxidized via AMO.

21

(Deleted: This

Formatted: Normal, No bullets or numbering

Formatted: Font: Times New Roman
Formatted: Font: Times New Roman
Deleted: ON

Deleted: 2016b

(Deleted:).

Deleted: WOA

The fraction ultimately oxidized by AMO is thus determined by the Michaelis-Menten parameterization of AMO, as well as the O_2 threshold for anammox.

Nitrite oxidation

5 The rates of NO₂⁻ oxidation (NXR) are dependent on the availability of NO₂⁻ as well as O₂. Similar to AMO, we parameterize O₂ dependence using Michaelis-Menten kinetics and a fixed half-saturation constant for O₂ (K_m^{NXR} , Table 1). K_m^{NXR} was taken to be 0.8 μ M O₂, based on kinetics experiments performed with natural populations of NO₂⁻ oxidizing bacteria (Bristow et al., 2016). Finally, we employ an optimized rate constant (¹⁴k_{NXR}, Table 2) to fit the available data.

10 16. $J_{14}^{NXR} = {}^{14}k_{NXR} [{}^{14}NO_2^-] \frac{[O_2]}{[O_2] + K_m^{NXR}}$

2.3.4 N sink processes

Nitrate and nitrite reduction

1	4
1	

 NO_3^- reduction (NAR) and NO_2^- reduction (NIR) are two processes within the stepwise reductive pathway of canonical denitrification. The end result of denitrification is the conversion of DIN to N_2 gas, rendering it bioavailable to only a restricted set of marine organisms. Although there are intermediate gaseous products between NO_2^- and N_2 , we treat <u>NIR</u> as the rate-limiting step in the denitrification pathway, where DIN is removed from the system.

For both NAR and NIR, we introduce a dependency on two state variables, their respective N substrates, and organic matter availability. Where NAR and NIR occur heterotrophically, they consume organic matter in addition to their main N

- 20 substrates/electron receptors. When NAR occurs chemoautotrophically, it would be dependent primarily on the presence of NO₃⁻ and an electron donor, such as hydrogen sulfide (Lavik et al., 2009). Since we do not model the production of reduced sulfur species in our model, our estimates of denitrification would not explicitly include the effects of this process. However, chemolithotrophic denitrification could be tacitly accounted for in the optimization process, since the rate constants that control the rates of NAR and NIR are optimized in order to best fit the observations, and the isotope effect for chemolithotrophic
- 25 denitrification is thought to be similar to that of heterotrophic denitrification (Frey et al., 2014). In order to maintain levels of <u>heterotrophic</u> NAR and NIR that are bounded both by the available NO₃⁻ or NO₂⁻ and the available organic matter in a linear model, it was necessary to run<u>organic N</u> and DIN equations separately, since it is not possible to include dependencies on two state variables (e.g. DON and NO₃⁻) in the linear system. Both NAR and NIR are dependent on the remineralization rate (*J*^{remin}₁) that is calculated in the <u>organic N</u> model run. In model boxes where NAR and NIR are occurring, some of the
- 30 remineralization is carried out with electron acceptors other than O₂. For simplicity, we assume that J^{remin}₁₄ does not depend on the choice of electron acceptor.

Deleted: µM

Deleted: as

Deleted: NO2" reduction

Deleted: . Since both NAR and NIR are heterotrophic processes, they consume organic matter in addition to their main N substrates/electron receptors. In order to maintain levels of

Deleted: ON

Deleted: ON

$$\begin{split} & 17. \; J_{14}^{NAR} = \eta_{NAR} \, {}^{14}k_{NAR} \big[{}^{14}NO_3^- \big] \, J_{14}^{remin} \\ & \underline{18.} \; J_{14}^{NIR} = \eta_{NIR} \, {}^{14}k_{NIR} \big[{}^{14}NO_2^- \big] \, J_{14}^{remin} \end{split}$$

The rate coefficients for NAR (${}^{14}k_{NAR}$) and NIR (${}^{14}k_{NIR}$) are optimized rather than fixed (Table 2). Further, the dependence 5 of J_{14}^{NAR} and J_{14}^{NIR} on J_{14}^{remin} means that k_{NAR} and k_{NIR} are not first order rate constants and have different units than k_{PON} , k_{DON} , and k_{NXR} (Table 2).

The inhibition of NAR and NIR by O₂, like AMX, is parameterized by a parameter η , which inhibits these processes when [O₂] is above their maximum threshold. Originally, we treated this term as a binary operator where it would be 0 if the

- 10 empirically-corrected 2013 <u>World Ocean Atlas annually averaged [O2]</u> is above the threshold for the process and 1 if [O2] is below the threshold. On further refinement, we wanted to account for the possibility of seasonal shifts in [O2] in ODZs. Thus, for each month, we assigned a value of 0 or 1 to each model grid box. These values were then averaged over the 12 months of the year to give a sliding value of η between 0 and 1 for each grid box. The O2 thresholds used to calculate η_{NAR} and η_{NIR} were fixed (7 µM and 5 µM, respectively; Table 1). Since we do not explicitly model O2, [O2] was predetermined using the
- 15 2013 World Ocean Atlas monthly O2 climatology (Garcia et al., 2014) interpolated to the model grid. We also applied an empirical correction that improves the fit of World Ocean Atlas [O2] data to observed suboxic measurements (Bianchi et al., 2012).

Anammox

20 Anammox (AMX) catalyzes the production of N₂ from NH₄⁺ and NO₂⁻. Since we do not use NH₄⁺ as a variable in our N cycling equations, we substituted remineralized DON (J_{14}^{remin}) as a proxy for NH₄⁺ availability. As described above in Section 2.3.3, remineralized DON is routed through either AMO or AMX depending on [O₂] and the O₂ dependencies of AMO and AMX.

19.
$$J_{14}^{AMX} = \eta_{AMX} \left(1 - \frac{[O_2]}{[O_2] + K_m^{AMO}}\right) J_{14}^{remin}$$

25

The O₂ threshold used to calculate η_{AMX} from monthly O₂ climatology is fixed (10 µM; Table 1). In order to maintain mass balance on remineralized DON, we do not include dependence on [NO₂⁻] in <u>Equation 19</u>, although f_{14}^{AMX} removes NO₂⁻ (<u>Equation 2</u>). This parameterization inherently assumes that AMX is limited primarily by [NH₄⁺] supply and not [NO₂], which may not always be correct (Bristow et al., <u>2016</u>). Anammox also produces 0.3 moles of NO₃⁻ via associated NXR for every 1

30 mole of N₂ gas produced (Strous et al., 1999). For this reason, anammox appears in the state equation for NO₃⁻ (Equation 1).

Deleted: WOA

Deleted: WOA

Deleted: WOA

(Deleted: the rate equation	
(Deleted: equation	
(Deleted: 2016a	

Sedimentary denitrification

Sedimentary (or benthic) denitrification (f_{14}^{sed}) is an important loss term for N in the marine environment, and in order to encapsulate it within the model grid we assume that it is occurring within the bottom depth box for any particular model water column. The parameterization for sedimentary denitrification is based on a transfer function described by Bohlen et al. (2012).

5 The original transfer function was dependent on bottom water [O₂], bottom water [NO₃⁻], and the rain rate of particulate organic carbon (RRPOC). <u>Here, RRPOC was calculated via a Martin curve (Martin et al., 1987) using the *P_e* ratio, net primary production (NPP), depth (*z*), euphotic zone depth (*z_{eu}*), and a Martin curve exponent (*b*):</u>

20. RRPOC = NPP * $P_e * (\frac{z}{z_{ev}})^b$

10

Net primary production is derived from the productivity modeling of Westberry et al. (2008) as described in Section 2.3.2. The P_{c} ratio is calculated as previously described in Section 2.3.2. The depth for any given model box is assumed to be the depth at the bottom of the box. The euphotic zone depth is the bottom depth of the 2nd box (73 m), since all production is assumed to be occurring in the top two boxes. As described above, the Martin curve exponent, *b*, is a fixed value in our model (p = -0.85). Table 1), the web this mean model above is a fixed value in our model (p = -0.85).

15 0.858; Table 1), though this may result in underestimation of the particulate matter reaching the seafloor below ODZs (Van Mooy et al., 2002).

The transfer function for sedimentary denitrification was originally described using a non-linear dependence of the rate on $([O_2] - [NO_3^-])$. In order for sedimentary denitrification to be properly implemented in our linear model, we broke the original

- 20 non-linear relationship into three roughly linear segments, to create a piecewise relationship between ([O₂] [NO₃⁻]) and sedimentary denitrification rate (Figure S2). We obtained three linear relationships between ([O₂] [NO₃⁻]) and sedimentary denitrification rate, each applicable across a given range of ([O₂] [NO₃⁻]) values. Due to the nature of our linear model, we needed to express the interval cutoff points that define the transition between the piecewise relationship segments in terms of O₂ rather than ([O₂] [NO₃⁻]). Therefore, a linear relationship between O₂ and ([O₂] [NO₃⁻]) was determined using the 2013
 25 World Ocean Atlas annually averaged data (Garcia et al., 2014; Figure \$3). The cutoff points were determined to be 75 µM
- O_2 and 175 μ M O_2 . The linear relationships were then rearranged in order to estimate sedimentary denitrification rate as a function of RRPOC, [O₂], and [NO₃⁻]. These equations were then further broken down into a component that is dependent on [NO₃⁻] and a component that is dependent on [O₂],
- 30 An additional term is introduced that reduces the sedimentary denitrification rate by 27% if the depth of the bottom model box is less than 1000 m. This term represents the potential for efflux of NH4⁺ into the water column from shallow, organic rich shelf sediments (Bohlen et al., 2012). This decreases overall sedimentary denitrification by approximately 6 Tg N/yr. This transfer function also assumes that all of the NH4⁺ efflux is immediately oxidized to NO3⁻ and does not alter its isotopic

Moved (insertion) [1] Moved up [1]: Here, RRPOC was calculated via a Martin cur (Martin et al., 1987) using the Deleted: pe ratio, NPP, depth (2), euphotic zone depth (z_{en}), at Martin curve exponent (b): Deleted: $pe * (\frac{z}{z_{en}})^{6}$ Deleted: NPP Deleted: pe Formatted: Font: Italic	Del	leted: as
Moved up [1]: Here, RRPOC was calculated via a Martin cu (Martin et al., 1987) using the Deleted: pe ratio, NPP, depth (z), euphotic zone depth (z_{en}), at Martin curve exponent (b): Deleted: $pe * (\frac{z}{z_{en}})^b$ Deleted: NPP Deleted: pe Formatted: Font: Italic		
(Martin et al., 1987) using the Deleted: pe ratio, NPP, depth (z), euphotic zone depth (z_{eu}), and Martin curve exponent (b): Deleted: $pe * (\frac{z}{z_{eu}})^b$ Deleted: NPP Deleted: pe Formatted: Font: Italic	Мо	ved (insertion) [1]
(Martin et al., 1987) using the Deleted: pe ratio, NPP, depth (z), euphotic zone depth (z_{eu}), and Martin curve exponent (b): Deleted: $pe * (\frac{z}{z_{eu}})^b$ Deleted: NPP Deleted: pe Formatted: Font: Italic		
(Martin et al., 1987) using the Deleted: pe ratio, NPP, depth (z), euphotic zone depth (z_{eu}), and Martin curve exponent (b). ⁴ Deleted: $pe * (\frac{z}{z_{eu}})^b$ Deleted: NPP Deleted: pe Formatted: Font: Italic		
Martin curve exponent (b): Deleted: $pe * (\frac{z}{z_{eu}})^b$ Deleted: NPP Deleted: pe Formatted: Font: Italic		
Deleted: NPP Deleted: pe Formatted: Font: Italic		
Deleted: pe Formatted: Font: Italic	Del	leted: $pe * (\frac{z}{z_{eu}})^b$
Formatted: Font: Italic	Del	leted: NPP
	Del	leted: pe
	For	matted: Font: Italic
Formatted: Font: Italic	For	rmatted: Font: Italic
	Del	leted:
Deleted:		
Deleted: .	Del	leted: (Figure S1).
Deleted: . Deleted: (Figure S1).		leted: our

Deleted: sediments

Deleted: WOA

Deleted	
Deleted	In organic-rich shelf sediments an
Deleted	a globally averaged
Deleteu	a globally averaged
C	
	1000 1 /

composition in bottom water. This is a conservative estimate of the effects of benthic N loss on water column NO₃⁻ isotopes, as several studies suggest that benthic N processes may contribute to water column nitrate ¹⁵N-enrichment (Lehmann et al., 2007; Granger et al., 2011; Somes et al., 2015; Brown et al., 2015). However, our current model parameterization does not require enhanced fractionation during benthic N loss to fit deep ocean δ^{15} NNo₃. Additionally, our spatial resolution does not

 $5 \quad \underline{\text{well represent regions where this effect might be significant on bottom water } \delta^{15}N_{NO3}, \text{ such as the shallow shelves.}$

2.4 N isotope implementation

10

In our model, we are interested in using the isotopic composition of NO₃⁻ and NO₂⁻ to constrain the rates of N cycling and loss from the global ocean. As DON and PON are ultimate substrates for NO₂⁻ and NO₃⁻ production, it is essential to track the ¹⁵N in the <u>organic N</u> pools as well. The matrix setup for ¹⁵N is similar to that for the ¹⁴N species, but the rates were changed as follows:

21.
$$J_{15}^{process} = 1/\alpha_{process} \frac{[{}^{15}N_{substrate}]}{[{}^{14}N_{substrate}]} J_{14}^{process}$$

 $J_{14}^{process}$ is the rate of each relevant ¹⁴N process as described above, and $J_{15}^{process}$ is the rate of each ¹⁵N process. $\alpha_{process}$ is the 15 fractionation factor for a given process, which is given by the ratio between the rate constants for ¹⁴N and ¹⁵N ($\alpha = ^{14}k/^{15}k$). A fractionation factor greater than 1 indicates a normal isotope effect and a fractionation factor less than 1 indicates an inverse isotope effect. Several of these fractionation factors are well known, but others are more poorly constrained, especially when values are calculated from *in situ* concentration and isotope ratio measurements (Hu et al., 2016; Casciotti et al., 2013; Ryabenko et al., 2012). For this reason, we ran several model cases with different fractionation factors for NAR, NIR, and

20 NXR during the optimization process (Section 2.6, Table 3). The other fractionation factors were fixed (Table 1). In order to produce the ¹⁵N concentrations of N species from our observations to constrain the model, we calculated ¹⁵N/¹⁴N from measured ⁵¹N, and assumed that [¹⁴N] ~ [¹⁴N] + [¹⁵N], the measured concentration of <u>each</u> modeled N species.

This simple ¹⁵N implementation was used with fixed fractionation factors for remineralization ($\alpha_{remin} = 1$), solubilization ($\alpha_{sol} = 1$), assimilation ($\alpha_{assim} = 1.004$), sedimentary denitrification ($\alpha_{sed} = 1$), and AMO ($\alpha_{AMO} = 1$) (Table 1). Isotope effects for NAR (ε_{NAR}), NIR (ε_{NIR}), and NXR (ε_{NXR}) were varied in different combinations during model optimization (Table 3). Distinct isotopic parameterizations were also required for atmospheric deposition, N₂ fixation and anammox, as described below.

Atmospheric deposition

30 For atmospheric deposition of N, we prescribe a constant δ^{15} N value of -4‰ (Table 1), which can be related to the fractional abundance of ¹⁴N, previously described in Section 2.3.2 as r_{14}^{dep} , and the fractional abundance of ¹⁵N (r_{15}^{dep}) in deposited N. We multiply r_{15}^{dep} by S^{dep}, the estimated rate of total N deposition, to obtain J_{15}^{dep} .

25

Deleted: ON

(Formatted: Font: Times New Roman
~(Formatted: Font: Times New Roman
)(Formatted: Font: Times New Roman
(Deleted: $\delta^{15}N$
Ì	Formatted: Font: Times New Roman
Y,	Deleted: the
()	Formatted: Font: Times New Roman
Y	Formatted: Font: Times New Roman

Nitrogen fixation

Similar to atmospheric deposition, newly fixed N has a constant prescribed δ^{15} N value (-1‰; Table 1). In Section 2.3.2 we described r_{14}^{fix} , the fractional abundance of ¹⁴N in newly fixed N. Here we multiply the fractional abundance of ¹⁵N, r_{15}^{fix} , by the other terms in the N₂ fixation equation (Equation 6) to obtain the rate of ¹⁵N fixation.

Anammox

5

Anammox is the most complicated <u>process</u> to parameterize isotopically because it has three different <u>N</u> isotope effects associated with it. There is an isotope effect on both substrates <u>that are</u> converted to N₂ (NO₂⁻ and NH₄⁺), as well as for the associated NO₂⁻ oxidation to NO₃⁻. We assume that the fractionation factor for ammonium oxidation via AMX ($\alpha_{AMX,NH4}$) is 1, setting it to match the fractionation factor for AMO (α_{AMO} ; Table 1), both with no expressed fractionation since NH₄⁺ does not accumulate in the model. Since all remineralized DON must be routed either through AMO or AMX, this simplifies the mass balance and ensures that all remineralized ¹⁴N and ¹⁵N is accounted for. ¹⁵NO₂⁻ is removed with the isotope effects of NO₂⁻ reduction ($\alpha_{AMX,NIR}$) and NO₂⁻ oxidation ($\alpha_{AMX,NXR}$), in the expected 1:0.3 proportion (Brunner et al., 2013).

15 2.5 Model inversion

Once our N cycle equations were set up as described above, we input them into MATLAB in block matrix form. The equations were of the general form Ax = b. All model ocean boxes (200,160 in total) are accounted for in the matrices. Matrix A (400,320 x 400,320) contained the rate constants and other parameters that are multiplied by the vector of state variables, x (400,320 x 1). Vector x contained the state variables (i.e. [NO₃⁻] and [NO₂⁻] or [DON] and [PON]) to be solved for by the linear solver.

- 20 Vector *b* (400,320 x 1) contained the rates that were independent of the state variables, such as N₂ fixation and N deposition. Let us consider, as an example, the DIN model setup. The top left corner of matrix *A* would contain rate constants for processes that produce and consume NO₃⁻ that are also dependent on [NO₃⁻]. The top right corner of matrix *A* would contain rate constants for processes that produce and consume NO₃⁻ but are dependent on [NO₂⁻]. The bottom left corner of matrix *A* would contain rate constants for processes that produce and consume NO₃⁻ but are dependent on [NO₂⁻]. The bottom left corner of matrix *A* would contain rate constants for processes that produce and consume NO₂⁻ but are dependent on [NO₃⁻]. The bottom right corner of matrix *A*
- 25 would contain rate constants that produce and consume NO_2^- and are also dependent on $[NO_2^-]$. The top half of vector *x* would be $[NO_3^-]$ for each model box, and the bottom half of vector *x* would be $[NO_2^-]$ for each model box. The top half of vector *b* would be independent processes that produce or consume $[NO_3^-]$, and the bottom half of vector *b* would be independent processes that produce or consume $[NO_2^-]$.
- 30 In MATLAB, we used METIS ordering, which is part of SuiteSparse (http://faculty.cse.tamu.edu/davis/suitesparse.html) to order our large, sparse matrix A. We then used the built-in function umfpack with METIS to factorize matrix A. The built-in matrix solver mldivide was then used with the factorized components of matrix A and matrix b to solve for x.

(Formatted: Font: Times New Roman

Deleted: 200,160

Formatted: Font: Courier New

Formatted: Font: Courier New

2.6 Parameter optimization

There are many parameters in the model that control the rates of the different N cycle processes (Tables 1-3). Some of these parameters are well constrained by literature values, (Table 1). Others, such as the rate constants, were objects of our investigation and were optimized against available observations, (Table 2). For our optimization, we compared model output

- 5 using different parameter values to a database of NO₃⁻ and NO₂⁻ concentrations and isotopes. The database was originally compiled by Rafter et al. (in prep.) and has been expanded to include some additional unpublished data (Table <u>\$2</u>). All of the database observations were binned and interpolated to the model grid. If multiple observations occurred within the same model grid box, the values were averaged and a standard deviation was calculated. The database was divided randomly into a training set, used for optimization, and a test set, used to assess model performance. The same number of grid points with observations 10 was used in the training and test sets.
 - The optimization procedure used the MATLAB function fminunc to obtain values for the non-fixed parameters that minimized a cost function (Equation 22). In each iteration of the optimization, the model system was solved by running the ¹⁴N-prganic N model, ¹⁵N-prganic N model, ¹⁴N-DIN model, and ¹⁵N-DIN model. The modeled output [NO₃⁻], [NO₂⁻], δ¹⁵N_{NO3},
- 15 and $\delta^{15}N_{NO2}$ were compared to values from the database training set. Though DON and PON observations were not used to optimize the model, the open ocean and deep water NO₃⁻ values were useful in constraining the parameters that control PON solubilization and DON remineralization. The entire model was run using a set of initial parameter values (Table 2) and the optimized to alter those starting parameters until a minimum in the cost function was attained. We optimized the logarithm of the parameter values rather than the original parameters themselves so the unconstrained
- 20 optimization returned positive values. The transformed starting parameters and subsequent modified parameter sets were then fed back into the model equations as e^x , where *x* denotes the log-transformed parameter. The cost function in the optimization procedure is as follows:

$$22. \text{ Cost} = \frac{w_{NO3}}{n_{NO3} sd_{NO3}} \sum ([NO_3^-]_{model} - [NO_3^-]_{training})^2 + \frac{w_{NO2}}{n_{NO2} sd_{NO2}} \sum ([NO_2^-]_{model} - [NO_2^-]_{training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + \frac{w_{\delta NO2}}{n_{\delta NO2} sd_{\delta NO2}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO2}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO2}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,model} - \delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{\delta NO3} sd_{\delta NO3}} \sum (\delta^{15} N_{NO3,training})^2 + 25 \frac{w_{\delta NO3}}{n_{$$

The *w* terms are weighting terms <u>introduced</u> to scale the contributions of the four observed parameters to equalize their contributions to the cost function. The *n* terms and standard deviation (*sd*) terms were used to normalize the contributions of each measurement type to the cost function. Each *n* term is equal to the number of each type of measurement in the training data set (e.g. the number of $[NO_3^-]$ data points = n_{NO3}). The *sd* term is equal to the standard deviation of all the measurements of a given type (e.g. the standard deviation of all the $[NO_3^-]$ data points within the training set).

Deleted:

Deleted:

Deleted: S1

Formatted: Font: Courier New	
Deleted:	
Deleted: ON	
Deleted: ON	

Deleted: intended

Deleted: in order

In order to account for error in our model parameter estimates, we also iterated over several possible values for three of the most important isotope effects for processes in ODZs: NAR, NIR, and NXR_{v} (Table 3). We chose to iterate over these parameters rather than optimize them since there is a large range of estimates from the literature as to what these parameters might be We assigned different possible values for each of these parameters (Table 3), resulting in 12 possible combinations.

5 The optimization protocol was performed for each of those combinations and unique optimized parameter sets were obtained. The parameter results were then averaged (final values, Table 2) and their spread is categorized as the error (error, Table 2).

3 Results

3.1 Global model-data comparison

The simulations of NO₂⁻ distribution and its isotopic composition are the most unique features of this model in comparison to existing global models of the marine N cycle. As such, NO₂⁻ accumulation in ODZs is a feature that should be well-represented by the model in order to use it to test hypotheses about processes that control N cycling and loss in ODZs. Overall, we see NO₂⁻ accumulating at <u>200 m</u> in the major ODZs of the Eastern Tropical North Pacific (ETNP), Eastern Tropical South Pacific (ETSP), and the Arabian Sea (AS) (Figure 2), which is consistent with observations and expected based on the low <u>O2</u> conditions found there. However, accumulation of NO₂⁻ in the model ETSP was lower than expected. The model also accumulated NO₂⁻ in the Bay of Bengal, which is a low-O₂ region off the east coast of India that does not generally accumulate

NO₂⁻ or support water column denitrification, but is thought to be near the "tipping point" for allowing N loss to occur (Bristow et al., <u>2016)</u>. Possible reasons for the underestimation of NO₂⁻ in the ETSP and overestimation in the Bay of Bengal will be discussed further in Section 4.2.

20 The model optimization described above yielded a set of isotope effects that best fit the global dataset of [NO₃⁻], [NO₂⁻], δ¹⁵N_{NO3} and δ¹⁵N_{NO2}. The best fit was achieved for isotope effects of 13‰ for NO₃⁻ reduction (ε_{NAR}), 0‰ for NO₂⁻ reduction (ε_{NIR}), and -13‰ for NO₂⁻ oxidation (ε_{NXR}). Figure 3 shows the test set comparison for the global best-fit set of isotope effects overlaid with a 1:1 line, which the data would follow if there was perfect agreement between model results and observations. There is general agreement between model and observations, with most of the data clustering near the 1:1 lines. Agreement between the observations and the training data are similar (Figure \$4), indicating that we did not overfit the training data.

In the test set, there were some low $[O_2]$ points where our model $[NO_3^-]$ exceeded observations (Figure 3a, filled black circles); these are largely within the ETSP. In contrast, the Arabian Sea tended to show slightly lower modeled $[NO_3^-]$ than expected. The $[NO_2^-]$ accumulation (Figure 3b) and $\delta^{15}N_{NO2}$ signals (Figure 3c) in the ETSP were also generally too low compared with

30 observations. These signals are likely tied to insufficient NO₃⁻ reduction <u>occurring</u> in the <u>model ETSP</u>. Another consideration is that there may be a mismatch in resolution between the model and the space and time scales needed to resolve the high NO₂⁻ accumulations observed sporadically (Anderson et al., 1982; Codispoti et al., 1985; 1986).

28

 Deleted:

Deleted: (Table 3).

Deleted: many other

Deleted: 200m

Formatted: Font: Times New Roman

(Deleted: 2016a). The

Deleted: $\delta^{15}N_{NO3}$
Deleted: $\delta^{15}N_{NO2}$
Formatted: Font: Times New Roman
Formatted: Font: Times New Roman
Formatted: Font: Times New Roman
Deleted: S3
Deleted: Eastern Tropical South Pacific (
Deleted:), where insufficient NO3 ⁻ reduction occurred in the model.
Deleted: In addition, the
Deleted: $\delta^{15}N_{NO3}$
Deleted: , although excessive consumption of NO2 [°] may also play a role
Deleted:

Overall, the representation of $\delta^{15}N_{NO3}$ was fairly good (RMSE = 2.4%), though there were a subset of points above $\delta^{15}N_{NO3}$ = 10% where the modeled $\delta^{15}N_{NO3}$ exceeded the observed $\delta^{15}N_{NO3}$ and others where modeled $\delta^{15}N_{NO3}$ was lower than observations (Figure 3c). Many of the points with overestimated $\delta^{15}N_{NO3}$ were located within the Arabian Sea ODZ, where

5 there may be too much NO_3^{-1} reduction occurring, leading to excess enrichment in ¹⁵N-NO₃⁻¹. As indicated above, the underestimated $\delta^{15}N_{NO3}$ points largely fell within the ETSP where we believe the model is underestimating NO₃⁻¹ reduction. The representation of $\delta^{15}N_{NO2}$ was also fairly good (RMSE = 8.6‰), though the modeled $\delta^{15}N_{NO2}$ was generally not low enough (Figure 3d), indicating an underestimated sink of 'heavy' NO₂⁻¹.

3.2 Oxygen deficient zone model-data comparison using station profiles

- 10 To further investigate the distribution of model N species within the three main ODZs, we selected representative <u>offshore</u> grid boxes within each ODZ that contained observations to directly compare with model results in station profiles. Overall, the modeled NO₃⁻ and NO₂⁻ concentration and isotope profiles in the AS and ETNP were consistent with the observations, with [NO₃⁻] slightly<u>underestimated</u> in the Arabian Sea ODZ and overestimated in the ETSP (Figure 4). As [O₂] goes to zero, the O₂-intolerant processes NAR, NIR, and AMX are released from inhibition. These processes result in a decrease in [NO₃⁻]
- 15 (via NAR) which corresponds to an increase in $\delta^{15}N_{NO3}$, since NAR has a normal isotope effect. NO2⁻ also starts to accumulate in the <u>secondary NO2⁻ maximum</u> as a result of NAR. $\delta^{15}N_{NO2}$ is lower than $\delta^{15}N_{NO3}$ since light NO2⁻ is preferentially created via NAR, and this discrepancy is further reinforced by the inverse isotope effect of NXR (Casciotti, 2009). These patterns are readily observed in the AS and ETNP, but were less apparent in the ETSP, where [NO3⁻] depletion and [NO2⁻] accumulation in the model were lower than observed. This could be due in part to the time-independent nature of this steady state inverse
- 20 model, which does not capture the effects of upwelling events in the ETSP on N supply and cycling (Canfield, 2006; Chavez and Messié, 2009).

In order <u>to</u> gauge the model results for N loss, we also calculated N*, a measure of the availability of DIN relative to $PO4^{3-}$ compared to Redfield ratio stoichiometry (N* = $[NO_3^-] + [NO_2^-] - 16 * [PO4^{3-}]$; Deutsch et al., 2001). Negative N* values are

- 25 associated with N loss due to AMX or NIR₆or release of PO4³⁻ from anoxic sediments (Noffke et al., 2012), while positive N* values are associated with input of new N through N₂ fixation (Gruber and Sarmiento, 1997). Although we did not model PO4³⁻, we used the modeled [NO₃⁻] and [NO₂⁻] together with World Ocean Atlas PO4³⁻ data interpolated to the model grid to calculate N* resulting from the model. Both the AS and ETNP showed a decrease in model N* in the ODZ, as expected for water column N loss. Below the ODZ, N* increased again and returned to expected deep water values. Modeled N* in the ETSP,
- 30 however, did not follow the observed trend, consistent with an underestimate of N loss in the model ETSP.

Though the global best fit isotope effects for NAR, NIR, and NXR produced good agreement to the data in general, the isotope effects that best fit individual ODZ regions differed when the cost function was restricted to observations from a given ODZ.

Deleted: under estimated

Deleted: SNM

Deleted: fits

(Deleted: the
	Formatted: Font: Times New Roman
	Deleted: ,

Deleted: WOA

For the ETSP, the best fit isotope effects were the same as the previously stated global best fit. For the AS, the best fit isotope effects were $\epsilon_{NAR} = 13\%$, $\epsilon_{NIR} = 0\%$, and $\epsilon_{NXR} = -32\%$. For the ETNP, the best fit isotope effects were $\epsilon_{NAR} = 13\%$, $\epsilon_{NIR} = 15\%$, and $\epsilon_{NXR} = -32\%$, though the performance is only marginally better than with $\epsilon_{NIR} = 0\%$. The lower (more inverse) value for ϵ_{NXR} resulted in higher $\delta^{15}N_{NO3}$ and lower $\delta^{15}N_{NO2}$, which better fit the ODZ $\delta^{15}N_{NO2}$ data compared to the global best fit

5 ϵ_{NXR} = -13‰. These results are consistent with earlier isotope modeling studies in the ETSP (Casciotti et al., 2013; Peters et al., 2016; Peters et al., 2016; Peters et al., 2018b) and in the Arabian Sea (Martin and Casciotti, 2017). Although, in the Arabian Sea, modeled $\delta^{15}N_{NO3}$ were too high in the ODZ, likely in part due to overpredicted rates of NAR, which also resulted in lower modeled [NO₃⁻] (Figure 4).

3.3 Model-data comparison in GEOTRACES sections

- 10 We also investigated the agreement between global best fit model concentration and isotope distributions with data from two GEOTRACES cruise sections: GP16 in the South Pacific, and GA03 in the North Atlantic. For GP16, we see that [NO₃⁻] is low in surface waters and increases to a mid-depth maximum between 1000-2000 m. The highest [NO₃⁻] are found at mid-depth in the eastern boundary of the section. The model reproduces the general patterns, matching observations fairly well in the surface waters, but <u>diverges</u> below 500 m (Figure 5). Although the patterns are generally correct, insufficient NO₃⁻ is
- 15 accumulated in the deep waters of the model Pacific, <u>This could be due to an underestimate of preformed NO₃⁻ (over estimate of assimilation in the Southern Ocean), or inadequate supply of organic matter to be remineralized at depth. In the Southern Ocean, model surface [NO₃⁻] are 5-10 <u>µlM</u> lower than observations (Figure <u>\$5</u>), which could be enough to explain the lower than expected [NO₃⁻] in the deep Pacific, which is largely sourced from the Southern Ocean (Rafter et al., 2013; Sigman et al., 2009; Peters et al., 2018a,b).</u>
- 20

In the GP16 section, we also see that there are elevated $\delta^{15}N_{N03}$ values in the model surface waters and in the ETSP ODZ (Figure 5d), as expected from observations (Figure 5c). However, we can also see that the insufficient depletion of NO₃⁻ and increase in $\delta^{15}N_{N03}$ in the ETSP ODZ (Figure 5b and d) extends beyond the single grid box highlighted earlier (Figure 4). The less than expected increase of $\delta^{15}N_{N03}$ in the ETSP ODZ and the upper thermocline in the eastern part of the section is consistent.

- 25 with an underestimate of NO₃⁻ reduction. In GP16 we were also able to compare modelled and observed [NO₂⁻] and δ¹⁵N_{NO2} (Figure S6). Patterns of modeled [NO₂⁻] and δ¹⁵N_{NO2} showed accumulation of NO₂⁻ in the ODZ, with an appropriate δ¹⁵N_{NO2} value (Figure S6). Although, generally lower modeled concentrations of NO₂⁻ in the ODZ also support an underestimate of NAR (Figure S6).
- 30 Surface δ¹⁵N_{N03} values were also not as high in the model as in the observations (Figure 5), which could result from insufficient NO₃⁻ assimilation or too low supplied δ¹⁵N_{N03} (Peters et al., 2018a). However, we do see a similar depth range for high surface δ¹⁵N_{N03} and a local δ¹⁵N_{N03} minimum between the surface and ODZ propagating westward in both the model and observations,

30

Deleted: modeled [NO₃⁻] and $\delta^{15}N_{NO3}$ **Deleted:** matches

(Deleted: diverged
(Deleted: 500m as well as at the eastern edge of the ETSP ODZ
•(Deleted: being
(Deleted: , which
(Deleted: µM
.(Deleted: S4

Deleted: the

Deleted: Along with this, there is also minimal	
Deleted: .	
Deleted: ,	

	indicating that the physical and biogeochemical processes affecting $\delta^{15}N_{NO3}$ are represented by the model. Additionally, the
	model shows slightly elevated $\delta^{15}N_{NO3}$ in the thermocline depths (200-500m) west of the ODZ, which is consistent with the
	observations (Figure 5c), though not of the correct magnitude. This is partly related to the muted ODZ signal as mentioned
	above and its lessened impact on thermocline $\delta^{15}N_{NO3}$ across the basin. Peters et al. (2018a) and Rafter et al. (2013) also
5	postulated that these elevated $\delta^{15}N_{NO3}$ values were in part driven by remineralization of organic matter with high $\delta^{15}N.$ The
	δ^{15} N of sinking PON in the model (6-10‰) was similar to those observed in the south Pacific (Raimbault et al., 2008), as well
	as those predicted from other N isotope studies (Rafter et al., 2013; Peters et al., 2018). The model also shows slightly elevated
	$\delta^{15}N_{NO3} \text{ in the intermediate depths (500-1500m), which is consistent with observations, again reflecting remineralization of }$
	PON with $\delta^{15}N$ greater than mean ocean $\delta^{15}N_{NO3}$. Overall, the patterns of $\delta^{15}N_{NO3}$ are correct but the magnitudes of isotopic
10	variation are muted, largely due to the lack of N loss in the ODZ and modeled surface $\delta^{15}N_{\text{NO3}}$ values that are lower than
	$observations. \ The \ simplification \ of \ NH_4^+ \ dynamics \ in \ the \ model \ could \ also \ contribute \ to \ underestimation \ of \ \delta^{15}N_{NO3} \ values \ if \ NH_4^+ \ dynamics \ in \ the \ model \ could \ also \ contribute \ to \ underestimation \ of \ \delta^{15}N_{NO3} \ values \ if \ herebox{}$
	there was a large flux of ¹⁵ N-enriched NH ₄ ⁺ from sediments (Granger et al., 2011), or if ¹⁵ N-depleted NH ₄ ⁺ was preferentially
	transferred to the N2 pool via anammox. While the isotope effect on NH4+ during anammox (Brunner et al., 2013) is indeed
	higher than that applied here, we chose to balance this with a low isotope effect during aerobic NH4 ⁺ oxidation (Table 1).

15 ×

In the north Atlantic along GEOTRACES section GA03, we see good agreement between the observed and modeled [NO3] (Figure 6). There is generally low surface [NO₃⁻] with a distinct area of high [NO₃⁻] propagating from near the African coast. Deep water (> 2000 m) [NO37] is lower than we see in the Pacific section, and the model matches well with the Atlantic observations. Again, there is not quite enough NO3⁻ present in Southern Ocean-sourced intermediate waters (500-1500 m;

- 20 Marconi et al., 2015). Modeled $\delta^{15}N_{NO3}$ values at first glance appear higher than observed values at the surface (Figure 6). However, many of the surface $[NO_3^-]$ were below the operating limit for $\delta_1^{15}N_{NO3}$ analysis and were not determined. Focusing on areas where both measurements and model results are present yields excellent agreement. For example, we do see low $\delta^{15}N_{NO3}$ values in upper thermocline waters in both the model and observations, likely corresponding to low $\delta^{15}N$ contributions from N2 fixation that is remineralized at depth and accumulated in North Atlantic Central Water (Marconi et al., 2015; Knapp
- 25 et al., 2008). The model input includes significant rates of N2 fixation in the North Atlantic that are consistent with this observation (Martin et al., in review). However, rates of N deposition in the North Atlantic are also fairly high and can contribute to the low δ^{15} N signal (Knapp et al., 2008). In our model, atmospheric N deposition contributed between 0-50% of N input along the cruise track.

Deleted: δ¹⁵N Deleted: δ¹⁵N_{NO3}

Deleted: $\delta^{15}N_{NO3}$

Deleted:

(Formatted: Font: Times New Roman

Deleted: high

Deleted: prep

4 Discussion

5

4.1 Assumption checks

As previously mentioned (Section 2.3), <u>organic N</u> and DIN were modeled separately in order to introduce dependence on both <u>organic N</u> and substrate availability for the heterotrophic processes NAR and NIR. <u>These</u> separate <u>model runs require</u> several assumptions to be made regarding the processes that impact both <u>organic N</u> and DIN, namely assimilation and remineralization.

The first assumption is that the rates of N assimilation are equal between the <u>organic N</u> and DIN model runs. The <u>organic N</u> model run<u>uses World Ocean Atlas</u> surface [NO₃⁻] to estimate the contribution of DIN assimilation to the production of <u>organic</u> N, whereas the DIN model run uses modeled [NO₃⁻] and [NO₂⁻] to estimate DIN removal via assimilation. Though these two

- 10 methods utilized the same rate constants for assimilation, differences in [DIN] could cause some discrepancies between the overall rates. Analysis of the results revealed that slightly more overall DIN assimilation occurred in the DIN model run than <u>organic N</u> produced in the organic N model (Figure <u>\$57</u>). This could be due in part to assimilation of NO₂⁻ in the top two boxes <u>of the DIN model</u>, since NO₂⁻ assimilation is unaccounted for in the <u>organic N</u> model. This is largely an issue in the oligotrophic gyres, where surface [NO₃⁻] is very low and NO₂⁻ accumulates to low but non-zero concentrations (Figure 2). Assimilation of
- 15 NO₂⁻ accounts for a significant fraction of DIN assimilation in these regions, but the overall assimilation rates there are low and the resulting influence on the whole system is also low. In other regions, modeled surface [NO₃⁻] may be higher than the <u>World Ocean Atlas</u> surface [NO₃⁻] that are supplied to the <u>organic N</u> model, which would result in higher assimilation rates in the DIN model run. Indeed, points at which the DIN assimilation rates are higher than the <u>organic N</u> production rates do tend to have modeled [NO₃⁻] that was higher than observed [NO₃⁻] (Figure <u>\$7</u>). Likewise, points with relatively lower DIN
- 20 assimilation had modeled [NO₃⁻] less than observed [NO₃⁻]. However, the majority of DIN assimilation estimates were within 10 μM/yr of the <u>organic N</u> production estimates, with an <u>average offset of approximately 3.5% compared to DIN assimilation</u>. The total global assimilation rates were within 0.4%, with some spatially variable differences due to offset between surface [NO₃⁻] and modeled [NO₃⁻]. However we find that the World Ocean Atlas surface NO₃⁻ values are fairly well represented by our modeled surface NO₃⁻ (Figure <u>\$5</u>). We conclude that though the assimilation rates are not identical in the <u>organic N</u> and
- 25 DIN model runs, the discrepancy in modeled DIN assimilation is less than 0.1%, and there is unlikely to be significant creation or loss of N as a result of the split model.

4.2 Model dependency on input O2

The modeled concentration and isotope profiles for the ETSP, unlike in the AS and ETNP, reflected <u>an</u> underestimation of water column denitrification in the best-fit model. In ETSP [NO₃⁻] measurements, there is a clear deficit in [NO₃⁻], coincident

30 with the <u>secondary NO₂⁻ maximum</u> and N* minimum (Figure 4). In our modeled profiles, this NO₃⁻ deficit is missing, and although a <u>secondary NO₂⁻ maximum</u> is present, its magnitude is lower than observed (Figure 4). The model also does not capture the negative N* excursion (Figure 4), which we think reflects a model underestimation of <u>NAR</u> and <u>NIR</u> in the ETSP.

Deleted: ON
Deleted: ON
Deleted: This
Deleted: modeling requires
Deleted: ON
Deleted: ON
Deleted: ON
Deleted: used WOA
Deleted: ON
Deleted: used
Deleted: ON
Deleted: S5
Deleted: ON

Deleted:	WOA
Deleted:	data
Deleted:	ON
Deleted:	ON
Deleted:	85
Deleted:	ON
Deleted:	overall
Deleted:	rate of 9235 µM/yr. We also found
Deleted:	WOA
Deleted:	S4
Deleted:	ON

-(Deleted: a significant
(Deleted: SNM
-(Deleted: SNM
-(Deleted: NO ₃
••(Deleted: NO2 ⁻ reduction

The cause of this missing denitrification is likely to be poor representation of the ETSP O₂ conditions in the model grid space. Since our model grid is fairly coarse $(2^{\circ}x2^{\circ})$, only a few boxes within the ETSP had averaged [O₂] below the thresholds that would allow processes such as NAR and NIR to occur. The anoxic region of the ETSP is adjacent to the coast and not as spatially extensive as in the AS and ETNP (Figure S8); therefore, this region in particular was less compatible with the model grid. In order to test whether the parameterization of O₂ dependence was the cause of the low N loss, we ran the model using the globally optimized parameters (Table 3) but with higher O2 thresholds (15 µM) for NAR, NIR, and AMX (Table 1). This extended the region over which ODZ processes could occur and resulted in an increase in water column N loss from 6 to 32 Tg N/yr in the ETSP, which is more consistent with previous estimates (DeVries et al., 2012; Deutsch et al., 2001). This change also stimulated the development of a NO3⁻ deficit, larger secondary NO2⁻ maximum, and N* minimum within the ODZ (Figure

10 7).

5

As previously mentioned (Section 3.1), modeled [NO2-] in the Bay of Bengal is higher than observations. The accumulation of NO2⁻ here in the model is likely due to O2 concentrations falling below the set threshold for NAR but above the threshold for NIR, so NO2⁻ can accumulate via NAR but cannot be consumed via NIR. Although AMX and NXR occur there, the 15 modeled rates of their NO2⁻ consumption are rather low, which in combination with high rates of NAR leads to more NO2⁻

being produced than consumed. This is in contrast to observations that NO2⁻ production is tightly matched with NO2 <u>oxidation</u> in the Bay of Bengal, which limits NO2 accumulation and N loss there (Bristow et al., 2016). The fact that the model over predicts NAR in the Arabian Sea may also be connected with over-prediction of NAR in the Bay of Bengal. It is possible that the oxygen thresholds for ODZ processes are not the same in all ODZs, and further work on oxygen sensitivities of N cycle 20

processes will be addressed in a companion study (Martin et al., in review),

5 Conclusions

A global inverse ocean model was modified to include $\downarrow^4 N$ and $\downarrow^5 N$ in both NO₂⁻ and NO₂⁻ as state variables. Adding the processes required to describe the cycling of NO2- in the global ocean, including oxic and anoxic processes, resulted in a globally representative distribution of [NO₃⁻], [NO₂⁻], δ^{15} N_{NO3}, and δ^{15} N_{NO2}. In particular, the patterns of variation in both oxic

25 and anoxic waters are generally consistent with observations, though some magnitudes of variation were somewhat muted by the model. This could be due to an underestimation of a process rate, due to parameterization or model resolution, or an underestimation of the isotope effect involved.

Importantly, we were able to generate a roughly balanced steady-state ocean N budget without the need for an artificial 30 restoring force. The [NO₃⁻] and [NO₂⁻] distributions that were required to achieve this roughly balanced budget are well within the range of observed values. Some interesting take-home messages from this work are: 1) a relatively low isotope effect for NO₃⁻ reduction ($\varepsilon_{NAR} = 13\%$) gives a good fit to $\delta^{15}N_{NO3}$ data, similar to that concluded in some recent studies (Marconi et al.,

Deleted: S6

Deleted: in Deleted: Deleted: Deleted: slow Deleted: via NAR Deleted: consumption via NXR Deleted: in the Bay of Bengal Deleted: 2016a Deleted: , leading to accumulation of NO2⁻ despite realistic NO2⁻ consumption rates. Further Deleted: prep Formatted: Font: Bold Deleted: Deleted: [NO₂⁻], δ¹⁵N_{NO3}, Deleted: $\delta^{15}N_{NO2}$

Deleted: , as well as adding

Formatted: Font: Times New Roman

2017; <u>Bourbonnais et al., 2015</u>; Casciotti et al., 2013), 2) low <u>O2</u> half-saturation constants for NO2⁻ oxidation allowing <u>NO2⁻</u> oxidation to occur in parallel with <u>NO3⁻</u> reduction, NO2⁻ reduction, and anammox were needed to achieve the correct distributions of NO3⁻, NO2⁻, and their isotopes, in the oceans water column ODZs.

5 Though we have been able to adequately represent and assess N cycling in ODZs, there are many areas in which this model could be improved in order to expand its usefulness. Improving resolution of the model, particularly in coastal regions where there are steep gradients in nutrient and O₂ concentrations, would improve the accuracy of the model in regions such as the ETSP. Further, in regions that have high seasonal or interannual variability, an annually averaged steady-state model may not represent some important temporal dynamics. While we attempted to account for seasonal variation in the strength of the ODZs
10 through use of monthly O₂ climatologies, we did not simulate seasonal variations in <u>net primary production and the strength</u> of the biological pump. Variations in these parameters are likely to drive variations in N loss (Kalvelage et al., 2013; Ward,

In addition to the dependency on external static nutrient and parameter fields, this N cycle model is highly dependent on isotope

2013; Babbin et al., 2014).

- 15 effects for N cycle processes. Previous work has shown that the laboratory-derived isotope effects for some N cycle processes are not the same as their expressed isotope effects in environmental samples or under conditions relevant to environmental samples (Casciotti et al., 2013; Bourbonnais et al., 2015; Buchwald et al., 2015; Kritee et al., 2012; Marconi et al., 2017a). Further probing the isotope effects using an inverse model such as this could provide insight into the expressed isotope effects that should be used in other modeling efforts involving field data. As presented in Section 3.1, the larger magnitude isotope
- 20 <u>effect for NO₂⁻ oxidation best fit the ETNP and Arabian Sea ODZs</u>, where most of the ODZ volume resided. However, most model $\delta^{15}N_{NO2}$ values still do not reach the lowest values observed on the edges of marine ODZs, indicating that further work is needed to understand the expression of these isotope effects.

The larger isotope effects resulted in better fits to observations of $\delta^{15}N$ and DIN concentrations with lower rates of N cycling.

25 This reinforces the importance of obtaining realistic isotope effect estimates for each process that are relevant on an environmental scale. Additionally, this highlights the need for critical consideration of isotope effects used in N cycle models that use isotope balance to predict N cycling rates. Though isotopes provide us a useful tool to assess the relative contributions of different processes, these estimates are highly subject to the isotope effects employed. Also, as illustrated by the regional optimizations, the isotope effect for a given process may vary, or be expressed differently, in different regions.

This model provides an excellent framework for <u>further</u> testing hypotheses about controls on the marine N inventory and cycling of N on a global scale. The distribution <u>and sensitivities</u> of N cycle rates resulting from this model will be explored in a companion manuscript (Martin et al., in <u>review</u>). Incorporation of <u>variable</u> environmental input data, such as temperature, productivity, and [O₂], could also help us predict how the N cycle might be affected by past and future environmental changes.

Deleted: NXR	
Formatted: Font: Times New Roman	
Deleted: NAR, NIR	
Deleted: AMX	

Deleted: NPP

Deleted: As discussed in Section 2.6, the rate parameters were optimized using a variety of different combinations of isotope effects for NAR, NIR, and NXR to explore model uncertainty. As presented in Section 3.1, the larger magnitude isotope effect for NXR best fit the ETNP and Arabian Sea ODZs, where most of the ODZ volume resided. The larger magnitude isotope effect also resulted in optimized rate constants for NAR, NIR, and NXR that were lower than the global best fit rate constants. The lower rate constants and larger isotope effects resulted in better fits to observations of $\frac{\partial^2 N}{\partial x}$ and OIN concentrations.

Moved down [2]: This reinforces the importance of obtaining realistic isotope effect estimates for each process that are relevant on an environmental scale.

Deleted:

Moved (insertion) [2]

Deleted: Additionally, this reinforces

Deleted: be

Deleted: prep
Deleted: modified

Code availability

Model code and model output from the three optimal ODZ isotope effect combinations, including the global best fit, are available in the Stanford Digital Repository (https://purl.stanford.edu/hr045dx8661).

Author contribution

5 KLC, FP, and TSM designed the study. TSM and FP constructed the model. TSM and KLC analysed and interpreted the results. TSM, KLC, and FP wrote the manuscript.

Competing interests

The authors declare that they have no conflict of interest.

Acknowledgements

10 Thanks to Patrick Rafter for sharing a pre-publication version of his NO₃⁻ isotope database. Thanks to Tim Davis for guidance on sparse matrix solvers. Thanks to Tim DeVries for helpful discussions about earlier versions of the inverse model. Thanks to Kevin Arrigo and Leif Thomas for comments on an earlier draft of this manuscript. This work was partly supported by NSF Chemical Oceanography grant 1657868 to KLC.

References

15	1.	Anderson, J. J., Okubo, A., Robbins, A. S, and Richards, F. A.: A model for nitrate distributions in oceanic oxygen Deleted: .,
•		minimum zones, Deep-Sea Res., 29, 1113-1140, doi:10.1016/0198-0149(82)90031-0, 1982.
[<u>2.</u>	Babbin, A. R., Keil, R. G., Devol, A. H. and Ward, B. B.: Organic Matter Stoichiometry, Flux, and Oxygen Control
		Nitrogen Loss in the Ocean, Science, 344, 406-408, doi:10.1126/science.1248364, 2014.
	3.	Babbin, A. R., Peters, B. D., Mordy, C. W., Widner, B., Casciotti, K. L. and Ward, B. B.: Multiple metabolisms Deleted: .,
20		constrain the anaerobic nitrite budget in the Eastern Tropical South Pacific, Global Biogeochem. Cy., 31, 258-271,
		doi:10.1002/2016GB005407, 2017.
	4.	Berelson, W. M.: Particle settling rates increase with depth in the ocean, Deep-Sea Res. Pt. II, 49, 237-251.
		doi:10.1016/S0967-0645(01)00102-3, 2002.
[5.	Bianchi, D., Dunne, J. P., Sarmiento, J. L. and Galbraith, E. D.: Data-based estimates of suboxia, denitrification, and Deleted: .,
25		N2O production in the ocean and their sensitivities to dissolved O2, Global Biogeochem. Cy., 26, 1-13,
		doi:10.1029/2011GB004209, 2012.

	6. Bohlen, L., Dale, A. W, and Wallmann, K.: Simple transfer functions for calculating benthic fixed nitrogen losses	Deleted: .,
I	and C:N:P regeneration ratios in global biogeochemical models, Global Biogeochem. Cy, 26, GB3029,	
	doi:10.1029/2011GB004198, 2012.	
	7. Bonin, P., Gilewicz, M. and Bertrand, J. C.: Effects of oxygen on each step of denitrification on Pseudomonas nautica,	Deleted: .,
5	Can. J. Microbiol., 35, 1061-1064, doi:10.1139/m89-177, 1989.	
	8. Bourbonnais, A., Altabet, M. A., Charoenpong, C. N., Larkum, J., Hu, H., Bange, H. W, and Stramma, L.: N-loss	Deleted: .,
I	isotope effects in the Peru oxygen minimum zone studied using a mesoscale eddy as a natural tracer experiment,	
	Global Biogeochem. Cy., 29, 793-811, doi:10.1002/2013GB004679, 2015.	
1	9. Brandes, J. A, and Devol, A. H.: Isotopic fractionation of oxygen and nitrogen in coastal marine sediments, Geochim.	Deleted: .,
10	Cosmochim. Ac., 61, 1793-1801, doi:10.1016/S0016-7037(97)00041-0, 1997.	Deleted: .,
	10. Brandes, J. A. and Devol, A. H.: A global marine-fixed nitrogen isotopic budget: Implications for Holocene nitrogen	Deleted: .,
I	cycling, Global Biogeochem. Cy., 16, 67-1-67-14, doi:10.1029/2001GB001856, 2002.	
	11. Bristow, L. A., Dalsgaard, T., Tiano, L., Mills, D. B., Bertagnolli, A. D., Wright, J. J., Hallam, S. J., Ulloa, O.,	
	Canfield, D. E., Revsbech, N. P. and Thamdrup, B.: Ammonium and nitrite oxidation at nanomolar oxygen	Deleted:
15	concentrations in oxygen minimum zone waters, P. Natl. Acad. Sci. USA, 113, 10601-10606,	
	doi:10.1073/pnas.1600359113, 2016.	
	12. Bristow, L. A., Callbeck, C. M., Larsen, M., Altabet, M. A., Dekaezemacker, J., Forth, M., Gauns, M., Glud, R. N.,	
	Kuypers, M. M. M., Lavik, G., Milucka, J., Naqvi, S. W. A., Pratihary, A., Revsbech, N. P., Thamdrup, B., Treusch,	
1	A. H _e and Canfield, D. E.: N ₂ production rates limited by nitrite availability in the Bay of Bengal oxygen minimum	Deleted:
20	zone, Nat. Geosci., 10, 24-29, doi:10.1038/ngeo2847, 2017.	,
1	13. Brown, Z. W., Casciotti, K. L., Pickart, R. S., Swift, J. H. and Arrigo, K. R.: Aspects of the marine nitrogen cycle of	
	the Chukchi Sea shelf and Canada Basin, Deep. Res. Part II Top. Stud. Oceanogr., 118, 73-87,	
	doi:10.1016/j.dsr2.2015.02.009, 2015.	
I	14. Brunner, B., Contreras, S., Lehmann, M. F., Matantseva, O., Rollog, M., Kalvelage, T., Klockgether, G., Lavik, G.,	
25		Deleted:
I	Acad. Sci. USA, 110, 18994-18999, doi:10.1073/pnas.1310488110, 2013.	
1	15. Bryan, B. A., Shearer, G., Skeeters, J. L, and Kohl, D. H.: Variable expression of the nitrogen isotope effect associated	Deleted: .,
I	with denitrification of nitrite, J. Biol. Chem., 258, 8613-8617, 1983.	
1	16. Buchwald, C, and Casciotti, K. L.: Oxygen isotopic fractionation and exchange during bacterial nitrite oxidation,	Deleted:
30	Limnol. Oceanogr., 55, 1064-1074, doi:10.4319/lo.2010.55.3.1064, 2010.	
		Deleted: .,
I	maximum of the eastern tropical North Pacific off Costa Rica, Global Biogeochem. Cy., 29, 2061-2081, doi:10.1002/	
	2015GB005187, 2015.	

	18. Buesseler, K. Q. and Boyd, P. W.: Shedding light on processes that control particle export and flux attenuation in the Delete	ed: .,
•	twilight zone of the open ocean, Limnol. Oceanogr., 54, 1210-1232, doi:10.4319/lo.2009.54.4.1210, 2009.	
	19. Buesseler, K. O., Trull, T. W., Steinberg, D. K., Silver, M. W., Siegel, D. A., Saitoh, SI., Lamborg, C. H., Lam, P.	
	J., Karl, D. M., Jiao, N. Z., Honda, M. C., Elskens, M., Dehairs, F., Brown, S. L., Boyd, P. W., Bishop, J. K. Br and Delete	ed: .,
5	Bidigare, R. R.: VERTIGO (VERtical Transport In the Global Ocean): A study of particle sources and flux attenuation	
	in the North Pacific, Deep-Sea Res. Pt. II, 55, 1522-1539, doi:10.1016/j.dsr2.2008.04.024, 2008.	
	20. Canfield, D. E.: Models of oxic respiration, denitrification and sulfate reduction in zones of coastal upwelling,	
	Geochim. Cosmochim. Acta, 70, 5753-5765, doi:10.1016/j.gca.2006.07.023, 2006.	
	21. Capone, D. G., Burns, J. A., Montoya, J. P., Subramaniam, A., Mahaffey, C., Gunderson, T., Michaels, A. F, and Delete	ed: .,
10	Carpenter, E. J.: Nitrogen fixation by Trichodesmium spp.: An important source of new nitrogen to the tropical and	
	subtropical North Atlantic Ocean, Global Biogeochem. Cy., 19, GB2024, doi:10.1029/2004GB002331, 2005.	
	22. Carpenter, E. J., Harvey, H. R., Brian, F. and Capone, D. G.: Biogeochemical tracers of the marine cyanobacterium	ed: .,
I	Trichodesmium, Deep-Sea Res. Pt. I, 44, 27-38, doi:10.1016/S0967-0637(96)00091-X, 1997.	
	23. Casciotti, K. L.: Inverse kinetic isotope fractionation during bacterial nitrite oxidation, Geochim. Cosmochim. Ac.,	
15	73, 2061-2076, doi:10.1016/j.gca.2008.12.022, 2009.	
	24. Casciotti, K. L., Trull, T. W., Glover, D. M. and Davies, D.: Constraints on nitrogen cycling at the subtropical North	ed: .,
I	Pacific Station ALOHA from isotopic measurements of nitrate and particulate nitrogen, Deep-Sea Res. Pt. II, 55,	
	1661-1672, doi:10.1016/j.dsr2.2008.04.017, 2008.	
	25. Casciotti, K. L., Buchwald, C. and McIlvin, M.: Implications of nitrate and nitrite isotopic measurements for the Delete	ed: .,
20	mechanisms of nitrogen cycling in the Peru oxygen deficient zone, Deep-Sea Res. Pt. I, 80, 78-93,	
	doi:10.1016/j.dsr.2013.05.017, 2013.	
	26. Chavez, F. P. and Messié, M.: A comparison of Eastern Boundary Upwelling Ecosystems, Prog. Oceanogr., 83, 80-	
	96, doi:10.1016/j.pocean.2009.07.032, 2009.	
	27. Chien, C-T., Mackey, K. R. M., Dutkiewicz, S., Mahowald, N. M., Prospero, J. M. and Paytan, A.: Effects of African Delete	ed: .,
25	dust deposition on phytoplankton in the western tropical Atlantic Ocean off Barbados, Global Biogeochem. Cy., 30,	
	716-734, doi:10.1002/2015GB005334, 2016.	
	28. Codispoti, L. A.: Phosphorus vs. Nitrogen limitation of new and export production, in: Productivity of the Oceans:	
	Present and Past. Vol 44., John Wiley & Sons Ltd., 377-394, 1989.	
	29. Codispoti, L. A, and Christensen, J. P.: Nitrification, denitrification and nitrous oxide cycling in the eastern tropical	ed: .,
30	South Pacific ocean, Mar. Chem., 16, 277-300, doi:10.1016/0304-4203(85)90051-9, 1985.	
	30. Codispoti, L. A., Friederich, G. E., Packard T. T., Glover, H. E., Kelly, P. J., Spinrad, R. W., Barber, R. T., Elkins, J.	
	W., Ward, B. B., Lipschultz, F _u and Lostaunau, N.: High Nitrite Levels off Northern Peru: A Signal of Instability in Delete	ed: .,
•	the Marine Denitrification Rate, Science, 233, 1200-1202, doi:10.1126/science.233.4769.1200, 1986.	

	31.	Codispoti, L. A., Brandes, J. A., Christensen, J. P., Devol, A. H., Naqvi, S. W. A, Paerl, H. W, and Yoshinari, T.: The Deleted: .,
		oceanic fixed nitrogen and nitrous oxide budgets: Moving targets as we enter the anthropocene?, Sci. Mar., 65, 80-
		105, doi:10.3989/scimar.2001.65s285, 2001.
	32.	Dalsgaard, T., Stewart, F. J., Thamdrup, B., De Brabandere, L., Revsbech, N. P., Ulloa, O., Canfield, D. E. and Deleted: .,
5		DeLong, E. F.: Oxygen at nanomolar levels reversibly suppresses process rates and gene expression in anammox and
		denitrification in the oxygen minimum zone off Northern Chile, MBio, 5, e01966-14, doi:10.1128/mBio.01966-14,
		2014.
	33.	Dentener, F., Drevet, J., Lamarque, J. F., Bey, I., Eickhout, B., Fiore, A. M., Hauglustaine, D., Horowitz, L. W., Krol,
		M., Kulshrestha, U. C., Lawrence, M., Galy-Lacaux, C., Rast, S., Shindell, D., Stevenson, D., Van Noije, T., Atherton,
10		C., Bell, N., Bergman, D., Butler, T., Cofala, J., Collins, B., Doherty, R., Ellingsen, K., Galloway, J., Gauss, M.,
		Montanaro, V., Müller, J. F., Pitari, G., Rodriguez, J., Sanderson, M., Solmon, F., Strahan, S., Schultz, M., Sudo, K.,
		Szopa, S <u>and</u> Wild, O.: Nitrogen and sulfur deposition on regional and global scales: A multimodel evaluation, Global Deleted: .,
		Biogeochem. Cy., 20, GB4003, doi:10.1029/2005GB002672, 2006.
	34.	Deutsch, C., Gruber, N., Key, R. M., Sarmiento, J. L _* and Ganachaud, A.: Denitrification and N ₂ fixation in the Pacific Deleted: .,
15		Ocean, Global Biogeochem. Cy., 15, 483-506, doi:10.1029/2000GB001291, 2001.
	35.	Deutsch, C., Sarmiento, J. L., Sigman, D. M., Gruber, N. and Dunne, J. P.: Spatial coupling of nitrogen inputs and Deleted: "
		losses in the ocean, Nature, 445, 163-167, doi:10.1038/nature05392, 2007.
	36.	DeVries, T _e and Primeau, F.: Dynamically and Observationally Constrained Estimates of Water-Mass Distributions Deleted: "
		and Ages in the Global Ocean, J. Phys. Oceanogr., 41, 2381-2401, doi:10.1175/JPO-D-10-05011.1, 2011.
20	37.	DeVries, T., Deutsch, C., Rafter, P. A, and Primeau, F.: Marine denitrification rates determined from a global 3-D Deleted: .,
		inverse model, Biogeosciences, 10, 2481-2496, doi:10.5194/bg-10-2481-2013, 2013.
	38.	Dunne, J. P., Armstrong, R. A., Gnanadesikan, A <u>and</u> Sarmiento, J. L.: Empirical and mechanistic models for the Deleted: .,
		particle export ratio, Global Biogeochem. Cy., 19, GB4026, doi:10.1029/2004GB002390, 2005.
	39.	Follows, M. J., Dutkiewicz, S., Grant, S. and Chisholm, S. W.: Emergent Biogeography of Microbial Communities Deleted: .,
25		in a Model Ocean, Science, 315, 1843-1846, doi:10.1126/science.1138544, 2007.
	<u>40.</u>	Frey, C., Hietanen, S., Jürgens, K., Labrenz, M. and Voss, M.: N and O isotope fractionation in nitrate during
		chemolithoautotrophic denitrification by Sulfurimonas gotlandica, Environ. Sci. Technol., 48(22), 13229-13237,
		doi:10.1021/es503456g, 2014.
	41.	Fuchsman, C. A., Devol, A. H., Casciotti, K. L., Buchwald, C., Chang, B. X, and Horak, R. E. A.: An N isotopic mass
30		balance of the Eastern Tropical North Pacific Oxygen Deficient Zone, Deep-Sea Res. Pt. II,
		doi:10.1016/j.dsr2.2017.12.013, 2017.
	42.	Galloway, J. N., Dentener, F. J., Capone, D. G., Boyer, E. W., Howarth, R. W., Seitzinger, S. P., Asner, G. P.,
		Cleveland, C. C., Green, P. A., Holland, E. A., Karl, D. M., Michaels, A. F., Porter, J. H., Townsend, A. R. and Deleted: .,

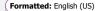
	Vörösmarty, C. J.: Nitrogen cycles: past, present, and future, Biogeochemistry, 70, 153-226, doi:10.1007/s10533-004-0370-0, 2004.			
	43. Garcia, H. E., Boyer, T. P., Locarnini, R. A., Boyer, T. P., Antonov, J. I., Mishonov, A. V., Baranova, O. K., Zweng,			
1	M. M., Reagan, J. R, and Johnson, D. R.: World Ocean Atlas 2013. Volume 3: dissolved oxygen, apparent oxygen		Deleted: .,	\supset
5	utilization, and oxygen saturation, NOAA Atlas NESDIS 75, 27 pp., 2013.			
1	44. Garcia, H. E., Locarnini, R. A., Boyer, T. P., Antonov, J. I., Baranova, O. K., Zweng, M. M., Reagan, J. R, and		Deleted: .,	\supset
I	Johnson, D. R.: World Ocean Atlas 2013. Volume 4: Dissolved inorganic nutrients (phosphate, nitrate, silicate),			
	NOAA Atlas NESDIS 76, 25 pp., 2013.			
	45. Gaye, B., Nagel, B., Dähnke, K., Rixen, T, and Emeis, K-C, Evidence of parallel denitrification and nitrite oxidation		Deleted: .,	\supset
10	in the ODZ of the Arabian Sea from paired stable isotopes of nitrate and nitrite. Global Biogeochem. Cy. 27, 1059-	(Deleted: ."	J
	1071, doi:10.1002/2011GB004115, 2013.			
1	46. Granger, J., Sigman, D. M., Lehmann, M. F. and Tortell, P. D.: Nitrogen and oxygen isotope fractionation during		Deleted: .,	
I	dissimilatory nitrate reduction by denitrifying bacteria, Limnol. Oceanogr., 53, 2533-2545,			
	doi:10.4319/lo.2008.53.6.2533, 2008.			
15	47. Granger, J., Sigman, D. M., Rohde, M. M., Maldonado, M. T. and Tortell, P. D.: N and O isotope effects during		Deleted: .,	\supset
I	nitrate assimilation by unicellular prokaryotic and eukaryotic plankton cultures, Geochim. Cosmochim. Ac., 74, 1030-			
	1040, doi:10.1016/j.gca.2009.10.044, 2010.			
1	48. Granger, J., Prokopenko, M. G., Sigman, D. M., Mordy, C. W., Morse, Z. M., Morales, L. V., Sambrotto, R. N. and			
	Plessen, B.: Coupled nitrification-denitrification in sediment of the eastern Bering Sea shelf leads to 15N enrichment			
20	of fixed N in shelf waters, J. Geophys. Res. Ocean., 116(11), 1-18, doi:10.1029/2010JC006751, 2011.			
I	49. Gruber, N. The Dynamics of the Marine Nitrogen Cycle and its Influence on Atmospheric CO ₂ Variations, in: The			
1	Ocean Carbon Cycle and Climate, Springer, Netherlands, 2004.			
Ĩ	50. Gruber, N. The Marine Nitrogen Cycle: Overview and Challenges, in: Nitrogen in the Marine Environment. Capone,			
1	D. G., Bronk, D. A., Mulholland, M. R., Carpenter, E. J., eds. Elsevier, 2008		Formatted: Font: 12 pt, English (US)	\supset
25	51. Gruber, N. and Galloway, J. N.: An Earth-system perspective of the global nitrogen cycle., Nature, 451(7176), 293-		Formatted: English (US)	5
	6, doi:10.1038/nature06592, 2008.	(Deleted: .,	\supset
	52. <u>Gruber, N. and Sarmiento, J. L.: Global patterns of marine nitrogen fixation and denitrification, Global Biogeochem.</u>			
Ĩ	Cy., 11, 235-266, doi:10.1029/97GB00077, 1997.			
1	53. Harding, K., Turk-Kubo, K. A., Sipler, R. E., Mills, M. M., Bronk, D. A. and Zehr, J. P.: Symbiotic unicellular			
30	cyanobacteria fix nitrogen in the Arctic Ocean, Proc. Natl. Acad. Sci., 201813658, doi:10.1073/pnas.1813658115,			
	<u>2018.</u>			
	54. Hastings, M. G., Jarvis, J. C, and Steig, E. J.: Anthropogenic Impacts on Nitrogen Isotopes of Ice-Core Nitrate,		Deleted: .,	\supset
I	Science, 324, 1288, doi:10.1126/science.1170510, 2009.			

	55.	Hoering, T. C. and Ford, H. T.: The Isotope Effect in the Fixation of Nitrogen by Azotobacter, J. Am. Chem. Soc.,	(Deleted: .,
1		82, 376-378. doi:10.1021/ja01487a031, 1960.	6	
	56.	Holl, C. M _e and Montoya, J. P.: Interactions between nitrate uptake and nitrogen fixation in continuous cultures of		Deleted: .,
		the marine diazotroph Trichodesmium (Cyanobacteria), J. Phycol., 41, 1178-1183, doi:10.1111/j.1529-		
5		8817.2005.00146.x, 2005.		
	57.	Hu, H., Bourbonnais, A., Larkum, J., Bange, H. W, and Altabet, M. A.: Nitrogen cycling in shallow low-oxygen		Deleted: .,
		coastal waters off Peru from nitrite and nitrate nitrogen and oxygen isotopes, Biogeosciences, 13, 1453-1468,		
		doi:10.5194/bg-13-1453-2016, 2016.		
	58.	Jensen, M. M., Kuypers, M. M. M., Lavik, G. and Thamdrup, B. Rates and regulation of anaerobic ammonium		Deleted: .,
10		oxidation and denitrification in the Black Sea, Limnol. Oceanogr., 53, 23-36, doi:10.4319/lo.2008.53.1.0023, 2008.		
	59.	Kalvelage, T., Jensen, M. M., Contreras, S., Revsbech, N. P., Lam, P., Günter, M., LaRoche, J., Lavik, G. and		Deleted: .,
		Kuypers, M. M. M.: Oxygen Sensitivity of Anammox and Coupled N-Cycle Processes in Oxygen Minimum Zones,		
		PLoS One, 6, e29299, doi:10.1371/journal.pone.0029299, 2011.		
	<u>60.</u>	Kalvelage, T., Lavik, G., Lam, P., Contreras, S., Arteaga, L., Loscher, C. R., Oschlies, A., Paulmier, A., Stramma, L.		
15		and Kuypers, M. M. M.: Nitrogen cycling driven by organic matter export in the South Pacific oxygen minimum		
		zone, Nat. Geosci., 6(3), 228-234, doi:10.1038/ngeo1739, 2013.		
	61.	Knapp, A. N., DiFiore, P. J., Deutsch, C., Sigman, D. M. and Lipschultz, F.: Nitrate isotopic composition between		Deleted: .,
		$Bermuda \ and \ Puerto \ Rico: \ Implications \ for \ N_2 \ fixation \ in \ the \ Atlantic \ Ocean, \ Global \ Biogeochem. \ Cy., \ 22, \ GB3014,$		
		doi:10.1029/2007GB003107, 2008.		
20	62.	Knapp, A. N., Sigman, D. M., Lipschultz, F., Kustka, A. B, and Capone, D. G.: Interbasin isotopic correspondence		Deleted: .,
1		between upper-ocean bulk DON and subsurface nitrate and its implications for marine nitrogen cycling, Global		
		Biogeochem. Cy., 25, GB4004, doi:10.1029/2010GB003878, 2011.		
	63.	Kritee, K., Sigman, D. M., Granger, J., Ward, B. B., Jayakumar, Ag and Deutsch, C.: Reduced isotope fractionation		Deleted: .,
1		by denitrification under conditions relevant to the ocean, Geochim. Cosmochim. Ac., 92, 243-259,		
25		doi:10.1016/j.gca.2012.05.020, 2012.		
	64.	Kuypers, M. M., Lavik, G., Woebken, D., Schmid, M., Fuchs, B. M., Amann, R., Jorgensen, B. B, and Jetten, M.		Deleted: .,
1		S. M.: Massive nitrogen loss from the Benguela upwelling system through anaerobic ammonium oxidation, P. Natl.		
		Acad. Sci. USA, 102, 6478-6483, doi:10.1073/pnas.0502088102, 2005.		
	<u>65.</u>	Landolfi, A., Kähler, P., Koeve, W. and Oschlies, A.: Global marine N2 fixation estimates: From observations to		
30		models, Front. Microbiol., 9(SEP), 1-8, doi:10.3389/fmicb.2018.02112, 2018.		
	<u>66.</u>	Lavik, G., Stührmann, T., Brüchert, V., Van Der Plas, A., Mohrholz, V., Lam, P., Mußmann, M., Fuchs, B. M.,		
		Amann, R., Lass, U. and Kuypers, M. M. M.: Detoxification of sulphidic African shelf waters by blooming		
		chemolithotrophs, Nature, 457(7229), 581-584, doi:10.1038/nature07588, 2009.		
I				

	67.	Lehmann, M. F., Bernasconi, S. M., Barbieri, A., Simona, M, and McKenzie, J. A.: Interannual variation of the Deleted: .,	\supset
		isotopic composition of sedimenting organic carbon and nitrogen in Lake Lugano: A long-term sediment trap study,	
		Limnol. Oceanogr., 49, 839-849, doi:10.4319/lo.2004.49.3.0839, 2004.	
	<u>68.</u>	Lehmann, M. F., Sigman, D. M., McCorkle, D. C., Granger, J., Hoffmann, S., Cane, G. and Brunelle, B. G.: The	
5		distribution of nitrate ¹⁵ N/ ¹⁴ N in marine sediments and the impact of benthic nitrogen loss on the isotopic composition	
		of oceanic nitrate, Geochim. Cosmochim. Acta, 71(22), 5384-5404, doi:10.1016/j.gca.2007.07.025, 2007.	
	69.	Locarnini, R. A., Mishonov, A. V., Antonov, J. I., Boyer, T. P., Garcia, H. E., Baranova, O. K., Zweng, M. M., Paver,	
		C. R., Reagan, J. R., Johnson, D. R., Hamilton, M. and Seidov, D.: World Ocean Atlas 2013. Vol. 1: Temperature, Deleted: .,	\supset
		NOAA Atlas NESDIS 73, 40 pp., 2013.	
10	70.	Marconi, D., Weigand, M. A., Rafter, P. A., McIlvin, M. R., Forbes, M., Casciotti, K. L _e and Sigman, D. M.: Nitrate Deleted: .,	\supset
		isotope distributions on the US GEOTRACES North Atlantic cross-basin section: Signals of polar nitrate sources and	
		low latitude nitrogen cycling, Mar. Chem., 177, 143-156, doi:10.1016/j.marchem.2015.06.007, 2015.	
	71.	Marconi, D., Kopf, S., Rafter, P. A, and Sigman, D. M.: Aerobic respiration along isopycnals leads to overestimation Deleted: .,	\supset
		of the isotope effect of denitrification in the ocean water column, Geochim. Cosmochim. Ac., 197, 417-432,	
15		doi:10.1016/j.gca.2016.10.012, 2017.	
	<u>72.</u>	Mariotti, A.: Atmospheric nitrogen is a reliable standard for natural ¹⁵ N abundance measurements, Nature, 303(5919),	
		<u>685–687, doi:10.1038/303685a0, 1983.</u>	
	73.	Mariotti, A., Germon, J. C., Hubert, P., Kaiser, P., Letolle, R., Tardieux, A. and Tardieux, P.: Experimental Deleted: .,	\supset
		determination of nitrogen kinetic isotope fractionation: Some principles; illustration for the denitrification and	
20		nitrification processes, Plant Soil, 62, 413-430, doi:10.1007/BF02374138, 1981	
	74.	Martin, J. H., Knauer, G. A., Karl, D. M, and Broenkow, W. W.: VERTEX: carbon cycling in the northeast Pacific, Deleted: .,	\supset
		Deep-Sea Res., 34, 267-285, doi:10.1016/0198-0149(87)90086-0, 1987.	
	75.	Martin, T. S. and Casciotti, K. L.: Nitrogen and oxygen isotopic fractionation during microbial nitrite reduction, Deleted: .,	\supset
		Limnol. Oceanogr., 61, 1134-1143, doi:10.1002/lno.10278, 2016.	
25	76.	Martin, T. S, and Casciotti, K. L.: Paired N and O isotopic analysis of nitrate and nitrite in the Arabian Sea oxygen Deleted: .,	\supset
		deficient zone, Deep-Sea Res. Pt. I, 121, 121-131, doi:10.1016/j.dsr.2017.01.002, 2017.	
	77.	Monteiro, F. M., Dutkiewicz, S _r and Follows, M. J.: Biogeographical controls on the marine nitrogen fixers, Global Deleted: .,	\supset
		Biogeochem. Cy., 25, GB2003, doi:10.1029/2010GB003902, 2011.	
	78.	Moore, J. K, and Doney, S. C.: Iron availability limits the ocean nitrogen inventory stabilizing feedbacks between Deleted: .,	\supset
30		marine denitrification and nitrogen fixation, Global Biogeochem. Cy., 21, GB2001, doi:10.1029/2006GB002762,	
		2007.	
	79.	Moore, J. K., Doney, S. C. and Lindsay, K.: Upper ocean ecosystem dynamics and iron cycling in a global three- Deleted: .,	\supset
		dimensional model, Global Biogeochem. Cy., 18, GB4028, 2004, doi:10.1029/2004GB002220, 2004.	

	<u>80.</u>	Nixon, S. W., Ammerman, J. W., Atkinson, L. P., Berousky, V. M., Billen, G., Boicourt, W. C., Boynton, W. R.,		
		Church, T. M., Ditoro, D. M., Elmgren, R., Garber, J. H., Giblin, A. E., Jahnke, R. A., Owens, N. J. P., Pilson, M. E.		
		Q. and Seitzinger, S. P.: The fate of nitrogen and phosphorus at the land-sea margin of the North Atlantic Ocean Five		
		major rivers with an average water flow exceeding 3000 m ³ s ⁻¹ discharge, Biogeochemistry, 35, 141-180, 1996.		
5	81.	Noffke, A., Hensen, C., Sommer, S., Scholz, F., Bohlen, L., Mosch, T., Graco, M. and Wallmann, K.: Benthic iron		
		and phosphorus fluxes across the Peruvian oxygen minimum zone, Limnol. Oceanogr., 57(3), 851-867,		
		doi:10.4319/lo.2012.57.3.0851, 2012.		
	82.	Peng, X., Fuchsman, C. A., Jayakumar, A., Warner, M. J., Devol, A. He and Ward, B. B.: Revisiting nitrification in		Deleted: .,
		the Eastern Tropical South Pacific: A focus on controls, J. Geophys. ResOceans, 121, 1667-1684,		
10		doi:10.1002/2015JC011455, 2016.		
	83.	Penn, J., Weber, T, and Deutsch, C.: Microbial functional diversity alters the structure and sensitivity of oxygen		Deleted: .,
		deficient zones, Geophys. Res. Lett., 43, 9773-9780, doi:10.1002/2016GL070438, 2016.		
	84.	Peters, B. D., Babbin, A. R., Lettmann, K. A., Mordy, C. W., Ulloa, O., Ward, B. B, and Casciotti, K. L.: Vertical		Deleted: .,
		modeling of the nitrogen cycle in the eastern tropical South Pacific oxygen deficient zone using high-resolution		
15		concentration and isotope measurements, Global Biogeochem. Cy., 30, 1661-1681, doi:10.1002/2016GB005415,		
		2016.		
	85.	Peters, B. D., Lam, P. J. and Casciotti, K. L .: Nitrogen and oxygen isotope measurements of nitrate along the US		Deleted: .,
		GEOTRACES Eastern Pacific Zonal Transect (GP16) yield insights into nitrate supply, remineralization, and water		
		mass transport, Mar. Chem., 201, 137-150, doi:10.1016/j.marchem.2017.09.009, 2018a.		
20	86.	Peters, B. D., Horak, R., Devol, A., Fuchsman, C., Forbes, M., Mordy, C. W, and Casciotti, K. L.: Estimating fixed		Deleted: .,
		nitrogen loss and associated isotope effects using concentration and isotope measurements of NO_3^- , NO_2^- , and N_2		
		from the Eastern Tropical South Pacific oxygen deficient zone, Deep-Sea Res. Pt. II, doi:10.1016/j.dsr2.2018.02.011,		
		2018b.		
	87.	Rafter, P. A, and Sigman, D. M .: Spatial distribution and temporal variation of nitrate nitrogen and oxygen isotopes		Deleted: .,
25		in the upper equatorial Pacific Ocean, Limnol. Oceanogr., 61, 14-31, doi:10.1002/lno.10152, 2016.		
	88.	Rafter, P. A., DiFiore, P. J. and Sigman, D. M.: Coupled nitrate nitrogen and oxygen isotopes and organic matter		Deleted: .,
		remineralization in the Southern and Pacific Oceans, J. Geophys. Res. Oceans, 118, 4781-4794,		
		doi:10.1002/jgrc.20316, 2013.		
	89.	Raimbault, P., Garcia, N. and Cerutti, F.: Distribution of inorganic and organic nutrients in the South Pacific Ocean:		Deleted: .,
30		evidence for long-term accumulation of organic matter in nitrogen-depleted waters, Biogeosciences, 5, 281-298,		
		doi:10.5194/bg-5-281-2008, 2008.	_	
	90.	Redfield, A. C., Ketchum, B. H, and Richards, F. A .: The Influence of Organisms on the Composition of Sea Water,		Deleted: .,
		Sea, 2, 26-77, 1963.		

	91. Ryabenko, E., Kock, A., Bange, H. W., Altabet, M. A. and Wallace, D. W. R.: Contrasting biogeochemistry of Deleted:
	nitrogen in the Atlantic and Pacific Oxygen Minimum Zones, Biogeosciences, 9, 203-215, doi:10.5194/bg-9-203-
	2012, 2012.
	92. Seitzinger, S. P. and Giblin, A. E.: Estimating denitrification in North Atlantic continental shelf sediments,
5	Biogeochemistry, 35(1), 235–260, doi:10.1007/BF02179829, 1996.
	93. Shiozaki, T., Bombar, D., Riemann, L., Hashihama, F., Takeda, S., Yamaguchi, T., Ehama, M., Hamasaki, K. and
	Furuya, K.: Basin scale variability of active diazotrophs and nitrogen fixation in the North Pacific, from the tropics
	to the subarctic Bering Sea, Global Biogeochem. Cycles, 31(6), 996-1009, doi:10.1002/2017GB005681, 2017.
10	94. Sigman, D. M., Altabet, M. A., McCorkle, D. C., Francois, R, and Fischer, G.: The $\delta^{15}N$ of nitrate in the Southern Deleted: .,
	Ocean: Consumption of nitrate in surface waters, Global Biogeochem. Cy., 13, 1149-1166, doi:10.1029/1999GB900038, 1999.
	95. Sigman, D. M., Granger, J., DiFiore, P. J., Lehmann, M. F., Ho, R., Cane, G. and van Geen, A.: Coupled nitrogen
	and oxygen isotope measurements of nitrate along the eastern North Pacific margin, Global Biogeochem. Cy., 19,
15	GB4022, doi:10.1029/2005GB002458, 2005.
	96. Sigman, D. M., DiFiore, P. J., Hain, M. P., Deutsch, C., Wang, Y., Karl, D. M., Knapp, A. N., Lehmann, M. F. and Deleted: .,
	Pantoja, S.: The dual isotopes of deep nitrate as a constraint on the cycle and budget of oceanic fixed nitrogen, Deep-
	Sea Res. Pt. I, 56, 1419-1439, doi:10.1016/j.dsr.2009.04.007, 2009.
	97. Somes, C. J., Schmittner, A., Galbraith, E. D., Lehmann, M. F., Altabet, M. A., Montoya, J. P., Letelier, R. M., Mix,
20	A. C., Bourbonnais, A, and Eby, M.: Simulating the global distribution of nitrogen isotopes in the ocean, Global Deleted: .,
	Biogeochem. Cy., 24, GB4019, doi:10.1029/2009GB003767, 2010.
	98. Somes, C. J. and Oschlies, A.: On the influence of "non-Redfield" dissolved organic nutrient dynamics on the spatial
	distribution of N ₂ fixation and the size of the marine fixed nitrogen inventory, Global Biogeochem. Cycles, 29(7),
	973-993, doi:10.1002/2014GB005050, 2015.
25	99. Strous, M., Fuerst, J. A., Kramer, E. H. M., Logemann, S., Muyzer, G., van de Pas-Schoonen, K. T., Webb, R.,
	Kuenen, J. G. and Jetten, M. S. M.: Missing lithotroph identified as new planctomycete, Nature, 400, 446-449, Deleted: .,
	doi:10.1038/22749, 1999
	100. Van Mooy, B. A. S., Keil, R. G. and Devol, A. H.: Impact of suboxia on sinking particulate organic carbon: Enhanced Deleted: .,
	carbon flux and preferential degradation of amino acids via denitrification, Geochim. Cosmochim. Ac., 66, 457-465,
30	doi:10.1016/S0016-7037(01)00787-6, 2002.
	101. Ward, B. B.: How Nitrogen Is Lost, Science, 341(6144), 352-353, doi:10.1126/science.1240314, 2013.
	102. Westberry, T., Behrenfeld, M. J., Siegel, D. A, and Boss, E.: Carbon-based primary productivity modeling with Deleted: .,
	vertically resolved photoacclimation, Global Biogeochem. Cy., 22, GB2024, doi:10.1029/2007GB003078, 2008.



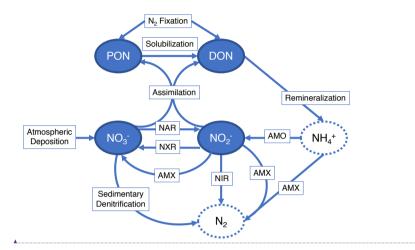
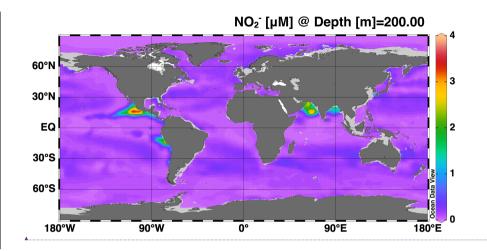


Figure 1: Diagram showing the nitrogen (N) cycle processes represented in the model. Two organic N pools are modeled: particulate organic N (PON) and dissolved organic N (DON). Two inorganic N pools are modeled: nitrate (NO₃⁻) and nitrite (NO₂⁻). N source processes are nitrogen (N₂) fixation and atmospheric deposition. N sink processes are sedimentary denitrification, NO₂⁻ reduction (NIR), and anammox (AMX). Internal cycling processes that transform N from one species to another are solubilization, remineralization, assimilation, NO₃⁻ reduction (NAR), ammonia oxidation (AMO), and NO₂⁻ oxidation (NXR). Neither ammonia (NH₃) nor ammonium (NH₄⁺) are tracked in this model, since they are assumed to not accumulate. N₂ is also not explicitly accounted for in the model.





Formatted: English (US)

Figure 2: Map showing the model-estimated accumulation of nitrite (NO2⁻) at 200 m depth.

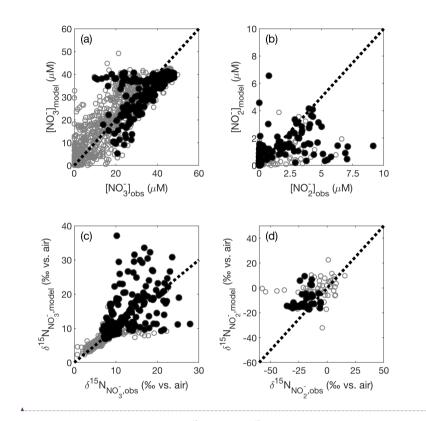
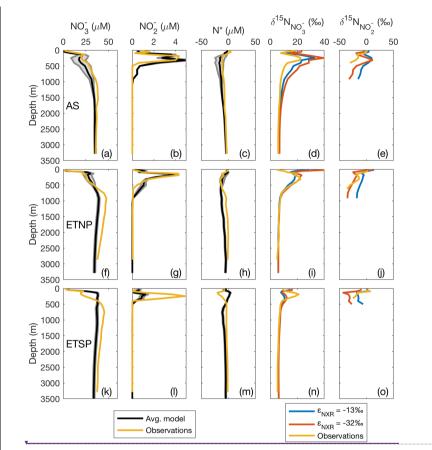


Figure 3: Modeled (a) [NO₃⁻], (b) [NO₂⁻], (c) $\delta^{15}N_{NO4e}$ and (d) $\delta^{15}N_{NO4e}$ are compared against the corresponding values from the database test set. Shown on each panel is a 1:1 line starting at the origin. Data in black have corresponding [O₂] < 10 μ M, and data in gray have [O₂] \geq 10 μ M.

Formatted: English (US)



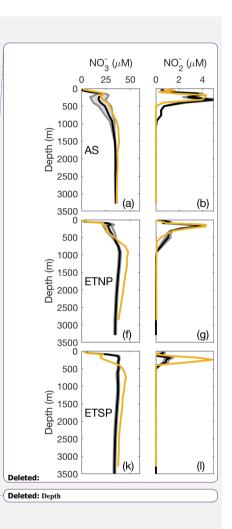


Figure 4: Offshore depth profiles comparing model results with binned and averaged database observations from a model water column. Results are shown for the three main oxygen deficient zones (ODZs): the Arabian Sea (a-e), the Eastern Tropical North Pacific (ETNP; f-j), and the Eastern Tropical South Pacific (ETSP; k-o). Average modeled nitrate concentration ([NO₃]), nitrite concentration ([NO₂]), and N* are shown in black. Gray error lines around the black line show the 2σ spread from the average from the 12 different optimized model results using the different combinations of isotope effects for nitrate reduction (ϵ_{NAR}), nitrite reduction (ϵ_{NIR}), and nitrite oxidation (ϵ_{NXR}). Observed data are shown in yellow in all panels. Modeled $\delta^{15}N_{NO4}$ and $\delta^{15}N_{NO4}$ are shown for three different combinations of isotope effect. The blue lines represent $\varepsilon_{NAR} = 13$, $\varepsilon_{NXR} = -13$, and $\varepsilon_{NIR} = 0$, which are the best fit isotope effects globally and in the ETSP. The red lines represent $\varepsilon_{NAR} = 13$, $\varepsilon_{NXR} = -32$, and $\varepsilon_{NIR} = 0$, which are the best fit isotope effects in the Arabian Sea.

10

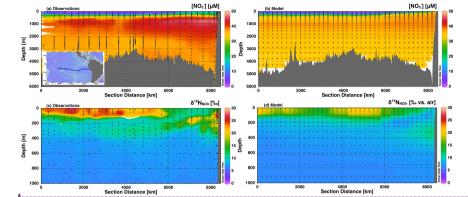
5

I

47

Deleted:

Deleted:



Formatted: English (US)

Section Distance [km]
Figure 5: Section profiles of NO₃⁻ concentrations and isotopes over the GP16 cruise track (panel (a) inset) in the South Pacific, Profiles are presented from east (right) to west (left). Comparison of (a) observed [NO₃⁻] to (b) modeled [NO₃⁻] is presented over the full depth range (0-6000m). Comparison of (c) observed δ¹⁵N_{NO2} to (d) modeled δ¹⁵N_{NO2} is presented over a shortened depth range (0-1000m) to better assess surface and ODZ values. GEOTRACES data are from Peters et al. (2018a) and available from BCO-DMO.

Deleted:

Deleted:

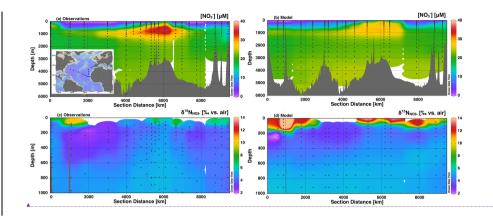
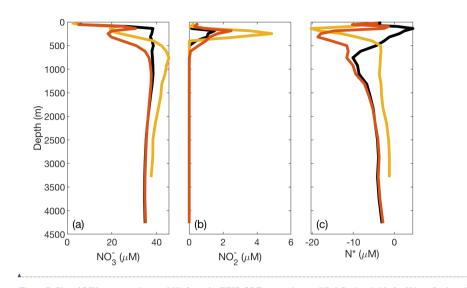


Figure 6: Section profiles of NO₃⁻ concentrations and isotopes over the GA03 cruise track (panel (a) inset) in the North Atlantic. Profiles are presented from east (right) to west (left) from 0-6000km section distance, and then from south to north. Comparison of (a) observed [NO₃] to (b) modeled [NO₃] is presented over the full depth range (0-6000m). Comparison of (c) observed δ^{15} No₃-to (d) modeled δ^{15} No₃-is presented over a shortened depth range (0-1000m) to better assess surface and the low δ^{15} N_{NO3}-contribution from N₂ fixation. GEOTRACES data are from Marconi et al. (2015) and available from BCO-DMO.

5

(Deleted: .	2
(Deleted: .	2
~(Deleted:	5

Formatted: English (US)



Formatted: English (US)

Figure 7: Plot of DIN concentrations and N* from the ETSP ODZ comparing modified O_2 thresholds for N loss. In the original optimized version of the model, there is insufficient N loss and NO₂⁻ accumulation in the ETSP. To demonstrate that this issue may be caused by issues with the gridded averages of O_2 and model grid size in the ETSP, we raised the O_2 thresholds for N loss-related processes (NAR, NIR, and AMX) to 15 μ M. This effectively lowers the observed $[O_2]$ in order to stimulate N loss. The resulting (a) [NO₂], (b) [NO₂], and (c) N* are shown with the observed values from the database (yellow), original optimized model values (black), and lowered O_2 threshold model values (red).

5

Parameter	Value	Reference
b	-0.858	Martin et al., 1987
F ₀	1.5 mmol N m ⁻³ yr ⁻¹	DeVries et al., 2013
		Capone et al., 2005
λ	10 mmol N m ⁻³	Holl & Montoya, 2005
T ₀	20 °C	DeVries et al., 2013
		Capone et al., 2005
K _{Fe}	4.4*10 ⁻⁵ mmol Fe m ⁻³	Follows et al., 2007
K _P	0.005 mmol PO4 ³⁻ m ⁻³	Moore & Doney, 2007
r _{C:N}	6.625	Redfield et al., 1963
$\mathrm{Km}^{\mathrm{AMO}}$	3.5 µM O ₂	Peng et al., 2016
K _m ^{NXR}	0.8 µM O ₂	Bristow et al., 2016b
O2 ^{NAR}	7 µM O ₂	Dalsgaard et al., 2014; Jensen et al., 2008; Kuypers et al.,
	15 μM O2 ^a	2005; Kalvelage et al., 2011
$O_2^{\rm NIR}$	5 µM O ₂	Bonin et al., 1989; Kalvelage et al., 2011
	$15 \ \mu M \ O_2{}^a$	
O2 ^{AMX}	10 µM O ₂	Dalsgaard et al., 2014; Jensen et al., 2008; Kuypers et al.,
	$15 \ \mu M \ O_2{}^a$	2005; Kalvelage et al., 2011
$\delta^{15}N_{dep}$	-4‰	Hastings et al., 2009
$\delta^{15} N_{\rm fix}$	-1‰	Hoering & Ford, 1960; Carpenter et al., 1997
α _{AMX,NO2}	1.016	Brunner et al., 2013
α _{AMX,NXR}	0.969	Brunner et al., 2013
α _{AMX,NH4}	1	
α _{AMO}	1	
α_{sed}	1	Brandes & Devol, 1997; Lehmann et al., 2004
α_{assim}	1.004	Granger et al., 2010
αremin	1	Casciotti et al., 2008; Möbius, 2013
α _{sol}	1	Knapp et al., 2011

a. Value used in test of ETSP N loss issue (Section 4.2)

Table 1. Non-optimized model parameters.

Parameter	Initial	Final (avg.)	Error (2σ)	Final (global best fit)
¹⁴ k _{PON} (yr ⁻¹)	3.9	3.9	0	3.9
¹⁴ k _{DON} (yr ⁻¹)	1.8	0.8	0.2	0.6
$^{14}k_{NXR}(yr^{-1})$	6.0	16.0	3.0	18.7
$^{14}k_{NAR}$ (μ M ⁻¹ DON)	2.5	1.6	0.8	2.3
¹⁴ k _{NIR} (µM ⁻¹ DON)	1.5	1.7	1.0	2.6

Table 2. Optimized model parameters.

Parameter	Values	References
ENAR	13‰, 25‰	Granger et al., 2008
		Kritee et al., 2012
		Casciotti et al., 2013
		Marconi et al., 2017
ENIR	0‰, 15‰	Casciotti et al., 2013
		Martin and Casciotti, 2016
ENXR	-32‰, -20‰, -13‰	Casciotti, 2009
		Buchwald and Casciotti, 2010
		Casciotti et al., 2013

Table 3: Isotope effect cases.

Section S1

Linearization of transfer function for sedimentary denitrification,

This section expands on the transformations applied to the non-linear transfer function for sedimentary denitrification presented by Bohlen et al. (2012) in order to use the transfer function in our linear N cycle model. The original function is as follows:

- 1. DIN loss = $(0.60 + 0.19*0.99^{(O2-NO3)bottom})*RRPOC$ for water depths >= 1000m
- 2. DIN loss = 0.73*(0.60 + 0.19*0.99^{(O2-NO3)bottom})*RRPOC for water depths < 1000m
- 10 The calculation of the rain rate of particulate organic carbon (RRPOC) follows the Martin curve is as described in Text S4 (Equation 20). (O2-NO3⁻)bottom is the difference between [O2] and [NO3⁻] at the bottom of the water column, where it interfaces with the sediments.

For every model grid box, the depth is taken to be the depth at the bottom of the box. Each box is then assigned a multiplier of 15 1 (if depth \geq 1000m) or 0.73 (if depth \leq 1000m) that will be multiplied by the sedimentary denitrification terms.

The next step is linearizing the (DIN loss)/RRPOC data presented by Bohlen et al. (2012) with respect to (O2-NO3⁻)_{bottom,} since we cannot use the exponential equation in our linear system. This was performed by selecting two (O2-NO3⁻)bottom cutoff points (29 µM and 141 µM), breaking the data into three groups. A piecewise linear regression was then performed on each of these sections (Figure S1), resulting in the following equations:

- 20
 - 3. (DIN loss)/RRPOC = $0.297 0.005(O_2 NO_3^{-})_{bottom}$ (O₂-NO₃⁻)_{bottom} <= 29 µM
 - 4. (DIN loss)/RRPOC = $0.222 0.001(O_2 NO_3^{-})_{bottom}$ 29 μ M < ($O_2 NO_3^{-})_{bottom} \le 141 \mu$ M
 - 5. (DIN loss)/RRPOC = 0.105 0.000006(O₂-NO₃⁻)_{bottom} $141 \ \mu M < (O_2 - NO_3)_{bottom}$

25

5

These (O2-NO3⁻)_{bottom} cutoff points were then converted to O2 cutoff points in order to use a simple N-independent mask to determine which of the relationships to apply to a given model grid box. A linear relationship between [O2] and (O2-NO3⁻)bottom was determined using The 2013 World Ocean Atlas interpolated data product for [O2] and [NO3⁻] (Garcia et al., 2014). The linear relationship is as follows and is also shown in Figure S2:

30

6. $(O_2 - NO_3)_{bottom} = 1.12[O_2]_{bottom} - 55.6$

The (O₂-NO₃⁻)_{bottom} cutoff points can then be expressed in [O₂] units as 75 and 175 µM.

55

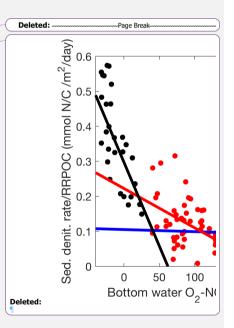
Formatted: Font: 10 pt, Bold

Formatted: Heading 1

Formatted: Font: 10 pt, Bold

The final step in modifying this transfer function for use in the linear model is to break the piecewise linear equations into a component that is independent of N. This facilitates the implementation of the equations in our linear system.

	7. Independent + dependent = (DIN loss)/RRPOC	
	 8. Independent = 0.297 - 0.005[O₂] 9. Dependent = 0.005[NO₃⁻] 	$O_2 \ll 75 \ \mu M$
10		
	10. Independent = $0.222 - 0.001[O_2]$	$75 \ \mu M < \mathrm{O_2} <= 175 \ \mu M$
	11. Dependent = $0.001[NO_3]$	
	12. Independent = $0.105 - 0.000006[O_2]$	175 µM < O ₂
15	13. Dependent = $0.000006[NO_3^-]_{\mu}$	··· r· ··-



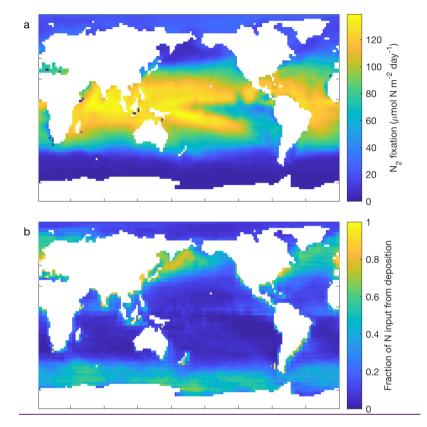


Figure S1

(a) Map of modeled areal N_2 fixation rates. (b) Map of the fraction of N input due to atmospheric deposition of DIN, with the remaining fraction due to N_2 fixation.



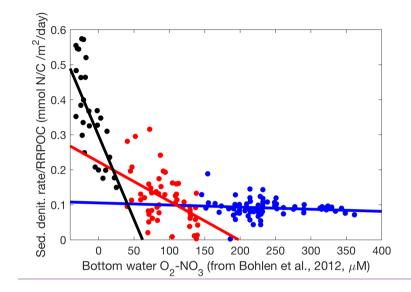


Figure S2. Piecewise division of the transfer function for sedimentary denitrification from Bohlen et al. (2012). In order to incorporate this into our liner model, we split the original non-linear relationship between sedimentary denitrification rate, rain
rate of particulate organic carbon (RRPOC), and bottom water ([O₂] – [NO₃⁻]) into three linear segments with cutoff points in terms of ([O₂] – [NO₃⁻]). These cutoff points were then converted to [O₂] cutoff points using a relation shown in Figure S2.

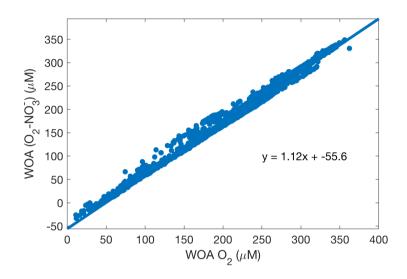


Figure §3. Plot of 2013 World Ocean Atlas [O₂] vs. ([O₂] – [NO₃⁻]) (Garcia et al., 2014). In order to express the sedimentary denitrification transfer function cutoff points (Figure S1) in terms of [O₂] rather than ([O₂] – [NO₃⁻]), we determined a linear 5 relationship between [O₂] and ([O₂] – [NO₃⁻]).

Deleted: S2

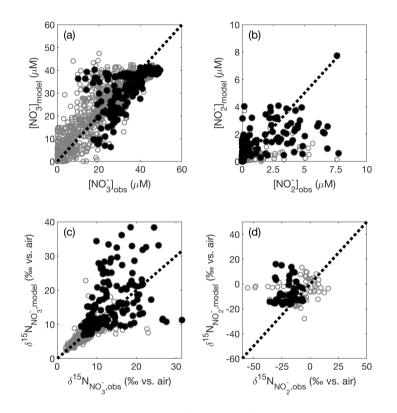


Figure <u>\$4</u>. Modeled (a) [NO₃⁻], (b) [NO₂⁻], (c) $\delta^{15}N_{NO3}$, and (d) $\delta^{15}N_{NO2}$ are compared against the corresponding values from the database training set. Shown on each panel is a 1:1 line starting at the origin. Data in black have corresponding [O₂] < 10 μ M, and data in gray have [O₂] \geq 10 μ M.

Deleted: S3

5

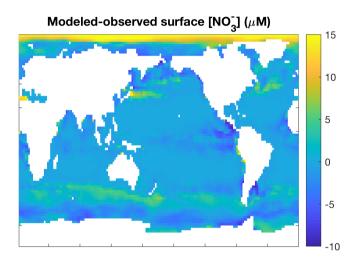
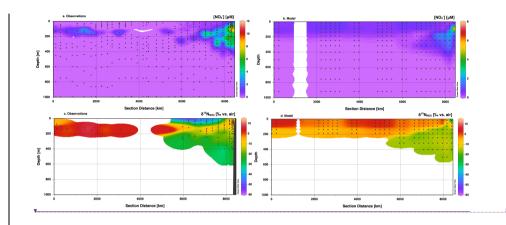


Figure 55. Map showing a comparison between modeled surface $[NO_3^-]$ for the top two model boxes and 2013 World Ocean <u>Atlas</u> $[NO_3^-]$ (Garcia et al., 2014) interpolated to the model grid for the same depths. Areas where the model overpredicts surface $[NO_3^-]$ are shown in yellow, and underprediction is shown in blue.

Deleted: S4

5



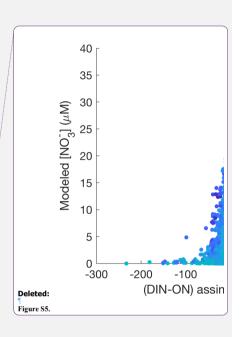


Figure S6. Section profiles of NO2⁻ concentrations and isotopes over the GP16 cruise track (panel (a) inset) in the South Pacific. Profiles are presented from east (right) to west (left). Comparison of (a) observed [NO2⁻] to (b) modeled [NO2⁻] is
presented over a shortened depth range (0-1000m) to better assess surface and ODZ values where NO2⁻ accumulates. GEOTRACES data are from Peters et al. (2018a) and available from BCO-DMO.

62

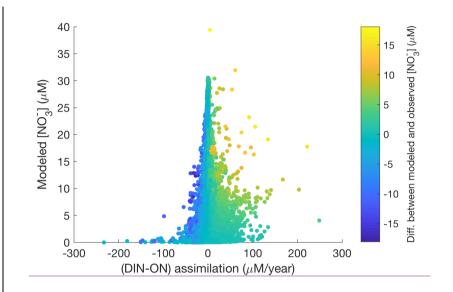


Figure S7. Difference in assimilation rates between the DIN and ON model runs plotted against the modeled [NO₃⁻]. Points are colored by the difference between modeled and observed [NO₃⁻].

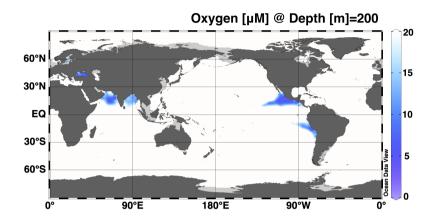


Figure <u>\$8.</u> Map of annual average 2013 World Ocean Atlas [O₂] @ 200m depth (Garcia et al., 2014) to demonstrate areas where O₂ is low enough for anoxic processes such as nitrate reduction, nitrite reduction, and anammox. The canonical ODZs

Deleted: S6

5 are visible in blue: the Arabian Sea, ETNP, and ETSP. Also shown in blue are the Bay of Bengal and the Black Sea.

Global					
N ₂ fixation rate (Tg N/yr)	Method of estimation	Reference			
<u>137</u>	Ocean circulation model of P*	Deutsch et al., 2007			
110 ± 40^{a}	<u>N* observations</u>	Gruber & Sarmiento, 1997			
<u>74 (51-110)</u>	Tracer incubations	Luo et al., 2014			
<u>131</u>	Global inverse N model	This study			
Pacific					
N ₂ fixation rate (Tg N/yr)	Method of estimation	Reference			
<u>95</u>	Ocean circulation model of P*	Deutsch et al., 2007			
59 ± 14	N* observations	Deutsch et al., 2001			
<u>37 (25-56)</u>	Tracer incubations	Luo et al., 2014			
<u>67</u>	Global inverse N model	This study			
Atlantic					
N ₂ fixation rate (Tg N/yr)	Method of estimation	Reference			
<u>20</u>	Ocean circulation model of P*	Deutsch et al., 2007			
30.5 ± 4.9	Nitrate isotope mass balance	Marconi et al., 2017			
27.6 ± 10	Tracer incubations	Fonseca-Batista et al., 2017			
<u>28</u> ^a	N* observations	Gruber & Sarmiento, 1997			
<u>13.6 (9.7-19.4)</u>	Tracer incubations	Luo et al., 2014			
<u>32</u>	Global inverse N model	This study			

a. Extrapolated from N. Atlantic estimate

b. Only from 10°N-50°N

Table S1. Global N2 fixation rate comparisons.

Location and year sampled	Data types	Reference
ETNP, 2003	[NO3 ⁻], [NO2 ⁻], δ^{15} Nn03-, δ^{15} Nn02-	Casciotti and McIlvin, 2007
ETNP, 2012	$[NO_{3}^{-}], [NO_{2}^{-}], \delta^{15}N_{NO3}^{-}, \\ \delta^{15}N_{NO2}^{-}.$	Casciotti, unpublished
ETSP, 2011	$[NO_{3}^{-}], [NO_{2}^{-}], \delta^{15}N_{NO3}^{-}, \\ \delta^{15}N_{NO2}^{-}.$	Casciotti, unpublished
ETSP, 2013	[NO3 ⁻], [NO2 ⁻], δ^{15} Nn03-, δ^{15} Nn02-	Peters et al., 2018

Table <u>\$2</u>. New additions to the database originally compiled by Rafter et al. (in prep.).

Deleted: S1

References

5

10

15

- Bohlen, L_g, Dale, A. W. and Wallmann, K_g: Simple transfer functions for calculating benthic fixed nitrogen losses and C:N:P regeneration ratios in global biogeochemical models, Global Biogeochem, Cycles, 26(3), doi:10.1029/2011GB004198, 2012.
- Casciotti, K. L. and McIlvin, M. R.: Isotopic analyses of nitrate and nitrite from reference mixtures and application to Eastern Tropical North Pacific waters, Mar. Chem., 107(2), 184–201, doi:10.1016/j.marchem.2007.06.021, 2007.
- Deutsch, C., Gruber, N., Key, R. M., Sarmiento, J. L. and Ganachaud, A.: Denitrification and N₂ fixation in the Pacific Ocean, Global Biogeochem. Cycles, 15(2), 483–506, 2001.
- Deutsch, C., Sarmiento, J. L., Sigman, D. M., Gruber, N. and Dunne, J. P.: Spatial coupling of nitrogen inputs and losses in the ocean, Nature, 445(7124), 163–167, doi:10.1038/nature05392, 2007.
- Fonseca-Batista, D., Dehairs, F., Riou, V., Fripiat, F., Elskens, M., Deman, F., Brion, N., Quéroué, F., Bode, M. and Auel, H.: Nitrogen fixation in the eastern Atlantic reaches similar levels in the Southern and Northern Hemisphere, J. Geophys. Res. Ocean., 122(1), 587–601, doi:10.1002/2016JC012335, 2017.
- Garcia, H. E., Boyer, T. P., Locarnini, R. A., Antonov, J. I., Mishonov, A. V. Baranova, O. K., Zweng, M. M., <u>Reagan, J. R. and Johnson, D. R.</u>: World Ocean Atlas 2013. Volume 3: <u>dissolved</u> oxygen, apparent oxygen utilization, and oxygen saturation, NOAA Atlas NESDIS 75, <u>3</u>, 27, 2013.
- Garcia, H. E., Locarnini, R. A., Boyer, T. P., Antonov, J. I., Baranova, O. K., Zweng, M. M., Reagan, J. R. and Johnson, D. R.: World Ocean Atlas 2013, Volume 4: Dissolved inorganic nutrients (phosphate, nitrate, silicate), NOAA Atlas NESDIS 76, 4, 27 pp, doi:10.1182/blood-2011-06-357442, 2013.
- 20 <u>Gruber, N. and Sarmiento, J. L.: Global patterns of marine nitrogen fixation and denitrification, Global</u> Biogeochem. Cycles, 11(2), 235, doi:10.1029/97GB00077, 1997.
 - Luo, Y. W., Lima, I. D., Karl, D. M., Deutsch, C. A. and Doney, S. C.: Data-based assessment of environmental controls on global marine nitrogen fixation, Biogeosciences, 11(3), 691–708, doi:10.5194/bg-11-691-2014, 2014.
 - Marconi, D., Sigman, D. M., Casciotti, K. L., Campbell, E. C., Weigand, M. A., Fawcett, S. E., Knapp, A. N.,
 - Rafter, P. A., Ward, B. B. and Haug, G. H.: Tropical dominance of N₂ fixation in the North Atlantic Ocean, Global Biogeochem. Cycles, 31, 1608-1623, doi:10.1002/2016GB005613, 2017.
 - Peters, B. D., Lam, P. J. and Casciotti, K. L.: Nitrogen and oxygen isotope measurements of nitrate along the US GEOTRACES Eastern Pacific Zonal Transect (GP16) yield insights into nitrate supply, remineralization, and water mass transport, Mar, Chem, 201, 137–150, doi:10.1016/j.marchem.2017.09.009, 2018.
- Deleted: AW, Deleted: Deleted: Deleted: 2012 Deleted:) Formatted: Font: 10 pt. Not Italic Formatted: Font: 10 pt, Not Italic Deleted: 2. Deleted: KL Deleted: MR. Deleted: Deleted: . 2007; Deleted:) Deleted: -Deleted: Formatted: Font: 10 pt. Not Italic Formatted: Font: 10 pt, Not Italic Deleted: 3 -Deleted: HE. Deleted: TP, Deleted: RA, et al. Deleted: Dissolved Deleted: Deleted: pp. Deleted: 4 Deleted: HE, Deleted: RA Deleted: TP. et al. Deleted:) Deleted: 25 pp. Deleted: 5. - Peters BD, Lam PJ, Casciotti KL Deleted: Formatted: Font: 10 pt, Not Italic Deleted: . 2018; Deleted: Deleted: -

Deleted: 1. -

Deleted:

Deleted:

30

25