Grazing related nitrous oxide emissions: from patch scale to field scale

Karl Voglmeier[1,2], Johan Six[2], Markus Jocher[1], Christof Ammann[1]

[1]Climate and Agriculture Group, Agroscope, Zürich, 8046, Switzerland

[2]Department of Environmental Systems Science, ETH Zurich, Zürich, 8092, Switzerland

Correspondence to: Karl Voglmeier (karl.voglmeier@agroscope.admin.ch)

Abstract.

Grazed pastures are strong sources of the greenhouse gas nitrous oxide (N_2O). The quantification of the N_2O emissions is challenging due to the strong spatial and temporal variabilities of the emission sources and so N_2O emission estimates are very uncertain. This study presents N_2O emission measurements from two grazing systems in western Switzerland over the grazing season of 2016. The 12 dairy cows of each herd were kept in an intensive rotational grazing management. The diet for the two herds of cows consisted of different protein to energy ratios (system G: grass only diet, system M: grass with additional maize silage) resulting in different nitrogen (N) excretion rates. The N in the excretion was estimated by calculating the animal nitrogen budget taking into account the measurements of feed intake, milk yield and body weight of the cow herds. Directly after the rotational grazing phases, background and urine patches were identified based on soil electric conductivity measurements while fresh dung patches were identified visually. The magnitude and temporal pattern of these different emission sources were measured with a fast-box (FB) chamber and the field scale fluxes were quantified using two eddy covariance (EC) systems. The FB measurements were finally up-scaled to field level and compared to the EC measurements for quality control by using EC footprint estimates of a backward Lagrangian stochastic dispersion model. The comparison between the two grazing systems was done during emission periods that were not influenced by fertilizer applications. This allowed the calculation of the excreta related N_2O emissions per cow and grazing hour and resulted in considerable higher emissions for system G compared to system M. Relating the found emissions to the excreta N resulted in excreta related emission factors (EF) of 0.74 ± 0.26 % for system M and 0.83 ± 0.29 % for system G. These EF values were thus significantly smaller compared to the default EF of 2 % provided by the IPCC guidelines for cattle excreta deposited on pasture. The measurements showed that urine patch emission dominated the field scale fluxes (57 %), followed by significant background emissions (38 %) and only a small contribution of dung patch emission (5 %). The resulting source specific EFs exhibited a clear difference between urine (1.12 ± 0.43 %) and dung (0.16 ± 0.06 %) supporting a disaggregation of the grazing related EFs by excreta type in emission inventories. The study also highlights the advantage of an N optimised diet which resulted in reduced N_2O emissions from animal excreta.

1 INTRODUCTION

Nitrous oxide (N_2O) is a strong greenhouse gas (GHG) with a 265 times stronger warming potential compared to CO_2 on a mass basis (IPCC, 2014). Typically an inert gas in the troposphere, N_2O has a strong potential to destroy the ozone layer in the stratosphere (Portmann et al., 2012). The largest share of N_2O emissions are attributed to nitrogen (N) fertilization in the agricultural sector, but also livestock grazing, especially by cows, can lead to significant direct and indirect N_2O emissions due to excreta from the animals (Luo et al., 2017; Reay et al., 2012). The nitrogen deposited by animal excreta often exceeds the N applied by fertilizer (Aarons et al., 2017). The available reactive N is used by microbial nitrification and denitrification processes where significant amounts of N_2O can be produced (Selbie et al., 2015). A non-linear response of N_2O emissions to N loading has been shown previously (Cardenas et al., 2010), and urine patches of cattle have exceptionally high N loading rates (up to 2000 kg N ha^{-1}) making them especially prone to high N_2O losses (Selbie et al., 2015).

For inventories and live cycle assessments, the magnitude of the N_2O emissions is usually calculated by applying emission factors (EF) related to the magnitude of N inputs to the agricultural fields (EF = emitted N_2O-N / N input). According to the guidelines of the Intergovernmental Panel on Climate Change (IPCC, 2006) for national emission reporting, a separation is made between (i) emissions related to excreta N deposited by the grazing animals and (ii) emissions related to fertiliser applications and other N inputs. While for fertiliser induced N_2O emissions, a default value of 1% is proposed by IPCC (2006), the default EF related to excreta of grazing cattle (denoted as $EF_{3PRP,CPP}$) is 2%. Most countries including Switzerland presently use this default values. However, the default $EF_{3PRP,CPP}$ value often overestimates observed pasture emissions (Bell et al., 2015; Chadwick et al., 2018) and does not take into account country specific conditions (climate, soil, management). Therefore, some countries have developed a country-specific EF (e.g. New Zealand, Saggar et al., 2015) which is still lacking for Switzerland. Additionally, it has been shown that separate EFs for urine and dung might be beneficial in describing the emissions and understanding the contributions of the different emission sources on a pasture (Bell et al., 2015). A better understanding of the individual contributions would also be very helpful to reduce the emissions, as e.g. dietary changes typically affect the excreted urine N which is mainly responsible for the high N_2O emission associated with excreta (Dijkstra et al., 2013). However, the range and thus the uncertainty of specific urine EFs is rather large (0-14%, n=40) as shown by Selbie et al. (2015) based on a survey of literature reports. Many of those studies measured the emissions on artificially applied urine or under laboratory conditions making these results questionable with regard to the applicability within greenhouse gas inventories.

The efficient use of fed N is essential to reduce the emissions associated to animal excreta. Studies have shown that an optimised feeding strategy can lead to less N excreted by the animals (e.g. Arriaga et al., 2010; Dijkstra et al., 2013; Yan et al., 2006). For this purpose, forage with a low N content (e.g. maize) can be used as a supplement to N rich grass and this subsequently leads to less N in the excreta, mainly in form of less urine N. A lower amount of N input to the pasture is supposed to produce less N_2O emissions, but corresponding emission experiments under real grazing conditions for a full season, to our knowledge, have not been reported hitherto.

1   Historically, most studies used static chambers to quantify N_2O emissions (Flechard et al., 2007). Chamber measurements are

2   ideal to quantify emissions on a small spatial scale and to attribute the measured fluxes to certain emission drivers, but for

3   excreta emissions these measurements were often performed on manually applied urine and dung patches (Bell et al., 2015;

4   Cai and Akiyama, 2016). Additionally, due to the strong heterogeneity of the emissions from a pasture (Cowan et al., 2015;

5   Flechard et al., 2007) chamber techniques are not ideal to compute field scale emissions for grazing systems. The eddy

6   covariance (EC) method overcomes this problem by integrating over multiple emission sources over a larger spatial domain.

7   The EC technique was already applied successfully to quantify N_2O emissions from pastures and grasslands (Jones et al.,

8   2011). Some studies also tried to compare different systems (e.g. intensive – extensive, different crops, land / lake) with one

9   EC tower (e.g. Biermann et al., 2014; Fuchs et al., 2018) by partitioning the fluxes based on wind direction and systems

10 geometry, but typically one tower for each system is preferable. In order to understand and quantify the emissions of a pasture,

11 the combined approach of EC measurements and chambers is regarded as the best solution (Cowan et al., 2015). The EC

12 systems can be used to quantify the field scale emissions while the chamber approach can be used to estimate the contributions

13 from single emission sources (urine patches, dung patches and other "background" areas).

14 In our experiment, we measured N_2O emissions from two neighbouring pastures simultaneously with the EC method over a

15 full grazing season. The two pastures differed in the energy to protein balance of the cows' diet. The small scale fluxes were

16 quantified with a fast-box chamber and up-scaled to match the EC flux footprints for comparison. Further on, we computed

17 the contribution of the different emission sources to the overall pasture emissions. The results were compared to default values

18 provided by IPCC and other literature values. The main goal of the study was to quantify the excreta related emission and the

19 corresponding EF for real grazing systems, and to analyse the specific contributions of dung and urine patches.

20 **2 MATERIAL AND METHODS**

21 **2.1 Experimental site**

22 The experiment was conducted at the research farm Agroscope Posieux in the Pre-Alps of Switzerland in the canton of Fribourg

23 (46°46´04´´N, 7°06´28´´E) during the grazing season of 2016 and already has been described in detail by Voglmeier et al.

24 (2018). The farm is located at an elevation of 642 m with an annual average temperature of 8.7 °C and a mean annual

25 precipitation sum of 1075 mm (MeteoSwiss, 2018). The soil consisted mainly of a stagnic Anthrosol with a loamy texture (see

26 Table 1). Soil profile samples for analysis of texture and other soil characteristics were taken at four locations on the pasture

27 in 2013 and 2016. The vegetation consisted of a grass-clover mixture typical for Swiss pastures (78 ± 12 % grasses and 15 ±

28 10 % legumes; main species: *Lolium perenne* and *Trifolium repens*, 10 sampling times between May and September). After

29 the last renovation treatment in 2007 the field had been used as an intensive pasture for cattle grazing with occasional grass

30 cuts for maintaining a homogenous sward. Beside the N input through excreta from the grazing animals, additional N had been

31 applied through fertiliser at a rate of about 120 kg N ha^{-1} per year between 2007 and 2015.

2.2 Experimental design

The experiment took place at a 5.5 ha pasture, which was divided into two separate systems differing in feeding strategy of the 12 cows per system (Fig. 1a). The northern system (system M) represented a N optimized feeding option where the diet of the cows consisted of grass with additional maize silage (roughly 20 % of the dry matter intake (DMI), fed in barn during milking periods) resulting in a demand optimized protein content in the diet (Arriaga et al., 2010; Yan et al., 2006). This was supposed to reduce the excreta N input to the pasture. The southern system (system G) represented a full grazing regime with no additional forage which resulted in a considerable protein surplus (see Table 2). Both systems were managed as a rotational grazing system with 11 paddocks (Fig. 1a) resulting in a typical rotation period of about 20 days. The size of the paddocks was adjusted for the different feeding strategies and resulted in typical sizes of 1700 m^2 for system M and 2200 m^2 for system G. The rotation of both systems was managed synchronously with a new rotation starting on the westerly paddocks (X.11 to X.16 with X indicating both systems) followed by the easterly ones (X.21 to X.25).

Grazing on the paddocks started with intermittent grazing phases in March and ended in early November with the main grazing season being between end of April and early October. During this time period eight full rotations took place. The cows typically spent 18 to 20 hours per day on the pasture and were brought to the barn twice a day (around 05:00 and 17:00 LT) for milking. However, in July and August the cows spent a longer time in the barn during daytime (up to six hours, see Fig. 2c) mainly due to high air temperatures and to a minor degree due to additional experiments of other research groups. Heavy rain events in June led to very wet soil conditions, which prevented grazing between the 8th of June and 4th of July and necessitated a grass cut on the 22nd and 27th of June (Fig. 2c).

2.3 N input to the pasture

During the grazing season, N input to the pasture mainly occurred in the form of excreta of the grazing animals and to a lesser extent as mineral fertilizer (Fig. 2d). The mineral fertilizer was ammonium nitrate (28 kg ha^{-1}) applied at the end of June and urea (42 kg ha^{-1}) with a split application between mid of August (western paddocks X.11–X.16) and early September (eastern paddocks X.21–X.25) due to concurrent grazing. In the present study we focus on the N input by grazing excreta and their effect on N_2O emissions. The comparison between the field-scale EC method and the small scale chamber measurements also required estimates of the number of dung and urine patches on the pasture. These numbers were calculated as described in Sect. 2.7 based on the excreted N amounts. N excretion cannot easily be measured in the field, but it can be calculated based on the energy demand of the cows and measured N in feeds and products (e.g. milk, body weight gain). We followed the approach described by Felber et al. (2016) to calculate the energy and N flows of the dairy cows in the experiment and to calculate daily values of excreted N per cow. Input parameters to the budget calculation were daily measurements of milk yield, milk N content and body weight gain as well as seasonal measurements of protein content of the grass (eight times between end of April and end of September) and of the maize silage (three times between beginning of May and beginning of

1 September). The breakdown of the excreted N in urine N and dung N was based on work by Bracher et al. (2011). For further

2 details see Voglmeier et al. (2018), where the corresponding uncertainty of the total N and urine / dung N was estimated to be

3 15 % (2σ) for the same experiment. Seasonal statistics of the input variables are given in Table 2.

4 **2.4 Small scale flux measurements**

5 **2.4.1 Excreta detection**

6 The localisation of fresh dung and urine patches was essential in this study to measure N_2O emissions attributable to specific

7 excreta sources. Intensive observation areas of 10 x 10 m or 15 x 15 m close to both EC towers in the paddocks X.11 and X.21,

8 respectively, (see Fig. 1a) were selected. Within these areas fresh dung and urine patches were mapped typically 1-3 days after

9 grazing of the respective paddock. Dung pats were mapped visually and labelled for subsequent chamber measurements. For

10 urine patches a direct visual identification was not possible. Bates et al. (2015) demonstrated the ability of surface-soil electrical

11 conductivity measurements to detect urine patches. Using this approach we mounted a soil probe (GS3, Meter Group, US; for

12 soil moisture, temperature and electrical conductivity measurements) on a hand-held stick and mapped the intensive

13 observation area on a 25 cm grid (Fig. 3). Based on pre-experimental tests, areas with conductivity values below a threshold

14 of 0.15 mS cm^{-1} (dark blue areas in Fig. 3a) were considered as 'background' without recent influence of excreta. Spots with

15 a conductivity above the threshold were marked as possible urine patches for the chamber measurements. Time series of

16 electrical conductivity measurements (Fig. 3b) on manually applied urine patches in 2017 illustrate the long term effect and

17 demonstrates the possibility to distinguish between background areas and urine patches more than 10 days after the application

18 of urine.

19 **2.4.2 Fast-box measurements**

20 Small scale emissions from urine and dung patches as well as background pasture areas were measured with a fast-box (FB)

21 chamber (Hensen et al., 2006). The measurements took place on the paddocks X.11 and X.21 (Fig. 1a) between beginning of

22 July and mid of October and were therefore taken mainly during dry soil conditions (Fig. 2a, periods with VWC < 0.4).

23 Measurements usually started after the excretion detection (Sect. 2.4.1) and about 1-2 days after the end of grazing (EOG).

24 The age of the excreta patches is important for the interpretation of the measured fluxes. However, the exact determination of

25 the excreta age was not possible. Thus, the time since EOG was used as excreta age for each FB measurement. The potential

26 age variability of a single excreta patch resulted from the sojourn time of the cows on the paddock which typically was in the

27 range of 1–1.5 days.

28 The manually-operated opaque 0.8 m x 0.8 m x 0.5 m box was connected to a fast response quantum cascade laser analyser

29 (QCL, Aerodyne Research Inc.) that was also used for the EC system on the respective field (see below Sect. 2.5.1). The

30 sample air was drawn continuously from the FB headspace through a 40 m 1/4" polyamide (PA) tube to the analyser, allowing

31 measurements within a radius of about 35 m on the paddocks X.11 and X.21 (see Fig. 1). The sample flow rate Q was typically

1   around $8 \, l \, min^{-1}$. The box was modified by using a defined vent to ambient air through a tube of 4 cm diameter and 1 m length.

2   The inlet of the vent tube was packed with a foam material over a length of 10 cm to avoid uncontrolled air exchange due to

3   wind induced pressure fluctuations. The chamber was also equipped with a GMP343 CO_2 probe (Vaisala, FI) to measure the

4   soil respiration, which was used for quality control purposes (Sect. 2.4.3). The increase in N_2O concentration after placing the

5   chamber on the soil with a flux F_{Cham} was recorded every three seconds for a time period of about 90 seconds (taking into

6   account the time delay due to tube sampling). The inflow of the background concentration C_{bg} into the chamber volume V

7   (with area A) through the vent lead to lower measured concentration values C. This can be described by the following

8   differential equation for the chamber headspace concentration $C(t)$:

$$V \frac{\delta C}{\delta t} = A \cdot F_{Cham} - Q(C - C_{bg}) \tag{1a}$$

11   This is a combination of the two equations for static chambers (right-hand term = 0) and for the dynamic chamber (left-hand

12   term = 0). Solving of the equation yields the explicit time function:

$$C(t) = \frac{A \cdot F_{Cham}}{Q}\left(1 - e^{-\frac{Q}{V}t}\right) + C_{bg} \tag{1b}$$

15   For small values of the exponent Q/V*t (slow chamber volume exchange of about 40 min and short measurement time) as

16   characteristic for the present fast-box measurements, the entire bracket term can be linearized with a series expansion to

17   (Q/V*t). Inserting the resulting function for C(t) into Eq. 1a yields:

$$V \frac{\delta C}{\delta t} = A \cdot F_{Cham}\left(1 + \frac{Q}{V}t\right) \tag{1c}$$

20   With the FB dimensions and sampling flow rate as given above and a maximum accumulation time $t \leq 2$ min, the deviation

21   from the ideal linear increase of a fully closed static chamber was $\leq 5\%$. The flux was finally calculated by using the HMR

22   package (Pedersen et al., 2010), which uses linear and non-linear regression to fit the measured concentration values. The

23   uncertainty of an individual box measurement is estimated to be around 20 % (Hensen et al., 2006).

24   In order to relate the measured fluxes to environmental driving parameters the following sensors were placed inside on the

25   chamber: a thermocouple (type K) for air temperature measurement within the chamber, a GS3 probe (see Sect. 2.4.1) for soil

26   moisture, soil temperature and soil conductivity measurements (c. 0-5 cm depth) and a ML3 Thetaprobe (Delta-T Devices Ltd,

27   UK) for soil moisture and soil temperature observations (c. 0-10 cm depth). All measured data values were stored on a data

28   logger mounted on top of the box and transferred to a computer in the nearby shelter or trailer. A customized LabView

29   (National Instruments, US) program allowed for online inspection of all measured data values including the gas concentrations.

2.4.3 Quality control and system comparison

FB fluxes were selected for post-processing after fulfilling certain quality criteria. In a first step, the R-squared value of any flux calculation had to exceed 0.9 (e.g. for N_2O flux either the R-squared value of N_2O, CH_4 or CO_2 had to exceed 0.9). For urine patches, the soil conductivity had to exceed 0.25 mS cm^{-1} at the beginning of the measurements (see also Fig. 3b) in order to exclude possible old urine patches (of previous management rotations). Presumable old patches were therefore rejected for further processing. Background fluxes were removed from further processing if the flux value exceeded 40 µg m^{-2} h^{-1} (=4 x median value) to ensure that undetected urine patches at the chamber surroundings did not influence the flux measurements. Finally, 360 and 293 flux measurements met the criteria on system M and G, respectively. These measurements were composed of 238 background fluxes, 242 urine patch fluxes and 173 dung fluxes.

For a direct comparison of the FB measurements on the two pasture systems, the fluxes obtained on the same day were ordered based on their magnitude for each system and source class. Due to the synchronous grazing regime, the fluxes represented the same excreta age (e.g. on day 3 after EOG). However, synchronous FB measurements on both systems were not always performed. Resulting numbers of data pairs are 46, 54 and 40 for background, urine and dung fluxes, respectively.

2.5 Field scale flux measurements

2.5.1 Eddy covariance system

For field scale flux measurements EC towers were installed in the middle of the two pasture fields to account for the predominant wind directions north-east and south-west (Fig. 1) and were fenced with a radius of 2-3 m to avoid unwanted animal contact. The measurement height was 2 m which enabled a good footprint coverage (Fig. 4, Sect. 2.5.4) of both fields and allowed to measure fields-scale fluxes of both systems.

The two EC systems were identically equipped with an ultra-sonic anemometer-thermometer (further on named sonic, HS-50, Gill Instruments Ltd., UK) to quantify the turbulent mixing by measuring the three dimensional wind velocity (u,v,w) and air temperature. Dry air mixing ratios of N_2O were measured with closed-path quantum cascade laser spectrometers (QCL, QC-TILDAS, Aerodyne Research Inc.) that analysed air samples drawn through a 25 m PA tube (inner diameter 6 mm) by a vacuum pump (Bluffton Motor Works, flow rate ca. 13 l min^{-1}). One filter at the inlet (AcroPak, Pall Corporation, 0.2 µm) and one before the instrument (Midisart 2000, Sartorius Stedim Biotech GmbH, 0.2 µm) were used for each system to filter out particles. The distance of the inlets of the QCL from the centre of the sonic head were around 20 cm and the QCL instruments were placed in a temperature controlled environment (trailer at system M, shelter at system G) about 20 meters north (system M) or south (system G) of the EC towers.

The sample frequency of the EC system was 10 Hz. A customized LabView (National Instruments, US) program was used to combine the data strings of the individual instruments and store them as binary raw data for offline analyses. Additionally the program visualized the measurements and fluxes of the N_2O concentrations and fluxes, calculated with an online flux calculation. The program also allowed to check the EC system by remote access.

2.5.2 Flux calculation

A customized program written in the statistical software R (R Core Team, 2016) was used to calculate EC fluxes for 30 min intervals (similar to Felber, 2015a; Felber et al., 2015b). The program is based on Ammann et al. (2006, 2007). In a first step, 10 Hz data outside a plausible physical range were identified and replaced by a running mean filter with a window size of 500 data points. In a next step, wind vector components were rotated into the mean wind direction using the double coordinate rotation technique (Kaimal and Finnigan, 1994), and concentration values were subject to linear detrending within an averaging interval of 5 min.

The EC flux is defined as the covariance of the vertical wind speed and the trace gas mixing ratio. Due to the long inlet tube the time series of the trace gas signals are delayed in relation to the wind measurements by a quasi-constant lag time of about six seconds for system M and seven seconds for system G. Thus, the trace gas signals have to be shifted to obtain the correct covariance flux (Langford et al., 2015). In a pre-evaluation, the 'default lag' was determined as the most frequent position of the maximum absolute value of the cross-covariance function over periods of weeks to months (depending on instrument maintenance). Then it was checked for each half-hour period whether the individual 'dynamic' lag was within a time window of 0.61 seconds around the default lag. If this was the case, the dynamic lag was used, otherwise the default lag was used. In order to minimize the effect of non-stationarities in the time series, the 30 min flux was finally calculated as average over six 5-min subinterval flux values. This caused a minor low-frequent spectral loss (1-5%) that was quantified (and corrected for) using Kaimal-cospectra and the theoretical transfer function for block averaging.

The fluxes measured by EC systems are also subject to different high-frequency losses due to sensor separation and in case of N_2O air transport through the inlet tubes (Foken et al., 2012). These damping effects can lead to a significant underestimation of the flux and must be corrected. Based on Ammann et al. (2006) the half-hourly high frequency losses were quantified using the 'ogive' method where the damping factor was calculated by fitting the normalized cumulative co-spectrum of N_2O to the one of the sensible heat at a frequency of 0.065 Hz. In a post processing step, these half-hourly damping factors were filtered for favourable conditions e.g. low noise level of the ogive and the flux. The selected values were used to compute a wind speed and stability dependent damping function which was finally used to estimate the damping factor. Depending mainly on the wind speed, a damping effect of 10 – 30 % was found and corrected for.

EC fluxes were measured continuously over the grazing season. Since the present study is focussed on N_2O emissions from grazing, time periods with strong influence of N_2O emissions from fertilization and harvest events (see Fig. 2c-d) were excluded for computation of cumulative emissions and for comparisons between field scale and small scale measurements. These exclusion periods were limited to the 15 d following fertilization or harvest and led to a rejection of 47 days during the grazing season. The criterion is based on observed EC fluxes (Sect. 3.1) and is in accordance with Jones et al. (2011). The time periods used for calculation of the cumulative grazing emissions are further on defined as grazing-only periods (GOP) and accumulated to 198 days.

2.5.3 Quality control and gap filling

EC flux measurements are subject to different sources of measurement problems and quality issues which often result in data loss or data rejection. These sources can be instrument specific like power failures or malfunctioning, environmental driven like measurements under non ideal conditions (e.g. low turbulence) or a combination of both (Papale, 2012). Power outage, instrument maintenance (only on system M) and delayed installation (only on system G) led to data losses during the GOP of 12 and 17 % for systems M and G, respectively. Data rejection due to low friction velocity ($u_* < 0.07$ m s^{-1}) and large vertical tilt angle (-2° to 6°) of the wind vector led to a further data loss of about 35 %. Because non-stationarity of the flux was already reduced by the short averaging/detrending interval of 5 min., a quality selection based on non-stationarity (Foken et al., 2012) had little effect and was therefore not used here. Additional rejection of wind sectors influenced by the farm facilities, trailer or shelter and to avoid cross-influences from the other pasture system (wind dir = 280° - 25° and wind dir = 97° - 195°) contributed to an overall data loss of 64 and 69 % for systems M and G. The resulting occurrence of data gaps showed a diurnal pattern with stronger data loss during the night, which was driven by the wind pattern with typically stronger wind speeds during daytime and calm nights.

The gaps in the flux time series needed to be filled in order to compute cumulative sums over a certain period of time. However, no well-established reference method for the gap filling of N$_2$O fluxes exists to date. We followed the evaluation of Mishurov and Kiely (2011) and used a lookup table method (LUT) with three parameters: one for the preceding cumulative rainfall of the last 12 hours with three classes (no rainfall, 0-2 mm, >2 mm), one for the percentiles of the soil temperature at 5 cm depth during the GOP with four classes (0-25th percentile, >25th percentile – median, >median – 75th percentile, >75th percentile), and one for the footprint-weighted (Sect. 2.5.4) averaged cow density (cows ha^{-1}) on the single paddocks over the preceding five days (0, 0 – 2, > 2 cows ha^{-1}). To check the sensitivity towards different gap filling methods three other techniques were compared to the LUT approach. [I] Running mean with a variable filter window size and at least 12 values; [II] Monthly mean diurnal variation (MDV, see Zhao and Huang, 2015) with a running half hourly window size of five in order to have more values during night-time, [III] seasonal MDV based on half-hourly values averaged over the whole grazing season. Due to the delayed installation of the EC tower on the southern field all values prior to the 14th of April on system G resulted from the gap filling routine. The uncertainty of gap filling for seasonal cumulative fluxes was estimated from the standard deviations of monthly cumulative fluxes retrieved with the different gap filling methods during GOP, which resulted in an uncertainty of 14 and 18 % for the system M and G, respectively (1 σ). It was assumed, that this uncertainty reflects the sum of all important individual uncertainties of the cumulative emissions (e.g. Sect. 3.3.1 and 4).

The experimental setup was expected to result in very similar systematic errors of the two EC systems, thus only the independent (or random) errors have to be considered for comparing the two neighbouring systems (Ammann et al., 2009). As the cumulative fluxes of both EC systems were by chance of similar magnitude (Sect. 3.1 and Sect. 3.3.1), the random uncertainty of the cumulative EC fluxes was determined from the differences between the cumulative, monthly EC fluxes of the two towers and resulted in a relative uncertainty of 5 % (1 σ).

1   **2.5.4 Footprint modelling**

2   EC measurements yield a spatially integrated flux over a certain area represented by the flux footprint (Schmid, 2002). In the

3   present study, this fooprint typically extends over multiple grazing paddocks depending on wind direction and turbulence

4   intensity. Therefore quantitative footprint information is needed for the comparison of the EC fluxes with the up-scaled FB

5   measurements (Sect. 2.7), and the footprint has to be checked for the spatial dimension to be sure that the measured flux is

6   mainly dominated by the area of the system and not contaminated by the neighbouring systems (either the other grazing system

7   or fluxes originating from surrounding fields). In this study an open source version of a backward Lagrangian Stochastic

8   dispersion footprint model (bLS) was used (Häni, 2017; Häni et al., 2018), based on Flesch et al. (2004). The flux to emission

9   ratio is calculated following Eq. 2

$$\frac{F_{EC}}{E_j} = \frac{2}{N} \sum_{i=1}^{n_j} \frac{w_{ini}^i}{w_o^i} \tag{2}$$

10   where F_{EC} is the measured EC flux, E_j the surface emission of paddock (source area) j, N the total number of released particles,

11   n_j the number of touchdowns within paddock j, w_{ini}^i the vertical release velocity and w_o^i the touchdown velocity of the

12   particles.

13   In order to calculate the footprint for a 30 min period, $N = 80'000$ fluid particles were released backwards in time using the

14   wind and turbulence parameters calculated from the sonic measurements of the EC systems. The systematic uncertainty of the

15   bLS model was estimated to about 10 % (Flesch and Wilson, 2005; Wilson et al., 2013). The half-hourly footprint fractions of

16   the individual paddocks were used to up-scale the small scale measurements to the EC flux footprint (Sect. 2.7) for inter-

17   comparison of the two flux measurement methods.

18   In addition, the seasonally integrated footprint extension was analysed, taking into account the wind direction and u∗ filtering

19   as described in Sect. 2.5.3. The analysis showed a distinct separation of the footprint distributions for the two systems (Fig. 4)

20   with only marginal contributions of the other system (<2.5 %). More than 80% of the footprint contributions was from the

21   actual rotation area (without the optional areas indicated in grey colour in Fig. 1a).

22   **2.6 Environmental parameters**

23   In order to relate the measured fluxes to meteorological driving parameters an automated weather station (with data logger

24   CR10X, Campell Scientific Ltd., UK) was installed at the northern field next to the Sonic. A WXT520 (Vaisala, Vantaa, FL)

25   measured the wind speed, precipitation, temperature and barometric pressure, and global radiation was measured with a

26   pyranometer (CNR1, Kipp&Zonen, Delft, NL).

27   Soil moisture and soil temperature were measured continuously with two repetitions on each pasture system close to the EC

28   towers with ML3 Thetaprobe (Delta-T Devices Ltd, UK) devices at a depth of 5, 10, 20 and 40 cm.

2.7 Up-scaling of chamber measurements to eddy covariance footprint

Pasture N_2O emissions result from a combination of 'hotspot' emissions from urine and dung patches and of 'background' emissions from the other pasture areas. Even though the FB measurements (Sect. 2.4.2) allowed for quantification of single emissions sources, quantifying the contributions to the overall pasture emission is challenging due to the inherent heterogeneous nature of these emissions (e.g. spatial dimension, emission strength, temporal behaviour, number of excreta patches). The EC method, on the other hand, allowed to measure the combination of all pasture sources by integrating over multiple paddocks (see footprint, Fig. 4).

FB measurements were up-scaled to the EC footprint to allow a direct comparison between the two measurement approaches and to compute the contributions of the different emission sources to the overall pasture emission. The up-scaling procedure is illustrated in Fig. 5. The number of urine and dung patches on the paddocks was estimated by using the daily N excretion rate (Sect. 2.3), the daily grazing duration of the cows, a N loading of 22 g N per urination event (Misselbrook et al., 2016) and of 12.5 g N per dung pad (Cardenas et al., 2016). During the grazing season, about 12.5 dung patches d^{-1} cow^{-1} and a ratio of dung to urine patches of 1.3 for system M and 1.1 for system G was calculated. This compares well to values from literature (Orr et al., 2012; Oudshoorn et al., 2008; Villettaz Robichaud et al., 2011). Due to very similar field scale N_2O emission pattern (Sect. 3.1) and comparable soil measurements (Fig. 2), it was assumed that soil parameters were homogenous on the pasture and that the soil measurements on system M were representative for the whole field.

The FB derived N_2O emissions for the different sources were analysed for the potential driving parameters excreta age, soil temperature and soil moisture. For this purpose various regression models (using the statistical software R; R Core Team, 2016) were tested using different predefined function types (linear, exponential, polynomial functions, sigmoidal). Based on goodness of fit and statistical significance of regression coefficients, the most suitable relationships were chosen and applied to produce continuous emission time series for the paddock areas (Fig. 5):

(I) Background fluxes were parametrized as a function of soil moisture at a depth of 5 cm using the soil profile information provided in Sect. 2.4 by using a logistic regression.

(II) Urine patch emissions were parametrized as an exponential decay function of excreta age. To account for different environmental conditions, the deviations of the single emissions to this temporal emission pattern was again parametrized as a function of soil temperature and moisture at a depth of 5 cm (Sect. 2.6). Up-scaling fluxes to the paddocks sizes involved additional information on the computed number density of urine patches (as mentioned previously).

(III) Dung patch emissions were parametrized as a second order polynomial function of excreta age. Paddock emissions were calculated by applying this function to the computed number of dung patches (as previously mentioned) per paddock.

The up-scaled paddock emissions were finally compared to the EC fluxes by applying the computed footprint fractions of the paddocks (Sect. 2.5.4) in order to validate the FB measurements and to quantify the uncertainty of the up-scaling process.

1   The area related N_2O emissions for urine and dung were also converted to emissions per cow and grazing hour. For this

2   purpose, the up-scaled paddock emissions were combined over all paddocks, accumulated for the GOP, multiplied by the

3   pasture area of each system and divided by the number of cows and the grazing duration (Fig. 5). The resulting emissions

4   associated to animal excreta were then related to the excreted N of the cows (Sect. 2.3) to obtain an excreta related EF that is

5   comparable to the one provided by the IPCC guidelines ($EF_{3PRP,CPP}$; IPCC, 2006).

6   **3 Results**

7   **3.1 EC fluxes**

8   Observed EC fluxes on both pasture systems showed an almost identical temporal pattern (Fig. 6). The half-hourly fluxes on

9   each system showed considerable variation during the grazing season with clear peaks after fertilization (grey shaded areas)

10 and after grazing phases in the nearby paddocks (e.g. peaks in May, beginning of August). The overall highest emissions (28.7

11 and 21.6 g N_2O-N ha^{-1} h^{-1} for system M and G) were measured directly after the fertilizer application, which followed a harvest

12 of hay at the end of June. This harvest event also led to an increase in the measured N_2O fluxes (0.5 – 3.0 g N_2O-N ha^{-1} h^{-1})

13 which lasted less than one day. The partial fertilizer application in mid of August resulted in higher fluxes compared to the

14 following one in early September. The relatively high emissions during the first full grazing event beginning of May were

15 characterized by high soil moisture contents (see also Fig. 2a) whereas the very wet soil conditions and the corresponding

16 grazing break during June resulted in low fluxes in both systems. The small observed fluxes from mid of March until end of

17 April resulted mainly from background fluxes and sporadic grazing (Fig. 2c). Occasional negative individual flux values

18 between 0 and -1.5 g N_2O-N ha^{-1} h^{-1} were observed in both systems (7-8% of the cases). However, these fluxes exclusively

19 occurred in cases, when no defined peak in the cross-covariance function could be identified (and thus the default lag was

20 used, Sect. 2.5.2). Thus it can be concluded that the negative fluxes were generally below the detection limit.

21 During the GOP (excluding the grey shaded fertilizer influenced time periods in Fig. 6), the fluxes were still very similar for

22 the two pasture systems M and G with a mean and standard deviation of 0.32 ± 0.36 vs 0.33 ± 0.37 g N_2O-N ha^{-1} h^{-1},

23 respectively. A mean diurnal cycle of the measured fluxes could be observed in both systems with highest values typically

24 occurring in the afternoon and, on average, about 10 – 20 % lower values during the night.

25 **3.2 Chamber fluxes**

26 **3.2.1 Comparison of pasture systems**

27 FB chamber fluxes of background and dung patches were considerably smaller compared to the fluxes of urine patches (Table

28 3, Fig. 7). Freshly deposited urine patches under 3 days old could result in N_2O emissions larger than 100 times the values of

29 background areas. The relative variability within the different source classes (urine, dung, background) were very high and

30 resulted in standard deviations larger than the associated mean values. The excreta fluxes measured on system G tended to be

1   somewhat higher in magnitude compared to system M, but no significant difference (p>0.05) was found. Also for the

2   background fluxes no significant (p>0.05) difference between the two pasture systems was observed. Therefore all FB fluxes

3   were combined for further processing without taking into account the different pasture systems.

4   **3.2.2 Dependence on excreta age**

5   The information on the temporal pattern of the excreta and background fluxes after grazing is important for the time integration

6   of the individual sources and for the comparison with the EC measurements. In order to analyse and parameterize the temporal

7   evolution of the emissions, the measured FB fluxes of each source class were averaged over 3-day periods and were related to

8   the excreta age Δt_{EOG} (Fig. 8), defined as days after EOG (Sect. 2.4.2).

9   Background fluxes were on average considerably smaller than excreta fluxes and showed small persisting emissions without

10   systematic dependence on time since grazing. In contrast, for urine patch fluxes a clear relation to Δt_{EOG} was found. Highest

11   fluxes were usually observed within the first days after the urination event. Afterwards, they rapidly decreased with time

12   although with a high variability that can partly be attributed to the influence of environmental conditions (see Sect. 3.2.3). The

13   age dependent evolution of urine patch emissions ($F_{U,age}$) was parameterised with an exponential decay function fitted to the

14   data points in Fig. 8:

16         $F_{U,age} = a_1 \cdot \exp^{b_1 \cdot \Delta t_{EOG}}$ (3)

17   The coefficients of Eq.3 – Eq.8 are presented in Table 4 and apply to fluxes in units of μg $N_2O\text{-}N$ m^{-2} h^{-1}.

18   Dung patch fluxes also showed a relation to excreta age (Fig. 8), however less pronounced compared to urine patches, and the

19   highest emissions were typically observed between 4 – 11 days after dung deposition. However they were still smaller on

20   average than the urine patch emissions during the entire observed age period. Because the evolution of dung emissions $F_{D,age}$

21   after the observed 20-day age period is unclear and a meaningful functional extrapolation was not possible, we decided to use

22   a simple 2nd order polynomial for parameterisation purposes. This allowed to reproduce the initial increase with age and a rapid

23   decrease to zero beyond the measured age range:

25         $F_{D,age} = a_2 + b_2 \cdot \Delta t_{EOG} - c_2 \cdot \Delta t_{EOG}{}^2$ (4)

26   The fitted polynomial function is only applicable up to $\Delta t_{EOG} \approx 25$ d, where it crosses the zero line.

28   **3.2.3 Dependence on environmental conditions**

29   Measured chamber fluxes were analysed in relation to driving soil parameters (Sect. 2.6). For dung patch emissions, no relation

30   to these parameters was found (thus $F_D = F_{D,age}$). For background fluxes no significant dependence on soil temperature (p<0.05),

31   but a clear dependence on the volumetric water content (*VWC*) at a depth of 5 cm was found. The background fluxes had a

large variability and could roughly be separated by three different VWC sectors (<0.27, 0.27-0.33, >0.33). In the sector below a VWC of 0.27, fluxes typically ranged between -3 µg N_2O-N m^{-2} h^{-1} and 15 µg N_2O-N m^{-2} h^{-1} whereas in the upper sector above a VWC of 0.33 the fluxes showed typical values between 0 µg N_2O-N m^{-2} h^{-1} and 30 µg N_2O-N m^{-2} h^{-1}. Nevertheless, the variability was especially pronounced in the VWC range between 0.27 and 0. 33 with fluxes ranging between 0 µg N_2O-N m^{-2} h^{-1} and 40 µg N_2O-N m^{-2} h^{-1}. Thus this VWC range also comprised of the overall highest background fluxes. However, averaging the fluxes by VWC intervals of 0.05 resulted in very similar values of about 12 ± 3 µg N_2O-N m^{-2} h^{-1} above a VWC of 0.3. Hence, the measured background fluxes could be parametrised with the following functional relationship:

$$F_{BG} = \frac{a_3}{1+exp^{(b_3-VWC)/c_3}} \tag{5}$$

This logistic regression curve has a strong effect below VWC values of 0.30 but stays fairly constant at higher VWC contents and converges to a flux of 12.6 µg N_2O-N m^{-2} h^{-1}. Below a VWC of 0.2 the logistic regression converges to a background flux of 0 µg N_2O-N m^{-2} h^{-1}.

Measured urine patch emissions not only showed a clear response to the excreta age as shown in Sect. 3.2.2 but also to changes in T_S and VWC. On a specific Δt_{EOG}, $F_{U,age}$ could vary significantly and correlated typically with soil conditions. The highest flux (5117 µg N_2O-N m^{-2} h^{-1}, $\Delta t_{EOG} = 6d$) was measured at a T_S of 18 °C and a VWC of 0.42 while the lowest measured flux (34 µg N_2O-N m^{-2} h^{-1}) on a similar Δt_{EOG} was measured at a low T_S (1°C) and a lower VWC (0.3). Maximum positive measured FB flux deviations (Sect. 2.7) from Eq. 3 were generally observed for wet (VWC > 0.45) and warm (>17 °C) soil conditions while low T_S and VWC resulted in negative flux deviations. Thus, the final regression model for urine patch emissions (Eq. 6) consists of multiple equations (Eq. 3, 7, 8) which relate the measured fluxes to the temporal decay (Eq. 3) and a deviation $\Delta F_{U,env}$ to it, where $\Delta F_{U,env}$ was parametrized as a function of environmental driving parameters T_S and VWC_U (Eq. 7 and 8, Fig. 9).

$$F_U = F_{U,age} + \Delta F_{U,env} \tag{6}$$

$$\Delta F_{U,env} = (a_4 + b_4 \cdot VWC_U + c_4 \cdot T_S) \cdot Corr_{U,env}(\Delta t_{EOG}) \tag{7}$$

$Corr_{U,env}$ corrects $\Delta F_{U,env}$ for different urine patch ages as the deviation can be larger for relatively new patches compared to older ones. This correction factor was found to be a linear relationship (p<0.01) between 1.35 for a Δt_{EOG} of 0 days (after the patch deposition) and 0.35 after 20 days. VWC_U (Eq. 8) accounts for different soil moisture conditions at the surface below an urine patch and nearby background areas and was parametrised as a function of background VWC and $\Delta t,_{EOG}$ (Eq. 8).

$$VWC_U = VWC + a_5 \cdot exp^{b_5 \cdot \Delta t_{EOG}} \tag{8}$$

3.3 Up-scaled chamber fluxes

3.3.1 Comparison between up-scaled chamber and EC fluxes

Generally the field scale fluxes represent the area integral of management related (excreta patches) and environmentally driven small scale fluxes. Therefore the relationships presented in Sect. 3.2.2 (dependency on excreta age) and Sect. 3.2.3 (environmental driving parameter) were applied to up-scale the FB measurements to the paddock size during the GOP.

As shown exemplary for an 18-day period in Fig. 10b, the magnitude of the management related up-scaled paddock fluxes depended mainly on the grazing duration on the single paddocks (similar slope for different paddocks M11–M14). The maximum of the emissions was typically calculated at the end of the grazing period on the respective paddocks. The lower limit of the fluxes was given by the estimated background fluxes, especially at the beginning of a new rotation and stayed therefore rather constant for VWC values above 0.3 (Eq. 5, Sect. 3.2.3). Variations in environmental conditions (mainly important for soil moisture) led to rapid changes in the emission level as long as significant urine patch emissions were present. These rapid variations occurred typically after stronger precipitation events (as shown in Fig. 10a for onsite meteorological and soil measurements).

Up-scaling the paddock fluxes to the EC footprint allowed a direct comparison with the EC fluxes on a half-hourly basis (Fig. 10c). The up-scaled FB fluxes compared well in magnitude with the measured EC fluxes and showed a similar temporal behaviour. While generally a response to variations in environmental driving parameter could be observed, it was less pronounced for the up-scaled FB fluxes in comparison to the EC fluxes.

Gapfilling of the EC fluxes (Sect. 2.5.3) allowed the calculation of the cumulative N_2O emissions during the GOP (solid lines in Fig. 11). These area related emissions were very similar between the two systems throughout the GOP with seasonal sums close to 1500 g N_2O-N ha^{-1}. Cumulating the N_2O emissions not only enabled a more quantitative comparisons between the systems, but also allowed a better comparison between the two measurement approaches (Fig. 11). The emissions of the up-scaled FB matched the EC emissions rather well with differences of the seasonal sums below 3 %. Distinct differences were mainly observed in May and June when FB derived emissions were significantly overestimated compared to EC. At the end of the grazing period slightly higher emissions were estimated from the up-scaling routine compared to the measured EC emissions. Monthly absolute differences between the cumulative EC and the up-scaled cumulative FB sums were normally distributed (p<0.05) with 1σ values of 26 % and 25 % for system M and G, respectively. Within this uncertainty range no difference between the two measurement approaches was observable.

3.3.2 Emission breakdown into contribution sources

The excellent match between the EC fluxes and the up-scaled chamber based fluxes showed that the used relationships with excreta age and environmental drivers (see Sect. 3.2) was reasonable and allowed the separation into single emission sources (Fig. 12). Except for the beginning of the grazing season when grazing rate was very low (see Fig. 2), the urine patch emissions dominated the field scale fluxes. In May, this effect was even more pronounced due to the wet soil conditions. Based on the

1 up-scaling, the averaged urine patch emission of both systems were responsible for about 57 % of the pasture emissions.

2 Background contributed to about 38 % and dung emissions to about 5 % to the overall field emissions. Both systems had very

3 similar contributions, with only 1 % difference in the dung contribution as a result of a different N excretion ha^{-1} on the pasture

4 by dung (Table 5). Background emissions were simulated to be constant for most of the GOP due to the weak sensitivity of

5 Eq. 5 to VWC and the undetected sensitivity towards soil temperature.

6 **4 Discussion**

7 **4.1 Area related and animal related emissions**

8 The EC and up-scaled FB emission results presented in Sect. 3.3.1 are normalized by area and showed the emissions for the

9 EC footprint (see also summary in Table 5). The good agreement with a relative difference below 1.5 % for yearly sums (which

10 is far below the uncertainty range, see Table 5) between the two independent approaches supports their quality (including the

11 up-scaling procedure) in this study. We assume that the EC fluxes are on average representative for the whole pasture system,

12 although the contribution of the central paddocks X.11, X.12 and X.21, X.22 to the EC footprint is generally higher than the

13 contribution of the other more distant paddocks (Fig. 4). We found no indication of significant differences between the

14 paddocks concerning soil conditions, vegetation productivity or other characteristics (data not shown). An alternative up-

15 scaling of the FB measurements to the entire pasture system (without taking the EC footprint into account) representing the

16 average emission over all rotation paddocks (Table 5, FB up-scaled to pasture system emissions) differed less than 4 % from

17 the EC footprint related emissions.

18 For assessing the effect of the N reduced diet on excreta related N_2O emissions, the emissions per cow and grazing hour were

19 compared, taking into account the different pasture sizes for system M and G (acc. to Sect. 2.7). The corresponding results in

20 Table 5 show about 25 % lower excreta related N_2O emissions per cow for the herd in system M than for the herd in system G

21 during the GOP. The difference is not statistically significant probably due to the considerable uncertainties resulting from the

22 FB up-scaling procedure. For comparing the two herds the parallel direct EC measurements are better suited as only random

23 uncertainties have to be taken into account (Sect. 2.5.3), yet they also include the background emissions. The EC based N_2O

24 emissions per cow were 0.20 ± 0.03 and 0.27 ± 0.05 g N_2O-N cow^{-1} h^{-1} for system M and system G, respectively, and resulted

25 in a significant difference of 0.07 ± 0.02 g N_2O-N cow^{-1} h^{-1} between the two herds. This indicates the ability of an N adjusted

26 forage to reduce the excreta N content and related emissions of N_2O. It has to be noted, that this evaluation does not comprise

27 the full N_2O emission of the pasture fields or of the milk production system but only the emissions related to grazing excreta

28 following the IPCC concept (EF$_{3PRP,CPP}$; IPCC, 2006). Any further N_2O emissions e.g. related to fertiliser application on the

29 pastures or for the supplement maize production were not taken into account here. A comparison of entire production systems

30 would require many additional assumptions outside the specific scope of this study. It also has to be considered that the N

31 optimisation of the diet is not necessarily linked to the supplemental feed of arable crops like maize, but may as well be

1 achieved with different feed strategies (e.g. grass varieties with a high content of water soluble carbohydrates; Misselbrook et

2 al., 2013).

4.2 Excreta related emission factor

4 Area or cow related emissions as described in Sect. 4.1 enabled the comparison of the different measurement approaches and

5 the discussion of the diet effects on N_2O emission. However, results presented in literature or used in national inventories

6 typically relate emissions to the N inputs within a given time period using EFs. The annual excreta related EF (Table 5) in the

7 present study was 0.74 ± 0.26 % for system M and 0.83 ± 0.29 % for system G. These EFs are based on the combined, up-

8 scaled FB measurements of urine and dung patches (see Sect. 3.3.2) relative to the N excreted on the pastures during the GOP

9 (Table 5). Their uncertainty is defined by the combined uncertainty of the up-scaling method (Sect. 3.3.1) and the N input

10 estimation (7.5 %). The difference in the EFs between the systems is therefore not statistically significant.

11 The resulting EFs were significantly smaller compared to the proposed default $EF_{3PRP,CPP}$ of the IPCC guidelines for cattle

12 excreta (2 %; IPCC, 2006), which makes the use of the latter in the Swiss national inventory questionable. The up-scaled FB

13 measurements also allowed to separately calculate the EFs for urine and dung. We found EFs of 1.12 ± 0.43 % and 0.16 ± 0.06

14 % for urine and dung, respectively (average of both systems due to small difference, see Table 5). These EFs are comparable

15 to the results of newer studies (0.59 % and 0.26 % for urine and dung patches combined from cattle and sheep; Cai and

16 Akiyama, 2016; 1.18 % and 0.31 % for cattle urine and dung; Krol et al., 2016). The large difference of the EFs for urine and

17 dung also supports the suggestion of Krol et al. (2016) to disaggregate the EF by excreta type in emission inventories. The

18 implementation of excreta specific EFs could allow for a more precise calculation of the grazing related N_2O emissions e.g. as

19 dietary effects regarding the N intake predominantly affect the excreted urine N, which is the main source for the high N_2O

20 emission associated to excreta (Dijkstra et al., 2013).

21 The background emissions measured by FB cannot be attributed to a specific N input in a quantitative way, but the annual sum

22 of 1.03 kg N_2O-N ha^{-1} yr^{-1} from this study (extrapolated using Eq. 5 and VWC data for the whole year) compares well with

23 background emissions reported by a meta study of Kim et al. (2013, median: 0.7 and mean 1.52 kg N_2O-N ha^{-1} yr^{-1}) for

24 agricultural lands. In agricultural systems, background emissions are usually determined as emissions from (managed) plots

25 receiving no fertilisation in the study year. Thus they still include N inputs from plant residues and atmospheric deposition.

26 Background emissions are also often regarded as a late effect of fertilization events from previous years (Bouwman, 1996; Gu

27 et al., 2009). On pastures, background emissions may additionally result from trampling of the cows that can further stimulate

28 the N_2O production via denitrification due to soil compaction (Bhandral et al., 2007).

4.3 Up-scaling of FB fluxes

30 Urine patch emissions were parametrized with an exponential decay and maximum initial emissions of about 600 µg N_2O-N

31 m^{-2} h^{-1} that is close to the maximum averaged emissions measured by Barneze et al. (2015) from manually applied urine in

32 laboratory conditions and on a grassland. A strong emission response to urine application was generally reported in the

1 literature, however, with a large range of different emission dynamics and magnitudes (e.g. two emission peaks due to

2 nitrification and denitrification; emission peak after a few days with near exponential decay afterwards; significant emissions

3 after weeks to month; Bell et al., 2015; Cardenas et al., 2016; Chadwick et al., 2018). Similar to our study, reported dung

4 patch emissions by those studies were much lower compared to urine induced emissions.

5 We found that pasture emissions were dominated by excreta related emissions during the GOP (about 60 %). On a seasonal

6 basis, the up-scaled aggregated fluxes compared well with the gap filled EC measurements, which also indicates the validity

7 of the source attribution in the up-scaled emissions. Especially during time periods where both FB fluxes and EC fluxes were

8 measured (July – October) the agreement between the systems was very good.

9 However, the parameterisations used for up-scaling resulted in a poor performance for certain soil conditions. The limited

10 sensitivity towards changes in VWC of the background fluxes is probably due to the fact that FB measurements were mainly

11 performed during dry soil conditions. We have no explanation why we did not find a significant sensitivity of the background

12 fluxes towards changes in Ts as reported by other studies (Butterbach-Bahl et al., 2013; Schindlbacher, 2004). Typically,

13 increasing soil temperature leads to increased soil respiration which subsequently can lead to a depletion of soil oxygen and

14 further to higher denitrification rates. In contrast to background fluxes, the urine patch emissions showed a clear response to

15 changes in Ts and VWC. This effect could be parametrised with a bi-linear regression (Eqs. 7 and 8). This regression led to

16 high up-scaled emissions from urine patches especially during wet soil conditions and subsequently to an overestimation of

17 the cumulative emissions in May and June compared to the EC systems. N_2O emissions often have an emission maximum

18 during moderately wet soil conditions (VWC between 0.40 and 0.45) while completely anaerobic conditions at saturated VWC

19 can lead to a complete denitrification with only marginal N_2O emissions (Butterbach-Bahl et al., 2013). Such conditions have

20 been very rare during the FB measurements (see Fig. 9) and therefore may not be adequately represented in the derived

21 parameterisation. A general trend towards lower emissions during very wet soil conditions was also observed by the EC

22 systems (not shown). However, in order to avoid mixing results of the different measurement systems and thus reducing the

23 explanatory power of the system inter-comparison we decided to base the environmental regression analysis (Sect. 3.2.3) only

24 on measured data by the FB.

26 **4.4 Advantages and problems of experimental setup**

27 The presented field campaign was designed to estimate the N_2O emissions of two parallel grazing systems and to compare

28 different feeding diets of the herds. Field scale emissions derived by the EC method resulted in a wide range of measured

29 emissions which were mainly driven by environmental and management related parameters. Nevertheless, the setup with two

30 towers allowed for a good comparison with a sufficient number of measured fluxes from both systems. Due to a delayed

31 installation of the EC tower at system G all fluxes prior mid of April had to be gap filled which resulted in a higher associated

32 uncertainty.

33 The excreta N input derived by the animal budget approach at a temporal resolution of 1 day was needed in order to quantify

34 the EF of the two systems and to up-scale FB chamber measurements to the field scale. Nevertheless, direct measurements

would have been preferable. However, as the N content in the excreta is highly variable (Betteridge et al., 2013) on a seasonal (e.g. due to variability in the N content of the fodder) and short term scale (e.g. different urine volume, different cows, difference between day and night) continuous measurements throughout the grazing period for a representative number of cows would have been needed. This is only possible with measurement equipment directly placed on the cow. Beside the still considerable uncertainty associated to these measurements, they are often limited regarding animal welfare and are not well established (Misselbrook et al., 2016). Thus, they were not used in this study.

The combined approach of EC and FB measurements allowed the quantification of the uncertainty of the up-scaling routine and the good match between the two measurement approaches also validates the resulting contributions of the different emission sources on the field scale. The uncertainty associated with the up-scaling mainly resulted from missing FB measurements during wet soil conditions (e.g. in spring), which prevented the use of a more complex parameterisation of environmental driver effects on background and urine emission. In summary, the experimental setup resulted in robust field scale emissions, allowed to compare the two pasture systems, and yielded source specific emission factors for dung and urine patches.

5 Concluding remarks

The temporal dynamics of background areas and excreta patches were observed by fast-box (FB) chamber measurements on the pasture. We found no significant temporal pattern of the background fluxes. Urine patch emissions were parametrised by an exponential decay with time whereas a less pronounced dependency on excreta age of dung emissions was observed. This relation was parametrised with a quadratic function and a maximum after about 10 days. On a field scale level, urine patch emissions dominated the pasture emissions during the grazing season. Nevertheless, background fluxes contributed significantly to the pasture emissions as well. The origin of these background fluxes is still uncertain and should be addressed in further studies.

The combined approach with EC and FB measurements proved to be appropriate to observe and quantify the magnitude of the pasture emissions and to calculate the contribution of the single emission sources. The different diet of the cows resulted in a excreta related N_2O emission difference of about 25 % between the two cow herds and revealed the large potential of an N optimised feeding strategy to reduce grazing related N_2O emissions. In this study, the N optimisation was achieved by feeding additional maize silage to the fodder in system M. However, a reduction in excreted N can potentially be realised by other means as well (e.g. grass varieties with a high content of water soluble carbohydrates). The excreta related EFs derived from the up-scaled FB measurements were 0.74 ± 0.26 % for system M and 0.83 ± 0.29 % for system G and were thus significantly lower compared to the current default EF of 2 % for cattle excreta provided by the guidelines of the IPCC. The findings also exhibited clear differences in the individual EFs for urine and dung (1.12 ± 0.43 % and 0.16 ± 0.06 %, respectively, averaged over system M and G) suggesting a corresponding disaggregation in emission inventories.

Data availability. Data obtained in this study will be online available at the time of publication from the data repository zenodo.org. (the DOI will be included in the final paper version).

Competing interests. The authors declare that they have no conflict of interest.

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References

Aarons, S. R., Gourley, C. J. P., Powell, J. M. and Hannah, M. C.: Estimating nitrogen excretion and deposition by lactating cows in grazed dairy systems, Soil Res., 55(6), 489, doi:10.1071/SR17033, 2017.

Ammann, C., Brunner, A., Spirig, C. and Neftel, A.: Technical note: Water vapour concentration and flux measurements with PTR-MS, Atmos Chem Phys, 9, 2006.

Ammann, C., Flechard, C. R., Leifeld, J., Neftel, A. and Fuhrer, J.: The carbon budget of newly established temperate grassland depends on management intensity, Agric. Ecosyst. Environ., 121(1–2), 5–20, doi:10.1016/j.agee.2006.12.002, 2007.

Ammann, C., Spirig, C., Leifeld, J. and Neftel, A.: Assessment of the nitrogen and carbon budget of two managed temperate grassland fields, Agric. Ecosyst. Environ., 133(3–4), 150–162, doi:10.1016/j.agee.2009.05.006, 2009.

Arriaga, H., Salcedo, G., Calsamiglia, S. and Merino, P.: Effect of diet manipulation in dairy cow N balance and nitrogen oxides emissions from grasslands in northern Spain, Agric. Ecosyst. Environ., 135(1–2), 132–139, doi:10.1016/j.agee.2009.09.007, 2010.

Barneze, A. S., Minet, E. P., Cerri, C. C. and Misselbrook, T.: The effect of nitrification inhibitors on nitrous oxide emissions from cattle urine depositions to grassland under summer conditions in the UK, Chemosphere, 119, 122–129, doi:10.1016/j.chemosphere.2014.06.002, 2015.

Bates, G., Quin, B. and Bishop, P.: Low-cost detection and treatment of fresh cow urine patches, in Moving farm systems to improved attenuation. (Eds L.D. Currie and L.L Burkitt), vol. 28, p. 12, Palmerston North, New Zealand. [online] Available from: http://flrc.massey.ac.nz/workshops/15/paperlist15.htm, 2015.

1 Bell, M. J., Rees, R. M., Cloy, J. M., Topp, C. F. E., Bagnall, A. and Chadwick, D. R.: Nitrous oxide emissions from cattle
2 excreta applied to a Scottish grassland: Effects of soil and climatic conditions and a nitrification inhibitor, Sci. Total Environ.,
3 508, 343–353, doi:10.1016/j.scitotenv.2014.12.008, 2015.

4 Betteridge, K., Costall, D. A., Li, F. Y., Luo, D. and Ganesh, S.: Why we need to know what and where cows are urinating –
5 a urine sensor to improve nitrogen models, Proc N. Z. Grassl. Assoc, 75, 119–124, 2013.

6 Bhandral, R., Saggar, S., Bolan, N. and Hedley, M.: Transformation of nitrogen and nitrous oxide emission from grassland
7 soils as affected by compaction, Soil Tillage Res., 94(2), 482–492, doi:10.1016/j.still.2006.10.006, 2007.

8 Biermann, T., Babel, W., Ma, W., Chen, X., Thiem, E., Ma, Y. and Foken, T.: Turbulent flux observations and modelling over
9 a shallow lake and a wet grassland in the Nam Co basin, Tibetan Plateau, Theor. Appl. Climatol., 116(1–2), 301–316,
10 doi:10.1007/s00704-013-0953-6, 2014.

11 Bouwman, A. F.: Direct emission of nitrous oxide from agricultural soils, Nutr. Cycl. Agroecosystems, 46(1), 53–70,
12 doi:10.1007/BF00210224, 1996.

13 Bracher, A., Schlegel, P., Münger, A., Stoll, W. and Menzi, H.: Möglichkeiten zur Reduktion von Ammoniakemissionen durch
14 Fütterungsmassnahmen beim Rindvieh (Milchkuh), SHL Agroscope Zollikofen Posieux, 2011.

15 Butterbach-Bahl, K., Baggs, E. M., Dannenmann, M., Kiese, R. and Zechmeister-Boltenstern, S.: Nitrous oxide emissions
16 from soils: how well do we understand the processes and their controls?, Philos. Trans. R. Soc. B Biol. Sci., 368(1621),
17 20130122–20130122, doi:10.1098/rstb.2013.0122, 2013.

18 Cai, Y. and Akiyama, H.: Nitrogen loss factors of nitrogen trace gas emissions and leaching from excreta patches in grassland
19 ecosystems: A summary of available data, Sci. Total Environ., 572, 185–195, doi:10.1016/j.scitotenv.2016.07.222, 2016.

20 Cardenas, L. M., Thorman, R., Ashlee, N., Butler, M., Chadwick, D., Chambers, B., Cuttle, S., Donovan, N., Kingston, H. and
21 Lane, S.: Quantifying annual N2O emission fluxes from grazed grassland under a range of inorganic fertiliser nitrogen inputs,
22 Agric. Ecosyst. Environ., 136(3–4), 218–226, doi:10.1016/j.agee.2009.12.006, 2010.

23 Cardenas, L. M., Misselbrook, T. M., Hodgson, C., Donovan, N., Gilhespy, S., Smith, K. A., Dhanoa, M. S. and Chadwick,
24 D.: Effect of the application of cattle urine with or without the nitrification inhibitor DCD, and dung on greenhouse gas
25 emissions from a UK grassland soil, Agric. Ecosyst. Environ., 235, 229–241, doi:10.1016/j.agee.2016.10.025, 2016.

26 Chadwick, D. R., Cardenas, L. M., Dhanoa, M. S., Donovan, N., Misselbrook, T., Williams, J. R., Thorman, R. E., McGeough,
27 K. L., Watson, C. J., Bell, M., Anthony, S. G. and Rees, R. M.: The contribution of cattle urine and dung to nitrous oxide
28 emissions: Quantification of country specific emission factors and implications for national inventories, Sci. Total Environ.,
29 635, 607–617, doi:10.1016/j.scitotenv.2018.04.152, 2018.

30 Cowan, N. J., Norman, P., Famulari, D., Levy, P. E., Reay, D. S. and Skiba, U. M.: Spatial variability and hotspots of soil N2O
31 fluxes from intensively grazed grassland, Biogeosciences, 12(5), 1585–1596, doi:10.5194/bg-12-1585-2015, 2015.

32 Dijkstra, J., Oenema, O., van Groenigen, J. W., Spek, J. W., van Vuuren, A. M. and Bannink, A.: Diet effects on urine
33 composition of cattle and N2O emissions, animal, 7(s2), 292–302, doi:10.1017/S1751731113000578, 2013.

34 Felber, R.: Bridging the gap between animal and ecosystem emissions: Performance of CH4 and CO2 eddy covariance
35 measurements over a grazed pasture, ETH Zurich., 2015.

1 Felber, R., Münger, A., Neftel, A. and Ammann, C.: Eddy covariance methane flux measurements over a grazed pasture: effect
2 of cows as moving point sources, Biogeosciences, 12(12), 3925–3940, doi:10.5194/bg-12-3925-2015, 2015.

3 Felber, R., Bretscher, D., Münger, A., Neftel, A. and Ammann, C.: Determination of the carbon budget of a pasture: effect of
4 system boundaries and flux uncertainties, Biogeosciences, 13(10), 2959–2969, doi:10.5194/bg-13-2959-2016, 2016.

5 Flechard, C. R., Ambus, P., Skiba, U., Rees, R. M., Hensen, A., van Amstel, A., Dasselaar, A. van den P., Soussana, J.-F.,
6 Jones, M., Clifton-Brown, J., Raschi, A., Horvath, L., Neftel, A., Jocher, M., Ammann, C., Leifeld, J., Fuhrer, J., Calanca, P.,
7 Thalman, E., Pilegaard, K., Di Marco, C., Campbell, C., Nemitz, E., Hargreaves, K. J., Levy, P. E., Ball, B. C., Jones, S. K.,
8 van de Bulk, W. C. M., Groot, T., Blom, M., Domingues, R., Kasper, G., Allard, V., Ceschia, E., Cellier, P., Laville, P.,
9 Henault, C., Bizouard, F., Abdalla, M., Williams, M., Baronti, S., Berretti, F. and Grosz, B.: Effects of climate and management
10 intensity on nitrous oxide emissions in grassland systems across Europe, Agric. Ecosyst. Environ., 121(1–2), 135–152,
11 doi:10.1016/j.agee.2006.12.024, 2007.

12 Flesch, T. K. and Wilson, J. D.: Estimating Tracer Emissions with a Backward Lagrangian Stochastic Technique, in Agronomy
13 Monograph, edited by J. L. Hatfield and J. M. Baker, American Society of Agronomy, Crop Science Society of America, and
14 Soil Science Society of America., 2005.

15 Flesch, T. K., Wilson, J. D., Harper, L. A., Crenna, B. P. and Sharpe, R. R.: Deducing Ground-to-Air Emissions from Observed
16 Trace Gas Concentrations: A Field Trial, J. Appl. Meteorol., 43(3), 487–502, doi:10.1175/1520-
17 0450(2004)043<0487:DGEFOT>2.0.CO;2, 2004.

18 Foken, T., Leuning, R., Oncley, S. R., Mauder, M. and Aubinet, M.: Corrections and Data Quality Control, in Eddy Covariance,
19 edited by M. Aubinet, T. Vesala, and D. Papale, pp. 85–131, Springer Netherlands, Dordrecht., 2012.

20 Fuchs, K., Hörtnagl, L., Buchmann, N., Eugster, W., Snow, V. and Merbold, L.: Management matters: Testing a mitigation
21 strategy for nitrous oxide emissions on intensively managed grassland, Biogeosciences Discuss., 1–43, doi:10.5194/bg-2018-
22 192, 2018.

23 Gu, J., Zheng, X. and Zhang, W.: Background nitrous oxide emissions from croplands in China in the year 2000, Plant Soil,
24 320(1–2), 307–320, doi:10.1007/s11104-009-9896-1, 2009.

25 Häni, C.: bLSmodelR – An atmospheric dispersion model in R. [online] Available from: http://www.agrammon.ch/
26 documents-to-download/blsmodelr/ (Accessed 24 October 2017), 2017.

27 Häni, C., Flechard, C., Neftel, A., Sintermann, J. and Kupper, T.: Accounting for Field-Scale Dry Deposition in Backward
28 Lagrangian Stochastic Dispersion Modelling of NH$_3$ Emissions, , doi:10.20944/preprints201803.0026.v1, 2018.

29 Hensen, A., Groot, T. T., van den Bulk, W. C. M., Vermeulen, A. T., Olesen, J. E. and Schelde, K.: Dairy farm CH4 and N2O
30 emissions, from one square metre to the full farm scale, Agric. Ecosyst. Environ., 112(2–3), 146–152,
31 doi:10.1016/j.agee.2005.08.014, 2006.

32 IPCC: 2006 IPCC Guidelines for National Greenhouse Gas Inventories, Prepared by the National Greenhouse Gas
33 Inventories Programme, Eggleston H.S., Buendia L., Miwa K., Ngara T. and Tanabe K. (eds). Published: IGES, Japan., 2006.

34 IPCC, 2014: Climate Change 2014: Synthesis Report . Contribution of Working Groups I, II and III to the Fifth
35 Assessment Report of the Intergovernmental Panel on Climate Change [Core Writing Team, R.K. Pachauri and L.A. Meyer
36 (eds.)], IPCC, Geneva, Switzerland., 2014.

1 Jones, S. K., Famulari, D., Di Marco, C. F., Nemitz, E., Skiba, U. M., Rees, R. M. and Sutton, M. A.: Nitrous oxide emissions
2 from managed grassland: a comparison of eddy covariance and static chamber measurements, Atmospheric Meas. Tech., 4(10),
3 2179–2194, doi:10.5194/amt-4-2179-2011, 2011.

4 Kaimal, J. C. and Finnigan, J. J.: Atmospheric boundary layer flows: their structure and measurement, Oxford University Press,
5 New York., 1994.

6 Kim, D.-G., Giltrap, D. and Hernandez-Ramirez, G.: Background nitrous oxide emissions in agricultural and natural lands: a
7 meta-analysis, Plant Soil, 373(1–2), 17–30, doi:10.1007/s11104-013-1762-5, 2013.

8 Krol, D. J., Carolan, R., Minet, E., McGeough, K. L., Watson, C. J., Forrestal, P. J., Lanigan, G. J. and Richards, K. G.:
9 Improving and disaggregating N 2 O emission factors for ruminant excreta on temperate pasture soils, Sci. Total Environ.,
10 568, 327–338, doi:10.1016/j.scitotenv.2016.06.016, 2016.

11 Langford, B., Acton, W., Ammann, C., Valach, A. and Nemitz, E.: Eddy-covariance data with low signal-to-noise ratio: time-
12 lag determination, uncertainties and limit of detection, Atmospheric Meas. Tech., 8(10), 4197–4213, doi:10.5194/amt-8-4197-
13 2015, 2015.

14 Luo, J., Wyatt, J., van der Weerden, T. J., Thomas, S. M., de Klein, C. A. M., Li, Y., Rollo, M., Lindsey, S., Ledgard, S. F.,
15 Li, J., Ding, W., Qin, S., Zhang, N., Bolan, N., Kirkham, M. B., Bai, Z., Ma, L., Zhang, X., Wang, H., Liu, H. and Rys, G.:
16 Potential Hotspot Areas of Nitrous Oxide Emissions From Grazed Pastoral Dairy Farm Systems, in Advances in Agronomy,
17 vol. 145, pp. 205–268, Elsevier., 2017.

18 MeteoSwiss: Climate   normals   Fribourg/Posieux,   [online]   Available   from:
19 www.meteoschweiz.admin.ch/product/output/climate-data/climate-diagrams-normal-values-station-
20 processing/GRA/climsheet_GRA_np8110_e.pdf (Accessed 31 January 2018), 2018.

21 Mishurov, M. and Kiely, G.: Gap-filling techniques for the annual sums of nitrous oxide fluxes, Agric. For. Meteorol., 151(12),
22 1763–1767, doi:10.1016/j.agrformet.2011.07.014, 2011.

23 Misselbrook, T., Del Prado, A. and Chadwick, D.: Opportunities for reducing environmental emissions from forage-based
24 dairy farms, Agric. Food Sci., 22(1), 93–107, doi:10.23986/afsci.6702, 2013.

25 Misselbrook, T., Fleming, H., Camp, V., Umstatter, C., Duthie, C.-A., Nicoll, L. and Waterhouse, T.: Automated monitoring
26 of urination events from grazing cattle, Agric. Ecosyst. Environ., 230, 191–198, doi:10.1016/j.agee.2016.06.006, 2016.

27 Orr, R. J., Griffith, B. A., Champion, R. A. and Cook, J. E.: Defaecation and urination behaviour in beef cattle grazing semi-
28 natural grassland, Appl. Anim. Behav. Sci., 139(1–2), 18–25, doi:10.1016/j.applanim.2012.03.013, 2012.

29 Oudshoorn, F. W., Kristensen, T. and Nadimi, E. S.: Dairy cow defecation and urination frequency and spatial distribution in
30 relation to time-limited grazing, Livest. Sci., 113(1), 62–73, doi:10.1016/j.livsci.2007.02.021, 2008.

31 Papale, D.: Data Gap Filling, in Eddy Covariance, edited by M. Aubinet and T. Vesala, pp. 159–172, Springer Netherlands,
32 Dordrecht., 2012.

33 Pedersen, A. R., Petersen, S. O. and Schelde, K.: A comprehensive approach to soil-atmosphere trace-gas flux estimation with
34 static chambers, Eur. J. Soil Sci., 61(6), 888–902, doi:10.1111/j.1365-2389.2010.01291.x, 2010.

35 Portmann, R. W., Daniel, J. S. and Ravishankara, A. R.: Stratospheric ozone depletion due to nitrous oxide: influences of other
36 gases, Philos. Trans. R. Soc. B Biol. Sci., 367(1593), 1256–1264, doi:10.1098/rstb.2011.0377, 2012.

1 R Core Team: R: A Language and Environment for Statistical Computing, R Foundation for Statistical Computing, Vienna,
2 Austria. [online] Available from: https://www.R-project.org/, 2016.

3 Reay, D. S., Davidson, E. A., Smith, K. A., Smith, P., Melillo, J. M., Dentener, F. and Crutzen, P. J.: Global agriculture and
4 nitrous oxide emissions, Nat. Clim. Change, 2(6), 410–416, doi:10.1038/nclimate1458, 2012.

5 Saggar, S., Giltrap, D. L., Davison, R., Gibson, R., de Klein, C. A., Rollo, M., Ettema, P. and Rys, G.: Estimating direct N2O
6 emissions from sheep, beef, and deer grazed pastures in New Zealand hill country: accounting for the effect of land slope on
7 the N2O emission factors from urine and dung, Agric. Ecosyst. Environ., 205, 70–78, doi:10.1016/j.agee.2015.03.005, 2015.

8 Schindlbacher, A.: Effects of soil moisture and temperature on NO, NO_2, and N_2O emissions from European forest soils, J.
9 Geophys. Res., 109(D17), doi:10.1029/2004JD004590, 2004.

10 Schmid, H. P.: Footprint modeling for vegetation atmosphere exchange studies: a review and perspective, Agric. For.
11 Meteorol., 113(1–4), 159–183, doi:10.1016/S0168-1923(02)00107-7, 2002.

12 Selbie, D. R., Buckthought, L. E. and Shepherd, M. A.: The Challenge of the Urine Patch for Managing Nitrogen in Grazed
13 Pasture Systems, in Advances in Agronomy, vol. 129, pp. 229–292, Elsevier., 2015.

14 Villettaz Robichaud, M., de Passillé, A. M., Pellerin, D. and Rushen, J.: When and where do dairy cows defecate and urinate?,
15 J. Dairy Sci., 94(10), 4889–4896, doi:10.3168/jds.2010-4028, 2011.

16 Voglmeier, K., Jocher, M., Häni, C. and Ammann, C.: Ammonia emission measurements of an intensively grazed pasture,
17 Biogeosciences Discuss., 1–32, doi:10.5194/bg-2018-86, 2018.

18 Wilson, J. D., Flesch, T. K. and Crenna, B. P.: Estimating Surface-Air Gas Fluxes by Inverse Dispersion Using a Backward
19 Lagrangian Stochastic Trajectory Model, in Geophysical Monograph Series, edited by J. Lin, D. Brunner, C. Gerbig, A. Stohl,
20 A. Luhar, and P. Webley, pp. 149–162, American Geophysical Union, Washington, D. C., 2013.

21 Yan, T., Frost, J. P., Agnew, R. E., Binnie, R. C. and Mayne, C. S.: Relationships among manure nitrogen output and dietary
22 and animal factors in lactating dairy cows, J. Dairy Sci., 89(10), 3981–3991, 2006.

23 Zhao, X. and Huang, Y.: A Comparison of Three Gap Filling Techniques for Eddy Covariance Net Carbon Fluxes in Short
24 Vegetation Ecosystems, Adv. Meteorol., 2015, 1–12, doi:10.1155/2015/260580, 2015.

1 **Table 1: Near-surface soil parameters (5-10 cm depth) averaged over four locations on the pasture. The measurements are given as**
2 **mean ± 1 standard deviation.**

Parameter	Value
Pore volume (%)	57 ± 4
Bulk density (g cm^{-3})	1.09 ± 0.11
pH (-)	6.0 ± 0.3
Sand (%)	42.6 ± 2.5
Clay (%)	18.7 ± 1.7
Silt (%)	33.0 ± 1.3
Soil organic matter (%)	5.7 ± 0.3
Total N (%)[#]	0.38 ± 0.03
Total C (%)[#]	3.76 ± 0.20

3 [#]were measured at a depth of 0-10 cm

Table 2: Measured averages ± standard deviation of observed cow properties (ECM: energy corrected milk) and feed protein contents used by the dairy cow nitrogen budget approach for both pasture systems during the grazing season 2016.

Input parameter (units)	System M	System G
Number of cows	12	12
Milk yield, ECM (kg cow^{-1} day^{-1})	25.1 ± 2.9	24.2 ± 3.7
Animal weight (kg)	633 ± 14	633 ± 10
Grass crude protein (g kg-DM^{-1})	195 ± 23	196 ± 23
Maize crude protein (g kg-DM^{-1})	84 ± 8	n.a.

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1 **Table 3: Flux measurements using the FB technique (mean ± std) of background and excreta patches averaged over 20 days following**
2 **a grazing phase.**

Measurement location	System M	System G
Background (μg N_2O-N m^{-2} h^{-1})	8 ± 8	5 ± 8
Urine (μg N_2O-N m^{-2} h^{-1})	121 ± 130	162 ± 190
Dung (μg N_2O-N m^{-2} h^{-1})	16 ± 18	35 ± 60

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Table 4: Coefficients (a-c), corresponding indices i and significance levels for the equations presented in Sect. 3.2. The equation coefficients were fitted using FB chamber measurements and yield fluxes in units of µg N_2O-N m^{-2} h^{-1}. The input quantities are the soil temperature T_S (in units of °C), time since end of grazing Δt_{EOG} (in units of days) and volumetric water content VWC (as dimensionless fraction).

Equation	I	a_i	b_i	c_i
Eq. 3	1	587 ***	-0.082 **	
Eq. 4	2	23 *	5.4 *	-0.25 *
Eq. 5	3	12.6 ***	0.267 ***	0.012 *
Eq. 7	4	-1490 ***	2900 ***	23.9 **
Eq. 8	5	0.098 ***	-0.086 **	

***Significant at level $p < 0.001$; **Significant at level $p < 0.01$; *Significant at level $p < 0.05$

1 **Table 5: Summary of cumulated grazing related emissions for both pasture systems during the GOP 2016. The table shows the**
2 **emissions per area (first part), pasture area and excreta N input (second part) and the emissions per cow and grazing hour as well**
3 **as the calculated EFs (third part). The uncertainties are given as 1σ.**

Parameter	System M	System G
EC emission (kg N_2O-N ha^{-1})	1.49 ± 0.21	1.49 ± 0.27
FB emissions up-scaled to EC footprint (kg N_2O-N ha^{-1})	1.51 ± 0.39	1.50 ± 0.37
FB emissions up-scaled to pasture system (kg N_2O-N ha^{-1}) [a]	1.48 ± 0.38	1.48 ± 0.37
Pasture system area (ha)	1.88	2.51
Excreta N total (g N cow^{-1} h^{-1})	16.5 ± 1.2	19.6 ± 1.5
Urine N (g N cow^{-1} h^{-1})	10.2 ± 1.1	13.0 ± 1.3
Dung N (g N cow^{-1} h^{-1})	6.3 ± 0.7	6.5 ± 0.7
FB excreta emissions (g N_2O-N cow^{-1} h^{-1})	0.12 ± 0.04	0.16 ± 0.05
EF excreta total (%) [b]	0.74 ± 0.26	0.83 ± 0.29
EF urine (%) [b]	1.09 ± 0.43	1.15 ± 0.43
EF dung (%) [b]	0.16 ± 0.06	0.17 ± 0.06

4 [a] average emissions over all paddocks

5 [b] based on FB emissions up-scaled to pasture system

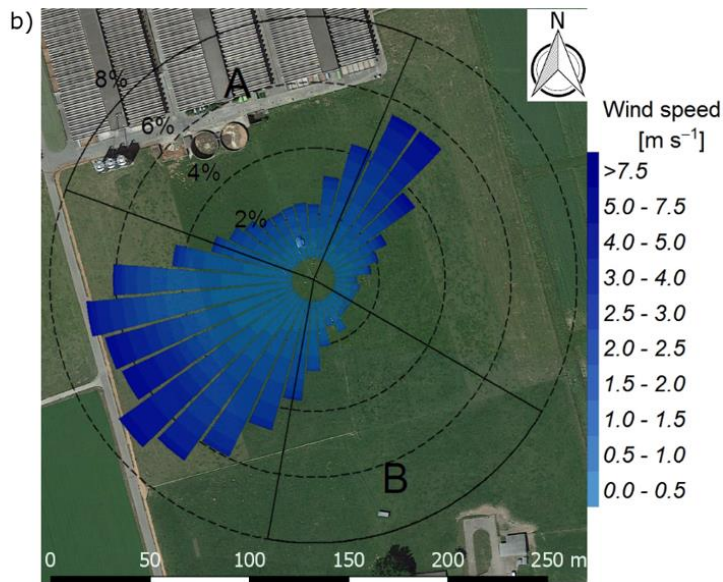
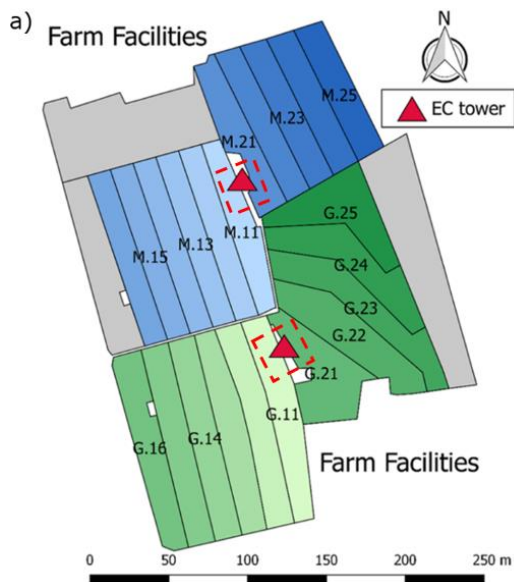
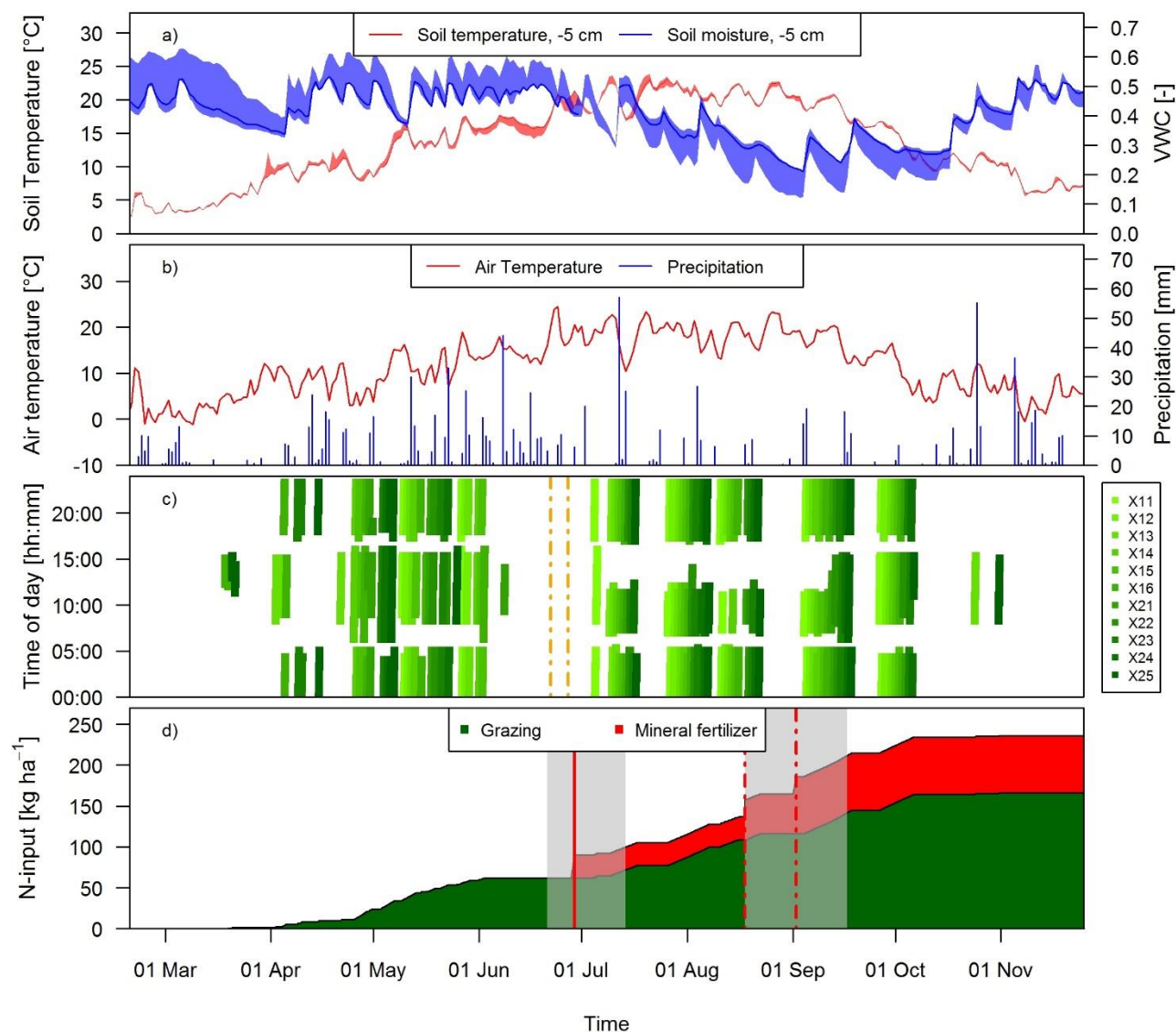
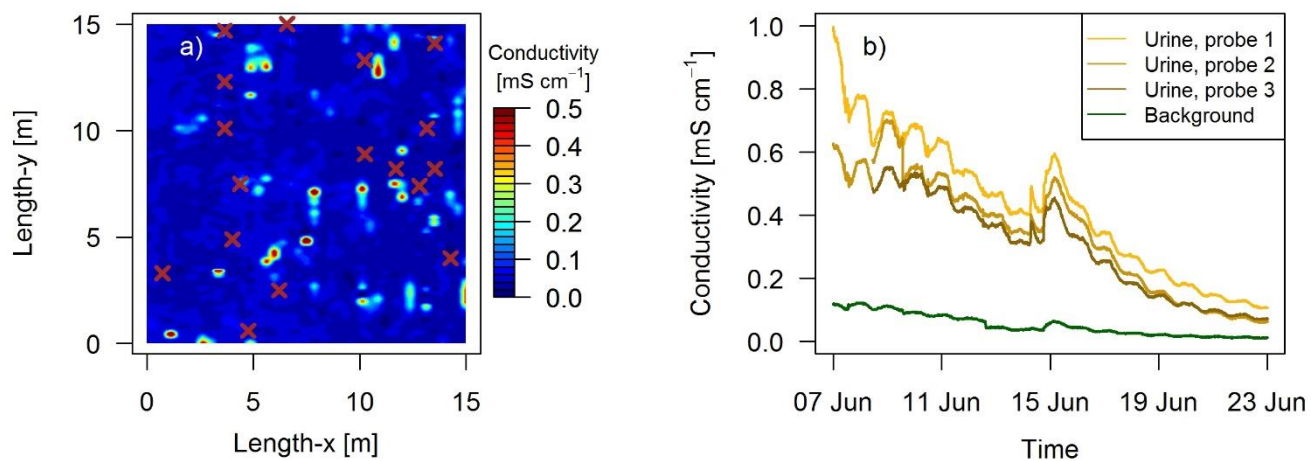
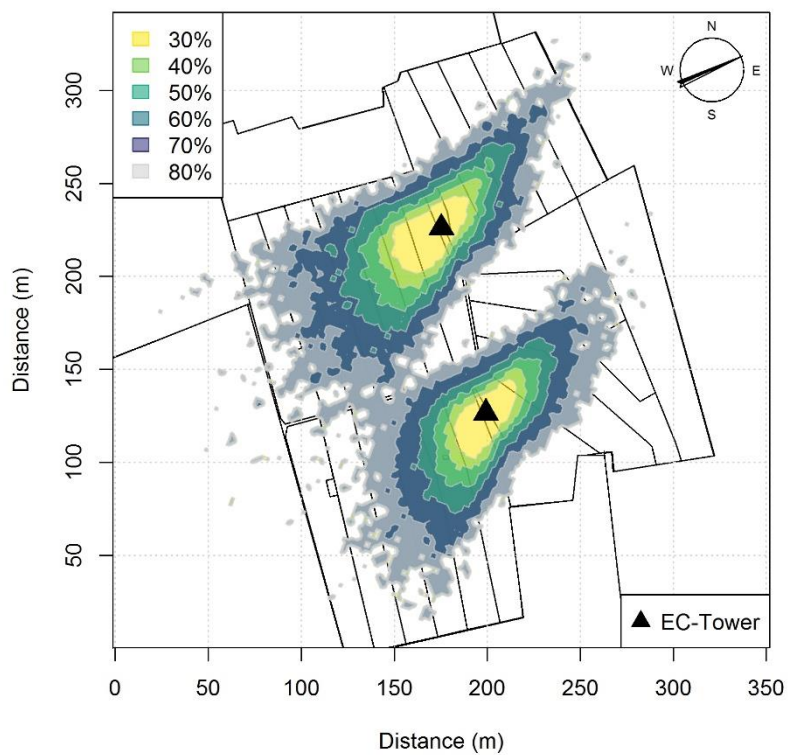



Figure 1: a): Measurement site with the pastures for the two herds (blue: grass diet with additional maize silage; green: full grazing regime; grey: optional pasture areas) and the division into the paddocks (M.11–M.25, G.11–G.25). Additionally the location of the two EC towers (triangles) and the area of the chamber measurements (red dashed rectangles) are shown. b) Wind distribution for the northern sonic anemometer with the corresponding sector contributions (black dotted circles) for the period May – October 2016. The areas A and B indicate wind sectors from which advection from nearby farm building can occur. The wind distribution was overlaid on a Google Earth image of the experimental area (Map data: Google, DigitalGlobe)

2 **Figure 2: Time series of a) daily averaged soil temperature and moisture at a depth of 5 cm measured at system M (solid lines) and**
3 **spread of the four measurement locations, b) daily air temperature at 2 m above ground and precipitation at the measurement site,**
4 **c) grazing duration on the single paddocks of the pasture (X: both pasture systems M and G) for the study year 2016. The dashed**
5 **vertical orange lines indicate the harvest event (split between X.11-X.16 and X.21-X.25) d) N input to system M during the main**
6 **grazing season in 2016. Fertilizer was applied two times (vertical red lines). The second application (dashed lines) in August was**
7 **split in two parts due to concurrent rotational grazing. The grey shaded areas indicate time periods influenced by fertilization or**
8 **harvest events as explained in Sect. 2.5.2.**

3 **Figure 3: a) Measured conductivity within a quadratic 15 x 15 m intensive observation area on the 3rd of October, 2016 in system G.**
4 **High values (>0.15 mS cm^{-1}) indicate urine patch locations and brown crosses indicate observed dung pats. b) Conductivity measured**
5 **continuously during a field experiment in 2017 with four GS3 sensors.**

2 **Figure 4: Footprint climatology for both EC towers averaged for the time period between 15th March 2016 and 15th November 2016.**
3 **The legend values indicate the percentage of the total footprint weight.**

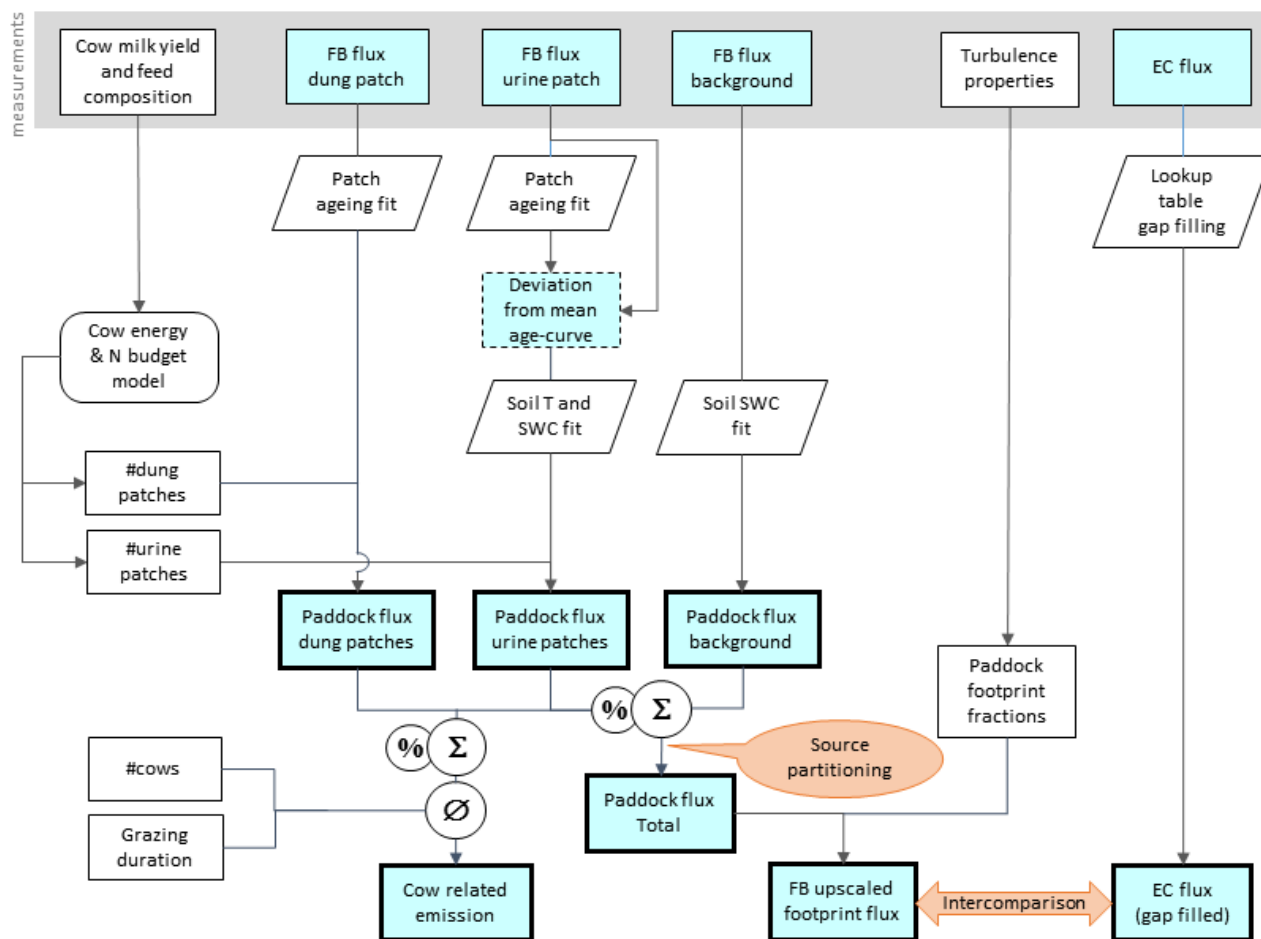
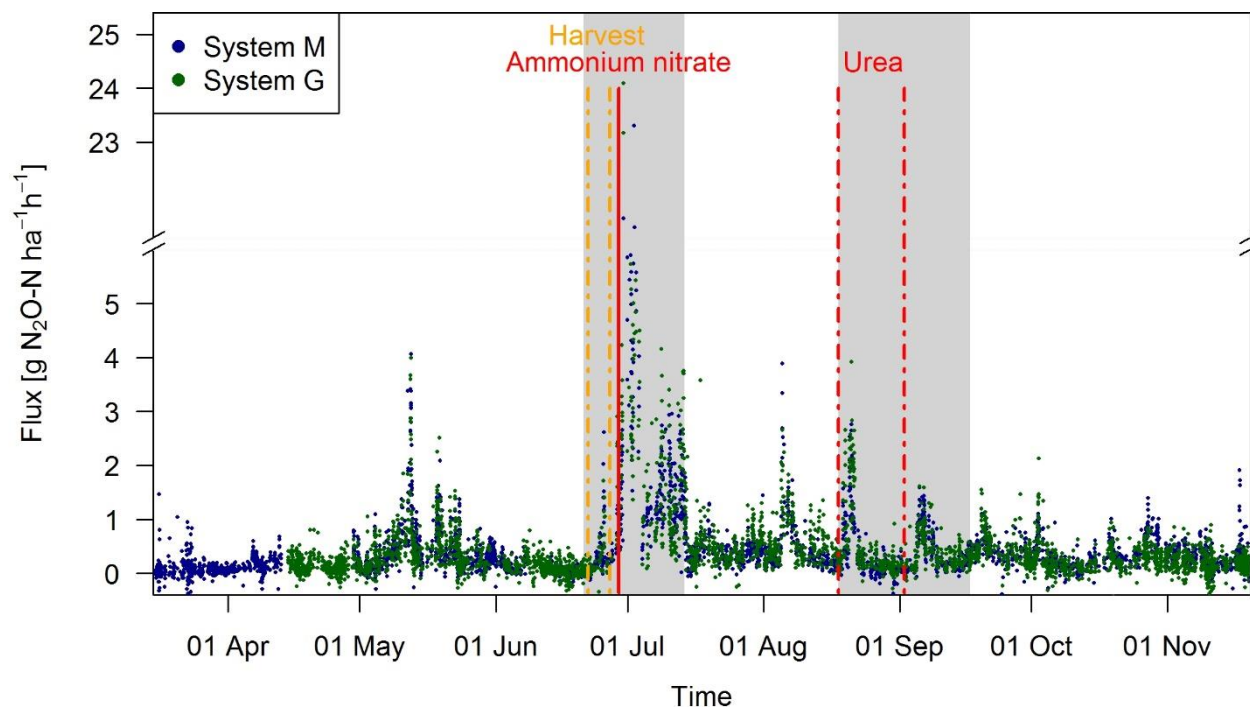
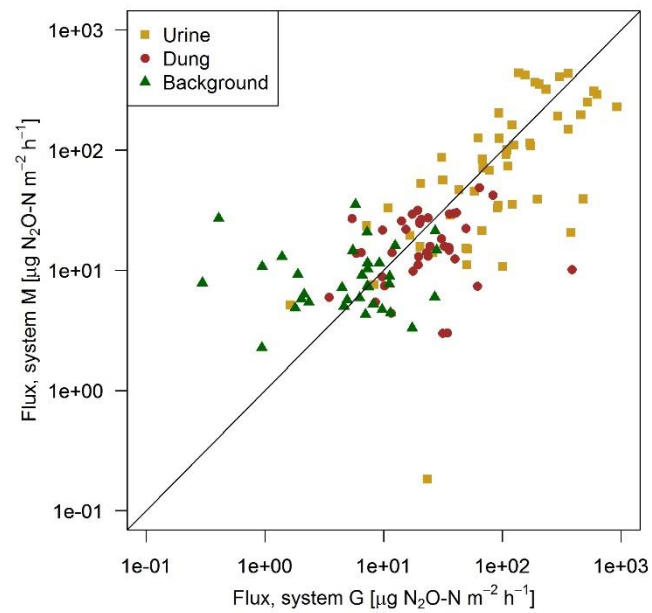
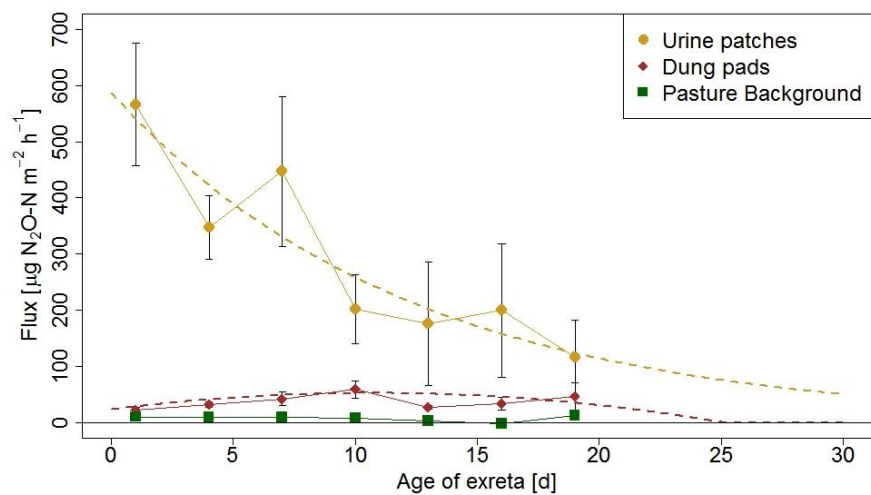
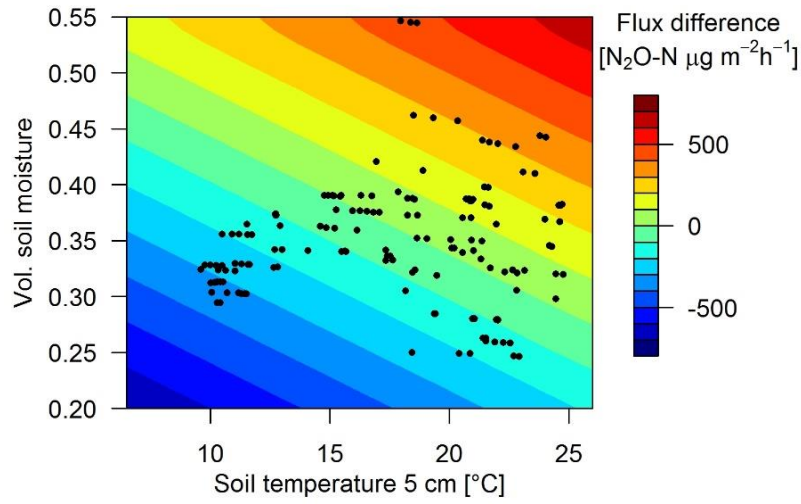



Figure 5: Flowchart of up-scaling procedure to compare small scale chamber fluxes with EC fluxes and to estimate the contribution of excreta emissions to the overall pasture emission. Rectangular shapes indicate data sets / time series data. Time series data with thin frames have gaps whereas bold frames indicate complete data sets. The light blue colour specifies N$_2$O flux data. Other shapes show operations (e.g. fit or gap-filling routines).

2 **Figure 6: Time series of half-hourly EC flux measurements in both systems during the grazing season 2016. For the analysis of**
3 **grazing related emissions, only the non-shaded periods (GOP) were used. The vertical lines show the timings of fertilization (red)**
4 **and harvest (orange) events. Dashed lines indicate that harvest and urea application were split for western (X.11-X.16) and eastern**
5 **(X.21-X.25) part. The shaded areas indicating time periods influenced by fertilization events or harvest were excluded for the**
6 **evaluation of grazing excreta related emissions. One flux value (28.7 g N_2O-N ha^{-1} h^{-1} on system M, 30.06.2016) was skipped for**
7 **better readability.**

2 **Figure 7: Scatterplot shows the comparison of near-simultaneous fluxes for different sources measured with the fast-box on the two**
3 **pasture systems. The black line indicates the 1:1 line.**

2 **Figure 8: N₂O flux evolution with time for urine patches, dung pats and background areas. The fluxes were measured with the fast-**
3 **box and averaged over 3-day periods and the error bars show the standard error of the measurements. The standard errors for the**
4 **background fluxes are smaller than the symbols. The dotted lines show the fitted curves through the averaged values of urine and**
5 **dung patch emissions (see also Eqs. 3 and 4).**

2 **Figure 9: Surface plot shows the estimated N₂O flux deviation (Eq. 6, 7; *Corr_{U,env}* = 0) from the exponential fit (Eq. 3) for urine**
3 **patches depending on soil moisture *VWC_U* and temperature at a depth of 5 cm. The black dots indicate the conditions under which**
4 **flux measurements with the FB were obtained.**

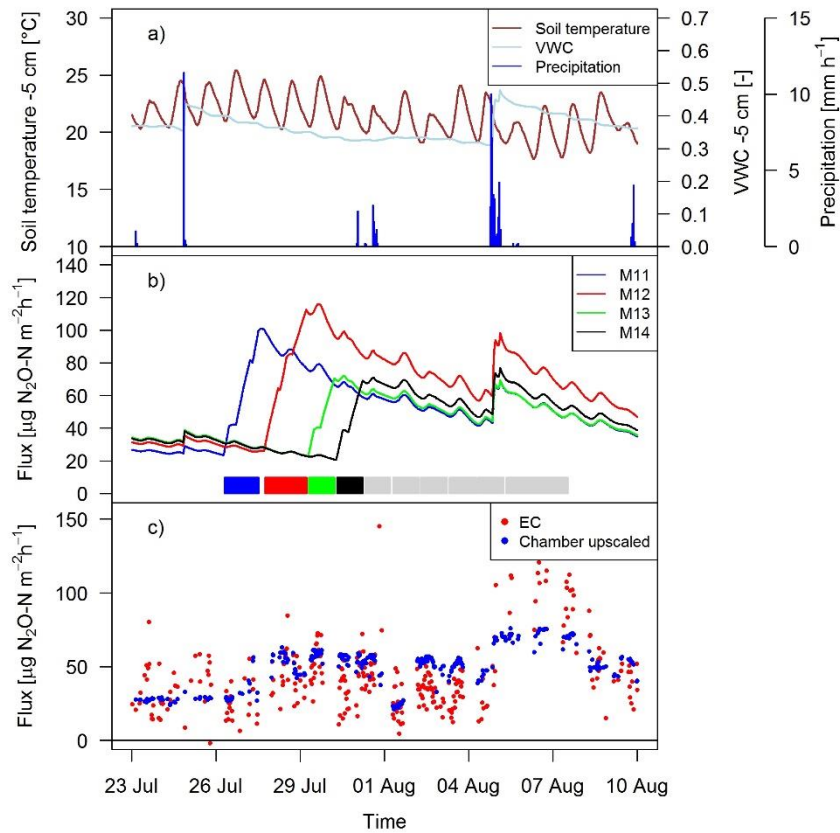



Figure 10: Time series of a) environmental parameters and b) up-scaled FB fluxes (Sect. 2.7) for different paddocks (M11-M14) in system M. The coloured rectangles at the bottom show the grazing phases on the four considered paddocks (grey colours indicating grazing on the remaining paddocks). c) N$_2$O fluxes by EC and up-scaled FB during a full rotation between 23th September and 10th August 2016 for system M.

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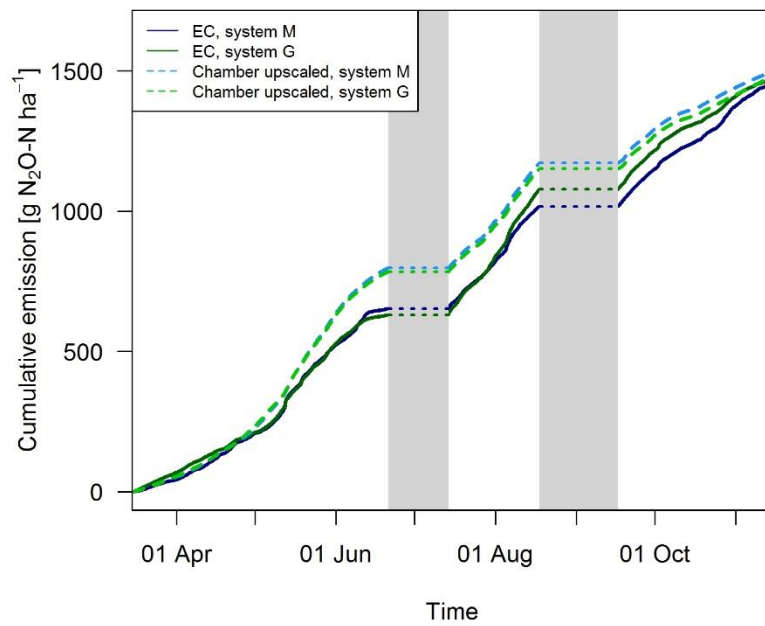
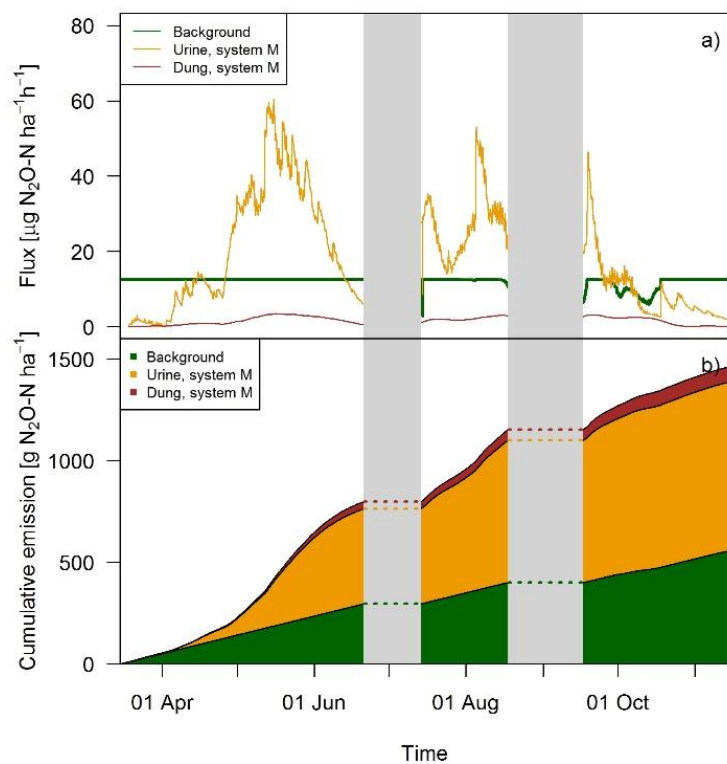

2 **Figure 11: a) Cumulative emissions for both systems obtained with FB and EC technique during GOP 2016. The grey shaded bars**
3 **indicate time periods which were excluded due to significant overlapping N₂O emissions from fertilization / harvest and grazing**
4 **(Sect. 2.5, 3.1).**

2 **Figure 12: a) Time series of up-scaled FB fluxes averaged over all paddocks of system M for all three emission sources during the**
3 **grazing season 2016, and b) retrieved cumulative emission contribution of the emission sources to the overall field emission.**