

RESPONSE TO REVIEWERS on “Particulate trace metal dynamics in response to increased CO₂ and iron availability in a coastal mesocosm experiment” by M. Rosario Lorenzo et al.

Anonymous Referee #1

1)... Generally, the topic of the change of particle trace metal during the marine environment changing such as Ocean acidification (OA) and different Fe availability is very interesting for ocean biogeochemists. I believe the data set is valuable in this field. I feel that, however, authors need to regard more about how they can present their data set to induce conclusion above, which they claimed in conclusion section in this paper. The present contents of this manuscript are not well organized for presenting their data set to conclude the claimed conclusions.

General comments: In construction of this manuscript, "results" section is not constructed only by result, and "discussion" section is not well explained by this study's result (data) (“Discussions” are only like a review of previous knowledge). I recommend that authors should re-construct and re-organized whole part of the manuscript. "Results" section should be used some "Figures" for presenting their data. It makes more easily to understand for readers. "Discussion" section should be related more to data from this study, including which data induce which conclusion more clearly.

In view of both referee comments we have substantially change the Ms attending to their comments. Abstract has been modified, as well as the results and discussion sections. We would like to pinpoint that specifically the Discussion section has been fully re-structured and re-discussed. We believe it is more focused now in order to get a good conclusion. Specific answers to the comments raised follow below.

2)The effect of CO₂ did not follow a clear trend in this study, as authors mentioned in the text. The effect of controlled Fe availability by DFB addition/non-addition to phytoplankton bloom is also not clear. How authors induced these their claimed results is not clearly understandable for readers.

This has been modified accordingly in 2.1 Experimental set-up section, Lns 119-124: “*To induce changes in Fe availability, and analyse its effects on the plankton community, 70 nM (final concentration) of the siderophore desferrioxamine B (DFB) (+DFB and –DFB treatments) (Figure 1b) was added to half of the mesocosms on Day 7, when the community was already acclimated to high CO₂. The initial dFe concentration before DFB addition was about 4.5 nM. Even though DFB is a strong Fe-binding organic ligand often used to induce iron limitation in phytoplankton (Wells 1999), DFB additions may also increase the dissolved Fe pool in environments with high concentrations of colloidal and/or particulate Fe, such as fjords (Kuma et al. 1996, Öztürk et al. 2002). By day 17, dissolved iron concentrations were significantly higher (by ~3-fold) in the high CO₂ and DFB treatments than in the control (Segovia et al. 2017). These results support an increase in the solubility of Fe in seawater by either lowering its pH (Millero 1998; Millero et al. 2009) and/or the addition of DFB (Chen et al. 2004).*”

We have added Figure 1, comprising 3 panels referring to CO₂ concentration, dFe and dCu concentrations to better understand the chemical scenario that lead the experiment to the obtained results.

3) For discussion OA influence, I think authors should focus on to show “How particle trace metal concentrations and its ratio changed by CO₂ concentrations” by more well presented their own data set.

We have changed the tables by bar plots (now figures 3 and 4) according to reviewer’s suggestions and we think it is much clearer now. We have re-written section 3.3-The effects of increased CO₂ and the DFB addition on particulate metal concentrations. Lns 233-242. *“Increased CO₂ and the DFB addition did not significantly affect the concentrations of particulate Al, Ti, Cu, and Pb (Tables 1 and S2). Similarly, the addition of DFB did not directly influence particulate concentrations of Fe, but high CO₂ had a significant negative impact on particulate Fe (Tables 1 and S2, Figure 3). Particulate Cd concentrations were also inversely affected by CO₂, but only in the presence of DFB (CO₂; and CO₂ x DFB effect, Tables 1 and S2, Figure 3). All other elements (P, Co, Zn, Mn and Mo) exhibited significant effects by CO₂ and by DFB, but there was also a significant interaction between these two factors (Table 1, S2). This indicates that, for example, particulate Mn, Zn, Mo, Co, and P concentrations were significantly decreased by high CO₂, but only in the +DFB treatments (Figure 3, Table 1, S2,). Similarly, the addition of DFB significantly increased pZn and pMn, but only at ambient CO₂ levels (Figure 3, Table 1, S2)”*

Data are now supported by new Fig 3, Table 1 and Suppl.Table S2.

4) For Fe availability, they need to discuss that “did DFB addition influence positive/negative to Fe availability?”. It depends on natural dissolved Fe concentrations.

See answer to point 2 above, and point 5 below.

5) Additionally, authors should show more clearly about relationship between Fe availability and *E. huxleyi* bloom response, with figure etc. It is very difficult for readers to understand the relation only from the “Tables” number.

We have substituted the Table by a new figure 2 with permission from Segovia et al. Mar. Ecol. Prog. Ser. 2017 for better understanding. This figure shows the temporal development of chlorophyll a ($\mu\text{g L}^{-1}$) and phytoplankton biomass ($\mu\text{g C L}^{-1}$) in the mesocosms exposed to different CO₂ and dissolved iron (dFe) treatments. (a) Chlorophyll a, (b) *Emiliania huxleyi* (5–10 μm), (c) *Synechococcus* (0.6–2 μm), (d) picoeukaryotes (0.1–2 μm), (e) small nanoeukaryotes (prasinophytes, small haptophytes, 2–7 μm), (f) large nanoeukaryotes (small single-celled diatoms and flagellated forms, 6–20 μm), (g) diatoms (chain-forming *Skeletonema* sp. 20–> 500 μm), (h) dinoflagellates (20–200 μm).

This is also discussed in a new 4.1 Section: *“The effects of CO₂ and dFe in the plankton community*

*In this experiment we investigated changes in particulate trace metal concentrations, in response to increased CO₂ and/or an addition of the siderophore DFB in a coastal mesocosm experiment. For a better understanding of the processes affecting these stressors, we briefly summarise the mesocosm experiment results originating from Segovia et al. (2017). High CO₂, as well as the DFB addition increased dFe concentration. The higher dFe concentrations were sustained in the DFB treatments. A bloom of the coccolithophore *Emiliana huxleyi* was observed in the ambient CO₂ treatments, and was especially massive in the one with the addition of DFB (LC+DFB). On the contrary, the biomass of *E. huxleyi* was negatively affected by increased CO₂. However, increased dFe partially mitigated the negative effect of elevated CO₂, indicating that the coccolithophore was able to acclimate better to ocean acidification when Fe availability was high. High dFe also had a positive effect on the cyanobacterium *Synechococcus sp.*, while the rest of the plankton food web did not respond to the treatments (Segovia et al. 2017)*

6) DFB addition inducing more dissolved fraction of TM is artificial response. This is different story from Fe bio-availability. Important for bio-availability is how much free Fe exist under each condition. As one of author well know that DFB-Fe uptake by phytoplankton need very complex mechanisms. Authors should discuss more detail about this part.

This was fully discussed in Segovia et al. 2017 and it is not the aim of this paper which is focused in pMe. However, we suggest to consult Segovia et .2017 for further details. See Lns 423-427. *“Under control conditions at present CO₂ concentration (LC, 380 μatm) and no DFB amendment, the globally important coccolithophore *Emiliana huxleyi* was experiencing Fe limitation (Segovia et al. 2017). The shift between particulate and dissolved Fe promoted a massive bloom of *E. huxleyi* in the treatments with ambient CO₂ due to increased Fe bioavailability (for further details on Fe-bioavailability in *E. huxleyi* please see Segovia et a. 2017)”*.

7)Line 116-117:, 171 Table 2 etc.: Authors described that “The biological and chemical variables analysed were phytoplankton abundance and species composition, dissolved Fe and Cu, nutrient concentration, and particle trace metals concentration”. They only show these data in Tables. Figures which present time variation are easier for readers to understand the data variation during the experiments. Please prepare Figures. I can imagine the particle trace metal data was only collected on d12, d17, and d21. But for grasp biological response and chemical environment change, sampling should be done more frequently. If authors have more frequent data for nutrient, cell number for *E. huxleyi*, etc., it should be plotted to the Figures. Authors described time changing of “diatom” with nutrient concentrations. Authors should make a plot of “day since day0” vs. “diatom cell number”, vs. “pigment”, vs. “*E. huxlei*” and vs. “nutrient concentrations” in each mesocosm. It is helpful for reader.

It is indeed done. We have included Chla and phytoplankton biomass in the new Fig.2. Dissolved Fe and Cu in Fig. 1, and Nitrate, Ammonium, Silicate and Phosphate in new Supplemental Figure S1. Additionally reader are encouraged to consult Segovia et al. 2017.

There is no point of re-publishing already published results. Only those that are really essential

8)Line 188-189, 3.2: Authors described “The only metal that showed a time-dependent decrease in its particulate concentrations was Fe.” “dMetals”, “pMetals”. These also should be appeared by figures. 176: Authors described “An increase in dFe was observed in all treatments between day 7 and 17.” In Table 2, there are no data from d7, it should be appeared in the Table 2. And d17 LC-DFB data is decrease. So “all treatments” in this sentence is not correct.

This is already solved by the new figures and the explanation in results section. In addition we have included Supplemental Figure S2 showing the Fe partition coefficients (the molar ratio between particulate and dissolved concentrations) in the different mesocosm treatments; LC: ambient CO₂ (390 µatm); HC: increased CO₂ (900 µatm); -DFB: no DFB addition; +DFB: with a 70 nM DFB addition; on day 12 and day.

9)Line 195-196: Authors indicate that “high CO₂ had negative impact on particulate Fe” and “Cd concentrations were also inversely affected by CO₂”. These parts are difficult to understand which data indicate this fact. Line 197-198: Authors described “All other elements (P, Co.these two factors was clear (Table 5)”. Please explain how clear as like Cd description in previous sentence. Only showing Table 5 is not enough explanation to reader.

We have rewritten 3.3-The effects of increased CO₂ and the DFB addition on particulate metal concentrations as follows: *“Increased CO₂ and the DFB addition did not significantly affect the concentrations of particulate Al, Ti, Cu, and Pb (Tables 1 and S2). Similarly, the addition of DFB did not directly influence particulate concentrations of Fe, but high CO₂ had a significant negative impact on particulate Fe (Tables 1 and S2, Figure 3). Particulate Cd concentrations were also inversely affected by CO₂, but only in the presence of DFB (CO₂; and CO₂ x DFB effect, Tables 1 and S2, Figure 3). All other elements (P, Co, Zn, Mn and Mo) exhibited significant effects by CO₂ and by DFB, but there was also a significant interaction between these two factors (Table 1, S2). This indicates that, for example, particulate Mn, Zn, Mo, Co, and P concentrations were significantly decreased by high CO₂, but only in the +DFB treatments (Figure 3, Table 1, S2,). Similarly, the addition of DFB significantly increased pZn and pMn, but only at ambient CO₂ levels (Figure 3, Table 1, S2)”*

We believe is much clearer now. Note that Table 5 has been substituted by figure 3 to make it easier and that Table 1 and S2 also support these data.

10)For “3. Results” section, all subtitle is not well organized. Some contents can be compiled to one (For example, 3.1 and 3.2 can merge for “biological chemical response in mesocosms”. And particle trace metal variation in different treatment in 3.2, 3.3, and 3.4 can merge to one section.

Some of he sub-headings have reorganised. We have accepted the suggestion of “3.1 biological chemical response in mesocosms”. However, we have not merged the following sections as we think it will be rather confusing to the reader.

11) Title and contents of subsection in 3.6 and 3.7 are part of “discussion”.

These results parts have been re-structured and part of the text has been moved to Discussion section.

12) Discussion Line 247-248: “Our results demonstrate that in the studied fjord, particulate Ti and Fe concentrations were dominated by lithogenic material.”. Authors need explanation how they judged this. The explanation is appeared in section 3.6 result (actually this is discussion). Please indicate clearly “this data is shown in Figure 1”. Line 248-250: “In contrast, particulate Cu, Co, Mn, Zn, Mo and Cd concentrations were correlated with P concentrations, as well as phytoplankton biomass, suggesting strong biogenic influence on their distribution (Table 6)”. Authors need explanation how they judged this. The explanation is appeared in section 3.7 result (actually this is discussion). Please indicate clearly “this data is shown in Figure 1”. Only showing Table 6 is not kind for reader. This part is overlapped to 4.2 section. It should be in to 4.2 section with detailed explanations with Tables and figures.

This one of the parts that have changed most. This section is supported by Fig 5 and Table 4. Thus, we have fully restructured and re-discussed 4.2 Particulate Fe and Ti are associated with lithogenic sources, while particulate Co, Cu, Zn, Cd, Mo and Mn are associated with biogenic sources, as follows, Lns 265-326:

“The particulate trace metal concentrations (nM, mean of all treatments and dates) during the experiment were highest for Al, Fe and Zn, and lowest for Co and Cd, following this trend: Al ≈ Fe ≈ Zn > Ti > Cu ≈ Mn > Mo ≈ Pb > Co > Cd. Lithogenic particles are enriched in Al and low in P (average crustal Al and P content is 2.9 mmol Al and 0.034 mmol P g⁻¹ dry weight, Taylor 1964), while biogenic particles are enriched in P and low in Al (average plankton Al and P content is 0.001 mmol Al and 0.26 mmol P g⁻¹ dry weight, Bruland et al. 1991). Therefore, the distinct high abundance of Al and P in lithogenic and biogenic particles, respectively, can be used to evaluate the relative contribution of lithogenic and biogenic material in our particulate samples. In order to do this, first, it is important to establish that the vast majority of the measured particulate P is associated the biogenic fraction. In this study, the abiotic P was estimated using particulate Al concentrations (nM) and the P:Al ratio in crustal material, and was calculated to be negligible (< 1% of the total measured particulate P). In addition, a significant correlation (p < 0.003) was found between particulate P concentrations and phytoplankton biomass (Table 4). Therefore, we assume a constant trace metal composition in biogenic particles (assuming they are rich in phytoplankton) and lithogenic particles (assuming they are rich in crustal material). We then calculated the expected metal concentrations in the particulate samples assuming that all the P measured in the particles is associated with a biogenic fraction, and that all Al in the particles is associated with the lithogenic fraction. Thus, for a given trace metal, its expected particulate trace metal concentration in seawater (mol L⁻¹) can be calculated as the sum of the contribution from biogenic and lithogenic particles, so that:

$$[Me] = a [P] + b [Al]$$

where $[Me]$ is the total concentration of the metal (mol L^{-1}) expected in the particulate sample; $[P]$ is the P concentration measured in the particles (mol L^{-1}); $[Al]$ is the Al concentration measured in the particles (nM L^{-1}); a is the average, well-known metal content in biogenic particles, normalized to P (i.e. mol Me: mol P in marine plankton) and b is the average, well known metal content in lithogenic particles, normalized to Al (mol Me: mol Al in the Earth crust). For example, on day 21 in the HC-DFB treatment, the concentrations of particulate Al and P were 8.22 and 134.8 nM, respectively (Table S2). Assuming a constant 0.0051 mol Fe: mol P in biogenic particles and 0.331 mol Fe: mol Al in lithogenic particles (Table 2), we calculated an expected particulate Fe concentration of 3.41 nM, where 80% was associated with lithogenic material and 20% with biogenic material. Similar calculations were made for the bioactive metals Mn, Co, Cu, Zn, Cd, and Mo (Table 5). Our calculations indicate that on average, particulate Fe was dominated by the lithogenic component (accounting for an average of 78% of the total expected particulate Fe), while for particulate Co, Cu, Zn, Cd, and Mo the biogenic fraction dominated (accounting for 94, 95, 99, 94 and 98%, respectively, of the total expected concentration; Table 5). Particulate concentrations of Mn were also dominated by the biogenic fraction (65% of the total), but the lithogenic fraction was also significant (35%). Moreover, the expected particulate Mn and Fe concentrations closely matched the particulate Mn and Fe concentration we measured (accounting for an average of $\sim 71\%$ of the measured Mn, and 115% of the measured Fe). For other metals (i.e. Cu, Mo and Zn), the expected particulate concentrations (nM) were lower than measured (23% of the measured pCu, and 8% of measured pZn; Table 5). This suggests that the particles were enriched in Cu, Mo, and Zn relative to what is expected based on natural marine plankton metal quotas (Bruland et al. 1991) and crustal ratios (Taylor 1964).

To further establish the lithogenic or biogenic source of the pMe in the particles, the particulate metal concentrations were normalized to the concentrations of particulate P and Al (Figure 4, and Table 2). These ratios were then compared with well-known molar ratios of metal to Al in the crust (Taylor 1964) and of metal to P ratios in marine plankton samples (Ho 2006) and cultures (Ho et al. 2003) (Table 2). The average Fe: Al (506 mmol Fe: mol Al) and Ti:Al ratios (119 mmol Ti: mol Al, Table 2) were relatively similar to crustal molar ratios (331 mmol Fe: mol Al and 39 mmol Ti: mol Al; Taylor 1964). Additional evidence for the significant lithogenic component in particulate Ti and Fe was gathered from Figure 5, where we plotted the molar ratios of the metals relative to P in the collected particles against the Al:P ratios measured in those same particles. The slope of these data $[(\text{Fe:P})/(\text{Al:P}) = \text{mol Me: mol Al}]$ is the ratio of Me:Al in the particles and can be compared to well-known Me:Al crustal ratio. Visually, if the data nicely fit the Me:Al line for crustal material, these metals are mainly associated with the lithogenic component, as evident for Fe and Ti (Figure 5). These combined results suggest that in our experiment, particulate Fe and Ti concentrations were enriched by lithogenic material. In support of this finding, we also found no significant correlation between particulate Fe and Ti concentrations (nM) and either the total plankton (phytoplankton and microzooplankton) or *E. huxleyi* biomass ($\mu\text{g C L}^{-1}$; Table 4).

In contrast, when the P-normalized metal ratios in the particles collected from the mesocosms were plotted against the Al:P ratios in these particles, there were no correlations for the following metals Co, Cu, Zn, Cd, Mn and Mo (Figure 5), indicating

that these particulate metals were not enriched in lithogenic material. Our measured metal: P ratios were comparable to plankton ratios in natural samples and in cultures (Table 5). The concentrations (mol L⁻¹) of these metals (i.e. Cu, Co, Zn, Cd, Mn, Mo), as well as P, also showed significant correlations with the biomass (µgC L⁻¹) of E. huxleyi and that of total plankton cells (p < 0.05, Table 4), supporting a significant influence of the phytoplankton in the distribution of these particulate elements.

13)Line 251-252: “Changes in CO₂ and/or Fe levels affected total particulate and biogenic metal concentrations for some metals.”. This part of results is not well presented in manuscript overall. Authors should regard to present some figures which can compare particle and biogenic metals concentrations among each treatment.

We have included Figure 3 and 4, and eliminated the corresponding tables that were unclear.

14)Line 255-263, 4.1 Efficacy of the oxalate-EDTA wash removing lithogenic trace metals from particles: First half part of this section is should be move to “results”. Especially from line 260-263, “In general, the concentrations of Fe and Co in the particles were decreased the least by the oxalate wash (by ~ 25%), while Mo and Pb concentrations were decreased the most (by ~70%). The concentrations of particulate Cu, Zn, Cd and Mn were reduced by 50% by the oxalate wash. As shown previously (Sanudo-Wilhelmy et al. 2004), the oxalate reagent also removed extracellular P (by ~20%).”.

This has been moved to material and methods and now reads as follows. Lns 176-195:

“2.3.3 The effect of oxalate-EDTA wash on particulate trace metal concentrations

To better estimate the biogenic fraction of the particulate metals, the filters were washed with an oxalate-EDTA solution, which removes extracellular metals and oxyhydroxides (Tovar-Sanchez et al., 2003; Tang and Morel, 2006). In our study, the oxalate wash significantly decreased the concentration of all particulate metals, with the exception of Al and Ti (Tables S2 and S3), as observed by Rauschenberg and Twining (2015). The quantity of metal remaining after the oxalate wash (i.e. biogenic fraction) varied among elements (Tables S2 and S3). In general, the concentrations of Fe and Co in the particles were decreased the least by the oxalate wash by ~ 25%, while Mo and Pb concentrations were decreased the most by ~70%. The concentrations of particulate Cu, Zn, Cd and Mn were reduced by 50% by the oxalate wash. As shown previously (Sanudo-Wilhelmy et al. 2004), the oxalate reagent also removed extracellular P (by ~20%, Table S2 & S3). Compared to Rauschenberg and Twining (2015), the estimates of the biogenic fraction, after the oxalate wash, were in agreement for Co, Cu and P, and lower for Fe, Mn, Zn and Cd concentrations.

However, the efficacy of the oxalate wash to dissolve Fe, and other metals, from lithogenic particles is not well constrained (Frew et al. 2006, Rauschenberg and Twining, 2015, King et al., 2012). Therefore, the results obtained after the oxalate-EDTA wash should be interpreted with caution because we do not know whether the removed metal fraction is a) only lithogenic; b) mainly lithogenic but some biogenic fraction is also removed, or c)

whether metals absorbed onto particles are equally labile to the wash on biogenic and lithogenic particles. Given that many of the trends we observed were identical for the oxalate-EDTA washed and non-washed particles [i.e. higher Me concentrations in the LC+DFB treatments (Table S2 & S3) and positive correlations between phytoplankton biomass and Me concentrations (Lorenzo-Garrido 2016)], below we present and discuss only the non-oxalate wash results”

So that, oxalate-wash is not further discussed. Yet, it is important to maintain this paragraph in the Ms.

15) Line 282:” Me:P ratios we measured in the particles are similar to those of natural phytoplankton assemblages (Ho, 2006) and of *Emiliana huxleyi* cultures (Ho et al., 2003).”. If authors want to compare their filed data to previous reported data by Ho, 2006, and Ho et al., 2003, authors should show the previous study’s number with their data on to Tables or Figures with citation. Otherwise, authors just state “similar” to natural plankton but did not show any evidence.

This has all been re-organised and re-discussed as required by both reviewers. Please see point 12 above.

16) Line 311-312, 319-320: “Interestingly, we also found a putative ZIP-transporter gene. ZIP-transporters are., such as tRNA synthetase, reverse transcriptase, metallo-carboxypeptidase, ABC-Zn-transporter and CDF-Zn-transporter..”. If authors want to say “we found”, they should show their data and discuss with using their data. If this “gene part” is part of other study, they should cite the other study appropriately. This discussion section is very strange for this aspect. It is written like author’s original data for this study.

We have removed the paragraph relating to referee comments. We fully agree with the comment.

17) Line 329- : Discussion on Cu:P should construct by using their data, what their data’s characteristics, what their data indicate, what is authors claim from the data, which previous knowledge supports their claims. This section 4.2 is like just a review of other papers.

We have re-written this part and better discussed our own data as follows, Lns 352-362:

*Similarly, the Cu:P ratios in the collected particles were relatively elevated (1.4 ± 0.8 mmol Cu: mol P) compared to those of other phytoplankton, including *E. huxleyi* (Table 2). The dissolved (7.7 ± 0.41 nM Cu, Figure 1) and particulate Cu concentrations (0.35 ± 0.25 nM, Table S2) in our experiment were high, and similar to those previously measured in this fjord (Muller et al., 2005). Rain events (or wet deposition of anthropogenic aerosols) in this fjord result in high dissolved Cu and the active production of strong organic ligands by *Synechococcus*—to lower the free Cu concentrations (Muller et al., 2005). Therefore, high Cu might be a general condition in this fjord, and indigenous plankton might have developed physiological mechanisms to deal with high Cu, such as the production of organic ligands to prevent uptake (Vraspir and Butler, 2009), or of heavy-metal-binding*

peptides (phytochelatin) to lower Cu toxicity inside the cell (Ahner and Morel, 1995; Ahner et al., 1995; Knauer et al., 1998). Since we measured high particulate Cu, and Cu:P in our experiment, E. huxleyi might have been relying mainly on phytochelatin to buffer high intracellular Cu (Ahner et al., 2002).

18) Line 344: “The Cd:P were significantly lower than those found in phytoplankton and E. huxleyi.”. Reader can not understand clearly which data they compared. Is this sentence mean that “The Cd:P were significantly lower than those found in individual phytoplankton and E. huxleyi which was reported by previous studies (Ho, 2006, Ho et al., 2003)”?. If so, they should show the comparable data from previous study.

We have clarified this in section 4.3 Particulate metals with a strong biogenic component: their P-normalized ratios, Lns 364-372 :” *The Cd:P ratios (average 0.024 ± 0.01 mmol Cd:mol P, Figure 4 and 6) were significantly lower than those in phytoplankton and E. huxleyi (0.36 mmol Cd:mol P, Figure 4 and 6). This was surprising, because Cd quotas are normally higher in coccolithophores than in diatoms and chlorophytes (Sunda and Huntsman, 2000; Ho et al., 2003). High Cd quotas in coccolithophores have been suggested to result from accidental uptake through Ca transporters and channels (Ho et al., 2009). The low Cd quotas here may be explained by the antagonistic interaction between Mn and Cd or Zn and Cd under high Mn and Zn, respectively (Sunda and Huntsman, 1998, 2000; Cullen and Sherrell, 2005). Since high Zn:P ratios were common in this study (34.02 ± 18.05 mmol Zn:mol P, Figure 4 and 6), we hypothesize that high Zn levels antagonistically interacted with Cd, resulting in low Cd:P ratios in the particles”*

19)Line 377: “The decrease in particulate Fe might have been due to enhanced solubility of Fe- oxides at low pH.”. The author should show scientific basis. They have to show relation between pFe and PH in each treatment.

It is now shown in Figure 1 and Figure S2 and discussed in 4.4 The effects of increased CO₂ and the DFB addition on particulate metal concentrations and P-normalized ratios, Lns 373-395:

“Iron enrichment is common in coastal waters, due to sediment resuspension, rivers input, aeolian deposition and mixing or upwelling of deep water. Indeed, Fe was the essential metal with the highest particulate concentrations in our study (Figure 3, Table 3). Furthermore, in this study particulate Fe was characterized by a strong lithogenic component, and was not correlated with phytoplankton biomass. Iron was also unique, in that it was the only trace element whose particulate concentration was significantly and uniquely affected by CO₂, regardless of the presence or absence of DFB (no interaction between CO₂ and DFB, Table 1). Furthermore, particulate Fe concentrations (nM) decreased steadily between days 12 and 21, with the exception of the control treatment (LC-DFB; Figure 3, Table 2S). This suggests that the increase in CO₂ and/or the DFB addition reduce the concentration of pFe, despite the phytoplankton bloom. Such a decrease in pFe (range 2.3-fold in LC-DFB, vs. 13.7-fold in HC+DFB; Table S2) might be mediated by the dissolution of particulate Fe by low pH or by the presence of strong organic chelators as observed in this very experiment (Segovia et al. 2017 and references therein). where dFe notably increased in treatments with high CO₂ and/or the addition of DFB (Figure 1). Furthermore, the dissolution of particulate Fe in the treatments with high

CO₂ and/or the addition of DFB was evident in the Fe partitioning coefficients—the molar ratio between particulate and dissolved concentrations. On day 21, the Fe partitioning coefficients varied by 22-fold between the highest for the control (LC-DFB: 1.039) and lowest for the HC+DFB treatments (HC+DFB: 0.047; Figure S2). Thus, either the DFB addition or high CO₂ promoted the dissolution of pFe. However, at the end of the experiment, high dFe concentrations were only observed in the treatments with the DFB additions, suggesting that the presence of strong organic Fe chelators, such as DFB, mediated the maintenance of high dissolved Fe concentrations, as previously observed (Segovia et al. 2017). Thus, in our future oceans, high CO₂ (low pH) will increase dissolved Fe concentrations in regions rich in particulate Fe, and in strong organic Fe chelators.”

20)Line 378: “the concentration of the elements P, Co, Zn, Mn and Mo were influenced by CO₂ and Fe levels”. Which data indicate those results? Authors should present with their dataset. Line 380-381: “where the addition of DFB resulted in higher dissolved Fe, and optimal pH enhanced E. huxleyi growth.”. Authors should present this relation, between dissolved Fe, pH and E. huxleyi growth, with figures which are constructed by their dataset.

This is now solved by inclusion of Fig 1, Fig 2 , by changing Tables to Figs 3 and 4 and answered in points 3,6,9 and 19.

21) conclusion remarks Please consider for my “general comment”. It is necessary to describe more specifically what was understood in each argument (claim) a)-d).Authors should present what are difference/similarity of their data among four mesocosms treatment more clearly, and what they can find from the difference/similarity? How they induced the conclusion of this study form the difference/similarity? This aspect is not clear overall in this manuscript.

We believe that this requirement is now met attending to the deep-structural changes we have done in the ms.

22) Others Authors used “pFe”, “particulate Fe”, “particulate iron”, “dFe”, and “dissolved Fe” in the text. They should use same words through the manuscript.

Changed accordingly

Anonymous Referee #2

I found it very hard to follow the description of the data in the Results, as most of them have been presented in tables, which is especially not good for presenting the time dependent changes in for example trace metal concentrations. I also found that the Discussion for the most part was on trace metal chemistry and physiology in general, but not specifically relative to the key objective of the study, i.e., the effects of CO₂ and Fe availability on particulate trace metal dynamics.

See point 1, 21 and others to Referee #1

Specific comments. Line 91. “(Hutchings, 2011)”, which was not included in the References.

Now included

Line 108. 10 uM nitrate: 0.3 uM PO₄ = 33:1 - was there a particular reason to use such a P limited nutrient condition? How may this affect the observed particulate trace metal concentration?

Yes, we used this specific ratio because we aimed at a *Emiliana huxleyi* bloom. This has been clarified as follows, Lns 117-119:

At the beginning of the experiment, nitrate (10 μM final concentration) and phosphate (0.3 μM final concentration) were added to induce a bloom of the coccolithophore Emiliana huxleyi, as recommended by Egge & Heimdal (1994).

Results: I would strongly suggest that the data should be presented as figures instead of tables. In addition, albeit statistical analyses were conducted and presented together in Table 5, I would suggest they should also be presented in each individual table (or figure, if the authors decide to follow my suggestion above in revising the manuscript).

Done

Lines 170-181. “days 1-10, phase I”, “day 7” and “After day 10” were mentioned when describing the data, but none of them can be found in Table 2. Line 221. “(Figure 5)” should be Table 5.

This has now changed with the new Ms organisation.

Discussion: Again the Discussion mostly did not center around the influence of acidification and/or Fe availability on trace metal dynamics, except for the last, very short section 4.4. I thus encourage the authors to considerably revise the Discussion, focusing on how the chemistry and utilization of trace metals were affected by changes in CO₂/pH and Fe levels and how these may be related to the proliferation of *Ehux* in the mesocosm.

This has now changed with the new Ms organisation. See comment to Referee 1.

We thank the reviewers for their comments and their time, and hope that our responses are satisfactory

Yours sincerely,

Maria Segovia & Maite Maldonado

Particulate trace metal dynamics in response to increased CO₂ and iron availability in a coastal mesocosm experiment

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Abstract. Rising concentrations of atmospheric carbon dioxide are causing ocean acidification and will influence marine processes and trace metal biogeochemistry. In June 2012, in Raunefjord (Bergen, Norway) we performed a mesocosm experiment, comprised of a fully factorial design of ambient and elevated pCO₂ and/or an addition of the siderophore desferrioxamine B (DFB). In addition, the macronutrient concentrations were manipulated to enhance a bloom of the coccolithophore *Emiliana huxleyi*. We report here the changes in particulate trace metal (pMe) concentrations during this experiment. Our results show that particulate Ti and Fe were dominated by lithogenic material while particulate Cu, Co, Mn, Zn, Mo and Cd had a strong biogenic component. Furthermore, significant correlations were found between particulate concentrations (mol L⁻¹) of Cu, Co, Zn, Cd, Mn, Mo, and P in seawater and phytoplankton biomass (µgC L⁻¹), supporting a significant influence of the bloom in the distribution of these particulate elements. The concentrations of these biogenic metals (mol L⁻¹) in the *E. huxleyi* bloom were ranked as: Zn > Cu ≈ Mn > Mo > Co > Cd. Changes in CO₂ and/or DFB affected total particulate concentrations (mol L⁻¹) and biogenic metal ratios (Me:P) for some metals. Variations in CO₂ had the most clear, and significant effect on particulate Fe concentrations (mol L⁻¹), decreasing its concentration under high CO₂. Similarly, high CO₂ decreased the Co, Zn and Mn: P ratios, while increased the Cu: P ratios. In contrast, the addition of DFB had no significant effect on any of the biogenic metal ratios, whilst high concentrations of dissolved Fe will only be maintained by the presence of strong organic ligands. Future predicted high CO₂ levels are expected to change the relative concentrations of particulate and dissolved metals, due to the differential effects of high CO₂ on trace metal solubility, speciation, adsorption and toxicity, as well as on the growth of different phytoplankton taxa, and their elemental trace metal composition. These processes will also be mediated by the presence of strong organic ligands in areas where particulate Fe inputs are important, since the effectiveness of some natural chelators such as siderophores, in dissolving Fe from oxyhydroxides and/or by enhancing the photoinduced redox cycle of Fe, will be increased. This study demonstrates the utility and robustness of combining trace metal analyses of particles in a controlled mesocosm experiment with manipulations of CO₂ and Fe concentrations using natural assemblages of marine phytoplankton in order to understand future ocean dynamics.

Key words: Global change, iron, CO₂, particulate trace metals, dissolved trace metals, mesocosms, *Emiliana huxleyi*, phytoplankton

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Eliminado: The importance of the combined impacts of elevated CO₂ and changes in trace metal availability on marine plankton remain largely unknown. A mesocosm experiment was performed to study changes in particulate trace metal concentrations during a bloom dominated by the coccolithophore *Emiliana huxleyi*. We employed a full-factorial experimental design, comprising all combinations of ambient and elevated pCO₂ and dissolved iron (dFe). Particulate metal concentrations (Fe, Cu, Zn, Co, Mn, Cd, Mo, Ti and Pb) were determined by high-resolution inductively coupled plasma mass spectrometry (HR-ICPMS). We examined biogenic and lithogenic sources of particulate metals, and their evolution during the experiment. Biogenic metal concentrations were estimated from bulk particle measurements by comparing phosphorus (P)-normalised quotas with published ratios, as well as concentrations of particulate trace metals in the presence and absence of an oxalate-EDTA wash. Our results demonstrate that particulate Ti and Fe concentrations were dominated by lithogenic material in the fjord. In contrast, particulate Cu, Co, Mn, Zn, Mo and Cd concentrations correlated with P concentrations and phytoplankton biomass, indicative of their strong biogenic character. Furthermore, ocean acidification changed the relative concentrations of particulate metals; a result mainly driven by the effects of ocean acidification on the growth of different phytoplankton phyla.

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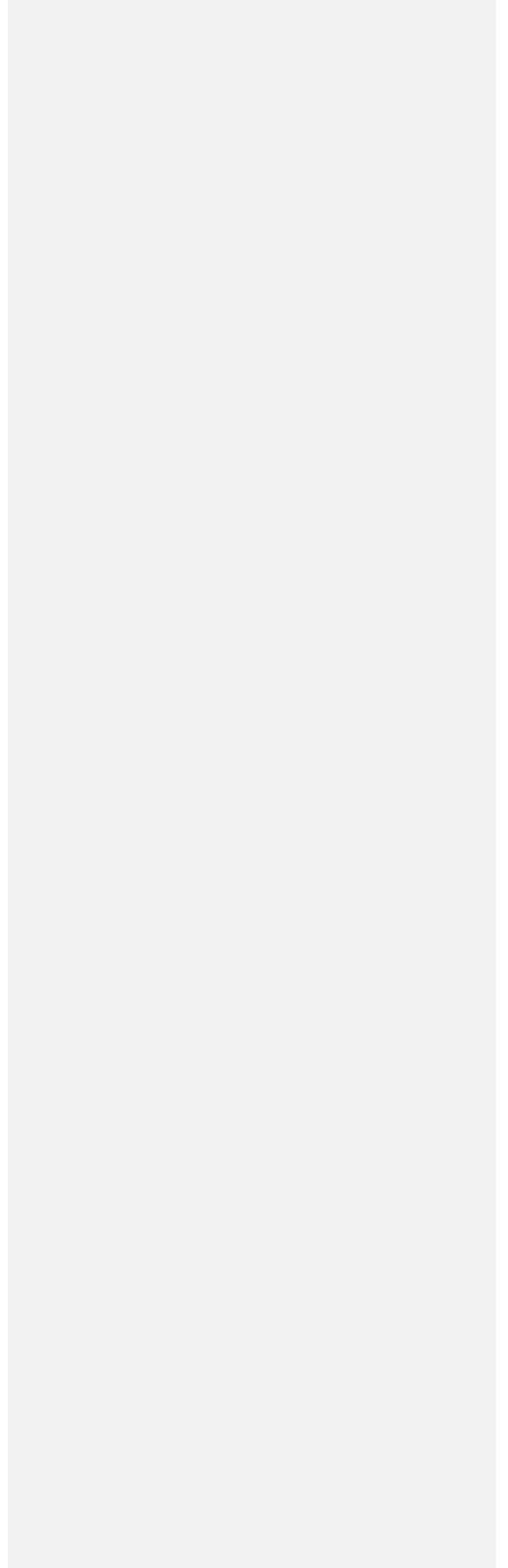
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72 **1. Introduction**

73 Marine **phytoplankton** contribute half of the world's total primary productivity, sustaining marine food webs and
74 driving the biogeochemical cycles of carbon and nutrients (Field et al., 1998). Annually, phytoplankton incorporate
75 approximately 45 to 50 billion metric tons of inorganic carbon (Field et al., 1998), removing a quarter of the CO₂
76 emitted to the atmosphere by anthropogenic activities (Canadell et al., 2007). Yet, the atmospheric CO₂ concentration
77 has increased by 40 % since pre-industrial times as a result of anthropogenic CO₂ emissions, producing rapid changes
78 in the global climate system (Stocker et al., 2013). The dissolution of anthropogenic CO₂ in seawater, causes shifts in
79 the carbonate chemical speciation, and leads to ocean acidification (OA). Marine ecosystems are sensitive to changes in
80 pH because pH strongly affects chemical and physiological reactions (Hoffman et al., 2012). Increased CO₂ in seawater
81 may enhance or diminish phytoplankton productivity (Mackey et al., 2015), decrease the CaCO₃ production in most
82 planktonic calcifiers (Riebesell and Tortell 2011), and/or inhibit organic nitrogen and phosphorus acquisition (Hutchins
83 et al., 2009). Thus, the biogeochemical cycling of nutrients is predicted to be highly affected by OA (Hutchins et al.
84 2009), **as well as** the distribution and speciation of trace metals in the ocean (Millero et al., 2009).

85 ▲
86 Trace metals, including Fe, Zn, Mn, Cu, Co and Mo, are essential for biological functions (e.g. photosynthesis,
87 respiration and macronutrient assimilation), and Cd can supplement these functions. **Trace metals** availability can
88 influence phytoplankton growth and community structure (Morel and Price, 2003). In turn, plankton control the
89 distribution, chemical speciation, and cycling of trace metals in the sea (Sunda, 2012), by, for example, releasing
90 organic compounds that dominate the coordination chemistry of metals, internalizing trace elements into the cells, and
91 reducing and/or oxidizing metals at the cell surface. The chemistry of redox speciation of active trace **metals** is highly
92 dependent on pH. For instance, Fe occurs in two main redox states in the environment: oxidized ferric Fe (Fe (III)),
93 which is poorly soluble at circumneutral pH; and reduced ferrous Fe (Fe (II)), which is easily soluble and therefore
94 more bioavailable. Fe speciation and bio-availability are dynamically controlled by the prevalent changing redox
95 conditions. Also, as the ocean becomes more acidic, reduction of Cu (II) will increase, as the ionic form of Cu (II) is
96 reduced to Cu (I) (Millero et al., 2009). The effect of higher concentrations of Cu (I) in surface waters on biological
97 systems is not well known. Therefore, while the effects of OA on inorganic metal speciation will be more pronounced
98 for metals that form strong complexes with carbonates (e.g. copper) or hydroxides (e.g. iron and aluminium), those that
99 form stable complexes with chlorides (e.g. cadmium) will not be greatly affected. **pH mediated changes in**
100 concentrations and/or speciation could possibly enhance **trace metals** limitation and/or toxicity to marine plankton
101 (Millero et al., 2009).

102 ▲
103 Iron is crucial for phytoplankton growth because of its involvement in many essential physiological processes, such as
104 photosynthesis, respiration, and nitrate assimilation (Behrenfeld and Milligan, 2013). The decrease in seawater pH in
105 response to OA may increase Fe solubility (Millero et al., 2009), but it may also result in unchanged or lower Fe
106 bioavailability, depending of the nature of the strong organic Fe ligands (Shi et al., 2010). Consequently, changes in
107 iron bioavailability due to ocean acidification can affect positively or negatively ocean productivity and CO₂
108 drawdown. Copper is an essential micronutrient but may be toxic at high concentrations (Semeniuk et al., 2016). An
109 increase in free cupric ion concentrations in coastal areas due to ocean acidification (Millero et al., 2009) could result in
110 negative effects on phytoplankton. From the open-ocean to coastal areas, the concentration of metals differ, as well as
111 the trace metal requirements of phytoplankton (Sunda and Huntsman, 1995a), and their tolerance to metal toxicity.
112 Accordingly, changes in pH may promote an increase in Cu toxicity in coastal phytoplankton, or enhance Fe limitation
113 in the open ocean. Given that **trace metals** are essential for phytoplankton productivity, and that are actively

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120 internalized during growth, it is important to study the impacts of ocean acidification in the trace metal content of
121 ecologically significant plankton species.

122
123 In a rapidly changing global environment, generated by anthropogenic CO₂ emissions, it is critical to gain adequate
124 understanding about ecosystem responses. Due to the complex interactions in aquatic ecosystems such predictions have
125 so far not been possible to do based upon observational data and modelling alone. However, direct empirical studies on
126 natural communities offer a robust tool to analyse interactive effects of multiple stressors. Specifically, mesocosm
127 experiments allow perturbation studies with a high degree of realism compared to other experimental systems
128 (Riebesell et al., 2010, Stewart et al., 2013, Riebesell and Gattuso, 2015).

129
130 In the present work a bloom of the coccolithophorid *Emiliania huxleyi* was induced in a mesocosm experiment to
131 examine the interactive effects of increased CO₂ and/or dissolved iron on its growth and physiology (Segovia et al.,
132 2017, Segovia et al., 2018, Lorenzo et al., 2018). *Emiliania huxleyi* is the most cosmopolitan and abundant
133 coccolithophore in the modern ocean (Paasche, 2002). Coccolithophores play a key role in the global carbon cycle
134 because they produce photosynthetically organic carbon, as well as particulate inorganic carbon through calcification.
135 These two processes foster the sinking of particulate organic carbon to the deep ocean carbon export (Hutchings, 2011)
136 and impact organic carbon burial in marine sediments (Archer, 1991, Archer and Maier-Reimer., 1994). However,
137 ocean acidification will disproportionately affect the abundance of coccolithophores, as well as their rates of
138 calcification and organic carbon fixation (Zondervan et al., 2007). The aim of the present study was to characterize the
139 changes in particulate trace metal concentrations during the bloom of *E. huxleyi* given realistic changes in CO₂ and Fe
140 bioavailability.

141 2. Materials and methods

142 2.1 Experimental set-up

143 The experimental work was carried out in June 2012 in the Raunefjord, off Bergen, Norway as described in detail by
144 Segovia et al., (2017). Twelve mesocosms (11 m³ each) were set-up in a fully factorial design with all combinations of
145 ambient and elevated pCO₂ and dFe in three independent replicate mesocosms. The mesocosms were covered by lids
146 (both transparent to PAR and UVR) and filled with fjord water from 8 m depth. We achieved two CO₂ levels
147 corresponding to present (390 ppm, LC) and those predicted for 2100 (900 ppm, HC) by adding different quantities of
148 pure CO₂ gas (Schulz et al., 2009). The specific CO₂ concentration and the CO₂ inlet flows in the mesocosms were
149 measured by non-dispersive infrared analysis by using a Li-Cor (LI-820) CO₂ gas analyser (Li-COR, Nebraska, USA).
150 CO₂ concentrations in the mesocosms were calculated from pH and total alkalinity measurements using the CO₂ SYS
151 software (Robbins et al., 2010). At the beginning of the experiment, nitrate (10 µM final concentration) and phosphate
152 (0.3 µM final concentration) were added to induce a bloom of the coccolithophore *Emiliania huxleyi*, as recommended
153 by Egge & Heimdal (1994). To induce changes in Fe availability, and analyse its effects on the plankton community, 70
154 nM (final concentration) of the siderophore desferrioxamine B (DFB) (+DFB and -DFB treatments) (Figure 1b) was
155 added to half of the mesocosms on Day 7, when the community was already acclimated to high CO₂. The initial dFe
156 concentration before DFB addition was about 4.5 nM. Even though DFB is a strong Fe-binding organic lig- and often
157 used to induce iron limitation in phyto- plankton (Wells 1999), DFB additions may also in- crease the dissolved Fe pool
158 in environments with high concentrations of colloidal and/or particulate Fe, such as fjords (Kuma et al. 1996, Öztürk et
159 al. 2002). By day 17, dissolved iron concentrations were significantly higher (by ~3-fold) in the high CO₂ and DFB
160 treatments than in the control (Segovia et al. 2017). These results support an increase in the solubility of Fe in seawater
161 by either lowering its pH (Millero 1998; Millero et al. 2009) and/or the addition of DFB (Chen et al. 2004). The
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178 multifactorial experimental design consisted of triplicate mesocosms per treatment and the combinations of high and
179 ambient pCO₂ and dFe levels, resulted in a total of 12 mesocosms: LC-DFB (control), LC+DFB, HC+DFB and HC-
180 DFB. Water samples from each mesocosm were taken from 2 m depth by gentle vacuum pumping of 25 L volume into
181 acid-washed carboys that were quickly transported to the onshore laboratory. The biological and chemical variables
182 analysed were phytoplankton abundance and species composition, dissolved Fe and Cu concentrations (dFe, dCu),
183 nutrient concentrations (nitrate, phosphate, silicic acid and ammonium) and particulate trace metal concentrations.

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185 2.2 Dissolved copper (dCu)

186 Low density polyethylene (LDPE) bottles were cleaned with 1% alkaline soap solution for one week, then filled with 6
187 M trace metal grade HCl and submerged in a 2 M HCl bath for one month. For transport, they were filled with 1 M
188 trace metal grade HCl (Fisher Chemicals) for one more month and kept double bagged. In between each acid treatment
189 the bottles were rinsed with Milli-Q water (Millipore; hereafter referred to as MQ). Before sampling, the bottles were
190 rinsed three times with filtered seawater. Seawater was collected from each mesocosm, filtered through 0.2 µm
191 AcroPak Supor membrane capsule filters into the trace metal clean LDPE bottles, and acidified with ultra-clean HCl
192 (Seastar) in a Class 100 laminar flow hood. Total dissolved Cu concentrations were measured following Zamzow et al.
193 (1998) using a flow injection analysis chemiluminescence detection system (CL-FIA, Waterville Analytical). Total
194 dissolved Fe concentrations were measured as described in Segovia et al., (2017) for this very experiment.

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196 2.3 Particulate metals (pMe)

197 2.3.1 Sampling

198 All equipment used during this study was rigorously acid-washed under trace metal clean conditions. Filters were
199 precleaned with 10% trace metal hydrochloric acid (Fisher, trace metal grade) at 60°C overnight and were rinsed with
200 Milli-Q (MQ) water. Seawater samples (1-3.5 L) were filtered gently onto 0.45 µm acid washed Supor®-450 filters
201 (within a trace metal clean Swinnex filter holder) on days 12, 17 and 21 of the experiment. Four technical replicates
202 were taken from each mesocosm. Two filters were analysed without manipulation and the other two were individually
203 washed with oxalate-EDTA reagent to remove extracellular Fe, as well as other metals (Tang and Morel, 2006).
204 Immediately following filtration, the treated filters were soaked with 20 mL EDTA-oxalate solution, added to the
205 headspace of the Swinnex holders, with an acid-washed polypropylene syringe. After 10 min, vacuum was applied to
206 remove the oxalate solution and 10 mL of 0.2 µM filtered chelexed synthetic oceanic water (SOW) solution was passed
207 through the filter to rinse off any remaining oxalate solution. Replicate filters that were not treated with oxalate solution
208 were transferred directly to centrifuge tubes for storage. The filters with particles were frozen in acid-washed 2 mL PP
209 tubes and then, dried and stored until analysis.

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211 2.3.2 Analytical methods

212 Filters were digested in 7-mL acid-washed Teflon (Teflon, Rochester, NY, USA) vials. Teflon vials were also
213 precleaned using 10% trace metal hydrochloric acid (Fisher, trace metal grade) during two days and then, with nitric
214 acid (Fisher, trace metal grade) at 70 °C during three days. In between each acid treatment, the bottles were rinsed with
215 MQ. Samples were digested in 3 mL of HNO₃ and 0.5 mL of HF (Fisher, trace metal grade) with lids on for 1 h on a
216 hot plate at 200 °C. The lids were then removed to evaporate HF at 200°C. After this, 1.5 mL of HNO₃ were added and
217 the samples were heated with lids on overnight at 150 °C. Finally, 2.25 mL of HClO₄ (Fisher, Optima grade) were
218 added and the samples were heated for 4 h at 200 °C. After complete digestion, the samples were dried on hot plates at
219 200°C. The dried samples were dissolved in 1% nitric acid with 1 ppb in internal standard. The analysis was performed

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229 using a high-resolution inductively coupled plasma-mass spectrometer (ICP-MS, Element XR, Thermo Scientific) and
230 the described instrumental settings (Table S1). Filter blanks were collected and subjected to the same storage, digestion,
231 dilution, and analysis processes, and these blank values were subtracted from sample measurements. Particulate
232 samples for ICPMS analysis were processed in a trace metal-clean laboratory under a trace metal-clean laminar flow
233 fume hood.

235 2.3.3 The effect of oxalate-EDTA wash on particulate trace metal concentrations

236 To better estimate the biogenic fraction of the particulate metals, the filters were washed with an oxalate-EDTA
237 solution, which removes extracellular metals and oxyhydroxides (Tovar-Sanchez et al., 2003; Tang and Morel, 2006).
238 In our study, the oxalate wash significantly decreased the concentration of all particulate metals, with the exception of
239 Al and Ti (Tables S2 and S3), as observed by Rauschenberg and Twining (2015). The quantity of metal remaining after
240 the oxalate wash (i.e. biogenic fraction) varied among elements (Tables S2 and S3). In general, the concentrations of Fe
241 and Co in the particles were decreased the least by the oxalate wash by ~25%, while Mo and Pb concentrations were
242 decreased the most by ~70%. The concentrations of particulate Cu, Zn, Cd and Mn were reduced by 50% by the oxalate
243 wash. As shown previously (Sanudo-Wilhelmy et al. 2004), the oxalate reagent also removed extracellular P (by ~20%,
244 Table S2 & S3). Compared to Rauschenberg and Twining (2015), the estimates of the biogenic fraction, after the
245 oxalate wash, were in agreement for Co, Cu and P, and lower for Fe, Mn, Zn and Cd concentrations.

247 However, the efficacy of the oxalate wash to dissolve Fe, and other metals, from lithogenic particles is not well
248 constrained (Frew et al. 2006, Rauschenberg and Twining, 2015, King et al., 2012). Therefore, the results obtained after
249 the oxalate-EDTA wash should be interpreted with caution because we do not know whether the removed metal
250 fraction is a) only lithogenic; b) mainly lithogenic but some biogenic fraction is also removed, or c) whether metals
251 absorbed onto particles are equally labile to the wash on biogenic and lithogenic particles. Given that many of the
252 trends we observed were identical for the oxalate-EDTA washed and non-washed particles [i.e. higher Me
253 concentrations in the LC+DFB treatments (Table S2 & S3) and positive correlations between phytoplankton biomass
254 and Me concentrations (Lorenzo-Garrido 2016)], below we present and discuss only the non-oxalate wash results.

256 2.4. Statistical analyses

257 Data were checked for normality (by Shapiro-Wilks' test), homoscedasticity (by Levene's test) and sphericity (by
258 Mauchly's test). All data met the requirements to perform parametric tests. Statistical significance of treatment effects
259 was carried out using Split-Plot ANOVA followed by post-hoc Sidak and Bonferroni tests (considering $P < 0.05$ as
260 significant). All analyses were performed using the General Linear Model (GLM) procedure. The correlation between
261 variables was analysed by Pearson's product-moment multiple comparisons (considering $P < 0.05$ as significant).
262 Statistical analyses were carried out using SPSS v22 (IBM statistics) and Sigmaplot 12 (Svstat Software, Chicago,
263 USA).

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676 **3. Results**

677 **3.1 Biological and chemical characteristics during the bloom**

678 Plankton community dynamics and their response to the applied treatments in the mesocosms are described in detail by
679 Segovia *et al.* (2017). Briefly, at the beginning of the experiment (days 1-10) a bloom of large chain-forming diatoms
680 was observed, which declined by day 7 (Figure 2). This diatom bloom was associated with a sharp decrease in nitrate
681 and silicic acid concentrations (Figure S1-supplemental material). Picoeukaryotes, dominated the phytoplankton
682 community on day 8 (Figure 2). During the first 10 days of the experiment, there were no significant differences in the
683 chemical variables measured between the treatments (Figures 1 and S1). On day 7, half of the mesocosms were
684 amended by adding DFB (+DFB treatments). Between day 7 and 17, an increase in dFe was observed in all treatments,
685 except in the control (Figure 1). This increase in dFe was sustained for the entire experiment in the DFB treatments
686 (Figure 1). Dissolved Cu concentrations were not affected by the different treatments (Figure 1). After day 10, a
687 massive bloom of the coccolithophore *Emiliana huxleyi* developed under LC +DFB condition (Figure 2), out-
688 competing the rest of the plankton groups (Figure 2). This bloom was not observed either in the control treatment (LC-
689 DFB) or in the HC treatments, although *E. huxleyi* was still the most abundant species in all treatments, with the
690 exception of the HC-DFB treatment (Figure 2).

691 **3.2 Particulate metal concentrations during the mesocosm experiment**

692 The pMe concentrations (nM, mean of all treatments and dates) during the experiment were highest for Al, Fe and Zn,
693 and lowest for Cd, following this trend: Al \approx Fe \approx Zn > Ti > Cu \approx Mn > Mo \approx Pb > Co > Cd (Figure 3, Table S2).
694 Significant changes over time were observed for all particulate trace metal concentrations (Fe, Cu, Co, Zn, Cd, Mn, Mo
695 and Pb), except for Ti and Al (Figure 3, Table 1). The only metal that showed a significant time-dependent decrease in
696 its particulate concentration was Fe (Figure 3, Table 1). In general, the treatments with the highest particulate metals
697 concentrations also exhibited the highest particulate P, except for Al, Ti, Fe, and Pb (Figure 3, Table S2). On days 12
698 and 17, the highest particulate metals concentrations were observed in the LC+DFB treatment, while on day 21, they
699 were observed in both LC treatments (Figure 3, Table S2).

700 **3.3 The effects of increased CO₂ and the DFB addition on particulate metal concentrations**

701 Increased CO₂ and the DFB addition did not significantly affect the concentrations of particulate Al, Ti, Cu, and Pb
702 (Tables 1 and S2). Similarly, the addition of DFB did not directly influence particulate concentrations of Fe, but high
703 CO₂ had a significant negative impact on particulate Fe (Tables 1 and S2, Figure 3). Particulate Cd concentrations were
704 also inversely affected by CO₂, but only in the presence of DFB (CO₂; and CO₂ x DFB effect, Tables 1 and S2, Figure
705 3). All other elements (P, Co, Zn, Mn and Mo) exhibited significant effects by CO₂ and by DFB, but there was also a
706 significant interaction between these two factors (Table 1, S2). This indicates that, for example, particulate Mn, Zn, Mo,
707 Co, and P concentrations were significantly decreased by high CO₂, but only in the +DFB treatments (Figure 3, Table 1,
708 S2). Similarly, the addition of DFB significantly increased pZn and pMn, but only at ambient CO₂ levels (Figure 3,
709 Table 1, S2).

710 **3.4 Phosphorous-normalized metal ratios in particles collected from the mesocosms and the effects of increased
711 CO₂ and the DFB addition on these ratios**

712 The P-normalized metal ratios (Figure 4 and means in Table 2) were highest for Al and Fe (mean: 70 \pm 38 mmol Al:
713 mol P, and 39 \pm 34 mmol Fe: mol P), and lowest for Cd and Co (mean 0.02 \pm 0.01 mmol Cd: mol P, and 0.07 \pm 0.02
714 mmol Co: mol P). Iron:P and Ti:P were not significantly affected by increased CO₂ and/or the DFB addition, but
715 showed a significant decrease over time (Table 3). The P-normalized Cu, Co and Zn ratios changed significantly over
716
717
718

Movido (inserción)[3]

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Movido (inserción)[4]

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Con formato: Fuente: Sin Negrita, Inglés (británico)

719 time (Table 3). Increased CO₂ significantly decreased Co, Zn and Mn:P ratios, while it increased Cu:P ratios (Figure 4,
720 Table 3). DFB did not affect the Me:P ratios of any of these bioactive elements (Table 3).

721 4. Discussion

723 4.1 The effects of CO₂ and dFe in the plankton community

724 In this experiment we investigated changes in particulate trace metal concentrations, in response to increased CO₂
725 and/or an addition of the siderophore DFB in a coastal mesocosm experiment. For a better understanding of the
726 processes affecting these stressors, we briefly summarise the mesocosm experiment results originating from Segovia et
727 al. (2017). High CO₂, as well as the DFB addition increased dFe concentration. The higher dFe concentrations were
728 sustained in the DFB treatments. A bloom of the coccolithophore *Emiliania huxleyi* was observed in the ambient CO₂
729 treatments, and was especially massive in the one with the addition of DFB (LC+DFB). On the contrary, the biomass of
730 *E. huxleyi* was negatively affected by increased CO₂. However, increased dFe partially mitigated the negative effect of
731 elevated CO₂, indicating that the coccolithophore was able to acclimate better to ocean acidification when Fe
732 availability was high. High dFe also had a positive effect on the cyanobacterium *Synechococcus sp.*, while the rest of the
733 plankton food web did not response to the treatments (Segovia et al. 2017).

734 4.2 Particulate Fe and Ti are associated with lithogenic sources, while particulate Co, Cu, Zn, Cd, Mo and Mn 735 are associated with biogenic sources

736 The particulate trace metal concentrations (nM, mean of all treatments and dates) during the experiment were highest
737 for Al, Fe and Zn, and lowest for Co and Cd, following this trend: Al ≈ Fe ≈ Zn > Ti > Cu ≈ Mn > Mo ≈ Pb > Co > Cd.
738 Lithogenic particles are enriched in Al and low in P (average crustal Al and P content is 2.9 mmol Al and 0.034 mmol
739 P g⁻¹ dry weight, Taylor 1964), while biogenic particles are enriched in P and low in Al (average plankton Al and P
740 content is 0.001 mmol Al and 0.26 mmol P g⁻¹ dry weight, Bruland et al. 1991). Therefore, the distinct high abundance
741 of Al and P in lithogenic and biogenic particles, respectively, can be used to evaluate the relative contribution of
742 lithogenic and biogenic material in our particulate samples. In order to do this, first, it is important to establish that the
743 vast majority of the measured particulate P is associated the biogenic fraction. In this study, the abiotic P was estimated
744 using particulate Al concentrations (nM) and the P:Al ratio in crustal material, and was calculated to be negligible (<
745 1% of the total measured particulate P). In addition, a significant correlation (p< 0.003) was found between particulate
746 P concentrations and phytoplankton biomass (Table 4). Therefore, we assume a constant trace metal composition in
747 biogenic particles (assuming they are rich in phytoplankton) and lithogenic particles (assuming they are rich in crustal
748 material). We then calculated the expected metal concentrations in the particulate samples assuming that all the P
749 measured in the particles is associated with a biogenic fraction, and that all Al in the particles is associated with the
750 lithogenic fraction. Thus, for a given trace metal, its expected particulate trace metal concentration in seawater (mol L⁻¹
751) can be calculated as the sum of the contribution from biogenic and lithogenic particles, so that:

$$753 [Me] = a [P] + b [Al]$$

754
755 where [Me] is the total concentration of the metal (mol L⁻¹) expected in the particulate sample; [P] is the P
756 concentration measured in the particles (mol L⁻¹); [Al] is the Al concentration measured in the particles (nM L⁻¹); *a* is
757 the average, well-known metal content in biogenic particles, normalized to P (i.e. mol Me: mol P in marine plankton)
758 and *b* is the average, well known metal content in lithogenic particles, normalized to Al (mol Me: mol Al in the Earth
759 crust). For example, on day 21 in the HC-DFB treatment, the concentrations of particulate Al and P were 8.22 and
760 134.8 nM, respectively (Table S2). Assuming a constant 0.0051 mol Fe: mol P in biogenic particles and 0.331 mol Fe:

Movido (inserción)[5]

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Con formato: Fuente: Negrita, Inglés (británico)

Eliminado: Cu, Co, Zn, Cd, Mn and Mo) were

762 mol Al in lithogenic particles (Table 2), we calculated an expected particulate Fe concentration of 3.41 nM, where 80%
 763 was associated with lithogenic material and 20% with biogenic material. Similar calculations were made for the
 764 bioactive metals Mn, Co, Cu, Zn, Cd, and Mo (Table 5). Our calculations indicate that on average, particulate Fe was
 765 dominated by the lithogenic component (accounting for an average of 78% of the total expected particulate Fe), while
 766 for particulate Co, Cu, Zn, Cd, and Mo the biogenic fraction dominated (accounting for 94, 95, 99, 94 and 98%,
 767 respectively, of the total expected concentration; Table 5). Particulate concentrations of Mn were also dominated by the
 768 biogenic fraction (65% of the total), but the lithogenic fraction was also significant (35%). Moreover, the expected
 769 particulate Mn and Fe concentrations closely matched the particulate Mn and Fe concentration we measured
 770 (accounting for an average of ~71% of the measured Mn, and 115% of the measured Fe). For other metals (i.e. Cu, Mo
 771 and Zn), the expected particulate concentrations (nM) were lower than measured (23% of the measured pCu, and 8% of
 772 measured pZn; Table 5). This suggests that the particles were enriched in Cu, Mo, and Zn relative to what is expected
 773 based on natural marine plankton metal quotas (Bruland et al. 1991) and crustal ratios (Taylor 1964).

775 To further establish the lithogenic or biogenic source of the pMe in the particles, the particulate metal concentrations
 776 were normalized to the concentrations of particulate P and Al (Figure 4, and Table 2). These ratios were then compared
 777 with well-known molar ratios of metal to Al in the crust (Taylor 1964) and of metal to P ratios in marine plankton
 778 samples (Ho 2006) and cultures (Ho et al. 2003) (Table 2). The average Fe: Al (506 mmol Fe: mol Al) and Ti:Al ratios
 779 (119 mmol Ti: mol Al, Table 2) were relatively similar to crustal molar ratios (331 mmol Fe: mol Al and 39 mmol Ti:
 780 mol Al; Taylor 1964). Additional evidence for the significant lithogenic component in particulate Ti and Fe was
 781 gathered from Figure 5, where we plotted the molar ratios of the metals relative to P in the collected particles against
 782 the Al:P ratios measured in those same particles. The slope of these data [(Fe:P)/(Al:P) = mol Me: mol Al] is the ratio
 783 of Me:Al in the particles and can be compared to well-known Me:Al crustal ratio. Visually, if the data nicely fit the
 784 Me:Al line for crustal material, these metals are mainly associated with the lithogenic component, as evident for Fe and
 785 Ti (Figure 5). These combined results suggest that in our experiment, particulate Fe and Ti concentrations were
 786 enriched by lithogenic material. In support of this finding, we also found no significant correlation between particulate
 787 Fe and Ti concentrations (nM) and either the total plankton (phytoplankton and microzooplankton) or *E. huxleyi*
 788 biomass ($\mu\text{g C L}^{-1}$; Table 4).

790 In contrast, when the P-normalized metal ratios in the particles collected from the mesocosms were plotted against the
 791 Al:P ratios in these particles, there were no correlations for the following metals Co, Cu, Zn, Cd, Mn and Mo (Figure
 792 5), indicating that these particulate metals were not enriched in lithogenic material. Our measured metal: P ratios were
 793 comparable to plankton ratios in natural samples and in cultures (Table 5). The concentrations (mol L^{-1}) of these metals
 794 (i.e. Cu, Co, Zn, Cd, Mn, Mo), as well as P, also showed significant correlations with the biomass ($\mu\text{g C L}^{-1}$) of *E.*
 795 *huxleyi* and that of total plankton cells ($p < 0.05$, Table 4), supporting a significant influence of the phytoplankton in the
 796 distribution of these particulate elements.

798 4.3 Particulate metals with a strong biogenic component: their P-normalized ratios

799 The concentrations of particulate bioactive metals (mol L^{-1}), with a significant biogenic component (i.e. excluding Fe),
 800 in the studied *Emiliania huxleyi* bloom were ranked as: $\text{Zn} > \text{Cu} \approx \text{Mn} > \text{Mo} > \text{Co} > \text{Cd}$ (Figure 3, Table S3), similar to
 801 those reported in indigenous phytoplankton populations: $\text{Fe} \approx \text{Zn} > \text{Cu} \approx \text{Mn} \gg \text{Co} \approx \text{Cd}$, (Twining and Baines, 2013).
 802 The only treatment where *E. huxleyi* did not dominate the community was the HC-DFB; in this treatment the ranking of
 803 these biogenic particulate trace metals was the same as that of LC+DFB (with the massive *Emiliania huxleyi* bloom).

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Eliminado: phytoplankton

Con formato: Fuente: Sin Negrita, Inglés (británico)

Movido (inserción)[6]

Con formato: Inglés (británico)

Eliminado: Certain particulate elements (P, Cu, Co, Zn, Cd, Mn and Mo) were clearly biogenic. Three lines of evidence are presented in support of this. First, the total biomass of phytoplankton exhibited a significant positive correlation with particulate P (Table 6), suggesting that most particulate P was biogenic, as shown previously (Ho et al., 2007, Ho et al., 2009). The concentrations of Cu, Co, Zn, Cd, Mn and Mo also exhibited positive significant correlations with the biomass of total cells (phytoplankton and microzooplankton) or *E. huxleyi* (Table 6), indicating that these particulate metals were also associated with phytoplankton. Second, the Me:P ratios are not similar to crustal ratios. Third, the Me:P ratios we measured in the particles are similar to those of natural phytoplankton assemblages (Ho, 2006) and of *Emiliania huxleyi* cultures (Ho et al., 2003).

Eliminado: biogenic metals

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Eliminado: 3

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Eliminado: $\approx \text{Ni} \approx \text{Cu}$

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Eliminado: Particulate Zn concentrations were especially high in the LC+DFB treatment, where the highest *E.*

Movido hacia abajo[7]: *huxleyi* biomass was observed.

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Eliminado: . In

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Eliminado: ,

Con formato: Inglés (británico)

Eliminado: particulate trace metal

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Con formato: Inglés (británico)

Eliminado: ,

Con formato: Inglés (británico)

832 but their concentrations were lower than those in LC+DFB. At the end of the experiment, the concentrations of these
833 biogenic metals were, in general, comparable in both HC treatments, and lower than those in the LC treatments (Figure
834 3, Table S3). Therefore, high CO₂ had a tendency to decrease particulate metal concentrations, especially on day 21.
835 Given the strong correlation between concentrations of these particulate bioactive metals and phytoplankton biomass,
836 the lower particulate concentrations in high CO₂ were mainly due to low phytoplankton biomass.

837
838 Particulate Zn concentrations were especially high in the LC+DFB treatment (Figure 3), where the highest *E. huxleyi*
839 biomass was observed. *Emiliania huxleyi* is well known for its high Zn cellular requirements (~ 1-10 for *E. huxleyi* vs.
840 1-4 mmol Zn: mol P for other phytoplankton; Sunda and Hunstman 1995, Sunda 2013). But, the Zn: P ratios in the
841 LC+DFB treatment (range 45-69 mmol Zn: mol P; Figure 4, Table S2), as well as in all the other treatment (range 16-
842 34 mmol Zn: mol P; Figure 4, Table S2) were significantly higher than these published ratios. This could be explained
843 by, the adsorption of these metals to the outside of the cells, and/or anthropogenic inputs of Zn into the fjord. The Zn:P
844 ratios in the samples washed with the oxalate-EDTA were still high (range 28-57 for LC+DFB and 16-33 mmol Zn:
845 mol P in all other treatments, Table S3), thus adsorption might have not been significant. We hypothesize that
846 anthropogenic aerosols which are rich in anthropogenic particulate metals, such as Zn and Cu (Perry et al. 1999; Narita
847 et al. 1999), and have high percentage of Zn and Cu dissolution (ref.), might be the source of these high Zn
848 concentrations and ratios in the particles.

849
850 Similarly, the Cu:P ratios in the collected particles were relatively elevated (1.4 ± 0.8 mmol Cu: mol P) compared to
851 those of other phytoplankton, including *E. huxleyi* (Table 2). The dissolved (7.7 ± 0.41 nM Cu, Figure 1) and particulate
852 Cu concentrations (0.35 ± 0.25 nM, Table S2) in our experiment were high, and similar to those previously measured in
853 this fjord (Muller et al., 2005). Rain events (or wet deposition of anthropogenic aerosols) in this fjord result in high
854 dissolved Cu and the active production of strong organic ligands by *Synechococcus*—to lower the free Cu
855 concentrations (Muller et al., 2005). Therefore, high Cu might be a general condition in this fjord, and indigenous
856 plankton might have developed physiological mechanisms to deal with high Cu, such as the production of organic
857 ligands to prevent uptake (Vraspir and Butler, 2009), or of heavy-metal-binding peptides (phytochelatins) to lower Cu
858 toxicity inside the cell (Ahner and Morel, 1995; Ahner et al., 1995; Knauer et al., 1998). Since we measured high
859 particulate Cu, and Cu:P in our experiment, *E. huxleyi* might have been relying mainly on phytochelatins to buffer high
860 intracellular Cu (Ahner et al., 2002).

861
862 The Cd:P ratios (average 0.024 ± 0.01 mmol Cd: mol P, Figure 4 and 6) were significantly lower than those in
863 phytoplankton and *E. huxleyi* (0.36 mmol Cd: mol P, Figure 4 and 6). This was surprising, because Cd quotas are
864 normally higher in coccolithophores than in diatoms and chlorophytes (Sunda and Huntsman, 2000; Ho et al., 2003).
865 High Cd quotas in coccolithophores have been suggested to result from accidental uptake through Ca transporters and
866 channels (Ho et al., 2009). The low Cd quotas here may be explained by the antagonistic interaction between Mn and
867 Cd or Zn and Cd under high Mn and Zn, respectively (Sunda and Huntsman, 1998, 2000; Cullen and Sherrell, 2005).
868 Since high Zn:P ratios were common in this study (34.02 ± 18.05 mmol Zn: mol P, Figure 4 and 6), we hypothesize that
869 high Zn levels antagonistically interacted with Cd, resulting in low Cd:P ratios in the particles.

870
871 **4.4 The effects of increased CO₂ and the DFB addition on particulate metal concentrations and P-normalized**
872 **ratios**

873 Iron enrichment is common in coastal waters, due to sediment resuspension, rivers input, aeolian deposition and mixing
874 or upwelling of deep water. Indeed, Fe was the essential metal with the highest particulate concentrations in our study

- Eliminado: higher
- Con formato ... [38]
- Eliminado: HC
- Con formato ... [39]
- Eliminado: on day 17
- Con formato ... [40]
- Eliminado: metal
- Con formato ... [41]
- Eliminado: in both HC treatments
- Con formato ... [42]
- Con formato ... [43]
- Eliminado: and, similar effect of
- Con formato ... [44]
- Eliminado: factors (increased CO₂ and the addition of
- Con formato ... [46]
- Movido (inserción)[7]
- Con formato ... [47]
- Eliminado: *Emiliania huxleyi* is well known for its high Zn
- Con formato ... [49]
- Eliminado: ¶
- Con formato ... [50]
- Con formato ... [51]
- Eliminado: Given that
- Con formato ... [52]
- Con formato ... [53]
- Eliminado: but comparable to previous studies (Muller
- Con formato ... [54]
- Con formato ... [55]
- Eliminado: or on other detoxification mechanisms
- Con formato ... [56]
- Eliminado: Indeed, in the genome of *E. huxleyi*, we found
- Con formato ... [58]
- Eliminado: Alternatively, Cu might be particularly
- Con formato ... [60]
- Con formato ... [61]
- Con formato ... [62]
- Eliminado: found
- Con formato ... [63]
- Con formato ... [64]
- Eliminado: .
- Con formato ... [65]
- Eliminado: The high Zn:P ratios in this study indicate high
- Con formato ... [67]
- Movido hacia arriba[6]: ¶
- Con formato ... [68]
- Con formato ... [69]
- Eliminado: Fe in bulk particles was mainly lithogenic¶
- Con formato ... [70]
- Eliminado: The
- Con formato ... [71]

1002 (Figure 3, Table 3). Furthermore, in this study particulate Fe was characterized by a strong lithogenic component, and
 1003 was not correlated with phytoplankton biomass. Iron was also unique, in that it was the only trace element whose
 1004 particulate concentration was significantly and uniquely affected by CO₂, regardless of the presence or absence of DFB
 1005 (no interaction between CO₂ and DFB, Table 1). Furthermore, particulate Fe concentrations (nM) decreased steadily
 1006 between days 12 and 21, with the exception of the control treatment (LC-DFB; Figure 3, Table 2S). This suggests that
 1007 the increase in CO₂ and/or the DFB addition reduce the concentration of pFe, despite the phytoplankton bloom. Such a
 1008 decrease in pFe (range 2.3-fold in LC-DFB, vs. 13.7-fold in HC+DFB; Table S2) might be mediated by the dissolution
 1009 of particulate Fe by low pH or by the presence of strong organic chelators as observed in this very experiment (Segovia
 1010 et al. 2017 and references therein), where dFe notably increased in treatments with high CO₂ and/or the addition of DFB
 1011 (Figure 1). Furthermore, the dissolution of particulate Fe in the treatments with high CO₂ and/or the addition of DFB
 1012 was evident in the Fe partitioning coefficients—the molar ratio between particulate and dissolved concentrations. On
 1013 day 21, the Fe partitioning coefficients varied by 22-fold between the highest for the control (LC-DFB: 1.039) and
 1014 lowest for the HC+DFB treatments (HC+DFB: 0.047; Figure S2). Thus, either the DFB addition or high CO₂ promoted
 1015 the dissolution of pFe. However, at the end of the experiment, high dFe concentrations were only observed in the
 1016 treatments with the DFB additions, suggesting that the presence of strong organic Fe chelators, such as DFB, mediated
 1017 the maintenance of high dissolved Fe concentrations, as previously observed (Segovia et al. 2017). Thus, in our future
 1018 oceans, high CO₂ (low pH) will increase dissolved Fe concentrations in regions rich in particulate Fe, and in strong
 1019 organic Fe chelators. The deleterious effects of OA on the development of ecologically important species sensitive to
 1020 increased CO₂ such as *E. Huxleyi*, will be more relevant in high-Fe environments than in Fe-limited ones.

1021 In contrast to the findings for Fe, particulate Cu concentrations a) were not affected by either high CO₂ or the DFB
 1022 addition; b) were dominated by a biogenic component and c) were significantly correlated with phytoplankton biomass
 1023 (Table 4 and 5). Furthermore, unique to Cu was a significant increase in Cu:P ratios by day 21 in the high CO₂
 1024 treatments, especially when no DFB was added. Since the Cu partitioning coefficients only varied by 3.25 fold among
 1025 treatments on day 21 (LC-DFB: 0.065 vs. HC+DFB: 0.047; data not shown), we hypothesize that high CO₂ did not
 1026 affect the partitioning between particulate and dissolved Cu, but instead, it affected the speciation of dissolved Cu,
 1027 increasing its bioavailability. This resulted in the highest Cu:P ratios in the high CO₂ treatments, despite the low
 1028 phytoplankton biomass. This increase in bioavailability under lower pH is typical of metals that form strong inorganic
 1029 complexes with carbonates, such as Cu²⁺ (Millero et al., 2009). Thus in our future oceans, high CO₂ (low pH) will shift
 1030 the speciation of dissolved Cu towards higher abundance of free ionic species, increasing its bioavailability and likely
 1031 its toxicity.

1032
 1033 Similarly to Cu, particulate Co, Zn and Mn were correlated with biomass and were dominated by the biogenic
 1034 component. But in contrast to Cu, these metals particulate concentrations were affect by increased CO₂ and/or the DFB
 1035 addition. However, the effects of high CO₂ and/or DFB were very complex because significant interactions between
 1036 these 2 factors were observed (Table 1); and further studies are required before we are able to discern and conclude a
 1037 significant trend. Yet, the P-normalized ratios of Co, Zn and Mn were significantly affected by CO₂ (Table 3),
 1038 exhibiting moderately lower ratios under high CO₂, when phytoplankton biomass was lowest. These results imply that
 1039 the bioavailability of these metals was not enhanced under acidic conditions. This suggests that under high CO₂ (low
 1040 pH) the free ionic species of these metals will not significantly increase in the future, as shown for metals that occur
 1041 predominantly as free ionic species in seawater (Millero et al., 2009).

1042
 1043 **5. Concluding remarks**

Eliminado: was Fe (

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Eliminado: and Ti were the only metals with positive correlations with Al and without correlation with phytoplankton biomass (Figure 1, Table 5), indicating

Eliminado: . The similarity between our values Fe:P values and that of the crustal ratio (Figure 1) also supports this finding. Indeed, the Fe:P ratios were significantly higher than those of indigenous plankton assemblages

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Eliminado: cultures (Figure 1). We believe

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Eliminado: the lithogenic fraction of Fe in the bulk particles in our experiment masked the biogenic signal, as proposed by King et al. (2012). Interestingly, the particulate Fe ...

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Eliminado: (nmol L⁻¹) decreased between days 12 and 21 (Table 3 and 5); this could be due to the solubilisation of [72]

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Eliminado: the increase in dissolved Fe

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Eliminado: Table 1). The sinking of pFe could have... [73]

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Eliminado: dissolved phase (i.e. partition coefficient), [74]

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Eliminado: Fe from the

Con formato: Inglés (británico) ... [76]

Eliminado: to the soluble pool (Segovia et al., 2017). In [77]

Con formato: Inglés (británico) ... [78]

Eliminado: our experiment. [79]

Con formato: Inglés (británico) ... [80]

Eliminado: dissolved Fe levels on particulate metal [81]

Movido hacia abajo[8]: 2009).

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Eliminado: The decrease in particulate Fe might have [82]

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Eliminado: resulted in higher dissolved Fe, and optimal [83]

Con formato: Inglés (británico) ... [84]

Eliminado: Me:P ratios

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Eliminado: Cu, Co, Zn, and Mn. For example, while Z [86]

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Movido (inserción)[8]

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1114 The results presented here show that in the fjord where we carried out the present experiment, particulate Fe was
1115 dominated by lithogenic material, and was significantly decreased by future predicted CO₂ concentrations (HC, 900
1116 uatm) and DFB addition. This condition may well be comparable to most coastal ecosystems in the future ocean.
1117 Indeed, high CO₂ and/or DFB promoted the dissolution of particulate Fe, and the presence of this strong organic
1118 complex helped maintaining high dissolved Fe. Under control conditions at present CO₂ concentration (LC, 380 uatm)
1119 and no DFB amendment, the globally important coccolithophore *Emiliana huxleyi* was experiencing Fe limitation
1120 (Segovia et al. 2017). The shift between particulate and dissolved Fe promoted a massive bloom of *E. huxleyi* in the
1121 treatments with ambient CO₂ due to increased Fe bioavailability (for further details on Fe-bioavailability in *E. huxleyi*
1122 please see Segovia et al. 2017). Moreover, the negative effects of high CO₂ were mitigated by enhanced dFe. During the
1123 mentioned bloom, the concentrations of particulate metals with a strong biogenic component (Cu, Co, Zn, Cd, Mn, and
1124 Mo) were a) highly dynamic, b) positively correlated with plankton biomass, and c) influenced by growth requirements.
1125 Furthermore, high CO₂ decreased the Me:P ratios of Co, Zn and Mn, mainly due to low phytoplankton biomass, while
1126 increased the Cu:P ratios. In contrast DFB had no effects on these ratios. According to our results, high CO₂ may
1127 decrease particulate Fe and increase dissolved Fe, but high concentrations of dissolved Fe will only be maintained by
1128 the presence of strong organic ligands. The decrease in particulate Fe may affect the sinking flux of other metals
1129 associated with terrestrial material/dust in open ocean settings. Furthermore, ocean acidification will decrease *E.*
1130 *huxleyi* abundance, and as a result, the concentration of many biogenic particulate metals. Moreover, the Me:P ratios of
1131 metals that are predominately present in an ionic free form in seawater (e.g. Co, Zn and Mn) will likely decrease or stay
1132 constant. However, the high pZn observed will possibly be the result of anthropogenic aerosols, and the responsible for
1133 the low pCd registered, most likely due to the antagonistic interaction between Zn and Cd. In contrast, high CO₂ is
1134 predicated to shift the speciation of dissolved metals associated with carbonates, such as Cu, increasing their
1135 bioavailability, and resulting in higher Me:P ratios. We suggest that high Cu might be putative in this fjord, and
1136 autochthonous plankton might be adapted to cope with high Cu levels by developing specific physiological
1137 mechanisms. Future predicted high CO₂ levels are expected to change the relative concentrations of particulate and
1138 dissolved metals, due to the differential effects of high CO₂ on trace metal solubility, speciation, adsorption and
1139 toxicity, as well as on the growth of different phytoplankton taxa, and their elemental trace metal composition. In the
1140 future ocean, this will have great implications in the carbon cycle and the biological pump, consequently affecting the
1141 physic, chemical and biological aspects, i.e. marine systems dynamics.

Eliminado: Knowledge of the trace metal composition of in situ marine phytoplankton may allow us to estimate how microalgae in the future will influence the relative distribution and vertical transport of trace metals in the ocean. The results presented here showed that except for Ti and Fe, the trace metal concentrations of marine particles during a bloom of *Emiliana huxleyi* were a) highly dynamic, b) positively correlated with plankton biomass, c) influenced by growth requirements, and d) strongly affected by changes in CO₂ and dissolved Fe. According to our results, ocean acidification will decrease *E. huxleyi* abundance, and as a result, the concentration of some particulate trace metals that are especially high in *E. huxleyi*, such as Zn. Most importantly, OA is expected to change the relative concentrations of particulate metals, due to the differential effects of OA on the growth of marine phytoplankton species, and the contrasting metal requirements of phytoplankton phyla. OA might also affect the sinking flux of particulate Fe which would have an impact on the sinking of particulate metals associated with terrestrial material/dust in open ocean settings. Therefore, as suggested by Twining and Baines (2013), we require the development of ecophysiological models that link trace element composition of phytoplankton to physiological performance, as well as ecological models that are able to predict plankton physiological strategies and metal composition in a changing ocean, and the resulting effects of phytoplankton on the biogeochemical cycles of metals under a rapidly changing ocean.¶

1142 ▲ **Acknowledgments**

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1145 grant from the Ministry for Education (Spain) and by fellowships associated to the mentioned above research grants to
1146 carry out a short-stay at MTM and JTC laboratories to analyze dissolved and particulate metals. We thank all the
1147 participants of the PHYTOSTRESS experiment for their collaboration, and the MBS (Espesrend, Norway) staff for
1148 logistic support during the experiment.
1149

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Fuente: Times, 12 pto, Inglés (británico)

531 **Table 1.** Statistical analyses (Split-plot ANOVA) of the effects of high CO₂, the addition of DFB, and their interaction, as well as the effect of
 532 time, on the concentrations of particulate metals (mmol L⁻¹, data in Table S2, and Figure 3) in particles collected from the different mesocosms
 533 treatments.

Factor	Al	Ti	P	Fe	Cu	Co	Zn	Cd	Mn	Mo	Pb
CO ₂	ns	ns	**	*	ns	**	***	***	**	***	ns
DFB	ns	ns	*	ns	ns	*	**	ns	*	*	ns
CO ₂ x DFB	ns	*	**	ns	ns	*	**	*	**	**	ns
Time	ns	ns	ns	***	*	***	***	***	***	***	**

534 ns: not significant; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$

Eliminado: Supplemental Table S1. Instrumental conditions of ICP-MS and measurement parameters used during determination of trace elements concentrations. [Instrument conditions](#) ... [1]
Eliminado: mesocosm

540 **Table 2.** The average metal ratios in the particles collected in this study (without oxalate wash) using the data reported in Table S2. The P-
 541 normalized ratios (mmol : mol P, Figure 4) are compared to previous estimates in marine plankton samples and phytoplankton cultures (A).
 542 The Al-normalized ratios (mmol : mol Al) are compared to crustal ratios (B).

543 A)

(mmol : mol P)	Mn:P	Fe:P	Co:P	Cu:P	Zn:P	Cd:P	MocP	Al:P	Reference
Phytoplankton _{LAB}	3.8	7.5	0.19	0.38	0.8	0.21	0.03		Ho et al. 2003
Marine Plankton _{real}	0.68±0.54	5.1±1.6	0.15±0.06	0.41±0.16	2.1±0.88				Ho 2006
<i>E. huxleyi</i> _{LAB}	7.1±0.36	3.5±0.07	0.29±0.02	0.07±0.013	0.38±0.002	0.36±0.01	0.022±0.0003		Ho et al. 2003
This study	1.65±0.41	39.2±34.3	0.07±0.02	1.41±0.55	34.02±18.05	0.02±0.01	0.42±0.12	70±38	
Crust ratio	510	29,738	13	25	32	0.05	0.46	89,972	Taylor 1964

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545 B)

(mmol : mol Al)	Mn:Al	Fe:Al	Co:Al	Cu:Al	Zn:Al	Cd:Al	Moc:Al	Pb:Al	Ti:Al
Crustal ratio	5.7	331	0.14	0.27	0.35	0.001	0.005	0.02	39
This study	35±28	506±342	1.5±1.2	26.5±15	795±865	0.5±0.4	8.6±6.5	4.9±3.9	119±47.6

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547

548 **Table 3.** Statistical analyses (Split-plot ANOVA) of the effects of CO₂, DFB, and their interaction, as well as the effect of time, on the P-
 549 normalized metal quotas (mmol: mol P, data in Figure 4, and Table S2) in particles collected from the different mesocosm treatments.

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Factor	Fe:P	Cu:P	Co:P	Zn:P	Cd:P	Mn:P	Mo:P	Pb:P	Ti:P
CO ₂	ns	*	***	**	ns	*	ns	ns	ns
DFB	ns								
CO ₂ x DFB	ns								
Time	***	***	***	***	ns	ns	ns	ns	***

551 *ns: not significant; * p < 0.05; ** p < 0.01; *** p < 0.001*

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Con formato: Espacio Después: 10 pto, Control de líneas viudas y huérfanas, Ajustar espacio entre texto latino y asiático, Ajustar espacio entre texto asiático y números

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Table 4. The relationship (Pearson correlations, $p < 0.05$) between particulate metals concentrations (mmol L^{-1} , no oxalate wash, reported in Table S2) and the biomass ($\mu\text{gC L}^{-1}$) of *Emiliania huxleyi* and total cells (phytoplankton and microzooplankton) collected from the different mesocosm treatments.

		P	Fe	Cu	Co	Zn	Cd	Mn	Mo	Pb	Ti
<i>E. huxleyi</i>	Correlation coefficient	0.622									
	P-value	0.003	ns	0.614	0.756	0.747	0.818	0.686	0.825	ns	ns
Total cells	Correlation coefficient	0.641									
	P-value	0.002	ns	0.51	0.644	0.889	0.802	0.598	0.53	ns	ns

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Comentado [mt4]: this table needs to be fixed. I removed the oxalate washed data...but the lines are still there...

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Página 1: [1] Eliminado	María Segovia	11/6/19 19:50:00
Página 4: [2] Eliminado	María Segovia	11/6/19 19:50:00

27 **Supplemental Figures**

Con formato: Numeración: Continua

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30 **Particulate trace metal dynamics in response to increased CO₂ and**
31 **iron availability in a coastal mesocosm experiment**

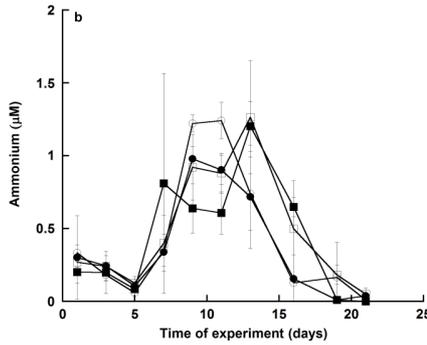
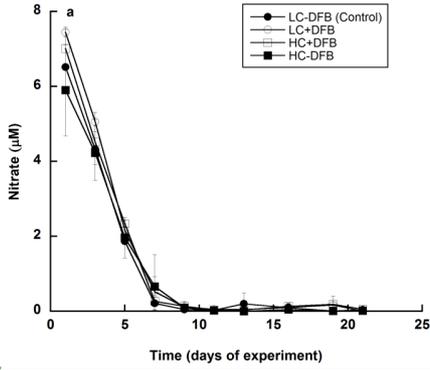
32 M. Rosario Lorenzo¹, María Segovia¹, Jay T. Cullen², and María T. Maldonado³

33 *Correspondence to:* María Segovia (segovia@uma.es) and María T. Maldonado (mmaldonado@coas.ubc.ca)

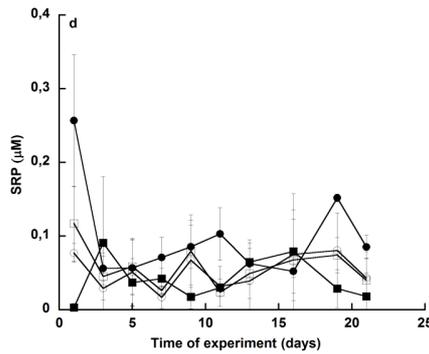
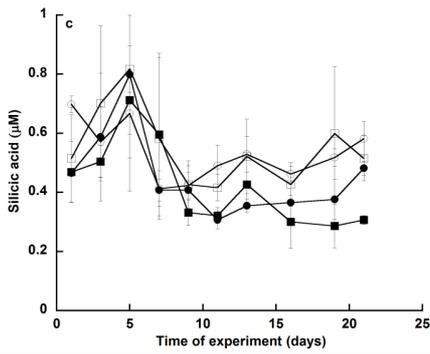
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Supplemental Figure S1. Temporal development of major nutrient concentrations within the mesocosms in the different treatments (LC: ambient CO₂ (390 μatm); HC: increased CO₂ (900 μatm); -DFB: no DFB addition; +DFB: with a 70 nM DFB addition): (a) nitrate, (b) ammonium, (c) silicic acid, (d) soluble reactive phosphate (SRP).



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Comentado [mt1]: the numbers in the Y-axis of the SRP graph need to be changed from 1,5 to 1.5 (please change the commas to dots).....

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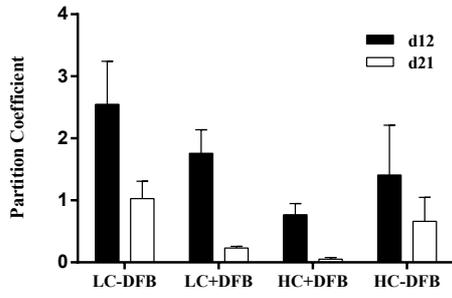
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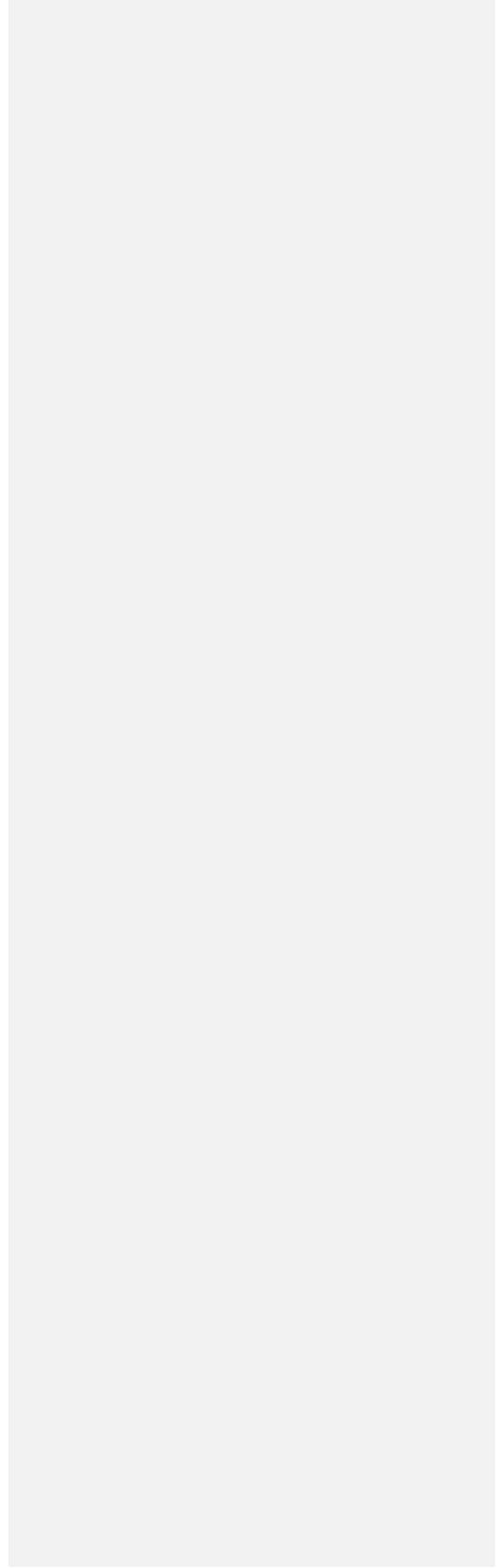
85 **Supplemental Figure S2.** The Fe partition coefficients (the molar ratio between particulate and dissolved
86 concentrations) in the different mesocosm treatments; LC: ambient CO₂ (390 μatm); HC: increased CO₂ (900 μatm); -
87 DFB: no DFB addition; +DFB: with a 70 nM DFB addition; on day 12 and day [21](#).
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Con formato: Fuente: 10 pto

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Supplemental Tables

Particulate trace metal dynamics in response to increased CO₂ and iron availability in a coastal mesocosm experiment

M. Rosario Lorenzo¹, María Segovia¹, Jay T. Cullen², and María T. Maldonado³

Correspondence to: María Segovia (segovia@uma.es) and María T. Maldonado (mmaldonado@eos.ubc.ca)

Supplemental Table S1. Instrumental conditions of ICP-MS and measurement parameters used during determination of trace elements concentrations.

<u>Instrument conditions</u>	<u>ELEMENT XR</u>
<u>Torch</u>	<u>Fassel type</u>
<u>Spray chamber</u>	<u>Glass cyclonic spray chamber</u>
<u>Nebuliser</u>	<u>ESI microflow ST nebuliser (selfaspirating)</u>
<u>Cones</u>	<u>Standard Ni sampler and skimmer</u>
<u>RF Power (W)</u>	<u>1120</u>
<u>Cooling gas flow rate (L min⁻¹)</u>	<u>16</u>

Definición de estilo: Normal

Definición de estilo: Revisión

Eliminado: Supplemental Table S1. Instrumental conditions of ICP-MS and measurement parameters used during determination of trace elements concentrations. [↩](#)

<u>Auxiliary gas flow rate (L.min⁻¹)</u>	<u>0.9</u>
<u>Sample gas flow rate (L.min⁻¹)</u>	<u>1.2</u>
<u>Sample matrix</u>	<u>1% nitric acid</u>

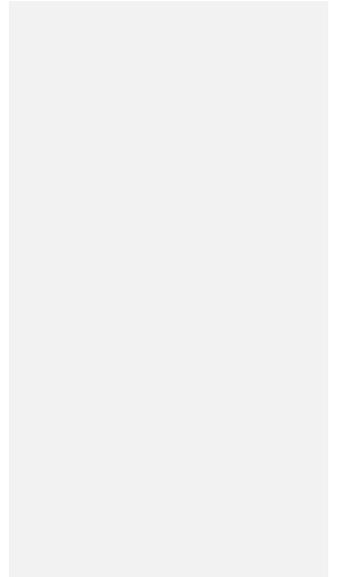
Method acquisition parameters

<u>Scan type</u>	<u>E-scan</u>	
<u>Spectral resolution</u>	<u>Low (nominal m/Δm-300)</u>	<u>Medium (nominal m/Δm-3000)</u>
<u>Isotopes of interest</u>	<u>⁹³Mo ⁸⁸Mo ¹¹¹Cd ¹¹⁴Cd ²⁰⁶Pb ²⁰⁸Pb</u>	<u>²⁷Al ³¹P ⁴⁷Ti ⁴⁹Ti ⁵⁵Mn ⁵⁶Fe ⁵⁹Co ⁶³Cu ⁶⁵Cu ⁶⁶Zn ⁶⁸Zn</u>
<u>Internal standard</u>	<u>¹¹⁵In</u>	<u>¹¹⁵In</u>
<u>Mass window (%)</u>	<u>40</u>	<u>125</u>
<u>Samples/peak</u>	<u>10</u>	<u>20</u>
<u>Samples time (ms)</u>	<u>10</u>	<u>10</u>
<u>Runs</u>	<u>3</u>	<u>3</u>
<u>Passes</u>	<u>10</u>	<u>10</u>

Supplemental Table S2. The concentration of particulate metals (without oxalate wash) in seawater (nM) in the different mesocosm treatments (LC: ambient CO₂; HC: increased CO₂ (900 natm); -DFB: no DFB addition; +DFB: with a 70 nM DFB addition) during the development of a bloom of *Emiliania huxleyi* (especially in treatment LC+DFB). Data are means of measurements in 3 independent mesocosms (n = 3) except for LC+DFB where n = 2. Error bars indicate SD. Data is plotted in Figure 3. Note that the concentrations of Co, Pb and Cd were much lower than the rest of the metals, so their values were multiplied by either (10) or (100) in the table (e.g. Co, Cd, and Pb concentrations on d12 in LC+DFB were 0.01, 0.0028 and 0.025 nmol L⁻¹, respectively).

Treatment	Al	Fe	P	Fe	Cu	Co (x10)	Zn	Cd (x100)	Mn	Mg	Pb (x10)
d12											
LC-DFB	9.16(1.16)	1.30(0.72)	131.8(27.05)	13.5(0.88)	0.24(0.01)	0.10(0.06)	3.24(0.15)	0.28(0.06)	0.24(0.06)	0.08(0.01)	0.25(0.01)
LC+DFB	29.2(6.00)	3.16(0.52)	329.3(107.8)	14.8(1.78)	0.30(0.01)	0.22(0.05)	14.81(2.69)	0.91(0.22)	0.54(0.14)	0.09(0.02)	0.56(0.05)
HC-DFB	11.0(7.04)	1.23(0.54)	120.1(45.49)	2.29(0.41)	0.32(0.09)	0.07(0.01)	3.13(0.55)	0.26(0.21)	0.17(0.06)	0.04(0.02)	0.12(0.05)
HC+DFB	18.1(8.53)	2.28(0.53)	193.7(66.43)	11.7(4.43)	0.29(0.05)	0.11(0.07)	4.48(0.38)	0.23(0.03)	0.29(0.13)	0.07(0.01)	0.85(0.51)
d17											
LC-DFB	27.1(14.83)	0.27(0.14)	171.6(20.1)	17.1(8.08)	0.10(0.04)	0.07(0.00)	2.87(1.23)	0.45(0.32)	0.20(0.04)	0.08(0.05)	0.28(0.11)
LC+DFB	29.2(19.2)	4.63(2.84)	972.8(563)	12.7(9.14)	1.02(0.50)	0.68(0.42)	62.7(38.2)	2.38(0.87)	2.36(1.49)	0.37(0.08)	0.77(0.41)
HC-DFB	5.94(4.38)	0.59(0.34)	134.1(42.7)	1.98(0.76)	0.13(0.07)	0.05(0.02)	2.53(0.49)	0.19(0.03)	0.14(0.04)	0.06(0.02)	0.14(0.05)
HC+DFB	35.4(17.9)	4.11(1.86)	322.7(253)	9.34(7.29)	0.50(0.06)	0.19(0.02)	5.88(3.78)	0.98(0.65)	0.56(0.42)	0.09(0.06)	1.42(0.37)
d21											
LC-DFB	19.2(1.01)	2.95(0.06)	241.9(20.1)	2.83(1.81)	0.48(0.02)	0.35(0.03)	15.5(0.97)	1.13(0.26)	0.66(0.06)	0.10(0.02)	2.07(0.26)
LC+DFB	9.18(5.35)	1.53(0.55)	380.9(45.3)	2.52(0.35)	0.44(0.06)	0.37(0.07)	26.2(2.96)	1.41(0.25)	0.88(0.09)	0.20(0.05)	1.23(0.21)
HC+DFB	2.64(1.58)	0.49(0.40)	95.9(12.5)	0.53(0.32)	0.15(0.06)	0.09(0.04)	3.24(1.96)	0.20(0.16)	0.14(0.05)	0.05(0.01)	0.19(0.05)

HC-SPH 8.22(2.05) 0.87(0.20) 134.7(22.1) 3.19(1.21) 0.26(0.05) 0.12(0.02) 3.47(0.97) 0.27(0.11) 0.22(0.08) 0.08(0.01) 0.58(0.13)



1 **Supplemental Table S3.** The concentration of particulate metals (with oxalate wash) in seawater (nmol L⁻¹) in the
 2 different mesocosm treatments; (LC: ambient CO₂; HC: increased CO₂ (900 μatm); -DFB: no DFB addition; +DFB:
 3 with a 70 nM DFB addition) during the development of a bloom of *Emiliania huxleyi* (especially in treatment
 4 LC+DFB). Data are means of measurements in 3 independent mesocosms (n = 3) except for LC-DFB where n = 2.
 5 Error bars indicate SD. Note that the concentrations of Co, Pb and Cd were much lower than the rest of the metals, so
 6 their values were multiplied by either (10) or (100) in the table (e.g. Co, Cd, and Pb concentrations on d12 in LC-DFB
 7 were 0.007, 0.0009 and 0.01 nmol L⁻¹, respectively). The percentage (%) indicates the mean quantity of metal
 8 remaining after the oxalate wash. Statistically significant differences are indicated with asterisk (* if p < 0.05; ** if p
 9 < 0.01 and *** if p < 0.001; ns: not significant).

Treatment	Al	Ti	P	Fe	Cu	Co (-10)	Zn	Cd (-100)	Mn	Mo	Pb (-10)
<i>d12</i>											
LC-DFB	11.6 (2.8)	1.32 (0.34)	117 (3.27)	12.52 (0.78)	0.16 (0.03)	0.07 (0.00)	1.92 (0.86)	0.09 (0.06)	0.15 (0.02)	0.02 (0.00)	0.10 (0.00)
LC+DFB	28.3 (12)	4.49 (1.91)	258 (46.1)	14.67 (3.35)	0.23 (0.08)	0.19 (0.00)	7.16 (1.29)	0.51 (0.14)	0.41 (0.06)	0.03 (0.01)	0.20 (0.11)
HC+DFB	15.9 (2.3)	2.52 (0.66)	139 (14.2)	8.05 (1.08)	0.22 (0.06)	0.09 (0.01)	2.39 (0.93)	0.20 (0.09)	0.21 (0.07)	0.03 (0.01)	0.11 (0.06)
HC-DFB	11.6 (8.8)	1.66 (0.68)	178 (66.3)	9.79 (3.75)	0.19 (0.08)	0.08 (0.03)	2.84 (0.52)	0.22 (0.06)	0.28 (0.08)	0.02 (0.01)	0.19 (0.08)
<i>d17</i>											
LC-DFB	6.42 (2.9)	0.85 (0.35)	97 (41.6)	1.23 (0.56)	0.11 (0.07)	0.09 (0.05)	2.86 (1.45)	0.26 (0.09)	0.18 (0.07)	0.03 (0.00)	0.09 (0.05)
LC+DFB	7.53 (4.7)	1.85 (0.63)	245 (136)	1.28 (0.68)	0.24 (0.11)	0.22 (0.08)	12.1 (3.78)	1.20 (0.69)	0.54 (0.29)	0.05 (0.03)	0.18 (0.09)
HC+DFB	4.48 (0.2)	1.29 (0.01)	131 (5.31)	1.55 (0.19)	0.14 (0.01)	0.07 (0.00)	3.03 (0.90)	0.21 (0.06)	0.14 (0.02)	0.03 (0.00)	0.10 (0.05)
HC-DFB	12.8 (2.7)	1.98 (0.74)	233 (162)	5.31 (0.99)	0.29 (0.11)	0.18 (0.06)	5.03 (3.06)	0.35 (0.13)	0.43 (0.33)	0.06 (0.01)	
<i>d21</i>											
LC-DFB	13.9 (3.2)	1.54 (0.48)	257 (20.9)	3.76 (0.75)	0.29 (0.06)	0.26 (0.01)	8.59 (0.69)	0.74 (0.31)	0.35 (0.04)	0.05 (0.02)	
LC+DFB	4.36 (0.4)	1.01 (0.41)	253 (47.6)	2.04 (0.63)	0.23 (0.02)	0.20 (0.01)	14.3 (1.32)	0.67 (0.09)	0.43 (0.05)	0.05 (0.01)	
HC+DFB	2.49 (0.9)	0.62 (0.17)	79 (19.6)	0.33 (0.07)	0.11 (0.02)	0.07 (0.03)	2.36 (1.38)	0.09 (0.06)	0.09 (0.03)	0.01 (0.00)	
HC-DFB	2.56 (1.2)	0.98 (0.30)	74 (20.7)	1.03 (0.18)	0.12 (0.03)	0.05 (0.01)	1.01 (0.35)	0.05 (0.02)	0.07 (0.01)	0.02 (0.00)	
%	ns	ns	80*	75*	60*	70*	55**	45***	55**	35***	

Movido hacia abajo[1]:
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Supplemental Table

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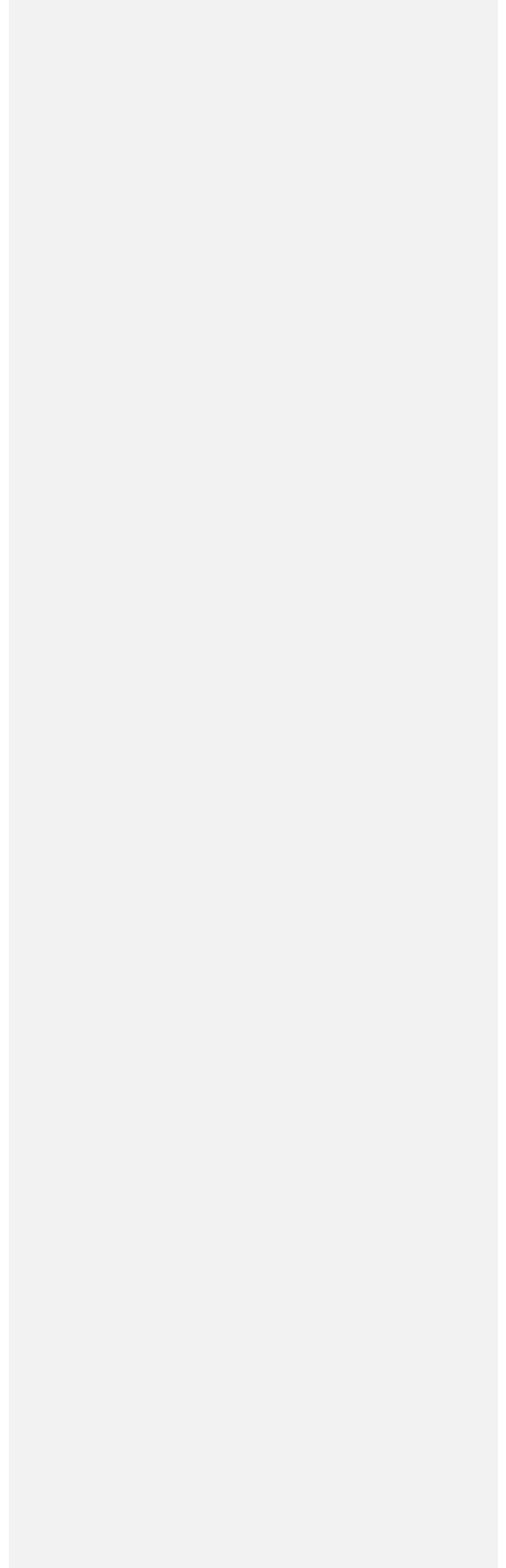
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Página 5: [6] Celdas eliminadas **María Segovia** **11/6/19 20:05:00**

Celdas eliminadas

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Página 5: [6] Celdas eliminadas **María Segovia** **11/6/19 20:05:00**

Celdas eliminadas

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Página 5: [7] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto

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Página 5: [7] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto

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Página 5: [7] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto

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Página 5: [7] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto

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Página 5: [7] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto

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Página 5: [7] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto

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Página 5: [7] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto

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Página 5: [7] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto



Página 5: [7] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto



Página 5: [7] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto



Página 5: [8] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto



Página 5: [8] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto



Página 5: [8] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto



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Fuente: 10 pto



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Fuente: 10 pto



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Fuente: 10 pto



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Fuente: 10 pto



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Fuente: 10 pto



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Fuente: 10 pto



Página 5: [8] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto



Página 5: [8] Con formato	Maria Segovia	11/6/19 20:05:00
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Fuente: 10 pto



Página 5: [8] Con formato	Maria Segovia	11/6/19 20:05:00
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Fuente: 10 pto



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Fuente: 10 pto



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Fuente: 10 pto



Página 5: [8] Con formato	María Segovia	11/6/19 20:05:00
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Fuente: 10 pto



Página 5: [9] Con formato	María Segovia	11/6/19 20:05:00
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Fuente: 10 pto, Sin Cursiva



Página 5: [9] Con formato	María Segovia	11/6/19 20:05:00
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Fuente: 10 pto, Sin Cursiva



Página 5: [10] Celdas eliminadas	María Segovia	11/6/19 20:05:00
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Celdas eliminadas



Página 5: [10] Celdas eliminadas	María Segovia	11/6/19 20:05:00
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Celdas eliminadas



Página 5: [10] Celdas eliminadas	María Segovia	11/6/19 20:05:00
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Celdas eliminadas



Página 5: [10] Celdas eliminadas	María Segovia	11/6/19 20:05:00
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Celdas eliminadas



Página 5: [10] Celdas eliminadas	María Segovia	11/6/19 20:05:00
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Celdas eliminadas



Página 5: [10] Celdas eliminadas	María Segovia	11/6/19 20:05:00
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Celdas eliminadas



Página 5: [10] Celdas eliminadas	María Segovia	11/6/19 20:05:00
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Celdas eliminadas



Página 5: [10] Celdas eliminadas **María Segovia** **11/6/19 20:05:00**

Celdas eliminadas

Página 5: [10] Celdas eliminadas **María Segovia** **11/6/19 20:05:00**

Celdas eliminadas

Página 5: [10] Celdas eliminadas **María Segovia** **11/6/19 20:05:00**

Celdas eliminadas

Página 5: [10] Celdas eliminadas **María Segovia** **11/6/19 20:05:00**

Celdas eliminadas

Página 5: [11] Con formato **María Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

Página 5: [11] Con formato **María Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

Página 5: [11] Con formato **María Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

Página 5: [11] Con formato **María Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

Página 5: [11] Con formato **María Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

Página 5: [11] Con formato **María Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

Página 5: [11] Con formato **María Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

Página 5: [11] Con formato **María Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

Página 5: [11] Con formato **María Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

Página 5: [11] Con formato **María Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro



Página 5: [11] Con formato **Maria Segovia** **11/6/19 20:05:00**

Fuente: 10 pto, Sin Cursiva, Color de fuente: Negro

