

RESPONSES TO REVIEWERS COMMENTS FOR BG-2018-448 by Lorenzo et al.

Referee #1

The authors have done an acceptable job in addressing my comments as well as those of other reviewers. Therefore, I would recommend publication.

Some minor comments.

Lines 62-65. “For instance, Fe occurs...more bioavailable.”: first, references are needed for this sentence; second, what described here actually cannot serve as examples for the dependence of trace metal chemistry on seawater pH.

We removed the clause about Fe II being more bioavailable, and added that Fe II is more soluble but rapidly oxidize according to Millero et al 1987: *“Iron occurs in two main redox states in the environment: oxidized ferric Fe (Fe (III)), which is poorly soluble at circumneutral pH; and reduced ferrous Fe (Fe (II)), which is more soluble in natural seawater, but becomes rapidly oxidized (Millero et al. 1987) “*

Line 84. “, and that are actively...” should be “, and that they are actively...”

Corrected

Line 85. the impact on, not in, the trace metal content...

Corrected

Line 97. “E. huxleyi”...please also check the rest of the manuscript

Checked

Line 100. 1) Delete “organic”, as CaCO₃ is not organic, 2) delete “carbon export”, and 3) Hutchings, 2011 is not in the reference list.

Done

Line 104. What “realistic” refers to here?

We have clarified this point, it now reads as: *“Specifically, mesocosm experiments allow perturbation studies with a high degree of realism compared to other experimental systems such as in the laboratory (high controlled conditions usually far from reality) or in situ in the ocean (where not all the interactions are contemplated) (Riebesell et al., 2010, Stewart et al., 2013, Riebesell and Gattuso, 2015)”*

Line 123. Replace “iron” with Fe, and also check the rest of the manuscript.

Done

Line 127. Should be “resulting in...”

Done

Line 135. Please provide the vendor of HCl in the first place.

It was said in Ln 200: Fisher chemicals. We have moved this to the line above.

Line 252. The effects of...on, not in, the plankton community

Done

Line 393. E. huxleyi.

Done, now Ln 403

Lines 401-402. At low pH, CO₃ concentration decreases, which should lead to a decrease in CuCO₃.

Ok

Although the diatom bloom is not the primary focus of this study, I would suggest the authors briefly discuss how the change in CO₂ may affect the biogenic fraction of particulate Co, Zn, and Cd at the beginning of the mesocosm experiment.

We do not agree about talking about diatoms. The paper is already pretty complex. And we do not want to speculate on diatoms, they were all gone (by day 6-7) before trace metal dynamics started to be affected. This was an Ehux bloom and so this is the main focus.

Referee #2

“Particulate trace metal dynamics in response to increased CO₂ and iron availability in a coastal mesocosm experiment”

Lorenzo et al.

- General comments

Authors have responded to most of my previous comments.

I have found significant improving of this manuscript, especially for presenting their results with adequate “Figures” which is more understandable for readers. However, I still found some issues for this manuscript which have to concern before publications. Please consider for following comments.

Since authors present their data with some “Figures”, it became more clearly that parts of presentation in this manuscript (MS) are overlapped to previous published paper, Segovia et al. (2017). For instance, Figure 1a, 1b and 2 are totally same figures as reported by Segovia et al. (2017). Is it really possible to publish only with the statement as “Figure reproduced with permission from Segovia et al. Mar. Ecol. Prog. Ser. 2017” in the caption? It might be better to place in “supplementary Figures”. I would like to ask about this aspect to Editor.

The copyright is all right. We have transferred them to supplemental material and re-numbered all figures. The partition coefficients that were in Suppl. Figs, is now Fig 4 in the main text.

Also, authors should make clear in MS that “what is original purpose of this MS” in introduction, “which is original findings from this MS” in “Abstract”. Original findings of this MS are not clear in the “Abstract”(see below).

See answer above please, in the abstract section.

Table 1 showed the result from statistically analyses of the particulate(P) trace metal (TM) data obtained from this study. This statistically analyses are one of the important points for discussion in the MS. However, the detail for the statistically analyses are not clear for reader. For instance, which “day’s” data did authors used for evaluating the difference among treatments for each P-TM (d12, d17, d21??)? Which day’s data did authors used for evaluating the time difference (is this means significant difference between d12 and d21??)? Please explain more clearly in the text (2.4 section) and in the caption of Table 1. Same for Table 3.

We used all the days because we performed a Split-Plot ANOVA (or mixed model) which integrates fixed factors (Co₂ and Fe) and a repeated measures factor (time) by using the post-hoc Bonferroni, thus, saying that the statistical treatment was a split-plot ANOVA+Bonferroni, compulsory means that time was fully considered during the whole experimental period. We have revised the text though see if this is unclear, and we believe it is not.

- Abstract

Line 30-38; From “Future predicted high..... future ocean dynamics.”. This sentence should be supported by results of this study which is described in the one sentence before. Author should state here “what is the original important result and major findings from this study”. Present sentence is not clear in this point. I think, main important results and findings from this study, which should be claimed in abstract, are described in line 376-404, and “conclusion”. Authors should not include speculation in the “Abstract”. The findings should strongly be supported by this study’s own results.

We rewrote the abstract removing most speculations. We tried to be very careful in the abstract as well as in the discussion and conclusions when we talked about particulate metal concentrations (mol L⁻¹) and ratios (Me:P). The abstract has then been modified as follows: *“Rising concentrations of atmospheric carbon dioxide are causing ocean acidification and will influence marine processes and trace metal biogeochemistry. In June 2012, in Raunefjord (Bergen, Norway) we performed a mesocosm experiment, comprised of a fully factorial design of ambient and elevated pCO₂ and/or an addition of the siderophore desferrioxamine B (DFB). In addition, the macronutrient concentrations were manipulated to enhance a bloom of the coccolithophore *Emiliania huxleyi*. We report here the changes in particulate trace metal concentrations during this experiment. Our results show that particulate Ti and Fe were dominated by lithogenic material while particulate Cu, Co, Mn, Zn, Mo and Cd had a strong biogenic component. Furthermore, significant correlations were found between particulate concentrations (mol L⁻¹) of Cu, Co, Zn, Cd, Mn, Mo, and P in seawater and phytoplankton biomass (µgC L⁻¹), supporting a significant influence of the*

bloom in the distribution of these particulate elements. The concentrations of these biogenic metals (mol L⁻¹) in the E. huxleyi bloom were ranked as: Zn > Cu ≈ Mn > Mo > Co > Cd. Changes in CO₂ affected total particulate concentrations (mol L⁻¹) and biogenic metal ratios (Me:P) for some metals, while the addition of DFB only affected significantly the concentrations of some particulate metals (mol L⁻¹). Variations in CO₂ had the most clear, and significant effect on particulate Fe concentrations (mol L⁻¹), decreasing its concentration under high CO₂. Indeed, high CO₂ and/or DFB promoted the dissolution of particulate Fe, and the presence of this siderophore helped maintaining high dissolved Fe. This shift between particulate and dissolved Fe concentrations, in the presence of DFB, promoted a massive bloom of E. huxleyi in the treatments with ambient CO₂. Furthermore, high CO₂ decreased the Me:P ratios of Co, Zn and Mn, while increased the Cu:P ratios. These findings support theoretical predictions that the Me:P ratios of metals whose seawater dissolved speciation is dominated by free ions (e.g. Co, Zn and Mn) will likely decrease or stay constant under ocean acidification. In contrast, high CO₂ is predicated to shift the speciation of dissolved metals associated with carbonates, such as Cu, increasing their bioavailability, and resulting in higher Me:P ratios”

- Introduction

Line 74-86; I found that some parts of contents in this sentence are overlapped to previous sentence (line 57-71). To make more concise introduction, please compile and reconstruct these two sentences and avoid repetition.

We have made this more concise and focused and removed a couple of sentences.

Line 95-105; This sentence is very important for explaining the aim of this MS. In the present description, authors explain “importance of E. huxleyi bloom for biogeochemistry in the ocean”. This is the research motivation for study of Segovia et al. (2017), but this is not motivation for investigating particulate trace metals in this MS. Whereas, Authors only describe the aim of this study as “The aim of the present study was CO₂ and Fe bioavailability.” in line 104-105. I think author should add more explanation about “why the measurement of particle trace metals and characterize the change of particle trace metals during E. huxleyi bloom are important?”.

Thanks for this insightful comment. We have clarified the text as follows: “*In the present work a bloom of the coccolithophorid *Emiliana huxleyi* was induced in a mesocosm experiment in a Norwegian fjord, where the speciation of particulate and dissolved trace metals is very dynamic (e.g. Fe; Ozturk et al. 2002). We aimed to examine and characterize the change of particle trace metals during the E. huxleyi bloom under the interactive effects of increased CO₂ and/or dissolved Fe. *Emiliana huxleyi* is the most cosmopolitan and abundant coccolithophore in the modern ocean (Paasche, 2002) and its growth and physiology has been studied under these experimental conditions (Segovia et al., 2017, Segovia et al., 2018, Lorenzo et al., 2018). Furthermore, E. huxleyi has unique trace metal requirements relative to other abundant phytoplankton taxa (ie. diatoms or dinoflagellates; Ho et al. 2003). Coccolithophores play a key role in the global carbon cycle because they produce photosynthetically organic carbon, as well as particulate inorganic carbon through calcification. These two processes foster the sinking of particulate organic carbon—and trace metals—and contribute to deep ocean carbon export (Hutchings, 2011) and ultimately*

to organic carbon burial in marine sediments (Archer, 1991, Archer and Maier-Reimer., 1994). However, ocean acidification will disproportionally affect the abundance of coccolithophores, as well as their rates of calcification and organic carbon fixation (Zondervan et al., 2007). The aim of the present study was to characterize the changes in particulate trace metal concentrations—in both lithogenic and biogenic particles—during a bloom of E. huxleyi under realistic changes in CO₂ and Fe bioavailability expected by 2100.”

Line 97; I could not find “Segovia et al., 2018” and “Lorenzo et al., 2018” in the reference list in end of MS. Please check all reference in the text and reference list in the MS.

Done

- Material and method

Line 114, Authors described that CO₂ concentration in the mesocosms were measured by NDIR analysis system. Whereas, in line 116-117, authors described that CO₂ concentrations in the mesocosms were calculated from pH and alkalinity. Which is correct? According to Segovia et al.2017, “CO₂ concentration in inlet flow” is only measured by NDIR.

Both are correct. The CO₂ concentration in the inlet airflow was measured with an IRGA, and the C speciation in seawater was checked by modelling by using the CO₂Calc package from alk and pH according to Segovia et al. 2017.

Line 121, Why the DFB concentration was set to 70 nM? This value is important for this study because particle dissolution for some trace metal was strongly influenced by the DFB concentrations as authors described in discussion.

The rational of adding 10-20 times higher DFB concentrations relative to the dFe is very well explained in Marchetti and Maldonado 2016. We have added this reference.

Line 128, “gentle vacuum pumping”. Is this pumping system clean for level of trace metal work? Detailed description is needed for explaining sampling system, cleaning procedure, because these are critical part of trace metal work.

This has been included as follows in lns217-224: *“All equipment and sampling material used during this study was rigorously acid-washed under trace metal clean conditions and protocols according to GEOTRACES. The material was cleaned with Milli-Q water (MQw) with 10 % Extran (Fisher Chemicals) at 60°C for 6h, followed by 3 thorough rinses with MilliQ water at room T. The material was then cleaned with 10 % HPLC grade HCl (Sigma-Aldrich) at 60°C or 12h and then rinsed thoroughly 5 times with MQw at room T. The material was then covered by plastic and transported to the raft. Sampling in the raft was carried out under a mobile plastic cover hood. Filters were precleaned with 10% trace metal grade hydrochloric acid (Seastar, Fisher Chemicals), at 60°C overnight and were rinsed with MQw”.*

Line 139, How did authors collect the seawater (by using pumping as described in line 128?).

Yes

“0.2 uM” should be change to “0.2um”.

Done

Line 147; “trace metal hydrochloric acid” should be “trace metal grade hydrochloric acid”.

Done

Line 154; “0.2 uM” should be change to “0.2um”.

Done

Line 156; Is this “centrifuge tubes” same tube as “2mL PP” tubes?? Reader will be confusing.

No. They are 2 mL polypropylene tubes. It has been clarified in Ln 232.

Line 225-249; Section 3.2, 3.3 and 3.4. Authors need to explain detail about statistically analysis for Table 1 and Table 3, see general comment.

Please see answer above: We used all the days because we performed a Split-Plot ANOVA (or mixed model) which integrates fixed factors (Co₂ and Fe) and a repeated measures factor (time) by using the post-hoc Bonferroni, thus, saying that the statistical treatment was a split-plot ANOVA+Bonferroni, compulsory means that time was fully considered during the whole experimental period. We have revised the text though see if this is unclear, and we believe it is not.

- Discussion

Line 286 “well-known metal content in biogenic particles” and line 287 “normalized to Al (mol Me: mol Al in the Earth crust)”. Please indicate the number which was used for this calculation and indicate reference for the number.

We added the appropriate reference in a parenthesis. Also, we believe these are in the legends. We can't add all the ratios in a parenthesis, they too many.

Line 289; “0.0051 for mol Fe:molP”, “0.331 mol Fe : mol Al”. How did authors select this number? Which information did author refer? Maybe, “Ho et al., 2006” in Table 2 (A) for “0.0051” and Table 2 (B) for “0.331”. Table 2 (B) crustal ratios need reference (Taylor 1964?). Add ref. to “Table 2 (B)”.

The Me:P are those of marine plankton (Ho 2006). We added this to the text.

Line 321; I could not find “Table 5” in MS.

Table 5 does not exist anymore, is Table 4. This has been corrected.

Line 347; “Zn and Cu dissolution (ref.)”. Please add reference.

Added

Line 353-355; “Rain events..... (Muller et al., 2005)”. Do authors have some information on rain event before or during the experiment? If they have, it is important additional information for this interpretation.

We have added a comment saying that this may be of importance due to the rainy nature of this geographical location

Line 355-358; “, and indigenous plankton.....Knauer et al., 1998).”. This sentence is too speculative, and not main discussion for particulate Cu:P ratio. I feel author can delete this sentence.

Deleted

Line 362, 363; I can not find “Figure 6” in the MS.

Fig 6 does not exist anymore. It was a mistake.

Line 376; “in that it was the only trace element whose particulate concentration significantly and uniquely affected.....”. I think this is not true. According to Table 1, other metals (Co, Zn, Cd Mn Mo) were also affected by CO₂.

What we said is true. Particulate Fe (mol L⁻¹) was the only one with a clear CO₂ effect, without any interaction with DFB.

Line 384-387; Authors discussed about partitioning of DFe and PFe with using Figure S2. I think this results and discussion are one of very important finding of this MS. This result is base of predicting future ocean in the following discussion. I recommend that Figure S2 should be placed for the normal Figures (not in Supplementary).

We thank reviewer for this suggestion. We have included the figure in the main text now.

Line 396; I could not find Table 5 in the MS.

Table 5 does not exist anymore. It was a mistake; it is Table 4.

Line 424; “due to increased Fe bioavailability”. How authors judged the increasing Fe bioavailability in the LC+DFB treatment? Is there any evidence which indicate that DFB-bounded Fe is available for *E. huxleyi*? Please discussed more carefully about the changing Fe bioavailability in “discussion”.

We removed the bioavailability and avoided any speculation.

We thank the reviewers for their constructive comments and their time, and we hope that our responses are satisfactory

Yours sincerely,

Maria Segovia and Maite Maldonado

1 **Particulate trace metal dynamics in response to increased CO₂ and iron availability in a coastal mesocosm**
2 **experiment**

3
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13
14 **Abstract.** Rising concentrations of atmospheric carbon dioxide are causing ocean acidification and will influence
15 marine processes and trace metal biogeochemistry. In June 2012, in Raunefjord (Bergen, Norway) we performed a
16 mesocosm experiment, comprised of a fully factorial design of ambient and elevated pCO₂ and/or an addition of the
17 siderophore desferrioxamine B (DFB). In addition, the macronutrient concentrations were manipulated to enhance a
18 bloom of the coccolithophore *Emiliania huxleyi*. We report here the changes in particulate trace metal concentrations
19 during this experiment. Our results show that particulate Ti and Fe were dominated by lithogenic material while
20 particulate Cu, Co, Mn, Zn, Mo and Cd had a strong biogenic component. Furthermore, significant correlations were
21 found between particulate concentrations (mol L⁻¹) of Cu, Co, Zn, Cd, Mn, Mo, and P in seawater and phytoplankton
22 biomass (µgC L⁻¹), supporting a significant influence of the bloom in the distribution of these particulate elements. The
23 concentrations of these biogenic metals (mol L⁻¹) in the *E. huxleyi* bloom were ranked as: Zn > Cu ≈ Mn > Mo > Co >
24 Cd. Changes in CO₂ affected total particulate concentrations (mol L⁻¹) and biogenic metal ratios (Me:P) for some
25 metals, while the addition of DFB only affected significantly the concentrations of some particulate metals (mol L⁻¹).
26 Variations in CO₂ had the most clear, and significant effect on particulate Fe concentrations (mol L⁻¹), decreasing its
27 concentration under high CO₂. Indeed, high CO₂ and/or DFB promoted the dissolution of particulate Fe, and the
28 presence of this siderophore helped maintaining high dissolved Fe. This shift between particulate and dissolved Fe
29 concentrations, in the presence of DFB, promoted a massive bloom of *E. huxleyi* in the treatments with ambient CO₂.
30 Furthermore, high CO₂ decreased the Me:P ratios of Co, Zn and Mn, while increased the Cu:P ratios. These findings
31 support theoretical predictions that the Me:P ratios of metals whose seawater dissolved speciation is dominated by free
32 ions (e.g. Co, Zn and Mn) will likely decrease or stay constant under ocean acidification. In contrast, high CO₂ is
33 predicated to shift the speciation of dissolved metals associated with carbonates, such as Cu, increasing their
34 bioavailability, and resulting in higher Me:P ratios.

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37 **Key words:** Global change, Fe, CO₂, particulate trace metals, dissolved trace metals, mesocosms, *Emiliania huxleyi*,
38 phytoplankton

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44 1. Introduction

45 Marine phytoplankton contribute half of the world's total primary productivity, sustaining marine food webs and
46 driving the biogeochemical cycles of carbon and nutrients (Field et al., 1998). Annually, phytoplankton incorporate
47 approximately 45 to 50 billion metric tons of inorganic carbon (Field et al., 1998), removing a quarter of the CO₂
48 emitted to the atmosphere by anthropogenic activities (Canadell et al., 2007). Yet, the atmospheric CO₂ concentration
49 has increased by 40 % since pre-industrial times as a result of anthropogenic CO₂ emissions, producing rapid changes
50 in the global climate system (Stocker et al., 2013). The dissolution of anthropogenic CO₂ in seawater, causes shifts in
51 the carbonate chemical speciation, and leads to ocean acidification (OA). Marine ecosystems are sensitive to changes in
52 pH because pH strongly affects chemical and physiological reactions (Hoffman et al., 2012). Increased CO₂ in seawater
53 may enhance or diminish phytoplankton productivity (Mackey et al., 2015), decrease the CaCO₃ production in most
54 planktonic calcifiers (Riebesell and Tortell 2011), and/or inhibit organic nitrogen and phosphorus acquisition (Hutchins
55 et al., 2009). Thus, the biogeochemical cycling of nutrients is predicted to be highly affected by OA (Hutchins et al.
56 2009), as well as the distribution and speciation of trace metals in the ocean (Millero et al., 2009).

58 Trace metals, including Fe, Zn, Mn, Cu, Co and Mo, are essential for biological functions (e.g. photosynthesis,
59 respiration and macronutrient assimilation), and Cd can supplement these functions. Trace metals availability can
60 influence phytoplankton growth and community structure (Morel and Price, 2003). In turn, plankton control the
61 distribution, chemical speciation, and cycling of trace metals in the sea (Sunda, 2012), by, for example, releasing
62 organic compounds that dominate the coordination chemistry of metals, internalizing trace elements into the cells, and
63 reducing and/or oxidizing metals at the cell surface. The chemistry of redox speciation of active trace metals is highly
64 dependent on pH. Fe occurs in two main redox states in the environment: oxidized ferric Fe (Fe (III)), which is poorly
65 soluble at circumneutral pH; and reduced ferrous Fe (Fe (II)), which is more soluble in natural seawater, but becomes
66 rapidly oxidized (Millero et al. 1987). Fe speciation and bio-availability are dynamically controlled by the prevalent
67 changing redox conditions. Also, as the ocean becomes more acidic, reduction of Cu (II) will increase, as the ionic form
68 of Cu (II) is reduced to Cu (I) (Millero et al., 2009). The effect of higher concentrations of Cu (I) in surface waters on
69 biological systems is not well known. Therefore, while the effects of OA on inorganic metal speciation will be more
70 pronounced for metals that form strong complexes with carbonates (e.g. copper) or hydroxides (e.g. Fe and aluminium),
71 those that form stable complexes with chlorides (e.g. cadmium) will not be greatly affected. pH mediated changes in
72 concentrations and/or speciation could possibly enhance trace metals limitation and/or toxicity to marine plankton
73 (Millero et al., 2009).

75 Fe is crucial for phytoplankton growth because it is involved in many essential physiological processes, such as
76 photosynthesis, respiration, and nitrate assimilation (Behrenfeld and Milligan, 2013). The decrease in seawater pH in
77 response to OA may increase Fe solubility (Millero et al., 2009), but it may also result in unchanged or lower Fe
78 bioavailability, depending on the nature of the strong organic Fe ligands (Shi et al., 2010). Consequently, changes in Fe
79 bioavailability due to ocean acidification can affect positively or negatively ocean productivity and CO₂ drawdown.
80 Copper is an essential micronutrient but may be toxic at high concentrations (Semeniuk et al., 2016). An increase in
81 free cupric ion concentrations in coastal areas due to ocean acidification (Millero et al., 2009) could result in negative
82 effects on phytoplankton. Given that trace metals are essential for phytoplankton productivity, and that they are
83 actively internalized during growth, it is important to study the impacts of ocean acidification on the trace metal content
84 of ecologically significant plankton species.

86 In a rapidly changing global environment, generated by anthropogenic CO₂ emissions, it is critical to gain adequate

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110 understanding about ecosystem responses. Due to the complex interactions in aquatic ecosystems, such predictions have
 111 so far not been possible to do based upon observational data and modelling alone. However, direct empirical studies on
 112 natural communities offer a robust tool to analyse interactive effects of multiple stressors. Specifically, mesocosm
 113 experiments allow perturbation studies with a high degree of realism compared to other experimental systems **such as in**
 114 **the laboratory (high controlled conditions usually far from reality) or *in situ* in the ocean (where not all the interactions**
 115 **are contemplated)** (Riebesell et al., 2010, Stewart et al., 2013, Riebesell and Gatusso, 2015).

117 In the present work a bloom of the coccolithophorid *Emiliania huxleyi* was induced in a mesocosm experiment **in a**
 118 **Norwegian fjord, where the speciation of particulate and dissolved trace metals is very dynamic (e.g. Fe; Ozturk et al.**
 119 **2002). We aimed to examine and characterize the change of particle trace metals during an *E. huxleyi* bloom under the**
 120 interactive effects of increased CO₂ and/or dissolved Fe. *Emiliania huxleyi* is the most cosmopolitan and abundant
 121 coccolithophore in the modern ocean (Paasche, 2002) **and its growth and physiology has been studied under this**
 122 **experimental conditions (Segovia et al., 2017, Segovia et al., 2018, Lorenzo et al., 2018). Furthermore, *E. huxleyi* has**
 123 **unique trace metal requirements relative to other abundant phytoplankton taxa (ie. diatoms or dinoflagellates; Ho et al.**
 124 **2003).** Coccolithophores play a key role in the global carbon cycle because they produce photosynthetically organic
 125 carbon, as well as particulate inorganic carbon through calcification. These two processes foster the sinking of
 126 particulate organic carbon **—and trace metals—and contribute to deep ocean carbon export (Hutchings, 2011) and**
 127 **ultimately to organic carbon burial in marine sediments (Archer, 1991, Archer and Maier-Reimer., 1994). However,**
 128 ocean acidification will disproportionately affect the abundance of coccolithophores, as well as their rates of
 129 calcification and organic carbon fixation (Zondervan et al., 2007). The aim of the present study was to characterize the
 130 changes in particulate trace metal concentrations **—in both lithogenic and biogenic particles—during a bloom of *E.***
 131 ***huxleyi* under realistic changes in CO₂ and Fe bioavailability, expected by 2100.**

134 2. Materials and methods

135 2.1 Experimental set-up

136 The experimental work was carried out in June 2012 in the Raunefjord, off Bergen, Norway as described in detail by
 137 Segovia et al., (2017). Twelve mesocosms (11 m³ each) were set-up in a fully factorial design with all combinations of
 138 ambient and elevated pCO₂ and dFe in three independent replicate mesocosms. The mesocosms were covered by lids
 139 (both transparent to PAR and UVR) and filled with fjord water from 8 m depth. We achieved two CO₂ levels
 140 corresponding to present (390 ppm, LC) and those predicted for 2100 (900 ppm, HC) by adding different quantities of
 141 pure CO₂ gas (Shculz et al., 2009). The specific CO₂ concentration and the CO₂ inlet flows in the mesocosms were
 142 measured by non-dispersive infrared analysis by using a Li-Cor (LI-820) CO₂ gas analyser (Li-COR, Nebraska, USA).
 143 CO₂ concentrations in the mesocosms were calculated from pH and total alkalinity measurements using the CO₂ SYS
 144 software (Robbins et al., 2010). At the beginning of the experiment, nitrate (10 µM final concentration) and phosphate
 145 (0.3 µM final concentration) were added to induce a bloom of the coccolithophore *Emiliania huxleyi*, **according to Egge**
 146 **& Heimdahl (1994). Following recommendations by Marchetti and Maldonado (2016), to induce changes in Fe**
 147 **availability, and analyse its effects on the plankton community, 70 nM (final concentration) of the siderophore**
 148 **desferrioxamine B (DFB) (+DFB and -DFB treatments) (Figure S1b-supplemental material) was added to half of the**
 149 **mesocosms on Day 7, when the community was already acclimated to high CO₂. The initial dFe concentration before**
 150 **DFB addition was about 4.5 nM. Even though DFB is a strong Fe-binding organic ligand and often used to induce Fe**
 151 **limitation in phytoplankton (Wells 1999), DFB additions may also increase the dissolved Fe pool in environments**
 152 **with high concentrations of colloidal and/or particulate Fe, such as fjords (Kuma et al. 1996, Öztürk et al. 2002). By day**

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198 17, dissolved Fe concentrations were significantly higher (by ~3-fold) in the high CO₂ and DFB treatments than in the
199 control (Segovia et al. 2017). These results support an increase in the solubility of Fe in seawater by either lowering its
200 pH (Millero 1998; Millero et al. 2009) and/or the addition of DFB (Chen et al. 2004). The multifactorial experimental
201 design consisted of triplicate mesocosms per treatment and the combinations of high and ambient pCO₂ and dFe levels,
202 resulted in a total of 12 mesocosms: LC-DFB (control), LC+DFB, HC+DFB and HC-DFB. Water samples from each
203 mesocosm were taken from 2 m depth by gentle vacuum pumping of 25 L volume into acid-washed carboys that were
204 quickly transported to the onshore laboratory. The biological and chemical variables analysed were phytoplankton
205 abundance and species composition, dissolved Fe and Cu concentrations (dFe, dCu), nutrient concentrations (nitrate,
206 phosphate, silicic acid and ammonium) and particulate trace metal concentrations. ▲

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208 2.2 Dissolved copper (dCu)

209 Low density polyethylene (LDPE) bottles were cleaned with 1% alkaline soap solution for one week, then filled with 6
210 M trace metal grade HCl (Seastar, Fisher Chemicals) and submerged in a 2 M HCl bath for one month. For transport,
211 they were filled with 1 M trace metal grade HCl for one more month and kept double bagged. In between each acid
212 treatment, the bottles were rinsed with Milli-Q water (Millipore; hereafter referred to as MQ). Before sampling, the
213 bottles were rinsed three times with filtered seawater. Seawater was collected from each mesocosm, filtered through 0.2
214 µm AcroPak Supor membrane capsule filters into the trace metal clean LDPE bottles, and acidified with ultra-clean
215 trace metal grade HCl in a Class 100 laminar flow hood. Total dissolved Cu concentrations were measured following
216 Zamzow et al., (1998) using a flow injection analysis chemiluminescence detection system (CL-FIA, Waterville
217 Analytical). Total dissolved Fe concentrations were measured as described in Segovia et al., (2017) for this very
218 experiment.

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220 2.3 Particulate metals (pMe)

221 2.3.1 Sampling

222 All equipment and sampling material used during this study was rigorously acid-washed under trace metal clean
223 conditions and protocols according to GEOTRACES. The material was cleaned with Milli-Q water (MQw) with 10 %
224 Extran (Fisher Chemicals) at 60°C for 6h, followed by 3 thorough rinses with MQw at room T. The material was then
225 cleaned with 10 % HPLC grade HCl (Sigma-Aldrich) at 60°C or 12h and then rinsed thoroughly 5 times with MQw at
226 room T. The material was then covered by plastic and transported to the raft. Sampling in the raft was carried out under
227 a mobile plastic cover hood. Filters were precleaned with 10% trace metal grade hydrochloric acid (Seastar, Fisher
228 Chemicals), at 60°C overnight and were rinsed with MQw. Seawater samples (1-3.5 L) were filtered gently onto 0.45
229 µm acid washed Supor®-450 filters (within a trace metal clean Swinnex filter holder) on days 12, 17 and 21 of the
230 experiment. Four technical replicates were taken from each mesocosm. Two filters were analysed without manipulation
231 and the other two were individually washed with oxalate-EDTA reagent to remove extracellular Fe, as well as other
232 metals (Tang and Morel, 2006). Immediately following filtration, the treated filters were soaked with 20 mL EDTA-
233 oxalate solution, added to the headspace of the Swinnex holders, with an acid-washed polypropylene syringe. After 10
234 min, vacuum was applied to remove the oxalate solution and 10 mL of 0.2 µm filtered chelexed synthetic oceanic water
235 (SOW) solution was passed through the filter to rinse off any remaining oxalate solution. Replicate filters that were not
236 treated with oxalate solution were transferred directly to centrifuge tubes for storage. The filters with particles were
237 frozen in acid-washed 2 mL polypropylene tubes and then, dried and stored until analysis.

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239 2.3.2 Analytical methods

253 Filters were digested in 7-mL acid-washed Teflon (Teflon, Rochester, NY, USA) vials. Teflon vials were also
254 precleaned using 10% trace metal hydrochloric acid (Fisher, trace metal grade) during two days and then, with nitric
255 acid (Fisher, trace metal grade) at 70 °C during three days. In between each acid treatment, the bottles were rinsed with
256 MQ. Samples were digested in 3 mL of HNO₃ and 0.5 mL of HF (Fisher, trace metal grade) with lids on for 1 h on a
257 hot plate at 200 °C. The lids were then removed to evaporate HF at 200°C. After this, 1.5 mL of HNO₃ were added and
258 the samples were heated with lids on overnight at 150 °C. Finally, 2.25 mL of HClO₄ (Fisher, Optima grade) were
259 added and the samples were heated for 4 h at 200 °C. After complete digestion, the samples were dried on hot plates at
260 200°C. The dried samples were dissolved in 1% nitric acid with 1 ppb in internal standard. The analysis was performed
261 using a high-resolution inductively coupled plasma-mass spectrometer (ICP-MS, Element XR, Thermo Scientific) and
262 the described instrumental settings (Table S1). Filter blanks were collected and subjected to the same storage, digestion,
263 dilution, and analysis processes, and these blank values were subtracted from sample measurements. Particulate
264 samples for ICP-MS analysis were processed in a trace metal-clean laboratory under a trace metal-clean laminar flow
265 fume hood.
266

267 2.3.3 The effect of oxalate-EDTA wash on particulate trace metal concentrations

268 To better estimate the biogenic fraction of the particulate metals, the filters were washed with an oxalate-EDTA
269 solution, which removes extracellular metals and oxyhydroxides (Tovar-Sanchez et al., 2003; Tang and Morel, 2006).
270 In our study, the oxalate wash significantly decreased the concentration of all particulate metals, with the exception of
271 Al and Ti (Tables S2 and S3), as observed by Rauschenberg and Twining (2015). The quantity of metal remaining after
272 the oxalate wash (i.e. biogenic fraction) varied among elements (Tables S2 and S3). In general, the concentrations of Fe
273 and Co in the particles were decreased the least by the oxalate wash by ~25%, while Mo and Pb concentrations were
274 decreased the most by ~70%. The concentrations of particulate Cu, Zn, Cd and Mn were reduced by 50% by the oxalate
275 wash. As shown previously (Sanudo-Wilhelmy et al. 2004), the oxalate reagent also removed extracellular P (by ~20%,
276 Table S2 & S3). Compared to Rauschenberg and Twining (2015), the estimates of the biogenic fraction, after the
277 oxalate wash, were in agreement for Co, Cu and P, and lower for Fe, Mn, Zn and Cd concentrations.
278

279 However, the efficacy of the oxalate wash to dissolve Fe, and other metals, from lithogenic particles is not well
280 constrained (Frew et al. 2006, Rauschenberg and Twining, 2015, King et al., 2012). Therefore, the results obtained
281 after the oxalate-EDTA wash should be interpreted with caution because we do not know whether the removed metal
282 fraction is a) only lithogenic; b) mainly lithogenic but some biogenic fraction is also removed, or c) whether metals
283 absorbed onto particles are equally labile to the wash on biogenic and lithogenic particles. Given that many of the
284 trends we observed were identical for the oxalate-EDTA washed and non-washed particles [i.e. higher Me
285 concentrations in the LC+DFB treatments (Table S2 & S3) and positive correlations between phytoplankton biomass
286 and Me concentrations (Lorenzo-Garrido 2016)], below we present and discuss only the non-oxalate wash results.
287

288 2.4 Statistical analyses

289 Data were checked for normality (by Shapiro-Wilks' test), homoscedasticity (by Levene's test) and sphericity (by
290 Mauchly's test). All data met the requirements to perform parametric tests. Statistical significance of treatment effects
291 was carried out using Split-Plot ANOVA followed by post-hoc Sidak and Bonferroni tests (considering $P < 0.05$ as
292 significant). All analyses were performed using the General Linear Model (GLM) procedure. The correlation between
293 variables was analysed by Pearson's product-moment multiple comparisons (considering $P < 0.05$ as significant).
294 Statistical analyses were carried out using SPSS v22 (IBM statistics) and Sigmaplot 12 (Systat Software, Chicago,
295 USA).

296 **3. Results**

297 **3.1 Biological and chemical characteristics during the bloom**

298 Plankton community dynamics and their response to the applied treatments in the mesocosms are described in detail by

299 Segovia *et al.* (2017). Briefly, at the beginning of the experiment (days 1-10) a bloom of large chain-forming diatoms

300 was observed, which declined by day 7 (Figure S1g- supplemental material). This diatom bloom was associated with a

301 sharp decrease in nitrate and silicic acid concentrations (Figure S2- supplemental material). Picoeukaryotes, dominated

302 the phytoplankton community on day 8 (Figure S1d). During the first 10 days of the experiment, there were no

303 significant differences in the chemical variables measured between the treatments (Figures S3 and S2- supplemental

304 material). On day 7, half of the mesocosms were amended by adding DFB (+DFB treatments). Between day 7 and 17,

305 an increase in dFe was observed in all treatments, except in the control (Figure S3). This increase in dFe was sustained

306 for the entire experiment in the DFB treatments (Figure S3). Dissolved Cu concentrations were not affected by the

307 different treatments (Figure S3). After day 10, a massive bloom of the coccolithophore *E. huxleyi* developed under LC

308 +DFB condition (Figure S1b), out-competing the rest of the plankton groups (Figure S1). This bloom was not observed

309 either in the control treatment (LC-DFB) or in the HC treatments, although *E. huxleyi* was still the most abundant

310 species in all treatments; with the exception of the HC-DFB treatment (Figure S1b).

311

312 **3.2 Particulate metal concentrations during the mesocosm experiment**

313 The pMe concentrations (nM, mean of all treatments and dates) during the experiment were highest for Al, Fe and Zn,

314 and lowest for Cd, following this trend: Al \approx Fe \approx Zn > Ti > Cu \approx Mn > Mo \approx Pb > Co > Cd (Figure 1, Table S2).

315 Significant changes over time were observed for all particulate trace metal concentrations (Fe, Cu, Co, Zn, Cd, Mn, Mo

316 and Pb), except for Ti and Al (Figure 1, Table 1). The only metal that showed a significant time-dependent decrease in

317 its particulate concentration was Fe (Figure 1, Table 1). In general, the treatments with the highest particulate metals

318 concentrations also exhibited the highest particulate P, except for Al, Ti, Fe, and Pb (Figure 1, Table S2). On days 12

319 and 17, the highest particulate metals concentrations were observed in the LC+DFB treatment, while on day 21, they

320 were observed in both LC treatments (Figure 1, Table S2).

321

322 **3.3 The effects of increased CO₂ and the DFB addition on particulate metal concentrations**

323 Increased CO₂ and the DFB addition did not significantly affect the concentrations of particulate Al, Ti, Cu, and Pb

324 (Tables 1 and S2). Similarly, the addition of DFB did not directly influence particulate concentrations of Fe, but high

325 CO₂ had a significant negative impact on particulate Fe (Tables 1 and S2, Figure 1). Particulate Cd concentrations were

326 also inversely affected by CO₂, but only in the presence of DFB (CO₂; and CO₂ x DFB effect, Tables 1 and S2, Figure

327 1). All other elements (P, Co, Zn, Mn and Mo) exhibited significant effects by CO₂ and by DFB, but there was also a

328 significant interaction between these two factors (Table 1, S2). This indicates that, for example, particulate Mn, Zn, Mo,

329 Co, and P concentrations were significantly decreased by high CO₂, but only in the +DFB treatments (Figure 1, Table 1,

330 S2). Similarly, the addition of DFB significantly increased pZn and pMn, but only at ambient CO₂ levels (Figure 1,

331 Tables 1, S2).

332

333 **3.4 Phosphorous-normalized metal ratios in particles collected from the mesocosms and the effects of increased**

334 **CO₂ and the DFB addition on these ratios**

335 The P-normalized metal ratios (Figure 2 and means in Table 2) were highest for Al and Fe (mean: 70 \pm 38 mmol Al:

336 mol P, and 39 \pm 34 mmol Fe: mol P), and lowest for Cd and Co (mean 0.02 \pm 0.01 mmol Cd: mol P, and 0.07 \pm 0.02

337 mmol Co: mol P). Fe:P and Ti:P were not significantly affected by increased CO₂ and/or the DFB addition, but showed

338 a significant decrease over time (Table 3). The P-normalized Cu, Co and Zn ratios changed significantly over time

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(Table 3). Increased CO₂ significantly decreased Co, Zn and Mn:P ratios, while it increased Cu:P ratios (Figure 2, Table 3). DFB did not affect the Me:P ratios of any of these bioactive elements (Table 3).

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4. Discussion

4.1 The effects of CO₂ and dFe on the plankton community

In this experiment we investigated changes in particulate trace metal concentrations, in response to increased CO₂ and/or an addition of the siderophore DFB in a coastal mesocosm experiment. For a better understanding of the processes affecting these stressors, we briefly summarise the mesocosm experiment results originating from Segovia et al. (2017). High CO₂, as well as the DFB addition increased dFe concentration. The higher dFe concentrations were sustained in the DFB treatments. A bloom of the coccolithophore *Emiliania huxleyi* was observed in the ambient CO₂ treatments, and was especially massive in the one with the addition of DFB (LC+DFB). Indeed, the biomass of *E. huxleyi* was negatively affected by increased CO₂. However, increased dFe partially mitigated the negative effect of elevated CO₂, indicating that the coccolithophore was able to acclimate better to ocean acidification when Fe availability was high. High dFe also had a positive effect on the cyanobacterium *Synechococcus sp.*, while the rest of the plankton food web did not response to the treatments (Segovia et al. 2017).

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4.2 Particulate Fe and Ti are associated with lithogenic sources, while particulate Co, Cu, Zn, Cd, Mo and Mn are associated with biogenic sources

The particulate trace metal concentrations (nM, mean of all treatments and dates) during the experiment were highest for Al, Fe and Zn, and lowest for Co and Cd, following this trend: Al ≈ Fe ≈ Zn > Ti > Cu ≈ Mn > Mo ≈ Pb > Co > Cd. Lithogenic particles are enriched in Al and low in P (average crustal Al and P content is 2.9 mmol Al and 0.034 mmol P g⁻¹ dry weight, Taylor 1964), while biogenic particles are enriched in P and low in Al (average plankton Al and P content is 0.001 mmol Al and 0.26 mmol P g⁻¹ dry weight, Bruland et al. 1991). Therefore, the distinct high abundance of Al and P in lithogenic and biogenic particles, respectively, can be used to evaluate the relative contribution of lithogenic and biogenic material in our particulate samples. In order to do this, first, it is important to establish that the vast majority of the measured particulate P is associated the biogenic fraction. In this study, the abiotic P was estimated using particulate Al concentrations (nM) and the P:Al ratio in crustal material, and was calculated to be negligible (< 1% of the total measured particulate P). In addition, a significant correlation (p < 0.003) was found between particulate P concentrations and phytoplankton biomass (Table 4). Therefore, we assume a constant trace metal composition in biogenic particles (assuming they are rich in phytoplankton) and lithogenic particles (assuming they are rich in crustal material). We then calculated the expected metal concentrations in the particulate samples assuming that all the P measured in the particles is associated with a biogenic fraction, and that all Al in the particles is associated with the lithogenic fraction. Thus, for a given trace metal, its expected particulate trace metal concentration in seawater (mol L⁻¹) can be calculated as the sum of the contribution from biogenic and lithogenic particles, so that:

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$$[Me] = a [P] + b [Al]$$

where [Me] is the total concentration of the metal (mol L⁻¹) expected in the particulate sample; [P] is the P concentration measured in the particles (mol L⁻¹); [Al] is the Al concentration measured in the particles (nM L⁻¹); *a* is the average, well-known metal content in biogenic particles, normalized to P (i.e. mol Me: mol P in marine plankton; Ho 2006) and *b* is the average, well known metal content in lithogenic particles, normalized to Al (mol Me: mol Al in the Earth crust; Taylor 1964). For example, on day 21 in the HC-DFB treatment, the concentrations of particulate Al and P were 8.22 and 134.8 nM, respectively (Table S2). Assuming a constant 0.0051 mol Fe: mol P in biogenic

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particles (Ho 2006) and 0.331 mol Fe: mol Al in lithogenic particles (Taylor 1964; Table 2), we calculated an expected particulate Fe concentration of 3.41 nM, where 80% was associated with lithogenic material and 20% with biogenic material. Similar calculations were made for the bioactive metals Mn, Co, Cu, Zn, Cd, and Mo (Table 2). Our calculations indicate that on average, particulate Fe was dominated by the lithogenic component (accounting for an average of 78% of the total expected particulate Fe), while for particulate Co, Cu, Zn, Cd, and Mo the biogenic fraction dominated (accounting for 94, 95, 99, 94 and 98%, respectively, of the total expected concentration; Table 2). Particulate concentrations of Mn were also dominated by the biogenic fraction (65% of the total), but the lithogenic fraction was also significant (35%). Moreover, the expected particulate Mn and Fe concentrations closely matched the particulate Mn and Fe concentration we measured (accounting for an average of ~ 71% of the measured Mn, and 115% of the measured Fe). For other metals (i.e. Cu, Mo and Zn), the expected particulate concentrations (nM) were lower than measured (23% of the measured pCu, and 8% of measured pZn; Table 2). This suggests that the particles were enriched in Cu, Mo, and Zn relative to what is expected based on natural marine plankton metal quotas (Bruland et al. 1991) and crustal ratios (Taylor 1964).

To further establish the lithogenic or biogenic source of the pMe in the particles, the particulate metal concentrations were normalized to the concentrations of particulate P and Al (Figure 2 and Table 2). These ratios were then compared with well-known molar ratios of metal to Al in the crust (Taylor 1964) and of metal to P ratios in marine plankton samples (Ho 2006) and cultures (Ho et al. 2003) (Table 2). The average Fe: Al (506 mmol Fe: mol Al) and Ti:Al ratios (119 mmol Ti: mol Al, Table 2) were relatively similar to crustal molar ratios (331 mmol Fe: mol Al and 39 mmol Ti: mol Al; Taylor 1964). Additional evidence for the significant lithogenic component in particulate Ti and Fe was gathered from Figure 3, where we plotted the molar ratios of the metals relative to P in the collected particles against the Al:P ratios measured in those same particles. The slope of these data $[(\text{Fe:P})/(\text{Al:P}) = \text{mol Me: mol Al}]$ is the ratio of Me:Al in the particles and can be compared to well-known Me:Al crustal ratio. Visually, if the data nicely fit the Me:Al line for crustal material, these metals are mainly associated with the lithogenic component, as evident for Fe and Ti (Figure 3). These combined results suggest that in our experiment, particulate Fe and Ti concentrations were enriched by lithogenic material. In support of this finding, we also found no significant correlation between particulate Fe and Ti concentrations (nM) and either the total plankton (phytoplankton and microzooplankton) or *E. huxleyi* biomass ($\mu\text{g C L}^{-1}$; Table 4).

In contrast, when the P-normalized metal ratios in the particles collected from the mesocosms were plotted against the Al:P ratios in these particles, there were no correlations for the following metals Co, Cu, Zn, Cd, Mn and Mo (Figure 4), indicating that these particulate metals were not enriched in lithogenic material. Our measured metal: P ratios were comparable to plankton ratios in natural samples and in cultures (Table 2). The concentrations (mol L^{-1}) of these metals (i.e. Cu, Co, Zn, Cd, Mn, Mo), as well as P, also showed significant correlations with the biomass ($\mu\text{g C L}^{-1}$) of *E. huxleyi* and that of total plankton cells ($p < 0.05$, Table 4), supporting a significant influence of the phytoplankton in the distribution of these particulate elements.

4.3 Particulate metals with a strong biogenic component: their P-normalized ratios

The concentrations of particulate bioactive metals (mol L^{-1}), with a significant biogenic component (i.e. excluding Fe) in the studied *E. huxleyi* bloom were ranked as: $\text{Zn} > \text{Cu} \approx \text{Mn} > \text{Mo} > \text{Co} > \text{Cd}$ (Figure 1, Table S3), similar to those reported in indigenous phytoplankton populations: $\text{Fe} \approx \text{Zn} > \text{Cu} \approx \text{Mn} \gg \text{Co} \approx \text{Cd}$, (Twining and Baines, 2013). The only treatment where *E. huxleyi* did not dominate the community was the HC-DFB; in this treatment the ranking of these biogenic particulate trace metals was the same as that of LC+DFB (with the massive *E. huxleyi* bloom), but their

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concentrations were lower than those in LC+DFB. At the end of the experiment, the concentrations of these biogenic metals were, in general, comparable in both HC treatments, and lower than those in the LC treatments (Figure 1, Table S3). Therefore, high CO₂ had a tendency to decrease particulate metal concentrations, especially on day 21. Given the strong correlation between concentrations of these particulate bioactive metals and phytoplankton biomass, the lower particulate concentrations in high CO₂ were mainly due to low phytoplankton biomass.

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Particulate Zn concentrations were especially high in the LC+DFB treatment (Figure 1), where the highest *E. huxleyi* biomass was observed. *Emiliania huxleyi* is well known for its high Zn cellular requirements (~ 1-10 for *E. huxleyi* vs. 1-4 mmol Zn: mol P for other phytoplankton; Sunda and Hunstman 1995, Sunda 2013). But, the Zn: P ratios in the LC+DFB treatment (range 45-69 mmol Zn: mol P; Figure 2, Table S2), as well as in all the other treatment (range 16-34 mmol Zn: mol P; Figure 2, Table S2) were significantly higher than these published ratios. This could be explained by, the adsorption of these metals to the outside of the cells, and/or anthropogenic inputs of Zn into the fjord. The Zn:P ratios in the samples washed with the oxalate-EDTA were still high (range 28-57 for LC+DFB and 16-33 mmol Zn: mol P in all other treatments, Table S3), thus adsorption might have not been significant. We hypothesize that anthropogenic aerosols which are rich in anthropogenic particulate metals, such as Zn and Cu (Perry et al. 1999; Narita et al. 1999), and have high percentage of Zn and Cu dissolution (ref.), might be the source of these high Zn concentrations and ratios in the particles.

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Similarly, the Cu:P ratios in the collected particles were relatively elevated (1.4 ± 0.8 mmol Cu: mol P) compared to those of other phytoplankton, including *E. huxleyi* (Table 2). The dissolved (7.7 ± 0.41 nM Cu, Figure S3) and particulate Cu concentrations (0.35 ± 0.25 nM, Table S2) in our experiment were high, and similar to those previously measured in this fjord (Muller et al., 2005). Rain events (or wet deposition of anthropogenic aerosols) in this fjord result in high dissolved Cu and the active production of strong organic ligands by *Synechococcus*—to lower the free Cu concentrations (Muller et al., 2005). Therefore, high Cu might be a general condition in this fjord due to the rainy nature of the geographical location, and indigenous plankton might have developed physiological mechanisms to deal with high Cu, such as the production of organic ligands to prevent uptake (Vraspir and Butler, 2009), or of heavy-metal-binding peptides (phytochelatins) to lower Cu toxicity inside the cell (Ahner and Morel, 1995; Ahner et al., 1995; Knauer et al., 1998). Since we measured high particulate Cu, and Cu:P in our experiment, *E. huxleyi* might have been relying mainly on phytochelatins to buffer high intracellular Cu (Ahner et al., 2002).

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The Cd:P ratios (average 0.024 ± 0.01 mmol Cd:mol P, Figure 2) were significantly lower than those in phytoplankton and *E. huxleyi* (0.36 mmol Cd:mol P, Figure 2). This was surprising, because Cd quotas are normally higher in coccolithophores than in diatoms and chlorophytes (Sunda and Huntsman, 2000; Ho et al., 2003). High Cd quotas in coccolithophores have been suggested to result from accidental uptake through Ca transporters and channels (Ho et al., 2009). The low Cd quotas here may be explained by the antagonistic interaction between Mn and Cd or Zn and Cd under high Mn and Zn, respectively (Sunda and Huntsman, 1998, 2000; Cullen and Sherrell, 2005). Since high Zn:P ratios were common in this study (34.02 ± 18.05 mmol Zn:mol P, Figure 2), we hypothesize that high Zn levels antagonistically interacted with Cd, resulting in low Cd:P ratios in the particles.

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4.4 The effects of increased CO₂ and the DFB addition on particulate metal concentrations and P-normalized ratios

Fe enrichment is common in coastal waters, due to sediment resuspension, rivers input, aeolian deposition and mixing or upwelling of deep water. Indeed, Fe was the essential metal with the highest particulate concentrations in our study

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(Figure 1, Table 3). Furthermore, in this study particulate Fe was characterized by a strong lithogenic component, and was not correlated with phytoplankton biomass. Fe was also unique, in that it was the only trace element whose particulate concentration was significantly and exclusively affected by CO₂ (no interaction between CO₂ and DFB), regardless of the presence or absence of DFB (Table 1). Furthermore, particulate Fe concentrations (nM) decreased steadily between days 12 and 21, with the exception of the control treatment (LC-DFB; Figure 1, Table 2S). This suggests that the increase in CO₂ and/or the DFB addition reduce the concentration of pFe, despite the phytoplankton bloom. Such a decrease in pFe (range 2.3-fold in LC-DFB, vs. 13.7-fold in HC+DFB; Table S2) might be mediated by the dissolution of particulate Fe by low pH or by the presence of strong organic chelators as observed in this very experiment (Segovia et al. 2017 and references therein), where dFe notably increased in treatments with high CO₂ and/or the addition of DFB (Figure S3). Furthermore, the dissolution of particulate Fe in the treatments with high CO₂ and/or the addition of DFB was evident in the Fe partitioning coefficients—the molar ratio between particulate and dissolved concentrations (Figure 4). On day 21, the Fe partitioning coefficients varied by 22-fold between the highest for the control (LC-DFB: 1.039) and lowest for the HC+DFB treatments (HC+DFB: 0.047; Figure 4). Thus, either the DFB addition or high CO₂ promoted the dissolution of pFe. However, at the end of the experiment, high dFe concentrations were only observed in the treatments with the DFB additions, suggesting that the presence of strong organic Fe chelators, such as DFB, mediated the maintenance of high dissolved Fe concentrations, as previously observed (Segovia et al. 2017). Thus, in our future oceans, high CO₂ (low pH) will increase dissolved Fe concentrations in regions rich in particulate Fe, and in strong organic Fe chelators. The deleterious effects of OA on the development of ecologically important species sensitive to increased CO₂ such as *E. Huxleyi*, will be more relevant in high-Fe environments than in Fe-limited ones.

In contrast to the findings for Fe, particulate Cu concentrations a) were not affected by either high CO₂ or the DFB addition; b) were dominated by a biogenic component and c) were significantly correlated with phytoplankton biomass (Table 4). Furthermore, unique to Cu was a significant increase in Cu:P ratios by day 21 in the high CO₂ treatments, especially when no DFB was added. Since the Cu partitioning coefficients only varied by 3.25 fold among treatments on day 21 (LC-DFB: 0.065 vs. HC+DFB: 0.047; data not shown), we hypothesize that high CO₂ did not affect the partitioning between particulate and dissolved Cu, but instead, it affected the speciation of dissolved Cu, increasing free Cu (Cu²⁺) and thus, its bioavailability. This resulted in the highest Cu:P ratios in the high CO₂ treatments, despite the low phytoplankton biomass. This increase in bioavailability under lower pH is typical of metals that form strong inorganic complexes with carbonates, such as Cu²⁺ (Millero et al., 2009). Thus in our future oceans, high CO₂ (low pH) will shift the speciation of dissolved Cu towards higher abundance of free ionic species, increasing its bioavailability and likely its toxicity.

Similarly to Cu, particulate Co, Zn and Mn were correlated with biomass and were dominated by the biogenic component. But in contrast to Cu, these metals particulate concentrations were affect by increased CO₂ and/or the DFB addition. However, the effects of high CO₂ and/or DFB were very complex because significant interactions between these 2 factors were observed (Table 1); and further studies are required before we are able to discern and conclude a significant trend. Yet, the P-normalized ratios of Co, Zn and Mn were significantly affected by CO₂ (Table 3), exhibiting moderately lower ratios under high CO₂, when phytoplankton biomass was lowest. These results imply that the bioavailability of these metals was not enhanced under acidic conditions. This suggests that under high CO₂ (low pH) the free ionic species of these metals will not significantly increase in the future, as shown for metals that occur predominantly as free ionic species in seawater (Millero et al., 2009).

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574 5. Concluding remarks

575 The results presented here show that in the fjord where we carried out the present experiment, particulate Fe was
 576 dominated by lithogenic material, and was significantly decreased in the treatments with high CO₂ concentrations
 577 and/or the DFB addition. Indeed, high CO₂ and/or DFB promoted the dissolution of particulate Fe, and the presence of
 578 this strong organic complex helped maintaining high dissolved Fe. This shift between particulate and dissolved Fe, in
 579 the presence of DFB, promoted a massive bloom of *E. huxleyi* in the treatments with ambient CO₂ due to increased
 580 dissolved Fe. During the bloom of *E. huxleyi*, the concentrations of particulate metals (mol L⁻¹) with a strong biogenic
 581 component (Cu, Co, Zn, Cd, Mn, and Mo) were a) highly dynamic, b) positively correlated with plankton biomass, and
 582 c) influenced by growth requirements. Furthermore, high CO₂ decreased the Me:P ratios of Co, Zn and Mn, while
 583 increased the Cu:P ratios. In contrast DFB had no effects on these ratios. According to our results, high CO₂ may
 584 decrease particulate Fe and increase dissolved Fe, but high concentrations of dissolved Fe will only be maintained by
 585 the presence of strong organic ligands. The decrease in particulate Fe may affect the sinking flux of other metals
 586 associated with Fe oxides in open ocean settings. Furthermore, ocean acidification will decrease *E. huxleyi* abundance,
 587 and as a result, the sinking of particulate metals enriched in *E. huxleyi*. Moreover, the Me:P ratios of metals that are
 588 predominately present in an ionic free form in seawater (e.g. Co, Zn and Mn) will likely decrease or stay constant. In
 589 contrast, high CO₂ is predicated to shift the speciation of dissolved metals associated with carbonates, such as Cu,
 590 increasing their bioavailability, and resulting in higher Me:P ratios. We suggest that high Cu might be a common
 591 condition in this fjord, and autochthonous plankton might be able to cope with high Cu levels by developing specific
 592 physiological mechanisms. Future high CO₂ levels are expected to change the relative concentrations of particulate and
 593 dissolved metals, due to the differential effects of high CO₂ on trace metal solubility, speciation, adsorption and
 594 toxicity, as well as on the growth of different phytoplankton taxa, and their elemental trace metal composition.

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 602 logistic support during the experiment.

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Table 1. Statistical analyses (Split-plot ANOVA) of the effects of high CO₂, the addition of DFB, and their interaction, as well as the effect of time, on the concentrations of particulate metals (mmol L⁻¹, data in Table S2, and Figure 3) in particles collected from the different mesocosms treatments.

Factor	Al	Ti	P	Fe	Cu	Co	Zn	Cd	Mn	Mo	Pb
CO ₂	ns	ns	**	*	ns	**	***	***	**	***	ns
DFB	ns	ns	*	ns	ns	*	**	ns	*	*	ns
CO ₂ x DFB	ns	*	**	ns	ns	*	**	*	**	**	ns
Time	ns	ns	ns	***	*	***	***	***	***	***	**

*ns: not significant; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$*

Table 2. The average metal ratios in the particles collected in this study (without oxalate wash) using the data reported in Table S2. The P-normalized ratios (mmol : mol P, Figure 4) are compared to previous estimates in marine plankton samples and phytoplankton cultures (A). The Al-normalized ratios (mmol : mol Al) are compared to crustal ratios (B).

A)

(mmol : mol P)	Mn:P	Fe:P	Co:P	Cu:P	Zn:P	Cd:P	Mo:P	Al:P	Reference
Phytoplankton _{Lab}	3.8	7.5	0.19	0.38	0.8	0.21	0.03		Ho et al. 2003
Marine Plankton _{Field}	0.68±0.54	5.1±1.6	0.15±0.06	0.41±0.16	2.1±0.88				Ho 2006
<i>E. huxleyi</i> _{Lab}	7.1±0.36	3.5±0.07	0.29±0.02	0.07±0.013	0.38±0.002	0.36±0.01	0.022±0.0003		Ho et al. 2003
This study	1.65±0.41	39.2±34.3	0.07±0.02	1.41±0.55	34.02±18.05	0.02±0.01	0.42±0.12	70±38	
Crust ratio	510	29,738	13	25	32	0.05	0.46	89,972	Taylor 1964

B)

(mmol : mol Al)	Mn:Al	Fe:Al	Co:Al	Cu:Al	Zn:Al	Cd:Al	Mo:Al	Pb:Al	Ti:Al
Crustal ratio	5.7	331	0.14	0.27	0.35	0.001	0.005	0.02	39
This study	35±28	506±342	1.5±1.2	26.5±15	795±865	0.5±0.4	8.6±6.5	4.9±3.9	119±47.6

Table 3. Statistical analyses (Split-plot ANOVA) of the effects of CO₂, DFB, and their interaction, as well as the effect of time, on the P-normalized metal quotas (mmol: mol P, data in Figure 4, and Table S2) in particles collected from the different mesocosm treatments.

Factor	Fe:P	Cu:P	Co:P	Zn:P	Cd:P	Mn:P	Mo:P	Pb:P	Ti:P
CO ₂	ns	*	***	**	ns	*	ns	ns	ns
DFB	ns	ns	ns	ns	ns	ns	ns	ns	ns
CO ₂ x DFB	ns	ns	ns	ns	ns	ns	ns	ns	ns
Time	***	***	***	***	ns	ns	ns	ns	***

*ns: not significant; * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$*

Table 4. The relationship (Pearson correlations, $p < 0.05$) between particulate metals concentrations (nmol L⁻¹, no oxalate wash, reported in Table S2) and the biomass (µgC L⁻¹) of *Emiliania huxleyi* and total cells (phytoplankton and microzooplankton) collected from the different mesocosm treatments.

		P	Fe	Cu	Co	Zn	Cd	Mn	Mo	Pb	Ti
<i>E. huxleyi</i>	Correlation coefficient	0.622	ns	0.614	0.756	0.747	0.818	0.686	0.825	ns	ns
	P-value	0.003		0.003	$7.35 \cdot 10^{-5}$	$1.01 \cdot 10^{-4}$	$6.02 \cdot 10^{-6}$	$5.93 \cdot 10^{-4}$	$4.20 \cdot 10^{-6}$		
Total cells	Correlation coefficient	0.641	ns	0.51	0.644	0.889	0.802	0.598	0.53	ns	ns
	P-value	0.002		0.02	$1.62 \cdot 10^{-3}$	$7.03 \cdot 10^{-8}$	$1.23 \cdot 10^{-5}$	$4.18 \cdot 10^{-3}$	$1.35 \cdot 10^{-2}$		

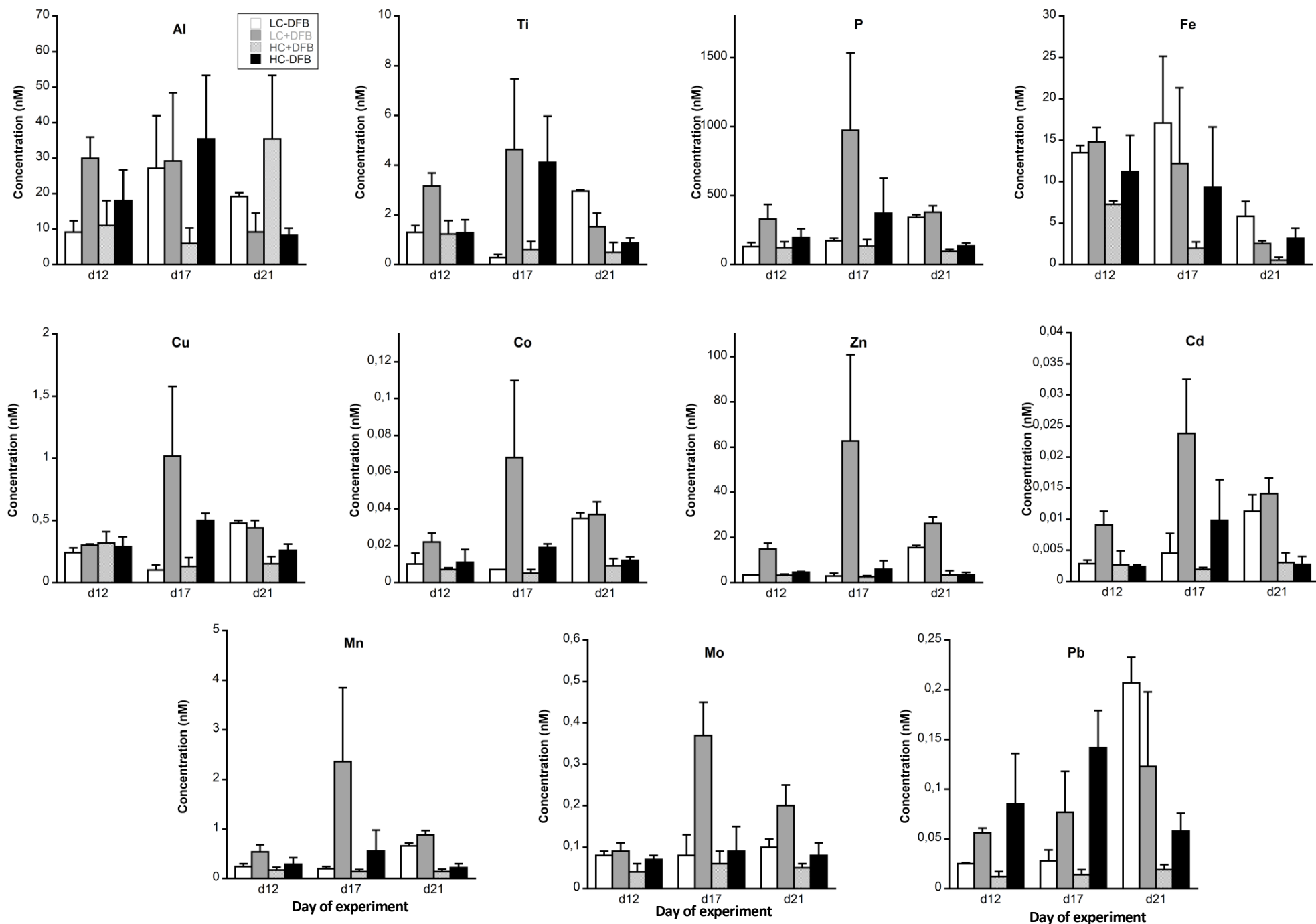


Fig. 1. The concentration of particulate metals in seawater (nM) in the different treatments; LC: ambient CO₂ (390 μatm); HC: increased CO₂ (900 μatm); -DFB (ambient dFe); +DFB (increased dFe) during the development of a bloom of *Emiliania huxleyi*. Bars are means of measurements in 3 independent mesocosms (n = 3) except for LC–DFB where n = 2. Error bars indicate SD.

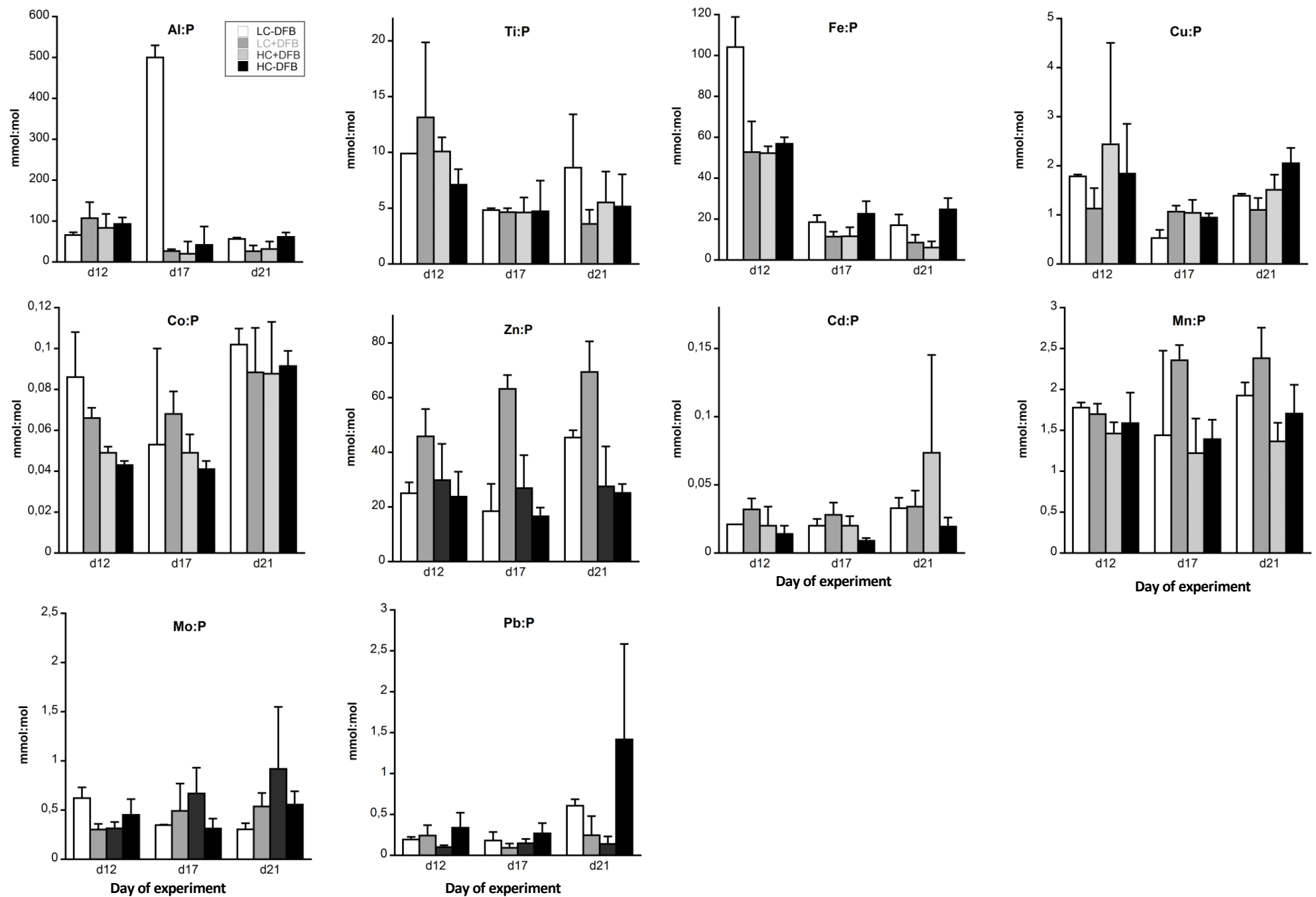


Fig. 2. P-normalized metal quotas (mmol:mol P) of particles from different treatments; LC: ambient CO₂ (390 μ atm); HC: increased CO₂ (900 μ atm); -DFB (ambient dFe); +DFB (increased dFe) during the development of a bloom of *Emiliana huxleyi*. Bars are means of measurements in 3 independent mesocosms (n = 3) except for LC-DFB where n = 2. Error bars indicate SD.

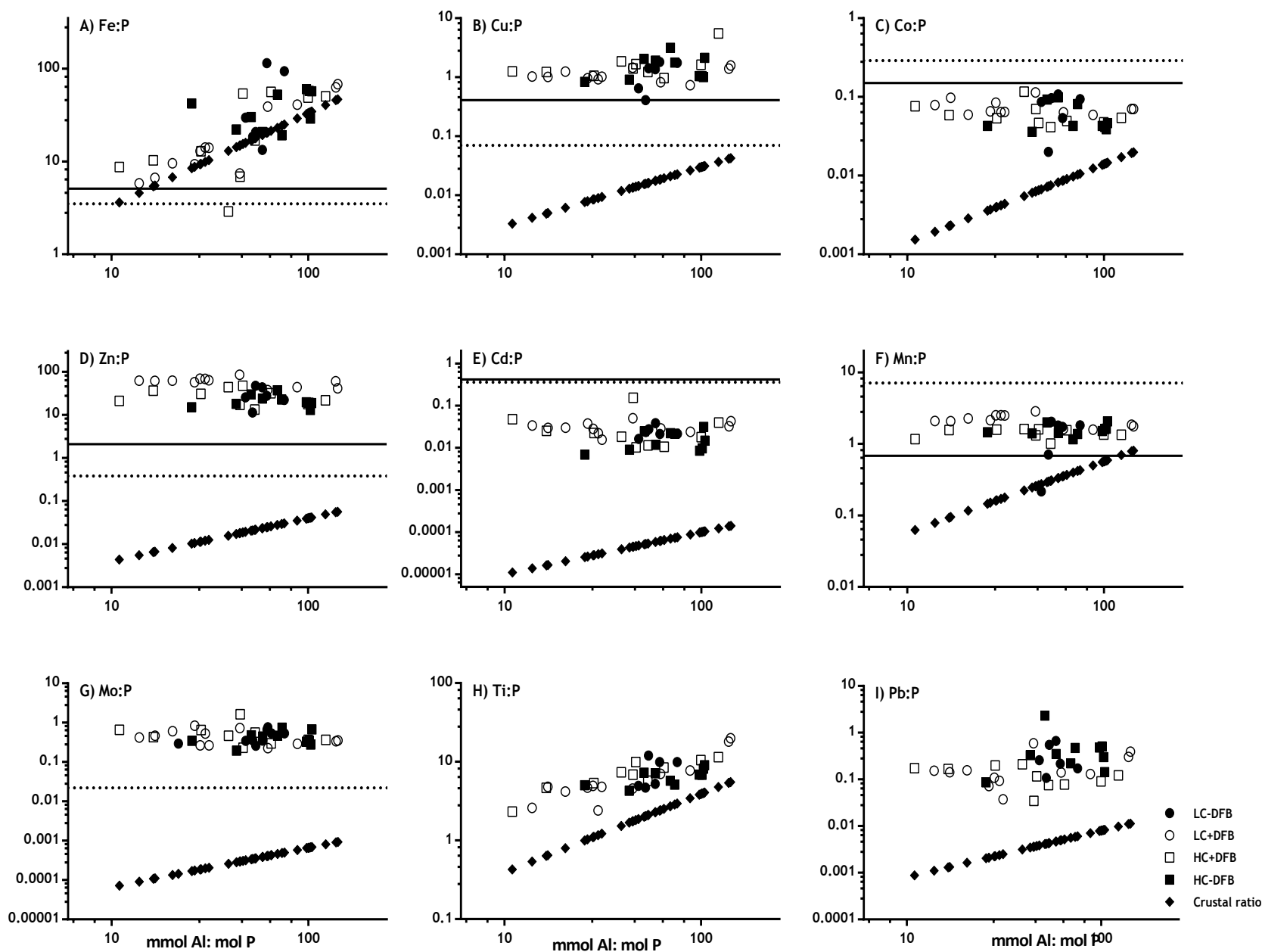


Fig. 3. Comparison of P-normalized metal ratios in particles (mmol:mol P) against mmol Al:mol P ratios in the same particles (without oxalate wash) collected from the different mesocosm treatments (LC: ambient CO₂; HC: increased CO₂ (900 μ atm); -DFB: no DFB addition; +DFB: with a 70 nM DFB addition) during the development of a bloom on day 12, 17 and 21 (original data reported in Table S2). The x-axis parallel solid and dotted lines represent the average metal quotas obtained from marine plankton assemblages (Ho 2006) and from cultures of *Emiliania huxleyi* (Ho et al. 2003). The slope of the line with the \blacklozenge symbols indicates the average metal : Al (mol:mol) in crustal material (Taylor, 1964). (A) Fe:P, (B) Cu:P, (C) Co:P, (D) Zn:P, (E) Cd:P, (F) Mn:P, (G) Mo:P, (H) Ti:P, (I) Pb:P.

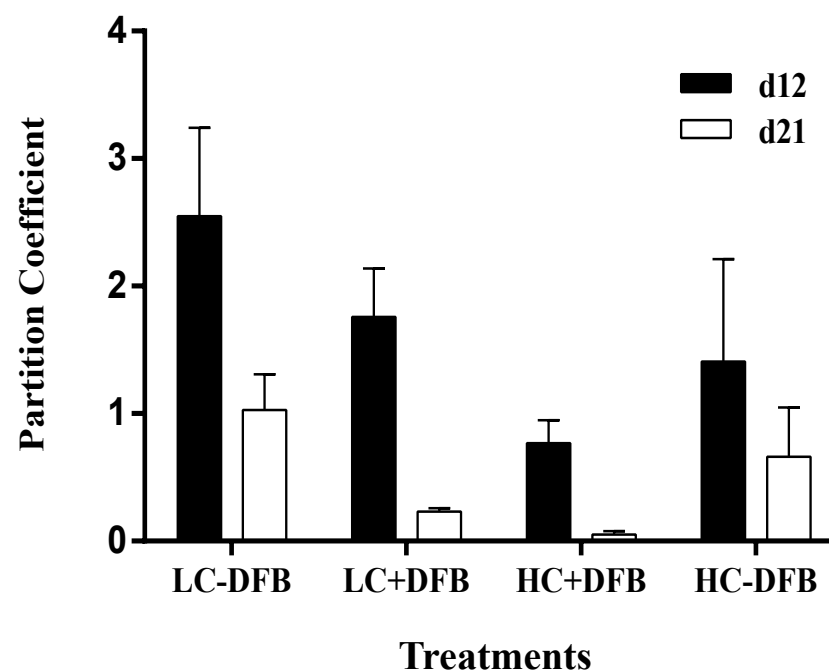


Fig. 4. The Fe partition coefficients (the molar ratio between particulate and dissolved concentrations) in the different mesocosm treatments; LC: ambient CO₂ (390 μ atm); HC: increased CO₂ (900 μ atm); -DFB: no DFB addition; +DFB: with a 70 nM DFB addition; on day 12 and day 21. . Bars are means of measurements in 3 independent mesocosms (n = 3) except for LC-DFB where n = 2. Error bars indicate SD.